

Measuring net ecosystem exchange of CO₂ using EC and chamber techniques

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Comparison between eddy covariance and automatic chamber techniques for measuring net ecosystem exchange of carbon dioxide in cotton and wheat fields

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Abstract

Static and transparent automatic chamber (AC) technique is a necessary choice for measuring net ecosystem exchange (NEE) of carbon dioxide (CO₂) in circumstances where eddy covariance (EC) technique is not applicable. However, a comparison of the two techniques for measurements on croplands has seldom been undertaken. We carried out NEE observations in a cotton field (for one year) and a winter wheat field (for one cropping season) using both AC and EC techniques, to (a) compare the NEE fluxes measured using each technique, and (b) test the NEE measurement performance of an automatic chamber system (AMEG), which was designed for simultaneous flux measurements of multiple gases. The half-hourly NEE fluxes measured with the two techniques were in approximate agreement, with the AC fluxes being 0.78 (cotton) and 1.06 (wheat) times those of the EC. When integrated to daily timescale, the fluxes of the two techniques were in better agreement, showing an average ratio of 0.94 and 1.00 for the cotton and wheat, respectively. During the periods with comparable field conditions and normal performance of both instruments, the cumulative NEE fluxes revealed small differences between the two techniques (−9.0 ~ 6.7%, with a mean of 0.1%). The measurements resulted in annual cumulative NEE of −40 gCm^{−2}yr^{−1} (EC) and −42 gCm^{−2}yr^{−1} (AC) in the cotton field and seasonal cumulative NEE of −251 gCm^{−2} (EC) and −205 gCm^{−2} (AC) in the wheat field. Our results indicate that, for cropland populated by short plants, the AMEG system and the data processing procedures applied in this study are able to provide NEE estimates comparable to those from EC measurements, although either technique may lead to an overestimation of the loss rate (or underestimation of the gain rate) of the soil organic carbon stock of an ecosystem, in particular with calcareous soils exposed to increasing atmospheric acid deposition.

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1 Introduction

The net ecosystem exchange (NEE) of carbon dioxide (CO₂) is the difference between carbon assimilation by photosynthesis and release via ecosystem respiration. CO₂ uptake by the ecosystem is represented by a negative flux and CO₂ loss is represented by a positive flux (e.g., Chapin III et al., 2006). Direct observation of the NEE is a very important step to take in the quantification of the carbon source or sink flux of a terrestrial ecosystem.

There are two main approaches for measuring NEE fluxes: the eddy covariance (EC) technique and the chamber technique. Each of them has advantages and disadvantages. The EC technique is based on micrometeorological theories. It allows for continuous and non-disturbing measurements and provides spatially averaged fluxes on scales of a few hectares to several square kilometers (Baldocchi, 2003). It has been widely used in different ecosystems (e.g., Goulden et al., 1996; Lafleur et al., 1997; Miyata et al., 2000). However, there are restrictions for its usage on small-scale field plots and complex terrains due to its theoretical assumptions (Lee et al., 2004). In addition, data gaps are inevitable with EC measurements due to extensive quality checks, particularly under low-turbulence mixing conditions (Foken and Wichura, 1996). The technique of static chamber in combination with a close-path gas analyzer can also be used for NEE measurement in ecosystems with low-stature canopies, such as grasslands and croplands (e.g., Maljanen et al., 2001; Steduto et al., 2002; Langensiepen et al., 2012). The chamber technique offers the advantages of lower cost and simplicity in principle. Also the chambers are portable and well-suited for small-scale (e.g., 10²–10⁴ m²) studies. In this regard, the chamber technique is appropriate for replicated measurements in multiple small plots of field trails, which are absolutely necessary for identifying the impacts of natural or anthropogenic driving forces upon the NEE and thus net ecosystem carbon balance. However, chamber measurements are prone to a variety of potential errors, such as modifications in the enclosed microclimate, pressure artifacts, and spatial heterogeneity (Livingston and Hutchinson, 1995; Welles

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et al., 2001; Davidson et al., 2002). These drawbacks have limited the application of the chamber technique for NEE measurements; nevertheless, it is an alternative method that can be used when the EC method is not suitable (e.g., Burkart et al., 2007).

Studies comparing the NEE fluxes measured using the EC and chamber techniques are necessary to highlight the potential sources of errors and to test the reliability of chamber fluxes. To date, however, only a few researchers have conducted simultaneous comparison studies. Laine et al. (2006) found that the two techniques had good agreement in measuring the NEE fluxes in a peatland. In a scrub-oak ecosystem, Dore et al. (2003) reported that the NEE fluxes measured by the chambers were generally higher than those measured by the EC by 8–26%. A study by Fox et al. (2008) found an even larger (up to 60%) difference between the two techniques in a heterogeneous Arctic tundra site, for which the difference depended on the up-scaling approaches of the chamber measurements. However, comparison studies, especially on the croplands, are still lacking.

Simultaneous measurements are required to quantify the aggregate emissions of multiple greenhouse gases (CO₂, CH₄, and N₂O) and the fluxes of other gases species (like nitric oxide) and to investigate how their relationships vary with environmental factors and management practices. An instrumental system, AMEG (automatic chamber measuring system for emissions of carbon and nitrogen trace gases), was designed to meet these requirements. The system was based on the static, transparent automatic chamber (AC) technique. It had been proven able to measure the N₂O fluxes successfully from cotton and wheat fields (Liu et al., 2010, 2011, 2012; Wang et al., 2013) but its performance for measuring the fluxes of NEE and other gas species still needed to be tested prior to this study.

In this study, we attempted to compare the AC and EC techniques for measuring NEE fluxes from croplands cultivated with cotton and winter wheat. The comparison was undertaken to (a) assess the reliability of chamber NEE measurements on cropland ecosystems with suitable plant heights for chamber application and (b) test the performance of the AMEG system for measuring NEE fluxes.

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2 Materials and methods

2.1 Site description and agronomic management

The experimental site (34° 55' 30" N, 110° 42' 35" E) was located in a western suburb of Yuncheng city, Shanxi Province, China. This region is subject to a warm temperate continental monsoon climate. The annual mean air temperature and precipitation from 1998 to 2008 were 14.7°C and 508 mm, respectively (National Meteorological Information Center of China Meteorology Administration, <http://cdc.cma.gov.cn>). The dominating wind directions at the site were east and west (Wang et al., 2013). The surface soil (0–6 cm) had a pH of 8.7 (water extract). More details of the soil properties and environmental characteristics of the site were described by Liu et al. (2010, 2011).

The field layout and instrument locations are shown in Fig. 1. A 5 m wide path divided the experimental site into the cotton area (south) and the wheat area (north). A temporary laboratory (length × width × height = 5 m × 2 m × 2.5 m) was built on the path, in which the gas samples from the chambers were analyzed. The automatic chambers for the measurements in the cotton and wheat fields were situated south and north of the lab house, respectively. The EC systems in the cotton and wheat fields were both located 60 m away from the border of the two areas. Four manual chambers were installed near the automatic chambers for intermittent measurements during the non-growing season of the cotton field.

In the southern area, cotton had been continuously cultivated since 2004. The cotton residue from the previous growing season was cut into pieces and plowed into the soil at depths of 0–30 cm by machine on 6 November 2008. The field was then fallow until cotton was sown on 10 April 2009. The cotton seeds were sown by machine with row spacing of 80 cm. Fertilizer was applied on 28 June 2009 at rates of 75 kg N ha⁻¹ (90 % urea, 10 % di-ammonium phosphate), 9 kg P ha⁻¹, and 10 kg K ha⁻¹. The field was sprinkled with underground water on 8 April (36.5 mm), 10 July (85.3 mm) and 21 August 2009 (28.7 mm). The mature cotton seeds and cotton fibers were manually

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harvested on 1, 10 and 23 September and 5 October. All of the residue that remained in the field was incorporated into the soil later.

In the northern area, winter wheat and summer maize had been cultivated in rotation since 2005. Our comparison study was conducted only in wheat season. The maize straws of the previous growing season were cut off and plowed into the soil at depths of 0–20 cm in the area of the automatic chambers and 0–5 cm in the ambient area on 18 October 2009. The difference in the depths resulted from malfunction of the machine used for deep plowing. The wheat seeds were sown on 19 October at a row spacing of 20 cm. Fertilizers were applied at rates of 60 kg N ha⁻¹ (urea), 60 kg P ha⁻¹ and 30 kg K ha⁻¹ before sowing (18 October 2009) and 120 kg N ha⁻¹ (urea) in the spring (18 March 2010). The field was irrigated on 9 January, 23 March, 10 April and 4 May 2010. The wheat was harvested on 13 June 2010.

2.2 Eddy covariance measurements and data processing

2.2.1 Instrumentation

The NEE measurements using the EC method were conducted from 8 November 2008 to 7 November 2009 in the cotton field and from 21 October 2009 to 13 June 2010 in the wheat field. During the cotton measurement period, the CO₂ concentration was measured with an open-path infrared gas analyzer (LI-7500, LI-COR Inc., USA); the wind velocities in three dimensions were measured with a CSAT-3 sonic anemometer (Campbell Scientific Inc., USA) from 7 November 2008 to 13 January 2009 and a USA-1 anemometer (METEK GmbH, Germany) from 14 January 2009 to 15 October 2009. During the wheat measurement period, a LI-7500 analyzer and a CSAT-3 anemometer were used. The sensors of each EC system were installed 2 m above the ground. The raw data were saved on a CR5000 data logger (Campbell Scientific, Inc., USA) at a frequency of 10 Hz. The detection limit of the EC system was estimated at 0.001 mg C m⁻² s⁻¹ following the equation in Wang et al. (2013).

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2.2.2 Flux calculation

The NEE fluxes from EC measurements (NEE_{ec}) were determined using Eq. (1) (Lee et al., 2004), where F_{ec} is the turbulent flux and F_s is the storage flux. The former term was calculated following Eq. (2), where ρ denotes the density of the air ($g\ m^{-3}$), w' and c' denote the instantaneous deviations of the vertical wind velocity (ms^{-1}) and CO_2 concentration ($\mu mol\ mol^{-1}$) from the mean values, respectively, and the overbar indicates time average over 30 min.

$$NEE_{ec} = F_{ec} + F_s \quad (1)$$

$$F_{ec} = \rho \overline{w'c'} \quad (2)$$

Before calculating F_{ec} , spikes in the raw data time series were removed following Vickers and Mahrt (1997), and gaps were filled by interpolating valid data points. The double rotation method (Kaimal and Finnigan, 1994) was performed on the wind components so that the mean vertical wind velocity was reduced to zero for each half-hour period. The raw turbulent fluxes were corrected for the spectral loss (Moore, 1986) and the density fluctuations caused by correlated heat and water vapor fluctuations (WPL-correction) (Webb et al., 1980). The storage flux (F_s) resulting from changes in the CO_2 concentration in the air column below the measurement height was computed using the single point method (Hollinger et al., 1994).

The calculated NEE_{ec} were quality controlled using the following four steps. First, the fluxes during episodes of precipitation and sprinkler irrigation were discarded. Second, during nighttime or fallow periods, the NEE_{ec} smaller than $-0.001\ mg\ C\ m^{-2}\ s^{-1}$ (the lower detection limit of EC) was rejected. Third, the stationarity and integral turbulence characteristic tests (Foken and Wichura, 1996) were applied, and quality flags from 1 to 9 were assigned to each half-hourly flux; the fluxes flagged with 4–9 were regarded as bad data. Finally, to ensure that the measured fluxes were representative of the exchange rate of the interested fields, an analytical footprint model (Horst and Weil,

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1995) was used to investigate the flux source area; the fluxes were rejected if the contribution from the field of interest was less than 80 %.

2.2.3 Estimation of cumulative flux

Gap-filling was performed to obtain the cumulative EC fluxes. The data gaps less than four hours were directly filled by linear interpolation. Larger gaps were filled using empirical models based on Eqs. (3)–(5) when meteorological data were available, and otherwise using the mean diurnal variation method (Falge et al., 2001).

The NEE was the residual ecosystem respiration (ER) and photosynthesis (i.e., gross primary productivity, GPP), as described by Eq. (3) (e.g., IPCC, 2006).

$$NEE = ER - GPP \quad (3)$$

Data gaps during the nighttime or fallow periods were filled using the exponential relationship (Eq. 4) between ER and temperature (T) (Lloyd and Taylor, 1994), as ER was equal to NEE for these cases. In Eq. (4), a and b are fitting parameters, and T denotes soil (5 cm) temperature (T_s) during the fallow periods and air temperature (T_a) during the vegetation periods. The daytime NEE gaps during the vegetation periods were filled using the residual daytime ER and the GPP, which were estimated using Eqs. (4) and (5) (Falge et al., 2001; Zheng et al., 2008a), respectively. In Eq. (5), PAR is the photosynthesis active radiation ($\mu\text{mol photon m}^{-2} \text{s}^{-1}$), α is the apparent quantum yield ($\text{mg C } \mu\text{mol}^{-1} \text{ photon}$), and GPP_{max} is the maximum assimilation rate ($\text{mg C m}^{-2} \text{s}^{-1}$) at saturation PAR.

$$ER = a \cdot \exp(b \cdot T) \quad (4)$$

$$GPP = \frac{\alpha \cdot PAR \cdot GPP_{\text{max}}}{\alpha \cdot PAR + GPP_{\text{max}}} \quad (5)$$

As the parameters in Eqs. (4) and (5) varied in time due to seasonal changes in crop biomass and soil microbial activity, the cotton and wheat measurement periods were

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wheat with a row spacing of ~ 23 cm. The following operations were adopted to minimize the chamber effects on the inside microclimate, the plant growth and thus the measured fluxes. First, when the chamber lid was open during measurement intervals, the chambers were designed to leave an open space between the chamber and the ground, which could improve the air exchange between inside and outside (Fig. 2). Second, each chamber was switched between two base collars every week (growing period) or every month (fallow period). Third, whenever a difference of plant growth between inside and outside was visible, the base collars and the chambers were moved to new locations nearby. Only one chamber was closed for sampling each hour. The four chambers were then closed in sequence, and they made a measurement cycle every four hours. Therefore, each chamber provided six flux data each day.

An automatic chamber enclosure lasted for 36–42 min. The first 2 min were used for the NEE measurement, while the remaining data was used for CH₄, N₂O and nitric oxide measurements (Liu et al., 2010, 2011, 2012; Wang et al., 2013). During the first 2 min period, the air samples were drawn from the chamber headspace using a vacuum pump (N89KNE, KNF, Germany) and transported to the LI-6262 at a flow rate of 600 mL min⁻¹ through the Teflon tubes. The samples were analyzed every 5 s. The CO₂ signals were recorded in voltages and converted to concentrations using monthly calibration curves, which were derived from six standard gases with concentrations ranging from 0 to 1000 $\mu\text{mol mol}^{-1}$. The precision of the LI-6262 for CO₂ analysis was $\pm 0.1 \mu\text{mol mol}^{-1}$ with a response time of 5 s. By assuming a noise-to-signal ratio of 3, this precision level led to detection limits of ± 0.002 and $\pm 0.004 \text{ mg C m}^{-2} \text{ s}^{-1}$ for the 45 cm and 90 cm high chambers, respectively.

During the fallow period of the cotton field, soil CO₂ effluxes were manually measured every week using four static opaque chambers (Fig. 1). Five gas samples were collected during each enclosure period at intervals of 6–10 min and were analyzed using a gas chromatograph (HP5890, Hewlett-Packard Inc., USA). A detailed description of the sampling operation and instrumental configurations is provided in Zheng et al. (2008b) and Wang et al. (2013).

2.3.2 Flux calculation

The NEE fluxes from the automatic and manual chamber measurements (F , in $\text{mg C m}^{-2} \text{ s}^{-1}$) were calculated using the following equation:

$$F = \frac{dC/dt \cdot \rho \cdot h \cdot T_0 \cdot P \cdot 10^{-3}}{T \cdot P_0} \quad (6)$$

5 where dC/dt is the initial change rate of CO_2 concentration ($\mu\text{mol mol}^{-1} \text{ s}^{-1}$) during the chamber enclosure, h is the chamber height (m), ρ is the CO_2 density (g m^{-3}) under standard atmospheric conditions ($T = 273\text{K}$, $P = 1013\text{hPa}$), T is the air temperature (K) and P is the pressure (hPa) of the chamber headspace. The dC/dt was determined using either the nonlinear or the linear method described by Wang et al. (2013), in
10 which this scheme was used to calculate the nitrous oxide fluxes. In this study, dC/dt of each AC measurement was derived from the 12 concentration data measured during the second minute of each enclosure period. The data measured during the first minute were discarded due to the time lag induced by the tubing system. We only accepted fluxes for which there was a significant correlation between the CO_2 concentration and
15 the enclosure time ($\rho < 0.05$).

To obtain the final fluxes (NEE_{ac}), two correction steps were applied to the raw AC fluxes (F_{ac}). The first one was performed to correct the biases in the ER and GPP components caused by the differences in temperature and PAR, respectively, between inside and outside the chamber. This step was conducted by taking two correction terms, ΔER and ΔGPP , into account following Eqs. (7)–(9), where f is a function of temperature based on Eq. (4), g is a function of PAR based on Eq. (5), and the subscripts “out” and “in” denote outside and inside of the chambers, respectively. The PAR_{in} was estimated with the PAR_{out} and the transmittance rate (90%) of the chamber walls for visible light. The parameters in both functions for each chamber during each
20 sub-period were obtained from the fittings according to Eqs. (4) and (5). The second correction was performed to correct the housing effect caused by the chambers during

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the fallow period, as the long-term employment of the automatic chamber over a surface of bare soil would possibly introduce bias in the measured fluxes. This step was conducted by adding a coefficient, c , to Eq. (7). The value of c was determined by the ratio of the monthly mean manual-chamber fluxes to the monthly mean AC fluxes corrected after the earlier steps. This correction was not performed, and thus, c was set as 1.0, during the vegetation period.

$$NEE_{ac} = c \cdot (F_{ac} + \Delta ER - \Delta GPP) \quad (7)$$

$$\Delta ER = f(T_{out}) - f(T_{in}) \quad (8)$$

$$\Delta GPP = g(PAR_{out}) - g(PAR_{in}) \quad (9)$$

2.3.3 Estimation of cumulative flux

Because the AC fluxes were calculated with the data during the second minute of each enclosure period, they were regarded as the NEE for the corresponding half hour. Thereafter, each AC flux was referred to as a half-hourly flux. To obtain the cumulative fluxes of individual chambers, the data gaps were filled as described below.

The regular gaps within each measurement interval (length = 3.5 h) were filled using linear interpolation, as each chamber was designed to provide a NEE flux every four hours. If the half-hourly flux was very close to the detection limit of the AC system, the missing measurements before or after it were filled with a random number between zero and the detection limit. In particular, this treatment was applied for the measurement gaps in the nighttime of cold winter, when the actual fluxes were very small. Data gaps larger than 4 h were filled using the empirical models used for the EC gaps.

There were two big gaps in the cotton data, from 25 April to 22 July and from 16 October to 7 November, respectively. The two gaps are referred to as C2 and C4. Thus, the periods before and between them are called C1 (8 November 2008–24 April 2009) and C3 (23 July 23–15 October 2009), respectively. To obtain the annual NEE, the cumulative fluxes of C2 and C4 were roughly estimated. Gap C2 was caused by a malfunction of the LI-6262 analyzer. It covered the seedling stage and the fast-growth

stage of the cotton. Gap C4 occurred during the maturity stage with relatively stable aboveground biomass. Different gap-filling strategies were applied to the three stages.

The gaps during the seedling and maturity stages were filled using the arithmetic means of the results simulated with two models. One was a function of soil temperature (Eq. 4), in which the parameters were obtained by fitting the soil CO₂ effluxes and soil temperature during the fallow period. The other model was a function of air temperature and PAR (Eqs. 3–5), in which the parameters were obtained from the nearest 10 days' measurements. We also employed the second empirical model to fill the gaps during the fast-growth stage. Considering that the aboveground biomass increased quickly during the seedling and fast-growth stages, when applying the fittings of the second model, the NEE fluxes were divided by the aboveground biomass. As a result, the modeled results needed to be multiplied by the aboveground biomass of the corresponding gap days. The aboveground biomass data used in this step were modeled with a growth curve function, based on the weekly measurements at the ambient fields.

Due to the different tillage depths between the chamber locations and the ambient field, we divided the entire wheat season into periods W1 (21 October–30 November 2009) and W2 (1 December 2009–13 June 2010), assuming that the effects of tillage difference only impacted period W1.

2.4 Auxiliary measurements

The incoming PAR and global radiation were measured every half hour using a quantum sensor (LI-190SA, LI-COR Inc., USA) and a Pyranometer (CM 6B, Kipp & Zonen, the Netherlands), respectively. The air temperature at a height of 2 m was recorded every half-hour using a Vaisala probe (HMP45C, Vaisala, Finland). The field soil temperature was measured every 10 min using a TidbiT temperature data logger (Onset, USA) at a depth of 5 cm. The air and soil (5 cm depth) temperatures inside the chambers were logged every minute using thermocouples (JWB series, Kunlun Coast Sensing Technology Center, Beijing, China). A tipping bucket rain gauge was used to measure hourly precipitation. The volumetric moisture of the top 6 cm of soil was manually

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displayed a clear seasonal variation pattern with small but continuously positive values in the fallow period (from November of 2008 to middle April of 2009) and much larger negative (daytime) or positive (nighttime) values during the vegetation period. During the fallow period, the NEE was equal to the CO₂ efflux from soil respiration, with a mean flux of 0.017 ± 0.015 (± 1 standard deviation, SD) $\text{mg C m}^{-2} \text{s}^{-1}$. Nearly 84 % of the variation in NEE during this period can be explained by temperature ($p < 0.01$). In response to the germination of cotton, the half-hourly NEE started to display a weak diurnal variation in late April, with lower values at daytime and higher values at nighttime. This diurnal pattern became more notable during the vegetation period due to enhanced daytime photosynthesis and nighttime respiration. The daily NEE fluxes ranged from -9.39 to $3.78 \text{ g C m}^{-2} \text{ d}^{-1}$. The minimum daily flux was observed on 23 July 2009. The EC measurement in the cotton field resulted in an annual NEE of $-40 \text{ g C m}^{-2} \text{ yr}^{-1}$ and a cumulative NEE of 190, -75 , -151 and -4 g C m^{-2} during the C1, C2, C3 and C4 periods, respectively (Table 1).

In the wheat field, we obtained 5172 valid half-hourly EC fluxes (Fig. 4b). The data coverage was 46 %. These fluxes ranged from -0.641 to $0.175 \text{ mg C m}^{-2} \text{ s}^{-1}$ and showed a significant seasonal variation pattern. The wheat germinated after a rainfall that occurred on 31 October 2009. Afterwards, we observed diurnal variation of NEE in November and December, which showed slight negative fluxes at daytime. From mid-December 2009 to early February 2010, the wheat experienced winter dormancy due to freezing. During this period, the average daytime and nighttime half-hourly fluxes were -0.004 and $0.009 \text{ mg C m}^{-2} \text{ s}^{-1}$, respectively. As the soil became warmer in the spring, the NEE magnitude gradually increased (Fig. 4a). The daily NEE fluxes of the wheat field ranged from -11.69 to $5.74 \text{ g C m}^{-2} \text{ d}^{-1}$ (Fig. 5b). The minimum daily flux was observed on 28 April 2010 and the maximum on 8 June 2010.

The cumulative NEE of the entire wheat measurement period was estimated at -251 g C m^{-2} based on the EC data. During the periods W1 and W2, the cumulative estimates were 67 and -317 g C m^{-2} , respectively (Table 1).

3.3 Chamber NEE

Using the AC system, we obtained 2571 and 2932 valid NEE fluxes in the cotton and the wheat fields, respectively, which corresponded to data coverage of 42 % and 52 %. During the fallow period, the temperature correction reduced the AC fluxes by 6 % during daytime but had very little influence on the nighttime fluxes. During the vegetation periods, the overlap effect of the correction on temperature and PAR increased the AC fluxes by 1 % (cotton) or 7 % (wheat) during nighttime and by 15 % (cotton) or 11 % (wheat) during daytime.

The gap period C2 separated the cotton measurement period into fallow (C1) and vegetation (C3) periods. The half-hourly AC fluxes ranged from 0.002 to 0.099 mgCm⁻²s⁻¹ during C1 and from -0.516 to 0.157 mgCm⁻²s⁻¹ during C3 (Fig. 4c). The daily NEE of period C1 ranged from 0.23 to 3.32 gCm⁻²d⁻¹ (Fig. 5a) and resulted in an average cumulative flux of 194 gCm⁻². The daily fluxes of period C3 varied between -10.27 and 5.46 gCm⁻²d⁻¹ (Fig. 5a) and the cumulative NEE was -138 gCm⁻² on average (Table 1).

The half-hourly AC fluxes from the wheat field ranged from -0.644 to 0.187 mgCm⁻²s⁻¹ (Fig. 4d). Significant positive NEE fluxes were measured in the first five weeks. In December and January, the data coverage was relatively low (< 20 %). The cold weather had both inhibited the CO₂ emission from respiration and the uptake by photosynthesis. The fluxes were usually at marginal levels during these two months. By contrast, during the remainder of the time during the wheat season, the data rejection rate was much smaller. The daily NEE fluxes varied between -12.46 and 8.40 gCm⁻²d⁻¹ (Fig. 5b). The cumulative NEE from the AC measurements were estimated at -205 gCm⁻² for the entire wheat measurement period, and 134 gCm⁻² and -339 gCm⁻² for the periods of W1 and W2, respectively (Table 1).

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3.4 Comparison between chamber and eddy covariance NEE

In both the cotton and wheat fields, the half-hourly NEE fluxes measured by the EC and AC methods, as well as their daily estimates, displayed similar seasonal variation patterns (Figs. 4 and 5). Figure 6 shows significant zero-intercept linear regressions ($p < 0.001$) between the half-hourly fluxes simultaneously measured by the two methods during the measurement periods on the cotton (Fig. 6a) and the wheat (Fig. 6b) fields. The slopes indicate that the AC fluxes were 0.78 and 1.06 times the EC fluxes from the two fields, respectively. Data from the W1 period were not included in the comparison, because the different tillage depths between the chamber locations and the ambient area might have contributed to the significant difference in the NEE fluxes during this period (AC higher by 100%, $p < 0.001$).

The comparison of the daily NEE fluxes between the two methods showed better agreement (Fig. 6c, d). The significant correlations ($p < 0.001$) indicate that the daily AC fluxes were on average 0.94 (cotton) and 1.00 (wheat) times the daily EC fluxes. This finding suggests that the AC-based AMEG system can yield NEE estimates in good agreement with those from the EC techniques.

During the periods for which there was normal performance of both instruments and comparable field conditions, the differences in the cumulative NEE between the two methods ranged from -9.0% to 6.7% , with a mean value of 0.1% (Table 1).

4 Discussion

4.1 Uncertainties in eddy covariance NEE

The random uncertainty in the EC flux mainly originated from the statistical errors associated with the EC method (Moncrieff et al., 1996). It decreased with increasing data set size. During the periods C1, C2, C3, C4, W1 and W2, this uncertainty was estimated at

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2.2 %, 0.9 %, 0.7 %, 0.8 %, 3.3 % and 0.6 %, respectively. For the annual cotton period and the entire wheat period, it decreased to 0.6 % and 0.5 %, respectively.

The systematic uncertainties in the EC fluxes included the flux losses due to insufficient coverage of the high or low frequency contribution to the turbulent fluxes (Moore, 1986), the biases caused by fluctuations of temperature and water vapor (Webb et al., 1980), and the flux underestimation due to insufficient turbulence development (Foken et al., 2004). These errors were reduced as much as possible by applying the corrections and quality controls as described in Sect. 2.2.2. However, the systematic errors may still remain due to incorrect application of these post-field processing treatments (Moncrieff et al., 1996) and other potential errors, such as the bias due to the surface heating effect of the open-path analyzer (Burba et al., 2008). In our study the LI-7500 analyzers were mounted at an angle (45°) from the vertical. Unfortunately, the currently available techniques for correcting the surface heating effect are only suitable for vertical sensors, which would overestimate the bias if applying to tilted situations. Besides, at present directly applying this correction seems not warranted because its performance varied greatly among different experimental sites (Burba et al., 2008; Wohlfahrt et al., 2008; Haslwanter et al., 2009). For these reasons, we assumed that the self-heating effect was of minimal importance in our case. In previous studies (e.g., Sottocornola and Kiely, 2005; Mauder et al., 2013), the lack of surface energy balance was considered as an indirect approach for estimating the systematic errors in EC fluxes. Considering that this parameter was not available at our site, we accessed the reports in literature on energy balance deficits at agricultural sites (Foken et al., 2010; Lei and Yang, 2010) and made a rough estimate of -30 ~ -20 % (mean: -25 %) for the systematic uncertainty in the NEE fluxes at our site. This underestimation was not applied in the EC fluxes.

Combining the random and systematic components, we obtained the total relative uncertainties of the cumulative NEEs. They were estimated at -25 % (-32 ~ -18 %) during period C1, -25 % (-31 ~ -19 %) during periods C2, C3 and C4, -25 % (-33 ~ -17 %) during period W1, and -25 % (-31 ~ -19 %) during period W2. During the

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annual period of the cotton field, the total uncertainty was estimated at -25% ($-31\% \sim -19\%$).

4.2 Uncertainties in automatic chamber NEE

The random uncertainties in the half-hourly AC fluxes were mainly introduced by instrumental detection noise. They were estimated at ± 0.002 and $\pm 0.004 \text{ mg C m}^{-2} \text{ s}^{-1}$ for the 45 and 90 cm chambers, respectively. According to Eq. (10), the random uncertainties in the daily NEE estimates were 8 %, 7 %, 8 % and 7 % during the periods C1, C3, W1 and W2, respectively. The random errors in the cumulative NEE of these four periods were then estimated at 0.6 %, 0.8 %, 1.3 % and 0.5 %, respectively.

Because chamber deployment can lead to a nonlinear increase or decrease of CO_2 concentration over time within the chamber headspace (Hutchinson et al., 2000; Kutzbach et al., 2007; Pedersen et al., 2010, etc.), the nonlinear regression method was involved in flux calculation in this study. However, significant nonlinearity could not always be detected, which could lead to flux underestimation. This failure rate was 48 % for the cotton and 56 % for the wheat. Meanwhile, we compared the fluxes calculated with the nonlinear and the linear procedures using the data from the nonlinear cases. This revealed an underestimation of 22 % by the linear method, which was within the range (4–40 %) reported in previous studies (Kutzbach et al., 2007; Schneider et al., 2009; Jassal et al., 2012). Consequently, the systematic underestimation due to detection failure for significant nonlinearity was estimated at $-11\% \sim 0$ (median: -5%) and $-12\% \sim 0$ (median: -6%) for the cotton and wheat fields, respectively (Table 1).

Spatial heterogeneity is always a problem for AC measurement, as the measured fluxes only represent the limited area where the chambers are placed, and it can be another source of systematic uncertainty. During the fallow period, this error was determined by comparing the mean EC fluxes from the automatic chamber directions ($30\text{--}60^\circ$ and $120\text{--}150^\circ$ for the cotton and wheat, respectively) with those from all wind directions, and the differences were -13% and 21% for the cotton and wheat fields, respectively. During the vegetation period, we performed the same approach only using the

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nighttime fluxes, as the large variation of daytime NEE fluxes might bias this analysis. The results showed that the fluxes from the chamber direction area were 15 % lower (cotton) and 2 % higher (wheat) than the fluxes from all wind directions. The biomass difference between inside and outside the chambers due to chamber effect was negligible as the measurement locations were changed frequently during the growing periods of cotton and wheat.

The total relative uncertainties in the cumulative NEEs were obtained by summing up the random and systematic parts. They were estimated at -19 % (-25 ~ -12 %) during period C1, -21 % (-27 ~ -14 %) during period C3, 15 % (8 ~ 22 %) during period W1, and -4 % (-11 ~ 3 %) during period W2 (Table 1). On the timescale of these periods, the uncertainties in the chamber-based NEE were predominantly systematic, while the random part was negligible. The flux uncertainties of gap C2 and C4 were roughly treated the same as those of period C3 (Table 1), because periods C2 ~ C4 were all during the growing stage of cotton and their sources of error were considered the same. Therefore, the uncertainties were dominated by the systematic part, as the random part could become very small due to the large number of dataset.

4.3 Comparison between chamber and eddy covariance NEE

In this study, the simultaneous NEE measurements with the EC and AC techniques covered nearly 8 months in the two fields. The datasets enabled comparisons between the two methods under a wide range of environmental conditions.

On average, the daily NEE fluxes derived from the two methods agreed with each other (Fig. 6c, d). By comparing the cumulative NEE of individual periods, we found the greatest difference during period W1, for which the AC result was 100 % higher (Table 1). This large discrepancy can most likely be attributed to the different tillage depths between the chamber locations (20 cm) and the ambient field (5 cm), as deeper tillage might have stimulated more CO₂ emissions from soil respiration (Bauer et al., 2006; Moraru et al., 2012) (Fig. 5b). For this reason, the period W1 data were omitted from further comparisons between the two techniques. In contrast to the data for W1,

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the cumulative NEE of AC and EC showed much smaller differences during the periods C1, C3 and W2 (Table 1). We compared the cumulative NEEs for the three periods using a zero-intercept linear regression and found a significant relationship between the two methods (slope = 1.04, $n = 3$, $R = 0.997$, $p < 0.05$). The slope indicates that the AC estimate was 4 % higher on average, suggesting that the AC technique was able to provide NEE estimates comparable to those derived from the EC measurements, particularly on seasonal or annual timescales. This level of difference is similar to that reported by Laine et al. (2006) but much smaller than those in Dore et al. (2003) and Fox et al. (2008).

The cumulative AC fluxes for the gap periods C2 and C4 were roughly estimated at -104 and 6 gCm^{-2} , respectively, following the gap-filling procedure described in Sect. 2.3.3. Therefore, we were able to obtain an annual NEE estimate of $-42 \text{ gCm}^{-2} \text{ yr}^{-1}$ for the cotton field. This result matched very well with that (-40 gCm^{-2}) derived from the EC measurements (Table 1). When the cumulative NEE from periods C2 and C4 were included in the zero-intercept linear regression, we again observed that there was a significant correlation between the two methods (slope = 1.05, $n = 5$, $R = 0.996$, $p = 0.000$). This finding further indicates that the AMEG system can provide NEE estimates similar to those of the EC measurements.

We compared the cumulative NEEs of periods C1, C3, and W2 after they were corrected by the values of mean uncertainty in Table 1 and found that the two methods were still linearly correlated, but with a decreased slope of 0.86. This difference suggested that the energy balance deficit (25 %) adapted from literature may have been an overestimate of the negative systematic biases of the EC fluxes, provided that the errors in the AC fluxes were accurately estimated. Therefore, simultaneous measurement of the lack of energy balance closure is necessary to provide supporting evidence. On the other hand, more research should be conducted to develop direct methods for estimating systematic uncertainties for EC measurements.

The gain-loss method uses net biome production (NBP, the residual of $-NEE$ and carbon loss from an ecosystem via, e.g., harvest of products) or net ecosystem carbon

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balance (NECB, the residual of carbon input and output of an ecosystem) as an indicator of the change in organic carbon stock of a terrestrial ecosystem (Chapin III et al., 2006; IPCC, 2006). Both of these measurements can be estimated using the NEE and the other carbon flows. NBP or NECB can be regarded as measures of the annual increase in soil organic carbon stock (Δ SOC) for an ecosystem that has grown annual non-woody plants for over 20 consecutive years. Positive and negative values of Δ SOC indicate the accumulation and loss rates, respectively, for soil organic carbon (IPCC, 2006). In the case of cotton for this study, the gain-loss method could simply estimate the annual Δ SOC with $-NEE - C_h$ due to absence of organic manure, removal of crop residues, and other significant disturbances to the carbon balance (the term C_h was the carbon amount in harvested cotton seeds and fiber, which was measured at $202 \text{ gC m}^{-2} \text{ yr}^{-1}$). Then, based on the ranges of annual NEE given in Table 1, the annual Δ SOC were estimated at $-153 \sim -160$ (AC) and $-140 \sim -154$ (EC) $\text{gC m}^{-2} \text{ yr}^{-1}$. Using another method proposed by Huang et al. (2007), the annual Δ SOC was estimated at $20 \text{ gC m}^{-2} \text{ yr}^{-1}$ by multiplying the amount of organic carbon in the incorporated cotton residues (measured at $\sim 180 \text{ gC m}^{-2} \text{ yr}^{-1}$ for the aboveground residue and the roots) with the conversion efficiency from incorporated residue carbon to soil organic carbon ($\sim 11\%$, adapted from the long-term field data in the North China Plain region with similar soil type, climate and management practices). The inconsistent values of Δ SOC estimated using these two methods likely indicates the presence of additional errors in our flux measurements that were not discussed above. These errors might include the soil CO₂ releases from urea hydrolysis (estimated at only $3 \text{ gCO}_2 - \text{C m}^{-2}$ for our cotton case) and other inorganic processes related to soil pH changes (such as nitrification following urea hydrolysis or directly application of ammonium-based fertilizers, exudation of acid substrates from roots, and atmospheric acid deposition). Thus, these emissions, which are related to inorganic processes in the calcareous soil of our site, might have led to an overestimation of the ER and thus the NEE to a large extent. However, neither the AC nor the EC technique could distinguish this flux component from the others, such as photosynthesis, autotrophic respiration,

and heterotrophic respiration. This may suggest that measurements using either the AC or EC technique likely lead to an overestimation of the SOC loss rate (or underestimate the SOC gain rate) of an ecosystem, particularly with a calcareous soil exposed to increasing atmospheric acid deposition (e.g., Wang et al., 2012).

5 Conclusions

The net ecosystem exchange (NEE) fluxes of carbon dioxide (CO₂) were simultaneously measured using static, transparent automatic chamber (AC) and eddy covariance (EC) techniques in a cotton field for a year and in a winter wheat field for an entire cropping season. Based on a comparison between the NEE fluxes measured using the two techniques, the following conclusions can be drawn. There is good agreement between the NEE measured using the AC and EC techniques at half-hourly, daily, seasonal and annual timescales, as long as the biases due to the modification of headspace temperature and radiation are corrected in data processing, and as long as on-site operation (such as alternation of chamber locations) is frequently carried out to minimize chamber effects on growth of the enclosed plants. Such data agreement has confirmed that the AC technique is a necessary alternative to the EC technique, if plant heights are suitable for chamber operation. These results also demonstrate the satisfying performance of AMEG (an AC system designed for monitoring fluxes of multiple carbon and nitrogen trace gases) for measuring NEE fluxes of short-plant croplands. In an ecosystem with calcareous soil, especially one that is exposed to increasing atmospheric acid deposition, CO₂ emissions related to inorganic processes likely lead to an overestimation of ecosystem respiration and thus an overestimation of NEE flux, no matter which measurement technique is employed. If such a positive bias is involved in estimating the change rate of soil organic carbon stock, it will overestimate the loss rate (or underestimate the gain rate).

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Table 1. The cumulative NEEs (gCm⁻²) and the relative uncertainties (%) derived from the measurements using the automatic chamber (AC) and eddy covariance (EC) techniques.

Period ^a	AC ^b	EC	D ^e
Cotton			
C1	194 (−19%, −25 ~ −12%) ^c	190 (−25%, −32 ~ −18%)	2.1 %
C2	−104 ^d (−21%, −27 ~ −14%)	−75 (−25%, −31 ~ −19%)	
C3	−138 (−21%, −27 ~ −14%)	−151 (−25%, −31 ~ −19%)	−9.0%
C4	6 ^d (−21%, −27 ~ −14%)	−4 (−25%, −31 ~ −19%)	
Annual	−42 (−20%, −26 ~ −13%)	−40 (−25%, −31 ~ −19%)	
Wheat			
W1	134 (15%, 8 ~ 22%)	67 (−25%, −33 ~ −17%)	
W2	−339 (−4%, −11 ~ 3%)	−317 (−25%, −31 ~ −19%)	6.7%

^a C1, C2, C3 and C4 represent the periods of 8 November 2008–24 April 2009, 25 April–22 July 2009, 23 July–15 October 2009 and 16 October–7 November 2009 of the cotton field, respectively. W1 and W2 represent the periods of 21 October–30 November 2009 and 1 December 2009–13 June 2010 of the wheat field, respectively.

^b Estimates of AC were the averages of the four chambers.

^c Figures outside of the parentheses represent the cumulative fluxes, and those inside represent the mean and the range of the relative uncertainties.

^d Estimates fully derived from gap-filling.

^e Differences between the cumulative AC and EC fluxes, calculated following $(AC - EC)/(AC/2 + EC/2) \times 100\%$.

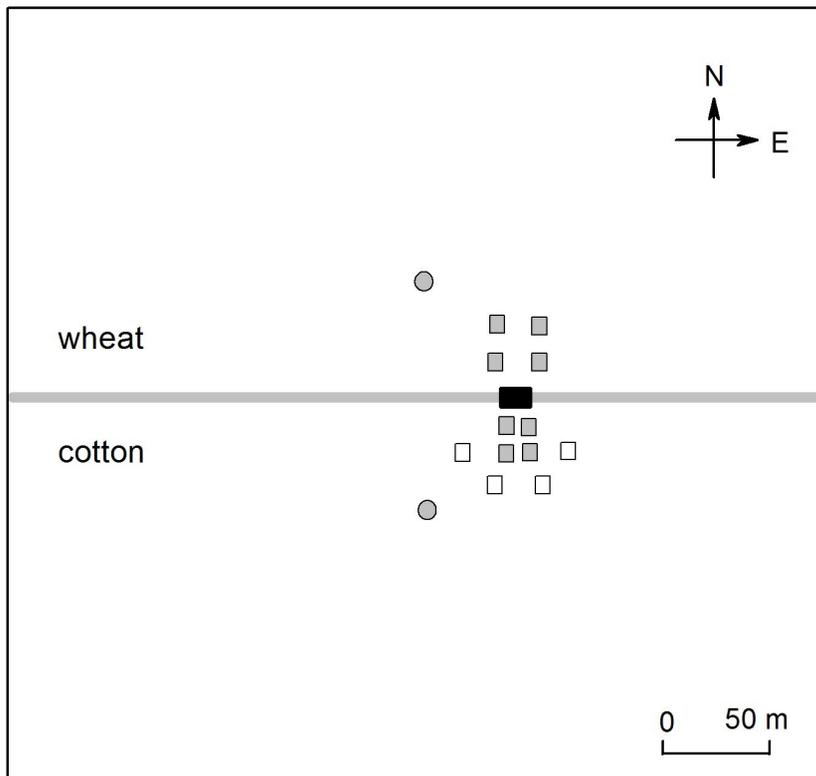


Fig. 1. Layout of the experimental site. The horizontal course line indicates the path dividing the experimental area into the cotton (north) and wheat (south) fields. The grey circles, grey squares, empty squares and black rectangle indicate the locations of the eddy covariance towers, automatic chambers, manual chambers, and the temporary laboratory, respectively.

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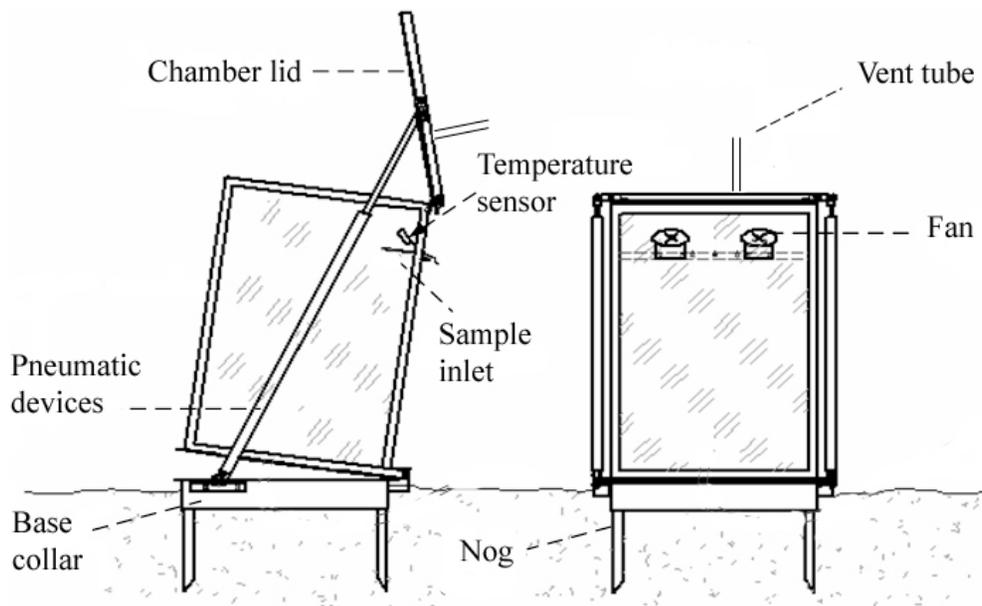


Fig. 2. Schematic illustration of the automatic chambers.

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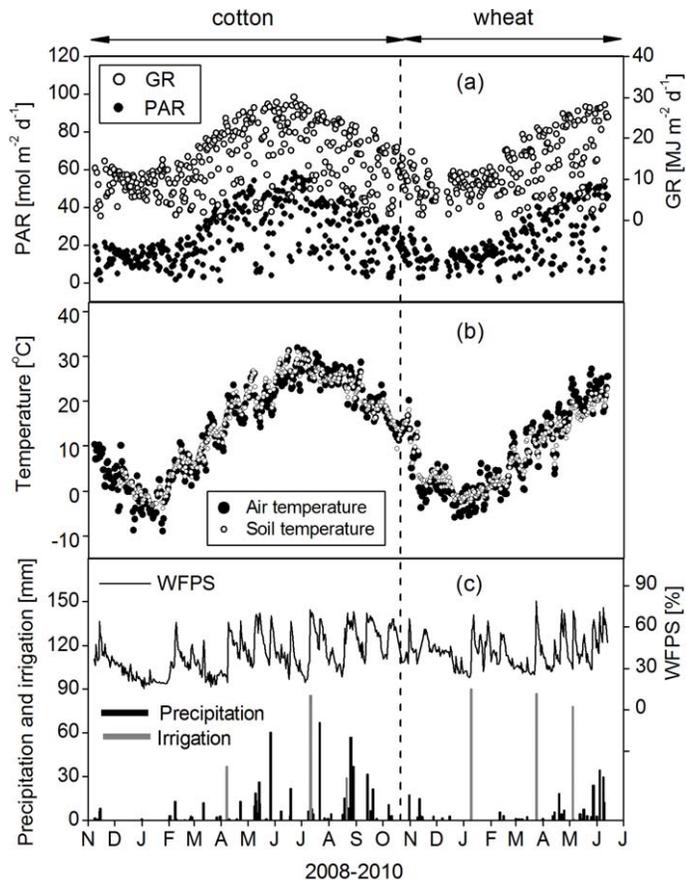


Fig. 3. (a) The daily cumulative global radiation (GR) and photosynthesis active radiation (PAR), (b) the daily mean air temperature and soil temperature at a depth of 5 cm, (c) the daily precipitation, the water amount of each irrigation event and the daily mean soil moisture (water-filled pore space, i.e. WFPS) at depths of 0–6 cm during the entire measurement period.

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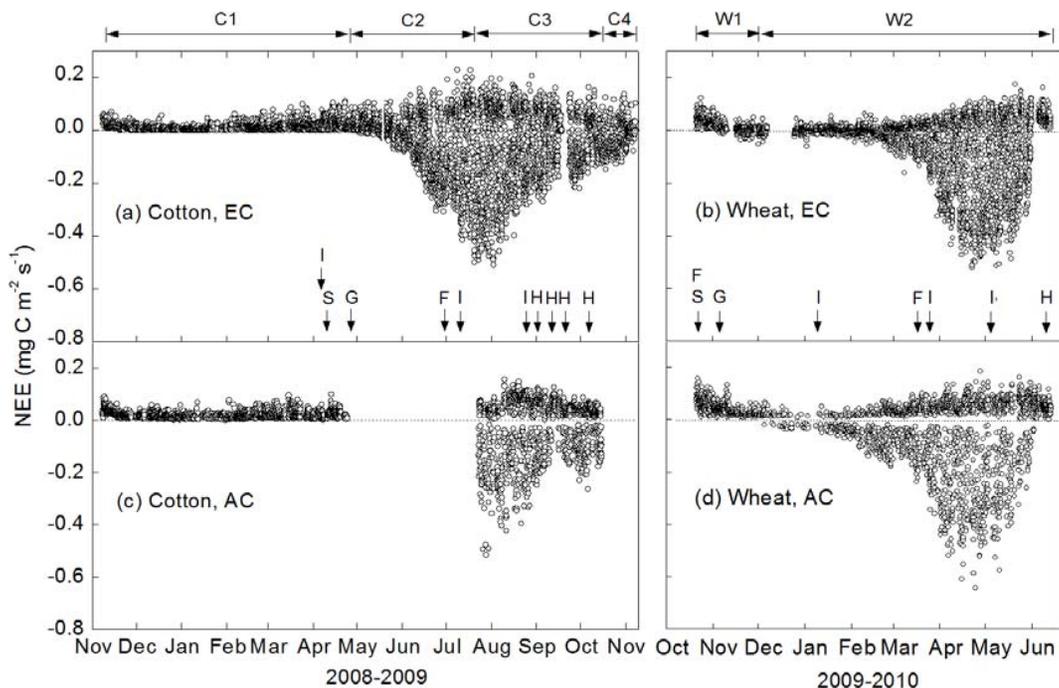


Fig. 4. Half-hourly net ecosystem exchange (NEE) measured using the eddy covariance (EC) and automatic static chamber (AC) techniques in the cotton and wheat fields. The arrows with “S”, “G”, “F”, “I” and “H” indicate the dates of sewing, germination, fertilization, irrigation and harvest, respectively.

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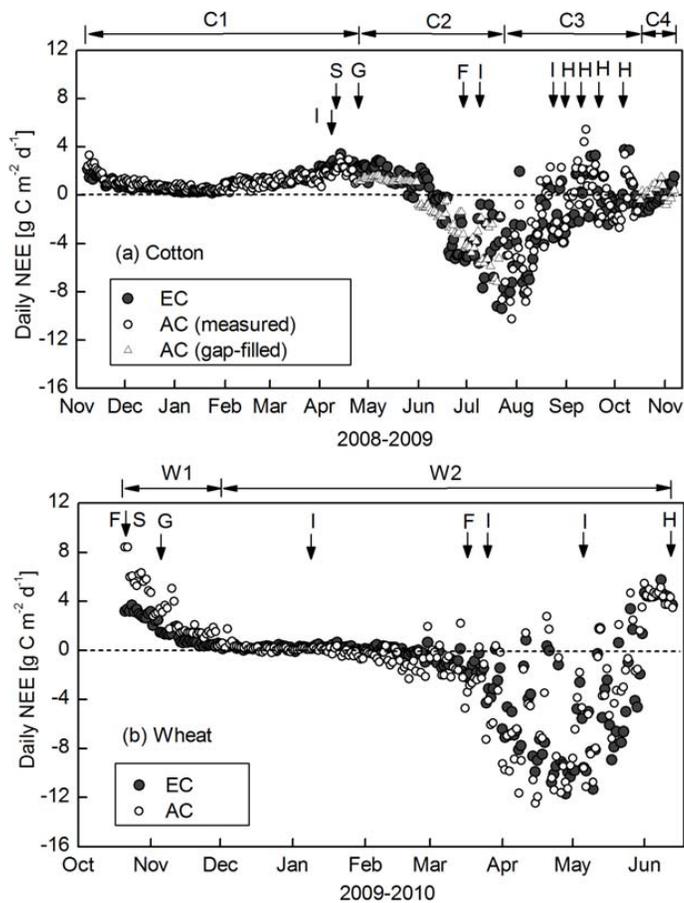


Fig. 5. Daily NEE fluxes measured using the eddy covariance (EC) and automatic static chamber (AC) techniques in the (a) cotton and (b) wheat fields.

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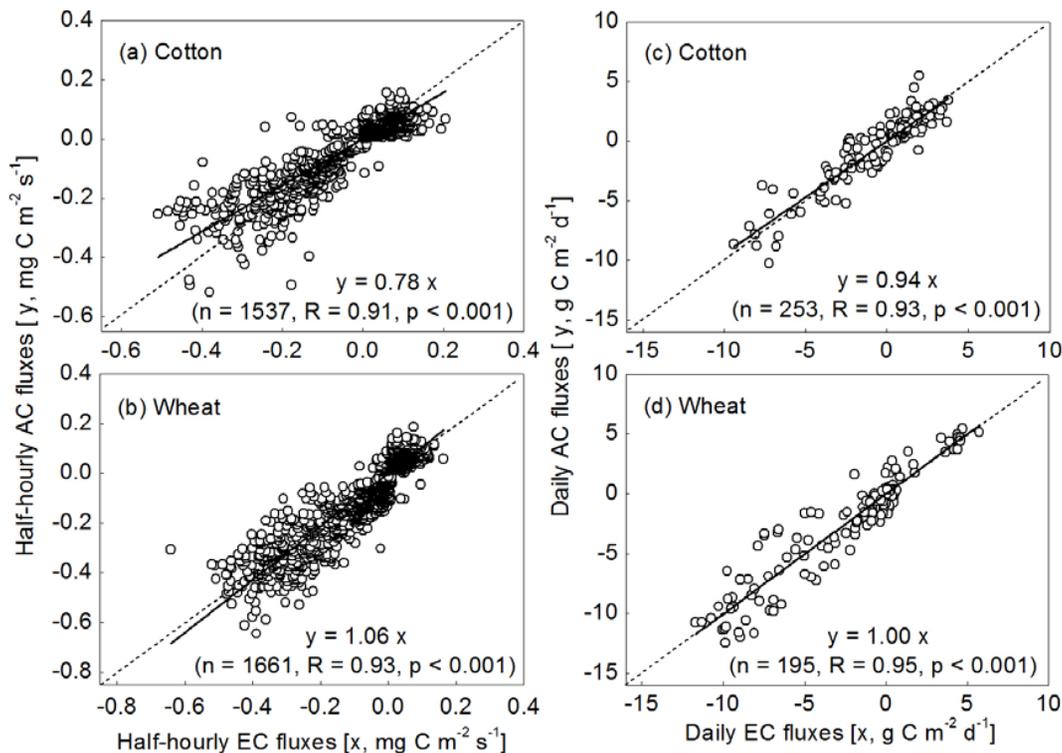


Fig. 6. Comparison of the half-hourly (a, b) and daily (c, d) NEE fluxes between the automatic chamber (AC) and eddy covariance (EC) techniques. Panels (a) and (c) present the data from periods C1 and C3. Panels (b) and (d) show the data from period W2. Definitions of periods C1, C3 and W2 are given in the footnotes of Table 1 as well as in the text.

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