

Nitrate source identification in the Baltic Sea using its isotopic ratios in combination with a Bayesian isotope mixing model

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Abstract

Nitrate (NO_3^-) is the major nutrient responsible for coastal eutrophication worldwide and its production is related to intensive food production and fossil-fuel combustion. In the Baltic Sea NO_3^- inputs have increased four-fold over the last decades and now remain constantly high. NO_3^- source identification is therefore an important consideration in environmental management strategies. In this study focusing on the Baltic Sea, we used a method to estimate the proportional contributions of NO_3^- from atmospheric deposition, N_2 fixation, and runoff from pristine soils as well as from agricultural land. Our approach combines data on the dual isotopes of NO_3^- ($\delta^{15}\text{N}-\text{NO}_3^-$ and $\delta^{18}\text{O}-\text{NO}_3^-$) in winter surface waters with a Bayesian isotope mixing model (Stable Isotope Analysis in R, SIAR). Based on data gathered from 47 sampling locations over the entire Baltic Sea, the majority of the NO_3^- in the southern Baltic was shown to derive from runoff from agricultural land (33–100%), whereas in the northern Baltic, i.e., the Gulf of Bothnia, NO_3^- originates from nitrification in pristine soils (34–100%). Atmospheric deposition accounts for only a small percentage of NO_3^- levels in the Baltic Sea, except for contributions from northern rivers, where the levels of atmospheric NO_3^- are higher. An additional important source in the central Baltic Sea is N_2 fixation by diazotrophs, which contributes 49–65% of the overall NO_3^- pool at this site. The results obtained with this method are in good agreement with source estimates based upon $\delta^{15}\text{N}$ values in sediments and

1 a three-dimensional ecosystem model, ERGOM. We suggest that this approach can be easily
2 modified to determine NO_3^- sources in other marginal seas or larger near-coastal areas where
3 NO_3^- is abundant in winter surface waters when fractionation processes are minor.

4 5 **1 Introduction**

6 Throughout the world, anthropogenic reactive N currently exceeds natural production
7 (Galloway et al., 2003; Gruber and Galloway, 2008). Consequently, riverine nitrogen (N)
8 fluxes have doubled in recent years, which has strongly impacted the marine N cycle and
9 ecosystem health, both at regional and global scales. In coastal ecosystems, the adverse
10 effects of these excess N loads include eutrophication, hypoxia, loss of biodiversity, and
11 habitat destruction (Galloway et al., 2003; Villnäs et al., 2013). For the shallow, brackish,
12 semi-enclosed Baltic Sea, where intense anthropogenic nutrient loadings have been
13 documented since the 1950s (Elmgren, 2001), riverine and atmospheric nutrient inputs are
14 now at least four-fold higher than a century ago, when anthropogenic influence was low
15 (Schernewski and Neumann, 2005; Stålnacke et al., 1999). Furthermore, cyanobacterial
16 blooms, which can fix N_2 , and thus add nutrients to the surface waters are regular large scale
17 phenomenon each summer (Finni et al., 2001; Vahtera et al., 2007) and the overall increase in
18 nutrient input has supported the expansion of hypoxic zones (Conley et al., 2009, 2011).

19 A main component of the N pool and the one most readily available is nitrate (NO_3^-) (Nestler
20 et al., 2011; Vitousek et al., 1997), which derives from a wide variety of sources. These can
21 be identified by analysis of the N and oxygen (O) isotopes ($\delta^{15}\text{N}-\text{NO}_3^-$ and $\delta^{18}\text{O}-\text{NO}_3^-$) since
22 the isotopic ratios of NO_3^- from different sources fall within distinct ranges (Kendall, 1998;
23 Kendall et al., 2007). For example, NO_3^- inputs from forested catchments can be
24 discriminated from those coming from agricultural runoff, and the NO_3^- signature of microbial
25 nitrification differs from that of atmospheric deposition (Kendall, 1998; Kendall et al., 2007;
26 Mayer et al., 2002). Source attribution is, however, complicated by N-transformation
27 processes such as denitrification, nitrification, and assimilation, each of which gives rise to
28 significant isotope fractionation. Since heavier isotopes are sequestered more slowly than
29 lighter ones, the reaction product will be isotopically depleted compared to the original NO_3^-
30 source (Kendall, 1998). Alterations of isotope values because of microbial fractionation
31 processes can be minimized by collecting the samples in winter, when low water temperatures
32 reduce microbial activity (Pfenning and McMahon, 1997).

1 Nonetheless, source attribution is still complicated when there are more than three sources but
2 only two isotopes that describe them (Fry, 2013). SIAR (Stable Isotope Analysis in R), a
3 Bayesian isotope mixing model originally developed to infer diet composition from the stable
4 isotope analysis of samples taken from consumers and their food sources (Moore and
5 Semmens, 2008), was already successfully applied for NO_3^- source identification. Xue et al.
6 (2012, 2013) were able to estimate the proportional contributions of five potential NO_3^-
7 sources in a small watershed in Flanders (Belgium). Based on their determinations of the
8 isotopes of nitrogen and oxygen they could show that manure and sewage were the major
9 sources of NO_3^- .

10 In the Baltic Sea the NO_3^- pool present in the surface waters in spring originates from the
11 previous growth season and is consumed during the onset of the phytoplankton spring bloom,
12 in February/March. Stratification in summer hinders circulation down to the halocline, thus
13 atmospheric deposition and N_2 fixation are the major N sources, whereas in coastal areas
14 riverine discharge dominates (Radtke et al., 2012; Voss et al., 2011). Yet, to what extent the
15 various NO_3^- sources add to the overall pool of NO_3^- in the Baltic as a whole is still a matter
16 of debate. In this study, a source attribution for four major sources is presented. Taking the
17 Baltic Sea as an example we will show, that the use of the isotopic composition of NO_3^-
18 ($\delta^{15}\text{N}-\text{NO}_3^-$ and $\delta^{18}\text{O}-\text{NO}_3^-$) in combination with SIAR can be used elsewhere for source
19 identification on an ecosystem scale level.

20

21 **2 Material and Methods**

22 **2.1 Field sampling**

23 Surface water samples from the Baltic Sea were collected in February 2008 (n=22) and 2009
24 (n=17) before the onset of the phytoplankton spring bloom aboard the RV *Alkor* and in
25 November 2011 (n=1) aboard the RV *Meteor* using a Seabird CTD system with attached
26 water bottles. Samples from the Nemunas River (55°18'5.5 N, 21°22'53.9 E; 55°41'25.6 N,
27 21°7'58.4 E; n=4) and Kalix River (65°56'4.2 N, 22°53'9.2 E; n=1) (Fig.1) were taken
28 between November 2009 and February 2010. Values for NO_3^- in which atmospheric
29 deposition was the source were obtained from wet deposition samples collected at three
30 stations around the Baltic Sea: Warnemünde, Germany (54°10' N, 12°5' E,); Majstre, Sweden
31 (57°30' N, 18°31' E); and Sännen, Sweden (56°13'N, 15°17'E) from December 2009 until
32 February 2010 (Table 1). In Warnemünde, precipitation was collected on an event basis, and
33 retrieved daily to limit microbial degradation, using a sampler consisting of a plastic funnel

1 (diameter: 24 cm) connected to a 1-L polyethylene bottle. At the two Swedish stations,
2 rainwater was sampled monthly by the Swedish Environmental Research Institute (IVL) as
3 part of the Swedish national long-term monitoring program. Here, the sampler consisted of a
4 plastic funnel (diameter 20.3 cm) connected to an 8-L polyethylene bag. All samples were
5 filtered through pre-combusted Whatman GF/F filters (4 h at 400°C) and stored frozen until
6 further analysis.

7

8 **2.2 Nutrient concentrations and dual isotope analysis of NO₃⁻**

9 Samples were analyzed following a standard protocol for the determination of NO₃⁻ and nitrite
10 (NO₂⁻) (Grasshoff et al., 1983); the precision of the method is ±0.02 μmol l⁻¹. Dual isotope
11 analysis of NO₃⁻ (δ¹⁵N-NO₃⁻ and δ¹⁸O-NO₃⁻) was carried out using the denitrifier method
12 (Casciotti et al., 2002; Sigman et al., 2001), in which NO₃⁻ and NO₂⁻ are quantitatively
13 converted to nitrous oxide (N₂O) by *Pseudomonas aureofaciens* (ATTC 13985), a bacterial
14 strain that lacks N₂O reductase activity. In brief, N₂O is removed from the sample vials by
15 purging with helium and then concentrated and purified in a GasBench II prior to analysis
16 with a Delta Plus mass spectrometer (ThermoFinnigan). NO₂⁻ was not removed since its
17 concentrations were always less than 2% (referring to the procedure described in Casciotti et
18 al., 2007). N and O isotope measurements of roughly 30% of the samples were replicated in
19 separate batch analyses. Two international standards, IAEA-N3 (δ¹⁵N=4.7‰ vs. N₂; δ¹⁸O
20 25.6‰ vs. VSMOW) and USGS 34 (δ¹⁵N -1.8‰ vs. N₂; δ¹⁸O -27.9‰ vs. VSMOW) (Böhlke
21 et al., 2003), were measured with each batch of samples. Samples with NO₃⁻/ NO₂⁻
22 concentrations as low as 1 μmol l⁻¹ were analyzed. The sample size for the actual stable
23 isotope measurements was 20 nmol for samples with concentrations >3.5 μmol l⁻¹ and 10
24 nmol for those with concentrations <3.5 μmol l⁻¹. Isotope values were corrected after Sigman
25 et al. (2009) for δ¹⁸O-NO₃⁻; single point correction was referred to IAEA-N3 for δ¹⁵N-NO₃⁻.
26 The precision was <0.2 ‰ for δ¹⁵N and <0.6 ‰ for δ¹⁸O. Together with the samples, a culture
27 blank was analyzed to which no sample was added. The isotope ratios are reported using the
28 delta notation in units of per mil (‰).

29

30 **2.3 NO₃⁻ sources**

31 To estimate the contribution of different NO₃⁻ sources, two isotopes δ¹⁵N-NO₃⁻ and δ¹⁸O-NO₃⁻
32 (j=2) from the four major NO₃⁻ sources: (1) atmospheric deposition, (2) runoff from pristine
33 soils, (3) runoff from agricultural land and (4) N₂ fixation were applied (Table 2). In this

1 context, N_2 fixation was defined as NO_3^- originating from the degradation and remineralization
2 of nitrogen fixers and therefore carried their low isotopic signal. Thus, for NO_3^- from N_2
3 fixation, $\delta^{15}N$ values of ~ -2 to 0‰ were assumed, since N_2 fixation produces organic material
4 that is only slightly N depleted against air nitrogen (Carpenter et al., 1999, 1997; Montoya et
5 al., 2002). The $\delta^{18}O$ values were estimated to be between -3.8‰ and 2.0‰ , based on
6 measurements in the subtropical northeast Atlantic where N_2 fixation was the main source of
7 N (Bourbonnais et al., 2009) ($\delta^{18}O\text{-}NO_3^- = 2\text{‰}$) and the estimated $\delta^{18}O$ of NO_3^- deriving from
8 N_2 fixation by Sigman et al. (2009) ($\delta^{18}O\text{-}NO_3^- = -0.2\text{‰}$) and Bourbonnais et al. (2012) ($\delta^{18}O\text{-}$
9 $NO_3^- = -3.8\text{‰}$).

10 To expand the dataset, we included NO_3^- isotope data from river water samples, ground water
11 samples, and samples from tile drain outlets collected in 2003 and published in Deutsch et al.
12 (2006). In that study, the Warnow River ($n=2$) was sampled twice, in January and February
13 2003. These sources were likewise sampled in winter, since marked seasonal shifts in the
14 isotopic composition of NO_3^- can occur due to shifts in the origins of the sources (Knapp et
15 al., 2005). Samples from tile drain outlets were used to represent NO_3^- from agricultural
16 runoff and were obtained from the catchment of the Warnow River, whose waters are strongly
17 influenced by agricultural land use (Pagenkopf, 2001). High $\delta^{15}N\text{-}NO_3^-$ values of $9.9\pm 1.5\text{‰}$
18 and lower $\delta^{18}O\text{-}NO_3^-$ values of $4.6\pm 1.0\text{‰}$ are typical for areas that are influenced by
19 agricultural activities and are similar to studies of Wankel et al. (2006) and Johannsen et al.
20 (2008). Johannsen et al. (2008) found in the rivers Rhine, Elbe, Weser and Ems, with
21 comparable high agricultural activities, $\delta^{15}N\text{-}NO_3^-$ values between 8.2 and 11.2‰ and $\delta^{18}O\text{-}$
22 NO_3^- values from 0.4 to 0.9‰ in winter. However, a differentiation between NO_3^- from
23 mineral fertilizers and sewage/manure was not done; rather a mixed signal from rivers that are
24 mainly influenced by agricultural activities was taken. Groundwater samples were used as the
25 source of NO_3^- from pristine land (Deutsch et al., 2006). Their $\delta^{15}N\text{-}NO_3^-$ and $\delta^{18}O\text{-}NO_3^-$
26 values significantly differed from those of agricultural runoff ($p<0.05$) but were similar to the
27 values of other areas, such as Biscuit Brook (Burns et al., 2009) and the San River (Koszelnik
28 and Gruca-Rokosz, 2013), where pristine soils were sampled and reflect nitrification activity
29 in soils unaffected by human activity.

30 The dual isotopes of NO_3^- values presented in Deutsch et al. (2006) were analyzed according
31 to Silva et al., (2000). In this method, NO_3^- is chemically converted via anion exchange resins
32 to $AgNO_3^-$ and the $\delta^{15}N\text{-}NO_3^-$ and $\delta^{18}O\text{-}NO_3^-$ values are measured via pyrolysis and isotopic
33 ratio mass spectrometry (for a detailed description, see Deutsch et al. 2006). A normal

1 distribution of the isotopic data from the four sources was confirmed by applying the Shapiro-
 2 Wilk normality test. $\delta^{15}\text{N-NO}_3^-$ and $\delta^{18}\text{O-NO}_3^-$ values from NO_3^- from atmospheric deposition
 3 of $0.3\pm 1.4\text{‰}$ and $76.7\pm 6.8\text{‰}$, respectively, are also in line with literature values. The $\delta^{15}\text{N}$
 4 values of atmospheric NO_3^- are usually between -15 to +15‰ and the $\delta^{18}\text{O}$ between 63 and
 5 94‰ (Kendall et al., 2007).

6 Six regions within the catchment of the Baltic Sea were investigated for their potential NO_3^-
 7 sources (Fig. 1). According to the topography of the Baltic Sea, the samples were assigned to
 8 four major areas: Western Baltic Sea, Baltic Proper, Gulf of Finland, and Gulf of Bothnia.
 9 Additionally, three rivers differing in their degree of anthropogenic impact were included in
 10 this study and divided into two groups: northern and southern rivers. Rivers with high nutrient
 11 loads drain mainly into the southern Baltic Proper and were represented here by the Nemunas
 12 and Warnow Rivers, whose NO_3^- concentrations in winter can be as high as $260 \mu\text{mol l}^{-1}$
 13 (Deutsch et al, 2006, Pilkaityte and Razinkovas 2006). The Gulf of Bothnia receives large
 14 amounts of fresh water from rivers represented by the Kalix River. These rivers drain mainly
 15 pristine, forested land and have maximum NO_3^- concentrations of around $20 \mu\text{mol l}^{-1}$
 16 (Sferratore et al, 2008).

17

18 2.4 SIAR mixing model

19 The applied mixing model is described by the following equations:

$$20 X_{ij} = \sum_{k=1}^K p_k (s_{jk} + c_{jk}) + \varepsilon_{ij} \quad (1)$$

$$21 s_{jk} \sim \text{N}(\mu_{jk}, \omega_{jk}^2) \quad (2)$$

$$22 c_{jk} \sim \text{N}(\lambda_{jk}, \tau_{jk}^2) \quad (3)$$

$$23 \varepsilon_{ij} \sim \text{N}(0, \sigma_j^2) \quad (4)$$

24 where X_{ij} is the observed isotope value j of the mixture i ; $i=1,2,3,\dots,I$ are individual
 25 observations; and $j=1,2,3,\dots,J$ are isotopes. s_{jk} is the source value k of isotope j ($k = 1,2,3,\dots,$
 26 K) and is normally distributed, with a mean of μ_{jk} and a standard deviation of ω_{jk} . p_k is the
 27 proportion of source k that needs to be estimated by the model. c_{jk} is the fractionation factor
 28 for isotope j on source k and is normally distributed, with a mean of λ_{jk} and a standard
 29 deviation of τ_{jk} . ε_{ij} is the residual error representing additional unquantified variations between
 30 mixtures and is normally distributed, with a mean of 0 and a standard deviation of σ_j . Detailed
 31 descriptions of the model can be found in Jackson et al. (2009), Moore and Semmens (2008),
 32 and Parnell et al. (2010). As noted above, by collecting samples between November and

1 February we minimized the influence of fractionation processes such as assimilation and
2 denitrification that can alter the isotopic signal of NO_3^- . Therefore in Eq. (1) we assumed that
3 $c_{jk} = 0$.

4 Two different runs of the SIAR model were performed. In the first, for the Western Baltic
5 Sea, Baltic Proper, and Gulf of Finland, all four sources were included in the calculation. In
6 the second, for the Gulf of Bothnia, the southern rivers, and the northern rivers, N_2 fixation as
7 a potential NO_3^- source was excluded since in these areas there is no N_2 fixation by
8 diazotrophs because the Gulf of Bothnia is phosphorus limited, in contrast to the Baltic Proper
9 (Graneli et al., 1990).

10

11 **3 Results**

12 **3.1 NO_3^- concentrations and isotopes**

13 Winter (Nov.-Feb.) surface NO_3^- concentrations ranged from a minimum of $2.6 \mu\text{mol l}^{-1}$ in the
14 open Baltic Sea to a maximum of $259 \mu\text{mol l}^{-1}$ close to the estuaries of the most nutrient-rich
15 rivers, i.e., the Nemunas and Warnow Rivers (Fig. 2, Supplement 1). In most basins of the
16 Baltic Sea, the NO_3^- concentrations in winter were almost identical, with the exception of the
17 Gulf of Finland, where concentrations were about two-fold higher ($7.6 \pm 0.9 \mu\text{mol l}^{-1}$; Fig. 2).
18 In the western Baltic Sea, the Baltic Proper, and the Gulf of Bothnia NO_3^- concentrations were
19 similar with 3.3 ± 0.6 , 3.4 ± 0.8 , and $3.7 \pm 0.4 \mu\text{mol l}^{-1}$, respectively.

20 Highest nitrate concentrations in the Nemunas River also corresponded to the highest $\delta^{15}\text{N}$ -
21 NO_3^- with 10.0 ‰ and vice versa, with lowest concentrations and nitrogen isotope values in
22 the Baltic Proper (1.5 ‰). The $\delta^{18}\text{O}$ - NO_3^- values ranged from -2.8 ‰ in the Gulf of Bothnia
23 to 10.6 ‰ in the Northern River, Kalix (Fig. 2, Supplement 1).

24

25 **3.2 Sources of NO_3^-**

26 SIAR calculated that in the southern Baltic Sea, agricultural runoff was the main NO_3^- source
27 with the highest contribution in the western Baltic Sea with up to 67 % (mean $53.5 \pm 3.2\%$) and
28 in the southern rivers with up to 100% (mean $93.5 \pm 4.1\%$) (Table 3, Fig. 1). NO_3^- from
29 atmospheric deposition was negligible with 3.5% (mean $1.1 \pm 0.5\%$) and NO_3^- from pristine
30 soils lower with up to 42% (mean $7.5 \pm 5.9\%$) in the western Baltic Sea (Table 3, Fig. 1). In the
31 Baltic Proper, NO_3^- from N_2 fixation was the dominant NO_3^- source with up to 65.3% (mean
32 $58.8 \pm 2.0\%$) (Table 3, Fig. 1). In the northern Baltic Sea NO_3^- from atmospheric deposition is
33 only important in the northern rivers with a contribution of up to 23.4% (mean $11.8 \pm 1.5\%$)

1 (Table 3, Fig. 1). NO_3^- from pristine soils is mainly transported by the northern rivers
2 ($75.3 \pm 7.9\%$) to the Gulf of Bothnia where SIAR calculated that $99.0 \pm 0.9\%$ stems from the
3 runoff from pristine soils (Table 3, Fig. 1).

4

5 **3.3 Comparison of isotope patterns in the water column and sediments**

6 The $\delta^{15}\text{N}$ values from surface water correlated significantly with those from surface
7 sediments, as reported in Voss et al. (2005) ($p < 0.001$) (Fig. 3). Stations for sediment sampling
8 were in close vicinity to stations from water column sampling (Fig 4). In the Baltic Proper,
9 the $\delta^{15}\text{N}$ of the surface water NO_3^- was indistinguishable from the $\delta^{15}\text{N}$ of the sediment
10 surface ($3.6 \pm 1.0\text{‰}$ and $3.5 \pm 0.6\text{‰}$, respectively; Table 4). In the near-coastal areas of the
11 Baltic Proper and the Gulf of Finland, the $\delta^{15}\text{N}$ of surface water NO_3^- was $7.9 \pm 1.8\text{‰}$, slightly
12 higher than the surface sediment value for the same area of $7.3 \pm 2.1\text{‰}$ (data in Voss et al.,
13 2005) but still not significant different ($p < 0.01$) (Table 4).

14

15 **4 Discussion**

16 **4.1 NO_3^- in the Baltic Sea**

17 The measured winter surface water concentrations of up to $259 \mu\text{mol l}^{-1}$ are typical for
18 eutrophied systems and similar values have been reported from the Chesapeake Bay and the
19 coastal areas of the North Sea (Dähnke et al., 2010; Francis et al., 2013). The concentrations
20 of nutrients in the sub-basins of the Baltic Sea reflect the densities of the human populations
21 in the vicinity of the adjacent sub-catchments. Thus, in the near-coastal area of the southern
22 Baltic Proper, NO_3^- concentrations were higher than in the northern parts, since the catchment
23 areas of Germany, Poland, and the Baltic States are much more densely populated (>500
24 inhabitants km^{-2}) and the land is intensively used for agricultural purposes. The northern
25 regions are dominated by boreal forests and less populated (<10 inhabitants km^{-2}) (Lääne et
26 al., 2005; Stepanauskas et al., 2002; Voss et al., 2011). Consequently, for the southern Baltic
27 Proper a relationship between fluvial NO_3^- loads and NO_3^- concentrations in coastal waters
28 could be established that indicates a direct impact of riverine nutrients on coastal waters (Voss
29 et al. 2011, HELCOM, 2009). However, there was no similar correlation between riverine N
30 loads and nutrient concentrations either for the coastal areas of the Gulf of Bothnia or for the
31 open waters of the Baltic Proper (Voss et al. 2011). The Gulf of Bothnia is the only sub-basin
32 in which the effects of eutrophication are so far minor, although Lundberg et al. (2009) and
33 Conley et al. (2011) reported a degradation in the water quality from north to south and from

1 the outer to the inner coastal area of the Gulf, with seasonal hypoxia at many sites. Trends of
2 increasing nutrient levels should be interpreted as a warning signal for the future and highlight
3 the need for management approaches based on sound knowledge of the many potential
4 sources of NO_3^- .

5 In the Gulf of Finland, which is regarded as the most heavily eutrophied sub-basin of the
6 Baltic Sea, a consequence of high receiving nutrient loads from the Neva River and the city of
7 St. Petersburg (Lundberg et al. 2005), NO_3^- concentrations were about two-fold higher
8 ($7.6 \pm 0.9 \mu\text{mol l}^{-1}$) compared to the rest of the Baltic Sea sub-basins, where concentrations in
9 winter were almost identical. This shows that NO_3^- concentrations alone cannot be used to
10 identify NO_3^- sources for the sub-basins; rather, stable NO_3^- isotopes values allow for accurate
11 source determination, as we will show in the following sections.

12

13 **4.2 Sources of NO_3^-**

14 The use of NO_3^- stable isotopes for source identification is complicated when the mixing of
15 multiple N sources with overlapping isotopic ranges occurs together with microbial processes
16 such as nitrification, assimilation, and denitrification (Kendall, 1998; Wankel et al., 2006). In
17 this study, we assumed that the effects of fractionation by microbial processes were negligible
18 because all our samples were collected in winter, at a mean temperature of $3.1 \pm 1.3 \text{ }^\circ\text{C}$ (data
19 not shown), when microbial activity is low (Pfenning and McMahon, 1997), as confirmed in a
20 study of nitrification in the Baltic Sea by Jäntti et al. (2011). They showed that in the Gulf of
21 Finland although nitrification potentials may be high during cold months, in situ nitrification
22 is undetectable whereas the rate increases progressively towards the summer.

23 We are aware that the variability of the source signals must be taken into account in source
24 attributions. Both Xue et al. (2012, 2013) and Yang et al. (2013) showed that SIAR can be
25 applied in NO_3^- source identification, although the resolution of this model is largely
26 determined by the uncertainty of the isotopic composition of the sources. In the studies of
27 both groups, the means and variances of the sources were calculated mostly from literature
28 values, which were not obtained in the investigation areas, nevertheless they received
29 consistent results. In contrast, in our study, the isotopic composition of the sources, except
30 NO_3^- from N_2 fixation, was determined from samples obtained within the study area. In our
31 calculations we considered the impact of the variability of the sources and report not only
32 mean values and error estimates but also minimum and maximum contributions, as suggested
33 by Fry (2013) (Table 3).

1

2 **4.2.1 NO₃⁻ from agricultural runoff**

3 The isotopic values of riverine NO₃⁻ were previously shown to be enriched when agricultural
4 land is the source of inputs (Johannsen et al., 2008; Mayer et al., 2002; Voss et al., 2006).
5 Catchments with high percentages of agricultural and/or urban land use export NO₃⁻ with
6 δ¹⁵N-NO₃⁻ values of around 7‰. In the same study, the oxygen isotope ratios of NO₃⁻ were
7 almost uniformly 13±1‰ (Mayer et al. 2002). Johannsen et al. (2008) measured δ¹⁵N-NO₃⁻
8 values of 11.3‰ in highly eutrophied rivers draining into the North Sea, whereas the highest
9 δ¹⁸O-NO₃⁻ value was 2.2‰. In the Oder River outflow, a main NO₃⁻ contributor to the Baltic
10 Sea, δ¹⁵N-NO₃⁻ of 7.6‰ and δ¹⁸O-NO₃⁻ of 2.9‰ were determined (Korth et al., 2013). Our
11 measurements for the Warnow and Nemunas Rivers fall in the expected range, with a mean
12 δ¹⁵N-NO₃⁻ of 9.2‰ and a mean δ¹⁸O-NO₃⁻ of 3.1‰, and are consistent with the high
13 percentages of agricultural land in the river catchment areas: 50% for the Warnow River
14 (Pagenkopf, 2001) and 50% for the Nemunas River (Christoph Humborg, personal
15 communication, 2011). For both, SIAR calculations indicated that 75.2–100% (mean
16 93.5±4.2%) of the NO₃⁻ pool is from agricultural runoff. NO₃⁻ with this signature seems to be
17 transported to the central Baltic Sea, since SIAR-based estimates showed significant
18 percentages of agriculturally derived NO₃⁻ in the Western Baltic Sea (41.0–66.5%; mean:
19 53.5±3.2%), the Baltic Proper (32.8–45.5%; mean: 39.0±1.6%), and the Gulf of Finland
20 (40.9–63.4%; mean: 51.9±3.0%). However, high percentages were only expected for the Gulf
21 of Finland and the Western Baltic Sea, where large N loads from agricultural land have been
22 documented (Hong et al., 2012). Indeed, for the Baltic Proper, the sizeable contribution of
23 agricultural NO₃⁻ (39.0±1.6%) was surprising and contrasted with previous findings that
24 nearly excluded riverine NO₃⁻ as a major nutrient source for the central Baltic Sea (Voss et al.,
25 2005, 2011). However, Neuman (2000) estimated that 13% of the N input of the Oder River is
26 transported to the central Baltic Sea, while Radtke et al. (2012) could show, using a source
27 attribution technique in the three-dimensional ecosystem model ERGOM (Ecological
28 ReGional Ocean Model), that at least a part of the dissolved inorganic nitrogen (DIN) load
29 from the Vistula River, the main NO₃⁻ contributor to the Baltic Sea (Wulff et al., 2009), enters
30 the Baltic Proper. This 3D model comprises a circulation model, a thermodynamic ice model,
31 and a biogeochemical model and utilizes the Modular Ocean Model, MOM3.1 (Radtke et al.,
32 2012).

1 Another explanation for the high estimated agricultural influence in our study could be the
2 intrusion of water containing NO_3^- with similar NO_3^- isotope values as our agricultural NO_3^-
3 source during mixing/advection from below the halocline. Deep-water NO_3^- in the Baltic Sea
4 has a $\delta^{15}\text{N}$ of about 7‰ (Frey et al. unpubl. data), which is higher than the average deep-water
5 ocean NO_3^- signature of 5‰ (Sigman et al., 2000). This elevated $\delta^{15}\text{N}$ in NO_3^- mainly comes
6 from water column denitrification in the oxic-anoxic interface in water at a depth of about 100
7 m (Dalsgaard et al., 2013). However, the year-to-year variations in DIN due to vertical mixing
8 and advection from below the halocline are sensitive to hydrographic conditions. When the
9 halocline is weak and well ventilated, oxygen conditions improve, resulting in higher DIN
10 concentrations in deep waters and greater advection and/or mixing (Vahtera et al, 2007) such
11 that the NO_3^- contribution from below the halocline is difficult to estimate.

12 Overall, the range of 32.8–45.5% (mean: $39.0 \pm 1.6\%$) determined for NO_3^- presumably
13 originating from agricultural runoff has to be considered with caution, because the former
14 imprint of deep water column denitrification and mixing/advection of this isotopically
15 enriched NO_3^- from below the halocline with the residual winter surface NO_3^- pool could have
16 resulted in an overestimation of the percentage of NO_3^- from agricultural runoff in the Baltic
17 Proper.

18

19 **4.2.2 NO_3^- from N_2 fixation**

20 The average $\delta^{15}\text{N}$ - NO_3^- value of 3.6‰ for the Baltic Proper is slightly lower than the ocean
21 average of around 5‰ (Sigman et al., 2000) and presumably reflects the influence of N_2
22 fixation. This is because the $\delta^{15}\text{N}$ of newly fixed N is between -2 and 0‰ such that NO_3^- has
23 slightly lower $\delta^{15}\text{N}$ values (Knapp et al., 2005; Liu et al., 1996). The $\delta^{18}\text{O}$ - NO_3^- value of -
24 0.5‰ in the Baltic Proper is also slightly lower than the ocean average of 1.5‰, and close to
25 our theoretical considered value of $-0.7 \pm 2.9\%$ after the degradation and remineralization of
26 N_2 fixers.

27 N_2 fixers are abundant in summer, reflecting the stimulation of their growth by the low N/P
28 ratios. N in the cyanobacterial biomass is remineralized over the winter months and the
29 resulting NO_3^- remains in the water masses down to the halocline. Our results show that the
30 contribution of N_2 fixation by diazotrophs to the NO_3^- pool is 49.3–65.3% (mean $58.8 \pm 2.0\%$).
31 This is slightly higher compared to the data reported by Wasmund et al. (2001), who
32 estimated that 39% ($370 \text{ ktons yr}^{-1}$) of a total input of 955 ktons N yr^{-1} (HELCOM, 2002)
33 stems from N_2 fixations in the central Baltic Sea. Both Radtke et al. (2012) and Voss et al.

1 (2005) concluded that N_2 fixation was the main NO_3^- source in the Baltic Proper. Using an
2 independent approach, we were able to confirm the contribution of N_2 fixation in this area. In
3 addition, we found that N_2 fixation is also a major source of NO_3^- in the Western Baltic Sea
4 and the Gulf of Finland (respectively, 11.0–51.9% (mean $37.9 \pm 5.1\%$) and 32.7–59.0% (mean
5 $45.5 \pm 3.2\%$)). This finding is consistent with our current understanding of N_2 fixation in the
6 Gulf of Finland (Vahtera et al, 2005) whereas the western Baltic Sea is rather perceived as an
7 area with no N_2 Fixation activity (Stal et al., 2003). In summary, our results provide important
8 evidence that N_2 fixation by cyanobacteria is a significant N source not only in the Baltic
9 Proper but also in the Western Baltic Sea and Gulf of Finland.

10

11 **4.2.3 NO_3^- from atmospheric deposition**

12 NO_3^- from atmospheric deposition is generally heavily enriched in ^{18}O ($>60\%$) because of
13 reactions involving ozone (O_3), which is anomalously enriched in heavy oxygen isotopes
14 (Durka et al., 1994; Kendall et al., 2007). This is consistent with the $\delta^{18}\text{O}$ measurements at the
15 three stations around the Baltic Sea, where the averaged isotope value in winter was 77%
16 (Table 1).

17 Our results show that direct inputs of atmospheric deposition contribute less NO_3^- than all
18 other sources. Indeed, among all basins of the Baltic Sea, that has a total area of 415.266 km^2 ,
19 the maximum mean contribution was in the Western Baltic Sea 0 to 3.5% (mean $1.1 \pm 0.5\%$).
20 Moreover, using a dataset from Michaels et al. (1993), Duce et al. (2008) estimated that even
21 an extremely rare and large atmospheric deposition event distributed over a 25-m mixed-layer
22 depth would increase the reactive N concentration only by around $0.045 \mu\text{mol l}^{-1}$. A study in
23 the Kattegat estimated an input of 52 kt N yr^{-1} from atmospheric deposition, which implied
24 rather limited nutritional support for phytoplankton (Spokes et al., 2006). Taking into account
25 that in the Baltic Proper, with an area of 211.069 km^2 , in winter the mixed-layer depth is 80–
26 100 m and that the residual NO_3^- pool, with a concentration of $3.6 \mu\text{mol l}^{-1}$, has a $\delta^{18}\text{O}-\text{NO}_3^-$ of
27 -0.5% , a similar rain event with a $\delta^{18}\text{O}$ of 76.7% (Table 1) would increase the $\delta^{18}\text{O}-\text{NO}_3^-$ of
28 the residual NO_3^- pool only by 0.2–0.3%, which is within our analytical error. Even though
29 several rain events typically occur during winter, their influence seems to be too low to leave
30 a detectable isotopic imprint. Additionally, the NO_3^- from atmospheric deposition is
31 presumably intensively cycled through the organic N pool in spring and summer such that
32 after several mineralization cycles its origin is difficult to recognize isotopically (Mayer et al.,
33 2002).

1 In the Kalix River $\delta^{18}\text{O}\text{-NO}_3^-$ was clearly enriched (10.6‰) compared to the values
2 determined for the Baltic Sea. We calculated that in this river up to 23.4% (mean $11.8\pm 1.5\%$)
3 of the NO_3^- originates from atmospheric deposition. Mayer et al. (2002) compared the isotopic
4 NO_3^- signature of 16 watersheds in the USA and were able to show that riverine NO_3^- derived
5 from atmospheric NO_3^- deposition and not from nitrification in soils is the dominant N input
6 in predominantly forested watersheds, when riverine NO_3^- concentrations are generally low.
7 Therefore only in the Kalix River, where up to 97% of the catchment with a size of 18.130
8 km^2 is covered by forests and NO_3^- concentrations are low during winter (Voss et al., 2011),
9 was the imprint of NO_3^- from atmospheric deposition visible; by contrast, in the southern
10 Baltic Sea and the rivers draining into it, the anthropogenic influence due to agriculture is
11 very high and therefore masks atmospheric contributions. However, NO_3^- loads to the
12 northern Baltic Sea from the Kalix River and other, similar boreal rivers are small, comprising
13 only about 20% of the sea's total N load (Voss et al., 2011). Thus, overall, we assume that
14 atmospheric deposition is a very minor source of NO_3^- in the Baltic Sea.

15

16 **4.2.4 NO_3^- from pristine soils**

17 In general, in rivers such as the Kalix River, whose catchments include pristine vegetation,
18 $\delta^{15}\text{N}\text{-NO}_3^-$ values are low while those of $\delta^{18}\text{O}\text{-NO}_3^-$ are high (Voss et al., 2006). This finding
19 was confirmed in the present study, in which $\delta^{15}\text{N}\text{-NO}_3^-$ and $\delta^{18}\text{O}\text{-NO}_3^-$ values of 1.6‰ and
20 10.6‰, respectively, were determined. In the Kalix River, the NO_3^- contribution from the
21 runoff of pristine soils as determined by SIAR is 33.8–92.8% (mean $75.3\pm 7.9\%$). In pristine
22 soils the isotopic NO_3^- signal is mainly derived from nitrification, which is in agreement with
23 previous studies of small catchments, where much of the NO_3^- was shown to be of microbial
24 origin (Campbell et al., 2002; Kendall et al., 2007; Mayer et al., 2002). Similar $\delta^{15}\text{N}\text{-NO}_3^-$
25 values were reported for areas where pristine soils were also sampled. For example, $\delta^{15}\text{N}\text{-NO}_3^-$
26 NO_3^- and $\delta^{18}\text{O}\text{-NO}_3^-$ values of 1.9 and 2.8‰ were determined for Biscuit Bay (Burns et al.,
27 2009) and 2.9 and 2.8‰ for the San River (Koszelnik and Gruca-Rokosz, 2013), respectively.
28 The higher $\delta^{18}\text{O}\text{-NO}_3^-$ values of the Kalix River can, as discussed above, be attributed to
29 atmospheric deposition.

30 For the Gulf of Bothnia, where the catchment is dominated by pristine areas like forests
31 (50%) and shrubs (20%), NO_3^- from pristine soils contributes 91.7–100% ($99.0\pm 0.9\%$).
32 However, for the Baltic Proper the NO_3^- contribution from pristine soils is negligible, because
33 the NO_3^- derived from nitrification is very low in concentrations and remains in the Gulf

1 because of the cyclonic circulation in the Bothnian Sea and Bothnian Bay (Humborg et al.,
2 2003) and the high residence time of the water (7.4 years) which results in a rather slow
3 exchange with the rest of the Baltic Sea (Myrberg and Andrejev, 2006).

4 5 **4.3 Comparison of isotope patterns in the water column and sediments**

6 Correlations between $\delta^{15}\text{N}$ values from the water column and surface sediment is a common
7 feature in coastal basins, like Cariaco Basin (Thunell et al., 2004), Guaymas Basin, Monterey
8 Bay, and San Pedro Basin (Altabet et al., 1999). This occurs when NO_3^- in the surface mixed
9 layer is fully consumed, which is the case in the Baltic Proper during the spring bloom, when
10 the only significant loss comes from the sinking of particulate nitrogen (Altabet et al., 1999).
11 Moreover, high organic matter preservation seems to stimulate the similarity in the $\delta^{15}\text{N}$ in the
12 surface water and sediments as seen in other depositional environments (Thunell et al., 2004).

13 Overall, the comparison with the sediment data set from Voss et al. (2005) shows that the
14 isotopic signature of NO_3^- in the euphotic layer of the Baltic Sea is directly transferred to the
15 particulate organic nitrogen pool and is subsequently found in the sediment surface as
16 detritus, thus conserving information about the origin of this NO_3^- source. Additionally, we
17 could show how consistent the nitrogen input to the sediments is over the years. Even though,
18 our surface water samples were sampled from 2008 to 2011, the surface sediment samples
19 from 1993 to 2003 and deposited in the period of approximately 10 years before collection,
20 the comparison of the $\delta^{15}\text{N}$ values showed that there is no significant difference. Coastal areas
21 preserve the isotope signature of riverine sources while the open Baltic Sea sediments indeed
22 mirror the nitrogen input dominated by N_2 fixation. Moreover the data demonstrate that no
23 change over time in the input of NO_3^- sources has occurred.

24 25 **5 Conclusions**

26 By combining dual isotope data of winter NO_3^- ($\delta^{15}\text{N}\text{-NO}_3^-$ and $\delta^{18}\text{O}\text{-NO}_3^-$) in surface waters
27 with a Bayesian isotope mixing model (SIAR), we estimated the contribution of four major
28 NO_3^- sources for the different basins of the Baltic Sea. A clear shift in the source of NO_3^-
29 inputs, from agricultural sources in the south to runoff from pristine soils in the north, was
30 identified. However, we could not fully determine how much of the agriculturally derived
31 NO_3^- entering the Baltic Sea finally ends up in the open waters of its central region, where the
32 addition of deep-water NO_3^- with similar isotope values might falsely, indicate a higher

1 contribution. However, we were able to show that N_2 fixation is an important NO_3^- source in
 2 the central Baltic Sea while the contribution of NO_3^- from atmospheric deposition is only a
 3 minor one.

4 Because they are particularly sensitive to human pressure and global climate change, marginal
 5 seas, including the Baltic Sea, will no doubt be affected by the increases in temperature and
 6 precipitation predicted for the near future (BACC, 2008). Indeed, increasing atmospheric
 7 depositions of NO_3^- in the world's oceans have already been reported, by Duce et al. (2008)
 8 and Kim et al. (2011) and, may impact northern catchments of the Baltic Sea to a larger
 9 extent. Additionally, in coastal waters under increasing eutrophication pressure the efficiency
 10 of NO_3^- removal was shown to be reduced (Lunau et al., 2013; Mulholland et al., 2008), and
 11 this additional NO_3^- may alter the biogeochemical cycle. Therefore, the identification of NO_3^-
 12 sources, especially as anticipated in response to global climate change, is important for future
 13 environmental management strategies for the Baltic Sea and other marine environments. We
 14 suggest that with an adaption of the potential sources the approach used in this study can
 15 easily be applied in other environments where NO_3^- is a major N contributor.

16

17 6 Supplement 1

18 Table S1: Overview of the sampling stations (location and selected chemical parameters).

Area	Date	Latitude	Longitude	Salinity	$NO_3^-+NO_2^-$ [μ Mol]	$\delta^{15}N$ (‰)	$\delta^{18}O$ (‰)
Western Baltic Sea	11.02.2008	12,4498	54,6503	8,87	3,79	4,8	3,4
	11.02.2008	12,7056	54,6959	9,22	3,81	4,3	3,5
	11.02.2008	13,0583	54,7950	8,51	4,26	4,7	4,4
	11.02.2008	13,2770	54,8598	8,20	2,71	5,0	5,2
	12.02.2008	13,9460	54,7099	7,73	2,81	4,7	3,0
	12.02.2008	13,9886	55,0624	7,73	3,04	5,0	3,1
	12.02.2008	14,1578	54,0763	5,98	46,25	8,0	1,8
Baltic Proper	12.02.2008	14,2827	54,6338	7,74	2,97	3,9	0,0
	25.02.2009	15,5695	55,5169	7,76	2,80	3,3	4,5
	25.02.2009	17,5808	57,2238	7,1	3,44	1,5	-0,9
	26.02.2009	17,3519	57,7003	7,01	3,52	3,1	0,1
	26.02.2009	18,2335	58,5837	6,45	3,79	2,0	-0,4
	26.02.2009	19,8832	59,7502	5,65	3,70	2,5	-0,8
	12.02.2008	14,5376	55,4046	7,71	3,13	4,4	-0,8
	13.02.2008	14,7156	55,4659	7,68	3,23	3,9	0,3
	13.02.2008	15,3344	55,3835	7,51	3,07	3,6	-0,4
	13.02.2008	15,6326	55,4564	7,48	2,95	3,5	0,0
13.02.2008	15,9834	55,2501	7,62	3,54	4,0	-1,0	

	19.02.2008	15,9837	55,2498	7,64	3,27	4,3	-1,1
	19.02.2008	17,0665	55,2168	7,45	3,24	3,5	-1,0
	19.02.2008	18,2351	55,3266	7,37	2,91	3,4	-1,6
	19.02.2008	18,4013	55,5502	7,34	2,61	4,0	-1,6
	18.02.2008	18,6002	55,6339	7,37	2,76	3,8	-1,3
	18.02.2008	18,8658	55,8413	7,37	2,88	4,8	1,6
	18.02.2008	19,1677	56,0841	7,37	2,71	4,3	2,0
	18.02.2008	19,5833	56,6334	7,34	3,46	3,8	-1,7
	18.02.2008	19,8289	57,0713	7,34	3,34	3,4	-2,0
	16.02.2008	20,0506	57,3196	7,35	3,38	2,6	-2,7
Gulf of Finland	04.03.2009	24,8500	59,8036	5,24	6,82	4,1	-0,2
	04.03.2009	25,6325	59,8659	5,09	8,12	6,4	-1,4
	04.03.2009	23,9989	59,6836	5,35	8,77	6,6	-1,4
	04.03.2009	22,9002	59,4836	6,11	6,95	6,4	-1,5
Gulf of Bothnia	27.02.2009	19,1177	60,1913	5,42	3,10	2,9	0,7
	27.02.2009	19,1464	60,6967	5,45	3,81	2,9	-1,1
	27.02.2009	19,2836	61,2265	5,54	3,96	5,8	-0,1
	27.02.2009	19,4678	61,7032	5,57	3,40	2,4	-1,1
	27.02.2009	19,7182	62,1441	5,45	3,49	3,2	-1,8
	28.02.2009	19,9682	62,5872	5,52	3,66	1,8	-2,2
	28.02.2009	20,4862	63,0417	5,54	4,31	4,0	-0,7
	20.11.2011	21,584	63,834	3,1	5,53	1,83	-2,79
Southern Rivers	15.11.2009	21,38187	55,30128	0	145,03	10,0	1,5
	01.12.2009	21,38187	55,30128	0	179,18	9,0	1,3
	07.12.2009	21,13293	55,69043	0	114,87	10,0	1,7
	22.02.2010	21,13293	55,69043	0	140,47	8,8	1,3
	17.01.2003	12,1429	54,03246	0	259,00	8,2	6,5
	13.02.2003	12,1429	54,03246	0	135,00	9,3	6,5
Northern rivers	11.11.2009	22,8415	65,9335	0	7,41	1,6	10,6

1

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1 **References**

- 2 Altabet, M. A., Pilskalns, C., Thunell, R., Pride, C., Sigman, D., Chavez, F. and Francois, R.:
3 The nitrogen isotope biogeochemistry of sinking particles from the margin of the Eastern
4 North Pacific, *Deep Sea Res. Part I Oceanogr. Res. Pap.*, 46(4), 655–679, doi:10.1016/S0967-
5 0637(98)00084-3, 1999.
- 6 BACC: Assessment of climate change for the Baltic Sea Basin, edited by H.-J. Bolle, M.
7 Menenti, and I. Rasool, Springer, Berlin Heidelberg., 2008.
- 8 Böhlke, J. K., Mroczkowski, S. J. and Coplen, T. B.: Oxygen isotopes in nitrate: new
9 reference materials for $^{18}\text{O}:^{17}\text{O}:^{16}\text{O}$ measurements and observations on nitrate-water
10 equilibration, *Rapid Commun. Mass Spectrom.*, 17(16), 1835–1846, doi:10.1002/rcm.1123,
11 2003.
- 12 Bourbonnais, A., Lehmann, M.F., Waniek, J.J., Schulz-Bull, D.E.: Nitrate isotope anomalies
13 reflect N_2 fixation in the Azores Front region (subtropical NE Atlantic), *J. Geophys. Res.*,
14 114, C03003, <http://dx.doi.org/10.1029/2007JC004617>, 2009.
- 15 Bourbonnais, A., Lehmann, M.F., Butterfield, D.A., Juniper, S.K.: Subseafloor nitrogen
16 transformations in diffuse hydrothermal vent fluids of the Juan de Fuca Ridge evidenced by
17 the isotopic composition of nitrate and ammonium, *Geochem. Geophys. Geosyst.*, 13 (1),
18 Q02T01, <http://dx.doi.org/10.1029/2011GC003863>, 2012.
- 19 Burns, D., Boyer, E., Elliott, E. and Kendall, C.: Sources and transformations of nitrate from
20 streams draining varying land uses: evidence from dual isotope analysis, *J. Environ. Qual.*, 38,
21 1149–1159, doi:10.2134/jeq2008.0371, 2009.
- 22 Campbell, D., Kendall, C., Chang, C. C. Y., Silva, S. R. and Tonnessen, K. A.: Pathways for
23 nitrate release from an alpine watershed: Determination using $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$, *Water Resour.*
24 *Res.*, 38(5), 1–9, 2002.
- 25 Carpenter, E. J., Harvey, H. R., Fry, B. and Capone, G. G.: Biogeochemical tracers of the
26 marine cyanobacterium *Trichodesmium*, *Deep. Res. I*, 44(1), 27–38, 1997.
- 27 Carpenter, E., Montoya, J., Burns, J., Mulholland, M. R., Subramaniam, A. and Capone, D.
28 G.: Extensive bloom of a N_2 -fixing diatom/cyanobacterial association in the tropical Atlantic
29 Ocean, *Mar. Ecol. Prog. Ser.*, 185(1977), 273–283, 1999.

1 Casciotti, K. L., Sigman, D. M., Hastings, M. G., Böhlke, J. K. and Hilkert, A.: Measurement
2 of the Oxygen Isotopic Composition of Nitrate in Seawater and Freshwater Using the
3 Denitrifier Method, *Anal. Chem.*, 74, 4905–4912, 2002.

4 Casciotti, K. L., Böhlke, J. K., McIlvin, M. R., Mroczkowski, S. J. and Hannon, J. E.: Oxygen
5 isotopes in nitrite: analysis, calibration, and equilibration, *Anal. Chem.*, 79(6), 2427–36,
6 doi:10.1021/ac061598h, 2007.

7 Conley, D. J., Carstensen, J., Aigars, J., Axe, P., Bonsdorff, E., Eremina, T., Haahti, B.-M.,
8 Humborg, C., Jonsson, P., Kotta, J., Lännegren, C., Larsson, U., Maximov, A., Medina, M.
9 R., Lysiak-Pastuszek, E., Remeikaitė-Nikienė, N., Walve, J., Wilhelms, S. and Zillén, L.:
10 Hypoxia is increasing in the coastal zone of the Baltic Sea., *Environ. Sci. Technol.*, 45(16),
11 6777–83, doi:10.1021/es201212r, 2011.

12 Conley, D. J., Carstensen, J., Vaquer-Sunyer, R. and Duarte, C. M.: Ecosystem thresholds
13 with hypoxia, *Hydrobiologia*, 629, 21–29, 2009.

14 Dalsgaard, T., De Brabandere, L. and Hall, P. O. J.: Denitrification in the water column of the
15 central Baltic Sea, *Geochim. Cosmochim. Acta*, 106, 247–260, 2013.

16 Dähnke, K., Emeis, K., Johannsen, a and Nagel, B.: Stable isotope composition and turnover
17 of nitrate in the German Bight, *Mar. Ecol. Prog. Ser.*, 408, 7–18, doi:10.3354/meps08558,
18 2010.

19 Deutsch, B., Mewes, M., Liskow, I. and Voss, M.: Quantification of diffuse nitrate inputs into
20 a small river system using stable isotopes of oxygen and nitrogen in nitrate, *Org. Geochem.*,
21 37, 1333–1342, 2006.

22 Duce, R. A., LaRoche, J., Altieri, K., Arrigo, K. R., Baker, A. R., Capone, D. G., Cornell, S.,
23 Dentener, F., Galloway, J., Ganeshram, R. S., Geider, R. J., Jickells, T., Kuypers, M. M.,
24 Langlois, R., Liss, P. S., Liu, S. M., Middelburg, J. J., Moore, C. M., Nickovic, S., Oschlies,
25 A., Pedersen, T., Prospero, J., Schlitzer, R., Seitzinger, S., Sorensen, L. L., Uematsu, M.,
26 Ulloa, O., Voss, M., Ward, B. and Zamora, L.: Impacts of Atmospheric Anthropogenic
27 Nitrogen on the Open Ocean, *Science* (80-.), 320, 893–897, 2008.

28 Durka, W., Schulze, E., Gebauer, G. and Voerkeliust, S.: Effects of forest decline on uptake
29 and leaching of deposited nitrate determined from ¹⁵N and ¹⁸O measurements, *Nature*, 372,
30 765–767, 1994.

1 Elmgren, R.: Understanding Human Impact on the Baltic Ecosystem: Changing Views in
2 Recent Decades, *Ambio*, 30(4-5), 222–231, 2001.

3 Finni, T., Kononen, K., Olsonen, R. and Wallström, K.: The history of cyanobacterial blooms,
4 *Ambio*, 30(4-5), 172–178, 2001.

5 Francis, C. a, O’Mullan, G. D., Cornwell, J. C. and Ward, B. B.: Transitions in nirS-type
6 denitrifier diversity, community composition, and biogeochemical activity along the
7 Chesapeake Bay estuary., *Front. Microbiol.*, 4, 237, doi:10.3389/fmicb.2013.00237, 2013.

8 Fry, B.: Alternative approaches for solving underdetermined isotope mixing problems, *Mar.*
9 *Ecol. Prog. Ser.*, 472, 1–13, doi:10.3354/meps10168, 2013.

10 Galloway, J. N., Aber, J. D., Erisman, J. W., Seitzinger, S. P., Howarth, R. W., Cowling, E. B.
11 and Cosby, B. J.: The Nitrogen Cascade, *Bioscience*, 53(4), 341–356, 2003.

12 Graneli, E., Wallstrom, K., Larsson, U., Graneli, W. and Elmgren, R.: Nutrient Limitation of
13 Primary Production in the Baltic Sea Area, *Ambio*, 19(3), 142–151, 1990.

14 Grasshoff, K., Ehrhardt, M. and Kremling, K.: *Methods of Seawater Analysis*, Verlag
15 Chemie., 1983.

16 Gruber, N. and Galloway, J. N.: An Earth-system perspective of the global nitrogen cycle,
17 *Nature*, 451, 2008.

18 HELCOM: Fourth periodic assessment of the state of the marine environment of the Baltic
19 Sea area 1994–1998. *Balt. Sea Environ. Proc. No. 82B*, 2002.

20 HELCOM: Eutrophication in the Baltic Sea – An integrated thematic assessment of the
21 effects of nutrient enrichment and eutrophication in the Baltic Sea region. *Balt. Sea Environ.*
22 *Proc. No. 115B*, 2009.

23 Hong, B., Swaney, D. P., Mörth, C.-M., Smedberg, E., Eriksson Hägg, H., Humborg, C.,
24 Howarth, R. W. and Bouraoui, F.: Evaluating regional variation of net anthropogenic nitrogen
25 and phosphorus inputs (NANI/NAPI), major drivers, nutrient retention pattern and
26 management implications in the multinational areas of Baltic Sea basin, *Ecol. Modell.*, 227,
27 117–135, doi:10.1016/j.ecolmodel.2011.12.002, 2012.

28 Humborg, C., Danielsson, A., Sjöberg, B. and Green, M.: Nutrient land–sea fluxes in
29 oligotrophic and pristine estuaries of the Gulf of Bothnia, Baltic Sea, *Estuar. Coast. Shelf*
30 *Sci.*, 56, 781–793, 2003.

1 Jackson, A. L., Inger, R., Bearhop, S. and Parnell, A.: Erroneous behaviour of MixSIR, a
2 recently published Bayesian isotope mixing model: a discussion of Moore & Semmens
3 (2008)., *Ecol. Lett.*, 12(3), E1–5, doi:10.1111/j.1461-0248.2008.01233.x, 2009.

4 Jäntti, H., Stange, F., Leskinen, E. and Hietanen, S.: Seasonal variation in nitrification and
5 nitrate-reduction pathways in coastal sediments in the Gulf of Finland, Baltic Sea, *Aquat.*
6 *Microb. Ecol.*, 63(2), 171–181, doi:10.3354/ame01492, 2011.

7 Johannsen, A., Dähnke, K. and Emeis, K.: Isotopic composition of nitrate in five German
8 rivers discharging into the North Sea, *Org. Geochem.*, 39(12), 1–12, 2008.

9 Kendall, C.: Tracing nitrogen sources and cycling in catchments, in *Isotope tracers in*
10 *catchment hydrology*, edited by C. Kendall and J. J. McDonnell, pp. 521–576, Elsevier,
11 Amsterdam., 1998.

12 Kendall, C., Elliott, E. and Wankel, S.: Tracing anthropogenic inputs of nitrogen to
13 ecosystems, in *Stable isotopes in ecology*, edited by K. L. R.H. Michener, pp. 375–449,
14 Blackwell Publishing., 2007.

15 Kim, T.-W., Lee, K., Najjar, R. G., Jeong, H.-D. and Jeong, H. J.: Increasing N abundance in
16 the northwestern Pacific Ocean due to atmospheric nitrogen deposition., *Science*, 334(6055),
17 505–9, doi:10.1126/science.1206583, 2011.

18 Knapp, A. N., Sigman, D. M. and Lipschultz, F.: N isotopic composition of dissolved organic
19 nitrogen and nitrate at the Bermuda Atlantic Time-series Study site, *Global Biogeochem.*
20 *Cycles*, 19(GB1018), 1–15, 2005.

21 Korth, F., Fry, B., Liskow, I. and Voss, M.: Nitrogen turnover during the spring outflows of
22 the nitrate-rich Curonian and Szczecin lagoons using dual nitrate isotopes, *Mar. Chem.*, 154,
23 1–11, doi:10.1016/j.marchem.2013.04.012, 2013.

24 Koszelnik, P. and Gruca-Rokosz, R.: Determination of nitrate isotopic signature in waters of
25 different sources by analysing the nitrogen and oxygen isotopic ratio, *Environ. Sci. Process.*
26 *Impacts*, 15(4), 751, doi:10.1039/c3em30920g, 2013.

27 Lääne, A., Kraav, E. and Titova, G.: Baltic Sea, GIWA regional assessment 17, University of
28 Kalmar on behalf of United Nations Environment Programme, Kalmar, Sweden., 2005.

29 Liu, K.-K., Su, M.-J., Hsueh, C.-R. and Gong, G.-C.: The nitrogen isotopic composition of
30 nitrate in the Kuroshio Water northeast of Taiwan: evidence for nitrogen fixation as a source

1 of isotopically light nitrate, *Mar. Chem.*, 54(3-4), 273–292, doi:10.1016/0304-
2 4203(96)00034-5, 1996.

3 Lunau, M., Voss, M., Erickson, M., Dziallas, C., Casciotti, K. and Ducklow, H.: Excess
4 nitrate loads to coastal waters reduces nitrate removal efficiency: mechanism and implications
5 for coastal eutrophication., *Environ. Microbiol.*, 15(5), 1492–504, doi:10.1111/j.1462-
6 2920.2012.02773.x, 2013.

7 Lundberg, C., Lönnroth, M., von Numers, M. and Bonsdorff, E.: A multivariate assessment of
8 coastal eutrophication. Examples from the Gulf of Finland, northern Baltic Sea, *Mar. Pollut.*
9 *Bull.*, 50(11), 1185–1196, 2005.

10 Lundberg, C., Jakobsson, B. M. and Bonsdorff, E.: The spreading of eutrophication in the
11 eastern coast of the Gulf of Bothnia, northern Baltic Sea - An analysis in time and space,
12 *Estuar. Coast. Shelf Sci.*, 82(1), 152–160, 2009.

13 Mayer, B., Boyer, E. W., Goodale, C., Jaworski, N. A., Breemen, N. van, Howarth, R. W.,
14 Seitzinger, S., Billen, G., Lajtha, K., Nadelhoffer, K., Dam, D. van, Hetling, L. J., Nosal, M.
15 and Paustian, K.: Sources of nitrate in rivers draining sixteen watersheds in the northeastern
16 U.S.: Isotopic constraints, *Biogeochemistry*, 57/58, 171–197, 2002.

17 Michaels, A. F., Siegel, D. A., Johnson, R. J., Knap, A. H. and Galloway, J. N.: Episodic
18 inputs of atmospheric nitrogen to the sargasso sea: contributions to new production and
19 phytoplankton blooms, *Global Biogeochem. Cycles*, 7(2), 339–351, 1993.

20 Montoya, J., Carpenter, E. and Capone, D.: Nitrogen fixation and nitrogen isotope abundances
21 in zooplankton of the oligotrophic North Atlantic, *Limnol. Oceanogr.*, 47(6), 1617–1628,
22 2002.

23 Moore, J. W. and Semmens, B. X.: Incorporating uncertainty and prior information into stable
24 isotope mixing models., *Ecol. Lett.*, 11(5), 470–80, doi:10.1111/j.1461-0248.2008.01163.x,
25 2008.

26 Mulholland, P. J., Helton, A. M., Poole, G. C., Hall, R. O., Hamilton, S. K., Peterson, B. J.,
27 Tank, J. L., Ashkenas, L. R., Cooper, L. W., Dahm, C. N., Dodds, W. K., Findlay, S. E. G.,
28 Gregory, S. V, Grimm, N. B., Johnson, S. L., McDowell, W. H., Meyer, J. L., Valett, H. M.,
29 Webster, J. R., Arango, C. P., Beaulieu, J. J., Bernot, M. J., Burgin, A. J., Crenshaw, C. L.,
30 Johnson, L. T., Niederlehner, B. R., O'Brien, J. M., Potter, J. D., Sheibley, R. W., Sobota, D.

1 J. and Thomas, S. M.: Stream denitrification across biomes and its response to anthropogenic
2 nitrate loading., *Nature*, 452(7184), 202–5, doi:10.1038/nature06686, 2008.

3 Myrberg, K. and Andrejev, O.: Modelling of the circulation, water exchange and water age
4 properties of the Gulf of Bothnia, *Oceanologia*, 48, 55–74, 2006.

5 Nestler, A., Berglund, M., Accoe, F., Duta, S., Xue, D. M., Boeckx, P. and Taylor, P.:
6 Isotopes for improved management of nitrate pollution in aqueous resources: review of
7 surface water field studies, *Environ. Sci. Pollut. Res.*, 18(4), 519–533, doi:10.1007/s11356-
8 010-0422-z, 2011.

9 Neumann, T.: Towards a 3D-ecosystem model of the Baltic Sea, *J. Mar. Syst.*, 25(3-4), 405–
10 419, doi:10.1016/S0924-7963(00)00030-0, 2000.

11 Pagenkopf, W.: Aktuelle Nährstoffbilanzierung für Teilgebiete des Einzugsgebiets der
12 Warnow. Geodaten Integration & Analyse, Berlin, 2001.

13 Parnell, A. C., Inger, R., Bearhop, S. and Jackson, A. L.: Source partitioning using stable
14 isotopes: coping with too much variation., *PLoS One*, 5(3), e9672,
15 doi:10.1371/journal.pone.0009672, 2010.

16 Pfenning, K. and McMahon, P.: Effect of nitrate, organic carbon, and temperature on potential
17 denitrification rates in nitrate-rich riverbed sediments, *J. Hydrol.*, 187, 283–295, 1997.

18 Pilkaityté, R. and Razinkovas, A.: Factors controlling phytoplankton blooms in a temperate
19 estuary: nutrient limitation and physical forcing, *Hydrobiologia*, 555, 41–48, 2006.

20 Radtke, H., Neumann, T., Voss, M. and Fennel, W.: Modeling pathways of riverine nitrogen
21 and phosphorus in the Baltic Sea, *J. Geophys. Res.*, 117(C9), C09024,
22 doi:10.1029/2012JC008119, 2012.

23 Schernewski, G. and Neumann, T.: The trophic state of the Baltic Sea a century ago: a model
24 simulation study, *J. Mar. Syst.*, 53, 109–124, 2005.

25 Sferratore, A., Billen, G., Garnier, J., Smedberg, E., Humborg, C. and Rahm, L.: Modelling
26 nutrient fluxes from sub-arctic basins: Comparison of pristine vs. dammed rivers, *J. Mar.*
27 *Syst.*, 73, 236–249, 2008.

- 1 Sigman, D. M., Altabet, M. A., Mccorkle, D. C., Francois, R. and Fischer, G.: The $\delta^{15}\text{N}$ of
2 nitrate in the Southern Ocean : Nitrogen cycling and circulation in the ocean interior, *J.*
3 *Geophys. Res.*, 105, 19599–19614, 2000.
- 4 Sigman, D. M., Casciotti, K. L., Andreani, M., Barford, C., Galanter, M. and Böhlke, J. K.: A
5 Bacterial Method for the Nitrogen Isotopic Analysis of Nitrate in Seawater and Freshwater,
6 *Anal. Chem.*, 73, 4145–4153, 2001.
- 7 Sigman, D. M., DiFiori, P. J., Hain, M. P., Deutsch, C., Wang, Y., Karl, D. M., Knapp, A. N.,
8 Lehmann, M. F., and Pantoja, S.: The dual isotopes of deep nitrate as a constraint on the cycle
9 budget of oceanic fixed nitrogen, *Deep Sea Res., Part I*, 56, 1419–1439, 2009.
- 10 Silva, S. R., Kendall, C., Wilkison, D. H., Ziegler, A. C., Chang, C. C. Y. and Avanzino, R.
11 J.: A new method for collection of nitrate from fresh water and the analysis of nitrogen and
12 oxygen isotope ratios, *J. Hydrol.*, 228, 22–36, 2000.
- 13 Spokes, L., Jickells, T., Weston, K., Gustafsson, B.G., Johnsson, M., Liljebladh, B., Conley,
14 D., Ambelas-Skjødth, C., Brandt, J., Carstensen, J., Christiansen, T., Frohn, L., Geernaert, G.,
15 Hertel, O., Jensen, B., Lundsgaard, C., Markager, S., W. Martinsen, Møller, B., Pedersen, B.,
16 Sauerberg, K., Sørensen, L.L., Hasager, C.C., Sempreviva, A.M., Pryor, S.C., Lund, S.W.,
17 Larsen, S., Tjernström, M., Svensson, G., Zagar, M.: MEAD: An interdisciplinary study of
18 the marine effects of atmospheric deposition in the Kattegat, *Environmental Pollution* 140,
19 1453-1462, 2006.
- 20 Stal, L.J., Albertano, P., Bergman, B., von Bröckel, K., Gallon, J.R., Hayes, P.K., Sivonen,
21 K., Walsby, A.E.: BASIC: Baltic Sea Cyanobacteria. An investigation of the structure and
22 dynamics of water blooms of cyanobacteria in the Baltic Sea - Responses to a changing
23 environment, *Cont Shelf Res* 23, 1695-1714, 2003.
- 24 Stålnacke, P., Grimvall, A., Sundblad, K. and Tonderski, A.: Estimation of riverine loads of
25 nitrogen and phosphorus to the Baltic Sea, 1970-1993, *Environ. Monit. Assess.*, 58(2), 173–
26 200, 1999.
- 27 Stepanauskas, R., Jorgensen, N. O. G., Eigaard, O. R., Žvikas, A., Tranvik, L. J. and
28 Leonardson, L.: Summer Inputs of Riverine Nutrients to the Baltic Sea: Bioavailability and
29 Eutrophication Relevance, *Ecol. Monogr.*, 72(4), 579–597, 2002.
- 30 Thunell, R. C., Sigman, D. M., Muller-Karger, F., Astor, Y. and Varela, R.: Nitrogen isotope
31 dynamics of the Cariaco Basin, Venezuela, *Global Biogeochem. Cycles*, 18, 1–13, 2004.

- 1 Vahtera, E., Laanemets, J., Pavelson, J., Huttunen, M., Kononen, K.: Effect of upwelling on
2 the pelagic environment and bloom-forming cyanobacteria in the western Gulf of Finland,
3 Baltic Sea, *Journal of Marine Systems* 58, 67-82, 2005.
- 4 Vahtera, E., Conley, D. J., Gustafsson, B. G., Kuosa, H., Pitkänen, H., Savchuk, O. P.,
5 Tamminen, T., Viitasalo, M., Voss, M., Wasmund, N. and Wulff, F.: Internal ecosystem
6 feedbacks enhance nitrogen-fixing cyanobacteria blooms and complicate management in the
7 Baltic Sea., *Ambio*, 36(2-3), 186–94, 2007.
- 8 Villnäs, A., Norkko, J., Hietanen, S., Josefson, A., Lukkari, K. and Norkko, A.: The role of
9 recurrent disturbances for ecosystem multifunctionality, *Ecology*, 94(10), 2275–2287, 2013.
- 10 Vitousek, P. M., Aber, J. D., Howarth, R. W., Likens, G. E., Matson, P. A., Schindler, D. W.,
11 Schlesinger, W. H. and Tilman, D. G.: Human alteration of the global nitrogen cycle: Sources
12 and consequences, *Ecol. Appl.*, 7(3), 737–750, 1997.
- 13 Voss, M., Deutsch, B., Elmgren, R., Humborg, C., Kuuppo, P., Pastuszak, M., Rolff, C. and
14 Schulte, U.: Source identification of nitrate by means of isotopic tracers in the Baltic Sea
15 catchments, *Biogeosciences*, 3, 663–676, 2006.
- 16 Voss, M., Dippner, J. W., Humborg, C., Hürdler, J., Korth, F., Neumann, T., Schernewski, G.
17 and Venohr, M.: History and scenarios of future development of Baltic Sea eutrophication,
18 *Estuar. Coast. Shelf Sci.*, 92(3), 307–322, 2011.
- 19 Voss, M., Emeis, K.-C., Hille, S., Neumann, T. and Dippner, J. W.: Nitrogen cycle of the
20 Baltic Sea from an isotopic perspective, *Global Biogeochem. Cycles*, 19(GB3001), 1–15,
21 2005.
- 22 Wankel, S. D., Kendall, C., Francis, C. A. and Paytan, A.: Nitrogen sources and cycling in the
23 San Francisco Bay Estuary: A nitrate dual isotopic composition approach, *Limnol. Ocean.*,
24 51(4), 1654–1664, 2006.
- 25 Wasmund, N., Voss, M. and Lochte, K.: Evidence of nitrogen fixation by non-heterocystous
26 cyanobacteria in the Baltic Sea and re-calculation of a budget of nitrogen fixation, *Mar. Ecol.*
27 *Prog. Ser.*, 214, 1–14, 2001.
- 28 Wulff, F., Humborg, C., Medina, M. R., Mörth, C.-M., Savchuk, O. and Sokolov, A.:
29 Revision of the country allocation of nutrient reductions in the Baltic Sea Action Plan, Baltic
30 Nest Institute, Stockholm., 2009.

- 1 Xue, D., De Baets, B., Van Cleemput, O., Hennessy, C., Berglund, M. and Boeckx, P.: Use of
2 a Bayesian isotope mixing model to estimate proportional contributions of multiple nitrate
3 sources in surface water., *Environ. Pollut.*, 161, 43–9, doi:10.1016/j.envpol.2011.09.033,
4 2012.
- 5 Xue, D., De Baets, B., Van Cleemput, O., Hennessy, C., Berglund, M. and Boeckx, P.:
6 Classification of Nitrate Polluting Activities through Clustering of Isotope Mixing Model
7 Outputs., *J. Environ. Qual.*, 42(5), 1486–97, doi:10.2134/jeq2012.0456, 2013.
- 8 Yang, L., Han, J., Xue, J., Zeng, L., Shi, J., Wu, L. and Jiang, Y.: Nitrate source
9 apportionment in a subtropical watershed using Bayesian model., *Sci. Total Environ.*, 463-
10 464, 340–7, doi:10.1016/j.scitotenv.2013.06.021, 2013.
- 11

- 1 Table 1. NO_3^- concentrations and $\delta^{15}\text{N-NO}_3^-$ and $\delta^{18}\text{O-NO}_3^-$ values of wet atmospheric
 2 deposition. Data are from Warnemünde (Germany), Sängen (Sweden), and Majstre (Sweden).

Location	Date	NO_3^- ($\mu\text{mol L}^{-1}$)	$\delta^{15}\text{N-NO}_3^-$ (‰)	$\delta^{18}\text{O-NO}_3^-$ (‰)
Warnemünde	21.12.2009	52.7	2.1	75.6
Warnemünde	04.01.2010	51.2	1.1	68.3
Warnemünde	19.01.2010	104.4	0.2	84.6
Warnemünde	01.02.2010	50.8	0.8	65.8
Warnemünde	19.02.2010	94.4	0.6	79.5
Warnemünde	22.02.2010	106.8	2.1	81.8
Sängen	Dec. 2009	12.1	-0.3	69.2
Sängen	Jan. 2010	60.4	-1.1	81.8
Sängen	Feb. 2010	69.3	-2.1	77.0
Majstre	Dec. 2009	30.7	-0.8	83.8

3

4

1 Table 2. Means and standard deviations of the $\delta^{15}\text{N-NO}_3^-$ and $\delta^{18}\text{O-NO}_3^-$ values of the NO_3^-
 2 sources used in the SIAR mixing model. For further details, see Material and Methods, SIAR
 3 mixing model.

Source	$\delta^{15}\text{N-NO}_3^-$ (mean \pm SD)	$\delta^{18}\text{O-NO}_3^-$ (mean \pm SD)	n	Origin	Reference
NO_3^- from atmospheric deposition	0.3 \pm 1.4	76.7 \pm 6.8	10	Warnemünde (Germany), and Sängen and Majstre (Sweden)	This study
NO_3^- from pristine soils	1.3 \pm 1.4	1.5 \pm 0.9	5	Groundwater	Deutsch et al., 2006
NO_3^- from agricultural runoff	9.9 \pm 1.5	4.6 \pm 1.0	21	Tile-drain outlets, Warnow River	Deutsch et al., 2006
NO_3^- from N_2 fixation	-1.0 \pm 1.0	-0.7 \pm 2.9	0	Estimated	Carpenter et al., 1999, 1997; Bourbonnais et al., 2009, 2012; Montoya et al., 2002; Sigman et al. 2009

4 .

- 1 Table 3. Source attribution results: Mean, standard deviation, and minimum and maximum values for the potential contributions of four potential
 2 NO₃⁻ sources for the areas Western Baltic Sea, Baltic Proper, Gulf of Finland, Gulf of Bothnia, southern rivers, and northern rivers.

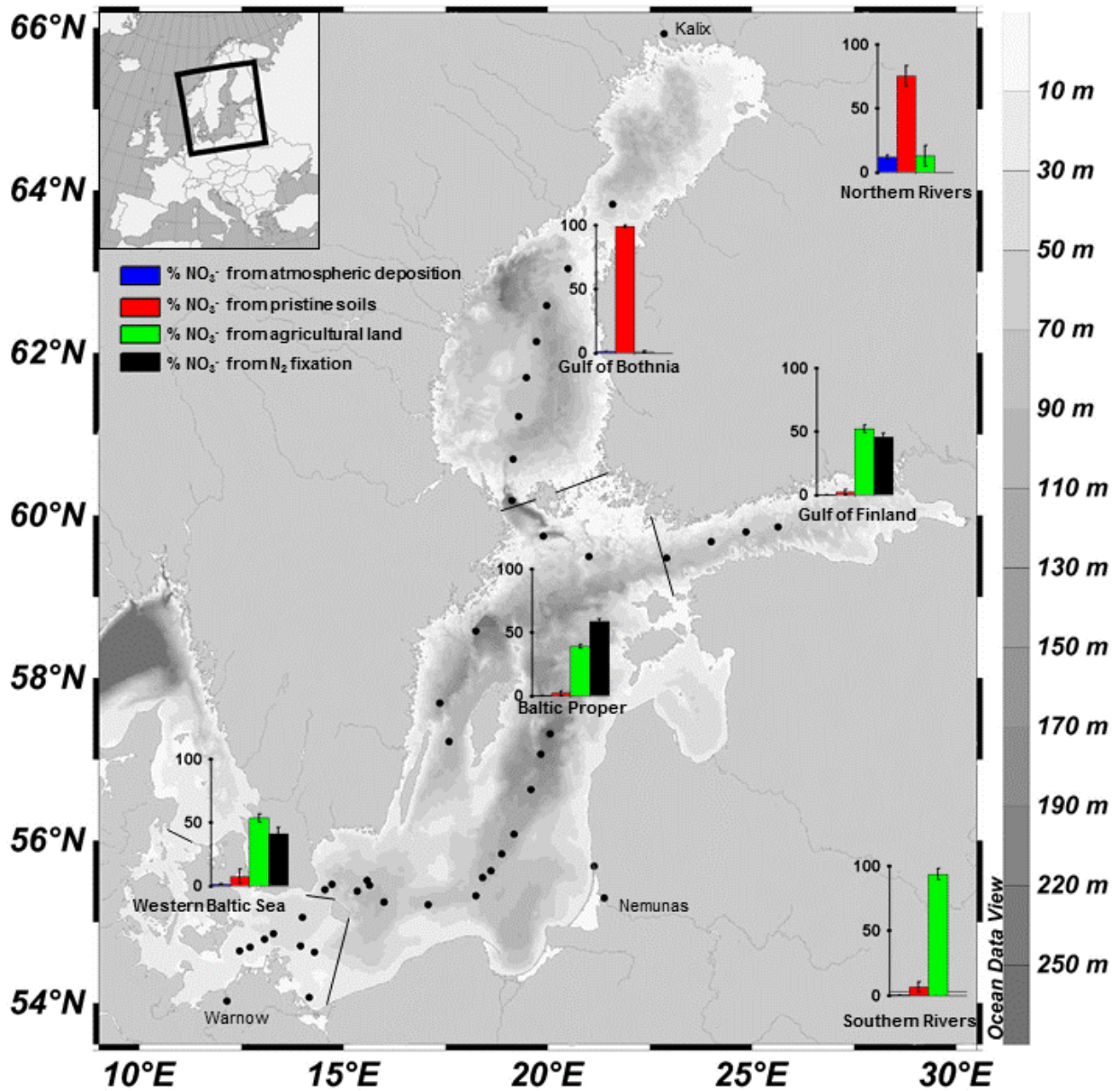
Area	NO ₃ ⁻ from atmospheric deposition		NO ₃ ⁻ from pristine soils		NO ₃ ⁻ from agricultural runoff		NO ₃ ⁻ from N ₂ fixation	
	Mean±SD	Min-Max	Mean±SD	Min-Max	Mean±SD	Min-Max	Mean±SD	Min-Max
Western Baltic Sea	1.1±0.5	0.0–3.5	7.5±5.9	0.0–42.0	53.5±3.2	41.0–66.5	37.9±5.1	11.0–51.9
Baltic Proper	0.1±0.1	0.0–0.9	2.1±1.9	0.0–14.7	39.0 ±1.6	32.8–45.5	58.8±2.0	49.3–65.3
Gulf of Finland	0.2±0.2	0.0–2.0	2.4±2.1	0.0–24.3	51.9±3.0	40.9–63.4	45.5±3.2	32.7–59.0
Gulf of Bothnia	0.1±0.1	0.0–0.5	99.0±0.9	91.7–100.0	1.0±0.9	0.0–8.2	-	-
Southern rivers	0.2±0.1	0.0–1.3	6.4±4.2	0.0–24.5	93.5±4.2	75.2–100.0	-	-
Northern rivers	11.8±1.5	6.6–23.4	75.3±7.9	33.8–92.8	12.9±8.1	0.0–57.2	-	-

1 Table 4. Comparison of $\delta^{15}\text{N-NO}_3^-$ values from surface water samples and $\delta^{15}\text{N}$ values from
 2 sediments samples in sub-regions of the Baltic Sea.

	Baltic southern coastal areas/ Gulf of Finland	Central Baltic Proper	
$\delta^{15}\text{N}$ sediments (‰)	7.3 ± 2.1	3.5 ± 0.6	Voss et al. 2005
$\delta^{15}\text{N-NO}_3^-$ surface water column (‰)	7.9 ± 1.8	3.6 ± 1.0	This study

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4



1

2 Figure 1. Station Map of the Baltic Sea and percent contribution of the four nitrate sources,
 3 NO_3^- from atmospheric deposition (blue), pristine soils (red), agricultural runoff (green), and
 4 N_2 fixation (black), for the Western Baltic Sea, Baltic Proper, Gulf of Finland, Gulf of
 5 Bothnia, southern rivers, and northern rivers. Stations are indicated as black dots. For more
 6 details see Suppl. Table 1.

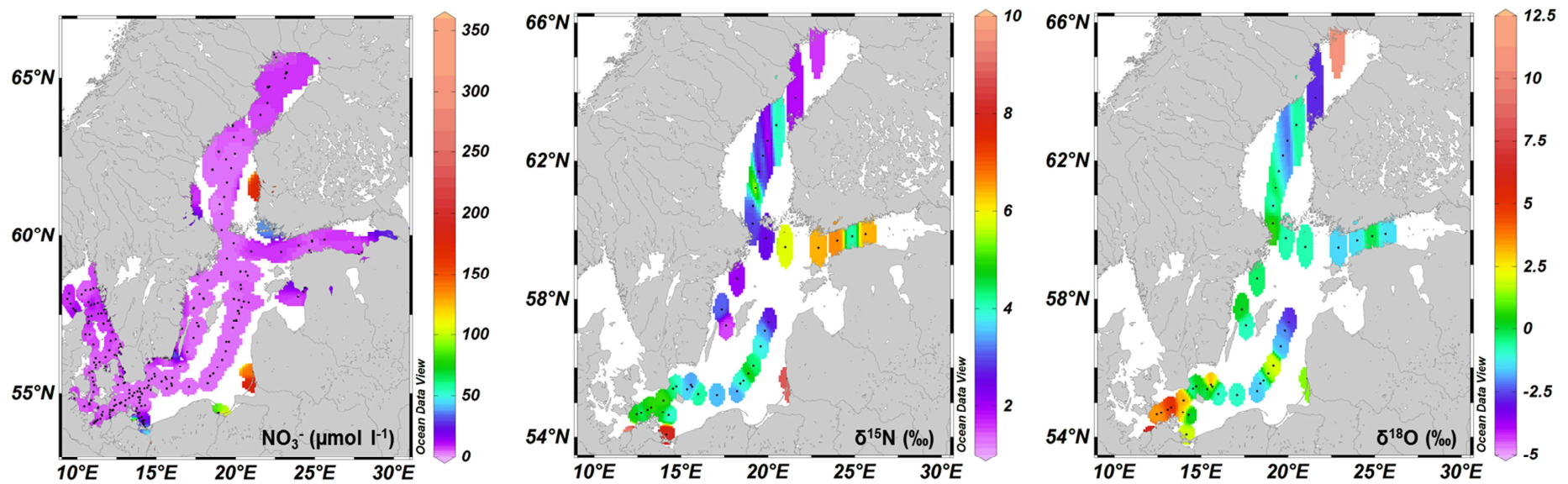
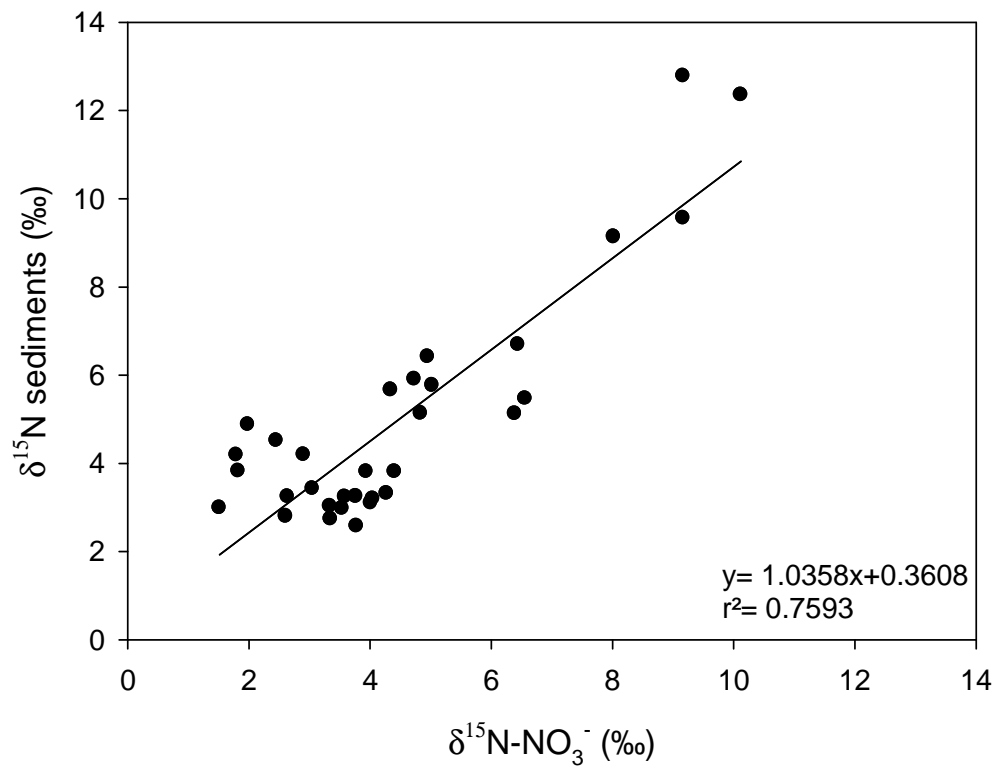
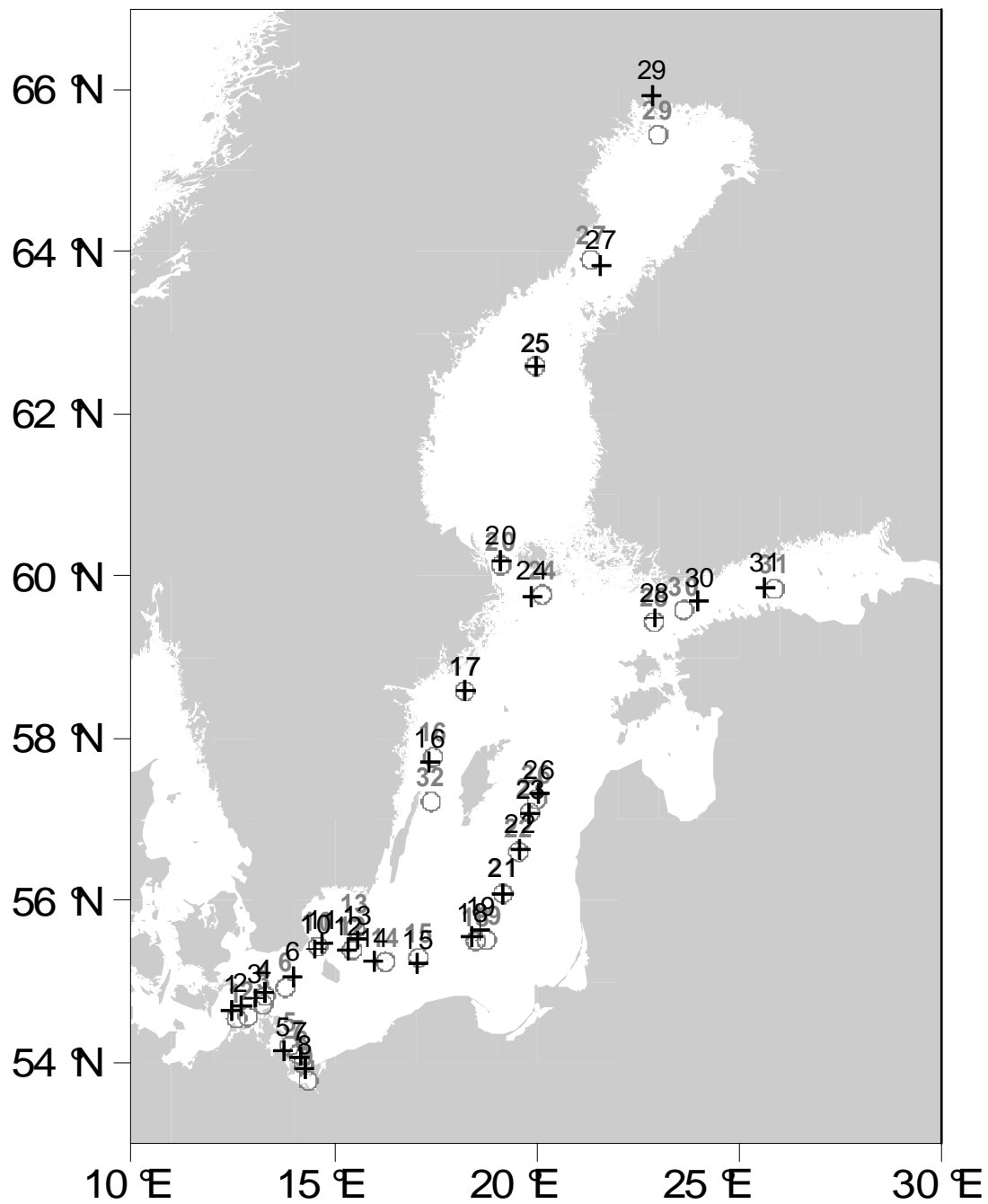


Figure 2. Surface water column NO_3^- concentrations (A), $\delta^{15}\text{N-NO}_3^-$ values (B), and $\delta^{18}\text{O-NO}_3^-$ values (C) for the Baltic Sea. Stations are indicated as black dots. Additional NO_3^- concentrations were obtained from the Data Assimilation System (DAS) (<http://nest.su.se/das/>) in winter (Nov.–Feb.) of the years 2000 to 2012.



1
 2 Figure 3. $\delta^{15}\text{N}$ from sediment samples vs. $\delta^{15}\text{N-NO}_3^-$ from surface water samples. $\delta^{15}\text{N}$
 3 values from sediments were taken from Voss et al. (2005). The positive slope suggests a tight
 4 coupling between $\delta^{15}\text{N-NO}_3^-$ in surface waters and $\delta^{15}\text{N}$ in sediment samples.



5

6 Figure 4. Station map for the comparison of isotope patterns in the water column and
 7 sediments. Gray circles are the stations referred to in Voss et al. (2005) and black crosses are
 8 those from this study. Isotope values were compared at stations with the same number.

9