1	Partial coupling and differential regulation of biologically and photo-chemically labile dissolved organic
2	carbon across boreal aquatic networks
3	
4	JF. Lapierre ^{1*} , P. A. del Giorgio ¹
5	
6	¹ Groupe de Recherche Interuniversitaire en Limnologie et en Environnement Aquatique (GRIL),
7	Département des Sciences Biologiques, Université du Québec à Montréal, Case Postale 8888, succursale
8	Centre-Ville, Montréal, QC, H3C 3P8, Canada
9	*Corresponding author: jfrancoislapierre@gmail.com
10	Keywords: dissolved organic carbon, lability, boreal, aquatic

12 Abstract

13 Despite the rapidly increasing volume of research on the biological and photochemical degradation of 14 DOC in aquatic environments, little is known on the large-scale patterns in biologically and photo-15 chemically degradable DOC (Bd-DOC and Pd-DOC, respectively) in continental watersheds, and on the 16 links that exist between these two key properties that greatly influence the flow of carbon from 17 continents to oceans. Here we explore the patterns in the concentrations and proportions of Bd- and 18 Pd-DOC across hundreds of boreal lakes, rivers and wetlands spanning a large range of system trophy 19 and terrestrial influence, and compared the drivers of these two reactive pools of DOC at the landscape 20 level. Using standardized incubations of natural waters, we found that the concentrations of Bd- and Pd-21 DOC co-varied across all systems studied but were nevertheless related to different pools of dissolved 22 organic matter (DOM, identified by fluorescence analyses) in ambient waters. Concentrations of nutrients and protein-like fluorescent DOM (FDOM) explained nearly half of the variation in Bd-DOC, 23 24 whereas Pd-DOC was exclusively predicted by DOM optical properties, consistent with the 25 photochemical degradability of specific FDOM pools that we experimentally determined. The 26 concentrations of colored DOM (CDOM), which we use here as a proxy of terrestrial influence, almost 27 entirely accounted for the observed relationship between FDOM and the concentrations of both Bd- and 28 Pd-DOC. The concentrations of CDOM and of the putative bio-labile fluorescence component shifted 29 from complete decoupling in clear-water environments to strong coupling in darker streams and 30 wetlands. This suggests a baseline autochthonous Bd-DOC pool fuelled by internal production that is gradually overwhelmed by land-derived Bd-DOC as terrestrial influence increases across landscape 31 32 gradients. The importance of land as a major source of both biologically and photo-chemically 33 degradable DOC for continental watersheds resulted in a partial coupling of those carbon pools in 34 natural freshwaters, despite fundamental contrasts in terms of their composition and regulation.

35 1. Introduction

36 The movement of terrestrial dissolved organic carbon (DOC) from the land-water interface to 37 the oceans is mediated by its transit through complex and highly heterogeneous continental freshwater 38 networks, where DOC from different sources and origins is simultaneously produced and removed via 39 biotic and abiotic processes (Massicotte and Frenette, 2011;Cole et al., 2007;Battin et al., 2008). The 40 overall role that DOC plays on aquatic ecosystems is well recognized (Findlay and Sinsabaugh, 2002), but there is still debate regarding the direct role of terrestrially derived organic carbon as a substrate for 41 42 ecological and biogeochemical processes in freshwater ecosystems. More specifically, it is generally 43 accepted that terrestrial DOC may support foodwebs at various trophic levels (Berggren et al., 44 2010b;Karlsson et al., 2003;Jansson et al., 2007), as well as fuel CO_2 evasion from these systems 45 (Algesten et al., 2004;Lapierre et al., 2013;Berggren et al., 2012), but the relative importance for those 46 processes compared to that of autochthonous sources of organic C (Brett et al., 2009;Wenzel et al., 47 2012), or terrestrially derived CO₂ (Wallin et al., 2013;Butman and Raymond, 2011), respectively, is still 48 questioned. This questioning largely emerges from unresolved issues concerning terrestrial organic 49 carbon degradability in aquatic ecosystems, which then determines the ability of this carbon to enter 50 aquatic foodwebs and biogeochemical cycles.

51 Microbial and photochemical degradation are the two main pathways by which terrestrial DOC 52 may influence aquatic metabolism (del Giorgio et al., 1999;Obernosterer and Benner, 2004;Bertilsson 53 and Tranvik, 1998) and gas dynamics (Molot and Dillon, 1997; Jonsson et al., 2001; Lapierre et al., 2013), 54 and both processes operate simultaneously in ambient waters. Their magnitude and relative 55 contribution in natural systems depend on environmental factors, such as light availability, water residence time and dissolved ions (Soumis et al., 2007), as well as on factors that are intrinsic to DOC 56 57 (Guillemette and del Giorgio, 2011). Microbial degradation is thought to preferentially target freshly 58 produced, low-molecular weight molecules with low aromaticity (Amon and Benner, 1996;Wickland et

al., 2007), whereas photochemical degradation mainly acts on colored, photo-reactive molecules
generally associated to high molecular weight and aromaticity (Benner and Kaiser, 2011;Bertilsson and
Tranvik, 1998;Stubbins et al., 2010). These broad chemical properties have thus been associated to
distinct pools of DOC with distinct sources (i.-e. the process that imported DOC in the environment, e.g.
leaching, exudation) and origins (i.-e. the environment where these processes take place, e.g. terrestrial,
riverine, marine).

65 Transposing the geochemical evidence described above to natural landscapes would suggest a 66 shift in the relative importance of these DOC degradation pathways, from a dominance of biological 67 degradation in systems dominated by autochthonous carbon sources, to a dominance of photo-chemical 68 degradation in environments with high terrestrial influence. This scenario, which is based on the 69 simplistic assumption that biological degradability is mostly linked to autochthonous DOC, and that 70 photochemical degradability is mostly associated to terrestrially derived DOC, would lead to a 71 compensatory dynamic wherein the overall (biological + photochemical) degradability of DOC would be 72 to some extent buffered across different environments and along a gradient of terrestrial influence. 73 Although the above assumption may hold for specific aquatic environments, it is unlikely that it 74 applies at the landscape level, in part because there is increasing evidence that land exports to 75 continental waters not only photo-chemically degradable DOC (Molot and Dillon, 1997; Jonsson et al., 76 2001; Weyhenmeyer et al., 2012), but also significant amounts of biologically degradable DOC (Berggren 77 et al., 2010a; Fellman et al., 2008; Guillemette et al., 2013). For example, significant concentrations of 78 protein-like DOC and small organic acids, typically attributed to autochthonous and bio-labile material, 79 have been measured in soils and headwater streams (Berggren et al., 2010a;Fellman et al., 2009), and 80 carbon pools that are considered recalcitrant from a geochemical perspective (based on their molecular 81 properties and degree of prior processing) may actually be biologically degradable under the right 82 environmental conditions, in soils (Schmidt et al., 2011) or in the water (Marín-Spiotta et al., 2014;Ward

et al., 2013). This implies that there may be faster turnover of terrestrially derived DOC in inland waters
than what was previously thought, yet there is a large volume of literature documenting the
recalcitrance of this carbon in marine or riverine environments with high residence time (see synthesis
by Marín-Spiotta et al., 2014). This would suggest that the degradability of terrestrial DOC may be very
different in contrasting aquatic environments, and in particular, that it becomes increasingly stable with
time as it circulates through continental watersheds, where the labile portions of the DOC pool
imported from upstream are gradually processed and removed from the bulk.

90 It thus appears that land delivers to inland waters a DOC pool that, depending on the local 91 environmental context, may be both biologically and photo-chemically degradable. If indeed biologically 92 and photo-chemically degradable DOC share, to a certain degree, similar sources and origins, one could 93 expect both functional pools of DOC to co-vary across aquatic environments. This would lead to an 94 additive rather than compensatory dynamic, wherein the overall degradability of DOC would increase as 95 a function of increasing terrestrial export, with major implications for the aquatic carbon cycle. Previous 96 evidence has shown that both Bd-DOC and Pd-DOC tend to increase in boreal freshwaters with 97 increasing terrestrial influence (Lapierre et al., 2013), but the respective drivers of those pools and the 98 degree to which they are coupled across a large diversity of freshwater environments, remain unclear; 99 few if any studies have simultaneously assessed the magnitude and regulation of biological and 100 photochemical degradability across gradients representative of aquatic ecosystems at a landscape level. 101 Here we explore the patterns in the concentrations and proportions of biologically and photo-chemically 102 degradable DOC across boreal lakes, rivers and wetlands, and the degree to which those distinct pools of 103 DOC co-vary in natural waters. We targeted this diversity of aquatic ecosystems because they represent 104 the existing gradient in terms of connectivity with land and therefore in the extent of terrestrial 105 influence on aquatic DOC dynamics. We then use a combination of optical and chemical properties in 106 order to explore how intrinsic DOC composition and extrinsic environmental properties respectively

- drive the concentrations of Bd- and Pd-DOC in boreal natural waters. Finally, we explore how a
- 108 continuous gradient of terrestrial influence (using CDOM as a proxy, see Lapierre et al., 2013) may
- 109 underlie the patterns in the concentrations of degradable DOC across diverse and complex freshwater
- 110 networks.
- 111

112 2.Methods

113 2.1 Study regions and sampling

114 Over the summer period from 2009 to 2013 we sampled a wide range of lakes (236), streams 115 (204), rivers and wetlands (83; mostly beaver ponds). Those systems span over a very large geographic 116 range (Fig. 1), which translates into very diverse climate, landscape and limnological properties. In 117 particular, mean annual temperature and precipitation ranged from -5.7 to 4.8 °C and from 334 to 1289 118 mm, respectively, and catchment vegetation ranged from mixed conifer and deciduous in the south to 119 black spruce moss forest in the north; tundra-type vegetation covered the northern-most, highest 120 altitude sites. Aquatic environmental properties spanned over most of the freshwater gradients 121 reported in the literature, with DOC, CDOM (a440 nm, naperian units), TN, TP and Chl a concentrations ranging from 0.4 - 123.9 mg L⁻¹, 0.05 - 30.0 m⁻¹, 0.03 - 1.90 mg L⁻¹, 2.24-248.2 μ g L⁻¹ and 2.2 - 248.2 μ g L⁻¹, 122 123 respectively, across the sampling sites (Lapierre et al. 2013). Hydrology and morphometry were also very 124 different across the systems studied, with rivers ranging from Strahler order 1 to 6, and lakes and wetlands ranging from 0.01 to 2300 km² (Lapierre et al., 2013), as well as mean depth and discharge 125 126 ranging over several orders of magnitude (Lapierre and del Giorgio, 2012;Campeau et al., 2014). The 127 different sites were seldom part of the same catchment except for the largest lakes and rivers that 128 drained vast territories.

Water was sampled at 0.5 m from the surface from the deepest measured point of lakes. It was sampled near the shore of streams, rivers and wetlands and in all cases, it was immediately filtered (0.45 µm) and stored in acid-washed glass vials for DOC, optical analyses and inorganic N. Samples for total nitrogen (TN) and phosphorus (TP) were also stored in acid-washed glass vials but were not filtered. All samples were immediately stored in cold and dark conditions. Analyses were typically performed within a month; optical measurements and inorganic N were typically performed within two weeks.

135 2.2 Biological and chemical analyses

136 DOC concentration was measured on an OI 1010 TIC-TOC (TX, USA) analyzer following sodium 137 persulfate digestion. We analyzed TP spectrophotometrically after persulfate digestion and TN was 138 analyzed as nitrate following alkaline persulfate digestion and measured on an Alpkem FlowSolution IV 139 autoanalyzer; ambient nitrites and nitrates were measured individually prior to persulfate digestion 140 (APHA, 1998). Ammonium was measured by fluorescence following Holmes et al. (1999). Chl a samples 141 were analyzed spectrophotometrically after filtration on Whatman (GF/F) filters and hot ethanol (90 %) 142 extraction (Marker and Nusch, 1980). Filters were sonicated prior to extraction. Each of those variables 143 has been collected in duplicates in the field; we present the mean here.

144 2.3 Optical analyses

145 The absorbance of colored dissolved organic matter (CDOM) was measured on nanopure-146 corrected samples of filtered ambient water from 230 to 700 nm, using a UV-visible Ultrospec 2100 147 spectrometer (Biochrom, Cambridge, UK) and a 2-cm quartz cuvette. We report CDOM as the absorption coefficient at 440 nm (in m⁻¹, naperian units), calculated by dividing the optical absorbance at 440 nm by 148 149 the path length in meters and multiplying by 2.303 (Cuthbert and del Giorgio, 1992). We also calculated 150 proxies of DOC aromaticity and molecular weight by estimating the DOC specific absorbance at 254 nm 151 (SUVA₂₅₄) and the ratio of absorbance at 250 nm to 365 nm (Spencer et al., 2009a), respectively. The 152 former was calculated by dividing absorption coefficients at 254 nm by DOC concentration and the latter 153 was directly obtained from the ratio of absorption coefficients at these specified wavelengths. 154 Fluorescence intensity was measured on a Shimadzu RF5301 PC (Shimadzu, Kyoto, Japan), 155 across excitation wavelengths of 275-450nm (5 nm increments) and emission wavelengths of 280-600 156 nm (2nm increments) in order to build excitation-emission matrices (EEMs). A parallel factor (PARAFAC) 157 model (Stedmon et al., 2003) was developed to identify and quantify groups of DOM that shared similar 158 optical properties and distribution across the sampled sites. Fluorescence data were corrected for inner-

159 filter effect and standardized to Raman units using the FDOMcorr 1.6 toolbox (Murphy et al., 2010) in 160 MATLAB (MathWorks, Natick, MA, USA). The model was based on 1349 samples from ambient waters 161 and from photochemical degradation experiments, and was performed on corrected data using the 162 DOMfluor 1.7 toolbox (Stedmon and Bro, 2008); only part of the data are reported here. The initial 163 dataset comprised 1577 samples; samples for which absorbance within the 1 cm cuvette was higher 164 than 0.6 at 254 nm were excluded due to potential for ineffective inner filter effect correction (Miller et 165 al., 2010), which could bias the final model. We express the "concentrations" of fluorescence 166 components as the maximum fluorescence intensity at the peak. These represent relative 167 concentrations that may be used to explore the cross-sample patterns for a given component, but this 168 does not necessarily mean that one unit of a given component represents an equal amount of DOM 169 compared to one unit of any other component.

170 2.4 Biological and photochemical DOC degradation

171 We carried out standardized biological and photochemical degradation experiments in order to 172 derive a concentration of degradable DOC that did not depend on varying incubation conditions and 173 would thus be fully comparable across ecosystems and in time (Lapierre et al., 2013). The experiments were started on the same day that the samples were collected. Biological degradation experiments were 174 175 carried out in water that was filtered through 2.8 µm nominal pore size GF/D filters (Whatman) in order 176 to maintain the *in situ* bacterial community and thus to avoid re-inoculation. Water samples were 177 incubated in the dark at a fixed incubation time (14 days) and temperature (20°C). The results presented 178 in Figs. 2 and 3 correspond to the concentrations in biologically degradable DOC (Bd-DOC), expressed as 179 the absolute amount of DOC removed over a period of 14 days, calculated as the difference in 180 concentration before and after degradation. Percent degradable DOC was calculated as the fraction 181 represented by the difference between DOC concentration at the start of the experiment and Bd-DOC, 182 multiplied by 100.

183 A subset of the filtered (2.8 µm) water was used for photochemical degradation experiments, 184 which were carried out in a solar simulator (Qsun XE1-BC, Qlab, FL, USA) under a standard light dose (0.68 W m⁻² at 340 nm, spectrum representative of natural sunlight) in 24 mm diameter glass tubes 185 186 disposed horizontally. Given the strong light dose and the small cross-section of the tubes, there was a negligible effect of the CDOM concentrations on the effective light dose inside the tubes, even for the 187 188 most colored samples (See Appendix A for details). The amount of light energy available in the vial for 189 wavelengths comprised between 300 and 450 nm, responsible for most photochemical processing of DOC (Vähätalo et al., 2000), averaged 130.8 W m⁻² and did not vary substantially across samples due to 190 self-shading (std.dev. = 9.2 W m^{-2} ; 10th and 90th percentiles = 118.4 and 140.4 W m⁻², respectively) 191 compared to the range of variation measured in the concentrations of photo-chemically degradable 192 193 DOC (Pd-DOC), which spanned several orders of magnitude. Consequently, standardizing the 194 concentrations in Pd-DOC for the corrected light dose in the corresponding tubes could in no way alter 195 the patterns and conclusions of the current study (see Figure A2). We therefore report the uncorrected 196 values (in mg C L⁻¹ of photo-chemically degradable DOC). We assume that most bacteria could not 197 survive the very strong UV dose and thus significantly contribute to the DOC loss in irradiation 198 experiments; even if that were not the case, the DOC loss rates (expressed per day) were always at least 199 one order of magnitude higher in photochemical degradation experiments compared to biological 200 degradation experiments conducted in the dark (Lapierre et al., 2013).

201 Photo-exposure time (24 hours) and temperature (24 °C) were the same for all experiments 202 (Lapierre et al., 2013). The data presented in Figs. 2 and 3 correspond to the amount of DOC removed in 203 1 day under those conditions; considering diel light cycles and the natural light intensity at the studied 204 latitudes, this would roughly correspond to six days of natural light exposure in the sampled regions. 205 Irradiated samples were stored in 40 ml glass tubes and directly analyzed for DOC concentration. We

- also measured DOM optical properties before and after irradiation for a subset (n = 187) of the samples.
- 207 The fluorescence data were included in the PARAFAC model.

208 2.5 Statistical analyses

- 209 Analyses of variance, simple and multiple linear regressions and principal components analyses
- 210 (PCA) were performed on log-transformed data in JMP 7.0.1 software. Data were centered and
- 211 standardized before performing the PCA. The best multiple linear regression (MLR) models were
- 212 identified using forward step-wise selection based on change in Akaike information criterion (AIC.)
- 213 Moving window regressions (MWR) of fluorescence component C6 against CDOM concentrations were
- 214 performed successively on subsets of 250 samples (about half of the sample size) sorted by increasing
- 215 CDOM.
- 216

217 **3. Results**

218 3.1 Patterns in the amounts and proportions of degradable DOC

The amounts of DOC removed in these standardized incubations ranged from 0.05 to 15.2 mg L⁻¹ 219 and from almost 0 to 26.0 mg L^{-1} , for biological and photochemical degradation, respectively (Fig. 2). 220 221 This represented 0.13% to 54.3%, and from 0.04 to 72.6% of the ambient DOC pool where those 222 samples were collected, respectively (Fig. 2). These numbers represent potential degradation under 223 standardized conditions and depending on environmental conditions may not be expressed in situ; they 224 should not be considered as representative of ambient rates. They do allow, however, to isolate DOC 225 degradability while minimizing the influence of site-specific conditions, such as temperature or 226 irradiation, and thus to explore the patterns and drivers of this degradability over a large range of DOC 227 properties. 228 The amounts and proportions of degradable DOC were typically lower in lakes than in rivers and 229 wetlands (Fig. 2). Both the amounts and proportions of Bd-DOC were highest in wetlands, whereas rivers contained the highest proportions of Pd-DOC, but similar amounts of Pd-DOC compared to 230 231 wetlands (Fig. 2). There was no relationship at all between the percent Bd-DOC and Pd-DOC across the 232 ensemble of almost 300 lakes, rivers and wetlands for which those measurements were available (Fig. 233 3a). There was, however, a significant positive relationship between the absolute amounts of Bd-DOC 234 and Pd-DOC in lakes (, rivers and wetlands separately. There was thus a significant coupling between 235 both pools of DOC when all the systems were considered together , and this coupling was typically 236 stronger in rivers and wetlands than in lakes (Fig. 3b). There was a very large scatter around the 237 regression, however, and concentrations of Pd-DOC ranged by more than one order of magnitude for 238 any given amount of Bd-DOC, and vice-versa.

239 3.2 The PARAFAC model

240 The PARAFAC model identified six fluorescence components (Fig. 4), corresponding to humic-, 241 fulvic- and protein-like material that are widely reported in freshwaters. In particular, quantitative 242 comparison of our components to published components (using OpenFluor database, (Murphy et al., 243 2014)) revealed that components C1 and C2 reported here matched very well previously reported 244 humic-like, terrestrially derived material identified in a variety of aquatic environments (Stedmon et al., 245 2003; Shutova et al., 2014). Component C5 and C6 matched widely reported components (Cory and 246 McKnight, 2005;Fellman et al., 2008;Kothawala et al., 2013b, among others), which have been 247 associated to microbially derived humic-like and freshly produced protein-like material, respectively. 248 The whole spectra of components C3 and C4 did not show a strong match with any fluorescence 249 component published in the OpenFluor database, but had fluorescence peaks comparable to other 250 published humic- and fulvic like material (Guillemette and del Giorgio 2012, Lu et al 2013); these 251 components had excitation spectra ranging in the UV-B and UV-A regions (Fig. 4), indicating that they 252 strongly absorb light at those wavelengths.

253 Although the model was not run with samples with the highest absorbance, as described in the 254 Methods section, it was subsequently used to estimate the concentration of fluorescence components 255 in highly colored samples for which the inner-effect correction may not optimal. Hence, for the 43 samples with absorbance ranging from 0.6 to up to 1.2 at 254 nm (cm⁻¹), the concentration of 256 257 tryptophan-like component C6 may have been slightly under-estimated whereas the concentration of 258 the remaining fulvic- and humic-like components may have been over-estimated by up to 20% (Miller et 259 al., 2010). Recent findings, however, suggest that the correction that we have performed adequately corrects for inner-filter effect for absorbance up to 1.5 cm⁻¹, such that the bias, if any, would actually be 260 261 much smaller (Kothawala et al., 2013a) Similar to the range of variation in Pd-DOC, the concentrations 262 in the different fluorescence components varied by several orders of magnitude across the sampled

sites, and thus the small bias that may be present in these 43 highly colored samples is unlikely toinfluence the patterns reported here.

265 3.3 Linking patterns in degradable DOC to DOM composition and nutrients

266 There were significant differences in the amounts and proportions of degradable DOC across 267 different types of systems (Figure 2), and a principal component analysis (PCA) further shows that lakes, 268 rivers and wetlands tend to group not only in terms of concentrations of total and degradable DOC, but 269 also in terms of DOM composition and nutrient concentrations (Fig. 5). The concentrations of DOC, 270 CDOM, the individual fluorescence components identified by the PARAFAC model, and nutrients all 271 tended to co-vary along a "concentration" axis (Component 1 of the PCA plot), which explained 45.4% of 272 the total variability in the measured variables (Fig. 5). Fluorescence components C1, C2 and C3, as well 273 as CDOM had the highest loadings on axis 1, and rivers and wetlands tended to have the highest scores 274 on this axis. The absolute amounts of Bd-DOC and Pd-DOC were also positively associated to this axis, 275 implying that systems with higher amounts of DOM and nutrients tended to have overall more 276 degradable DOC.

277 The links between DOC degradability and composition are captured by the second axis of the 278 PCA, which explained 23.7% of the variability. Concentrations of nutrients, as well as concentrations and 279 proportions of protein-like C5 and C6 are all situated in the top half of the PCA, along with the a250:365 280 absorbance ratio (proxy of low molecular weight) and Bd-DOC, whereas the amounts and proportions of 281 humic- or fulvic-like C1, C2 and C4, as well as SUVA254 and Pd-DOC, clustered in the lower half of the PCA 282 (Fig. 5); there was no clear system-specific distinction along this "composition" axis. These results 283 suggest that whereas the absolute amounts of biological and photochemical degradability are both 284 associated to the total amount of DOC, these two aspects of degradability are clearly linked to distinct pools within the bulk DOC. 285

286 3.4 The drivers of the concentrations of biologically and photo-chemically degradable DOC

287 The differential regulation of Bd-DOC and Pd-DOC is more clearly expressed by their direct 288 relationships with nutrients and the various DOM components (Table 1); pH and the concentration of 289 Chl a were not significant predictors when the variables presented in Table 1 were included. Likewise, 290 the inclusion of a categorical variable for system type (Lake, River, Wetland), or for the sampling region 291 (see Fig. 1) did not improve the fit in multiple regression models predicting either component of 292 degradability. There was thus no systematic ecosystem- or region-specific differences in the relationship 293 between Bd-DOC and its predictors, suggesting that the amount of biologically degradable DOC in the 294 studied systems is mainly a function of DOM composition and nutrients regardless of the type of aquatic 295 environment and the region where they lie.

296 Bd-DOC was strongly related to both DOM and nutrient concentrations (Fig. 5, Table 1), and TN 297 was the strongest single predictor (Table 1). Among the fluorescence components, the concentration of 298 protein-like component C6 was the strongest predictor of Bd-DOC (Model a, Table 1); no other 299 component had a significant effect on Bd-DOC once C6 was included. In contrast to Bd-DOC, DOM 300 optical properties played a greater role as predictors of Pd-DOC. The best MLR models for Pd-DOC 301 included a positive relationship with both the abundance and the relative proportions of component C3, 302 and negative relationships with the proportions of components C5 and C6 (Table 1). The EEMs analysis 303 allowed to identify specific pools of DOM that are particularly linked to ambient DOC biological or 304 photochemical degradability, but the concentration aspect was mostly captured by CDOM (measured as 305 absorbance at 440 nm, see methods) alone in both cases (Table 1, model c, g). In particular, when 306 concentrations of CDOM were included in MLR, no other variable associated to concentration of DOM 307 was significant in predicting Pd-DOC, but qualitative aspects of DOM, such as the percent contribution 308 of C3 and C6 (the latter negatively related) still significantly improved the model. 309 3.5 Factors influencing the relationship between the concentrations of biologically and photo-chemically

310 degradable DOC

The above patterns in nutrients and DOM concentration and composition were reflected in the residuals of the relationship between Bd-DOC and Pd-DOC. Sites with higher concentrations of CDOM and proportions of the very photo-chemically degradable fluorescence component C3 (see below) tended to contain more Pd-DOC per unit Bd-DOC (Table 1, model h). Conversely, sites with higher proportions of protein-like component C6 and higher nutrient concentrations tended to have a DOC pool that was more biologically degradable per unit Pd-DOC.

317 3.6 Susceptibility of DOM components to photochemical degradation

318 We further explored the photochemical degradability of specific pools of DOM in a subset of the 319 irradiation experiments. Component C3, which emerged as one of the main drivers of Pd-DOC, was 320 extremely photosensitive and was often completely removed after 12 to 24 hours of irradiation. In 321 contrast, components C5 and C6, the latter being a major driver of Bd-DOC (Table 1), appeared to be 322 largely un-reactive to irradiation, whereas the concentration of component C4 systematically increased 323 following irradiation, suggesting the photo-chemically mediated production of this component. 324 Components 1 and 2 showed an intermediate photochemical degradability, with an average loss of 24% 325 and 57%, respectively, of their initial fluorescence during incubations (Fig. 6). It is interesting to note 326 that these patterns of selective photochemical degradation of specific DOC components were very 327 consistent among ecosystem types and regions (see error bars in Fig. 6). 328 3.7 Patterns of biologically and photo-chemically degradable DOC across gradients of terrestrial 329 influence 330 Despite the fact that different pools of DOM were linked to the concentrations of Pd-DOC vs Bd-331 DOC (Table 1), CDOM itself accounted for as much variability in both Pd-DOC and Bd-DOC as any 332 combination of fluorescence components, in terms of concentration. Coherent with the PCA (Fig. 5), this

- 333 suggests that environments with high terrestrial influence also have higher concentrations and
- proportions of specific DOC pools associated to both Pd-DOC or Bd-DOC. Figure 5 shows the very strong

335 relationships that exist between CDOM and the photo-sensitive fluorescence components C2 and C3. 336 The relationship between CDOM and component C6, which was linked to Bd-DOC, was less obvious in 337 the PCA (Fig. 5). In this regard, there was an overall weak positive relationship between CDOM and C6 338 (Fig. 7a). Interestingly, the shape of this relationship varied greatly over a gradient of terrestrial influence (using CDOM as a proxy, see (Lapierre et al., 2013)). An analysis of discrete portions of this 339 340 gradient using moving window regression (MWR) showed that the relationship between C6 and CDOM 341 was not significant in low-CDOM environments (Fig. 7b), but became significant at CDOM (a440) concentrations of approximately 3 m⁻¹. Beyond this point, both the r² and the slope of the relationship 342 343 consistently increased with CDOM concentration until the slope stabilized when the lowest CDOM in the MWR was around 4 m⁻¹. There was thus a strong link between CDOM and C6 in systems with elevated 344 CDOM concentrations ranging from 4 to 30 m⁻¹. These results indicate that the amount of bio-labile 345 346 DOC, as reflected by the C6 component of DOM, is completely uncoupled to CDOM at low levels of 347 water color, but that the two become strongly coupled in systems with stronger terrestrial influence.

349 4. Discussion

350 4.1 Origin, age, freshness and degradability of DOC

351 The core result that we report here is that the concentrations of biologically and photo-352 chemically degradable DOC are uncoupled within any particular site, but that they nevertheless tend to 353 positively co-vary across large geographical and environmental gradients, and in particular, along a 354 gradient of terrestrial influence (Fig. 3b). These two functional groups of DOC are associated to 355 compositionally distinct pools that are subject to differential regulation across the landscape, and their 356 coupling is only apparent when both aspects of DOC degradability are assessed simultaneously over very 357 broad gradients of terrestrial influence and across a wide range of aquatic ecosystems, as we have done 358 in this study. This pattern likely reflects the expression of the different intrinsic DOC properties as well as 359 environmental conditions (Guillemette and del Giorgio, 2011; Marín-Spiotta et al., 2014) that are 360 encountered across the very diverse natural freshwater environments studied here.

361 The prevailing view on aquatic DOC biological degradability has been based on the notion that 362 material from autochthonous origin is typically fresh and highly degradable whereas material from 363 terrestrial origin tends to be old and highly altered, and thus does not contribute much to microbial 364 processes (Hedges et al., 1988;Dittmar and Kattner, 2003;Amon and Benner, 1996). Origin and age, 365 however, have different implications and different links to the notion of "freshness" (defined as the time 366 relative to when DOM left its site of production, e.g. soil, aquatic organisms, and was imported into the 367 aquatic environment) in different types of systems. For instance, in the open-ocean, freshly produced is 368 synonymous with autochthonous and young, whereas terrestrially derived implies leftovers of a pool 369 that entered the aquatic network months, years or even centuries before (Anderson and Williams, 370 1999), and is therefore both old and diagenetically altered. The terrestrially derived DOC pool in such 371 systems is not expected to contain significant amounts of highly labile (consumed within hours to days), 372 or even semi-labile (consumed within months to years, see Carlson (2002)) compounds. In contrast,

most continental aquatic environments are directly connected to a terrestrial source that delivers DOC
within a time frame of hours or days (Müller et al., 2013), and terrestrial DOC is therefore freshly
imported, but may still be either young or old (Mayorga et al., 2005;Raymond et al., 2007;Marín-Spiotta
et al., 2014). Recent work suggests that freshly imported terrestrial organic carbon can be readily
degraded in lakes and rivers even if it is extremely old (McCallister and del Giorgio, 2012;Kleber et al.,
2011), thus suggesting that age and origin may not be the best predictors of DOC biological degradability
across continental and marine waters.

380 Freshness thus appears to be a better common driver than age or origin to place apparently 381 differing patterns of DOC biological degradability on a common gradient that is independent of the type 382 of aquatic ecosystem studied. In this regard, studies in arctic rivers have shown that DOC is typically 383 more labile during the spring freshet (Mann et al., 2012;Holmes et al., 2008), when DOC has been 384 freshly imported into the aquatic environment. Likewise, DOC was more labile in temperate streams 385 characterized by low temperatures, presumably because it has been less altered than the DOC found in 386 warmer streams and thus conserved in a "fresher" state (Lu et al., 2013). Our results suggest that the 387 same basic pattern may apply not only seasonally, but also across the whole boreal aquatic network: 388 Close to the land-water interface, most of the biologically labile DOC appears to be of terrestrial origin, 389 as suggested by the increasingly tight coupling between CDOM (measured as absorbance at 440 nm, see 390 methods) and bio-labile component C6 as CDOM increases (Fig. 7). Furthermore, the highest 391 concentrations and proportions of both biologically and photo-chemically labile DOC are found in low 392 order rivers and wetlands (Fig. 2), supporting the notion that these are major hotspots for 393 biogeochemical processes in continental waters (Denfeld et al., 2013; Campeau et al., 2014). Systems 394 with greater CDOM content thus tend to contain higher concentrations of biologically and photo-395 chemically degradable DOC (Lapierre et al., 2013), because in such environments the DOM pool is 396 fresher and contains high amounts of photo-sensitive DOM (Fig. 6, Table 1), as well as bio-labile,

397 protein-like DOM (Fig. 7, Table 1), which interestingly, closely resembles DOM from autochthonous

398 origin (Stedmon et al., 2007;Lapierre and Frenette, 2009, Fig. 7).

399 4.2 The underlying basis of patterns in degradable DOC

400 The patterns in the concentrations of biologically and photo-chemically degradable DOC 401 reported here allow to identify DOM pools and environmental factors that are linked to DOC 402 degradability across boreal freshwater networks. In our study the amounts of Bd-DOC and Pd-DOC were 403 measured separately, but there are known synergistic effects between the biological and photochemical 404 processing of DOC in natural environments (Miller and Moran, 1997), wherein biological and 405 photochemical processes may operate on the same DOM pool (Koehler et al., 2012). The overall 406 degradation potential of DOC could thus be different in ambient waters where these processes actually 407 co-occur, as opposed to our experimental conditions where no such interaction was possible. The 408 objective of this study, however, was not to estimate the total concentration of degradable DOC, or the 409 realized expression of this potential degradability in situ, but rather to explore the large-scale patterns in 410 Bd-DOC and Pd-DOC and their respective drivers across a wide variety of aquatic environments in the 411 most comparable manner possible.

412 Both intrinsic DOM properties and environmental conditions appeared to play a role in determining the concentrations and relative proportions of degradable DOC observed in these boreal 413 414 freshwater environments. Bd-DOC was linked to nutrient concentration, and to DOM concentration and 415 composition (Fig. 5, Table 1). The influence of TN (and TP) on Bd-DOC may be threefold: 1) it may be 416 related to increased primary production and thus to the production of an autochthonous Bd-DOC 417 component (Descy et al., 2002;Demarty and Prairie, 2009); 2) it may be related to the stimulating effect 418 of nutrients on DOC degradation rates (Wickland et al., 2012;Guillemette et al., 2013); 3) it may be 419 related to DOM stoichiometry (Sun et al., 1997;Fellman et al., 2009). Chlorophyll a concentration, 420 however, was unrelated to Bd-DOC, and although we did not determine organic N, our measurements

421 show that the sum of ammonium, nitrates and nitrites represented on average 8% of TN, and never 422 more than 50%. These results thus suggest that an important fraction of the N in our systems was in the 423 form of DON and contained within the DOM itself along with DOC. The same is probably true for 424 phosphorus, although we do not have measurements of inorganic P to support this contention. It would 425 seem that the inclusion of N as a predictor of Bd-DOC is mostly (although not exclusively) reflecting 426 intrinsic DOM properties, i.-e. the stoichiometry and therefore quality of the DOM that was originally 427 loaded from land. This potential effect of DOM guality was reflected in absorbance and fluorescence 428 analyses, as protein-like C6 was the only DOM optical property that was related to Bd-DOC, consistent 429 with previous studies (Guillemette and del Giorgio, 2011;Fellman et al., 2008). The percent contribution 430 of C6 to total fluorescence was further strongly and positively related to the a250:a365 ratio, and 431 negatively to SUVA₂₅₄ (Fig. 5), consistent with previously reported relationships of those absorbance 432 properties with microbial processes such as bacterial production and growth efficiency (Berggren et al., 433 2008).

434 The patterns in Pd-DOC, on the other hand, were exclusively explained by DOM optical properties (Table 1). Extrinsic chemical factors are known to drive the photochemical degradation of 435 436 DOC (Porcal et al., 2014), and Pd-DOC was indeed significantly correlated with pH ($r^2 = 0.13$, p < 0.001, n 437 = 391) and iron concentration (r^2 = 0.35, p < 0.001, n = 216). These variables, however, were not 438 significant when included in multiple linear regression models containing DOM concentration or 439 composition, and CDOM entirely accounted for the concentration-effect of FDOM (Table 1), suggesting 440 that water color integrates several key biochemical properties that collectively determine 441 photochemical reactivity. The connection of Pd-DOC with CDOM or terrestrial influence in general is 442 intuitive, as land-derived DOM is typically associated to highly photo-reactive material with high humic 443 and lignin contents (Spencer et al., 2009b;Hernes et al., 2009;Stubbins et al., 2010). The average loss of 444 DOC in our incubations was 13%, whereas on average 41% of CDOM was lost during the same

445 experiments. This supports the widely accepted notion that colored DOM is preferentially removed by 446 photochemical oxidation (Graneli et al., 1996;Weyhenmeyer et al., 2012), and more importantly, 447 suggests that specific DOM pools (as shown by fluorescence components) contribute more than others 448 to the photochemical losses of DOC (Lu et al., 2013): the degradability of the various fluorescence 449 components (Fig. 6) mostly reflects the degree to which they absorb light in the UVA region (Fig. 4). 450 These results thus suggest a strong top-down control of DOM composition by photochemical processes, 451 which is reflected in the large-scale patterns of DOM composition across boreal freshwater ecosystems. 452 Certain fluorescence components were extremely photosensitive (Fig. 6); in particular, component C3 453 was typically completely degraded by light within 24h (Fig. 6). The ambient waters differ from 454 experimental conditions in terms of the light climate, but these results nonetheless suggest that there 455 are substantial losses of these fluorescence components in natural environments, and clearly indicate 456 that the photochemical loss rates of C3 are much faster than those of other components. This implies 457 that there must be a constant replenishment of this very reactive DOM across continental watersheds in 458 order to maintain the observed concentrations, such that the contribution of this highly photosensitive 459 DOM (e.g. C3) to aquatic biogeochemical processes is probably greater than what is suggested by its 460 ambient concentrations alone.

461 4.3 Linking DOM optical properties to degradability patterns

Previous work has shown that the links between the patterns of DOC consumption and the dynamics of specific DOM pools identified on the basis of fluorescence are much more complex than what is generally recognized, because most of these components appear to be both consumed and produced by biotic and abiotic processes across diverse aquatic environments (Guillemette and del Giorgio, 2012;Romera-Castillo et al., 2011;Kothawala et al., 2013b). In this regard, Fig. 6 shows a consistent behaviour of the various fluorescence components in response to irradiation across lakes, rivers and wetlands with very different environmental properties. It also shows that concentrations of

469	humic/fulvic component C4 systematically increased after exposure to sunlight, yet this component co-
470	varies positively and significantly with Pd-DOC ($r^2 = 0.30$, p < 0.001). Photosensitive component C3 (Fig.
471	5) also co-varies positively with Pd-DOC (Table 1), but clearly the mechanism underlying both
472	relationships is completely different.
473	Component C4 has an emission spectrum comparable to that of very photo-reactive C3, but the
474	excitation spectrum shifts towards shorter wavelengths (Fig. 4), suggesting that the former may be a
475	photoproduct of the latter. Given its excitation spectrum (Fig. 4), however, it is unlikely that component
476	C4 is unreactive to irradiation; Figure 6 most likely reflects the net balance of co-occurring
477	photochemical production and degradation processes rather than true reactivity, suggesting that DOC
478	that is potentially photo-reactive may be produced autochthonously in aquatic ecosystems via
479	photochemical processes. Likewise, it has been shown that photo-reactive components such as
480	component C3 reported here are also systematically produced by microbial processes during controlled
481	incubations (Guillemette and del Giorgio, 2012). To add complexity to this framework, not only may
482	biologically labile DOC be produced from the photochemical alteration of terrestrially derived DOC
483	(Bertilsson et al., 1999;Cory et al., 2013), but it can also be rendered more recalcitrant to microbial
484	degradation upon exposure to solar radiation (Tranvik and Bertilsson, 2001). It is difficult to quantify
485	those interactions, in particular the biotic (Guillemette and del Giorgio, 2012) and abiotic (Fig. 6)
486	production of Pd-DOC, but those different results nonetheless highlight the multiple processes, on land
487	and in the water, which fuel the aquatic pools of both Bd-DOC and Pd-DOC. Linking DOC degradability
488	measurements with DOM optical properties allows to uncover some of those interactions between
489	production and removal processes, which may not be detected purely based on the characterization of
490	the DOM found in ambient waters.
491	In the present study we have identified specific fluorescent DOM components associated to

492 either the biological or photochemical degradability of the ambient DOC pool across hundreds of lakes,

rivers and wetlands (Table 1, Figs. 3, 5). Our own work in this regard (Stubbins et al., submitted) suggests
that these fluorescent components involve not only the molecules responsible for the fluorescence
properties, but also that they tend to co-vary with a host of other, non-fluorescent compounds. The
molecules responsible for the actual fluorescence of these components may be acting both as
substrates, and perhaps more importantly, as tracers of a variety of non-fluorescent pools that share
similar sources and sinks, and which may themselves be the main substrates of these microbial
reactions.

500 In this regard, the very strong photochemical degradability of component C3 (Fig. 5), combined 501 with published evidence of consistent losses of a comparable component over a gradient of land cover 502 and water retention time in the landscape (Kothawala et al., 2013b;Lu et al., 2013), suggest that it is 503 indeed acting as a substrate for photochemical degradation, and that C3 is a causal driver of Pd-DOC in 504 freshwater environments. Our measurements, however, cannot confirm whether component C6 is 505 mostly driving Bd-DOC as a substrate for microbial processes or acting as a tracer of co-varying bio-labile 506 molecules. Regardless, our results indicate that Bd-DOC and Pd-DOC pools may be adequately tracked 507 by specific DOM properties across a large diversity of freshwater environments, presumably because 508 optical measurements allow to target meaningful DOM pools comprising different molecules which 509 share common production, importation, and removal processes.

510 4.4 Land as a source of both biologically and photo-chemically degradable DOC

Although it may appear intuitive that both Bd-DOC and Pd-DOC increase with overall DOC, the coupling between the concentrations of Bd-DOC and Pd-DOC implies that they share at least some basic sinks, or more likely, sources, in boreal watersheds. The biological and photochemical processes responsible for the removal of DOC from those boreal freshwaters are driven by different extrinsic variables and target different DOM pools (Table 1, Fig. 6), and the biological and photochemical degradability of DOC, expressed as the fraction of DOC that is biologically or photo-chemically

degradable in standardized conditions, are completely decoupled (Fig. 3a). The collective evidence
discussed above thus rather suggests that Bd-DOC and Pd-DOC partly share land as a common origin,
and the relatively weak coupling between both DOC pools suggest that the relative contribution of landderived DOC to Bd-DOC differs greatly across systems.

521 In this regard, autochthonous processes such as phytoplankton and macrophyte primary 522 production are known to generate fresh, protein-like DOM in freshwaters (Demarty and Prairie, 523 2009; Lapierre and Frenette, 2009), and more importantly, may supply the Bd-DOC pool independently 524 of terrestrial sources. This appears to be the case in several systems studied here, as the coupling 525 between Pd-DOC and Bd-DOC was weaker in lakes (Fig. 3b), and the amount of bio-labile DOM, 526 expressed in the patterns of component C6, was completely independent from CDOM in the lower 527 portion of the CDOM gradient (Fig. 7) where most lakes and some large rivers were found. A comparable 528 pattern has been reported in Swedish lakes (Kothawala et al., 2013b) where the equivalent protein-like 529 FDOM component appeared to be relatively invariable across gradients of lake DOC and water retention 530 time. Contrary to that study, however, we observed a stronger relationship between C6 and CDOM as 531 CDOM increased (Fig. 7), such that highly colored systems (mostly streams and wetlands) contained 532 much higher concentrations of component C6. The inclusion of rivers ranging from Strahler order 1 to 6, 533 and of a diversity of wetlands, greatly expands the range of terrestrial influence relative to what can be 534 observed based on lakes alone, and may explain the differences between our two studies. This may be a 535 combination of low macrophyte cover and algal biomass in these typically oligotrophic boreal systems, 536 the inhibition of autochthonous processes by CDOM-induced light limitation (Karlsson et al., 2009), and 537 perhaps more importantly, the high rate of supply of terrestrial labile DOC in highly colored systems 538 (Berggren et al., 2010a).

539 There would thus appear to be a baseline of autochthonous Bd-DOC that is always present in 540 those aquatic environments and varies more or less randomly across gradients of CDOM in response to

simultaneously increasing nutrients (Lapierre and del Giorgio, 2012) and light limitation (Karlsson et al., 541 542 2009). This baseline may be reflected in the significant intercept of the relationship between Bd-DOC 543 and Pd-DOC (Fig. 3b), which would suggest that in systems with very low terrestrial influence and almost 544 no Pd-DOC, there is still a significant pool of Bd-DOC that is likely from autochthonous origin. 545 Autochthonous sources may indeed be the main driver of Bd-DOC across narrow CDOM gradients, as 546 previously reported (Kritzberg et al., 2004;Guillemette et al., 2013;McCallister and del Giorgio, 2008), but the land-derived Bd-DOC might be the one pool which varies consistently across gradients of 547 548 terrestrial influence and may thus better explain the patterns in DOC degradability across continental 549 watersheds. These patterns collectively suggest that terrestrially derived Bd-DOC increases 550 proportionately faster than or independently from autochthonous Bd-DOC along a gradient of increasing 551 CDOM, such that land becomes an important, and even the main source of Bd-DOC as terrestrial 552 influence increases. This in turn may explain why bio-labile component C6 (and the molecules it tracks) 553 may actually accumulate and reach high concentrations in wetlands and rivers with very high terrestrial 554 influence, whereas in lakes it is typically decoupled from CDOM (Fig. 7), as well as from DOC 555 concentrations and water retention time (Kothawala et al., 2013b), since in the latter systems 556 production, importation and removal may be closer to steady-state.

557 Our results highlight the apparent links that tie terrestrial influence, water retention time and 558 DOC freshness with DOC composition and degradability across boreal freshwaters. In a previous study 559 we had shown that the concentrations of both Bd-DOC and Pd-DOC tended to increase with CDOM 560 content (Lapierre et al., 2013), resulting in increases in CO_2 flux. This result also suggested an additive 561 dynamic of DOC degradability along a gradient of terrestrial influence, and here we show that the role of 562 land as a major source of both biologically and photo-chemically degradable DOC to continental watersheds results in a pattern of co-variation between these two key pools of carbon across boreal 563 564 aquatic networks, despite fundamental contrasts in terms of their composition and regulation. The

- 565 potential of terrestrially derived organic carbon to contribute to ecological and biogeochemical
- 566 processes in boreal freshwaters is thus apparently not limited by its degradability, at least in
- 567 environments with high terrestrial influence where this carbon has been freshly imported.

569 Appendix A

570 Standardizing Pd-DOC based on average light dose in the experimental tubes

571 One of the main objective of this study was to assess the patterns in the concentrations of 572 photo-chemically degradable DOC, in the most standardized way possible. While the photochemical degradation experiments have been conducted in standardized conditions in terms of incoming light 573 574 dose, exposure time and temperature, the average light dose within the experimental tubes varied 575 depending on CDOM content which attenuated a fraction of the incoming light and resulted in self-576 shading. We tested the potential significance of this self-shading effect on the reported patterns by 577 comparing the uncorrected Pd-DOC values to light-standardized concentrations, which were obtained by 578 dividing each Pd-DOC concentration by the average light energy available (accounting for self-shading, 579 see below) in the corresponding tube in which the experiment was conducted.

580 Absorbance scans were obtained for half of an empty glass tube (to account for the energy lost 581 when passing through the glass, see Fig. A1) as well as for the colored dissolved organic matter (CDOM) 582 contained in each sample, as described in the Methods section. The tubes were round, horizontally 583 disposed in the solar simulator and had an inner diameter of 24 mm, so we calculated the proportion of 584 each wavelength that was remaining in ten different 2.4 mm-thick sections of the tube. The solar simulator light had a total light dose of 750 W m⁻² and a spectrum representative of the sunlight (Fig. 585 A1), which was used to calculate the total amount of light energy (in W m^{-2}) available in each section of 586 587 the experimental tube by multiplying the incoming light dose by (1 - fraction absorbed (by glass and 588 CDOM)). Only the 300-450 nm range of wavelengths was used. The lamp emitted substantial amounts of 589 longer wavelengths, which were not significantly absorbed by either the glass or CDOM, thus although 590 those wavelengths (450-750 nm) contribute a large fraction of the total energy delivered, they typically 591 do not contribute much to photochemical processes (Vähätalo et al., 2000). Including these wavelengths 592 would thus tend to attenuate the cross-sample differences in average light dose.

593 We then calculated a weighted average in the amount of light energy contained in the ten layers 594 based on the relative volume that each occupies in the tube. The average light dose varied little in the 595 tubes (see section 2.4 in Methods) but the measured concentrations in Pd-DOC among samples varied 596 by more than three orders of magnitude (Fig. 3b). As a result, although standardizing the raw Pd-DOC 597 values by the average effective light dose within the corresponding experimental tube resulted in a 598 certain bias when comparing the lowest to the highest CDOM samples, when these individual 599 differences are put in perspective of the whole set of Pd-DOC data (which ranges over several orders of 600 magnitude) the little amount of variation that is generated the relationship is barely noticeable. (Fig. 601 A2). There was a small amount of variation for the highest CDOM samples, but this potential variation is 602 arguably smaller than measurement error in other reported variables against which Pd-DOC is correlated. We thus chose to present uncorrected Pd-DOC values, which are more intuitively 603 comparable to the concentrations of Bd-DOC (in mg L^{-1}) that we report in this study. 604 605 606 **Appendix A Figure legend**

Figure A1. Comparing the lamp spectrum to light absorption from the glass and from CDOM. The same
scan was used to correct for absorption of light by glass for all the samples, whereas the corresponding
CDOM scan was used for each sample; the illustrated CDOM scan corresponds to a representative river
sample.

Figure A2. Comparing uncorrected *vs* light-standardized concentrations of Pd-DOC based on CDOM selfshading for wavelengths ranging 300-450 nm.

613

614 Acknowledgements

- This work is part of the Carbon Biogeochemistry in Boreal Aquatic Systems (CarBBAS) research chair, co-
- funded by the Natural Science and Engineering Research Council of Canada (NSERC) and Hydro-Québec.
- 617 We thank A. St-Pierre, A. Parkes and the CarBBAS team for the contribution to the field and laboratory
- 618 components of this study, and we thank P. Massicotte, T. Rasilo, C. Ruiz-Gonzalez, D. Vachon and F.
- 619 Guillemette for constructive discussions and comments on early versions of the manuscript.

620 References

- Algesten, G., Sobek, S., Bergstrom, A. K., Agren, A., Tranvik, L. J., and Jansson, M.: Role of lakes for
- organic carbon cycling in the boreal zone, Glob. Change Biol., 10, 141-147, 2004.
- Amon, R. M. W., and Benner, R.: Bacterial utilization of different size classes of dissolved organic matter,
- 624 Limnol. Oceanogr., 41, 41-51, 1996.
- Anderson, T. R., and Williams, P. J. I. B.: A one-dimensional model of dissolved organic carbon cycling in
- the water column incorporating combined biological-photochemical decomposition, Glob. Biogeochem.
- 627 Cycle, 13, 337-349, 10.1029/1999gb900013, 1999.
- APHA, A.: Standard methods for the examination of water and wastewater, 20, 1998.
- 629 Battin, T. J., Kaplan, L. A., Findlay, S., Hopkinson, C. S., Marti, E., Packman, A. I., Newbold, J. D., and
- 630 Sabater, F.: Biophysical controls on organic carbon fluxes in fluvial networks, Nat. Geosci., 1, 95-100,
- 631 10.1038/ngeo101, 2008.
- 632 Benner, R., and Kaiser, K.: Biological and photochemical transformations of amino acids and lignin
- phenols in riverine dissolved organic matter, Biogeochemistry, 102, 209-222, 10.1007/s10533-010-94354, 2011.
- Berggren, M., Laudon, H., and Jansson, M.: Hydrological Control of Organic Carbon Support for Bacterial
 Growth in Boreal Headwater Streams, Microb. Ecol., in press, 2008.
- 637 Berggren, M., Laudon, H., Haei, M., Strom, L., and Jansson, M.: Efficient aquatic bacterial metabolism of
- dissolved low-molecular-weight compounds from terrestrial sources, Isme J., 4, 408-416,
- 639 10.1038/ismej.2009.120, 2010a.
- 640 Berggren, M., Strom, L., Laudon, H., Karlsson, J., Jonsson, A., Giesler, R., Bergstrom, A. K., and Jansson,
- 641 M.: Lake secondary production fueled by rapid transfer of low molecular weight organic carbon from
- terrestrial sources to aquatic consumers, Ecol. Lett., 13, 870-880, 10.1111/j.1461-0248.2010.01483.x,
 2010b.
- 644 Berggren, M., Lapierre, J.-F., and del Giorgio, P. A.: Magnitude and regulation of bacterioplankton
- respiratory quotient across freshwater environmental gradients, ISME J, 6, 984-993,
- 646 10.1038/ismej.2011.157, 2012.
- 647 Bertilsson, S., and Tranvik, L. J.: Photochemically produced carboxylic acids as substrates for freshwater 648 bacterioplankton, Limnol. Oceanogr., 43, 885-895, 1998.
- 649 Bertilsson, S., Stepanauskas, R., Cuadros-Hansson, R., Graneli, W., Wikner, J., and Tranvik, L.:
- 650 Photochemically induced changes in bioavailable carbon and nitrogen pools in a boreal watershed,
- 651 Aquat. Microb. Ecol., 19, 47-56, 1999.

- 652 Brett, M. T., Kainz, M. J., Taipale, S. J., and Seshan, H.: Phytoplankton, not allochthonous carbon,
- sustains herbivorous zooplankton production, Proceedings of the National Academy of Sciences, 106,
 21197-21201, 10.1073/pnas.0904129106, 2009.
- 655 Butman, D., and Raymond, P. A.: Significant efflux of carbon dioxide from streams anr rivers in the 656 United States, Nat. Geosci., 4, 839-842, 10.1038/NGE01294, 2011.
- 657 Campeau, A., Lapierre, J.-F., Vachon, D., and del Giorgio, P. A.: Regional contribution of CO2 and CH4
- fluxes from the fluvial network in a lowland boreal landscape of Québec, Glob. Biogeochem. Cycle, 28,
- 659 57-69, 10.1002/2013gb004685, 2014.
- 660 Carlson, C. A.: Production and removal processes, in: Biogeochemistry of marine dissolved organic
- matter, edited by: Hansell, D. A., and Carlson, C. A., Elsevier, 2002.
- 662 Cole, J. J., Prairie, Y. T., Caraco, N. F., McDowell, W. H., Tranvik, L. J., Striegl, R. G., Duarte, C. M.,
- 663 Kortelainen, P., Downing, J. A., Middelburg, J. J., and Melack, J.: Plumbing the global carbon cycle:
- Integrating inland waters into the terrestrial carbon budget, Ecosystems, 10, 171-184, 10.1007/s10021006-9013-8, 2007.
- 666 Cory, R. M., and McKnight, D. M.: Fluorescence spectroscopy reveals ubiquitous presence of oxidized
- and reduced quinones in dissolved organic matter, Environ. Sci. Technol., 39, 8142-8149,
- 668 10.1021/es0506962, 2005.
- 669 Cory, R. M., Crump, B. C., Dobkowski, J. A., and Kling, G. W.: Surface exposure to sunlight stimulates CO2
- release from permafrost soil carbon in the Arctic, Proceedings of the National Academy of Sciences, 110,
 3429-3434, 10.1073/pnas.1214104110, 2013.
- 672 Cuthbert, I. D., and del Giorgio, P.: Toward a strandard method of measuring color in fresh-water,
- 673 Limnol. Oceanogr., 37, 1319-1326, 1992.
- del Giorgio, P. A., Cole, J. J., Caraco, N. F., and Peters, R. H.: Linking planktonic biomass and metabolism
- to net gas fluxes in northern temperate lakes, Ecology, 80, 1422-1431, 1999.
- 676 Demarty, M., and Prairie, Y. T.: In situ dissolved organic carbon (DOC) release by submerged
- macrophyte-epiphyte communities in southern Quebec lakes, Can. J. Fish. Aquat. Sci., 66, 1522-1531,
 10.1139/f09-099, 2009.
- Denfeld, B. A., Frey, K. E., Sobczak, W. V., Mann, P. J., and Holmes, R. M.: Summer CO 2 evasion from
 streams and rivers in the Kolyma River basin, north-east Siberia, Polar Research, 2013.
- 681 Descy, J.-P., Leporcq, B., Viroux, L., François, C., and Servais, P.: Phytoplankton production, exudation
- and bacterial reassimilation in the River Meuse (Belgium), J. Plankton Res., 24, 161-166,
- 683 10.1093/plankt/24.3.161, 2002.
- Dittmar, T., and Kattner, G.: The biogeochemistry of the river and shelf ecosystem of the Arctic Ocean: a
- 685 review, Mar. Chem., 83, 103-120, http://dx.doi.org/10.1016/S0304-4203(03)00105-1, 2003.
- 686 Fellman, J. B., D'Amore, D. V., Hood, E., and Boone, R. D.: Fluorescence characteristics and
- 687 biodegradability of dissolved organic matter in forest and wetland soils from coastal temperate
- 688 watersheds in southeast Alaska, Biogeochemistry, 88, 169-184, 10.1007/s10533-008-9203-x, 2008.
- 689 Fellman, J. B., Hood, E., D'Amore, D. V., Edwards, R. T., and White, D.: Seasonal changes in the chemical
- 690 quality and biodegradability of dissolved organic matter exported from soils to streams in coastal
- temperate rainforest watersheds, Biogeochemistry, 95, 277-293, 10.1007/s10533-009-9336-6, 2009.
- 692 Findlay, S., and Sinsabaugh, R. L.: Aquatic ecosystems: Interactivity of dissolved organic matter Aquatic
- 693 Ecology, Academic press, San Diego, 2002.
- 694 Graneli, W., Lindell, M., and Tranvik, L.: Photo-oxidative production of dissolved inorganic carbon in
- lakes of different humic content, Limnol. Oceanogr., 41, 698-706, 1996.
- 696 Guillemette, F., and del Giorgio, P. A.: Reconstructing the various facets of dissolved organic carbon
- bioavailability in freshwater ecosystems, Limnol. Oceanogr., 56, 734-748, 10.4319/lo, 2011.

- 698 Guillemette, F., and del Giorgio, P. A.: Simultaneous consumption and production of fluorescent
- dissolved organic matter by lake bacterioplankton, Environmental Microbiology, 14, 1432-1443,
- 700 10.1111/j.1462-2920.2012.02728.x, 2012.
- Guillemette, F., McCallister, S. L., and del Giorgio, P. A.: Differentiating the degradation dynamics of algal
- and terrestrial carbon within complex natural dissolved organic carbon in temperate lakes, Journal of
- 703 Geophysical Research: Biogeosciences, 118, 963-973, 10.1002/jgrg.20077, 2013.
- Hedges, J. I., Clark, W. A., and Cowie, G. L.: Fluxes and reactivities of organic-matter in a coastal marine
- 705 bay, Limnol. Oceanogr., 33, 1137-1152, 1988.
- Hernes, P. J., Bergamaschi, B. A., Eckard, R. S., and Spencer, R. G. M.: Fluorescence-based proxies for
- lignin in freshwater dissolved organic matter, J. Geophys. Res.-Biogeosci., 114, 10.1029/2009jg000938,
 2009.
- Holmes, R. M., Aminot, A., Kérouel, R., Hooker, B. A., and Peterson, B. J.: A simple and precise method
- for measuring ammonium in marine and freshwater ecosystems, Can. J. Fish. Aquat. Sci., 56, 1801-1808,
 10.1139/f99-128, 1999.
- Holmes, R. M., McClelland, J. W., Raymond, P. A., Frazer, B. B., Peterson, B. J., and Stieglitz, M.: Lability
- of DOC transported by Alaskan rivers to the arctic ocean, Geophys. Res. Lett., 35, 5,
- 714 L0340210.1029/2007gl032837, 2008.
- Jansson, M., Persson, L., De Roos, A. M., Jones, R. I., and Tranvik, L. J.: Terrestrial carbon and
- 716 intraspecific size-variation shape lake ecosystems, Trends Ecol. Evol., 22, 316-322,
- 717 10.1016/j.tree.2007.02.015, 2007.
- Jonsson, A., Meili, M., Bergstrom, A. K., and Jansson, M.: Whole-lake mineralization of allochthonous
- and autochthonous organic carbon in a large humic lake (Ortrasket, N. Sweden), Limnol. Oceanogr., 46,
 1691-1700, 2001.
- 721 Karlsson, J., Jonsson, A., Meili, M., and Jansson, M.: Control of zooplankton dependence on
- allochthonous organic carbon in humic and clear-water lakes in northern Sweden, Limnol. Oceanogr.,

723 48, 269-276, 2003.

- Karlsson, J., Bystrom, P., Ask, J., Ask, P., Persson, L., and Jansson, M.: Light limitation of nutrient-poor
 lake ecosystems, Nature, 460, 506-509, 10.1038/nature08179, 2009.
- 726 Kleber, M., Nico, P. S., Plante, A., Filley, T., Kramer, M., Swanston, C., and Sollins, P.: Old and stable soil
- 727 organic matter is not necessarily chemically recalcitrant: implications for modeling concepts and
- temperature sensitivity, Glob. Change Biol., 17, 1097-1107, 2011.
- 729 Koehler, B., Wachenfeldt, E., Kothawala, D., and Tranvik, L. J.: Reactivity continuum of dissolved organic
- carbon decomposition in lake water, Journal of Geophysical Research: Biogeosciences (2005–2012), 117,
 2012.
- 732 Kothawala, D. N., Murphy, K. R., Stedmon, C. A., Weyhenmeyer, G. A., and Tranvik, L. J.: Inner filter
- correction of dissolved organic matter fluorescence, Limnol. Oceanogr.: Methods, 11, 616-630, 2013a.
- 734 Kothawala, D. N., Stedmon, C. A., Müller, R. A., Weyhenmeyer, G. A., Köhler, S. J., and Tranvik, L. J.:
- 735 Controls of dissolved organic matter quality: Evidence from a large-scale boreal lake survey, Glob.
- 736 Change Biol., 20, 1101-1114, 10.1111/gcb.12488, 2013b.
- 737 Kritzberg, E. S., Cole, J. J., Pace, M. L., Graneli, W., and Bade, D. L.: Autochthonous versus allochthonous
- carbon sources of bacteria: Results from whole-lake C-13 addition experiments, Limnol. Oceanogr., 49,
 588-596, 2004.
- 740 Lapierre, J. F., and Frenette, J. J.: Effects of macrophytes and terrestrial inputs on fluorescent dissolved
- 741 organic matter in a large river system, Aquat. Sci., 71, 15-24, 10.1007/s00027-009-9133-2, 2009.
- Lapierre, J. F., and del Giorgio, P. A.: Geographic and environmental drivers of regional differences in the
- 743 lake *p*CO₂ versus DOC relationship across northern landscapes, J. Geophys. Res.-Biogeosci., 117,
- 744 10.1029/2012JG001945, 2012.

- Lapierre, J. F., Guillemette, F., Berggren, M., and del Giorgio, P.: Increases in terrestrially derived carbon
- stimulate organic carbon processing and CO2 emissions in boreal aquatic ecosystems, Nature
- 747 communications, 10.1038/ncomms3972, 2013.
- 748 Lu, Y., Bauer, J. E., Canuel, E. A., Yamashita, Y., Chambers, R. M., and Jaffé, R.: Photochemical and
- 749 microbial alteration of dissolved organic matter in temperate headwater streams associated with
- different land use, Journal of Geophysical Research: Biogeosciences, 118, 566-580, 10.1002/jgrg.20048,
 2013.
- 752 Mann, P. J., Davydova, A., Zimov, N., Spencer, R. G. M., Davydov, S., Bulygina, E., Zimov, S., and Holmes,
- 753 R. M.: Controls on the composition and lability of dissolved organic matter in Siberia's Kolyma River
- basin, J. Geophys. Res.-Biogeosci., 117, 15, G01028
- 755 10.1029/2011jg001798, 2012.
- 756 Marín-Spiotta, E., Gruley, K. E., Crawford, J., Atkinson, E. E., Miesel, J. R., Greene, S., Cardona-Correa, C.,
- 757 and Spencer, R. G. M.: Paradigm shifts in soil organic matter research affect interpretations of aquatic
- carbon cycling: transcending disciplinary and ecosystem boundaries, Biogeochemistry, 117, 279-297,
- 759 10.1007/s10533-013-9949-7, 2014.
- 760 Marker, A., and Nusch, E.: Rai. H. & B. Riemann. 1980. The measurement of photosynthetic pigments in
- 761 freshwaters and standardization of methods: conclusion and recommendation, Arch. Hydrobiol. Beih.
- 762 Ergebn. Limnol, 14, 91-106, 1980.
- Massicotte, P., and Frenette, J.-J.: Spatial connectivity in a large river system: resolving the sources and
 fate of dissolved organic matter, Ecol. Appl., 21, 2600-2617, 2011.
- 765 Mayorga, E., Aufdenkampe, A. K., Masiello, C. A., Krusche, A. V., Hedges, J. I., Quay, P. D., Richey, J. E.,
- and Brown, T. A.: Young organic matter as a source of carbon dioxide outgassing from Amazonian rivers,
 Nature, 436, 538-541, 10.1038/nature03880, 2005.
- 768 McCallister, S. L., and del Giorgio, P. A.: Direct measurement of the delta C-13 signature of carbon
- respired by bacteria in lakes: Linkages to potential carbon sources, ecosystem baseline metabolism, and
- 770 CO2 fluxes, Limnol. Oceanogr., 53, 1204-1216, 2008.
- 771 McCallister, S. L., and del Giorgio, P. A.: Evidence for the respiration of ancient terrestrial organic C in
- northern temperate lakes and streams, Proceedings of the National Academy of Sciences, 109, 1696316968, 10.1073/pnas.1207305109, 2012.
- Miller, M. P., Simone, B. E., McKnight, D. M., Cory, R. M., Williams, M. W., and Boyer, E. W.: New light on
 a dark subject: comment, Aquat. Sci., 72, 269-275, 10.1007/s00027-010-0130-2, 2010.
- 776 Miller, W. L., and Moran, M. A.: Interaction of photochemical and microbial processes in the degradation
- of refractory dissolved organic matter from a coastal marine environment, Limnol. Oceanogr., 42, 13171324, 1997.
- Molot, L. A., and Dillon, P. J.: Photolytic regulation of dissolved organic carbon in northern lakes, Glob.
 Biogeochem. Cycle, 11, 357-365, 1997.
- 781 Müller, R. A., Futter, M. N., Sobek, S., Nisell, J., Bishop, K., and Weyhenmeyer, G. A.: Water renewal
- along the aquatic continuum offsets cumulative retention by lakes: implications for the character of
- 783 organic carbon in boreal lakes, Aquat. Sci., 75, 535-545, 10.1007/s00027-013-0298-3, 2013.
- 784 Murphy, K. R., Butler, K. D., Spencer, R. G. M., Stedmon, C. A., Boehme, J. R., and Aiken, G. R.:
- 785 Measurement of Dissolved Organic Matter Fluorescence in Aquatic Environments: An Interlaboratory
 786 Comparison, Environ. Sci. Technol., 44, 9405-9412, 10.1021/es102362t, 2010.
- 787 Murphy, K. R., Stedmon, C. A., Wenig, P., and Bro, R.: OpenFluor–an online spectral library of auto-
- fluorescence by organic compounds in the environment, Anal. Methods, 6, 658-661, 2014.
- 789 Obernosterer, I., and Benner, R.: Competition between biological and photochemical processes in the
- mineralization of dissolved organic carbon, Limnol. Oceanogr., 49, 117-124, 2004.

- 791 Porcal, P., Dillon, P. J., and Molot, L. A.: Interaction of extrinsic chemical factors affecting
- photodegradation of dissolved organic matter in aquatic ecosystems, Photochem. Photobiol. Sci., 13,
 799-812, 10.1039/c4pp00011k, 2014.
- 793 799-812, 10.1039/c4pp00011k, 2014.
- Raymond, P. A., McClelland, J., Holmes, R., Zhulidov, A., Mull, K., Peterson, B., Striegl, R., Aiken, G., and
- 795 Gurtovaya, T.: Flux and age of dissolved organic carbon exported to the Arctic Ocean: A carbon isotopic
- study of the five largest arctic rivers, Glob. Biogeochem. Cycle, 21, 10.1029/2007GB002934, 2007.
- 797 Romera-Castillo, C., Sarmento, H., Álvarez-Salgado, X. A., Gasol, J. M., and Marrasé, C.: Net production
- and consumption of fluorescent colored dissolved organic matter by natural bacterial assemblages
- growing on marine phytoplankton exudates, Applied and environmental microbiology, 77, 7490-7498,2011.
- Schmidt, M. W., Torn, M. S., Abiven, S., Dittmar, T., Guggenberger, G., Janssens, I. A., Kleber, M., Kögel-
- Knabner, I., Lehmann, J., and Manning, D. A.: Persistence of soil organic matter as an ecosystem
 property, Nature, 478, 49-56, 2011.
- 804 Shutova, Y., Baker, A., Bridgeman, J., and Henderson, R. K.: Spectroscopic characterisation of dissolved
- 805 organic matter changes in drinking water treatment: From PARAFAC analysis to online monitoring
- 806 wavelengths, Water Res., 54, 159-169, http://dx.doi.org/10.1016/j.watres.2014.01.053, 2014.
- 807 Soumis, N., Lucotte, M., Larose, C., Veillette, F., and Canuel, R.: Photomineralization in a boreal
- 808 hydroelectric reservoir: a comparison with natural aquatic ecosystems, Biogeochemistry, 86, 123-135,
- 809 10.1007/s10533-007-9141-z, 2007.
- Spencer, R. G. M., Aiken, G. R., Butler, K. D., Dornblaser, M. M., Striegl, R. G., and Hernes, P. J.: Utilizing
- 811 chromophoric dissolved organic matter measurements to derive export and reactivity of dissolved
- 812 organic carbon exported to the Arctic Ocean: A case study of the Yukon River, Alaska, Geophys. Res.
- 813 Lett., 36, 10.1029/2008gl036831, 2009a.
- Spencer, R. G. M., Stubbins, A., Hernes, P. J., Baker, A., Mopper, K., Aufdenkampe, A. K., Dyda, R. Y.,
- 815 Mwamba, V. L., Mangangu, A. M., Wabakanghanzi, J. N., and Six, J.: Photochemical degradation of
- dissolved organic matter and dissolved lignin phenols from the Congo River, J. Geophys. Res.-Biogeosci.,
- 817 114, 10.1029/2009jg000968, 2009b.
- 818 Stedmon, C. A., Markager, S., and Bro, R.: Tracing dissolved organic matter in aquatic environments
- using a new approach to fluorescence spectroscopy, Mar. Chem., 82, 239-254, 10.1016/s03044203(03)00072-0, 2003.
- 821 Stedmon, C. A., Markager, S., Tranvik, L., Kronberg, L., Slatis, T., and Martinsen, W.: Photochemical
- 822 production of ammonium and transformation of dissolved organic matter in the Baltic Sea, Mar. Chem.,
- 823 104, 227-240, 10.1016/j.marchem.2006.11.005, 2007.
- 824 Stedmon, C. A., and Bro, R.: Characterizing dissolved organic matter fluorescence with parallel factor
- analysis: a tutorial, Limnol. Oceanogr. Meth., 6, 572-579, 2008.
- 826 Stubbins, A., Spencer, R. G., Chen, H., Hatcher, P. G., Mopper, K., Hernes, P. J., Mwamba, V. L.,
- 827 Mangangu, A. M., Wabakanghanzi, J. N., and Six, J.: Illuminated darkness: Molecular signatures of Congo
- 828 River dissolved organic matter and its photochemical alteration as revealed by ultrahigh precision mass
- spectrometry, Limnol. Oceanogr., 55, 1467, 2010.
- Stubbins, A., Lapierre, J. F., Berggren, M., Prairie, Y. T., Dittmar, T., and del Giorgio, P. A.: What's in an
 EEM? Molecular signatures of dissolved organic matter fluorophores, submitted.
- 832 Sun, L., Perdue, E., Meyer, J., and Weis, J.: Use of elemental composition to predict bioavailability of
- dissolved organic matter in a Georgia river, Limnol. Oceanogr., 42, 714-721, 1997.
- 834 Tranvik, L. J., and Bertilsson, S.: Contrasting effects of solar UV radiation on dissolved organic sources for
- 835 bacterial growth, Ecol. Lett., 4, 458-463, 2001.
- 836 Vähätalo, A. V., Salkinoja-Salonen, M., Taalas, P., and Salonen, K.: Spectrum of the quantum yield for
- 837 photochemical mineralization of dissolved organic carbon in a humic lake, Limnol. Oceanogr., 45, 664-
- 838 676, 2000.

- Wallin, M. B., Grabs, T., Buffam, I., Laudon, H., Ågren, A., Öquist, M. G., and Bishop, K.: Evasion of CO2
- from streams The dominant component of the carbon export through the aquatic conduit in a boreal
 landscape, Glob. Change Biol., 19, 785-797, 10.1111/gcb.12083, 2013.
- 842 Ward, N. D., Keil, R. G., Medeiros, P. M., Brito, D. C., Cunha, A. C., Dittmar, T., Yager, P. L., Krusche, A. V.,
- and Richey, J. E.: Degradation of terrestrially derived macromolecules in the Amazon River, Nature
- 844 Geosci, 6, 530-533, 10.1038/ngeo1817, 2013.
- 845 Wenzel, A., Bergström, A.-K., Jansson, M., and Vrede, T.: Poor direct exploitation of terrestrial
- particulate organic material from peat layers by Daphnia galeata, Can. J. Fish. Aquat. Sci., 69, 1870-1880,
 10.1139/f2012-110, 2012.
- 848 Weyhenmeyer, G. A., Fröberg, M., Karltun, E., Khalili, M., Kothawala, D., Temnerud, J., and Tranvik, L. J.:
- 849 Selective decay of terrestrial organic carbon during transport from land to sea, Glob. Change Biol., 18,
- 850 349-355, 10.1111/j.1365-2486.2011.02544.x, 2012.
- Wickland, K., Aiken, G., Butler, K., Dornblaser, M., Spencer, R., and Striegl, R.: Biodegradability of
- dissolved organic carbon in the Yukon River and its tributaries: Seasonality and importance of inorganic
- 853 nitrogen, Glob. Biogeochem. Cycle, 26, 10.1029/2012GB004342, 2012.
- Wickland, K. P., Neff, J. C., and Aiken, G. R.: Dissolved organic carbon in Alaskan boreal forest: Sources,
- chemical characteristics, and biodegradability, Ecosystems, 10, 1323-1340, 10.1007/s10021-007-9101-4,
- 856 2007.

Table 1. Multiple linear regression models predicting the concentrations of photo-chemically and
biologically degradable DOC across boreal lakes, rivers and wetlands. Abbreviations are defined in the
text. Fluorescence components in square brackets correspond to the absolute concentration of the
specified component. The sign in parentheses denotes the sign of the coefficient in the MLR model.

Response variable	Model	Predictors	n	adj. r²	AIC
Bd-DOC	а	(+)[C6]	323	0.35	-6864
	b	(+)TN	319	0.40	-683
	с	(+)TN, (+)CDOM	317	0.45	-696
	d	(+)TN, (+)[C6]	317	0.45	-693
	е	(+)TN, (+)[C6], (+)TP	316	0.46	-699
Pd-DOC	f	(+) [C3], (-) %C5, (-) %C6	392	0.67	-818
	g	(+) CDOM, (+) %C3, (-) %C6	392	0.73	-887
Residuals of Pd-DOC					
vs Bd-DOC	h	(+)%C3, (+)CDOM, (-)%C6, (-)TP	264	0.27	-406

865 Figure legends

Figure 1. Distribution of the sampling sites across seven boreal regions (James Bay, Abitibi, Laurentians,
Saguenay, Chibougamau, Schefferville) of Quebec, Canada. Grey lines and area represent large rivers
and lakes; green lines delimit the main watersheds in Quebec.

Figure 2. Amounts and proportions of biologically and photo-chemically degradable DOC across the

three main types of systems studied. Lines represent the 25, 50 and 75 percentiles, points, 5 and 95

percentiles. Different letters denote statistically different (p < 0.01) means across groups, based on

ANOVA and Tukey-Kramer post-hoc analyses. Bd-DOC (percent and absolute) : n = 201 for lakes, n = 71

873 for rivers, n = 53 for wetlands. Pd-DOC (percent and absolute): n = 193 for lakes, n = 136 for rivers, n =

874 67 for wetlands.

Figure 3. Relationship between the proportions (a) and amounts (b) of biologically and photo-chemically

degradable DOC across boreal lakes, rivers and wetlands. a) n = 262, not significant. b) lakes: log(Bd-

877 DOC) = 0.16log(Pd-DOC) - 1.67, p = 0.0003, r² = 0.11, n = 161; rivers: (log(Bd-DOC) = 0.88log(Pd-DOC) -

1.59, p < 0.0001, r² = 0.33, n = 58); wetlands (log(Bd-DOC) = 0.78log(Pd-DOC) - 1.34, p < 0.0001, r² = 0.23,

879 n = 50; overall relationship: log(Bd-DOC) = 0.71 log(Pd-DOC) -1.48, p < 0.001, n = 268.

880 Figure 4. Fluorescence signatures of the components identified by the PARAFAC model. Fluorescence

peaks are observed at the following wavelengths (nm), for excitation and emission, respectively

882 (secondary peak in parentheses): C1: 275, 424; C2: 275(340), 484; C3: 275(345), 436; C4: 334, 446; C5:

883 300, 388; C6: 275, 334

884 Figure 5. Principal component analysis of concentrations of degradable DOM, DOM optical properties

and nutrients across boreal lakes, rivers and wetlands. Abbreviations are defined in the text.

886 Fluorescence components in square brackets correspond to the absolute concentration of the specified887 component.

888	Figure 6. Average loss of specific fluorescence components based on the difference in concentration
889	before and after irradiation for samples originating from boreal lakes, rivers and wetlands. Rates have
890	been reported on a per day basis based on the average of the FDOM loss measured after 12 and 24
891	hours; this allowed to better quantify the photochemical susceptibility of C3 which was commonly
892	completely degraded after 24 hours, hence the average per day loss of over 100 %. n = 121
893	Figure 7. Concentrations of protein-like C6 as a function of CDOM in lakes, rivers and wetlands. a: r ² =
894	0.32, p < 0.001, n = 519. b: Parameters of moving window regressions performed on discrete portions of
895	the dataset based on increasing CDOM. Total sample size was 519, but individual regressions were
896	performed on subsets of 250 samples; a total of 270 regressions were thus performed. "x" signs denote
897	non-significant regressions. X axis shows the lowest CDOM concentration included in the regression for
898	which the parameters are reported. The first regression was thus performed on samples for which
899	CDOM ranged from 0.12 to 3.11 m ⁻¹ , whereas the last regression was performed on samples for which
900	CDOM ranged from 5.55 to 30.0. Note that the slope characterizes the log(C6) vs log(CDOM)
901	relationship.



904 Figure 1.



906 Figure 2.















917Fluorescence component concentration (R.u.)918Figure 6.

















927 Figure A2.