CO₂ and nutrient-driven changes across multiple levels of organization in *Zostera noltii* ecosystems

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9 Abstract

10 Increasing evidence emphasizes that the effects of human impacts on ecosystems must 11 be investigated using designs that incorporate the responses across levels of biological 12 organization as well as the effects of multiple stressors. Here we implemented a 13 mesocosm experiment to investigate how the individual and interactive effects of CO₂ 14 enrichment and eutrophication, scale-up from changes in primary producers at the 15 individual- (biochemistry) or population-level (production, reproduction, and/or 16 abundance) to higher levels of community (macroalgae abundance, herbivory, and 17 global metabolism) and ecosystem organization (detritus release and carbon sink 18 capacity). The responses of Zostera noltii seagrass meadows growing in low- and high-19 nutrient field conditions were compared. In both meadows, the expected CO₂ benefits 20 on Z. noltii leaf production were suppressed by epiphyte overgrowth, with no direct CO_2 21 effect on plant biochemistry or population-level traits. Multi-level meadow response to 22 nutrients was faster and stronger than to CO₂. Nutrient enrichment promoted the 23 nutritional quality of Z. noltii (high N, low C:N and phenolics), the growth of epiphytic 24 pennate diatoms and purple bacteria, and shoot mortality. In the low-nutrient meadow, 25 individual effects of CO₂ and nutrients separately resulted in reduced carbon storage in 26 the sediment, probably due enhanced microbial degradation of more labile organic 27 matter. These changes, however, had no effect on herbivory nor on community 28 metabolism. Interestingly, individual effects of CO₂ or nutrient addition on epiphytes, 29 shoot mortality, and carbon storage were attenuated when both nutrients and CO₂ acted 30 simultaneously. This suggests CO₂-induced benefits on eutrophic meadows. In the high-31 nutrient meadow, a striking shoot decline caused by amphipod overgrazing masked the

response to CO_2 and nutrient additions. Our results reveal that under future scenarios of CO₂, the responses of seagrass ecosystems will be complex and context dependent, being mediated by epiphyte overgrowth rather than by direct effects on plant biochemistry. Overall, we found that the responses of seagrass meadows to individual and interactive effects of CO_2 and nutrient enrichment varied depending on interactions among species and connections between organization levels.

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39 **1** Introduction

40 Understanding community and ecosystem responses to human impacts is a challenge 41 that requires integrating not only the organism-level responses across populations and 42 entire systems (Russell et al., 2012), but also synergistic or antagonistic effects of 43 multiple stressors (Woodward et al., 2010). A large number of articles has been 44 published on the effects of ocean acidification, and reviewed among others by Doney et 45 al. (2009) and Kroeker et al. (2010). This body of research has revealed that ocean acidification can be detrimental to most marine calcifying organisms, while increasing 46 47 carbon dioxide (CO₂) concentration can benefit primary productivity of phytoplankton, cyanobacteria, fleshy algae, and seagrasses. Our current understanding of these effects 48 49 is largely based on the species-specific responses of individuals or populations. 50 However, the broad variability in responses among organisms may influence species 51 interactions and drive unforeseen impacts on marine communities and ecosystems 52 (Hall-Spencer et al., 2008; Kroeker et al., 2013a).

53 The interactive effect of multiple stressors on ecological communities remains largely 54 unknown (Crain et al., 2008). Atmospheric CO₂ concentration has increased from 55 preindustrial levels of approximately 280 ppm to 397 ppm in 2013 (NOAA, Mauna Loa 56 Observatory, Hawaii), leading to a rise in the CO_2 absorbed by the ocean with an 57 associated pH decrease of 0.1 units. An additional pH decrease of 0.07-0.31 units is expected by the end of the 21th century based on the `Intergovernmental Panel on 58 59 Climate Change' predictions (IPCC, 2013). In the marine environment, ocean 60 acidification can locally interact with excess nutrients from coastal eutrophication to 61 accelerate changes in ecosystem structure and functioning (Russell et al., 2009).

62 Human impacts that alter the availability of environmental resources are shifting the 63 nutritional quality of primary producers through changes at the biochemical or 64 individual levels of the biological organization (e.g. allocation of resources to growth, 65 storage, and chemical defences). In addition, changes in environmental resources can favour different types of producers and alter the inter-species competitiveness and 66 producers' abundances (Kroeker et al., 2013b). Overall, this may affect ecological 67 68 interactions and fluxes, leading to shifts at community and ecosystem levels. Under 69 elevated nutrient concentrations, aquatic and terrestrial ecosystems with higher producer 70 nutritional quality often support higher rates of herbivory, more rapid decomposition 71 rates and recycling of nutrients, and lower net accumulation of soil carbon (Wardle et 72 al., 2004; Cebrian et al., 2009). The effects of nutrient enrichment have been widely 73 described on terrestrial, freshwater, and marine ecosystems, whereas the scaling up of 74 elevated CO₂ effects has been mostly studied in terrestrial plants. Under elevated CO₂ 75 levels, and especially if nutrient availability is limiting to growth, terrestrial plants 76 typically increase the accumulation of carbohydrates and/or carbon-based secondary 77 compounds (mostly phenolics). This increases C:N ratios ("nitrogen dilution" effect) 78 and sometimes leaf toughness through increasing indigestible polymers such as 79 cellulose and lignin (Zvereva and Kozlov 2006; Lindroth, 2010; Robinson et al., 2012). 80 Herbivores usually compensate for this lower food quality by eating more (Stiling and 81 Cornelissen, 2007). In addition, CO_2 enrichment may shift the biomass and composition 82 of soil microbial communities, directly through different responses of microbial groups 83 to high CO₂ / low pH (Krause et al., 2012; Lidbury et al., 2012) or indirectly through 84 reducing foliar and detritus quality (Drigo et al., 2007). Overall, high CO₂ levels may 85 have both positive and negative consequences on the decomposition of soil organic 86 matter and nutrient recycling (Lindroth, 2010).

87 In this study, we use seagrass meadows as model ecosystems to investigate the scaling 88 up of the effects of elevated CO_2 and nutrient levels on marine coastal environments. 89 Seagrass beds are widely distributed habitats that host high biodiversity and provide 90 valuable ecosystem services (Orth et al., 2006). A rich epiphyte community usually 91 colonizes seagrass leaves, thus providing a useful system for studying how changes in 92 environmental resources can favour different types of producers (e.g. non-calcareous vs. 93 calcareous). They are highly productive systems that sequester larger amounts of carbon 94 per area than tropical forests, providing for a long-term removal of carbon dioxide from 95 the atmosphere (Pidgeon, 2009; Fourqurean et al., 2012). The maintenance of the key 96 services provided by seagrass ecosystems under global change is thus of prime97 importance for human well-being.

98 A mesocosm experiment was conducted to assess: (1) how CO_2 and nutrient 99 enrichments affect primary producers' at the individual- (plant biochemistry including 100 CN and allocation of resources to carbohydrate reserves and carbon-based chemical 101 defences) or population-level (plant allocation of resources to biomass and 102 reproduction, and composition and abundance of seagrass epiphytes), and (2) whether 103 these changes propagate to the community (macroalgae abundance, meso-herbivory, 104 whole-community metabolism) and to the ecosystem (detritus production and organic 105 carbon storage in sediment). The responses of meadows of the seagrass Zostera noltii 106 Horneman developing in low- and high- nutrient conditions in the field were compared 107 to assess if they react differently.

108

109 2 Methods

110 2.1 Study meadows

111 Samples of Z. noltii community were collected from two meadows separated 5.5 km 112 from each other within the Ria Formosa lagoon (South Portugal). This shallow 113 mesotidal lagoon is dominated by monospecific beds of the seagrass Z. noltii that 114 occupy ca. 45% of the intertidal area. One meadow was developing under prior field conditions of low nutrient levels (36°59'40''N 7°58'00''W; hereafter low-nutrient 115 116 meadow) and the other under high nutrient levels (37°01'15''N 8°00'56.50''W; 117 hereafter high-nutrient meadow). Low-intertidal samples exposed to a small emersion 118 period only during low spring tides were selected. Table 1 presents seawater nutrient 119 concentrations and seagrass meadow traits that reveal the substantial initial differences 120 between meadows.

121 2.2 Mesocosm experiment

The study was conducted in an outdoor mesocosm system at the Ramalhete field station of the Centre of Marine Sciences, which is located at the Ria Formosa lagoon. To assess the effects of CO_2 and nutrients on *Z. noltii* meadows an enrichment experiment was conducted for 6 weeks during August-September 2011, after 4 days of acclimation to 126 the experimental mesocosms. This time span is sufficient to detect any treatment-driven 127 changes in physiological, morphological and population traits of this fast-growing 128 species (e.g. Peralta et al., 2002). Core samples of Z. noltii community, including 129 sediment and algal, faunal and microbial components, were randomly collected from 130 each donor meadow and allocated to flowerpots of 20 cm of diameter and height. Three 131 flowerpots were placed in each of the 16 experimental mesocosms (tanks of 110 L), 132 which were exposed to combinations of two CO₂ and two nutrient levels in a crossed 133 design with two replicates.

134 Experimental levels of CO_2 encompassed present (pH 8.00+0.02, equivalent to ca. 400 ppm CO₂) and future conditions (pH 7.83+0.01, equivalent to ca. 800 ppm CO₂) in Ria 135 136 Formosa lagoon. The mesocosms received sand-filtered seawater from two head tanks of 1000 L at a rate of 240 L h^{-1} . In one of the head tanks, the water CO₂ was 137 manipulated to ensure fixed pH differences between treatment means within the range 138 139 predicted for 2100 by the IPCC (pH decline = -0.18 units) following a commonly used 140 method (e.g. Alsterberg et al., 2013). The CO₂ injection was controlled by an auto-141 analyzer (Yokogawa, EXAxt 450, Tokyo, Japan), which continuously monitored the water pH and temperature. Total alkalinity, pH, temperature and salinity within the 142 143 mesocosms, as well as the seawater DIC and carbon speciation are provided on 144 Appendix B (Supplement).

Water nutrient levels encompassed the natural values found in the lagoon and the values of highly eutrophic conditions (N: 45x and P: 11x natural, see Appendix B, Supplement). The nutrient enrichment was obtained by adding a solubilised mixture of the fertilizers ammonium nitrate and monoammonium phosphate directly into the water column of each enriched mesocosm using a multi-channel dosing pump. Water samples were collected weekly to analyze nutrient concentrations using a loop-flow analyzer $(\mu$ Mac-1000; Systea, Anagni, Italy).

The water within mesocosms was homogenized using a submersible circulation pump placed at leaf height. Pumps were stopped 2 h twice per day to simulate tidal currents. Twice a week, the epiphytes growing in the mesocosm walls were removed and the position of flowerpots within each mesocosm was reassigned to minimize potential spatial differences. Natural settlement and growth of leaf epiphytes and small animals were allowed throughout the experiment.

158 **2.3 Producers' traits at the individual- or population-level**

159 Changes on producers' at the individual or population levels were assessed by 160 measuring: (1) plant biochemistry and allocation of resources to biomass and 161 reproduction; and (2) the composition and abundance of seagrass epiphytes.

162 Allocation of plant resources to biomass and reproduction at the population-level was 163 monitored almost every week. Shoot recruitment or mortality were quantified within 164 each flowerpot excluding shoots growing around the border to avoid edge effects. 165 Allocation to reproduction was quantified as density of flowering shoots. Five shoots at 166 the beginning and at the end of the experiment and three shoots in between sampling 167 events were randomly chosen within each flowerpot to quantify the number of leaves 168 and the leaf area index (LAI) as indicators of aboveground productivity. To estimate 169 LAI, leaf area was measured on these shoots, averaged, multiplied by the number of 170 shoots within the pot, and scaled per surface area.

After four weeks of experiment, leaf epiphyte composition was determined in the oldest
leaves of three randomly chosen shoots. The surface covered by each taxon was
quantified under a microscope and standardized per 10 cm² of leaf area.

174 After six weeks, all shoots from each mesocosm were harvested and plant traits 175 quantified in each flowerpot. Belowground productivity was estimated from the vertical 176 or horizontal rhizome length and from the total root length (number of roots multiplied 177 by the average root length) of five shoots per pot. The above- and below-ground 178 biomass allocation was quantified after drying at 60 °C until constant weight. The 179 pooled epiphyte load of three shoots was removed using a glass slide and quantified as 180 relative to leaf area after drying at 60 °C until constant weight. Pooled material of five 181 shoots was separated into leaves (without epiphytes) and rhizomes, freeze-dried, 182 weighted, ground to fine powder and used in subsequent analyses of plant biochemistry. 183 Carbon and nitrogen concentrations were analyzed using an elemental analyzer (Carlo-184 Erba, Milan, Italy). Total non-structural carbohydrates were measured in rhizomes using 185 the phenol-sulfuric acid colorimetric method (Dubois et al., 1956) with glucose as 186 standard, after sugar extraction in hot ethanol and enzymatic conversion of starch to 187 glucose equivalents (Smith and Zeeman, 2006). Total phenolics were quantified as 188 indicators of plant allocation of resources to chemical defences. Phenolics were 189 extracted from leaf material with methanol 50% for 24 h under constant agitation at 4

°C and determined with a spectrophotometer using chlorogenic acid as standard
following a modified Folin-Ciocalteu method (Bolser et al., 1998).

192 2.4 Community- and ecosystem-level traits

193 The response of the seagrass community to CO₂ and nutrient enrichments was 194 quantified weekly by: (1) the percentage of flowerpot surface covered by *Ulva* spp.; (2) 195 the feeding activity of mesograzers (percentage of leaves showing bite marks in the 196 same shoots used to measure the morphological traits); and (3) the whole-community 197 metabolism quantified from the oxygen evolution within benthic chambers of 17 cm 198 diameter (4.8+0.01 L incubated) fitted to the flowerpots for 30-45 minutes at midday 199 (12-14 h). A transparent acrylic chamber to estimate net production and a dark chamber 200 to estimate respiration were simultaneously deployed within each mesocosm. Dissolved 201 oxygen concentration was measured by spectrophotometry using the Winkler method 202 (Labasque et al., 2004) in three water samples collected before and after incubations 203 into 12 mL soda glass vials. Community metabolism was estimated from the net change 204 in oxygen concentration during incubations integrated by the chamber volume and 205 standardized by incubation time and bottom area. There were no effects of enclosure on 206 the water temperature within chambers (measured with onset HOBO loggers, Southern 207 MA, USA). All incubations were run under irradiances of photosynthetically active radiation (PAR) averaging 283+8.6 µmol quanta m⁻² s⁻¹ (measured with a Li-192SA 208 209 underwater PAR quantum sensor, Li-Cor, USA), when Z. noltii photosynthesis is light 210 saturated and not photoinhibited (Peralta et al., 2002).

At the ecosystem level, detritus production (fresh weight of all floating material collected during a 24h period) was quantified almost every week. Organic matter in the sediment (loss of dry weight after combustion at 450 °C, 4h) was measured at the end of the experiment as indicator of the carbon sink capacity.

215 2.5 Statistical analyses

The effects of CO_2 and nutrient treatments throughout the experiment were tested using three-way repeated-measures analyses of variance (RM ANOVA). The subject repeatedly sampled was the mesocosm, CO_2 and nutrients were the among-subject factors (two fixed crossed factors) and Time the within-subject factor. To avoid the masking effect of the strong initial differences between meadows on the responses to experimental treatments, data for the low- and high-nutrient meadows were analyzed separately. Data were checked for parametric assumptions and transformed where needed. When sphericity was not met, corrected degrees of freedom from Greenhouse-Geisser adjustment were used (Quinn and Keough, 2002).

225 The effects of CO₂ and nutrient treatments on variables measured at the end of the 226 experiment were tested using two-way ANOVAs (two fixed crossed factors) after 227 testing parametric assumptions. A normal distribution with unequal variances was found 228 for all variables, which is usual when the sample size is small. Following 229 recommendation by Quinn and Keough (2002), we proceeded with the analyses but 230 making significance level more restrictive to minimize the possibility of Type I error 231 (mistakenly detection of differences). Welch t tests that are robust against unequal 232 variances were used to interpret significant interactions. Again, data for the low- and 233 high-nutrient meadows were analyzed separately.

234 To assess the ordination of treatments based on differences in the composition of leaf 235 epiphyte assemblages, a non-metric Multi-Dimensional Scaling analysis (NMDS) with 236 Brav-Curtis distances was carried out. Because NMDS axes are arbitrary, the final 237 solution was rotated using a Principal Component Analysis (PCA) to align the largest 238 variance in the first axis. The significance of the effect of CO₂ and nutrient treatments 239 on assemblage composition was tested with a two-way permutational analysis of 240 variance (PERMANOVA; two fixed crossed factors). To perform the test, Bray-Curtis 241 distances were calculated from untransformed data and 999 permutations were used 242 under a reduced model.

Finally, two PCAs, one for each meadow, were performed to assess links among the several traits and the trajectory of treatment responses through time. Traits showing the highest correlation with the components ($r \ge 0.7$) were selected for interpretations. Since our variables were not dimensionally homogeneous, principal components were computed from the correlation matrix.

249 **3 Results**

250 **3.1** Responses of meadow traits measured through time

251 The response of the low-nutrient meadow showed a threshold at the third week of the 252 experiment, when most variables responded differently from the first two weeks (Fig. 1, 253 left). Shoot recruitment occurred mostly in unfertilized but also in CO₂-enriched 254 conditions until the third week (Fig. 1a), after which shoot mortality progressively 255 increased. Figure 1d suggests that the Z. noltii leaf area index (LAI) tended to increase 256 with CO₂ enrichment until the third week of the experiment. A positive, significant 257 effect of the CO₂ enrichment was observed on detritus production throughout the 258 experiment (Fig. 1c). Nutrient addition increased shoot mortality (Fig. 1a), whereas it 259 decreased leaf number and LAI (Fig. 1b and d). Shoot mortality induced by the nutrient 260 enrichment was attenuated by the simultaneous addition of CO₂, especially from the 261 third week onwards (Fig. 1a). A treatment and time interaction was detected on the 262 community production and respiration (Fig. 1e and f). These variables showed high 263 variability with similar ranges of variation in unfertilized and enriched conditions. No 264 treatment effects were detected throughout the experiment on shoot flowering or meso-265 herbivory (see Appendix C, Supplement).

266 The responses of the high-nutrient meadow to CO₂ enrichment included an increased 267 shoot mortality during the second and third weeks and an increased detritus production 268 at the end of the experiment (Fig. 1g and i). Nutrient addition decreased the number of 269 leaves per shoot and increased detritus production throughout the experiment (Fig. 1h 270 and i). CO₂ enrichment interacted with nutrients to alleviate the nutrient-induced 271 reduction of the number of leaves (Fig. 1h). No effects of CO₂ or nutrient enrichment 272 were observed through time on LAI, meso-herbivory, Ulva spp. cover, shoot flowering, 273 and community production or respiration (Figs. 1j-l and C, Supplement). Independently 274 of the experimental treatments, overgrazing by the herbivorous amphipod Cymadusa 275 filosa Savigny severely affected the plants from the high-nutrient meadow causing 276 massive shoot mortality (Fig. 1k and g). At the end of the experiment a mean (+se) of 277 89 (+3.7)% of shoots died, 81 (+9.1)% of the seagrass leaves showed bite marks and leaf area was reduced from 5.0 (+0.2) to 1.0 (+0.4) cm² shoot⁻¹. Similarly, *Ulva* spp. 278 279 cover progressively decreased, being close to 0% in all treatments after six weeks (Fig. 280 11). At the end of the experiment, all Z. noltii shoots and Ulva spp. fronds disappeared

from three flowerpots (one unfertilized and two CO₂-and-nutrient-enriched). However, net production and respiration in these pots were within the range observed in the other pots (Fig. Cd and e, Supplement), indicating that the metabolism of the sediment microbial community was similar to that of the *Z. noltii* community.

285 We did not detect any significant effect of CO₂ or nutrient enrichment on plant damage 286 by meso-herbivory in the low- or high-nutrient meadows. However, at the end of the 287 experiment plants from the high-nutrient meadow showed 81 (+9.1)% of leaves with 288 bite marks, compared to only 6.9 (+3.2)% in the low-nutrient meadow (Figs. 1k and Cb, 289 Supplement). These between meadow differences, as well as the link between shoot 290 mortality and meso-herbivory in masking the enrichment effects in the high-nutrient 291 meadow, were further confirmed by a PCA of responsive variables from the low- and 292 high-nutrient meadows at the end of the experiment (see Appendix E, Supplement).

293 **3.2** Responses of meadow traits measured at the end of the experiment

294 In plants from the low-nutrient meadow (Fig. 2, grey bars), nutrient enrichment 295 enhanced the leaf nutritional quality (high leaf nitrogen and low leaf C:N ratio, Fig. 2a 296 and c) and the accumulation of nitrogen in rhizomes (high rhizome nitrogen 297 concentration and low rhizome C:N ratio, Fig. 2d and b), whereas it had a negative 298 impact on the accumulation of leaf phenolics (Fig. 2f). A significant interaction of CO₂ 299 and nutrient additions was detected for epiphyte load and sediment organic matter (Fig. 300 2g and h). The leaf epiphyte load increased significantly under CO₂ addition, whereas 301 nutrient enrichment and especially the interactive CO₂ and nutrient additions had a 302 lower and not significant effect (t test comparisons in Fig. 2g). Similarly, CO₂ and 303 nutrient interaction resulted in maintenance of the organic matter content in the 304 sediment, which tended to decrease with separated CO_2 and nutrient additions (t test 305 comparisons in Fig. 2h).

In plants from the high-nutrient meadow (Fig. 2, black bars), CO_2 enrichment decreased rhizome C:N (Fig. 2b) and increased epiphyte loads (Fig. 2g). The CO₂-induced increase of the epiphyte load was maintained under the simultaneous addition of nutrients. Nutrient addition enhanced the leaf nutritional quality (high leaf nitrogen concentration, Fig. 2a). A reduction of leaf C:N ratio and phenolics was detected apparently in response to CO_2 and/or nutrient enrichments (Fig. 2c and f), but this was actually caused by an increase of these traits in the unfertilized plants at the end of the

- experiment in relation to the initial field conditions (Table 1). A synergistic interaction between CO_2 and nutrient additions caused an increase of the rhizome length (Fig. 2e).
- 315 Variables for which no significant effects of CO_2 or nutrient addition were detected are 316 shown in Appendix D (Supplement).

317 3.3 Responses of Z. noltii epiphytes

318 Both, CO₂ and nutrient additions altered the relative abundance of epiphyte populations, 319 whereas elevated nutrient levels also modified the epiphyte composition (Fig. 3a). In the 320 unfertilized plants, the epiphyte cover was low and the most abundant leaf epiphytes 321 were the fanlike diatoms Licmophora spp. The second-most-abundant epiphyte in plants 322 from the low-nutrient meadow was the encrusting coralline algae Melobesia 323 membranacea, whereas in plants from the high-nutrient meadow it was the 324 cyanobacterium Microcoleus spp. The response to the CO₂ enrichment in both, low- and 325 high-nutrient meadows was a great increase of epiphyte cover, mostly due to a bloom of 326 Microcoleus spp. (73% of the total cover) that outcompeted the diatoms Licmophora 327 spp. and the encrusting corallines. Under nutrient-enrichment pennate diatom 328 populations dominated by *Navicula* spp. outcompeted the other taxa. In the nutrient- and 329 CO₂-and-nutrient- treatments the composition of epiphyte assemblages was similar, but 330 with a reduced replacement of Licmophora spp. by pennate diatoms in the CO₂-and-331 nutrient- treatment. Chlorophytes (mainly Ulva prolifera) and filamentous rhodophytes 332 (mainly Bangia spp. and Stylonema alsidii) were also present in all treatments. 333 Temporal changes in epiphyte abundances within the enriched mesocosms involved a 334 shift from relatively low epiphyte loads until the second week to increasing epiphyte 335 loads from the third week onwards, with the occurrence of purple bacteria in nutrient-336 and CO₂-and-nutrient-enriched treatments during the fourth week.

337 NMDS ordination of treatments based on the epiphyte composition showed clear CO₂ 338 effects (Fig. 3b). CO₂ treatments were separated along axis I (51% of variance 339 explained), whereas the other treatments were ordered along axis II (49% of variance 340 explained) from unfertilized to CO₂-, CO₂-and-nutrient-, and nutrient-enriched. 341 Separation of CO₂ enrichments along axis I was due to a higher epiphyte cover (mean +se: 22+2.3 cm² per 10 cm² of leaf) than the unfertilized, nutrient-enriched and CO₂-342 and-nutrient-enriched treatments (7.6+1.4, 11+1.3 and 8.4+3.2 cm² per 10 cm² of leaf, 343 344 respectively). Treatments of both, low- and high-nutrient meadows were nearby in the ordination diagram, reflecting minor differences among meadows in the response of the
epiphyte assemblage. NMDS pattern was further confirmed by the PERMANOVA
results, which showed significant effects of CO₂, nutrients and their interaction (Fig.
348 3b).

The above-mentioned PCA of traits from the low- and high-nutrient meadows further confirmed the increased epiphyte load and the change in epiphyte composition as main drivers of the meadow responses to CO_2 and nutrient enrichments (see Appendix E, Supplement).

353 **3.4 Response trajectories through time**

354 The first two PCA components of the low-nutrient meadow traits measured through 355 time, explained 41% (component I) and 20% (component II) of the variance. The Z. 356 noltii traits that highly correlated with component I were the LAI and the number of 357 leaves, which were negatively correlated with shoot mortality and herbivory (Fig. 4a, 358 right graph). Flowering, community production and community respiration highly 359 correlated with component II (variable loadings are presented in Table E2, Supplement). 360 The variability of all treatment scores on the component I during the first week and of 361 unfertilized and CO₂-enriched treatments during the second and third weeks were within 362 the initial range of natural variability (week 0, grey rectangle in Fig. 4a, left graph). At 363 this time, Z. noltii plants showed higher LAI and higher number of leaves. The time 364 series ordination of the rest of treatments along component I revealed that the effects of 365 nutrient addition started during the second week, when the scores of nutrient- and CO₂-366 and-nutrient-treatments suddenly shifted to higher values. These nutrient effects were 367 dominated by high mortality of Z. noltii shoots and to a less extent by high meso-368 herbivory. The highest scores on component I were attained by the nutrient-treatment 369 during the fourth to sixth weeks. The system response to the CO₂ treatment was slower 370 (starting at week 4) and of lower magnitude than the response to nutrient- and CO₂-and-371 nutrient-treatments. This analysis supported the previous indication of a temporal 372 threshold for the meadow responses, which was the second week for elevated nutrients 373 and the fourth week for elevated CO₂. No clear ordination of treatments was detected 374 along component II, indicating that traits highly correlated with this component were 375 substantially influenced by natural variability.

376 The first two PCA components of the high-nutrient meadow traits measured through 377 time explained 64% (component I) and 12% (component II) of the variance. The Z. 378 noltii traits that highly correlated with component I were the LAI, the number of leaves, 379 the community production and the abundance of Ulva, which were negatively correlated 380 with mortality, herbivory and detritus production (Fig. 4b, right graph). Community 381 respiration highly correlated with component II (see variable loadings in Table E2, 382 Supplement). The range of initial natural variability of all treatment scores was narrower than for the low-nutrient meadow (Fig. 4b, left graph). The system was 383 384 initially dominated by high seagrass LAI and number of leaves, cover of Ulva spp., and 385 community production. This progressively shifted to a later stage (week 6) dominated 386 by high Z. noltii mortality, herbivory and detritus production. Contrary to the response 387 of the low-nutrient meadow, there were no relevant differences in the time course and in 388 the final stage attained by PCA scores of both unfertilized and enriched treatments. The 389 shoot mortality of Z. noltii was positively correlated with meso-herbivore activity and 390 detritus production, and negatively correlated with LAI, number of leaves and Ulva 391 cover. No clear ordination of treatments was detected along component II.

392

393 4 Discussion

4.1 Effects of CO₂ enrichment in low-nutrient meadows

395 The CO₂ enrichment had no direct effects on Z. noltii biochemistry (Fig. 5a). We found no evidence of increased nonstructural carbohydrates and subsequent nitrogen dilution 396 397 effect (increased C:N ratio) as has been previously observed in the seagrass Thalassia 398 hemprichii (Jiang et al., 2010) and T. testudinum (Campbell and Fourgurean, 2013). As 399 well, there was no increase of phenolic contents as predicted by the carbon-nutrient 400 balance hypothesis and no propagation to susceptibility to herbivory. Several studies in 401 terrestrial plants reveal that this lack of response is not uncommon (reviewed by 402 Peñuelas and Estiarte, 1998 and Bidart-Bouzat and Imeh-Nathaniel, 2008). The lack of 403 accumulation of carbohydrates and phenolics that we observed could be explained by 404 the trade-off between secondary metabolism and plant growth predicted by the growth-405 differentiation balance hypothesis under no light and nutrient limitation (review by 406 Stamp, 2003). However, we found no significant increase of seagrass productivity under 407 CO₂ enrichment to support this trade-off, probably due to light limitation induced by

epiphyte overgrowth from the third week of experiment onwards. Photosynthesis 408 409 enhancements have been reported under CO2 addition in Z. noltii (Alexandre et al., 410 2012) and Z. marina (Zimmerman et al. 1997), but they do not always translate into 411 seagrass growth since other determinant factors such as light and nutrient availability 412 are also at play (Palacios and Zimmerman, 2007; Alexandre et al., 2012; Campbell and 413 Fourgurean, 2013). In accordance with the previously cited terrestrial studies, our 414 results suggest that the seagrass responses to elevated CO₂ levels are highly context-415 and species-specific, and are not as readily consistent and predictable as resource 416 availability hypotheses would suggest.

417 The most striking response of Z. noltii meadows to the CO₂ enrichment was the increase 418 of the epiphyte load, with changes in the relative abundance but not in the identity of the 419 main epiphyte taxa. Interestingly, epiphyte-induced shading did not cause seagrass 420 mortality as occurred under nutrient enrichment (see below). This suggests attenuation 421 of the negative effect of reduced light availability by increased CO₂ availability, which 422 may reduce the energy cost of carbon uptake (Koch et al., 2013). The epiphyte bloom 423 was mostly caused by the proliferation of the colonial and filament-forming 424 cyanobacterium *Microcoleus* spp. at the expense of a reduction of coralline algae crusts 425 of *Melobesia membranacea* and fanlike diatoms *Licmophora* spp. This is in accordance 426 with previous studies that showed elevated CO_2 / low pH to stimulate cyanobacteria 427 growth and photosynthesis (Liu et al., 2010) and to decrease abundance of coralline 428 algae (Hall-Spencer et al., 2008; Martin et al., 2008; Kuffner et al., 2008, Campbell and 429 Fourgurean, 2014). As well, Hervé et al. (2012) reported negative effects of low pH on 430 diatom valve formation and porosity, which were alleviated by a simultaneous nutrient 431 addition. We found that the activity of mesograzers was insufficient to regulate the 432 epiphyte proliferation in response to increased CO₂ levels, despite their recognized 433 capacity of controlling epiphyte biomass (Hughes et al., 2004) and particularly 434 cyanobacteria blooms (Neckles et al., 1993). Likely explanations are that the feeding 435 capacity of the most abundant mesograzer in the experiment, the amphipod C. filosa, 436 was exceeded by the cyanobacterium overgrowth or that the amphipod was not targeting 437 these particular epiphytes.

438 Epiphyte overgrowth resulted in increased detritus production and decreased organic 439 matter accumulated in the sediment. This suggests that bacterial decomposition in the 440 sediment was accelerated due to the highly labile organic matter of epiphytes, as 441 reported under nutrient enrichment (see below). The acceleration of bacterial 442 degradation of organic matter polysaccharides at low pH reported by Piontek et al. 443 (2010) would also support this explanation. Our findings may have relevant 444 implications, since the organic carbon produced in seagrass meadows sustains important 445 detritus-based food webs (Pergent et al., 1994; Moore and Fairweather, 2006) and 446 provides a major global carbon sink (Pidgeon, 2009; Fourqurean et al., 2012).

447 4.2 Nutrient enrichment and interaction with CO₂ in low-nutrient 448 meadows

449 We found that nutrient enrichment had a faster and greater effect than CO₂ addition on 450 meadows developing in low-nutrient conditions. Nutrient enrichment enhanced leaf 451 nutritional quality (high nitrogen and low C:N ratio) and reduced the accumulation of 452 phenolic compounds (Fig. 5a). Both, the overall increase of plant nitrogen (e.g. Cabaço 453 et al., 2008; Invers et al., 2004) and the decrease of phenolics (e.g. van Katwijk et al., 454 1997; Goecker et al., 2005) have been widely described in seagrasses as a result of 455 nutrient additions. The nitrogen increase was higher in leaves than in rhizomes, as 456 expected for this fast-growing species that acquires preferentially ammonium through 457 the leaves and shows minimal translocation of nitrogen to belowground tissues 458 (Alexandre et al., 2011).

459 A reduction of the number of Z. noltii leaves and of leaf area index, and an increase of 460 shoot mortality were observed after the second week in response to nutrient addition. 461 These effects can be linked to ammonium toxicity, which has previously been reported 462 in Z. noltii (Brun et al., 2002) and other seagrass species (Santamaría et al., 1994, van 463 Katwijk et al., 1997). In addition, nutrient-induced changes in the epiphyte assemblage 464 may also contribute to shoot mortality by reducing light availability to seagrass leaves. 465 This seems supported by the abrupt increase of shoot mortality after the third week, 466 coinciding with the shift of the epiphyte assemblage from coralline algae and fanlike 467 diatoms to a dense layer of pennate diatoms (mostly of the genus Navicula), with a 468 purple bacteria biofilm developing as well during the fourth week. Towards the end of 469 the experiment, excess organic matter was released within the system due to increased 470 shoot mortality and epiphyte shifts. Coincidently, the accumulation of organic carbon in 471 the sediments decreased, suggesting that an accelerated microbial decomposition was 472 promoted by the higher nutritional quality of producers as reported elsewhere for

473 terrestrial systems (Wardle et al., 2004) and seagrass beds (López et al., 1998; Holmer
474 et al., 2004; Spivak et al., 2007).

475 The simultaneous addition of CO₂ and nutrients did not modify the individual effects of 476 nutrient enrichment on plant biochemistry, but attenuated the proliferation of certain 477 epiphyte taxa (also occurring under high CO₂) and subsequent nutrient-induced Z. noltii 478 mortality. The interactive attenuation of epiphyte overgrowth may result from an 479 increase in the interspecific competition between the species that dominated the 480 epiphyte community under elevated CO₂ (i.e. the cyanobacterium *Microcoleus* spp.) and 481 under elevated nutrient levels (i.e. diatoms of the genus Navicula). Our findings are in 482 agreement with the negative effects of interspecific competition on the involved species 483 (i.e. symmetrical competition) that have long been reported in ecological studies 484 (Connel, 1983). Together, attenuation by simultaneous CO₂ and nutrient additions of the 485 overgrowth of certain epiphytes and of nutrient-induced Z. noltii mortality, reduced the 486 amount of more labile organic matter reaching the sediments compared to the individual 487 CO₂ or nutrient enrichment. This probably resulted in the maintenance at control levels 488 of the bacterial decomposition rates and of the sediment capacity to store organic 489 matter. To our knowledge, this is the first report of the interactive effect of CO₂ and 490 nutrient enrichments on the meadow carbon sink capacity. Overall, we found that under 491 simultaneous addition of CO₂ and nutrients, species interactions attenuated the direct 492 effects of individual stressors on Z. noltii and on sensitive epiphyte species or 493 taxonomic groups.

494 **4.3** High- vs. low- nutrient meadows

495 Our results revealed that the expected benefits of high CO₂ predicted for the end of the 496 century on seagrass productivity might be restrained by epiphyte overgrowth and by the 497 interaction with local eutrophication. In both low- and high-nutrient meadows, CO2 498 effects were more important in epiphyte populations than in the seagrass Z. noltii. These 499 findings strengthen the increasingly recognized importance of species interactions in 500 modulating the direct effects of eutrophication or acidification in single species, 501 populations, and ultimately in ecosystem functioning (Orth et al., 2006; Kroeker et al., 502 2013b). The effect of nutrient enrichment was greater in the low- than in the high-503 nutrient meadow (Fig. 5a vs. b), with nutrient-induced mortality of Z. noltii only 504 appearing in the former. When CO₂ and nutrient enrichments interacted, an increase of

505 epiphyte load was observed in the high-nutrient meadow as opposed to the low-nutrient 506 meadow. These results highlight the context-dependence of the effects of multiple 507 stressors in agreement with the meta-analysis of Crain et al. (2008). We observed that 508 the accumulation of phenolics and carbohydrates was higher under lower nutrient 509 regimes in the initial field conditions, and also in the experimental conditions for 510 phenolics. This suggests that nutrient deficiency rather than a direct effect of high CO₂ 511 drives the accretion of carbon-based compounds in Z. noltii. These observations are in 512 agreement with previous studies in both, terrestrial plants (Lambers, 1993; Peñuelas and 513 Estiarte, 1998) and seagrasses (Campbell et al., 2012), and reinforce the idea of the 514 context-dependence of seagrass responses to CO₂ enrichment.

515 Interestingly, we detected little evidence that CO₂ or nutrient addition affected seagrass 516 herbivory by mesograzers. However, clear differences in herbivory between meadows 517 were observed. Plants from the high-nutrient meadow experienced a 12-fold higher 518 amphipod grazing than plants from the low-nutrient meadow, which resulted in a 519 massive loss of shoots. Blooms of the amphipod C. filosa may occur in warmer months 520 (Appadoo and Myers, 2004). This tube-building amphipod is widely distributed and 521 uses macroalgae for feeding (Ceh et al., 2005) and shelter (Appadoo and Myers, 2003). 522 To our knowledge, these observations constitute the first report of C. filosa using the 523 seagrass Z. noltii for both feeding and shelter-construction. We found that the 524 consequences of plant-specific vulnerability to grazing on seagrass meadows can be 525 stronger than the effects of CO₂ and eutrophication. This result concurs with findings by 526 Alsterberg et al. (2013), which showed that the presence of grazers masked the response 527 of benthic microalgae to ocean acidification and warming. Further studies aiming to 528 identify the factors underlying the plant-specific seagrass vulnerability to grazers are 529 thus of vital importance.

530 Our results showed that separated CO₂ or nutrient enrichment individually result in a 531 loss of the carbon sink capacity of the low-nutrient meadow, as opposed to the high-532 nutrient meadow. This loss contrasts with results of previous studies conducted in situ 533 with other seagrass species, which found that the meadow carbon sequestration capacity 534 was unaffected by nutrient addition (Antón et al., 2011) or increased due to CO₂ 535 enrichment (Russell et al., 2013). In both studies, the whole-community metabolism 536 was used as indicator of the carbon storage capacity of seagrass meadows. In our study, 537 however, the loss of carbon storage occurred without a significant response of the whole-community metabolism, indicating a metabolic compensation between the *Z*. *noltii* populations and the leaf epiphyte and sediment microbial communities. A similar dynamic global balance has also been reported for marine pelagic systems under CO_2 addition (Silyakova et al., 2013).

Overall, we found that shifts in the community dynamics of leaf epiphytes or sediment 542 543 bacteria mediated the multi-level responses of Z. noltii meadows to independent CO2 or 544 nutrient addition. They also modulated the attenuation of individual effects under 545 simultaneous CO₂ and nutrient enrichment. Overgrazing masked the response to CO₂ 546 enrichment and eutrophication but only in the high-nutrient meadow. Our findings 547 highlight the importance of integrative multi-level and ecosystem-based approaches 548 considering not only species interactions and connections between organization levels, 549 but also the effect of interactive stressors, to anticipate the evolution of seagrass 550 meadows in the near future and to endorse conservation efforts.

551

552 Appendices

553 Appendix A. Full comparative description of the low- and high-nutrient meadows of 554 *Zostera noltii* in June-August 2011, prior to the start of the experiment.

555 Appendix B. Seawater chemistry within the experimental mesocosms.

556 Appendix C. Full results of the response to CO₂ and nutrient additions of Zostera noltii

557 plant-, community-, and ecosystem-level traits measured through time.

558 Appendix D. Full results of the response to CO₂ and nutrient additions of Zostera noltii

559 plant-, community-, and ecosystem-level traits measured at the end of the experiment.

- 560 Appendix E. Results of principal component analyses.
- 561

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Table 1. Environmental and *Zostera noltii* community traits that revealed significant differences between the low- and high-nutrient donor meadows during June-August 2011, prior to the start of the experiment, tested using unpaired t-tests. ^aSqrttransformed data to meet normality. ^bMann-Whitney rank tests were conducted for variables that did not meet normality even after transformation. All measured traits and methods are shown in Appendix A (Supplement).

	Low-nutrient meadow	High-nutrient meadow
Leaf phenolics (mg (gDW) ⁻¹)	48 <u>+</u> 1.2	29 <u>+</u> 3.2
Leaf nitrogen (mg gDW ⁻¹)	21 <u>+</u> 0.8	25 <u>+</u> 0.6
Leaf C : N	19 <u>+</u> 0.7	16 <u>+</u> 0.4
Rhizome starch (mgGlu (gDW) ⁻¹)	473 <u>+</u> 14	355 <u>+</u> 27
Rhizome TNC (mg Glu g (DW) ⁻¹)	668 <u>+</u> 18	532 <u>+</u> 28
Shoot area (cm ² shoot ⁻¹) ^a	7.8 <u>+</u> 0.6	4.5 <u>+</u> 0.7
Z. noltii density (shoots m ⁻²)	5517 <u>+</u> 755	2664 <u>+</u> 411
<i>Z. noltii</i> cover (% of sediment surface) ^b	96 <u>+</u> 2.2	18 <u>+</u> 9.8
<i>Ulva</i> spp. cover (% of sediment surface) ^b	absent	38 <u>+</u> 12
Seawater nitrate $(\mu M)^{b}$	< 0.01	1.1 <u>+</u> 0.2
Seawater ammonium (µM)	0.7 <u>+</u> 0.2	3.0 <u>+</u> 0.4
Seawater phosphate (μM)	0.5 <u>+</u> 0.1	1.2 <u>+</u> 0.1

782 Figure captions

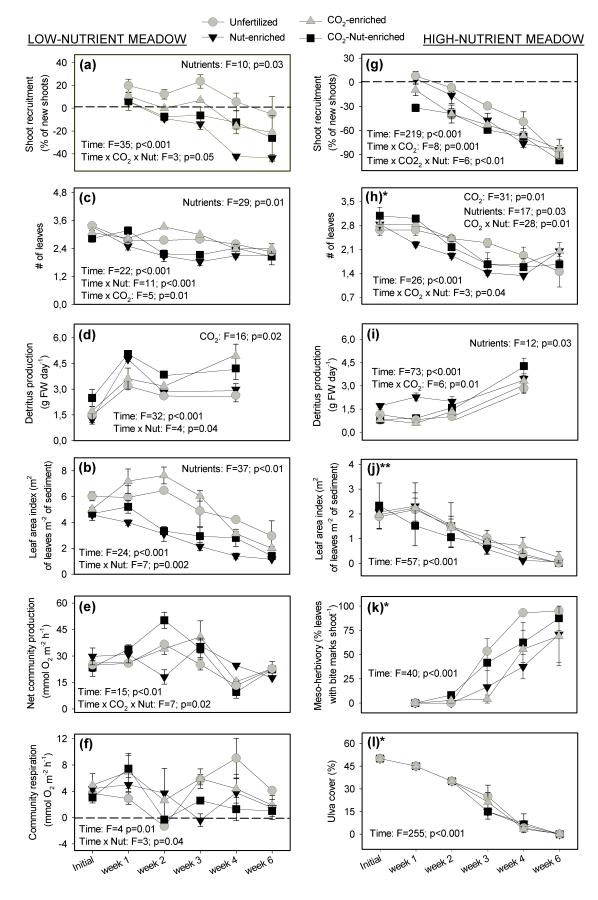
783 Fig. 1. Effects of CO₂ and nutrient additions on Zostera noltii plant-, community-, and 784 ecosystem-level traits from the low-nutrient (left graphs) and high-nutrient meadow (right graphs) through time. Symbols are means (\pm se, n = 2). F statistics and p levels 785 786 from RM ANOVA tests are shown for among-subject factors (CO₂; Nut: nutrients) and 787 their interaction (CO₂ x Nut), and for within-subject factor (time) and interactions (Time 788 x CO₂, Time x Nut, Time x CO₂ x Nut). Only the significant effects (p < 0.05) and useful traits for results interpretation are shown here; the non-significant effects are 789 790 shown in Appendix C (Supplement). *Variables that did not meet normality after transformation, for which the significance level was more restrictive (p < 0.03) to 791 792 minimize the possibility of Type I error. **Variable sqrt-transformed to meet normality.

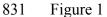
793 Fig. 2. Effects of CO₂ and nutrient additions on Zostera noltii plant-, community-, and 794 ecosystem-level traits measured at the end of the experiment from the low-nutrient (grey 795 bars) and high-nutrient meadow (black bars). Bars are means (+se, n = 2). The F 796 statistics and p levels from two-way ANOVA tests are shown for each fixed crossed 797 factor (CO₂; Nut: nutrients) or interaction (CO₂ x Nut). Lowercase letters above bars 798 show significant differences between treatments for significant interactions (t tests, see 799 Methods). Only restrictive significant effects (p < 0.03) selected to minimize the 800 possibility of Type I error due to unequal variances are shown here; the non-significant 801 effects are shown on Appendix D (Supplement).

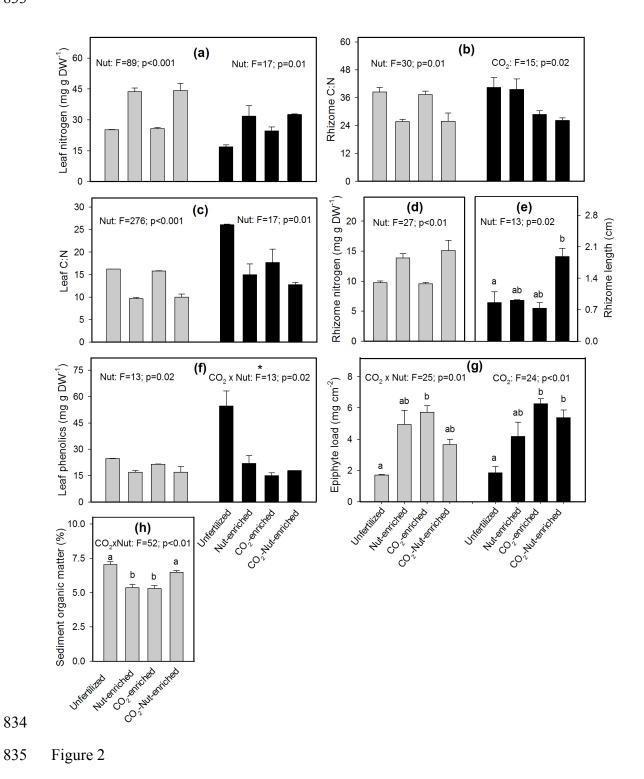
802 Fig. 3. Responses of Zostera noltii epiphyte populations to experimental addition of 803 CO₂ and nutrients: (a) Relative abundance of the main epiphyte taxa of plants exposed 804 to experimental treatments; and (b) NMDS ordination of experimental treatments (see 805 symbol legend below treatment names in Fig. 3a) based on leaf epiphytes of plants from 806 the low-nutrient (L; grey symbols) and high-nutrient meadow (H; black symbols). 807 Pseudo-F statistics and p levels from two-way PERMANOVA test are shown in the 808 NMDS diagram for each fixed crossed factor (CO₂; Nut: nutrients) and their interaction 809 $(CO_2 \times Nut)$.

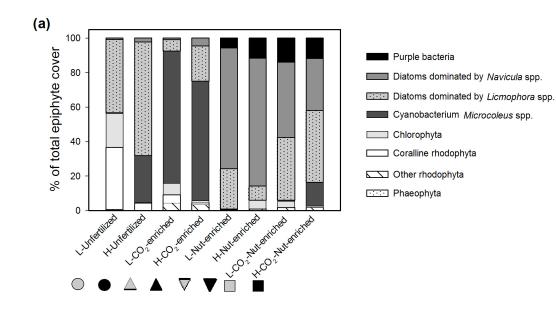
Fig. 4. Principal components analysis of *Zostera noltii* plant-, community, and ecosystem- level responses to treatments through time: (a) low-nutrient and (b) highnutrient meadow. Numbers inside the symbols indicate sampling weeks from 0 to 6. The initial variability (week 0) along component I is incorporated within a grey rectangle to highlight the range of initial natural variability. Variable loadings on the
two principal components are depicted in right graphs. LAI refers to leaf area index,
NCP to net community production and CR to community respiration.

817 Fig. 5. Summary of the effects of CO₂ (blue line) and nutrient (green line) additions, and 818 when significant of their interaction (red line), on Zostera noltii plant- community and 819 ecosystem- level traits of low-nutrient (a) and high-nutrient (b) meadows. Solid lines 820 indicate significant effects on variables measured at the end of the experiment and 821 dashed lines on variables measured through time. Dotted lines indicate no significant 822 effects. Letters below the x-axis denote unfertilized (U) and enriched (E) treatment, 823 which is the pooled mean response to the respective enrichment over the other when 824 there was no significant interaction (no red line). For shoot mortality and net community production (NCP) in the low-nutrient meadow a significant Time x CO₂ x Nutrients 825 interaction was detected and values were represented as for CO2 x Nutrients significant 826 827 interactions. * Apparent effect due to a drastic increase in unfertilized plants in relation 828 to the initial field conditions (see results).

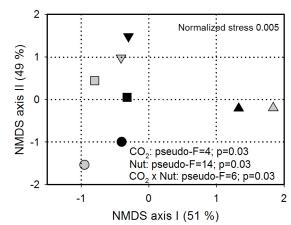








(b)



836

837 Figure 3

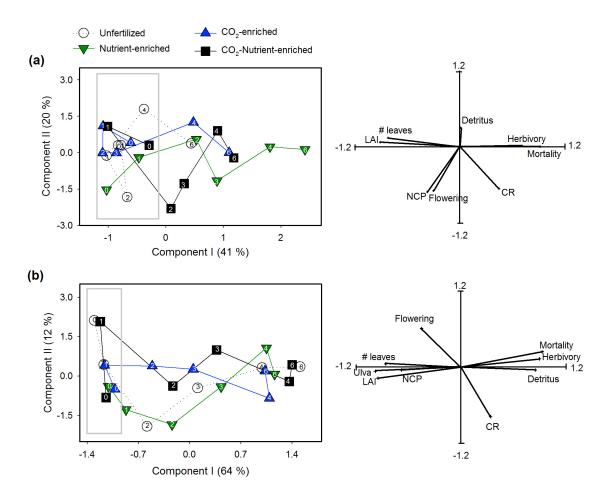
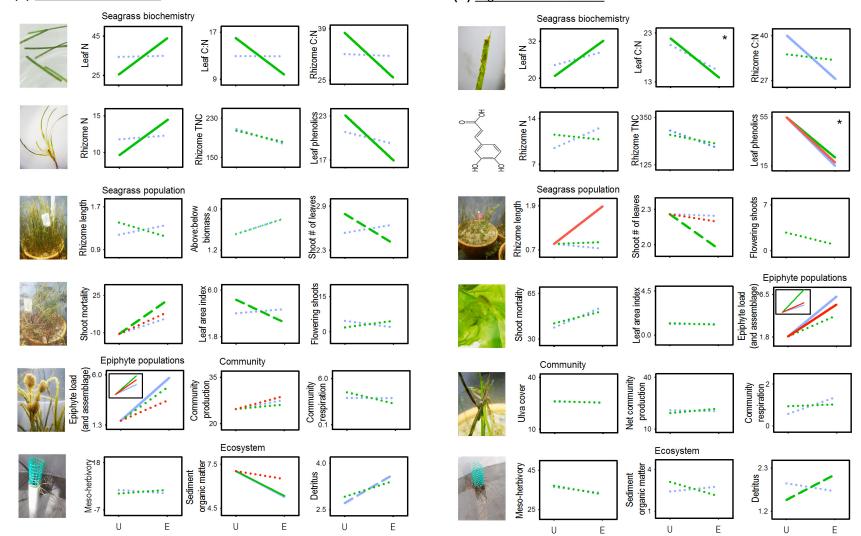




Figure 4.

(a) Low-nutrient meadow

(b) <u>High-nutrient meadow</u>



841 Figure 5