



*Supplement of*

## **Boron incorporation in the foraminifer *Amphistegina lessonii* under a de-coupled carbonate chemistry**

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## SUPPLEMENT

## MATERIAL AND METHODS

## 1.1 Determination of the B isotopic composition of the culture media

5 A detailed description of the method used for the analysis of the  $\delta^{11}\text{B}$  of the culture media is  
6 given in Misra et al. (2014). All samples and standards were analyzed in 0.3M HF matrix to facilitate  
7 rapid boron wash out. We used Savillex® self-aspirating C-flow nebulizers with sample uptake rate of  
8 50 $\mu\text{l}/\text{min}$  for sample aspiration. High performance extraction cones (Jet sampler and ‘X’ skimmer)  
9 were used to boost boron sensitivity. The instrumental plasma conditions and mass calibration were  
10 appropriately adjusted to eliminate any artificial mass bias caused by  $^{20}\text{Ne}^{2+}$  and  $^{40}\text{Ar}^{4+}$  peak tailing  
11 into the  $^{10}\text{B}^+$  peak. Before each analytical session the instrument was optimized for maximum boron  
12 sensitivity (1 M-cps / ppb on  $^{11}\text{B}$ ) and samples were analyzed at a signal to blank intensity ratio of  $> 500$ . An optimal analyses time of  $\sim 12$  minutes per sample was adopted to obtain  $\geq 2500$   $^{11}\text{B}/^{10}\text{B}$  ratios  
13 to achieve the desired external precision of  $\leq 0.5\%$  per quintuplicate analysis ( $2\sigma/\sqrt{n}$ ). Based on  
14 sample aspiration rate and analysis time at total of  $\sim 3$  ng B was consumed per analysis. Instrumental  
15 settings are given in table S2.

17 Prior to mass spectrometric analysis boron was separated from the sample matrix by a single  
18 step micro-distillation method, modified after (Gaillardet et al., 2001) and (Wang et al., 2010) as  
19 described in Misra et al. (2014). Savillex® Teflon® fin legged 5 ml beakers with conical interior were  
20 used as the distillation reservoir. Samples were loaded on to the cap of a pre-cleaned beaker – the  
21 beaker was tightly closed to avoid boron loss – set on a hotplate at 95°C with the conical end pointing  
22 up. The distillation process was carried out for 15 to 18 hours to achieve a quantitative distillation of  
23 boron from the load into the distillate. Sample load volume was kept below 50 µl to avoid the  
24 accumulated droplet at the conical end from dropping back onto the cap. Post distillation, beakers  
25 were taken off the hotplate, allowed to cool for ~ 15 minutes, then 0.5 ml of 0.3 M HF was added and  
26 the beakers were capped with pre-cleaned caps.

27

28      **1.2 Boron analysis of the tests**

29              Detailed information about the simultaneous determination of B/Ca and  $\delta^{11}\text{B}$  of foraminifer  
30              using LA-MC-ICP-MS and OES are given in Kaczmarek et al. (2015).

31      *1.2.1 Laser Ablation*

32              The in-house built laser ablation system (Solstice Newport/Spectra Physics) is based on a 100  
33              femtosecond Ti-sapphire regenerative amplifier system operating at a fundamental wavelength of 777  
34              nm in the infrared spectrum. Subsequent harmonic generations produce the wavelengths 389 nm in the  
35              second, 259 nm in the third and 194 nm in the fourth harmonic. The pulse energies measured with a  
36              pyroelectric sensor (Molelectron, USA) are 3.2mJ/pulse at 777nm, 0.7 mJ/pulse at 259 nm, and 0.085  
37              mJ/pulse at 194 nm. After the fourth harmonic generation stage the 194 nm beam is streered by eight  
38              dichronic mirrors into a 8x objective (NewWave-Research, USA) and focussed onto the sample. Spot  
39              size was set to 50  $\mu\text{m}$  for standard and samples. Within this spot an energy density of 2  $\text{J}/\text{cm}^2$  is  
40              maintained.

41              Table S3 shows different laser efficiencies applied to foraminifers and NISTSRM 610 from  
42              different treatments. Laser ablation on NISTSRM 610 was performed in raster mode setting a  $\sim 150$   
43               $\times 150\mu\text{m}$  raster with a spot size of 50  $\mu\text{m}$ . For foraminifers the raster mode was applied, too. However,  
44              the size of the raster varied dependent upon the individual size of each foraminifer. Laser ablation of  
45              foraminifers was always performed on the so-called knob, a massive calcite without pores, located in  
46              the middle on the spiral side of the foraminifer. Figure S1 shows an example of a foraminifer from the  
47               $\text{pH\_8.1}^{160}$  treatment after laser ablation.

48      *1.2.2 Isotope Analysis - Acquisition parameters*

49              All measurements were carried out in low mass resolution ( $\Delta\text{m}/\text{m}=350$  where  $\text{m}$  is the mass of  
50              the ion of interest and  $\Delta\text{m}$  is the mass difference between its 5 and 95% peak height). Compact  
51              discrete dynode multipliers (CDD, Thermo) were attached to faraday cups at the low site on L4 and  
52              the high site on H4. The low resolution mode is sufficient enough to resolve potential interferences  
53              from doubly charged ions due to the intrinsic high resolution in the low mass region. Possible

54 interferences are the clusters of  $^{40}\text{Ar}^{4+}$  or  $^{20}\text{Ne}^{2+}$  which are well resolved to the background level.  
 55 Working with ion counters it is necessary to determine the detector dead time especially for isotope  
 56 ratios with large isotope abundance differences. The dead time corrections have been performed by  
 57 measuring the  $^{238}\text{U}/^{235}\text{U}$  ratio using SRM 981 in a multidynamic measuring sequence. Subsequently,  
 58 the dead time has been checked prior to every analytical session by analysing NISTSRM 610 using  
 59 different repetition rates of laser, resulting in a counting range between 300000 – 1000000 cps. Prior  
 60 to each analytical session the instrument was tuned for optimal peak shape. All measurements were  
 61 performed at plateau voltage of the CDDs which was checked prior to every analytical session. Before  
 62 the beginning of samples analysis measurements of NISTSRM 610 were continued until instrumental  
 63 drift due to warm-up was less than 300 ppm over a bracketing sequence duration of twelve minutes.  
 64 Boron signal intensities of the reference material, NISTSRM 610, and samples were matched within  
 65 10% in signal intensity by adapting the laser repetition rate. For analysis we adopt the standard sample  
 66 bracketing procedure using NISTSRM 610 as reference material containing 351 ppm B and 8.45 %  
 67 Ca. The acquisition parameters in static mode for analysis of NISTSRM 610 and samples were set to  
 68 acquire 200 cycles of 1 s integrations each. During the first 40 cycles the background signal was  
 69 acquired whereas the remaining cycles represent the sum of the background + reference material, or  
 70 background + sample signals. The B isotopic composition is reported using the delta notation:

$$71 \quad \delta^{11}\text{B}_{\text{sample}} (\text{\%}) = \left[ \frac{\left( \frac{^{11/10}\text{B}}{^{11/10}\text{B}} \right)_{\text{sample}}}{\left( \frac{^{11/10}\text{B}}{^{11/10}\text{B}} \right)_{\text{NIST610}}} - 1 \right] \times 1000 \quad (S2)$$

72 The  $^{11/10}\text{B}$  ratio of NIST 610 represents a mean value calculated from the NIST 610 measurements  
 73 performed before and after the sample. The errors of  $\delta^{11}\text{B}$  of the foraminifers were calculated by  
 74 propagating the  $\text{SE}/^{11/10}\text{B}$  for NIST610 and samples respectively according to the standard bracketing:

$$2\text{RSE}_{\delta^{11}\text{B}} (\text{\%}) = \sqrt{\left( \frac{\text{SE}}{\frac{^{11/10}\text{B}}{^{11/10}\text{B}}} \right)_{\text{NIST-1}}^2 + \left( \frac{\text{SE}}{\frac{^{11/10}\text{B}}{^{11/10}\text{B}}} \right)_{\text{sample}}^2 + \left( \frac{\text{SE}}{\frac{^{11/10}\text{B}}{^{11/10}\text{B}}} \right)_{\text{NIST+1}}^2} \times 1000 \times 2 \quad (S3)$$

75 Where NIST-1 and NIST+1 refer to NIST610 measurements performed before and after the sample. It  
 76 should be noted that the 2RSE of foraminifers represents a criterion for the homogeneity of B isotope  
 77 distribution rather than an analytical uncertainty. The analytical uncertainty and external

78 reproducibility is given best by repeated measurements of a homogenous material such as NIST 610.

79 Delta  $^{11}\text{B}$  values of NIST 610 were calculated by:

80 
$$\delta^{11}\text{B} (\text{‰}) = \left[ \frac{\frac{^{11/10}\text{B}_0}{^{11/10}\text{B}_{-1} + ^{11/10}\text{B}_{+1}}}{2} - 1 \right] \times 1000 \quad (\text{S4})$$

81 Where the measurements of the  $(^{11/10}\text{B})_{-1}$  and  $(^{11/10}\text{B})_{+1}$  ratios of NIST 610 were performed before and  
82 after the measurement of  $(^{11/10}\text{B})_0$ , respectively. For the determination of the analytical uncertainty and  
83 external reproducibility all measurements of NIST 610 performed between each sample measurement  
84 in this study were taken into account and are on average  $0.4 \pm 0.5$  (2sd) ‰.

85 As shown by Fietzke et al. (2010) no matrix dependent offsets between silicate and carbonate  
86 matrices exist suggesting that silicate glass standards, as NIST 610, may serve as adequate reference  
87 material. Therefore, we consider the analytical uncertainty of NIST 610 as an adequate external error  
88 of the foraminiferal samples. Operating conditions for La-MC-ICP-MC are given in table S4.

89 *1.2.2 Determination of B/Ca of the test by Optical Emission Analysis*

90 Ocean Optics Maya2000 Pro is a high-sensitivity fiber optical spectrometer. It exhibits a  
91 measuring range of 250 to 460 nm with a resolution of 0.11 nm covering the first order emission lines  
92 of Mg II, Ca II, Sr II Ba II and Li II. It is equipped with a back-thinned 2D FFT-CCD detector, and a  
93 grating with a groove density of 1200 lines/mm. More technical information about Maya2000 Pro can  
94 be found on <http://www.oceanoptics.com/products/maya.asp>. The optical fiber used is two meters long  
95 (attenuation of the photon flux is length dependent) connecting the spectrometer with the coupling lens  
96 at the end of the plasma torch of the MC-ICP-MS. Ca II ion lines were measured at a wavelength of  
97 393.48 nm and 396.86. At these wavelengths the Ca spectra shows no detectable interferences for the  
98 matrices used. The acquisition parameters were set to acquire 220 cycles per analysis with an  
99 integration time of 1 s for each cycle. Because of the stable BG signal detected for the first 40 cycles  
100 BG correction was done by subtracting its intensity from the intensity of the reference material and  
101 samples. For the determination of the relative error of the B concentration and B/Ca we considered the  
102 B and Ca concentrations of SRMNIST 610 and their uncertainties known from literature (lit) and the

103 intensities of B and Ca and their uncertainties measured (m) in this study with respect to SRMNIST

104 610 and the samples where B cps were normalized to Ca cps:

$$105 RSD (\%) = \sqrt{\frac{\left(\frac{SD B conc_{NIST}}{B conc_{NIST}}\right)_{lit}^2 + \left(\frac{SD Ca conc_{NIST}}{Ca conc_{NIST}}\right)_{lit}^2 + \left(\frac{SD B cps_{NIST}}{B cps_{NIST}}\right)_m^2 + \left(\frac{SD Ca cps_{NIST}}{Ca cps_{NIST}}\right)_m^2 + \left(\frac{SD B cps_{sample}}{B cps_{sample}}\right)_m^2 + \left(\frac{SD Ca cps_{sample}}{Ca cps_{sample}}\right)_m^2}{\left(\frac{SD B cps_{sample}}{B cps_{sample}}\right)_m^2 + \left(\frac{SD Ca cps_{sample}}{Ca cps_{sample}}\right)_m^2}} \times 100 \quad (S5)$$

106 Based on all measurements of NIST 610 performed in this study the precision for the B  
107 concentration (and subsequently B/Ca) was 3 %. Measuring the B concentration of NIST 612  
108 (calibrated against NIST 610) yielded  $35 \pm 1$  ppm which is in excellent agreement with the value  
109 reported in literature (34 – 39 ppm, (Tiepolo et al., 2005; Jacob et al., 2006; Hu et al., 2009; Liu et al.,  
110 2008; Deschamps et al., 2010; Lazarov et al., 2012)).

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## TABLES

**Table S1**

Composition of the culture media. Shown are the calculated carbonate systems using different pairs of input parameters. m - measured, cal - calculated. For information about errors on measured parameters see paper text section 2.3.

Input pH and DIC	pH <sub>m</sub> total	DIC <sub>m</sub> μmol/kg	TA <sub>m</sub> μmol/kg	TA <sub>cal</sub> μmol/kg	CO <sub>3</sub> <sup>2-</sup> <sub>cal</sub> μmol/kg	HCO <sub>3</sub> <sup>-</sup> <sub>cal</sub> μmol/kg	B(OH) <sub>4</sub> <sup>-</sup> <sub>cal</sub> <sup>a</sup> μmol/kg	pCO <sub>2</sub> <sub>cal</sub> μatm	B <sub>m</sub> μmol/kg	Ca <sub>m</sub> μmol/kg	δ <sup>11</sup> B <sub>m</sub> ‰	δ <sup>11</sup> B B(OH) <sub>4</sub> <sup>-</sup> <sub>cal</sub> ‰
pH_8.1 <sup>160</sup>	8.115	1257	2501	2456	155	1096	1062	213	4233	8503	-9.22	-29.01
pH_8.1 <sup>260</sup>	8.111	2093	3232	3387	257	1826	1016	356	4063	9638	-9.00	-28.81
pH_8.1 <sup>540</sup>	8.142	4128	5626	5748	538	3573	1098	652	4177	9692	-8.88	-28.37
pH_8.1 <sup>640</sup>	8.109	5736	7392	7295	643	5064	1053	1006	4240	9360	-8.73	-28.59
pH_8.6 <sup>640</sup>	8.560	2261	5387	4925	638	1620	1904	113	3944	9688	-9.00	-22.75
pH_7.9 <sup>260</sup>	7.858	3890	4691	4789	280	3576	647	1254	4136	9580	-9.11	-31.34
Input pH and TA	pH <sub>m</sub> total	TA <sub>m</sub> μmol/kg	DIC <sub>m</sub> μmol/kg	DIC <sub>cal</sub> μmol/kg	CO <sub>3</sub> <sup>2-</sup> <sub>cal</sub> μmol/kg	HCO <sub>3</sub> <sup>-</sup> <sub>cal</sub> μmol/kg	B(OH) <sub>4</sub> <sup>-</sup> <sub>cal</sub> <sup>a</sup> μmol/kg	pCO <sub>2</sub> <sub>cal</sub> μatm	B <sub>m</sub> μmol/kg	Ca <sub>m</sub> μmol/kg	δ <sup>11</sup> B <sub>m</sub> ‰	δ <sup>11</sup> B B(OH) <sub>4</sub> <sup>-</sup> <sub>cal</sub> ‰
pH_8.1 <sup>160</sup>	8.115	2501	1257	1297	160	1131	1062	220	4233	8503	-9.22	-29.01
pH_8.1 <sup>260</sup>	8.111	3232	2093	1955	240	1706	1016	333	4063	9638	-9.00	-28.81
pH_8.1 <sup>540</sup>	8.142	5626	4128	4020	524	3479	1098	635	4177	9692	-8.88	-28.37
pH_8.1 <sup>640</sup>	8.109	7392	5736	5669	691	4952	1037	975	4240	9360	-8.73	-28.69
pH_8.6 <sup>640</sup>	8.560	5387	2261	2348	663	1682	1904	117	3944	9688	-9.00	-22.75
pH_7.9 <sup>260</sup>	7.858	4691	3890	3798	273	3491	647	1225	4136	9580	-9.11	-31.34
Input DIC and TA	pH <sub>m</sub> total	pH <sub>cal</sub> total	DIC <sub>m</sub> μmol/kg	TA <sub>m</sub> μmol/kg	CO <sub>3</sub> <sup>2-</sup> <sub>cal</sub> μmol/kg	HCO <sub>3</sub> <sup>-</sup> <sub>cal</sub> μmol/kg	B(OH) <sub>4</sub> <sup>-</sup> <sub>cal</sub> <sup>a</sup> μmol/kg	pCO <sub>2</sub> <sub>cal</sub> μatm	B <sub>m</sub> μmol/kg	Ca <sub>m</sub> μmol/kg	δ <sup>11</sup> B <sub>m</sub> ‰	δ <sup>11</sup> B B(OH) <sub>4</sub> <sup>-</sup> <sub>cal</sub> ‰
pH_8.1 <sup>160</sup>	8.115	8.136	1257	2501	162	1090	1100	202	4233	8503	-9.22	-28.77
pH_8.1 <sup>260</sup>	8.111	8.045	2093	3232	223	1858	901	424	4063	9638	-9.00	-29.54
pH_8.1 <sup>540</sup>	8.142	8.100	4128	5626	494	3614	1022	725	4177	9692	-8.88	-28.83
pH_8.1 <sup>640</sup>	8.109	8.086	5736	7392	667	5040	1012	1045	4240	9360	-8.73	-28.84
pH_8.6 <sup>640</sup>	8.560	8.689	2261	5387	783	1476	2195	76	3944	9688	-9.00	-20.81
pH_7.9 <sup>260</sup>	7.858	7.806	3890	4691	250	3601	714	1424	4136	9580	-9.11	-30.92

<sup>a</sup> Calculation of B(OH)<sub>4</sub><sup>-</sup> was performed using a pK<sub>B</sub> of 8.59 corrected for S and T according to Dickson 1990.

**Table S2**

ICP-MS settings for B isotopes determination of culture media and standards.

Parameter	Isotope mode
Plasma RF Power	1250 W
Nebulizer	Savillex™-50 µl (C-flow)
Spray Chamber	Teflon Scott type (single pass)
Injector	ESI™ Platinum (1.8 mm I.D)
Sample Cone	Ni - JET
Skimmer Cone	Ni- X
Extraction Voltage	-2000 V
Sample Matrix	0.3 M HF
Uptake time	60 s
Analysis time	210 s
Washout time	150 s
Mass Resolution	Low
Magner Mass	10.012 (fixed)
<sup>10</sup> B Mass Range	10.012 - 10.013
<sup>11</sup> B Mass Range	11.008 - 11.009
Samples per peak	200 ( <sup>10</sup> B and <sup>11</sup> B)
Runs	225
Passes	10
Detection mode	Analog

**Table S3**

Laser efficiencies applied to NISTSRM 610 and foraminifers.

treatment	pH_8.1 <sup>160</sup>	pH_8.1 <sup>260</sup>	pH_8.1 <sup>540</sup>	pH_8.1 <sup>640</sup>	pH_8.6 <sup>640</sup>	pH_7.9 <sup>260</sup>
Laser efficiency (Hz) NIST 610	15	10	8	15	10	10
Laser efficiency (Hz) foraminifers	5	10	12	20	5	14
Spot size sample (μm)	50	50	50	50	30	50

**Table S4**

Instrumental operating conditions for Neptune MC-ICP-MS and LA.

Extraction[V]:	-2000
Focus[V]:	-759.2
Source Quad1[V]:	229.1
Rot-Quad1[V]:	12.8
Foc-Quad1[V]:	-19.8
Rot-Quad2[V]:	6.1
Source Offset[V]:	-13
Matsuda Plate[V]:	0.1
Cool Gas[l/min]:	14.6
Aux Gas[l/min]:	1.2
Sample Gas[l/min]:	1.3
Add Gas[l/min]:	0.3
Org Gas[l/min]:	0
Operation Power[W]:	1268.9
X-Pos[mm]:	1.5
Y-Pos[mm]:	-1.3
Z-Pos[mm]:	-4
Ampl.-Temp[°C]:	46.6
Fore Vacuum[mbar]:	2.31E-04
High Vacuum[mbar]:	1.27E-07
IonGetter-Press[mbar]:	3.22E-08
Guard electrode	on

**Table S5** Results from single B measurements of foraminifers. Errors are calculated according to formulas S3 (error of  $\delta^{11}\text{B}$ ) and S5 (error of B/Ca).

Treatment pH 8.1 <sup>160</sup> N = 50				
Foram #	$\delta^{11}\text{B}$ (‰)	$\pm$ (‰)	B/Ca (mmol/l)	$\pm$ (mmol/mol)
1	-33.88	0.87	5.16	0.06
1	-32.08	1.04	4.55	0.18
2	-33.43	1.00	5.37	0.05
2	-33.27	0.97	5.60	0.07
3	-31.98	1.10	5.22	0.09
3	-32.84	0.95	5.95	0.08
3	-33.69	1.18	3.47	0.06
4	-32.11	1.20	3.18	0.02
5	-35.33	1.15	3.91	0.06
5	-29.75	1.03	3.80	0.02
6	-31.11	1.14	3.43	0.03
7	-33.77	1.05	6.48	0.35
7	-33.38	1.12	5.21	0.23
8	-33.91	1.17	5.59	0.30
8	-33.47	1.48	4.90	0.26
9	-32.44	1.19	4.63	0.20
10	-32.20	0.85	5.26	0.20
11	-32.05	0.80	4.27	0.15
12	-33.32	0.91	6.27	0.09
12	-33.29	0.82	5.70	0.05
13	-32.27	0.99	5.48	0.06
13	-32.75	1.08	5.61	0.08
14	-31.83	0.90	5.57	0.05
14	-33.87	0.96	6.85	0.08
15	-32.05	0.93	6.74	0.09
16	-29.83	0.98	5.24	0.06
16	-29.51	0.87	4.81	0.07
17	-32.46	0.98	5.80	0.05
17	-32.90	0.94	5.53	0.06
18	-32.87	0.90	6.75	0.17
19	-31.88	0.96	6.90	0.06
19	-33.51	0.88	7.20	0.06
20	-32.27	0.98	6.21	0.06
20	-29.75	1.08	6.26	0.06
21	-32.75	0.88	7.17	0.08
22	-32.60	0.94	4.61	0.07
22	-32.43	0.85	4.90	0.08
23	-33.29	0.82	5.33	0.05
24	-33.55	0.86	5.47	0.06
24	-35.05	0.80	4.93	0.10
25	-34.84	0.83	4.08	0.04
26	-32.28	1.18	3.53	0.18
26	-31.46	1.06	3.47	0.20
27	-33.54	1.00	4.03	0.19
27	-32.75	1.21	4.79	0.33
28	-32.31	1.83	4.26	0.17
28	-31.48	0.85	3.83	0.18
29	-33.57	1.59	5.39	0.27
30	-34.71	1.31	6.89	0.76
31	-33.67	1.32	5.81	0.32

Treatment pH 8.1 <sup>260</sup> N = 41				
Foram #	$\delta^{11}\text{B}$ (‰)	$\pm$ (‰)	B/Ca (mmol/l)	$\pm$ (mmol/mol)
32	-33.13	1.05	3.39	0.04
32	-32.11	0.97	2.79	0.02
32	-32.22	0.94	2.82	0.03
32	-32.89	1.80	4.60	0.10
33	-33.23	1.03	3.09	0.03
33	-33.10	0.95	3.28	0.04
33	-33.49	1.00	3.38	0.03
34	-30.58	1.11	2.51	0.05
34	-32.90	1.04	2.09	0.02
34	-32.36	1.05	2.52	0.04
35	-29.98	1.50	2.86	0.13
35	-32.97	1.37	3.35	0.27
36	-32.23	1.23	2.80	0.21
36	-32.37	1.29	3.06	0.20
37	-30.25	1.40	2.49	0.15
37	-31.77	0.99	2.36	0.10
38	-34.47	1.69	2.80	0.12
38	-32.88	1.38	2.73	0.13
39	-30.91	1.26	2.86	0.21
40	-31.87	0.94	3.38	0.19
41	-31.50	0.90	3.30	0.18
41	-31.09	0.91	3.11	0.11
42	-29.21	0.95	2.63	0.12
43	-32.44	1.01	3.88	0.33
44	-30.23	1.07	2.60	0.44
44	-31.50	0.91	2.52	0.06
44	-33.56	0.90	2.92	0.04
45	-33.31	0.84	3.84	0.06
45	-31.89	0.99	2.93	0.06
45	-31.56	0.88	2.84	0.03
46	-31.91	0.81	3.29	0.03
46	-30.53	0.84	2.54	0.04
46	-32.36	0.86	2.90	0.03
47	-30.21	0.91	3.41	0.16
47	-30.11	0.82	2.74	0.13
48	-33.77	0.99	3.85	0.17
49	-31.94	0.76	3.18	0.03
50	-31.13	0.62	2.84	0.09
51	-30.42	0.80	2.07	0.06
52	-30.83	1.13	1.91	0.04
53	-31.82	0.64	2.29	0.05

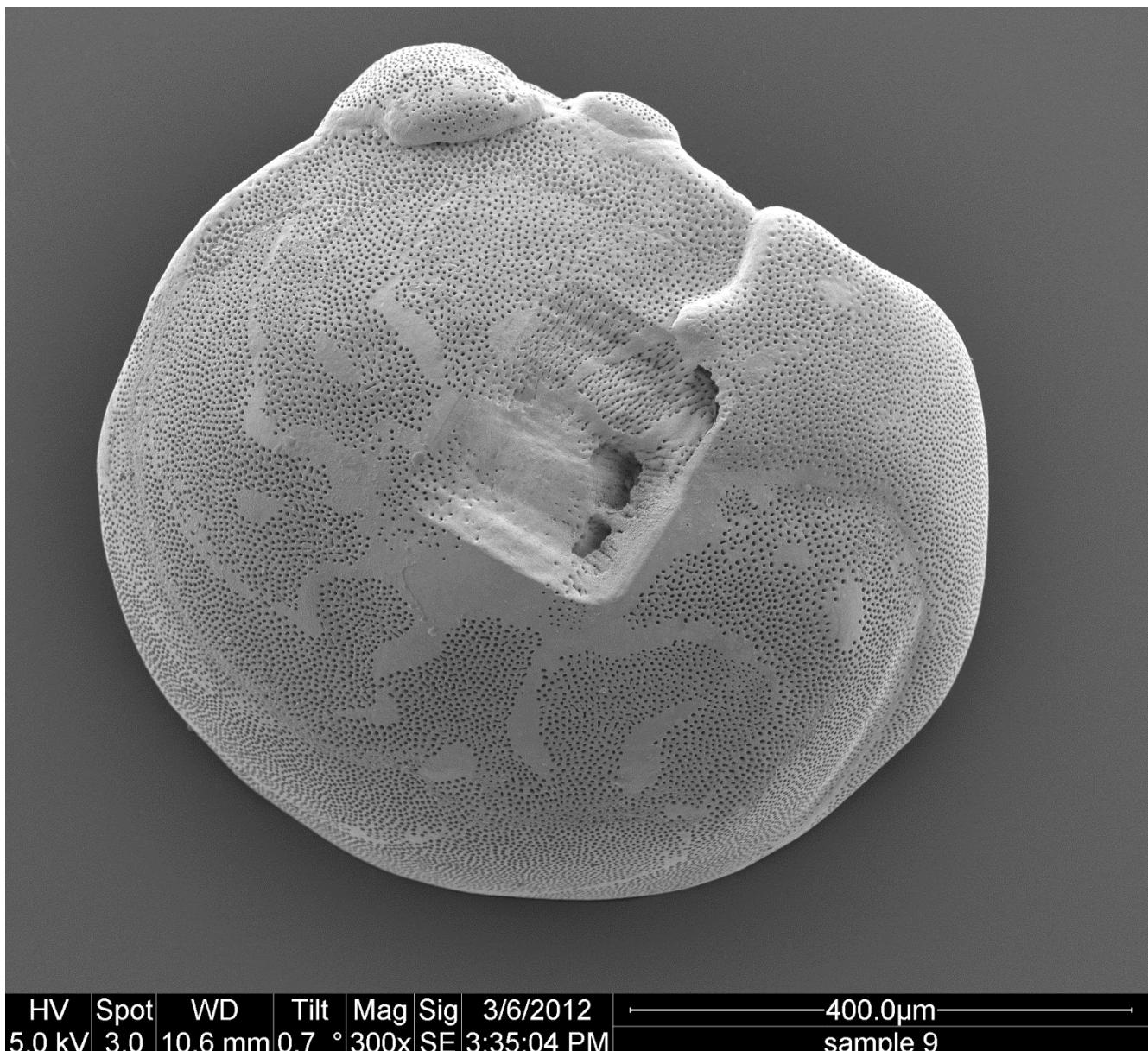
Treatment pH 8.1 <sup>540</sup> N = 15				
Foram #	$\delta^{11}\text{B}$ (‰)	$\pm$ (‰)	B/Ca (mmol/l)	$\pm$ (mmol/mol)
54	-29.19	0.98	1.59	0.01
54	-30.84	0.96	1.69	0.01
55	-31.09	0.97	1.85	0.01
55	-31.10	0.93	1.79	0.01
56	-31.69	0.92	1.64	0.01
56	-31.88	0.92	1.96	0.02
57	-33.08	0.98	1.74	0.01
57	-31.38	0.98	1.87	0.01
58	-32.26	0.95	1.73	0.01
58	-30.44	0.96	1.79	0.01
59	-31.23	1.02	1.84	0.02
59	-32.25	0.99	1.86	0.02
60	-34.40	1.25	1.65	0.03
61	-31.31	1.01	1.57	0.01
62	-33.14	1.13	1.71	0.04

Treatment pH 8.1 <sup>640</sup> N = 12				
Foram #	$\delta^{11}\text{B}$ (‰)	$\pm$ (‰)	B/Ca (mmol/l)	$\pm$ (mmol/mol)
63	-31.66	1.19	1.61	0.02
63	-29.72	1.22	1.36	0.02
64	-35.89	1.16	1.37	0.02
65	-31.76	1.70	1.42	0.16
66	-33.19	1.13	1.73	0.03
66	-32.87	1.00	1.72	0.03
67	-32.76	1.48	1.57	0.01
67	-31.66	0.91	1.60	0.02
68	-33.57	1.12	1.66	0.02
68	-32.97	1.29	1.67	0.08
69	-31.41	0.89	1.67	0.02
69	-31.97	0.92	1.58	0.03

Treatment pH 8.6 <sup>640</sup> N = 12				
Foram #	$\delta^{11}\text{B}$ (‰)	$\pm$ (‰)	B/Ca (mmol/l)	$\pm$ (mmol/mol)
70	-26.53	0.88	8.86	0.07
70	-25.57	0.97	8.11	0.08
71	-26.27	1.07	4.98	0.11
72	-24.14	1.09	7.37	0.10
73	-24.71	0.93	4.60	0.10
74	-23.56	0.89	6.67	0.05
74	-21.50	0.85	6.67	0.07
75	-23.79	0.88	7.07	0.09
75	-20.89	0.96	6.37	0.06
76	-24.12	0.95	4.77	0.04
77	-22.53	0.87	5.65	0.15
77	-20.13	0.87	5.22	0.08

<b>Treatment pH 7.9<sup>260</sup> N = 11</b>				
<b>Foram #</b>	<b>δ11B (‰)</b>	<b>±(‰)</b>	<b>B/Ca (mmol/l)</b>	<b>±(mmol/mol)</b>
78	-33.57	1.19	1.04	0.01
78	-34.89	1.36	1.18	0.01
79	-36.14	1.39	1.34	0.02
79	-35.08	1.29	1.26	0.03
80	-36.29	1.50	1.24	0.02
81	-38.64	1.21	1.19	0.02
81	-35.67	0.93	1.15	0.01
82	-35.28	1.05	1.10	0.01
82	-36.13	1.02	1.16	0.01
83	-34.78	1.19	1.27	0.02
83	-35.07	1.10	1.29	0.02

**Figure S1**



A foraminifer from the pH<sub>8.1</sub><sup>160</sup> treatment after laser ablation. The knob was measured twice.