

Interactive comment on “CO₂ and nutrient-driven changes across multiple levels of organization in *Zostera noltii* ecosystems” by B. Martínez-Crego et al.

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REPLY TO REVIEWER #1 COMMENTS

In this reply version, our answer to technical corrections on pages 5256 (L2-4), 5257 (L10-11), and 5251 (L13-14) has been modified to jointly address interlinked Reviewer#1 and #2 comments (see below). Technical corrections accepted without further clarifications are also listed.

We greatly appreciate the comments and detailed suggestions by the anonymous referee. The referee provided two major comments, two specific comments, and several technical corrections for manuscript improvement or for detail clarifications. Regarding the general comments, as the referee correctly points out, our experimental design allows for determining the nature of the interaction between stressors (synergistic or antagonistic) but not the existence of thresholds or linear responses. This latter point was out of the scope of our study. The referee acknowledges that, in spite of the low replication of treatments, significant differences were detected that provided a general insight on the seagrass ecosystem responses at several levels of the biological organization. This general view is the major aim of the present work and it suggests ways forward for future research.

Referee’s specific comment – “*The authors attribute the lack of grazer control of epiphyte growth to either an imbalance between epiphyte growth and grazing activity or a non-specialist grazer being the most abundant. Is it possible that the shift in epiphyte assemblages towards cyanobacteria which were less palatable for the grazers present?*”

This would be a particular case in which the most abundant mesograzer in our experiment, *Cymadusa filosa*, was not targeting the epiphytes that were mostly composed by cyanobacteria in the CO₂-enriched treatment. However, cyanobacteria are palatable and structurally accessible food for mesograzers (e.g. Vargas et al. 1998- J. Phycol. 34, 812–817; Nagarkar et al. 2004- Hydrobiologia 512: 89–95). At the same time, as stated in this paragraph, small invertebrate grazers (amphipods, isopods, gastropods) have been widely recognized as controllers of epiphyte overgrowth under nutrient enrichments (Hughes et al., 2004-Mar Ecol Prog Ser 282, 87-99), including cyanobacteria blooms (Neckles et al. 1993-Oecologia, 93, 285-295). We have revised the paragraph according the referee’s suggestion, considering the particular control of the cyanobacteria overgrowth by the amphipod *Cymadusa filosa* rather than a general epiphyte control by grazers (L539-545 in the marked-up manuscript version, see below).

Referee’s specific comment – “*Overall, the statistical analysis is sound, however I would question*

why t-tests were used instead of post-hoc analysis (e.g. SNK, Tukeys or an appropriate test if variances were heterogeneous)? The ANOVA is the appropriate test to use and does not need to be confirmed by t tests. Nor do t tests tell you anything about interactions.”

Regarding the first part of this remark, post hoc comparisons were not possible in the factorial design because we only have two levels (unfertilized and enriched) within each factor (CO₂, Nutrients). When significant interactions were detected, we needed another test to detect significant differences between the 4 treatments separately considered (Unfertilized, CO₂-enriched, Nut-enriched, and CO₂-Nut-enriched). Given the unequal variances found for all variables under this design (1 factor: treatment, with 4 levels) and that the homogeneity of variances is an important assumption in the ANOVA (see Quinn and Keough 2002), we chose to use Welch's t test (robust against unequal variances) instead SNK or Tukey's comparisons after one-way ANOVA.

We agree with the referee that the ANOVA is the appropriate test and does not need to be confirmed by t tests. We have revised this paragraph to incorporate the remarks made by the referee (L272-280 in the marked-up manuscript version). We have clarified that within the design of the two-way ANOVA (two fixed crossed factors), a normal distribution with unequal variances was found for all variables, which is usual when the sample size is small. Following recommendation by Quinn and Keough (2002), we proceeded with the analyses but making significance level more restrictive to minimize the possibility of Type I error (mistakenly detection of differences). Fig. 2 and 5 (and Fig.2 legend), Appendix D, and text in the manuscript have been modified accordingly (e.g. changes in L364-386).

Regarding the technical corrections suggested by the referee, we have either accepted all edits and minor corrections or have added most suggested clarifications. Clarifications are addressed in detail as follows:

“Page 5245, Line 18: It is not clear what three-five leaves means and why it is not standardised to a single number” - Five shoots were used to quantify the number of leaves and the LAI at the beginning and at the end of the experiment, while three shoots were used in between sampling events. We have revised this paragraph to clarify this referee's remark (L202-207).

“Page 5245, Line 28: How were epiphytes removed?” - Epiphytes were removed with a glass slide to minimize damage on leaf surface. This has been clarified in the revised version (L216).

“Page 5247, Line 8: I don't think export is the correct term. How do you know this carbon would not have settled back to the sediment within the bed?” – The referee is right. In seagrass meadows, detritus production can be exported by currents and waves to neighboring systems (e.g. deep systems or beaches), enter in the trophic web through decomposers or detritivores, or be buried under the meadow becoming part of the carbon sink (Pergent et al. 1994-Mar Ecol Prog Ser 106, 139-146; Cebrián and Duarte 2001-Aquatic Botany, 70, 295-309; see also references already cited in the manuscript: Pidgeon 2009, Fourqurean et al. 2012). To avoid any confusion, we have removed the

allusion of detritus production as indicator of carbon export (L258).

“Page 5247, Line 19: Greenhouse-Geisser adjustments actually correct the degrees of freedom, not the Significance levels (although significance levels will also change)” - The referee is right. We have corrected this sentence (L270).

“Page 5251, Lines 13-14: Should read “...with only purple bacteria appearing in them.” - This sentence has been altered to highlight the attenuation of nutrient-induced changes in the epiphyte assemblage by the simultaneous addition of CO₂ (L400-403).

“Page 5255, Line 13: Surely there are more recent papers than (Fenchel 1977) to cite.” - Fenchel 1977 has been replaced by more recent references that highlight the relevance of detritus-based food webs in seagrass meadows (i.e. Pergent et al. 1994-Mar Ecol Prog Ser 106, 139-146; Moore and Fairweather 2006-Oikos 114, 329-337).

“Page 5256, Lines 2-4: The sentence beginning with “To avoid toxicity, . . .” is confusing and the latter part needs revising for clarity.” - The paragraph containing this sentence (L604-609) has been rewritten in the revised version to address also Reviewer#2 remark about this paragraph (comment#10).

“Page 5257, Lines 10-11: “The effect of nutrient enrichment was higher in the low-nutrient than in the high-nutrient meadow (Fig. 5a vs. b).” This belongs in results section.” - Reviewer#2 also made a similar comment (#13). We have revised the entire paragraph to incorporate a proper discussion and contextualization of the results as Reviewer#1 and #2 requested (L677-712).

“Page 5258, Line 7: The sentence beginning with “Blooms of the, . . .” is confusing and the latter part needs revising for clarity.” - The sentence has been rewritten and the confusing part has been removed (L717-718).

List of technical corrections accepted without further clarifications:

Page 5243, Line 11: Should read “...assess if they react differently.”

Page 5245, Line 28: Should read “...five shoots per pot.”

Page 5247, Line 19: Greenhouse-Geisser adjustments actually correct the degrees of freedom, not the Significance levels (although significance levels will also change)

Page 5251, Line 28: Replace “low” with “minor” or “small”.

Page 5255, Line 19: Replace “stronger” with “greater”.

Page 5255, Line 27: Replace “a scarce” with “minimal”.

Page 5258, Line 19: Should read “...CO₂ or nutrient enrichment.” - This correction has also been incorporated through the manuscript (e.g. L387, 669, 713, 728)

Page 5258, Line 25: Replace “with no” with “without a”.

Page 5262, Line 1: Hughes et al. 2004 – List of minor authors has first names rather than surnames

listed (e.g. Susan, L. W. should be Williams, S. L.)

List of technical corrections that refer to paragraphs that were modified in the revised version following Reviewer#2 suggestions:

Page 5253, Line 26: Replace “evidenced” with “observed”.

Page 5254, Line 6: Remove “, however,”.

Page 5256, Line 5: Replace “since” with “as”.

Page 5257, Line 27: Should read “...meadow supports the hypothesis that nutrient.”

Page 5258, Line 20: Replace “of” with “in”.

REPLY TO REVIEWER #2 COMMENTS

We thank the reviewer for the useful comments, which helped to improve the manuscript by flushing out our oversights and clarifying key aspects. The comments are presented below, followed by our responses.

Referee’s specific comment #1 - *Lines 8-10, p5241: Please provide citations to support these statements.*

The reviewer refers to the sentence “This body of research has revealed that ocean acidification can be detrimental to most marine calcifying organisms, while increasing carbon dioxide (CO₂) concentration can benefit primary productivity of phytoplankton, cyanobacteria, fleshy algae, and seagrasses”. This sentence is connected to the previous one, in which the reviews by Doney et al. (2009) and Kroeker et al. (2010) are cited for supporting the statements. No change done.

Referee’s specific comment #2- *Lines 27-28, p5241: Please provide citations to support these statements.*

A reference has been added (Kroeker et al. 2013b-Nature Clim. Change 3, 156-159).

Referee’s specific comment #3 - *Line 1, p5254: Any reason why all effects on epiphytes and sediments were attenuated under both CO₂ and nutrient enrichment.*

Regarding changes in the epiphyte community, we found that the interactive effect resulted in the attenuation of both, the nutrient-induced changes in the epiphyte assemblages and the CO₂-induced bloom of the cyanobacterium *Microcoleus* spp. The most likely explanation is that the interspecific competition between the species that dominated the epiphyte community under high CO₂ (the cyanobacterium *Microcoleus* spp.) and under high nutrient levels (diatoms of the genus *Navicula*) resulted in the observed attenuation under simultaneous addition of CO₂ and nutrients of the direct effects of individual stressors on certain taxa. Negative effects of interspecific competition on the involved species (i.e. symmetrical competition) have long been reported by ecologists (Connel, 1983)

and support this explanation.

Regarding changes in the sediments of the low-nutrient meadow, we observed similar organic matter content in the sediment under simultaneous CO₂ and nutrient addition and control conditions, which were lower than under individual enrichments. The simultaneous addition of CO₂ and nutrients might keep microbial decomposition rates in the sediment at control levels, as opposed to the probable acceleration observed under independent CO₂ or nutrient enrichment. Our results under individual enrichments contrast with previous studies conducted in situ with different seagrass species. Antón et al. (2011) found that meadow carbon sequestration was unaffected by nutrient addition, whereas Russell et al. (2013) observed increased carbon sequestration in response to CO₂ enrichment. These authors, however, did not assess the interactive effect of nutrients and CO₂ on carbon sequestration. Moreover, they used whole-community metabolism instead organic matter in the sediment as indicator of carbon sequestration.

We have better explained the responses to the simultaneous CO₂ and nutrient additions in the Discussion, highlighting the key role of species interactions in the attenuation of direct effects of individual stressors. The first paragraph of the discussion, to which the reviewer refers here, has been removed from this position, incorporated into the Abstract and discussed in each appropriate subsection of the Discussion (see comments below and L657-675 and 728-788 in the marked-up manuscript version). In addition, we have highlighted the role of species interactions in mediating the individual and interactive effects of eutrophication and acidification in ecosystem functioning through the manuscript (Discussion, Results, and Abstract).

Referee's specific comments #4 and #8- *Line 11, p5254: However, this finding is in contradiction to much empirical research for seagrasses. Please address. (Jiang et al. 2010, Campbell & Fourqurean 2013).* - *Line 23, p.5254: Yet note that in a subsequent study Campbell 2013 Mar Biol document increases in carbohydrate content with elevated CO₂, along with Jiang et al 2010, Palacios 2007, and Zimmerman et al 1997. Clearly both nutrient regime and CO₂ levels can have an effect on carbohydrate levels and should be acknowledged. (Zimmerman et al. 1997, Jiang et al. 2010, Campbell & Fourqurean 2013)*

These two specific comments refer to the same paragraph and are interlinked, so we answered and addressed them together. We found that CO₂ enrichment had no direct effects on *Z. noltii* biochemistry, with no significant changes on the total carbohydrate reserves. This finding contrasts with observations in the seagrass *Thalassia hemprichii* (Jiang et al., 2010) and *T. testudinum* (Campbell and Fourqurean, 2013).

We have rewritten this entire paragraph to acknowledge that our results are different from others and to reinforce the idea that the plant responses to elevated CO₂ levels are highly context- and species-specific, and are not as readily consistent and predictable as resource availability hypotheses would suggest. References have been modified accordingly. The effects of nutrient regime on the

carbohydrate accumulation have been addressed in the sub-section 'High- vs. low- nutrient meadows', where we discuss that the accretion of carbon based-compounds in *Z. noltii* might be driven by nutrient deficiency, thus, reinforcing the idea of the context-dependence of seagrass response to high CO₂ (L705-712).

Referee's specific comments #5 to #7 - *Line 12, p.5254: So the seagrasses from the low-nutrient meadows were not nutrient limited? What evidence do you have to suggest a trade-off between phenolic production and growth? I currently don't see data to support this conclusion.* - *Line 16, p.5254: Did CO₂ enrichment actually enhance LAI? Is this supported by your statistical analyses.* - *Line 20, p.5254: Were your plants really not under any degree of light limitation? Didn't the excessive epiphyte loading reduce light levels?*

Again, these three specific comments refer to the same paragraph than the previous remarks and are interlinked. We agree with the referee. As stated in the Results section, Figure 1b suggests that the *Z. noltii* leaf area index (LAI) tended to increase with CO₂ enrichment until the third week of the experiment. However, this trend was not statistically significant (no significant CO₂ effect or CO₂ x Time interaction in the RM ANOVA). Consequently, our results did not provide significant arguments to support a trade-off between phenolic production and growth. As the referee suggests, the lack of significant CO₂-induced changes in *Z. noltii* productivity can be explained by nutrient or light limitation to seagrass growth. Given the huge epiphyte overgrowth induced by CO₂ enrichment, we pointed out to light limitation mediated by epiphyte shading as the most likely reason.

We have revised and rewritten this entire paragraph to accommodate these and previous remarks (L475-524). We highlight that we found no significant increase of seagrass productivity under CO₂ enrichment to support the trade-off between phenolic accumulation and plant growth.

Referee's specific comment #9 - *Line 2, p.5255: Cite (Campbell & Fourqurean 2014)*

The reference has been added.

Referee's specific comment #10 - *Line 5, p. 5256: But don't you document declines in shoot recruitment and LAI?*

The referee is right. The paragraph has been modified and combined with the next paragraph to explain that the increased shoot mortality under nutrient enrichment can be linked to ammonium toxicity and/or to a reduction in light availability caused by the dense epiphytic layer of pennate diatoms and purple bacteria (L604-613). References have been modified accordingly. This change also addresses Reviewer#1 remark about this paragraph.

Referee's specific comment #11 - *Line 17 p. 5256: Any chance that this excess organic matter was simply exported out of the mesocosms due to the experimental set up?*

No chance. Any export of organic matter should equally affect all treatments, but it was not apparent in the control conditions as reflected by the relatively high organic matter content in the sediment of

the unfertilized mesocosms.

Referee's specific comment #12 - *Line 22 p.5256: Any explanation for this statement? The first sentence of this paragraph needs clarification / explanation.*

This paragraph has been modified also in reply to the referee's comment #3. As explained above, the simultaneous addition of CO₂ and nutrients attenuated the nutrient-induced changes in the epiphyte assemblages and the CO₂-induced increase in the total epiphyte load through the alteration of competitive dynamics and species interactions within the epiphyte community.

Referee's specific comment #13 - *Line 4- 17 p.5257: This paragraph reads more like the results section. Please revise.*

Reviewer#1 also made a similar comment for lines 10-11 in p5257. We have revised the entire paragraph to incorporate a proper discussion and contextualization of the results as Reviewer#1 and #2 requested (L677-712).

Referee's specific comment #14 - *Line 19 p.5258: But if I understand correctly, combined CO₂ and nutrients had no effect on carbon sink capacity. Why might this be the case?*

See above answer to comment #3.

CO₂ and nutrient-driven changes across multiple levels of organization in *Zostera noltii* ecosystems

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Abstract

Increasing evidence emphasizes that the effects of human impacts on ecosystems must be investigated using designs that incorporate the responses across levels of biological organization as well as the effects of multiple stressors. Here we implemented a mesocosm experiment to investigate how the individual and interactive effects of CO₂ enrichment and eutrophication, scale-up from changes in primary producers at the individual- (biochemistry) or population-level (production, reproduction, and/or abundance) to higher levels of community (macroalgae abundance, herbivory, and global metabolism) and ecosystem organization (detritus release and carbon sink capacity). The responses of *Zostera noltii* seagrass meadows growing in low- and high-nutrient field conditions were compared. In both meadows, the expected CO₂ benefits on *Z. noltii* leaf production were suppressed by epiphyte overgrowth, with no direct CO₂ effect on plant biochemistry or population-level traits. Multi-level meadow response to nutrients was faster and stronger than to CO₂. Nutrient enrichment promoted the nutritional quality of *Z. noltii* (high N, low C:N and phenolics), the growth of epiphytic pennate diatoms and purple bacteria, and shoot mortality. In the low-nutrient meadow, individual effects of CO₂ and nutrients separately resulted in reduced carbon storage in the sediment, probably due enhanced microbial degradation of more labile organic matter. These changes, however, had no effect on herbivory nor on community metabolism. Interestingly, individual effects of CO₂ or nutrient addition on epiphytes, shoot mortality, and carbon storage were attenuated when both nutrients and CO₂ acted simultaneously. Thus reflecting CO₂-induced benefits on eutrophic meadows. In the high-nutrient meadow, a striking shoot decline caused by amphipod overgrazing

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50 masked the response to CO₂ and nutrient additions. Our results reveal that under future
51 scenarios of CO₂, the responses of seagrass ecosystems will be complex and context
52 dependent, being mediated by epiphyte overgrowth rather than by direct effects on plant
53 biochemistry. Overall, we found that the responses of seagrass meadows to individual
54 and interactive effects of CO₂ and nutrient enrichments varied depending on interactions
55 among species and connections between organization levels. ▾

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57 1 Introduction

58 Understanding community and ecosystem responses to human impacts is a challenge
59 that requires integrating not only the organism-level responses across populations and
60 entire systems (Russell et al., 2012), but also synergistic or antagonistic effects of
61 multiple stressors (Woodward et al., 2010). A large number of articles has been
62 published on the effects of ocean acidification, and reviewed among others by Doney et
63 al. (2009) and Kroeker et al. (2010). This body of research has revealed that ocean
64 acidification can be detrimental to most marine calcifying organisms, while increasing
65 carbon dioxide (CO₂) concentration can benefit primary productivity of phytoplankton,
66 cyanobacteria, fleshy algae, and seagrasses. Our current understanding of these effects
67 is largely based on the species-specific responses of individuals or populations.
68 However, the broad variability in responses among organisms may influence species
69 interactions and drive unforeseen impacts on marine communities and ecosystems
70 (Hall-Spencer et al., 2008; Kroeker et al., 2013a).

71 The interactive effect of multiple stressors on ecological communities remains largely
72 unknown (Crain et al., 2008). Atmospheric CO₂ concentration has increased from
73 preindustrial levels of approximately 280 ppm to 397 ppm in 2013 (NOAA, Mauna Loa
74 Observatory, Hawaii), leading to a rise in the CO₂ absorbed by the ocean with an
75 associated pH decrease of 0.1 units. An additional pH decrease of 0.07-0.31 units is
76 expected by the end of the 21th century based on the 'Intergovernmental Panel on
77 Climate Change' predictions (IPCC, 2013). In the marine environment, ocean
78 acidification can locally interact with excess nutrients from coastal eutrophication to
79 accelerate changes in ecosystem structure and functioning (Russell et al., 2009).

80 Human impacts that alter the availability of environmental resources are shifting the
81 nutritional quality of primary producers through changes at the biochemical or

90 individual levels of the biological organization (e.g. allocation of resources to growth,
91 storage, and chemical defences). In addition, changes in environmental resources can
92 favour different types of producers and alter the inter-species competitiveness and
93 producers' abundances (Kroeker et al., 2013b). Overall, this may affect ecological
94 interactions and fluxes, leading to shifts at community and ecosystem levels. Under
95 elevated nutrient concentrations, aquatic and terrestrial ecosystems with higher producer
96 nutritional quality often support higher rates of herbivory, more rapid decomposition
97 rates and recycling of nutrients, and lower net accumulation of soil carbon (Wardle et
98 al., 2004; Cebrian et al., 2009). The effects of nutrient enrichment have been widely
99 described on terrestrial, freshwater, and marine ecosystems, whereas the scaling up of
100 elevated CO₂ effects has been mostly studied in terrestrial plants. Under elevated CO₂
101 levels, and especially if nutrient availability is limiting to growth, terrestrial plants
102 typically increase the accumulation of carbohydrates and/or carbon-based secondary
103 compounds (mostly phenolics). This increases C:N ratios ("nitrogen dilution" effect)
104 and sometimes leaf toughness through increasing indigestible polymers such as
105 cellulose and lignin (Zvereva and Kozlov 2006; Lindroth, 2010; Robinson et al., 2012).
106 Herbivores usually compensate for this lower food quality by eating more (Stiling and
107 Cornelissen, 2007). In addition, CO₂ enrichment may shift the biomass and composition
108 of soil microbial communities, directly through different responses of microbial groups
109 to high CO₂ / low pH (Krause et al., 2012; Lidbury et al., 2012) or indirectly through
110 reducing foliar and detritus quality (Drigo et al., 2007). Overall, high CO₂ levels may
111 have both positive and negative consequences on the decomposition of soil organic
112 matter and nutrient recycling (Lindroth, 2010).

113 In this study, we use seagrass meadows as model ecosystems to investigate the scaling
114 up of the effects of elevated CO₂ and nutrient levels on marine coastal environments.
115 Seagrasses beds are widely distributed habitats that host high biodiversity and provide
116 valuable ecosystem services (Orth et al., 2006). A rich epiphyte community usually
117 colonizes seagrass leaves, thus providing a useful system for studying how changes in
118 environmental resources can favour different types of producers (e.g. non-calcareous vs.
119 calcareous). They are highly productive systems that sequester larger amounts of carbon
120 per area than tropical forests, providing for a long-term removal of carbon dioxide from
121 the atmosphere (Pidgeon, 2009; Fourqurean et al., 2012). The maintenance of the key

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125 services provided by seagrass ecosystems under global change is thus of prime
126 importance for human well-being.

127 A mesocosm experiment was conducted to assess: (1) how CO₂ and nutrient
128 enrichments affect primary producers' at the individual- (plant biochemistry including
129 CN and allocation of resources to carbohydrate reserves and carbon-based chemical
130 defences) or population-level (plant allocation of resources to biomass and
131 reproduction, and composition and abundance of seagrass epiphytes), and (2) whether
132 these changes propagate to the community (macroalgae abundance, meso-herbivory,
133 whole-community metabolism) and to the ecosystem (detritus production and organic
134 carbon storage in sediment). The responses of meadows of the seagrass *Zostera noltii*
135 Horneman developing in low- and high- nutrient conditions in the field were compared
136 to assess if they react differently.

137

138 2 Methods

139 2.1 Study meadows

140 Samples of *Z. noltii* community were collected from two meadows separated 5.5 km
141 from each other, within the Ria Formosa lagoon (South Portugal). This shallow
142 mesotidal lagoon is dominated by monospecific beds of the seagrass *Z. noltii* that
143 occupy ca. 45% of the intertidal area. One meadow was developing under prior field
144 conditions of low nutrient levels (36°59'40''N 7°58'00''W; hereafter low-nutrient
145 meadow) and the other under high nutrient levels (37°01'15''N 8°00'56.50''W;
146 hereafter high-nutrient meadow). Low-intertidal samples exposed to a small emersion
147 period only during low spring tides were selected. Table 1 presents seawater nutrient
148 concentrations and seagrass meadow traits that reveal the substantial initial differences
149 between meadows.

150 2.2 Mesocosm experiment

151 The study was conducted in an outdoor mesocosm system at the Ramalhete field station
152 of the Centre of Marine Sciences, which is located at the Ria Formosa lagoon. To assess
153 the effects of CO₂ and nutrients on *Z. noltii* meadows an enrichment experiment was
154 conducted for 6 weeks during August-September 2011, after 4 days of acclimation to

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163 the experimental mesocosms. This time span is enough to detect any treatment-driven
164 changes in physiological, morphological and population traits of this fast-growing
165 species (e.g. Peralta et al., 2002). Core samples of *Z. noltii* community, including
166 sediment and algal, faunal and microbial components, were randomly collected from
167 each donor meadow and allocated to flowerpots of 20 cm of diameter and height. Three
168 flowerpots were placed in each of the 16 experimental mesocosms (tanks of 110 L),
169 which were exposed to combinations of two CO₂ and two nutrient levels in a crossed
170 design with two replicates.

171 Experimental levels of CO₂ encompassed present (pH 8.00±0.02, equivalent to ca. 400
172 ppm CO₂) and future conditions (pH 7.83±0.01, equivalent to ca. 800 ppm CO₂) in Ria
173 Formosa lagoon. The mesocosms received sand-filtered seawater from two head tanks
174 of 1000 L at a rate of 240 L h⁻¹. In one of the head tanks, the water CO₂ was
175 manipulated to ensure fixed pH differences between treatment means within the range
176 predicted for 2100 by the IPCC (pH decline = -0.18 units) following a commonly used
177 method (e.g. Alsterberg et al., 2013). The CO₂ injection was controlled by an auto-
178 analyzer (Yokogawa, EXAxt 450, Tokyo, Japan), which continuously monitored the
179 water pH and temperature. Total alkalinity, pH, temperature and salinity within the
180 mesocosms, as well as the seawater DIC and carbon speciation are provided on
181 Appendix B (Supplement).

182 Water nutrient levels encompassed the natural values found in the lagoon and the values
183 of highly eutrophic conditions (N: 45x and P: 11x natural, see Appendix B,
184 Supplement). The nutrient enrichment was obtained by adding a solubilised mixture of
185 the fertilizers ammonium nitrate and monoammonium phosphate directly into the water
186 column of each enriched mesocosm using a multi-channel dosing pump. Water samples
187 were collected weekly to analyze nutrient concentrations using a loop-flow analyzer
188 (μ Mac-1000; Systea, Anagni, Italy).

189 The water within mesocosms was homogenized using a submersible circulation pump
190 placed at leaf height. Pumps were stopped 2 h twice per day to simulate tidal currents.
191 Twice a week, the epiphytes growing in the mesocosm walls were removed and the
192 position of flowerpots within each mesocosm was reassigned to minimize potential
193 spatial differences. Natural settlement and growth of leaf epiphytes and small animals
194 were allowed throughout the experiment.

195 2.3 Producers' traits at the individual- or population-level

196 Changes on producers' at the individual or population levels were assessed by
197 measuring: (1) plant biochemistry and allocation of resources to biomass and
198 reproduction; and (2) the composition and abundance of seagrass epiphytes.

199 Allocation of plant resources to biomass and reproduction at the population-level was
200 monitored almost every week. Shoot recruitment or mortality were quantified within
201 each flowerpot excluding shoots growing around the border to avoid edge effects.

202 Allocation to reproduction was quantified as density of flowering shoots. Five shoots at
203 the beginning and at the end of the experiment and three shoots in between sampling
204 events were randomly chosen within each flowerpot to quantify the number of leaves
205 and the leaf area index (LAI) as indicators of aboveground productivity. To estimate
206 LAI, leaf area was measured on these shoots, averaged, multiplied by the number of
207 shoots within the pot, and scaled per surface area.

208 After four weeks of experiment, leaf epiphyte composition was determined in the oldest
209 leaves of three randomly chosen shoots. The surface covered by each taxon was
210 quantified under a microscope and standardized per 10 cm² of leaf area.

211 After six weeks, all shoots from each mesocosm were harvested and plant traits
212 quantified in each flowerpot. Belowground productivity was estimated from the vertical
213 or horizontal rhizome length and from the total root length (number of roots multiplied
214 by the average root length) of five shoots per pot. The above- and below-ground
215 biomass allocation was quantified after drying at 60 °C until constant weight. The
216 pooled epiphyte load of three shoots was removed using a glass slide and quantified as
217 relative to leaf area after drying at 60 °C until constant weight. Pooled material of five

218 shoots was separated into leaves (without epiphytes) and rhizomes, freeze-dried,
219 weighted, ground to fine powder and used in subsequent analyses of plant biochemistry.

220 Carbon and nitrogen concentrations were analyzed using an elemental analyzer (Carlo-
221 Erba, Milan, Italy). Total non-structural carbohydrates were measured in rhizomes using
222 the phenol-sulfuric acid colorimetric method (Dubois et al., 1956) with glucose as
223 standard, after sugar extraction in hot ethanol and enzymatic conversion of starch to
224 glucose equivalents (Smith and Zeeman, 2006). Total phenolics were quantified as
225 indicators of plant allocation of resources to chemical defences. Phenolics were
226 extracted from leaf material with methanol 50% for 24 h under constant agitation at 4

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Deleted: The number of leaves of three-five randomly chosen shoots within each flowerpot and the leaf area index (LAI) were quantified as indicators of aboveground productivity. To estimate LAI, leaf area was measured on these three-five shoots, averaged, multiplied by the pot number of shoots and scaled per surface area

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236 °C and determined with a spectrophotometer using chlorogenic acid as standard
237 following a modified Folin-Ciocalteu method (Bolser et al., 1998).

238 **2.4 Community- and ecosystem-level traits**

239 The response of the seagrass community to CO₂ and nutrient enrichments was
240 quantified weekly by: (1) the percentage of flowerpot surface covered by *Ulva* spp.; (2)
241 the feeding activity of mesograzers (percentage of leaves showing bite marks in the
242 same shoots used to measure the morphological traits); and (3) the whole-community
243 metabolism quantified from the oxygen evolution within benthic chambers of 17 cm
244 diameter (4.8±0.01 L incubated) fitted to the flowerpots for 30-45 minutes at midday
245 (12-14 h). A transparent acrylic chamber to estimate net production and a dark chamber
246 to estimate respiration were simultaneously deployed within each mesocosm. Dissolved
247 oxygen concentration was measured by spectrophotometry using the Winkler method
248 (Labasque et al., 2004) in three water samples collected before and after incubations
249 into 12 mL soda glass vials. Community metabolism was estimated from the net change
250 in oxygen concentration during incubations integrated by the chamber volume and
251 standardized by incubation time and bottom area. There were no effects of enclosure on
252 the water temperature within chambers (measured with onset HOBO loggers, Southern
253 MA, USA). All incubations were run under irradiances of photosynthetically active
254 radiation (PAR) averaging 283±8.6 μmol quanta m⁻² s⁻¹ (measured with a Li-192SA
255 underwater PAR quantum sensor, Li-Cor, USA), when *Z. noltii* photosynthesis is light
256 saturated and not photoinhibited (Peralta et al., 2002).

257 At the ecosystem level, detritus production (fresh weight of all floating material
258 collected during a 24h period) was quantified almost every week. Organic matter in the
259 sediment (loss of dry weight after combustion at 450 °C, 4h) was measured at the end of
260 the experiment as indicator of the carbon sink capacity.

261 **2.5 Statistical analyses**

262 The effects of CO₂ and nutrient treatments throughout the experiment were tested using
263 three-way repeated-measures analyses of variance (RM ANOVA). The subject
264 repeatedly sampled was the mesocosm, CO₂ and nutrients were the among-subject
265 factors (two fixed crossed factors) and Time the within-subject factor. To avoid the
266 masking effect of the strong initial differences between meadows on the responses to

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268 experimental treatments, data for the low- and high-nutrient meadows were analyzed
269 separately. Data were checked for parametric assumptions and transformed where
270 needed. When sphericity was not met, corrected degrees of freedom from Greenhouse-
271 Geisser adjustment were used (Quinn and Keough, 2002).

272 The effects of CO₂ and nutrient treatments on variables measured at the end of the
273 experiment were tested using two-way ANOVAs (two fixed crossed factors) after
274 testing parametric assumptions. A normal distribution with unequal variances was found
275 for all variables, which is usual when the sample size is small. Following
276 recommendation by Quinn and Keough (2002), we proceeded with the analyses but
277 making significance level more restrictive to minimize the possibility of Type I error
278 (mistakenly detection of differences). Welch *t* tests that are robust against unequal
279 variances were used to interpret significant interactions. Again, data for the low- and
280 high-nutrient meadows were analyzed separately.

281 To assess the ordination of treatments based on differences in the composition of leaf
282 epiphyte assemblages, a non-metric Multi-Dimensional Scaling analysis (NMDS) with
283 Bray-Curtis distances was carried out. Because NMDS axes are arbitrary, the final
284 solution was rotated using a Principal Component Analysis (PCA) to align the largest
285 variance in the first axis. The significance of the effect of CO₂ and nutrient treatments
286 on assemblage composition was tested with a two-way permutational analysis of
287 variance (PERMANOVA; two fixed crossed factors). To perform the test, Bray-Curtis
288 distances were calculated from untransformed data and 999 permutations were used
289 under a reduced model.

290 Finally, two PCAs, one for each meadow, were performed to assess links among the
291 several traits and the trajectory of treatment responses through time. Traits showing the
292 highest correlation with the components ($r \geq 0.7$) were selected for interpretations.
293 Since our variables were not dimensionally homogeneous, principal components were
294 computed from the correlation matrix.

295

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Deleted: The effects of CO₂ and nutrient treatments on variables measured at the end of the experiment were tested using two-way ANOVAs (two fixed crossed factors) after checking parametric assumptions. *t* tests (Student for equal or Welch for unequal variances) were used to interpret significant interactions and to confirm ANOVA results due to unequal variances. When no significant interaction was detected, the main effect of a given factor was pooled over the levels of the other factor in the *t* tests ($n = 4$). When *t* tests did not confirm ANOVA results the significance level was more restrictive to minimize the possibility of Type I error (mistakenly detection of differences). Again, data for the low- and high-nutrient meadows were analyzed separately

312 3 Results

313 3.1 Responses of meadow traits measured through time

314 The response of the low-nutrient meadow showed a threshold at the third week of the
315 experiment, when most variables responded differently from the first two weeks (Fig. 1,
316 left). Shoot recruitment occurred mostly in unfertilized but also in CO₂-enriched
317 conditions until the third week (Fig. 1a), after which shoot mortality progressively
318 increased. Figure 1b suggests that the *Z. noltii* leaf area index (LAI) tended to increase
319 with CO₂ enrichment until the third week of the experiment. A positive, significant
320 effect of the CO₂ enrichment was observed on detritus production throughout the
321 experiment (Fig. 1d). Nutrient addition increased shoot mortality (Fig. 1a), whereas it
322 decreased LAI and leaf number (Fig. 1b and c). Shoot mortality induced by the nutrient
323 enrichment was attenuated by the simultaneous addition of CO₂, especially from the
324 third week onwards (Fig. 1a). A treatment and time interaction was detected on the
325 community production and respiration (Fig. 1e and f). These variables showed high
326 variability with similar ranges of variation in unfertilized and enriched conditions. No
327 treatment effects were detected throughout the experiment on shoot flowering or meso-
328 herbivory (see Appendix C, Supplement).

329 The responses of the high-nutrient meadow to CO₂ enrichment included an increased
330 shoot mortality during the second and third weeks and an increased detritus production
331 at the end of the experiment (Fig. 1g and i). Nutrient addition decreased the number of
332 leaves per shoot and increased detritus production throughout the experiment (Fig. 1h
333 and i). CO₂ enrichment interacted with nutrients to alleviate the nutrient-induced
334 reduction of the number of leaves (Fig. 1h). No effects of CO₂ or nutrient enrichment
335 were observed through time on LAI, meso-herbivory, *Ulva* spp. cover, shoot flowering,
336 and community production or respiration (Figs. 1j-l and C, Supplement). Independently
337 of the experimental treatments, overgrazing by the herbivorous amphipod *Cymadusa*
338 *filosa* Savigny severely affected the plants from the high-nutrient meadow causing
339 massive shoot mortality (Fig. 1k and g). At the end of the experiment a mean (\pm se) of
340 89 (\pm 3.7)% of shoots died, 81 (\pm 9.1)% of the seagrass leaves showed bite marks and
341 leaf area was reduced from 5.0 (\pm 0.2) to 1.0 (\pm 0.4) cm² shoot⁻¹. Similarly, *Ulva* spp.
342 cover progressively decreased, being close to 0% in all treatments after six weeks (Fig.
343 1l). At the end of the experiment, all *Z. noltii* shoots and *Ulva* spp. fronds disappeared

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346 from three flowerpots (one unfertilized and two CO₂-and-nutrient-enriched). However,
347 net production and respiration in these pots were within the range observed in the other
348 pots (Fig. Cd and e, Supplement), indicating that the metabolism of the sediment
349 microbial community was similar to that of the *Z. noltii* community.

350 We did not detect any significant effect of CO₂ or nutrient enrichment on plant damage
351 by meso-herbivory in the low- or high-nutrient meadows. However, at the end of the
352 experiment plants from the high-nutrient meadow showed 81 (+9.1)% of leaves with
353 bite marks, compared to only 6.9 (+3.2)% in the low-nutrient meadow (Figs. 1k and Cb,
354 Supplement). These between meadow differences, as well as the link between shoot
355 mortality and meso-herbivory in masking the enrichment effects in the high-nutrient
356 meadow, were further confirmed by a PCA of responsive variables from the low- and
357 high-nutrient meadows at the end of the experiment (see Appendix E, Supplement).

358 3.2 Responses of meadow traits measured at the end of the experiment

359 In plants from the low-nutrient meadow (Fig. 2, grey bars), nutrient enrichment
360 enhanced the leaf nutritional quality (high leaf nitrogen and low leaf C:N ratio, Fig. 2a
361 and c) and the accumulation of nitrogen in rhizomes (high rhizome nitrogen
362 concentration and low rhizome C:N ratio, Fig. 2d and b), whereas it had a negative
363 impact on the accumulation of leaf phenolics (Fig. 2f). A significant interaction of CO₂
364 and nutrient additions was detected for epiphyte load and sediment organic matter (Fig.
365 2g and h). The leaf epiphyte load increased significantly under CO₂ addition, whereas
366 nutrient enrichment and especially the interactive CO₂ and nutrient additions had a
367 lower and not significant effect (*t* test comparisons in Fig. 2g). Similarly, CO₂ and
368 nutrient interaction resulted in maintenance of the organic matter content in the
369 sediment, which tended to decrease with separated CO₂ and nutrient additions (*t* test
370 comparisons in Fig. 2h).

371 In plants from the high-nutrient meadow (Fig. 2, black bars), CO₂ enrichment decreased
372 rhizome C:N (Fig. 2b) and increased epiphyte loads (Fig. 2g). The CO₂-induced
373 increase of the epiphyte load was maintained under the simultaneous addition of
374 nutrients. Nutrient addition enhanced the leaf nutritional quality (high leaf nitrogen
375 concentration, Fig. 2a). A reduction of leaf C:N ratio and phenolics was detected
376 apparently in response to CO₂ and/or nutrient enrichments (Fig. 2c and f), but this was
377 actually caused by an increase of these traits in the unfertilized plants at the end of the

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385 experiment in relation to the initial field conditions (Table 1). A synergistic interaction
386 between CO₂ and nutrient additions caused an increase of the rhizome length (Fig. 2e).
387 Variables for which no significant effects of CO₂ or nutrient addition were detected are
388 shown in Appendix D (Supplement).

389 3.3 Responses of *Z. noltii* epiphytes

390 Both, CO₂ and nutrient additions altered the relative abundance of epiphyte populations,
391 whereas elevated nutrient levels also modified the epiphyte composition (Fig. 3a). In the
392 unfertilized plants, the epiphyte cover was low and the most abundant leaf epiphytes
393 were the fanlike diatoms *Licmophora* spp. The second-most-abundant epiphyte in plants
394 from the low-nutrient meadow was the encrusting coralline algae *Melobesia*
395 *membranacea*, whereas in plants from the high-nutrient meadow it was the
396 cyanobacterium *Microcoleus* spp. The response to the CO₂ enrichment in both, low- and
397 high-nutrient meadows was a great increase of epiphyte cover, mostly due to a bloom of
398 *Microcoleus* spp. (73% of the total cover) that outcompeted the diatoms *Licmophora*
399 spp. and the encrusting corallines. Under nutrient-enrichment pennate diatom
400 populations dominated by *Navicula* spp. outcompeted the other taxa. In the nutrient- and
401 CO₂-and-nutrient- treatments the composition of epiphyte assemblages was similar, but
402 with a reduced replacement of *Licmophora* spp. by pennate diatoms in the CO₂-and-
403 nutrient- treatment. Chlorophytes (mainly *Ulva prolifera*) and filamentous rhodophytes
404 (mainly *Bangia* spp. and *Stylonema alsidii*) were also present in all treatments.
405 Temporal changes in epiphyte abundances within the enriched mesocosms involved a
406 shift from relatively low epiphyte loads until the second week to increasing epiphyte
407 loads from the third week onwards, with the occurrence of purple bacteria in nutrient-
408 and CO₂-and-nutrient-enriched treatments during the fourth week.

409 NMDS ordination of treatments based on the epiphyte composition showed clear CO₂
410 effects (Fig. 3b). CO₂ treatments were separated along axis I (51% of variance
411 explained), whereas the other treatments were ordered along axis II (49% of variance
412 explained) from unfertilized to CO₂-, CO₂-and-nutrient-, and nutrient-enriched.
413 Separation of CO₂ enrichments along axis I was due to a higher epiphyte cover (mean
414 \pm se: 22 \pm 2.3 cm² per 10 cm² of leaf) than the unfertilized, nutrient-enriched and CO₂-
415 and-nutrient-enriched treatments (7.6 \pm 1.4, 11 \pm 1.3 and 8.4 \pm 3.2 cm² per 10 cm² of leaf,
416 respectively). Treatments of both, low- and high-nutrient meadows were nearby in the

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424 | ordination diagram, reflecting minor differences among meadows in the response of the
425 epiphyte assemblage. NMDS pattern was further confirmed by the PERMANOVA
426 results, which showed significant effects of CO₂, nutrients and their interaction (Fig.
427 3b).

428 The above-mentioned PCA of traits from the low- and high-nutrient meadows further
429 confirmed the increased epiphyte load and the change in epiphyte composition as main
430 drivers of the meadow responses to CO₂ and nutrient enrichments (see Appendix E,
431 Supplement).

432 **3.4 Response trajectories through time**

433 The first two PCA components of the low-nutrient meadow traits measured through
434 time, explained 41% (component I) and 20% (component II) of the variance. The *Z.*
435 *noltii* traits that highly correlated with component I were the LAI and the number of
436 leaves, which were negatively correlated with shoot mortality and herbivory (Fig. 4a,
437 right graph). Flowering, community production and community respiration highly
438 correlated with component II (variable loadings are presented in Table E2, Supplement).
439 The variability of all treatment scores on the component I during the first week and of
440 unfertilized and CO₂-enriched treatments during the second and third weeks were within
441 the initial range of natural variability (week 0, grey rectangle in Fig. 4a, left graph). At
442 this time, *Z. noltii* plants showed higher LAI and higher number of leaves. The time
443 series ordination of the rest of treatments along component I revealed that the effects of
444 nutrient addition started during the second week, when the scores of nutrient- and CO₂-
445 and-nutrient-treatments suddenly shifted to higher values. These nutrient effects were
446 dominated by high mortality of *Z. noltii* shoots and to a less extent by high meso-
447 herbivory. The highest scores on component I were attained by the nutrient-treatment
448 during the fourth to sixth weeks. The system response to the CO₂ treatment was slower
449 (starting at week 4) and of lower magnitude than the response to nutrient- and CO₂-and-
450 nutrient-treatments. This analysis supported the previous indication of a temporal
451 threshold for the meadow responses, which was the second week for elevated nutrients
452 and the fourth week for elevated CO₂. No clear ordination of treatments was detected
453 along component II, indicating that traits highly correlated with this component were
454 substantially influenced by natural variability.

456 The first two PCA components of the high-nutrient meadow traits measured through
457 time explained 64% (component I) and 12% (component II) of the variance. The *Z.*
458 *noltii* traits that highly correlated with component I were the LAI, the number of leaves,
459 the community production and the abundance of *Ulva*, which were negatively correlated
460 with mortality, herbivory and detritus production (Fig. 4b, right graph). Community
461 respiration highly correlated with component II (see variable loadings in Table E2,
462 Supplement). The range of initial natural variability of all treatment scores was
463 narrower than for the low-nutrient meadow (Fig. 4b, left graph). The system was
464 initially dominated by high seagrass LAI and number of leaves, cover of *Ulva* spp., and
465 community production. This progressively shifted to a later stage (week 6) dominated
466 by high *Z. noltii* mortality, herbivory and detritus production. Contrary to the response
467 of the low-nutrient meadow, there were no relevant differences in the time course and in
468 the final stage attained by PCA scores of both unfertilized and enriched treatments. The
469 shoot mortality of *Z. noltii* was positively correlated with meso-herbivore activity and
470 detritus production, and negatively correlated with LAI, number of leaves and *Ulva*
471 cover. No clear ordination of treatments was detected along component II.

472

473 4 Discussion

474 4.1 Effects of CO₂ enrichment in low-nutrient meadows

475 The CO₂ enrichment had no direct effects on *Z. noltii* biochemistry (Fig. 5a). We found
476 no evidence of increased nonstructural carbohydrates and subsequent nitrogen dilution
477 effect (increased C:N ratio) as has been previously observed in the seagrass *Thalassia*
478 *hemprichii* (Jiang et al., 2010) and *T. testudinum* (Campbell and Fourqurean, 2013). As
479 well, there was no increase of phenolic contents as predicted by the carbon-nutrient
480 balance hypothesis and no propagation to susceptibility to herbivory. Several studies in
481 terrestrial plants reveal that this lack of response is not uncommon (reviewed by
482 Peñuelas and Estiarte, 1998 and Bidart-Bouzat and Imeh-Nathaniel, 2008). The lack of
483 accumulation of carbohydrates and phenolics that we observed could be explained by
484 the trade-off between secondary metabolism and plant growth predicted by the growth-
485 differentiation balance hypothesis under no light and nutrient limitation (review by
486 Stamp, 2003). However, we found no significant increase of seagrass productivity under
487 CO₂ enrichment to support this trade-off, probably due to light limitation induced by

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Deleted: The response of the *Z. noltii* meadows to nutrients was faster and of higher magnitude than the response to CO₂ (Fig. 5a and b). Eutrophication and CO₂ caused significant shifts in primary producers' quality, which were mainly mediated by increased loads of diatoms or cyanobacteria epiphyte populations. Increased *Z. noltii* quality (high leaf nitrogen) was also a response to nutrient addition but not to CO₂ enrichment. In the meadow from low-nutrient field conditions, a decrease in sediment organic matter storage due to independent CO₂ and nutrient additions was evidenced at the ecosystem-level. Interestingly, all effects on epiphytes and sediment were attenuated when both nutrient and CO₂ acted simultaneously, as also occurred with the nutrient-induced shoot mortality. The higher plant quality induced by nutrients did not cascade up to an increase in herbivory, suggesting that the decrease of organic matter in the sediment was due to enhanced microbial activity. In the meadow from high-nutrient field conditions overgrazing by amphipods unrelated to CO₂ or nutrient enrichments drove, however, a striking seagrass decline. The connections between different organization levels assessed here can help to better understand the consequences of ocean acidification and eutrophication on ecosystems, as has been previously pointed out by other authors (Woodward et al., 2010).

516 epiphyte overgrowth from the third week of experiment onwards. Photosynthesis
517 enhancements have been reported under CO₂ addition in *Z. noltii* (Alexandre et al.,
518 2012) and *Z. marina* (Zimmerman et al. 1997), but they do not always translate into
519 seagrass growth since other determinant factors such as light and nutrient availability
520 are also at play (Palacios and Zimmerman, 2007; Alexandre et al., 2012; Campbell and
521 Fourqurean, 2013). In accordance with the previously cited terrestrial studies, our
522 results suggest that the seagrass responses to elevated CO₂ levels are highly context-
523 and species-specific, and are not as readily consistent and predictable as resource
524 availability hypotheses would suggest.

525 The most striking response of *Z. noltii* meadows to the CO₂ enrichment was the increase
526 of the epiphyte load, with changes in the relative abundance but not in the identity of the
527 main epiphyte taxa. Interestingly, epiphyte-induced shading did not cause seagrass
528 mortality as occurred under nutrient enrichment (see below). This suggests attenuation
529 of the negative effect of reduced light availability by increased CO₂ availability, which
530 may reduce the energy cost of carbon uptake (Koch et al., 2013). The epiphyte bloom
531 was mostly caused by the proliferation of the colonial and filament-forming
532 cyanobacterium *Microcoleus* spp. at the expense of a reduction of coralline algae crusts
533 of *Melobesia membranacea* and fanlike diatoms *Licmophora* spp. This is in accordance
534 with previous studies that showed elevated CO₂ / low pH to stimulate cyanobacteria
535 growth and photosynthesis (Liu et al., 2010) and to decrease abundance of coralline
536 algae (Hall-Spencer et al., 2008; Martin et al., 2008; Kuffner et al., 2008; Campbell and
537 Fourqurean, 2014). As well, Hervé et al. (2012) reported negative effects of low pH on
538 diatom valve formation and porosity, which were alleviated by a simultaneous nutrient
539 addition. We found that the activity of mesograzers was insufficient to regulate the
540 epiphyte proliferation in response to increased CO₂ levels, despite their recognized
541 capacity of controlling epiphyte biomass (Hughes et al., 2004) and particularly
542 cyanobacteria blooms (Neckles et al., 1993). Likely explanations are that the feeding
543 capacity of the most abundant mesograzer in the experiment, the amphipod *C. filosa*,
544 was exceeded by the cyanobacterium overgrowth or that the amphipod was not targeting
545 these particular epiphytes,

546 Epiphyte overgrowth resulted in increased detritus production and decreased organic
547 matter accumulated in the sediment. This suggests that bacterial decomposition in the
548 sediment was accelerated due to the highly labile organic matter of epiphytes, as

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Deleted: , with no significant changes on the carbon-rich phenolics and the total carbohydrate reserves. This result is in accordance with resource allocation hypotheses subsumed in the Growth-Differentiation Balance hypothesis (Stamp, 2003), which predicts that under no light and nutrient limitation carbon is allocated to growth instead to the production of secondary metabolites. In fact, we found that CO₂ addition tended to enhance *Z. noltii* leaf production (LAI) in the first three weeks of the experiment. However, the effect on leaf production disappeared after the third week when a massive proliferation of epiphytes reduced the light available for *Z. noltii* photosynthesis. Our results are in agreement with previous studies, which have reported that under no light limitation, nutrient deficiency rather than a direct effect of high CO₂ levels drives the accumulation of carbohydrates and phenolic compounds in both terrestrial plants (Lambers, 1993; Peñuelas and Estiarte, 1998) and seagrasses (Campbell et al., 2012). - ... [1]

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586 reported under nutrient enrichment (see below). The acceleration of bacterial
587 degradation of organic matter polysaccharides at low pH reported by Piontek et al.
588 (2010) would also support this explanation. Our findings may have relevant
589 implications, since the organic carbon produced in seagrass meadows sustains important
590 detritus-based food webs (Pergent et al., 1994; Moore and Fairweather, 2006) and
591 provides a major global carbon sink (Pidgeon, 2009; Fourqurean et al., 2012).

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592 4.2 Nutrient enrichment and interaction with CO₂ in low-nutrient 593 meadows

594 We found that nutrient enrichment had a faster and greater effect than CO₂ addition on
595 meadows developing in low-nutrient conditions. Nutrient enrichment enhanced leaf
596 nutritional quality (high nitrogen and low C:N ratio) and reduced the accumulation of
597 phenolic compounds (Fig. 5a). Both, the overall increase of plant nitrogen (e.g. Cabaço
598 et al., 2008; Invers et al., 2004) and the decrease of phenolics (e.g. van Katwijk et al.,
599 1997; Goecker et al., 2005) have been widely described in seagrasses as a result of
600 nutrient additions. The nitrogen increase was higher in leaves than in rhizomes, as
601 expected for this fast-growing species that acquires preferentially ammonium through
602 the leaves and shows minimal translocation of nitrogen to belowground tissues
603 (Alexandre et al., 2011).

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Deleted: Ammonium toxicity has been reported in *Z. noltii* (Brun et al., 2002) and other seagrass species (Santamaría et al., 1994, van Katwijk et al., 1997). To avoid toxicity, ammonium must be rapidly assimilated into amino acids at the expense of energy and carbon skeletons usually diverted from carbohydrate metabolism (Brun et al., 2002; Invers et al., 2004). In our experiment, ammonium toxicity was not apparent since nutrient addition had no significant effects on carbohydrate concentrations or plant survival. This finding concurs with previous studies, which showed that the effect of ammonium toxicity on carbohydrate reserves can be alleviated by high irradiance (van Katwijk et al., 1997) and simultaneous phosphate supply (Brun et al., 2008).

604 A reduction of the number of *Z. noltii* leaves and of leaf area index, and an increase of
605 shoot mortality were observed after the second week in response to nutrient addition.
606 These effects can be linked to ammonium toxicity, which has previously been reported
607 in *Z. noltii* (Brun et al., 2002) and other seagrass species (Santamaría et al., 1994, van
608 Katwijk et al., 1997). In addition, nutrient-induced changes in the epiphyte assemblage
609 may also contribute to shoot mortality by reducing light availability to seagrass leaves.
610 This seems supported by the abrupt increase of shoot mortality after the third week,
611 coinciding with the shift of the epiphyte assemblage from coralline algae and fanlike
612 diatoms to a dense layer of pennate diatoms (mostly of the genus *Navicula*), with a
613 purple bacteria biofilm developing as well during the fourth week. Towards the end of
614 the experiment, excess organic matter was released within the system due to increased
615 shoot mortality and epiphyte shifts. Coincidentally, the accumulation of organic carbon in
616 the sediments decreased, suggesting that an accelerated microbial decomposition was
617 promoted by the higher nutritional quality of producers as reported elsewhere for

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Deleted: A reduction of the number of *Z. noltii* leaves and of leaf area index, and an increase of shoot mortality were observed after the second week due to excess nutrients. Shoot mortality abruptly increased after the third week, coinciding with the shift of the epiphyte assemblage from coralline algae and fanlike diatoms to a dense layer of pennate diatoms (mostly of the genus *Navicula*). During the fourth week,

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655 terrestrial systems (Wardle et al., 2004) and seagrass beds (López et al., 1998; Holmer
656 et al., 2004; Spivak et al., 2007).

657 The simultaneous addition of CO₂ and nutrients did not modify the individual effects of
658 nutrient enrichment on plant biochemistry, but attenuated the proliferation of certain
659 epiphyte taxa (also occurring under high CO₂) and subsequent nutrient-induced *Z. noltii*
660 mortality. The interactive attenuation of epiphyte overgrowth may result from an
661 increase in the interspecific competition between the species that dominated the
662 epiphyte community under elevated CO₂ (i.e. the cyanobacterium *Microcoleus* spp.) and
663 under elevated nutrient levels (i.e. diatoms of the genus *Navicula*). Our findings are in
664 agreement with the negative effects of interspecific competition on the involved species
665 (i.e. symmetrical competition) that have long been reported in ecological studies
666 (Connel, 1983). Together, attenuation by simultaneous CO₂ and nutrient additions of the
667 overgrowth of certain epiphytes and of nutrient-induced *Z. noltii* mortality, reduced the
668 amount of more labile organic matter reaching the sediments compared to the individual
669 CO₂ or nutrient enrichment. This probably resulted in the maintenance at control levels
670 of the bacterial decomposition rates and of the sediment capacity to store organic
671 matter. To our knowledge, this is the first report of the interactive effect of CO₂ and
672 nutrient enrichments on the meadow carbon sink capacity. Overall, we found that under
673 simultaneous addition of CO₂ and nutrients, species interactions attenuated the direct
674 effects of individual stressors on *Z. noltii* and on sensitive epiphyte species or
675 taxonomic groups.

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Deleted: eutrophication effects

676 4.3 High- vs. low- nutrient meadows

677 Our results revealed that the expected benefits of high CO₂ predicted for the end of the
678 century on seagrass productivity might be restrained by epiphyte overgrowth and by the
679 interaction with local eutrophication. In both low- and high-nutrient meadows, CO₂
680 effects were more important in epiphyte populations than in the seagrass *Z. noltii*. These
681 findings strengthen the increasingly recognized importance of species interactions in
682 modulating the direct effects of eutrophication or acidification in single species,
683 populations, and ultimately in ecosystem functioning (Orth et al., 2006; Kroeker et al.,
684 2013b). The effect of nutrient enrichment was greater in the low- than in the high-
685 nutrient meadow (Fig. 5a vs. b), with nutrient-induced mortality of *Z. noltii* only
686 appearing in the former. When CO₂ and nutrient enrichments interacted, an increase of

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Deleted: reduced the nutrient-induced changes in the epiphyte assemblages and the CO₂-induced increase in epiphyte loads. This led to maintenance at control levels of the capacity of the sediments to store organic matter. Most studies of the interactive effect of CO₂ and nutrients in terrestrial systems are focused on nutrient levels that alleviate nutrient limitation and increase plant growth at the expense of carbon reserves as predicted by the Growth-Differentiation Balance hypothesis (Johnson and Lincoln, 1991; Coley et al., 2002). However, the potential alleviation by CO₂ of the plant mortality that may occur due to imbalances induced by excess nutrients has not, to our knowledge, been previously reported.

703 epiphyte load was observed in the high-nutrient meadow as opposed to the low-nutrient
704 meadow. These results highlight the context-dependence of the effects of multiple
705 stressors in agreement with the meta-analysis of Crain et al. (2008). We observed that
706 the accumulation of phenolics and carbohydrates was higher under lower nutrient
707 regimes in the initial field conditions, and also in the experimental conditions for
708 phenolics. This suggests that nutrient deficiency rather than a direct effect of high CO₂
709 drives the accretion of carbon-based compounds in *Z. noltii*. These observations are in
710 agreement with previous studies in both, terrestrial plants (Lambers, 1993; Peñuelas and
711 Estiarte, 1998) and seagrasses (Campbell et al., 2012), and reinforce the idea of the
712 context-dependence of seagrass responses to CO₂ enrichment,

713 Interestingly, we detected little evidence that CO₂ or nutrient addition affected seagrass
714 herbivory by mesograzers. However, clear differences in herbivory between meadows
715 were observed. Plants from the high-nutrient meadow experienced a 12-fold higher
716 amphipod grazing than plants from the low-nutrient meadow, which resulted in a
717 massive loss of shoots. Blooms of the amphipod *C. filosa* may occur in warmer months,
718 (Appadoo and Myers, 2004). This tube-building amphipod is widely distributed and
719 uses macroalgae for feeding (Ceh et al., 2005) and shelter (Appadoo and Myers, 2003).
720 To our knowledge, these observations constitute the first report of *C. filosa* using the
721 seagrass *Z. noltii* for both feeding and shelter-construction. We found that the
722 consequences of plant-specific vulnerability to grazing on seagrass meadows can be
723 stronger than the effects of CO₂ and eutrophication. This result concurs with findings by
724 Alsterberg et al. (2013), which showed that the presence of grazers masked the response
725 of benthic microalgae to ocean acidification and warming. Further studies aiming to
726 identify the factors underlying the plant-specific seagrass vulnerability to grazers are
727 thus of vital importance.

728 Our results showed that separated CO₂ or nutrient enrichment individually result in a
729 loss of the carbon sink capacity of the low-nutrient meadow, as opposed to the high-
730 nutrient meadow. This loss contrasts with results of previous studies conducted in situ
731 with other seagrass species, which found that the meadow carbon sequestration capacity
732 was unaffected by nutrient addition (Antón et al., 2011) or increased due to CO₂
733 enrichment (Russell et al., 2013). In both studies, the whole-community metabolism
734 was used as indicator of the carbon storage capacity of seagrass meadows. In our study,
735 however, the loss of carbon storage occurred without a significant response of the

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Deleted: Our findings revealed that the expected benefits of high CO₂ predicted for the end of the century on seagrass productivity can be restrained by epiphyte proliferation and by the interaction with local eutrophication. The most striking responses of *Z. noltii* meadows to the CO₂ enrichment were an increase in the total epiphyte load and changes in the relative abundance, but not in the identity, of the main epiphyte taxa. In both low- and high-nutrient meadows, CO₂ effects were more important in epiphyte populations than in the seagrass *Z. noltii*. The effect of nutrient enrichment was higher in the low-nutrient than in the high-nutrient meadow (Fig. 5a vs. b). In both meadows, elevated nutrient levels altered leaf nitrogen concentration and epiphyte composition in the same way, but contrary to the low-nutrient meadow, there was no significant increase of the shoot mortality in the high-nutrient meadow. When CO₂ and nutrient enrichments interacted, the increased epiphyte load induced by elevated CO₂ levels was maintained in the high-nutrient meadow. In addition, there was an increase of the *Z. noltii* belowground productivity (longer rhizomes) that decreased the rhizome C:N ratio. The investment in vegetative growth of belowground rhizomes for colonizing adjacent areas is coherent with the Growth-Differentiation Balance hypothesis in the absence of nutrient limitation. This response has been reported for this species in relation to sedimentary (Cabaço and Santos, 2007) and hydrodynamic disturbances (Peralta et al., 2005), and under a 1-year CO₂ enrichment for *Zostera marina* (Palacios and Zimmerman, 2007). In the high-nutrient meadow, we found a reduction of leaf C:N ratios and phenolics, apparently in response to CO₂ and/or nutrient enrichments. However, this was actually due to a drastic increase in the C:N ratio and phenolic concentration of the unfertilized plants (Fig. 2c and f) in relation to the original field conditions (Table 1). This, together with the low phenolics observed in the field for the high-nutrient meadow support that nutrient deficiency was driving phenolic accumulation as previously pointed out for the low-nutrient meadow.

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785 whole-community metabolism, indicating a metabolic compensation between the *Z.*
786 *noltii* populations and the leaf epiphyte and sediment microbial communities. A similar
787 dynamic global balance has also been reported for marine pelagic systems under CO₂
788 addition (Silyakova et al., 2013).

789 Overall, we found that shifts in the community dynamics of leaf epiphytes or sediment
790 bacteria mediated the multi-level responses of *Z. noltii* meadows to independent CO₂ or
791 nutrient addition. They also modulated the attenuation of individual effects under
792 simultaneous CO₂ and nutrient enrichments. Overgrazing masked the response to CO₂
793 enrichment and eutrophication but only in the high-nutrient meadow. Our findings
794 highlight the importance of integrative multi-level and ecosystem-based approaches
795 considering not only species interactions and connections between organization levels,
796 but also the effect of interactive stressors, to anticipate the evolution of seagrass
797 meadows in the near future and to endorse conservation efforts.

798

799 **Appendices**

800 Appendix A. Full comparative description of the low- and high-nutrient meadows of
801 *Zostera noltii* in June-August 2011, prior to the start of the experiment.

802 Appendix B. Seawater chemistry within the experimental mesocosms.

803 Appendix C. Full results of the response to CO₂ and nutrient additions of *Zostera noltii*
804 plant-, community-, and ecosystem-level traits measured through time.

805 Appendix D. Full results of the response to CO₂ and nutrient additions of *Zostera noltii*
806 plant-, community-, and ecosystem-level traits measured at the end of the experiment.

807 Appendix E. Results of principal component analyses.

808

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814 onset; J. Reis, P. Frias and M. Viegas for support during the mesocosm experiment; M.

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Deleted: Our study revealed that CO₂ or nutrient enrichments may lead to a loss of the carbon sink capacity of low-nutrient meadows. These results are contradictory with studies by Antón et al. (2011) and Russell et al. (2013), who found no effect and an increase in the meadow carbon sequestration capacity of other seagrass species due to in situ nutrient and CO₂ enrichments, respectively. They used the whole-community metabolism as indicator of the carbon storage capacity of seagrass meadows. However, in our study, the loss of carbon storage occurred with no response of the whole-community metabolism, which showed high variability. Our results suggest a dynamic balance of the community metabolism in response to high CO₂ and nutrients, in which the changes in the *Z. noltii* metabolism are compensated by the metabolism of fast-growing leaf epiphyte and sediment microbial communities. A similar dynamic global balance has also been reported for marine pelagic systems under CO₂ addition (Silyakova et al., 2013).

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Deleted: Overall, the role of herbivory in driving differences between low- and high-nutrient meadows, as well as the role of epiphyte and microbial communities in mediating meadow responses to CO₂ and nutrients, illustrated the importance of interactions among species and connections between organization levels in ecosystem-based studies. The alleviation of nutrient effects by high CO₂ highlighted the relevance of incorporating responses to multiple stressors. This integrative ecosystem-based approach is an important challenge to endorse future conservation efforts.

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854 SFRH/BPD/71129/2010, respectively).
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1099 Table 1. Environmental and *Zostera noltii* community traits that revealed significant
 1100 differences between the low- and high-nutrient donor meadows during June-August
 1101 2011, prior to the start of the experiment, tested using unpaired t-tests. ^aSqrt-
 1102 transformed data to meet normality. ^bMann-Whitney rank tests were conducted for
 1103 variables that did not meet normality even after transformation. All measured traits and
 1104 methods are shown in Appendix A (Supplement).

	Low-nutrient meadow	High-nutrient meadow
Leaf phenolics (mg (gDW) ⁻¹)	48±1.2	29±3.2
Leaf nitrogen (mg gDW ⁻¹)	21±0.8	25±0.6
Leaf C : N	19±0.7	16±0.4
Rhizome starch (mgGlu (gDW) ⁻¹)	473±14	355±27
Rhizome TNC (mg Glu g (DW) ⁻¹)	668±18	532±28
Shoot area (cm ² shoot ⁻¹) ^a	7.8±0.6	4.5±0.7
<i>Z. noltii</i> density (shoots m ⁻²)	5517±755	2664±411
<i>Z. noltii</i> cover (% of sediment surface) ^b	96±2.2	18±9.8
<i>Ulva</i> spp. cover (% of sediment surface) ^b	absent	38±12
Seawater nitrate (μM) ^b	< 0.01	1.1±0.2
Seawater ammonium (μM)	0.7±0.2	3.0±0.4
Seawater phosphate (μM)	0.5±0.1	1.2±0.1

1105

1106 **Figure captions**

1107 Fig. 1. Effects of CO₂ and nutrient additions on *Zostera noltii* plant-, community-, and
1108 ecosystem-level traits from the low-nutrient (left graphs) and high-nutrient meadow
1109 (right graphs) through time. Symbols are means (\pm se, $n = 2$). *F* statistics and *p* levels
1110 from RM ANOVA tests are shown for among-subject factors (CO₂; Nut: nutrients) and
1111 their interaction (CO₂ x Nut), and for within-subject factor (time) and interactions (Time
1112 x CO₂, Time x Nut, Time x CO₂ x Nut). Only the significant effects ($p < 0.05$) and
1113 useful traits for results interpretation are shown here; the non-significant effects are
1114 shown in Appendix C (Supplement). *Variables that did not meet normality after
1115 transformation, for which the significance level was more restrictive ($p < 0.03$) to
1116 minimize the possibility of Type I error. **Variable sqrt-transformed to meet normality.

1117 Fig. 2. Effects of CO₂ and nutrient additions on *Zostera noltii* plant-, community-, and
1118 ecosystem-level traits measured at the end of the experiment from the low-nutrient (grey
1119 bars) and high-nutrient meadow (black bars). Bars are means (\pm se, $n = 2$). The *F*
1120 statistics and *p* levels from two-way ANOVA tests are shown for each fixed crossed
1121 factor (CO₂; Nut: nutrients) or interaction (CO₂ x Nut). Lowercase letters above bars
1122 show significant differences between treatments for significant interactions (*t* tests, see
1123 Methods). Only ~~restrictive~~ significant effects ($p < 0.03$) ~~selected to minimize the~~
1124 ~~possibility of Type I error due to unequal variances~~ are shown here; the non-significant
1125 effects are shown on Appendix D (Supplement).

1126 Fig. 3. Responses of *Zostera noltii* epiphyte populations to experimental addition of
1127 CO₂ and nutrients: (a) Relative abundance of the main epiphyte taxa of plants exposed
1128 to experimental treatments; and (b) NMDS ordination of experimental treatments (see
1129 symbol legend below treatment names in Fig. 3a) based on leaf epiphytes of plants from
1130 the low-nutrient (L; grey symbols) and high-nutrient meadow (H; black symbols).
1131 Pseudo-*F* statistics and *p* levels from two-way PERMANOVA test are shown in the
1132 NMDS diagram for each fixed crossed factor (CO₂; Nut: nutrients) and their interaction
1133 (CO₂ x Nut).

1134 Fig. 4. Principal components analysis of *Zostera noltii* plant-, community, and
1135 ecosystem- level responses to treatments through time: (a) low-nutrient and (b) high-
1136 nutrient meadow. Numbers inside the symbols indicate sampling weeks from 0 to 6.
1137 The initial variability (week 0) along component I is incorporated within a grey

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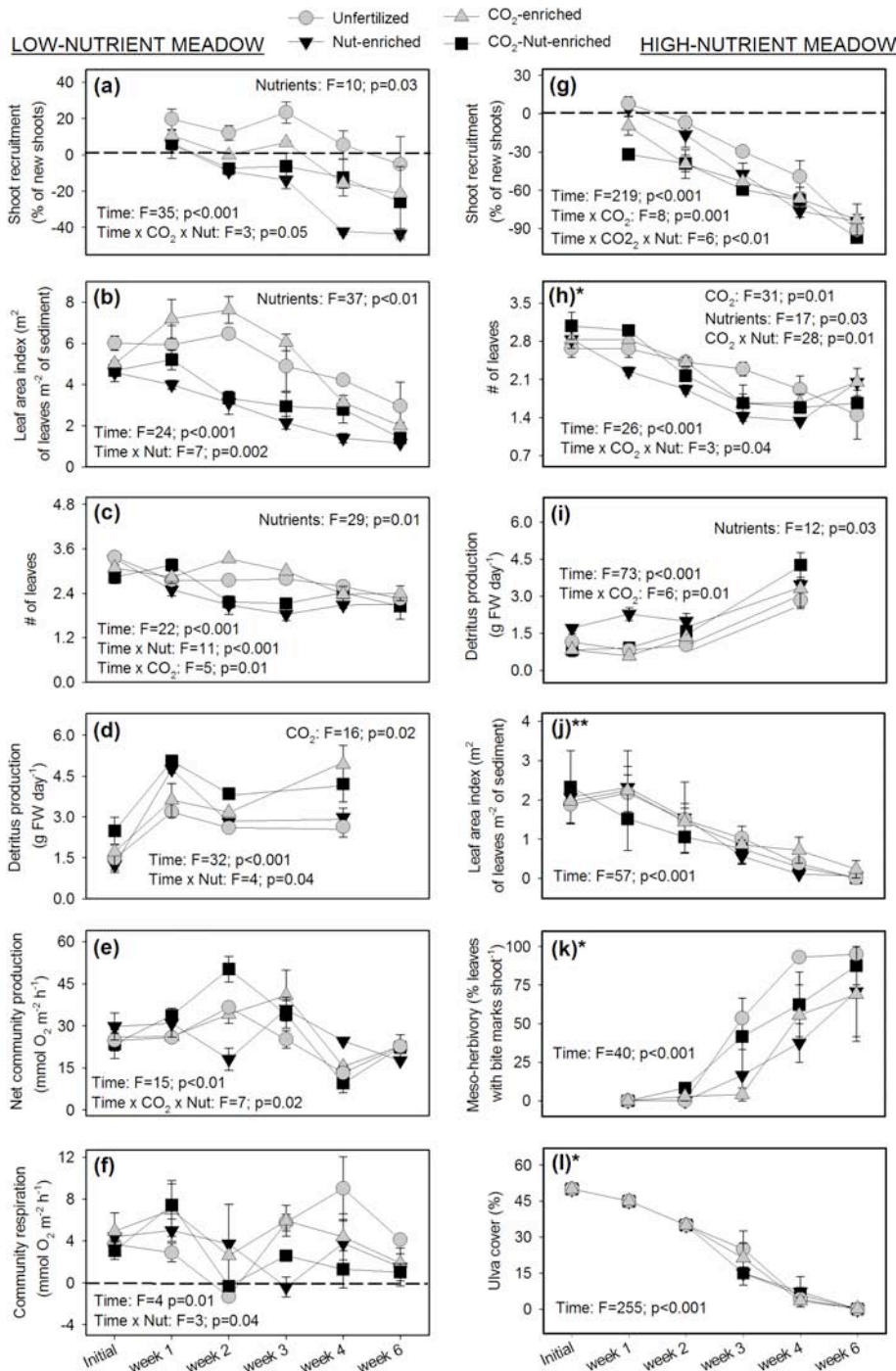
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1144 rectangle to highlight the range of initial natural variability. Variable loadings on the
1145 two principal components are depicted in right graphs. LAI refers to leaf area index,
1146 NCP to net community production and CR to community respiration.

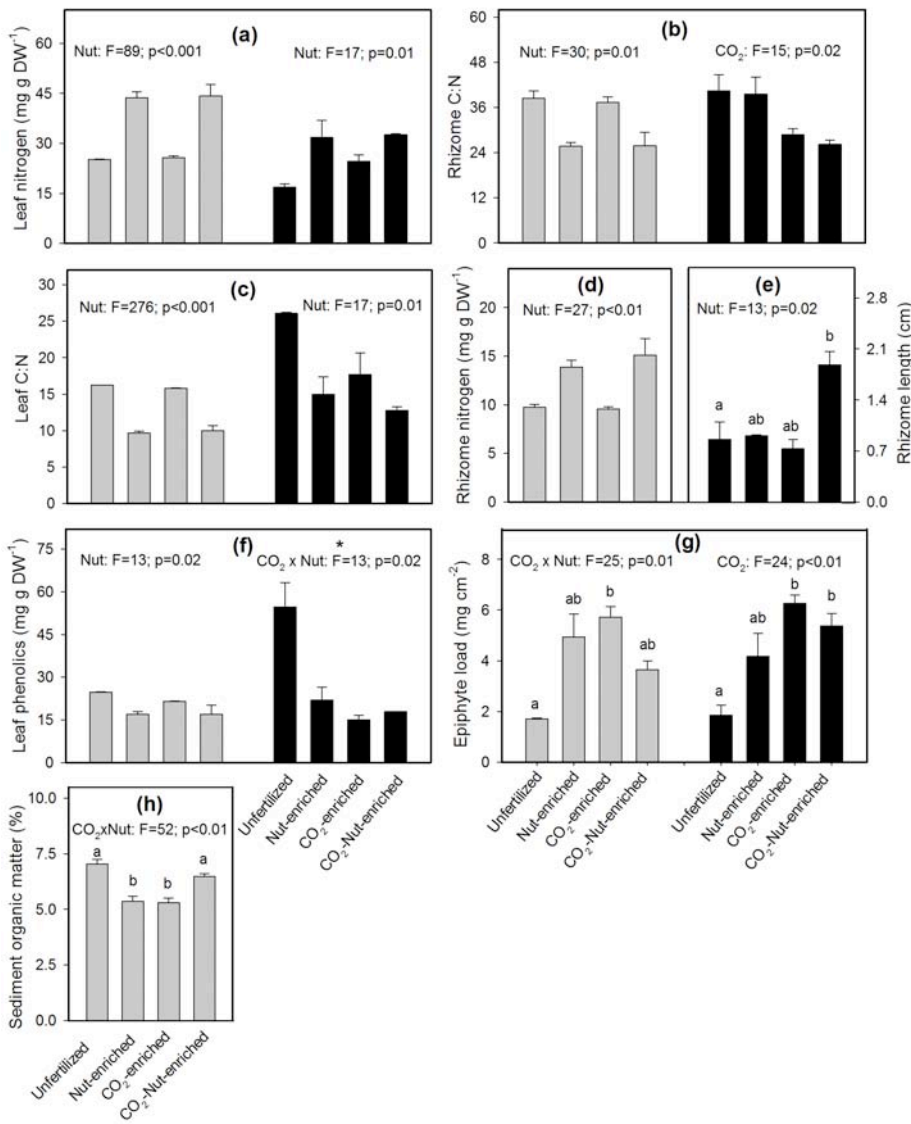
1147 Fig. 5. Summary of the effects of CO₂ (blue line) and nutrient (green line) additions, and
1148 when significant of their interaction (red line), on *Zostera noltii* plant- community and
1149 ecosystem- level traits of low-nutrient (a) and high-nutrient (b) meadows. Solid lines
1150 indicate significant effects on variables measured at the end of the experiment and
1151 dashed lines on variables measured through time. Dotted lines indicate no significant
1152 effects. Letters below the x-axis denote unfertilized (U) and enriched (E) treatment,
1153 which is the pooled mean response to the respective enrichment over the other when
1154 there was no significant interaction (no red line). For shoot mortality and net community
1155 production (NCP) in the low-nutrient meadow a significant Time x CO₂ x Nutrients
1156 interaction was detected and values were represented as for CO₂ x Nutrients significant
1157 interactions. * Apparent effect due to a drastic increase in unfertilized plants in relation
1158 to the initial field conditions (see results).
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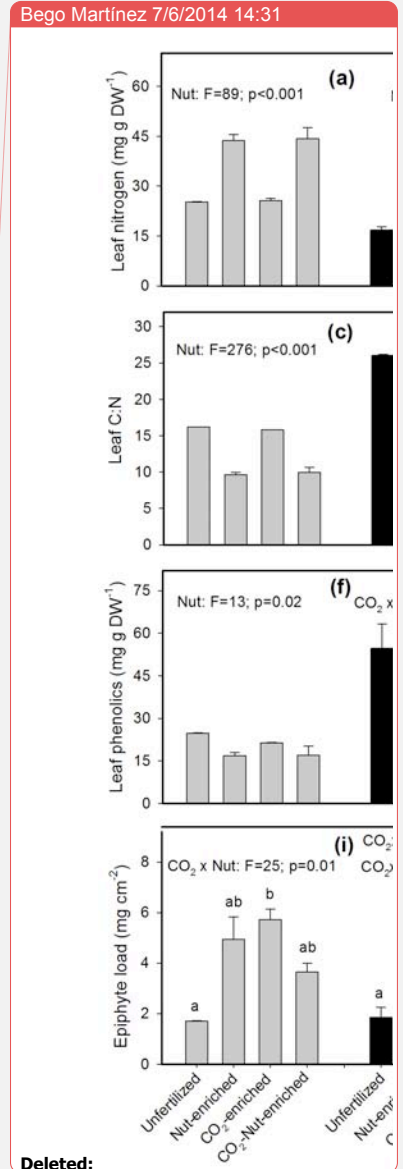
1161 Figure 1

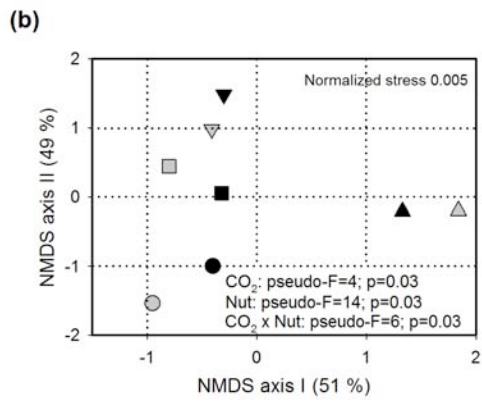
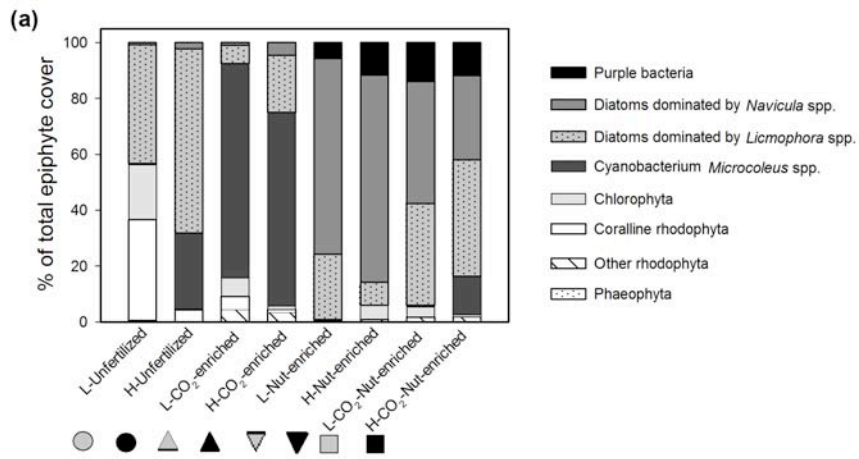
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1165 Figure 2

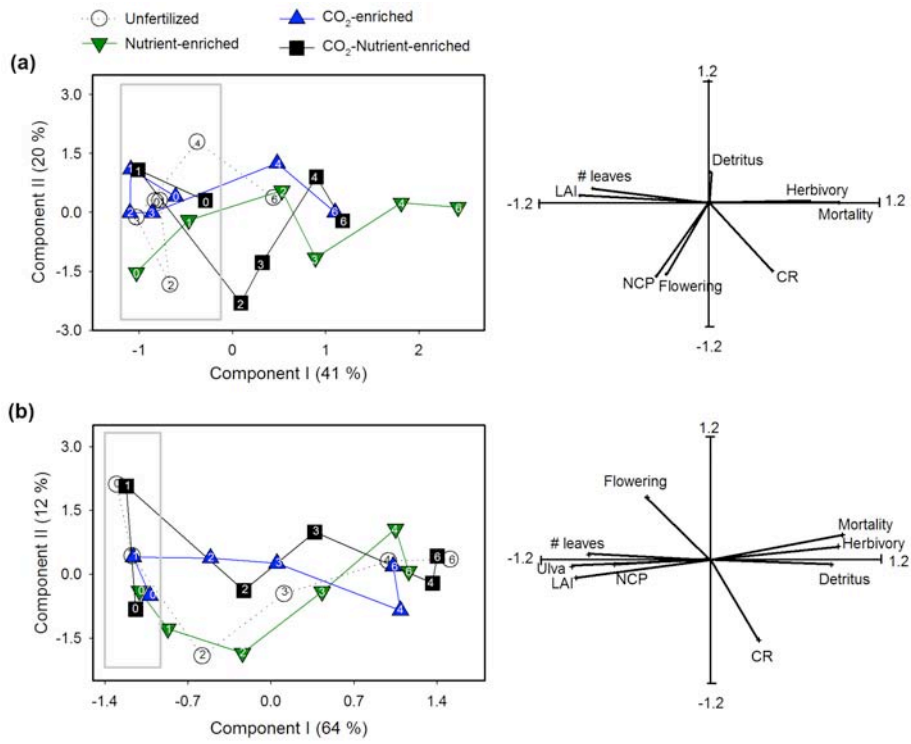




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1168 Figure 3

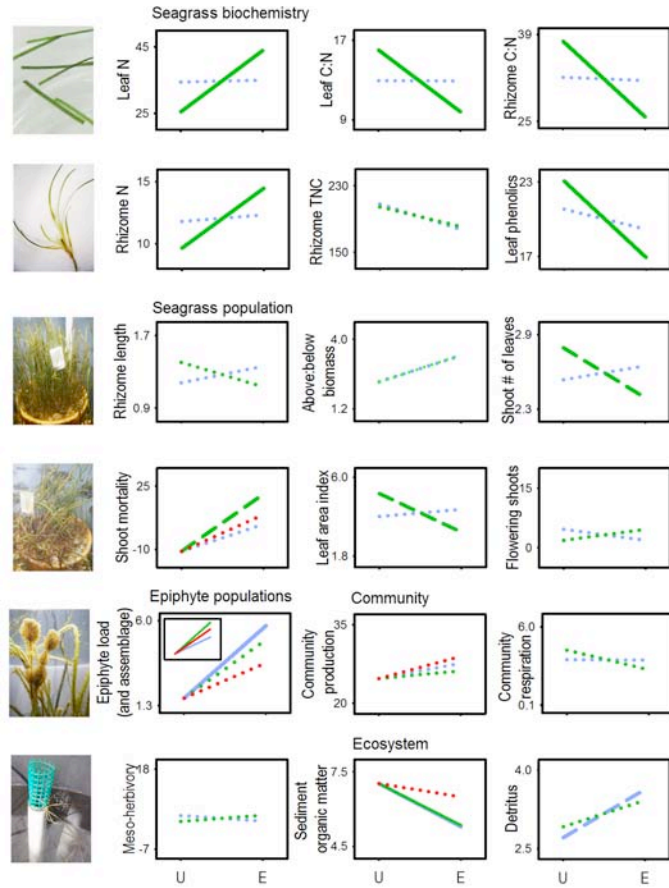
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1171 Figure 4.

(a) Low-nutrient meadow



(b) High-nutrient meadow

