

Basin-wide variations in foliar properties of Amazonian forest: phylogeny, soils and climate

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Received: 16 December 2008 – Published in Biogeosciences Discuss.: 6 April 2009

Revised: 14 October 2009 – Accepted: 5 November 2009 – Published: 27 November 2009

Abstract. We analysed 1040 individual trees, located in 62 plots across the Amazon Basin for leaf mass per unit area (M_A), foliar carbon isotopic composition ($\delta^{13}\text{C}$) and leaf level concentrations of C, N, P, Ca, Mg, K and Al. All trees were identified to the species level with the dataset containing 58 families, 236 genera and 508 species, distributed across a wide range of soil types and precipitation regimes. Some foliar characteristics such as M_A , [C], [N] and [Mg] emerge as highly constrained by the taxonomic affiliation of tree species, but with others such as [P], [K], [Ca] and $\delta^{13}\text{C}$ also strongly influenced by site growing conditions. By removing the environmental contribution to trait varia-

tion, we find that intrinsic values of most trait pairs coordinate, although different species (characterised by different trait suites) are found at discrete locations along a common axis of coordination. Species that tend to occupy higher fertility soils are characterised by a lower M_A and have a higher intrinsic [N], [P], [K], [Mg] and $\delta^{13}\text{C}$ than their lower fertility counterparts. Despite this consistency, different scaling patterns were observed between low and high fertility sites. Inter-relationships are thus substantially modified by growth environment. Analysing the environmental component of trait variation, we found soil fertility to be the most important predictor, influencing all leaf nutrient concentrations and $\delta^{13}\text{C}$ and reducing M_A . Mean annual temperature was negatively associated with leaf level [N], [P] and [K] concentrations. Total annual precipitation positively influences M_A ,



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[C] and $\delta^{13}\text{C}$, but with a negative impact on [Mg]. These results provide a first basis for understanding the relationship between the physiological functioning and distribution of tree species across Amazonia.

1 Introduction

Plants are the central link in the soil-plant-atmosphere continuum, utilising and cycling a range of atmospherically (C, H, O, and N) or geologically (P, Ca, K) derived elements (Jobbagy and Jackson, 2004). Plant growth is usually considered to be either nitrogen- or phosphorus-limited (Aerts and Chapin, 2000), but with less abundant nutrients also being important for discrete ecosystem processes (Hungate et al., 2004; Kaspari et al., 2008). Foliar ratios of leaf level nitrogen, phosphorus, calcium, and potassium concentrations can indicate the nature of nutrient limitation (Koerselman and Meuleman, 1996; Jobbagy and Jackson, 2004). It has been argued, for example, that leaf N:P > 12.5 indicate a limitation on ecosystem processes by P availability (Tessier and Raynal, 2003). But different hypotheses have been proposed regarding the factors controlling the metabolic tissue concentration of the main nutrients related to plant growth (Reich and Oleksyn, 2004), especially N and P (Niklas, 2006). These hypotheses summarize a physiological, temperature or geochemical driven background of N and P cycling and have gained particular interest recently, as foliar C:N:P stoichiometry may be an important factor controlling the growth rate of a wide range of plants (Elser et al., 2000; Kerkhoff et al., 2005; Niklas, 2006). Following on from the now widely accepted suggestion of Vitousek (1984) that tropical forests may generally be P rather than N limited, Reich and Oleksyn (2004) showed that tropical trees typically have higher N:P ratios than are observed at higher latitudes. Nevertheless, Townsend et al. (2007) also showed that for trees growing on the more fertile tropical soils, foliar N:P ratios are generally similar to those observed in the temperate and boreal zones. Examining a range of potential edaphic and environmental predictors, Quesada et al. (2009a) found total soil phosphorus to be the best predictor of above ground woody biomass growth rates for a wide range of Amazon forest types growing on a wide range of different soil types.

The Amazon Basin is highly diverse in terms of climate (Sombroek, 2001; Malhi and Wright, 2004), soil physical and chemical properties (Sombroek, 2000; Quesada et al., 2009b), and species composition (ter Steege et al., 2006). Such complexity hinders any attempt to accurately estimate significant biogeochemical fluxes (Townsend et al., 2008) or to predict the Amazon carbon balance (Cox et al., 2000). It is also now clear that a large scale gradient in Amazon forest tree dynamics exists, with forests growing on the generally more fertile soils of the western Amazon having lower wood densities (Baker et al., 2004), higher above-ground

growth rates (Malhi et al., 2004) and with higher rates of tree turnover (Phillips et al., 2004). But how do key foliar properties vary across evolutionary grouping of trees and relate to these different environmental gradients? This is an important question; both to help us understand the ecology and dynamics of Amazonian rain forests and also to rationally parameterise the next generation of coupled vegetation – climate models.

Recent developments in plant functional ecology have highlighted suites of plant traits such as per area leaf mass (M_A) and leaf nutrient concentrations, that can serve as predictors of individual plant growth and performance (Reich et al., 1991; Garnier et al., 2004; Poorter and Bongers, 2006). Although considerable variability in these characters has been reported, a global spectrum of coordination has also been proposed (Reich et al., 1997; Wright et al., 2004). One of the major axes of ecological variation is thought to be captured by both leaf longevity and M_A (or its inverse specific leaf area, SLA), with these two traits well correlated at both a global and a tropical scale (Reich et al., 1991, 1997). Species with low M_A tend to have short-lived leaves with high [N] and [P] (dry weight basis) and are usually found at the fast payback end of the “economic spectrum” (Wright et al., 2004). This combination of traits is usually found at species with fast growth rates (Poorter and Bongers, 2006). At the other end of the continuum, species characterised by high M_A , also generally have low leaf [N] and [P] and lower growth rates, thus being considered to represent a more conservative strategy of resource use and turnover (Reich et al., 2003). Wright et al. (2005b) examined the role of potassium within the “leaf economic spectrum”, concluding that although associated with M_A , N and P, [K] might be more closely associated with other cations such as calcium and magnesium.

All major cations are considered in this study, including [Al], along with an additional major potential component of the leaf physiological spectrum, *viz* the extent of discrimination against the heavier ^{13}C isotope during photosynthetic CO_2 assimilation, Δ . This is indicated by a leaf’s carbon isotopic composition, $\delta^{13}\text{C}$, and can provide a measure of a plant’s water use efficiency, WUE (Farquhar et al., 1989). It has, for example, been proposed that leaves with a high M_A should also have a lower WUE (Lamont et al., 2002).

Here, as well as considering $\delta^{13}\text{C}$ we also examine the integration into the plant physiological spectrum of leaf carbon content. Although often considered to be relatively invariant, differences in M_A should nevertheless be accompanied by differences in leaf carbon composition. For example, the greater proportion of dry matter invested in cell walls as expected in high M_A plants should also be associated with increased levels of carbon rich structural carbohydrate compounds such as lignin and cellulose (Niinemets, 1997). Although not reporting on the relationships between individual compounds or elements and M_A , when investigating leaf chemical variations for 45 different French Guiana rain forest

species, Hättenschwiler et al. (2008) reported considerable variation in foliar carbon contents (from 0.45 to 0.52 of leaf dry weight) with large inter-specific variations in different carbon constituent compounds also noted.

Along with the coordination of leaf functional characters, environmental factors, such as climate and/or soil may affect physiological linkages. Modifications of pair-wise relations (Wright et al., 2001) or systematic trends of a functional trait across environmental gradients should indicate trait plasticity and/or adaptive potential (Sultan, 2000). These may have important consequences for individual and community level processes. Evaluating this plasticity is thus a key issue in developing theoretical and computational schemes of the potential vegetation response to changing environmental conditions.

At a global level, Wright et al. (2005a) have shown how some photosynthetic tissue properties and their relationships also vary across climate, but for the selected traits (M_A , leaf longevity, foliar [N] and photosynthetic capacity) climate accounted for only 0.18 of the total variation. Nevertheless, such shifts were considered to be of significant importance for the global leaf economic spectrum. At regional scales and along rainfall and soil phosphorus gradients, significant strategic shifts in leaf properties and functioning have also been identified, with species found at drier sites exhibiting a lower photosynthetic capacity for a given foliar [N] and [P] and higher [N] and [P] concentrations at a given M_A (Fonseca et al., 2000; Wright et al., 2001). Niinemets and Kull (2003) suggested that the strength of the correlation between foliar nutrient concentrations and M_A were controlled by soil nutrient availability, with the strongest associations between M_A and the most limiting nutrient. Similar results were reported for Amazonian species, but with weak M_A -[N] relationships in what were considered to be P limited stands (Reich and Walters, 1994). At a pan-tropical scale Townsend et al. (2007) found that leaf N:P ratios vary with soil order, and suggested that species diversity may be an important factor controlling this ratio.

Different tropical tree species may have markedly different foliar nutrient concentrations (Drechsel and Zech, 1991; Townsend et al., 2007) and, at least at the landscape level, different species tend to be associated with soils of different fertility (Phillips et al., 2003; John et al., 2007). Given that even for the same species, natural variations in soil fertility may also serve to modify foliar seedling nutrient concentrations along with other leaf physiological characteristics such as M_A (Veenendaal et al., 1996; Kanowski, 2001), we were also interested to ascertain whether differences in foliar nutrient concentrations sampled across Amazonia represented directly different levels of soil fertility and/or intrinsic differences in physiological leaf traits of the different species growing in different plots. Likewise, although there may be large scale changes in physiological traits with rainfall or temperature when a range of species are considered (Santiago et al., 2004a; Wright et al., 2005a) it is important to

know if changes in traits observed at the community level arise solely as a consequence of changes in species composition or whether factors such as dry-season length directly influence physiological properties and their inter-relationships.

This plasticity of traits and the potential for tree species population to exhibit different trait combinations according to changing soil and climate conditions may be very important for understanding Amazon forest species distributions. So here we analyse nine key leaf traits of 1040 individuals positioned in sixty-two plots distributed across the Amazon Basin. The traits reported are M_A , $\delta^{13}\text{C}$, and leaf level concentrations of C, N, P, Ca, Mg, K and Al with 508 species sampled across a wide range of Amazon soil types and precipitation regimes. An accompanying paper will investigate structural traits such as maximum tree height, individual leaf area, ratio of supported leaf area to xylem cross sectional area, branch xylem density and seed size, also integrating the dataset presented here with those components, and across the edaphic and precipitation regimes observed (Patiño et al., 2009).

The aims of this paper are first to partition variation in foliar properties into genetic and environmental components. Second to identify the extent to which traits are conservative across evolutionary grouping of tree species, and how these traits differ across taxonomic affinities. Third to explore if there are differences in the trait combinations found under different environmental conditions. Ultimately, we attempt to link environmental components of trait variation with key soil and climate variables. We specifically hypothesised that:

1. Although genetically determined, many of the traits of interest would be modified by the growth environment – as has been demonstrated recently, for example, by Poorter et al. (2009) for M_A .
2. That species which occur in the faster growing stands on the more fertile soils of the Amazon Basin (Quesada et al., 2009a) would be characterised by leaf traits associated with a faster growth potential – for example a lower M_A and intrinsically higher [N] and [P] than species occurring on the less fertile substrates. Although it has long been assumed that this should be the case for plants in general (Lambers and Poorter, 1992), we know of few demonstrations that this is actually the case and none at a spatial scale similar to that of the Amazon Basin.
3. That the strong dependence of Amazon forest growth on soil available phosphorus concentrations (Quesada et al., 2009a) would also be reflected in much higher concentrations of foliar P (as well as possible other elements) for stands growing on more fertile soils – this being above and beyond that expected on the basis of differences in species composition (as in 2. above) alone.

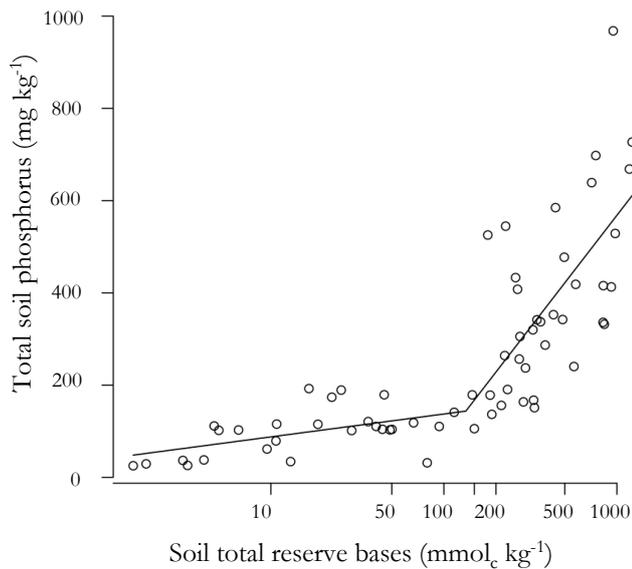


Fig. 1. Broken line regression and breakpoint identification in the linear relationship between soil total P content and \log_{10} -transformed total reserve bases. See also Quesada et al. (2009c).

2 Methods

2.1 Study sites and leaf minerals, xylem density and environmental data

For 62 of the RAINFOR network plots described in detail in Patiño et al. (2009), we utilised foliar nutrient and M_A data from upper canopy sun-exposed leaves from trees reliably identified at the species level, with data collected and analysed as described in detail in Lloyd et al. (2009). The elements of interest were leaf level concentrations of C, N, P, Ca, Mg, K, all expressed here in mg g^{-1} , with leaf mass per unit area, M_A , expressed in units of g m^{-2} and $\delta^{13}\text{C}$ as per mil (‰). As in Lloyd et al. (2009), [C] as shown and analysed here, has been adjusted for variations in mineral content by subtracting the measured concentrations of the major cations *viz* [Ca]+[Mg]+[K]+[Na]. This allows variations in [C] to be better interpreted in terms of variations in foliar carbohydrate chemistry, as opposed to variation in [C] simply reflecting differences in mineral concentrations.

Some sample plots included in Patiño et al. (2009) are not considered in this analysis, due to less than complete species identification (APG II, 2003). These include ALF-01, MAN-03, SIN-01, SUC-04, ZAR-01,02,03,04). Data from some plots have been aggregated (ex. TAP-01 & TAP-02 & TAP-03 \rightarrow TAP-123, CAX-03 & CAX-03.1 \rightarrow CAX-03) where they were located in close proximity and having all but identical topography, soils and climatic conditions (see Supplementary information I, Table S11: <http://www.biogeosciences.net/6/2677/2009/bg-6-2677-2009-supplement.zip>).

Previous work has shown tropical forest soil type to significantly influence foliar trait interactions, for example the slope of the foliar N versus foliar P relationship (Townsend et al., 2007). Similarly, Kattge et al. (2009) examined photosynthesis versus nitrogen relationships for tropical trees also finding distinctions between fertile (“oxisol”) versus infertile (“non-oxisol”) soils. To provide some linkage with these studies, soil chemical and physical data for 0–0.3 m depth, collected and analysed as described in Quesada et al. (2009c) were thus assembled to allow a classification of plots according to soil fertility groups and thus a differentiation of foliar traits and their relationships between low and high fertility sites. Our differentiation into low and high fertility soils was more quantitative, however, being based on the measured “total reserve bases” Σ_{RB} , from 0.0 to 0.3 m depth, which provides a quantitative estimate of the extent of soil weathering as described in Quesada et al. (2009c). A “breakpoint” in the linear relationships of different soil variables such as phosphorus content (Fig. 1), pH and others with $\log_{10}(\Sigma_{RB})$, was identified using broken line regression models (Muggeo, 2008), with this breakpoint almost inevitably occurring at a mean Σ_{RB} around 130 (mmol kg^{-1}). Using the World Reference Base for Soil Resources Classification System (IUSS Working Group WRB, 2006), $\Sigma_{RB} < 130 \text{ mmol kg}^{-1}$ encompasses nutrient poor soils such as arenosols, podzols and ferralsols along with any dystrophic acrisols, alisols, cambisols gleysols and plinthisols. On the other hand, $\Sigma_{RB} > 130 \text{ mmol kg}^{-1}$ defines the usually fertile cambisols, along with the more fertile alisols, nitisols, fluvisols, lixisols and plinthisols. Σ_{RB} is an easily measured soil property which is often determined in soil science studies, both in the tropics and elsewhere (e.g. Federer et al., 1998; Barthold et al., 2008; Quesada et al., 2009c).

Climate, temperature and precipitation datasets were obtained from the free access web site at <http://www.worldclim.org>. This set of global climate layers, “WorldClim”, includes annual time series with mean monthly data for precipitation, and mean, minimum and maximum temperatures obtained from over 4000 weather stations between 1950 and 2000 (Hijmans et al., 2005). Solar radiation data is from New et al. (2002).

2.2 Statistical analysis

The analysis here focuses on genetic and plot-environmental components of trait variation, as estimated from a multilevel model and discussed below.

Preliminary tests included: analysis of normality (Shapiro-Wilk), and homogeneity of variances (Fligner-Killeen) for each foliar property. This showed [C], [N], [P], [Ca], [Mg] [K] and M_A to all not be normally distributed and these parameters were thus \log_{10} -transformed prior to analyses. Foliar $\delta^{13}\text{C}$ was approximately normally distributed, but we \log_{10} -transformed the absolute values in order to consistently analyse the full dataset, taking the negative values of the

transform and then returning the values to the original sign. One way analysis of variance (ANOVA) was used to explore for differences between fertility groups, as well as for differences between families, genera within a family and species within a genus. All analyses were performed with the R statistical platform (R Development Core Team, 2008).

A multilevel model (Snijders and Bosker, 1999; McMahon and Diez, 2007) was first fitted for each foliar trait according to

$$T = \mu + p + f/g/s + \varepsilon, \quad (1)$$

where μ is the overall mean value of each trait (T), p is the plot effect, i.e. the effect of the location at which each individual was found (soil and climate), $f/g/s$ represents the genetic structure of the data, i.e. that each individual belongs to a species (s), nested in a genus (g), nested in a family (f), and ε is the residual term which includes both natural within-species variability as well as any measurement error. All parameters were estimated by the Residual Maximum Likelihood (REML) method with the *lme4* library (Bates and Sarkar, 2007). The multilevel model Eq. (1), in a similar way to taxonomically based nested ANOVAs, can be used to partition the variance from species up to the family level. It is particularly useful for not fully resolved plant supertrees (Kerckhoff et al., 2006; Watanabe et al., 2007) and it can be used to estimate group- or individual-level regression coefficients and their variation in unbalanced datasets (Gelman and Hill, 2006; pp. 246) with even one observation per group (Gelman and Hill, 2006; pp. 276). In our case the estimated components of variance can be distinguished into both a {"plot" – "environmental"} and a "genetic" term. Recognising that our study represents an incomplete and, to a large extent, under-represented sampling of the edaphic and climatic variability of Amazonia, as well as only a selection of the many trees species living there, all terms in the multilevel model were treated as random (as opposed to fixed) effects (McCulloch and Searle, 2001). Random effects were quantified through the best linear unbiased predictor (BLUP) method, providing shrunken estimates of the differences between terms and the overall means (Galwey, 2006). The random genetic and the plot effects were then used for further analysis as described below.

Due to the highly unbalanced nature of our dataset we verified the efficiency of the hierarchical/multilevel model in both partitioning the variance and in providing accurate estimates of the genetic and plot level dependencies of the observed foliar properties through a simulation study (Supplementary Information SII). In these simulations a predefined pattern of variance partitioning as well as species and plot level effects was imposed (along with significant residual effects) with the sampling of individual species undertaken in a highly unbalanced manner similar to that of our actual sample. We then retrieved the genetic and environmental effects for the artificially generated population using the REML approach, also testing the efficacy of the REML procedure in

partitioning the trait variation between species and plots in the presence of substantial residual "noise". As is shown in Supplementary Information SII, the hierarchical model was able to adequately extract both the variance structure and the magnitude of the species/plot effects. Most importantly, it also provided unbiased estimates of the slopes of the bivariate relationships existing between the various traits of interest for both the genetic and plot-environmental effects.

In what follows we consider the derived environmental term to represent the combined influences of climate, soil and location. The genetic term (estimated here as the sum of the family, genus and species effects) represents the phylogenetic structure of the dataset. We note that this taxonomically based multilevel model does not have as input values the species means for the one species calculated at single and/or multiple sites as is sometimes the case (e.g. Baker et al., 2004; Wright et al., 2004). But rather, it incorporates the full range of available information, taking into account the observed intra-specific variation in foliage characteristics and allowing for all traits to vary systematically across the different plots sampled, as well as allowing for intra-species variability within any one plot.

Bivariate relationships of foliar properties were first assessed with Pearson's correlation coefficient (r), and with Standardised Major Axis (SMA) line fits (Legendre and Legendre, 1998; Warton et al., 2006) subsequently applied where r was significantly different from zero. SMA regression lines represent the first axis of a principal component analysis (of a correlation matrix) and are often used in plant allometry studies. It is common for variables to be logarithmically transformed with the regression $\log(y) = \log(\beta) + \alpha \log(x)$, this expressing a power law of the form $y = \beta x^\alpha$. The slope or scaling exponent α , quantifies the rate of increase of y in relation to x , indicating an isometric ($\alpha \approx 1$) or allometric ($\alpha \neq 1$) scale. The intercept or elevation β of the regression line expresses the magnitude of y per unit of x (Kerckhoff and Enquist, 2006). SMA regressions were used for both the genetic ($f/g/s$) and the plot-environmental (p) component of different trait pairs. For the genetic component we further tested for differences in elevation and shift across the major axis for high and low fertility oriented species using the *R smatr* library (Warton et al., 2006).

The plot effect trait estimates were further explored with three different statistical methods; one of them presented here and two of them in Appendix A and Supplementary Information 3 (<http://www.biogeosciences.net/6/2677/2009/bg-6-2677-2009-supplement.zip>). Initially multiple linear regressions (OLS) of plot contribution to trait variation against a set of key soil and climate variables were performed. For this, a matrix of soil variables was assembled using data from Quesada et al. (2009c) i.e. total soil C and N concentration, exchangeable Ca, Mg, K, Na and Al, total extractable phosphorus, Σ_{RB} , effective cation exchange capacity, base saturation and the sand and clay fraction. To reduce dimensions and avoid multicollinearity we applied

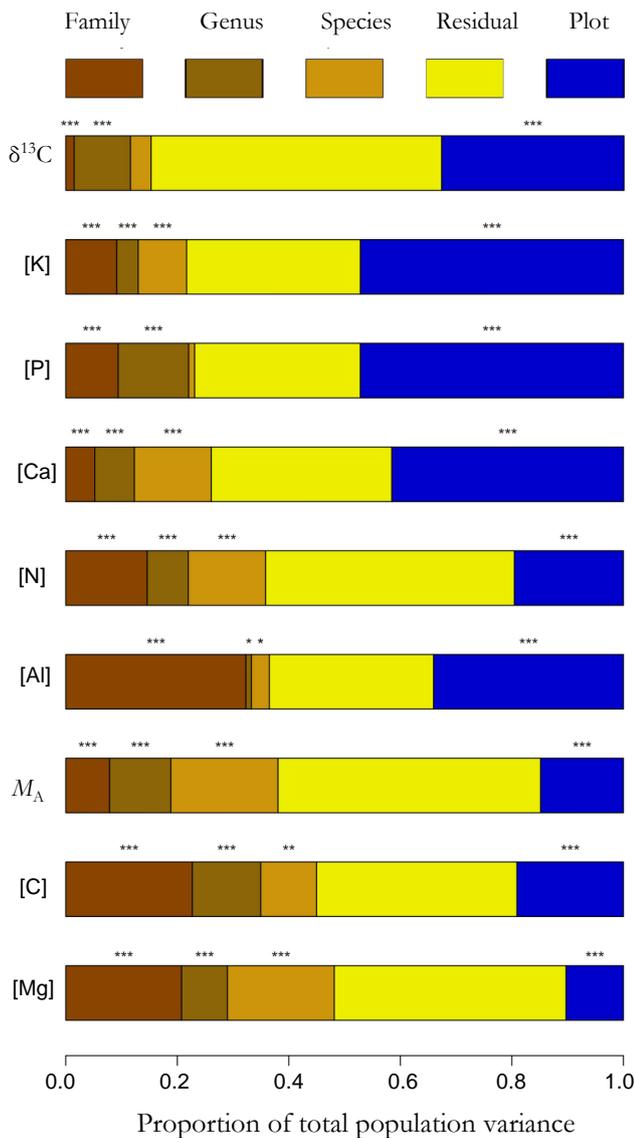


Fig. 2. Partitioning of the total variance for each foliar property into genetic (family/genus/species), environmental (plot) and an error (residual) components. Foliar properties are sorted from less to more phylogenetically constrained. Significance of each variance component was tested with a likelihood ratio test (Faraway, 2004; Galwey, 2006). Significance codes: ***<math><0.001</math>, **<math><0.01</math>, *<math><0.05</math>. M_A denotes leaf mass per unit area, $\delta^{13}\text{C}$ represents the dry matter $^{13}\text{C}/^{12}\text{C}$ composition (per mil) with all other symbols representing the elemental composition of the leaves (dry weight basis). All values had been \log_{10} transformed prior to analysis.

principal components analysis (PCA), using the R *ade4* library (Chessel et al., 2004), to the correlation table of the soil matrix with most variables log-transformed (Quesada et al., 2009b) and derived the main axes of variation. Multiple regressions for each trait's "plot effect" were then computed on the first three principal components (Legendre and Legendre, 1998) with four climate variables also included; annual

mean temperature, total annual precipitation, precipitation during the three driest months of the year and mean annual radiation (Appendix A). Simplification of the full model was explored following a stepwise elimination of the less significant terms, based on Akaike's Information Criterion (AIC) to derive the minimum adequate model (Crawley, 2007). Additionally, simultaneous autoregressive models (SAR) including a spatial error term at two scales were also fitted (Appendix A). Our spatial analysis was not designed to fully disentangle the scales where different environmental variables control the observed variation of the leaf properties studied. Rather, we have tried to filter the most important environmental predictors and to investigate if the trends identified by the OLS were biased and if the identified important environmental variables from OLS remained significant after accounting for spatial patterns in our models (Lichstein et al., 2002).

Finally as some plot effect versus environmental predictor relationships did not show a simple linear trend and/or normally distributed homoscedastic errors, we used Kendall's τ as a non-parametric measure of association. Though less common than Spearman's ρ , Kendall's τ has slightly better distributional properties and also has the advantage that it can be interpreted in terms of probabilities of observing concordant and discordant pairs (Conover, 1980). Specifically, $\tau = \pi_c - \pi_d$, where π_c is the probability of concordant pairs and π_d is the probability of discordant. For example, if $\tau=0.5$, then 0.75 of the ranked pairs are concordant and 0.25 are discordant. Kendall's τ also has the advantage that it can be generalised to a partial correlation coefficient (Legendre and Legendre, 1998; pp. 202). As is discussed in Legendre and Legendre (1998) it is, however, difficult to assess the statistical significance of the partial τ and so here we assess likely significance levels of our calculated partial τ by numerical simulation as described in Maghsoodloo and Laszlo Pallos (1981). Although not allowing spatial patterns to be taken implicitly into account, this approach does allow the inter-relationships between environmental predictors to be explicitly included in the analysis of environmental factors influencing the studied plant physiological properties in a non-parametric manner.

3 Results

3.1 Partitioning of the variance

Through fitting the multilevel model of Eq. (1), a partitioning of the variance to genetic and plot level components was achieved with results presented in Fig. 2. This shows that, not only does the proportion of the variance attributable to the nested taxonomy (genetic) component differ for different traits, but also that the level of genetic variation, particularly at the species level, contrasts greatly between traits. For example, for M_A the genetic component accounts for

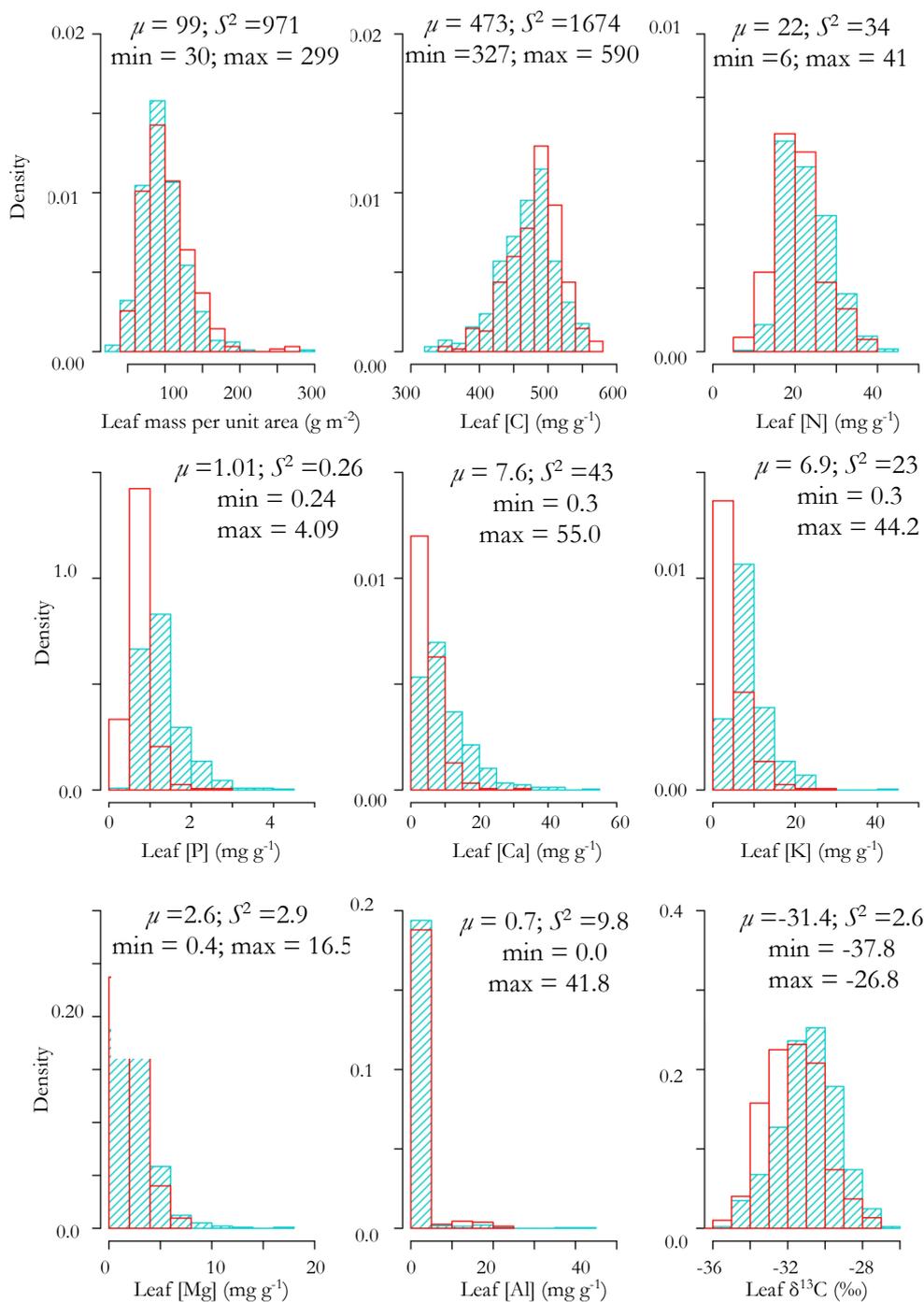


Fig. 3. Probability density histograms of raw data per fertility group. Red bars represent low and blue bars high soil fertility plots, as defined by the quantitative determinations of the level of total reserve bases from 0–30 cm depth (see Fig. 1 and text). Also given the overall mean, range and variance for each property.

approximately 0.38 of the total variance (with half of this attributable to species effect) and with the variability associated with tree location, the “plot effect”, being only 0.15 of the total variance. A little less than half the variance in the dataset is attributable to an error term. As mentioned previously, the “error term” represents the proportion of the

variance in the dataset attributable to intra-species variability as well as any measurement error.

In contrast to M_A , the principle source of variation in [P] was the “plot effect” (accounting for 0.47 of the total variance) with only 0.23 of the observed variance attributable to a genetic component and with the species

specific component of the variation being less significant. The “plot effect” was similarly very high for [Ca] and [K] (0.42 and 0.47 of the total variance respectively), whilst the lowest plot contribution of 0.1 was observed for [Mg]. Along with M_A , partitioning of the variance to the genetic component was highest for [Mg] (0.48) and [C] (0.40). Also of note is the high proportion of the variance in [Al] attributable at the family level (0.32), consistent with independent phylogenetic analyses (Jansens et al., 2002). For $\delta^{13}\text{C}$, the attributed genetic variation was less than for any of the elements or M_A , but still with an appreciable “plot effect”, suggesting that geographic variations in either soil or climate exert significant effects on the extent of photosynthetic ^{13}C discrimination across Amazonia.

The ability of the REML model to accurately retrieve the correct partitioning of the variance for the various traits is demonstrated in Supplementary Information II (<http://www.biogeosciences.net/6/2677/2009/bg-6-2677-2009-supplement.zip>) and, for the interested reader, some aspects of the inferred taxonomic variation of the examined foliar properties is presented and discussed in Appendix B, including a consideration of differences between N_2 -fixing and non- N_2 fixing members of the Fabaceae.

3.2 Statistical distribution of measured traits

Trait distributions for the complete dataset divided into the low and high soil-fertility groups are shown in Fig. 3, with the overall arithmetic mean, range and variances also shown. Significant differences across mean plot values were identified for all foliar properties as summarised in the Supplementary Information I, Table SI1. Figure 3 shows that although M_A and [C] had similar statistical distributions for both low and high fertility plots, for the other traits examined the distributions for trees growing on the more fertile soils were shifted to the right, apart from [Al] (shifted to the left). Mean leaf N:P ratio was 30.1 ± 9.5 for the low soil fertility plots, this being significantly higher ($F_{1,789}=388.6$, $p < 0.001$) than the 19.6 ± 5.6 observed on high fertility soils. Thus, the natural variations in rain forest soil fertility that occur within the Amazon Basin, exert a strong effects on plant nutrient concentrations and $\delta^{13}\text{C}$, but do not strongly influence M_A and [C].

3.3 Bivariate relationships (raw data)

A preliminary analysis of the raw data foliar properties showed significant correlations between most of the examined trait pairs as shown in Table 1 (left panel), where we also summarise the SMA regression estimates for the whole dataset (no separation into low and high fertility plots). Nevertheless, when low and high fertility sites were considered separately, most of the pairwise associations remained important, but with significant differences in slope or elevation and shift of the SMA axis being identified (see Supplemen-

tary Information I, Table SI2: <http://www.biogeosciences.net/6/2677/2009/bg-6-2677-2009-supplement.zip>). Interestingly, the [P] vs. [N] relationship and all bivariate relationships including [K], showed statistically significant differences in slope between low and high fertility sites (apart from [K] vs. [N]). In what follows we therefore retained our consideration of the effects of soil fertility on trait variation, probing differences between our fertile and infertile soil classes on the nature of the bivariate relationships for the genetic components of the overall variation observed. Some of the raw data bivariate relationships are presented in Fig. SI1.

3.4 Bivariate relationships (Genetic component)

The genetic component of the multilevel model (i.e. the *family+genus+species*) showed numerous significant correlations (Table 1 – middle panel). By separating species to low and high fertility oriented ones, most relationships identified as significant in the raw dataset, retained their importance (Table 2). As is also shown in Fig. 4a and b, using the convention $y \leftrightarrow x$ to denote bivariate relationships, a strong relationship was identified for $[\text{N}] \leftrightarrow M_A$ ($r = -0.403$ and -0.407 on low and high fertility soils respectively) and $[\text{P}] \leftrightarrow M_A$ ($r = -0.388$ and -0.433). There was also a reasonably strong relationship between M_A and [K] but this was considerably weaker for [Mg] for both soil types (Table 2, graph not shown). M_A showed a weak correlation with [Ca] and was stronger associated with [C] in species found at low soil fertility sites. The genetic component of $[\text{C}] \leftrightarrow [\text{N}]$ was only marginally significant on low fertility sites, while strong negative associations were found between leaf carbon and cations (Ca, K & Mg) content on both fertile and infertile plots.

Very strong correlations were also observed between some of the individual nutrients, with $[\text{P}] \leftrightarrow [\text{N}]$ ($r = 0.648$ and 0.698 on low and high fertility soils respectively) being especially well associated (Fig. 4c). The relationships between [K] and [P] ($r = 0.394$ and 0.507 ; Fig. 4e), [Mg] and [Ca] ($r = 0.643$ and 0.660 ; Fig. 4f) and between [K] and [Ca] ($r = 0.447$ and 0.450) are also of note, as is the observation that the relationship between [K] and [N] is substantially weaker than for $[\text{K}] \leftrightarrow [\text{P}]$. Though not shown in Fig. 4, of interest were the generally strong correlations between all three base cation pairs and with P.

A comparison of SMA slopes for all $M_A \leftrightarrow [\text{nutrient}]$ relationships showed no difference between low and high fertility sites (Table 2). This was also true for between nutrient pairs such as $[\text{P}] \leftrightarrow [\text{N}]$, $[\text{K}] \leftrightarrow [\text{P}]$, $[\text{Ca}] \leftrightarrow [\text{K}]$ and $[\text{Ca}] \leftrightarrow [\text{Mg}]$. This suggests that the intrinsic (genetically defined) way M_A is linked with leaf nutrient concentrations, as well as the way in which different nutrients are related to each other is to a large degree common for species found across the Amazon and independent of the fertility of the soil where they usually grow.

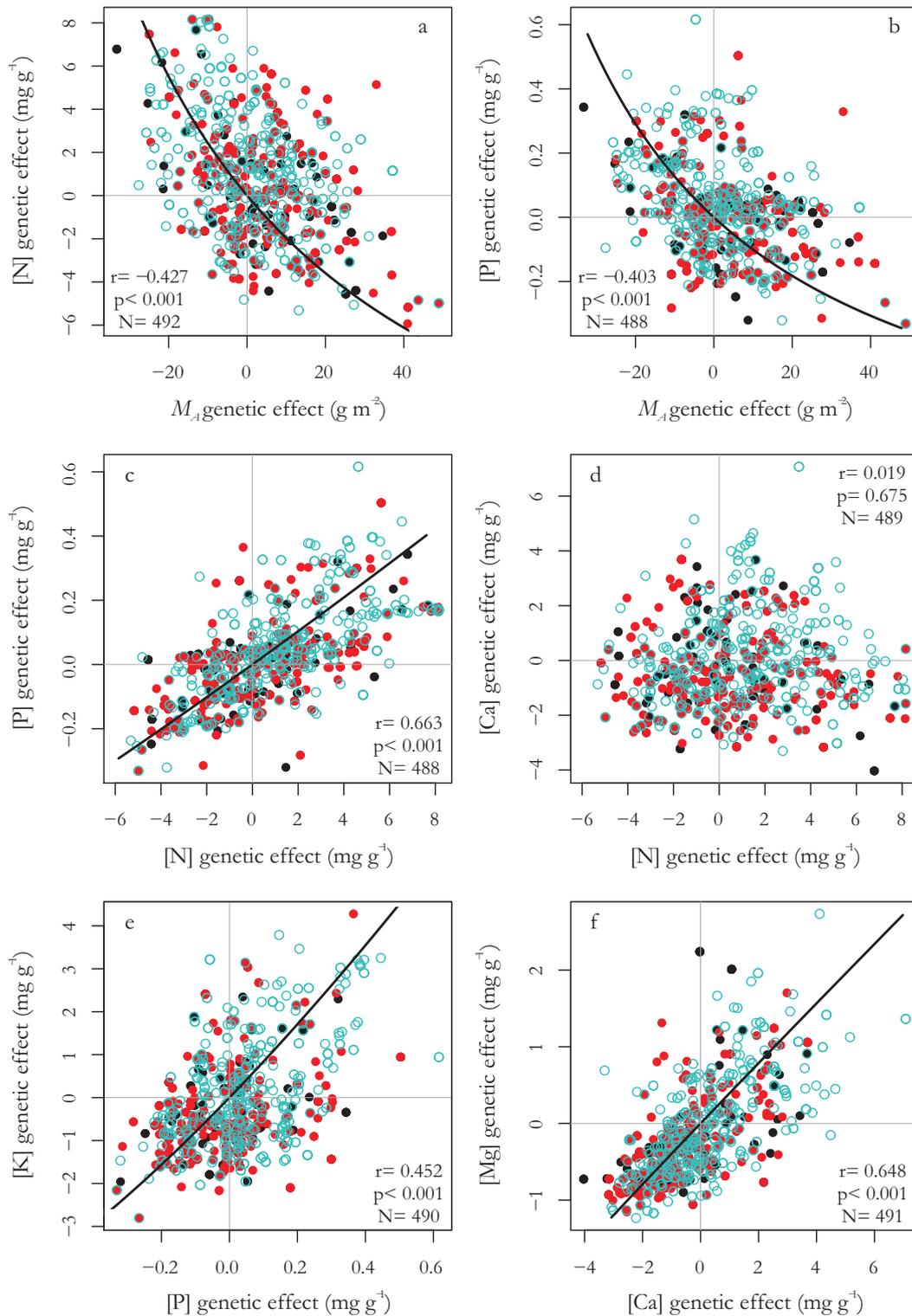


Fig. 4. Relationship between genetic effect terms for selected pairs of foliar properties. These curves are reconstructed following the footnote on Table 1 (analysis originally done on \log_{10} transformed variables, but here back transformed to the original scale) and with the overall geometric mean for each foliar property subtracted. Red dots indicate species found on low fertility sites and blue circles indicate species found on high fertility sites. Species found on both soil fertility groups are illustrated with black dots (see text for details). Common slope SMA lines are shown, though shifts along the SMA axis with fertility were also common (Table 2). M_A denotes leaf mass per unit area with other symbols representing the elemental composition of the leaves (dry weight basis).

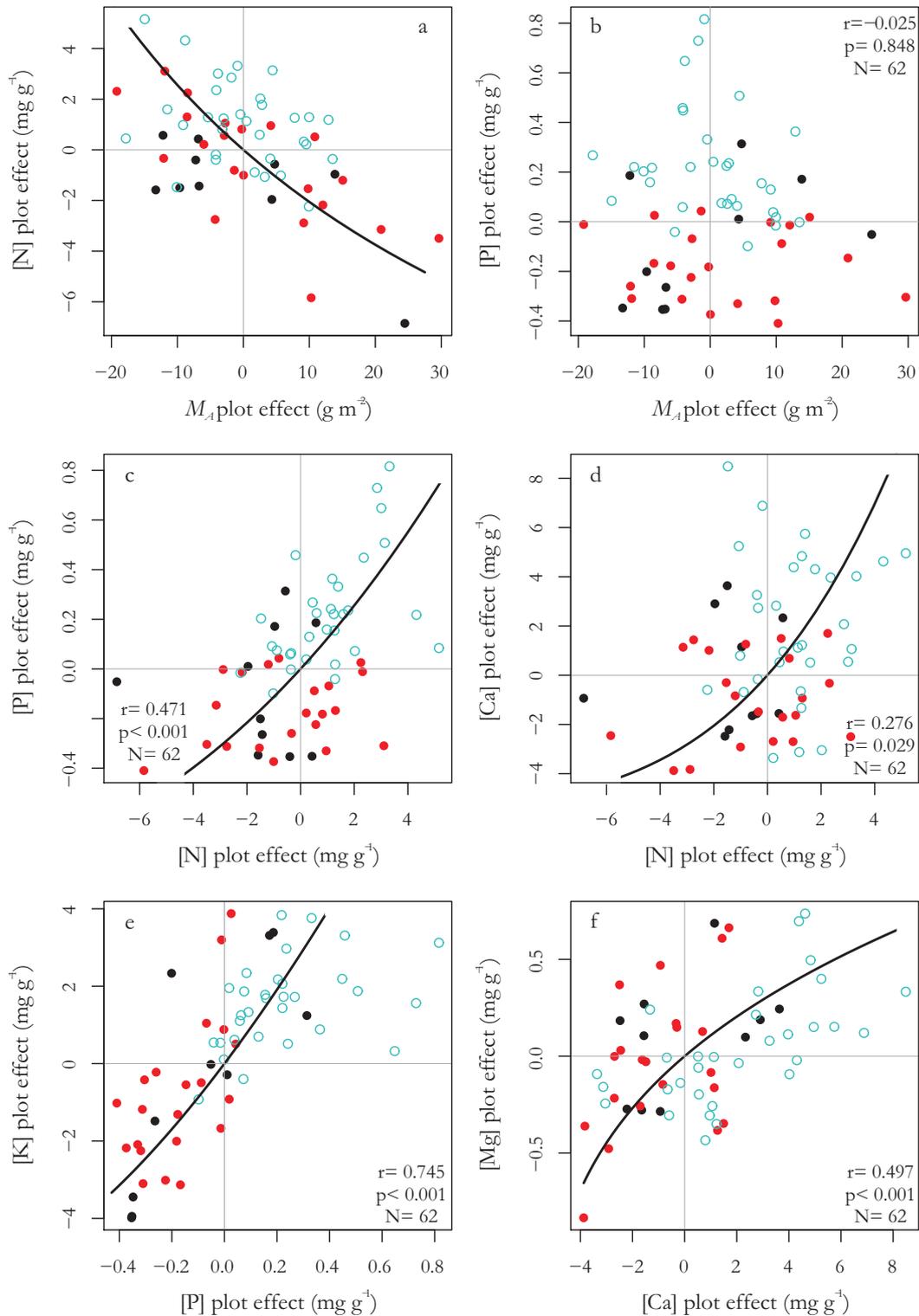


Fig. 5. Relationship between plot-environmental effects for selected pairs of foliar properties. These curves are reconstructed following the footnote on Table 1 (analysis originally done on \log_{10} transformed variables, but here back transformed to the original scale) with the overall geometric mean for each foliar property subtracted. Red dots indicate low fertility sites, blue circles indicate high fertility ones and black dots illustrate plots with no fertility classification. M_A denotes leaf mass per unit area with other symbols representing the elemental composition of the leaves (dry weight basis).

Table 1a. Pairwise relationships of key foliar properties using the raw data (left panel), pooled genetic (middle panel) and pooled plot-environmental effects (right panel). *Intercept* & *Slope* of the SMA regression, Pearson's *r* correlation coefficient, *sig* the significance of the correlation, and *n* the number of cases used. The (–) sign indicates that the respective estimates cannot be made. The genetic and plot effects lines are forced through (0,0). See note for reconstructing these lines. Significance codes: ***<0.001, **<0.01, *<0.05, –<0.01. M_A denotes leaf mass per unit area, $\delta^{13}\text{C}$ represents the $^{13}\text{C}/^{12}\text{C}$ composition (per mil) with all other symbols representing the elemental composition of the leaves (dry weight basis). All values had been \log_{10} transformed prior to analysis.

$\log_{10}(y)$	raw data				genetic effects				plot-environmental effects								
	intercept	inter-95%ci	slope	slope-95%ci	<i>r</i>	sig	<i>n</i>	slope	slope-95%ci	<i>r</i>	sig	<i>n</i>	slope	slope-95%ci	<i>r</i>	sig	<i>n</i>
[C]	2.100	(2.064 2.135)	0.290	(0.273 0.309)	0.233	***	957	0.375	(0.343 0.409)	0.139	**	487	0.318	(0.258 0.393)	0.561	***	62
M_A	3.078	(2.976 3.180)	-0.891	(-0.841 -0.944)	-0.408	***	971	-0.989	(-0.912 -1.073)	-0.407	***	492	-1.037	(-0.836 -1.286)	-0.517	***	63
[N]	2.886	(2.705 3.067)	-1.480	(-1.392 -1.574)	-0.232	***	964	-1.167	(-1.075 -1.267)	-0.423	***	488	(-)	(-)	-0.025	0.848	62
[P]	6.210	(5.865 6.555)	-2.768	(-2.599 -2.947)	-0.122	***	965	(-)	(-)	-0.069	0.127	489	-4.839	(-3.789 -6.179)	-0.264	*	62
[Ca]	5.470	(5.172 5.767)	-2.396	(-2.250 -2.550)	-0.143	***	965	-1.645	(-1.510 -1.791)	-0.270	***	489	(-)	(-)	0.015	0.910	62
[K]	3.845	(3.627 4.064)	-1.766	(-1.659 -1.879)	-0.164	***	965	-2.297	(-2.104 -2.508)	-0.127	**	489	-1.362	(-1.090 -1.703)	-0.475	***	62
[Mg]	(-)	(-)	(-)	(-)	-0.016	0.617	964	7.342	(6.720 8.022)	0.080	(-)	491	(-)	(-)	0.050	0.699	62
[Al]	-1.822	(-1.842 -1.801)	0.165	(0.155 0.176)	0.196	***	923	0.103	(0.095 0.113)	0.087	(-)	485	0.262	(0.206 0.334)	0.324	*	61
$\delta^{13}\text{C}$	9.562	(9.046 10.079)	-3.084	(-2.897 -3.284)	-0.079	*	977	2.654	(2.430 2.899)	0.077	(-)	489	-3.258	(-2.553 -4.157)	-0.275	*	62
[C]	13.544	(12.700 14.389)	-5.082	(-4.776 -5.407)	-0.150	***	977	(-)	(-)	-0.020	0.656	488	(-)	(-)	-0.102	0.432	62
[P]	25.975	(24.581 27.370)	-9.440	(-8.932 -9.975)	-0.476	***	978	-6.311	(-5.854 -6.804)	-0.513	***	489	-15.209	(-12.267 -18.857)	-0.553	***	62
[Ca]	22.967	(21.633 24.301)	-8.318	(-7.834 -8.832)	-0.297	***	978	-4.386	(-4.055 -4.744)	-0.452	***	489	(-)	(-)	-0.158	0.221	62
[K]	16.483	(15.613 17.354)	-6.033	(-5.716 -6.367)	-0.512	***	978	-6.131	(-5.675 -6.624)	-0.464	***	489	-4.282	(-3.606 -5.085)	-0.738	***	62
[Mg]	-3.076	(-3.178 -2.974)	0.590	(0.554 0.630)	0.072	*	926	(-)	(-)	0.028	0.536	482	0.274	(0.656 1.072)	0.274	*	61
$\delta^{13}\text{C}$	-2.211	(-2.327 -2.095)	1.648	(1.564 1.738)	0.532	***	976	1.174	(1.098 1.254)	0.663	***	488	2.691	(2.151 3.366)	0.471	***	62
[N]	-3.289	(-3.541 -3.037)	3.060	(2.876 3.256)	0.156	***	977	(-)	(-)	0.019	0.675	489	4.668	(3.659 5.956)	0.276	*	62
[P]	-2.811	(-3.027 -2.595)	2.690	(2.532 2.858)	0.259	***	977	1.653	(1.513 1.805)	0.178	***	489	4.151	(3.242 5.315)	0.221	*	62
[Ca]	-2.219	(-2.379 -2.058)	1.955	(1.838 2.080)	0.168	***	977	(-)	(-)	0.044	0.333	489	1.314	(1.029 1.679)	0.258	*	62
[K]	-1.739	(-1.755 -1.723)	0.185	(0.173 0.197)	0.157	***	936	0.104	(0.095 0.113)	0.237	***	486	0.250	(0.195 0.320)	0.240	(-)	61
[Mg]	0.814	(0.789 0.840)	1.856	(1.752 1.966)	0.397	***	985	2.023	(1.853 2.210)	0.140	***	490	1.735	(1.394 2.160)	0.506	***	62
$\delta^{13}\text{C}$	0.798	(0.781 0.814)	1.628	(1.555 1.704)	0.684	***	985	1.400	(1.292 1.517)	0.452	***	490	1.543	(1.302 1.829)	0.745	***	62
[Ca]	0.403	(0.385 0.422)	1.177	(1.108 1.251)	0.221	***	985	1.957	(1.792 2.137)	0.178	***	490	(-)	(-)	0.056	0.665	62
[K]	-1.492	(-1.493 -1.490)	0.112	(0.105 0.119)	0.365	***	931	0.089	(0.081 0.097)	0.273	***	482	0.093	(0.074 0.116)	0.492	***	61
[Mg]	-0.116	(-0.145 -0.088)	0.637	(0.605 0.672)	0.544	***	986	0.971	(0.909 1.038)	0.648	***	491	0.282	(0.226 0.351)	0.497	***	62
$\delta^{13}\text{C}$	-1.540	(-1.543 -1.536)	0.059	(0.056 0.063)	0.169	***	932	0.043	(0.040 0.047)	0.119	**	483	(-)	(-)	0.246	0.056	61
[Ca]	-0.093	(-0.144 -0.041)	1.138	(1.077 1.202)	0.467	***	986	1.439	(1.331 1.555)	0.456	***	491	1.125	(0.901 1.403)	0.490	***	62
[K]	-0.175	(-0.211 -0.140)	0.725	(0.684 0.769)	0.326	***	986	1.398	(1.302 1.500)	0.586	***	491	(-)	(-)	0.077	0.552	62
[Mg]	-1.547	(-1.550 -1.543)	0.069	(0.065 0.074)	0.219	***	932	(-)	(-)	0.058	0.205	483	0.061	(0.048 0.077)	0.311	*	61
$\delta^{13}\text{C}$	(-)	(-)	(-)	(-)	0.003	0.936	932	(-)	(-)	0.095	0.037	483	-0.195	(-0.152 -0.251)	-0.222	(-)	61

* The genetic and plot-environmental effect lines can be reconstructed using the following example and the table of geometric means of each foliar property (Table 1b). By definition, $\log_{10}(M_A) = (\log_{10}(M_A) + G_{\log_{10}(M_A)}) + G_{\log_{10}(M_A)}$ and $\log_{10}[\text{N}] = (\log_{10}[\text{N}] + G_{\log_{10}[\text{N}]}) + G_{\log_{10}[\text{N}]}$ where the term bracketed by $\langle \rangle$ is taken to represent its arithmetic mean value and $G_{\log_{10}(M_A)}$ and $G_{\log_{10}[\text{N}]}$ the genetic effects of the associated logged values. We then fit the bivariate relationship of $G_{\log_{10}[\text{N}]} = \alpha_G \circ G_{\log_{10}(M_A)}$, where α_G is the slope from the genetic SMA line fit, with no intercept needing to be estimated as according to the model in Eq. (1), the sum of all genetic effect terms must equal zero. To reconstruct the line, for example for $\log_{10}[\text{N}]$ as a function of $\log_{10}(M_A)$, we can write $\log_{10}[\text{N}] = (\log_{10}[\text{N}]) + \alpha_G \circ G_{\log_{10}(M_A)} = (\log_{10}[\text{N}]) + \alpha_G \circ [(\log_{10}(M_A) - (\log_{10}(M_A)) - (\log_{10}(M_A)))]$. In order to return to the non-log transformed values, for example for M_A , one simply raises the relevant term(s) to the power of 10, for example: $M_A = 10^{[(\log_{10}(M_A)) + G_{\log_{10}(M_A)}]} = (M_A)_g \circ 10^{G_{\log_{10}(M_A)}} = (M_A)_g \circ G_{M_A}$ where the “g” subscript is now used to denote the geometric mean and G_{M_A} (equal to $10^{G_{\log_{10}(M_A)}}$) represents the value by which the geometric mean must be multiplied to give the actual (model fitted) species dependent value of M_A . The plot-environmental SMA line can be reconstructed in a similar way, using the provided slope coefficient α_E , of the bivariate relationship between plot-environmental effects as in Table 2.

As there was no effect of soil fertility on the SMA slopes, for each bivariate relationship exhibiting a common scaling exponent, further tests for difference in elevation and/or shift were subsequently applied (Warton et al., 2006). Although in no case were significant differences in elevation (intercept) identified between SMA regression lines of low and high fertility oriented species, all of the $M_A \leftrightarrow [\text{nutrient}]$ and $[\text{P}] \leftrightarrow [\text{N}]$, $[\text{K}] \leftrightarrow [\text{P}]$, $[\text{Ca}] \leftrightarrow [\text{K}]$ pairs presented significant shifts along the M_A axis (Table 2). Thus species found on fertile soils are shifted towards the upper end of the M_A vs. [nutrient] scaling line, tending to have inherently lower M_A and inherently higher leaf nutrient concentrations (Fig. 4a, b). There were 33 species found at both low and high fertility sites, overlapping the two soil groups which were not included in the calculation of the above effects.

Finally for $\delta^{13}\text{C}$ there was a strong positive relationship with both [N] and [P] only and with soil fertility affecting the elevation of (rather than a shift in) the SMA regressions (Table 2). This suggests that although there is a general tendency for species with intrinsically higher [N] and [P] to also have an intrinsically lower photosynthetic discrimination against $^{13}\text{CO}_2$ (Δ), the magnitude of this effect is offset because species on the more fertile soils also have a higher Δ than those typically found under less fertile conditions.

3.5 Bivariate relationships (plot-environmental component)

The aggregated SMA regressions of the plot (environmental) component are summarised in the right panel of Table 1. Although a substantially smaller sample size was used (number of plots \ll number of species) many significant linear relationships were identified. However in some cases, relationships identified as important for the genetic component were weak for the plot level effects and vice versa. For example, although the relationships between $[\text{C}] \leftrightarrow M_A$ and $[\text{N}] \leftrightarrow M_A$ (Fig. 5a) were significant for the plot level effects and with a similar slope to the genetic level effects, this was not the case for the $[\text{P}] \leftrightarrow M_A$ (Fig. 5b) and the $[\text{K}] \leftrightarrow M_A$ pairs. This suggests that the often observed relationship between M_A and [P] does not reflect some sort of fundamental physiological imperative. The correlation between plot level [N] and [P] contribution was significant (Fig. 5c) and the same was true for the $[\text{Ca}] \leftrightarrow [\text{N}]$ pair (Fig. 5d), although for the latter no significant association was identified for the genetic component. The strongest association was found between $[\text{K}] \leftrightarrow [\text{P}]$ (Fig. 5e).

Another point of interest was the comparison of the SMA slopes, for the genetic and plot component which describe the functional relationship across and within species respectively (though note that our “residual” component must also contain some aspects of within-species variability). Out of the 17 cases where pairwise relationships were important both for the genetic and the plot component, seven had overlapping slope confidence intervals, i.e. similar slopes, and

ten had different slopes. For example the SMA slope for the $[\text{K}] \leftrightarrow [\text{P}]$ relationship was equal to 1.40, almost identical with the one estimated for the genetic component pair (1.54). On the other hand the slope for the $[\text{P}] \leftrightarrow [\text{N}]$ plot effect relationship was 2.69 as compared to 1.17 when genotype was the source of variation and with no overlapping confidence interval limits (Table 2). Clearly then, although M_A , [N] and [P] are closely linked in a consistent way when examined across different species, the relationships between M_A and [P] and between [N] and [P] for tropical trees can both be substantially modified by the environment in which they are growing.

3.6 Environmental predictors of the plot effects

The results from the ordination of the soil variables are illustrated in Table 3 with the first three PCA axes explaining 0.74 of the total variance. The first axis which accounted for 0.45 of the variance was mainly related with variations in exchangeable bases (*viz* Ca, K and Mg), total extractable P, total reserve bases (Σ_{RB}) and effective cation exchange capacity, thus reflecting variations in soil fertility (denoted φ_F). The second axis, φ_T , explained 0.18 of the variance and was mostly associated with variations in exchangeable Al and soil texture, with the third, φ_C , (accounting for 0.11 of the variance) mostly associated with variations in total soil C and N. These principal components were used as non-collinear predictor variables, along with mean annual temperature, annual total precipitation, precipitation during the dry months and incoming solar radiation in a multiple linear regression against the plot level effect as derived from the multilevel model of Eq. (1).

As is outlined in Appendix A, important environmental predictors of trait variation were identified using both ordinary regression and simultaneous autoregressive models (SAR). However, some OLS analyses (especially for [P], [Ca], [K], [Al] and $\delta^{13}\text{C}$) suggested that data normality and homoscedacity did not always occur. Thus, we utilised the rank-based Kendall's τ to evaluate the relationships between plot-level trait effects and environmental predictors (i.e. using the φ_F axis and the climatic variables) that retained the strongest significance in the OLS-SAR comparative analysis. These relationships are illustrated in Figs. 6 and 7 along with their associated τ and level of significance.

Bearing in mind that any spatial autocorrelation will lead to an overestimate of the level of significance, Fig. 6a thus suggests that the relationship showing a decline between M_A and φ_F , is weak, but that the decrease in [C] plot effect with φ_F (Fig. 6b) and the increasing [N] effect with φ_F (Fig. 6c) are both much more likely to be significant, with the very strong relationship between [P] plot effects and φ_F (Fig. 6d) even more so. Of the cations, the relationships between φ_F and both the [Ca] (Fig. 6e) and [K] plot effect (Fig. 6f) were both also quite strong, but as is shown in Fig. 6g this was much less the case for the [Mg], for which,

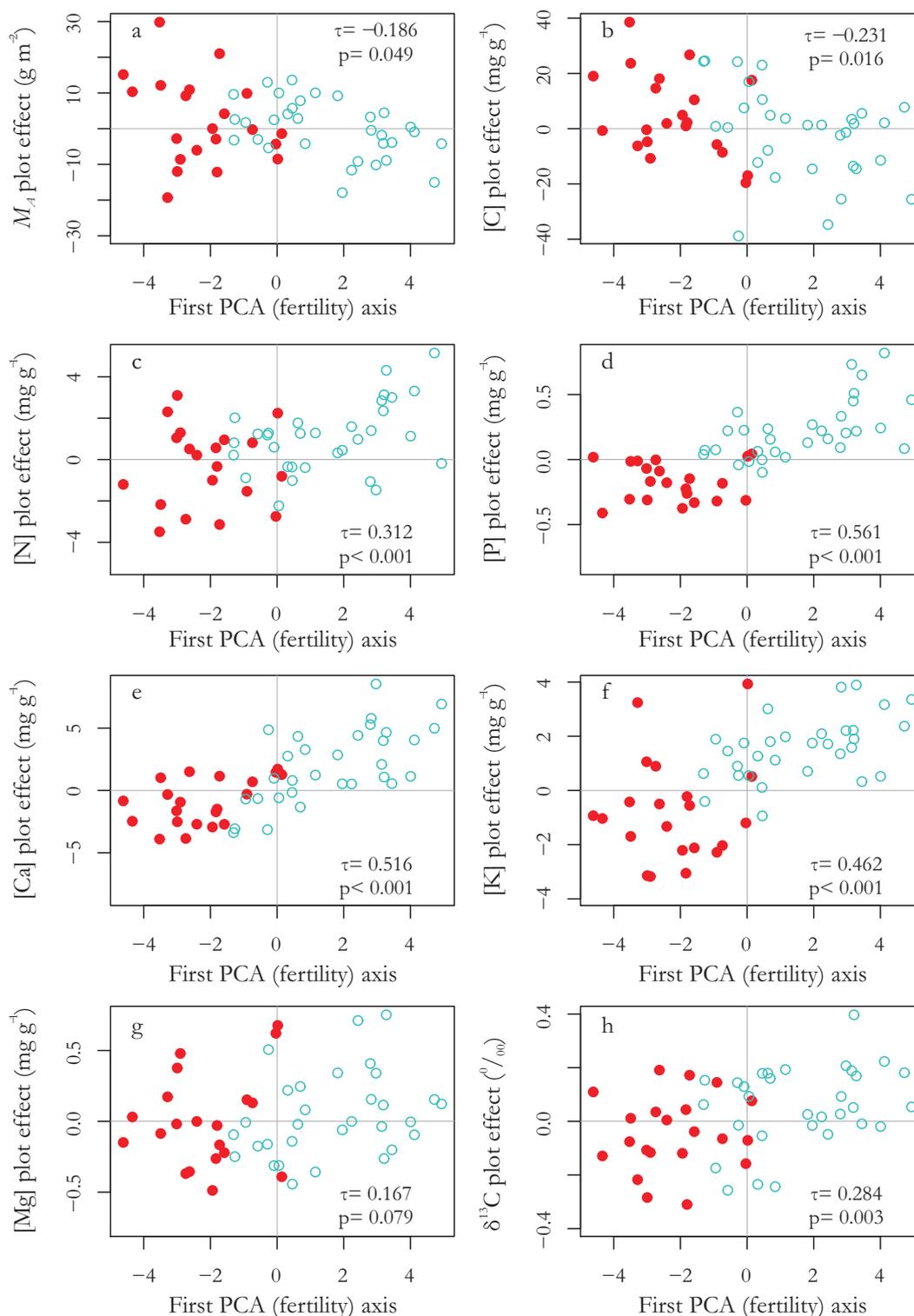


Fig. 6. Relationships between plot-environmental effects and the PCA first PCA (fertility) axis, φ_F for key foliar properties. Red dots indicate low fertility plots and blue circles indicate high fertility ones. Full Kendall's τ along with its significance is also given. M_A denotes leaf mass per unit area, $\delta^{13}\text{C}$ represents the dry matter $^{13}\text{C}/^{12}\text{C}$ composition (per mil) with all other symbols representing the elemental composition of the leaves (dry weight basis).

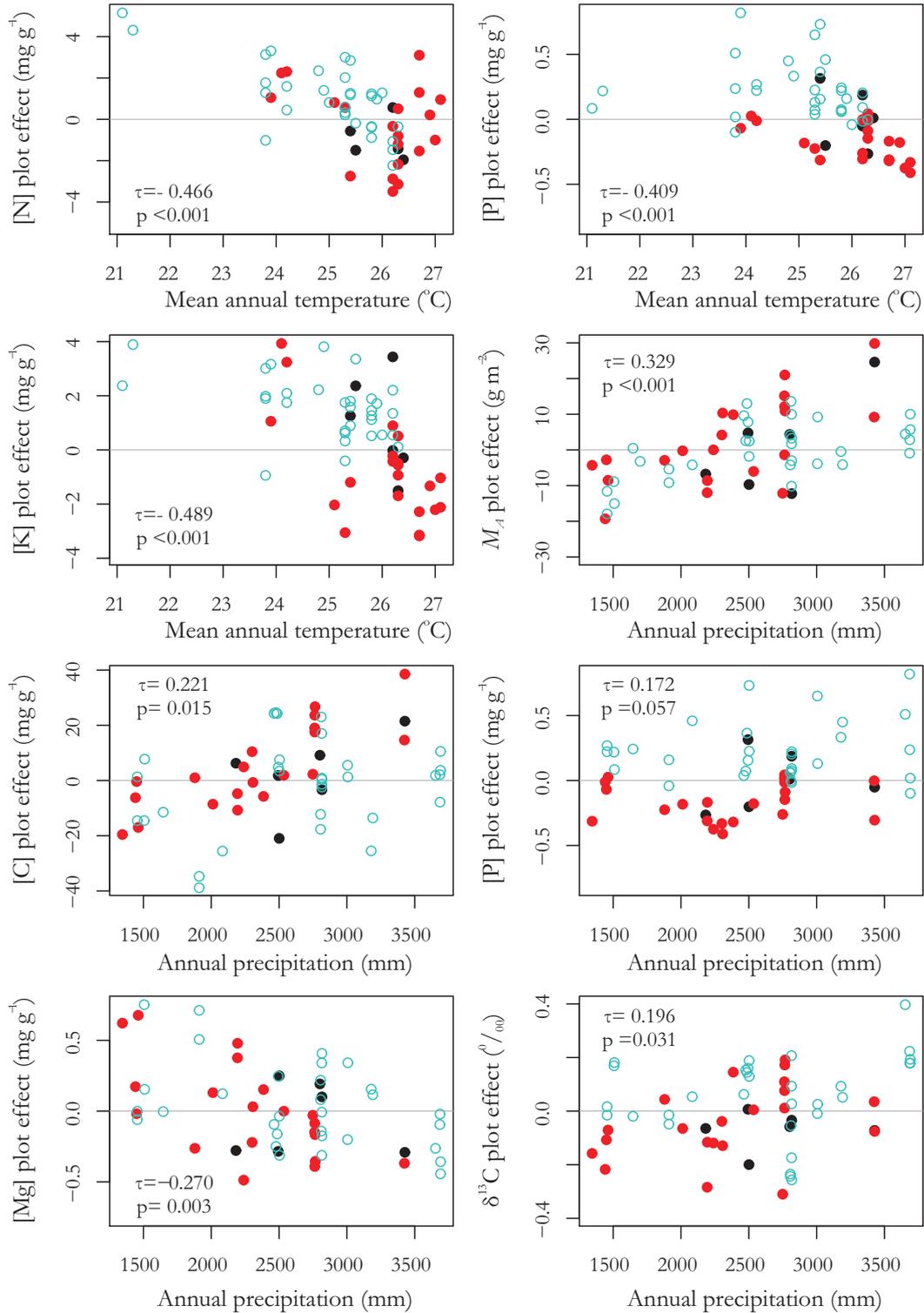


Fig. 7. Relationships between plot-environmental effects and mean annual temperature and precipitation for key foliar properties. Red dots indicate low fertility plots, blue circles indicate high fertility ones and black dots plots with no soil fertility data. Full Kendall's τ along with its significance is also given. M_A denotes leaf mass per unit area, $\delta^{13}\text{C}$ represents the dry matter $^{13}\text{C}/^{12}\text{C}$ composition (per mil) with all other symbols representing the elemental composition of the leaves (dry weight basis).

Table 1b. Table of geometric means.

	M_A (g m ⁻²)	[C] (mg g ⁻¹)	[N] (mg g ⁻¹)	[P] (mg g ⁻¹)	[Ca] (mg g ⁻¹)	[K] (mg g ⁻¹)	[Mg] (mg g ⁻¹)	[Al] (mg g ⁻¹)	$ \delta^{13}C $ (‰)
Overall geometric mean	94.85	471.53	20.69	0.91	5.48	5.38	2.26	0.044	31.33
(log ₁₀ estimate)	(1.97)	(2.67)	(1.31)	(-0.04)	(0.74)	(0.73)	(0.35)	(-1.357)	(1.50)

Table 2. Pairwise relationships between the genetic components of key foliar properties of species found in low and high fertility plots. The genetic component is computed by summing the *Family+Genus+Species* effect as estimated from the multilevel model. *Slope* of the SMA, Pearson's *r* correlation coefficient, *sig* the significance of the correlation, and *n* the number of cases used. Boldface indicates significant difference (*p* < 0.05) in slope or elevation and/or shift across the SMA axis. The (-) sign indicates that the respective test cannot be applied. sig: ***<0.001, **<0.01, *<0.05, <0.1. M_A denotes leaf mass per unit area, $\delta^{13}C$ represents the ¹³C/¹²C carbon isotopic composition with all other symbols representing the elemental composition of the leaves(dry weight basis). All values had been log₁₀ transformed prior to analysis.

		low fertility					high fertility					sig. of difference in		
log ₁₀ (y)	log ₁₀ (x)	slope	slope 95%ci	<i>r</i>	sig	<i>n</i>	slope	slope 95%ci	<i>r</i>	sig	<i>n</i>	slope	elevation	shift
[C]	M_A	0.353	(0.307 0.407)	0.228	**	188	0.392	(0.349 0.440)	0.099	-	289	0.267	0.623	< 0.001
[N]	M_A	-1.045	(-0.916 -1.192)	-0.403	***	189	-0.959	(-0.863 -1.065)	-0.407	***	293	0.313	0.994	0.003
[P]	M_A	-1.202	(-1.052 -1.373)	-0.388	***	188	-1.170	(-1.054 -1.298)	-0.433	***	290	0.751	0.525	0.001
[Ca]	M_A	-2.284	(-1.980 -2.635)	-0.134	-	188	(-)	(-)	-0.083	0.157	291	(-)	(-)	(-)
[K]	M_A	-1.616	(-1.408 -1.855)	-0.291	***	188	-1.703	(-1.527 -1.900)	-0.323	***	291	0.559	0.272	< 0.001
[Mg]	M_A	-2.082	(-1.806 -2.399)	-0.179	*	188	-2.194	(-1.957 -2.461)	-0.124	*	291	0.570	0.365	< 0.001
[Al]	M_A	(-)	(-)	0.087	0.232	190	6.775	(6.042 7.596)	0.133	*	291	(-)	(-)	(-)
$\delta^{13}C$	M_A	0.117	(0.101 0.135)	0.126	-	184	(-)	(-)	0.025	0.667	293	(-)	(-)	(-)
[N]	[C]	2.956	(2.562 3.411)	0.122	-	188	(-)	(-)	0.064	0.277	290	(-)	(-)	(-)
[P]	[C]	(-)	(-)	0.116	0.113	188	(-)	(-)	-0.086	0.146	289	(-)	(-)	(-)
[Ca]	[C]	-6.462	(-5.658 -7.381)	-0.387	***	188	-5.679	(-5.154 -6.257)	-0.546	***	290	0.123	0.182	< 0.001
[K]	[C]	-4.572	(-3.998 -5.228)	-0.368	***	188	-4.346	(-3.923 -4.814)	-0.467	***	290	0.555	0.536	< 0.001
[Mg]	[C]	-5.888	(-5.147 -6.737)	-0.359	***	188	-5.608	(-5.063 -6.212)	-0.470	***	290	0.571	0.598	< 0.001
$\delta^{13}C$	[C]	0.330	(0.286 0.381)	0.146	*	183	(-)	(-)	-0.059	0.315	290	(-)	(-)	(-)
[P]	[N]	1.150	(1.030 1.284)	0.648	***	188	1.217	(1.120 1.323)	0.698	***	289	0.419	0.387	0.002
[Ca]	[N]	(-)	(-)	-0.015	0.838	188	(-)	(-)	0.090	0.127	290	(-)	(-)	(-)
[K]	[N]	1.547	(1.340 1.784)	0.127	-	188	1.767	(1.579 1.978)	0.230	***	290	0.151	0.318	< 0.001
[Mg]	[N]	(-)	(-)	-0.055	0.454	188	2.280	(2.032 2.559)	0.103	-	290	(-)	(-)	(-)
$\delta^{13}C$	[N]	0.111	(0.097 0.128)	0.277	***	184	0.101	(0.090 0.113)	0.240	***	293	0.263	0.008	0.221
[Ca]	[P]	(-)	(-)	0.046	0.530	188	1.896	(1.694 2.121)	0.229	***	291	(-)	(-)	(-)
[K]	[P]	1.345	(1.178 1.535)	0.394	***	188	1.441	(1.305 1.592)	0.507	***	291	0.412	0.524	< 0.001
[Mg]	[P]	(-)	(-)	0.067	0.363	188	1.858	(1.661 2.079)	0.242	***	291	(-)	(-)	(-)
$\delta^{13}C$	[P]	-0.099	(-0.087 -0.114)	-0.349	***	183	-0.082	(-0.073 -0.092)	-0.242	***	290	0.033	(-)	(-)
[Mg]	[Ca]	0.911	(0.816 1.018)	0.643	***	188	0.987	(0.905 1.076)	0.660	***	292	0.266	0.311	< 0.001
$\delta^{13}C$	[Ca]	(-)	(-)	0.095	0.201	183	0.043	(0.038 0.048)	0.223	***	291	(-)	(-)	(-)
[Ca]	[K]	1.413	(1.242 1.608)	0.447	***	188	1.307	(1.179 1.449)	0.450	***	292	0.352	0.493	< 0.001
[Mg]	[K]	1.288	(1.140 1.455)	0.534	***	188	1.290	(1.175 1.415)	0.591	***	292	0.987	0.888	< 0.001
$\delta^{13}C$	[K]	(-)	(-)	0.106	0.154	183	(-)	(-)	0.085	0.150	291	(-)	(-)	(-)
$\delta^{13}C$	[Mg]	(-)	(-)	0.065	0.386	183	0.044	(0.039 0.049)	0.146	*	291	(-)	(-)	(-)

after taking into account the likely spatial autocorrelation discussed in Appendix A, is probably not significantly related to φ_F . Likewise for $\delta^{13}C$ the relationship with φ_F was relatively weak, though still of interest.

In a similar manner, Fig. 7 illustrates the strongest relationships found between the various plot effects and climate. Three likely significant plot effects related to mean annual temperature (T_a) were observed, namely a decline in leaf nitrogen (Fig. 7a), leaf phosphorus (Fig. 7b) and K (Fig. 7c) with increasing temperature. Figure 7d–h shows the most important relationships with mean annual precipi-

tation, P_a , with a strong positive relationship observed with M_A (Fig. 7d), [C] (Fig. 7e) and a negative slope for [Mg] (Fig. 7g). Figure 7f and h shows less dramatic, though potentially important relationships with precipitation for the [P] and $\delta^{13}C$ plot effects respectively.

Table 4 lists Kendall's partial rank coefficients (τ_P) for the various plot effects examined as related to φ_F , φ_T , T_a , P_a and mean annual radiation, Q_a . In all cases the coefficient given is for the one factor after controlling for the other four. Based on our numerical simulations of the partial τ sampling distribution quantiles and the issue of spatial autocorrelation

Table 3. Summary of the Principal Components Analysis for selected soil variables averaged over 0.0–0.3 m depth. Full details on methods used for soil analysis are provided in Quesada et al. (2009c).

Eigenvalue	6.227	2.537	1.597
Proportion of total variance	0.445	0.181	0.114
pH	–0.278	0.371	–0.082
log ₁₀ [total reserve bases]	– 0.328	–0.072	0.050
log ₁₀ [total extractable P] ^a	– 0.340	–0.103	–0.148
log ₁₀ [total N]	–0.290	–0.095	– 0.514
log ₁₀ [total C]	–0.169	–0.044	– 0.667
log ₁₀ [exchangeable Ca]	– 0.360	0.162	0.103
log ₁₀ [exchangeable Mg]	– 0.367	0.083	0.202
log ₁₀ [exchangeable K]	–0.286	–0.100	0.134
log ₁₀ [exchangeable Na]	–0.140	–0.135	–0.022
log ₁₀ [exchangeable Al]	0.161	– 0.359	–0.137
log ₁₀ (effective cation exchange capacity)	– 0.329	–0.202	0.109
log ₁₀ (base saturation)	–0.204	0.410	0.237
fraction sand	0.199	0.442	–0.261
fraction clay	–0.077	– 0.491	0.187
Axis definition	(in)fertility	texture & Al	C&N

^a Sum of both inorganic and organic fractions extracted by resin, bicarbonate and NaOH according to a modified Hedley extraction procedure as detailed in Quesada et al. (2009c).

Table 4. Kendall's partial correlation τ for the environmental contribution (plot effect estimate) of each foliar property with the set of environmental predictors. Kendall's τ are estimated as described in Legendre and Legendre (1998, pp. 202). Their significance is computed based on Maghsoodloo and Laszlo Pallos (1981). Bold values indicate a very strong correlation ($p < 0.001$) and italics indicate significant correlations at $p < 0.01$; see text for details. M_A denotes leaf mass per unit area, $\delta^{13}\text{C}$ represents the $^{13}\text{C}/^{12}\text{C}$ composition (per mil) with all other symbols representing the elemental composition of the leaves (dry weight basis).

	M_A	[C]	[N]	[P]	[Ca]	[K]	[Mg]	[Al]	$\delta^{13}\text{C}$
Fertility φ_F	–0.201	–0.233	0.204	0.475	0.475	0.337	0.220	0.097	0.203
Texture φ_T	0.048	0.103	0.115	0.043	– 0.272	–0.169	–0.178	–0.149	0.022
Temperature T_a	0.107	0.051	– 0.382	–0.256	–0.081	– 0.408	0.031	–0.176	–0.133
Precipitation P_a	0.329	0.300	–0.178	0.171	–0.010	0.113	– 0.306	0.016	<i>0.244</i>
Radiation Q_a	–0.058	0.152	0.018	0.117	–0.140	0.075	0.003	–0.139	0.119

discussed above, we suggest that $\tau_P > 0.23$ (approximately relating to the probability of a Type II error, P , being less than 0.01) should be taken as a minimum criterion for statistical significance with $\tau_P > 0.31$ almost certainly indicating a meaningful correlative relationship ($P < 0.001$). Nevertheless, relationships with $0.17 < \tau_P < 0.23$ cannot be entirely discounted.

Table 4 shows that very strongly related to φ_F were the plot effects for [P], [Ca] and [K], and, with the exception of [Al], the other parameters examined also had close to significant relationships with M_A , [C] declining somewhat with increased fertility and $\delta^{13}\text{C}$ increasing. On the other hand, only the [Ca] plot effect was significantly associated with φ_T although all other cations (including [Al]) did show trends in the same direction, *viz* a decline with increasing

soil sandiness and/or higher soil pH. Of the climatic parameters, Q_a , showed no meaningful associations, but [N], [P] and [K] were all negatively associated with T_a . There were several strong relationships detected with P_a ; in particular an increase in both M_A and [C] with increasing precipitation, and with significant declines in [Mg] and less negative $\delta^{13}\text{C}$ as P_a increased.

4 Discussion

Suites of plant traits are often used to infer the functioning and performance of different species (Westoby et al., 2002) as well as to identify potential evolutionary pathways of trait variation (Reich et al., 2003; Ackerly, 2003). Among the

range of functional characters some are regarded as highly conservative while others are considered more plastic. Identifying the variation of different functional traits among and within different evolutionary affinities and exploring the way these plant functional characteristics vary across environmental gradients should help us to understand the functioning of different ecosystems and their responses to global change.

4.1 Genetic and environmental variation in leaf foliar properties of Amazon trees

Analyses of global (Wright et al., 2004) and regional (Fonseca et al., 2000; Wright et al., 2001) datasets have highlighted that a large portion of the variation observed in M_A , [N] and [P] is found between species within a common environment. Data from north-west Amazonia presented a thirty-fold variation in leaf longevity and a three-fold variation in M_A and [N] in adjacent communities (Reich et al., 1991) with Townsend et al. (2007) highlighting the importance of local-scale species diversity as an important component controlling the variation of foliar N:P ratio. Our results place a special emphasis on this variability, covering considerable tree species diversity across an ecosystem-wide climate and soil gradient. A ten-fold range of variation was identified for M_A , covering a similar range (30 to 300 g m⁻²) as a recent worldwide review (Poorter et al., 2009). At the same time [N] and [P] presented a six-fold and a seventeen-fold range respectively. Significant differences in average M_A , [N] and [P] exist between many families as well as varying with genera within several of the more widely abundant ones (Figs. B1 and B2).

In our dataset some properties like M_A , and leaf [C], [N], [Mg] and [Al] seem to be more strongly genetically than environmentally constrained, with much of their variation attributable to their phylogenetic grouping (Fig. 2). For example approximately 0.07, 0.11 and 0.19 of the total variation in M_A , is apportioned to the family, genus and species component respectively (i.e. 0.37 of the total variance is attributable to genotype) and only 0.15 is due to the location of an individual. At the other end, traits such as [P], [Ca] and [K] had a much higher level of plasticity, with 0.47, 0.41 and 0.47 of the respective variation being attributed to the site at which the leaf was measured, and thus suggesting a strong environmental control.

For the cations, significant genetic variation has been observed before for both herbaceous (Thompson et al., 1997) and woody (Dauer et al., 2007) plant communities (see also Broadley et al., 2004). Much of the genetically controlled variation in [Ca] and [Mg] seems to be mediated through differences in cation exchange capacities of cell walls (Demarty et al., 1984; White and Broadley, 2003), which are themselves strongly influenced by genetic variations in pectin and galacturonic acid chemistry (Kirkby and Pilbeam, 1984; Sattelmacher, 2001). Nevertheless, even though the uptake of both cations is clearly genetically controlled and with some

evidence of similar transport mechanisms (Broadley et al., 2008), Mg²⁺-specific transport genes also exist (Gardner, 2003). This is consistent with the much greater plot effect for calcium observed here (Fig. 2) with Mg²⁺-uptake likely subject to a much tighter physiological regulation via symplastic transport pathways and hence the relatively lower environmental effect.

It is also intuitive that should an element be limiting for ecosystem function then foliar concentrations should show a strong correlation with variations in its availability within the soil and with the associated environmental component of variation likely being high compared to any genetic component. In this respect the strong environmental components observed for [P], [Ca] and [K] in Fig. 2 and their strong correlation with the fertility axis, φ_F (Fig. 6), are especially informative. It has, of course, long been argued that phosphorus may be the main element limiting tropical forest productivity (Vitousek, 1984) and analysing the relationship between above-ground growth rates and various soil fertility measures, Quesada et al. (2009b) have also found good evidence supporting this critical role for phosphorus in modulating tropical forest productivity. Nevertheless, from the data presented here it is clear that foliar potassium in particular seems to always correlate strongly with foliar phosphorus (Fig. 5), this also being observed for the genetically dependent relationship with a virtually identical slope (Fig. 4).

4.2 Scaling relationships and species distributions across the Amazon

Scaling relationships were identified between all $M_A \leftrightarrow$ [nutrient] raw data pairs with the only exception being the $M_A \leftrightarrow$ [Al] pair (Table 1). Additionally for the genetic $M_A \leftrightarrow$ [nutrient] pairs common slope SMA axes were identified, indicating similar scaling mechanisms regardless of edaphic conditions (Table 2). Thus at this level, general axes of leaf traits variation expressing the economic spectrum of fast and slow resource turnover are indeed valid for Amazonian forests (Table 1, middle panel).

However there is a clear distinction in the inherent physiological ranges that low and high fertility oriented species operate (Paoli, 2006; Kraft et al., 2008) and this is illustrated by the significant shifts across the common axis of variation identified for the genetic component (Table 2). Thus species which are found on fertile soils tend to be at the fast return (right hand) side of the intrinsic $M_A \leftrightarrow$ [nutrient] continuum (Fig. 3). In the raw dataset this distinction was expressed both with significant difference in the intercept and/or shifts of the principal axis or even more in some cases with difference is the SMA slope (Table S12). This was also shown for [N] vs. [P] by Townsend et al. (2007). Shifts in the intercept of the raw data SMA seem to be controlled by the environmental contribution of trait variation, as no such difference was identified in the genetic scaling. Thus the environment

within which a plant is growing clearly affects the nature of the $M_A \leftrightarrow [\text{nutrient}]$ relationship.

Across the Amazon Basin there are two well-recorded gradients of resource availability, namely a northeast to southwest gradient of soil fertility (Quesada et al., 2009b) and a northwest to southeast gradient in precipitation and dry season length (Sombroek, 2001; Malhi and Wright, 2004). Tree species distributions (ter Steege et al., 2006), diversity (ter Steege et al., 2006) and forest turnover (Phillips et al., 2004) all follow these gradients, with western forests being more dynamic and with faster growth rates (Malhi et al., 2004; Phillips et al., 2004; Quesada et al., 2009b). Results here suggest that these gradients are reflected in foliar characteristics as well as other functional properties (Patiño et al., in preparation; Baker et al., 2009). Specifically, species on richer soils tend to have intrinsically lower M_A , and intrinsically higher leaf nutrient concentrations compared with species on poor soils. Of special interest are the identified shifts across the principal axis of genetic variation, supporting the “habitat tracking” hypothesis (Ackerly, 2003). We suggest that specific trait dimensions systematically change along soil fertility gradients.

Our sampling strategy and subsequent analyses were not designed to specifically explore niche separation mechanisms occurring across the RAINFOR plots. Nevertheless, the clear shift in the genetic component of foliar traits associated with rich or poor soils implies that soil fertility exerts a fundamental role in modulating community composition across Amazonia. As is also discussed in Fyllas and Lloyd (2009), it seems potential resourced-based tree niche differentiation processes are accompanied by specific suites of functional foliar and other properties. The unbalanced nature of our data-sampling with more than half of the species (303) being measured only once, is unlikely to have given rise to biased species effect estimates driven by the environmental conditions of the site (Supplementary Information II: <http://www.biogeosciences.net/6/2677/2009/bg-6-2677-2009-supplement.zip>). However, these correlations could have arisen through a small number of ancient events, and subsequent diversification from common ancestors that colonised rich and poor soils respectively. Alternatively, habitat specialisation may have repeatedly driven diversification in a large range of clades, such as in the Burseraeae (Fine et al., 2005). Distinguishing these hypotheses will require mapping the genetic component of species traits onto their phylogenetic relationships and assessing whether the genetic component of these trait values show overdispersed or clustered distributions (Cavendar-Bares et al., 2006).

4.3 Environmental predictors of foliage properties variation

Although there have been several previous studies investigating effects of climate and/or soil conditions on foliar trait

combinations (e.g. Fonseca et al., 2000; Wright et al., 2004, 2005a; Townsend et al., 2007) all of these have treated the measured values of the traits examined as being genetically determined, often pooling several values into a single set of values for the one species. We have taken a different approach here, allowing for the traits observed for any given species to vary with location by utilising the REML methodology. A similar statistical approach has been independently undertaken by Watanabe et al. (2007) in an analysis of evolutionary controls of plant nutrient composition. They fitted a REML model of a similar form, also using random terms only (in their case “site + clade/family/species”), though in their study they were more interested in partialing out any site effects, rather than, as in our case, trying to quantify and understand them. Our approach of quantifying both the genetic and environmental components through REML is a valid one despite the strongly unbalanced nature of the data set employed (see Supplementary Information II) and occurs because about half of the species in the dataset occurred in more than one plot. This species/plot overlap allows for a direct estimate of plot-environmental effects as well as estimates of the genetic effects of these species occurring in more than one plot. These plot effect estimates, once subtracted from the raw observations, then allow for “genetic effects” to be estimated for those species occurring only once in the dataset. A nested genetic variance structure used within a REML context to analyse the genetic variations observed (in our case “family/genus/species”) has previously been recognised by other workers and extensively applied in phylogenetic analyses (Broadley et al., 2004).

By combining the non-spatial and spatially explicit regression models of the plot-level effects and then using this information to help us interpret the partial Kendall’s τ , soil fertility emerges as a key axis of association between leaf level nutrient variation and environmental factors. Soil fertility was positively related with leaf nutrient concentrations and negatively associated with M_A and with leaf [C] (Fig. 6). Based on studies with soil nutrients and/or investigations on the effects of variations in soil fertility on tropical tree leaf nutrient concentrations (Montagini, 2000; Webb et al., 2000; Specht and Turner, 2006), increased foliar nutrient concentrations for any given species on more fertile soils are not all that surprising. What is more interesting is the coordinated response of M_A and [C]. Clearly for any given species considerable plasticity exists. Leaf structure and physiology can vary together with the wider ranging species investing fewer resources into structural carbon and M_A under high nutrient conditions.

Plot level variation in M_A and leaf [C] also showed a strong positive trend with increasing precipitation, with a significant decline in leaf [Mg] also observed (Fig. 7). This increase in M_A with increased moisture availability contrasts with the generally accepted pattern for evergreen species where M_A declines with increasing rainfall (Wright et al., 2005a). Nevertheless, it might be explicable on the basis

of higher M_A leaves being structurally more robust and thus more resistant to the increased intensities of pathogen and herbivore attack which would be expected in the less seasonal and higher rainfall environments. Consistent with this idea is the increase in [C] also observed with increasing precipitation, pointing to an increased investment of carbon in constitutive compounds, as has been reported, for example to occur for lignin and phenolics (though in this case across different life forms and with a reduction rather than an increase in M_A) in northern Patagonia (Bertiller et al., 2006).

More difficult to explain is the observed decrease in [Mg] with increasing precipitation. Perhaps there are changes in cell wall carbohydrate chemistry associated with increased foliar defences (Vorwerk et al., 2004) at higher precipitation sites. This would likely be achieved through changes in cell wall structure as mediated, for example, by pectin methylesterase (Pelloux et al., 2007). If so, such differences would be expected to give rise to substantial changes in cell wall ionic properties and, in particular, associated changes in foliar magnesium contents (Pilling et al., 2004).

It is also interesting to note that the three foliar properties showing significant changes with precipitation, viz M_A , [C] and [Mg], are also those for which the overall environmental contribution to the observed variance were all but the lowest, but with a substantial genetic contribution to the variation observed (Fig. 2). This suggests that rather than being directly caused by precipitation per se, the “precipitation effect” as calculated might actually reflect phenotypically distinct populations. Such intraspecific variation has been observed before for woody species. For example, in a “common garden” experiment using fifteen distinct populations of red ironbark (*Eucalyptus sideroxylon* subsp. *tricarpa*) in Australia, Warren et al. (2005) found considerable between inter-population variability (i.e. phenotypic plasticity) for both M_A and [N] and with some of this variation attributable to seed-source precipitation regime. Despite the notoriously high biodiversity of Amazon forest (Hubbell et al., 2008) there is no reason why such intra-specific variation should not occur for at least some of the species growing there, as has been shown for different populations of Costa Rican *Cedrela odorata* for example (Gillies et al., 1997; Navarro et al., 2002).

Environmental effects on leaf N concentration were negatively related with annual mean temperature (Fig. 7). This result is in agreement with the global scale prediction of leaf nitrogen decreasing with temperature (Reich and Oleskyn, 2004), supporting the “Temperature-Plant Physiology Hypothesis” (Woods et al., 2003) which argues that plants at lower temperatures should show higher concentrations of physiologically relevant compounds in order to compensate for repressed rates of fundamental biochemical processes. Even after removing the genetic N component of species adapted and established to more fertile environments and accounting for the potential existence of spatial autocorrelation phenomena, the negative effect of temperature on foliar [N] remained highly significant (Table A1) as was also the case

for [P] from the partial Kendall’s τ analysis (Table 4).

4.4 Amazonian axes of trait coordination in a global context

Concentrating on the genetic component, it is interesting to compare the slopes of the bivariate relationships we have observed with those proposed to operate globally (Wright et al., 2004). Here we note that for the (aggregated) genetic component of trait variation we observed [N] \leftrightarrow M_A slopes (with confidence intervals in brackets) of -0.99 (-0.91 to -1.07) slightly steeper than that of Wright et al. (2004), who (taking reciprocal values from the $M_A \leftrightarrow$ [N] slopes, their Table 1) reported a value of -0.78 (-0.76 to -0.81). If we accept that the negative [N] \leftrightarrow M_A scaling relationship occurs because higher M_A leaves invest a greater proportion of their biomass in structural rather than metabolic compartments (Reich et al., 1999), then this suggests that higher M_A tropical tree species may retain a lesser amount of nitrogen in structural compartments such as cell walls. It is well known that cell wall N contents can vary substantially (Lampert, 1965; Takashima et al., 2004) with several classes of cell wall protein with an important role in disease resistance (Showalter, 1993; de Bruxelles and Roberts, 2001). It is, however less likely that this nitrogen-based defensive strategy, would be employed for species characteristic of the more infertile soils (including white sands) as some evidence for N-deficiency exists (Quesada et al., 2009c). According to the Carbon: Nutrient Balance Hypothesis (CNB: Bryant et al., 1993), C-based constitutive defences would be expected to be more prevalent as has shown to be the case by Fine et al. (2006). Consistent with this idea is the significant positive [C] \leftrightarrow M_A relationship observed for species found on low fertility soils. This may be accountable for in terms of high M_A leaves having a greater investment in carbon based defensive compounds such as phenols, lignin and tannin, all of which have a relatively high carbon content (Poorter and Villar, 1997). A critical discussion of the CNB and other relevant plant defence hypotheses as related to plant genotypic characteristics and edaphic limitations may be found in Stamp (2003).

In contrast to [N] \leftrightarrow M_A our genetic scaling slopes for [P] \leftrightarrow M_A , of -1.17 (-1.08 to -1.27) are all but identical to the Wright et al. (2004) global estimate of -1.22 (-1.16 to -1.28) suggesting stronger similarities between Amazon forest trees and other terrestrial plants. The steeper negative slope for the [P] \leftrightarrow M_A relationship as compared to that for [N] \leftrightarrow M_A may be explained by the presence of still appreciable amounts of N, but not P, in structural tissues such as cell walls (Showalter, 1993; Gabriel and Kesselmeir, 1999; White and Hammond, 2008), the proportion of which should generally increase as M_A increases. Also important may be the ability for low P requiring species to exhibit a more conservative use of phosphorus in their metabolically active tissues, for example, through employing organic acids rather than

inorganic phosphate to maintain ionic balance in the vacuole, and through substitution of phospholipids with galactolipids and sulpholipids in thylakoid and extraplastidic membranes (Amtmann et al., 2006; White and Hammond, 2008).

Research quantifying relationships between metabolically active tissue properties, for example [N] and [P], and linking these with the way annual growth rate scales with plant mass, has received some attention in recent years, with modellers in this area attempting to provide a modelling framework to predict the way trees accumulate resources (Niklas et al., 2005; Niklas, 2006, 2008). Recent modelling exercises (Kerkhoff et al., 2005, 2006; Kerkhoff and Enquist, 2006) are based on a perceived strong association between leaf [N] and [P], according to a 2/3 (Wright et al., 2004) or 3/4 power law (Niklas et al., 2005). Whilst not necessarily endorsing the generality of such exercises, it is of some interest to see how our observations fit with the conceptual constructs. Our results verify the strong [N]↔[P] relationship, on both fertile and infertile Amazonian soils. This scaling relationship was significant both in the raw data analyses (Tables 1 and SI2) and in the analyses of the multilevel effect estimates (Tables 1 and 2). The aggregated raw data estimate for the [N]↔[P] scaling exponent is (0.58–0.64), whereas, as was similarly found by Townsend et al. (2007), for low fertility sites the estimated N:P slope (in our case 0.77–0.93) is clearly higher than for high fertility sites (0.63–0.73). Scaling of the genetic component of leaf [N]↔[P] is calculated at (0.78–0.97) and (0.76–0.89) for species found on low and high fertility sites respectively with Table 1 suggesting a value of 0.85 (0.80, 0.91) for the population as a whole. Thus although the raw data analyses, with environmental effects contribution included, seem reasonably close to the 2/3 power law, the analyses of the genetic component clearly show that the 2/3 power law is not adequate for expressing the N:P scaling of Amazonian trees. Indeed, the exponent seems to even be a bit larger than 3/4. The above estimates, in conjunction with the large portion of [P] variance attributed to the plot level (Fig. 2), underline the importance of soil conditions in influencing N:P ratios as is discussed further below.

In addition to [N] and [P] we also examined the relationships of [Ca], [K], [Mg] and [Al] with M_A as well as relationships between the various foliar nutrient concentrations, again separating genetic versus environmental effects. Considering genetic effects first: As was reported by Wright et al. (2005b) we found a reasonably strong relationship ($r=0.45$) between [K] and [P] but our slope of 1.40 was nearly twice as high as their slope of 0.78. This is probably due to the relatively low [P] in Amazon forest leaves (Lloyd et al., 2009b), meaning that phosphate ions are less often employed as reserve anions in the vacuole (where they would be normally balanced by K^+ and other cations) and with a greater proportion of foliar P assigned to the photosynthetic apparatus than is usually the case (White and Hammond, 2008). This would also mean that [K] should also scale with

[P] with a higher exponent than is generally observed. Notably the [K]↔[N] genetic relationship was markedly less strong ($r = 0.178$), although our slope of 1.65 was closer to the 1.19 reported in Wright et al. (2005b) than was the case for phosphorus.

The very strong genetic [Ca]↔[Mg] association observed here has also been observed on other studies (Thompson et al., 1997; Broadley et al., 2004) and may be attributable to the chemical similarities between these two divalent cations and a general lack of selectivity during cation uptake by plants (White, 2001; Broadley et al., 2004). It is also likely that these two cations share, to a large extent, the same membrane transporters (Broadley et al., 2008). There were also very strong negative relationships between the concentration of [Ca], [Mg] and [K] with [C], with significant shifts existing between low and high fertility plants. This has been observed before for a range of species (Poorter and de Jong, 1999) and may reflect an underappreciated dimension of the leaf economic spectrum, which reflects a continuum of strategies for leaf construction, ranging from the use of relatively cheap components (minerals) to more expensive carbon based constituents such as lignin. The latter strategy would also likely be associated with increased carbon based defences and other factors associated with long leaf durability such as a high M_A .

Strong genetic based relationships were also seen between $\delta^{13}C$ and [N] and, to a stronger extent with foliar [P] (Table 1). It now seems clear that either N or P can limit photosynthetic metabolism in tropical tree species (Domingues et al., 2009) and this lower carbon isotope discrimination for the higher nutrient species suggesting, on average, a lower partial pressure of CO_2 in their chloroplasts (Farquhar et al., 1989). This indicates that higher photosynthetic capacities of species characterised by intrinsically high nutrient concentrations are not totally balanced by higher stomatal conductances (Farquhar and Sharkey, 1982), perhaps suggesting form of hydraulic constraint on maximum photosynthetic rates of tropical tree species, even in the absence of soil moisture deficits (Santiago et al., 2004b; Patiño et al., 2009)

5 Conclusions

Taken together our results highlight three important points regarding the biogeochemistry of the Amazon basin, as expressed through a set of key tree foliar properties. First, there exists a substantial variability at most levels of the evolutionary grouping of species. This variability depends on the studied foliar property: Some leaf traits are more phylogenetically constrained than others; traits such as foliar [P] showing strong associations with growing conditions and are possibly linked to variations in stand-level productivity. Second, these environmental effects on leaf level nutrient concentrations make the use of general scaling relationships difficult within the Amazon basin, especially if the soil fertility

variations are not implicitly taken into account. Nevertheless a third point is that the strong genetic correlations between M_A and leaf nutrient concentrations underlines the general existence of the “leaf economic spectrum” across the diverse group of Amazonian tree species studied. Although varying along the same trait coordination axes, Amazonian trees also clearly associate with the soil conditions most appropriate for their growth. Species are not randomly distributed across the basin. But, rather, they follow distribution patterns based on an association between genetic trait potential and the availability of environmental resources.

Appendix A

Multiple linear regressions and autoregressive models analysis of relationships between plot-environmental effect terms and environmental variables

To deal with spatial autocorrelation issues in the dataset, we explored the results of the multiple linear regressions through the inspection of correlograms and estimation of the global Moran's I (Legendre and Legendre, 1998). Here the presence of spatial autocorrelation could simply reflect a patterned functioning of the plot level effects, i.e. nearby plots contributing similarly to the variation observed. Such a spatially structured environmental effect could arise as a consequence of basin-wide gradients in soil age and fertility (Quezada et al., 2009a, c) and/or precipitation (Malhi and Wright, 2004). Alternatively, it could be driven by species distribution patterns, with a species specific (or at least a functional group specific) effect modulating plot effect response. These two effects point at exogenous or endogenous forces respectively (Legendre and Legendre, 1993), either of which has the potential to give rise to a violation of the assumption of independently distributed errors. This leads to an overestimation of the relevant degrees of freedom as well as to an overestimation of the importance of the environmental variables included in any such analyses (Lennon, 2000). Interpretation of the macroecological patterns through statistical analyses where spatial autocorrelation is present is an issue of active debate (Lennon, 2000; Diniz-Filho et al., 2003), with recent simulations showing that the existence of autocorrelated residuals may not seriously affect parameter estimates (Hawkins et al., 2007).

To address the above issues, in addition to the non-spatially explicit linear models (OLS), we also fitted two simultaneous autoregressive models (SAR) including a spatial error term at a “fine scale” and at a “medium scale” (Lichstein et al., 2002; Kissling and Carl, 2008). Previous spatial analyses of ecological datasets have revealed that in some cases small neighbourhood distances (first-order) SAR models are able to remove the spatial autocorrelation signal (Jetz and Rahbek, 2002), while other studies suggest that larger neighbourhood (higher-order) SARs should be used

(Tognelli and Kelt, 2004). The usual practice for selecting the neighbourhood size in SAR models is to identify, through the inspection of the OLS residuals, the maximum distance of a significant autocorrelation signal and use this distance as a neighbourhood size (Lichstein et al., 2002; Hawkins et al., 2007). Our data revealed discrete maximum distances of spatial signal for each plot effect (determined from the correlograms of the OLS residuals off each parameter of interest) and we thus used a variant neighbourhood size for the medium scale SARs and a common (50 km) scale for fine scale SARs. We checked the value of the autoregression coefficient (λ) in the SAR models; Specifically, if λ was found to be significantly different from zero after controlling for the environmental effect, then the autoregressive component (i.e. the neighborhood effect) was deemed important. Furthermore, by inspecting the SAR residual correlograms we identified their ability to remove spatial autocorrelation. At the same time we used a Monte Carlo (999) permutation test for the significance of Moran's I for the initial plot effect estimates and the residuals from the OLS and SAR models. In all cases coefficients for ten distance classes of equal widths (200 km) are reported – at this distance all classes have an approximately balanced (more than 100) number of pairs (Legendre and Legendre, 1998). For correlograms the local significance of Moran's I at the j -th class were corrected with a progressive Bonferroni procedure ($\alpha^*=a/j$, with α the original significance level set at $\alpha=0.05$), while the adjusted significance level for the global Moran's I was 0.005 (0.05/10 reflecting the ten distance classes used) for which more information is available in Legendre and Legendre (1998) and Lichstein et al. (2002). The above were implemented with the R libraries *ncf* (Bjornstad and Falck, 2001) and *spdep* (Bivand, 2006).

All foliar plot effect estimates had an important spatial pattern as illustrated in the respective maps and tested with the Monte Carlo permutation method at equally distant classes of 200km, and a global Bonferroni significance level α^* of 0.005 (Fig. SIII1: <http://www.biogeosciences.net/6/2677/2009/bg-6-2677-2009-supplement.zip>). This means that spatial autocorrelation needs to be taken into account in any analysis. The results of the non-spatial multiple OLS regression analyses are summarized in the top panel of Table A1, and in the second and third panel we present the results of the SAR models. Correlograms of residuals of all three models are shown in Fig. SIII 2 and the comparative consideration of these models gives some insights to the potential patterns of spatial autocorrelation of each foliar property of interest.

Both the OLS model residuals correlogram (Fig. SIII 2a) and the permutation method (Table A1 – OLS panel – “Moran's I ”) did not identify an important autocorrelation signal in the non-spatial regression of M_A , with the OLS accounting for 0.42 of the variation in the plot effect M_A contribution. The fertility PCA axis, φ_F , and total annual precipitation were the most important environmental predictors.

Table A1. Coefficient estimates from non-spatially multiple linear regressions (OLS), simultaneous autoregressive models at a common fine scale (SAR FS, FS=50 km) and at variant medium scale (SAR MS), for each foliar property on the set of the environmental predictors. FS SARs had a common neighbourhood size of 50 km, while the MS SARs were at 750 km for [C], 1000 km for [P] and $\delta^{13}\text{C}$, 1750 km for [Ca] and [K] and 1900 km for [Al]. See text and Fig. SIII 2 (<http://www.biogeosciences.net/6/2677/2009/bg-6-2677-2009-supplement.zip>) for selection of appropriate neighbourhood size. λ gives the autoregression coefficient for each SAR (boldface indicate its significant difference from 0, at $p=0.05$). Moran's I for each model's residuals tested at a global Bonferroni adjusted level (0.005), with 999 Monte Carlo permutations (bold values indicate the existence of spatial autocorrelation in the residuals). Significance levels for environmental predictors estimates: ***<0.001, **<0.01, *<0.05, -<0.1 and (-) non significant. M_A denotes leaf mass per unit area, $\delta^{13}\text{C}$ represents the $^{13}\text{C}/^{12}\text{C}$ composition (per mil) with all other symbols representing the elemental composition of the leaves (dry weight basis). All values had been \log_{10} transformed prior to analysis.

OLS	M_A	[C]	[N]	[P]	[Ca]	[K]	[Mg]	[Al]	$\delta^{13}\text{C}$
Intercept	-9.54E-02 ***	-5.78E-02 **	4.39E-01 ***	-3.09E-01 *	1.63E+00 *	7.34E-01 (-)	1.16E-01 ***	3.09E+00 *	3.78E-02 (-)
Fertility, φ_F	-1.53E-02 **	-5.98E-03 ***	1.02E-02 -	9.07E-02 ***	1.26E-01 ***	7.09E-02 ***	1.79E-02 *		3.11E-03 -
Texture, φ_T		2.88E-03 (-)	1.31E-02 **		-4.76E-02 *	-2.50E-02 (-)			
Soil Carbon, φ_C	-7.19E-03 (-)	2.58E-03 (-)			-7.05E-02 **		-1.79E-02 *		
Temperature			-1.66E-02 ***		-4.41E-02 *	-4.15E-02 **		-1.47E-01 **	-2.60E-03 -
Precipitation	3.88E-05 ***			5.23E-05 *			-4.75E-05 ***	-7.59E-04 **	1.59E-05 *
Dry Season Precipitation		1.26E-04 ***	-1.24E-04 -			6.61E-04 *		1.09E-02 ***	-9.62E-05 (-)
Radiation		2.63E-04 *		1.11E-03 -	-2.96E-03 **	1.60E-03 (-)		8.28E-03 *	
R^2	0.418	0.362	0.498	0.568	0.601	0.488	0.376	0.346	0.305
Moran's I	0.053	0.394	-0.049	0.173	0.425	0.460	0.143	0.477	0.469
SAR FS	M_A	[C]	[N]	[P]	[Ca]	[K]	[Mg]	[Al]	$\delta^{13}\text{C}$
Intercept		-2.73E-02 (-)		-1.69E-01 (-)	1.19E+00 -	-4.00E-01 (-)		1.34E+00 (-)	6.77E-02 -
Fertility, φ_F		-4.97E-03 ***		8.26E-02 ***	1.29E-01 ***	6.10E-02 ***			3.40E-03 **
Texture, φ_T		3.23E-03 *			-4.45E-02 *	-4.38E-02 ***			
Soil Carbon, φ_C		2.66E-03 (-)			-6.26E-02 *				
Temperature					-3.48E-02 (-)	5.68E-03 (-)		-8.91E-02 -	-3.87E-03 **
Precipitation				5.41E-05 *				-8.40E-04 **	1.68E-05 *
Dry Season Precipitation		8.47E-05 *				1.38E-03 ***		1.21E-02 ***	-9.29E-05 (-)
Radiation		9.83E-05 (-)		1.63E-04 (-)	-1.73E-03 (-)	4.02E-04 (-)		1.01E-02 **	
λ		0.483		0.421	0.460	0.761		0.461	0.566
Moran's I		0.033		0.018	0.030	0.023		0.154	-0.010
SAR MS	M_A	[C]	[N]	[P]	[Ca]	[K]	[Mg]	[Al]	$\delta^{13}\text{C}$
Intercept		-5.83E-02 **		-2.42E-01 -	1.71E+00 **	4.14E-01 (-)		3.54E+00 **	4.45E-02 (-)
Fertility, φ_F		-6.01E-03 ***		5.97E-02 ***	1.13E-01 ***	5.87E-02 ***			3.58E-03 *
Texture, φ_T		2.86E-03 *			-3.87E-02 -	-2.53E-02 -			
Soil Carbon, φ_C		2.60E-03 (-)			-6.81E-02 **				
Temperature					-4.50E-02 **	-2.99E-02 *		-2.03E-01 ***	-2.98E-03 *
Precipitation				5.55E-05 ***				-7.94E-04 ***	1.82E-05 **
Dry Season Precipitation		1.26E-04 ***				5.20E-04 (-)		1.25E-02 ***	-1.21E-04 *
Radiation		2.66E-04 **		6.51E-04 (-)	-3.33E-03 ***	1.56E-03 -		1.35E-02 ***	
λ		0.012		0.832	-2.311	0.849		-0.856	-0.622
Moran's I		0.392		-0.060	0.377	0.381		0.390	0.485

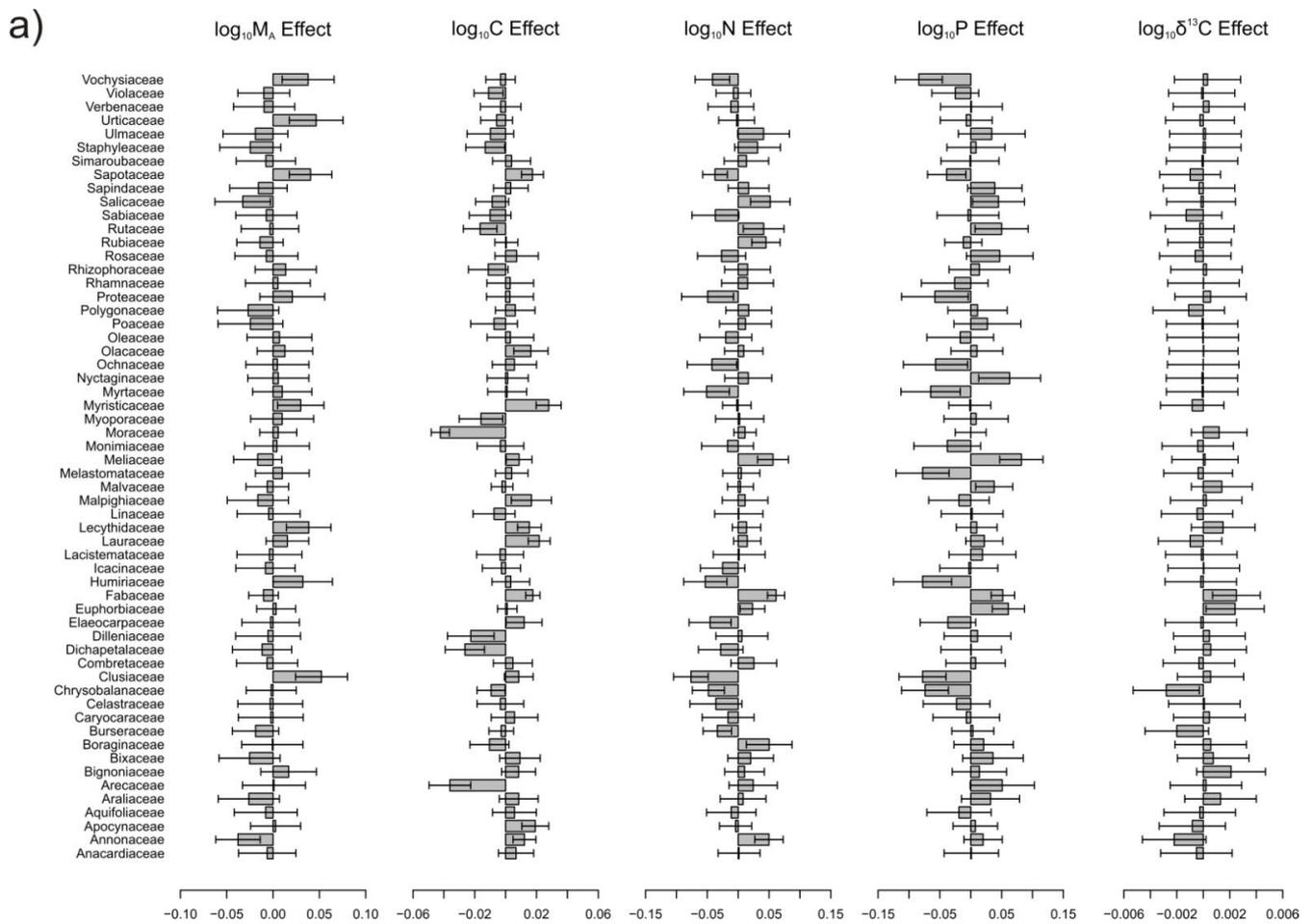


Fig. B1. Family effect \pm standard error estimates of the fitted multilevel model for leaf mass per unit area (M_A) expressed in g m^{-2} , elemental concentrations (all in mg g^{-1}) and $\delta^{13}\text{C}$ expressed in $[0/00]$. Note the \log_{10} scale used for all parameters.

As another example we discuss the results for [Ca] plot effect regressions. The OLS indicated the φ_F axis, the φ_T axis, the φ_C axis, annual temperature and solar radiation as important predictors. The residuals of the non-spatial model (Fig. SIII 2b-OLS) and the permutation method for the global Moran's I (Table A1) identified a significant spatial signal in the simple OLS. Thus we proceeded by fitting the fine and medium scale SAR models. The fine scale (50 km) SAR model removed the spatial autocorrelation from the residuals as indicated by both the respective correlogram (Fig. SIII 2b-SAR50) and the global Moran's I (Table A1 – SAR50 panel). However in this model annual temperature and solar radiation lost their significance, while the rest of the predictors retained their significance with a small difference in their coefficient estimates. Finally the medium scale SAR model seemed not to perform better as it did not manage to remove spatial autocorrelation from the residuals. We thus accepted as the better predictors for [Ca] plot effect the three soil axis.

A similar comparative consideration was undertaken for the OLS and SAR results for other foliar properties, includ-

ing $\delta^{13}\text{C}$. In summary, plot level nitrogen and magnesium effects showed no spatial signal in the residuals, and thus the OLS model was considered adequate to describe the main environmental predictors. Plot level [N] effects were mainly associated with the mean annual temperature and the soil texture axis with a less important contribution of fertility and dry season precipitation. The plot level [Mg] effect was mainly related with φ_F and the third φ_C axis, as well as with annual precipitation. These environmental predictors accounted for a high 0.50 and 0.38 of the total plot effect [N] and [Mg] variability respectively.

On the other hand, leaf [C] variation had a strong spatial pattern which affected the residuals of the OLS (Moran's $I=0.39$). Nevertheless, the fine scale (50 km) SAR model removed most of this strong spatial signal (Moran's $I=0.03$) although this was not the case for the medium scale (750 km) SAR model (0.39). Nevertheless, the main environmental predictors of the OLS model, namely soil fertility and dry season precipitation remained important in both SAR models, suggesting their valid influence.

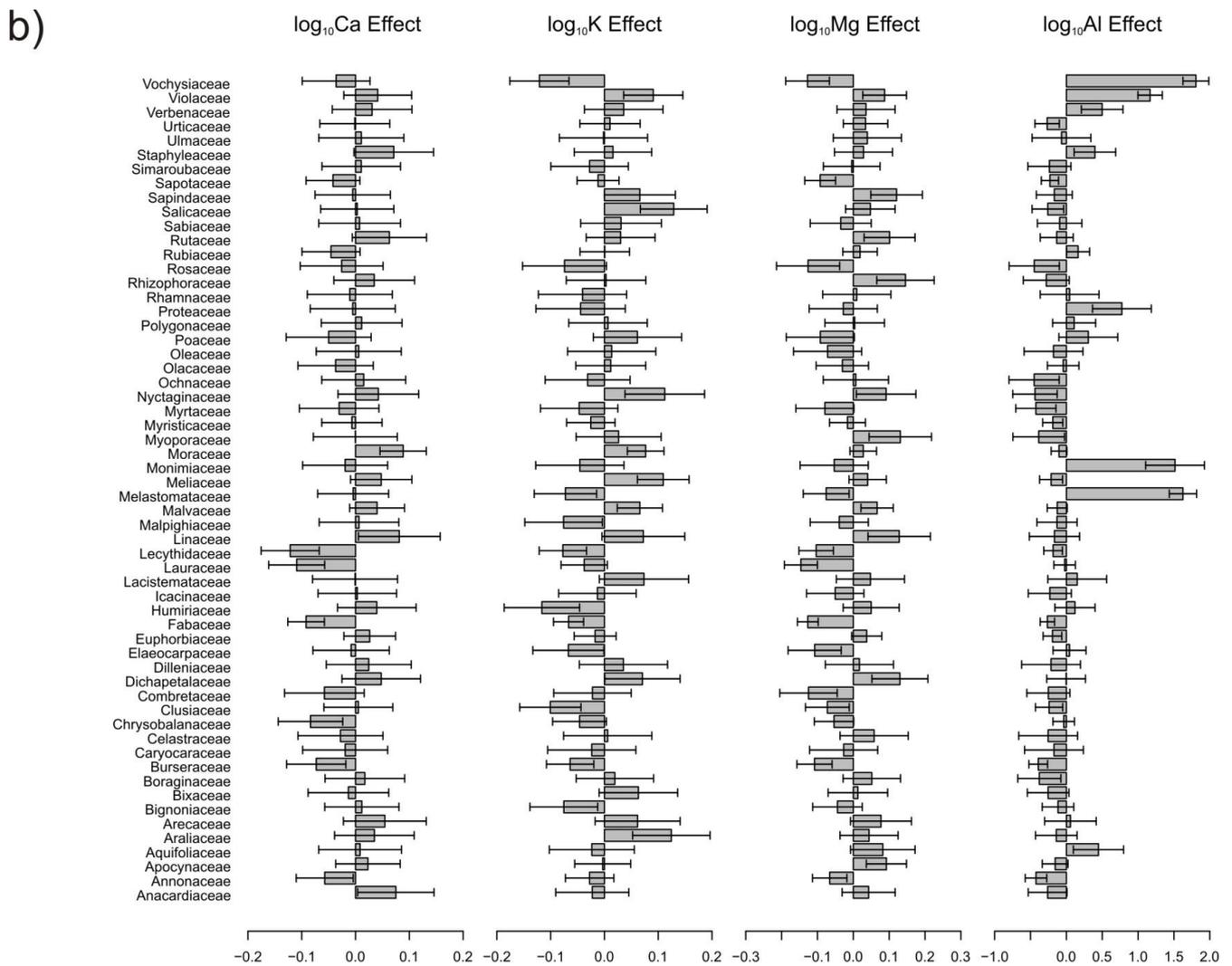


Fig. B1. Continued.

The residuals of the plot effect [P] non-spatial regression presented a spatial signal which was effectively removed by both the fine scale (50 km) and the medium scale (1000 km) SAR models (Fig. SIII2), and in both cases the autoregression coefficient λ was significantly different from zero (Table A1). Following our comparative approach we thus suggest a significant plot level [P] contribution with the soil fertility axis and a weaker one with total annual precipitation.

Plot level [K] and [Al] effects had autocorrelated residuals when space was not explicitly taken into account. However no spatial signal in the residuals of the fine scale SARs was identified, and the spatial component as expressed through λ was significant in all cases. Following again the comparative consideration of OLS and SAR results we retained as significant the possible associations between φ_F , φ_T and dry season precipitation with plot [K] effect and annual and dry season precipitation as well as solar radiation with plot [Al]

effect. Finally the spatial pattern in the OLS residuals for $\delta^{13}\text{C}$ was greatly removed by the fine scale SAR model but not from the medium scale model. The plot $\delta^{13}\text{C}$ effect was mainly related with φ_F , mean annual temperature and total annual precipitation. Summarizing the above we could suggest that the fine scale SAR models seemed more appropriate to account for the spatial patterns of our dataset, and that soil fertility, precipitation and to a lesser extent annual temperature were the main environmental predictors related with most plot effect contributions.

The simple spatial analysis applied, gave us some insights as to the effects of documented soil and climate gradients on the variation of the studied traits. The residuals of the OLS regression for the M_A , [N] and [Mg] plot level effects did not show any spatial autocorrelation (Fig. SIII 2, Table A1), suggesting that at these scales there was no specific fertility oriented pattern in the environmental contribution to the

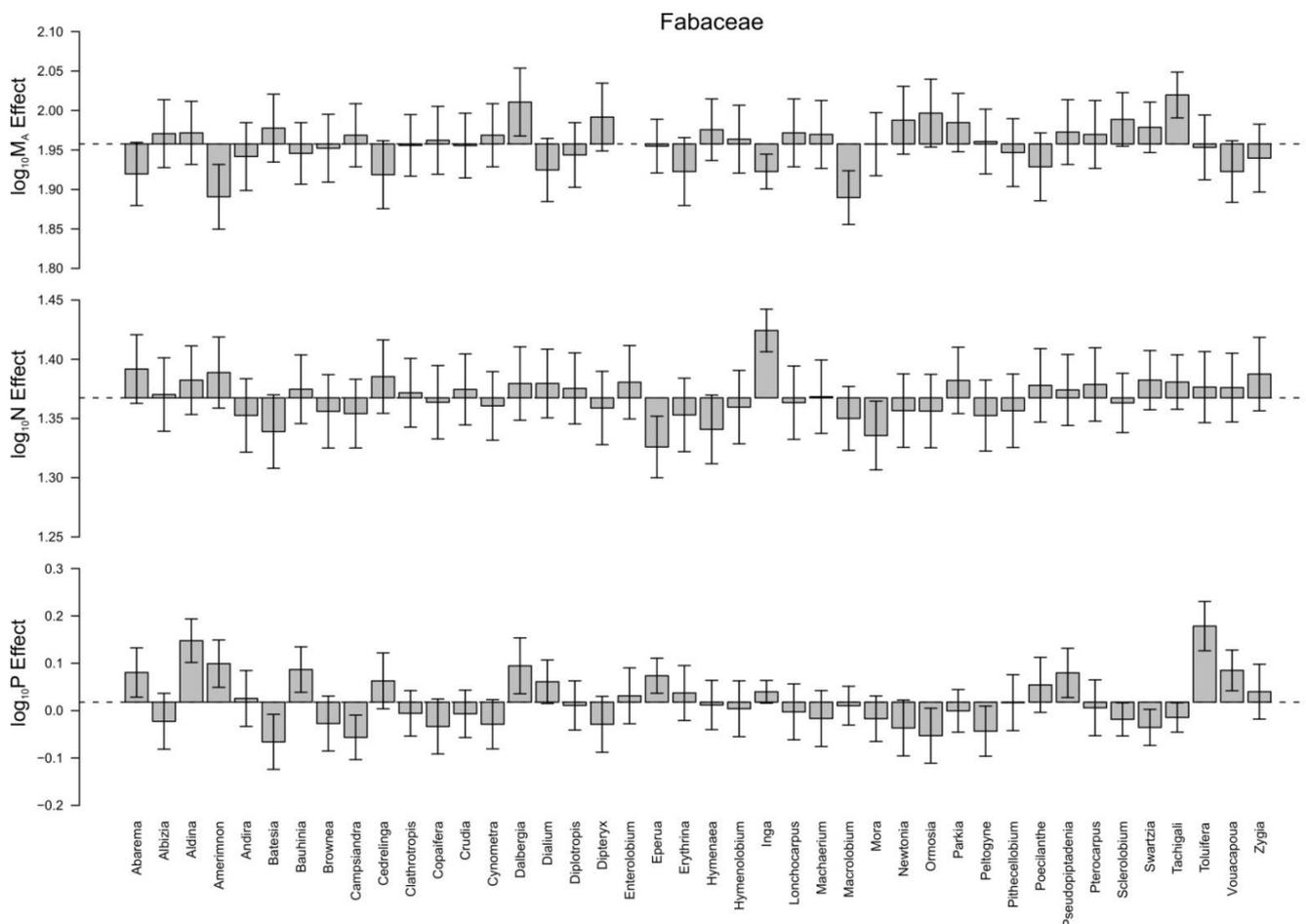


Fig. B2. Genus effect±standard error estimates from the multilevel model for sampled genera within Fabaceae. The dotted line illustrates the (mean) family effect for leaf mass per area (M_A) in g m^{-2} and leaf N and leaf P concentrations (mg g^{-1}). Note the \log_{10} used scale in all cases.

above variables. On the other hand, the residuals of the non-spatial regressions of [P], [Ca] and [K] plot effects on environmental predictors illustrated a significant spatial signal, which was effectively removed by the fine scale (50 km) SAR models. For these three nutrients, the spatial component was important ($\lambda \neq 0$, Table A1) highlighting once more the substantial influence of environmental conditions on trait variation. We note, however, that for the plot level [P] contribution, the medium scale (1000 km) SAR model was equally capable of removing the spatial signal with the spatial component being similarly important. Thus the spatial patterning of environmental contribution to leaf [P] variation may be realized at broader scales. The results of the spatial analysis for [C], $\delta^{13}\text{C}$ and [Al], presented a similar behaviour, with fine scale SAR models removing most of the autocorrelation in the residuals. Although correlograms showed a significant Moran's I for both plot [C] and $\delta^{13}\text{C}$ effects at the second distance class (Fig. SIII 2), the Monte Carlo permutation did not identify a globally significant spatial signal (Table A1).

Thus the comparative consideration of both the spatial and non-spatial regressions identified the most significant environmental factors contributing to trait variation.

Appendix B

Family, genera and species effects

In our dataset there were significant differences between families for all sampled traits (ANOVA results not shown) with Fig. B1 illustrating mean family effects and their standard error estimates from the multilevel model. This shows that some families like Vochysiaceae, Urticaceae, Sapotaceae, Myristicaceae, Lecythidaceae, Humiriaceae and Clusiaceae are characterised by higher than average M_A , while others like Salicaceae and Annonaceae have unusually low M_A . Although [N] and [P] also tend to be lower in families with high M_A such as Vochysiaceae, Sapotaceae, Humiriaceae and Clusiaceae, these concentrations were also

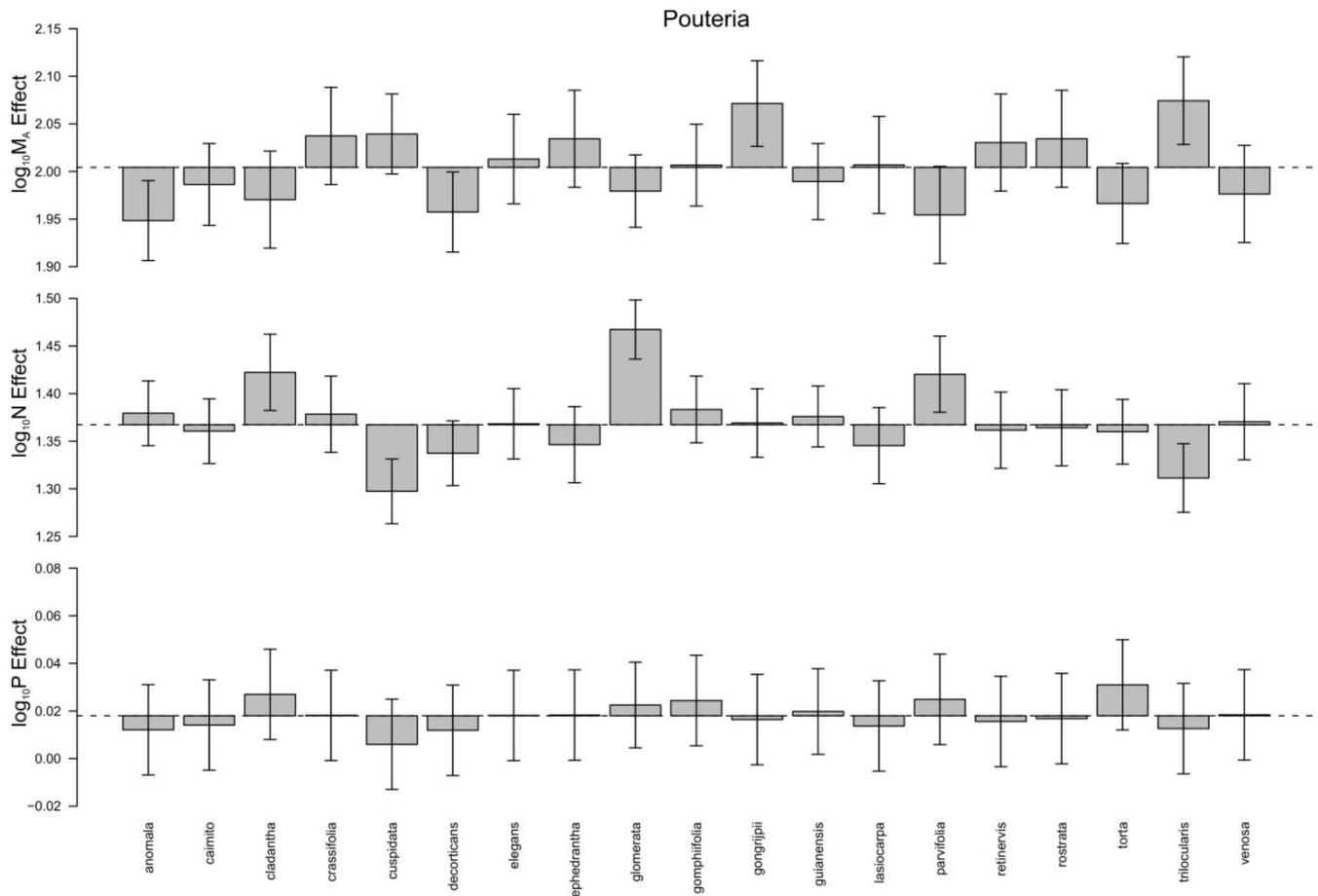


Fig. B3. Species effect \pm standard error estimates from the multilevel model for species within *Pouteria*. The dotted line illustrates the mean (family + genus) effect for leaf mass per area (M_A) in g m^{-2} and leaf P and leaf N concentrations (mg g^{-1}). Note the \log_{10} used scale in all cases.

Table B1. Mean genetic values (intercept + genetic effect) \pm standard deviation for taxa not belonging to Fabaceae, non-N-fixing Fabaceae and N-fixing Fabaceae. Values followed by different letters within a trait-column indicate significant differences at $p < 0.001$ for the three groups.

	M_A (g m^{-2})	[C] (mg g^{-1})	[N] (mg g^{-1})	[P] (mg g^{-1})	[Ca] (mg g^{-1})	[K] (mg g^{-1})	[Mg] (mg g^{-1})	[Al] (mg g^{-1})	$\delta^{13}\text{C}$ (‰)
non-Fabaceae	99.3 \pm 14.2 ^a	516.7 \pm 9.6 ^a	21.07 \pm 1.43 ^c	0.91 \pm 0.12 ^c	4.59 \pm 0.88 ^{a,b}	5.78 \pm 1.16 ^a	1.95 \pm 0.35 ^a	0.042 \pm 0.012 ^a	-31.21 \pm 0.33
Fabaceae									
non-N ₂ -Fixers	87.9 \pm 11.4 ^b	506.7 \pm 8.5 ^b	22.53 \pm 2.17 ^b	1.01 \pm 0.09 ^b	5.40 \pm 2.01 ^a	4.95 \pm 0.75 ^b	1.89 \pm 0.46 ^{a,b}	0.033 \pm 0.006 ^b	-31.16 \pm 0.46
N ₂ -Fixers	84.9 \pm 11.0 ^b	508.4 \pm 9.1 ^b	25.42 \pm 1.81 ^a	1.11 \pm 0.09 ^a	4.24 \pm 1.01 ^b	5.17 \pm 0.63 ^b	1.69 \pm 0.26 ^b	0.033 \pm 0.008 ^b	-31.2 \pm 0.32

lower for families like Proteaceae, Ochnaceae, Myrtaceae and Chrysobalanaceae which were not distinctive in terms of M_A . Likewise, some families that do not display a markedly lower genetic M_A component (Rutaceae, Meliaceae, Fabaceae and Euphorbiaceae) do have relatively high [N] and [P].

Looking at within family variation and using the Fabaceae as an example, genus level effects are shown for M_A , [P] and

[N] in Fig. B2. For this family, the most abundant within the dataset (160 observations), there were significant genus effects for M_A ($F_{40,111}=3.832$, $p < 0.001$), N ($F_{41,109}=4.788$, $p < 0.001$) and P ($F_{41,108}=4.095$, $p < 0.001$) but with M_A and [N] showing relatively less variability. Nevertheless, genera like *Amerimnon*, *Dalbergia*, *Inga* and *Tachigalia* all tended to differentiate in both M_A and [P] with *Inga* also being notable for unusually high [N].

For the other widely distributed families, genera within the Malvaceae differed in M_A , [N] and [P], with the Euphorbiaceae and Urticaceae showing differences in M_A and [N] only; Moraceae and Myristicaceae in [N] and [P] only; and the Burseraceae and Lecythidaceae giving between-genera variability only for [P]. Moreover, there were no differences in these foliar traits across genera in some families; for example the Chrysobalanaceae and Sapotaceae (results not shown). Generally speaking the highest within family variation was also observed for the more widespread families with the genus level analysis pointing to M_A , [P] and [N] not necessarily always varying in concert with genotype as the sole course of variation. For example, genera within the Moraceae were statistically indistinguishable in terms of M_A , but showed large variations for [P] with *Olmedia* species tending to have significantly higher [P] and *Pseudolmedia* and *Sahaqunia* significantly lower [P] than the family mean estimate.

Within *Eschweilera*, the most extensively sampled genus in our dataset ($n=60$), significant species-to-species variation were identified in M_A ($F_{14,44}=2.009$, $p=0.040$), and [P] ($F_{14,43}=2.591$, $p=0.008$). Species within *Pouteria* ($n=44$) showed differences in M_A ($F_{18,23}=2.728$, $p=0.012$), [N] ($F_{18,23}=4.157$, $p < 0.001$) and [P] ($F_{18,23}=4.217$, $p < 0.001$), but within *Inga* ($n=38$) no species specific differences were identified for either M_A ($F_{21,15}=1.051$, $p=0.470$), [N] ($F_{19,15}=1.876$, $p=0.110$) or [P] ($F_{19,15}=0.796$, $p=0.685$). Other well represented genera such as *Licania* showed significant variation in M_A and [P] only, but other genera for [N] only (*Pourouma*, *Protium*), or in many cases, with no difference at all (e.g. *Pseudolmedia*, *Virola*).

To illustrate such species effects, we summarize the M_A and [P] estimates from the multilevel model for all *Pouteria* species sampled in Fig. B3. It should be noted that in our analysis all random effect estimates are shrunk towards the overall mean with this shrinkage increasing with decreasing genetic variance and increasing environmental variance (Galwey, 2006). Thus random effect estimates show a greater variability for M_A and [N] compared with [P], which is realistic considering that only 0.01 of the total variance can be attributed to the species affiliation for [P], compared with the 0.19 for M_A and 0.14 for [N]. Most obvious here is the exceptionally high M_A for *P. gongripitii* and *P. tricularis*, the latter also being accompanied by an unusually low [N], as well as very high [N] for *P. glomerata*. Thus within-genera variability is not restricted to specific foliar properties or even to combinations of them.

We were also interested to see if there was a difference in the physiological traits examined between the nitrogen fixing and non-nitrogen fixing members of the Fabaceae. We therefore utilised a database summarising published records of the presence or absence of N_2 fixation in the Fabaceae (Patiño et al., 2008) taking a species as a N_2 fixer if the proportion of positive records was greater than 0.9 and as a non-fixer if the proportion of negative records was greater

than 0.9. Species with intermediate (conflicting) records of their N_2 -fixing status were not included in this analysis (Table B1). This showed that N_2 -fixing Fabaceae have significantly higher [N] and [P] than their non- N_2 -fixing relatives but with significantly lower [Ca] and no significant differences in M_A . In turn, the non- N_2 fixing Fabaceae have significantly higher [N] and [P] but lower [K] than is the average for the other non- N_2 fixing trees sampled across the Amazon Basin.

This result suggests that although [P] and [N] are indeed higher for the N_2 -fixing capable members of the Fabaceae, in accordance with the notion that N_2 fixers have a high [P] and a high [N] requiring “lifestyle” (Vitousek et al., 2002), members of the Fabaceae who cannot fix nitrogen also have elevated [N] and [P] compared to the Amazon tree population as a whole. But with [N] and [P] concentrations elevated to a lesser extent than for Fabaceae capable of N_2 fixation. It has recently been suggested that N_2 -fixing Fabaceae are abundant in tropical ecosystems through their high [N] status, giving them an ability to exude high level of phosphatase enzymes and hence acquire extra phosphorus (Houlton et al., 2008). But it also seems to be the case that most Fabaceae within Amazonia do not fix nitrogen, even when physiologically capable of doing so (Nardoto et al., 2008). The lower foliar [Ca] for the N_2 -fixing Fabaceae may be attributable to the high calcium requirement for *Rhizobia* growth in the nodulating rhizosphere (O’Hara, 2001).

Acknowledgements. Nikos Fyllas and much of the data analysis and production phase of this work were funded through the UK National Research Council QUEST (Querying the Earth System) initiative, subprogram QUERCC (Quantifying and Understanding Ecosystems Role in the Carbon Cycle). Shiela Wilson helped with the final manuscript presentation. Comments from J. Grace and an anonymous referee improved this paper.

Edited by: J. Grace

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