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Comparing soil carbon loss through respiration and leaching under extreme precipitation events in arid and semiarid grasslands

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Abstract. Respiration and leaching are two main processes responsible for soil carbon loss. While the former has received considerable research attention, studies examining leaching processes are limited, especially in semiarid grasslands due to low precipitation. Climate change may increase the extreme precipitation event (EPE) frequency in arid and semiarid regions, potentially enhancing soil carbon loss through leaching and respiration. Here we incubated soil columns of three typical grassland soils from Inner Mongolia and the Qinghai-Tibetan Plateau and examined the effect of simulated EPEs on soil carbon loss through respiration and leaching. EPEs induced a transient increase in CO₂ release through soil respiration, equivalent to 32 and 72 % of the net ecosystem productivity (NEP) in the temperate grasslands (Xilinhot and Keqi) and 7 % of NEP in the alpine grasslands (Gangcha). By comparison, leaching loss of soil carbon accounted for 290, 120, and 15 % of NEP at the corresponding sites, respectively, with dissolved inorganic carbon (DIC, biogenic DIC + lithogenic DIC) as the main form of carbon loss in the alkaline soils. Moreover, DIC loss increased with recurring EPEs in the soil with the highest pH due to an elevated contribution of dissolved CO2 from organic carbon degradation (indicated by DIC- δ^{13} C). These results highlight the fact that leaching loss of soil carbon (particularly in the form of DIC) is important in the regional carbon budget of arid and semiarid grasslands and also imply that SOC mineralization in alkaline soils might be underestimated if only measured as CO₂ emission from soils into the atmosphere. With a projected increase in EPEs under climate change, soil

carbon leaching processes and the influencing factors warrant a better understanding and should be incorporated into soil carbon models when estimating carbon balance in grassland ecosystems.

1 Introduction

Soils store approximately 2500 Pg of carbon (including organic and inorganic carbon) globally, equivalent to 3.3 and 4.5 times the carbon in the atmosphere (760 Pg) and terrestrial plants (560 Pg), respectively (Lal, 2004). Slight variations in the soil carbon pool will hence severely influence atmospheric CO₂ concentrations and have important implications for climate change (Davidson and Janssens, 2006; Trumbore and Czimczik, 2008). Respiration and leaching are two main processes responsible for soil carbon loss. While respiration has received considerable research attention (Raich and Schlesinger, 1992; Raich and Potter, 1995; Hoover et al., 2016; Burri et al., 2015; Escolar et al., 2015), leaching is relatively poorly constrained despite its importance in certain ecosystems (Cole et al., 2007; Battin et al., 2008; Liu et al., 2017). For instance, soil carbon leached from forests, grasslands, and croplands is estimated to be 15.1, 32.4, and 20.5 g C m⁻² yr⁻¹ across Europe, representing 4, 14, and 8 % of net ecosystem exchange (NEE), respectively (Kindler et al., 2011). Additionally, leaching of carbon previously preserved in surface litter and soil layers is believed to be a main source of dissolved organic and inorganic matter

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in inland waters (Spencer et al., 2008). In particular, soil inorganic carbon (SIC) that occurs widely in arid and semiarid regions is more prone to leaching than organic carbon during sporadic high precipitation events (Lal and Kimble, 2000). Despite the importance of leaching loss in regional soil carbon budgets, very little detailed data exists to investigate and compare the relative contribution of respiration and leaching processes to soil carbon loss.

Climate change is reported to increase the frequency as well as the intensity of extreme precipitation events (EPEs; Knapp et al., 2002; Goswami et al., 2006; Parry et al., 2007; Min et al., 2011; Reichstein et al., 2013), especially in arid regions (Donat et al., 2017). In northwestern China, the frequency and intensity of EPEs have shown an increasing trend in the most recent 50 years, constituting a much higher proportion of total precipitation than light precipitation events (Liu et al., 2005; Chen et al., 2012; Wang et al., 2012, 2014; Fu et al., 2013). Increasing EPEs will not only enhance soil carbon leaching but also affect soil respiration processes by increasing soluble substrates for microbial decomposition and potentially inducing hypoxic conditions (Knapp et al., 2002; Harper et al., 2005; Morel et al., 2009; Unger et al., 2010). Hence, it is critical to evaluate the effects of EPEs on soil respiration and leaching processes in order to better understand the impact of climate change on terrestrial carbon cycling, especially in arid and semiarid regions.

Grasslands, containing 20 % of the global soil carbon pool, are the most widespread ecosystems in arid and semiarid regions globally (Jobbagy and Jackson, 2000). The deposition rate of carbonate is relatively high in grassland soils with a high alkalinity and aridity (Lal, 2008; Yang et al., 2012), and hence SIC is the major form of soil carbon in many grasslands (Mi et al., 2008). SIC storage in China is approximately 53.3-77.9 Pg (Li et al., 2007; Mi et al., 2008), 54 % of which is mainly distributed in the temperate and alpine grasslands located in Inner Mongolia and the Qinghai-Tibetan Plateau (Mi et al., 2008). From the 1980s to 2000s, SIC in the topsoil of Chinese grasslands was estimated to decrease by 26.8 g C m⁻² yr⁻¹, mainly attributed to soil acidification (Yang et al., 2012). Alternatively, precipitation is one of the main factors influencing the distribution and storage of SIC in arid and semiarid regions (Batjes, 1998; Lal and Kimble, 2000). Mi et al. (2008) found that 84 % of SIC in China was distributed in areas with a mean annual precipitation (MAP) of <500 mm and that SIC content decreased significantly with the increase in MAP. Given the high leaching potential of SIC in grassland soils under altered precipitation patterns in the future, we hypothesize that EPEs may significantly enhance SIC loss through leaching processes and further reduce SIC storage in grasslands.

In this study, soils were collected from varied depths of three typical temperate and alpine grasslands in Inner Mongolia and the Qinghai–Tibetan Plateau to construct soil columns for a laboratory incubation study. Using simulated EPEs, we examined soil carbon loss through respiration and

leaching processes and compared their fluxes after EPEs. In addition, the leaf litter of a C_4 grass was added to the surface of one set of soil columns to compare soil carbon loss from bare versus litter-covered soils and to estimate the contribution of litter-derived carbon to soil respiration after EPEs. Our research objectives were (1) to investigate the influence of EPEs on soil respiration, (2) to quantify the loss of SIC and soil organic carbon (SOC) through leaching during EPEs, and (3) to compare the relative importance of respiration and leaching in EPE-induced soil carbon loss from grassland soils.

2 Materials and methods

2.1 Study area

For the incubation experiment, soils were collected from three different sites of temperate and alpine grasslands in China with varied environmental characteristics. Temperate grasslands were sampled near Xilinhot (XLHT; 116°22′ E, 44°8′N; mean elevation 1170 m) and Keqi (KQ; 117°15′E, 43° 18′ N; mean elevation 1250 m) within the arid and semiarid regions of Inner Mongolia (Fig. S1 in the Supplement) with MAP of 299 and 402 mm and mean annual temperature (MAT) of 1.2 and 0.4 °C, respectively. Soil in this region is mainly chestnut soil, classified as Calcic Chernozems according to the World Reference Base for Soil Resources (Steffens et al., 2008; IUSS working group WRB, 2015), with Stipa klemenzii, Stipa Goboca, Stipa breviflora, and Stipa glareosa as the dominating species (Sui and Zhou, 2013). The alpine grassland was sampled in Gangcha (GC; 100°7′ E, 37°19′ N; mean elevation 3500 m) located north of the Qinghai Lake on the northeastern edge of the Qinghai-Tibetan Plateau. The GC site has a MAT of 0.4 °C, a MAP of 370 mm, and a mean annual evaporation (MAE) of 607 mm. Soils at this site are mainly Gelic Cambisol (IUSS working group WRB, 2015), with Potentilla ansrina Rosaceae, Elymus nutans Griseb, and Deveuxia arundinacea as the dominant species.

Soils were collected by digging soil pits of $25 \, \mathrm{cm} \times 25 \, \mathrm{cm} \times 70 \, \mathrm{cm}$ from the temperate (XLHT and KQ) and alpine (GC) sites in October 2014 and August 2015, respectively. At each site, three plots $(200 \, \mathrm{m} \times 200 \, \mathrm{m})$ were selected (>200 m in between) with three random soil pits (distance of $\sim 5 \, \mathrm{m}$ in between) sampled within each plot. Soils from the same depth $(0{\text -}20, 20{\text -}40, \text{ and } 40{\text -}60 \, \mathrm{cm})$ of the three soil pits were mixed in situ for each plot, shipped back to the laboratory immediately, and stored at $4 \, ^{\circ}\mathrm{C}$ before the experiment started within 1 month. As a result, each sampling site had three "true" replicates from the field for the soil column experiment.

Table 1. Bulk properties of soil samples collected from the temperate and alpine grasslands for the soil column experiment (mean \pm standard error; n = 3).

Station	Depth	SOC	SIC	Z	SOC: N	Hd	δ^{13} C	FWC	Max. V	BD	Clay	Silt	
	(cm)	(%)	(%)	(%)	ratio		(%0)	(%)	(%)	(g cm 2)	(%)	(%)	(%)
Xilinhot (XLHT)	0-20	$0-20$ 1.48 ± 0.02	0.41 ± 0.01 0.18	0.18 ± 0.00	8.03 ± 0.18	8.98 ± 0.03	-24.1		47.12 ± 0.37	1.06 ± 0.02	0.4	64.6	
	20-40	1.00 ± 0.05	0.64 ± 0.00	0.13 ± 0.00	7.69 ± 0.22	9.09 ± 0.01	-24.1	6.48 ± 0.24	44.92 ± 0.25	1.24 ± 0.05	0.5	58.2	41.3
	40-60	0.67 ± 0.03	1.05 ± 0.01	0.09 ± 0.00	7.09 ± 0.22	9.09 ± 0.04	-23.7	5.56 ± 0.11	39.78 ± 0.39	1.31 ± 0.03	9.0	58.5	0.14 0.0
Keqi (KQ)	0-20	$0-20$ 3.36 ± 0.05	0.02 ± 0.00	0.29 ± 0.00	11.48 ± 0.24	7.79 ± 0.10	-26.0	19.59 ± 0.22	65.57 ± 0.82	1.14 ± 0.03	0.4	41.0	
	20-40	2.52 ± 0.04	0.01 ± 0.00	0.22 ± 0.00	11.59 ± 0.27	7.63 ± 0.04	-25.9	8.56 ± 0.05	53.59 ± 1.98	1.22 ± 0.01	0.2	55.7	44.1
	40–60	1.65 ± 0.03	0.02 ± 0.00	0.14 ± 0.00	11.49 ± 0.42	7.57 ± 0.12	-25.5	8.00 ± 0.27	42.92 ± 0.57	1.19 ± 0.01	0.2	61.6	38.1
Gangcha (GC)	0-20	$0-20$ 3.32 ± 0.23	0.34 ± 0.04	0.31 ± 0.03	10.70 ± 1.28	8.53 ± 0.07	-26.3	33.24 ± 0.68	60.79 ± 0.21	n.d.	1.3	75.9	22.8
	20-40	2.90 ± 0.18	0.44 ± 0.10	0.29 ± 0.01	9.93 ± 0.69	8.60 ± 0.03	-24.0	36.15 ± 0.52	62.03 ± 0.30	n.d.	6.0	75.8	23.3
	40–60	2.12 ± 0.22	0.52 ± 0.06	0.20 ± 0.02	10.55 ± 1.50	8.76 ± 0.10	-25.3	35.79 ± 0.91	62.85 ± 0.61	n.d.	9.0	64.0	35.4
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size < 20 µm; Sand <soil particle water-holding capacity; BD: bulk density; Clay: soil particle size < 0.2 μm; Silt: 0.2 μm maximum soil inorganic carbon; N: nitrogen; FWC: field water content; Max. WHC: particle size > 20 µm; n.d.:

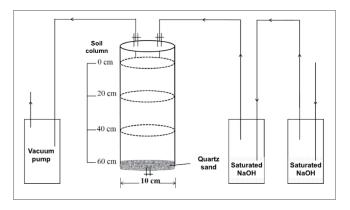


Figure 1. Design of the soil column experiment for monitoring soil respiration and leaching after simulated extreme precipitation events (EPEs).

2.2 Soil column experiment and simulated EPEs

For the laboratory experiment, we reconstructed soil columns of similar structures and texture under controlled conditions and used gravity to collect soil leachates. This approach is commonly used in process-related research (Hendry et al., 2001; Thaysen et al., 2014; Artiola and Walworth, 2009; Aslam et al., 2015) as it minimizes experimental errors and bias caused by unknown factors including soil heterogeneity and microbial community variations. It is also more favorable in terms of quantifying soil carbon leaching loss as it circumvents pore-water contamination by vacuum suction in the field. In particular, leachate sampling by gravity from soil columns prevents alterations to DIC concentrations, which may be caused by CO₂ outgassing using vacuum suction in field studies. Artificial soil columns were constructed in the laboratory with polymethyl methacrylate frames (diameter: 10 cm; height: 70 cm; Fig. 1). The bottom of each column had an aperture (inner diameter: 0.6 cm; height: 3 cm) for the collection of soil leachates, and the column top was fitted with an airtight lid connected to two tubes for gas exchange and collection. Empty columns were soaked in 0.1 mM hydrochloric acid (HCl) solutions for 12 h and rinsed with distilled water before use. Column bottoms were packed with pre-cleaned quartz sand (5 cm thick; soaked in 0.1 mM HCl and combusted at 450 °C for 6 h before use) with a layer of nylon net (pore size: 150 µm; diameter: 10 cm) on both sides to prevent the movement of soil particles. Subsequently, soils were passed through 2 mm sieves with roots removed and packed into each column at the corresponding depths (in the sequence of 40-60, 20-40, and 0-20 cm). Soils were compacted gently to maintain a similar bulk density as in the field (Table 1). The water content of each soil layer was separately adjusted to 60% of the maximum water-holding capacity (Table 1) to provide an ideal moisture condition for microbial growth (Howard and Howard, 1993; Rey et al., 2005).

There was a 10 cm headspace unfilled with soil for each col-

Six soil columns (one litter-amended and one nonamended column for each of the three sampling plots) were set up for each site as described above and pre-incubated for 2 weeks in the laboratory to allow for the recovery of microbial communities after disturbance. Subsequently, the leaf litter of a C4 grass, Cleistogenes squarrosa, a dominant species in the grasslands of northern China (Tian et al., 2015), was added to the surface of three columns in an amount equivalent to the aboveground biomass in the field (1.26 g for the XLHT and KQ sites and 1.59 g for the GC site; Bai et al., 2008). The isotopic signal of the leaf litter $(\delta^{13}$ C of -16.2%) allowed us to estimate the contribution of litter-derived CO₂ to total soil respiration. The columns were pre-incubated again for 7 days. The basal respiration rate was measured by collecting CO2 gas in the column headspace after 4h of incubation. Temperature was recorded every day during the whole incubation period (23 \pm 1 °C).

According to historical precipitation records (Fig. S2), more than 70 % of the annual precipitation occurs from June to August in the study area, mainly in the form of two to four heavy precipitation events. Therefore, a total of three EPEs were simulated over a period of 2 months for each soil using artificial rainwater prepared according to the rainwater composition at the corresponding sites (pH of 7.3; Table S1 in the Supplement; Tang et al., 2014; Zhang et al., 2013). A maximum rainfall intensity of ~ 100 mm per precipitation event has been recorded in the past 2 decades in the study area (Fig. S2) and is predicted to increase by 18.1 % in the late 21st century in north China (Chen et al., 2012). Hence, approximately 1 L of rainwater (rainfall of ~ 127 mm), comparable to 30 % of the MAP of the investigated sites, was added to the surface of each soil column over 3-4 h at rates of one drop per second using syringes and allowed to leach through the column to be collected with a clean beaker within 12-14 h. The leachates were weighed, filtered through a 0.45 µm PTFE syringe filter, and analyzed for dissolved organic carbon (DOC) and dissolved inorganic carbon (DIC) concentrations immediately. To monitor soil respiration every 1–2 days following each EPE, soil columns were first aerated for 1 h using CO₂-depleted air that had been passed through saturated sodium hydroxide (NaOH) solutions (twice; Fig. 1) and then incubated for 4h with lids closed. CO₂ gas in the column headspace was collected by gastight syringes for the subsequent measurement. After the collection of CO₂ gas, the lids were open to allow for exchange with the ambient air. Soil respiration was monitored for 30 days after the first EPE and observed to stabilize approximately on the 20th day (Fig. S3). Hence, the first, second, and third EPEs were conducted on the 1st, 31st, and 51st day of incubation, and the CO₂ measurement was conducted for approximately 30, 20, and 20 days after the first, second, and third EPEs, respectively. Basal respiration was considered to be represented by the stabilized respiration rate at the end of each EPE cycle.

In addition, due to constrained time and logistic reasons, the soil respiration after the second EPE in the KQ soils was not monitored, and the cumulative respiration after the second EPE was calculated as the average respiration after the first and third EPEs in the KQ soils.

2.3 Sample analyses

Soil pH was measured at a soil: water ratio of 1:2.5 (w:v)using a pH meter (Sartorius PB-10). Soil texture was examined by laser diffraction using a Malvern Mastersizer 2000 (Malvern Instruments Ltd., UK) after the removal of organic matter and calcium carbonates. Soil field water content was determined by the difference between moist and dried soils (dried at 105 °C for 8 h). Maximum water-holding capacity was estimated by weighing soils before and after the removal of redundant water from fully soaked soils (in water for 8 h). For SOC analysis, dried soils were decarbonated by exposure to concentrated HCl vapor for 72 h, followed by saturated NaOH solutions for 48 h to neutralize extra HCl, and then dried at 45 °C. Total soil carbon, SOC (after decarbonation), and nitrogen (N) contents were measured by combustion using an elemental analyzer (Vario EL III; Elementar, Hanau, Germany). SIC was calculated as the difference between total carbon and SOC contents. Small aliquots of the soil leachates were analyzed immediately on a Multi N/C 3100-TOC/TN Analyzer (Analytik Jena, Germany) for DIC and DOC concentrations (with the latter acidified to pH<2 with concentrated HCl before analysis). It should be mentioned that the DIC concentration may vary due to exchanges between dissolved and atmospheric CO₂ during leachate collection. However, the potential contribution from this process was < 7% owing to the low proportions of dissolved CO₂ in total DIC of our samples (Table S2) as calculated according to Ran et al. (2015). CO₂ concentration in the soil column headspace was determined by a gas chromatograph (Agilent 7890A, USA) coupled with a flame ionization detector (FID).

To examine the contribution of SOC- and litter-derived carbon to soil respiration, the $\delta^{13}C$ values of SOC and CO₂ gas were determined on an isotope ratio mass spectrometer (Deltaplus XP; Thermo, Germany) with a precision of $\pm 0.2\%$. To estimate the contribution of SOC degradation to leached DIC, the $\delta^{13}C$ values of DIC were determined on a Picarro isotopic CO₂ analyzer equipped with an automated DIC sample preparation system (AutoMate) based on a wavelength-scanned cavity ring-down spectroscopy technique (Picarro AM-CRDS, USA). The precision for the DIC- $\delta^{13}C$ measurement was $\pm 0.3\%$. Due to budget constraints and logistic reasons, we only measured the $\delta^{13}C$ of the respired CO₂ in the GC soils during the first EPE and the leached DIC in the XLHT soils.

2.4 Data analysis and statistics

The relative contribution of litter- and SOC-derived CO₂ to total respired CO₂ in the litter-amended soils was estimated using the following mass balance model:

$$f_{\text{litter-derived}} + f_{\text{SOC-derived}} = 1,$$
 (1)

 $f_{\text{litter-derived}} \times \delta^{13} C_{\text{litter-derived}} + f_{\text{SOC-derived}}$

$$\times \delta^{13}C_{SOC\text{-derived}} = \delta^{13}C_{respired\text{-CO}_{2}}, \tag{2}$$

where $f_{\text{litter-derived}}$ and $f_{\text{SOC-derived}}$ are the proportion of litter- and SOC-derived CO₂ in the total respired CO₂; $\delta^{13}C_{\text{litter-derived}}$ is the $\delta^{13}C$ value of litter-derived CO₂, equivalent to -16.25%; $\delta^{13}C_{\text{SOC-derived}}$ is the $\delta^{13}C$ value of SOC-derived CO₂, which assumes the same value as that in the non-amended soils at the beginning of incubation (-23.1%) according to Cerling et al. (1991); and $\delta^{13}C_{\text{respired-CO}_2}$ is the measured $\delta^{13}C$ of respired CO₂.

Similarly, the relative contribution of lithogenic carbonate and biogenic DIC derived from SOC degradation to leached DIC was assessed according to the following isotopic mass balance model:

$$f_{\text{carbonate}} + f_{\text{biogenic-DIC}} = 1,$$
 (3)

 $f_{\rm carbonate} \times \delta^{13} C_{\rm carbonate} + f_{\rm biogenic\text{-}DIC}$

$$\times \delta^{13} C_{\text{biogenic-DIC}} = \delta^{13} C_{DIC}, \tag{4}$$

where $f_{\text{carbonate}}$ and $f_{\text{biogenic-DIC}}$ are the proportion of carbonate and biogenic DIC in total DIC; $\delta^{13}C_{carbonate}$ is the δ^{13} C value of soil carbonate, equivalent to 0% (Edwards and Saltzman, 2016); and $\delta^{13}C_{\text{biogenic-DIC}}$ is the $\delta^{13}C$ value of biogenic carbonate and bicarbonate derived from the dissolution of CO₂ produced by SOC degradation, which is estimated to shift by approximately 8 % compared with the δ^{13} C value of soil-respired CO₂ (-24 ‰ here) due to isotope fractionation during CO₂ dissolution (Zhang et al., 1995). Hence, δ^{13} C_{biogenic-DIC} is estimated to be -16%. The δ^{13} C_{DIC} value is the measured δ^{13} C signature of leached DIC. The isotopic fractionation of leached DIC due to CO2 loss in an open system is insignificant when the partial pressure of CO_2 (pCO_2) in the solution is lower than twice that of the surrounding atmosphere (Hendy, 1971; Doctor et al., 2008). In the present study, pCO₂ in the XLHT leachates was low ($\sim 400 \,\mu atm$ assuming alkalinity equals to DIC concentration; Table S2) due to its high pH, low soil respiration, and dilution of dissolved CO₂ under an EPE. Thus, we considered the influence of CO₂ outgassing on the δ^{13} C of leached DIC to be negligible.

EPE-induced CO_2 release via respiration was assessed following two steps. First, cumulative respiration during the first 20 days after each EPE (until respiration rate stabilized) was calculated. Second, the difference between the measured cumulative respiration and that estimated using the stabilized basal respiration rate after each EPE was calculated as the EPE-induced CO_2 release.

Independent sample T tests (group size = 2) and one-way ANOVA analysis (group size > 2) were used to compare the dissolved carbon concentrations and fluxes among different columns. Linear regression analysis was used to assess correlations between leachate carbon flux and influencing factors (carbon content, soil pH, soil texture, etc.). All these analyses were performed using IBM SPSS Statistics 22. Differences and correlations are considered to be significant at a level of p < 0.05.

3 Results and discussion

3.1 Bulk properties of grassland soil samples

In the investigated grassland soils, SOC represented 59-99 % of soil carbon and exhibited δ^{13} C values typical of C₃ plant inputs (ranging from -24.1 to -26.3%; Table 1). The XLHT soil had much lower SOC and nitrogen (N) contents than the KQ and GC soils despite a similar soil texture (p < 0.05; Table 1). The SOC: N ratio was also lowest in XLHT (7.09-8.03), indicating a more decomposed state of soil organic matter (Weiss et al., 2016). Conversely, the SIC content was highest in XLHT and lowest in KQ, in line with soil pH variations at these sites, i.e., lowest pH in KQ and highest in XLHT. This correlation of SIC with soil pH is consistent with the results of Shi et al. (2012), showing that pH is the most important factor controlling SIC variation across the Mongolian and Tibetan grasslands. In terms of depth variations, soils became coarser with depth in XLHT and GC, but became finer with increasing depth in KQ. The SOC and N contents decreased with depth in all soils due to declining plant inputs (p < 0.05; Table 1), while the SOC: N ratio remained relatively similar (except a small decrease with depth in XLHT). By contrast, XLHT and GC soils showed an increasing SIC content with depth (p < 0.05; Table 1) because SIC, with a good solubility, is prone to leaching from the topsoil and subsequently precipitates in the deeper soil (Mi et al., 2008; Tan et al., 2014). The KQ soil, showing an almost neutral pH, had an invariant SIC content and pH with depth. Overall, the varied properties (including SOC, SIC, pH, etc.) of these soils allowed us to compare the effects of EPEs on soil respiration and leaching processes in different grassland soils.

3.2 EPE-induced changes to soil respiration

Shortly after each simulated EPE, soil respiration was similar to or lower than basal respiration (Fig. S3). The latter case may be attributed to hypoxic conditions induced by water saturation during EPEs (Hartnett and Devol, 2003; Jessen et al., 2017). Subsequently, soil respiration increased and peaked after approximately 1 week due to the recovery of microbial activity with improved soil aeration (Borken and Matzner, 2009). It then decreased to a constant level approximately 20 days after each EPE (Fig. S3). The transient increase

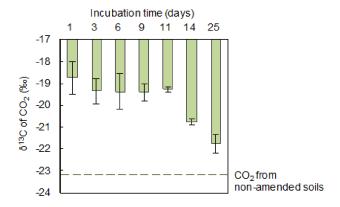


Figure 2. The δ^{13} C values of respired CO₂ in the litter-amended Gangcha (GC) soils after the first extreme precipitation event (EPE). Mean values are shown with standard error (n = 3).

in respiration was consistent with the "Birch effect" (Birch, 1964), i.e., a pulse of soil respiration after rewetting events due to resuscitation of microorganisms and improved diffusive transport of substrate and extracellular enzymes (Borken and Matzner, 2009; Navarro-García et al., 2012; Placella et al., 2012). The maximum soil respiration rates were 40.6 and 37.3 mg C m⁻² h⁻¹ after EPEs in the non-amended KQ and GC soils, respectively. These rates were significantly higher than in the XLHT soil (13.7 mg C m⁻² h⁻¹), likely related to the higher SOC content in the former soils. The maximum specific soil respiration rates normalized to SOC were 2.2, 2.6, and 2.0 μ g C g⁻¹ SOC h⁻¹ in the non-amended GC, KQ, and XLHT soils, respectively. Therefore, SOC degradability was quite similar in the alpine and temperate grassland soils.

Total respired CO₂ was higher in the litter-amended than non-amended soils before and after EPEs (Fig. S6). The cumulative respired CO₂ amounts in the litter-amended XLHT, KQ, and GC soils were 16.7, 54.8, and $44.6 \,\mathrm{g} \,\mathrm{C} \,\mathrm{m}^{-2}$ during three EPEs, which is 20, 22, and 15 % higher than the non-amended soils, respectively. Due to the wide presence of litter coverage in our studied soils, the litter effect on soil respiration should be considered when estimating carbon budgets for these grassland soils. The higher total respired CO₂ in litter-amended soils might be caused by one or both of the following: (1) the degradation of labile components in the fresh litter and/or (2) induced priming effects due to the addition of an easily available energy source (Fröberg et al., 2005; Ahmad et al., 2013). To distinguish the influences of the above two factors on total respired CO2 and further differentiate the contribution of litter (C₄) and SOC (C₃) to the respired CO₂, we examined the δ^{13} C values of CO₂ evolved from the GC soils after the first EPE. On the first day after EPE, CO2 from the non-amended and litteramended GC soils had a δ^{13} C value of -23.1 and -18.7%, respectively. The latter was close to the δ^{13} C signature of the added litter (-16.25%). Using the two-endmember mixing model of Eqs. (1) and (2), we calculated that litter contributed $\sim\!64\,\%$ of the respired CO_2 in the litter-amended GC soils. However, along with the consumption of labile carbon in litter, the $\delta^{13}C$ signature of CO_2 decreased from $-18.7\,\%$ on day 1 to $-21.8\,\%$ on day 25 after an EPE in the litter-amended soils (Fig. 2). Accordingly, the proportion of litter-derived CO_2 decreased from 64 to 20 %. The litter-derived CO_2 flux in litter-amended GC soils was estimated to range from 7.0 to 17.5 mg C m $^{-2}$ h $^{-1}$, while the SOC-derived CO_2 flux increased from 6.2 to 15.7 mg C m $^{-2}$ h $^{-1}$ after the first EPE (Fig. S4). Compared with the SOC-derived CO_2 flux in non-amended GC soils (ranging from 17.2 to 27.1 mg C m $^{-2}$ h $^{-1}$), litter addition had a negative priming effect on the degradation of native SOC while increasing total respiration through labile litter degradation.

Using the data shown in Figs. S3 and S5, we calculated that total EPE-induced CO2 release during three EPEs was higher in the KQ and GC soils than in the XLHT soil (p < 0.05; Fig. 3a) with a lower SOC content and a lower SOC: N ratio (Table 1). However, the specific EPE-induced CO₂ release normalized to SOC content showed no significant difference in the non-amended soils among the three sites (Fig. 3b), indicating that a similar proportion of SOC $(\sim 4\%)$ was subject to EPE-induced CO₂ release in the alpine and temperate grassland soils (Fig. 3b). The total EPEinduced CO₂ release was significantly higher in the litteramended KQ soils than the non-amended ones. Besides the availability of labile OC provided by litter, the higher total EPE-induced CO₂ in litter-amended KQ soils might be related to its relatively lower soil pH (\sim 7.7) that facilitates the release rather than the dissolution of respired CO₂ (from both SOC and litter mineralization) in soil solution. In addition, KQ has the highest mean sand content (46.9%) among the three soils (Table 1), i.e., the least possible mineral protection on labile OC dissolved from the litter, and this benefits the transport and mineralization of labile OC that meanwhile might induce positive priming effects on the SOC mineralization. We therefore conclude that the KQ soil, with a coarser texture and a lower pH (Table 1), may have provided less sorptive protection for the labile DOC components after EPEs (Kell et al., 1994; Nelson et al., 1994), allowed less dissolution of the respired CO₂, and hence showed a more responsive respiration to the precipitation events. Consequently, we deduced that the availability of labile organic carbon, soil texture, and pH are important factors influencing the total EPE-induced CO₂ release in temperate and alpine grassland soils.

3.3 EPE-induced leaching of soil carbon

During the first EPE, a total of 0.57, 0.56, and 0.73 L of leachates were collected from the XLHT, KQ, and GC soils, respectively. The leachate increased to 0.71, 0.94, and 0.87 L during the second EPE and was 0.69, 0.83, and 0.89 L during the third EPE, respectively (Fig. 4). Soil water content

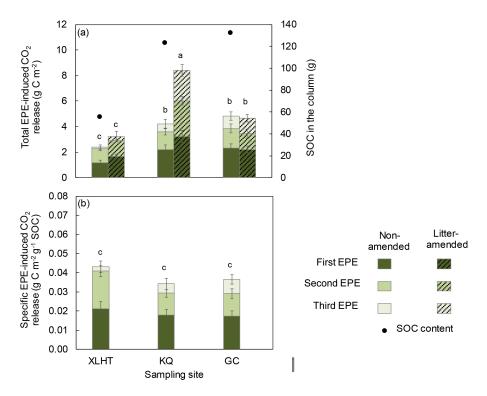


Figure 3. Total (a) and specific (b) EPE-induced CO₂ release in the litter-amended and non-amended grassland soils during three EPEs. Mean values are shown with standard deviation (n = 3). Lowercase letters (a, b, c) indicate significantly different levels among the litter-amended and non-amended soils determined by Duncan's multiple range test (one-way ANOVA, p < 0.05).

was set to $\sim 60\%$ of maximum WHC before the first EPE, and leaching did not occur until soil water reached saturation. Therefore, the leachate volume was lowest during the first EPE and similar for the second and third EPEs. There were some variations in the volume of leachates from different soils, possibly related to preferential flows created during EPEs in the soil columns (McGrath et al., 2009) and water evaporation between EPEs. DIC was the main form of carbon in the leachates from the alkaline soils with a high SIC content (XLHT and GC) but low from the KQ soil with a neutral pH and low SIC content (Fig. 4). The resulting DIC flux was much higher for the XLHT soils ($\sim 21.3 \,\mathrm{g} \,\mathrm{C} \,\mathrm{m}^{-2}$) than the other two $(2.9 \text{ g C m}^{-2} \text{ for KQ and } 7.4 \text{ g C m}^{-2} \text{ for}$ GC soils) during three EPEs, equivalent to 5 times its DOC flux $(3.8-4.2 \text{ g C m}^{-2}; \text{ Fig. 4})$. In contrast, DIC flux in the KQ soils was only one-third of its DOC flux during EPEs. The form of leached carbon was mainly linked to the amount of SOC and SIC in the columns (shown in Fig. S5).

Litter amendment did not increase DOC fluxes in any of the investigated soils but increased DIC fluxes leached from the KQ soil during the second and third EPEs and from the GC soil during the second EPE (p < 0.05; Fig. 4b–c). We postulate that while litter contribution to DOC was minor, CO₂ derived from litter degradation contributed to dissolved CO₂ in soils and hence increased DIC in the leachates (Monger et al., 2015). This effect was not evident during the first EPE

when litter decomposition just started and was not significant for the third EPE in the GC soil due to a high sample variability associated with the litter-amended soil (Fig. 4c). Due to the high SIC content in the XLHT soils (38.15 g per column) and the low litter OC amendment (0.7 g per column), there was no significant difference in DIC fluxes between the non-amended and litter-amended XLHT soils (Fig. 4a). However, for the KQ soil having a relatively low SIC content similar to the added litter OC (0.7 g per column; Table 1), litter amendment had a significant effect on the DIC flux (p < 0.05), increasing by 21 ± 13 and 15 ± 7 % relative to the non-amended KO soils during the second and third EPEs, respectively. There was also a $30 \pm 19\%$ increase in the DIC flux from the litter-amended GC soils relative to its non-amended counterpart during the second EPE. Therefore, litter amendment had a significant influence on DIC fluxes from soils with a relatively low SIC content (KQ and GC) under EPEs compared with the high-SIC XLHT soil.

Between different EPEs, leachate DOC fluxes did not vary in any of the investigated soils. By comparison, DIC fluxes increased in the XLHT soil from 4.5 g C m⁻² after the first EPE to 9.0 g C m⁻² after the third EPE (p<0.01; Fig. 4). This increase may be caused by (i) an increased contribution of SOC degradation to soil DIC and/or (ii) an elevated dissolution of soil carbonates induced by higher soil CO₂ concentrations with repeated EPEs (Gulley et al., 2014; Ren et

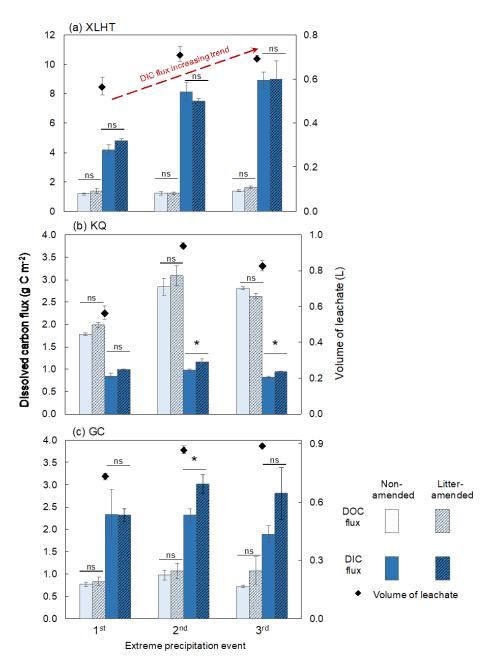


Figure 4. Fluxes of dissolved organic carbon (DOC) and dissolved inorganic carbon (DIC) and volume of leachates from soil columns after extreme precipitation events (EPEs). Mean values are shown with standard error (n = 3). The * and "ns" denote significant and no difference between the litter-amended and non-amended soils determined by independent sample T tests, respectively (p < 0.05).

al., 2015). To evaluate these contributions, the δ^{13} C values of DIC were measured for the non-amended XLHT soil. The δ^{13} C of leached DIC ranged from -10.0 to -6.6% during the first EPE. Based on the isotopic mass balance of Eqs. (3) and (4), lithogenic carbonate (with a δ^{13} C value of 0%) contributed 51% to the leached DIC, while biogenic DIC produced by SOC degradation contributed 48% (Fig. 5). The δ^{13} C value of leached DIC decreased to -12.3 and -13.5% during the second and third EPEs, corresponding to a contribution of 77 and 84% by biogenic sources in the total DIC,

respectively (Fig. 5). These results confirm our previous hypothesis that SOC decomposition contributed significantly to soil DIC fluxes. Combined with the total flux rate, we calculated that both lithogenic and biogenic DIC fluxes were $\sim 2.1~g~C~m^{-2}$ in the first EPE. Subsequently, lithogenic DIC flux decreased to $\sim 1.3~g~C~m^{-2}$, while biogenic DIC flux increased to $7.6~g~C~m^{-2}$ in the third EPE. This demonstrates that the increased DIC flux with repeated EPEs was mainly derived from an increased contribution of SOC mineralization. Interestingly, increasing DIC fluxes with repeated EPEs

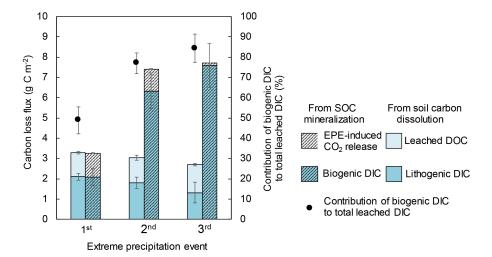


Figure 5. Carbon loss fluxes from soil organic carbon (SOC) mineralization in the non-amended XLHT soils. Fluxes include EPE-induced CO_2 release and leaching of biogenic dissolved inorganic carbon (DIC), dissolved organic carbon (DOC), and lithogenic DIC. Mean values are shown with standard error (n = 3).

were not observed in the KQ and GC soils (Fig. 4) despite their higher SOC contents (Table 1) and CO₂ release rates (Fig. S3). Given that the XLHT soil had the highest soil pH, the high alkalinity may have favored the retention of respired CO₂ in the soil solution compared with the other soils (Parsons et al., 2004; Yates et al., 2013; Liu et al., 2015), leading to its high contribution to DIC fluxes.

Regardless of its source, the EPE-induced leaching loss of inorganic carbon was 31.5 and 10.6 μ g DIC g⁻¹ soil from the alkaline XLHT and GC soils, respectively, which is approximately 3 and 5 times higher than the corresponding DOC leaching loss (5.9 and 3.9 µg DOC g⁻¹ soil, respectively). However, the KQ soil had a relatively lower EPE-induced DIC loss $(4.4 \,\mu g \, DIC \, g^{-1} \, soil)$ than the DOC leaching loss $(11.6 \,\mu g \, DOC \, g^{-1} \, soil)$ mainly due to its lower initial SIC content and relatively neutral soil pH value. Hence, total DIC (biogenic DIC + lithogenic DIC) was the main form of soil carbon loss in alkaline soils during EPEs. When the source of the leached DIC is taken into account, the dissolution of CO₂ produced by SOC mineralization (biogenic DIC) constituted more than half of the leached DIC (at least from the XLHT soils; Fig. 5), whose contribution increased with recurring EPEs (Fig. 5). This implies that SOC mineralization during the three EPEs was underestimated by approximately a factor of 8 when measured as CO2 gas flux from the soil into the column headspace only (Fig. 5). In addition, DIC loss exclusively resulting from SIC dissolution or weathering was also a significant fraction of soil carbon loss, equivalent to 219 % SOC loss in the form of EPE-induced CO₂ during EPEs (Fig. 5). These results collectively corroborate the evidence that inorganic carbon loss is the main form of soil carbon loss in alkaline soils during EPEs.

As for the influencing factors on soil carbon leaching loss, the DIC flux was positively correlated with the amount of SIC in the soil columns and soil pH (p < 0.05; Fig. 6a-b). These two relationships may be self-correlated due to a positive relationship between soil pH and SIC (Liu et al., 2016). By comparison, DOC flux was linked with the amount of SOC in the soil columns, but decreased with an increasing content of silt and clay (p < 0.05; Fig. 6c). This may be explained by the stronger retention of SOC on small-sized particles with more sorption sites (Barré et al., 2014; Mayer, 1994). Interestingly, neither DOC nor DIC fluxes showed any significant relationships with the volume of leachates during EPEs (Fig. 6e-f). This indicates that we used a sufficient amount of precipitation in this study to "scavenge" dissolved carbon from soils, and hence these fluxes represent the soil carbon leaching potential under EPEs. Overall, total soil carbon loss through leaching under EPEs was positively related to soil pH values (p < 0.05; Fig. 6d), suggesting that soil pH is a critical factor determining the magnitude of soil carbon loss under EPEs.

3.4 Main pathways of grassland soil carbon loss under EPEs

In this study, EPE-induced soil carbon loss was composed of three parts: leachate DIC including lithogenic and biogenic DIC, leached DOC, and EPE-induced CO₂ emission into the column headspace. Total DIC and DOC fluxes accounted for 90, 62, and 68% of EPE-induced total loss at XLHT, KQ, and GC, respectively, representing the major pathway of soil carbon loss in these grassland soils under EPEs. Soil carbon leaching fluxes were 25.3, 10.4, and 10.1 g C m⁻² yr⁻¹ in XLHT, KQ, and GC soils during three EPEs, respectively, with DIC as the dominant form in XLHT

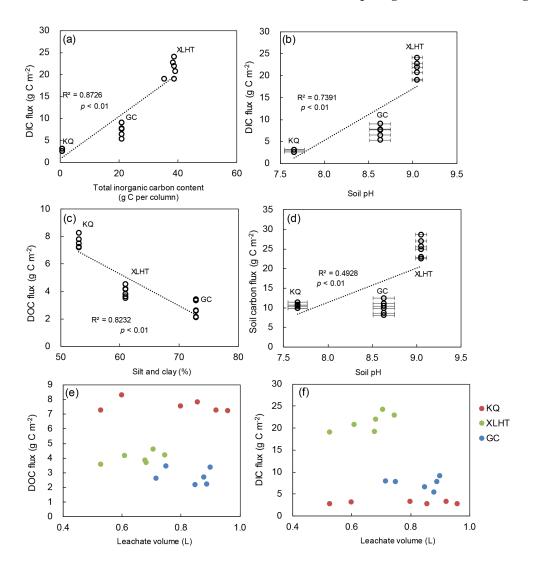


Figure 6. Relationship of dissolved inorganic carbon (DIC) and dissolved organic carbon (DOC) fluxes with soil properties: (a) DIC flux with total inorganic carbon in the soil columns; (b) DIC flux with soil pH; (c) DOC flux with silt and clay content of soils; (d) total soil carbon flux with soil pH; (e) DOC flux with leachate volume; (f) DIC flux with leachate volume. Mean pH values are shown with standard error (n = 3).

and GC soils. While DIC fluxes found for the KQ and GC soils generally fell within the range reported for grassland soils (1.3–47.8 g C m⁻² yr⁻¹; Parfitt et al., 1997; Brye et al., 2001; Kindler et al., 2011), the XLHT soil had a DIC flux higher than the majority (>50%) of the reported values (Fig. 7). This may be attributed to the higher SIC content and stronger dissolution of respired CO₂ in the XLHT soils due to its higher soil pH (9.1 \pm 0.1) relative to other grassland soils (pH: 5.4–7.5; Kindler et al., 2011) and the high intensity of our simulated EPEs (precipitation: 40 mm h⁻¹). Nonetheless, the DIC fluxes in grassland soils reported in this study and elsewhere (Brye et al., 2001; Kindler et al., 2011) were significantly higher than in forest and cropland ecosystems (p<0.05; Rieckh et al., 2014; Lentz and Lehrsch, 2014; Gerke et al., 2016; Herbrich et al., 2017; Siemens et al., 2012;

Walmsley et al., 2011; Wang and Alva, 1999; Kindler et al., 2011), highlighting the importance of leaching as a major pathway of soil carbon loss in grasslands. By contrast, DOC fluxes in this study $(4.8 \pm 2.5 \, \mathrm{g \, C \, m^{-2}})$ were lower than most of the reported values in forest and grassland ecosystems due to the low SOC contents in our soils (Fig. 7).

Net ecosystem production (NEP) in the temperate steppe of Inner Mongolia (XLHT and KQ) is $8.7 \,\mathrm{g}$ C m⁻² yr⁻¹ (Sui and Zhou, 2013). While the EPE-induced CO₂ release (2.8 ± 0.6 and $6.3 \pm 3.0 \,\mathrm{g}$ C m⁻²) accounted for 32 and 72 % of the NEP at XLHT and KQ, respectively, soil carbon leached during three EPEs was equivalent to 290 and 120 % of NEP, with total DIC loss accounting for 244 and 33 %, respectively. It is worth mentioning that biogenic DIC loss ($16.0 \pm 3.4 \,\mathrm{g}$ C m⁻²) caused by SOC degradation ac-

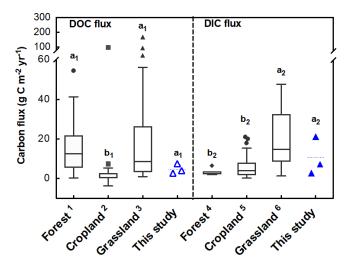


Figure 7. Leaching fluxes of dissolved organic carbon (DOC) and dissolved inorganic carbon (DIC) in this study compared with those reported in the literature. 1 n = 110; data from Brooks et al. (1999), Fröberg et al. (2005, 2006, 2011), Gielen et al. (2011), Kindler et al. (2011), Lu et al. (2013), Michalzik et al. (2001), Sanderman et al. (2009). 2 n = 33; data from Brye et al. (2001), Kindler et al. (2011), Siemens et al. (2012), Walmsley et al. (2011), Wang and Alva (1999), Gerke et al. (2016), Herbrich et al. (2017), Rieckh et al. (2014), Lenz (2014). 3 n = 46; data from Brooks et al. (1999), Brye et al. (2001), Ghani et al. (2010), Kindler et al. (2011), McTiernan et al. (2001), Parfitt et al. (2009), Sanderman et al. (2009), Tipping et al. (1999). $^4 n = 8$; data from Kindler et al. (2011). $^5 n = 32$; data from Kindler et al. (2011), Siemens et al. (2012), Walmsley et al. (2011), Wang and Alva (1999), Gerke et al. (2016), Herbrich et al. (2017), Rieckh et al. (2014), Lenz (2014). 6 n = 9; data from Brye et al. (2001), Kindler et al. (2011). Lowercase letters (a_1 , b_1 and a_2, b_2) represent significant difference levels of DOC and DIC fluxes, respectively, in different ecosystems determined by Duncan's multiple range test (one-way ANOVA, p < 0.05). Dashed lines represent mean values for the investigated soils.

counted for 184 % of NEP at XLHT, indicating the importance of biogenic DIC to leached inorganic carbon loss during EPEs. By comparison, NEP in the studied alpine grassland $(68.5 \text{ g C m}^{-2} \text{ yr}^{-1}; \text{ Fu et al., } 2009)$ is much higher than in typical temperate steppe. Hence, soil carbon loss through leaching and respired CO₂ release accounted for 15 % (DIC: 11%, DOC: 4%) and 7% of the NEP at GC, respectively. Nonetheless, the EPE-induced soil carbon loss relative to NEP was higher in this study than that estimated for grassland topsoil across Europe (12 % for DIC loss, 2 % for DOC loss; Kindler et al., 2011) where the net ecosystem exchange (NEE) reported by Kindler et al. was used as NEP according to the report of Kirschbaum et al. (2001). This was partially attributed to the lower NEP and higher SIC content in XLHT and KQ soils, underscoring the fact that soil carbon leaching is more important in fragile ecosystems with low productivity.

The uncertainty related to the importance of leaching processes in the overall carbon budget along the "soil-riverocean" continuum is in the ultimate downstream fate of the leached carbon. If part of this carbon is retained in the surrounding soils or carried along from the river to the ocean in the form of DIC without outgassing into the air, it will not constitute a source of atmospheric CO₂ on a relatively short term (over years or decades). However, soil columns used in our study have a depth (60 cm) typical of or even deeper than the average soil depth in the alpine grasslands of the Qinghai-Tibetan Plateau (Wang et al., 2001). Hence, we assume that the carbon leached in our experiments will have minimum retention in the soil. Furthermore, compared to DOC and DIC in the soil solution, the leached carbon is more likely to be subject to more intensified mineralization and outgassing during the land-ocean transfer given more intensified mixing processes, oxygen exposure, and photooxidation of terrestrial carbon upon releasing into the river (Hedges et al., 1997; Battin et al., 2009). Hence, we postulate that carbon leached from soils is more vulnerable to decomposition and/or release compared to that retained in the soil. That being said, it will be necessary to confirm our results and hypothesis using field-based leaching experiments to better understand the ultimate fate of leached soil carbon and whether it will be retained in the deeper soil or show higher degradability upon leaving the soil matrix. Such information will be complementary to our study and further elucidate the importance of leaching processes in terms of ecosystem carbon budget.

In summary, this study quantified and compared soil carbon loss through respired CO₂ release and leaching in three typical grassland soils of northern China under simulated EPEs. Soil CO₂ release was stimulated shortly after each EPE, leading to an EPE-induced CO₂ release equivalent to 32 and 72 % of the NEP at XLHT and KQ (temperate grasslands) and 7 % at GC (alpine grassland). By comparison, total soil carbon leaching fluxes accounted for 290, 120 and 15% of the NEP at XLHT, KQ, and GC, respectively, with DIC as the main form of carbon loss in the SIC-enriched XLHT and GC soils. In view of DIC sources, biogenic DIC loss derived from SOC mineralization contributed to more than half of the total leached DIC fluxes and accounted for 184 % of the NEP at XLHT. Moreover, DIC loss increased with recurring EPEs in the XLHT soil with the highest pH due to the increased dissolution of soil carbonates and an elevated contribution of dissolved CO₂ from SOC degradation. These results also imply that SOC mineralization in alkaline grassland soils during EPEs might be underestimated if measured only as CO₂ emission from soil into the atmosphere. Admittedly, our results are based on artificial soil columns that destroyed natural soil structures, hence potentially increasing the contact between pore water and soil particles through changing soil porosity. Also, soil water content was set to \sim 60 % of maximum WHC initially in our experiment, which is higher than that in the field of temperate grasslands

(XLHT and KQ). Thus, our measured DOC and DIC fluxes are likely to be higher than carbon leaching in the field due to greater water retention in drier soils. Hence, our estimate may represent an upper limit of soil carbon leaching potential under EPEs. Nonetheless, these results highlight the fact that leaching loss of soil carbon, especially in the form of DIC originated from biogenic and lithogenic carbonates, plays an important role in the regional carbon budget of grasslands located in arid and semiarid regions. Further research effort is needed to combine short-term laboratory experiments with long-term field measurements to fully assess the impacts of EPEs on the soil carbon budget in these areas. In addition, with a projected increase in EPEs under climate change, soil carbon leaching processes and the influencing factors warrant a better understanding and should be incorporated into soil carbon models when estimating carbon balance in grassland ecosystems.

Data availability. All data are available within this paper (Table 1) and in the Supplement (Dataset S1).

The Supplement related to this article is available online at https://doi.org/10.5194/bg-15-1627-2018-supplement.

Competing interests. The authors declare that they have no conflict of interest.

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