



*Supplement of*

**An analysis of the variability in  $\delta^{13}\text{C}$  in macroalgae from the Gulf of California: indicative of carbon concentration mechanisms and isotope discrimination during carbon assimilation**

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20 **Table List**

21 Table S1. The signal  $\delta^{13}\text{C}$  values in macroalgae of the Gulf of California. The asterisk in  
22 the specie's name means that specie is endemic or its origin is in the Gulf of California,  
23 according to Espinoza-Avalos, (1994). Strategies of dissolved inorganic carbon use are  
24 follows the classification by Díaz-Pulido e al., (2016) based on  $\delta^{13}\text{C}$  signatures: Strategy 1:  
25  $\delta^{13}\text{C} > 10\text{‰}$  (CCM-only (i.e.,  $\text{HCO}_3^-$  use only)); strategy 2:  $\delta^{13}\text{C} = -30$  to  $-10\text{‰}$  ( CCM  
26 activity by mix of  $\text{HCO}_3^-/\text{CO}_2$ ); strategy 3:  $< 30\text{‰}$  (Non-CCM:  $\text{CO}_2$  uptake by diffusion  
27 only); strategy 4: Calcifiers from three phyla, possibly with different carbon-use strategies  
28 related to different modes of calcification. Vertical habitat: S=Subtidal and Intertidal are  
29 divided in Rp= Rockpool, Ee= Eventually exposed. Coastal sectors: Continental coast: C1:  
30 south, C2: Central, C3: north and peninsula coast: P1: south, P2: Central and P3: north.  
31 Season: Rain (from june to october) and Dry (from november to may).

32 Table S2. Comparison of  $\delta^{13}\text{C}$  values in macroalgae species of the Gulf of California in  
33 different seasons and coastal regions.

34 Table S3. Comparison of  $\delta^{13}\text{C}$  values in macroalgae genus of the Gulf of California in  
35 different seasons and coastal regions.

36 Table S4. Comparison of different methods to identify DIC sources used by macroalgae.

37 Table S5. Temperature mean by regions and seasons in the Gulf of California based on time  
38 series data (1981 to 2016) by Escalante et al. (2013) and Robles-Tamayo (2018).

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 49 Vertical habitat: S=Subtidal and Intertidal are divided in Rp= Rockpool, Ee= Eventually exposed. Coastal  
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 51 P3: north. Season: Rain (from june to October) and Dry (from november to May).

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Specie	$\delta^{13}\text{C}$ Mean $\pm$ S.D.	Range: (Min to Max)	Strategies of dissolved inorganic carbon use	Habitat	Coastal region	Season and collecting year
<b>Chlorophyta</b>						
<i>Bryopsis</i> sp.	-15.3 $\pm$ 0.8‰	-15.9 to -14.7‰	Strategy 2	Rp, Ee	C2, C3	Rain (2013)
<i>Caulerpa cupressoides</i> (Vahl) C.Agardh	-16.3‰	-16.3‰	Strategy 2	Rp	C1	Dry (2009)
<i>Caulerpa mexicana</i> Sonder ex Kützing	-19.9‰	-19.91‰	Strategy 2	S	C1	Dry (2009)
<i>Caulerpa peltata</i> J.V. Lamouroux	-10.7 $\pm$ 4.5‰	-13.8 to -5.6‰	Strategies 1 and 2	Rp, Ee	C1	Rain (2013)
<i>Caulerpa sertularioides</i> (S.G.Gmelin) M.A.Howe	-18.5 $\pm$ 0.1‰	-18.5 to -18.3‰	Strategy 2	Ee	C2	Rain (2013)
<i>Chaetomorpha antennina</i> (Bory de Saint-Vincent) Kützing	-14.5 $\pm$ 1.07‰	-16.3 to -12.8‰	Strategy 2	Rp, Ev, S	C1, C3	Dry (2009), Rain (2013)
<i>Chaetomorpha linum</i> (O.F. Müller) Kützing	-16.8 $\pm$ 1.6‰	-18.4 to -14.6‰	Strategy 2	Ee	C1, C2	Dry (2009), Rain (2013)
<i>Chaetomorpha</i> sp.	-13.7 $\pm$ 0.8‰	-14.6 to -12.9‰	Strategy 2	Rp, Ee	C1, C2	Dry (2009), Rain (2013)
<i>Chaetomorpha</i> sp.2	-17.2‰	-17.24‰	Strategy 2	Ee	C2	Rain (2013)
<i>Cladophora albida</i> (Nees) Kützing	-19.4 $\pm$ 6.1‰	-31.3 to -13.8‰	Strategy 2	Ee	C2, C3	Dry (2009)
<i>Cladophora columbiana</i> F.S. Collins	-13.7‰	-13.7‰	Strategy 2	Rp	C3	Dry (2009)
<i>Cladophora microcladioides</i> F.S. Collins	-18.6 $\pm$ 3.6‰	-22.2 to -12.9‰	Strategy 2	Rp	C1	Dry (2009)
<i>Cladophora</i> sp.	-12.8 $\pm$ 2.6‰	-15.4 to -8.3‰	Strategies 1 and 2	Rc, Ee,S	C1,C2,C 3, P2	Dry (2008, 2009), Rain (2013)
<i>Cladophora</i> sp.2	-15.3‰	-15.3‰	Strategy 2	Ee	C2	Rain (2013)
<i>Cladophora</i> sp.3	-14.1‰	-14.1‰	Strategy 2	Ee	C2	Rain (2013)
<i>Cladophora</i> sp.4	-14.8 $\pm$ 1.4‰	-15.8 to -13.8‰	Strategy 2	Ee	C2	Rain (2013)
<i>Cladophoropsis</i> sp.	-12.2‰	-12.2‰	Strategy 2	Ee	C1	Rain (2013)
<i>Codium amplivesiculatum</i> * Setchell & N.L.Gardner	-14.4 $\pm$ 2.7‰	-20.4 to -11.3‰	Strategy 2	S	C2, P2, P3	Dry (2008, 2009)
<i>Codium brandegeei</i> Setchell & N.L.Gardner	-11.8 $\pm$ 1.2‰	-13.7 to -10.4‰	Strategy 2	S	C2, P2, P3	Dry (2009)
<i>Codium fragile</i> Setchell & N.L.Gardner	-13.0 $\pm$ 2.7‰	-14.8 to -9‰	Strategies 1 and 2	S	P2, P3	Dry (2009)
<i>Codium simulans</i> * Setchell & N.L.Gardner	-12.4 $\pm$ 2.2‰	-14.9 to -8.3‰	Strategies 1 and 2	S	C3, P1, P2, P3	Dry (2009)
<i>Codium</i> sp.	-11.6 $\pm$ 3.0‰	-14.1 to 6.7‰	Strategies 1 and 2	Ee, S	C1, C3, P2, P3	Dry (2009), Rain (2013)
<i>Phyllocladon robustum</i> Setchell & N.L.Gardner	-13.0 $\pm$ 2.0‰	-14.1 to -13.0‰	Strategy 2	Ee	P2, P3	Dry (2009)

<i>Rhizoclonium riparium</i> (Roth) Harvey	-19.6±1.9‰	-21.7 to -17.8‰	Strategy 2	<i>Ee</i>	C2	Dry (2008, 2009)
<i>Rhizoclonium</i> sp.	-13.7±2.7‰	-16.3 to -9.8‰	Strategies 1 and 2	<i>Ee, S</i>	C2, C3	Rain (2013)
<i>Struveopsis</i> sp.	-10.6±1.5‰	-14 to -8.8‰	Strategies 1 and 2	<i>Rp, Ee</i>	C3	Rain (2013)
<i>Struveopsis</i> sp2.	-10.4‰	-10.46‰	Strategy 2	<i>Ee</i>	C3	Rain (2013)
<i>Ulva acanthophora</i> (Kützting) Hayden Blomster, Maggs, P.C.Silva, Stanhope & J.R.Waaland	-15.8±1.7‰	-18.3 to -11.4‰	Strategy 2	<i>Ee</i>	P1, P2, P3	Dry (2008, 2009)
<i>Ulva clathrata</i> (Roth) C. Agardh	-16.4±2.0‰	-20.5 to -14.5‰	Strategy 2	<i>Ee</i>	C2, C3, P2	Dry (2008, 2009)
<i>Ulva compressa</i> Linnaeus	-17.8±2.4‰	-20.6 to -15.4‰	Strategy 2	<i>Ee</i>	C2, C3, P2	Dry (2009), Rain (2013)
<i>Ulva flexuosa</i> Wulfen	-16.0±3.7‰	-25.9 to -10.4‰	Strategy 2	<i>Ee</i>	C1, C2, P2	Dry (2009), Rain (2013)
<i>Ulva intestinalis</i> Linnaeus	-15.3±2.5‰	-20.3 to -8.9‰	Strategies 1 and 2	<i>Ee, S</i>	C1, C2, C3, P1, P2, P3	Dry (2008, 2009), Rain (2013)
<i>Ulva lactuca</i> Linnaeus	-14.1±3.1‰	-19.6 to -7.7	Strategies 1 and 2	<i>Ee, S</i>	C1, C2, C3, P1, P2, P3	Dry (2008, 2009), Rain (2013)
<i>Ulva linza</i> Linnaeus	-15.6±2.4‰	-19.4 to -13.2‰	Strategy 2	<i>Ee</i>	C2, C3, P1, P3	Dry (2009)
<i>Ulva lobata</i> (Kützting) Harvey	-13.2±1.9‰	-15.3 to -11.1‰	Strategy 2	<i>Ee</i>	C1, P1	Dry (2009)
<i>Ulva prolifera</i> O.F.Müller	-14.2±1.8‰	-15.5 to -12.2‰	Strategy 2	<i>Ee</i>	C3, P2	Dry (2008, 2009)
<i>Ulva rigida</i> C.Agardh	-13.7‰	-13.67‰	Strategy 2	<i>Ee</i>	P2	Dry (2009)
<i>Ulva</i> sp.	-14.0±3.9‰	-19.2 to -7.1‰	Strategies 1 and 2	<i>Ee, S</i>	C1, C2, P1, P2, P3	Dry (2009)
<i>Ulva stenophylla</i> Setchell & N.L.Gardner	-17.4‰	-17.4‰	Strategy 2	<i>Ee</i>	P2	Dry (2009)
<i>Valoniopsis pachynema</i> (G.Martens) Børgesen	-11.9‰	-11.9‰	Strategy 2	<i>Rp</i>	C3	Dry (2009)
<b>Ochrophyta</b>						
<i>Colpomenia ramosa</i> W.R. Taylor	-11.4±2.6‰	-13.8 to -7.8‰	Strategies 1 and 2	<i>Ee, S</i>	C1, P2	Dry (2008, 2009)
<i>Colpomenia sinuosa</i> (Mertens ex Roth) Derbès & Solier	-10.2±3‰	-16.3 to -7.2‰	Strategies 1 and 2	<i>Rp, Ee, S</i>	C2, C3, P2, P3	Dry (2008, 2009)
<i>Colpomenia</i> sp.	-11.0±3.7‰	-19.0 to -5.4‰	Strategies 1 and 2	<i>Ee</i>	C1, C2, C3, P2, P3	Dry (2009)
<i>Colpomenia tuberculata</i> De A. Saunders	-8.7 ±3.2‰	-19.3 to -2.2‰	Strategies 1 and 2	<i>Ee, S</i>	C1, C2, C3, P1, P2, P3	Dry (2008, 2009), Rain (2013)
<i>Cutleria hancockii</i> * E.Y.Dawson	-14.64‰	-14.6‰	Strategy 2	<i>S</i>	C3	Dry (2009)
<i>Dictyota concrescen</i> W.R.Taylor	-16.5‰	-16.5‰	Strategy 2	<i>Rp</i>	C1	Dry (2009)
<i>Dictyota coriacea</i> (Holmes) I.K.Wang, H. S.Kim & W.J.Lee	-18.1‰	-18.1‰	Strategy 2	<i>S</i>	C3	Dry (2009)
<i>Dictyota dichotoma</i> (Hudson) J.V. Lamouroux	-22.2±6.2‰	-15.2 to -6.5‰	Strategies 1 and 2	<i>Ee, S</i>	C3, P2	Dry (2009)
<i>Dictyota flabellata</i> (F.S.Collins) Setchell & N.L.Gardner	-16.7‰	-16.7‰	Strategy 2	<i>S</i>	P3	Dry (2009)
<i>Dictyota implexa</i> (Desfontaines) J.V.Lamouroux	-17.3±3.0‰	-17.8 to -16.9‰	Strategy 2	<i>S</i>	C3, P3	Dry (2009)
<i>Dictyota</i> sp.	-14.5±3.7‰	-21.5 to -7.0‰	Strategies 1 and 2	<i>Rp, Ee, S</i>	C1, C3, P2, P3	Dry (2009), Rain (2013)
<i>Dictyota</i> sp2.	-12.8‰	-12.8‰	Strategy 2	<i>Ee</i>	C3	Rain (2013)
<i>Ectocarpus</i> sp.	-15.4±1.9‰	-17.1 to -12.3‰	Strategy 2	<i>Rp, Ee, S</i>	C1, C2, C3	Dry (2009), Rain (2013)
<i>Endarachne</i> sp.	-12.2±0.5‰	-12.5 to -11.8‰	Strategy 2	<i>Ee</i>	P3	Dry (2009)

<i>Hydroclathrus clathratus</i> (C.Agardh) M.A.Howe in N.L.Britton & C.F.Millsbaugh	-7.4± 4.1‰	-15.6 to -4.4‰	Strategies 1 and 2	<i>Ee, S</i>	P1, P2	Dry (2008, 2009)
<i>Hydroclathrus</i> sp.	-7.2‰	-7.2‰	Strategy 1	<i>S</i>	P2	Dry (2009)
<i>Ishige foliacea</i> Okamura	-13.3‰	-13.3‰	Strategy 2	<i>S</i>	C3	Dry (2009)
<i>Pachydietyon coriaceum</i> (Holmes) Okamura	-18.2‰	-18.2‰	Strategy 2	<i>S</i>	C3	Dry (2009)
<i>Padina concrescens</i> Thivy in W.R. Taylor	-11.68‰	-11.68‰	Strategy 2	<i>S</i>	P2	Dry (2009)
<i>Padina crispata</i> Thivy in W.R. Taylor	-11.3±1.7‰	-12.5 to -10.1‰	Strategies 1 and 2	<i>Rp, S</i>	P2, P3	Dry (2009)
<i>Padina durvillei</i> Bory Saint-Vincent	-13.2±2.6‰	-20. To -9.2‰	Strategies 1 and 2	<i>Rp, Ee, S</i>	C1, C2, C3, P1, P2, P3	Dry (2008, 2009), Rain (2013)
<i>Padina</i> sp.	-11.1±1.5‰	-13.1 to -7.9‰	Strategies 1 and 2	<i>Rp, Ee, S</i>	C1, C3, P1, P2, P3	Dry (2008, 2009), Rain (2013)
<i>Rosenvingea intricata</i> (J.Agardh) Børgesen	-11.4±2.2‰	-12.9 to -9.8‰	Strategies 1 and 2	<i>Ee, S</i>	P2	Dry (2009)
<i>Rosenvingea</i> sp.	-16.0±0.04‰	-16.0 to -16.0‰	Strategy 2	<i>Ee, S</i>	C3, P3	Dry (2009)
<i>Sargassum brandegeei</i> Setchell & N.L.Gardner	-16.36‰	-16.4‰	Strategy 2	<i>S</i>	P2	Dry (2009)
<i>Sargassum herphorizum</i> * Setchell et Gardner	-13.7±1.6‰	-16.6 to -11.5‰	Strategy 2	<i>S</i>	C3, P2, P3	Dry (2009)
<i>Sargassum horridum</i> * Setchell & N.L.Gardner	-15.5±2.9‰	-19.17 to -9.5‰	Strategies 1 and 2	<i>S</i>	C3, P2, P3	Dry (2009)
<i>Sargassum johnstonii</i> Setchell & N.L.Gardner	-15.4±2.0‰	-17.7 to -11.8‰	Strategy 2	<i>S</i>	C3, P2, P3	Dry (2009)
<i>Sargassum lapazeanum</i> Setchell & N.L.Gardner	-14.5±1.6‰	-17.2 to -12.8‰	Strategy 2	<i>S</i>	C3, P2	Dry (2008, 2009)
<i>Sargassum liebmannii</i> J.Agardh	-13.7‰	-13.7‰	Strategy 2	<i>S</i>	C1	Dry (2009)
<i>Sargassum muticum</i>	-19.2‰	-19.2‰	Strategy 2	<i>S</i>	P2	Dry (2008)
<i>Sargassum sinicola</i> Setchell & N.L.Gardner	-15.1±2.4‰	-21.1 to -12.1‰	Strategy 2	<i>S</i>	C2, C3, P2, P3	Dry (2008, 2009)
<i>Sargassum sinicola subsp. camouii</i> (E.Y.Dawson) J.N.Norris & Yensen	-12.7±3.1‰	-14.8 to -10.7‰	Strategy 2	<i>S</i>	C3, P1	Dry (2009)
<i>Sargassum</i> sp.	-14.3±2.4‰	-18.7 to -8.0‰	Strategies 1 and 2	<i>Ee, S</i>	C2, C3, P1, P2, P3	Dry (2008, 2009), Rain (2013)
<i>Sargassum</i> sp.2	-16.5‰	-16.5‰	Strategy 2	<i>S</i>	C3	Rain (2013)
<i>Sargassum</i> sp.3	-15.3‰	-15.3‰	Strategy 2	<i>Ee</i>	C3	Rain (2013)
<i>Spatoglossum howellii</i>	-18.2±3.6‰	-20.8 to -15.7‰	Strategy 2	<i>S</i>	C1	Dry (2009)
<i>Sphacelaria californica</i> Setchell & N.L.Gardner	-10.8‰	-10.8‰	Strategy 2	<i>Ee</i>	C3	Dry (2009)
<i>Taonia lennebackerae</i> Farlow ex J.Agardh	-15.3‰	-15.3‰	Strategy 2	<i>S</i>	C3	Dry (2009), Rain (2013)
<i>Zonaria</i> sp.	-16.0± 1.6‰	-17.6 to -13.4‰	Strategy 2	<i>Ee, S</i>	C2, C3, P2	Dry (2009), Rain (2013)
<b>Rhodophyta</b>						
<i>Acanthophora</i> sp.	-16.3±0.5‰	-16.7 to -15.9‰	Strategy 2	<i>Rp, S</i>	P2, P3	Dry (2009)
<i>Acanthophora spicifera</i> (M.Vahl) Børgesen	-14.5‰	-14.5‰	Strategy 2	<i>Rp</i>	P1	Dry (2009)
<i>Ahnfeltiopsis leptophylla</i> (J. Agardh) P.C. Silva y DeCew	-14.4±2.4‰	-17.9 to -10.7‰	Strategy 2	<i>Rp, Ee</i>	C1, P2, P3	Dry (2008, 2009)
<i>Ahnfeltiopsis</i> sp.1	-15.9±2.6‰	-18.7 to -13.5‰	Strategy 2	<i>Ee, S</i>	C2, C3	Rain (2013)
<i>Ahnfeltiopsis</i> sp.2	-16.9‰	-16.9‰	Strategy 2	<i>S</i>	C3	Rain (2013)
<i>Amphiroa beauvoisii</i> J.V.Lamouroux	-8.6±0.4‰	-8.9 to -8.4‰	Strategy 1	<i>S</i>	C3, P3	Dry (2009)
<i>Amphiroa misakiensis</i> Yendo	-7.5‰	-7.5‰	Strategy 1	<i>S</i>	C1	Dry (2009)
<i>Amphiroa</i> sp. J.V.Lamouroux	-6.5± 1.1‰	-8.5 to 4.5‰	Strategy 1	<i>Rp, Ee, S</i>	C1, C2, C3, P3	Dry (2009), Rain (2013)
<i>Amphiroa</i> sp.2	-8.0±2.1‰	-10.0 to -5.8‰	Strategy 1	<i>Rp, Ee</i>	C1, C3	Rain (2013)
<i>Amphiroa</i> sp.3	-6.4‰	-6.4‰	Strategy 1	<i>Rp</i>	C1	Rain (2013)
<i>Bostrychia radicans</i>	-25.8‰	-25.8‰	Strategy 2	<i>Rp</i>	C1	Dry (2009)

<i>Centroceras clavulatum</i> (C.Agardh) Montagne	-15.3±2‰	-16.6 to -13.8‰	Strategy 2	S	C1	Dry (2009),
<i>Centroceras</i> sp.	-16.1±0.4‰	-16.3 to -15.8‰	Strategy 2	Rp, Ee	C1	Dry (2008, 2009)
<i>Ceramium</i> sp.	-12.5±2.7‰	-15.4 to -8.2‰	Strategies 1 and 2	Rp, Ee, S	C2, C3	Rain (2013)
<i>Ceratodictyon tenue</i> (Setchell & N.L.Gardner) J.N.Norris	-14.2±1.4‰	-15.1 to -12.6‰	Strategy 2	Rp, Ee	C1	Rain (2013)
<i>Champia</i> sp.	-14.9‰	-14.9‰	Strategy 2	S	P2	Dry (2009)
<i>Chondracanthus squarulosus</i> (Setchell & N.L.Gardner) J.R.Hughey, P.C.Silva & Hommersand	-12.6‰	-12.6‰	Strategy 2	S	P2	Dry (2009)
<i>Chondracanthus tepidus</i> (Hollenberg) Guiry	-17.6‰	-17.6‰	Strategy 2	S	P2	Dry (2009)
<i>Chondria acrorhizophora</i> Setchell & N.L.Gardner	-14.2‰	-14.2‰	Strategy 2	S	P2	Dry (2009)
<i>Chondria californica</i> (Collins) Kylin	-18.8‰	-18.8‰	Strategy 2	S	P2	Dry (2009)
<i>Corallina vancouverensis</i> Yendo	-8.3‰	-8.3‰	Strategy 1	S	P3	Dry (2009)
<i>Dasya bailloouviana</i> (S.G.Gmelin) Montagn	-18.1±2.9‰	-20.1 to -16.0‰	Strategy 2	S	P1, P2	Dry (2009)
<i>Dermoneia</i> sp.	-15.8‰	-15.8‰	Strategy 2	Rp	C1	Rain (2013)
<i>Digenia simplex</i> (Wulfen) C. Agardh	-11.5‰	-11.5‰	Strategy 2	Rp	P2	Dry (2009)
<i>Digenia</i> sp.	-15.8±1.1‰	-16.7 to -15.0‰	Strategy 2	S	P2	Dry (2009)
<i>Eucheuma</i> sp.	-17.9±1.4‰	-19.0 to -16.9‰	Strategy 2	Ee, S	P3	Dry (2009)
<i>Gelidium sclerophyllum</i> W.R. Taylor	-15.2±3.1‰	-11.7 to -10.6‰		Ee	C1, P3	Dry (2009)
<i>Gelidium</i> sp.	-14.3±2.0‰	-16.9 to -12.5‰	Strategy 2	Rp, Ee, S	C1, C3, P2	Dry (2009), Rain (2013)
<i>Gigartina</i> sp.	-16.9±6.6‰	-31.8 to -12.8‰	Strategies 2 and 3	Es, S	C2, C3, P3	Dry (2008, 2009)
<i>Gracilaria crispata</i> (Bory de Saint-Vincent) D.H.Kim.	-15.0±2.9‰	-19.1 to -10.1‰	Strategies 1 and 2	Ee, S	P2	Dry (2008, 2009)
<i>Gracilaria marcialana</i> * Dawson	-16.6‰	-16.6‰	Strategy 2	S	P3	Dry (2009)
<i>Gracilaria pacifica</i> . Kjellman	-16.5±1.6‰	-18.6 to -13.6‰	Strategy 2	S	C2, C3, P2	Dry (2009)
<i>Gracilaria</i> sp.	-15.5±2.4‰	-21.8 to -12.2‰	Strategy 2	Rp,Ee, S	C2, C3, P1, P2, P3	Dry (2008, 2009), Rain (2013)
<i>Gracilaria</i> sp.2	-14.4±3.7‰	-18.7 to -12.3‰	Strategy 2	Ee, S	C2, C3	Rain (2013)
<i>Gracilaria spinigera</i> * E.Y.Dawson	-14.9±3.8‰	-17.7 to -12.2‰	Strategy 2	S	C2	Dry (2009)
<i>Gracilaria subsecundata</i> Setchell & N.L.Gardner	-15.9±2.8‰	-20.3 to -12.8‰	Strategy 2	S	P1, P2, P3	Dry (2008, 2009)
<i>Gracilaria tepocensis</i> E.Y.Dawson	-15.1±1.9‰	-17.0 to -13.2‰	Strategy 2	S	P1, P3	Dry (2009)
<i>Gracilaria textorii</i> (Suringar) De Toni	-16.2±2.6‰	-18.0 to -14.3‰	Strategy 2	S	C1, P2	Dry (2008, 2009)
<i>Gracilaria textorii</i> var. <i>Textorii</i> (Suringar) De Toni	-15.1±0.2‰	-15.2 to -15.0‰	Strategy 2	S	P2	Dry (2009)
<i>Gracilaria turgida</i> E.Y.Dawson	-15.3±3.5‰	-20.7 to -12.0‰	Strategy 2	Rp, S	C3, P2, P3	Dry (2008, 2009)
<i>Gracilaria vermiculophylla</i> (Ohmi) Papenfuss	-15.9±3.8 ‰	-23.4 to -8.8‰	Strategies 1 and 2	Ee, S	C1,C2, C3, P1, P2, P3	Dry (2009), Rain (2013)
<i>Gracilariopsis lemaneiformis</i> (Bory de Saint-Vincent) E.Y.Dawson, Acleto & Foldvik	-16.0‰	-16.0‰	Strategy 2	S	P2	Dry (2008)
<i>Gracilariopsis longissima</i> (S.G.Gmelin) M.Steentoft, L.M.Irvine & W.F.Farnham	-16.8±1.8‰	-18.1 to -15.0‰	Strategy 2	S	P2, P3	Dry (2009)
<i>Grateloupia doryphora</i> (Montagne) M.A.Howe	-13.6±0.6‰	-13.9 to -13.1‰	Strategy 2	Ee	P2	Dry (2009)

<i>Grateloupia filicina</i> (J.V. Lamouroux) C. Agardh	-17.0±2.5‰	-19.9 to -14.1‰	Strategy 2	<i>Ee</i>	C1, C2, C3	Dry (2008, 2009), Rain (2013)
<i>Grateloupia Howei</i> Setchell y Gardner	-14.7±1.5‰	-15.8 to -13.7‰	Strategy 2	<i>Ee</i>	C1	Dry (2009)
<i>Grateloupia</i> sp.	-16.4±4.4‰	-22.6 to -12.6‰	Strategy 2	<i>S</i>	C3, P2, P3	Dry (2009)
<i>Grateloupia versicolor</i> (J. Agardh) J. Agardh	-12.6±1.3‰	-13.8 to -11.2‰	Strategy 2	<i>Ee</i>	C1	Dry (2009)
<i>Gymnogongrus guadalupensis</i> E.Y.Dawson	-13.1‰	-13.1‰	Strategy 2	<i>S</i>	C3	Dry (2009)
<i>Gymnogongrus</i> sp.	-14.1±3.8‰	-23.2 to -10.1‰	Strategies 1 and 2	<i>Rp, Ee, S</i>	C1, C3, P2, P3	Dry (2009), Rain (2013)
<i>Gymnogongrus</i> sp.2	-12.8‰	-12.8‰	Strategy 2	<i>Rp</i>	C3	Rain (2013)
<i>Gymnogongrus</i> sp.3	-13.1‰	-13.1‰	Strategy 2	<i>Rp</i>	C3	Rain (2013)
<i>Halymenia actinophysa</i> M.A.Howe	-24.5‰	-24.5‰	Strategy 2	<i>S</i>	P3	Dry (2009)
<i>Halymenia</i> sp.	-23.7±13.6‰	-33.4 to -14.1‰	Strategies 2 and 3	<i>S</i>	P2	Dry (2009)
<i>Hypnea johnstonii</i> Setchell y Gardner	-11.2±3.5‰	-13.8 to -6.5‰	Strategies 1 and 2	<i>Rp, Ee</i>	C1, P1, P2	Dry (2008, 2009)
<i>Hypnea pannosa</i> J. Agardh	-11.8±3.3‰	-15.0 to -6.4‰	Strategies 1 and 2	<i>Rp, Ee</i>	C1, P2	Dry (2009), Rain (2013)
<i>Hypnea</i> sp.	-14.9±2.3‰	-20.8 to -11.4‰	Strategy 2	<i>Rp, Ee, S</i>	C1, C2, C3, P2, P3	Dry (2009), Rain (2013)
<i>Hypnea</i> sp.2	-14.1‰	-14.1‰	Strategy 2	<i>Ee</i>	C3	Rain (2013)
<i>Hypnea spinella</i> (C.Agardh) Kützing	-16.4±1.8‰	-19.2 to -14.9‰	Strategy 2	<i>Rp, Ee, S</i>	C1, P2, P3	Dry (2008, 2009), Rain (2013)
<i>Hypnea valentiae</i> (Turner) Montagne	-15.2±2.3‰	-19.2 to -13.6‰	Strategy 2	<i>Ee, S</i>	C1, C3, P1, P2	Dry (2008, 2009)
<i>Jania</i> sp.	-9.5±1.0‰	-10.2 to -8.8‰	Strategy 1	<i>Ee, S</i>	C3	Rain (2013)
<i>Jania</i> sp.2	-10.7‰	-10.7‰	Strategy 1	<i>S</i>	C3	Rain (2013)
<i>Laurencia pacifica</i> Kylin	-14.9±2.2‰	-19.0 to -12.7‰	Strategy 2	<i>Ee, S</i>	P1, P2, P3	Dry (2009)
<i>Laurencia papillosa</i> * (C.Agardh)	-15.7±0.3‰	-15.9 to -15.6‰	Strategy 2	<i>Ee</i>	P1, P2	Dry (2008, 2009)
<i>Laurencia</i> sp.	-12.9±1.2‰	-14.7 to -10.5‰	Strategies 1 and 2	<i>Rp, Ee</i>	C1, C3, P2, P3	Dry (2009), Rain (2013)
<i>Lomentaria hakodatensis</i> Yendo	-12.8‰	-12.8‰	Strategy 2	<i>Ee</i>	P2	Dry (2009)
<i>Lomentaria</i> sp.	-15.7‰	-15.8‰	Strategy 2	<i>S</i>	P3	Dry (2009)
<i>Mazzaella leptorhynchus</i> (J.Agardh) Leister	-19.9‰	-19.9‰	Strategy 2	<i>S</i>	P3	Dry (2009)
<i>Neosiphonia johnstonii</i> Setchell y N.L Gardner	-13.4±6.0‰	-19.0 to -6.9‰		<i>Ee, S</i>	C3, P1, P2	Dry (2009)
<i>Neosiphonia paniculat</i> Montagne	-17.8±1.3‰	-18.7 to -16.9‰	Strategy 2	<i>S</i>	C2, P2	Dry (2009)
<i>Palisada paniculata</i> (Kützing) J.N.Norris	-12.4‰	-12.4‰	Strategy 2	<i>S</i>	P3	Dry (2009)
<i>Polysiphonia mollis</i> . J.D. Hooker y Harvey	-19.8±1.8‰	-21.4 to -15.8‰	Strategy 2	<i>Ee, S</i>	C2, C3, P1, P2, P3	Dry (2009)
<i>Polysiphonia nathaniellii</i>	-18.6‰	-18.6‰	Strategy 2	<i>S</i>	C1	Dry (2009)
<i>Polysiphonia pacifica</i> . Hollenberg	-14.0±7.9‰	-21.5 to -5.7‰		<i>Ee, S</i>	C2, P2	Dry (2009)
<i>Polysiphonia</i> sp.	-19.4±3.5‰	-23.0 to -15.6‰	Strategy 2	<i>Ee, S</i>	P2, C3	Dry (2009), Rain (2013)
<i>Prionitis abbreviata</i> Setchell & E.Y.Dawson	-13.9±2.7‰	-15.8 to -12.0‰	Strategy 2	<i>Rp</i>	C3, P3	Dry (2009), Rain (2013)
<i>Prionitis abbreviata</i> var <i>abbreviata</i> (E.Y.Dawson) E.Y Dawson	-13.4‰	-13.4‰	Strategy 2	<i>S</i>	C3	Dry (2009)
<i>Prionitis abbreviata</i> var <i>guaymanensis</i> (E.Y.Dawson) E.Y.Dawson	-11.4‰	-11.4‰	Strategy 2	<i>S</i>	C3	Dry (2009)
<i>Prionitis</i> sp.	-14.4±1.1‰	-15.7 to -13.6‰	Strategy 2	<i>Ee, S</i>	C3	Dry (2009), Rain (2013)
<i>Prionitis</i> sp.2	-13.8±1.1‰	-14.6 to -13.0‰	Strategy 2	<i>Rp, Ee</i>	P3	Dry (2009)

<i>Pterocladia</i> sp.	-13.5‰	-13.5‰	Strategy 2	S	P3	Dry (2009)
<i>Pyropia thuretii</i> (Setchell & E.Y.Dawson) J.E.Sutherland, L.E.Aguilar Rosas & R	-20.1‰	-20.1‰	Strategy 2	S	C3, P2, P3	Dry (2009)
<i>Rhodoglossum</i> sp.	-16.1±1.0‰	-17.1 to -15.0‰	Strategy 2	S	C3	Dry (2009)
<i>Rhodymenia</i> sp.	-18.7±2.4‰	-20.4 to -17.0‰	Strategy 2	S	P2	Dry (2009)
<i>Schizymenia pacifica</i> (Kylin) Kylin	-33.8±1.2‰	-34.6 to -33.0‰	Strategy 3	S	P2	Dry (2009)
<i>Scinaia johnstoniae</i> * Setchell	-19.0‰	-19.0‰	Strategy 2	S	C3	Dry (2009)
<i>Spyrida</i> sp.	-17.0±1.2‰	-19.1 to -16.1‰	Strategy 2	S	P1, P2	Dry (2009)
<i>Spyridia filamentosa</i> (Wulfen) Harvey	-15.9±3.8‰	-26.2 to -11.5‰	Strategy 2	Ee, S	C2, C3, P1, P2, P3	Dry (2008, 2009), Rain (2013)
<i>Tacanoosca uncinata</i> (Setchell & N.L.Gardner) J.N.Norris, P.W.Gabrielson & D.P.Cheney	-10.9±4.7‰	-16.3 to -7.9‰	Strategy 2	Ee, S	C3, P3	Dry (2009)
<i>Weeksia coccinea</i> . (Harvey) S.C.Lindstrom	-13.8‰	-13.9‰	Strategy 2	Rp	P3	Dry (2009)
<i>Zanardinula</i> sp.	-13.3‰	-13.3‰	Strategy 2	Rp	C3	Rain (2013)

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55 Table S2. Comparison of  $\delta^{13}\text{C}$  values in macroalgae species of the Gulf of California in different seasons and  
 56 coastal regions. Different letters indicate significant differences by kruskal wallis test at  $p < 0.05$ .

Taxa	Specie	Collecting season by year	Coastal region	$\delta^{13}\text{C}$ mean $\pm$ DS	n=
Chlorophyta	<i>Chaetomorpha antennina</i>	Dry (2009)	C1	-15.2 $\pm$ 1.1 <sup>a</sup> ‰	4
		Rain (2013)	C1	-13.9 $\pm$ 0.8 <sup>a</sup> ‰	5
Chlorophyta	<i>Ulva acanthophora</i>	Dry (2008)	P2	-16.7 $\pm$ 1.1 <sup>a</sup> ‰	4
		Dry (2009)	P2	-15.9 $\pm$ 1.6 <sup>a</sup> ‰	13
		Dry (2009)	P3	-14.3 $\pm$ 2.0 <sup>a</sup> ‰	5
Chlorophyta	<i>U. flexuosa</i>	Dry (2009)	C1	-14.1 $\pm$ 2.7 <sup>a</sup> ‰	4
		Dry (2009)	C2	-14.9 $\pm$ 2.0 <sup>a</sup> ‰	3
Chlorophyta	<i>U. intestinalis</i>	Dry (2009)	P2	-14.9 $\pm$ 1.5 <sup>a</sup> ‰	4
		Rain (2013)	C1	-17.6 $\pm$ 2.4 <sup>a</sup> ‰	3
Chlorophyta	<i>U. lactuca</i>	Dry (2008)	P2	-15.3 $\pm$ 3.3 <sup>a</sup> ‰	4
		Dry (2009)	P2	-14.5 $\pm$ 3.3 <sup>a</sup> ‰	9
		Dry (2009)	C1	-12.6 $\pm$ 2.7 <sup>a</sup> ‰	5
Ochrophyta	<i>Colpomenia tuberculata</i>	Dry (2008)	P1	-8.3 $\pm$ 3.8 <sup>a</sup> ‰	4
		Dry (2008)	P2	-7.2 $\pm$ 1.4 <sup>a</sup> ‰	4
		Dry (2009)	P2	-8.8 $\pm$ 4.0 <sup>a</sup> ‰	28
		Dry (2009)	P3	-9.7 $\pm$ 2.8 <sup>a</sup> ‰	13
		Dry (2009)	C1	-8.5 $\pm$ 1.9 <sup>a</sup> ‰	12
		Dry (2009)	C3	-7.55 $\pm$ 1.0 <sup>a</sup> ‰	3
Ochrophyta	<i>Sargassum sinicola</i>	Dry (2008)	P2	-14.2 $\pm$ 1.6 <sup>a</sup> ‰	3
		Dry (2009)	P2	-15.0 $\pm$ 2.7 <sup>a</sup> ‰	16
		Dry (2009)	P3	-15.3 $\pm$ 2.5 <sup>a</sup> ‰	5
		Dry (2009)	C2	-15.2 $\pm$ 0.7 <sup>a</sup> ‰	3
Ochrophyta	<i>Padina durvillaei</i>	Dry (2009)	P2	-12.5 $\pm$ 2.2 <sup>a</sup> ‰	10
		Dry (2009)	P3	-11.8 $\pm$ 1.2 <sup>a</sup> ‰	5
		Dry (2009)	C1	-12.7 $\pm$ 2.2 <sup>ab</sup> ‰	6
		Rain (2013)	C1	-15.8 $\pm$ 2.5 <sup>b</sup> ‰	7
Rhodophyta	<i>Spyridia filamentosa</i>	Dry (2009)	P1	-12.6 $\pm$ 1.0 <sup>a</sup> ‰	4
		Dry (2009)	P2	-14.8 $\pm$ 1.6 <sup>a</sup> ‰	4

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Table S3. Comparison of  $\delta^{13}\text{C}$  values in macroalgae genus of the Gulf of California in different seasons and coastal regions. Different letters indicate significant differences by kruskal wallis test at  $p < 0.05$ .

Taxa	Genus	Collecting season and year	Coastal region	$\delta^{13}\text{C}$ mean $\pm$ DS	n=
Chlorophyta	<i>Chaetomorpha</i>	Dry (2009)	C1	-14.2 $\pm$ 1.3 <sup>a</sup> ‰	8
		Rain (2013)	C1	-15.5 $\pm$ 1.3 <sup>a</sup> ‰	6
		Rain (2013)	C2	-15.9 $\pm$ 2.3 <sup>a</sup> ‰	4
Chlorophyta	<i>Cladophora</i>	Dry (2009)	C1	-17.27 $\pm$ 7.8 <sup>a</sup> ‰	3
		Rain (2013)	C2	-14.9 $\pm$ 0.8 <sup>a</sup> ‰	6
Chlorophyta	<i>Codium</i>	Dry (2009)	P2	-13.3 $\pm$ 1.4 <sup>a</sup> ‰	14
		Dry (2009)	P3	-9.8 $\pm$ 1.8 <sup>b</sup> ‰	9
		Dry (2009)	C3	-12.3 $\pm$ 1.4 <sup>a</sup> ‰	4
Chlorophyta	<i>Ulva</i>	Dry (2008)	P1	-15.6 $\pm$ 1.8 <sup>ab</sup> ‰	6
		Dry (2009)	P1	-15.6 $\pm$ 2.4 <sup>ab</sup> ‰	7
		Dry (2008)	P2	-16.6 $\pm$ 2.2 <sup>b</sup> ‰	10
		Dry (2009)	P2	-15.2 $\pm$ 2.4 <sup>ab</sup> ‰	35
		Dry (2009)	P3	-12.4 $\pm$ 3.2 <sup>a</sup> ‰	11
		Dry (2009)	C1	-13.5 $\pm$ 2.5 <sup>ab</sup> ‰	18
		Rain (2013)	C1	-17.4 $\pm$ 2.0 <sup>b</sup> ‰	4
		Dry (2009)	C2	-14.8 $\pm$ 1.6 <sup>ab</sup> ‰	13
		Dry (2009)	C3	-15.8 $\pm$ 2.7 <sup>b</sup> ‰	10
Ochrophyta	<i>Colpomenia</i>	Dry (2008)	P1	-8.4 $\pm$ 3.3 <sup>a</sup> ‰	5
		Dry (2008)	P2	-8.2 $\pm$ 2.5 <sup>a</sup> ‰	5
		Dry (2009)	P2	-8.8 $\pm$ 3.8 <sup>a</sup> ‰	32
		Dry (2009)	P3	-10.5 $\pm$ 3.3 <sup>a</sup> ‰	20
		Dry (2009)	C1	-9.8 $\pm$ 2.8 <sup>a</sup> ‰	16
		Dry (2009)	C2	-8.6 $\pm$ 1.6 <sup>a</sup> ‰	3
		Dry (2009)	C3	-7.8 $\pm$ 1.0 <sup>a</sup> ‰	5
Ochrophyta	<i>Dictyota</i>	Dry (2009)	P2	-12.3 $\pm$ 5.0 <sup>a</sup> ‰	3
		Dry (2009)	P3	-15.9 $\pm$ 2.4 <sup>a</sup> ‰	3
		Dry (2009)	C3	-16.7 $\pm$ 1.5 <sup>a</sup> ‰	3
		Rain (2013)	C3	-14.3 $\pm$ 1.0 <sup>a</sup> ‰	5
Ochrophyta	<i>Padina</i>	Dry (2009)	P2	-12.0 $\pm$ 2.2 <sup>a</sup> ‰	13
		Dry (2009)	P3	-11.8 $\pm$ 1.0 <sup>a</sup> ‰	9
		Dry (2009)	C1	-11.9 $\pm$ 2.3 <sup>a</sup> ‰	8
		Rain (2013)	C1	-15.9 $\pm$ 2.5 <sup>b</sup> ‰	7
		Rain (2013)	C3	-11.9 $\pm$ 0.9 <sup>a</sup> ‰	6
Ochrophyta	<i>Sargassum</i>	Dry (2008)	P2	-14.7 $\pm$ 2.4 <sup>a</sup> ‰	6
		Dry (2009)	P2	-14.9 $\pm$ 2.5 <sup>a</sup> ‰	38
		Dry (2009)	P3	-14.6 $\pm$ 2.4 <sup>a</sup> ‰	20
		Dry (2009)	C2	-14.9 $\pm$ 1.5 <sup>a</sup> ‰	14
		Dry (2009)	C3	-15.2 $\pm$ 2.6 <sup>a</sup> ‰	14
		Rain (2013)	C3	-15.2 $\pm$ 1.1 <sup>a</sup> ‰	5
Rhodophyta	<i>Gracilaria</i>	Dry (2009)	P1	-13.8 $\pm$ 1.7 <sup>a</sup> ‰	10
		Dry (2008)	P2	-14.3 $\pm$ 2.4 <sup>a</sup> ‰	4
		Dry (2009)	P2	-15.8 $\pm$ 2.2 <sup>a</sup> ‰	24
		Dry (2009)	P3	-16.0 $\pm$ 4.0 <sup>a</sup> ‰	8
		Rain (2013)	C2	-15.9 $\pm$ 2.7 <sup>a</sup> ‰	7
		Dry (2009)	C3	-17.0 $\pm$ 3.5 <sup>a</sup> ‰	10
		Rain (2013)	C3	-14.0 $\pm$ 1.4 <sup>a</sup> ‰	4
Rhodophyta	<i>Hypnea</i>	Dry (2009)	P2	-15.3 $\pm$ 2.4 <sup>a</sup> ‰	9
		Dry (2009)	P3	-15.6 $\pm$ 1.8 <sup>a</sup> ‰	4
		Dry (2009)	C1	-13.0 $\pm$ 3.8 <sup>a</sup> ‰	11



Table S4. Comparison of different methods to identify DIC sources used by macroalgae.

Taxa	Specie	Locality	Sampling date	Habitat	DIC source	Method	Comment	Source
Chlorophyta	<i>Bryopsis</i> sp.	Gulf of California	2013	Rockpool, subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	15.3±0.8(2)‰	This study
	<i>Bryopsis</i> sp.	Great barrier reef, Australia	2008-2012	Subtidal (0.5-5.2m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-15.2±6.9(5)‰	Lovelock et al., (2019)
	<i>Caulerpa brownii</i>	Shortlands Bluff, Australia	2002-2003	Exposed rock-pool faces and the rock-pool	CO <sub>2</sub>	pH drift experiments, δ <sup>13</sup> C signal	Final pH~8.9, -30.2‰	Kevekordes et al., (2006)
	<i>C. cactoides</i>	Shortlands Bluff, Australia	2002-2003	Exposed rock-pool faces and the rock-pool	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments, δ <sup>13</sup> C signal	Final pH>9, -21.7‰	Kevekordes et al., (2006)
	<i>C. cupressoides</i>	Gulf of California	2009	Intertidal (rockpool)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-16.3(1)‰	This study
	<i>C. cupressoides</i>	Great barrier reef, Australia	2008-2012		HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	-10.5±3.4(4)‰	Lovelock et al., (2019)
	<i>C. flexilis</i>	Shortlands Bluff, Australia	2002-2003	Rock-pool faces, partially sheltered	CO <sub>2</sub>	pH drift experiments, δ <sup>13</sup> C signal	Final pH~8.9, -30.2(1)‰	Kevekordes et al., (2006)
	<i>C. flexilis</i>	Tazmania	2013-2014	Subtidal	CO <sub>2</sub>	δ <sup>13</sup> C signal	-30.6±1.1(2)‰	Cornwall et al., (2015)
	<i>C.hodgkinsoniae</i>	Tazmania	2013-2014	Subtidal	CO <sub>2</sub>	δ <sup>13</sup> C signal	-25.6±1(2)‰	Cornwall et al., (2015)
	<i>C. longifolia</i>	Shortlands Bluff, Australia	2002-2003	Rock-pool faces, partially sheltered	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	pH drift experiments, δ <sup>13</sup> C signal	Final pH~8.9, -28.55(1)‰	Kevekordes et al., (2006)
	<i>C. mexicana</i>	Gulf of California	2009	Subtidal (<2m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-19.9(1)‰	This study
	<i>C. obscura</i>	Shortlands Bluff, Australia	2002-2003	Rock-pool faces, partially sheltered	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	pH drift experiments, δ <sup>13</sup> C signal	Final pH~8.9, -29.5(1)‰	Kevekordes et al., (2006)
	<i>C. peltata</i>	Gulf of California	2013	Subtidal (<2m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-10.7±4.5(3)‰	This study
	<i>C. prolifera</i>	Cádiz, southern Spain	2004-2005	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-12.4±0.4(3)‰	Mercado et al., (2009)
	<i>C. prolifera</i>	Mediterranean	2014	Subtidal (2m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-14.9±1.5(3)‰	Cornwall et al., (2017)
	<i>C. racemosa</i>	Egipt	2009	Subtidal (2m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-16.5±0.1(3) ‰	Marconi et al., (2011)
	<i>C. racemosa</i>	Mediterranean	2014	Subtidal (2m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-16.1±1.3(2)‰	Cornwall et al., (2017)
	<i>C. racemosa</i>	Great barrier reef, Australia	2008-2012	Intertidal (0m) to Subtidal (0.5-4.8m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-13.5±3.8(6)‰	Lovelock et al., (2019)
	<i>C. scalpelliformis</i>	Shortlands Bluff, Australia	2002-2003	Rock-pool faces, partially sheltered	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments. δ <sup>13</sup> C signal	Final pH~9, -19.4(1)‰	Kevekordes et al., (2006)
	<i>C. scalpelliformis</i>	Tazmania	2013-2014	Subtidal (5m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-17.8(1)‰	Cornwall et al., (2015)
	<i>C. serrulata</i>	Egypt	2013-2014	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-12.21±0.03(3)‰	Marconi et al., (2011)
	<i>C. serrulata</i>	Great barrier reef, Australia	2008-2012	Intertidal (0m) to Subtidal (0.5-5.5m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-12.0±3.3(20)‰	Lovelock et al., (2019)

<i>C. sertularioides</i>	Gulf of California	2013	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-18.5±0.1(2)‰	This study
<i>Caulerpa sp.</i>	Mexican Caribbean	2016	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-11.4±3.0(5)‰	Cabanillas-Teran et al., (2019)
<i>C. taxifolia</i>	Great barrier reef, Australia	2008-2012	Subtidal(10-25m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-18.31±0.7‰	Marconi et al., (2011)
<i>C. taxifolia</i>			Subtidal (5.2m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-12.1±5.1(6)‰	Lovelock et al., (2019)
<i>C. trifaria</i>	Shortlands Bluff, Australia	2002-2003		CO <sub>2</sub>	δ <sup>13</sup> C signal	-31.2(1)‰	Kevekordes et al., (2006)
<i>C. trifaria</i>	Southern, Astralia	2013-2014	Subtidal (5-15m)	CO <sub>2</sub>	δ <sup>13</sup> C signal	-33±1.7(4)‰	Cornwall et al., (2015)
<i>Chaetomorpha aerea</i>	Cádiz, southern Spain	1995		CO <sub>2</sub>	Measurement of the external Carbonic Anhydrase (CA) activity and photosynthetic rates with addition of acetazolamide (AZ) an inhibitor of CA activity.	The rate of O <sub>2</sub> evolution is ceased when CO <sub>2</sub> concentration dismiss. CA activity no detected	Mercado et al., (1997)
<i>C. antennina</i>	Gulf of California	2009,2913	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-14.5±1.07(10)‰	This study
<i>C. coliformis</i>	Southern, Astralia	2013-2014	Subtidal (5m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-20.6(1)‰	Cornwall et al., (2015)
<i>C. linun</i>	Denmark		upper sublittoral to 20 m	HCO <sub>3</sub> <sup>-</sup>	Photosynthetic rates at different pH	Low photosynthetic rates >9	Sand-Jensen and Gordon (1984)
<i>C. linun</i>	Kristineberg, Sweden		Subtidal (0.5-5m)	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>10	Axelsson and Uusitalo (1988)
<i>C. linun</i>	Gulf of California	2009,2013	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-16.8±1.6(4)‰	This study
<i>Chaetomorpha sp.</i>	Gulf of California	2009,2013	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-13.7±0.8(3)‰	This study
<i>Cladophora albida</i>	Gulf of California	2009	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-19.4±6.1(2)‰	This study
<i>C. columbiana</i>	Gulf of California	2009	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-13.7(1)‰	This study
<i>C. laetevirens</i>	Cádiz, southern Spain	2004-2005	Intertidal (rockpool)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-15.6±0.8(3)‰	Mercado et al., (2009)
<i>C. microcladioides</i>	Gulf of California	2009	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-18.6±3.6(2)‰	This study
<i>Cladophora sp.</i>	Gulf of California UK	2008,2009,2013	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-12.8± 2.6(7)‰	This study
<i>Cladophora sp.</i>				HCO <sub>3</sub> <sup>-</sup>	pH drift experiments, δ <sup>13</sup> C signal	Final pH>10 -14.7±1(15)‰	Marconi et al., (2011)
<i>C. glomerata</i>	northern Baltic Sea	2002	Subtidal (0.3m)	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>10	Choo et al (2005)

<i>C. rupestris</i>	Kristineberg, Sweden		Subtidal (0.5-5m)	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>10	Axelsson and Uusitalo (1988)
<i>C. rupestris</i>	Scotland		Subtidal	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>10	Maberly (1990)
<i>C. sericea</i>	Scotland		Subtidal	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>10	Maberly (1990)
<i>Cladophora</i> sp.	Southeast France	2013		HCO <sub>3</sub> <sup>-</sup>	pH compensation point	Final pH>10	Maberly et al., (2015)
<i>Codium adharens</i>	Cádiz, southern Spain	2005-2005	Intertidal	HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	-7.7±0.8‰	Mercado et al., (2009)
<i>C. amplivesiculatum</i>	Gulf of California	2008, 2009	Subtidal (<2m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-14.4±2.7(8)‰	This study
<i>C. brandegeei</i>	Gulf of California	2009	Subtidal (<2m)	HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	-11.8±1.2(7)‰	This study
<i>C. bursa</i>	Mediterranean	2014	Subtidal (2m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-21.2±1.2(3)‰	Cornwall et al., (2017)
<i>C. bursa</i>	Cádiz, southern Spain	2005-2005	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-12.4±0.8(3)‰	Mercado et al., (2009)
<i>C. decortiatum</i>	Cádiz, southern Spain	2005-2005	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-12.0±1.0(3)‰	Mercado et al., (2009)
<i>C. fragile</i>	Gulf of California	2009	Subtidal (2m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-13.0±2.7(4)‰	This study
<i>C. fragile</i>	Kristineberg, Sweden		Sublittoral (0.5-2m)	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>10	Axelsson and Uusitalo (1988)
<i>C. fragile</i>	Scotland		Subtidal	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>9.5	Maberly (1990)
<i>C. fragile</i>	Cádiz, southern Spain	1995	Intertidal and rockpools	HCO <sub>3</sub> <sup>-</sup>	Measurements of O <sub>2</sub> evolution curves vs. Ci concentration and measurement of the external Carbonic Anhydrase (CA) activity, with addition of acetazolamide (AZ) and/or 6-ethoxyzolamide (EZ) an inhibitors of CA activity.	The rate of O <sub>2</sub> evolution is ceased when CO <sub>2</sub> concentration dismiss. CA activity no detected	Mercado et al., (1997)
<i>C. muelleri</i>	Australia	2009	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-13.2±0.0(3) ‰	Marconi et al., (2011)
<i>C. simulans</i>	Gulf of California	2009	Subtidal (2m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-12.4±2.2(9)‰	This study
<i>Codium</i> sp.	Gulf of California	2009, 2013	Subtidal (2m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-11.6±3.0(5)‰	This study
<i>Codium</i> sp.	Great barrier reef, Australia	2008-2012	Subtidal (0.5-1.5m)	HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	-11.7±2.4(8)‰	Lovelock et al., (2019)

<i>Ulva acantophora</i>	Gulf of California	2008,2009	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	15.8±1.7(25)‰	This study
<i>U. clathrata</i>	Gulf of California	2008,2009	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-16.4±2.0(8)‰	This study
<i>U. compressa</i>	Cádiz, southern Spain	1995		HCO <sub>3</sub> <sup>-</sup>	Measurement of the external Carbonic Anhydrase (CA) activity and photosynthetic rates with addition of acetazolamide (AZ) an inhibitor of CA activity.	Photosynthesis depend of CO <sub>2</sub> and HCO <sub>3</sub> <sup>-</sup> with equal affinity. Low CA activity detected	Mercado et al., (1997)
<i>U. comglobata</i>	China		Intertidal	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>10	Zou (2014)
<i>U. flexuosa</i>	Lizard Island, Queensland, Australia	1973	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-21.20(1)‰	Black and Bender (1976)
<i>U. flexuosa</i>	Gulf of California	2008,2009	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-16.0±3.7(13)‰	This study
<i>U. intestinalis</i>	Kristineberg, Sweden		tidal pools (0-1m)	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>10	Axelsson and Uusitalo (1988)
<i>U. intestinalis</i>	Scotland		Subtidal	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>10	Maberly (1990)
<i>U. intestinalis</i>	Cádiz, southern Spain	2005-2005	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-16.01±0.01(3)‰	Mercado et al., (2009)
<i>U. intestinalis</i>	Gulf of California	2008,2009,2013	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-15.3±2.5(16)‰	This study
<i>U. lactuca</i>	Scotland		Subtidal	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>10	Maberly (1990)
<i>U. lactuca</i>	Denmark			HCO <sub>3</sub> <sup>-</sup>	Photosynthetic rates at different pH	Low photosynthetic rates at pH >9	Sand-Jensen and Gordon (1984)
<i>U. lactuca</i>	Argentina	2007-2008		HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>		-15.4±0.7(2)‰	Becherucci et al., (2019)
<i>U. lobata</i>	Gulf of California	2008,2009,2013	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-13.2±1.9(5)‰	This study
<i>U. procera</i>	China		Subtidal (0.3m)	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>10	Choo et al (2005)
<i>Ulva</i> sp.	Cádiz, southern Spain	2005-2005	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-18.8±0.1(3)‰	Mercado et al., (2009)
<i>Ulva</i> sp.	Portugal	2005-2006	Intertidal	HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	-12.5±1.0(16)	Baeta et al., (2009)
<i>Ulva</i> sp.				HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>10	Cornwall et al., (2012)
<i>Ulva</i> sp.	Great barrier reef, Australia	2008-2012	Intertidal (0m), subtidal (3m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>		-13.1±4.7(21)‰	Lovelock et al., (2019)
<i>Ulva</i> sp.			Subtidal (0.3m)	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>10	Marconit et al., (2011)

	<i>Ulva</i> sp. <i>U. rigida</i>	Ballina, Australia Cádiz, southern Spain			$\text{HCO}_3^-$ $\text{CO}_2 > \text{HCO}_3^-$	$\delta^{13}\text{C}$ signal Measurement of the external Carbonic Anhydrase (CA) activity and photosynthetic rates with addition of acetazolamide (AZ) an inhibitor of CA activity.	$-12.23 \pm 0.32(3)\text{‰}$ The evolution of $\text{O}_2$ evolution was higher with $\text{CO}_2$ concentrations than $\text{HCO}_3^-$ .	Carvalho and Eyre, (2011) Mercado et al., (1997)
	<i>U. rigida</i>	Cádiz, southern Spain	1995		$\text{CO}_2 = \text{HCO}_3^-$	Measurement of the external Carbonic Anhydrase (CA) activity and photosynthetic rates with addition of acetazolamide (AZ) an inhibitor of CA activity.	Photosynthesis depend of $\text{CO}_2$ and $\text{HCO}_3^-$ with equal affinity. Low CA activity detected	Mercado et al., (1997)
Ochrophyta	<i>U. rigida</i>	Gulf of California	2008,2009,2013	Intertidal	$\text{HCO}_3^- / \text{CO}_2$	$\delta^{13}\text{C}$ signal	$-13.7(1)\text{‰}$	This study
	<i>C. peregrina</i>	Cádiz, southern Spain	2004-2005			$\delta^{13}\text{C}$ signal	$-10.4 \pm 0.7(3)\text{‰}$	Mercado et al., (2009)
	<i>C. ramosa</i>	Gulf of California	2008,2009	Intertidal	$\text{HCO}_3^- / \text{CO}_2$	$\delta^{13}\text{C}$ signal	$-11.4 \pm 2.6(4)\text{‰}$	This study
	<i>Colpomenia</i> sp.	Great barrier reef, Australia	2008-2012	Intertidal (0.5m)	$\text{HCO}_3^-$	$\delta^{13}\text{C}$ signal	$-8.9 \pm 3.5(8)\text{‰}$	Lovelock et al., (2019)
	<i>C. sinuosa</i>	Cádiz, southern Spain	1995	Intertidal and rockpools	$\text{HCO}_3^-$	Measurements of $\text{O}_2$ evolution curves vs. $\text{C}_i$ concentration and measurement of the external Carbonic Anhydrase (CA) activity, with addition of acetazolamide (AZ) and/or 6-ethoxyzolamide	High initial slope of the $\text{O}_2$ evolution rate vs. $\text{C}_i$ curve	Mercado et al., (1998)



<i>C. sinuosa</i>	Gulf of California	2008,2009	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	(EZ) an inhibitors of CA activity. δ <sup>13</sup> C signal	-10.2±3(7)‰	This study
<i>C. tuberculata</i>	Gulf of California	2008,2009, 2013	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-8.7 ±3.2(65)‰	This study
<i>Dictyota concrescen</i>	Gulf of California	2009	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-16.5‰	This study
<i>Dictyota dichotoma</i>	Cádiz, southern Spain	1995	Intertidal and rockpools	CO <sub>2</sub> > HCO <sub>3</sub> <sup>-</sup>	Measurements of O <sub>2</sub> evolution curves vs. Ci concentration and measurement of the external Carbonic Anhydrase (CA) activity, with addition of acetazolamide (AZ) and/or 6-ethoxyzolamide (EZ) an inhibitors of CA activity.	Low O <sub>2</sub> evolution rates at pH 8.7 and low activity of CA detected	Mercado et al., (1998)
<i>D. dichotoma</i>	Mediterranean	2014	Subtidal (2m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-21.5±2.4(4)‰	Cornwall et al., (2017)
<i>D. dichotoma</i>	Cádiz, southern Spain	2004-2005	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-15.9±0.5(3)‰	Mercado et al., (2009)
<i>D. dichotoma</i>	Gulf of California	2009	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-22.2±6.2(2)‰	This study
<i>D. dichotoma implexa</i>	Cádiz, southern Spain	2004-2005	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-18.6±0.8(3)‰	Mercado et al., (2009)
<i>D. implexa</i>	Gulf of California	2009	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-17.3±3.0(2)‰	This study
<i>D. pinnatifida</i>	Florida, EEUU	2003, 2005	Subtidal (<1m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-16.2±0.6(87)‰	Lamb et al. 2012
<i>Dictyota</i> sp.	Mexican Caribbean	2016	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-15.9±0.4(6)‰	Cabanillas-Teran et al., (2019)
<i>Dictyota</i> sp.	Great barrier reef, Australia	2008-2012	Subtidal (1-12.6m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-14.7±3.2(35)‰	Lovelock et al., (2019)
<i>Hydroclathrus clathratus</i>	Great barrier reef, Australia	2008-2012	Intertidal (0m), Subtidal (0.5-3m)	HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	-10.4±3.2(13)‰	Lovelock et al., (2019))
<i>H. clathratus</i>	Gulf of California	2009	Intertidal	HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	-7.2(1)‰	This study
<i>Padina crispate</i>	Gulf of California	2008,2009, 2013	Intertidal	HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	-11.3±1.7(2)‰	This study
<i>P. durvillei</i>	Gulf of California	2008,2009, 2013	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-13.2±2.6(36)‰	This study
<i>P. tenuis</i>	Lizard Island, Queensland, Australia	1973	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-11.40(1)‰	Black and Bender (1976)
<i>P. pavonia</i>	Cádiz, southern Spain	1995	Intertidal and rockpools	CO <sub>2</sub> > HCO <sub>3</sub> <sup>-</sup>	Measurements of O <sub>2</sub> evolution	low O <sub>2</sub> evolution rates at pH 8.7	Mercado et al., (1998)

					curves vs. Ci concentration and measurement of the external Carbonic Anhydrase (CA) activity, with addition of acetazolamide (AZ) and/or 6-ethoxzolamide (EZ) an inhibitors of CA activity.	and No activity of CA detected		
	<i>Padina pavonia</i>	Mediterranean	2014	Subtidal (2m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-16.4±4.5(5)‰	Cornwall et al., (2017)
	<i>Padina</i> sp.	Great barrier reef, Australia	2008-2012	Intertidal (0m), Subtidal (0.5-12.6m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-10.7±4.4(51)‰	Lovelock et al., (2019))
	<i>Padina</i> sp.	Bolinao, Pangasinan	2020		HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH=9.89	Narvarte et al., (2020)
	<i>Padina</i> sp.	Gulf of California	2009	Intertidal	HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	-11.1±1.5(15)‰	This study
	<i>Sargassum flavicans</i>	Great barrier reef, Australia	2008-2012	Subtidal (0.5-6.7m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-15.5±2.3(16)‰	Lovelock et al., (2019))
	<i>S. henslowianum</i>	China	2007	Subtidal	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH=9.3	Zou et al., (2011)
	<i>S. lacerifolium</i>	Tasmania	2014	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-18.72±1.41(3)‰	Cornwall et al., (2015)
	<i>S. muticum</i>	Mediterranean	2014	Subtidal (2m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-19.7±0.5(6)‰	Cornwall et al., (2017)
	<i>Sargassum</i> sp.	Lizard Island, Queensland, Australia	1973	Subtidal	HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	-14.10(1)‰	Black and Bender (1976)
	<i>Sargassum</i> sp.	Great barrier reef, Australia	2008-2012	Intertidal (0m), Subtidal (0.5-3.3m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-14.9±2.4(27)‰	Lovelock et al., (2019))
	<i>Sargassum</i> sp.	Ballina, Australia			HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	-12.8±0.7(3) ‰	Carvalho and Eyre, (2011)
	<i>S. spinifex</i>	Great barrier reef, Australia	2008-2012	Subtidal (0.5m)		δ <sup>13</sup> C signal	-15.1±1.0(6)‰	Lovelock et al., (2019)
	<i>S. vulgare</i>	Cádiz, southern Spain	2004-2005	Subtidal		δ <sup>13</sup> C signal	-15.4±0.1(3)‰	Mercado et al., (2009)
Rhodophyta	<i>Acanthophora</i> sp.	Gulf of California	2009	Intertidal	HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	-16.3±0.5(2) ‰	This study
	<i>Acanthophora spicifera</i>	Lizard Island, Queensland, Australia	1973	Subtidal	HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	-10.01(1)‰	Black and Bender (1976)
	<i>A.spicifera</i>	Great barrier reef, Australia	2008-2012	Subtidal (0.5-6.6m)	HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	-12.1±2.8(11)‰	Lovelock et al., (2019)

<i>Acanthophora spicifera</i>	Gulf of California	2009	Intertidal	HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	-14.5(1) ‰	This study
<i>Ahnfeltiopsis leptophyllus</i>	San Juan Island, USA		Subtidal	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH=9.56	Murru and Sandgreen (2004)
<i>A. leptophyllus</i>	Gulf of California	2008, 2009	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-14.4±2.4(7)‰	This study
<i>Amphiroa anceps</i>	Australia			HCO <sub>3</sub> <sup>-</sup>	Measurement of photosynthetic rates at different pH	High photosynthetic rates at pH>9	Borowitzka (1981)
<i>A.beauvoisii</i>	Gulf of California	2009	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-8.6±0.4(2)‰	This study
<i>Amphiroa</i> sp.	Gulf of California	2009	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-6.5± 1.1(9)‰	This study
<i>Bostrychia radicans</i>	Gulf of California	2009	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	C1	-25.8‰	This study
<i>B. scorpioides</i>	Cádiz, southern Spain	1998	Subtidal	CO <sub>2</sub> > HCO <sub>3</sub> <sup>-</sup>	Measurement of the external Carbonic Anhydrase (CA) activity, with addition of acetazolamide (AZ) and/or 6-ethoxyzolamide (EZ) an inhibitors of CA activity.		Mercado and Niell (2000)
<i>B. scorpioides</i>	Cádiz, southern Spain	2004-2005	Subtidal	CO <sub>2</sub>		-30.9±0.4(3)‰	Mercado et al., (2009)
<i>Ceramium pacificum</i>	San Juan Island, USA		Subtidal	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH=10.15	Murru and Sandgreen (2004)
<i>Ceramium rubrum</i>	Kristineberg, Sweden		littoral to sublittoral (0.5-20 m)	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH= 9.6	Axelsson and Uusitalo (1988)
<i>Ceramium rubrum</i>	Roskilde Fjord			HCO <sub>3</sub> <sup>-</sup>	Photosynthetic rates at different pH	Low photosynthetic rates at pH>9	Sand-Jensen and Gordon (1984)
<i>Ceramium rubrum</i>	Scotland		Subtidal	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>9.7	Maberly (1990)
<i>Ceramium</i> sp.	west coast of Sweden			HCO <sub>3</sub> <sup>-</sup>	Photosynthetic rates and their effect (inhibition in percentages) by addition of acetazolamide (AZ), TRIS buffer at pH 8.7 and both.	Photosynthetic rate was inhibited by TRIS and TRIS + AZ.	Moulin et al., (2011)

<i>Ceramium</i> sp.	Gulf of California	2013	Subtidal and Intertidal	HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	-12.5±2.7‰	This study
<i>Chondracanthus acicularis</i>	France		upper inter-tidal and medium inter-tidal	HCO <sub>3</sub> <sup>-</sup>	Measurements of photosynthetic rates and their inhibition effect (%) by acetazolamide (AZ), TRIS buffer at pH 8.7 and both.	Photosynthetic rate was inhibited by TRIS and TRIS + AZ.	Moulin et al., (2011)
<i>C. acicularis</i>	Cádiz, southern Spain	2004-2005		HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-19.9±1.2(3)‰	Mercado et al., (2009)
<i>C. exasperatus</i>	San Juan Island, USA		Subtidal	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Ph=9.71	Murru and Sandgreen (2004)
<i>C. squarulosus</i>	Gulf of California	2009	Strategy 2	S	P2	-12.6‰	This study
<i>C. tepidus</i>	Gulf of California	2009	Strategy 2	S	P2	-17.6‰	This study
<i>Chondria incrassata</i>			Subtidal (15m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-20.7(1)‰	Cornwall et al., (2015)
<i>C. acrorhizophora</i>	Gulf of California	2009	Intertidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-14.2(2)‰	This study
<i>Corallina officinalis</i>	Cádiz, southern Spain	2004-2005		HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-15.8±0.5(3)‰	Mercado et al., (2009)
<i>Corallina officinalis</i>	Scotland		Subtidal	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>10	Maberly (1990)
<i>Corallina officinalis</i>	South Island, New Zealand	2009		HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	pH drift experiments	pH<9	Cornwall et al., (2012)
<i>Corallina sp.</i>	Cádiz, southern Spain		Upper intertidal and medium inter-tidal	HCO <sub>3</sub> <sup>-</sup>	Measurements of photosynthetic rates and their inhibition effect (%) by acetazolamide (AZ), TRIS buffer at pH 8.7 and both.	Photosynthetic rate was inhibited by TRIS and TRIS + AZ.	Moulin et al., (2011)
<i>Corallina elongata</i>	Cádiz, southern Spain	1995		CO <sub>2</sub> > HCO <sub>3</sub> <sup>-</sup>	Measurement of the external Carbonic Anhydrase (CA) activity, with addition of acetazolamide	The evolution of O <sub>2</sub> evolution was higher with CO <sub>2</sub> concentrations than HCO <sub>3</sub> <sup>-</sup> . The activity of CA was low	Mercado et al., (1997)

<i>C. vancouverensis</i>	Gulf of California	2008, 2009	Subtidal	$\text{HCO}_3^- / \text{CO}_2$	(AZ) an inhibitor of CA activity. $\delta^{13}\text{C}$ signal	-8.3(1)‰	This study
<i>Gelidium latifolium</i>	west coast of Sweden			$\text{HCO}_3^-$	Measurements of photosynthetic rates and their inhibition effect (%) by acetazolamide (AZ), TRIS buffer at pH 8.7 and both.	Photosynthetic rate was inhibited by TRIS and TRIS + AZ.	Moulin et al., (2011)
<i>G. pusillum</i>	Cádiz, southern Spain	1995	Intertidal and rockpools	$\text{CO}_2 > \text{HCO}_3^-$	Measurements of $\text{O}_2$ evolution curves vs. $\text{C}_i$ concentration and measurement of the external Carbonic Anhydrase (CA) activity, with addition of acetazolamide (AZ) and/or 6-ethoxyzolamide (EZ) an inhibitors of CA activity.	Low $\text{O}_2$ evolution rates at pH 8.7 and no activity of CA detected.	Mercado et al., (1998)
<i>G. sesquipedale</i>	Cádiz, southern Spain	1995	Intertidal and rockpools	$\text{CO}_2 > \text{HCO}_3^-$	Measurements of $\text{O}_2$ evolution curves vs. $\text{C}_i$ concentration and measurement of the external Carbonic Anhydrase (CA) activity, with addition of acetazolamide (AZ) and/or 6-ethoxyzolamide (EZ) an	Low $\text{O}_2$ evolution rates at pH 8.7 and no activity of CA detected.	Mercado et al., (1998)

<i>G. sclerophyllum</i> <i>G. spinosum</i>	Gulf of California Cádiz, southern Spain	2008, 2009 2004-2005	Subtidal Subtidal	HCO <sub>3</sub> <sup>-</sup> /CO <sub>2</sub> HCO <sub>3</sub> <sup>-</sup> /CO <sub>2</sub>	inhibitors of CA activity. δ <sup>13</sup> C signal δ <sup>13</sup> C signal	-15.2±3.1(2)‰ -17.3±0.7(3)‰	This study Mercado et al., (2009)
<i>Gigartina</i> <i>exasperata</i>	California			HCO <sub>3</sub> <sup>-</sup>	Comparing photosynthetic rates of O <sub>2</sub> evolution with the photosynthetic rate which could be supported by CO <sub>2</sub> arising from the uncatalysed dehydration of HCO <sub>3</sub> <sup>-</sup> in the medium	At pH 9, the rates of photosynthetic O <sub>2</sub> evolution exceeded the CO <sub>2</sub> supply rate	Cook et al., (1986)
<i>G. papillata</i>	California			HCO <sub>3</sub> <sup>-</sup>	Comparing photosynthetic rates of O <sub>2</sub> evolution with the photosynthetic rate which could be supported by CO <sub>2</sub> arising from the uncatalysed dehydration of HCO <sub>3</sub> <sup>-</sup> in the medium	At pH 9, the rates of photosynthetic O <sub>2</sub> evolution exceeded the CO <sub>2</sub> supply rate	Cook et al., (1986)
<i>Gigartina sp.</i>	Gulf of California	2008, 2009	Subtidal	HCO <sub>3</sub> <sup>-</sup> /CO <sub>2</sub>	δ <sup>13</sup> C signal	-16.9±6.6(9)‰	This study
<i>Gracilaria</i> <i>gracilis</i>	west coast of Sweden			HCO <sub>3</sub> <sup>-</sup>	Measurements of photosynthetic rates and their inhibition effect (%) by acetazolamide (AZ), TRIS	Photosynthetic rate was inhibited by TRIS and TRIS + AZ.	Moulin et al., (2011)

<i>G. lemaneiformis</i>	China			HCO <sub>3</sub> <sup>-</sup>	buffer at pH 8.7 and both. pH drift experiments	pH final = 9.58	Zou et al., (2004)
<i>G. pacifica</i>	San Juan Island, USA		Subtidal	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	pH final= 9.43	Murru and Sandgreen (2004)
<i>G. secundata</i>	Southern, Australia	2013-2014	Subtidal	CO <sub>2</sub>	δ <sup>13</sup> C signal	-32.4±0.7‰	Cornwall et al., (2015)
<i>G.subsecundata</i>	Gulf of California	2008,2009	Subtidal	HCO <sub>3</sub> <sup>-</sup> /CO <sub>2</sub>	δ <sup>13</sup> C signal	-15.9±2.8(8)‰	This study
<i>Gracilaria sp.</i>	Great barrier reef, Australia	2008-2012	Subtidal (0.5-4.9m)	HCO <sub>3</sub> <sup>-</sup> /CO <sub>2</sub>	δ <sup>13</sup> C signal	-19.8±1.5‰	Loveloock et al., (2019)
<i>G. vermiculophylla</i>	Gulf of California	2009,2013	Subtidal	HCO <sub>3</sub> <sup>-</sup> /CO <sub>2</sub>	δ <sup>13</sup> C signal	-15.9±3.8(23)‰	This study
<i>G. tenuistipitata</i>	Cádiz, southern Spain	1995		CO <sub>2</sub> > HCO <sub>3</sub> <sup>-</sup>	Measurement of the external Carbonic Anhydrase (CA) activity, with addition of acetazolamide (AZ) an inhibitor of CA activity.	The evolution of O <sub>2</sub> was higher with CO <sub>2</sub> concentrations than HCO <sub>3</sub> <sup>-</sup> . The activity of CA was low.	Mercado et al., (1997)
<i>Gymnogongrus crenulatus</i>	Cádiz, southern Spain	2004-2005	Subtidal	HCO <sub>3</sub> <sup>-</sup> /CO <sub>2</sub>	δ <sup>13</sup> C signal	-22.3±1.1‰	Mercado et al., (2009)
<i>G. guadalupensis</i>	Gulf of California	2009	Subtidal	HCO <sub>3</sub> <sup>-</sup> /CO <sub>2</sub>	δ <sup>13</sup> C signal	-13.1(1)‰	This study
<i>G. linearis</i>	California			HCO <sub>3</sub> <sup>-</sup>	Comparing photosynthetic rates of O <sub>2</sub> evolution with the photosynthetic rate which could be supported by CO <sub>2</sub> arising from the uncatalysed dehydration of HCO <sub>3</sub> <sup>-</sup> in the medium	At pH 9, the rates of photosynthetic O <sub>2</sub> evolution exceeded the CO <sub>2</sub> supply rate	Cook et al., (1986)
<i>Gymnogongrus spp.</i>	Cádiz, southern Spain	1995	Intertidal and rockpools	CO <sub>2</sub> > HCO <sub>3</sub> <sup>-</sup>	Measurements of O <sub>2</sub> evolution curves vs. Ci concentration and	Low O <sub>2</sub> evolution rates at pH 8.7 and low activity of CA detected.	Mercado et al., (1998)

<i>Gymnogongrus sp.</i>	Gulf of California	2009	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	measurement of the external Carbonic Anhydrase (CA) activity, with addition of acetazolamide (AZ) and/or 6-ethoxyzolamide (EZ) an inhibitors of CA activity. δ <sup>13</sup> C signal	-14.1±3.8(11)‰	This study
<i>Halymenia duruillaei</i>	Lizard Island, Queensland, Australia	1973	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-32.0(1)‰	Black and Bender (1976)
<i>H. gardneri</i>	San Juan Island, USA			HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	pH final=9.35	Murru and Sandgreen (2004)
<i>H. schizymenioides</i>	San Juan Island, USA			HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	pH final=9.38	Murru and Sandgreen (2004)
<i>Halymenia sp.</i>	Great barrier reef, Australia	2008-2012	Subtidal (1.5-5.1m)	CO <sub>2</sub>	δ <sup>13</sup> C signal	-27.7±6.8‰	Lovelock et al., (2019)
<i>Halymenia sp.</i>	Southern, Australia	2013-2014	Subtidal (15m)	CO <sub>2</sub>	δ <sup>13</sup> C signal	-35.1±0.3(2)‰	Cornwall et al., (2015)
<i>Halymenia sp.</i>	Gulf of California	2009	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-23.7±13.6(2)‰	This study
<i>Hypnea musciformis</i>	Cádiz, southern Spain	2004-2005			δ <sup>13</sup> C signal	-15.5±0.8(3)‰	Mercado et al., (2009)
<i>Hypnea pannosa</i>	Great barrier reef, Australia	2008-2012	Subtidal (0.5-4m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-14.4±2.8‰	Lovelock et al., (2019)
<i>Hypnea spinella</i>	Gulf of California	2009	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-16.4±1.8(6)‰	This study
<i>Hypnea sp.</i>	Great barrier reef, Australia	2008-2012	Intertidal (0m), Subtidal (0.5-3.7m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-18.2±2.4‰	Lovelock et al., (2019)
<i>H. valentiae</i>	Gulf of California	2009	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-15.2±2.3(6)‰	This study
<i>Jania sp.</i>	Great barrier reef, Australia	2008-2012	Subtidal (0.5-9.8m)	HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	8.5±2.0‰	Lovelock et al., (2019)
<i>Jania sp.</i>	Gulf of California	2009	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-9.5±1.0(2)‰	This study
<i>Laurencia pacifica</i>	Gulf of California	2009	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-14.9±2.2(8)‰	This study
<i>L. pinnafitida</i>	Scotland		Subtidal	HCO <sub>3</sub> <sup>-</sup>	pH drift experiments	Final pH>9.7	Maberly (1990)
<i>Laurencia sp.</i>	Great barrier reef, Australia	2008-2012	Subtidal (0.5-7.1m)	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-16.0±4.7‰	Lovelock et al., (2019)
<i>Polysiphonia elongata</i>	west coast of Sweden		upper inter-tidal and medium inter-tidal	HCO <sub>3</sub> <sup>-</sup>	Measurements of photosynthetic rates and their	Photosynthetic rate was inhibited by TRIS and TRIS + AZ.	Moulin et al., (2011)



<i>P. fucooides</i>	west coast of Sweden		upper inter-tidal and medium inter-tidal	HCO <sub>3</sub> <sup>-</sup>	inhibition effect (%) by acetazolamide (AZ), TRIS buffer at pH 8.7 and both. Measurements of photosynthetic rates and their inhibition effect (%) by acetazolamide (AZ), TRIS buffer at pH 8.7 and both.	Photosynthetic rate was inhibited by TRIS and TRIS + AZ.	Moulin et al., (2011)
<i>P. mollis</i> <i>P. lanosa</i>	Gulf of California Scotland	2009	Subtidal Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub> HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal pH drift experiments	-14.0±7.9(3)‰ Final pH>10	This study Maberly (1990) Marconi et al., (2011)
<i>P. pacifica</i> <i>P. stricta</i>	Gulf of California west coast of Sweden	2009	Subtidal upper inter-tidal and medium inter-tidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub> HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal Measurements of photosynthetic rates and their inhibition effect (%) by acetazolamide (AZ), TRIS buffer at pH 8.7 and both.	-19.8±1.8(7)‰ Photosynthetic rate was inhibited by TRIS and TRIS + AZ.	This study Moulin et al., (2011)
<i>P. violaceae</i>	Denmark			HCO <sub>3</sub> <sup>-</sup>	Photosynthetic rates at different pH	Low Photosynthetic rates >9	Sand-Jensen and Gordon (1984)
<i>Prionitis lanceolata</i> <i>P. lyalli</i>	San Juan Island, USA San Juan Island, USA		Subtidal Subtidal	HCO <sub>3</sub> <sup>-</sup> HCO <sub>3</sub> <sup>-</sup>	pH drift experiments pH drift experiments	pH final=9.46 pH final=9.55	Murru and Sandgreen (2004) Murru and Sandgreen (2004)
<i>P. abbreviata</i> var <i>guaymanensis</i> <i>Prionitis</i> sp.	Gulf of California Gulf of California	2009 2009	Subtidal Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub> HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal δ <sup>13</sup> C signal	-11.4(1)‰ -14.4±1.1(2)‰	This study This study
<i>Rhodoglossum affine</i>	California			HCO <sub>3</sub> <sup>-</sup>	Comparing photosynthetic rates of O <sub>2</sub> evolution with the photosynthetic	At pH 9, the rates of photosynthetic O <sub>2</sub> evolution exceeded the CO <sub>2</sub> supply rate	Cook et al., (1986)

						rate which could be supported by CO <sub>2</sub> arising from the uncatylsed dehydration of HCO <sub>3</sub> <sup>-</sup> in the medium		
<i>Rhodoglossum sp.</i>	Gulf of California	2009	Subtidal	HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-16.1±1.0‰	This study	
<i>Rhodymenia palmata</i>	California			HCO <sub>3</sub> <sup>-</sup>	Comparing photosynthetic rates of O <sub>2</sub> evolution with the photosynthetic rate which could be supported by CO <sub>2</sub> arising from the uncatylsed dehydration of HCO <sub>3</sub> <sup>-</sup> in the medium	At pH 9, the rates of photosynthetic O <sub>2</sub> evolution exceeded the CO <sub>2</sub> supply rate	Cook et al., (1986)	
<i>R. asutralis</i>	Australia	1999-2000		CO <sub>2</sub>	δ <sup>13</sup> C signal	-32.2±0.7‰	Raven et al., (2002a)	
<i>Rhodymenia sp.</i>	Italy			CO <sub>2</sub>	δ <sup>13</sup> C signal	-29.2±0.5‰	Marconi et al., (2011)	
<i>Rhodymenia sp.</i>	Gulf of California	2009	Subtidal	HCO <sub>3</sub> <sup>-</sup>	δ <sup>13</sup> C signal	18.7±2.4(2)‰	This study	
<i>Schizymenia pacifica</i>	San Juan Island, USA		Subtidal	CO <sub>2</sub>	pH drift experiments	pH final=9.02	Murru and Sandgreen (2004)	
<i>S. pacifica</i>	Catalina Island, CA, USA	2000		HCO <sub>3</sub> <sup>-</sup> / CO <sub>2</sub>	δ <sup>13</sup> C signal	-18.46(1) ‰	Raven et al., (2002a)	
<i>S. pacifica</i>	Gulf of California	2009	Subtidal	CO <sub>2</sub>	δ <sup>13</sup> C signal	-33.8±1.2(2)‰	This study	

Table S5. Temperature mean by regions and seasons in the Gulf of California based on time series data (1981 to 2016) by Escalante et al. (2013) and Robles-Tamayo (2018).

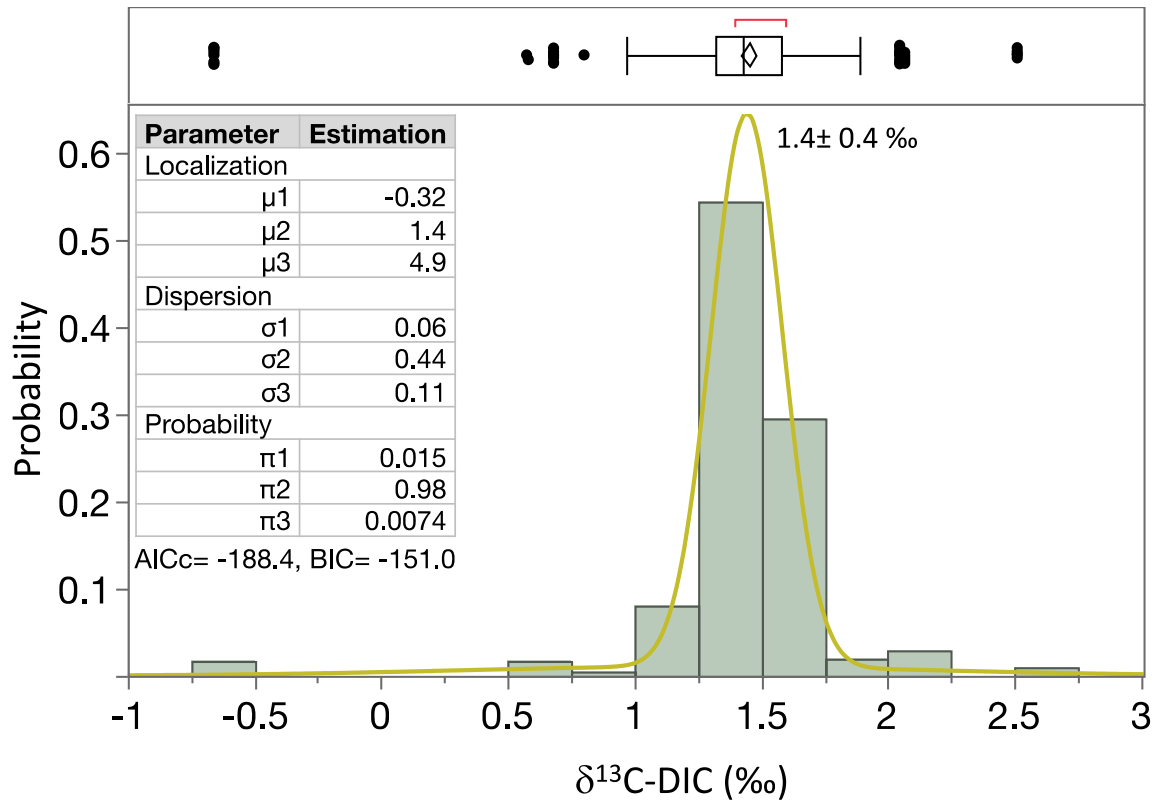
Region	South	Central	Midriff Islands	North	GC	Source
Annual mean of both GC coast	25.6±0.2°C	24.5±0.4°C	23.2±0.4°C		24.5±0.7°C	Escalante et al., (2013)
Annual mean of eastern GC coast	25.8±4.3°C	24.8±4.8 °C	23.7±5.7°C	23.5±5.0°C	24.4±1.1°C	Robles-Tamayo (2018)
Annual T°C range of eastern GC coast	32.4 to 16.7°C	32.4 to 15.7°C	36.3 to 11.9°C	31.98 to 15.5°C		Robles-Tamayo (2018)
Winter (GC)	24.1±0.2°C	19.3±0.3°C	17.7±0.4°C		20.4±1.9°C	Escalante et al., (2013)
Spring (GC)	24.3±0.2°C	21.6±0.3°C	19.5±0.3°C		21.8±1.4°C	Escalante et al., (2013)
Summer (GC)	28.7±0.2°C	29.3±0.3°C	28.6±0.3°C		28.9±0.2°C	Escalante et al., (2013)
Autumm (GC)	29.1±0.2°C	28.2±0.3°C	27.0±0.3°C		28.1±0.6°C	Escalante et al., (2013)
Cold period time. (eastern coast GC)	5 months	6 months	7 months	8 months		Robles-Tamayo (2018)
Cold period T°C mean. (eastern coast GC)	22.1±2.7°C	21.1±3.0°C	20.1±4.0°C	20.7±3.5°C		Robles-Tamayo (2018)
Cold period T°C range. (eastern coast GC)	28.0 to 16.7°C	30.8 to 15.7°C	34.8 to 11.9°C	29.5 to 15.5°C		Robles-Tamayo (2018)
T°C mean during season transitions	25°C	25°C	25°C	25°C	25°C	Robles-Tamayo (2018)
Warm period time (eastern coast GC)	7 months	6 months	5 months	4 months		Robles-Tamayo (2018)
Warm period T°C mean (eastern coast GC)	28.5±3.0°C	28.5±2.9°C	28.7±3.3°C	29.0±2.2°C		Robles-Tamayo (2018)
Warm period T°C range (eastern coast GC)	32.4 to 19.5°C	32.4 to 18.9°C	36.3 to 13.9°C	31.8 to 24°C		Robles-Tamayo (2018)

1 **Figure List**

2 Fig. S1. Histogram representing the distribution of  $\delta^{13}\text{C}$ -DIC values in surface seawater in the Gulf  
3 of California.

4 Fig. S2. Histograms representing the distribution of  $\Delta^{13}\text{C}$ -macroalgal in macroalgae collected in the  
5 Gulf of California for Phyla a) Chlorophyta, Rhodophyta, and Ochrophyta.

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9 Fig. S1. Histogram representing the distribution of  $\delta^{13}\text{C-DIC}$  values in surface seawater in the Gulf  
10 of California.

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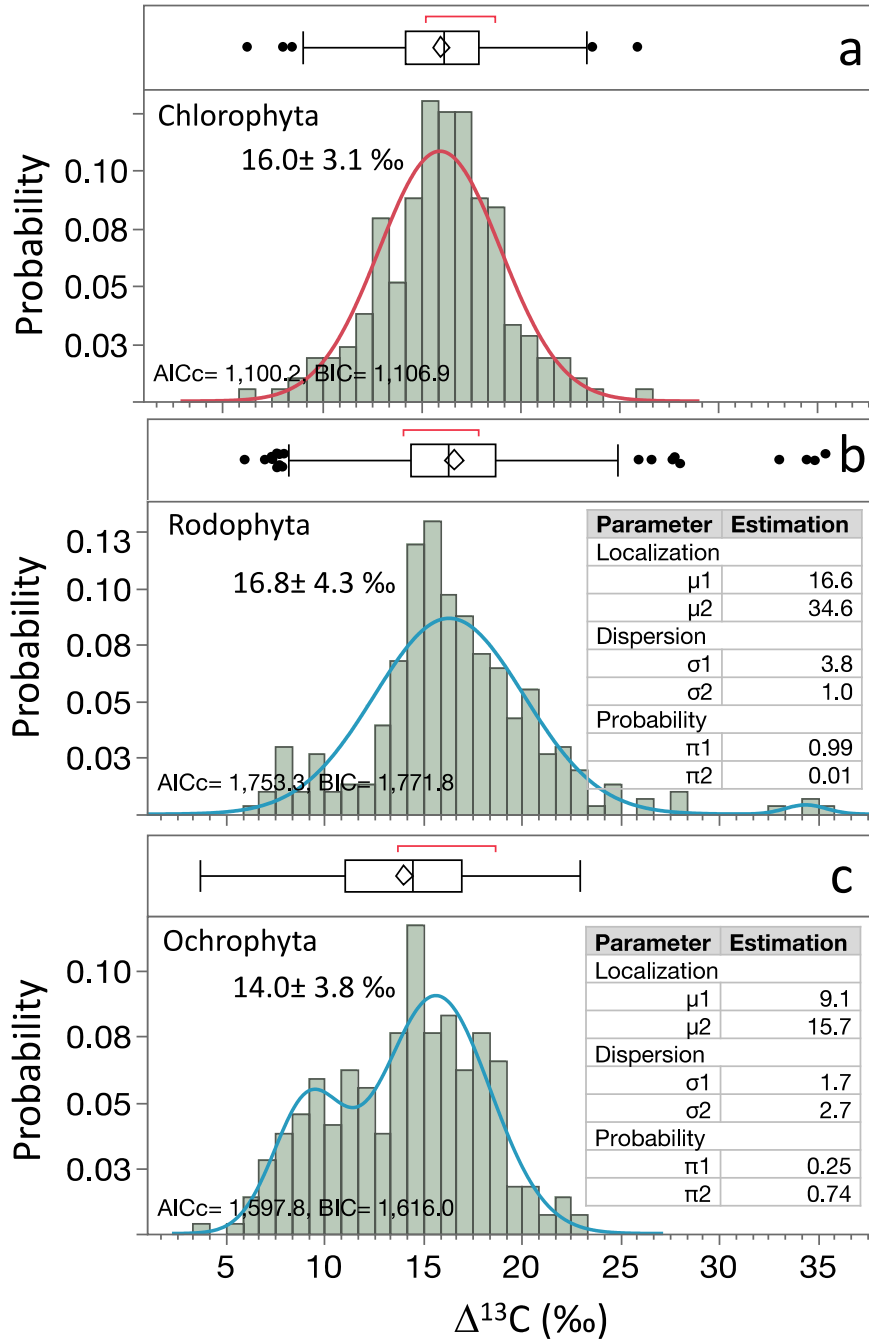


Fig. SI-2. Histograms representing the distribution of  $\Delta^{13}\text{C}$ -macroalgal in macroalgae collected in the Gulf of California for Phyla a) Chlorophyta, Rhodophyta, and Ochrophyta.

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