



# Temporal dynamics of surface ocean carbonate chemistry in response to natural and simulated upwelling events during the 2017 coastal El Niño near Callao, Peru

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**Abstract.** Oxygen minimum zones (OMZs) are characterized by enhanced carbon dioxide (CO<sub>2</sub>) levels and low pH and are being further acidified by uptake of anthropogenic atmospheric CO<sub>2</sub>. With ongoing intensification and expansion of OMZs due to global warming, carbonate chemistry conditions may become more variable and extreme, particularly in the eastern boundary upwelling systems. In austral summer (February–April) 2017, a large-scale mesocosm experiment was conducted in the coastal upwelling area off Callao (Peru) to investigate the impacts of ongoing ocean deoxygenation on biogeochemical processes, coinciding with a rare coastal El Niño event. Here we report on the temporal dynamics of carbonate chemistry in the mesocosms and surrounding Pacific waters over a continuous period of 50 d with high-temporal-resolution observations (every second day). The mesocosm experiment simulated an upwelling event in the mesocosms by addition of nitrogen (N)-deficient and CO<sub>2</sub>-enriched OMZ water. Surface water in the mesocosms was acidified by the OMZ water addition, with pH<sub>T</sub> lowered by 0.1–0.2 and *p*CO<sub>2</sub> elevated to above 900 μatm. Thereafter, surface *p*CO<sub>2</sub> quickly dropped to near or below the atmospheric level (405.22 μatm in 2017; Dlugokencky and Tans, 2021; NOAA/Global Monitoring Laboratory (GML)) mainly due to enhanced phytoplankton production with rapid CO<sub>2</sub> consumption. Further observations revealed that the dominance of the dinoflagellate *Akashiwo sanguinea* and contamination of bird excrements played important roles in the dynamics of carbonate chemistry in the mesocosms. Compared

to the simulated upwelling, natural upwelling events in the surrounding Pacific waters occurred more frequently with sea-to-air CO<sub>2</sub> fluxes of 4.2–14.0 mmol C m<sup>-2</sup> d<sup>-1</sup>. The positive CO<sub>2</sub> fluxes indicated our site was a local CO<sub>2</sub> source during our study, which may have been impacted by the coastal El Niño. However, our observations of dissolved inorganic carbon (DIC) drawdown in the mesocosms suggest that CO<sub>2</sub> fluxes to the atmosphere can be largely dampened by biological processes. Overall, our study characterized carbonate chemistry in nearshore Pacific waters that are rarely sampled in such a temporal resolution and hence provided unique insights into the CO<sub>2</sub> dynamics during a rare coastal El Niño event.

## 1 Introduction

One of the most extensive oxygen minimum zones (OMZs) in the global ocean can be found off central/northern Peru (4–16° S; Chavez and Messié, 2009). High biological productivity is stimulated by permanent upwelling of cold, nutrient-rich water to the surface, supporting remarkable fish production off Peru (Chavez et al., 2008; Montecino and Lange, 2009; Albert et al., 2010). The high primary production also leads to enhanced remineralization of sinking organic matter in subsurface waters which depletes dissolved oxygen (O<sub>2</sub>) and creates an intense and shallow OMZ (Chavez et al., 2008). The depletion of O<sub>2</sub> in OMZs plays an impor-

tant role in the global nitrogen (N) cycle, accounting for 20%–40% of N loss in the ocean (Lam et al., 2009; Paulmier and Ruiz-Pino, 2009). Denitrification and anammox processes that occur in O<sub>2</sub>-depleted waters remove biologically available N from the ocean and produce an N deficit and hence phosphorus (P) excess with respect to the Redfield ratio (C : N : P = 106 : 16 : 1) in the water column (Redfield, 1963; Deutsch et al., 2001, 2007; Hamersley et al., 2007; Galán et al., 2009; Lam et al., 2009). Upwelling of this N-deficient water has been found to control the surface water nutrient stoichiometry and thus influence phytoplankton growth and community compositions (Franz et al., 2012; Hauss et al., 2012).

Apart from being N-deficient, the OMZ waters are also characterized by enhanced carbon dioxide (CO<sub>2</sub>) concentrations and low pH from respiratory processes and are further acidified by increasing anthropogenic atmospheric CO<sub>2</sub> (Feely et al., 2008; Friederich et al., 2008; Paulmier et al., 2008, 2011). Accordingly, surface water carbonate chemistry is influenced by upwelling of CO<sub>2</sub>-enriched OMZ water (Van Geen et al., 2000; Capone and Hutchins, 2013). The upwelled CO<sub>2</sub>-enriched OMZ water can give rise to surface CO<sub>2</sub> levels > 1000 µatm, pH values as low as 7.6, and undersaturation for the calcium carbonate mineral aragonite (Feely et al., 2008; Hauri et al., 2009). As a result, there is a significant flux of CO<sub>2</sub> from the ocean to the atmosphere off Peru, which is further facilitated by surface ocean warming, making the Peruvian upwelling region a year-round CO<sub>2</sub> source to the atmosphere (Friederich et al., 2008). In contrast, rapid utilization of upwelled CO<sub>2</sub> and nutrients by phytoplankton can occasionally deplete surface CO<sub>2</sub> below atmospheric equilibrium and dampen the CO<sub>2</sub> outgassing (Van Geen et al., 2000; Friederich et al., 2008; Loucaides et al., 2012). The enhanced primary production in turn contributes to increasing export of organic matter, enhanced bacterial respiration, O<sub>2</sub> consumption, and CO<sub>2</sub> production at depth. Such a positive feedback may determine the intensity of the underlying OMZ and promote carbon (C) preservation in marine sediments (Dale et al., 2015).

In response to reduced O<sub>2</sub> solubility and enhanced stratification induced by global warming, OMZs have been intensifying and expanding over the past decades (Stramma et al., 2008, 2010; Fuenzalida et al., 2009). Based on regional observations and model projections, a decline in dissolved O<sub>2</sub> concentrations has been reported for most regions of the global ocean (Matear et al., 2000; Matear and Hirst, 2003; Whitney et al., 2007; Stramma et al., 2008; Keeling et al., 2009; Bopp et al., 2013; Schmidt et al., 2017; Oschlies et al., 2018). The vertical expansion of OMZs represents shoaling of CO<sub>2</sub>-enriched seawater, which has become further enriched by oceanic uptake of anthropogenic CO<sub>2</sub> (Doney et al., 2012; Gilly et al., 2013; Schulz et al., 2019). Since biogeochemical processes in OMZs are directly linked to the C cycle and control surface nutrient stoichiometry, with ongoing ocean warming and acidification, the deoxygenation

may have cascading effects on plankton productivity and composition, C uptake, and food web functioning (Keeling et al., 2009; Gruber, 2011; Doney et al., 2012; Gilly et al., 2013; Levin and Breitburg, 2015). Therefore, it is important to monitor the changes in CO<sub>2</sub> when investigating the effects of deoxygenation on marine ecosystems.

To investigate the potential impacts of upwelling on pelagic biogeochemistry and natural plankton communities in the Peruvian OMZ, a large-scale in situ mesocosm study was carried out in the coastal upwelling area off Peru. An upwelling event was simulated in the mesocosms by addition of OMZ waters collected from two different locations where the OMZ was considered to contain different nutrient concentrations and N : P ratios. The ecological and biogeochemical responses in the mesocosms were monitored and compared with those influenced by natural upwelling events in the ambient coastal water surrounding the mesocosms. As part of this collaborative research project, questions specific to the present paper were as follows: (1) how does surface water carbonate chemistry respond to an upwelling event? And (2) how does upwelled OMZ water with different chemical signatures modulate surface water carbonate chemistry? The current study will mainly focus on the temporal changes in surface water carbonate chemistry within the individual mesocosms, including observations made in the ambient Pacific water and a local estimate of air–sea CO<sub>2</sub> exchange, together with the influence of a rare coastal El Niño event (Garaud, 2018). This provides first insights into how inorganic C cycling links to chemical signatures of OMZ waters in a natural plankton community and its implications for ongoing environmental changes.

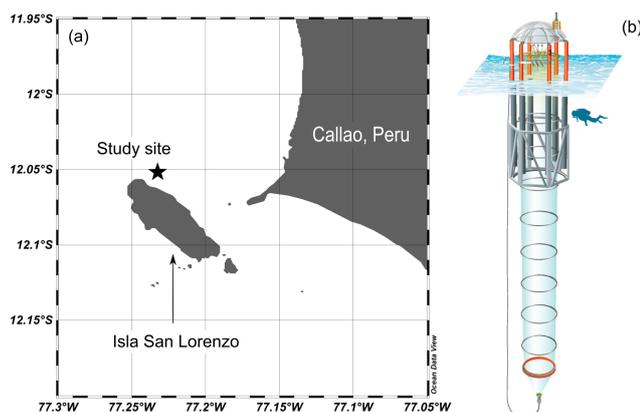
## 2 Material and methods

### 2.1 Study site

The experiment was conducted in the framework of the Collaborative Research Center 754 “Climate-Biogeochemistry Interactions in the Tropical Ocean” (<https://www.sfb754.de/en>, last access: 16 June 2020) and in collaboration with the Instituto del Mar del Perú (IMARPE) in Callao, Peru (Fig. 1a). The coastal area off Callao lies within the Humboldt Current System and is influenced by wind-induced coastal upwelling (Bakun and Weeks, 2008).

### 2.2 Mesocosm setup

Eight Kiel Off-Shore Mesocosms for Future Ocean Simulations (KOSMOS) units (M1–M8), extending 19 m below the sea surface, were deployed by the research vessel Buque Armada Peruana (BAP) *Morales* and moored at 12.06° S, 77.23° W in the coastal upwelling area off Callao, Peru (Fig. 1a) on 23 February 2017 (late austral summer). The technical design of these seagoing mesocosms is described by Riebesell et al. (2013). For a more detailed description



**Figure 1.** The study site of the mesocosm experiment (a) created and modified using Ocean Data View (Reiner Schlitzer, Ocean Data View, <https://odv.awi.de/>, last access: 6 April 2021) and a schematic illustration of a KOSMOS mesocosm unit (b). We acknowledge reprint permission from the American Geophysical Union as parts of this drawing were used for a publication by Bach et al. (2016). The star symbol marks the approximate location of mesocosm deployment.

of the mesocosm deployment and maintenance in this study, please refer to Bach et al. (2020a).

The mesocosm bags were filled with surrounding seawater through the upper and lower openings. Both openings were covered by screens with a mesh size of 3 mm to avoid enclosing larger organisms such as fish. The mesocosm bags were left open below the water surface for 2 d, allowing free exchange with surrounding coastal water. On 25 February, mesocosm bags were closed with the screens removed, tops pulled above the sea surface, and bottoms sealed with 2 m long conical sediment traps (Fig. 1b). The experiment started with the closure of the mesocosms (day 0) and lasted for 50 d. Each mesocosm bag enclosed a seawater volume of  $\sim 54 \text{ m}^3$ . After the bags were closed, daily or every-second-day sampling was performed to monitor the initial conditions of the enclosed water before simulating an upwelling event on day 11 and 12 (see Sect. 2.4 for details).

### 2.3 Simulated upwelling and salt addition

To simulate an upwelling event in the mesocosms, OMZ-influenced waters were collected from the nearby coastal area and added to the mesocosms. Two OMZ water masses were collected at Station 1 ( $12^\circ 01.70' \text{ S}$ ,  $77^\circ 13.41' \text{ W}$ ) at a depth of  $\sim 30 \text{ m}$  and at Station 3 ( $12^\circ 02.41' \text{ S}$ ,  $77^\circ 22.50' \text{ W}$ ) at a depth of  $\sim 70 \text{ m}$ , respectively, using a deep-water collection system as described by Taucher et al. (2017). These two water masses were sampled for chemical and biological variables as done in the mesocosms (see Sect. 2.4). The OMZ water collected from Station 3 had a dissolved inorganic nitrogen (DIN) concentration of  $4.3 \mu\text{mol L}^{-1}$  (denoted as “Low DIN” in this paper) and was added to M2, M3,

M6, and M7. The OMZ water from Station 1 had a DIN of  $0.3 \mu\text{mol L}^{-1}$  (denoted as “Very low DIN” in this paper) and was added to M1, M4, M5, and M8. Before OMZ water addition, approximately  $9 \text{ m}^3$  of seawater was removed from 11–12 m of each mesocosm on 5 March (day 8). During the night of 8 March (day 11),  $\sim 10 \text{ m}^3$  of OMZ water was added to 14–17 m of each mesocosm. On 9 March (day 12),  $\sim 10 \text{ m}^3$  of seawater was removed from 8–9 m followed by an addition of  $\sim 12 \text{ m}^3$  OMZ water to 0–9 m of each mesocosm.

To maintain a low- $\text{O}_2$  bottom layer in the mesocosms and avoid convective mixing induced by heat exchange with the surrounding Pacific, 69 L of a concentrated sodium chloride (NaCl) brine solution was added to the bottom of each mesocosm (10–17 m) on day 13, which increased the bottom salinity by  $\sim 0.7$  units. Following that, turbulent mixing induced by sampling activities continuously interrupted the artificial halocline. Hence, on day 33, 46 L of the NaCl brine solution was added again to the bottom of each mesocosm (12.5–17 m), which increased the bottom salinity by  $\sim 0.5$  units. At the end of the experiment after the last sampling (day 50), 52 kg of NaCl brine was added again to each mesocosm to calculate the enclosed seawater volume from a measured salinity change by  $\sim 0.2$  units (see Czerny et al., 2013a, and Schulz et al., 2013, for details). The average final volume for each mesocosm bag was calculated at  $\sim 54 \text{ m}^3$ . With known sampling volumes and deep-water addition volumes during the experiment, the enclosed volumes of each mesocosm on each sampling day could be calculated. The NaCl solution for the halocline establishments had been prepared in Germany by dissolving 300 kg of food industry grade NaCl (free of anti-caking agents) in 1000 L of deionized water (Milli-Q, Millipore) and purified with ion exchange resin (Lewatit® MonoPlus TP 260, Lanxess, Germany) to minimize potential contaminations with trace metals (Czerny et al., 2013a). The NaCl solution for the volume determination was produced on-site using locally purchased table salt. For a more detailed description of OMZ water and salt additions, please refer to Bach et al. (2020a).

### 2.4 Sampling procedures and CTD operations

Sampling was carried out in the morning (07:00–11:00 local time) daily or every second day throughout the entire experimental period. Depth-integrated samples were taken from the surface (0–10 m for day 3–28) and bottom layer (10–17 m for day 3–28) of the mesocosms and the surrounding coastal water (named “Pacific”) using a 5 L integrating water sampler (IWS, HYDRO-BIOS, Kiel). Due to the deepening of the oxycline as observed from the CTD profiles, the sampling depth for the surface was adjusted to 0–12.5 m while that for the bottom was changed to 12.5–17 m from day 29 until the end of the experiment (day 50).

For gas-sensitive variables such as pH and dissolved inorganic carbon (DIC), 1.5 L of seawater from each integrated

depth in each mesocosm was taken directly from the fully filled 5 L integrating water sampler. Clean polypropylene sampling bottles (rinsed with deionized water in the laboratory; Milli-Q, Millipore) were pre-rinsed with sample water immediately prior to sampling. Bottles were filled from bottom to top using pre-rinsed Tygon tubing with overflow of at least one sampling bottle volume (1.5 L) to minimize the impact of CO<sub>2</sub> air–water gas exchange. Nutrient samples were collected into 250 mL polypropylene bottles using pre-rinsed Tygon tubing (see Bach et al., 2020a, for details). Sample containers were stored in cool boxes for ~3 h and protected from sunlight and heat before being transported to the shore. Once in the lab, sample water was sterile-filtered by gentle pressure using syringe filters (0.2 µm pore size), Tygon tubing, and a peristaltic pump to remove particles that may cause changes to seawater carbonate chemistry (Bockmon and Dickson, 2014). For DIC measurements, the water was filtered from the bottom of the 1.5 L sample bottle into 100 mL glass-stoppered bottles (DURAN) with an overflow of at least 100 mL to minimize contact with air. Once the glass bottle was filled with sufficient overflow, it was immediately sealed without headspace using a round glass stopper. This procedure was repeated to collect a second bottle (100 mL) of filtered water for pH measurements. The left-over seawater was directly filtered into a 500 mL polypropylene bottle for total alkalinity (TA) measurements (non-gas-sensitive). Filtered DIC and pH samples were stored at 4 °C in the dark, and TA samples were at room temperature in the dark until further analysis. Samples were analyzed for DIC and pH on the same day of sampling, while TA was determined overnight (see Sect. 2.5 for analytical procedures).

CTD casts were performed with a multiparameter logging probe (CTD60M, Sea & Sun Technology) in the mesocosms and Pacific on every sampling day. From the CTD casts, profiles of salinity, temperature, pH, dissolved O<sub>2</sub>, chlorophyll *a* (chl *a*), and photosynthetically active radiation were obtained (see Schulz and Riebesell, 2013, and Bach et al., 2020a, for details).

## 2.5 Carbonate chemistry and nutrient measurements

Total alkalinity was determined at room temperature (22–32 °C) by a two-stage open-cell potentiometric titration using a Metrohm 862 Compact Titrosampler, Aquatrode Plus (Pt1000), and 907 Titrand unit in the IMARPE laboratory following Dickson et al. (2003). The acid titrant was prepared by preparing a 0.05 mol kg<sup>-1</sup> hydrochloric acid (HCl) solution with an ionic strength of ca. 0.7 mol kg<sup>-1</sup> (adjusted by NaCl). Approximately 50 g of sample water from each sample was weighed into the titration cell with the exact weight recorded (precision 0.0001 g). After the two-stage titration, the titration data between a pH of ~3.5 and 3 were fitted to a modified non-linear Gran approach described in Dickson et al. (2007) using MATLAB (The MathWorks). The results were calibrated against certified reference material (CRM)

batch 142 (Dickson, 2010) measured on each measurement day. In this paper, measured TA values refer to the measured values that have been calibrated against the CRM.

Seawater pH<sub>T</sub> (total scale) was determined spectrophotometrically by measuring the absorbance ratios after adding the indicator dye *m*-cresol purple (mCP) as described in Carter et al. (2013). Before measurements, samples were acclimated to 25.0 °C in a thermostatted bath. The absorbance of samples with mCP was determined on a Varian Cary 100 double-beam spectrophotometer (Varian), scanning between 780 and 380 nm at a 1 nm resolution. During the spectrophotometric measurement, the temperature of the sample was maintained at 25.0 °C by a water bath connected to the thermostatted 10 cm cuvette. The pH<sub>T</sub> values were calculated from the baseline-corrected absorbance ratios and corrected for in situ salinity (obtained from CTD casts) and pH change caused by dye addition (using the absorbance at the isosbestic point, i.e., 479 nm) as described in Dickson et al. (2007). To minimize potential CO<sub>2</sub> air–water gas exchange, a syringe pump (Tecan Cavro XLP) was used for sample and dye mixing and cuvette injection (see Schulz et al., 2017, for details). For the dye correction, a batch of sterile filtered seawater of known salinity was prepared. The pH<sub>T</sub> was determined once for an addition of 7 µL of dye and once for an addition of 25 µL at five pH levels (raised to 7.95 with NaOH and lowered to 7.74, 7.58, 7.49, and 7.36 with HCl stepwise). The pH change resulting from the dye correction addition was calculated from the change in measured absorbance ratio for each pair of dye additions (see Clayton and Byrne, 1993, and Dickson et al., 2007, for details). The dye-corrected pH<sub>T</sub> values measured at 25.0 °C and atmospheric pressure were then re-calculated for in situ temperature and pressure as determined by CTD casts (averaged over 0–10/12.5 m for surface and 10/12.5–17 m for bottom). For carbonate chemistry speciation calculations (see Sect. 2.6), the dye-corrected pH<sub>T</sub> values were used as one of the input parameters.

Dissolved inorganic carbon was measured by infrared absorption using a LI-COR LI-7000 on an AIRICA system (MARIANDA, Kiel; see Taucher et al., 2017, and Gafar and Schulz, 2018, for details). The results were calibrated against CRM batch 142 (Dickson, 2010). Unfortunately, due to a malfunctioning of the AIRICA system, we obtained measured DIC data only up to 7 March (day 10). Therefore, measured TA and pH<sub>T</sub> were used for calculations of carbonate system parameters at in situ temperature and salinity, but we used DIC measurements from day 3–10 for consistency checks of calculated carbonate chemistry parameters. In this paper, measured DIC values refer to the measured values that have been calibrated against the CRM.

Inorganic nutrients were analyzed colorimetrically (NO<sub>3</sub><sup>-</sup>, NO<sub>2</sub><sup>-</sup>, PO<sub>4</sub><sup>3-</sup>, and Si(OH)<sub>4</sub>) and fluorimetrically (NH<sub>4</sub><sup>+</sup>) using a continuous-flow analyzer (QuAatro AutoAnalyzer with integrated photometers, SEAL Analytical) connected to a fluo-

rescence detector (FP-2020, JASCO). All colorimetric methods were conducted according to Murphy and Riley (1962), Mullin and Riley (1955a, b), and Morris and Riley (1963) and corrected following the refractive index method developed by Coverly et al. (2012). For details of the quality control procedures, see Bach et al. (2020a).

## 2.6 Carbonate chemistry speciation calculations and propagated uncertainties

Calculations of carbonate chemistry parameters (in situ  $\text{pH}_T$ , DIC,  $p\text{CO}_2$ , and calcium carbonate saturation state for calcite and aragonite) were performed with the Excel version of CO2SYS (Version 2.1; Pierrot et al., 2006) using  $K_1$  and  $K_2$  dissociation constants from Mehrbach et al. (1973) which were refitted by Lueker et al. (2000). The dissociation constants for  $\text{KHSO}_4$  from Dickson (1990) and for total boron from Uppström (1974) were applied in the calculations (see Orr et al., 2015, for details). The observed  $\text{pH}_T$  and TA as well as inorganic nutrient concentration (phosphate and silicic acid) were used as input  $\text{CO}_2$  system parameters. In situ salinity and temperature were obtained by CTD casts and averaged over surface (0–10 or 0–12.5 m) and bottom (10–17 or 12.5–17 m) waters for each sampling day. In situ pressure was approximated for surface (5 dbar) and bottom (13.5 or 14.75 dbar) waters. For details of calculation procedures and choices of constants, see Lewis et al. (1998) and Orr et al. (2015).

To evaluate the performance of  $\text{pH}_T$  and TA measurements, quality control procedures were performed. First, standard deviations of  $\text{pH}_T$  measurements were graphed over time. Measured TA values of a control sample (CRM batch 142; Dickson, 2010) were plotted over time and compared to the warning and control limits calculated from their mean and standard deviation (for details please see Dickson et al., 2007) as well as the certified value of the CRM. To compute a range control chart for the evaluation of measurement repeatability, the absolute difference between duplicate measurements of CRMs on each sampling day was calculated and plotted over time and compared to the warning and control limits calculated from their mean and standard deviation (for details see Dickson et al., 2007).

We used the R package “seacarb” with a Gaussian approach and an input variable pair ( $\text{pH}_T$ , TA) to calculate uncertainties for calculated  $\text{CO}_2$  system parameters (Orr et al., 2018; Gattuso et al., 2020). The contributions of input uncertainties in nutrient concentrations and in situ salinity and temperature to the uncertainties in the CO2SYS-based calculations are often small ( $< 0.1\%$ ; Orr et al., 2018), so they were not considered in our propagation. The input uncertainties in  $\text{pH}_T$  and TA were estimated based on our measurements (Table 1). Standard uncertainties include random and systematic errors. For TA, systematic errors were removed by calibrating the measured results using CRMs (see Sect. 2.5). Hence, the random error in TA, estimated by the

averaged standard deviation of all the CRM measurements ( $4.4 \mu\text{mol kg}^{-1}$ ;  $n = 62$ ), was used as the standard uncertainty. For  $\text{pH}_T$ , an uncertainty of 0.01 was used as the standard uncertainty. Due to the unavailability of CRMs that correct for systematic error in pH measurements, the standard deviations of repeated measurements ( $0.0012$ ;  $n = 377$ ) only accounted for the random components of standard uncertainties (Orr et al., 2018). Therefore, we used 0.01 in our uncertainty propagation as an approximation of the total standard uncertainty for  $\text{pH}_T$ , which has been used in previous assessments (Orr et al., 2018).

The air–sea flux of  $\text{CO}_2$  ( $\text{FCO}_2$ ,  $\text{mmol C m}^{-2} \text{d}^{-1}$ ) in the Pacific was determined based on

$$\text{FCO}_2 = k K_0 \Delta p\text{CO}_2, \quad (1)$$

where  $k$  is the gas transfer velocity parameterized as a function of wind speed,  $K_0$  is the solubility of  $\text{CO}_2$  in seawater dependent on in situ salinity and temperature (Weiss, 1974), and  $\Delta p\text{CO}_2$  is the difference between  $p\text{CO}_2$  in the surface water and in the atmosphere (Wanninkhof 2014). Wind data were averaged over 2 sampling days for the sampling location from a satellite-derived gridded dataset (Global Land Data Assimilation System model, near-surface wind speed,  $0.25 \times 0.25^\circ$ , 3 h temporal resolution, 12.375 to 11.875° S, 77.375 to 76.875° W), obtained from NASA Giovanni (Rodell et al., 2004; Beaudoin and Rodell, 2020). In situ salinity and temperature were obtained from the CTD casts (see Sect. 2.4). Calculated  $p\text{CO}_2$  based on ( $\text{pH}_T$ , TA) and an estimated atmospheric  $p\text{CO}_2$  of  $405.22 \mu\text{atm}$  (referenced to year 2017; Dlugokencky and Tans, 2021; NOAA/GML) were used in the air–sea flux estimation.

## 3 Results

### 3.1 Responses of surface layer nutrient concentrations

The OMZ-influenced water masses were collected from two locations and added to the mesocosms to simulate an upwelling event (see Sect. 2.3). The two water masses were named Low DIN and Very low DIN, respectively, based on their DIN concentrations (Table 2). Both water masses shared similar silicic acid (Si) and phosphate ( $\text{PO}_4^{3-}$ ) concentrations but differed in DIN concentration. The Low DIN water had a DIN concentration of  $4.3 \mu\text{mol L}^{-1}$ , 14 times as high as that of the Very low DIN water ( $0.3 \mu\text{mol L}^{-1}$ , Table 2).

On day 10 before OMZ water addition, the average surface DIN concentrations of the two treatment groups were similar ( $3.4 \mu\text{mol L}^{-1}$ ) but lower than that in the Pacific ( $9.8 \mu\text{mol L}^{-1}$ , Table 3). Surface layer DIN concentrations in the mesocosms ranged between 2.0 and  $6.0 \mu\text{mol L}^{-1}$  before OMZ water addition (Fig. 2a). The addition of OMZ water elevated surface DIN in the Low DIN mesocosms to  $3.6$ – $6.4 \mu\text{mol L}^{-1}$  but lowered that in the Very low DIN to  $0.9$ – $2.0 \mu\text{mol L}^{-1}$ . The average surface DIN concentration

**Table 1.** Standard uncertainties in  $\text{pH}_T$  and TA estimated based on our measurements are denoted by  $\mu(\text{pH}_T)$  and  $\mu(\text{TA})$ . Based on  $\mu(\text{pH}_T)$  and  $\mu(\text{TA})$ , propagated uncertainties were estimated for each data point in R and averaged for each reported variable ( $\mu$ ), with standard deviation ( $\sigma$ ), minimum (min), and maximum (max) values presented. The relative percentages (%) of propagated standard uncertainties were calculated by dividing the propagated uncertainty by the corresponding data point and averaged for each reported variable ( $\mu$ ), with  $\sigma$ , min, and max values presented.

$\mu(\text{pH}_T)$	$\mu(\text{TA})$ $\mu\text{mol kg}^{-1}$	$\Delta p\text{CO}_2$		$\Delta\text{DIC}$		$\Delta\Omega_{\text{Ar}}$		$\Delta\Omega_{\text{Ca}}$		
		$\mu\text{atm}$	%	$\mu\text{mol kg}^{-1}$	%	%	%	%	%	
0.01	4.4	$\mu$	35.94	3.8	6.63	0.3	0.08	5.1	0.13	5.1
		$\sigma$	12.60	0.3	0.80	0.0	0.03	0.3	0.05	0.3
		min	15.07	3.2	5.88	0.3	0.04	4.4	0.07	4.4
		max	62.84	4.8	8.72	0.4	0.16	5.8	0.24	5.8

**Table 2.** Inorganic nutrient concentrations of the two collected deep-water masses. Please note that DIN is the sum of nitrate, nitrite, and ammonium. P is phosphate. Si is silicic acid.

Water mass	Si ( $\mu\text{mol L}^{-1}$ )	DIN ( $\mu\text{mol L}^{-1}$ )	$\text{PO}_4^{3-}$ ( $\mu\text{mol L}^{-1}$ )	N : P ratio (mol : mol)
Low DIN	19.6	4.3	2.5	1.7
Very low DIN	17.4	0.3	2.6	0.1

in the Very low DIN decreased to  $1.6 \mu\text{mol L}^{-1}$  while the Low DIN slightly increased to  $4.7 \mu\text{mol L}^{-1}$  (Table 3), followed by a sharp depletion on day 16 except for M3. M3 received the highest input of DIN ( $6.4 \mu\text{mol L}^{-1}$ ) and was not depleted until day 24. Despite several small peaks in M3, M4, M5, and M6 ( $\leq 1.6 \mu\text{mol L}^{-1}$ ), surface DIN concentrations in the mesocosms were at around the limits of detection (LODs –  $\text{NH}_4^+$ ,  $0.063 \mu\text{mol L}^{-1}$ ;  $\text{NO}_2^-$ ,  $0.054 \mu\text{mol L}^{-1}$ ;  $\text{NO}_3^-$ ,  $0.123 \mu\text{mol L}^{-1}$ ) most of the time after depletion. A slight rise could be observed from day 44 towards the last sampling day (day 48). In the Pacific, surface layer DIN concentration was mostly greater than  $5 \mu\text{mol L}^{-1}$  (except on day 16 and 18) and became considerably higher during the second half of the experiment ( $> 10 \mu\text{mol L}^{-1}$  for day 26–44, Fig. 2a).

Surface layer  $\text{PO}_4^{3-}$  concentrations in the mesocosms initially ranged between  $1.1$  and  $1.5 \mu\text{mol L}^{-1}$  and were elevated by OMZ water addition to around  $1.9 \mu\text{mol L}^{-1}$  (Fig. 2b). Thereafter,  $\text{PO}_4^{3-}$  exhibited a slow but steady decline until the end of the study with a slightly higher decrease in Low DIN mesocosms (blue symbols, Fig. 2b). Throughout the study,  $\text{PO}_4^{3-}$  in the mesocosms was never lower than  $1.1 \mu\text{mol L}^{-1}$ . Surface layer  $\text{PO}_4^{3-}$  in the Pacific was generally higher, fluctuating between  $1.4$  and  $2.9 \mu\text{mol L}^{-1}$ . In the mesocosms, enhanced chl *a* concentrations were observed at depths shallower than 5 m and below 15 m before OMZ water addition (Fig. 2c). Following OMZ water addition, a chl *a* maximum occurred at  $\sim 10$  m and persisted until day 40, except for M3 and M4 with a 1-week-delayed increase in the former and a lack of bloom in the latter (Fig. 2c). After day 40, chl *a* concentrations in all mesocosms (except for

M4) increased to  $12$ – $38 \mu\text{g L}^{-1}$  with a bloom occurring in 0–10 m (Fig. 2c). Throughout the study, a chl *a* maximum was continuously observed above 10 m in the Pacific (Fig. 2c).

### 3.2 Temporal dynamics of carbonate chemistry

Before OMZ water addition, surface layer  $\text{pH}_T$  in the mesocosms ranged between  $7.80$ – $7.94$  with a slight decline by  $\sim 0.1$  over time (Fig. 3a). The initial surface layer TA ranged between  $2310$  and  $2330 \mu\text{mol kg}^{-1}$  (Fig. 3b, day 3–12). Surface layer  $p\text{CO}_2$  and DIC ranged from  $541$  to  $749 \mu\text{atm}$  and  $2119$  to  $2180 \mu\text{mol kg}^{-1}$ , respectively (Fig. 3c, d).

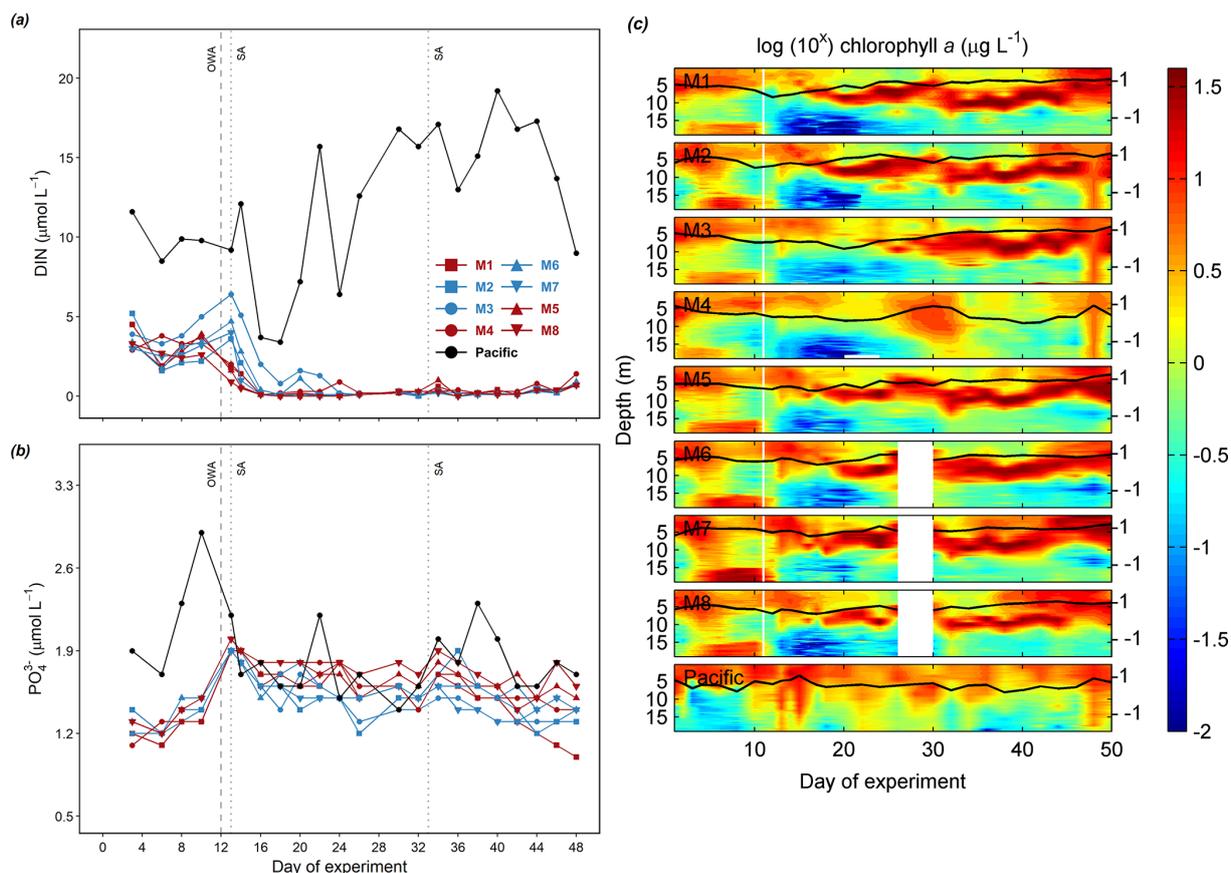
The two collected OMZ water masses shared similar carbonate chemistry properties despite the differences in DIN concentrations. In both water masses,  $\text{pH}_T$  was  $\sim 7.48$ , DIC was  $\sim 2305$ – $2310 \mu\text{mol kg}^{-1}$ , TA was  $\sim 2337 \mu\text{mol kg}^{-1}$ , and  $p\text{CO}_2$  was between  $1700$  and  $1780 \mu\text{atm}$  (Table 4).

Surface DIC and  $p\text{CO}_2$  were elevated from  $\sim 2150 \mu\text{mol kg}^{-1}$  and  $\sim 600 \mu\text{atm}$  to  $\sim 2200 \mu\text{mol kg}^{-1}$  and  $\sim 900 \mu\text{atm}$  (except M7), respectively, by OMZ water addition without distinct differences between the two treatments (Mann–Whitney *U* test,  $p > 0.05$ ; Fig. 3c). Following OMZ water addition, surface  $p\text{CO}_2$  in the mesocosms decreased quickly and reached minima at  $340$ – $500 \mu\text{atm}$  (except M3 and M4) on day 24 and 26. These minima corresponded with DIC minima at  $2040$ – $2110 \mu\text{mol kg}^{-1}$  and  $\text{pH}_T$  maxima at  $7.9$ – $8.1$  (except M3 and M4; Fig. 3c, d). After reaching the minima, surface layer  $p\text{CO}_2$  exhibited a steady increase to  $410$ – $680 \mu\text{atm}$  from day 24 to day 38 and later declined in M3, M5, and M7 while the rest remained relatively stable until day 42 (Fig. 3c). Interestingly, and unlike the other mesocosms, after OMZ water addition,  $p\text{CO}_2$  in M3 steadily declined from  $928$  to  $342 \mu\text{atm}$  until the end of the experiment while that in M4 remained constantly higher than the other mesocosms ( $> 700 \mu\text{atm}$ ), with a slightly decreasing trend to  $645 \mu\text{atm}$  towards the end of the study (Fig. 3c).

In the Pacific, much lower surface  $\text{pH}_T$  and higher surface  $p\text{CO}_2$  and DIC were observed compared to the mesocosms, with an average of  $7.7$  ( $7.6$ – $7.8$ ),  $1078 \mu\text{atm}$  ( $775$ – $1358 \mu\text{atm}$ ), and  $2221 \mu\text{mol kg}^{-1}$  ( $2173$ – $2269 \mu\text{mol kg}^{-1}$ ;

**Table 3.** DIN concentration ( $\mu\text{mol L}^{-1}$ ) in the surface layer of each mesocosm (M1–M8) and the average DIN concentration ( $\mu\text{mol L}^{-1}$ ) for each treatment (Low DIN and Very low DIN;  $n = 4$ ) before (t10) and after deep-water addition (t13). The DIN concentration in the surface Pacific water is also shown.

	M1	M2	M3	M4	M5	M6	M7	M8	Low DIN	Very low DIN	Pacific
t10	3.7	2.2	5.0	3.3	3.9	3.4	3.2	2.6	$3.4 \pm 1.2$	$3.4 \pm 0.5$	9.8
t13	1.8	3.6	6.4	2.0	1.6	4.7	4.0	0.9	$4.7 \pm 1.3$	$1.6 \pm 0.5$	9.2



**Figure 2.** Temporal dynamics of depth-integrated surface DIN concentration (a) and  $\text{PO}_4^{3-}$  concentration (b) and vertical distribution of chl *a* concentration determined by CTD casts (c). The solid black lines on top of the colored contours represent the average values over the entire water column, with the corresponding additional y axes on the right. The vertical white lines represent the day when OMZ water was added to the mesocosms. Color codes and symbols denote the respective mesocosms. Abbreviations: OWA, OMZ water addition; SA, salt addition. The dataset is available at <https://doi.pangaea.de/10.1594/PANGAEA.923395> (Bach et al., 2020b).

minimum-to-maximum range in parentheses; Fig. 3c, d), respectively. TA in the Pacific was initially similar to that in the mesocosms, fluctuating between  $2310$  and  $2330 \mu\text{mol kg}^{-1}$ , and later decreased to  $\sim 2310 \mu\text{mol kg}^{-1}$  for the rest of the study.

Surface waters in the mesocosms and the Pacific were always saturated with respect to calcite and aragonite throughout the entire experimental period, with lower values observed in the Pacific (Fig. 4a, c). Bottom waters in the mesocosms and Pacific were always saturated with respect to calcite during the experiment (Fig. 4b), while bottom waters in

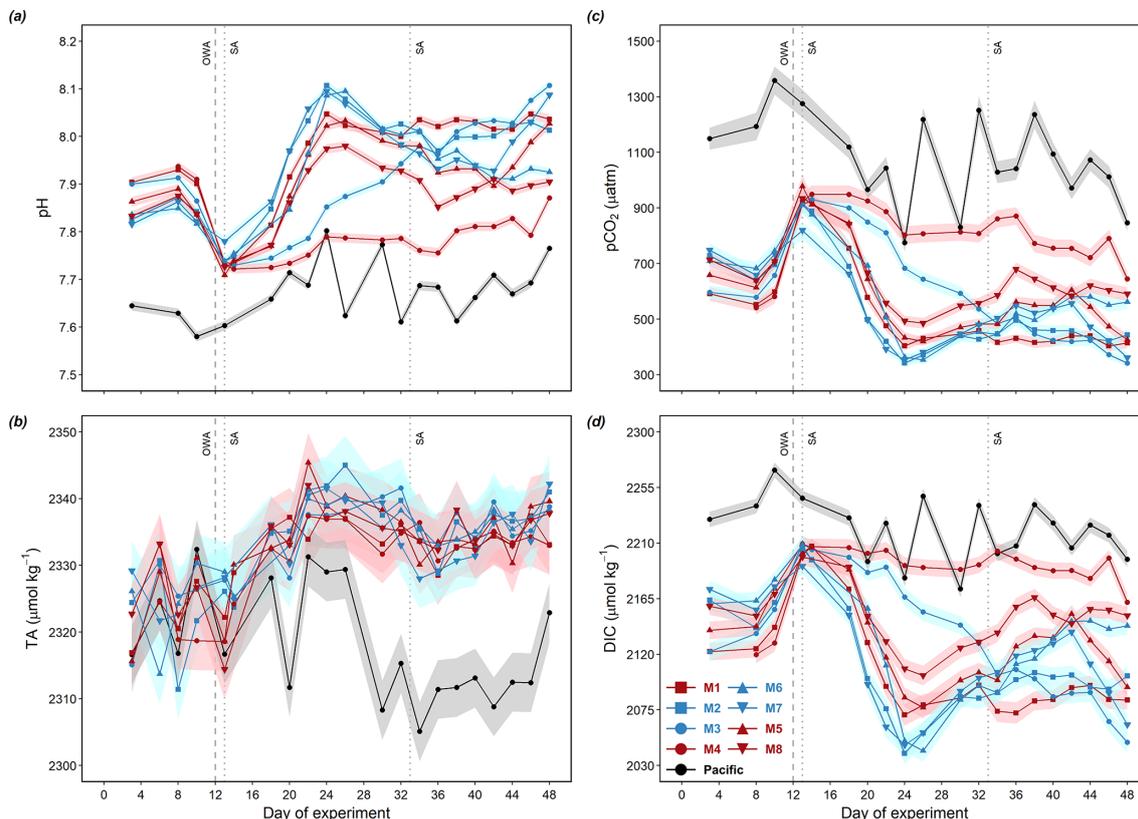
the Pacific were under-saturated with respect to aragonite before day 13 ( $0.88$ – $0.99$ ) and had  $\Omega_{\text{Ar}}$  values slightly above  $1.0$  for the rest of the study period (Fig. 4d).

### 3.3 Air–sea $\text{CO}_2$ fluxes in the Pacific

Positive  $\text{FCO}_2$  values indicate  $\text{CO}_2$  outgassing from the surface waters to the atmosphere, while negative values indicate a  $\text{CO}_2$  flux from the atmosphere to the ocean. The air–sea  $\text{CO}_2$  flux in the Pacific was constantly positive throughout our study, fluctuating from  $4.2$  to  $14.0 \text{ mmol C m}^{-2} \text{ d}^{-1}$  over time (Fig. 5a). The minima of  $\text{FCO}_2$  occurred on day 26

**Table 4.** The in situ  $\text{pH}_T$ , TA, DIC,  $p\text{CO}_2$ ,  $\Omega_{\text{Ar}}$ , and  $\Omega_{\text{Ca}}$  of the two collected OMZ water masses.

Water mass	$\text{pH}_T$	TA ( $\mu\text{mol kg}^{-1}$ )	DIC ( $\mu\text{mol kg}^{-1}$ )	$p\text{CO}_2$ ( $\mu\text{atm}$ )	$\Omega_{\text{Ar}}$	$\Omega_{\text{Ca}}$
Low DIN	7.49	2336.5	2305.4	1707.5	0.90	1.38
Very low DIN	7.47	2338.2	2312.1	1775.3	0.87	1.34

**Figure 3.** Temporal dynamics of measured depth-integrated surface  $\text{pH}_T$  (a) and TA (b) and calculated  $p\text{CO}_2$  (c) and DIC (d). The error ribbons present measurement and propagated standard uncertainties in the calculations, respectively. Color codes and symbols denote the respective mesocosms. Abbreviations: OWA, OMZ water addition; SA, salt addition.

and 30, while the maximum occurred on day 32 when near-surface wind was the highest ( $2.89 \text{ m s}^{-1}$ , Fig. 5b), corresponding to the minima and maxima of surface  $p\text{CO}_2$ . Co-occurring with a decrease in surface temperature to below  $19^\circ\text{C}$  after day 36 (Fig. 5c),  $\text{FCO}_2$  slightly declined from  $\sim 10$  to  $\sim 6 \text{ mmol C m}^{-2} \text{ d}^{-1}$  (Fig. 5a).  $\text{FCO}_2$  was positively correlated with near-surface wind speed ( $r^2 = 0.4$ ). No correlation was found between  $\text{FCO}_2$  and temperature ( $r^2 = 0$ ).

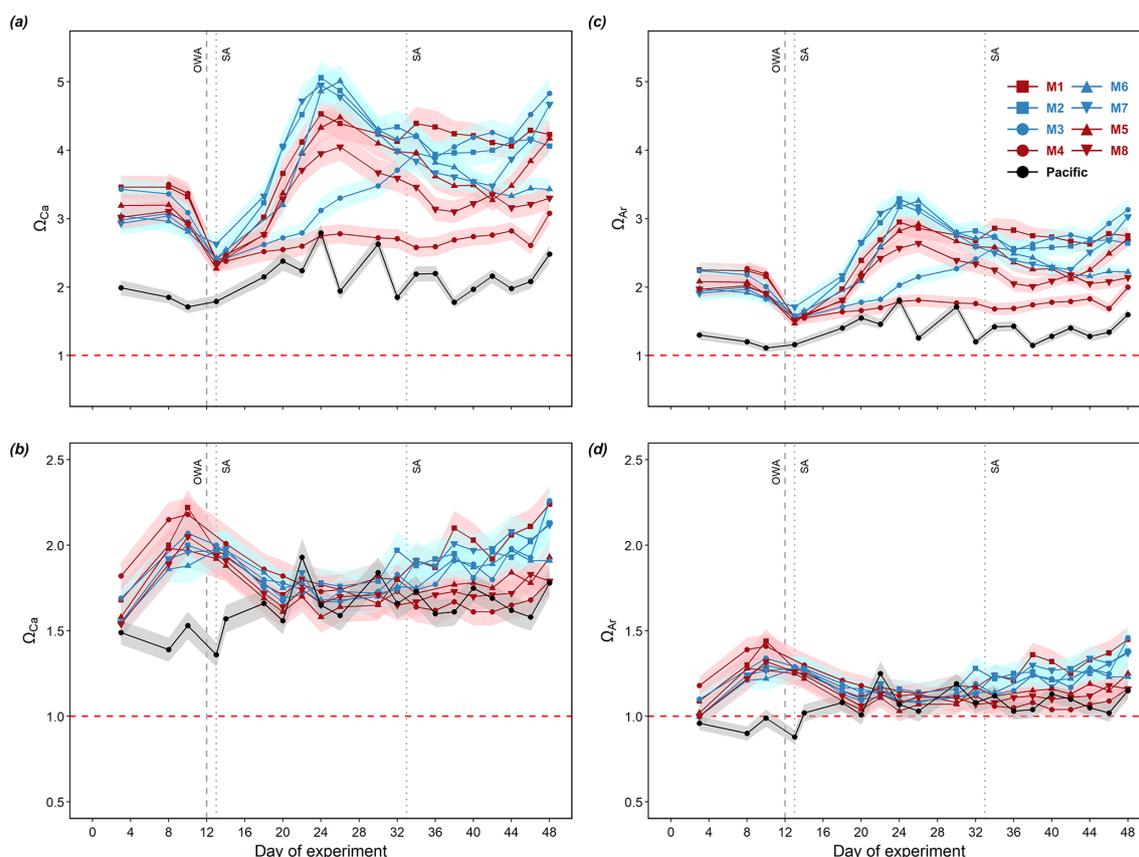
## 4 Discussion

### 4.1 Quality control and propagated uncertainties

To compare the sensitivity of different calculated variables to uncertainties in the input variables, the propagated uncertainties were averaged for each calculated variable, reported

in numerical values and percentages relative to the calculated values of each variable (Table 1). Among the 4 reported variables,  $\Omega_{\text{Ca}}$  and  $\Omega_{\text{Ar}}$  were the most sensitive to uncertainties in  $\text{pH}_T$  and TA with an average uncertainty of 5.1%. This adds ambiguity to whether the bottom water (10–17 m for day 3–28, 12.5–17 m for day 29–50) in the Pacific was undersaturated with respect to aragonite when  $\Omega_{\text{Ar}}$  was oscillating near 1 (Fig. 4d). The propagated uncertainty in  $p\text{CO}_2$  was slightly lower (3.8%), while DIC was the least sensitive (0.3%).

We examined the internal consistency between DIC measurements and calculations. DIC was measured from day 3 until the malfunction of the instrument on day 10. In total, 53 sets of measured DIC and calculated DIC (from measured  $\text{pH}_T$  and TA) values were obtained from day 3 to day 10 and compared to test their consistency (Fig. 6a). The calculated DIC values were generally in agreement with the



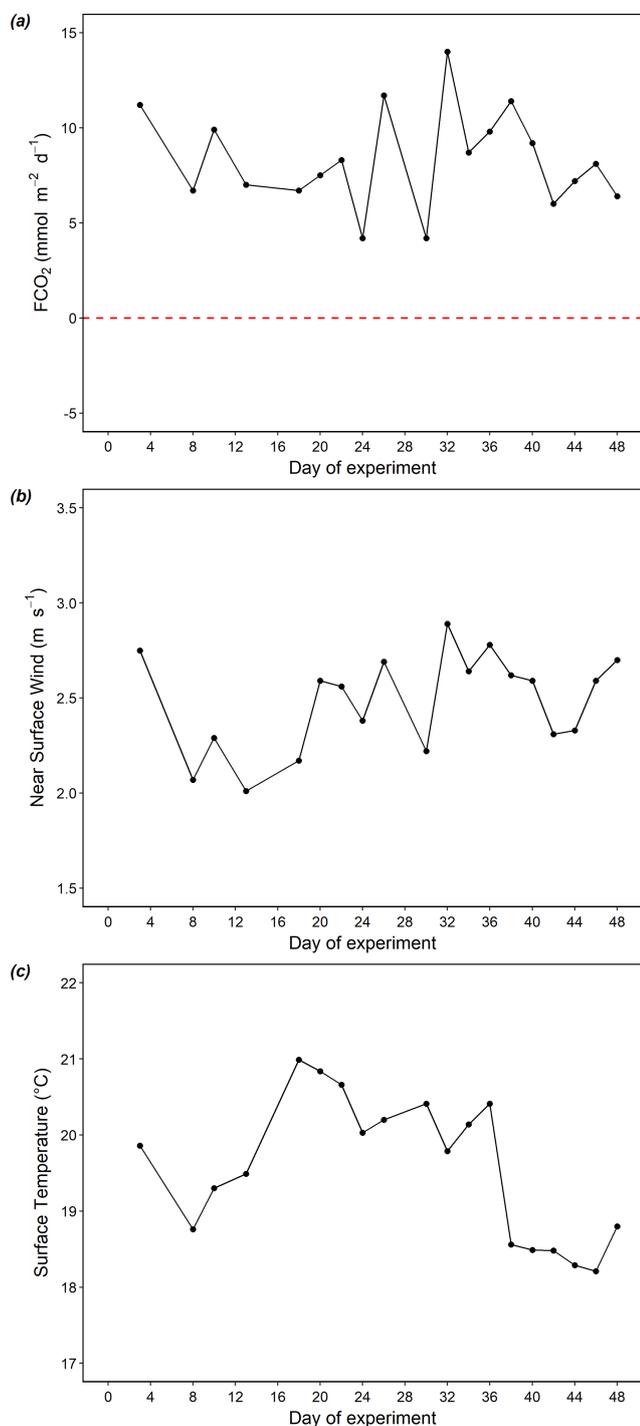
**Figure 4.** Temporal dynamics of depth-integrated surface calcite saturation state (a), bottom calcite saturation state (b), surface aragonite saturation state (c), and bottom aragonite saturation state (d) in the mesocosms and the surrounding Pacific. The error ribbons present the propagated standard uncertainties in the calculations. When  $\Omega > 1$  (above dashed red line), seawater is supersaturated for calcium carbonate. When  $\Omega < 1$  (below dashed red line), seawater is under-saturated for calcium carbonate. Color codes and symbols denote the respective mesocosms. Abbreviations: OWA, OMZ water addition; SA, salt addition.

measured values ( $r^2 = 0.985$ ;  $p < 0.005$ ), showing that the calculations made an overall good prediction for the measured DIC values. The average of the residuals (calculated DIC – measured DIC) was  $-8.27 \pm 6.9 \mu\text{mol kg}^{-1}$ , indicating an underestimation of calculated DIC. This result is consistent with a previous observation of underestimated calculated DIC ( $\text{pH}_T$ , TA) compared with measured DIC when applying the same set of constants ( $-6.6 \pm 7.9 \mu\text{mol kg}^{-1}$ ; Raimondi et al., 2019). The reasons for such underestimation have not been addressed in previous studies and remain unclear. No significant relationships with input variables  $\text{pH}_T$  and TA ( $r^2 = 0.12$  for both) and temperature ( $r^2 = 0.30$ ) were found in the DIC residuals (salinity remained the same from day 3 to day 10). The lack of correlation with  $\text{pH}_T$  and TA indicated that the underestimation in calculated DIC was not a result from changes in  $\text{pH}_T$  and TA. Although dissociation constants are known to be salinity- and temperature-dependent, the lack of correlation between DIC residuals and temperature may be attributed to the relatively narrow ranges of temperature in the mesocosms ( $17.9\text{--}20.9^\circ\text{C}$  from day 3–

10). The offsets were typically larger at lower temperatures (e.g., samples from the Arctic; Chen et al., 2015).

To assess the quality of carbonate chemistry measurements in this study, the stability and performance of measurements were evaluated. The standard deviation of triplicate  $\text{pH}_T$  measurements varied by up to 0.003 with an average of 0.0012 throughout the whole experiment (Fig. 6b). The average standard deviation was in agreement with reported analytical precisions of pH (0.003, Orr et al., 2018; 0.002, Raimondi et al., 2019; Ma et al., 2019).

For TA, triplicate measurements of CRM distributed before and after the sample measurements were carried out on each measuring day to monitor the stability of the measurement process and the performance of the system. Based on the offsets, a correction factor was applied to the measured values of samples on each sampling day to calibrate for instrument drift. As shown in Fig. 6c, 90.5 % of the measured TA values of CRM fell between warning limits (upper warning limit, UWL, and lower warning limit, LWL) with one data point falling outside the control limits (upper control limit, UCL, and lower control limit, LCL),



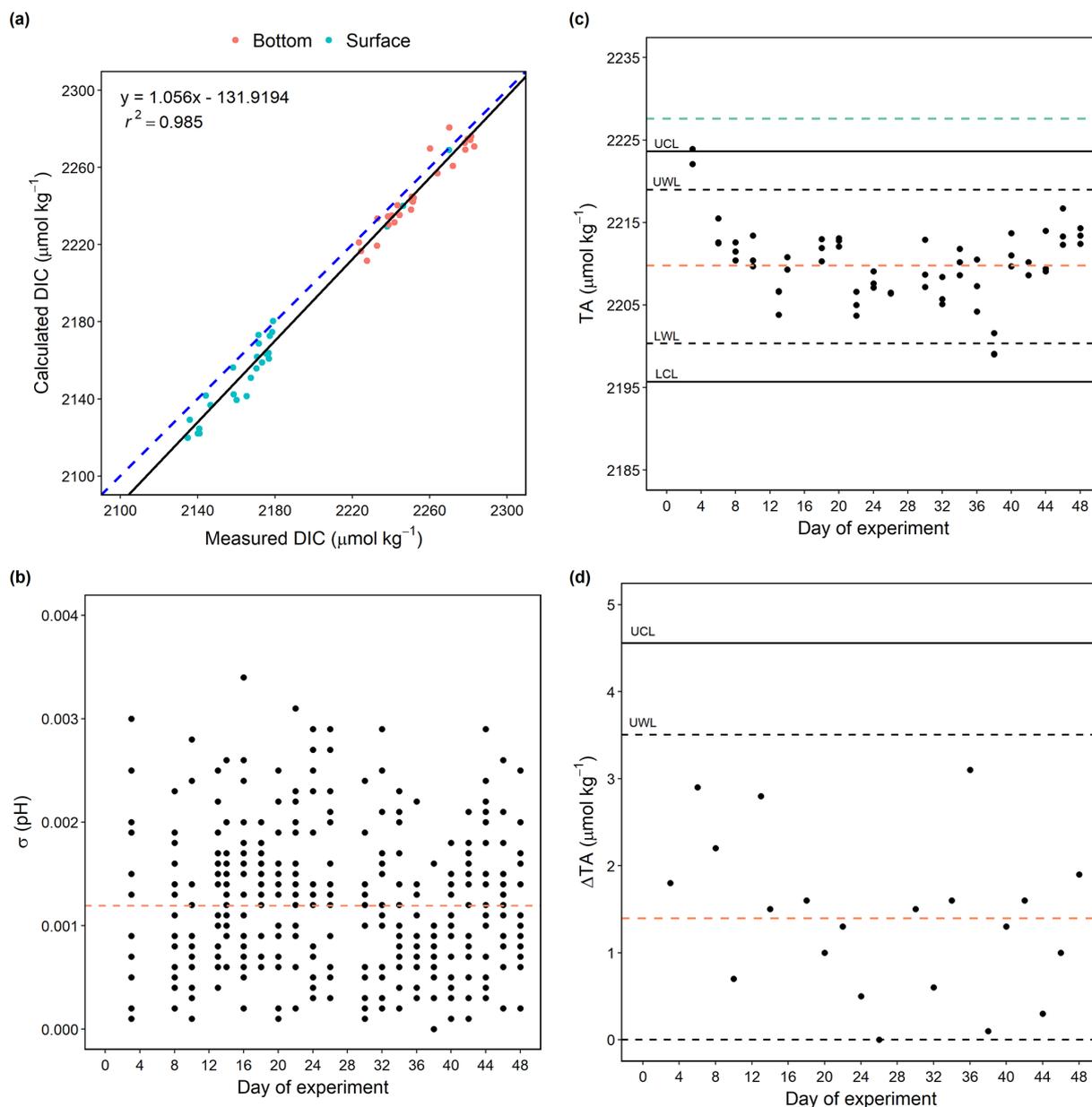
**Figure 5.** Temporal dynamics of surface air–sea CO<sub>2</sub> flux (a), near-surface wind speed (b), and surface temperature (c) in the Pacific. FCO<sub>2</sub> > 0 (above dashed red line) indicates CO<sub>2</sub> outgassing from the sea surface to the atmosphere. FCO<sub>2</sub> < 0 (below dashed red line) indicates a CO<sub>2</sub> flux from the atmosphere to the sea.

overall suggesting a relatively stable measurement system. The average measured TA was 2209.9  $\mu\text{mol kg}^{-1}$ , which was 17.69  $\mu\text{mol kg}^{-1}$  lower than the certified concentration of the CRM (2227.59  $\mu\text{mol kg}^{-1}$ ), indicating a relatively poor accuracy (compared to the suggested bias of less than 2  $\mu\text{mol kg}^{-1}$ ; Dickson et al., 2003, 2007). The poor accuracy could be attributed to the fact that the concentration of the acid titrant was not checked after being prepared, as suggested in the protocol (Dickson et al., 2003). A range control chart was computed based on duplicate measurements of CRM made prior to the sample measurements on each sampling day to evaluate the consistency of the offset between measured and certified TA values over the course of the study (Fig. 6d; Dickson et al., 2007). The absolute difference (range) between the repeated CRM measurements was on average 1.4  $\mu\text{mol kg}^{-1}$ . All the range values fell below the UWL (3.50  $\mu\text{mol kg}^{-1}$ , Fig. 6d), suggesting a relatively good precision of the measurement system.

#### 4.2 CO<sub>2</sub> responses to the simulated upwelling event

At the beginning of the experiment, surface  $p\text{CO}_2$  levels in the mesocosms were > 500  $\mu\text{atm}$  (Fig. 3c). This suggests that we initially enclosed an upwelled water mass that was enriched with respiratory CO<sub>2</sub>. The addition of OMZ water with high concentrations of CO<sub>2</sub> to the mesocosms reduced the surface pH<sub>T</sub> by 0.1–0.2 and increased the surface  $p\text{CO}_2$  to > 900  $\mu\text{atm}$  (except for M7, which was 819.4  $\mu\text{atm}$  on day 13). The simulated upwelling substantially reduced the variability in CO<sub>2</sub> between mesocosms because OMZ water addition replaced  $\sim 20 \text{ m}^3$  of seawater in each mesocosm (out of  $\sim 54 \text{ m}^3$ ). The enhanced  $p\text{CO}_2$  level is comparable with our observations in the ambient Pacific water (> 775  $\mu\text{atm}$ , Fig. 3c). These values also agree with reported observations for our study area in 2013 (> 1200  $\mu\text{atm}$  in the upper 100 m and > 800  $\mu\text{atm}$  at the surface; Bates, 2018).

In the days after OMZ water addition, surface  $p\text{CO}_2$  in the mesocosms dropped near or below the atmospheric level (405.22  $\mu\text{atm}$ ; Dlugokencky and Tans, 2021; NOAA/GML) with a decline in DIC by  $\sim 100 \mu\text{mol kg}^{-1}$  (except M3 and M4; Fig. 3c, d). The declining  $p\text{CO}_2$  could be partially attributed by CO<sub>2</sub> outgassing due to a high CO<sub>2</sub> gradient from the sea surface to the air. Due to a rare coastal El Niño event (Garreaud, 2018), the CO<sub>2</sub> loss process may have been enhanced by a rapid surface warming (19.8–21.0 °C from day 14 to 36, Fig. 5) which reduced surface CO<sub>2</sub> solubility (Zeebe and Wolf-Gladrow, 2001). However, air–sea gas exchange could not explain surface CO<sub>2</sub> under-saturation in relation to the atmosphere, as observed in response to OMZ water addition in some mesocosms (Van Geen et al., 2000; Friederich et al., 2008; Fig. 3c). Biological production typically has impacts on CO<sub>2</sub> drawdown that are 1 to 4 times greater than air–sea gas exchange in the equatorial Pacific where surface waters are exposed to local wind stress (Feely et al., 2002). This interpretation is supported by the continu-



**Figure 6.** Comparison of calculated values of DIC ( $\text{pH}_T$ , TA) and measured values (a). The black line is the regression line, with the corresponding equation and  $r^2$  shown in the top-left corner. The dashed blue line shows the regression line forced through the origin. Standard deviations are of all the triplicate  $\text{pH}_T$  measurements on each sampling day over the study period. The dashed orange line shows the average ( $n = 377$ ) of the standard deviations (b). TA values of CRM measurements on each sampling day over the study period. The dashed orange line shows the average ( $n = 62$ ) of the measured values, and the dashed green line indicates the certified value of the CRM (c). The absolute difference in TA values between duplicate CRM measurements (range) on each sampling day over the study period. The dashed orange line shows the average ( $n = 21$ ) of the ranges (d). Abbreviations: UCL, upper control limit; LWL, lower warning limit; LCL, lower control limit.

ously high DIC in M4 where photosynthetic biomass buildup was substantially lower (Fig. 3d). Hence, the depletion of nutrients (Fig. 2a, b) and increase in chl *a* concentration (Fig. 2c; Bach et al., 2020a) strongly suggest that the loss of DIC (except M4) was primarily driven by biological uptake and phytoplankton growth. Nevertheless, it is difficult to dis-

sect how much  $\text{CO}_2$  was outgassed and how much was taken up photosynthetically as we did not measure air–sea gas exchange in the mesocosms (please note that equations from Wanninkhof, 2014, are not applicable to mesocosms; Czerny et al., 2013b). In previous mesocosm studies,  $\text{N}_2\text{O}$  addition has been a common practice to monitor air–sea gas exchange

in the mesocosms (Czerny et al., 2013b). However, this was not carried out in our study because it would have interfered with  $^{15}\text{N}$  label incubations to determine N loss processes (Schulz et al., 2021). Due to high variability in DOC data and the poorly constrained gas exchange of  $\text{CO}_2$ , C budgeting often comes with high uncertainties and large errors, even for a relatively simple dataset (Czerny et al., 2013b; Boxhammer et al., 2018). It becomes even more difficult for the current dataset because the water column was not homogeneously mixed like in previous studies. With the inability to estimate  $\text{CO}_2$  gas exchange, it was impossible to calculate a reasonable C budget for this study. Before OMZ water addition, dissolved inorganic N : P ratios in the mesocosms ranged from 1.6 to 3.5 (data not shown), indicating N is the limiting nutrient in the water column (Bach et al., 2020a). Not surprisingly, the uptake of DIC was higher in the Low DIN mesocosms, which received more input of DIN from OMZ water addition, with on average  $41.0 \mu\text{mol kg}^{-1}$  higher drawdown compared to the Very low DIN from day 13 to day 24 (excluding M3 and M4; Mann–Whitney  $U$  test,  $p = 0.05$ ; Table S1). This observation agrees with the general expectations that the addition of limiting nutrients to the water column should enhance biological biomass buildup. Such differences in DIC uptake, however, were not reflected in the buildup of particulate organic carbon (POC) in the mesocosms (excluding M3 and M4; Mann–Whitney  $U$  test,  $p > 0.1$ ). As mentioned above, the differences in OMZ water DIN between the two treatments were minor, and hence, their potential to trigger treatment differences was small. Also, *Akashiwo sanguinea* was persistent in the water column in the mesocosms (except M4 where it never bloomed), retaining the biomass in the water column and not sinking out until the very end of the experiment (Bach et al., 2020a). Due to the developing N limitation after the biomass buildup there, much of the consumed DIC could have been channeled to the dissolved organic carbon (DOC) pool. Indeed, we observed a pronounced increase in DOC following OMZ water addition (except for M4; Igarza et al., 2022). The increase in DOC may be attributed to extracellular release by phytoplankton due to nutrient limitation or cellular lysis of phytoplankton cells by bacteria (Mykkestad, 2000; Igarza et al., 2022).

After day 24, variability in carbonate chemistry between individual mesocosms increased, with a general trend of recovering from  $\text{CO}_2$ -under-saturated conditions during the peak of the bloom (except for M3 and M4, Fig. 3c). One factor that may have controlled the differences in  $\text{CO}_2$  increase are the mesocosm-specific phytoplankton succession patterns. A shift from a diatom-dominated community to a dominance of dinoflagellates (in particular *Akashiwo sanguinea*) occurred when DIN was exhausted, which was absent in M3 and M4 (Bach et al., 2020a). The different succession patterns in the plankton community are the most likely explanation of why M3 and M4 behaved differently from the others in terms of surface layer productivity and hence carbonate chemistry. Although the rates of DIN depletion in M3

and M4 were similar to the others, the reduction in  $p\text{CO}_2$  in M3 experienced a 1-week delay, which is consistent with the delayed buildup of chl *a* biomass (Figs. 2c, 3c). On the other hand, the  $p\text{CO}_2$  level in M4 remained constantly elevated throughout the experiment, indicative of a lack of a phytoplankton bloom (Figs. 2c, 3c). M4 was the only mesocosm where *A. sanguinea* remained undetectable, whereas a delayed and reduced contribution by *A. sanguinea* was observed in M3. This strongly suggests that *A. sanguinea* was a key factor driving the trend of carbonate chemistry in the mesocosms.

Near the end of the experiment, a slight decline in  $p\text{CO}_2$  became apparent in the mesocosms, which co-occurred with a second phytoplankton bloom observed in the uppermost layer of the water column (Figs. 2c, 3c). This bloom was likely fueled by surface eutrophication due to defecating sea birds. During the last part of our experiment, Inca terns (*Larosterna inca*) were frequently observed to rest on the roofs and the edges of the mesocosms (Bach et al., 2020a). Bird excrements, dropped into the mesocosms, are known to be enriched in inorganic nutrients (Bedard et al., 1980). They generally contain 60 % water, 7.3 % N and, 1.5 % P, and the main form of N is uric acid and ammonium, which makes them slightly acidic (De La Peña-Lastra, 2021). Therefore, the droppings may lower the pH of the surface water. However, this should have been visible in decreasing alkalinity, which was not the case. The excrements may also be high in dissolved organic nitrogen (DON), evidenced by a substantial increase in DON concentrations in the mesocosm surface from day 38 onward (Igarza et al., 2022). The triggered surface eutrophication and phytoplankton blooms were noticeable from an accumulation of chl *a* biomass above the mixed layer in the mesocosms near the end of the study (Fig. 2c). As a result, another drawdown of DIC could be observed in the mesocosms except for M4, M6, and M8. While the buildup of chl *a* was comparable with that triggered by OMZ water addition, the drawdown in DIC was less pronounced, potentially counteracted by the release of  $\text{CO}_2$  by enhanced respiration and remineralization following the previous bloom. Also, the second bloom occurred in the top 2 meter in the mesocosms (Fig. 2c), where gas exchange can quickly replete the DIC drawdown during photosynthesis and biomass build up.

There are potential complications when monitoring the carbonate chemistry dynamics in an enclosed mesocosm. First, the brine addition to the mesocosms could have influenced the community composition which played an important role in driving the carbonate chemistry. However, the effects of the brine on the enclosed organisms have been discussed in past studies and considered negligible for a salinity increase of less than 1 ( $\sim 0.7$  and  $\sim 0.5$  increase for both salt additions in our study, respectively; Czerny et al., 2013a). The difference in salinity in the mesocosms from the Pacific was less than 1 throughout the study, so we believe salinity was not a stressor to the system (Bach et al., 2020a). Sec-

ond, multiple factors may introduce variabilities into air–sea gas exchange in the mesocosms. With the water surface sheltered from direct wind forcing by the 2 m high plastic walls of the mesocosm bag, air–sea gas exchange could be very low (but not zero due to other energy inputs such as thermal convection and surrounding wave movements; Czerny et al., 2013b). On the other hand, the extensive daily sampling that actively perturbs the mesocosm surface may enhance the gas exchange (Czerny et al., 2013b). With a high variability in DOC data and a lack of direct measurements of CO<sub>2</sub> gas exchange, it is impossible to estimate a reasonable C budget for our dataset (Czerny et al., 2013b; Boxhammer et al., 2018). Other complications include heterogenous initial conditions in the mesocosms and light limitation due to self-shading (please see Bach et al., 2020a, for an extensive discussion). It would have been ideal to have had control mesocosms that were treated the same way except in terms of the OMZ water addition to rule out the effects induced by mesocosm manipulations and changes in hydrodynamics. Not having this was a compromise to ensure enough replicate numbers for both treatments despite the enormous cost of mesocosm experimentation. Nevertheless, a previous study has examined impacts of different mixing techniques in outdoor mesocosms and found no effects on phytoplankton biomass and minor effects on phytoplankton and zooplankton community composition (Striebel et al., 2013). In our study, various measures were also taken to minimize the mixing (brine additions and slow casting of CTD).

### 4.3 Temporal changes in carbonate chemistry in the coastal Pacific near Callao

According to estimations by Takahashi et al. (2009) of global air–sea CO<sub>2</sub> fluxes, our study site in the equatorial Pacific (14° N–14° S) is a major source of CO<sub>2</sub> to the atmosphere. Our near-coastal location showed high *p*CO<sub>2</sub> levels over the study period (with an average of 1078 μatm), with a sea-to-air CO<sub>2</sub> flux of 4.2–14.0 mmol C m<sup>-2</sup> d<sup>-1</sup> (Fig. 5). Compared to the criterion of high CO<sub>2</sub> fluxes (5 mmol C m<sup>-2</sup> d<sup>-1</sup> or more) as proposed by Paulmier et al. (2008), our study site was a strong CO<sub>2</sub> source to the atmosphere most of the time. These results of air–sea CO<sub>2</sub> fluxes were slightly higher than observations by Friederich et al. (2008) along the coast of Peru in February 2004–2006 (0.85–4.54 mol C m<sup>-2</sup> yr<sup>-1</sup>, spatially averaged for 5–15° S along the coast of Peru). This is not surprising because Friederich et al. (2008) averaged the air–sea CO<sub>2</sub> fluxes for 0–200 km from the shore, where much lower *p*CO<sub>2</sub> levels were observed offshore (< 600 μatm) compared to our nearshore study site. The decline in *p*CO<sub>2</sub> with increasing distance from shore was driven by biological uptake and outgassing to the atmosphere (Friederich et al., 2008; Loucaides et al., 2012). However, when compared to the magnitude of DIC drawdown triggered by upwelling events in the mesocosms, the flux of CO<sub>2</sub> to the atmosphere was insignificant. Assuming a

10 m mixed layer in the Pacific with a DIC concentration of 2200 μmol kg<sup>-1</sup>, the DIC content below a 1 m<sup>2</sup> surface area would be ~ 22 mol m<sup>-2</sup>. With an upper bound outgassing of 14.2 mmol C m<sup>-2</sup> d<sup>-1</sup> over 10 d (day 13–24), the loss of CO<sub>2</sub> would only be 0.142 mol m<sup>-2</sup>. On the other hand, the average DIC drawdown of 118.2 μmol kg<sup>-1</sup> in the Very low DIN and 160.3 μmol kg<sup>-1</sup> in the Low DIN mesocosms (M3 and M4 excluded) during this period accounts for 1.18 and 1.60 mol m<sup>-2</sup>, respectively, over the same water column. This shows that biological processes drawing down CO<sub>2</sub> are stronger than loss by air–sea gas exchange.

During our study, we experienced a coastal El Niño event which was the strongest on record (compared to those recorded in 1891 and 1925) and induced rapid sea surface warming of ~ 1.5 °C and enhanced stratification (Garreaud, 2018). Previous investigations showed that the impact of reduced upwelling on CO<sub>2</sub> fluxes is pronounced for upwelling areas (Feely et al., 1999, 2002). A decline in upwelling of CO<sub>2</sub>-enriched OMZ water results in a decrease in sea-to-air CO<sub>2</sub> fluxes. For example, during the 1991–1994 El Niño year, a total reduction in CO<sub>2</sub> fluxes to the atmosphere was reported for the equatorial Pacific. They were only 30%–80% of that of a non-El Niño year (Feely et al., 1999, 2002). This is likely to be the case for our study location. Most studies have investigated air–sea CO<sub>2</sub> fluxes at larger timescales and regional scales (Feely et al., 1999; Friederich et al., 2008; Takahashi et al., 2009). Therefore, it is difficult to conclude the magnitude of the coastal El Niño influence on the local CO<sub>2</sub> fluxes in our study by comparing our results with previous observations. Nevertheless, our observations can serve as the first evidence of carbonate chemistry dynamics in the coastal Peruvian upwelling system during a coastal El Niño event. Observations of sea surface carbonate chemistry with a high temporal resolution (every second day) in nearshore waters are scarce, as they are rarely covered by typical research expeditions in the open ocean (Takahashi et al., 2009; Franco et al., 2014), especially during such an extremely rare coastal El Niño event. Comparisons of our data with previous or future observations may enhance our understanding of how inorganic carbon cycling interacts with extreme climate events in upwelling systems.

CO<sub>2</sub>-enriched OMZ water has been occasionally reported to be under-saturated with respect to aragonite (Feely et al., 2008; Fassbender et al., 2011). In our study, calcite under-saturation did not occur in the mesocosms or in the Pacific (Fig. 4). Aragonite under-saturation, however, was observed below the surface (10–17 m for day 3–28, 12.5–17 m for day 29–50) of the Pacific at the start of the experiment (Fig. 4d), when *p*CO<sub>2</sub> was the highest (*p*CO<sub>2</sub> > 1100 μatm, Fig. 3c). Aragonite under-saturation was also observed in the two deep water masses collected at deeper depths (30 and 70 m) in the Pacific (Table 4). Throughout the study period, the aragonite saturation state fluctuated close to around 1 below the surface (Fig. 4d). Considering the water column we sampled in the Pacific still belonged to the upper surface

ocean, we could expect deeper and more CO<sub>2</sub>-enriched water in the underlying OMZ to be most likely under-saturated with respect to calcite and aragonite. Hence, our observations of aragonite under-saturation in the Pacific suggest a potential risk of dissolution for marine calcifiers in response to the ongoing intensification and expansion of acidified OMZ water (Comeau et al., 2009; Lischka et al., 2011; Maas et al., 2012).

## 5 Conclusions

Our observations in the mesocosms revealed that, following the addition of two OMZ water masses with different nutrient signatures, there was a higher drawdown of DIC in response to slightly more DIN input from the OMZ water addition but no difference in the buildup of POC and chl *a* (Figs. 2a and c, 3d). The timing of the first phytoplankton bloom was consistent with a shift from a diatom-dominated community to *A. sanguinea* dominance in most mesocosms, indicating that *A. sanguinea* was a key factor driving the changes in carbonate chemistry under N-limited conditions. A second phytoplankton bloom was triggered by defecation of Inca terns, which eased the N limitation in the mesocosms (Fig. 2c). These findings provide improved insights into the links between upwelling-induced N limitation, phytoplankton community shifts, and carbonate chemistry dynamics in the Peruvian upwelling system.

The surrounding Pacific waters at the study site were characterized by constantly high *p*CO<sub>2</sub> levels (with an average of 1078.1  $\mu$ atm). Most CO<sub>2</sub> flux estimates have been conducted in the open ocean, and few studies have surveyed coastal regions (Takahashi et al., 2009; Franco et al., 2014). Our study site was a strong CO<sub>2</sub> source to the atmosphere most of the time (4.2–14.2 mmol C m<sup>-2</sup> d<sup>-1</sup>), despite a rare coastal El Niño event. However, evidence from our mesocosm experiment suggests biological responses that draw down DIC can quickly turn a CO<sub>2</sub> source into a sink in the upwelling system. The influence of the co-occurring coastal El Niño event on the local CO<sub>2</sub> fluxes remains unclear. Nevertheless, future carbonate chemistry fluctuations are expected to be enhanced by expanding and intensifying ocean deoxygenation, as well as by reducing buffer factors (Schulz et al., 2019). Hence, it is essential to improve our understanding of the mechanisms driving the inorganic carbon cycling in upwelling systems. As a unique dataset that characterized nearshore carbonate chemistry with a high temporal resolution during a rare coastal El Niño event, our study gives important insights into the carbonate chemistry responses to extreme climate events in the Peruvian upwelling system.

**Data availability.** The dataset is now available under <https://doi.org/10.1594/PANGAEA.933337> (Chen et al., 2021) and <https://doi.org/10.1594/PANGAEA.923395> (Bach et al., 2020b).

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**Author contributions.** UR, KGS, and LTB designed the experiment. All authors contributed to the sampling. SMC measured, calculated, and analyzed the carbonate chemistry. LTB and KGS supervised the carbonate chemistry analysis. KGS carried out the CTD casts and data analyses. EvdE and EPA measured and analyzed nutrients. SMC wrote the manuscript with input from all the co-authors.

**Competing interests.** The contact author has declared that neither they nor their co-authors have any competing interests.

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