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# Microbial respiration per unit microbial biomass depends on soil litter carbon-to-nitrogen ratio

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## Abstract

Soil microbial respiration is a central process in the terrestrial carbon (C) cycle. In this study I tested the effect of the carbon-to-nitrogen (C:N) ratio of soil litter layers on microbial respiration in absolute terms and per unit microbial biomass C. For this purpose, a global dataset on microbial respiration per unit microbial biomass C – termed the metabolic quotient ( $q\text{CO}_2$ ) – was compiled from literature data. It was found that the  $q\text{CO}_2$  in the soil litter layers was positively correlated with the litter C:N ratio and negatively related with the litter nitrogen (N) concentration. The positive relation between  $q\text{CO}_2$  and litter C:N ratio resulted from an increase in respiration with the C:N ratio in combination with no significant effect of the litter C:N ratio on the soil microbial biomass C concentration. The results suggest that soil microorganisms respire more C both in absolute terms and per unit microbial biomass C when decomposing N-poor substrate. Thus, the findings indicate that atmospheric N deposition, leading to decreased litter C:N ratios, might decrease microbial respiration in soils.

## 1 Introduction

Large amounts of organic carbon (C) are transformed, stored and respired by microorganisms in soil. Hence, gaining insight into the factors controlling the respiration rate per unit soil microbial biomass is crucial to understand the terrestrial C cycle. The respiration rate per unit microbial biomass C – termed the metabolic quotient ( $q\text{CO}_2$ ) – is as a measure for the ecophysiological status of soil microorganisms (Anderson and Domsch, 1993). Although a large number of studies on the  $q\text{CO}_2$  has been published (reviewed by Brookes, 1995; Bastida et al., 2008; Anderson and Domsch, 2010), little is known about how the  $q\text{CO}_2$  is affected by soil C:N:P stoichiometry.

The soil microbial biomass shows a relatively well constrained stoichiometry similarly to the Redfield ratio found for planktonic biomass (Redfield, 1934). Although the stoichiometry of individual phylogenetic groups may vary, the molar C:N ratio of the

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soil microbial biomass at a global scale converges towards 6–8 (Cleveland and Liptzin, 2007; Xu et al., 2013). The C : N ratio of soil litter layers is in the range of 12–80 (Berg and McLaugherty, 2003). Thus, microorganisms decomposing litter with a high C : N ratio are confronted with a surplus of C in relation to N. Compared to other ecosystems, microorganisms in forests face extreme substrate imbalances since the C : N ratios of woody plants are extremely high compared to the microbial biomass C : N ratio. While, for example, in phytoplankton and magroalgae the C : N ratio amounts to approximately 10, woody plants have a C : N ratio of up to 400 (Cebrian, 1999; Sterner and Elser, 2002).

When growing on N-poor substrate, microorganisms have not enough N to build up as much biomass as the C concentration would allow. Thus, it has been argued that microorganisms can dispose of C via overflow respiration as CO<sub>2</sub> to make the substrate meet their nutritional demands (Manzoni et al., 2008, 2010; Sinsabaugh et al., 2013). Overflow respiration is thought to be respiration without the production of energy. The concept of overflow respiration has recently been criticized by several studies. It has been argued, first, that for disposing C via the respiratory chain, N for the proteins of the respiratory chain has to be invested and, second, that the energy lost by disposing of C could be invested into storage, anti-viral defense or other processes, which increase the fitness of the organism (Hessen and Anderson, 2008; Hessen et al., 2013). Hence, while overflow respiration seems to be likely from a perspective of stoichiometric models, the existence of this process is still under discussion.

The objective of this study was to use data of published studies on the  $q\text{CO}_2$  in soil litter layers to learn about how litter C : N stoichiometry affects the respiration rate per unit decomposer biomass. Following stoichiometric theory, I tested the hypothesis that the  $q\text{CO}_2$  increases with litter C : N ratio and decreases with litter N concentration. For this purpose, data from literature on the  $q\text{CO}_2$  in soil litter layers and litter properties was compiled.

## 2 Material and methods

Literature searches were conducted using Google Scholar, Web of Science, and Scopus in November and December 2013. I searched for the word “metabolic quotient” in combination with the following terms “litter decomposition”, “litter layer”, “leaf decomposition”, “needle decomposition”, “microbial activity”, “forest floor”, “microbial respiration”, “tropical forest”, “temperate forest”, “boreal forest”, “mediterranean forest”, “plantation”.

Based on the literature search, I selected studies that reported the  $q\text{CO}_2$  measured in laboratory incubations on litter collected from the soil litter layer of forests, tree and palm plantations and heathlands. Studies that mixed litter with mineral soil were excluded because it is assumed that stabilization of the soil organic matter by sorption and aggregation possibly obscures relations between element concentrations and the  $q\text{CO}_2$ . If results for different treatments were reported, only the data for the control treatment were extracted. If time series were reported, I only extracted the first data point of the series in order to avoid pseudo-replication. In order to prevent confounding results due to different methods, the following criteria were applied for data selection. The  $q\text{CO}_2$  had to be reported in unambiguous units as the rate of C mineralization per unit of microbial biomass C. Basal respiration had to be determined during incubations based on  $\text{CO}_2$  measurements by gas chromatography or titration (but not, for example,  $\text{O}_2$  consumption), and the microbial biomass C had to be determined by the fumigation-extraction method. Additionally, the studies had to report either the C:N ratio of the litter or both the C and N concentration. Besides the metabolic quotient, microbial biomass C, basal respiration, and the C:N ratio of the litter, the following parameters were collected if reported in the studies: latitude and mean annual temperature of the study site, classification of the litter layer, litter pH, plant species from which the litter was derived, microbial biomass N, litter P, microbial biomass P, and temperature and water holding capacity at which the respiration measurement had been

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performed. In case data was reported in the form of graphs, numbers were extracted using the open-source software DataThief (Tummers, 2006).

Units were converted to obtain microbial biomass C in mg (g litter)<sup>-1</sup>, basal respiration in μg CO<sub>2</sub>-C (g litter-C)<sup>-1</sup> h<sup>-1</sup>, *q*CO<sub>2</sub> in μg CO<sub>2</sub>-C (mg microbial-C)<sup>-1</sup> h<sup>-1</sup>, and the C : N ratio in mol mol<sup>-1</sup>. For all analyses including latitude, only the degree of latitude was considered, but no differentiation between Southern and Northern Hemisphere was made. The Pearson's correlation coefficients were calculated, and the significance of the correlation was tested by the Pearson test. All data analysis was conducted in R (R Core Team, 2013).

### 3 Results

Fourteen studies were found that met the above-mentioned criteria, resulting in 48 observations. The studies covered the tropical, temperate, and boreal climate zone, and included data on the *q*CO<sub>2</sub> measured on litter derived from seven tree genera. Additionally, two studies reported data on litter of mixed forests with non-characterized species composition, and two studies reported results on litter derived from a palm and legumes and a forb (Table 1).

The *q*CO<sub>2</sub> was positively related to the C : N ratio of the litter (slope = 0.14, *R* = 0.78, *p* < 0.001, Fig. 1) and negatively to the litter N concentration (slope = 0.30, *R* = -0.72, *p* < 0.001, Fig. 2). The positive relation between litter C : N ratio and *q*CO<sub>2</sub> resulted from a positive relation between respiration and the C : N ratio (slope = 1.47, *R* = 0.71, *p* < 0.001, Fig. 3), and no effect of the litter C : N ratio on the microbial biomass C concentration (*R* = 0.16, *p* > 0.05, Table 2). The incubation temperatures, at which the respiration rates had been determined, ranged from 14 to 25 °C. Some of the variation in the *q*CO<sub>2</sub> was due to the different incubation temperatures and the positive correlation between incubation temperature and *q*CO<sub>2</sub> (slope = 0.25, *R* = 0.55, *p* < 0.001, Table 2). Moreover, the latitude was negatively related with the litter N concentration (*R* = -0.51, *p* < 0.001, Table 2). Other statistically significant correlations, such as be-

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tween respiration rate and  $q\text{CO}_2$ , and N concentration and C : N ratio (Table 2), are due to the intrinsic dependence of the variables. No significant relation between the litter C : N ratio and the microbial C : N ratio was found ( $R = 0.11$ ,  $p > 0.05$ , Table 2). Unfortunately, only very few studies reported litter P or microbial P concentrations, making the inclusion of these parameters into the analysis impossible.

## 4 Discussion

Here it was found that soil microbial respiration both in absolute terms and per unit microbial biomass is positively correlated with the soil litter C : N ratio. The findings are in accordance with previous studies that reported a positive correlation between litter C : N ratio and respiration (Othonen, 1994; Gødde et al., 2002; Michel and Matzner, 2002), and a negative relation between respiration and available N (Craine et al., 2007). The findings also agree with results from litterbag studies on litter decomposition in relation to litter C : N ratio (Berg and Matzner, 1997; Berg and McLaugherty, 2003). Several explanations for this negative relationship between respiration and C : N ratio have been proposed. A first explanation might be that microorganisms mine litter for N, i.e., they burn readily available C in order to gain energy to acquire N from more recalcitrant forms of organic matter (Craine et al., 2007) or in order to have physical access to the N incorporated in organic compounds. A second explanation is based on stoichiometry theory. It states that excess C is burned through “overflow respiration”, which means that microorganisms uncouple respiration from energy production and only respire C to dispose it (Russel and Cook et al., 1995; Manzoni et al., 2008, 2010). However, this argument has been criticized for two reasons (Hessen and Anderson, 2008). First, microorganisms may use C that is in surplus to their demands of somatic growth for promoting their fitness by C storage, buildup of structural defenses, viral repellents or establishment of symbiosis. Second, it seems that the disposal of C via respiration may need nutrients to maintain the proteins of the respiratory chain (Hessen and Anderson, 2008). A third explanation for decreased respiration at low litter C : N

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ratios could be that the activity of oxidative enzymes involved in the degradation of aromatic compounds decreases with N concentration (Carreiro et al., 2000; Saya-Cork et al., 2002; Michel and Matzner, 2003; Gallo et al., 2004). Decreased lignolytic activity might decrease microbial respiration in litter with low C : N ratios (Carreiro et al., 2000; Eiland et al., 2001; Saya-Cork et al., 2002).

One further way in which microorganisms can react to imbalanced substrate stoichiometry, is to gradually adapt the microbial biomass stoichiometry as recently shown for microorganisms in tropical litter (Fanin et al., 2013). However, in this study, I did not find a significant relation between the litter C : N ratio and the microbial C : N ratio, indicating that the microbial community did not adapt its biomass composition to the litter stoichiometry.

The positive relation between  $q\text{CO}_2$  and litter C : N ratio resulted from an increase in respiration with the C : N ratio in combination with no significant effect of the litter C : N ratio on the soil microbial biomass C concentration. The findings of this study indicate that atmospheric N deposition, leading to decreased litter C : N ratios, might decrease microbial respiration in soil litter layers both in absolute terms and per unit microbial biomass. This is in accordance with studies reporting that long-term N deposition and fertilization, resulting in decreases in plant litter C : N ratios, increased soil C sequestration in forests (Magnani et al., 2007; Pregitzer et al., 2008; Janssens et al., 2010). Pregitzer et al. (2008) and Janssens et al. (2010) found that the major reason for the positive effect of N deposition on C sequestration is reduced respiration with decreasing soil C : N ratio. This study suggests that this reduction in respiration rates is not due to a lower microbial biomass concentration, but due to a reduced respiration rate per unit microbial biomass.

## 5 Conclusions

This analysis of literature data shows that microbial respiration per unit microbial biomass in litter layers increases with the litter C : N ratio, highlighting the importance

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of soil stoichiometry for microbial mineralization processes. The findings indicate that atmospheric N deposition, leading to decreased litter C : N ratios, might decrease microbial respiration in soils.

**The Supplement related to this article is available online at  
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## References

- Anderson, T. H. and Domsch, K. H.: The metabolic quotient for CO<sub>2</sub> ( $q\text{CO}_2$ ) as a specific activity parameter to assess the effects of environmental conditions, such as pH, on the microbial biomass of forest soils, *Soil Biol. Biochem.*, 25, 393–395, 1993.
- Anderson, T. H. and Domsch, K. H.: Soil microbial biomass: the eco-physiological approach, *Soil Biol. Biochem.*, 42, 2039–2043, 2010.
- Bastida, F., Zsolnay, A., Hernández, T., and García, C.: Past, present and future of soil quality indices: a biological perspective, *Geoderma*, 147, 159–171, 2008.
- Berg, B. and Matzner, E.: Effect of N deposition on decomposition of plant litter and soil organic matter in forest systems, *Environ. Rev.*, 5, 1–25, 1997.
- Berg, B. and McClaugherty, C.: *Plant Litter: Decomposition, Humus Formation, Carbon Sequestration*, 1st edn., Springer-Verlag, Berlin, Germany, 2003.
- Brookes, P. C.: The use of microbial parameters in monitoring soil pollution by heavy-metals, *Biol. Fert. Soils*, 19, 269–279, 1995.
- Carreiro, M. M., Sinsabaugh, R. L., Repert, D. A., and Parkhurst, D. F.: Microbial enzyme shifts explain litter decay responses to simulated nitrogen deposition, *Ecology*, 81, 2359–2365, 2002.
- Cebrian, J.: Patterns in the fate of production in plant communities, *Am. Nat.*, 154, 449–468, 1999.





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- Cleveland, C. C. and Liptzin, D.: C : N : P stoichiometry in soil: is there a “Redfield ratio” for the microbial biomass?, *Biogeochemistry*, 85, 235–252, 2007.
- Craine, J. M., Morrow, C., and Fierer, N.: Microbial nitrogen limitation increases decomposition, *Ecology*, 88, 2105–2113, 2007.
- 5 Eiland, F., Klamer, M., Lind, A. M., Leth, M., and Baath, E.: Influence of initial C/N ratio on chemical and microbial composition during long term composting of straw, *Microb. Ecol.*, 41, 272–280, 2001.
- Fanin, N., Fromin, N., Buatois, B., and Hättenschwiler, S.: An experimental test of the hypothesis of non-homeostatic consumer stoichiometry in a plant litter–microbe system, *Ecol. Lett.*, 16, 764–772, 2013.
- 10 Gallo, M., Amonette, R., Lauber, C., Sinsabaugh, R. L., and Zak, D. R.: Microbial community structure and oxidative enzyme activity in nitrogen-amended north temperate forest soils, *Microb. Ecol.*, 48, 218–229, 2004.
- Gödde, M., David, M. B., Christ, M. J., Kaupenjohann, M., and Vance, G. F.: Carbon mobilization from the forest floor under red spruce in the northeastern USA, *Soil Biol. Biochem.*, 28, 1181–1189, 1996.
- 15 Hessen, D. O. and Anderson, T. R.: Excess carbon in aquatic organisms and ecosystems: physiological, ecological, and evolutionary implications, *Limnol. Oceanogr.*, 53, 1685–1696, 2008.
- 20 Hessen, D. O., Elser, J. J., Sterner, R. W., and Urabe, J.: Ecological stoichiometry: an elementary approach using basic principles, *Limnol. Oceanogr.*, 58, 2219–2236, 2013.
- Janssens, I., Dieleman, W., Luysaert, S., Subke, J.-A., Reichstein, M., Ceulemans, R., Ciais, P., Dolman, A. J., Grace, J., Matteucci, G., Papale, D., Piao, L., Schulze, E. D., Tang, J., and Law, B. W.: Reduction of forest soil respiration in response to nitrogen deposition, *Nat. Geosci.*, 3, 315–322, 2010.
- 25 Magnani, F., Mencuccini, M., Borghetti, M., Berbigier, P., Berninger, F., Delzon, S., Grelle, A., Hari, P., Jarvis, P. G., Kolari, P., Kowalski, A. S., Lankreijer, H., Law, B. E., Lindroth, A., Loustau, A., Manca, G. M., Moncrieff, J. B., Rayment, M., Tedeschi, C., Valentini, R., and Grace, J.: The human footprint in the carbon cycle of temperate and boreal forests, *Nature*, 447, 849–851, 2007.
- 30 Manzoni, S., Jackson, R. B., Trofymow, J. A., and Porporato, A.: The global stoichiometry of litter nitrogen mineralization, *Science*, 321, 684–686, 2008.

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- Manzoni, S., Trofymow, J. A., Jackson, R. B., and Porporato, A.: Stoichiometric controls on carbon, nitrogen, and phosphorus dynamics in decomposing litter, *Ecol. Monogr.*, 80, 89–106, 2010.
- Michel, K. and Matzner, E.: Nitrogen content of forest floor Oa layers affects carbon pathways and nitrogen mineralization, *Soil Biol. Biochem.*, 34, 1807–1813, 2002.
- Michel, K. and Matzner, E.: Response of enzyme activities to nitrogen addition in forest floors of different C-to-N ratios, *Biol. Fert. Soils*, 38, 102–109, 2003.
- Ohtonen, R.: Accumulation of organic matter along a pollution gradient: application of Odum's theory of ecosystem energetics, *Microb. Ecol.*, 27, 43–55, 1994.
- Pregitzer, K. S., Burton, A. J., Zak, D. R., and Talhelm, A. F.: Simulated chronic nitrogen deposition increases carbon storage in Northern Temperate forests, *Glob. Change Biol.*, 14, 142–153, 2008.
- R Core Team: R: a Language and Environment for Statistical Computing, R Foundation for Statistical Computing, Vienna, Austria, 2013.
- Redfield, A. C.: On the proportions of organic derivations in sea water and their relation to the composition of plankton, in: James Johnstone Memorial Volume, edited by: Daniel, R. J., University Press of Liverpool, Liverpool, 177–192, 1934.
- Russell, J. B. and Cook, G. M.: Energetics of bacterial growth: balance of anabolic and catabolic reactions, *Microbiol. Rev.*, 59, 48–62, 1995.
- Saiya-Cork, K. R., Sinsabaugh, R. L., and Zak, D. R.: The effects of long term nitrogen deposition on extracellular enzyme activity in an *Acer saccharum* forest soil, *Soil Biol. Biochem.*, 34, 1309–1315, 2002.
- Sinsabaugh, R. L., Manzoni, S., Moorhead, D. L., and Richter, A.: Carbon use efficiency of microbial communities: stoichiometry, methodology and modelling, *Ecol. Lett.*, 16, 930–939, 2013.
- Sterner, R. W. and Elser, J. E.: *Ecological Stoichiometry: the Biology of Elements from Molecules to the Biosphere*, Princeton University Press, Princeton, 1–43, 2002.
- Tummers, B.: DataThief III, available at: <http://datathief.org/>, last access: 20 January 2014, 2006.
- Xu, X., Thornton, P. E., and Post, W. M.: A global analysis of soil microbial biomass carbon, nitrogen and phosphorus in terrestrial ecosystems, *Global Ecol. Biogeogr.*, 22, 737–749, 2013.

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**Table 1.** References considered in the analysis together with the latitude of the study site, the plant genus from which the litter was derived and the number of data points gained from each reference. A detailed list of the publications, from which data was extracted is given in the Supplement.

Reference	Latitude	Plant	Data points
Chang and Trofymow (1996)	50° N	<i>Cedrus</i>	3
Chapman et al. (2003)	57° N	<i>Pinus</i>	1
Dinesh et al. (2006)	10° S	<i>Cocos</i> and <i>Legumes</i>	10
Fisk and Fahey (2001)	44° N	<i>Fagus</i> and <i>Betula</i>	1
Karneva and Smolander (2007)	66° N	<i>Picea</i> , <i>Pinus</i> , <i>Betula</i>	8
van Meeteren et al. (2007)	52° N	Forbs	1
Ndaw et al. (2009)	21° S	Various broadleaf trees, <i>Eucalyptus</i>	4
Pietikainen and Fritze (1996)	65° N	<i>Picea</i>	3
Ross and Sparling (1993)	36° S	<i>Pinus</i>	4
Ross and Tate (1993)	36° S	<i>Fagus</i>	2
Ross et al. (1996)	43° S	<i>Fagus</i>	2
Ross et al. (1999a)	38° S	Various trees, <i>Pinus</i>	4
Ross et al. (1999b)	61° N, 42° S, 40° S, 36° S	<i>Pinus</i>	4
Schimel et al. (1999)	64° N	<i>Betula</i>	1

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**Table 2.** Spearman's correlation coefficient of the latitude of the study site, the  $\text{pH}_{\text{H}_2\text{O}}$  of the soil litter layer, the C and N concentration and the C : N ratio of the soil litter layer, the microbial biomass C and N concentration ( $C_{\text{mic}}$  and  $N_{\text{mic}}$ ), the microbial biomass C : N ratio, the incubation temperature at which the respiration rate was determined (Temp), the respiration rate (Resp), and the metabolic quotient ( $q\text{CO}_2$ ). <sup>a</sup>, <sup>b</sup>, <sup>c</sup> denote levels of significance at  $p < 0.05$ , 0.01 and 0.001.

	Latitude	$\text{pH}_{\text{H}_2\text{O}}$	C	N	C : N	$C_{\text{mic}}$	$N_{\text{mic}}$	$C_{\text{mic}} : N_{\text{mic}}$	Temp	Resp	$q\text{CO}_2$
Latitude											
$\text{pH}_{\text{H}_2\text{O}}$	-0.39 <sup>a</sup>										
C	0.52 <sup>c</sup>	-0.16									
N	-0.51 <sup>c</sup>	-0.14	0.00								
C : N	0.38 <sup>b</sup>	0.17	0.51 <sup>a</sup>	-0.81 <sup>c</sup>							
$C_{\text{mic}}$	0.22	-0.12	0.24	-0.01	0.16						
$N_{\text{mic}}$	-0.01	0.25	0.13	-0.20	0.22	0.08					
$C_{\text{mic}} : N_{\text{mic}}$	0.04	-0.07	0.18	0.00	0.11	0.54 <sup>c</sup>	-0.39 <sup>a</sup>				
Temp	-0.42 <sup>b</sup>	0.39 <sup>a</sup>	0.17	-0.38 <sup>a</sup>	0.30 <sup>a</sup>	-0.06	0.40 <sup>b</sup>	0.03			
Resp	0.17	0.19	0.35 <sup>a</sup>	-0.56 <sup>c</sup>	0.71 <sup>c</sup>	0.52 <sup>c</sup>	0.38 <sup>a</sup>	0.07	0.33 <sup>a</sup>		
$q\text{CO}_2$	0.13	0.36 <sup>a</sup>	0.26	-0.72 <sup>c</sup>	0.78 <sup>c</sup>	0.01	0.22	0.05	0.55 <sup>c</sup>	0.64 <sup>c</sup>	

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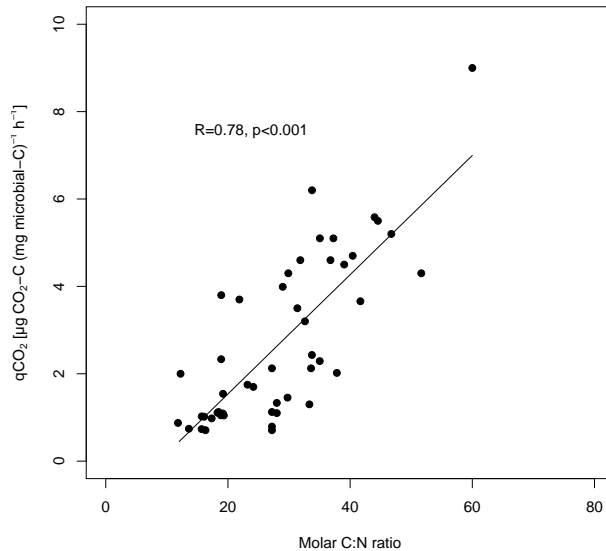
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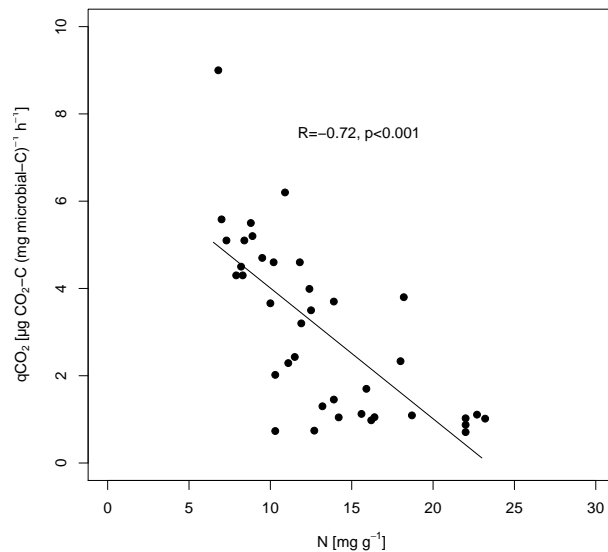
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**Figure 1.** Correlation between the metabolic quotient ( $q_{CO_2}$ ) and the molar carbon-to-nitrogen ratio (C : N) of the soil litter layer.

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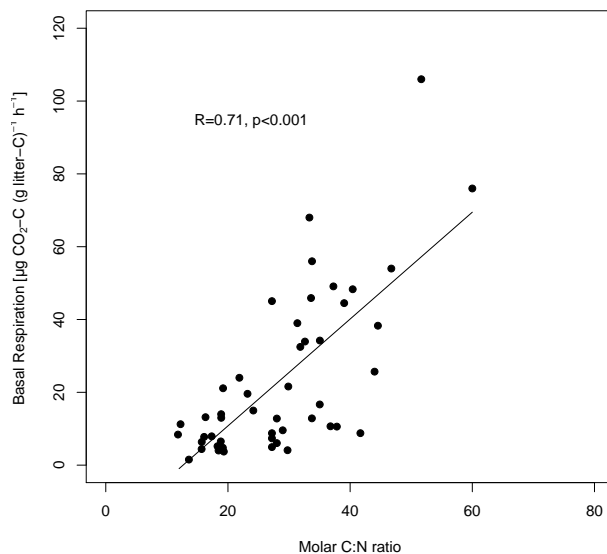


**Figure 2.** Correlation between the metabolic quotient ( $q_{CO_2}$ ) and the soil litter layer nitrogen (N) concentration.

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**Figure 3.** Correlation between the basal respiration rate and the molar carbon-to-nitrogen ratio (C : N) of the soil litter layer.

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