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Meiofauna winners and losers of coastal hypoxia: case study harpacticoid copepods

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Abstract

The impact of anoxia on meiobenthic copepod species was assessed by means of a field experiment. Four plexiglass chambers were deployed in situ in 24 m depth to simulate an anoxic event of 9 days, 1 month, 2 months and 10 months. From normoxic to anoxic conditions, we recorded a drop in copepod density and species richness. With increasing duration of anoxia the relative abundance of the individuals of the family Cletodidae increased, and they survived the 1 month and 2 month anoxia, the latter with few specimens. They were the true “winners” of the experimentally induced anoxia. Dominance did not increase throughout all deployments because; not one, but several species from this family were tolerant to anoxia. The overall rate of survival was the same for males and females, but no juvenile stages of copepods survived in anoxia. During a recovery phase of 7 days after a short-term anoxia of 9 days, harpacticoid copepod density did not increase significantly, and there was only a slight increase in species diversity. We concluded that there was no substantial colonisation from the surrounding sediment. The survivors, however, showed a high potential for recovery according to the number of gravid females, whose number increased significantly once the oxygen was available again. These findings imply that a substantial amount of energy is allocated to reproduction in the recovery phase.

1 Introduction

One key threat to the marine interstitial benthos is the shortage of oxygen. Consequently, their vertical distribution is governed by the oxygen penetration into the sediment, and oxygen is considered to be a major structuring factor of metazoan communities in marine sediments (Glud, 2008, within this special issue). With worldwide more than 400 oxygen-depleted systems recognized, covering a total area of > 245 000 km² (Diaz and Rosenberg, 2008), hypoxia (defined here as DO levels ≤ 2 mL L⁻¹) and

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anoxia (no oxygen) are amongst the most pressing threats for shallow coastal seas (UNEP, 2006) and thus for the organisms in the sediments.

Benthic organisms are typically classified in two size classes based on the mesh size used to extract them from the sediment. Meiofauna are metazoans that pass through a 1 mm sieve but are retained on a 38 μ m sieve, whereas macrofaunal organisms are larger than 1 mm. Beyond this methodological classification they also differ considerably in their functioning and oxygen demand. Meiofauna have lower oxygen requirements and hence are generally more resistant to hypoxia (Giere, 2009). They all lack respiratory organs and thus depend on diffusive oxygen uptake (Wetzel et al., 2001). Consequently, slender, worm-shaped organisms, for example, benefit from a higher surface : volume ratio to gather the required oxygen (Powell, 1989; Wetzel et al., 2001), which is then consumed in the body in seconds (Braeckman et al., 2013).

Within the meiofauna, different levels of sensitivities and tolerances to hypoxia and anoxia have been described. Generally, harpacticoid copepods (Crustacea, Copepoda) are more sensitive to low oxygen concentrations, while nematodes (Nematoda; roundworms) are more tolerant (Vernberg and Coull, 1975; Hicks and Coull, 1983; Murrell and Fleeger, 1989; Wetzel et al., 2001). The latter, worm shape benefits from a higher surface : volume ratio (see above). Moreover, species-specific responses within copepods and nematodes are reported (Vopel et al., 1998; Modig and Ólafsson, 1998; Wetzel et al., 2001). Thus, an in-depth response study should be based on species-level to accurately interpret the community-level response to hypoxia/anoxia. Furthermore, the local environment, i.e. sediment characteristics or food availability (De Troch et al., 2013) will govern the final response of a benthic community. (Hicks and Coull, 1983; Wetzel et al., 2001) suggested that also the evolutionary history, i.e. the frequency of hypoxia, might support the establishment of a more tolerant community, such as the one found in the Skagerrak area (Josefson and Widbom, 1988). This calls for integrating, not only the resistance but also the resilience of a community. A holistic approach encompassing the overall meiofauna community effects to hypoxia/anoxia should therefore also include the post-perturbation reactions, i.e. the recovery phase.

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The individual responses of species will determine “winners” (surviving, more tolerant species) and “losers” (death of sensitive taxa), with the resilience of the survivors being essential for the post-impact community composition. A fast recovery (i.e. weeks) can be achieved through a fast reproduction (Harris, 1973; Coull and Vernberg, 1975; Gee and Warwick, 1984) requiring information on often neglected population dynamics.

The present study is designed to provide a more holistic picture of the response of harpacticoid copepods to hypoxia/anoxia in the natural environment. Hypoxia and anoxia were experimentally induced in 24 m depth in the Gulf of Trieste, Northern Adriatic Sea (Mediterranean) with four plexiglass underwater chambers (for details see Stachowitsch et al., 2007). They were deployed for 9 days, 1 month, 2 months and 10 months. Harpacticoid copepods were selected in view of their lowest sensitivity to anoxia. Note that Langlet et al. (2013a, b) analysed the benthic foraminiferans, typically among the most tolerant groups to hypoxia/anoxia, to complement our taxon. Our community analysis is based on species composition and focuses on the individual response of the most resistant species. The hypothesis is that despite their low resistance to hypoxia, harpacticoid copepods show a high resilience. This resilience can be explained by short generation times (Hicks and Coull, 1983; Giere, 2009), low food selectivity in certain species (De Troch et al., 2006) and by an opportunistic nature resulting in fast colonisation (De Troch et al., 2005). These features will allow them to establish a new community as soon as oxygen becomes available again. For this purpose we evaluated copepod community recovery after 9 days of hypoxia/anoxia in the impacted site. Also, we evaluated the population dynamics and sex ratio during the hypoxic/anoxic event as well as during the recovery phase. This yielded additional information on the condition of the survivors, because this copepod community was found to stop feeding during hypoxia (see laboratory experiment De Troch et al., 2013), although it remained unknown whether they allocate energy to reproduction during low oxygen conditions.

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2 Material and methods

2.1 Experimental set-up

A field experiment was performed on a poorly sorted silty sand bottom in the Northern Adriatic Sea (Mediterranean) (45°32.90' N, 13°33.00' E) at 24 m depth. Artificial hypoxia and anoxia were created with 4 underwater chambers (each 50 cm × 50 cm × 50 cm). While almost all chambers were deployed at the same time, the duration of the respective deployments differed (i.e. 9 days, 1 month, 2 months, 10 months, see below). The chamber used in the 9 day deployment was originally designed to document macroepi- and infauna behaviour during oxygen decline (Riedel et al., 2013). The separate lid houses a time-lapse camera (images taken in 6 min intervals), 2 flashes, battery packs, a microsensor array (dissolved oxygen, hydrogen sulphide, temperature) plus datalogger (Unisense®). In this deployment, the sensors were positioned 2 cm above the sediment (measurements taken in 1 min intervals). For more details on the experimental design see Stachowitsch et al. (2007). The other 3 chambers were not instrument-equipped. pH was recorded at the beginning and end of deployments with an external WTW TA 197-pH sensor. All chambers were positioned at macroepifauna-free sites, without major traces and structures (mounds or pits) that would indicate the presence of larger infauna species. 2–3 brittle stars were placed on the sediment inside each chamber to visually better follow and verify the oxygen decline based on their behaviour (Riedel et al., 2008).

A triplicate of normoxic reference cores (length 20 cm, inner $\varnothing = 4.6$ cm, surface = 16.6 cm²) were taken on 3 August 2010 (treatment: *N*). Four underwater chambers were deployed, one on 2 August 2010, that was sampled on 11 August, named 9 days anoxic chamber. After this chamber was sampled and removed, the sediment was marked and sampled again (on the intact site) to monitor the recovery on 18 August 2010 (9 days recovery). The second chamber was deployed on 27 July and sampled on 25 August, namely the 1 month anoxic chamber. The third chamber was deployed on 27 July and sampled on 23 September, namely the 2 months anoxic chamber,

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and the last chamber was deployed on 24 September 2010 and sampled on 5 August of the following year, the 10 month chamber. Those samples are referred to as treatments A9d, R7d, A1m, A2m, A10m.

In the A9d-deployment, the oxygen concentration started to decrease as soon as the chamber was closed, and anoxia was reached after five days, that is two days before the samples were taken from the 9 days anoxic chamber (A9d). According to the geochemical data (Metzger et al., 2013) and the pore water nutrients (Koron et al., 2013), anoxia persisted in all the other chambers as well.

2.2 Copepod extraction and identification

Meiofauna cores were taken by scuba divers. Three cores were randomly taken outside the chamber (normoxia control treatments, see above) and another three cores were taken inside the chambers before the experiments were terminated (treatments anoxia). Moreover, after short-term anoxia, we followed the short-term recovery and took triplicate cores in the sediment which was reoxygenated. Cores were immediately transported to the laboratory in cooling boxes and transferred into a thermostatic room with in situ temperature (i.e. 18 °C). The cores were sliced in 0.5 cm intervals for the first 2 cm and 1 cm intervals down to 5 cm depth. The sliced sediment was placed into separate 250 mL containers.

The overlying seawater from the cores (10 mL) was added into each 250 mL container including the sediment slices. The fluorescent CellTracker Green (CTG) label (1 mg stored at -20 °C) was dissolved in 1 mL dimethyl sulfoxide (DMSO). The final concentration of CTG/DMSO solution was ~ 1 μ M, corresponding roughly to ~ 5 μ L CTG/DMSO per 10 mL of sediment and liquid together. Overall, 10 μ L of the CTG/DMSO solution was added to the sediment-overlying water with a micropipette. The containers were incubated at in situ temperature in the dark for 12 h. Grego et al. (2013) demonstrated that CTG is more accurate to enumerate living meiofauna after anoxia impact than for example Rose Bengal. The samples were subsequently fixed in 4 % borax-buffered (5 gL⁻¹) formalin.

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The samples were further processed following the standard meiofauna extraction protocol (De Jonge, 1977; Mc Intyre and Warwick, 1984). Formalin-fixed sediment samples were washed with tap water to eliminate formalin and clay by pouring them on a 38 μm sieve. The sediment recovered on the 38 μm sieve was transferred into 1 dl centrifuges. Levasil[®] (-distilled water) solution (specific density = 1.17 g cm^{-3}) was added and gently mixed with the sediment prior to centrifugation. After centrifugation, the copepods were retained in the floating phase and were separated from the sediment phase (containing Foraminifera, see Langlet et al., 2013a, b). As copepods were absent in the deeper sediment layers, only copepods from the uppermost cm sediment layers (0–0.5 cm, 0.5–1 cm, 1–1.5 cm and 1.5–2 cm) were counted under a binocular (Nikon SMZ 800 with a Nikon INTENSILIGHT C-HGFI for UV production and A 488 filter) and picked out with a needle. CTG samples containing a large number of animals were split into several petri dishes to avoid long exposure of animals to UV light, in order not to lose fluorescence due to bleaching. The specimens were picked out from the sample and then mounted in toto on slides containing glycerol. The species were identified to “working species” and enumerated as sp. 1, sp. 2, etc. within a particular genus (as for example in Janssen et al., 2013). Species identification was done with an Olympus BX-51 microscope equipped with a drawing tube (main identification features). The copepod reference collection and observation notes with sketches from this study are available at the Marine Biology Station Piran, Slovenia. Identification keys used include (Lang, 1948; Boxshall and Hasley, 2004; Wells, 2007). The different stages (copepodite, male, female, ovigerous female) and number of egg sacs were enumerated.

2.3 Data analysis

Data on copepod densities, ratio of male/female, and diversity indices were checked for distribution patterns prior to ANOVA calculations (Sokal, 1995; Dytham, 2003; Zuur et al., 2010), which was performed using the R statistical software package (Team, 2010). The graphs were done in Microsoft Office Excel.

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For multivariate analyses and for the calculation of diversity indices, Primer v6 (PRIMER-E, 2006) (Clarke and Warwick, 2001; Clarke and Gorley, 2006) was used. Based on the species densities (not transformed) the copepod communities were plotted in a non-metric Multi-Dimensional Scaling (nMDS) to demonstrate similarity distances based on the Bray–Curtis similarity index. In addition, an analysis of similarity (ANOSIM) between the groups of samples (factor treatment: N, A9d, R7d, A1m, A2m, A10m) was performed and a SIMPER analysis was used to investigate which species were responsible for dissimilarities among groups of samples.

A cluster analysis of the copepod species densities per different sediment depth layers (absolute densities, not transformed) was calculated based on Bray–Curtis similarity.

Diversity of the harpacticoid copepod communities was calculated at species levels using three indices: species richness, Shannon diversity index and Pielou’s evenness index by means of Primer v6 (Clarke and Warwick, 2001; Clarke and Gorley, 2006). Copepodite (CI–CV) life stages were excluded from the diversity measures.

3 Results

3.1 Copepod assemblage in different oxidic conditions

This study yielded 47 species (overall 1038 individuals identified) of harpacticoids.

The copepod density in the sediment cores was highest in normoxic conditions and gradually dropped with the duration of anoxia (Fig. 1). The density (\pm STD per 16.6 cm^2) in normoxic samples was significantly higher (198 ± 57.3) than that in all anoxic samples (A9d: 56.3 ± 11 ; A1m: 16.7 ± 5 ; A2m: 2 ± 1.4 , A10m: 0) as well as in the R7d-samples (73.7 ± 18.5). The one way ANOVA (factor oxygenation) revealed a significant difference ($p < 0.001$) among samples, and the Tukey HSD posthoc test clarified the pairwise differences (i.e. normoxic samples always differed from the other samples: $p < 0.001$ or $p < 0.01$). Seven days after termination of the A9d-deployment, poten-

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tial recolonisation from the surroundings was determined. Except for one replicate in which the density was higher than in anoxic samples, the densities were similar to anoxia, suggesting that no recolonisation took place.

The multidimensional scaling-MDS based on absolute copepod species abundances showed a high similarity among replicates of normoxia (Fig. 2). Similarly, a low spread among the replicates of 9 day anoxia and 7 day recovery, respectively, was found (Fig. 2). The 1 month anoxia replicates formed the third group, while the 2 month anoxia replicates were most different from the rest (Fig. 2). The latter showed also more variance among the replicates. Note that one replicate of the 2 month anoxia deployment as well as the 10 month anoxia replicates were not plotted because no live copepods were present. ANOSIM for all groups (N, A9d, R7d, A1m, A2m) revealed a significant separation of the samples (global $R = 0.846$, $p = 0.01$). The pairwise ANOSIMs showed a clear difference among all groups except for no difference between the 9 day anoxia and the 7 day recovery group ($R = 0.222$, $p = 0.3$).

The SIMPER analysis (Table 1) showed the species that contributed most to the dissimilarity among treatments. After short-term anoxia (9 days), the species that were mostly impacted (see average abundance in Table 1) were *Pseudobradia* sp. 2, *Heteropsyllus* sp. 1, *Nitocrella* sp. 1 and *Longipedia* sp. 2. In contrast, the density of *Stylicletodes* sp. 2 was highest after 9 days of anoxia (meiofauna patchiness), whereas the density and the number of six species of the genus *Cletodes* remained at the same level as in normoxia. With increasing anoxia duration (1 month), however, the number of *Stylicletodes* sp. 2 and *Cletodes* sp. 2 drastically dropped. This was also the main reason for the dissimilarity among samples because both species contributed 31.9% to the dissimilarity with the other treatments. Interestingly, while *Acrenhydrosoma* sp. 1 and *Enhydrosoma* sp. 4 were absent from the 9 day anoxic samples, the same species were present in the 1 month anoxia samples. This may reflect the patchiness of meiofauna communities, that is not always (for all species) assessed with three replicates. Finally, the prolongation of anoxia to 2 months yielded killed most harpacticoid cope-

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pods, with individuals of only three species (two *Cletodes* species, one *Enhydrosoma*) surviving.

Overall, the number of species decreased significantly ($p = 0.004$) by half already after short-term (9 days) anoxia (Fig. 3a), and it continued decreasing with the prolongation of anoxia. Similarly, the Shannon diversity significantly dropped ($p < 0.001$); only a slight increase was measured in the 7dR (Fig. 3b). Interestingly, in all anoxic deployments the evenness index remained stable (Fig. 3c), indicating that there was never one species that dominated the community.

3.2 Copepod population composition

Regardless of the prevailing oxygen conditions, the male/female ratio remained relatively stable. The ratio is not balanced as the relative abundance of females was always significantly higher than that of males (Table 2) (2-way ANOVA with factors sex and treatment, $p < 0.001$).

In all treatments, including normoxia, the highest number of gravid females was recorded in the family Cletodidae (Table 2). The high values for this family were independent of the relative abundance of the family because the relative share of Cletodidae to the overall community was highly similar to the shares of Ameiridae, Canthocamptidae, Longipedidae, Ectionsomatidae and Miraciidae (Fig. 4). Anoxia resulted in a drastic decrease in the number of gravid females in all families (Table 2). The level of reduction corresponded with the increasing duration of oxygen depletion (i.e. greatest drop in A2m). The number of gravid females (mostly family Cletodidae and also some Miraciidae) significantly increased in the 7d-recovery treatment. The ANOVA calculated from the density of gravid females per treatment (triplicated samples) N, A9d, R7d, and A1m, showed significant differences among treatments ($p < 0.001$), and Tukey HSD revealed that those differences are significant among pairs: N with all the anoxia treatments, and R7d with all the anoxia treatments.

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Likewise, the number of juvenile copepodite stages (I–V) dropped drastically (Table 2) with 9 days anoxia, but copepodites were completely absent with further increase of anoxia (1 month, 2 months and 10 months).

3.3 Copepod assemblage in different depths and oxic conditions

5 Four main clusters were defined based on the absolute copepod species composition in the different sediment layers (0–0.5 cm, 0.5–1 cm, 1–1.5 cm, 1.5–2 cm) (Fig. 5). In the first cluster the most similar were the top layers (0–0.5 cm) of 9 days anoxia and 9 days recovery, that grouped close to the second (0.5–1 cm) sediment layer of normoxia. The top layer of anoxia is also in this cluster. The second cluster is formed from deeper
10 layers of 9 days anoxia, and 7 days recovery and the top layers of 1 month anoxia. In the last two clusters the remaining deeper sediment layers clustered together with the top layer (0–0.5 cm) of the long-term (2 months) anoxia sample.

4 Discussion

This field experiment documents the individual response of harpacticoid copepods to
15 anoxia. By including different incubation deployment times, we were able to illustrate more precisely than ever before in the field the sensitivity of harpacticoid copepods to oxygen depletion, a recurring event in the Gulf of Trieste (Stachowitsch, 1984, 1991; Faganeli et al., 1991; Justić et al., 1993). Although copepods are known to be among the most sensitive meiofauna groups to low oxygen concentrations (see review by Wetzel
20 et al., 2001; De Troch et al., 2013), our results show that the densities of harpacticoids dropped considerably after 9 days of anoxia, but that still one fourth of the community survived. A longer duration of anoxia (up to 2 and 10 months) was necessary to remove almost all specimens. This sheds new light on the anoxia-sensitivity/tolerance of harpacticoid copepods, i.e. they may be less sensitive than generally stated in the literature (Hicks and Coull, 1983; Murrell and Fleeger, 1989; Modig and Ólafsson, 1998;
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Wetzel et al., 2001). Alternatively, benthic diatoms may have produced enough pulses of oxygen – not recorded by the sensor – to allow a few species to survive the extreme conditions for 1–2 months. Emili et al. (2012) analysed the light penetration through a similar benthic chamber (same size and plexiglass thickness – 0.6 cm) and recorded
5 92 % penetration of photosynthetically active radiation. Those were ideal conditions: our chambers, however, soon became shadowed by the sedimented material and animals (e.g. brittle stars) climbing onto them. The initial lack of such shading, however, coupled with primary production, would be one potential explanation of the survival of few copepods in the 2 month anoxia treatment. Other strategies, such as the attachment of epibiontic diatoms on the ventral part of the pelagic copepod *Acartia italica*,
10 have also been forwarded to explain survivals in anoxia (Kršinić et al., 2000).

According to stable fluxes of pore water nutrients (see Koron et al., 2013) there was no bioturbation in normoxic or recovery cores, although such bioturbation was recorded after 9 days and 1 month of anoxia, especially in the top sediment layers. This is probably
15 due to the vertical migration of infauna, yet this bioturbation has apparently not substantially reoxygenated the chamber. No bioturbation was recorded in the 2 month and 10 month anoxic chambers, which is in accordance to our results, i.e. only 4 copepods alive after 2 months of chamber closure and all copepods dead after 10 months.

The copepod community composition differed substantially after various periods of
20 anoxia, clearly demonstrating that anoxia governs community composition and that species-specific responses provide the baseline for the overall response. Analysing the copepod community at morphospecies level was therefore a rigorous and valid approach. The genera *Pseudobradya*, *Heteropsyllus*, *Nitocrella* and *Longipedia* showed a fast mortality after 9 days of anoxia. The sensitivity of *Pseudobradya* was previously described by Modig and Ólafsson (1998) from the Baltic Sea. Representatives
25 of the family Cletodidae, in contrast, were the most resistant to anoxia, dominating the community in all anoxia treatments. In terms of morphology, species of the genera *Cletodes*, *Stylicletodes*, *Enhydrosoma* and *Acrenhydrosoma* are characterised by a slender habitus (Boxshall and Hasley, 2004). A slender body shape favours a fast

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turnover between ambient oxygen concentrations and copepod “internal” oxygen concentration, a process taking a few seconds in nematodes (Braeckman et al., 2013). Moreover, certain species may survive due to a reduced metabolism in suboxic conditions, and consequently have a longer life span (Braeckman et al., 2013). Cletodids are active mud-burrowers known mainly from shallow and sublittoral marine habitats, whereas some cletodid genera (e.g. *Pyrocletodes*, *Mesocletodes*) also occur in the deep sea (Coull, 1973; Boxshall and Hasley, 2004). The latter points to an evolutionary line of adaptation to environments with low oxygen input. Both environments – shallow hypoxic coastal seas and deep sea – have in common that (1) oxygen in the interstitial spaces is a limiting factor, (2) fine-grained sediments dominate and (3) the food input largely depends on allochthonous material. The genera *Cletodes* and *Enhydrosoma* that were identified as last survivors (2 month anoxia) in the present study are typical for muddy sediments (Hicks and Coull, 1983). In addition, the genus *Asellopsis* (family Laophontidae) survived as anoxia was induced (including 1 month of anoxia).

With increasing duration of anoxia, the overall density (per 16.6 cm²) dropped from 56.3 ± 11 (9 day anoxia) to 16.7 ± 5 (1 month anoxia) but the dominance remained stable (Fig. 3c). This shows that no single species dominated the community. This differs from the diversity changes resulting from impacts such as eutrophication (Pearson and Rosenberg, 1978); fish farming, harbour pollution and sewage outfall (Vrišer, 1984; Moore and Bett, 1989; Sandulli and De Nicola, 1991; Mazzola et al., 1999; Gee, 2005; Tomassetti et al., 2009). Even if the family Cletodidae clearly dominated the communities in anoxic treatments, the Cletodidae species (6 species) were still equally represented by similar number of individuals (see stable evenness). As expected, the overall species richness decreased significantly from normoxia to 9 day anoxia, to 1 month and 2 month anoxia. This shows that certain species are more tolerant to both low oxygen and hydrogen sulphide (see Metzger et al., 2013). This helps interpret the recovery results of the copepod community soon after re-oxygenation (i.e. termination of the deployment): the surviving species have the potential to create a new community, because colonisation was minimal.

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The resilience of the remaining species, and thus the resilience of the community, will largely depend on the reproductive success of these species. Our results did in fact show a substantial and rapid increase in the number of gravid females after 7 days of recovery. An organism’s ability to increase its reproductive potential is known to be positively affected by extrinsic factors such as temperature, nutrition and salinity (see studies cited in Hicks and Coull, 1983). Our results indicate that also the increase in oxygen levels after anoxia stimulated the reproduction of copepods, in particular in Cletodidae. Even though they were not dominant in normoxia, they had the largest number of gravid females compared to other families in all treatments, including normoxia. Reproduction in harpacticoids generally occurs year-round (Hicks and Coull, 1983), but this suggests that our experiments coincided with their reproduction peak. In contrast, two sympatric species of *Enhydrosoma* (Cletodidae) were found to have a different cycle of seasonal abundance, with one having the peak in summer and the other in autumn and spring (Ivester and Coull, 1977). Independent of the overall density of Cletodidae, the number of gravid females increased considerably during recovery and was even higher than in the normoxic samples. According to Pearson and Rosenberg (1978), those species that have a potential to colonise empty spaces are those that can reproduce rapidly.

Autoecologically, this could mean that the most resistant species are also characterised by a high resilience because they adapt quickly to a changing environment, i.e. rise in the oxygen level. In this case, resilience included rapid reproduction in order to establish a new population. Those species possessing this strategy will largely govern overall community resilience. In the present study, only species of the family Cletodidae survived and reproduced quickly. Elsewhere, the eggs of the planktonic *Acartia bifilosa* were also found to survive long periods of anoxia in dormancy, and started to hatch when the oxygen concentration rose > 0.17 mL L⁻¹ (Katajisto, 2004). Fast reproduction rates have been reported for other copepod species (Hicks, 1984; Matias-Peralta et al., 2005; Zaleha and Busra, 2012, and references therein), but not in combination with a strong resistance to stressors such as anoxia. This is a unique and important

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combination in terms of the functional consequences of low biodiversity after an anoxic event. Also, only few surviving copepodites were found in anoxia, and slightly more in the recovery phase, which points to their higher sensitivity compared to the adults. These results are supported by Veit-Köhler et al. (2009), who found fewer copepodites and no nauplii in oxygen minimum zones.

In addition to rapid reproduction, harpacticoid copepods also need a specific feeding ecology in order to (1) cope with low oxygen levels and (2) gain sufficient energy as soon as anoxic conditions end. Several recent isotope measurements of Cletodidae (De Troch et al., unpubl. data) showed remarkably low and thus very depleted ^{13}C values (as low as $\delta^{13}\text{C} = -30$ to -40‰). This suggests that Cletodidae are specialists feeding almost exclusively on chemoautotrophic bacteria. Additionally, Cletodidae may use a trophic pathway completely separate from the trophic cascade (typically depending on phototrophic primary production). Chemoautotrophic bacteria are mainly known from symbiosis with deep-sea animals (Cavanaugh, 1983; Jannasch and Mottl, 1985), but are also ectosymbionts on the benthic marine nematode *Leptonemella* in the North Sea (Riemann et al., 2003). These symbioses are interpreted as a strategy to migrate across the sulfur chemocline and may play a nutritional role for the host as well (Ott et al., 1991). Chemoautotrophic bacteria are also predominant in close proximity of intertidal macrofaunal burrows (Aller et al., 1983) and meiofaunal burrows (Reise, 1981). Here, the dependence on chemoautotrophic bacteria again interlinks our results with certain features of the deep sea. This makes specimens of the family Cletodidae – an evolutionary line represented in both the deep sea and shallow seas – clearly the “winners” of anoxia due to their specific feeding ecology and the reproductive success/rate as soon as oxygen levels increase again.

Beyond the Cletodidae, other harpacticoid copepod families were also found live after 7 d recovery. In terms of composition, however, this community resembled that of 9 day anoxia rather than of normoxia. We interpret this to reflect no species colonising from the nearby undisturbed, normoxic sediment. This is a surprising result, and may be due to a low motility of copepods from the nearby sediment and/or because the im-

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pacted sediment remained toxic for other species due to high sulphide concentrations (Bagarinao, 1992; Middelburg and Levin, 2009; Metzger et al., 2013). However, De Troch et al. (2013) showed that the reduced grazing pressure during anoxia promotes post-anoxia food availability (e.g. elevated concentrations of the fatty acid 16:1 ω 7, typical for diatoms). This organic matter and the bacteria decomposing it represent an important food supply for organisms in the recovery phase. The recovery data suggest that only the surviving specimens benefited from this because the number of colonisers is negligible. Similarly, Langlet et al. (2013b) found an increased abundance of benthic Foraminifera in the 1 month anoxia as a consequence of increased food availability due to (a) macrofauna mortality (b) reduced grazing pressure of survivors (see De Troch et al., 2013). Clearly, regardless of the mechanism and strategy, recovery in harpacticoid copepods and foraminifera is much more rapid than in the macrofauna (Blasnig et al., 2013).

In terms of population dynamics, the male/female ratio remained relatively stable, independent of oxygen level. This demonstrates similar survival levels in both sexes. Females, representing > 60 %, dominated in all treatments. Females also typically outnumber males in field samples (see Hicks and Coull, 1983 and references herein), and anoxia apparently does not influence this.

From a shorter (5 days) but similar experimental set-up in 2009, De Troch et al. (2013) concluded that harpacticoid copepod diversity on the family level was unaffected by anoxic conditions. The families Ectinosomatidae and Cletodidae were again most abundant in both normoxic and anoxic samples. The same authors also observed an increasing trend in relative abundance of Cletodidae in anoxia. At species level (this study), however, there were significant differences between normoxia and anoxia, possibly due to the lengthier anoxia (and by identifying all the copepods rather than a subsample, yielding higher diversity indices).

The copepod community also differed with depth. With increasing duration of anoxia, however, communities (diversity and density) typical for deeper layers of oxygenated sediments become similar to communities typical for higher layers of less oxygenated

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sediments. For example, the copepod assemblage in the second (0.5–1 cm) layer of normoxia was most similar to that from the first (0–0.5 cm) layer of 9 day anoxia and 7 day recovery. Also, the 0.5–1 cm depth layer of 9 day anoxia (and 7 day recovery) was similar to the top (0–0.5 cm) layer of 1 month anoxia. We interpret this with the vertical upward migration of copepods in the sediment as oxygen depletion continues.

Interestingly, despite frequent oxygen depletion events in the Northern Adriatic, the region is characterised by a high diversity of harpacticoid copepods (Vrišer, 2000a, b), one of the highest in European seas (Veit-Kohler et al., 2010). This highly diverse (in terms of sensitivity and tolerance) community is apparently able to survive shorter anoxic events relatively well and shows a high degree of resilience, whereby species of the family Cletodidae are best adapted to decreasing oxygen concentrations.

5 Conclusions

The in situ deployment of four plexiglass chambers during 9 days, 1 month, 2 months and 10 months allowed us to study in detail the response of harpacticoid copepods to anoxic events in the natural setting. After a short-term (9 days) anoxic event, several species (*Pseudobryadia* sp. 2, *Heteropsyllus* sp. 1, *Nitocrella* sp. 1 and *Longipedia* sp. 2.) died and were therefore classified as the “losers” in anoxia. This led to a very fast drop in harpacticoid density and species richness. As anoxia persisted, the relative abundance of Cletodidae increased, with some representatives surviving 1 and even 2 months. Based on the diversity patterns and stable evenness, all the species of this family were tolerant to anoxia (9 days and 1 month). This differs from the typical one-species communities after other impacts (e.g. eutrophication), underlining the strength of a species-based response analysis.

Combination with current knowledge on the ecology of Cletodidae, we conclude that these “winners” are very well adapted to anoxia in terms of (1) high resistance (see survival), (2) specific feeding ecology (stable isotope data, De Troch unpubl.) and (3) high resilience (rapid reproduction).

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In terms of recovery, we recorded no significant increase in density but a slight increase in species diversity. This points to minimal colonisation from the nearby sediment: mainly the survivors guaranteed the recovery (based on the rapid increase in the number of gravid females). This will be important in the larger-scale anoxias that characterize most so-called dead zones. Our findings also underline that, in the recovery phase, a substantial amount of energy is allocated to reproduction.

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Table 1. SIMPER analysis – average abundances of species within a group (treatment) and species cumulative contribution to dissimilarity. N = normoxia, A = anoxia, duration of treatment: 9 d (9 days), 1 m (1 month), 2 m (2 months).

Species	Group normoxia Av. Abund	Group 9d anoxia Av. Abund	Cum.%
<i>Pseudobradia</i> sp. 2	18	1.67	10.84
<i>Heteropsyllus</i> sp. 1	13.67	1.67	19.23
<i>Nitocrella</i> sp. 1	9.67	0.67	25.52
<i>Longipedia</i> sp. 2	6.33	0	30.29
<i>Stylicletodes</i> sp. 2	3.33	9	34.76
<i>Normanella</i> sp. 1	7.67	1.33	39.22
<i>Idyanthe</i> sp. 1	6.33	0	43.62
<i>Cletodes</i> sp. 4	6.33	7	47.62
<i>Pseudobradia</i> sp. 1	5.67	0.33	51.2
<i>Stenhelia (Delavalia)</i> sp. 1	5	0	54.76
<i>Asellopsis</i> sp. 1	5.67	1	58.25
<i>Mesochra</i> sp. 1	4.67	0.33	61.15
<i>Cletodes</i> sp. 2	7	9.33	63.83
<i>Proameira</i> sp. 1	4	0	66.51
<i>Ectinosoma</i> sp. 1	3.33	0	69.06
<i>Longipedia</i> sp. 1	3.67	0	71.61
<i>Cletodes</i> sp. 1	1	4	73.7
<i>Ameira</i> sp. 1	2.33	0	75.63
<i>Cletodes</i> sp. 3	3.67	4.33	77.55
<i>Haloschizopera</i> sp. 2	4	1.33	79.47
<i>Robertsonia</i> sp. 1	3.67	1	81.35
<i>Proameira</i> sp. 2	2.33	0	83.04
<i>Rhizothrix</i> sp. 1	2.67	0.67	84.66
<i>Vibriopsyllus</i> sp. 1	3	1.33	86.09
<i>Stylicletodes</i> sp. 1	2.33	1	87.45
<i>Enhydrosoma</i> sp. 2	0.33	2	88.61
<i>Typhlamphiascus</i> sp. 1	2	1.33	89.72
<i>Cletodes</i> sp. 6	3	2.33	90.81

12408

Table 1. Continued.

Species	Group 9d anoxia Av. Abund	Group 1 m anoxia Av. Abund	Cum.%
<i>Stylicletodes</i> sp. 2	9	0.33	17.86
<i>Cletodes</i> sp. 2	9.33	2.67	31.88
<i>Cletodes</i> sp. 4	7	2.33	41.07
<i>Cletodes</i> sp. 1	4	0.33	48.01
<i>Asellopsis</i> sp. 1	1	3.33	54.17
<i>Enhydrosoma</i> sp. 2	2	0	58.35
<i>Cletodes</i> sp. 6	2.33	1	62.48
<i>Cletodes</i> sp. 3	4.33	3.33	65.96
<i>Enhydrosoma</i> sp. 4	0	1.67	69.3
<i>Pseudobradya</i> sp. 2	1.67	0	72.4
<i>Heteropsyllus</i> sp. 1	1.67	0	75.31
<i>Typhlamphiascus</i> sp. 1	1.33	0	78.04
<i>Vibriopsyllus</i> sp. 1	1.33	0	80.77
<i>Haloschizopera</i> sp. 2	1.33	0	83.32
<i>Normanella</i> sp. 1	1.33	0.67	85.7
<i>Robertsonia</i> sp. 1	1	0	87.88
<i>Acrenhydrosoma</i> sp. 1	0	1	89.98
<i>Stylicletodes</i> sp. 1	1	0	91.62

12409

Table 1. Continued.

Species	Group 1 m anoxia Av. Abund	Group 2 m anoxia Av. Abund	Cum.%
<i>Cletodes</i> sp. 3	3.33	0	19.13
<i>Asellopsis</i> sp. 1	3.33	0	35.11
<i>Cletodes</i> sp. 4	2.33	0	50.4
<i>Cletodes</i> sp. 2	2.67	0.5	61.1
<i>Enhydrosoma</i> sp. 4	1.67	0	71.17
<i>Acrenhydrosoma</i> sp. 1	1	0	78.32
<i>Cletodes</i> sp. 6	1	0.5	85.28
<i>Enhydrosoma</i> sp. 2	0	1	91.35

12410

Table 2. Number of gravid females (sum of all cores) of various families treatments, and number of copepodite stages in different treatments. Gravid females absent in 2 months and 10 months anoxia treatments. Relative amount of females (F) and males (M) averaged over three replicates is also given (note in 2 month anoxia calculated from 4 individuals present in two replicates). N = normoxia, A = anoxia, R = recovery, duration of treatment: 7 d (7 days), 9 d (9 days), 1 m (1 month), 2 m (2 months).

	Normoxia	Anoxia 9d	Recovery 7d	Anoxia 1m	A2m
Ameiridae	3		1		
Argestidae					
Canthocamptidae	1				
Canuellidae					
Cletodidae	15	3	22	2	
Dactylopusiidae					
Ectinosomatidae	1				
Ectinosomatidae	2				
Idyanthidae	1				
Laophonthopsidae					
Laophontidae					
Longipedidae	1				
Miraciidae	3		2		
Normanellidae	1				
Rhizothricidae					
Zosimeidae					
copepodites	135	7	17		
F (%)	0.63	0.60	0.71	0.72	0.60
M (%)	0.37	0.40	0.29	0.28	0.40

12411

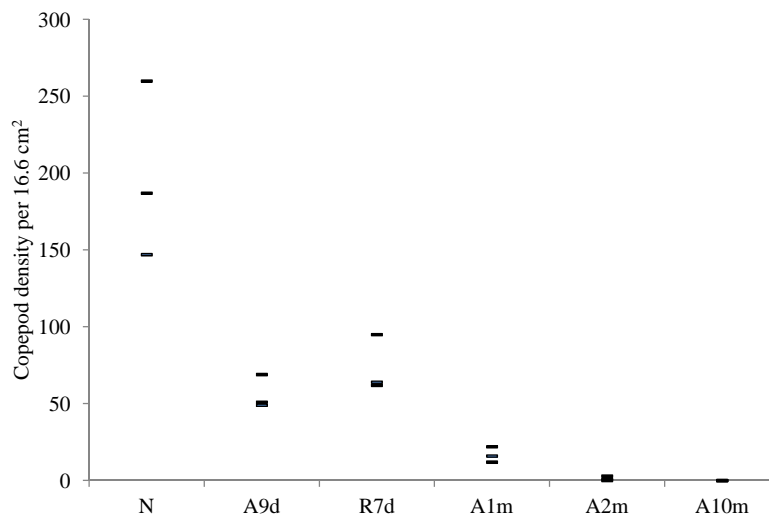


Fig. 1. Absolute copepod densities (ind. per 16.6 cm⁻²) in all samples (pooled depths, copepodites included).

12412

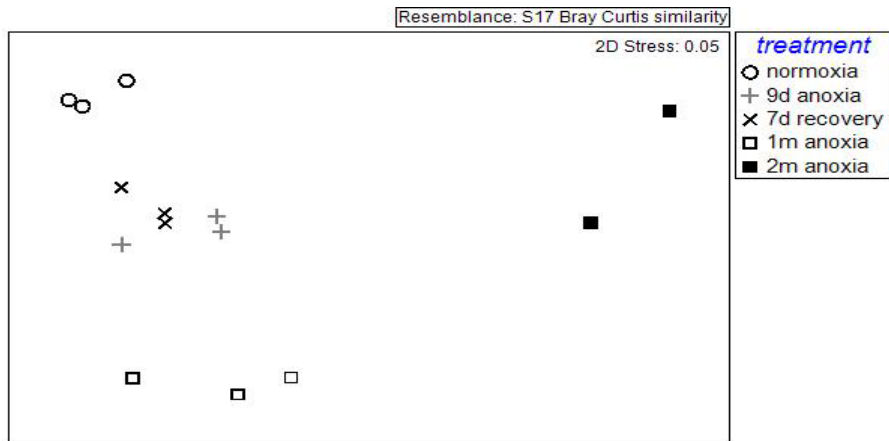


Fig. 2. MDS based on absolute harpacticoid copepod composition (depth pooled, without copepodites).

12413

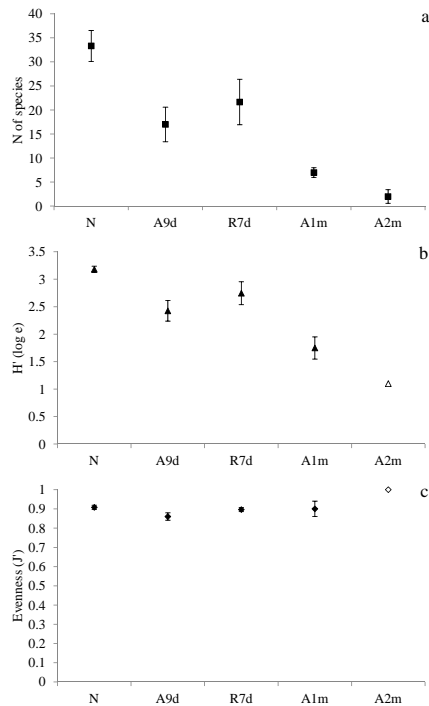


Fig. 3. Diversity indices **(a)** Species richness, **(b)** Shannon diversity index (A2m; calculated only from one replicate containing 3 specimens as in the other replicate a single individual was present) **(c)** Pielou's evenness index (A2m; same as for Shannon diversity index). Depth was pooled, copepodites excluded from calculations. N = normoxia, A = anoxia, R = recovery, duration of treatment: 7 d (7 days), 9 d (9 days), 1 m (1 month), 2 m (2 months).

12414

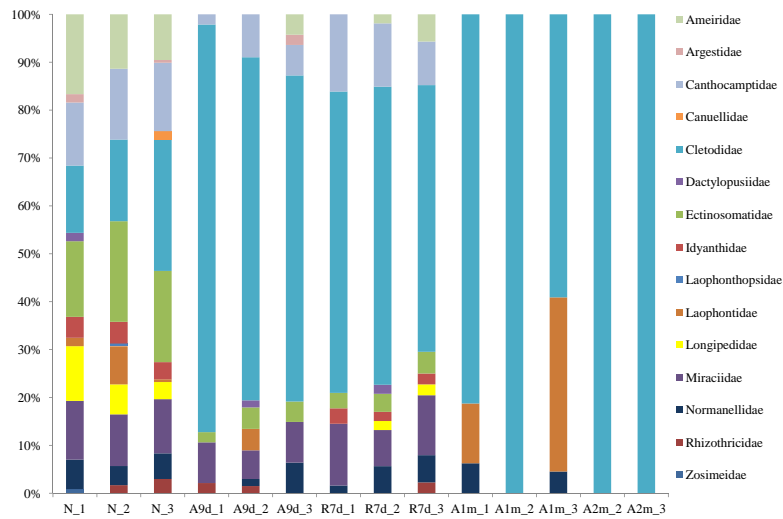


Fig. 4. Relative composition of copepod families in the different replicates (three per treatment, only 2 replicates for the anoxia 2 month treatment). N = normoxia, A = anoxia, R = recovery, duration of treatment: 7 d (7 days), 9 d (9 days), 1 m (1 month), 2 m (2 months), replicates indicated by 1, 2, 3. Families listed alphabetically, and plotted in the column in same order.

12415

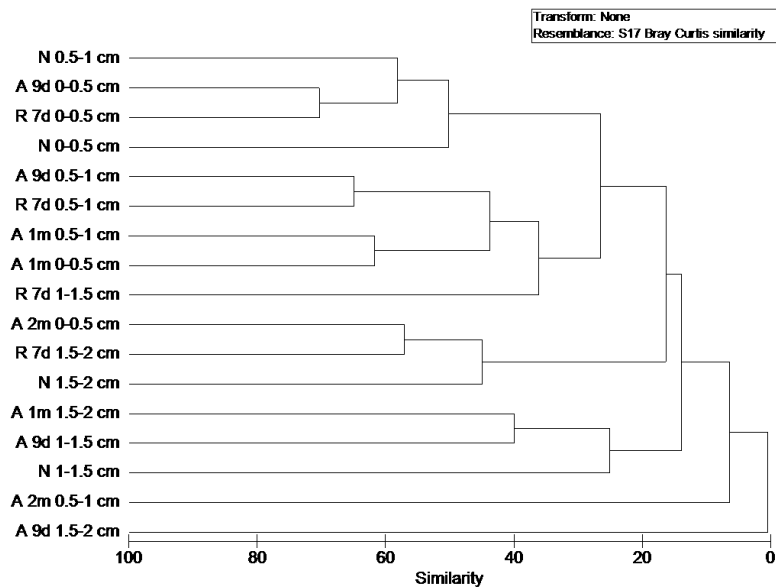


Fig. 5. Dendrogram of the Bray–Curtis similarity between samples (different treatments and different depths). N = normoxia, A = anoxia, R = recovery, duration of treatment: 7 d (9 days), 1 m (1 month), 2 m (2 months). Triplicates averaged per depth layer.

12416