

Biogeochemical factors contributing to enhanced carbon storage following afforestation of a semi-arid shrubland

J. M. Grünzweig¹, I. Gelfand², Y. Fried¹, and D. Yakir²

¹Robert H. Smith Institute of Plant Sciences and Genetics in Agriculture, Faculty of Agricultural, Food and Environmental Quality Sciences, the Hebrew University of Jerusalem, Rehovot 76100, Israel

²Department of Environmental Sciences and Energy Research, Weizmann Institute of Science, Rehovot 76100, Israel

Received: 23 May 2007 – Published in Biogeosciences Discuss.: 2 July 2007

Revised: 17 September 2007 – Accepted: 11 October 2007 – Published: 24 October 2007

Abstract. Ecosystems in dry regions are generally low in productivity and carbon (C) storage. We report, however, large increases in C sequestration following afforestation of a semi-arid shrubland with *Pinus halepensis* trees. Using C and nitrogen (N) inventories, based in part on site-specific allometric equations, we measured an increase in the standing ecosystem C stock from 2380 g C m⁻² in the shrubland to 5840 g C m⁻² in the forest after 35 years, with no significant change in N stocks. Carbon sequestration following afforestation was associated with increased N use efficiency as reflected by an overall increase in C/N ratio from 7.6 in the shrubland to 16.6 in the forest. The C accumulation rate in the forest was particularly high for soil organic C (SOC; increase of 1760 g C m⁻² or 50 g C m⁻² yr⁻¹), which was associated with the following factors: 1) Analysis of a small ¹³C signal within this pure C₃ system combined with size fractionation of soil organic matter indicated a significant addition of new SOC derived from forest vegetation (68% of total forest SOC) and a considerable portion of the old original shrubland SOC (53%) still remaining in the forest. 2) A large part of both new and old SOC appeared to be protected from decomposition as about 60% of SOC under both land-use types were in mineral-associated fractions. 3) A short-term decomposition study indicated decreased decomposition of lower-quality litter and SOC in the forest, based on reduced decay rates of up to 90% for forest compared to shrubland litter. 4) Forest soil included a significant component of live and dead roots (12% of total SOC). Our results suggest a role for increased N use efficiency, enhanced SOC protection and reduced decomposition rates in the large C sequestration potential following afforestation in semi-arid regions. These results are particularly relevant in light of persistent predictions of drying trends in the Mediterranean and other regions.

1 Introduction

The Kyoto Protocol of the United Nations Framework Convention on Climate Change encourages management of terrestrial ecosystems that leads to increased carbon (C) sequestration. One of the ways to achieve this is through afforestation, which was discussed as an instrument for reducing global CO₂ emissions and stabilizing the climate (Jackson and Schlesinger, 2004; Pacala and Socolow, 2004). Afforestation is defined as establishment of forests on lands which historically have not contained forests (Houghton et al., 1996) or, alternatively, as lands which have been without forest for a period of several decades and have previously been under a different land use (Watson et al., 2000). In addition to C storage, afforestation can be a tool for restoring degraded dryland soils and ecosystems (Lal, 2004).

Afforestation often results in increased ecosystem-scale C stocks, mainly as a consequence of the build-up of above-ground tree biomass (Thuille and Schulze, 2006). Consequences of forest establishment for soil C are more complex, and depend on the former land use, on forest age and on the environmental conditions. Afforestation of former cropland resulted in a mean increase of 18% in soil organic C (SOC) stocks across various climatic conditions and forest ages, while planting forests on former native forest or grassland resulted in a mean decrease of 10–13% in SOC stocks (Guo and Gifford, 2002). Those losses of SOC were smaller or no longer present at a later stage of forest development (30 to 40 years after afforestation) and below ~1200 mm mean annual precipitation (MAP) (Guo and Gifford, 2002; Paul et al., 2002). Woody encroachment, the invasion of woody vegetation into deserts and grasslands, resulted in reduction of SOC at moist sites with 660–1070 mm MAP, but to increased SOC stocks at dry sites below ~350 mm MAP (Jackson et al., 2002; Smith and Johnson, 2003). Therefore, dry areas appear to have a potential for net C sequestration in soils following a few decades of establishment of woody vegetation.

Correspondence to: J. M. Grünzweig
(jose@agri.huji.ac.il)

For example, the SOC sequestration potential of leguminous trees in subtropical semi-arid regions of the globe was estimated at 6.2 Pg (Geesing et al., 2000).

A first estimate of C stocks in a semi-arid plantation of Aleppo pine (*Pinus halepensis* Mill.) in Israel showed that both ecosystem and SOC storage were increased by afforestation of former shrubland (Grünzweig et al., 2003). However, the biogeochemical mechanisms enabling increased C storage in this and similar forests remained unclear. The main processes that control C storage following afforestation are accumulation of C in vegetation, C emission by decomposition, and litter incorporation into the soil (Quideau et al., 2001; Farley et al., 2004). These processes are controlled by litter quality, nutrient use efficiency (particularly nitrogen, N) at which C input is used, stabilization of SOC stocks by physical protection, microbial composition and activity, and microclimatic variables, such as soil temperature and moisture (Conant et al., 1998; Post and Kwon, 2000; Chapela et al., 2001; Six et al., 2002; Farley et al., 2004).

The primary objectives of the current study were to evaluate key processes in the C cycle, such as C partitioning, SOC turnover and litter decomposition, and to assess potential biogeochemical mechanisms for increased C storage following afforestation in a hot semi-arid region. A small isotopic signal and physical fractionation of SOC were used to fractionate the forest SOC stock and to calculate shrubland SOC turnover. A litter bag assay and C/N analyses of the main C stocks in forest and shrubland were performed. In addition, the preliminary estimate of C stocks (Grünzweig et al., 2003) was complemented by an N stock and was improved by developing site-specific allometric equations for tree biomass stocks.

2 Materials and methods

2.1 Site details

This study was conducted in and around Yatir Forest (31°20' N, 35°03' E), a pine plantation established at the edge of the Negev Desert between the years 1965 and 1969 (for the purpose of this study, we assumed a mean forest age of 35 years at the time of main soil sampling). The site is located at the transition between sub-humid and arid Mediterranean climatic zones, with mean diurnal (24-h) air temperature and relative humidity of 10°C and 65% in January and 25°C and 50% in July. The semi-arid climate of the northern Negev is characterized by an extended rainless period between May and October or November, and a main rainy season from December to March or April. The forest is dominated by *P. halepensis*, with smaller proportions of other pine species (mostly *Pinus brutia* Ten. and *Pinus pinea* L.), Italian cypress (*Cupressus sempervirens* L.), and little understory vegetation (peak understory aboveground biomass < 100 g m⁻²).

Tree density in 2004 was 300 trees ha⁻¹, mean tree diameter was 17.5 cm and mean tree height was 10 m. The trees were neither irrigated nor fertilized at any time. The surrounding native vegetation, which was the land cover prior to afforestation, consists of sparse shrubland with patches dominated by the dwarf shrub *Sarcopoterium spinosum* (L.) Spach and patches of herbaceous annuals and perennials (maximum vegetation height varied around 30–50 cm). The shrubland has been under moderate to heavy grazing during the last decades, and, like the entire region, was probably under grazing for several millennia. Herbaceous vegetation in the forest was removed mechanically or chemically during the first 12 years after afforestation, and since then is subjected to a moderate to heavy grazing regime (Abed Abualkewan, personal communication). The main soil type in forest and shrubland is light Rendzina (Haploxeroll) above chalk and limestone, with a deep ground-water table (> 100 m). Main plant activity is concentrated during the winter and spring season between December and April.

2.2 Field sampling and methodology

Five 30×30 m plots were chosen in the central part of the forest. Four of those five plots were stocked with *P. halepensis* only, while in the fifth plot, 65% of individuals and 85% of aboveground biomass were of *P. halepensis*, the rest being *C. sempervirens*. Five 30×30 m plots were chosen in the native shrubland around the forest. All shrubland and forest plots were at an altitude of 580–650 m a.s.l.

Standing aboveground tree biomass and biomass of the belowground stem-root transition section in the forest (see below) were estimated from measurements of stem diameter at 1.3 m and tree height using allometric equations (Appendix A, Table A1). For the establishment of allometric equations (see e.g. Scarascia-Mugnozza et al., 2000), 28 *P. halepensis* trees were harvested in 2003. Trees were selected along a curve of tree height vs. stem diameter which was determined in a preliminary inventory of 256 trees. The selected trees spanned a stem diameter range of 3–26 cm, and different age classes were bulked. After felling, the stem was divided into sections of 1–3 m, and the crown was separated into two to three equal compartments. Branches were separated into three different diameter classes, and representative branches were selected for separation into three different subclasses, cones and leaves. Five trees were also assessed for determination of belowground biomass of the stem-root transition section (belowground part of the stem + coarse roots protruding below the stem) excavated to a distance of 50–100 cm from the stem and a depth of about 50 cm. Samples of all components were collected for determination of the dry matter content at 80°C. For determination of aboveground tree biomass of *C. sempervirens*, stem-volume equations developed by Israel's forestry agency were used (KKL-JNF, 1999). Those equations cover the range of tree sizes in Yatir Forest, and tree biomass was estimated

using measured stem-wood density of 900 kg m^{-3} , dry matter content of 60.1%, and a ratio of stem/total biomass of 0.64 derived from the literature for the congeneric species *Cupressus torulosa* D. Don (Rathore et al., 1997). During periodic thinning campaigns, part of the aboveground biomass was harvested by the forestry agency. The amount of harvested biomass was estimated from diameter measurements of the generally well preserved stumps after converting to stem diameter at 1.3 m (KKL-JNF, 1999) and applying a set of allometric equations using only stem diameter (Table A1). Harvested stems $>10.2 \text{ cm}$ diameter were processed by industry into products of medium lifetime, with the remaining harvested wood used for heating (Abed Abualkean, personal communication). Aboveground herbaceous biomass in the forest and shrubland was assumed zero, since grazing removed virtually all herbaceous plant material by the end of the drought period. Herbaceous root biomass and grazed biomass replaced by defecation of domestic animals were included in the belowground C stocks. Shrub biomass in the shrubland was determined by destructive harvest of two subplots of 8-m diameter adjacent to each plot. The biomass of the stem-root transition section of shrubs was calculated by a general relationship with aboveground biomass established by a destructive harvest (data not shown).

For C and N concentrations and stable isotopes, aboveground plant material was collected from five trees or shrubs per plot at the end of the dry period in 2001. Leaves included all living leaf cohorts on a twig, and tree stem properties were measured on stem cores at 1.3 m height. Soil in the forest was sampled from five cores (5 cm diameter) per plot during 2001 and 2002. Soil in each shrubland plot was sampled from five cores under *S. spinosum* shrubs (shrub microsite) and five cores in the interspaces between shrubs (intershrub microsite). The two microsites typically differ in soil chemical and other properties (e.g. Charley and West, 1975). At both sites, the mineral soil was separated into the following layers, according to Harrison et al. (2000): 0–5, 5–10, 10–20 and 20–50 cm. Roots were included in carbon stocks of mineral soils, since it was impossible to manually separate myriads of fine root segments from dry soil particles. However, in 2007, fine roots ($<2 \text{ mm}$ diameter) and coarse roots ($<5 \text{ mm}$; $\sim 15\%$ of root C stock) were sampled from five cores (4.8 cm diameter) per plot, and were washed from soil over a series of sieves. The percentage of roots per total SOC was averaged across three sampling dates in 2007. The shallow litter layer was sampled within a grid of $40 \times 40 \text{ cm}$ in the forest and at the intershrub microsite, and within a grid of $10 \times 10 \text{ cm}$ under shrubs. Litter C and N stocks were presented as part of the soil.

2.3 Laboratory methodology

Soil samples were air-dried, sieved (2 mm) and mixed. Litter was separated from mineral soil, coarsely ground and mixed. All cores in a plot were then combined to one

composite sample per plot and depth. Soil texture was analyzed by the hydrometer method after dispersion with sodium hexametaphosphate (Sheldrick and Wang, 1993). Soil pH was measured by a glass electrode in the supernatant of a 1:10 soil/water suspension. For elemental and isotopic analyses, soil and litter was ground to pass a $250\text{-}\mu\text{m}$ sieve. Concentrations of total C and N were determined in an elemental analyzer (EA 1108, Carlo-Erba, Milan, Italy). SOC was measured by the EA 1108 following removal of carbonates by treating ground soil samples with 1N HCl for 24 h according to Midwood & Boutton (1998). Stable carbon isotope analyses were conducted on CO_2 samples in an isotope-ratio mass spectrometer (Optima, Micromass, Manchester, UK) after quantitative combustion in the EA 1108. Stable C isotope ratios were expressed as $\delta^{13}\text{C} (\text{‰}) = (\text{R}_{\text{sample}}/\text{R}_{\text{standard}} - 1) \times 1000$, where $\text{R} = {}^{13}\text{C}/{}^{12}\text{C}$. The standard was V-PDB, and the precision of the ${}^{13}\text{C}$ analysis was $\pm 0.1\text{‰}$. Inorganic C concentration was determined as the difference between total and organic C, and was expressed as CaCO_3 -equivalent C. Coarse organic matter ($>2 \text{ mm}$) was added to the fine fraction to determine soil organic C concentration and stocks. In plots with shallow soil ($<50 \text{ cm}$ depth), samples were taken to 20 cm only, and C and N stocks were corrected to the maximal depth in the plot (30–40 cm) as determined by extensive probing with a metal rod. Carbon and N stocks were corrected for differences in bulk density by the method of equivalent soil mass (Grünzweig et al., 2004). To calculate total shrubland C and N stocks, stocks obtained for the shrub and intershrub microsites were weighted with the respective percentage ground cover (19% shrub cover, 61% covered by intershrub spaces on soil, the remaining being bare rock). All area-based values of C and N inventories were corrected for ground cover by rocks.

Using a small isotopic signal introduced by afforestation (see Results) in combination with physical fractionation of soil organic matter enabled partitioning of forest SOC into its components, i.e. SOC derived from forest trees vs. SOC remaining from the original shrubland. Organic matter in the top 10 cm of forest and shrubland soil was fractionated into different size classes, generally following methods of Cambardella and Elliott (1992) and Sollins et al. (1999). Briefly, 20 g of dry soil was dispersed in 50 ml of 5% (w/v) sodium hexametaphosphate in a reciprocal shaker for 18 h. The suspension was passed through sieves to generate fractions $>250 \mu\text{m}$ (particulate organic C), $>50 \mu\text{m}$ (intermediate C fraction), and $<50 \mu\text{m}$ (mineral-associated C). The fractions were dried at 80°C and then treated to remove carbonates as described above. The smaller the particle size, the more microbially degraded and recalcitrant the organic matter was assumed to be, which was confirmed by the $\delta^{13}\text{C}$ data (see Results). Fractionation at various densities of sodium polytungstate did not yield a significant separation of SOC pools.

Because the small isotopic signal in this C_3 -all system might limit the SOC analysis, we compared the results of

two isotope mixing models that use different parameters for the computation of the fraction of forest-derived C in forest soil. The common mixing model (Balesdent and Mariotti, 1996; Six and Jastrow, 2002) was slightly modified by substituting $\delta^{13}\text{C}$ of bulk SOC of the native soil with $\delta^{13}\text{C}$ of the mineral-associated SOC fraction in that soil, as follows:

$$C_{f-d}/C_f = (\delta^{13}\text{C}_f - \delta^{13}\text{C}_{s-ma}) / (\delta^{13}\text{C}_{fr} - \delta^{13}\text{C}_{s-ma}), \quad (1)$$

where C_{f-d} is the forest-derived C stock in forest soil, C_f is the total forest SOC stock, $\delta^{13}\text{C}_f$ and $\delta^{13}\text{C}_{fr}$ are the isotopic composition of the total forest SOC stock and of forest roots, respectively, and $\delta^{13}\text{C}_{s-ma}$ is the isotopic composition of the mineral-associated SOC fraction in the shrubland. A second version of the mixing model was modified from the alternative model of Balesdent & Mariotti (1996) as follows:

$$C_{f-d}/C_f = (\delta^{13}\text{C}_f - \delta^{13}\text{C}_{s-po}) / (\delta^{13}\text{C}_{f-po} - \delta^{13}\text{C}_{s-po}), \quad (2)$$

where $\delta^{13}\text{C}_{f-po}$ and $\delta^{13}\text{C}_{s-po}$ are the isotopic composition of the particulate organic C fraction in the forest and the shrubland soil, respectively. In this version of the mass balance, $\delta^{13}\text{C}$ of the vegetation (Balesdent and Mariotti, 1996) was substituted by $\delta^{13}\text{C}$ of assumingly most labile SOC.

Mean residence time (MRT) of shrubland SOC in the forest was calculated as follows (Six and Jastrow, 2002):

$$\text{MRT} = t / \ln(C_s / C_{s-d}), \quad (3)$$

where t is the time since afforestation, C_s is the total SOC stock in the shrubland and C_{s-d} is the shrubland-derived SOC in the forest soil. C_{s-d} was determined by the difference between C_f and C_{f-d} (from Eqs. 1 and 2).

2.4 Litter-decomposition assay

Local and standard litters were placed in litter bags for short-term decomposition in the forest and the shrubland during the wet and dry seasons using standard methodology (Harmon et al., 1999; Grünzweig et al., 2003). Local litter in the forest consisted of needles (C/N ratio=155.8) collected in litter traps during the 2004–2005 season and of roots (C/N ratio=60.0) collected from three plots to a depth of 10 cm at the end of the dry season in 2005. Local litter in the shrubland consisted of standing leaf litter (C/N ratio=39.0) and of roots to a depth of 10 cm (C/N ratio=36.0) collected from three plots under shrubs and in the intershrub microsite at the end of the dry season in 2005. Wheat straw served as standard litter (C/N ratio=102.9), and was obtained from M. Sternberg and Y. Navon (Tel Aviv University, Tel Aviv, Israel). Initial litter dry mass (d.m.) was 3.0 ± 0.2 g (target d.m. \pm planned and weighed deviation from target) in all litter bags. Leaf and standard litter to be placed on the soil surface were weighed into 5×10 cm litter bags made of nylon with a mesh size of 0.5×1 mm facing down and 2×4 mm facing up. Root and standard litter to be placed vertically at 0–10 cm in the soil were weighed into litter bags of the same size and

material as the leaf-litter bags with a mesh size of 0.5×1 mm at both sides. Litter was placed in the field in January 2006 for decomposition of 4 months during the wet season, and in June 2006 for decomposition of 4 months during the dry season. Litter bags that were placed in the field and immediately retrieved served as control.

3 Results

3.1 Soil properties

Carbonate concentration at all sites and depths ranged from 19 to 31%, with the forest being depleted in carbonate compared with the native shrubland in the top 20 cm of the soil profile (Table 1). Soil pH was dominated by the high carbonate content, and was significantly lower under shrubs than in the open interspace between shrubs. Its increase with depth reflected the parallel increase in carbonate content. Clay content ranged from 32 to 53% and increased with depth, with no differences among sites.

3.2 Plant carbon and nitrogen relations

Nitrogen concentrations in plant parts and fresh litter from pine were lower by 10–60% than those from *S. spinosum* (Table 2). Consequently, C/N ratios were wider in pine than in *S. spinosum* tissue, particularly in tree stem and branch wood. The intershrub plant community had N concentrations and C/N ratios that were closer to shrub than to pine tissues. Pine tissues were significantly enriched in $\delta^{13}\text{C}$ by 2–4‰ compared with plant parts of *S. spinosum* and by 4–6‰ compared with the intershrub community. $\delta^{13}\text{C}$ in pine did not change from living leaves to leaf litter, and was higher by about 1‰ in needles compared with roots.

3.3 Soil carbon and nitrogen concentrations and $\delta^{13}\text{C}$

Concentrations of organic C were higher in the mineral soil of the forest and the shrub microsite in the shrubland compared with intershrub soil (Fig. 1, Table 3). The difference between forest and intershrub soil extended over the entire soil profile, while forest soil was not significantly different from soil under shrubs. Total N concentration was higher under shrubs than in intershrub soil, with no difference between forest soil and either of the shrubland microsities. Both SOC and N concentrations decreased typically with depth at all sites and microsities. The forest had a significantly wider C/N ratio than the shrubland microsities for both the litter layer (57 vs. 29–31) and the mineral soil (10.7–11.9 vs. 6.0–8.6). SOC concentration did not correlate with either pH or clay content (data not shown).

Forest soil was significantly enriched in ^{13}C compared with both shrubland microsities for the mineral soil (Fig. 2, Table 3) and the litter layer (Fig. 2). Higher forest soil $\delta^{13}\text{C}$ reflected the difference in $\delta^{13}\text{C}$ between forest and shrubland

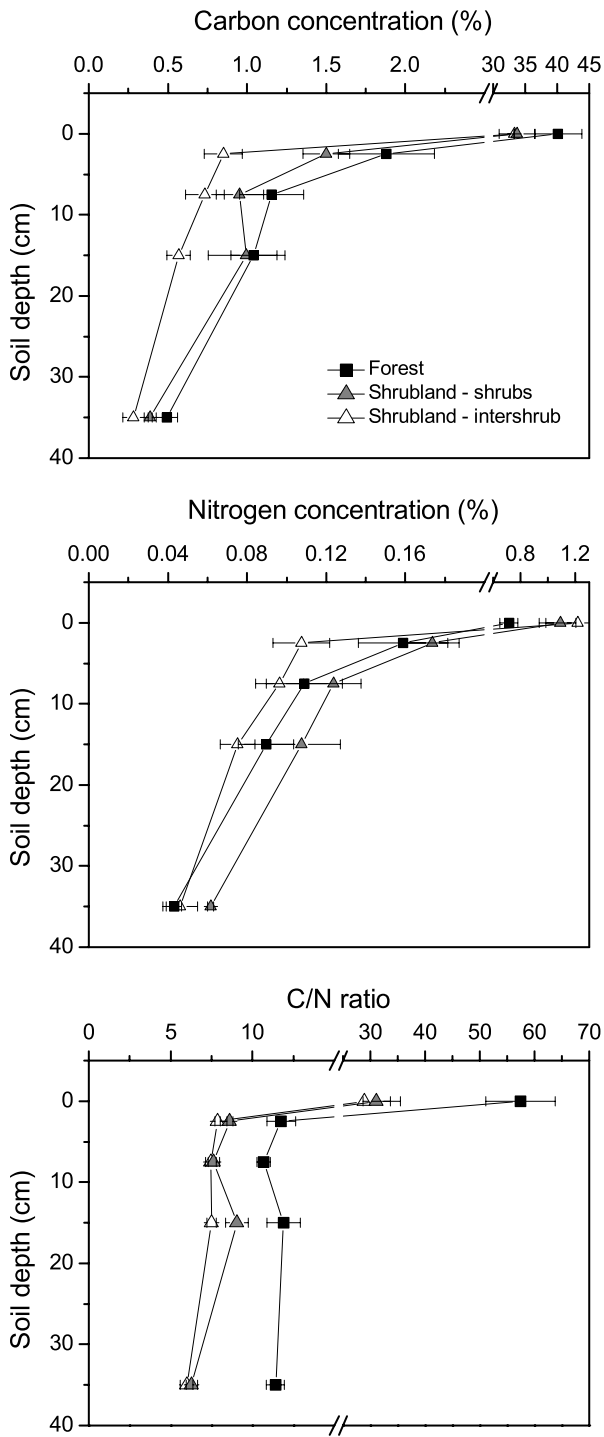


Fig. 1. Soil C and N concentrations and C/N ratio in the forest and the shrubland microsites. The litter layer was set at 0 cm soil depth. Mean \pm s.e., $n=3-5$ plots.

vegetation and fresh litter (Table 2). Forest soil at 0–20 cm had 0.5–0.7‰ and 0.9–1.3‰ higher $\delta^{13}\text{C}$ than intershrub and shrub soils, respectively. $\delta^{13}\text{C}$ increased with increasing depth in the mineral soil at all sites and microsites.

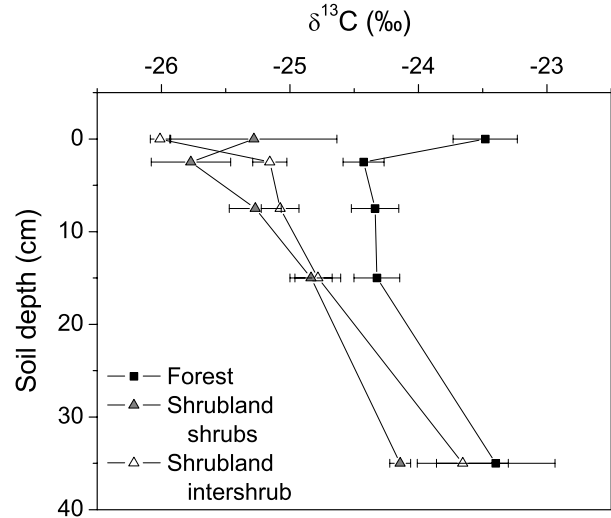


Fig. 2. $\delta^{13}\text{C}$ in SOC in the forest and the shrubland microsites. The litter layer was set at 0 cm soil depth. Mean \pm s.e., $n=3-5$ plots.

3.4 Carbon and nitrogen stocks

Organic C stocks were significantly larger in the forest than in the shrubland in part of the tested soil layers and in the soil profile as a whole (Table 4). Afforestation resulted in an increase of 1760 g C m^{-2} or 75% in the SOC stock (including litter layer and roots, but not the stem-root transition section) after 35 years. This increase in SOC translated into a mean annual SOC accumulation rate of $50 \text{ g C m}^{-2} \text{ yr}^{-1}$. A large part of SOC under both land-use types was concentrated at 0–20 cm in the mineral soil (58–64%), and only a small fraction was found in the litter layer (1–3%). The live and total C stocks of mainly fine roots were also determined separately from SOC for the top 20 cm of the mineral soil and contributed $10 \pm 2\%$ and $12 \pm 2\%$ (mean \pm s.e., $n=5$), respectively, to the total forests SOC stock.

Naturally, the C stock in standing aboveground biomass was much larger in the forest than in the shrubland, and amounted to 27% and 2.5% of their respective total ecosystem C stocks. The total ecosystem organic C stock in the forest was almost 2.5 times the equivalent C stock in the shrubland, an increase of 3460 g C m^{-2} or $99 \text{ g C m}^{-2} \text{ yr}^{-1}$. Forty-three percent of the increase in total ecosystem organic C originated from increased aboveground C stocks and 57% from increased belowground C stocks. Periodic thinning in the forest removed a large amount of total aboveground biomass that was of the same order of magnitude as standing biomass (Table 4). Adding harvested stems >10.2 cm diameter ($770 \pm 236 \text{ g C m}^{-2}$; assumingly processed into products of medium life time) to standing ecosystem C stocks increased the total amount of medium- to long-term C stores generated by afforestation to 4230 g C m^{-2} or $121 \text{ g C m}^{-2} \text{ yr}^{-1}$.

Table 1. Soil properties in forest and shrubland microsites. Clay content and pH were determined over 0–10 cm. Mean (s.e.), $n=2-5$ plots.

Soil depth per site and microsite	Carbonate concentration (% CaCO ₃ -equiv.)	pH	Clay content (%)
Forest			
0–5 cm	19.4 (1.3)	8.34 (0.02)	36.5 (2.4)
5–10 cm	19.2 (1.9)	8.32 (0.05)	40.5 (0.9)
10–20 cm	27.0 (1.5)	8.37 (0.07)	41.7 (1.7)
Shrubland, shrub microsite			
0–5 cm	23.8 (4.2)	8.09 (0.06)	31.7 (7.1)
5–10 cm	22.5 (4.0)	8.30 (0.03)	32.5 (7.5)
10–20 cm	23.1 (3.2)	8.36 (0.05)	52.5 (2.5)
Shrubland, intershrub microsite			
0–5 cm	24.9 (2.1)	8.44 (0.02)	35.0 (2.9)
5–10 cm	25.3 (2.3)	8.39 (0.04)	38.0 (3.7)
10–20 cm	27.2 (3.0)	8.57 (0.07)	50.3 (2.3)
ANOVA			
Site/microsite	0.019	<0.001	0.755
Depth	0.103	0.007	<0.001
Site/microsite × depth	0.991	0.054	0.230

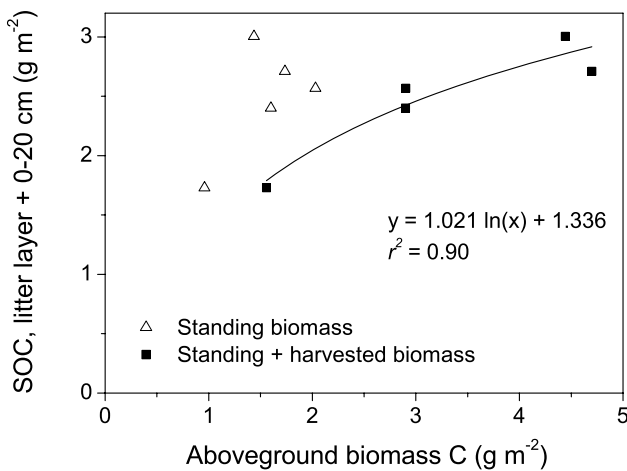


Fig. 3. Effect of harvested biomass on the relationship between aboveground tree biomass and SOC in five forest plots. Regression analysis has been presented where statistically significant.

Variation in standing aboveground biomass among forest plots did not correlate with the SOC stocks in those plots, although C addition to the forest soil should originate from tree biomass (Fig. 3). However, adding harvested to standing biomass in each plot resulted in a significant loga-

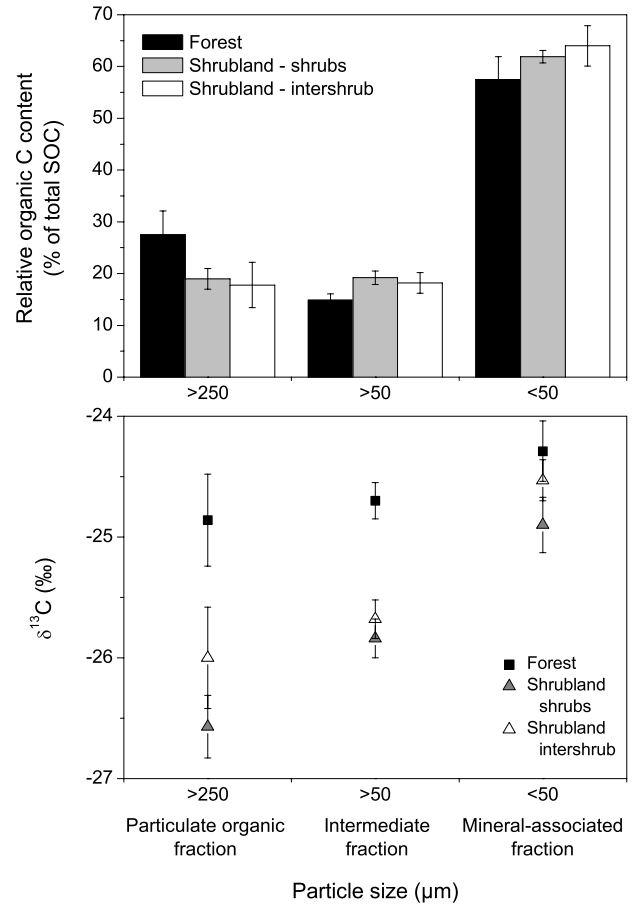


Fig. 4. Relative content and ¹³C composition of SOC fractions in the top 10 cm of forest and shrubland soil. The two-way ANOVA for relative content of SOC fractions for site/microsite, particle size and their interaction (top panel) resulted in respective *P* values of 0.99, <0.001 and 0.12. The two-way ANOVA for ¹³C for site/microsite, particle size and their interaction (bottom panel) resulted in respective *P* values of <0.001, <0.001 and 0.31.

arithmic relationship between aboveground C and SOC to a depth of 20 cm (Fig. 3), and a linear relationship between aboveground C and SOC in the 0–10 cm layer ($r^2=0.93$, $P=0.008$). This shows that the legacy of harvested biomass is apparent in forest SOC.

Total N stocks in the various soil layers did not differ significantly between forest and shrubland, except for more N in the forest litter layer than in the equivalent layer in the shrubland (Table 4). The aboveground forest N stock was larger than the equivalent shrubland N stock, mainly as a consequence of relatively large amounts of N in tree branches and leaves. The ecosystem N stock in the forest was 12% larger than the shrubland N stock, a mean annual increase rate of $1.06 \text{ g N m}^{-2} \text{ yr}^{-1}$, but this difference was not statistically significant ($P=0.26$). N stored in aboveground structures contributed only a minor part of total ecosystem N stocks (2.5% in the forest, 0.3% in the shrubland). Therefore,

Table 2. N concentration, C/N ratio and C isotopic composition in plant organs of the main forest and shrubland taxa. Mean (s.e.), $n=3-5$ plots.

Plant organ	N conc. (%)			C/N ratio			$\delta^{13}\text{C}$ (‰)		
	Forest <i>P. halepensis</i>	Shrubland <i>S. spinosum</i>	Intershrub community	Forest <i>P. halepensis</i>	Shrubland <i>S. spinosum</i>	Intershrub community	Forest <i>P. halepensis</i>	Shrubland <i>S. spinosum</i>	Intershrub community
Leaves	0.96 (0.05)	1.81 (0.13)	1.52 (0.04) ^z	53 (3)	22.2 (1.6)	33.8 (0.8) ^z	-22.9 (0.2)	-26.8 (0.5)	-28.6 (0.1) ^z
Branches	0.29 (0.02)	0.73 (0.03)		169 (12)	64.8 (2.6)		-23.3 (0.1)	-25.5 (0.3)	
Stem	0.07 (0.003)			682 (30)			-22.3 (0.1)		
Fine roots	1.09 (0.06)	n.a.	1.34 ^y	34.3 (1.6)	n.a.	29.5 ^y	-24.3 (0.1)	n.a.	-28.1 ^y
Fresh leaf litter	0.49 (0.03)	0.63 (0.02)	1.59 (0.27)	102 (6)	63.5 (1.5)	26.8 (4.8)	-23.0 (0.2)	-26.3 (0.1)	-28.3 (0.4)

^z Data from (Grünzweig and Körner, 2001)^y Data from herbaceous roots in one forest plot**Table 3.** Analysis of variance and multiple comparisons of C and N concentrations in the mineral soil of the forest and the shrubland microsites.

Variable	Source of variation	df	F ratio	P value	Site/microsite differences (Tukey HSD) ^z F S I
C	Site/microsite	2	10.66	<0.001	a a b
	Soil depth	3	14.28	<0.001	
	Site/microsite × soil depth	6	1.196	0.330	
N	Site/microsite	2	4.503	0.018	ab a b
	Soil depth	3	16.46	<0.001	
	Site/microsite × soil depth	6	0.742	0.619	
C/N	Site/microsite	2	60.63	<0.001	a b b
	Soil depth	3	3.927	0.016	
	Site/microsite × soil depth	6	0.911	0.498	
$\delta^{13}\text{C}$	Site/microsite	2	14.30	<0.001	a b b
	Soil depth	3	17.73	<0.001	
	Site/microsite × soil depth	6	0.774	0.596	

^z Tukey's honestly significant difference test for *posthoc* multiple comparisons; F = forest, S = shrubland under shrubs, I = shrubland intershrub soil

harvesting aboveground biomass during thinning campaigns removed only a small amount of N (Table 4).

3.5 Soil organic carbon fractions and dynamics

The major part (58–64%) of the total SOC content in the top 10 cm of the soil profile was in the mineral-associated fraction (<50 μm), with no significant difference between sites and microsites (Fig. 4, top panel). Particulate organic C per total SOC was slightly but statistically not significantly higher in the forest (28%) than in the shrubland (18–19%). Because of a higher total SOC stock, the absolute values of all SOC pools were higher in the forest than in the shrubland (data not shown).

SOC fractions in the forest were significantly enriched in ^{13}C compared with the shrubland microsites ($P < 0.001$; Fig. 4, bottom panel). $\delta^{13}\text{C}$ ranged from -24.9 to -24.3‰ in forest soil and from -26.6 to -24.9‰ in soil under shrubs. The ^{13}C signature increased with decreasing particle size for all sites and microsites.

Applying two different mass balances to total SOC and its fractions in the forest topsoil allowed estimating the portion of SOC that originated from afforestation as compared with SOC remaining from the native shrubland. Thirty-five years after afforestation, 68% on average of organic C in forest soil was derived from forest vegetation (Table 5). Assuming that the SOC stock prior to afforestation was equivalent to current

Table 4. Organic C and total N stocks in forest and shrubland. To calculate total shrubland C and N stocks, stocks obtained for the shrub and intershrub microsites were weighted with the respective percentage ground cover. Mean (s.e.), $n=5$ plots. *, **, *** statistically significant differences between forest and shrubland at $P \leq 0.05, 0.01, 0.001$, respectively.

Compartment	Organic carbon stocks (g m^{-2})		Total nitrogen stocks (g m^{-2})	
	Forest	Shrubland	Forest	Shrubland
1. Standing stock				
<i>Aboveground</i>				
Stem	805 (80)		1.19 (0.07)	
Branches ^y	565 (83)		3.37 (0.44)	
Foliage	158 (13)		3.00 (0.23)	
Total	1553 (178)***	60 (20)	7.55 (0.73)***	0.93 (0.33)
<i>Belowground</i>				
Stem-root transition section	215 (16)***	7.6 (2.5)	0.32 (0.01)**	0.12 (0.04)
<i>Soil^y</i>				
Litter layer	125 (21)***	23 (6)	2.5 (0.4)**	0.9 (0.3)
0–5 cm	825 (110)*	450 (58)	70.7 (6.2)	54.3 (6.2)
5–10 cm	540 (66)	391 (66)	50.7 (6.1)	49.2 (5.9)
10–20 cm	993 (106)	648 (118)	86.1 (9.0)	76.5 (8.4)
20–50 cm	1589 (218)*	803 (220)	132.8 (14.0)	131.7 (34.6)
Total soil	4072 (353)**	2316 (183)	342.8 (13.3)	312.6 (28.5)
<i>Ecosystem</i>	5839 (437)***	2383 (159)	350.7 (12.5)	313.7 (25.4)
2. Harvested tree stock	1676 (532)	n.a.	8.43 (2.66)	n.a.

^y Branches include twigs and cones, mineral soil includes live and dead root stocks.

Table 5. SOC dynamics for the 0–10 cm layer in the forest soil. Mean residence time (MRT; applying Eq. 3 to the outcome of Eqs. 1 and 2) relates to the original shrubland SOC in forest soil.

Mass balance	Forest-derived SOC in forest soil		Shrubland-derived SOC in forest soil			
	Stock (g C m^{-2})	Fraction of total forest SOC (%)	Stock (g C m^{-2})	Fraction of total forest SOC (%)	Fraction of original shrubland SOC (%)	MRT of original shrubland SOC (yr)
Eq. (1)	980	72	385	28	46	45
Eq. (2)	864	63	501	37	59	67
Mean	922	68	443	32	53	56

shrubland SOC indicated that 53% of the original native SOC was still present in the forest. Mean MRT of the native shrubland SOC in the forest soil was 56 years as calculated by a first-order decay model (Table 5).

The C inventory and the SOC partitioning enabled estimating total C produced by the forest as 6250 g C m^{-2} or on av-

erage $179 \text{ g C m}^{-2} \text{ yr}^{-1}$. This approximation of “gross forest production” used total harvested biomass and assumed that all C stocks in the forest were newly forest-derived C, except for 32% of the mineral soil which was comprised of native shrubland SOC (obtained for the 0–10 cm layer and applied to a depth of 50 cm; Table 5).

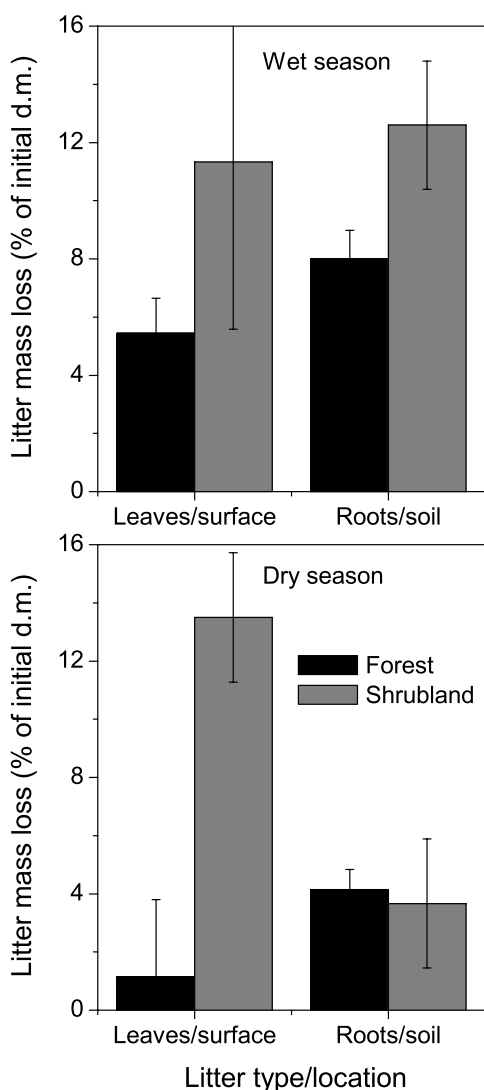


Fig. 5. Mass loss of local litter at forest and shrubland sites during the wet and the dry seasons. Since at both shrubland microsites, a common mixture of shrub + herbaceous leaf litter and a common mixture of root litter were used, litter mass loss in the shrubland was presented across both microsites (after weighting for the relative spatial cover of each microsites). Mean \pm s.e., $n=3-5$ plots.

3.6 Litter decomposition

Litter-bag assays with local litter revealed significantly lower decomposition rates of forest compared with shrubland litter across seasons and litter type (Fig. 5, Table 6). Short-term leaf- and root-litter mass loss was 37–52% lower in the forest than in the shrubland during the wet season. During the dry season, leaf-litter decomposition was surprisingly high in the shrubland, even higher than litter decomposition during the wet season. Decay rate of needles in the forest during the dry season was low, 92% lower than decay rate of leaf litter in the shrubland.

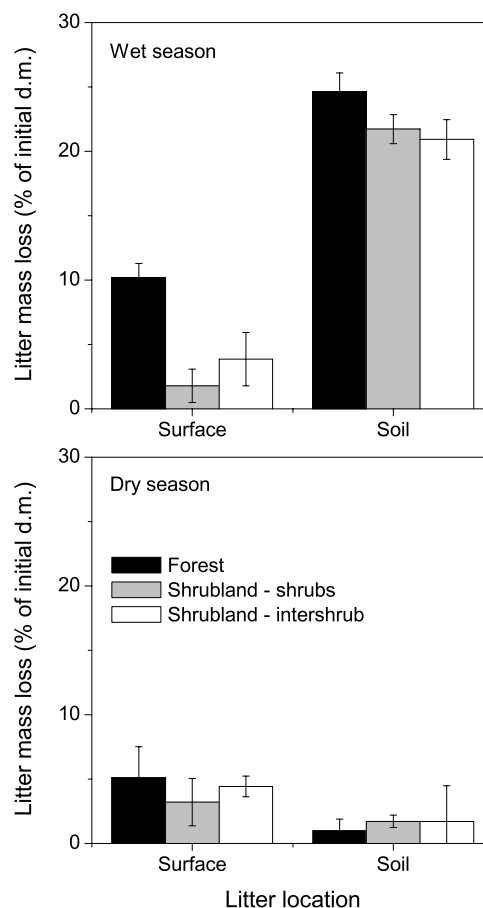


Fig. 6. Mass loss of standard litter at forest and shrubland microsites during the wet and the dry seasons. Mean \pm s.e., $n=4-5$ plots.

Standard litter decomposition was significantly higher in the forest than in the two shrubland microsites during the wet season, particularly at the soil surface (Fig. 6, Table 6). Mass loss at the soil surface (location of leaf litter) was 3.6 times greater in the forest than in the two microsites of the shrubland. Mass loss of standard litter in the soil (location of root litter) was greater than at the surface, and was 15% greater in the forest than in the shrubland. Standard-litter decomposition was low in the dry season, with no significant differences between sites and between litter locations.

4 Discussion

Afforestation of sparse shrubland in this hot semi-arid region increased standing ecosystem C stocks by 3460 g C m^{-2} or on average $99 \text{ g C m}^{-2} \text{ yr}^{-1}$ over 35 years. This integrative value for decades of C sequestration takes biomass harvest into account, and can be considered, therefore, equivalent to long-term net biome production (Schulze et al., 2000). The long-term C stocks including C stored in wood products (Houghton, 1995) amounted to 4230 g C m^{-2} or

Table 6. Analysis of variance of the litter-bag assays in the forest and the shrubland during the wet and dry seasons. Mass loss of local litter in the shrubland was combined for both microsites, mass loss for standard litter was analyzed separately for the microsites (see Figs. 5 and 6). Standard litter was analyzed with two-way ANOVA after interactions among variables in the three-way analysis were statistically significant.

Litter source	Time period	Source of variation	df	F ratio	P value
Local litter	Both seasons	Season	1	3.37	0.077
		Litter type/location	1	0.14	0.713
		Site	1	7.57	0.010
		Season x litter type/location	1	1.71	0.201
		Season x site	1	0.03	0.865
		Site x litter type/location	1	3.02	0.093
		Season x site x litter type/location	1	2.01	0.167
Standard litter	Wet season	Litter location	2	9.66	< 0.001
		Site/microsite	1	216.5	< 0.001
		Litter location x site/microsite	2	1.91	0.172
	Dry season	Litter location	2	4.20	0.053
		Site/microsite	1	0.09	0.915
		Litter location x site/microsite	2	0.33	0.725

on average $121 \text{ g C m}^{-2} \text{ yr}^{-1}$, while “gross forest production”, i.e. total C stocks generated by the forest, was estimated at 6250 g C m^{-2} or on average $179 \text{ g C m}^{-2} \text{ yr}^{-1}$.

The rate of C gain by the forest was lower than many records of annual increase in ecosystem C stocks following afforestation in temperate regions (Hooker and Compton, 2003; Paul et al., 2003), mainly because of lower aboveground C stocks. However, tree biomass is a transient and periodically fluctuating C store, and C needs to be allocated to the soil to enable greater continuity of storage.

Soil organic C sequestration of 1760 g C m^{-2} or $50 \text{ g C m}^{-2} \text{ yr}^{-1}$ was remarkably high compared with most forests established by afforestation or abandonment of agricultural land in temperate regions (Schlesinger, 1990; Post and Kwon, 2000; Thuille et al., 2000; Vesterdal et al., 2002; Hooker and Compton, 2003; Paul et al., 2003) or by woody encroachment of dry grassland (Jackson et al., 2002). Dry shrubland can accumulate large amounts of SOC following afforestation with pine, such as planting *Pinus coulteri* B. Don in large lysimeters filled with shrubland soil ($49 \text{ g C m}^{-2} \text{ yr}^{-1}$; Ulery et al., 1995), or planting *Pinus ponderosa* (Dougl.) Laws in a cold semi-arid shrubland ($107 \text{ g C m}^{-2} \text{ yr}^{-1}$; Noretto et al., 2006). However, when pine replaced dense productive vegetation, such as in Mediterranean macchia shrubland, SOC stocks often decreased following afforestation with *P. halepensis* (Maestre and Cortina, 2004; Goberna et al., 2007). Carbon sequestration following afforestation or woody encroachment was mainly, or sometimes entirely, contributed by aboveground biomass increase in temperate regions (Hooker

and Compton, 2003; Jackson et al., 2002; Paul et al., 2003; Thuille and Schulze, 2006). However, in semi-arid regions, belowground C accumulation contributed 50% or more to the ecosystem C gain in the forest or woodland (Jackson et al., 2002; Noretto et al., 2006; the current study).

Various mechanisms for increased SOC stocks following afforestation have been discussed. Root biomass accumulation was an important factor in C sequestration following afforestation and shrub expansion (Connin et al., 1997; Usiri et al., 2006), and the sole factor for SOC accumulation in a *P. ponderosa* plantation under a cold semi-arid climate (Noretto et al., 2006). In the present study, root biomass was included in the measure of SOC because of the impracticality of separating myriads of fine root segments from soil organic matter. Figure 3 indicates that harvested trees leave their legacy in the soil, which suggests a significant contribution of roots to SOC over the period of forest development. The live plus dead root C stock contributed 12% to total forest SOC, a value that increased to 34% when comparing root C to the added SOC by the forest relative to shrubland SOC. It appears, therefore, that roots contribute significantly to the SOC stock and to the increase in SOC by afforestation.

Enhanced C sequestration in N-limited systems requires either increased net N input to the ecosystem or increased N use efficiency. The latter can be achieved by an increase in the C/N ratio and/or by redistribution of N from soil with a narrow C/N ratio to vegetation which normally has a wider C/N ratio (Shaver et al., 1992; Halliday et al., 2003). Despite considerable C sequestration, accumulation of N was low and statistically not significant, suggesting no or negligible

net ecosystem N input to the forest. On the other hand, the C/N ratio was considerably wider in *P. halepensis* trees than in shrubland vegetation, which translated into a wider C/N ratio also in forest soil organic matter. In addition, redistribution of an albeit small amount of N from soil to trees was obvious from a significantly larger N stock in forest biomass and litter as compared with the shrubland. It seems, therefore, that C accumulation in this semi-arid forest was associated with increased N use efficiency, via widening the C/N ratio of soil organic matter, a buildup of biomass with a wide C/N ratio, such as woody biomass, and redistributing some N from soil to trees. Similarly, C accumulation with no net N input in pine forests following afforestation or agricultural abandonment was attributed to a wider C/N ratio and/or a transfer of N from soil to tree biomass (Hooker and Compton, 2003; Ussiri et al., 2006).

To investigate organic C dynamics in forest and shrubland soils and to partition SOC, we combined size fractionation with the analysis of a small isotopic signal introduced by relatively ^{13}C -rich pine roots. Such SOC partitioning was commonly performed using the much larger ^{13}C signal of a conversion of C_3 to C_4 vegetation or *vice versa*, and entering $\delta^{13}\text{C}$ of soils and vegetation into an isotope mixing model (Balesdent and Mariotti, 1996; Six and Jastrow, 2002). Despite the small isotopic signal in our C_3 -only system, the two mixing models (Eqs. 1 and 2) produced only moderately different values for SOC partitioning and MRT (Table 5). Our approach indicated three potential factors involved in SOC accumulation in the pine forest. First, net C input into forest soil seemed to be large, with new forest-derived SOC contributing a major fraction of total SOC (68%). This value was higher than new organic C in agricultural soil afforested with broad-leaved deciduous species in a temperate region after 20 years (43%) (Del Galdo et al., 2003). Second, 53% of the original shrubland SOC stock remained in the forest soil after 35 years. Third, a significant amount of SOC was protected in the mineral-associated fraction ($<50\ \mu\text{m}$; Fig. 4), thus enhancing long-term storage in the soil. Similar protection of both new and old SOC in mineral-associated pools were found following afforestation of agricultural soil (Del Galdo et al., 2003). Protection of old and/or new SOC in microaggregates was reported as an additional factor contributing to long-term SOC storage (Six et al., 2002; Del Galdo et al., 2003), a factor that was not tested in the current study.

The investigation of short-term decomposition indicated an additional mechanism for organic C storage in the forest soil. The experiment with standard litter suggested that the forest provided a better environment for litter decomposition than the shrubland, particularly during the wet season. Reduced runoff following afforestation (Farley et al., 2005; Safriel and Moshe, 2005) and slowed dehydration under the forest canopy (Safriel and Moshe, 2005; I. Gelfand, unpub. res.) apparently provided more moisture to the forest soil, thus enhancing litter decomposition. Despite improved conditions in the forest compared with the shrubland, tree lit-

ter decomposed more slowly than shrubland litter. This was probably caused by lower tissue quality, as indicated by a wider C/N ratio of tree leaf and root litter. Moreover, shrubland litter appeared to be adapted to decomposition in the dry environment. Considering the lack of moisture during the dry season, mass-loss was surprisingly high for local shrubland leaf litter compared with standard litter on the soil surface. This might be explained by an assumingly high susceptibility of local litter to photodegradation by UV radiation, a mechanism active in drought-stressed and high-radiation regions (Austin and Vivanco, 2006). Standard litter seemed to be less prone to degradation by UV radiation, and forest litter was less exposed to short-wave radiation by about 60% under the tree canopy during the dry season (E. Rotenberg, personal communication).

5 Conclusions

In conclusion, afforestation of a hot semi-arid shrubland resulted in significant C sequestration, particularly below-ground, which can have an economic potential in the framework of the Kyoto Protocol, e.g. for poorer countries (Perez et al., 2007). In contrary to the global cooling potential of the CO_2 uptake, it should also be considered that afforestation can result in increased heat uptake as a consequence of a decrease in albedo (Betts, 2000; Bala et al., 2007). Enhanced C sequestration could be attributed to the following factors: (1) increase in N use efficiency reflected by widening of the soil and plant C/N ratio and translocation of N from soil organic matter to tree biomass; (2) production of considerable stocks of live and dead root C; (3) large input of new forest-derived C into the overall forest SOC, and protection of both new and old SOC in mineral-associated fractions; (4) reduced decay rates of tree leaf and root litter, probably as a consequence of reduced litter quality. Compared with C sequestration under a more humid climate, soils in semi-arid regions have a large potential for C storage. This potential could be used by proper management for increased C sequestration in dry regions of the world.

Appendix A

Table A1. Allometric equations for biomass (kg d.m.) of *P. halepensis* trees.

Purpose	Tree part	<i>n</i>	Equation form ^z	<i>a</i>	<i>b</i>	<i>r</i> ²
Standing biomass	Total aboveground biomass	28	$a (d^2 h)^b$	0.030553	1.031064	0.987
	Stem	28	$a (d^2 h)^b$	0.039508	0.918943	0.995
	Branches, twigs, cones	28	$a (d^2 h)^b$	0.001348	1.291356	0.956
	Branches, twigs	28	$a (d^2 h)^b$	0.023916	0.913090	0.975
	Cones	28	$a (d^2 h)^b$	0.001170	1.155894	0.797
	Foliage	28	$a (d^2 h)^b$	0.031062	0.741424	0.941
	Belowground stem-root transition section	5	$a (d^2 h)^b$	0.219842	0.539662	0.997
Harvested biomass	Total aboveground biomass	28	$a d^b$	0.082228	2.540675	0.988
	Stem	28	$(a + b d)^2$	-0.925181	0.505739	0.990
	Branches, twigs, cones	28	$a d^b$	0.005591	3.125069	0.961
	Foliage	28	$(a + b d)^2$	0.333570	0.175186	0.933

^z *a* and *b* are regression coefficients, *d* is stem diameter at 1.3 m height (cm), *h* is tree height (m), *n* is the number of trees used for the generation of the equation.

Acknowledgements. We thank Y. Moshe and the JNF for cooperation and logistics at the field site, D. Hemming for assistance with the allometric equations, and M. Sternberg and Y. Navon for assistance with the litter-decomposition study. We also acknowledge the technical help of E. Negreanu, P. Harel, O. Migdal, N. Kaufmann, N. Zehariah and S. Ben Ari in various parts of this study.

Edited by: F. X. Meixner

References

- Austin, A. T. and Vivanco, L.: Plant litter decomposition in a semi-arid ecosystem controlled by photodegradation, *Nature*, 442, 555–558, 2006.
- Balesdent, J. and Mariotti, A.: Measurement of soil organic matter turnover using ¹³C natural abundance, in: *Mass Spectrometry of Soils*, edited by: Boutton, T. W. and Yamasaki, S.-I., 83–111, Marcel Dekker, New York, 1996.
- Bala, G., Caldeira, K., Wickert, M., Phillips, T. J., Lobell, D. B., Delire, C., and Mirin, A.: Combined climate and carbon-cycle effects of large-scale deforestation, *Proc. Natl. Acad. Sci. U. S. A.*, 104, 6550–6555, 2007.
- Betts, R. A.: Offset of the potential carbon sink from boreal forestation by decreases in surface albedo, *Nature*, 408, 187–190, 2000.
- Cambardella, C. A. and Elliott, E. T.: Particulate soil organic-matter changes across a grassland cultivation sequence, *Soil Sci. Soc. Am. J.*, 56, 777–783, 1992.
- Chapela, I. H., Osher, L. J., Horton, T. R., and Henn, M. R.: Ectomycorrhizal fungi introduced with exotic pine plantations induce soil carbon depletion, *Soil Biol. Biochem.*, 33, 1733–1740, 2001.
- Charley, J. L. and West, N. E.: Plant-induced soil chemical patterns in some shrub-dominated semi-desert ecosystems of Utah, *J. Ecol.*, 63, 945–963, 1975.
- Collins, H. P., Blevins, R. L., Bundy, L. G., Christenson, D. R., Dick, W. A., Huggins, D. R., and Paul, E. A.: Soil carbon dynamics in corn-based agroecosystems: results from carbon-13 natural abundance, *Soil Sci. Soc. Am. J.*, 63, 584–591, 1999.
- Conant, R. T., Klopatek, J. M., Malin, R. C., and Klopatek, C. C.: Carbon pools and fluxes along an environmental gradient in northern Arizona, *Biogeochemistry*, 43, 43–61, 1998.
- Connin, S. L., Virginia, R. A., and Chamberlain, C. P.: Carbon isotopes reveal soil organic matter dynamics following arid land shrub expansion, *Oecologia*, 110, 374–386, 1997.
- Del Galdo, I., Six, J., Peressotti, A., and Cotrufo, M. F.: Assessing the impact of land-use change on soil C sequestration in agricultural soils by means of organic matter fractionation and stable C isotopes, *Glob. Change Biol.*, 9, 1204–1213, 2003.
- Farley, K. A., Jobbágy, E. G., and Jackson, R. B.: Effects of afforestation on water yield: a global synthesis with implications for policy, *Glob. Change Biol.*, 11, 1565–1576, 2005.
- Farley, K. A., Kelly, E. F., and Hofstede, R. G. M.: Soil organic carbon and water retention after conversion of grasslands to pine plantations in the Ecuadorian Andes, *Ecosystems*, 7, 729–739, 2004.
- Geesing, D., Felker, P., and Bingham, R. L.: Influence of mesquite (*Prosopis glandulosa*) on soil nitrogen and carbon development: Implications for global carbon sequestration, *J. Arid Environ.*, 46, 157–180, 2000.
- Goberna, M., Sánchez, J., Pascual, G., and García, C.: *Pinus halepensis* Mill. plantations did not restore organic carbon, microbial biomass and activity levels in a semi-arid Mediterranean

- soil, *Appl. Soil Ecol.*, 36, 107–115, 2007.
- Grünzweig, J. M. and Körner, C.: Growth, water and nitrogen relations in grassland model ecosystems of the semi-arid Negev of Israel exposed to elevated CO₂, *Oecologia*, 128, 251–262, 2001.
- Grünzweig, J. M., Lin, T., Rotenberg, E., Schwartz, A., and Yakir, D.: Carbon sequestration in arid-land forest, *Glob. Change Biol.*, 9, 791–799, 2003.
- Grünzweig, J. M., Sparrow, S. D., and Chapin III, F. S.: Impact of forest conversion to agriculture on carbon and nitrogen mineralization in subarctic Alaska, *Biogeochemistry*, 64, 271–296, 2003.
- Grünzweig, J. M., Sparrow, S. D., Yakir, D., and Chapin III, F. S.: The impact of agricultural land-use change on carbon storage in boreal Alaska, *Glob. Change Biol.*, 10, 452–472, 2004.
- Guo, L. B. and Gifford, R. M.: Soil carbon stocks and land use change: a meta analysis, *Glob. Change Biol.*, 8, 345–360, 2002.
- Halliday, J. C., Tate, K. R., McMurtrie, R. E., and Scott, N. A.: Mechanisms for changes in soil carbon storage with pasture to *Pinus radiata* land-use change, *Glob. Change Biol.*, 9, 1294–1308, 2003.
- Harmon, M. E., Nadelhoffer, K. J., and Blair, J. M.: Measuring decomposition, nutrient turnover, and stores in plant litter, in: *Standard Soil Methods for Long-Term Ecological Research*, edited by: Robertson, G. P., Coleman, D. C., Bledsoe, C. S., and Sollins, P., pp. 202–240, Oxford University Press, New York, 1999.
- Harrison, A. F., Harkness, D. D., Rowland, A. P., Garnett, J. S., and Bacon, P. J.: Annual carbon and nitrogen fluxes in soils along the European forest transect, determined using the ¹⁴C-bomb, in: *Carbon and Nitrogen Cycling in European Forest Ecosystems*, edited by: Schulze, E.-D., 257–275, Springer Verlag, Berlin, 2000.
- Hooker, T. D. and Compton, J. E.: Forest ecosystem carbon and nitrogen accumulation during the first century after agricultural abandonment, *Ecol. Appl.*, 13, 299–313, 2003.
- Houghton, J. T., Meira Filho, L. G., Lim, B., Treanton, K., Mamaty, I., Bonduki, Y., Griggs, D. J., and Callender, B. A.: Revised 1996 IPCC Guidelines for National Greenhouse Gas Inventories, Reporting Instructions, UK Meteorological Office, Bracknell, UK, 1996.
- Houghton, R. A.: Changes in the storage of terrestrial carbon since 1850, in: *Soils and Global Change*, edited by: Lal, R., Kimble, J., Levine, E., and Stewart, B. A., 45–65, CRC Lewis Publishers, Boca Raton, 1995.
- Jackson, R. B., Banner, J. L., Jobbágy, E. G., Pockman, W. T., and Wall, D. H.: Ecosystem carbon loss with woody plant invasion of grasslands, *Nature*, 418, 623–626, 2002.
- Jackson, R. B. and Schlesinger, W. H.: Curbing the U.S. carbon deficit, *Proc. Natl. Acad. Sci. U. S. A.*, 101, 15 827–15 829, 2004.
- KKL-JNF: A Guide for Wood Volume of the Main Species in Israel, Keren Kayemeth Leisrael-Jewish National Fund, Kiryat Haim, (in Hebrew), 1999.
- Lal, R.: Carbon sequestration in dryland ecosystems, *Environ. Manage.*, 33, 528–544, 2004.
- Maestre, F. T. and Cortina, J.: Are *Pinus halepensis* plantations useful as a restoration tool in semiarid Mediterranean areas?, *Forest Ecol. Manag.*, 198, 303–317, 2004.
- Midwood, A. J. and Boutton, T. W.: Soil carbonate decomposition by acid has little effect on δ¹³C of organic matter, *Soil Biol. Biochem.*, 30, 1301–1307, 1998.
- Nosetto, M. D., Jobbágy, E. G., and Paruelo, J. M.: Carbon sequestration in semi-arid rangelands: Comparison of *Pinus ponderosa* plantations and grazing exclusion in NW Patagonia, *J. Arid Environ.*, 67, 142–156, 2006.
- Pacala, S. W. and Socolow, R.: Stabilization wedges: solving the climate problem for the next 50 years with current technologies, *Science*, 305, 968–972, 2004.
- Paul, E. A., Morris, S. J., Six, J., Paustian, K., and Gregorich, E. G.: Interpretation of soil carbon and nitrogen dynamics in agricultural and afforested soils, *Soil Sci. Soc. Am. J.*, 67, 1620–1628, 2003.
- Paul, K. I., Polglase, P. J., Nyakuengama, J. G., and Khanna, P. K.: Change in soil carbon following afforestation, *Forest Ecol. Manag.*, 168, 241–257, 2002.
- Perez, C., Roncoli, C., Neely, C., and Steiner, J. L.: Can carbon sequestration markets benefit low-income producers in semi-arid Africa? Potentials and challenges, *Agr. Syst.*, 94, 2–12, 2007.
- Post, W. M., and Kwon, K. C.: Soil carbon sequestration and land-use change: processes and potential, *Glob. Change Biol.*, 6, 317–327, 2000.
- Quideau, S. A., Chadwick, O. A., Trumbore, S. E., Johnson-Maynard, J. L., Graham, R. C., and Anderson, M. A.: Vegetation control on soil organic matter dynamics, *Org. Geochem.*, 32, 247–252, 2001.
- Rathore, S. K. S., Singh, S. P., Singh, J. S., and Tiwari, A. K.: Changes in forest cover in a central Himalayan catchment: inadequacy of assessment based on forest area alone, *J. Environ. Manage.*, 49, 265–276, 1997.
- Safriel, U. N. and Moshe, I.: Monitoring and Evaluating Afforestation in a Semiarid Watershed (Yatir Region, Israel), Annual Scientific Report, Middle East Watershed Monitoring and Evaluation Project, US-AID MERC, 2005.
- Scarascia-Mugnozza, G., Bauer, G. A., Persson, H., Matteucci, G., and Masci, A.: Tree biomass, growth and nutrient pools, in: *Carbon and Nitrogen Cycling in European Forest Ecosystems*, edited by: Schulze, E.-D., 49–62, Springer Verlag, Berlin, 2000.
- Schlesinger, W. H.: Evidence from chronosequence studies for a low carbon-storage potential of soils, *Nature*, 348, 232–234, 1990.
- Schulze, E.-D., Wirth, C., and Heimann, M.: CLIMATE CHANGE: Managing forests after Kyoto, *Science*, 289, 2058–2059, 2000.
- Shaver, G., Billings, W., Chapin, F. I., Giblin, A., Nadelhoffer, K., Oechel, W., and Rastetter, E.: Global change and the carbon balance of arctic ecosystems, *Bioscience*, 42, 433–441, 1992.
- Sheldrick, B. H. and Wang, C.: Particle size distribution, in: *Soil Sampling and Methods of Analysis*, edited by: Carter, M. R., 499–511, Canadian Society of Soil Science, Lewis Publishers, Boca Raton, 1993.
- Six, J., Callewaert, P., Lenders, S., De Gryze, S., Morris, S. J., Gregorich, E. G., Paul, E. A., and Paustian, K.: Measuring and understanding carbon storage in afforested soils by physical fractionation, *Soil Sci. Soc. Am. J.*, 66, 1981–1987, 2002.
- Six, J. and Jastrow, J. D.: Organic matter turnover, in: *Encyclopedia of Soil Science*, edited by: Lal, R., 936–942, Marcel Dekker, New York, 2002.
- Smith, D. L. and Johnson, L. C.: Expansion of *Juniperus virginiana* L. in the Great Plains: Changes in soil organic carbon dynamics, *Global Biogeochem. Cy.*, 17, 1062, 2003.
- Sollins, P., Glassman, C., Paul, E. A., Swanston, C., Lajtha, K.,

- Heil, J. W., and Elliott, E. T.: Soil carbon and nitrogen. Pools and fractions, in: *Standard Soil Methods for Long-Term Ecological Research*, edited by: Robertson, G. P., Coleman, D. C., Bledsoe, C. S., and Sollins, P., 89–105, Oxford University Press, New York, 1999.
- Thuille, A., Buchmann, N., and Schulze, E.-D.: Carbon stocks and soil respiration rates during deforestation, grassland use and subsequent Norway spruce afforestation in the Southern Alps, Italy, *Tree Physiol.*, 20, 849–857, 2000.
- Thuille, A. and Schulze, E.-D.: Carbon dynamics in successional and afforested spruce stands in Thuringia and the Alps, *Glob. Change Biol.*, 12, 325–342, 2006.
- Ulery, A. L., Graham, R. C., Chadwick, O. A., and Wood, H. B.: Decade-scale changes of soil carbon, nitrogen and exchangeable cations under chaparral and pine, *Geoderma*, 65, 121–134, 1995.
- Ussiri, D. A. N., Lal, R., and Jacinthe, P. A.: Soil properties and carbon sequestration of afforested pastures in reclaimed mine-soils of Ohio, *Soil Sci. Soc. Am. J.*, 70, 1797–1806, 2006.
- Vesterdal, L., Ritter, E., and Gundersen, P.: Change in soil organic carbon following afforestation of former arable land, *Forest Ecol. Manag.*, 169, 137–147, 2002.
- Watson, R. T., Noble, I. R., Bolin, B., Ravindranath, N. H., Verardo, D. J., and Dokken, D. J. (Eds.): *Land Use, Land-use Change, and Forestry*, Intergovernmental Panel on Climate Change – Cambridge University Press, Cambridge, 2000.