# A halocarbon survey from a seagrass dominated subtropical lagoon, Ria Formosa (Portugal): Flux pattern and isotopic composition

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#### 12 Abstract

Here we report fluxes of chloromethane (CH<sub>3</sub>Cl), bromomethane (CH<sub>3</sub>Br), iodomethane 13 14 (CH<sub>3</sub>I), and bromoform (CHBr<sub>3</sub>) from two sampling campaigns (summer and spring) in the 15 seagrass dominated subtropical lagoon Ria Formosa, Portugal. Dynamic flux chamber 16 measurements were performed when seagrass patches were either air-exposed or submerged. 17 Overall, we observed highly variable fluxes from the seagrass meadows and attributed them to diurnal cycles, tidal effects, and the variety of possible sources and sinks in the seagrass 18 meadows. Highest emissions with up to 130 nmol  $m^{-2} h^{-1}$  for CH<sub>3</sub>Br were observed during 19 20 tidal changes from air exposure to submergence and conversely. Furthermore, during the 21 spring campaign, the emissions of halocarbons were significantly elevated during tidal 22 inundation as compared to air exposure.

Accompanying water sampling during both campaigns revealed elevated concentrations of CH<sub>3</sub>Cl and CH<sub>3</sub>Br indicating productive sources within the lagoon. Stable carbon isotopes of halocarbons from the air and water phase along with source signatures were used to allocate the distinctive sources and sinks in the lagoon. Results suggest CH<sub>3</sub>Cl rather originating from seagrass meadows and water column than from salt marshes. Aqueous and atmospheric CH<sub>3</sub>Br was substantially enriched in <sup>13</sup>C in comparison to source signatures for seagrass meadows and salt marshes. This suggests a significant contribution from the water phase on
the atmospheric CH<sub>3</sub>Br in the lagoon.

3 A rough global upscaling yields annual productions from seagrass meadows of 2.3-4.5 Gg yr

4  $^{1}$ , 0.5-1.0 Gg yr<sup>-1</sup>, 0.6-1.2 Gg yr<sup>-1</sup>, and 1.9-3.7 Gg yr<sup>-1</sup> for CH<sub>3</sub>Cl, CH<sub>3</sub>Br, CH<sub>3</sub>I, and CHBr<sub>3</sub>

5 respectively. This suggests a minor contribution from seagrass meadows to the global

6 production of CH<sub>3</sub>Cl and CH<sub>3</sub>Br with about 0.1 % and 0.7 %, respectively. In comparison to

7 the known marine sources for CH<sub>3</sub>I and CHBr<sub>3</sub>, seagrass meadows are rather small sources.

8

# 9 1 Introduction

The halocarbons chloromethane (CH<sub>3</sub>Cl), bromomethane (CH<sub>3</sub>Br), iodomethane (CH<sub>3</sub>I), and bromoform (CHBr<sub>3</sub>) are prominent precursors of reactive halogens which affect the oxidative capacity of the atmosphere and initiate stratospheric ozone destruction (Saiz-Lopez and von Glasow, 2012 and references therein). Therefore, during the last decades, the sources and sinks of these trace gases have been intensively studied.

For CH<sub>3</sub>Cl, recent atmospheric budget calculations suggest that the known sinks can be balanced by large emissions from tropical terrestrial sources (Saito and Yokouchi, 2008; Xiao et al., 2010). Nevertheless, these calculations still incorporate large uncertainties. The atmospheric budget of CH<sub>3</sub>Br remains still unbalanced, with the known sinks exceeding known sources by about 30% (Yvon-Lewis et al., 2009). The current emission estimates for CH<sub>3</sub>I and CHBr<sub>3</sub> are assigned with even larger uncertainties (Bell et al., 2002; Quack and Wallace, 2003).

Stable carbon isotopes of halocarbons have been applied to further elucidate their sources and sinks by using individual source signatures (Keppler et al., 2005). While this was primarily done for CH<sub>3</sub>Cl, first isotopic source signatures of naturally-produced CH<sub>3</sub>Br were recently reported (Bill et al., 2002; Weinberg et al., 2013). Moreover, the biogeochemical cycling of halocarbons underlies various transformation processes which can be studied by the stable carbon isotope approach in addition to flux and/or concentration measurements.

28 Coastal zones are reported as being important source regions of halocarbons. In these salt

29 water affected systems, halocarbon producers comprise phytoplankton (Scarratt and Moore,

30 1998), macroalgae (Gschwend et al., 1985), salt marshes (Rhew et al., 2000), and mangroves

31 (Manley et al., 2007).

With a net primary production of 1211 g C m<sup>-2</sup> yr<sup>-1</sup>, seagrass meadows are one of the most 1 2 productive ecosystems with a similar global abundance as mangroves and salt marshes 3 (Duarte et al., 2005). They cover huge areas of the intertidal and subtidal zone in temperate 4 and subtropical/tropical regions. Thus, they may represent an additional source for 5 halocarbons to the atmosphere which is not yet sufficiently studied. Seagrass meadows are highly diverse ecosystems with respect to potential halocarbon producers. Along with the 6 7 seagrass itself, they comprise epiphytes such as microalgae and diatoms, and sediment 8 reassembling microphytobenthos and bacteria communities. All these constituents of the 9 benthic community have been reported to produce halocarbons (Amachi et al., 2001; Blei et al., 2010; Manley et al., 2006; Moore et al., 1996; Rhew et al., 2002; Tokarczyk and Moore, 10 11 1994; Urhahn, 2003). While first evidence for the release of halocarbons from seagrass was obtained by incubation experiments (Urhahn 2003), we could recently confirm this production 12 13 potential in a field study of a temperate seagrass meadow in Northern Germany (Weinberg et 14 al., 2013).

In order to refine these results, we conducted two field campaigns in the subtropical lagoon Ria Formosa, Portugal in 2011 and 2012. Here we report the results of these campaigns comprising dynamic flux chamber measurements for halocarbons over seagrass meadows during air exposure and tidal inundation. Using the flux and isotopic data, we present first insights into the environmental controls of halocarbon dynamics within this ecosystem.

20

#### 21 2 Materials and methods

#### 22 2.1 Sampling site

The Ria Formosa, covering an area of 84 km<sup>2</sup>, is a mesotidal lagoon at the South-eastern coast 23 24 of the Algarve, Portugal (Fig. 1). It is separated from the Atlantic Ocean by a series of barrier 25 islands and two peninsulas. About 80% of the lagoon is intertidal with a semi-diurnal tidal 26 regime and tidal ranges between 1.3 m during neap tides and 3.5 m during spring tides 27 (Cabaco et al., 2012). Due to negligible inflow of fresh water and high exchange of water 28 with the open Atlantic during each tidal cycle, the salinity within the lagoon is 35 to 36 PSU 29 year round, except for periods of heavy rainfalls. About a quarter of the intertidal area (13.04 30 km<sup>2</sup>) is covered by dense stands of Zostera noltii Hornem (Guimarães et al., 2012).Further, 31 but much less abundant, seagrass species in the lagoon are Zostera Marina L. and Cymodocea nodosa (Ucria) Ascherson which are mainly located in shallow parts of the subtidal areas
 (Santos et al., 2004). About 30% of the lagoon's area is covered with salt marsh communities
 (Rui Santos, pers. comm.).

#### 4 2.2 Sampling

5 We conducted two sampling campaigns in the western part of the lagoon at the Ramalhete 6 research station (Centre of Marine Sciences (CCMAR), Universidade do Algarve) in the vicinity of Faro (37.0°N, 7.6 W) (Fig. 1). The sampling was carried out from July 23<sup>rd</sup> -7 August 7<sup>th</sup> 2011 and April 17<sup>th</sup> – April 28<sup>th</sup> 2012 coinciding with the beginning (2012 8 9 campaign) and peak (2011 campaign) of the seagrass reproductive season. Ambient air temperatures were distinctively different between both campaigns ranging from 21 to 27°C 10 (mean 24°C) with almost entirely clear weather in summer and 13 to 23°C (mean 17°C) in 11 spring with frequent strong cloud cover. Mean water temperatures were 25.9°C (summer) and 12 17.5°C (spring). The prevailing wind direction during both campaigns was West South-West 13 to with rather low average wind speeds of 4 m s<sup>-1</sup> during summer and 5 m s<sup>-1</sup> during spring. 14

15 During the two campaigns we used different dynamic flux chamber systems. During the 2011 16 campaign, we measured the halocarbon fluxes during air exposure using a quartz-glass chamber  $(0.1m^2 \text{ surface area, 7 L enclosure volume})$  as described in Weinberg et al. (2013) 17 with some adjustments. For this study a permanent backup flow  $(3 \pm 0.2 \text{ Lmin}^{-1})$  through the 18 19 flux chamber during sampling and the change of cryotraps was applied to ensure sufficient mixing. Further, to overcome analytical problems with the high humidity in the sampled air, 20 21 the water content was reduced using a condenser (-15°C). Briefly, the quartz-glass flux 22 chamber was placed on the seagrass patch and sealed with surrounding sediment. Two sampling systems were operated simultaneously measuring inlet and outlet air of the flux 23 chamber (flow rate  $1 \pm 0.2 \text{ Lmin}^{-1}$ ). Prior to sampling, the flux chamber was flushed for about 24 25 10 min ensuring sufficient equilibration of compounds in the chamber air.

During the 2012 campaign, we used a dynamic flux chamber system (0.037 m<sup>2</sup> bottom surface area, 8 L enclosure volume) suitable for flux measurements during both periods of air exposure and tidal immersion. The properties and setup of this dynamic chamber system is described in detail elsewhere (Bahlmann et al., 2014). Since this system acts as an ordinary purge and trap system, the extraction efficiencies were simulated using halocarbon equilibrated artificial seawater. While the results from these tests revealed that 1 monohalomethanes were almost completely extracted ( $\geq$ 90%), the purge efficiencies for 2 CHBr<sub>3</sub> were only 33%. Thus the reported CHBr<sub>3</sub> fluxes determined from seagrass meadows 3 using the submergible chamber system represent an under-estimate.

Based on the sampling system for the determination of stable carbon isotopes of halocarbons (Bahlmann et al., 2011), we modified the cryogenic trapping system for the measurements of halocarbon mixing ratios, in order to establish a better temporal resolution by reducing the analysis time. This results in a final air volume of 28± 5 L of air at the inlet and the outlet of the chambers, respectively. The specifications along with the results from test surveys are given in the Supplementary Materials.

10 The seagrass species sampled was exclusively Z. Noltii. The seagrass patches sampled had an area coverage of >95% and were free of visible epiphytes such as macroalgae. In this low to 11 12 medium intertidal region the epiphytes of Z .Noltii are almost exclusively diatoms whose 13 contribution ranges from 0.5 to 4% of the total seagrass biomass (Cabaco et al., 2009). We 14 further determined the fluxes from an adjacent bare sediment spot during the 2011 campaign. On 2 August 2011, these chamber-based measurements were complemented by atmospheric 15 sampling at a nearby beach (Praia de Faro, upwind site) about 3 km distant from the lagoon 16 during the summer campaign 2011 (Fig. 1). At this time the wind direction was south-17 18 westerly reflecting background air from the coastal ocean.

19 Discrete water samples for the determination of dissolved halocarbons concentration and isotopic composition at high tide were taken during both campaigns. The samples were taken 20 21 directly above the studied seagrass meadow using Duran glass bottles (1-2 L volume). Air and 22 sediment intrusions during water sampling were avoided. The water depth was between 0.3 m and 1 m. On April 24<sup>th</sup> 2012, a transect cruise through the middle and western part of the 23 lagoon was conducted during rising waters (Fig. 1). The water samples were taken from a 24 25 water depth of 1 m. Dissolved halocarbons were extracted from seawater using a purge and trap system. Seawater was purged with helium 5.0 (purge flow 1L min<sup>-1</sup>) for 30 minutes. 26 27 After water vapour reduction of the purge gas, the compounds were enriched on cryotraps 28 (submerged in a dry shipper). The shape of the cryotraps used here was the same as those for 29 flux chamber and atmospheric samples. The water samples were usually processed within 30 30 minutes after sampling. Samples from the transect cruise were stored in the dark at 4°C and 31 analyzed within eight hours. Purge efficiencies of monohalomethanes from lagoon water were  $\geq$ 95% (1 L and 2 L samples). However, the less volatile CHBr<sub>3</sub> was only extracted with 50% 32

(1 L samples) and 30% (2 L samples). Therefore, the results of water concentration were
 corrected for the respective purge efficiency for this compound.

#### 3 **2.3 Measurement and quantification**

4 The measurement procedure is described in detail in the Supplementary Materials. Briefly, 5 compounds adsorbed on the cryotraps, were thermally desorbed and transferred to Peltier-6 cooled adsorption tubes. The analytes were further desorbed from the adsorption tubes and refocused cryogenically before injection to the GC-MS system. Air and water samples were 7 8 measured on-site at Ramalhete research station using a GC-MS system (6890N/5975B, 9 Agilent, Germany) equipped with a CP-PorabondQ column (25 m, 0.25 µm i.d., Varian, Germany). The GC-MS was operated in the electron impact mode. Identification of 10 compounds was executed by retention times and respective mass spectra. Aliquots of gas 11 12 standard (Scott EPA TO 15/17, 65 compounds, 1 ppm each in nitrogen, Sigma Aldrich, 13 Germany) containing CH<sub>3</sub>Cl, CH<sub>3</sub>Br, and CHBr<sub>3</sub> were applied to quantify the target 14 compounds. During onsite measurements, CH<sub>3</sub>I was quantified using the response factor 15 against CH<sub>3</sub>Br. The response factor was determined prior to the campaign. Equivalent 16 amounts of CH<sub>3</sub>I and CH<sub>3</sub>Br from single gas standards were analysed together for the response factor calculation. The analytical limit of detection was 0.3 ppt (pmol  $mol^{-1}$ ) for the 17 18 halocarbons. The accuracy of the entire sampling method (sampling, sample treatment, 19 measurement) was derived from test samples in triplicates. The deviation between the 20 individual samples for CH<sub>3</sub>Cl, CH<sub>3</sub>Br, CH<sub>3</sub>I, and CHBr<sub>3</sub> was 5.4%, 6.3%, 15.4% and 6.7%, 21 respectively. A series of procedural blanks (cryotraps and adsorption tubes) were taken during 22 the sampling campaigns. The occasionally detected blanks of CH<sub>3</sub>Cl and CH<sub>3</sub>Br from these 23 determinations were  $\leq 3\%$  to the "real" samples taken from the seagrass meadows during sampling campaigns. Therefore, the halocarbon fluxes were not blank corrected. 24

Air and water samples for determining the isotopic composition of halocarbons were transferred to adsorption tubes and stored at -80°C until measurements. The analysis was conducted using the GC-MS-IRMS system at our home laboratory (Bahlmann et al., 2011). Additional transport and storage blanks were processed which revealed no contamination for all halocarbons studied.

#### 1 2.4 Calculations

The fluxes were determined with dynamic flux chambers. The chamber is positioned on the a sampling spot and flushed continuously with ambient air. The mixing ratios of compounds at the inlet and outlet air are then measured. The difference of mixing ratios of compounds between inlet and outlet air along with the flushing rate and the surface area are used for the flux calculation ( $F_{Net}$ , nmol m<sup>-2</sup> h<sup>-1</sup>):

7 
$$F_{Net} = \frac{Q \times (C_{out} - C_{in})}{A \times V \times 1000}$$
(1)

8 Here, Q is the flushing rate of air through the chamber (L h<sup>-1</sup>),  $C_{out}$  and  $C_{in}$  are the mixing 9 ratios of target compounds (ppt) at the outlet and the inlet of the flux chamber. A is the 10 enclosed surface area of the flux chamber (m<sup>2</sup>) and V is the molar volume (L) at 1013.25 mbar 11 and 298.15 K.

For calculation of the sea-air fluxes from the lagoon water, the inlet samples of the flux chamber were used which reflect the air mixing ratios. Where no corresponding inlet sample was available, the campaign means were applied. After conversion of the air mixing ratios to pmol  $L^{-1}$  using temperature data and the respective molar volume of the ambient air, the seaair fluxes (F, nmol m<sup>-2</sup> h<sup>-1</sup>) of halocarbons were calculated by the equation:

17 
$$F = k_w \times (C_w - C_a \times H^{-1})$$
 (2)

where  $k_w$  is the gas exchange velocity (m h<sup>-1</sup>),  $C_w$  and  $C_a$  the water concentration and air 18 concentration (pmol  $L^{-1}$ ), respectively, and H the dimensionless and temperature dependent 19 20 Henry's law constant taken from Moore (2000) for CH<sub>3</sub>Cl, Elliott and Rowland (1993) for CH<sub>3</sub>Br and CH<sub>3</sub>I, and Moore et al. (1995) for CHBr<sub>3</sub>. Several approximations emerged to 21 22 estimate the relationship between the gas exchange velocity k and the wind speed u for open and coastal oceans (e.g. Nightingale et al., 2000; Wanninkhof, 1992). These estimations rely 23 on assumptions that trace gas exchange is based on wind-driven turbulence. This is not 24 25 applicable in shallow estuarine and riverine systems where the sea-air gas exchange is 26 additionally driven by wind-independent currents and the bottom turbulence and thus water 27 depth and current velocities further play a major role (Raymond and Cole, 2001). Studying the sea-air exchange in the Ria Formosa, these additional factors have to be considered in 28 29 addition to wind driven outgassing. Therefore, we used the parameterization of  $k_w$  with the assumption that wind speed and water current driven turbulence are additive (Borges et al.,
 2004):

(3)

3 
$$k_w = 1.0 + 1.719 \times w^{0.5} \times h^{-0.5} + 2.58 \times u$$

where w is the water current (cm s<sup>-1</sup>), h the water depth (m) and u the wind speed (m s<sup>-1</sup>). For 4 the calculations of the sea-air flux in the lagoon, a mean water depth of 1.5m (Tett et al., 5 2003) and a mean water current of 24 cm s<sup>-1</sup> (Durham, 2000) was used. The Schmidt number 6 (Sc) expresses the ratio of transfer coefficients of the kinematic viscosity of water and gas 7 diffusivity of interest. The gas exchange velocity  $k_w$  for each gas was then normalized to a 8 Schmidt number of 660, assuming a proportionality to  $Sc^{-0.5}$  (Borges et al., 2004). The 9 individual Schmidt numbers were obtained from Tait (1995) for CH<sub>3</sub>Cl, De Bruyn and 10 11 Saltzman (1997) for CH<sub>3</sub>Br and CH<sub>3</sub>I, and Quack and Wallace (2003) for CHBr<sub>3</sub>.

12

## 13 3 Results

#### 14 **3.1** Halocarbons in the atmosphere and lagoon water

15 The air mixing ratios in the lagoon were adopted from the inlets of the flux chambers at 1 m above ground during both campaigns. The results of these measurements and those of the 16 17 upwind site outside the lagoon (Praia de Faro) are presented in Table 1. In summer, elevated 18 air mixing ratios of the monohalomethanes were observed during periods of easterly winds 19 when air masses at the sampling site had presumably passed over major parts of the lagoon. These mixing ratios reached up to 1490 ppt for CH<sub>3</sub>Cl, 61 ppt for CH<sub>3</sub>Br, and 11 ppt for CH<sub>3</sub>I 20 reflecting a potent source in this system. The mixing ratios at the upwind site (Praia de Faro) 21 22 were distinctively lower with mean values of 613 ppt (CH<sub>3</sub>Cl), 13 ppt (CH<sub>3</sub>Br), 1 ppt (CH<sub>3</sub>I), 23 and 8 ppt (CHBr<sub>3</sub>) further indicating a source inside the lagoon. In spring 2012, the mean air 24 mixing ratios in the lagoon were significantly lower than during summer with 654 ppt for CH<sub>3</sub>Cl, 12 ppt for CH<sub>3</sub>Br, 1 ppt for CH<sub>3</sub>I, and 2 ppt for CHBr<sub>3</sub>. 25

Discrete water samples were taken above the studied seagrass meadow during tidal inundation (summer n=9; spring n=10). The results are presented in Table 1. In summer, concentrations ranged from 158 to 301 pmol  $L^{-1}$  (CH<sub>3</sub>Cl), 5 to 11 pmol  $L^{-1}$  (CH<sub>3</sub>Br), 4 to 18 pmol  $L^{-1}$  (CH<sub>3</sub>I), and 67 to 194 pmol  $L^{-1}$  (CHBr<sub>3</sub>). During the spring campaign, the water concentrations were 1 101 to 267 pmol  $L^{-1}$  for CH<sub>3</sub>Cl, 6 to 28 pmol  $L^{-1}$  for CH<sub>3</sub>Br, 2 to 16 pmol  $L^{-1}$  for CH<sub>3</sub>I, and 39 2 to 133 pmol  $L^{-1}$  for CHBr<sub>3</sub>.

The results obtained from samples of the transect cruise covered in 2012 (Fig. 1) are given in 3 Table 2. We observed an about two-fold increase of concentration for CH<sub>3</sub>Cl (from 121 to 4 241 pmol  $L^{-1}$ ) and CHBr<sub>3</sub> (from 26 to 55 pmol  $L^{-1}$ ) between position 1 (Faro-Olhão inlet) and 5 position 2 (near to the seagrass meadows studied). The increase was less pronounced for 6 CH<sub>3</sub>Br (5 to 7 pmol L<sup>-1</sup>) and not notable for CH<sub>3</sub>I. The seawater at positions 6 and 7, the 7 nearest to the Ancão inlet, revealed rather low concentrations for all compounds. We further 8 9 observed rising concentrations for all halocarbons along positions 3, 4, and 5 with increasing distance to the Ancão inlet. They increased from 96 to 180 pmol L<sup>-1</sup> for CH<sub>3</sub>Cl, from 9 to 19 10 pmol L<sup>-1</sup> for CH<sub>3</sub>Br, 2 to 14 pmol L<sup>-1</sup> for CH<sub>3</sub>I, and 21 to 95 pmol L<sup>-1</sup> for CHBr<sub>3</sub>. 11

#### 12 **3.2** Fluxes from seagrass meadows, sediment, and sea-air exchange

The mean fluxes and ranges of CH<sub>3</sub>Cl, CH<sub>3</sub>Br, CH<sub>3</sub>I, and CHBr<sub>3</sub> from seagrass meadows,
sediment, and from sea-air exchange calculations obtained from the two sampling campaigns
are given in Table 3.

16 During the summer campaign (air exposure), we observed highly variable emission and deposition fluxes ranging from -49 to 74 nmol  $m^{-2} h^{-1}$  and -5.7 to 130 nmol  $m^{-2} h^{-1}$  for CH<sub>3</sub>Cl 17 and CH<sub>3</sub>Br, respectively. The variability was less pronounced for CH<sub>3</sub>I (0.5 to 2.8 nmol m<sup>-2</sup> h<sup>-</sup> 18 <sup>1</sup>) and CHBr<sub>3</sub> (-0.6 to 5.7 nmol  $m^{-2} h^{-1}$ ) where predominantly emissions were measured. 19 Strongly elevated fluxes up to 130 nmol m<sup>-2</sup> h<sup>-1</sup> for CH<sub>3</sub>Br were recorded in conjunction with 20 tidal change from air exposure to inundation and conversely. These high fluxes were 21 substantiated by a concurrent enhanced atmospheric mixing ratios ranging from 23 ppt to 118 22 ppt (campaign median 14 ppt). Omitting these compound-specific tidal phenomena, the fluxes 23 of CH<sub>3</sub>Cl and CH<sub>3</sub>Br were positively correlated ( $R^2 0.55$ , p< 0.05). There were no significant 24 25 correlations between CH<sub>3</sub>I and CHBr<sub>3</sub> and the other investigated halocarbons. Due to the inherent high variability of the fluxes, halocarbon fluxes were poorly correlated with solar 26 radiation ( $R^2 \le 0.20$ ). 27

- 28 The flux chamber measurements over the sediment during air exposure revealed 29 predominantly emissions of all four halocarbons (n=5). These fluxes were  $3.6\pm4.3$  nmol m<sup>-2</sup> h<sup>-</sup>
- $30^{-1}$  (CH<sub>3</sub>Cl), 0.6±0.5 nmol m<sup>-2</sup> h<sup>-1</sup> (CH<sub>3</sub>Br), 0.3±0.2 nmol m<sup>-2</sup> h<sup>-1</sup> (CH<sub>3</sub>I), and 0.8±1.0 nmol m<sup>-2</sup>
- 31 h<sup>-1</sup> (CHBr<sub>3</sub>). Except for CH<sub>3</sub>I, the halocarbon fluxes were statistically significant different

1 from zero (Mann-Whitney-U test; p <0.05). Hence, the bare sediment may contribute to the

2 overall emissions above the seagrass by about 10 to 20% for CH<sub>3</sub>Cl and CH<sub>3</sub>Br, and 45% for

3 CHBr<sub>3</sub>.

4 During the 2012 spring campaign the halocarbon fluxes from seagrass meadows were determined during both, periods of air exposure and periods of tidal immersion. Furthermore, 5 6 the measurements were complemented by other trace gases including hydrocarbons and sulphur containing compounds (Bahlmann et al., 2014). As in the summer campaign, the 7 seagrass meadows were a net source for all halocarbons studied, but on a lower level. The 8 individual ranges of air exposure measurements were -30 to 69 nmol m<sup>-2</sup> h<sup>-1</sup> (CH<sub>3</sub>Cl), -0.8 to 9 3.9 nmol m<sup>-2</sup> h<sup>-1</sup> (CH<sub>3</sub>Br), -0.6 to 2.6 nmol m<sup>-2</sup> h<sup>-1</sup> (CH<sub>3</sub>I), and -0.5 to 1.3 nmol m<sup>-2</sup> h<sup>-1</sup> 10 (CHBr<sub>3</sub>). On average, the seagrass meadows were a net source also under submerged 11 conditions ranging from -58 to 100 nmol m<sup>-2</sup> h<sup>-1</sup> for CH<sub>3</sub>Cl, -1.6 to 8.3 nmol m<sup>-2</sup> h<sup>-1</sup> for 12 CH<sub>3</sub>Br, 0.1 to 8.0 nmol  $m^{-2}$  h<sup>-1</sup> for CH<sub>3</sub>I, and -0.4 to 10.6 nmol  $m^{-2}$  h<sup>-1</sup> for CHBr<sub>3</sub>. Due to the 13 14 low purge efficiency of CHBr<sub>3</sub> during high tide measurements, the fluxes determined with the submergible chamber are underestimated for this compound. Despite this high variability in 15 16 production/decomposition during air exposure and inundation, the monohalomethanes were significantly correlated to each other ( $R^2 > 0.50$ ). These correlations were enhanced compared 17 to those found when the seagrass meadows were air-exposed ( $R^2 \ge 0.50$ ). In this case, only 18 CH<sub>3</sub>I and CH<sub>3</sub>Br were significantly correlated (R<sup>2</sup> 0.51). CHBr<sub>3</sub> was only slightly correlated 19 to the monohalomethanes. 20

While deposition fluxes of CH<sub>3</sub>Cl and CH<sub>3</sub>Br of air-exposed seagrass meadows occurred predominantly during periods of low irradiance in summer, no obvious relation to the time of day and/or solar radiation was observed during spring when deposition fluxes were frequently detected. For CH<sub>3</sub>I and CHBr<sub>3</sub>, uptake was only occasionally observed and situations of emission clearly dominated.

As in the summer campaign, we observed some remarkable tidal effects on halocarbon fluxes during the spring campaign. Firstly, the highest fluxes of all halocarbons were measured when the lagoon water was just reaching the sampling site. Occasionally this was also observed from air exposure to tidal inundation, although less pronounced. However, these short-timed effects were not as strong as during the summer campaign. Secondly, at tidal maximum we observed deposition fluxes for CH<sub>3</sub>Cl and CH<sub>3</sub>Br and deposition fluxes or very weak emissions for CH<sub>3</sub>I and CHBr<sub>3</sub>. Before and after this period, emission fluxes during incoming
 tide and ebb flow dominated.

The lagoon water was a net source for all investigated halocarbons to the atmosphere during both campaigns. In summer, the flux ranges were 13-45 nmol  $m^{-2} h^{-1}$  (CH<sub>3</sub>Cl), 0.6-1.7 nmol  $m^{-2} h^{-1}$  (CH<sub>3</sub>Br), 0.5-3.2 nmol  $m^{-2} h^{-1}$  (CH<sub>3</sub>I), and 1.0-8.0 nmol  $m^{-2} h^{-1}$  (CHBr<sub>3</sub>). The respective fluxes in spring were 3.5-32 (CH<sub>3</sub>Cl), 0.5-4.1 (CH<sub>3</sub>Br), 0.3-3.7 (CH<sub>3</sub>I), 3.8-24 (CHBr<sub>3</sub>).

#### 8 **3.3** Stable carbon isotopes of halocarbons

9 Stable carbon isotope ratios of halocarbons were determined for selected samples of both 10 campaigns (Table 4). Isotopic source signatures from seagrass meadows for  $CH_3Cl$  and 11  $CH_3Br$  were calculated using a coupled isotope and mass balance without integration of a 12 possible sink function (Weinberg et al., 2013).

In 2011, the difference in atmospheric mixing ratios of CH<sub>3</sub>Cl and CH<sub>3</sub>Br between within the lagoon and the upwind position (Praia de Faro) was accompanied by a shift of  $\delta^{13}$ C values. More <sup>13</sup>C depleted values were found for CH<sub>3</sub>Cl in the lagoon (-42±2‰) compared to the upwind position (-39±0.4‰). In contrast, the  $\delta^{13}$ C values of CH<sub>3</sub>Br were significantly enriched in <sup>13</sup>C by about 10‰ inside the lagoon (-29±5‰) as compared to the upwind site (-38±3). These  $\delta^{13}$ C values found in air samples in the lagoon roughly correspond to the  $\delta^{13}$ C values of CH<sub>3</sub>Cl (-43±3‰) and CH<sub>3</sub>Br (-23±3‰) found in samples of lagoon waters.

Atmospheric CH<sub>3</sub>Cl and CH<sub>3</sub>Br were on average more enriched in <sup>13</sup>C in spring than in summer by 4 and 6‰, respectively. While the  $\delta^{13}$ C values of CH<sub>3</sub>Cl in the lagoon water were quite similar between both periods of the year, those of CH<sub>3</sub>Br were on average more depleted in <sup>13</sup>C during spring suggesting certain changes in production/decomposition processes. The isotopic composition of CH<sub>3</sub>I in lagoon water was quite similar between summer (-39± 9‰) and spring (mean -37±7‰). As for CH<sub>3</sub>Br, the  $\delta^{13}$ C values of CHBr<sub>3</sub> were more enriched in <sup>13</sup>C in summer when compared with those of the spring campaign.

The difference in concentration along the transect cruise was accompanied by variations in the carbon isotopic composition of all compounds (Table 1, Figure 2). The most <sup>13</sup>C depleted values of CH<sub>3</sub>Cl, CH<sub>3</sub>Br, and CH<sub>3</sub>I were detected at the position furthest from the inlet. Interestingly, CHBr<sub>3</sub> showed the opposite trend with more <sup>13</sup>C enriched values in the lagoon (-25.8‰ vs. ~ -18‰).

Using the fluxes and  $\delta^{13}$ C values from the inlet and outlet of the flux chamber, we were able 1 2 to calculate the source signatures of seagrass covered areas. The resulting source signatures of 3 CH<sub>3</sub>Cl from seagrass meadows were similar during both campaigns ( $-51\pm6\%$  and  $-56\pm2\%$ ). 4 respectively) and independent from the strength of emission. For CH<sub>3</sub>Br, we observed most depleted  $\delta^{13}$ C values of -53‰ and -58‰ at increased emission fluxes in summer, but values 5 of -26‰ and -29‰ during periods of low emission. This corroborates the findings of 6 7 isotopically heavy CH<sub>3</sub>Br produced within the seagrass meadows (-29‰) in spring 2012 8 when all samples analysed for the isotopic composition were taken at situations of low 9 emission.

10

# 11 4 Discussion

#### 12 4.1 Dissolved halocarbons

13 A comparison of halocarbon concentrations in the lagoon water to other measurements of the 14 coastal Atlantic found in the literature is displayed in Table 5. The lagoon waters appeared to be highly enhanced in CH<sub>3</sub>Cl. Except one early study of Tait et al. (1994), our measurements 15 gave the most elevated concentrations for this compound. Enhanced concentrations in the 16 lagoon waters were also found for CH<sub>3</sub>Br. Given the mean concentrations from other coastal 17 18 Atlantic studies (Baker et al., 1999; Carpenter et al., 2000; Hu et al., 2010), we recorded 19 higher concentration by a factor of 2 to 3 at our sampling site. The average water 20 concentrations in the lagoon of CH<sub>3</sub>I were in the same range as reported from other parts of the Atlantic (Moore and Groszko 1999; Zhou et al., 2005). However, especially those regions 21 22 where macroalgae are the dominating source organisms possess higher maximum values (Bravo-Lineares and Mudge, 2009; Jones et al., 2009). This is even more pronounced for 23 24 CHBr<sub>3</sub>, for which the seawater concentration within or in the vicinity of macroalgae beds are strongly elevated (Bravo-Lineares and Mudge, 2009; Carpenter et al., 2000; Jones et al., 25 2009). The area occupied by the prevalent macroalgae species Enteromorpha spp. and Ulva 26 *spp.* in the Ria Formosa is estimated to be 2.5  $\text{km}^2$  (Duarte et al., 2008), considerably below 27 that of other abundant sources such as seagrass meadows. We cannot exclude that 28 29 phytoplankton contributes significantly to the water concentration of halocarbons, but the predominantly low chlorophyll a concentrations (3.06  $\mu$ g L<sup>-1</sup> from long-term measurements, 30 Brito et al., 2012) and low water volumes seem to limit the impact from this source. 31

Despite the short residence time of the lagoon water masses of which 50-75% is exchanged during one tidal cycle (Brito et al., 2010), the transect cruise along the main channels revealed a successive enrichment of halocarbon concentration in the water with increasing distance from the main inlets (Fig. 1 and Table 2). Therefore, the net halocarbon production in the lagoon appears to clearly exceed that outside the lagoon. This is supported by the distinctively increased air mixing ratios of halocarbons in the lagoon as compared to the upwind site (Table 1).

8 Overall, the lagoon seems to comprise highly potent halocarbon sources in the water column
9 for CH<sub>3</sub>Cl and CH<sub>3</sub>Br but not for CH<sub>3</sub>I and CHBr<sub>3</sub>.

#### 10 **4.2** Flux pattern from seagrass meadows

11 The halocarbon fluxes from seagrass meadows were characterized by a high variability with deposition and emission fluxes occurring at all sampling spots. The same was observed within 12 other studies investigating halocarbon fluxes in coastal environments (e.g. Blei et al., 2010; 13 Manley et al., 2006; Rhew et al., 2000). Halocarbon dynamics in coastal systems where 14 15 multiple sources and sinks interact are complex, and it should be noted that the fluxes discussed here refer to the entire benthic community constituting the seagrass meadows. Thus, 16 17 some variability may relate to the activity of distinct source organisms which may be 18 stimulated by different environmental factors. To gain insights into the common 19 environmental controls for this ecosystem we discuss the following factors i) diurnal variations ii) tidal effects and iii) seasonal dependence. 20

21 i) Diurnal variations. The correlation analysis with solar radiation resulted in only a weak 22 association with the magnitude of fluxes. However, after grouping by daytime, our data provide some indication for a diurnal pattern (Fig. 2). For CH<sub>3</sub>Cl, there was the most obvious 23 relationship between time of day and actual emissions. Highest emissions were observed 24 during day periods with increased sunlight (midday and afternoon). In contrast, deposition 25 26 fluxes were exclusively recorded during periods of low radiation and nighttimes. The same 27 was also observed for CH<sub>3</sub>Br. However, highest mean emissions of this compound seemed to be shifted towards the afternoon. CH<sub>3</sub>I was constantly emitted from the seagrass covered spot 28 29 revealing a weak diurnal dependence. The emissions did not cease during periods of low irradiance and darkness. Nevertheless, elevated mean emissions were observed in the 30

afternoon. Except one occasion, CHBr<sub>3</sub> was emitted throughout the sampling periods. Mean
 emissions were higher around midday and afternoon as during night.

3 Several studies from salt marshes reported a diurnal trend of halocarbon emissions initiated 4 by irradiance (Dimmer et al., 2001; Rhew et al., 2000, 2002; Drewer et al., 2006). The flux data of halocarbons from the summer campaign with elevated fluxes during midday and 5 6 afternoon suggest a similar pattern also in seagrass meadows. However, this was more 7 obvious for CH<sub>3</sub>Cl and CH<sub>3</sub>Br than for CH<sub>3</sub>I and CHBr<sub>3</sub>. The lower production of CH<sub>3</sub>I 8 during the time of highest light intensity cannot fully be explained. Blei et al. (2010) reported 9 that the main environmental association in salt marsh emissions of CH<sub>3</sub>Cl and CH<sub>3</sub>Br was 10 with ambient temperature rather than light. However, during the summer campaign, temperature variations (day/night) were too low to explain the observed emission/deposition 11 12 pattern of CH<sub>3</sub>Cl and CH<sub>3</sub>Br.

13 It is known that coastal sediments can act as a sink for CH<sub>3</sub>Cl and CH<sub>3</sub>Br mainly due to 14 microbial degradation (Miller et al., 2004; Oremland et al. 1994). This would support our findings of the deposition fluxes during night times where production above the sediment is 15 presumably lower than during daytime (summer campaign). While, in general, the deposition 16 fluxes of CH<sub>3</sub>Cl and CH<sub>3</sub>Br occurred more frequently during the spring campaign, they did 17 18 not exhibit a day-night-relationship. Moreover, the dependence of light intensity on the 19 magnitude of emission fluxes of halocarbons seemed to have a minor effect during this period 20 of the year.

21 ii) Tidal effects. During the spring campaign, mean fluxes derived from submerged seagrass 22 meadows were elevated by factors of 17 (CH<sub>3</sub>Cl), 5 (CH<sub>3</sub>Br), 3 (CH<sub>3</sub>I), and 8 (CHBr<sub>3</sub>) when 23 compared to the average fluxes during air exposure. This clearly higher production of halocarbons under submerged conditions was quite unexpected, as in general it is believed 24 25 that the production of trace gases during low tide exceeds that during inundation. For halocarbons this was suggested for example by Carpenter et al. (1999) and Jones et al. (2009) 26 27 from atmospheric measurements over intertidal macroalgae beds in Mace Head, Ireland. 28 Nevertheless, in accordance with our results from halocarbon measurements we also observed 29 higher primary productivity by increased CO<sub>2</sub> uptake during submerged conditions 30 (Bahlmann et al., 2014). Therefore, the higher productivity may reflect higher enzymatic 31 activity (e.g. methyltransferases) within the organisms of the seagrass community, by which 32 monohalomethanes are presumably formed. Furthermore, the correlation analysis revealed a

different behaviour of halocarbons between the two tidal states with stronger correlations between monohalomethanes during tidal inundation than air exposure. Obviously the change in environmental conditions was accompanied with a shift in the halocarbon productiondecomposition pattern of the benthic community and/or different source organisms were stimulated.

6 An interesting outcome of both campaigns is the observation of strongly elevated halocarbon 7 fluxes during tidal change from air exposure to submergence and reversely (Table 3). 8 Continuous high-time resolution CO<sub>2</sub> and methane flux measurements performed in spring 9 2012 (Bahlmann et al., 2014) principally support this observation. At the particular moment when the water reached the sampling site, we observed a distinct peak flux of methane and 10 CO<sub>2</sub>. This may be evidence for processes in the sediments attributable to changes in 11 hydrodynamic pressures resulting in the release of trace gases trapped in sedimentary pore 12 13 spaces (Bahlmann et al., 2014). The remarkable deposition flux of CH<sub>3</sub>Cl and CH<sub>3</sub>Br during 14 the maximum water level (Table 3) was accompanied by highest emissions of other trace 15 gases such as methanethiol and hydrogen sulfide as discussed by Bahlmann et al. (2014). 16 These compounds are effective nucleophiles which could have contributed to the degradation 17 of halocarbons as described in Barbash and Reinhard (1989). This suggests a significantly different biogeochemistry during this period compared with tide and ebb flow. 18

Overall, while there is evidence for a tidal control on halocarbon production anddecomposition, additional research is needed to further elucidate these phenomena.

21 iii) Seasonal dependence. There are considerable differences between the results from spring 22 and summer campaign. We observed elevated mixing ratios for all halocarbons in ambient air 23 as well as higher water concentrations for CH<sub>3</sub>Cl, CH<sub>3</sub>I, and CHBr<sub>3</sub> compounds in summer (Table 1). This observed signal of general increased halocarbon production in the lagoon 24 25 during summer might be attenuated by enhanced degradation in the water phase and sediments at higher temperatures. Nevertheless, given the calculated sea-air flux there is only 26 27 little evidence for a pronounced seasonal relationship in halocarbon volatilisation to the 28 atmosphere from the lagoon water. While the fluxes of CH<sub>3</sub>Cl appeared to be enhanced in 29 summer, those of CH<sub>3</sub>Br and CH<sub>3</sub>I seemed to be quite similar between spring and summer. 30 CHBr<sub>3</sub> emissions were actually higher in spring than in summer due to higher water 31 concentrations. Comparing the data obtained from air-exposed sites during the two 32 campaigns, the fluxes in summer were strongly enhanced by factors of 16 (CH<sub>3</sub>Cl and

1 CH<sub>3</sub>Br), 2 (CH<sub>3</sub>I), and 5 (CHBr<sub>3</sub>). Moreover, the halocarbon fluxes showed a distinct diurnal 2 cycle during summer but not during spring. The differences of ambient conditions between 3 the campaigns with lower air temperatures and lower solar radiation in spring may have 4 contributed to the differences in the emission patterns of halocarbons. That these 5 environmental conditions can substantially influence the magnitude of fluxes was reported from other ecosystems such as salt marshes (Blei et al., 2010; Manley et al., 2006). However, 6 7 further studies covering the entire seasoning are necessary to fully unravel the annual 8 halocarbon emissions from seagrass meadows.

## 9 4.3 Halocarbons sources in the lagoon: an isotopic perspective

The results from the atmospheric sampling of Praia de Faro air (upwind) and lagoon air 10 revealed differences regarding the mixing ratios and  $\delta^{13}$ C values of CH<sub>3</sub>Cl and CH<sub>3</sub>Br (Tables 11 1 and 4). We observed elevated concentrations in the lagoon for both compounds, whereby 12 13 the higher concentrations were accompanied with shifts towards isotopically light CH<sub>3</sub>Cl but 14 heavy CH<sub>3</sub>Br. Sources other than the studied seagrass meadows, for example the abundant 15 salt marshes, may have contributed substantially to the elevated mixing ratios. Assuming atmospheric stable conditions with negligible sinks in the atmosphere, the difference of air 16 mixing ratios and  $\delta^{13}$ C values between upwind air and lagoon air should reflect the isotopic 17 18 source signature within the lagoon. Therefore, as a first approach, an isotope mass balance 19 was used by integrating mean data from both sampling sites (Tables 1 and 4). The resulting 20 source signatures within the lagoon are -49‰ for CH<sub>3</sub>Cl and -16‰ for CH<sub>3</sub>Br.

21 Isotopic source signatures of CH<sub>3</sub>Cl from seagrass meadows during chamber incubations (air exposure) in the Ria Formosa were -51±6‰ (summer) and -56±2‰ (spring). During the 22 summer campaign, CH<sub>3</sub>Cl emissions from the salt marsh plant Spartina maritima were 23 determined with  $\delta^{13}$ C values of -66 and -72‰. These values are in good agreement with those 24 25 of Bill et al. (2002) from a Californian salt marsh (-69 to -71‰, daytime values). 26 Unfortunately, we do not have isotopic data for the inundated periods from seagrass meadows, but the  $\delta^{13}$ C values of CH<sub>3</sub>Cl in the water phase (-42±2‰) come close to those 27 measured in the atmosphere. An abiotic production mechanism has been reported for CH<sub>3</sub>Cl 28 from senescent plant material (Hamilton et al., 2003). While we cannot generally exclude 29 30 additional CH<sub>3</sub>Cl generation via this pathway, the isotopic data obtained in the Ria Formosa do not mirror strongly <sup>13</sup>C depleted values ( $\delta^{13}$ C of -135±12‰, Keppler et al., 2004) as 31 32 expected for compounds from this production mechanism. Overall, this rather indicates a 1 stronger imprint of the seagrass meadows and/or water column on the atmospheric  $CH_3Cl$ 

2 than from salt marshes or abiotic processes.

With  $\delta^{13}$ C values of -42±17‰ the source signature of CH<sub>3</sub>Br from seagrass meadows tend to 3 be more depleted in <sup>13</sup>C compared with the calculated source signature from the atmospheric 4 samples. It should be noted that the  $\delta^{13}$ C values for this compound were more depleted in  ${}^{13}$ C 5 during periods of increased emission (-55%) than during low emissions (-28%). This shift 6 7 can most likely be explained by degradation processes in the sediments which occurred 8 simultaneously. This corroborates our observations from Northern Germany with subsequent 9 recalculation of a sedimentary sink function from accompanying sediment measurements 10 (Weinberg et al., 2013). Reported source signatures of CH<sub>3</sub>Br from salt marshes range from -59 to -65‰ (day time values, Bill et al., 2002). Our own measurements in the Ria Formosa 11 indicate similar  $\delta^{13}$ C values (-65‰) or even more depleted ones (unpublished data). In any 12 case, neither source signatures from seagrass meadows nor salt marshes seem to match the 13 14 overall source signature estimated from the atmospheric samples. Therefore, it is most likely 15 that the atmospheric CH<sub>3</sub>Br is strongly influenced by CH<sub>3</sub>Br emissions from the surface waters ( $\delta^{13}$ C values in water phase (summer): -23±3%). Even during periods of low tide the 16 water remains in the deep channels which may be sufficient to have an impact on the local 17 18 atmosphere. Thus, despite the sources in the lagoon presumably producing isotopically light CH<sub>3</sub>Br,  $\delta^{13}$ C values in the atmosphere strongly reflect decomposed CH<sub>3</sub>Br whose residual 19 fraction is actually enriched in <sup>13</sup>C. Accordingly, aqueous CH<sub>3</sub>Br appears to become rapidly 20 21 degraded by biotic/abiotic processes such as hydrolysis, transhalogenation, and microbial 22 degradation with strong isotopic fractionation (King and Saltzman, 1997; Miller et al., 2004). 23 These decomposition mechanisms are temperature dependent with increasing destruction with increasing seawater temperature (King and Saltzman, 1997). This is most likely the reason 24 why the  $\delta^{13}$ C values in the lagoon waters in summer are more enriched in  $^{13}$ C compared with 25 those those from the spring campaign. 26

To the best of our knowledge, this is the first report of  $\delta^{13}$ C values of CH<sub>3</sub>I in the water phase. As shown by the water samples from the transect cruise, the sources in the lagoon may produce isotopic light CH<sub>3</sub>I. Given this, CH<sub>3</sub>I seems to some extent to follow the  $\delta^{13}$ C values of CH<sub>3</sub>Cl. These sources may be biotic by e.g. phytoplankton, seagrass meadows, or bacteria. On the other hand, Moore and Zafirou (1994) reported a photochemical source for CH<sub>3</sub>I by radical recombination of iodine with seawater dissolved organic matter. Due to the lack of isotopic source signatures and fractionation factors for production (and consumption), it is
 difficult to draw conclusions from the data.

The  $\delta^{13}$ C values of CHBr<sub>3</sub> were more depleted in  $^{13}$ C from the lagoon inlet towards the parts 3 deeper inside. This suggests a different combination of sources in water masses coming from 4 5 the Atlantic. Moreover, this potential variation of source contribution can be further assumed 6 by the certain change between summer and spring where e.g. macroalgae are more abundant in the latter period (Anibal et al., 2007). Already reported source signatures of phytoplankton, 7 8 macroalgae, and seagrass meadows cover the range of -10% to -23% (Auer et al., 2006; 9 Weinberg et al., 2013), thus demonstrating differences in their isotopic fingerprint. We cannot exclude that degradation might also have an effect on the  $\delta^{13}$ C values determined in lagoon 10 waters. As for CH<sub>3</sub>I, there is still need for further research on the CHBr<sub>3</sub> cycling utilizing 11 12 stable carbon isotopes.

# 4.4 Magnitude of fluxes and comparison to other coastal measurements and first estimate of global source strength

15 The areal based fluxes of CH<sub>3</sub>Cl, CH<sub>3</sub>Br, and CH<sub>3</sub>I from seagrass meadows in comparison to emission data of other coastal sources are presented in Fig. 3. In comparison to the emissions 16 17 from a temperate seagrass meadow in late summer in Northern Germany (Weinberg et al., 18 2013), fluxes were elevated in the subtropical lagoon in summer during air exposure. This 19 was more pronounced for CH<sub>3</sub>Br (factor 33) than for CH<sub>3</sub>Cl (factor 2), CH<sub>3</sub>I (factor 2), and 20 CHBr<sub>3</sub> (factor 5). In contrast, fluxes from air-exposed seagrass meadows recorded during 21 spring are comparable to those determined in Northern Germany. Thus, the difference between fluxes from temperate and subtropical regions is less pronounced than reported for 22 salt marshes with emissions from subtropical regions exceeding those from temperate regions 23 by up to two orders of magnitude for CH<sub>3</sub>Cl and CH<sub>3</sub>Br (Blei et al. 2010; Cox et al., 2004; 24 25 Dimmer et al., 2001; Drewer et al., 2006; Manley et al., 2006; Rhew and Mazéas, 2010; Rhew et al., 2000, 2014; Valtanen et al., 2009). Beside this regional (climatic) difference several 26 27 authors attributed this to a highly species dependent emission potential.

Average emissions of CH<sub>3</sub>Cl from the air-exposed seagrass meadows in summer are in the same range than those determined in temperate salt marshes (Blei et al. 2010; Cox et al., 2004; Dimmer et al., 2001; Drewer et al., 2006; Valtanen et al., 2009). In contrast, subtropical counterparts of these macrophytes are distinctively stronger emitters of this compound by at least one order of magnitude (Manley et al., 2006; Rhew and Mazéas, 2010; Rhew et al.,
 2000, 2014). Greenhouse grown mangroves produce significantly more CH<sub>3</sub>Cl than seagrass
 meadows revealing a higher emission potential for these plants species on a per area basis
 (Manley et al., 2007).

5 Fluxes of CH<sub>3</sub>Br from subtropical seagrass meadows during air exposure exceed those of 6 temperate macroalgae from Mace Head, Ireland (Carpenter et al., 2000) and temperate salt 7 marshes (Blei et al. 2010; Cox et al., 2004; Dimmer et al., 2001; Drewer et al., 2006; Valtanen 8 et al., 2009). However, the CH<sub>3</sub>Br fluxes from seagrass meadows are distinctively lower than 9 those of subtropical salt marsh plants (Manley et al., 2006; Rhew and Mazéas, 2010; Rhew et 10 al., 2000). Mangroves seem to have a similar emission potential as seagrass meadows 11 (Manley et al., 2007).

For CH<sub>3</sub>I, seagrass meadows are a minor source in comparison to the high release of macroalgae in subtropical areas (Leedham et al. 2013). Except for salt marshes from Tasmania (Cox et al., 2004), plant-related communities such as mangroves (Manley et al., 2007) and salt marshes (Dimmer et al., 2001) are more pronounced emission sources of this compound. The same holds true for CHBr<sub>3</sub>, where macroalgae communities from temperate and subtropical/tropical regions dominate the emissions of polyhalomethanes on a per area basis (e.g. Carpenter et al., 2000; Gschwend et al., 1985; Leedham et al., 2013).

19 Many uncertainties arise from a limited number of emission data to estimate the global 20 relevance of seagrass meadows. Those may be high variation in space and time, high 21 heterogeneity of seagrass meadows, species dependent emission potential, and errors 22 regarding the global seagrass abundance. Therefore, the scale-up of our data gives only a first 23 rough approximation; it was undertaken as follows. Since we did not measure a full annual 24 cycle, we assumed that seagrass measurements during the summer campaign represent 25 emissions from the reproductive season (May - September). The remaining period of the year (October - April) was calculated with emission data from the spring campaign. The emission 26 27 data were weighted to tidal states using 8 hours and 16 hours per day as durations when 28 seagrass meadows are air-exposed or submerged, respectively. Due to the lack of flood tide 29 emission data in summer, we used those derived from the sea-air exchange. The resulting average annual emissions from seagrass meadows of 150 µmol m<sup>-2</sup> yr<sup>-1</sup> (CH<sub>3</sub>Cl), 18 µmol m<sup>-2</sup> 30 yr<sup>-1</sup> (CH<sub>3</sub>Br), 14  $\mu$ mol m<sup>-2</sup> yr<sup>-1</sup> (CH<sub>3</sub>I), and 25  $\mu$ mol m<sup>-2</sup> yr<sup>-1</sup> (CHBr<sub>3</sub>) were scaled-up with the 31

1 current estimates of a global seagrass area ranging from  $0.3 \times 10^{12}$  m<sup>2</sup> (Duarte et al., 2005) to 2  $0.6 \times 10^{12}$  m<sup>2</sup> (Charpy-Roubaud and Sournia, 1990).

The tentative estimate yields annual emissions of 2.3-4.5 Gg yr<sup>-1</sup> for CH<sub>3</sub>Cl, 0.5-1.0 Gg yr<sup>-1</sup> 3 for CH<sub>3</sub>Br, 0.6-1.2 Gg yr<sup>-1</sup> for CH<sub>3</sub>I, and 1.9-3.7 Gg yr<sup>-1</sup> for CHBr<sub>3</sub>. Based on the recent global 4 5 budget calculations (Xiao et al., 2010; Montzka and Reimann, 2011), these ranges are 6 equivalent to 0.06-0.11% and 0.45-0.89%, for CH<sub>3</sub>Cl and CH<sub>3</sub>Br, respectively. Seagrass 7 meadows would therefore cover a portion of 1.4-2.8% of the missing sources for CH<sub>3</sub>Br reported in the most recent WMO report (36.1 Gg yr<sup>-1</sup>; Montzka and Reimann, 2011). Given 8 9 the emissions from oceanic sources (e.g. Butler et al., 2007; Quack and Wallace, 2003 and references therein), CH<sub>3</sub>I and CHBr<sub>3</sub> emissions from seagrass meadows are rather 10 11 insignificant on a global scale.

12

#### 13 **5** Conclusions

14 Our data are the first to report detailed halocarbon fluxes from seagrass meadows. The fluxes of halocarbons were highly variable with increased fluxes when the seagrass meadows were 15 16 submerged, and distinct emission peaks when lagoon waters were just arriving or leaving the sampling site. For CH<sub>3</sub>Cl and CH<sub>3</sub>Br we observed a diurnal dependence on the fluxes with 17 18 increased emissions during midday/afternoon and deposition fluxes during periods of low 19 radiation. Generally, diurnal variations (during air exposure), atmospheric mixing ratios, and 20 emission rates of halocarbons were smaller in spring than in summer, suggesting a seasonal 21 dependence. Our results indicate that on a global scale, seagrass meadows are a minor source 22 of halocarbons, but that they will have an imprint on the local and regional budgets, 23 particularly on subtropical coastlines, where seagrass meadows belong to the most abundant ecosystems. 24

Our stable carbon isotope results suggest that CH<sub>3</sub>Cl originates predominantly from the water column and/or seagrass meadows, rather than from adjacent salt marshes or abiotic formation processes. Atmospheric and aqueous CH<sub>3</sub>Br in the lagoon was substantially enriched in <sup>13</sup>C pointing towards degradation processes and re-emission into the atmosphere.

Future studies should focus on halocarbon emissions from seagrass-based systems from different regions in order to refine the global relevance. Since the magnitudes of fluxes are often species-dependent, budget calculations would benefit from a more detailed investigation of fluxes from different seagrass species. More work is also required to identify other
elements of these ecosystems, such as the sediments, which are capable of acting as both a
sink and a source of halocarbons.

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Table 1: Summary of air mixing ratios and water concentrations of halocarbons in the Ria Formosa and at the background site (Praia de Faro) for the sampling campaigns in summer 2011 and spring 2012. Values are given as means (bold) and ranges (in parentheses). Samples from the Ria Formosa are data from the inlet of the flux chambers with a sampling height of 1 m above ground (summer: n=36; Praia de Faro: n=5; spring n=47). Given water concentrations refer to n=8 (summer) and n=10 (spring).

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	Air mixing ratio	Air mixing ratio	Water concentration
	Ria Formosa (ppt)	Praia de Faro (ppt)	Ria Formosa (pmol L <sup>-1</sup> )
summer 2011			
CH <sub>3</sub> Cl	<b>828</b> (503-1490)	<b>613</b> (498-685)	<b>220</b> (158-301)
CH <sub>3</sub> Br	<b>22</b> (8-118)	<b>13</b> (9-19)	<b>8</b> (5-11)
CH <sub>3</sub> I	<b>3</b> (2-11)	1 (0.8-2)	<b>12</b> (4-18)
CHBr <sub>3</sub>	<b>15</b> (6-31)	<b>8</b> (6-9)	<b>102</b> (67 -194)
spring 2012			
CH <sub>3</sub> Cl	<b>654</b> (484-976)	-	<b>166</b> (101-267)
CH <sub>3</sub> Br	<b>12</b> (4-40)	-	10 (6-28)
CH <sub>3</sub> I	1 (0.4-4.8)	-	7 (2-16)
CHBr <sub>3</sub>	<b>2</b> (0.4-10)	-	<b>62</b> (39 - 133)

8

1 Table 2: Water concentration (pmol L<sup>-1</sup>) and stable carbon isotope ratios of halocarbons (‰)

2 obtained from a two-hours transect cruise on 24<sup>th</sup> April 2012 (see Fig. 1 for sampling

3 positions).

Sample	Time	CH <sub>3</sub> Cl		CH <sub>3</sub> Br		CH <sub>3</sub>	I	CHBr <sub>3</sub>	
	(CET)	pmol L <sup>-1</sup>	‰						
1	15:09	121	-40.9	5	-25.6	5	-20.0	26	-25.8
2	15:50	241	-42.3	7	-21.2	5	-31.1	55	-18.3
3	15:58	96	-	9	-	2	-	21	-
4	16:10	106	-	11	-	5	-	31	-
5	16:21	180	-44.3	19	-35.9	14	-44.5	95	-18.9
6	16:46	72	-	5	-	3	-	18	-
7	16:50	82	-	4	-	5	-	14	

4

Table 3: Mean net fluxes (bold) and ranges (parentheses) of halocarbons from flux chamber experiments seagrass meadows and sediments as well as those from sea-air exchange calculations. Data were obtained during the summer 2011 and spring 2012 campaigns in the Ria Formosa.

	n	CH <sub>3</sub> Cl	CH <sub>3</sub> Br	CH <sub>3</sub> I	CHBr <sub>3</sub>
		nmol m <sup>-2</sup> h <sup>-1</sup>			
Summer 2011					
air exposure	28	<b>15.6</b> (-49 - 74)	<b>6.5</b> (-5.7 - 130)	<b>1.2</b> (0.5 - 2.8)	<b>1.8</b> (-0.6 - 5.7)
air exposure (sediment)	5	<b>3.6</b> (-1.9 - 8.1)	<b>0.6</b> (-0.2 - 1.1)	<b>0.3</b> (0.1 - 0.6)	<b>0.8</b> (-0.3 - 1.9)
Sea-air exchange	8	<b>29.8</b> (13 - 45)	<b>1.3</b> (0.6 - 1.7)	<b>2.2</b> (0.5 - 3.2)	<b>4.7</b> (1.0 - 8.0)
Spring 2012					
air exposure	17	<b>1.0</b> (-30 - 69)	<b>0.4</b> (-0.8 - 3.9)	<b>0.6</b> (-0.6 - 2.6)	<b>0.4</b> (-0.5 - 1.3)
tidal inundation	18	<b>16.6</b> (-58 - 100)	<b>1.8</b> (-1.6 - 8.3)	<b>1.9</b> (0.1 - 8.0)	<b>3.0</b> (-0.4 - 11)
tidal change	5	<b>40.1</b> (-14- 100)	<b>2.7</b> (0.1 - 8.3)	<b>3.3</b> (0.1 - 8.0)	<b>2.9</b> (0.2 - 11)
incoming tide	6	<b>11.4</b> (-15 - 37)	<b>1.8</b> (0.2 - 3.3)	<b>1.6</b> (0.1 - 2.9)	<b>2.8</b> (0.2 - 5.1)
tidal maximum	2	-18, -58	-0.5, -1.6	0.1, 0.1	0.5, -0.1
ebb flow	5	<b>21.3</b> (-14 - 46)	<b>2.1</b> (0.1 - 4.4)	<b>1.5</b> (0.2 - 3.0)	<b>4.5</b> (-0.4 - 8.6)
Sea-air exchange	10	<b>15.2</b> (3.5 - 32)	<b>1.4</b> (0.5 - 4.1)	<b>1.3</b> (0.3 - 3.7)	<b>8.3</b> (3.8 - 24)

Table 4: Compilation of stable carbon isotope values of halocarbons (%) from the two
sampling campaigns. Source signatures of seagrass meadows were calculated using a coupled
mass and isotope balance (Weinberg et al., 2013).

	Atmosphere Atmosphere			source signature				
	Ria Formosa (‰)	n	Praia de Faro (‰)	n	lagoon water (‰)	n	seagrass meadow (‰)	n
summer 201	1							
CH <sub>3</sub> Cl	$-42 \pm 2$	7	$-39 \pm 0.4$	5	$-43 \pm 3$	7	$-51 \pm 6$	5
CH <sub>3</sub> Br	$-29 \pm 5$	7	$-38 \pm 3$	5	$-23 \pm 3$	7	$-42 \pm 17$	4
CH <sub>3</sub> I	-	-	-	-	$-39 \pm 9$	7	-	-
CHBr <sub>3</sub>	-	-	-	-	$-13 \pm 1$	7	-	-
spring 2012								
CH <sub>3</sub> Cl	$-38 \pm 1$	3	-	-	$-42 \pm 1$	5	$-56 \pm 2$	3
CH <sub>3</sub> Br	$-23 \pm 10$	3	-	-	$-33 \pm 8$	5	-26; -33	2
CH <sub>3</sub> I	-	-	-	-	$-37 \pm 7$	5	-	
CHBr <sub>3</sub>	-	-	-	-	$-18 \pm 1$	5	-	

- 1 Table 5: Mean concentrations (bold) and ranges (parentheses) of dissolved halocarbons (pmol
- 2 L<sup>-1</sup>) from the subtropical lagoon Ria Formosa in summer 2011 (n=9) and spring 2012 (n=10)
- 3 in comparison to published data from coastal Atlantic waters.

location	CH <sub>3</sub> Cl	CH <sub>3</sub> Br	CH <sub>3</sub> I	CHBr <sub>3</sub>
Faro, Portugal (summer) <sup>1</sup>	<b>220</b> (158 - 301)	8 (5-11)	<b>12</b> (4 - 18)	<b>102</b> (67 -194)
Faro, Portugal (spring) <sup>1</sup>	<b>166</b> (101 - 267)	<b>10</b> (6 - 28)	7 (2 - 16)	<b>62</b> (39 - 133)
2.4				
East Atlantic <sup>2,#</sup>	-	-	-	<b>68.3</b> (36.6 - 102.0)
Roscoff, France <sup>3, #</sup>	-	-	<b>12.9</b> (9.0 - 31.8)	<b>217.4</b> (124.8 - 519.4)
Greenland, NW Atlantic <sup>4</sup>	104 - 260	-	0.2 - 16.1	-
Norfolk, UK <sup>5</sup>	-	<b>3.2</b> (1.7 - 8.7)	-	-
Menai Strait, UK <sup>6,#</sup>	-	-	<b>6.7</b> (0.0 - 80.0)	<b>214.2</b> (3.0 - 3588.4)
Mace Head, Ireland <sup>7, #</sup>	-	<b>3.7</b> (1.7 - 5.7)	<b>15.3</b> (10.9 - 19.2)	<b>388.0</b> (221.8 - 554.3)
West Atlantic <sup>8</sup>	<b>88.4</b> (61.5 - 179.0)	<b>1.9</b> (0.8-5)	-	-
North West Atlantic <sup>9</sup>	<b>71.0</b> (55.0 - 106.0)	-	-	-
Nova Scotia, Canada <sup>10</sup>	-	-	4 - 6	-
Gulf of Maine, UK <sup>11, #</sup>	-	-	8 -18	40 - 1240

<sup>1</sup> this study; <sup>2</sup> Carpenter et al. (2009); <sup>3</sup> Jones et al. (2009); <sup>4</sup> Tait et al. (1994); <sup>5</sup> Baker et al. (1999); <sup>6</sup> Bravo-Linares and Mudge (2009); <sup>7</sup> Carpenter et al. (2000); <sup>8</sup> Hu et al. (2010); <sup>9</sup> MacDonald and Moore (2007); <sup>10</sup> Moore and Groszko (1999); <sup>11</sup> Zhou et al. (2005); <sup>#</sup> macroalgae dominated

32

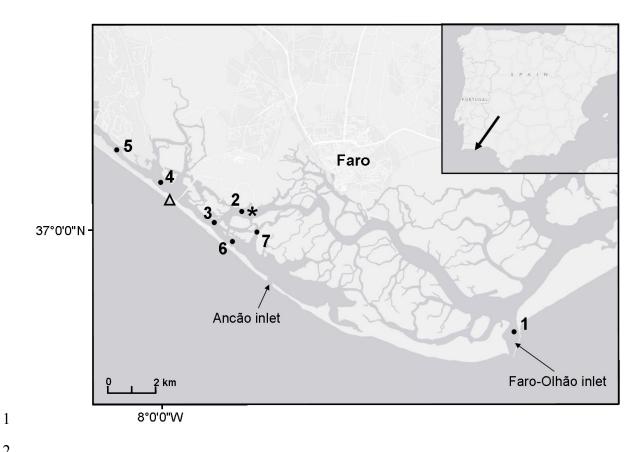




Fig. 1: Map of the lagoon Ria Formosa, Portugal. Asterisk: site of seagrass meadow studies; triangle: sampling site on the Praia de Faro (upwind position). Dots with numbers represent sampling points during the transect cruise. 

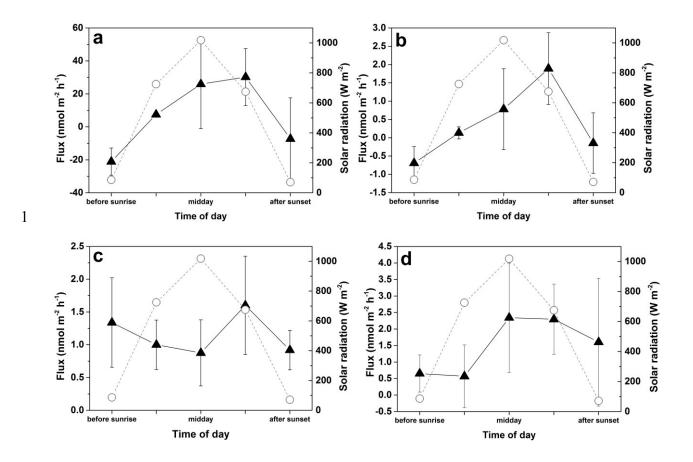
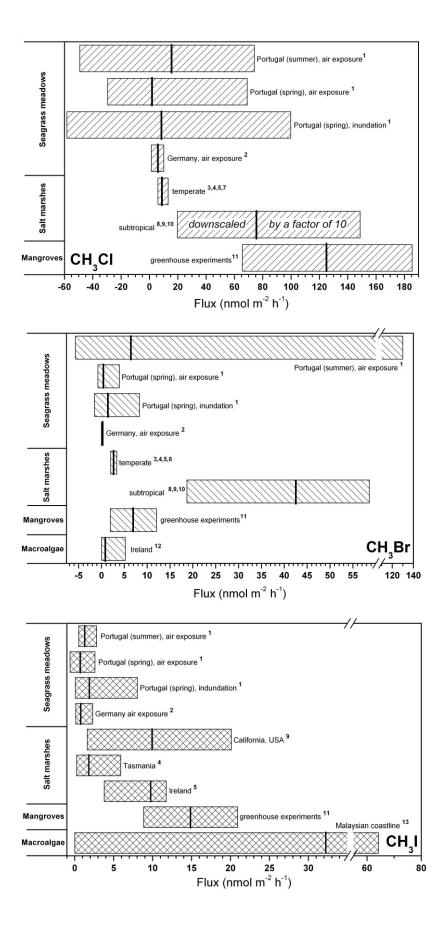


Fig. 2a-d: Diurnal variation of mean halocarbon fluxes (triangles) from seagrass meadows
during periods of air exposure in summer 2011 (a: CH<sub>3</sub>Cl, b: CH<sub>3</sub>Br, c: CH<sub>3</sub>I, d: CHBr<sub>3</sub>).
Error bars refer to standard deviations. Circles are solar radiation values. Note that the scales
on y-axis are different for each compound.





1 Fig. 3: Compilation of mean emissions (bold black vertical lines) and ranges from different sources in coastal environments for CH<sub>3</sub>Cl (upper panel). CH<sub>3</sub>Br (middle panel) and CH<sub>3</sub>I 2 (lower panel). Note the different scales. Published data adopted from: <sup>1</sup> this study; <sup>2</sup> Weinberg 3 et al. (2013); <sup>3</sup> Blei et al. (2010); <sup>4</sup> Cox et al., (2004); <sup>5</sup> Dimmer et al. (2001); <sup>6</sup> Drewer et al. 4 (2006); <sup>7</sup> Valtanen et al. (2009); <sup>8</sup> Rhew and Mazéas (2010); <sup>9</sup> Manley et al. (2006); <sup>10</sup> Rhew 5 et al. (2000); <sup>11</sup> Manley et al. (2007); <sup>12</sup> Carpenter et al. (2000); <sup>13</sup> Leedham et al. (2013). Note 6 that the data of CH<sub>3</sub>Cl from subtropical salt marshes are downscaled by a factor of 10 for 7 8 visualization reasons. Where multiple references were used, the individual study means were averaged and presented along with the resulting ranges. Thus, ranges of halocarbon fluxes in 9 each single study are not covered. Studies reporting a strong species dependency in magnitude 10 11 of fluxes were averaged over all species for simplicity reasons. Macroalgae emissions given in g fresh weight per hour were converted by using the species' fresh weights and spatial 12 13 coverage in the coastal belt in Mace Head, Ireland for CH<sub>3</sub>Br (Carpenter et al., 2000) and the Malaysian coastline for CH<sub>3</sub>I (Leedham et al., 2013), respectively. 14