



Corrigendum to “Seasonal variability in methane and nitrous oxide fluxes from tropical peatlands in the western Amazon basin” published in Biogeosciences, 14, 3669–3683, 2017

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Published: 21 June 2018

In the paper “Seasonal variability in methane and nitrous oxide fluxes from tropical peatlands in the western Amazon basin” by Teh et al. (Biogeosciences, 14, 3669–3683, 2017), the following error occurred: methane (CH₄) and nitrous oxide (N₂O) fluxes were calculated using incorrect values for the surface area of the flux chambers, leading to an underestimation of the CH₄ and N₂O fluxes. Application of the correct surface area resulted in fluxes that were approximately 5× higher than values reported in the paper, except for forested vegetation, where fluxes increased by approximately 16×.

Importantly, statistical re-analysis indicates that none of the principal trends or major findings reported in the original paper were affected by this scaling error. For example, CH₄ fluxes from the plant communities sampled in this paper are still large, implying that the Pastaza-Marañón foreland basin (PMFB) is a major regional source of atmospheric CH₄, as we stated before. Likewise, the revised N₂O flux is still negligible when compared against high-emitting ecosystems such as agro-ecosystems or lowland terra firme forest. Lastly, the divergent seasonal trends in CH₄ flux also still remain; i.e. forested vegetation and mixed palm swamp show significantly greater emissions during the dry season compared to the wet season, whereas forested (short pole) vegetation and *Mauritia flexuosa* palm swamp show the reverse trend. This last result is important because it suggests that the two different sets of ecosystems experience different environmental controls. Moreover, the seasonally asynchronous CH₄ fluxes from different vegetation types may partially explain the weak seasonality in CH₄ flux observed across the

Amazon basin as a whole, as observed by investigators such as Wilson et al. (2016).

However, correction of this error has resulted in changes to some of the minor findings. For instance, forested vegetation now shows the highest diffusive CH₄ flux, followed by mixed palm swamp, *M. flexuosa* palm swamp, and forested (short pole) vegetation. Second, the revised estimates of CH₄ fluxes now suggest that emissions from the PMFB region greatly exceed the flux observed from comparable peatland ecosystems in Southeast Asia. This result is more generally consistent with other empirical studies from central and South America, which indicate that tropical peatlands in the Americas tend to show greater CH₄ emission potentials than their analogues in Southeast Asia. Third and last, revised estimates of CH₄ fluxes now suggest that there is no qualitative trend in ebullition among different vegetation types, which we speculated upon in the original paper.

In order to correct for this error, we present a fully revised version of the paper and Supplement here. The revised paper includes updated values for CH₄ and N₂O fluxes in the body of the text, revised statistical outputs, and corrected figures and tables. Selected parts of the text have also been revised to account for changes to some of the minor findings that we have reported in this corrigendum.

Abstract. The Amazon plays a critical role in global atmospheric budgets of methane (CH₄) and nitrous oxide (N₂O). However, while we have a relatively good understanding of the continental-scale flux of these greenhouse gases (GHGs), one of the key gaps in knowledge is the specific contribution of peatland ecosystems to the regional budgets of these GHGs. Here we report CH₄ and N₂O fluxes from lowland tropical peatlands in the Pastaza-Marañón foreland basin (PMFB) in Peru, one of the largest peatland complexes in the Amazon basin. The goal of this research was to quantify the range and magnitude of CH₄ and N₂O fluxes from this region; assess seasonal trends in trace gas exchange; and determine the role of different environmental variables in driving GHG flux. Trace gas fluxes were determined from the most numerically dominant peatland vegetation types in the region: forested vegetation, forested (short pole) vegetation, *Mauritia flexuosa*-dominated palm swamp, and mixed palm swamp. Data were collected in both wet and dry seasons over the course of four field campaigns from 2012 to 2014. Diffusive CH₄ emissions averaged 204.8 ± 23.5 mg CH₄-C m⁻² d⁻¹ across the entire dataset, with diffusive CH₄ flux varying significantly among vegetation types and between seasons. Net ebullition of CH₄ averaged 4625.2 ± 752.1 mg CH₄-C m⁻² d⁻¹, and did not vary significantly among vegetation types or between seasons. Diffusive CH₄ flux was greatest for forested vegetation (469.3 ± 179.4 mg CH₄-C m⁻² d⁻¹), followed by mixed palm swamp (279.5 ± 82.0 mg CH₄-C m⁻² d⁻¹), *M. flexuosa* palm swamp (171.1 ± 20.1 mg CH₄-C m⁻² d⁻¹), and forested (short pole) vegetation (149.2 ± 21.9 mg CH₄-C m⁻² d⁻¹). Diffusive CH₄ flux also showed marked seasonality, with divergent seasonal patterns among ecosystems. Forested vegetation and mixed palm swamp showed significantly higher dry season (751.5 ± 290.4 and 457.6 ± 132.9 mg CH₄-C m⁻² d⁻¹, respectively) compared to wet season emissions (31.9 ± 4.9 and 24.3 ± 12.9 mg CH₄-C m⁻² d⁻¹, respectively). In contrast, forested (short pole) vegetation and *M. flexuosa* palm swamp showed the opposite trend, with dry season flux of 45.3 ± 12.3 and 97.2 ± 10.3 mg CH₄-C m⁻² d⁻¹, respectively, versus wet season flux of 488.1 ± 64.4 and 285.1 ± 39.1 mg CH₄-C m⁻² d⁻¹, respectively. These divergent seasonal trends may be linked to very high water tables (> 1 m) in forested vegetation and mixed palm swamp during the wet season, which may have constrained CH₄ transport across the soil–atmosphere interface. Diffusive N₂O flux was low (7.90 ± 4.93 µg N₂O-N m⁻² d⁻¹), and did not vary significantly among ecosystems or between seasons. We conclude that peatlands in the PMFB are large and regionally significant sources of atmospheric CH₄ that need to be better accounted for in regional emissions inventories. In contrast, N₂O flux was negligible, suggesting that this region does not make a significant contribution to regional atmospheric budgets of N₂O. The divergent seasonal pattern in CH₄ flux among vegetation types challenges our underlying assump-

tions of the controls on CH₄ flux in tropical peatlands, and emphasizes the need for more process-based measurements during high water table periods.

1 Introduction

The Amazon basin plays a critical role in the global atmospheric budgets of carbon (C) and greenhouse gases (GHGs) such as methane (CH₄) and nitrous oxide (N₂O). Recent basin-wide studies suggest that the Amazon as a whole accounts for approximately 7% of global atmospheric CH₄ emissions (Wilson et al., 2016). N₂O emissions are of a similar magnitude, with emissions ranging from 2 to 3 Tg N₂O-N yr⁻¹ (or approximately 12–18% of global atmospheric emissions) (Huang et al., 2008; Saikawa et al., 2013, 2014). While we have a relatively strong understanding of the role that the Amazon plays in regional and global atmospheric budgets of these gases, one of the key gaps in knowledge is the contribution of specific ecosystem types to regional fluxes of GHGs (Huang et al., 2008; Saikawa et al., 2013, 2014). In particular, our understanding of the contribution of Amazonian wetlands to regional C and GHG budgets is weak, as the majority of past ecosystem-scale studies have focused on *terra firme* forests and savannas (D’Amelio et al., 2009; Saikawa et al., 2013; Wilson et al., 2016; Kirschke et al., 2013; Nisbet et al., 2014). Empirical studies of GHG fluxes from Amazonian wetlands are more limited in geographic scope and have focused on three major areas: wetlands in the state of Amazonas near the city of Manaus (Devol et al., 1990; Bartlett et al., 1988, 1990; Keller et al., 1986), the Pantanal region (Melack et al., 2004; Marani and Alvalá, 2007; Lienggaard et al., 2013), and the Orinoco River basin (Smith et al., 2000; Lavelle et al., 2014). Critically, none of the ecosystems sampled in the past were peat-forming ones; rather, the habitats investigated were non-peat-forming (i.e. mineral or organo-mineral soils), seasonally inundated floodplain forests (i.e. *varzea*), rivers or lakes.

Peatlands are one of the major wetland habitats absent from current bottom-up GHG inventories for the Amazon basin, and are often grouped together with non-peat-forming wetlands in regional atmospheric budgets (Wilson et al., 2016). Unlike their Southeast Asian counterparts, most peatlands in the Amazon basin are unaffected by human activity at the current time (Lähteenoja et al., 2009a, b; Lähteenoja and Page, 2011), except for ecosystems in the Madre de Dios region in southeastern Peru, which are impacted by gold mining (Householder et al., 2012). Because we have little or no data on ecosystem-level land–atmosphere fluxes from Amazonian peatlands (Lähteenoja et al., 2012, 2009b; Kirschke et al., 2013; Nisbet et al., 2014), it is difficult to ascertain whether rates of GHG flux from these ecosystems are similar to or different from mineral soil wetlands (e.g. *varzea*). Given that underlying differences in plant community composition and soil properties are known to modulate the cy-

cling and flux of GHGs in wetlands (Limpens et al., 2008; Melton et al., 2013; Belyea and Baird, 2006; Sjögersten et al., 2014), expanding our observations to include a wider range of wetland habitats is critical in order to improve our understanding of regional trace gas exchange, and also to determine whether aggregating peat and mineral soil wetlands together in bottom-up emissions inventories is appropriate for regional budget calculations. Moreover, Amazonian peatlands are thought to account for a substantial land area (i.e. up to 150 000 km²) (Schulman et al., 1999; Lähteenoja et al., 2012), and any differences in biogeochemistry among peat and mineral/organo-mineral soil wetlands may therefore have important implications for understanding and modelling the biogeochemical functioning of the Amazon basin as a whole.

Since the identification of extensive peat-forming wetlands in the north (Lähteenoja et al., 2009a, b; Lähteenoja and Page, 2011) and south (Householder et al., 2012) of the Peruvian Amazon, several studies have been undertaken to better characterize these habitats, investigating vegetation composition and habitat diversity (Draper et al., 2014; Kelly et al., 2014; Householder et al., 2012; Lähteenoja and Page, 2011), vegetation history (Lähteenoja and Roucoux, 2010), C stocks (Lähteenoja et al., 2012; Draper et al., 2014), hydrology (Kelly et al., 2014), and peat chemistry (Lähteenoja et al., 2009a, b). Most of the studies have focused on the Pastaza-Marañón foreland basin (PMFB), where one of the largest stretches of contiguous peatlands has been found (Lähteenoja et al., 2009a; Lähteenoja and Page, 2011; Kelly et al., 2014), covering an estimated area of $35\,600 \pm 2133$ km² (Draper et al., 2014). Up to 90 % of the peatlands in the PMFB lie in flooded backwater river margins on floodplains and are influenced by large, annual fluctuations in the water table caused by the Amazonian flood pulse (Householder et al., 2012; Lähteenoja et al., 2009a). These floodplain systems are dominated by peat deposits that range in depth from ~ 3.9 (Lähteenoja et al., 2009a) to ~ 12.9 m (Householder et al., 2012). The remaining 10 % of these peatlands are not directly influenced by river flow and form domed (i.e. raised) nutrient-poor bogs that likely only receive water and nutrients from rainfall (Lähteenoja et al., 2009b). These nutrient-poor bogs are dominated by large, C-rich forests (termed “pole forests”) that represent a very high density C store (total pool size of 1391 ± 710 Mg C ha⁻¹, which includes both above- and below-ground stocks), exceeding in fact the C density of nearby floodplain systems (Draper et al., 2014). Even though the peats in these nutrient-poor bogs have a relatively high hydraulic conductivity, they act as natural stores of water because of high rainwater inputs (> 3000 mm a⁻¹), which help to maintain high water tables, even during parts of the dry season (Kelly et al., 2014).

CH₄ flux in tropical soils is regulated by the complex interplay among multiple factors that regulate CH₄ production, oxidation, and transport. Key factors include redox/water table depth (Couwenberg et al., 2010, 2011; Silver et al., 1999;

Teh et al., 2005; von Fischer and Hedin, 2007), plant productivity (von Fischer and Hedin, 2007; Whiting and Chanton, 1993), soil organic matter lability (Wright et al., 2011), competition for C substrates among anaerobes (Teh et al., 2008; Teh and Silver, 2006; von Fischer and Hedin, 2007), and the presence of plants capable of facilitating atmospheric egress (Pangala et al., 2013). Of all these factors, fluctuation in soil redox conditions, as mediated by variations in water table depth, is perhaps most critical in regulating CH₄ dynamics (Couwenberg et al., 2010, 2011), because of the underlying physiology of the microbes that produce and consume CH₄. Methanogenic archaea are obligate anaerobes that only produce CH₄ under anoxic conditions (Conrad, 1996); as a consequence, they are only active in stably anoxic soil microsites or soil layers, where they are protected from the effects of strong oxidants such as oxygen or where competition for reducing equivalents (e.g. acetate, H₂) from other anaerobic microorganisms is eliminated (Teh et al., 2005, 2008; Teh and Silver, 2006; von Fischer and Hedin, 2002, 2007). CH₄ oxidation, on the other hand, is thought to be driven primarily by aerobic methanotrophic bacteria in tropical soils (Hanson and Hanson, 1996; Teh et al., 2005, 2006; von Fischer and Hedin, 2002, 2007), with anaerobic CH₄ oxidation playing a quantitatively smaller role (Blazewicz et al., 2012). Thus, fluctuations in redox or water table depth play a fundamental role in directing the flow of C among different anaerobic pathways (Teh et al., 2008; Teh and Silver, 2006; von Fischer and Hedin, 2007), and shifting the balance between production and consumption of CH₄ (Teh et al., 2005; von Fischer and Hedin, 2002). Moreover, water table or soil moisture fluctuations are also thought to profoundly influence CH₄ transport dynamics throughout the soil profile, changing the relative partitioning of CH₄ among different transport pathways such as diffusion, ebullition, and plant-facilitated transport (Whalen, 2005; Jungkunst and Fiedler, 2007).

Controls on N₂O flux are also highly complex (Groffman et al., 2009), with N₂O originating from as many as four separate sources (e.g. bacterial ammonia oxidation, archaeal ammonia oxidation, denitrification, dissimilatory nitrate reduction to ammonium), each with different environmental controls (Baggs, 2008; Morley and Baggs, 2010; Firestone and Davidson, 1989; Firestone et al., 1980; Pett-Ridge et al., 2013; Silver et al., 2001; Prosser and Nicol, 2008). Key factors regulating soil N₂O flux include redox, soil moisture content or water table depth, temperature, pH, labile C availability, and labile N availability (Groffman et al., 2009). As is the case for CH₄, variations in redox/water table depth play an especially prominent role in regulating N₂O flux in tropical peatland ecosystems, because all of the processes that produce N₂O are redox-sensitive, with bacterial or archaeal ammonia oxidation occurring under aerobic conditions (Prosser and Nicol, 2008; Firestone and Davidson, 1989; Firestone et al., 1980), whereas nitrate-reducing processes (i.e. denitrification, dissimilatory nitrate reduction to ammonium) occur under anaerobic ones (Firestone and

Davidson, 1989; Firestone et al., 1980; Morley and Baggs, 2010; Silver et al., 2001). Moreover, for nitrate-reducing processes, which are believed to be the dominant source of N_2O in wet systems, the extent of anaerobiosis also controls the relative proportion of N_2O or N_2 produced during dissimilatory metabolism (Firestone and Davidson, 1989; Firestone et al., 1980; Morley and Baggs, 2010; Silver et al., 2001).

In order to improve our understanding of the biogeochemistry and rates of GHG exchange from Amazonian peatlands, we conducted a preliminary study of CH_4 and N_2O fluxes from forested peatlands in the PMFB. The main objectives of this are to

1. quantify the magnitude and range of soil CH_4 and N_2O fluxes from a sub-set of peatlands in the PMFB that represent dominant vegetation types;
2. determine seasonal patterns of trace gas exchange; and
3. establish the relationship between trace gas fluxes and environmental variables.

Sampling was concentrated on the four most dominant vegetation types in the area, based on prior work by the investigators (Lähteenoja and Page, 2011). Trace gas fluxes were captured from both floodplain systems and nutrient-poor bogs in order to account for underlying differences in biogeochemistry that may arise from variations in hydrology. Sampling was conducted during four field campaigns (two wet seasons, two dry seasons) over a 27-month period, extending from February 2012 to May 2014.

2 Materials and methods

2.1 Study site and sampling design

The study was carried out in the lowland tropical peatland forests of the PMFB, between 2 and 35 km south of the city of Iquitos, Peru (Lähteenoja et al., 2009a, b) (Fig. 1, Table 1). The mean annual temperature is 26 °C, annual precipitation is c. 3100 mm, relative humidity ranges from 80–90 %, and altitude ranges from ca. 90 to 130 m a.s.l. (above sea level) (Marengo et al., 1998). The northwestern Amazon basin near Iquitos experiences pronounced seasonality, which is characterized by consistently high annual temperatures, but marked seasonal variation in precipitation (Tian et al., 1998), and an annual river flood pulse linked to seasonal discharge from the Andes (Junk et al., 1989). Precipitation events are frequent, intense and of significant duration during the wet season (November to May) and infrequent, intense and of short duration during the dry season (June to August). September and October represent a transitional period between dry and wet seasons, where rainfall patterns are less predictable. Catchments in this region receive no less than 100 mm of rain per month (Espinoza Villar et al., 2009a, b) and >3000 mm of rain per year. River discharge varies

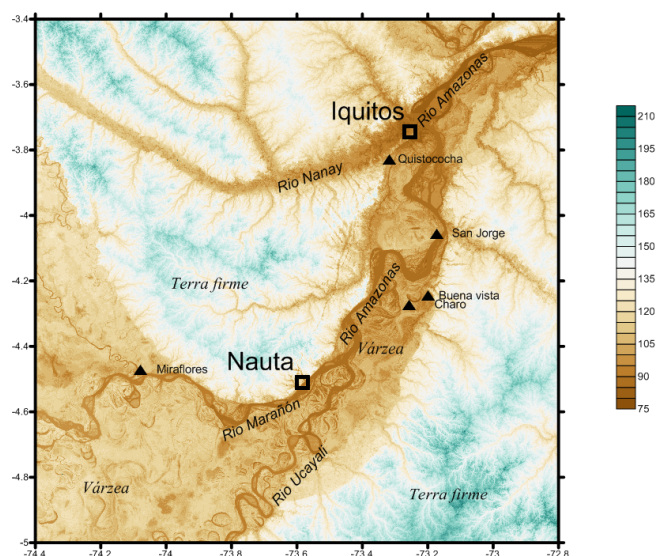


Figure 1. Map of the study region and field sites. The colour scale to the right of the map denotes elevation in metres above sea level (m a.s.l.). Tan and brown tones indicate areas in which peatlands are found; however, not all of these areas are peatland-dominated.

by season, with the lowest discharge between the dry season months of August and September. Peak discharge from the wet season flood pulse occurs between April and May, as recorded at the Tamshiyaku River gauging station (Espinoza Villar et al., 2009b).

Histosols form the dominant soil type for peatlands in this region (Andriese, 1988; Lähteenoja and Page, 2011). Study sites are broadly classified as nutrient-rich, intermediate, or nutrient-poor (Lähteenoja and Page, 2011), with pH ranging from 3.5 to 7.2 (Lähteenoja and Page, 2011; Lähteenoja et al., 2009a, b). More specific data on pH for our plots are presented in Table 3. Nutrient-rich (i.e. minerotrophic) sites tend to occur on floodplains and river margins, and account for at least 60 % of the peatland cover in the PMFB (Lähteenoja and Page, 2011; Draper et al., 2014). They receive water, sediment, and nutrient inputs from the annual Amazon River flood pulse (Householder et al., 2012; Lähteenoja and Page, 2011), leading to higher inorganic nutrient content, of which Ca and other base cations form major constituents (Lähteenoja and Page, 2011). Many of the soils in these nutrient-rich areas are fluvaquentic Tropofibrists (Andriese, 1988), and contain thick mineral layers or minerogenic intrusions, reflective of episodic sedimentation events in the past (Lähteenoja and Page, 2011). In contrast, nutrient-poor (i.e. oligotrophic) sites tend to occur further inland (Lähteenoja and Page, 2011; Draper et al., 2014). They are almost entirely rain-fed, and receive low or infrequent inputs of water and nutrients from streams and rivers (Lähteenoja and Page, 2011). These ecosystems account for 10 to 40 % of peatland cover in the PMFB, though precise estimates vary depending on the land classification scheme employed (Lähteenoja

Table 1. Site characteristics including field site location, nutrient status, plot, and flux chamber replication.

Vegetation type	Site name	Nutrient status*	Latitude (S)	Longitude (W)	Plots	Flux chambers
Forested	Buena Vista	Rich	4°14′45.60″ S	73°12′0.20″ W	21	105
Forested (short pole)	San Jorge (centre)	Poor	4°03′35.95″ S	73°12′01.13″ W	6	28
Forested (short pole)	Miraflores	Poor	4°28′16.59″ S	74°4′39.95″ W	41	204
<i>M. flexuosa</i> palm swamp	Quistococha	Intermediate	3°49′57.61″ S	73°12′01.13″ W	135	668
<i>M. flexuosa</i> palm swamp	San Jorge (edge)	Intermediate	4°03′18.83″ S	73°10′16.80″ W	18	86
Mixed palm swamp	Charo	Rich	4°16′21.80″ S	73°15′27.80″ W	18	90

* Following Householder et al. (2012) and Lahteenoja et al. (2009a, b).

and Page, 2011; Draper et al., 2014). Soil Ca and base cation concentrations are significantly lower in these sites compared to nutrient-rich ones, with similar concentrations to that of rainwater (Lähteenoja and Page, 2011). Soils are classified as typic or hydric Tropofibrists (Andriess, 1988). Even though Ca and base cations themselves play no direct role in modulating CH₄ and N₂O fluxes, underlying differences in soil fertility may indirectly influence CH₄ and N₂O flux by influencing the rate of labile C input to the soil, the decomposability of organic matter, and the overall throughput of C and nutrients through the plant–soil system (Firestone and Davidson, 1989; Groffman et al., 2009; von Fischer and Hedin, 2007; Whiting and Chanton, 1993).

We established 239 sampling plots (~30 m² per plot) within five tropical peatland sites that captured four of the dominant vegetation types in the region (Draper et al., 2014; Householder et al., 2012; Kelly et al., 2014; Lähteenoja and Page, 2011), and which encompassed a range of nutrient availabilities (Fig. 1, Table 1) (Lähteenoja and Page, 2011; Lähteenoja et al., 2009a). These four dominant vegetation types included forested vegetation (nutrient-rich; $n = 21$ plots), forested (short pole) vegetation (nutrient-poor; $n = 47$ plots), *Mauritia flexuosa*-dominated palm swamp (intermediate fertility; $n = 153$ plots), and mixed palm swamp (nutrient-rich; $n = 18$ plots) (Table 1). Four of the study sites (Buena Vista, Charo, Miraflores, and Quistococha) were dominated by only one vegetation type, whereas San Jorge contained a mixture of *M. flexuosa* palm swamp and forested (short pole) vegetation (Table 1). As a consequence, both vegetation types were sampled in San Jorge to develop a more representative picture of GHG fluxes from this location. Sampling efforts were partially constrained by issues of site access; some locations were difficult to access (e.g. centre of the San Jorge peatland) due to water table height and navigability of river channels; as a consequence, sampling patterns were somewhat uneven, with higher sampling densities in some peatlands than in others (Table 1).

In each peatland site, transects were established from the edge of the peatland to its centre. Each transect varied in length from 2 to 5 km, depending on the relative size of the peatland. Randomly located sampling plots (~30 m² per

plot) were established at 50 or 200 m intervals along each transect, from which GHG fluxes and environmental variables were measured concomitantly. The sampling interval (i.e. 50 or 200 m) was determined by the length of the transect or size of the peatland, with shorter sampling intervals (50 m) for shorter transects (i.e. smaller peatlands) and longer sampling intervals (200 m) for longer transects (i.e. larger peatlands).

2.2 Quantifying soil–atmosphere exchange

Soil–atmosphere fluxes (CH₄, N₂O) were determined in four campaigns over a 2-year annual water cycle: February 2012 (wet season), June–August 2012 (dry season), June–July 2013 (dry season), and May–June 2014 (wet season). The duration of the campaign for each study site varied depending on its size. Each study site was generally sampled only once for each campaign, except for a sub-set of plots within each vegetation type where diurnal studies were conducted to determine whether CH₄ and N₂O fluxes varied over daily time steps. Gas exchange was quantified using a floating static chamber approach (Livingston and Hutchinson, 1995; Teh et al., 2011). Static flux measurements were made by enclosing a 0.0476 m² area with a dark, single-component, vented 10 L flux chamber. No chamber bases (collars) were used due to the highly saturated nature of the soils. In most cases, a standing water table was present at the soil surface, so chambers were placed directly onto the water. In the absence of a standing water table, a weighted skirt was applied to create an airtight seal. Under these drier conditions, chambers were placed carefully on the soil surface. In order to reduce the risk of pressure-induced ebullition or disruption to soil gas concentration profiles caused by the investigators' footfall, flux chambers were lowered from a distance of 2 m away using a 2 m long pole. Gas samples were collected with syringes using > 2 m lengths of Tygon[®] tubing, after thoroughly purging the dead volumes in the sample lines. To promote even mixing within the headspace, chambers were fitted with small computer fans (Pumpanen et al., 2004). Headspace samples were collected from each flux chamber at five intervals over a 25 min enclosure period using a gas-tight syringe. Gas samples were stored in evacuated Exetainers[®] (Labco Ltd., Lam-

peter, UK), shipped to the UK, and subsequently analysed for CH₄, CO₂ and N₂O concentrations using Thermo TRACE GC Ultra (Thermo Fischer Scientific Inc., Waltham, Massachusetts, USA) at the University of St. Andrews. Chromatographic separation was achieved using a Porapak-Q column, and gas concentrations determined using a flame ionization detector (FID) for CH₄, a methanizer-FID for CO₂, and an electron capture detector (ECD) for N₂O. Instrumental precision, determined from repeated analysis of standards, was < 5 % for all detectors.

Diffusive fluxes were determined by using the JMP IN version 11 (SAS Institute, Inc., Cary, North Carolina, USA) statistical package to plot best-fit lines to the data for headspace concentration against time for individual flux chambers, with fluxes calculated from linear or non-linear regressions depending on the individual concentration trend against time (Teh et al., 2014). Gas mixing ratios (ppm) were converted to areal fluxes by using the ideal gas law to solve for the quantity of gas in the headspace (on a mole or mass basis) and normalized by the surface area of each static flux chamber (Livingston and Hutchinson, 1995). Ebullition-derived CH₄ fluxes were also quantified in our chambers where evidence of ebullition was found. This evidence consisted of either (i) rapid, non-linear increases in CH₄ concentration over time; (ii) abrupt, stochastic increases in CH₄ concentration over time; or (iii) an abrupt stochastic increase in CH₄ concentration, followed by a linear decline in concentration. For observations following pattern (i), flux was calculated by fitting a quadratic regression equation to the data ($P < 0.05$), and CH₄ flux determined from the initial steep rise in CH₄ concentration. For data following pattern (ii), the ebullition rate was determined by calculating the total CH₄ production over the course of the bubble event, in line with prior work conducted by the investigators (Teh et al., 2011). Last, for data following pattern (iii), a best-fit line was plotted to the CH₄ concentration data after the bubble event, and a net rate of CH₄ uptake calculated from the gradient of the line. While observations (i)–(iii) all reflect the effects of ebullition, only observations following patterns (i) and (ii) indicate net emission to the atmosphere, whereas observations following pattern (iii) indicate emission followed by net uptake. As a consequence, patterns (i) and (ii) were categorized as “net ebullition” (i.e. net efflux), whereas observations following pattern (iii) were categorized as “ebullition-driven CH₄ uptake” (i.e. net influx).

2.3 Environmental variables

To investigate the effects of environmental variables on trace gas fluxes, we determined air temperature, soil temperature, chamber headspace temperature, soil pH, soil electrical conductivity (EC; $\mu\text{S m}^{-2}$), dissolved oxygen concentration of the soil porewater (DO; measured as percent saturation, %) in the top 15 cm of the peat column, and water table position concomitant with gas sampling. Air temperature (mea-

sured 1.3 m above the soil) and chamber headspace temperature were measured using a Checktemp[®] probe and meter (Hanna Instruments LTD, Leighton Buzzard, UK). Peat temperature, pH, DO and EC were measured at a depth of 15 cm below the peat surface and recorded in situ with each gas sample using a HACH[®] rugged outdoor HQ30D multi meter and pH, LDO or EC probe. At sites where the water level was above the peat surface, the water depth was measured using a metre rule. Where the water table was at or below the peat surface, the water level was measured by auguring a hole to 1 m depth and measuring water table depth using a metre rule.

2.4 Statistical analyses

Statistical analyses were performed using JMP IN version 11 (SAS Institute, Inc., Cary, North Carolina, USA). Box–Cox transformations were applied where the data failed to meet the assumptions of analysis of variance (ANOVA); otherwise, non-parametric tests were applied (e.g. a Wilcoxon signed-rank test). ANOVA and analysis of co-variance (ANCOVA) were used to test for relationships between gas fluxes and vegetation type, season, and environmental variables. When determining the effect of vegetation type on gas flux, data from different study sites (e.g. San Jorge and Miraflores) were pooled together. Mean comparisons were tested using Fisher’s least significant difference (LSD) test.

3 Results

3.1 Differences in gas fluxes and environmental variables among vegetation types

All vegetation types were net sources of CH₄, with an overall mean (\pm standard error) diffusive flux of 204.8 ± 23.5 and a mean net ebullition flux of 4625.2 ± 752.1 mg CH₄–C m^{−2} d^{−1} (Fig. 2, Table 2). We also saw examples of ebullition-driven CH₄ uptake (i.e. a sudden or stochastic increase in CH₄ concentration, followed immediately by a rapid linear decline in concentration), with a mean rate of -2494.9 ± 386.8 mg CH₄–C m^{−2} d^{−1} (Table 2). Diffusive fluxes of CH₄ accounted for the majority of observations (87 %), while ebullition fluxes accounted for a much smaller proportion of observations (13 % of all fluxes; see Table 2 for a description of the relative proportion of ebullition fluxes for each vegetation type).

Diffusive CH₄ flux varied significantly among the four vegetation types sampled in this study (two-way ANOVA with vegetation, season and their interaction, $F_{7, 997} = 15.6$, $P < 0.0001$; Fig. 2a). However, the effect of vegetation was weaker than the effect of the vegetation by season interaction or the effect of season alone (see ANCOVA results in the section “Relationships between gas fluxes and environmental variables”). A means comparison test on the pooled data was unable to determine

Table 2. Proportion of observations for each vegetation type that showed evidence of ebullition, mean rates of ebullition, and ebullition-driven CH₄ uptake. Values represent means and standard errors.

Vegetation type	Percentage of observations (%)	Net ebullition (mg CH ₄ -C m ⁻² d ⁻¹)		Ebullition-driven (mg CH ₄ -C m ⁻² d ⁻¹)	
		Wet season	Dry season	Wet season	Dry season
Forested	1.9	0	0	0	-644.3 ± 0.4
Forested (short pole)	7.0	4696.6 ± 1384.5	2420.1 ± 722.4	-1159.4 ± 231.1	-452.4
<i>M. flexuosa</i> palm swamp	16.8	5628.7 ± 1443.6	4695.4 ± 1120.4	-4105.2 ± 1250.3	-1895.3 ± 282.9
Mixed palm swamp	11.0	0	4106.3 ± 1313.0	0	-4410 ± 1926.4

which means differed significantly from the others (Fisher's LSD, $P > 0.05$). For the pooled data, the overall numerical trend was that forested vegetation (469.3 ± 179.4 mg CH₄-C m⁻² d⁻¹) showed the highest mean diffusive flux, followed by mixed palm swamp (279.5 ± 82.0 mg CH₄-C m⁻² d⁻¹), *M. flexuosa* palm swamp (171.1 ± 82.0 mg CH₄-C m⁻² d⁻¹), and forested (short pole) vegetation (149.2 ± 21.9 mg CH₄-C m⁻² d⁻¹). In contrast to diffusive flux, CH₄ ebullition (i.e. net ebullition and ebullition-driven uptake) did not vary significantly among vegetation types or between seasons (Table 2). Broadly speaking, however, we saw a greater frequency of ebullition in the *M. flexuosa* palm swamp, followed by mixed palm swamp, forested (short pole) vegetation, and forested vegetation (Table 2).

These study sites were also a weak net source of N₂O, with a mean diffusive flux of 7.90 ± 4.93 μg N₂O-N m⁻² d⁻¹. We saw only limited evidence of ebullition of N₂O, with only 3 chambers out of 1181 (0.3 % of observations) showing evidence of N₂O ebullition. These data were omitted from the analysis of diffusive flux of N₂O. Because of the high variance in diffusive N₂O flux among plots, analysis of variance indicated that mean diffusive N₂O flux did not differ significantly among vegetation types (two-way ANOVA, $P > 0.5$, Fig. 2b). However, when the N₂O flux data were grouped by vegetation type, we see that some vegetation types tended to function as net atmospheric sources, while others acted as atmospheric sinks (Fig. 2b). For example, the highest N₂O emissions were observed from *M. flexuosa* palm swamp (13.46 ± 7.62 μg N₂O-N m⁻² d⁻¹). In contrast, forested (short pole) vegetation, forested vegetation and mixed palm swamp were weak sinks for N₂O, with a mean flux of -0.06 ± 3.94 , -3.14 ± 3.64 and -0.21 ± 0.70 μg N₂O-N m⁻² d⁻¹, respectively.

Soil pH varied significantly among vegetation types (data pooled across all seasons; ANOVA, $P < 0.0001$, Table 3). Multiple comparison tests indicated that mean soil pH was significantly different for each of the vegetation types (Fisher's LSD, $P < 0.0001$, Table 3), with the lowest pH in forested (short pole) vegetation (4.10 ± 0.04), followed by *M. flexuosa* palm swamp (5.32 ± 0.02), forested vegetation (6.15 ± 0.06), and the mixed palm swamp (6.58 ± 0.04).

Soil dissolved oxygen (DO) content varied significantly among vegetation types (data pooled across all seasons; Kruskal-Wallis, $P < 0.0001$, Table 3). Multiple comparison tests indicated that mean DO was significantly different for each of the vegetation types (Fisher's LSD, $P < 0.05$, Table 3), with the highest DO in the forested (short pole) vegetation (25.2 ± 2.1 %), followed by the *M. flexuosa* palm swamp (18.1 ± 1.0 %), forested vegetation (11.8 ± 2.8 %), and the mixed palm swamp (0.0 ± 0.0 %).

Electrical conductivity (EC) varied significantly among vegetation types (data pooled across all seasons; Kruskal-Wallis, $P < 0.0001$, Table 3). Multiple comparison tests indicated that mean EC was significantly different for each of the vegetation types (Fisher's LSD, $P < 0.05$, Table 3), with the highest EC in the mixed palm swamp (170.9 ± 6.0 μS m⁻²), followed by forested vegetation (77.1 ± 4.2 μS m⁻²), *M. flexuosa* palm swamp (49.7 ± 1.4 μS m⁻²) and the forested (short pole) vegetation (40.9 ± 3.5 μS m⁻²).

Soil temperature varied significantly among vegetation types (data pooled across all seasons; ANOVA, $P < 0.0001$, Table 3). Multiple comparison tests indicated that soil temperature in forested (short pole) vegetation was significantly lower than in the other vegetation types (Table 3), whereas the other vegetation types did not differ in temperature amongst themselves (Fisher's LSD, $P < 0.05$, Table 3).

Air temperature varied significantly among vegetation types (data pooled across all seasons; ANOVA, $P < 0.0001$, Table 3). Multiple comparison tests indicated that air temperature in *M. flexuosa* palm swamp was significantly lower than in the other vegetation types, whereas the other vegetation types did not differ in temperature amongst themselves (Fisher's LSD, $P < 0.05$, Table 3).

Water table depths varied significantly among vegetation types (data pooled across all seasons; ANOVA, $P < 0.0001$, Table 3). The highest mean water tables were observed in mixed palm swamp (59.6 ± 9.3 cm), followed by forested vegetation (34.0 ± 6.9 cm), *M. flexuosa* palm swamp (17.4 ± 1.2 cm), and forested (short pole) vegetation (3.5 ± 1.0 cm) (Fisher's LSD, $P < 0.0005$).

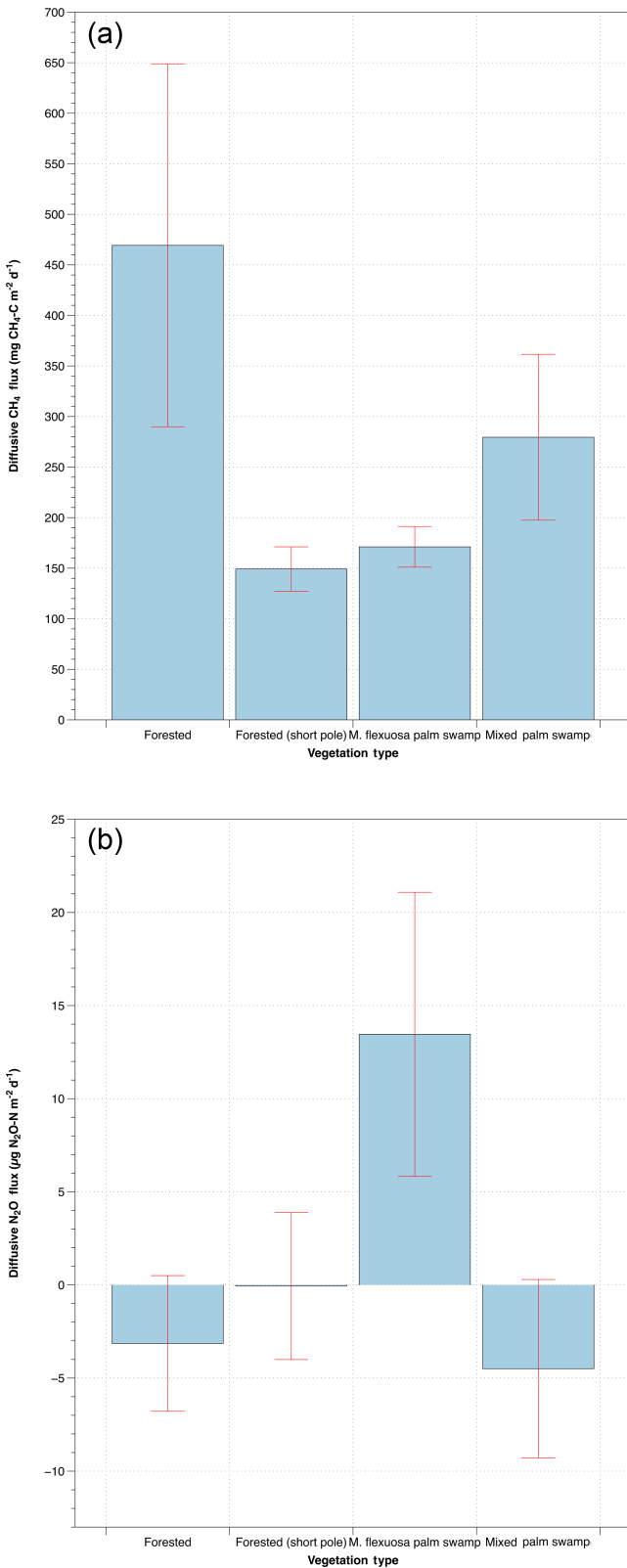


Figure 2. Net diffusive (a) methane (CH_4) and (b) nitrous oxide (N_2O) fluxes by vegetation type. Error bars denote standard errors.

Table 3. Environmental variables for each vegetation type for the wet and dry seasons. Values reported here are means and standard errors. Lowercase letters indicate significant differences among vegetation types within the wet or dry season (Fisher's LSD, $P < 0.05$).

Vegetation type	Peat temperature ($^{\circ}\text{C}$)		Air temperature ($^{\circ}\text{C}$)		Conductivity ($\mu\text{S m}^{-2}$)		Dissolved oxygen (%)		Water table level (cm)		pH	
	Wet season	Dry season	Wet season	Dry season	Wet season	Dry season	Wet season	Dry season	Wet season	Dry season	Wet season	Dry season
Forested	26.1 \pm 0.1a	24.7 \pm 0.0a	28.8 \pm 0.7a	26.4 \pm 0.3a	79.0 \pm 5.9a	75.9 \pm 5.7a	0.2 \pm 0.1a	18.9 \pm 4.4a	110.8 \pm 9.3a	-1.3 \pm 0.7a	5.88 \pm 0.15a	6.31 \pm 0.04a
Forested (short pole)	25.2 \pm 0.0b	24.8 \pm 0.1a	27.6 \pm 0.1b	27.5 \pm 0.1b	21.0 \pm 0.0b	48.5 \pm 4.8b	4.4 \pm 0.0a	33.1 \pm 2.6b	26.9 \pm 0.5b	-4.7 \pm 0.4b	4.88 \pm 0.01b	3.8 \pm 0.03b
<i>M. flexuosa</i> palm swamp	25.6 \pm 0.0c	25.3 \pm 0.1b	26.3 \pm 0.1c	26.4 \pm 0.1a	45.9 \pm 2.1c	51.9 \pm 1.8b	19.4 \pm 1.3b	17.3 \pm 1.5a	37.2 \pm 1.7c	6.1 \pm 1.3c	5.04 \pm 0.03c	5.49 \pm 0.03c
Mixed palm swamp	26.0 \pm 0.0a	25.0 \pm 0.1ab	26.1 \pm 0.1c	28.2 \pm 0.3b	100.0 \pm 0.2d	206.4 \pm 4.2c	0.0 \pm 0.0a	0.0 \pm 0.0c	183.7 \pm 1.7d	-2.4 \pm 0.3b	6.1 \pm 0.03a	6.82 \pm 0.02d

Table 4. Trace gas fluxes for each vegetation type for the wet and dry seasons. Values reported here are means and standard errors. Uppercase letters indicate significant differences in gas flux between seasons within a vegetation type (*t*-test, $P < 0.05$), while lowercase letters indicate significant differences among vegetation types within a season (Fisher's LSD, $P < 0.05$).

Vegetation type	Methane flux ($\text{mg CH}_4\text{-C m}^{-2} \text{d}^{-1}$)		Nitrous oxide flux ($\mu\text{g N}_2\text{O-N m}^{-2} \text{d}^{-1}$)	
	Wet season	Dry season	Wet season	Dry season
Forested	31.9 ± 4.9 Aa	751.5 ± 290.4 Ba	0.89 ± 1.53	-5.47 ± 5.68
Forested (short pole)	488.1 ± 64.4 Ab	45.3 ± 12.3 Bb	1.89 ± 0.86	-0.78 ± 5.39
<i>M. flexuosa</i> palm swamp	244.3 ± 44.8 Ab	120.6 ± 13.7 Bc	0.82 ± 0.41	21.11 ± 12.23
Mixed palm swamp	24.3 ± 12.9 Ab	457.6 ± 132.9 Ba	1.45 ± 0.79	-0.80 ± 0.79

3.2 Temporal variations in gas fluxes and environmental variables

The peatlands sampled in this study showed pronounced seasonal variability in diffusive CH_4 flux, with different plant communities showing divergent trends in diffusive CH_4 flux between seasons (two-way ANOVA, $F_{7, 997} = 15.6$, $P < 0.0001$; Table 4). For ebullition of CH_4 and ebullition-driven uptake of CH_4 , mean fluxes varied between seasons, but high variability meant that these differences were not statistically significant (two-way ANOVA, $P > 0.8$; Table 2). Diffusive N_2O flux showed no seasonal trends (two-way ANOVA, $P > 0.5$; Table 4), and therefore will not be discussed further here. Diurnal studies suggest that diffusive fluxes of neither CH_4 nor N_2O varied over the course of a 24 h period.

For diffusive CH_4 flux, the overall trend was towards higher wet season ($236.4 \pm 32.2 \text{ mg CH}_4\text{-C m}^{-2} \text{d}^{-1}$) compared to dry season ($186.3 \pm 32.1 \text{ mg CH}_4\text{-C m}^{-2} \text{d}^{-1}$) flux, although this effect was only significant at the $P < 0.08$ level (*t*-test with data pooled across all vegetation types; Table 4). However, when diffusive CH_4 flux was disaggregated by vegetation type, very different seasonal trends emerged (two-way ANOVA $F_{7, 997} = 15.6$, $P < 0.0001$, Table 4). For example, both forested vegetation and mixed palm swamp showed significantly greater diffusive CH_4 flux during the *dry season*, with net fluxes of 751.5 ± 290.4 and $457.6 \pm 132.9 \text{ mg CH}_4\text{-C m}^{-2} \text{d}^{-1}$, respectively (Fisher's LSD, $P < 0.05$, Table 4). In contrast, *wet season* fluxes were 19–24× lower, with net fluxes of 31.9 ± 4.9 and $24.3 \pm 12.9 \text{ mg CH}_4\text{-C m}^{-2} \text{d}^{-1}$, respectively (Fisher's LSD, $P < 0.05$, Table 4). On the other hand, forested (short pole) vegetation and *M. flexuosa* palm swamp showed seasonal trends consistent with the pooled data set; i.e. significantly higher flux during the wet season (488.1 ± 64.4 and $244.3 \pm 44.8 \text{ mg CH}_4\text{-C m}^{-2} \text{d}^{-1}$, respectively) compared to the dry season (45.3 ± 12.3 and $120.6 \pm 13.7 \text{ mg CH}_4\text{-C m}^{-2} \text{d}^{-1}$, respectively) (Fisher's LSD, $P < 0.05$, Table 3).

Even though seasonal trends in CH_4 ebullition were not statistically significant, we will briefly describe the over-

all patterns for the different vegetation types as they varied among ecosystems (Table 2). Forested vegetation only showed evidence of ebullition during the dry season, where ebullition-driven uptake was observed. For forested (short pole) vegetation and *M. flexuosa* palm swamp, net ebullition and ebullition-driven uptake were generally greater during the wet season. Lastly, for mixed palm swamp, net ebullition was greater during the dry season, while ebullition-driven uptake was greater during the dry season.

For the environmental variables, soil pH, DO, EC, water table depth, and soil temperature varied significantly between seasons, whereas air temperature did not. Thus, for the sake of brevity, air temperature is not discussed further here. Mean soil pH was significantly lower during the wet season (5.18 ± 0.03) than during the dry season (5.31 ± 0.04) (data pooled across all vegetation types; *t*-test, $P < 0.05$, Table 3). When disaggregated by vegetation type, the overall trend was found to hold true for all vegetation types except forested (short pole) vegetation, which displayed higher pH during the wet season compared to the dry season (Table 3). A two-way ANOVA on Box–Cox transformed data using vegetation type, season and their interaction as explanatory variables indicated that vegetation type was the best predictor of pH, with season and vegetation type by season playing a lesser role ($F_{7, 1166} = 348.9$, $P < 0.0001$).

For DO, the overall trend was towards significantly lower DO during the wet season ($13.9 \pm 1.0\%$) compared to the dry season ($19.3 \pm 1.2\%$) (data pooled across all vegetation types; Wilcoxon test, $P < 0.0001$, Table 3). However, when the data were disaggregated by vegetation type, we found that individual vegetation types showed distinct seasonal trends from each other. Forested vegetation, forested (short pole) vegetation and mixed palm swamp were consistent with the overall trend (i.e. lower wet season compared to dry season DO), whereas *M. flexuosa* palm swamp displayed the reverse trend (i.e. higher *wet season* compared to *dry season* DO) (Table 3). A two-way ANOVA on Box–Cox transformed data using vegetation type, season and their interaction as explanatory variables indicated that vegetation type was the best predictor of DO, followed by a strong vegetation by season interaction; season itself played a lesser role than

either of the other two explanatory variables ($F_{7, 1166} = 57.0$, $P < 0.0001$).

For EC, the overall trend was towards lower EC in the wet season ($49.4 \pm 1.8 \mu\text{S m}^{-2}$) compared to the dry season ($65.5 \pm 2.2 \mu\text{S m}^{-2}$) (data pooled across all vegetation types; Wilcoxon test, $P < 0.05$, Table 3). When the data were disaggregated by vegetation type, this trend was consistent for all the vegetation types except for forested vegetation, where differences between wet and dry seasons were not statistically significant (Wilcoxon, $P > 0.05$, Table 3).

Water table depths varied significantly between seasons (data pooled across all vegetation types; Wilcoxon test, $P < 0.0001$, Table 3). Mean water table level was significantly higher in the wet (54.1 ± 2.7 cm) than dry (1.3 ± 0.8 cm) seasons. When disaggregated by vegetation type, the trend held true for individual vegetation types (Table 3). All vegetation types had negative dry season water tables (i.e. below the soil surface) and positive wet season water tables (i.e. water table above the soil surface), except for *M. flexuosa* palm swamp that had positive water tables in both seasons. Two-way ANOVA on Box–Cox transformed data using vegetation type, season and their interaction as explanatory variables indicated that all three factors explained water table depth, but that season accounted for the largest proportion of the variance in the model, followed by vegetation by season, and lastly by vegetation type ($F_{7, 1157} = 440.1$, $P < 0.0001$).

For soil temperature, the overall trend was towards slightly higher temperatures in the wet season ($25.6 \pm 0.0^\circ\text{C}$) compared to the dry season ($25.1 \pm 0.0^\circ\text{C}$) (t -test, $P < 0.0001$). Analysis of the disaggregated data indicates this trend was consistent for individual vegetation types (Table 3). Two-way ANOVA on Box–Cox transformed data using vegetation type, season and their interaction as explanatory variables indicated that all three variables played a significant role in modulating soil temperature, although season accounted for the largest proportion of the variance, whereas the other two factors accounted for a similar proportion of the variance ($F_{7, 1166} = 21.3$, $P < 0.0001$).

3.3 Relationships between gas fluxes and environmental variables

To explore the relationships between environmental variables and diffusive gas fluxes, we conducted an analysis of covariance (ANCOVA) on Box–Cox transformed gas flux data, using vegetation type, season, vegetation by season, and environmental variables as explanatory variables. We did not analyse trends between ebullition and environmental variables because of the limitations in the sampling methodology and the limited number of observations.

For diffusive CH_4 flux, ANCOVA revealed that the vegetation by season interaction was the strongest predictor of CH_4 flux, followed by the effect of season and vegetation alone ($F_{13, 933} = 9.7$, $P < 0.0001$). Other significant drivers

included water table depth, soil temperature, and a weak effect of air temperature (only significant at $P < 0.09$). However, it is important to note that each of these environmental variables was only weakly correlated with CH_4 flux even if the relationships were statistically significant. For example, when individual bivariate regressions were calculated, the r^2 values were less than 0.005 for each plot or were not statistically significant (see Supplement Figs. S1 and S2).

For diffusive N_2O flux, ANCOVA indicated that the best predictors of flux rates were electrical conductivity, dissolved oxygen and soil temperature ($F_{13, 1004} = 2.1$, $P < 0.0139$). As was the case for CH_4 , when the relationships between these environmental variables and N_2O flux were explored using individual bivariate regressions, r^2 values were found to be very low (e.g. less than $r^2 < 0.004$) or not statistically significant (see Figs. S3 and S4).

4 Discussion

4.1 Large and asynchronous CH_4 fluxes from peatlands in the Pastaza-Marañón foreland basin

The ecosystems sampled in this study were strong atmospheric sources of CH_4 . Diffusive CH_4 flux, averaged across all vegetation types, was $204.8 \pm 23.5 \text{ mg CH}_4\text{-C m}^{-2} \text{ d}^{-1}$, spanning a range from -471 to $17\,076 \text{ mg CH}_4\text{-C m}^{-2} \text{ d}^{-1}$. This mean falls within the range of diffusive flux observed in other Amazonian wetlands ($7.1\text{--}390.0 \text{ mg CH}_4\text{-C m}^{-2} \text{ d}^{-1}$) (Bartlett et al., 1990, 1988; Devol et al., 1990, 1988), but exceeds the flux observed in Indonesian peatlands ($3.7\text{--}87.8 \text{ mg CH}_4\text{-C m}^{-2} \text{ d}^{-1}$) (Couwenberg et al., 2010). Although the ebullition data must be treated with caution because of the sampling methodology (see below), we observed a mean net ebullition flux of $4625.2 \pm 752.1 \text{ mg CH}_4\text{-C m}^{-2} \text{ d}^{-1}$, spanning a range of 127 to $38\,168 \text{ mg CH}_4\text{-C m}^{-2} \text{ d}^{-1}$. While data on ebullition from Amazonian wetlands are sparse, these values are broadly in line with riverine and lake ecosystems sampled elsewhere (Bastviken et al., 2010; Smith et al., 2000; Sawakuchi et al., 2014). Ebullition-driven CH_4 uptake is not a commonly reported phenomenon in other peatland studies because it is likely an artefact of chamber sampling methods; as a consequence, we do not discuss these data further here. To summarize, these data on diffusive CH_4 flux and ebullition suggest that peatlands in the Pastaza-Marañón foreland basin are strong contributors to the regional atmospheric budget of CH_4 , given that the four vegetation types sampled here represent the dominant cover types in the PMFB (Draper et al., 2014; Householder et al., 2012; Kelly et al., 2014; Lähteenoja and Page, 2011).

The overall trend in the diffusive flux data was towards greater temporal (i.e. seasonal) variability in diffusive CH_4 flux rather than strong spatial (i.e. inter-site) variability. For the pooled dataset, diffusive CH_4 emissions were significantly greater during the wet season than the dry season,

with emissions falling by approximately 21 % from one season to the other (i.e. 236.4 ± 32.2 to 186.3 ± 32.0 mg CH₄–C m^{–2} d^{–1}). This is in contrast to the data on diffusive CH₄ flux among study sites, where statistical analyses indicate that there was a weaker effect of vegetation type on CH₄ flux. For the ebullition data, there was no significant difference among vegetation types or between seasons.

On face value, these data on diffusive CH₄ flux suggest two findings: first, the weaker effect of vegetation type on diffusive CH₄ flux implies that patterns of CH₄ cycling are broadly similar among study sites. Second, the strong *overall* seasonal pattern suggests that – on the whole – these systems conform to our normative expectations of how peatlands function with respect to seasonal variations in hydrology and redox potential, i.e. enhanced CH₄ emissions during a more anoxic wet season (i.e. when water tables rise), and reduced CH₄ emissions during a more oxic dry season (i.e. when water tables fall). However, closer inspection of the data reveals that different vegetation types showed contrasting seasonal emission patterns (Table 4), challenging our basic assumptions about how these ecosystems function. For example, while forested (short pole) vegetation and *M. flexuosa* palm swamp conformed to expected seasonal trends for methanogenic wetlands (i.e. higher wet season compared to dry season emissions), forested vegetation and mixed palm swamp showed the opposite pattern, with significantly greater CH₄ emissions during the dry season. The disaggregated data thus imply that the process-based controls on CH₄ fluxes may vary significantly among these different ecosystems, rather than being similar, leading to a divergence in seasonal flux patterns.

What may explain this pattern of seasonal divergence in CH₄ flux? One explanation is that CH₄ emissions from forested vegetation and mixed palm swamp, compared to the other two ecosystems, may be more strongly transport-limited during the wet season than the dry season. This interpretation is supported by the field data; forested vegetation and mixed palm swamp had the highest wet season water table levels, measuring 110.8 ± 9.3 and 183.7 ± 1.7 cm, respectively (Table 3). In contrast, water table levels for forested (short pole) vegetation and *M. flexuosa* palm swamp in the wet season were 3–7× lower, measuring only 26.9 ± 0.5 and 37.2 ± 1.7 cm, respectively (Table 3). Moreover, a scatter plot of diffusive CH₄ flux against water table depth shows a peak in diffusive CH₄ emissions when water tables are between 30 and 40 cm above the surface, after which CH₄ emissions decline precipitously (Fig. S2). Thus, the greater depth of overlying water in forested vegetation and mixed palm swamp may have exerted a much greater physical constraint on gas transport compared to the other two ecosystems. This interpretation is broadly consistent with studies from other ecosystems, which indicate that high or positive water tables may suppress CH₄ emissions from wetlands above a system-specific threshold (Couwenberg et al., 2010, 2011).

However, transport limitation alone does not fully explain the difference in dry season CH₄ emissions among vegetation types. Forested vegetation and mixed palm swamp showed substantially higher dry season CH₄ emissions (751.5 ± 290.4 and 131.3 ± 31.4 mg CH₄–C m^{–2} d^{–1}, respectively) compared to forested (short pole) vegetation and *M. flexuosa* palm swamp (45.3 ± 12.3 and 97.2 ± 10.3 mg CH₄–C m^{–2} d^{–1}, respectively), pointing to underlying differences in CH₄ production and oxidation among these ecosystems. One possibility is that dry season methanogenesis in forested vegetation and mixed palm swamp was greater than in the other two ecosystems, potentially driven by higher rates of C flow (Whiting and Chanton, 1993). This is plausible given that forested vegetation and mixed palm swamp tend to occur in more nutrient-rich parts of the Pastaza-Marañón foreland basin, whereas forested (short pole) vegetation and *M. flexuosa* palm swamp tend to dominate in more nutrient-poor areas (Lähteenoja et al., 2009a), leading to potential differences in rates of plant productivity and below-ground C flow. Moreover, it is possible that the nutrient-rich vegetation may be able to utilize the higher concentration of nutrients, deposited during the flood pulse, during the Amazonian dry season (Morton et al., 2014; Saleska et al., 2016), with implications for overall ecosystem C throughput and CH₄ emissions. Of course, this interpretation does not preclude other explanations, such as differences in CH₄ transport rates among ecosystems (e.g. due to plant-facilitated transport or ebullition) (Panagala et al., 2013), or varying rates of CH₄ oxidation (Teh et al., 2005). However, these other possibilities cannot be explored further without recourse to more detailed process-level experiments. Forthcoming studies on the regulation of GHG fluxes at finer spatial scales (e.g. investigation of environmental gradients within individual study sites) or detailed diurnal studies of GHG exchange (Murphy et al., 2017) will further deepen our understanding of the process controls on soil GHG flux from these peatlands, and shed light on these questions.

Finally, while the trends described here are intriguing, it is important to acknowledge some of the potential limitations of our data. First, given the uneven sampling pattern, it is possible that the values reported here do not fully represent the entire range of diffusive flux rates, especially for the more sparsely sampled habitats. However, given the large and statistically significant differences in CH₄ emissions between seasons, it is likely that the main trends that we have identified will hold true with more spatially extensive sampling. Second, the data are a conservative underestimate of CH₄ emissions, because the low-frequency, static chamber sampling approach that we utilized was unable to fully capture erratic ebullition events representatively (McClain et al., 2003). Although we attempted to quantify CH₄ ebullition within our static flux chambers, the sampling approach that we utilized was not the best suited for representatively quantifying ebullition. Given the erratic or stochastic nature of ebullition, automated chamber measurements or an inverted

“flux funnel” approach would have provided better estimates of ebullition (Strack et al., 2005). However, we lacked the resources to apply these techniques here. We also did not measure CH₄ emissions from the stems of woody plants, even though woody plants have been recently identified as an important point of atmospheric egress (Pangala et al., 2013). We did not have enough data on floristic composition or individual plant identities within our plots to develop a sampling design that would adequately represent plant-mediated fluxes from our study sites, or the resources to implement a separate study of stem fluxes. Third and last, our data probably underestimate net CH₄ fluxes for the PMFB because we chose to include fluxes with strong negative values (i.e. more than $-10 \text{ mg CH}_4\text{-C m}^{-2} \text{ d}^{-1}$) in our calculation of mean diffusive flux rates. These observations are more negative than other values typically reported elsewhere in the tropical wetland literature (Bartlett et al., 1990, 1988; Devol et al., 1990, 1988; Couwenberg et al., 2010). However, they represent only a small proportion of our dataset (i.e. 13 %, or only 130 out of 997 measurements), and inspection of our field notes and the data themselves did not produce convincing reasons to exclude these observations (e.g. we found no evidence of irregularities during field sampling, and any chambers that showed statistically insignificant changes in concentration over time were removed during our quality control procedures). While headspace concentrations for these measurements were often elevated above mean tropospheric levels ($> 2 \text{ ppm}$), this in itself is not unusual in reducing environments that contain strong local sources of CH₄ (Baldocchi et al., 2012). We did not see this as a reason to omit these values as local concentrations of CH₄ are likely to vary naturally in methanogenic forest environments due to poor mixing in the understory and episodic ebullition events. Importantly, exclusion of these data did not alter the overall statistical trends reported above, and only produced slightly higher estimates of diffusive CH₄ flux (i.e. $251.3 \pm 26.6 \text{ mg CH}_4\text{-C m}^{-2} \text{ d}^{-1}$ versus $204.8 \pm 23.5 \text{ mg CH}_4\text{-C m}^{-2} \text{ d}^{-1}$).

4.2 Western Amazonian peatlands as weak atmospheric sources of nitrous oxide

The ecosystems sampled in this study were negligible atmospheric sources of N₂O, emitting only $7.90 \pm 4.93 \text{ } \mu\text{g N}_2\text{O-N m}^{-2} \text{ d}^{-1}$, suggesting that peatlands in the Pastaza-Marañón foreland basin make little or no contribution to regional atmospheric budgets of N₂O. This is consistent with N₂O flux measurements from other forested tropical peatlands, where N₂O emissions were also found to be relatively low (Inubushi et al., 2003; Couwenberg et al., 2010). No statistically significant differences in N₂O flux were observed among study sites or between seasons, suggesting that these different peatlands may have similar patterns of N₂O cycling. Interestingly, differences in N₂O fluxes were not associated with the nutrient status of the peatland; i.e. more nutrient-rich ecosystems, such as forested vegetation and mixed palm

swamp, did not show higher N₂O fluxes than their nutrient-poor counterparts, such as forested (short pole) vegetation and *M. flexuosa* palm swamp. This may imply that N availability, one of the principal drivers of nitrification, denitrification, and N₂O production (Groffman et al., 2009; Werner et al., 2007), may not be greater in nutrient-rich versus nutrient-poor ecosystems in this part of the western Amazon. Alternatively, it is possible that even though N availability and N fluxes may differ between nutrient-rich and nutrient-poor systems, N₂O yield may also vary such that net N₂O emissions are not significantly different among study sites (Teh et al., 2014).

One potential source of concern are the negative N₂O fluxes that we documented here. While some investigators have attributed negative fluxes to instrumental error (Cowan et al., 2014; Chapuis-Lardy et al., 2007), others have demonstrated that N₂O consumption – particularly in wetland soils – is not an experimental artefact, but occurs due to the complex effects of redox, organic carbon content, nitrate availability, and soil transport processes on denitrification (Ye and Horwath, 2016; Yang et al., 2011; Wen et al., 2016; Schlesinger, 2013; Teh et al., 2014; Chapuis-Lardy et al., 2007). Given the low redox potential and high carbon content of these soils, it is plausible that microbial N₂O consumption is occurring, because these types of conditions have been found to be conducive to N₂O uptake elsewhere (Ye and Horwath, 2016; Teh et al., 2014; Yang et al., 2011).

5 Conclusions

Our data suggest that peatlands in the Pastaza-Marañón foreland basin are strong sources of atmospheric CH₄ at a regional scale, and need to be better accounted for in CH₄ emissions inventories for the Amazon basin as a whole. In contrast, N₂O fluxes were negligible, suggesting that these ecosystems are weak regional sources at best. A divergent or asynchronous seasonal emissions pattern for CH₄ among different vegetation types was intriguing, and challenges our underlying expectations of how tropical peatlands function. These data highlight the need for greater wet season sampling, particularly from ecosystems near river margins that may experience very high water tables (i.e. $> 40 \text{ cm}$). Moreover, these data also emphasize the need for more spatially extensive sampling across both the Pastaza-Marañón foreland basin and the wider Amazon region as a whole, in order to establish whether these asynchronous seasonal emission patterns are commonplace or specific to peatlands in the PMFB region. If CH₄ emission patterns for different peatlands in the Amazon are in fact asynchronous and decoupled from rainfall seasonality, then this may partially explain some of the heterogeneity in CH₄ sources and sinks observed at the basin-wide scale (Wilson et al., 2016).

Data availability. These data are publicly available through the UK Natural Environment Research Council's (NERC's) Centre for Environmental Data Analysis (CEDA), with <https://doi.org/10.5072/a3614fb00ff74999a5187d3a3767d96d> (Teh et al., 2017).

Author contributions. YAT secured the funding for this research, assisted in the planning and design of the experiment, and took the principal role in the analysis of the data and preparation of the manuscript. WAM planned and designed the experiment, collected the field data, analysed the samples, and took a secondary role in data preparation, data analysis, and manuscript preparation. JCB, AB, and SEP supported the planning and design of the experiment, and provided substantive input into the writing of the manuscript. AB in particular took a lead role in developing the maps of our study sites in the PMFB.

Competing interests. The authors declare that they have no conflict of interest.

Acknowledgements. The authors would like to acknowledge the UK Natural Environment Research Council for funding this research (NERC award number NE/I015469). We would like to thank MINAG and the Ministerio de Turismo in Iquitos for permits to conduct this research, the Instituto de Investigaciones de la Amazonía Peruana (IIAP) for logistical support, Peruvian rainforest villagers for their warm welcome and acceptance, Hugo Vasquez, Pierro Vasquez, Gian Carlo Padilla Tenazoa, and Yully Rojas Reátegui for fieldwork assistance, Outi Lähteenoja and Ethan Householder for fieldwork planning, and Paul Beaver of Amazonia Expeditions for lodging and logistical support. Our gratitude also goes to Alex Cumming for fieldwork support and laboratory assistance, and Bill Hickin, Gemma Black, Adam Cox, Charlotte Langley, Kerry Allen, and Lisa Barber of the University of Leicester for all of their continued support. Thanks are also owed to Graham Hambley (St. Andrews), Angus Calder (St. Andrews), Viktoria Oliver (Aberdeen), Torsten Diem (Aberdeen), Tom Kelly (Leeds), and Freddie Draper (Leeds) for their help in the laboratory and with fieldwork planning. Torsten Diem, Viktoria Oliver, and two anonymous referees provided very helpful and constructive comments on earlier drafts of the paper. This publication is a contribution from the Scottish Alliance for Geoscience, Environment and Society (<http://www.sages.ac.uk>, last access: 19 June 2018) and the UK Tropical Peatland Working Group (<https://tropicalpeat.wordpress.com>, last access: 19 June 2018).

Edited by: Ivonne Trebs

Reviewed by: Edzo Veldkamp and one anonymous referee

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