



# Supplement of

## Modelling temporal variability of in situ soil water and vegetation isotopes reveals ecohydrological couplings in a riparian willow plot

Aaron Smith et al.

*Correspondence to:* Aaron Smith (aaron.smith@igb-berlin.de)

The copyright of individual parts of the supplement might differ from the article licence.



Figure S1: Conceptual diagram of the (a) energy and (b) water balance within EcH<sub>2</sub>O-iso, and the (c) carbon uptake and allocation within vegetation to roots, stem, and leaves. Modified from Smith et al., 2020 and Maneta, 2021.

### Text S1: Detailed Description of In-situ Measurements

#### **Destructive Sampling**

Replicate samples of soil isotopes were collected monthly (June – Oct) using bulk soil water sampling with a soil auger. Samples were taken at increments of 0-10, 10-20, 20-40, 40-70, and 70-100cm. Samples were sealed in a
metalized bag, equilibrated from 48hours prior to analysis. Analysis was conducted using the direct-equilibrium method from Wassenaar et al., (2008) and described in more detail for the IGB laboratory in Kleine et al. (2020). Correction of water samples included nine 10ml standard samples 180 (-10.3, -7.68, 2.91 or 1.53‰) and 2H (-72.81, -56.70, 0.78 or 16.74‰), utilized for all sampling periods.

Destructive vegetation samples were conducted monthly (July – Oct) using samples from three (unique) sun exposed branches on each willow tree. Only branches where bark was unimpaired were sampled. Collected
 branches had the bark and phloem removed to prevent the interference of water sources. Samples were frozen and
 stored in the lab until cryogenic vacuum extraction (as per Koeniger et al. (2011) using 60-90min extraction time
 per sample). Extracted water was measured using CRDS (Picarro L2130-i).

#### **Insitu Sampling**

20 Collected soil and xylem vapour samples were attached to bottles filled with desiccant (Drierite W. A. Hammond DRIERITE Co. LTD, Xenia, OH, USA) to dry incoming air to the laser spectrometer (Picarro L2130-i). Vapour sampling of each soil or xylem location was conducted for 10 min periods to establish stable vapour concentration (plateaus) to the laser spectrometer. Measurement for each location was conducted at 2-hour intervals.

#### 25 Text S2: Energy Balance and EcH2O development

Plant hydraulics are resolved using the Soil Plant Atmosphere Continuum module (SPAC) which tracks leaf water potential and conductivity limitations due to cavitation (Simeone et al., 2019). The SPAC module includes supply-demand functions in the rhizosphere (as a function of soil hydraulic conductivity, root area index, and pore-disconnectedness index), and stem and leaf (function of vegetation conductivity and leaf area index). The supply-demand further regulates transpiration by soil and vapour pressure deficits. Initial testing of the study site using the SPAC module did not reveal sensitivity due to high continued water use of the willows during the growing season and was therefore deactivated prior to calibration.

5

30

35 Table S1: Calibrated soil parameters for below Willow and Grass, presented as the mean, standard deviation (±), and skew. Note: field capacity is not directly calibration, but is a function of Brooks-Corey, air entry pressure, porosity, and residual moisture content. SPAC module was evaluated prior to calibration (insensitive) and was deactivated for calibration (parameters not shown).

	Wil	low	Grass			
		Soil P	ameters			
	Mean ± std	Skew	Mean (std)	Skew		
Albedo	$0.25\pm0.11$	-0.01	$0.22\pm0.06$	-0.39		
Brooks-Corey						
Lambda	$3.76\pm0.68$	0.46	$3.78\pm0.92$	-0.09		
Air entry pressure						
[m]	$0.24 \pm 0.12$	0.56	$0.14 \pm 0.12$	0.96		
Field Capacity	$0.2 \pm 0.01$	-0.09	$0.16 \pm 0.01$	0.02		
Porosity [m <sup>3</sup> /m <sup>3</sup> ]	$0.41\pm0.02$	-0.06	$0.4 \pm 0.02$	-0.39		
Residual Moisture						
Content [m <sup>3</sup> /m <sup>3</sup> ]	$0.01\pm0.01$	0.11	$0.01 \pm 0$	0.03		
Vertical						
Conductivity [m/s]	$1.1E-4 \pm 9E-5$	1.01	$6.8E-4 \pm 2E-4$	0.24		
		Vegetatio	n Parameters			
	Mean ± std	Skew	Mean (std)	Skew		
Root Aspect Ratio						
(horizontal)	$2.73\pm0.47$	0.93	$0.57 \pm 0.28$	-0.2		
Canopy Water						
Storage [m/LAI]	$0.74E3 \pm 0.14E3$	1.23	$1.39E3 \pm 0.3E3$	0.42		
Maximum stomatal						
conducance [m/s]	$0.49E-2 \pm 0.13E-2$	2.35	$1.64E-2 \pm 0.61E-2$	1.39		
Stomatal						
conducance light						
coefficient	$512.44 \pm 121.87$	1.34	$323.3 \pm 119.96$	-0.39		
Stomatal						
conducance VPD						
coefficient [Pa]	$1.35E4 \pm 1.29E4$	0.79	$0.23E4 \pm 0.25E4$	1.26		
Root Distribution						
Parameter (K <sub>root</sub> )	$13.33 \pm 3.98$	0.79	$8.21 \pm 1.05$	0.35		
Maximum LWP for						
stomatal						
conductance	$0.11 \pm 0.12$	1.57	$0.48 \pm 0.3$	0.08		
Wilting Point						
[m <sup>3</sup> /m <sup>3</sup> ]	$0.04 \pm 0.01$	0.59	$0.04 \pm 0.01$	-0.08		
		Vegetation Bi	omass Parameters			
	Mean ± std	Skew	Mean (std)	Skew		
Vegetation Water	$0.82 \pm 0.11$	-0.10	$0.76 \pm 0.10$	0.74		
Use Parameter 1						
Vegetation Water	$4.17 \pm 2.76$	0.25	$5.23 \pm 2.52$	-0.07		
Use Parameter 2						
Water Use	1.6E4 ±1.2E4	1.25	$2.54E3 \pm 7.99E3$	4.59		
Efficiency [gCm <sup>-1</sup> ]						

Foliage allocation coefficient a	$0.06\pm0.06$	1.04	N/A	N/A
$[m^2KgC^{-1}]$				
Foliage allocation	$0.91\pm0.02$	1.17	N/A	N/A
coefficient b				
$[m^2KgC^{-1}]$				
Stem allocation	$0.25\pm0.33$	1.81	N/A	N/A
coefficient a				
Minimum tree	$5.07\pm3.01$	-0.08	N/A	N/A
height to stem ratio				
Wood Density	$6.36E4 \pm 3.39E4$	0.31	N/A	N/A
[gCm <sup>-2</sup> ]				
Specific Leaf Area	$2.74\pm0.76$	-0.30	$3.06 \pm 0.93$	-1.20
$[m^2KgC^{-1}]$				
Leaf Turnover [s <sup>-1</sup> ]	$5.27E-8 \pm 2.65E-8$	0.73	$5.16E-7 \pm 5.00E-8$	-0.23
Maximum Leaf	$5.65E-7 \pm 1.58E-7$	0.62	N/A	N/A
Turnover Due To				
Water Stress [s <sup>-1</sup> ]				
Water Stress	$5.31 \pm 2.87$	-0.22	N/A	N/A
Parameter				
Maximum Leaf	$5.68E-7 \pm 5.73E-7$	1.29	N/A	N/A
Turnover Due To				
Temperature Stress				
[s <sup>-1</sup> ]				
Temperature Stress	$6.61 \pm 3.47$	0.35	N/A	N/A
Parameter				
Dry Leaf Turnover	N/A	N/A	$1.01E-6 \pm 2.94E-7$	1.22
Rate [s <sup>-1</sup> ]				
Dry Leaf Turnover	N/A	N/A	$5.25 \pm 2.72$	1.12
Adjustment				
Parameter				



40 Figure S2: Climate data used within EcH<sub>2</sub>O-iso for the study site between January and November 2020. (a) Hourly precipitation, (b) cumulative precipitation, (c) mean hourly temperature, (d) mean diel temperature (and range) during the growing season (May –September), (e) mean hourly relative humidity, (f) mean diel humidity (and range) during the growing season (May – September), (g) mean hourly wind speed, (h) mean diel wind speed (and range) during the growing season (May –September), (i) mean hourly shortwave radiation, and (j) mean diel radiation (and range) during the growing season (May –September).

Table S2: Average and standard deviation of the goodness-of-fit criteria for modelled time-series of soil, vegetation, and energy balance datasets. \*Dataset was not used in calibration. \*\*Isotope measurement uncertainty shows the average range of sub-daily variability (average measurement uncertainty in parentheses) – for processes not included in EcH2O-iso – and uncertainty of the LMWL regression (lc-excess). "Sub-discretized soil moisture at 10cm.

			Site A		Site B	
			MAE	Measurement	MAE	Measurement
				Uncertainty**		Uncertainty**
	Layer 1 (0-	VMC [m3/m3]	$0.03\pm0^{\scriptscriptstyle \rm H}$	0.025	$0.02 \pm 0$	0.025
	10cm)	δ <sup>2</sup> H [‰]	$8.6 \pm 0.1$	11.9 (0.7)	$5.7 \pm 0.2$	2 (0.7)
		lc-excess		9	$4.5\pm0$	6.9
		[‰]*	$9.3\pm0.7$			
lic	Layer 2 (10-	VMC [m <sup>3</sup> /m <sup>3</sup> ]	$0.02 \pm 0$	0.025	$0.04\pm0.01$	0.025
x	40cm)	δ <sup>2</sup> H [‰]	$4.2 \pm 1.4$	1.3 (0.7)	$2.4 \pm 0.4$	1.74 (0.7)
		lc-excess [‰]	$2.8 \pm 0.3$	5.6	$2.1 \pm 0$	6.1
	Layer 3 (40-	VMC [m <sup>3</sup> /m <sup>3</sup> ]	$0.06\pm0.05$	0.025	$0.01\pm0.01$	0.025
	100cm)	δ <sup>2</sup> H [‰]	$6.8 \pm 1.5$	0.8 (0.7)	$1.3 \pm 0.1$	1.6 (0.7)
		lc-excess [‰]	$1.8 \pm 0$	5.7	$4.6 \pm 0.1$	6
	Flux	Sapflow [m <sup>3</sup> /s]	$0.023 \pm 0.003$	0.016	NA	NA
	Biomass	LAI (Willow	$0.5\pm0.2$	0.9	NA	NA
		1) $[m^2/m^2]$				
Vegetation		LAI (Willow	$0.5\pm0.2$	0.9	NA	NA
		2) $[m^2/m^2]$				
		LAI (Grass)	NA	NA	$0.1\pm0$	NA
		$[m^2/m^2]$	15.07	0.6	NT A	NT A
		Basal Area	$1.5 \pm 0.7$	0.6	NA	NA
		(willow I)				
		Epili Basal Area	$2.2 \pm 1$	0.6	ΝA	NA
		(Willow 2)	$2.2 \pm 1$	0.0		INA
		[µm]				
	Soil	10 cm [°C]	$2 \pm 0.3$	0.4	$1.2 \pm 0.1$	0.3
► 0	Temperature*	40 cm [°C]	$2.4 \pm 0.5$	0.2	$1.9 \pm 0.2$	0.2
rgy		100 cm [°C]	$1.4 \pm 0.5$	0.2	$2.8 \pm 0.2$	0.2
lne	Surface Tempera	ature [°C]	NA	NA	$1.8 \pm 0.2$	0.6
E F	Latent Heat W/	m <sup>2</sup> ]	NA	NA	$20.5\pm0.7$	13.9
	Sensible Heat [V	V/m <sup>2</sup> ]	NA	NA	$4.5 \pm 0.2$	7.1



Figure S3: Measured and simulated soil  $\delta$ 18O at Site A and B at 10cm (a and b), 40cm (c and d), and 100cm (e and f).



55 Figure S4: Simulated transpiration rate (mm/day) for (a) Site A (Willow only) and (b) Site B (Grass). Simulated and measured soil temperature at 10cm at (c) Site A, and (d) Site B, and at 100cm at (e) Site A, and (f) Site B.



Figure S5: Cumulative proportion of water in storage and in root-uptake from each month since the beginning of the simulation (January). White regions indicate water is older than the simulation

60

	Willow										
	М	ay 1 - June	15	June 15-July31			Aug1 - Oct31				
	Layer 1	Layer 2	RWU	Layer 1	Layer 2	RWU	Layer 1	Layer 2	RWU		
$\geq Jan$	0.00	0.27	0.07	0.00	0.25	0.05	0.00	0.21	0.05		
$\geq$ Feb	0.02	0.80	0.21	0.00	0.75	0.13	0.00	0.64	0.08		
$\geq Mar$	0.13	0.97	0.33	0.00	0.91	0.16	0.00	0.77	0.09		
$\geq Apr$	0.42	0.99	0.56	0.01	0.93	0.17	0.00	0.79	0.09		
$\geq May$	0.89	1.00	0.91	0.04	0.95	0.20	0.00	0.81	0.09		

Table S3: Proportion of water in storage or flux for the willows older than or equal to precipitation in each month (e.g. older than or equal to January precipitation). Proportions are displayed for each period of the growing season. N/A indicates not applicable.

≥June	1.00	1.00	1.00	0.46	1.00	0.53	0.00	0.85	0.09
$\geq July$	N/A	N/A	N/A	1.00	1.00	1.00	0.19	0.85	0.24
$\geq Aug$	N/A	N/A	N/A	N/A	N/A	N/A	0.55	0.87	0.57
$\geq$ Sept	N/A	N/A	N/A	N/A	N/A	N/A	0.86	0.96	0.87
$\geq Oct$	N/A	N/A	N/A	N/A	N/A	N/A	1.00	1.00	1.00

Table S4: Proportion of water in storage or flux for the grass older than or equal to precipitation in each month (e.g. older than or equal to January precipitation). Proportions are displayed for each period of the growing season. N/A indicates not applicable.

	Grass									
	М	ay 1 - June	15	June 15-July31			Aug1 - Oct31			
	Layer 1	Layer 2	RWU	Layer 1	Layer 2	RWU	Layer 1	Layer 2	RWU	
$\geq Jan$	0.00	0.29	0.19	0.00	0.21	0.14	0.00	0.12	0.11	
$\geq$ Feb	0.03	0.76	0.42	0.00	0.54	0.29	0.00	0.30	0.20	
$\geq Mar$	0.18	0.92	0.57	0.01	0.66	0.35	0.00	0.37	0.24	
$\geq Apr$	0.48	0.95	0.73	0.03	0.71	0.38	0.00	0.39	0.25	
$\geq May$	0.93	0.99	0.96	0.11	0.80	0.46	0.00	0.45	0.28	
$\geq$ June	1.00	1.00	1.00	0.59	0.94	0.75	0.03	0.56	0.33	
$\geq July$	N/A	N/A	N/A	1.00	1.00	1.00	0.28	0.72	0.50	
$\geq Aug$	N/A	N/A	N/A	N/A	N/A	N/A	0.59	0.91	0.75	
$\geq$ Sept	N/A	N/A	N/A	N/A	N/A	N/A	0.86	0.98	0.92	
$\geq Oct$	N/A	N/A	N/A	N/A	N/A	N/A	1.00	1.00	1.00	



Figure S6: Hourly variability of standardized (0 to maximum transpiration) of (a) measured sap flow, and (b) simulated transpiration.



Figure S7: Normalized daily basal diameter for (a) Willow 1 and (b) Willow 2

Table S5: Average ( $\pm$  standard deviation) transit time in hours of root water in each layer to 1m measurement height in Willow 1 and Willow 2 and the average ( $\pm$  standard deviation) of root length for each soil layer

		Root Length			
	Simulat	( <b>m</b> )			
Soil Layer	Willow 1 (hr)	Willow 2 (hr)	Willow 1 (hr)	Willow 2 (hr)	
Layer 1	$160.0 \pm 21.2$	$140.5 \pm 3.8$	$217.3\pm40.0$	$196.0\pm44.8$	$4.0 \pm 0.5$
Layer 2	205.0 + 97.3	$82.1 \pm 12.1$	$224.0 \pm 127.4$	$160.7 \pm 23.5$	$1.6 \pm 0.5$
Layer 3	$85.2\pm26.8$	$78.8\pm8.4$	$85.6 \pm 1.0$	$132.8\pm10.3$	$0.5 \pm 0.2$

75

85

70

#### References

Kleine, L., Tetzlaff, D., Smith, A., Wang, H., Soulsby, C.: Using water stable isotopes to understand evaporation, moisture stress, and rewetting in catchment forest and grassland soils of the summer drought of 2018, Hydrology and Earth System Sciences, 24(7), 3737-3752, https://doi.org/10.5194/hess-24-3737-2020, 2020.

80 Koeniger, P., Marshall, J. D., Link, T., Mulch, A.: An inexpensive, fast, and reliable method for vacuum extraction of soil and plant water for stable isotope analyses by mass spectrometry, Rapid Communications in Mass Spectrometry, 25(20), 3041-3048, https://doi.org/10.1002/rcm.5198, 2011.

Simeone, C., Maneta, M., Holden, Z. A., Sapes, G., Sala, A., and Dobrowski, S. Z.: Coupled ecohydrology and plant hydraulics modeling predicts ponderosa pine seedling mortality and lower treeline in the US Northern Rocky Mountains, New Phytologist, 221, 1814-1830, doi: 10.1111/nph.15499, 2019.

Wassenaar, L. I., Hendry, M. J., Chostner, V. L., Lis, G. P.: High resolution pore water δ 2H and δ 18 655 O measurements by H2O (liquid)- H2O (vapor) equilibration laser spectroscopy, Environmental science & technology, 42(24), 9262-9267, https://doi.org/10.1021/es802065s, 2008.