

Symbiosis increases coral tolerance to ocean acidification

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Symbiosis increases coral tolerance to ocean acidification

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Abstract

Increasing the acidity of ocean waters will directly threaten calcifying marine organisms such as reef-building scleractinian corals, and the myriad of species that rely on corals for protection and sustenance. Ocean pH has already decreased by around 0.1 pH units since the beginning of the industrial revolution, and is expected to decrease by another 0.2–0.4 pH units by 2100. This study mimicked the pre-industrial, present, and near-future levels of $p\text{CO}_2$ using a precise control system ($\pm 5\%$ $p\text{CO}_2$), to assess the impact of ocean acidification on the calcification of recently-settled primary polyps of *Acropora digitifera*, both with and without symbionts, and adult fragments with symbionts. The increase in $p\text{CO}_2$ of $100\ \mu\text{atm}$ between the pre-industrial period and the present had more effect on the calcification rate of adult *A. digitifera* than the anticipated future increases of several hundreds of micro-atmospheres of $p\text{CO}_2$. The primary polyps with symbionts showed higher calcification rates than primary polyps without symbionts, suggesting that (i) primary polyps housing symbionts are more tolerant to near-future ocean acidification than organisms without symbionts, and (ii) corals acquiring symbionts from the environment (i.e. broadcasting species) will be more vulnerable to ocean acidification than corals that maternally acquire symbionts.

1 Introduction

As humans are continuing to burn fossil fuels at an unprecedented rate, the concentration of CO_2 in the atmosphere is presently higher than it has been for the last 420 000 yr (Hoegh-Guldberg et al., 2007; IPCC, 2007). The oceans uptake a large proportion of that CO_2 , forcing them toward more acidic conditions (i.e., with high $p\text{CO}_2$), threatening the very foundation of calcifying marine organisms and coral reefs (Kleypas et al., 2006; Orr et al., 2005; Raven et al., 2005). Indeed, coral reefs support a wealth of calcifying organisms, of which scleractinians corals have been the most essential reef builder since the Triassic (Stanley and Fautin, 2001).

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Since the pre-industrial period, we have witnessed a steady increase in $p\text{CO}_2$ concentrations around $100 \mu\text{atm}$, which is predicted to reach $200\text{--}700 \mu\text{atm}$ above present values ($400 \mu\text{atm}$) by 2100 (IPCC, 2007). Such an increase in $p\text{CO}_2$ concentrations reduce both the pH and the concentration of carbonate ions in the water column, and increase the availability of bicarbonate ions (Kleypas et al., 1999). Several studies have found that coral calcification rates are directly related to the concentration of carbonate ions in the water column (Anthony et al., 2008; Gattuso et al., 1998; Kleypas et al., 2006; Marubini et al., 2008), whereas other studies have shown a positive relationship between coral growth rates and the availability of bicarbonate ions (Jury et al., 2010). Therefore, the response of coral growth and the state of the ocean's carbonate chemistry is under intensive investigation (Pandolfi et al., 2011).

Moreover, the oceans are not homogeneous, and the temperature gradient from the tropics to the poles sets carbonate ion concentrations naturally higher in the tropics where coral reefs occur. Nevertheless, the decrease in carbonate ion concentrations from the pre-industrial period to the present has been greater in the tropics ($\sim 29 \mu\text{molkg}^{-1}$) than in the Southern Ocean ($\sim 18 \mu\text{molkg}^{-1}$) (Orr et al., 2005). Yet, symbiosis is prolific in the tropics, and the self-extending symbiosis theory tells us that organisms harboring symbionts should be more tolerant to environmental change than organisms without symbionts (i.e., aposymbiotic organisms) (Kitano, 2004; Kitano and Oda, 2006). These assertions lead to two pertinent questions: (i) will calcifying coral species survive in high $p\text{CO}_2$ seawater? And (ii) are juvenile corals, without symbionts, more vulnerable to high $p\text{CO}_2$ seawater than juveniles and adult corals with symbionts?

Recent experiments that have mimicked the near-future $p\text{CO}_2$ conditions on coral reefs have either adjusted the pH of seawater by adding an acid or a base, or by bubbling CO_2 through the seawater in experimental chambers (Atkinson and Cuet, 2008). Adding an acid or a base results in seawater with different alkalinity, bicarbonate, and carbonate ion concentrations than when CO_2 is bubbled through seawater (Atkinson and Cuet, 2008). Although bubbling CO_2 through the seawater more closely reflects near-future conditions than adding acids, it is nevertheless difficult to achieve a stable

$p\text{CO}_2$ environment, especially in flow-through systems (e.g. Leclercq et al., 2002; Suwa et al., 2010; Takahashi and Kurihara, 2013). To overcome these problems, our research group developed a system that produced stable $p\text{CO}_2$ concentrations in flow-through conditions (Fujita et al., 2011).

5 Using this system, we examined the effect of $p\text{CO}_2$ -adjusted seawater on the calcification rates of *Acropora digitifera*, one of the most common corals in the Pacific Ocean. Calcification was examined in five $p\text{CO}_2$ treatments: (i) pre-industrial $p\text{CO}_2$, < 300 μatm , (ii) present-day $p\text{CO}_2$, 400 μatm , and at three near-future conditions, (iii) 600 μatm , (iv) 800 μatm , and (v) 1000 μatm . Within these treatments, we investigated
10 the response of: (1) primary aposymbiotic coral polyps (i.e., without symbionts), (2) primary symbiotic polyps, and (3) adult symbiotic fragments. It was hypothesized that the calcification process of symbiotic corals was more tolerant to $p\text{CO}_2$ adjustments than aposymbiotic corals.

2 Materials and methods

15 2.1 Experimental setup

To produce $p\text{CO}_2$ -adjusted seawater, we used a precise $p\text{CO}_2$ control system (Fujita et al., 2011). This system was used to generate five different $p\text{CO}_2$ levels, including one lower than the present level of atmospheric $p\text{CO}_2$: (i) pre-industrial, < 300 μatm , (ii) present-day $p\text{CO}_2$, 400 μatm , and at three near-future conditions, (iii) 600 μatm , (iv) 800 μatm , and (v) 1000 μatm . The $p\text{CO}_2$ -adjusted seawater was supplied to duplicate
20 flow-through (150 mL min⁻¹) aquaria systems (12 L). The seawater temperature was maintained at 27 °C, with a 12:12 h light:dark photoperiod (of 75 $\mu\text{mol m}^{-2} \text{s}^{-1}$) under metal-halide lamps (Funnel2 150 W, Kamihata, Japan) throughout all treatments. The aragonite saturation state of the seawater was estimated using the CO₂SYS program
25 (Lewis and Wallace, 1998) and the variables: temperature, pH, mean salinity, and total

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alkalinity were measured repeatedly during the experiments. The chemical and physical conditions of each $p\text{CO}_2$ treatment are summarized in Tables 1 and 2.

2.2 Primary polyp experiment

Several 20 cm *A. digitifera* colonies were collected from a fringing reef of Sesoko Island. Gametes from two colonies, which spawned on 29 May 2010 were combined in a flow-through aquarium, from which we derived several hundred planulae larvae. Primary polyps were prepared following the methods outlined in our previous report (Suwa et al., 2010) using 13 day old planulae. To prepare the symbiotic primary polyps, primary polyps of *A. digitifera* were infected with the dinoflagellate *Symbiodinium* that were derived from the giant clam *Tridacna crocea* (using a solution of 4×10^5 cells mL^{-1}) because the primary polyps could acquire algae from this bivalve more efficiently than from other hosts, including *Acropora* species (Hirose et al., 2008). Four days after inducing metamorphosis, primary polyps were exposed to the symbiont solution for one day. Three days after exposure to the symbiont solution, we confirmed symbiont infection using a dissecting microscope. The primary polyps, both with and without symbionts, were subjected to four $p\text{CO}_2$ treatments: (i) pre-industrial, $< 300 \mu\text{atm}$, (ii) present day $p\text{CO}_2$, $400 \mu\text{atm}$, (iii) $800 \mu\text{atm}$, and (iv) $1000 \mu\text{atm}$.

Eight 6-well culture plates, containing the settled primary polyps, were placed into each aquarium (i.e., 4 plates for aposymbiotic primary polyps, and 4 plates for symbiotic primary polyps) during 10 days. Twenty polyps per treatment were used to evaluate skeletal growth of polyps. At the end of the experiment, soft tissues were removed from each polyp with a water-pik. The dry weight of each polyp skeleton was measured according to Inoue et al. (2011). The dry weight (μg) of the polyp skeleton, at the end of the experiment, was used to represent the amount of growth of each coral during the experiment.

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2.3 Adult-coral-fragment experiment

Five > 30 cm colonies of *Acropora digitifera* were collected in August 2009 from a shallow (2 m) fringing reef at Sesoko Island, Okinawa, Japan. The colonies which were growing at least 10 m apart were haphazardly selected. The *A. digitifera* colonies were kept in a flow-through aquarium for 3 weeks under natural light conditions at Sesoko Station, Tropical Biosphere Research Center, University of the Ryukyus (Okinawa, Japan). Fifty, 2–3 cm fragments were cut from each parent colony and attached to plastic bolts with superglue. The fragments were kept in a flow-through aquarium for 2 weeks under natural light conditions until the coral tissues started to spread over the surfaces of the plastic bolts. Five of these fragments, from each parent colony, were maintained for 6 weeks in each of ten aquaria to which $p\text{CO}_2$ -adjusted seawater was supplied using the flow-through system (two aquaria per $p\text{CO}_2$ treatment).

The weight of each colony was measured as buoyant weight (Spencer-Davies, 1989), which directly reflects skeletal weight (Anthony et al., 2008). The calcification rate was calculated as the percentage change in final weight relative to the initial weight, during the 6-week experiment. During the adult fragment experiment, 29 fragments died and were excluded from the calcification analysis. To evaluate the photosynthetic fitness of zooxanthellae in the adult fragments, the symbionts' maximum photosynthetic quantum yields (F_v/F_m) was measured after 6 weeks using a Diving-PAM Underwater Fluorometer (Walz, Germany) after at least 1 h of darkness.

2.4 Data analysis

The dry weights of the primary polyp skeleton were analyzed using an ANOVA model with $p\text{CO}_2$ as fixed-effect factors. The subsequent pairwise comparisons among different $p\text{CO}_2$ levels were performed using Tukey's HSD tests ($\alpha = 0.01$). In order to perform statistical analysis on adult calcification, we used a general linear model to estimate the response of calcification to: $p\text{CO}_2$ (fixed-effect factor), aquarium (nested within $p\text{CO}_2$; fixed-effect factor), colony (fixed-effect factor), initial weight (covariate),

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and their interactions ($p\text{CO}_2 \times$ initial weight, colony \times initial weight, colony $\times p\text{CO}_2$, colony $\times p\text{CO}_2 \times$ initial weight). The result of the F -tests (based on type-III sum of squares) and stepwise backward model selection, suggested that only $p\text{CO}_2 \times$ initial weight, and colony \times initial weight remained as statistically significant interactions (each $\alpha = 0.05$). To remove the variation of covariates, we calculated the adjusted mean final weights relative to the mean initial weight for each colony, assuming that their regression lines were heterogeneous among all the combinations of colony and $p\text{CO}_2$. The adjusted final weight (W_{AFIN}) for each colony was independently analyzed using an ANOVA model with $p\text{CO}_2$ (fixed-effect factor) and aquarium (nested within $p\text{CO}_2$; fixed-effect factor) as the independent fixed factors. Statistically significant factors ($\alpha = 0.01$) were subjected to pairwise comparisons (Tukey's HSD tests; $\alpha = 0.01$) to specify significant combinations of treatment levels. Arcsine transformed F_v/F_m values of adult fragments were analyzed using an ANOVA model with $p\text{CO}_2$ as fixed-effect factors. The subsequent pairwise comparisons among different $p\text{CO}_2$ levels were performed using Tukey's HSD tests ($\alpha = 0.01$).

3 Results

At 300 and 400 μatm the skeletal weights of aposymbiotic primary polyps were significantly heavier (posteriori Tukey HSD test, $p < 0.01$) than those at future-level treatments (at 800 and 1000 μatm). The primary polyps with symbionts showed a different response. Firstly, the skeletal weights of primary polyps with symbionts were heavier than those for primary polyps without symbionts. Secondly, the skeletal weights of primary polyps with symbionts were heaviest in the pre-industrial, 300 μatm treatment, and significant difference was observed only between 300 and 1000 μatm treatments (posteriori Tukey HSD test, $p < 0.01$), whereas aposymbiotic primary polyps showed no differences between pre-industrial and contemporary treatments (Fig. 1).

We evaluated the calcification rates of adult fragments of *Acropora digitifera* under five $p\text{CO}_2$ treatments. ANOVA on the adult fragment weight adjusted for initial size

variation indicated that a higher $p\text{CO}_2$ leads to significantly slower growth rates in four out of five colonies (Colony b–e; Fig. 2; Table S3). The analysis also suggested that the potential environmental differences between two replicate aquaria were negligible in all five colonies (all $p > 0.05$). The subsequent Tukey's HSD tests indicated that the mean final weight of adult fragments, reared at $300 \mu\text{atm}$, was significantly greater than those at the other $p\text{CO}_2$ conditions in all of the four colonies, showing significant $p\text{CO}_2$ effects (Fig. 2; Table S3). The maximum photosynthetic efficiencies of the adult fragments were approximately constant, and did not differ significantly among $p\text{CO}_2$ treatments (Fig. 2; Table S4).

4 Discussion

The differences in the skeletal weights between primary polyps with and without symbionts might reflect the difficulty aposymbiont corals have in acquiring resources or acquiring a substrate medium for calcification. Yet, why would the primary polyps with symbionts be more responsive to pre-industrial treatments than aposymbiotic primary polyps? The increase in calcification in the pre-industrial $p\text{CO}_2$ treatment only occurred in corals that housed symbionts. Indeed, the adult colonies showed the same response as primary polyps with symbionts, clearly increasing calcification rates in low $p\text{CO}_2$ treatments. Moreover, the calcification rates of symbiotic adult *A. digitifera* fragments were higher in the pre-industrial seawater $p\text{CO}_2$ treatment than in the present-day $p\text{CO}_2$ treatment.

Higher calcification in the pre-industrial $p\text{CO}_2$ treatment was most likely attributed to a change in skeletal precipitation by the coral host, because there was no evidence of any differences in the maximum photosynthetic quantum yield among the symbionts in the different $p\text{CO}_2$ treatments (Fig. 2, Table S4). Still, there were no differences in calcification rates between present day and near-future concentrations (Fig. 2). We note that this lack of difference in calcification between present day and anticipated future $p\text{CO}_2$ treatments was not apparent for primary polyps (Fig. 1). These differences

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suggest a number of potential mechanisms that are not mutually exclusive. First, an increase in calcification in low $p\text{CO}_2$ environments was only apparent in the presence of symbionts. Therefore, such phenotypic plasticity in calcification potential was most likely attributed to the presence of the symbionts. Second, the adult colonies did not respond to higher $p\text{CO}_2$ environments, whereas the primary polyps with symbionts did show reduced calcification rates at high $p\text{CO}_2$. Such results suggest a hierarchical response in tolerance to $p\text{CO}_2$ environments depending on the density of symbionts, from adult colonies with symbionts as the most tolerant, to symbiotic primary polyps showing some tolerance, to primary polyps without symbionts being the least tolerant to high $p\text{CO}_2$ treatments.

There is mounting evidence that symbiotic dinoflagellates facilitate calcification within corals through a positive feedback system between the host and the symbionts (Allemand et al., 2004; Muscatine, 1990; Yellowlees et al., 2008). The glycerol and oxygen produced by the symbionts facilitate calcification through mitochondrial respiration and ATP production which could be used for ion transport (Allemand et al., 2004; Colombo-Pallotta et al., 2010). CO_2 uptake by photosynthesis is also thought to stimulate calcification by changing the equilibrium of dissolved inorganic carbon (DIC) in coral tissue, although the mechanisms are unresolved (Allemand et al., 2004). Our results also indicate that the primary polyps with symbionts grow faster than aposymbiotic polyps (Fig. 1). Although the primary polyps with symbionts seem to be more sensitive to acidified seawater than aposymbiotic polyps (Fig. 1), faster growth induced by symbiosis could compensate for the decrease of calcification by acidified seawater. The reason why coral-algal symbiosis enhances coral calcification is not only attributed to algal photosynthesis but is potentially related to the removal of substances inhibiting calcification, such as phosphates (Allemand et al., 2004).

Previous research indicates that acidified seawater increases the concentration of HCO_3^- , possibly followed by the activation of photosynthesis in coral symbionts (Jury et al., 2010; Marubini et al., 2008). In our experiments, however, there was no evidence that acidified seawater activates the photosynthesis of *Acropora digitifera*. The

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reason why the acidified seawater, with high $p\text{CO}_2$ concentration (1000 μatm), did not affect adult coral calcification and photosynthetic activity is unknown. We suspect that there were obvious advantages from symbiosis. For example, the removal of phosphates would facilitate calcification even in acidified seawater. Irrespective of the cellular mechanism involved, our results clearly showed that corals without symbionts were most vulnerable to $p\text{CO}_2$ increases, whereas corals that housed symbionts were more tolerant.

These results are particularly troublesome because globally $\sim 80\%$ of the scleractinian corals are spawners that acquire symbionts from the “wild” after settlement (Baird et al., 2009). Therefore, one serious, and up to now overlooked, repercussion of ocean acidification is that reef corals are vulnerable to ocean acidification upon first settlement and therefore reefs will experience extensive declines in coral recruitment in the near future. This inference may be particularly evident in the Indian and Pacific Oceans where most corals are spawners that horizontally transfer symbionts (Harrison and Wallace, 1990), acquiring them after settlement. By contrast, newly settled corals may do better in the Caribbean where most corals are brooders and symbionts are maternally (i.e., vertically) acquired, and the planulae are symbiotic (Harrison and Wallace, 1990).

The degree of selective pressure by ocean acidification on newly settled polyps may therefore depend on how rapidly corals are able to support symbionts. Such selective filtering could lead to relative shifts in coral species abundance, changing reefs from those that primarily support spawners, to reefs that primarily support brooders (that maternally acquire symbionts). Similar shifts in species composition have occurred in the Oligocene, when rapidly cooling oceans favored brooding corals over spawning corals in the Caribbean (Edinger and Risk, 1995).

In summary, the increase in $p\text{CO}_2$ of just 100 μatm , between the pre-industrial period and the present, had more effect on the calcification rate of adult *A. digitifera* than the anticipated future increases of several hundreds of micro-atmospheres of $p\text{CO}_2$. Our results also suggest that ocean acidification has been ongoing for over two centuries,

and is therefore not only a near-future problem but ocean acidification is a direct and *present* threat to ocean ecosystems (Talmage and Gobler, 2010). In conclusion, this study showed that the apparent sensitivity of primary polyps to near-future ocean acidification was a consequence of not housing symbionts, and those organisms harboring symbionts, at any life-history stage, are more tolerant to ocean acidification than organisms without symbionts.

Supplementary material related to this article is available online at:
**[http://www.biogeosciences-discuss.net/10/7013/2013/
bgd-10-7013-2013-supplement.pdf](http://www.biogeosciences-discuss.net/10/7013/2013/bgd-10-7013-2013-supplement.pdf)**

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References

- Allemand, D., Ferrier-Pagès, C., Furla, P., Houlbrèque, F., Puvarel, S., Reynaud, S., Tambutté, É., Tambutté, S., and Zoccola, D.: Biomineralisation in reef-building corals: from molecular mechanisms to environmental control, *C. R. Palevol.*, 3, 453–467, doi:10.1016/j.crpv.2004.07.011, 2004.
- Anthony, K. R. N., Kline, D. I., Diaz-Pulido, G., Dove, S., and Hoegh-Guldberg, O.: Ocean acidification causes bleaching and productivity loss in coral reef builders, *P. Natl. Acad. Sci. USA*, 105, 17442–17446, doi:10.1073/pnas.0804478105, 2008.
- Atkinson, M. J. and Cuët, P.: Possible effects of ocean acidification on coral reef biogeochemistry: topics for research, *Mar. Ecol.-Prog. Ser.*, 373, 249–256, doi:10.3354/meps07867, 2008.

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Baird, A. H., Guest, J. R., and Willis, B. L.: Systematic and biogeographical patterns in the reproductive biology of scleractinian corals, *Annu. Rev. Ecol. Evol. S.*, 40, 551–571, doi:10.1146/annurev.ecolsys.110308.120220, 2009.

Colombo-Pallotta, M. F., Rodriguez-Román, A., and Iglesias-Prieto, R.: Calcification in bleached and unbleached *Montastraea faveolata*: evaluating the role of oxygen and glycerol, *Coral Reefs*, 29, 899–907, doi:10.1007/s00338-010-0638-x, 2010.

Edinger, E. N. and Risk, M. J., Preferential survivorship of brooding corals in a regional extinction, *Paleobiology*, 21, 200–219, 1995.

Fujita, K., Hikami, M., Suzuki, A., Kuroyanagi, A., Sakai, K., Kawahata, H., and Nojiri, Y.: Effects of ocean acidification on calcification of symbiont-bearing reef foraminifers, *Biogeosciences*, 8, 2089–2098, doi:10.5194/bg-8-2089-2011, 2011.

Gattuso, J. P., Frankignoulle, M., Bourgeb, I., Romaine, S., and Buddemeier, R. W.: Effect of calcium carbonate saturation of seawater on coral calcification, *Global Planet. Change*, 18, 37–46, doi:10.1016/S0921-8181(98)00035-6, 1998.

Harrison, P. L. and Wallace, C. C.: Reproduction, dispersal and recruitment of scleractinian corals, *Coral Reefs*, 133–207, 1990.

Hoegh-Guldberg, O., Mumby, P. J., Hooten, A. J., Steneck, R. S., Greenfield, P., Gomez, E., Harvell, C. D., Sale, P. F., Edwards, A. J., Caldeira, K., Knowlton, N., Eakin, C. M., Iglesias-Prieto, R., Muthiga, N., Bradbury, R. H., Dubi, A., and Hatziolos, M. E.: Coral-reefs under rapid climate change and ocean acidification, *Science*, 318, 1737–1742, doi:10.1126/science.1152509, 2007.

Hirose, M., Yamamoto, H., and Nonaka, M.: Metamorphosis and acquisition of symbiotic algae in planula larvae and primary polyps of *Acropora spp.*, *Coral Reefs*, 27, 247–254, doi:10.1007/s00338-007-0330-y, 2008.

Inoue, M., Suwa, R., Suzuki, A., Sakai, K., and Kawahata, H.: Effects of seawater pH on growth and skeletal U/Ca ratios of *Acropora digitifera* coral polyps, *Geophys. Res. Lett.*, 38, L12809, doi:10.1029/2011GL047786, 2011.

IPCC: Climate Change 2007: The Physical Science Basis, Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change, edited by: Solomon, S., Qin, D., Manning, M., Chen, Z., Marquis, M., Averyt, K. B., Tignor, M., and Miller, H. L., Cambridge University Press, Cambridge, UK and New York, NY, USA, 996 pp., 2007.

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Jury, C. P., Whitehead, R. F., and Szmant, A. M.: Effects of variations in carbonate chemistry on the calcification rates of *Madracis auretenra* (= *Madracis mirabilis* sensu Wells, 1973): bicarbonate concentrations best predict calcification rates, *Glob. Change Biol.*, 16, 1632–1644, doi:10.1111/j.1365-2486.2009.02057.x, 2010.

5 Kitano, H.: Biological robustness, *Nat. Rev. Genet.*, 5, 826–837, doi:10.1038/nrg1471, 2004.

Kitano, H. and Oda, K.: Self-extending symbiosis: a mechanism for increasing robustness through evolution, *Biological Theory*, 1, 61–66, doi:10.1162/biot.2006.1.1.61, 2006.

Kleypas, J. A., Feely, R. A., Fabry, V. J., Langdon, C., Sabine, C. L., and Robbins, L. L.: Impacts of Ocean Acidification on Coral Reefs and Other Marine Calcifiers: a Guide for Future Research, Report of a Workshop held 18–20 April 2005, St. Petersburg, FL, sponsored by NSF, NOAA, and the US Geological Survey, 88 pp., 2006.

10 Leclercq, N., Gattuso, J. P., and Jaubert, J.: Primary production, respiration, and calcification of a coral reef mesocosm under increased CO₂ partial pressure, *Limnol. Oceanogr.*, 47, 558–564, doi:10.4319/lo.2002.47.2.0558, 2002.

15 Lewis, E. and Wallace, D. W. R.: Program developed for CO₂ system calculations, ORNL/CDIAC-105, Carbon Dioxide Information Analysis Center, Oak Ridge National Laboratory, Oak Ridge, 1998.

Marubini, F., Christine, A. E., Ferrier-Page's, A. E., Furla, P., and Allemand, D.: Coral calcification responds to seawater acidification: a working hypothesis towards a physiological mechanism, *Coral Reefs*, 27, 491–499, doi:10.1007/s00338-008-0375-6, 2008.

20 Muscatine, L.: The role of symbiotic algae in carbon and energy flux in reef corals, *Coral Reefs*, 25, 75–87, 1990.

Orr, J. C., Fabry, V. J., Aumont, O., Bopp, L., Doney, S. C., Feely, R. A., Gnanadesikan, A., Gruber, N., Ishida, A., Joss, F., Key, R. M., Lindsay, K., Maier-Reimer, E., Matear, R., Monfray, P., Mouchet, A., Najjar, R. G., Plattner, G. K., Rodgers, K. B., Sabine, C. L., Sarmiento, J. L., Schlitzer, R., Slater, R. D., Totterdell, I. J., Weirig, M. F., Yamanaka, Y., and Yool, A.: Anthropogenic ocean acidification over the twenty first century and its impact on calcifying organisms, *Nature*, 437, 681–686, 2005.

25 Pandolfi, J. M., Connolly, S. R., Marshall, D. J., and Cohen, A. L.: Projecting coral reef futures under global warming and ocean acidification, *Science*, 333, 418–422, doi:10.1126/science.1204794, 2011.

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Raven, J., Caldeira, K., Elderfield, H., Hoegh-Guldberg, O., Liss, P., Riebesell, U., Shepherd, J., Turley, C., and Watson, A.: Ocean acidification due to increasing atmospheric carbon dioxide, Policy Document 12/05, Royal Society, London, 2005.

Spencer-Davies, P.: Short-term growth measurements of corals using an accurate buoyant weighing technique, *Mar. Biol.*, 101, 389–395, doi:10.1007/BF00428135, 1989.

Stanley Jr., G. D. and Fautin, D. G.: The origins of modern corals, *Science*, 291, 1913–1914, doi:10.1126/science.1056632, 2001.

Suwa, R., Nakamura, M., Morita, M., Shimada, K., Iguchi, A., Sakai, K., and Suzuki, A.: Effects of acidified seawater on early life stages of scleractinian corals (Genus *Acropora*), *Fish. Sci.*, 76, 93–99, doi:10.1007/s12562-009-0189-7, 2010.

Takahashi, A., Kurihara, H.: Ocean acidification does not affect the physiology of the tropical coral *Acropora digitifera* during a 5-week experiment, *Coral Reefs*, 32, 305–314, doi:10.1007/s00338-012-0979-8, 2013.

Talmage, S. C. and Gobler, C. J.: Effects of past, present, and future ocean carbon dioxide concentrations on the growth and survival of larval shellfish, *P. Natl. Acad. Sci. USA*, 107, 17246–17251, doi:10.1073/pnas.0913804107, 2010.

Yellowlees, D., Rees, T. A. V., and Leggat, W.: Metabolic interactions between algal symbionts and invertebrate hosts, *Plant Cell Environ.*, 31, 679–694, doi:10.1111/j.1365-3040.2008.01802.x, 2008.

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Table 1. Summary of mean physical and chemical conditions in each $p\text{CO}_2$ treatment of the primary polyp experiment. Standard deviation is shown for pH_T and $p\text{CO}_2$. The mean salinity and total alkalinity measured repeatedly during the experiments were 34.6 and 2257 mol kg^{-1} , respectively. The value of Ω_{arg} was calculated using total alkalinity and $p\text{CO}_2$. (i) Pre-industrial $p\text{CO}_2$, $< 300 \mu\text{atm}$, (ii) present-day $p\text{CO}_2$, $400 \mu\text{atm}$, and at two near-future conditions, (iii) $800 \mu\text{atm}$, and (iv) $1000 \mu\text{atm}$.

Treatment	Temperature ($^{\circ}\text{C}$)	pH_T at 25°C	$p\text{CO}_2$ (μatm)	Ω_{arg}
i) Pre-industrial	26.9	8.180 ± 0.009	242 ± 13	4.60
ii) Present	27.2	8.032 ± 0.008	390 ± 21	3.54
iii) $800 \mu\text{atm}$	27.2	7.801 ± 0.006	777 ± 9	2.22
iv) $1000 \mu\text{atm}$	27.3	7.743 ± 0.003	944 ± 13	1.93

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Table 2. Summary of mean physical and chemical conditions in each $p\text{CO}_2$ treatment of the adult fragment experiment. Standard deviation is shown for pH_T and $p\text{CO}_2$. The mean salinity and total alkalinity measured repeatedly during the experiments were 34.7 and 2236 mol kg^{-1} , respectively. The value of Ω_{arg} was calculated using total alkalinity and $p\text{CO}_2$. (i) Pre-industrial $p\text{CO}_2$, $< 300 \mu\text{atm}$, (ii) present-day $p\text{CO}_2$, $400 \mu\text{atm}$, and at three near-future conditions, (iii) $600 \mu\text{atm}$, (iv) $800 \mu\text{atm}$, and (v) $1000 \mu\text{atm}$.

Treatment	Temperature ($^{\circ}\text{C}$)	pH_T at 25°C	$p\text{CO}_2$ (μatm)	Ω_{arg}
i) Pre-industrial	27.0	8.143 ± 0.014	279 ± 13	4.21
ii) Present	27.1	8.040 ± 0.015	391 ± 18	3.47
iii) $600 \mu\text{atm}$	27.1	7896 ± 0.033	621 ± 24	2.56
iv) $800 \mu\text{atm}$	27.1	7.793 ± 0.022	842 ± 33	2.05
v) $1000 \mu\text{atm}$	27.1	7.719 ± 0.029	1048 ± 44	1.73

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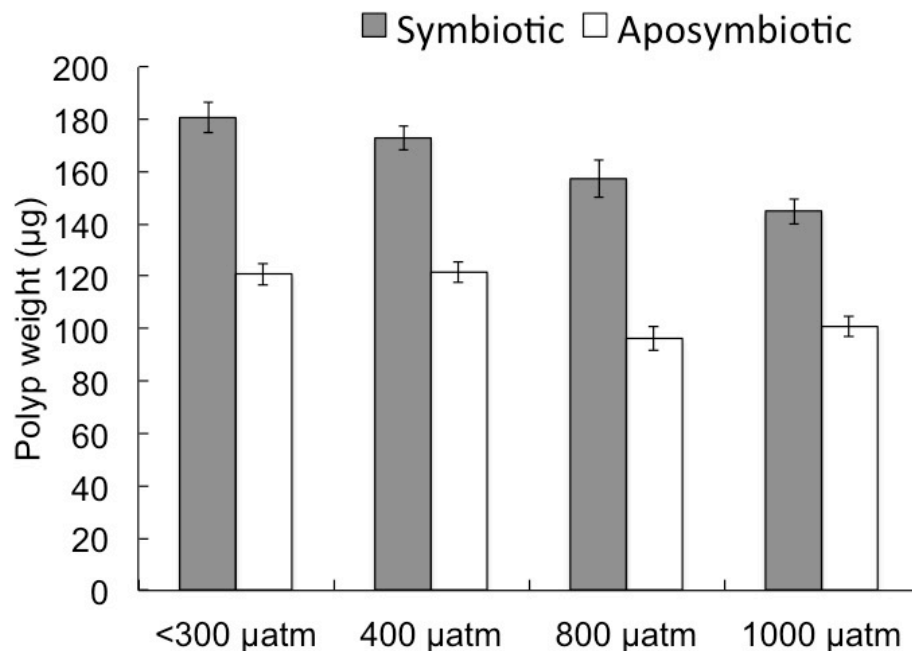


Fig. 1. Skeletal weights of primary polyps of *Acropora digitifera* in the (i) pre-industrial $p\text{CO}_2$, <math><300 \mu\text{atm}</math>, (ii) present-day $p\text{CO}_2$, $400 \mu\text{atm}$, and at two near-future conditions, (iii) $800 \mu\text{atm}$, and (iv) $1000 \mu\text{atm}$. Bars show \pm S.E.

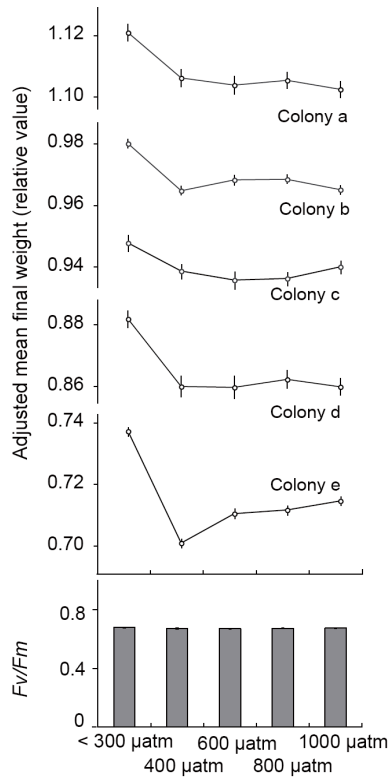


Fig. 2. Adjusted mean final weights of coral fragments and mean F_v/F_m values from five colonies (Colonies a–e) of *Acropora digitifera* in the five $p\text{CO}_2$ treatments (i) pre-industrial $p\text{CO}_2$, <math>< 300 \mu\text{atm}</math>, (ii) present-day $p\text{CO}_2$, $400 \mu\text{atm}$, and at three near-future conditions, (iii) $600 \mu\text{atm}$, (iv) $800 \mu\text{atm}$, and (v) $1000 \mu\text{atm}$. Bars show \pm S.E.

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