



Supplement of

Shifts in organic matter character and microbial assemblages from glacial headwaters to downstream reaches in the Canadian Rocky Mountains

Hayley F. Drapeau et al.

Correspondence to: Suzanne E. Tank (suzanne.tank@ualberta.ca)

The copyright of individual parts of the supplement might differ from the article licence.

Table S1: Site characteristics describing distance range, glacier source of headwaters, name of river, coordinates (in degrees, decimal minutes), approximate distance of sampling site from glacier terminus (estimated from Google Earth), and the number of times each site was visited in this study. Distance from glacier terminus and catchment coverage are as in Serbu et al. (2024).

Distance Range	Glacier headwaters	River	Latitude	Longitude	Distance from glacier terminus (km)	Number of times visited	Area	Catchment coverage (snow+ice / rock / forest)
Headwater	Athabasca	Sunwapta	52°12'25.8"	-117°14'05.9"	0.2	16	22.7	50.7 / 46.8 / 0.0
Near	Athabasca	Sunwapta	52°12'59.5"	-117°14'01.9"	1.7	15	29.3	41.8 / 54.2 / 0.3
Mid	Athabasca	Sunwapta	52°18'38.1"	-117°19'57.3"	15.5	17	198	19.1 / 55.3 / 12.8
Far	Athabasca	Sunwapta	52°31'58.7"	-117°38'39.2"	52.9	17	731	6.6 / 50.2 / 25.1
Far	Athabasca	Athabasca	52°34'59.6"	-117°44'21.9"	63.1	14	1635	12.2 / 43.2 / 27.3
Far	Athabasca	Athabasca	52°39'46.5"	-117°52'51.7"	73.9	18	1956	10.8 / 42.8 / 29.0
Far	Athabasca	Athabasca	52°48'43.4"	-118°02'33.2"	97.8	16	3020	8.4 / 41.2 / 32.0
Near	Saskatchewan	NSR	52°10'10.1"	-117°04'34.9"	5.6	18	76.2	54.7 / 35.5 / 3.6
Mid	Saskatchewan	NSR	52°4'9.1"	-116°54'54.9"	24.6	16	616	19.1 / 40.8 / 24.4
Mid	Saskatchewan	NSR	51°58'14.0"	-116°43'16.0"	46.3	16	1551	18.8 / 37.4 / 29.7
Near	Bow	Bow	51°39'42.3"	-116°29'13.0"	2.4	14	21.4	41.5 / 47.7 / 4.8
Mid	Bow	Bow	51°37'53.0"	-116°20'6.6"	17.1	15	105	11.1 / 42.0 / 31.4
Mid	Bow	Bow	51°25'43.2"	-116°11'20.4"	51.3	17	422	9.6 / 35.7 / 41.0
Far	Bow	Bow	51° 17' 5.82"	- 115° 59' 0.6"	75.4	13	1104	5.6 / 38.1 / 41.3

Table S2: Detection limits and methodological references for parameters assessed in this study. See also Serbu et al. (2024) for details.

Analyte	Detection limit	Method reference
TN	3 $\mu\text{g L}^{-1}$	EPA 353.2 (U.S. EPA, 1993b)
TP	1 $\mu\text{g L}^{-1}$	
NH ₄ ⁺	3 $\mu\text{g L}^{-1}$	SM 4500-NH3 (SM-APHA _b)
TDN	3 $\mu\text{g L}^{-1}$	
TDP	1 $\mu\text{g L}^{-1}$	SM 4500-P (SM-APHA _c)
SRP	1 $\mu\text{g L}^{-1}$	
NO ₂ ⁻ +NO ₃ ⁻	2 $\mu\text{g L}^{-1}$	SM 4500-SiO ₂ (SM-APHA _d)
Si	0.02 mg L ⁻¹	
Mg ²⁺	0.05 mg L ⁻¹	EPA 200.7 (U.S. EPA, 1994), SM 3125 (SM-APHA _a)
Na ⁺	0.05 mg L ⁻¹	
Ca ²⁺	0.05 mg L ⁻¹	
K ⁺	0.05 mg L ⁻¹	
Cl ⁻	0.03 mg L ⁻¹	
SO ₄ ²⁻	0.04 mg L ⁻¹	EPA 300.1 (U.S. EPA, 1993a)
Al ³⁺	0.49 $\mu\text{g L}^{-1}$	EPA 200.7, (U.S. EPA, 1994), SM 3125 (SM-APHA _a)
Ba	0.05 $\mu\text{g L}^{-1}$	
Cr	0.02 $\mu\text{g L}^{-1}$	
Mn	0.04 $\mu\text{g L}^{-1}$	
Mo	0.01 $\mu\text{g L}^{-1}$	
Ni	0.02 $\mu\text{g L}^{-1}$	
Sr	0.04 $\mu\text{g L}^{-1}$	

Table S3: Description of characteristics for various the absorbance- and fluorescence-based metrics used in this study.

Measurement	Calculation	Purpose
SUVA ₂₅₄	Normalization of UV absorbance at 254 nm to DOC	Increasing values indicate increasing aromaticity (Weishaar et al. 2003)
S ₂₇₅₋₂₉₅	Slope over the 275nm-295nm wavelength range	Decreasing values indicates increasing molecular weight (Helms et al. 2008)
BIX	The ratio of emission at 380nm by 430nm at excitation of 310nm	Biological Index: Increasing values indicate recently produced organic carbon (Huguet et al. 2008)
HIX	Fluorescence intensity over 300-340nm divided by the combined fluorescence in the 300-345nm and 435-480nm	Humification Index; Increasing values indicate increasing humification (Ohno 2002)
FI	The ratio of emission at 400nm to 500nm at an excitation of 370nm	Fluorescence Index: Higher values indicate microbially derived fulvic acids, lower values indicate terrestrially derived fulvic acids (McKnight et al. 2001)

Table S4: Outputs of mixed effects models showing random effects and the fixed effect of distance based on the full model structure of *constituent ~ distance * melt.season + (1|river) + (1|year)*. Fixed effects of melt season and the interaction effect are not shown directly, but are reflected in the analysis of deviance. For the analysis of deviance outputs, probabilities <0.05 are highlighted in bold, and probabilities <0.10 are italicized. Where the effect of melt season was significant, post-hoc contrasts are shown in Table S5.

Constituent	Random Effects						Fixed Effects			Analysis of Deviance								
	River (Intercept)		Year (Intercept)		Residual		Distance			Distance			Melt Season			Interaction		
	Variance	SD	Variance	SD	Variance	SD	Estimate	SE	t-value	ChiSq	df	pr>ChiSq	ChiSq	df	pr>ChiSq	ChiSq	df	pr>ChiSq
d-excess	1.15E-01	3.39E-01	2.22E-03	4.71E-02	6.43E-01	8.02E-01	-1.91E-02	2.70E-03	-7.099	58.746	1	0.000	34.401	3	0.000	2.035	3	0.565
O-18	6.23E-02	2.50E-01	4.18E-02	2.05E-01	3.57E-01	5.98E-01	-1.16E-03	2.00E-03	-0.579	0.005	1	0.944	30.805	3	0.000	9.053	3	0.029
DOC	5.99E-03	7.74E-02	8.72E-03	9.34E-02	1.15E-01	3.39E-01	5.46E-03	1.05E-03	5.184	44.865	1	0.000	60.328	3	0.000	17.063	3	0.001
POC	1.13E-02	1.06E-01	3.89E-03	6.24E-02	1.60E-01	4.00E-01	5.34E-04	1.27E-03	0.420	0.569	1	0.451	5.195	3	0.158	1.194	3	0.754
%POC	9.71E-03	9.86E-02	2.77E-03	5.27E-02	2.47E-02	1.57E-01	-2.43E-03	5.59E-04	-4.353	18.308	1	0.000	23.152	3	0.000	3.242	3	0.356
Humic	2.43E-03	4.93E-02	3.08E-04	1.76E-02	3.77E-02	1.94E-01	3.68E-03	6.21E-04	5.929	43.959	1	0.000	22.602	3	0.000	0.734	3	0.865
13C-DOC	0	0	1.77E+00	1.33E+00	4.85E+00	2.20E+00	1.64E-02	6.88E-03	2.378	3.569	1	<i>0.059</i>	9.683	3	0.022	2.688	3	0.442
13C-POC	1.14E-09	3.37E-05	6.06E+00	2.46E+00	3.03E+00	1.74E+00	7.99E-03	4.83E-03	1.653	6.232	1	0.013	1.580	3	0.664	10.703	3	0.013
14C-DOC	2.84E-05	3.26E-04	6.56E+03	8.10E+01	1.44E+04	1.20E+02	3.81	2.12	1.795	6.033	1	0.014	23.704	3	0.000	1.285	3	0.733
14C-POC	0	0	1.01E+04	1.01E+02	1.08E+04	1.04E+02	-1.89	3.39	-0.557	0.461	1	0.497	3.283	2	0.194	2.656	3	0.265

Table S5: Post-hoc contrasts following significant effect of melt season in the models presented in Table S4.

				Estimate	SE	df	t-ratio	p-value
d-excess	Melt	-	PostMelt	0.53227	0.265	37.7	2.009	0.203
	Melt	-	PreMelt	1.09275	0.218	94.9	5.021	<.001
	Melt	-	Winter	0.00836	0.507	163.6	0.016	1.000
	PostMelt	-	PreMelt	0.56048	0.296	164	1.894	0.235
	PostMelt	-	Winter	-0.52391	0.528	182.4	-0.992	0.754
	PreMelt	-	Winter	-1.08439	0.525	184.3	-2.065	0.169
O-18	Melt	-	PostMelt	-0.56	0.194	174	-2.891	0.022
	Melt	-	PreMelt	0.613	0.153	187	3.995	0.001
	Melt	-	Winter	-0.371	0.375	187	-0.989	0.756
	PostMelt	-	PreMelt	1.173	0.217	187	5.393	<.001
	PostMelt	-	Winter	0.189	0.393	185	0.481	0.963
	PreMelt	-	Winter	-0.984	0.389	187	-2.531	0.058
DOC	Melt	-	PostMelt	0.2232	0.11	164	2.030	0.181
	Melt	-	PreMelt	-0.4948	0.081	188	-6.112	<.001
	Melt	-	Winter	0.1704	0.145	192	1.173	0.645
	PostMelt	-	PreMelt	-0.7179	0.121	190	-5.913	<.001
	PostMelt	-	Winter	-0.0528	0.174	192	-0.304	0.990
	PreMelt	-	Winter	0.6652	0.156	190	4.261	<.001
%POC	Melt	-	PostMelt	0.17317	0.0659	160	2.628	0.046
	Melt	-	PreMelt	0.17064	0.0413	166	4.134	<.001
	Melt	-	Winter	0.10376	0.1033	165	1.004	0.747
	PostMelt	-	PreMelt	-0.00253	0.0721	166	-0.035	1.000
	PostMelt	-	Winter	-0.06941	0.1216	166	-0.571	0.941
	PreMelt	-	Winter	-0.06689	0.1079	165	-0.620	0.926
Humic	Melt	-	PostMelt	0.1426	0.0641	86	2.223	0.125
	Melt	-	PreMelt	-0.1662	0.049	149	-3.394	0.005
	Melt	-	Winter	0.0864	0.1029	178	0.840	0.835
	PostMelt	-	PreMelt	-0.3088	0.075	135	-4.120	<.001
	PostMelt	-	Winter	-0.0561	0.1215	136	-0.462	0.967
	PreMelt	-	Winter	0.2527	0.1078	184	2.343	0.092
13C-DOC	Melt	-	PostMelt	-2.168	0.791	129	-2.740	0.035
	Melt	-	PreMelt	0.659	0.741	129	0.889	0.811
	Melt	-	Winter	4.236	9.642	129	0.439	0.972
	PostMelt	-	PreMelt	2.826	1.087	129	2.600	0.050
	PostMelt	-	Winter	6.404	9.673	130	0.662	0.911
	PreMelt	-	Winter	3.577	9.66	129	0.370	0.983
14C-DOC	Melt	-	PostMelt	3.72	99.8	39.9	0.037	1.000
	Melt	-	PreMelt	-9.69	68	38.9	-0.142	0.999
	Melt	-	Winter	408.70	96	38.5	4.257	0.001
	PostMelt	-	PreMelt	-13.4	121.3	40.5	-0.111	1.000
	PostMelt	-	Winter	404.99	139.1	40.1	2.912	0.029
	PreMelt	-	Winter	418.39	105.7	37.2	3.959	0.002

Table S6: Excitation and emission maxima associated with PARAFAC components and associated DOM character as assessed by cross reference with the Open Fluor database.

Component	Emission peak	Excitation peak	Character	Number of Open Fluor Matches
C1	271	230	Terrestrial / humic (Shutova et al., 2014)	118
C2	409	230	Terrestrial / humic (Catalán et al., 2018; Coulson et al., 2022; Shutova et al., 2014)	105
C3	340	230	Microbial/tryptophan like (Murphy et al., 2008; Stedmon & Markager, 2005)	27
C4	300	230, 270-275	Microbial/tyrosine like(Graeber et al., 2012)	14

Table S7: perMANOVA test outputs showing the variation between proportion of microbial communities by distance range, hydrological season, year, and river. perMANOVA pairwise comparisons with holm adjusted p values are shown. Significant interactions are bolded.

Factor	R²	significance	PERMANOVA pairwise comparison	p (Holms adjusted)
Distance Range	0.09	< 0.001	Headwater - Near	0.1659
			Headwater - Far	0.0044
			Near - Far	0.0003
Year	0.08	<0.001	2019-2020	0.0003
			2019-2021	0.0024
			2020-2021	0.0068
Season	0.06	0.002	PreMelt – Melt	0.0039
			PreMelt – PostMelt	0.0602
			Melt – PostMelt	0.2125
River	0.08	<0.001	Sunwapta-Athabasca	0.0027
			Sunwapta-NSR	0.0003
			Athabasca-NSR	0.0003

Table S8: Potential OC sources to glacially-influenced streams, and their associated expected fluorescence signature and $\delta^{13}\text{C-OC}$ and $\Delta^{14}\text{C-OC}$ ranges.

Source	$\delta^{13}\text{C-OC}$	$\Delta^{14}\text{C-OC}$	Expected DOM fluorescence
Vegetation	-26 to -28‰ (Peterson and Fry, 1987)	Downstream (contemporary) vegetation will be modern whereas subglacial over-ridden vegetation likely represents an aged OC source (Bhatia et al., 2013)	Humic-like (Gabor et al., 2014)
Soils and associated pore water	-26 to -28‰ (Peterson and Fry, 1987)	Modern at the surface with increasing age with depth (Shi et al., 2020)	Humic-like with decreasing humification and increasing protein-like fluorescence with depth and increased soil residence time (Gabor et al., 2014) (McDonough et al., 2022)
Phytoplankton	-22 to -30‰ (Chanton & Lewis, 1999)	Ranging from modern to slightly aged depending on CO_2 source	Protein-like (Fellman et al., 2010)
Benthic algae	Greater than -7 to -15‰ due to decreased isotopic fractionation, following carbon limitation associated with benthic boundary layers (Hecky & Hesslein, 1995)	Ranging from modern to slightly aged depending on CO_2 source	Protein-like (Fellman et al., 2010)
Fossil fuel deposits	-27‰ (Peterson & Fry, 1987)	Ancient (radiocarbon dead)	Variable (Mladenov et al., 2010)
Wildfire derived soot	-26 to -28‰ (Peterson & Fry, 1987)	Modern (Masiello & Druffel, 2003)	Variable (Mladenov et al., 2010)
Chemosynthesis	Very depleted -30 to -80‰, via sulfur oxidizing and methanogenic microbes (Blaser & Conrad, 2016; Rau & Hedges, 1979; Ruby et al., 1987)	Ranging from modern to slightly aged depending on the CO_2 source. Notably, subglacial CO_2 can represent an aged C source	Protein-like (Fellman et al., 2010)

Table S9: Overview of all sample codes used in this study, with corresponding river (Sunwapta, Athabasca, North Saskatchewan, Bow), distance downstream, and month and year of sampling. “E” and “L” refers to samples collected early, and late, in June 2020.

Sample ID	River	Distance	MMYY	Sample ID	River	Distance	MMYY
R19_04	S	52.9	0519	R20_61	S	52.9	0920
R19_05	A	73.9	0519	R20_71	A	97.8	1020
R19_06	A	97.8	0519	R20_73	S	1.7	1020
R19_09	N	5.6	0519	R20_75	S	52.9	1020
R19_25	A	63.1	0719	R21_01	S	52.9	0121
R19_29	S	1.7	0719	R21_03	A	73.9	0121
R19_31	S	52.9	0719	R21_05	A	63.1	0521
R19_39	A	63.1	0819	R21_06	A	73.9	0521
R19_41	A	97.8	0819	R21_07	A	97.8	0521
R19_43	S	1.7	0819	R21_08	S	0.2	0521
R19_45	S	52.9	0819	R21_09	S	1.7	0521
R19_46	N	5.6	0819	R21_11	S	52.9	0521
R19_57	S	1.7	1019	R21_12	N	5.6	0521
R19_59	S	52.9	1019	R21_18	A	63.1	0621
R20_02	S	1.7	0620E	R21_19	A	73.9	0621
R20_04	S	52.9	0620E	R21_20	A	97.8	0621
R20_05	A	63.1	0620E	R21_21	S	0.2	0621
R20_06	A	73.9	0620E	R21_22	S	1.7	0621
R20_07	A	97.8	0620E	R21_24	S	52.9	0621
R20_08	N	5.6	0620E	R21_25	N	5.6	0621
R20_15	A	73.9	0620L	R21_34	A	97.8	0721
R20_16	A	97.8	0620L	R21_38	S	52.9	0721
R20_19	S	52.9	0620L	R21_39	N	5.6	0721
R20_20	N	5.6	0620L	R21_45	A	63.1	0821
R20_28	A	73.9	0720	R21_46	A	73.9	0821
R20_29	A	97.8	0720	R21_47	A	97.8	0821
R20_30	S	0.2	0720	R21_48	S	0.2	0821
R20_31	S	1.7	0720	R21_49	S	1.7	0821
R20_33	S	52.9	0720	R21_51	S	52.9	0821
R20_34	N	5.6	0720	R21_52	N	5.6	0821
R20_42	A	73.9	0820	R21_61	A	97.8	0921
R20_47	S	52.9	0820	R21_62	S	0.2	0921
R20_48	N	5.6	0820	R21_63	S	1.7	0921
R20_55	A	63.1	0920	R21_65	S	52.9	0921
R20_56	A	73.9	0920	R21_66	N	5.6	0921
R20_58	S	0.2	0920	R21_snow	Sn		

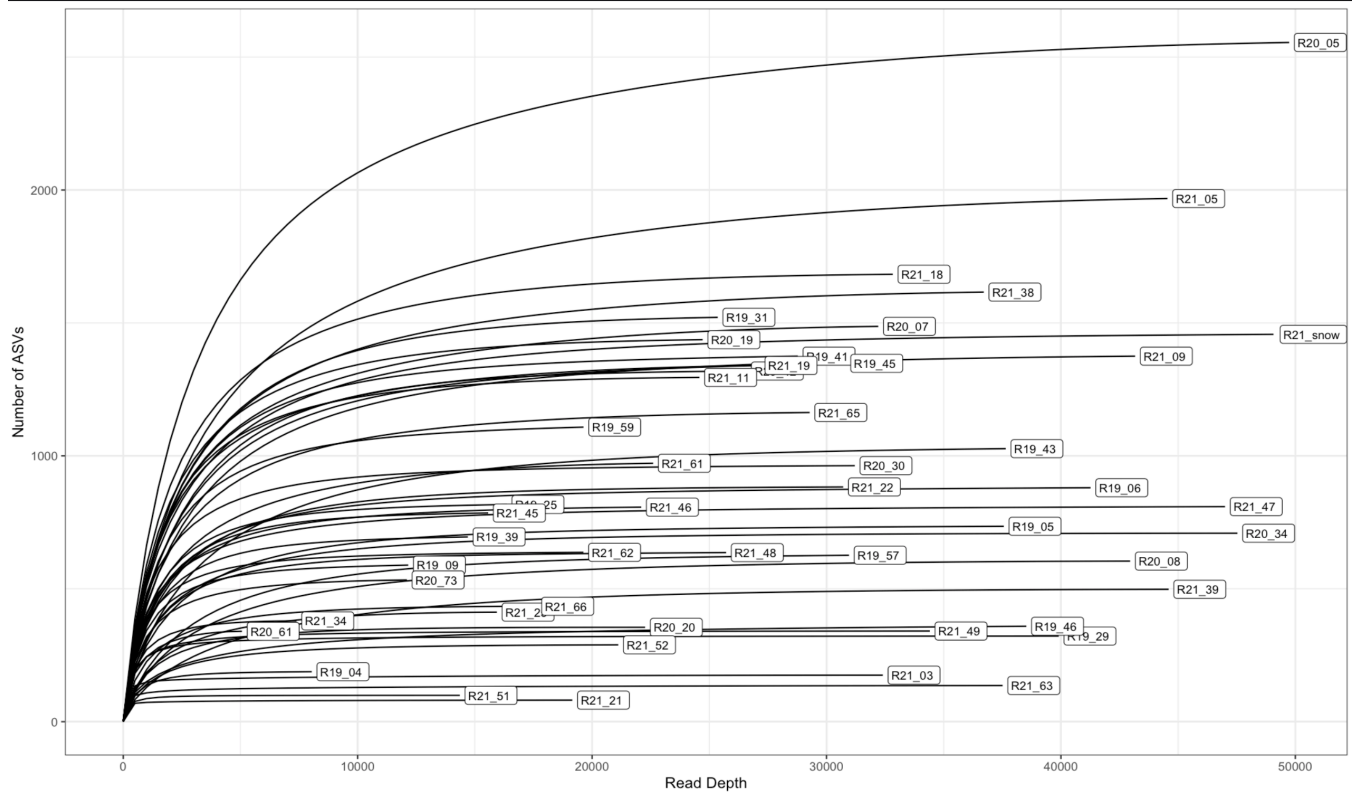
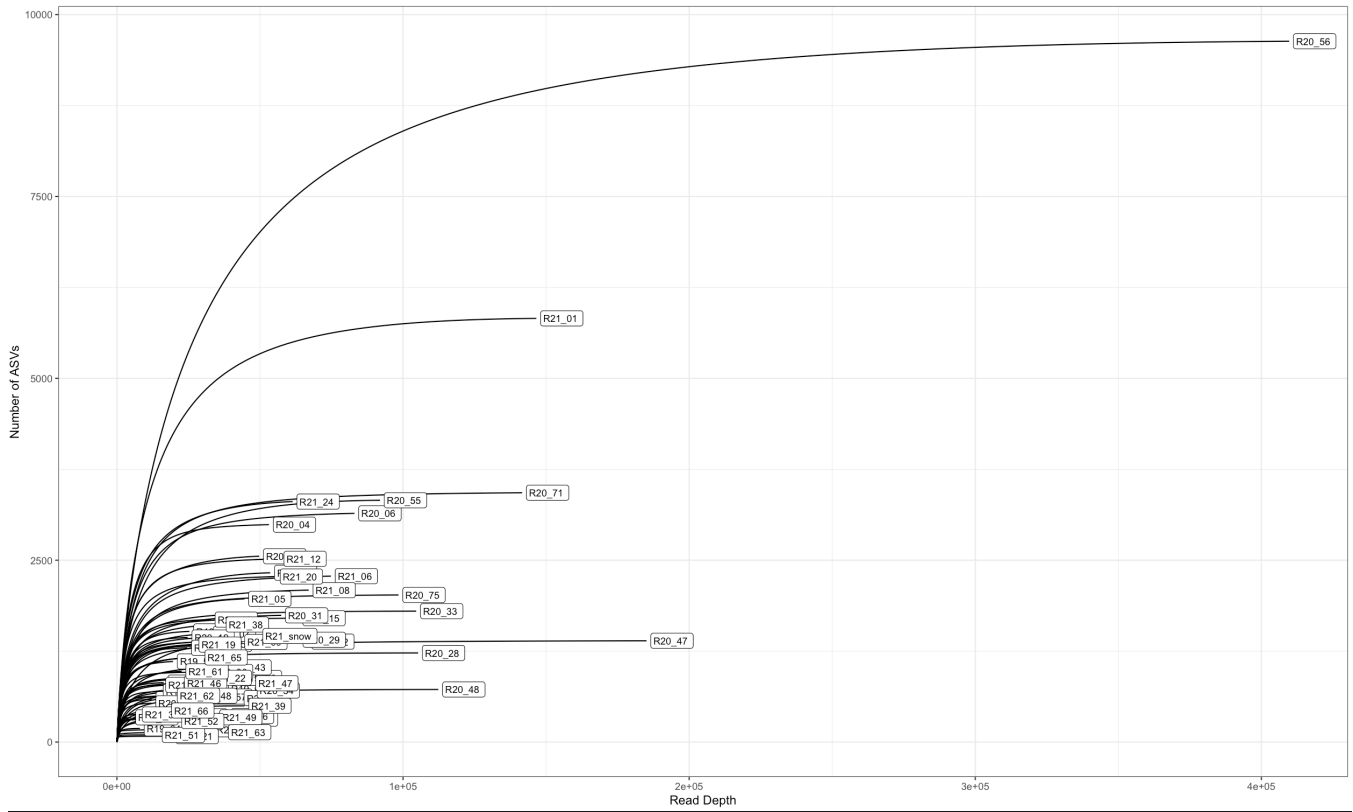


Figure S1: Rarefaction curves to show read depth relative to ASV count for individual samples collected during this study. The top panel shows all samples. The bottom panel shows only samples with a read depth of less than 50,000. Samples with read depths of less than 5,000 were excluded, and are therefore not shown in these plots. Sample codes correspond with rivers and dates in Table S9.

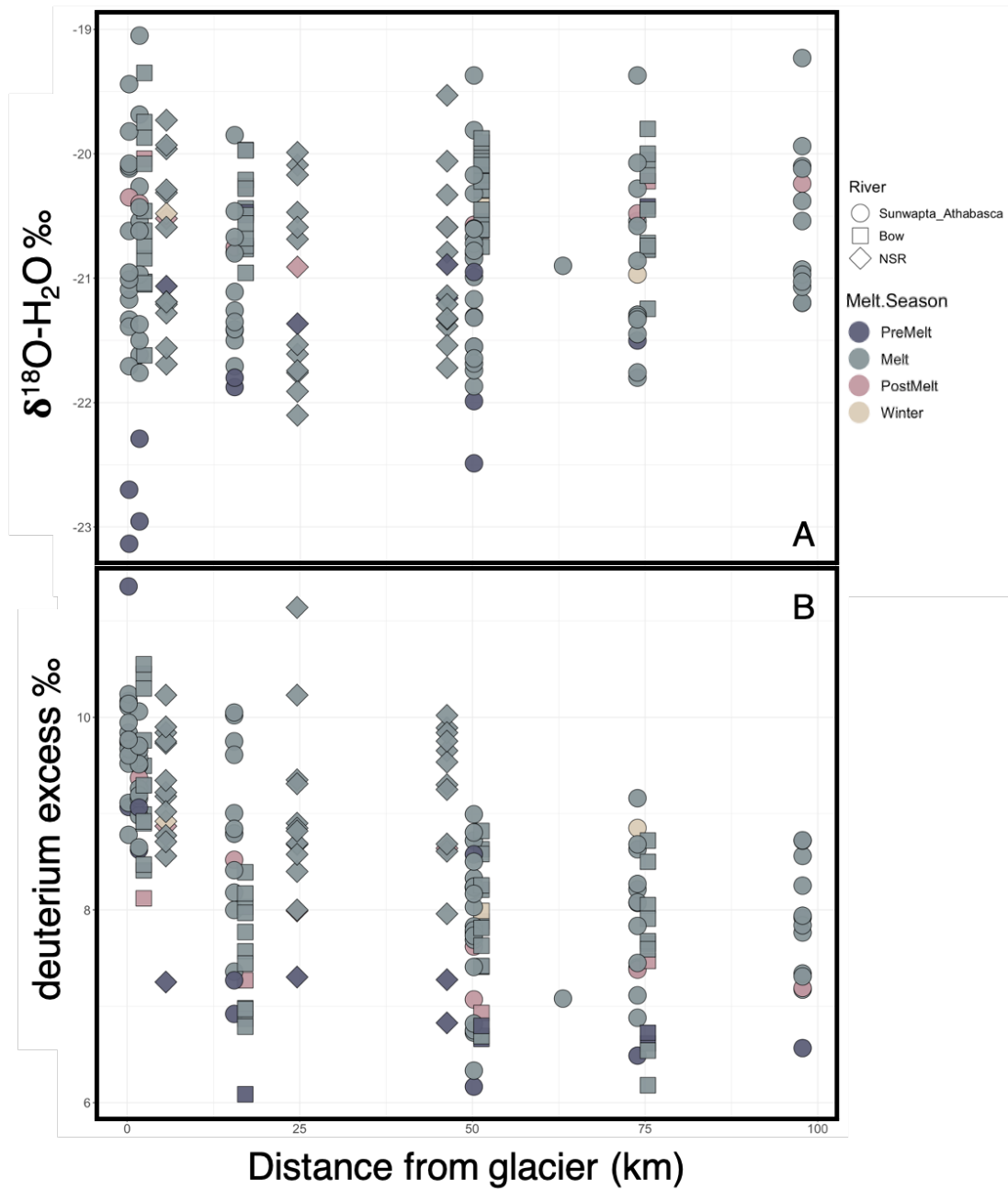


Figure S2: Plots of (A) $\delta^{18}\text{O}-\text{H}_2\text{O}$, and (B) deuterium excess with increasing distance downstream from glacial terminus. Melt season and river are indicated.

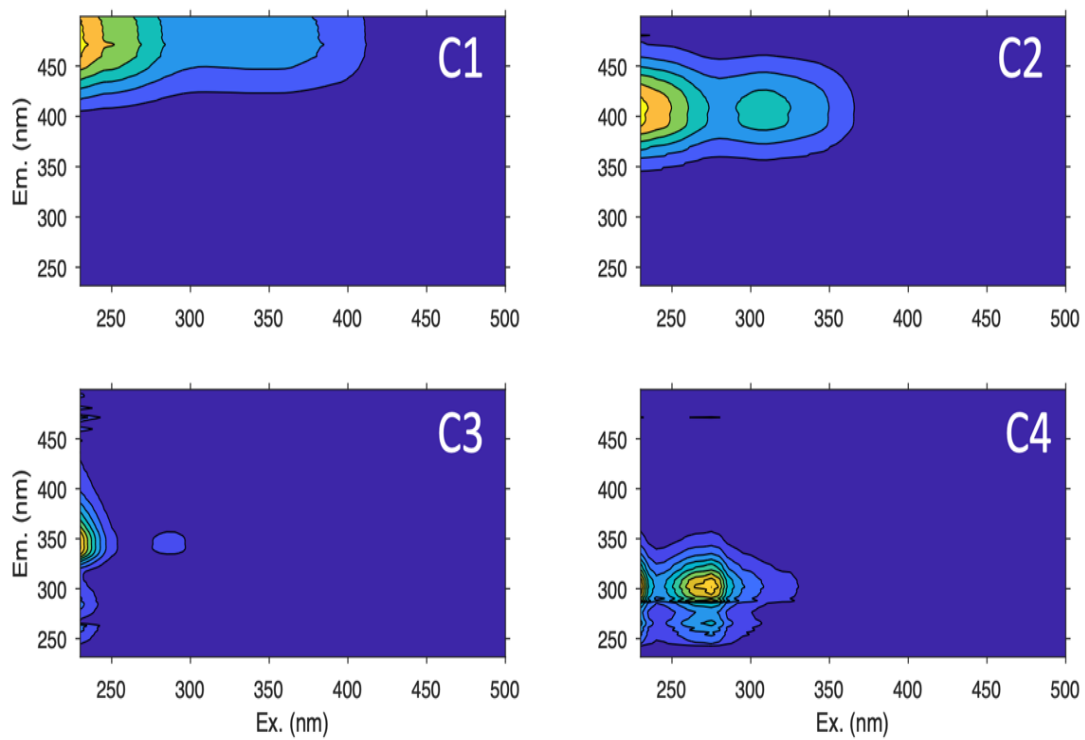


Figure S3: Component plots for the four-component PARAFAC model

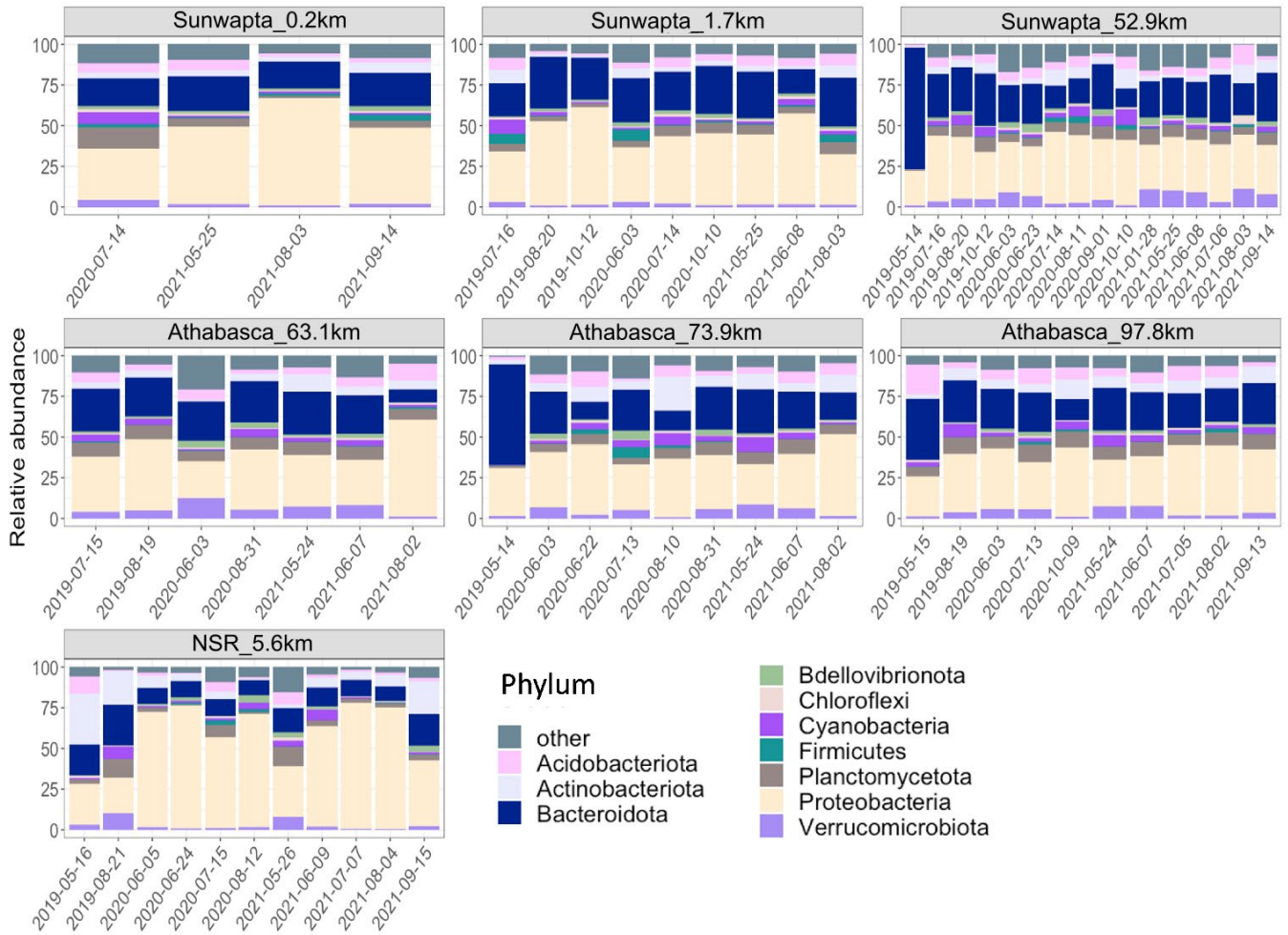


Figure S4: Relative abundance of the top ten most abundant phyla across all sampling dates at headwater, near and far sites in the Sunwapta-Athabasca and North Saskatchewan rivers. The category “Other” represents classes not included in the top ten.

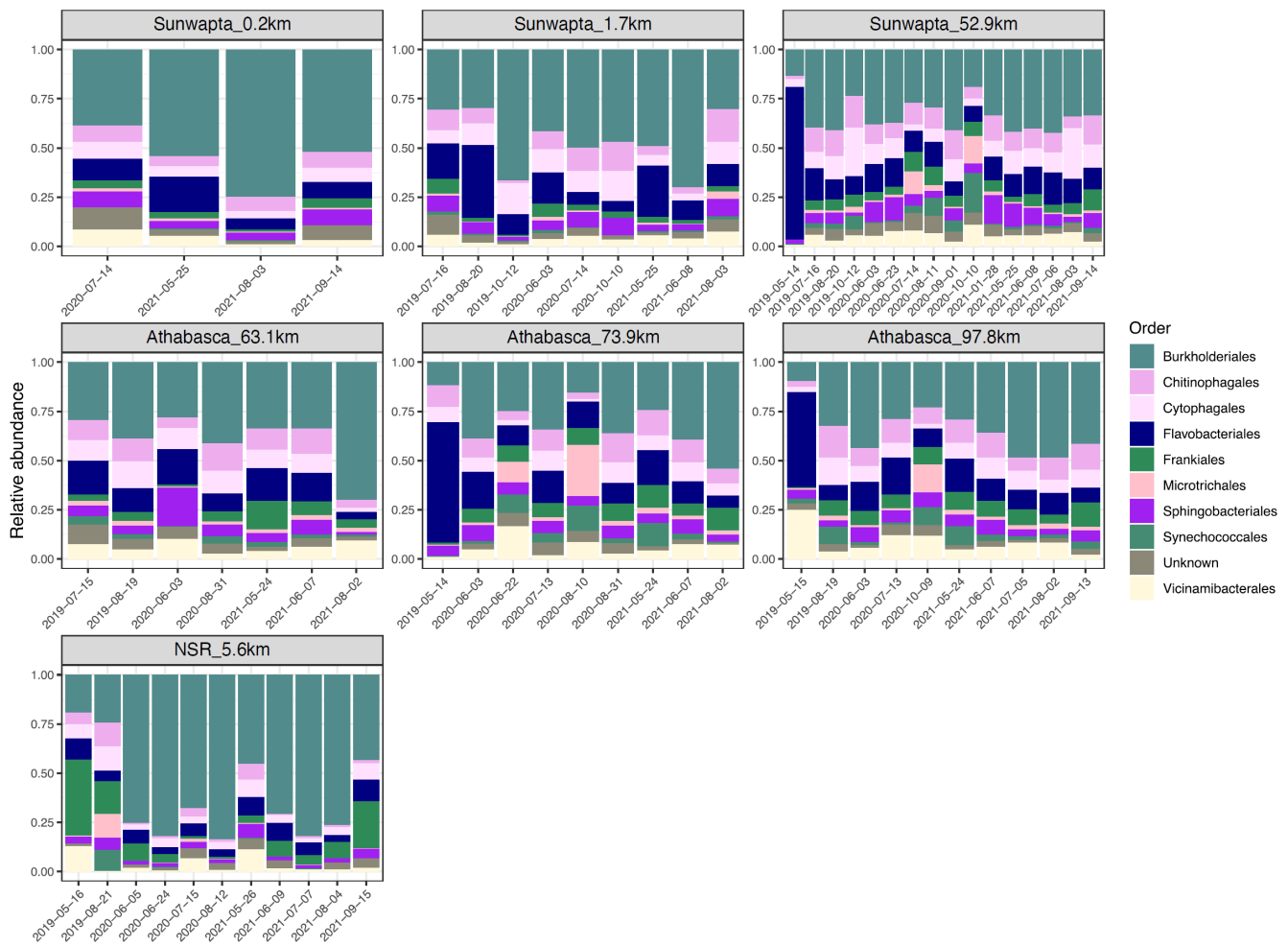


Figure S5: Relative abundance of the top ten most abundant orders across all sampling dates at the near and far sites in the Sunwapta-Athabasca and North Saskatchewan rivers. The category “Other” represents classes not included in the top ten.

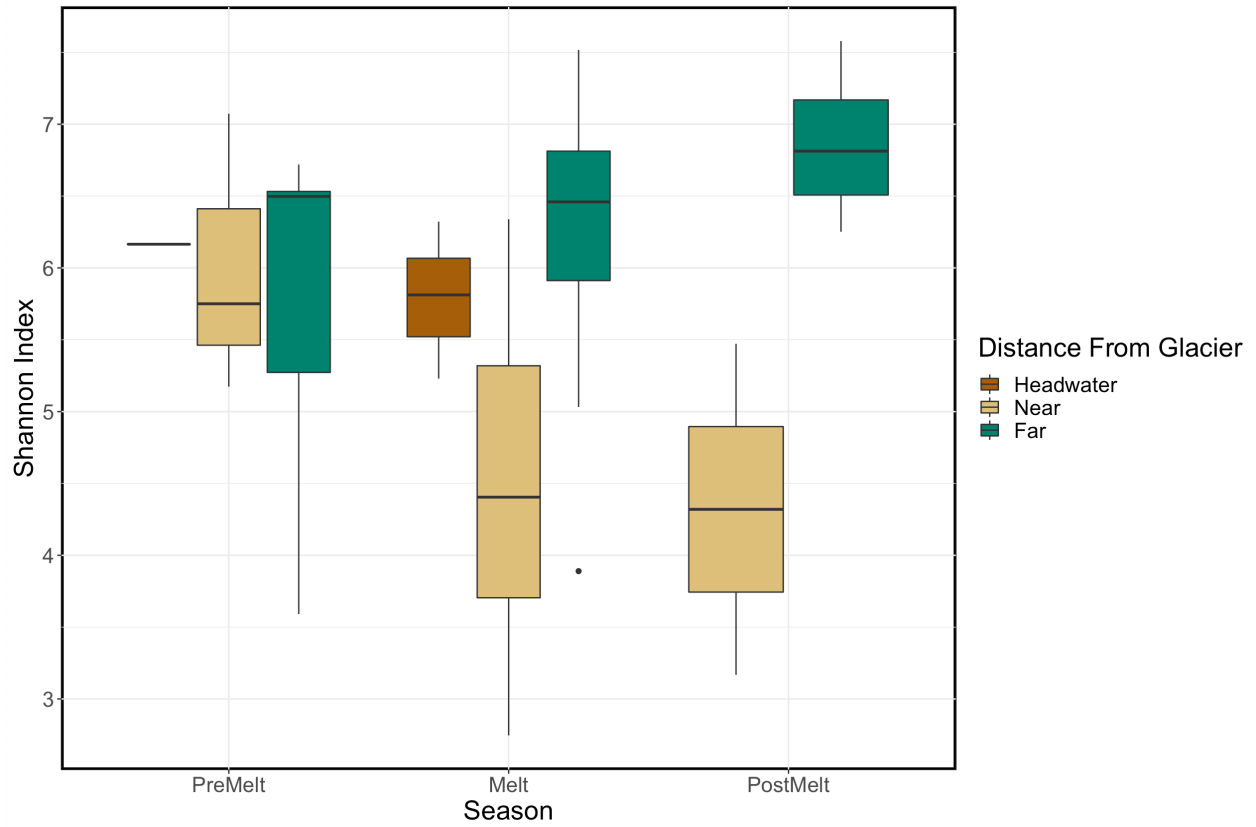


Figure S6: Boxplots showing variation in alpha diversity across hydrological periods and distance range, using the Shannon diversity index. The boxes represent the interquartile range, the black line represents the median value, and the points represent outliers.

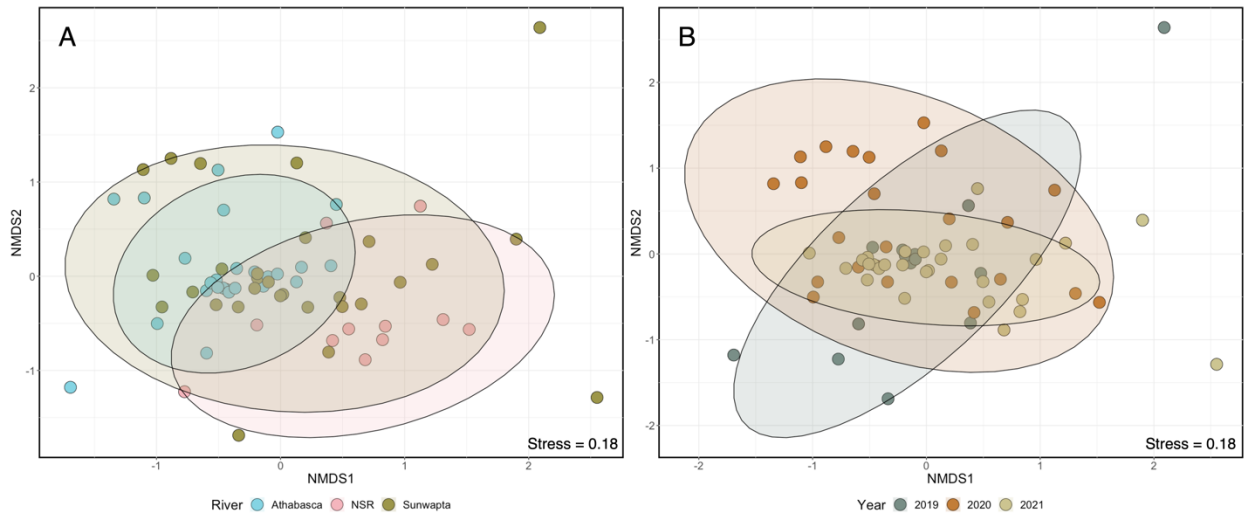


Figure S7: NMDS of microbial assemblage composition based on Bray Curtis distances of Hellinger transformed ASV data. Colour represents different (A) study rivers and (B) sampling years. Shaded circles represent the 95% confidence interval for significantly different groupings (pairwise perMANOVA $p < 0.01$ (Holm's adjusted) see Table S7).

References:

- Bhatia, M. P., Das, S. B., Xu, L., Charette, M. A., Wadham, J. L., & Kujawinski, E. B. (2013). Organic carbon export from the Greenland ice sheet. *Geochimica et Cosmochimica Acta*, *109*, 329–344. <https://doi.org/10.1016/j.gca.2013.02.006>
- Blaser, M., & Conrad, R. (2016). Stable carbon isotope fractionation as tracer of carbon cycling in anoxic soil ecosystems. *Current Opinion in Biotechnology*, *41*, 122–129. <https://doi.org/10.1016/j.copbio.2016.07.001>
- Chanton, J. P., & Lewis, F. G. (1999). Plankton and dissolved inorganic carbon isotopic composition in a river-dominated estuary: Apalachicola Bay, Florida. *Estuaries*, *22*(3), 575–583. <https://doi.org/10.2307/1353045>
- Gabor, R. S., Baker, A., McKnight, D. M., & Miller, M. P. (2014). Fluorescence Indices and Their Interpretation. In A. Baker, D. M. Reynolds, J. Lead, P. G. Coble, & R. G. M. Spencer (Eds.), *Aquatic Organic Matter Fluorescence* (pp. 303–338). Cambridge University Press. <https://doi.org/10.1017/CBO9781139045452.015>
- Fellman, J. B., Hood, E., & Spencer, R. G. M. (2010). Fluorescence spectroscopy opens new windows into dissolved organic matter dynamics in freshwater ecosystems: A review. *Limnology and Oceanography*, *55*(6), 2452–2462. <https://doi.org/10.4319/lo.2010.55.6.2452>
- Hecky, R. E., & Hesslein, R. H. (1995). Contributions of Benthic Algae to Lake Food Webs as Revealed by Stable Isotope Analysis. *Journal of the North American Benthological Society*, *14*(4), 631–653. <https://doi.org/10.2307/1467546>
- Masiello, C. A., & Druffel, E. R. M. (2003). Organic and black carbon ¹³C and ¹⁴C through the Santa Monica Basin sediment oxic-anoxic transition. *Geophysical Research Letters*, *30*(4). <https://doi.org/10.1029/2002GL015050>
- McDonough, L. K., Andersen, M. S., Behnke, M. I., Rutledge, H., Oudone, P., Meredith, K., O'Carroll, D. M., Santos, I. R., Marjo, C. E., Spencer, R. G. M., McKenna, A. M., & Baker, A. (2022). A new conceptual framework for the transformation of groundwater dissolved organic matter. *Nature Communications*, *13*(1), Article 1. <https://doi.org/10.1038/s41467-022-29711-9>
- Mladenov, N., Reche, I., Olmo, F. J., Lyamani, H., & Alados-Arboledas, L. (2010). Relationships between spectroscopic properties of high-altitude organic aerosols and Sun photometry from ground-based remote sensing. *Journal of Geophysical Research: Biogeosciences*, *115*(G1). <https://doi.org/10.1029/2009JG000991>
- Peterson, B. J., & Fry, B. (1987). Stable Isotopes in Ecosystem Studies. *Annual Review of Ecology and Systematics*, *18*(1), 293–320. <https://doi.org/10.1146/annurev.es.18.110187.001453>
- Rau, G. H., & Hedges, J. I. (1979). Carbon-13 depletion in a hydrothermal vent mussel: Suggestion of a chemosynthetic food source. *Science (New York, N.Y.)*, *203*(4381), 648–649. <https://doi.org/10.1126/science.203.4381.648>
- Ruby, E. G., Jannasch, H. W., & Deuser, W. G. (1987). Fractionation of Stable Carbon Isotopes during Chemoautotrophic Growth of Sulfur-Oxidizing Bacteria. *Applied and Environmental Microbiology*, *53*(8), 1940–1943.
- Serbu, J. A., St. Louis, V. L., Emmerton, C. A., Tank, S. E., Criscitiello, A. S., Silins, U., et al. (2024). A comprehensive biogeochemical assessment of climate-threatened glacial river headwaters on the eastern slopes of the Canadian Rocky Mountains. *Journal of Geophysical Research: Biogeosciences*, *129*, e2023JG007745. <https://doi.org/10.1029/2023JG007745>
- Standard Methods Committee of the American Public Health Association, American Water Works Association, & Water Environment Federation. (2018a). Method 3125: Metals by Inductively

Coupled Plasma-Mass Spectrometry (ICP-MS). In W. C. Lipps, T. E. Baxter, & E. Braun-Howland (Eds.), *Standard Methods for the Examination of Water and Wastewater*. APHA Press. <https://doi.org/10.2105/smww.2882.048>

Shi, Z., Allison, S. D., He, Y., Levine, P. A., Hoyt, A. M., Beem-Miller, J., Zhu, Q., Wieder, W. R., Trumbore, S., & Randerson, J. T. (2020). The age distribution of global soil carbon inferred from radiocarbon measurements. *Nature Geoscience*, 13(8), Article 8. <https://doi.org/10.1038/s41561-020-0596-z>

Standard Methods Committee of the American Public Health Association, American Water Works Association, & Water Environment Federation. (2018b). Method 4500-NH₃ (Ammonia). In W. C. Lipps, T. E. Baxter, & E. Braun-Howland (Eds.), *Standard Methods for the Examination of Water and Wastewater*. APHA Press. <https://doi.org/10.2105/SMWW.2882.087>

Standard Methods Committee of the American Public Health Association, American Water Works Association, & Water Environment Federation. (2018c). Method 4500-P (Phosphorus). In W. C. Lipps, T. E. Baxter, & E. Braun-Howland (Eds.), *Standard Methods for the Examination of Water and Wastewater*. APHA Press.

Standard Methods Committee of the American Public Health Association, American Water Works Association, & Water Environment Federation. (2018d). Method 4500-SiO₂ (Silica). In W. C. Lipps, T. E. Baxter, & E. Braun-Howland (Eds.), *Standard Methods for the Examination of Water and Wastewater*. APHA Press. <https://doi.org/10.2105/SMWW.2882.095>

U.S. EPA. (1993a). Method 300.1: Determination of anions in drinking water by ion chromatography. In *Methods for the Determination of Organic and Inorganic Compounds in Drinking Water* (EPA 815/R-00-014). U.S. EPA Federal Registry.

U.S. EPA. (1993b). Method 353.2: Determination of nitrate-nitrite nitrogen by automated colorimetry. In *Methods for Chemical Analysis of Water and Wastes* (EPA 600/4-79-020). U.S. EPA Federal Registry.

U.S. EPA. (1994). Method 200.7, Revision 4.4: Determination of metals and trace elements in water and wastes by Inductively Coupled Plasma-Atomic Emission Spectrometry (ICP-AES). U.S. EPA Federal Registry.