

1 **X-ray fluorescence mapping of mercury on suspended mineral particles and**
2 **diatoms in a contaminated freshwater system**

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10
11 **Abstract**

12 Mercury (Hg) bioavailability and geochemical cycling is affected by its partitioning between the
13 aqueous and particulate phases. We applied a synchrotron-based X-ray fluorescence (XRF)
14 microprobe to directly visualize and quantify the spatial localization of Hg and its correlations
15 with other elements of interest on suspended particles from a Hg contaminated freshwater
16 system. Up to 175 $\mu\text{g/g}$ Hg is found on suspended particles, but $< 0.01\%$ of which is in the form
17 of methylmercury. Mercury is heterogeneously distributed among phytoplankton (e.g., diatoms)
18 and mineral particles that are rich in iron oxides and natural organic matter (NOM). The diatom-
19 bound Hg is mostly found on outer surfaces of the cells, suggesting passive sorption of Hg on
20 diatoms. Our results indicate that localized sorption of Hg onto suspended particles, including
21 diatoms and NOM-coated oxide minerals, may play an important role in affecting the
22 partitioning, reactivity, and biogeochemical cycling of Hg in natural aquatic environments.

1 **1 Introduction**

2 Mercury (Hg) is a global pollutant. Certain anaerobic bacteria can transform inorganic Hg into
3 neurotoxic monomethylmercury (MeHg) (Hu et al., 2013b; Parks et al., 2013), which is the most
4 worrisome because MeHg can be readily bioaccumulated in food chains to levels harmful to both
5 humans and wildlife (Barkay and Wagner-Dobler, 2005; Mason et al., 1995; Watras and Bloom,
6 1992). Suspended particulates including both colloidal minerals and phytoplankton (such as
7 diatoms) are important carriers of Hg and MeHg in freshwater ecosystems (Adams et al., 2009;
8 Balogh et al., 2008; Choe et al., 2003; Pickhardt and Fisher, 2007; Plourde et al., 1997). Nearly
9 90% of total Hg in water has been reported to be suspended-particle bound (Balogh et al., 2008;
10 Choe et al., 2003; Pickhardt and Fisher, 2007), but the exact localization and chemical
11 characteristics of particulate-Hg have been poorly studied to date. Similarly in the contaminated
12 East Fork Poplar Creek (EFPC) at the U.S. Department of Energy's (DOE) Y-12 National
13 Security Complex (NSC) in Oak Ridge, Tennessee, about 75–95% of the Hg is associated with
14 suspended particles (Brooks and Southworth, 2011) (Supplement 1), although the range of
15 particle-bound Hg varies with season, flow conditions, and the distance from the contamination
16 source. In fresh water lakes and stream systems, studies have also shown that Hg mainly
17 associates with particulate natural organic matter (NOM) and iron and aluminum oxyhydroxides
18 (Adams et al., 2009; Quemerais et al., 1998), resulting in elevated Hg concentrations on these
19 particles since Hg sorption on phyllosilicate minerals is relatively low (Hintelmann and Harris,
20 2004; Senevirathna et al., 2011). The association of Hg with riverine particles is usually
21 attributed to passive adsorption, and both abiotic and biological processes are important for this
22 process (Mason et al., 1996; Pickhardt and Fisher, 2007). However, the accumulation of MeHg
23 by phytoplankton is attributed primarily to an active uptake process, resulting in the
24 biomagnification of Hg in aquatic food chains from phytoplankton to fish (Barkay and Wagner-

1 Dobler, 2005; Mason et al., 1995; Watras and Bloom, 1992). For example, in a study of the
2 bioaccumulation of inorganic Hg and MeHg by freshwater phytoplankton, Pickhardt and Fisher
3 (2007) found that the volume concentration factors (VCFs) for the inorganic Hg on
4 phytoplankton ranged from 0.5 to 5×10^4 , whereas VCFs for MeHg were about 20–30 times
5 higher, ranging from 1.3 to 15×10^5 . Additionally, these authors showed that about 84 to 91% of
6 inorganic Hg is associated with cell surfaces (Pickhardt and Fisher, 2007). Similarly, Mason et
7 al. (1996) reported that Hg(II) is principally bound to phytoplankton membranes, whereas MeHg
8 is largely accumulated inside the cell, in the cytoplasm.

9 Hg association with phytoplankton cell membranes has been traditionally determined by the
10 difference between total Hg in the whole cell and that in the cytoplasmic fraction of lysed cells
11 that are obtained following mechanical grinding or sonification and separation (Mason et al.,
12 1996; Pickhardt and Fisher, 2007). This is partly because the very low Hg concentrations in
13 natural water that result in a relatively low level of Hg on cell surfaces; thus to assay the low
14 cell-surface bound Hg, highly sensitive techniques such as cold-vapor atomic absorption or
15 fluorescence spectroscopy are needed for quantification. However, complications may occur in
16 such assays due to possible interactions between cell wall-associated Hg and cytoplasmic Hg,
17 resulting in re-partitioning (sorption or desorption) of Hg on cell membranes. More importantly,
18 these techniques cannot discern the localization of sorbed Hg in natural samples consisting of
19 both mineral and phytoplankton particles. Whether the sorbed Hg is evenly or heterogeneously
20 distributed on the particles is unknown. Techniques such as X-ray fluorescence (XRF)
21 microprobes could potentially provide direct and accurate localization and characterization of Hg
22 on suspended particles of mixed mineral or organic origins. Such techniques have been used to
23 map two-dimensional (2D) and 3D elemental distributions over length scales from sub-

1 micrometers to tens of millimeters, covering microbial cells, soil, and plant roots (Bernaus et al.,
2 2006; Blute et al., 2004; De Jonge et al., 2010; Kemner et al., 2004; Terzano et al., 2010). The
3 spatial resolution and elemental sensitivity of X-ray fluorescence extend below 100 nm and are
4 able to detect, for example, arsenic at 30–1200 $\mu\text{g/g}$ on cattail root plaques (Blute et al., 2004)
5 and as low as 1 $\mu\text{g/g}$ within a single bacterial cell (Kemner et al., 2004). Furthermore, the
6 technique allows simultaneous detection of multiple elements and can provide a powerful tool to
7 map their spatial distribution and correlations.

8 In this study, we used XRF microprobes to examine Hg distributions and its correlations with
9 multiple elements of interest on both diatoms and mineral particles. To our knowledge, this is the
10 first time XRF microprobes have been used to map the localization of Hg on suspended particles
11 in a contaminated freshwater ecosystem. We hypothesized that localized sorption of Hg occur
12 on suspended particles with different binding affinities and that particulate organic matter or
13 NOM-coated iron oxyhydroxides sorb more Hg than those of phyllosilicate minerals.
14 Complementary techniques including inductively coupled plasma mass spectrometry (ICP-MS),
15 Fourier transform infrared (FTIR), scanning electron microscopy (SEM) and elemental mapping
16 were used to determine the bulk loading of Hg and MeHg on particles and the association of
17 NOM with minerals. Results of this study are expected to have important implications in
18 determining Hg partitioning, geochemical cycling, and bioavailability in the natural aquatic
19 environment.

20

21 **2 Materials and methods**

22 Suspended particles from the contaminated EFPC water in Oak Ridge, Tennessee, were obtained
23 during two sampling campaigns along a 2.5 km stretch from its headwater within the Y-12 NSC

1 (Brooks and Southworth, 2011; Miller et al., 2009). Past industrial operations and processes at
2 site resulted in the release of more than 200 metric tons of Hg into the water and sediments
3 (Barnett et al., 1997; Brooks and Southworth, 2011; Miller et al., 2009). The total Hg
4 concentration in the headwater remains high at ~1000 ng/L. The water in EFPC is a Ca-Mg-
5 HCO₃ type composition with a slightly alkaline pH ranging from 7.4 to 8.0. Creek water was
6 first collected in plastic bottles and transported to the laboratory in a cooler on ice. During the
7 first sampling in early spring, suspended solids were collected at three locations including
8 EFK25.9, EFK25.1, and EFK23.5, which are approximately 0.1, 0.9, and 2.5 km, respectively,
9 from the headwater, with a total Hg concentration in water decreasing from ~1000 to 200 ng/L
10 (Brooks and Southworth, 2011; Miller et al., 2009). The total particle concentration in water was
11 determined by collecting and weighing the particulate matter on a 0.2- μ m membrane filter after
12 drying. Suspended particles were separated in two different size fractions (between 0.2 and 3
13 μ m, and >3 μ m) using membrane filtration. The >3 μ m fraction contained mostly diatoms and
14 large mineral particles or aggregates, whereas the < 3 μ m fraction consisted of mostly mineral
15 particles. During the second sampling in late fall, samples were collected only at the EFK23.5
16 location, and particles were separated by centrifugation at 4000 rpm in 250-mL bottles and then
17 resuspended in a small amount of the creek water in 3 mL vials. They were stored in a freezer (-
18 20°C) until use. Total Hg and MeHg concentrations on suspended solids were analyzed using
19 established procedures (described below).

20 Size and morphological properties of suspended particles were subsequently evaluated with a
21 Zeiss Merlin scanning electron microscopy (SEM) operated at 3 kV. Particulate samples were
22 mounted by allowing a small drop of the suspension to evaporate either on a silicon nitride
23 membrane or a copper transmission electron microscopy (TEM) grid with carbon-Formvar

1 coatings. Energy-dispersive X-ray (EDX) spectra of selected diatoms and mineral particles were
2 recorded at 20 kV with a Quantax microanalysis system (Bruker-AXS Microanalysis GmbH,
3 Berlin, Germany), and electron backscattering elemental analysis was also performed with the
4 same system. Fourier transform infrared (FTIR) spectroscopy of selected samples in the size
5 fraction between 0.2 and 3 μm was performed with a Nicolet Magna 760 spectrophotometer
6 (Nicolet Instrument Corp.). A small droplet of particulate samples was placed onto a ZnSe
7 window and allowed to dry before the window was inserted in the IR beam for analysis. The
8 spectral resolution was 4 cm^{-1} .

9 Localization and 2D XRF elemental maps were determined on two sets of selected diatom and
10 particulate samples, with the first set (collected in the spring at EFK23.5) used as is and the other
11 collected in the late fall at the same location but amended with Hg(II) (as HgCl_2 , 2.9 μM) at pH
12 7.8 (same as in the creek water) for comparisons. The final sorbed Hg concentration was 617
13 $\mu\text{g/g}$ dry wt, which is about 3–10 times higher than that observed in natural EFPC water. The
14 XRF analysis was performed at beamline 2ID-D at the Advanced Photon Source at Argonne
15 National Laboratory, United States (Cai et al., 2000; Kemner et al., 2004). Fresnel zone plates
16 are used as focusing optics in hard X-ray microprobes at energies typically between 6 and 30
17 keV. Highly focused X-rays are used to quantify and map trace elements in heterogeneous
18 materials with low detection limits at the sub-micron spatial resolution. The microprobe excites
19 the specimen with an intense X-ray beam and measures the energies and intensities of emitted X-
20 rays, allowing quantification of the concentration of elements in a heterogeneous sample matrix.

21 The X-ray probe energy was set to 12,300 eV (16 eV above Hg L_{III} edge) and the sample
22 chamber was flushed with He for the X-ray measurement, as described previously (Glasauer et
23 al., 2007). The size of the beam at the focal point was 150 nm (V) \times 200 nm (H) at FWHM (full

1 width at half maximum), determined by scans with a Cr knife-edge. Step size for the final raster
2 scans of the diatoms and particulates were 100 nm. Incident and transmitted beam intensities
3 were monitored with air-filled ionization detectors. XRF signals from the samples were detected
4 by a single-element Ge detector placed perpendicular to the incident beam direction in the plane
5 of polarization.

6 The XRF concentration maps of elements from Si to Hg were obtained, although only those of
7 Si, Ca, S, Fe, Mn, Zn, Hg, P, and Cl were presented because of their relatively high
8 concentrations. The XRF peaks of interest were fitted by modified Gaussian profiles to separate
9 the XRF signal from background in the spectrum. Quantitative elemental analysis in several
10 regions of each sample was done by averaging the multi-channel analyzer (MCA) spectra
11 collected within these regions, subtracting the averaged MCA spectra for regions outside the
12 diatoms or mineral particles (representative of an experimental background signal) and then
13 integrating the counts within each region of interest. The integrated MCA counts were converted
14 to area concentrations of elements in each sample by comparing the XRF intensities between the
15 samples and thin glass film standards 1832 and 1833 [obtained from National Institute of
16 Standards and Technology (NIST)] measured under the same experimental conditions and
17 normalized to incident X-ray beam intensity (Cai et al., 2000; Kemner et al., 2004). Under given
18 operation conditions, the estimated detection limits were about 10^{-4} $\mu\text{g}/\text{cm}^2$ for Hg, $\sim 3 \times 10^{-4}$ to
19 2×10^{-3} $\mu\text{g}/\text{cm}^2$ for Mn, Fe, Ni, and Zn, and ~ 2 $\mu\text{g}/\text{cm}^2$ for Si, P, and Cl (Ortega et al., 2004;
20 Twining et al., 2003). We note however that the detection limit is affected by factors such as the
21 incident energy used to probe the sample, the atomic number of the element, and the sample
22 matrix.

1 For the determination of bulk total Hg, MeHg, and elemental compositions on particles, the first
2 set of samples were separated into two size fractions of (i) greater than 3 μm and (ii) greater than
3 0.2 μm but less than 3 μm with corresponding membrane filters (Supor and GF/C Glass Fiber).
4 Particles were digested in a 1:1 mixture of concentrated nitric and hydrochloric acids. Bromine
5 monochloride (BrCl) was added to half of the digestion mixture for the determination of total Hg
6 by cold vapor atomic fluorescence spectrometry (CVAFS) (Miller et al., 2009; Zheng et al.,
7 2012). Briefly, hydroxyammonium hydrochloride ($\text{NH}_2\text{OH}\cdot\text{HCl}$) was added to the digest to
8 remove free halogens followed by the addition of stannous chloride (SnCl_2) to reduce Hg(II) to
9 gaseous Hg(0) , which was trapped onto a gold trap and subsequently thermally desorbed into a
10 N_2 gas stream and analyzed by CVAFS (Gu et al., 2011; Hu et al., 2013b). The remaining digest
11 was diluted and used to determine the total elemental compositions including major ions of Fe,
12 Al, Mn, Ca, Mg, and Zn, by inductively coupled plasma-mass spectrometry (ICP-MS, Perkin-
13 Elmer). MeHg concentrations were determined based on a modified U.S. EPA Method 1630
14 involving the distillation of samples followed by ethylation, gas chromatographic separation, and
15 detection by ICP-MS (Hu et al., 2013b; Parks et al., 2013).

16

17 **3 Results and discussion**

18 Total suspended particles ($>0.2 \mu\text{m}$) collected from the EFPC ranged from about 3 mg/L to 5.5
19 mg/L under steady flow conditions. These particles comprised a mixture of diatoms (usually >5
20 μm) and mineral particulates (Fig. 1a) and their dominant elemental compositions were Si, Al,
21 O, and C as determined by EDX analysis (Fig. 1b) (Note that Cu was an artifact resulting from
22 the use of copper grid sample holders). Using acid digestion coupled with ICP-MS analysis, we
23 determined the dominant metal ions in samples were Fe, Al, Mg, Mn, and Zn (Fig. 1c), with the

1 smaller size fraction (between 0.2 and 3 μm) showing slightly higher amounts of Fe and Al on a
2 dry weight basis. The presence of zinc is consistent with the industrial effluents entering the
3 headwaters of EFPC. As expected, a large percentage of Hg (>80%) is associated with particles,
4 and the highest Hg concentration observed was $\sim 175 \mu\text{g/g}$ dry weight (Fig. 1d). The smaller size
5 fraction (between 0.2 and 3 μm) also showed slightly higher levels of sorbed total Hg and MeHg
6 than those of the larger size fraction (>3 μm), except samples collected 0.9 km downstream of
7 the headwater (EFK25.1). The MeHg concentrations in these samples were generally low or
8 below the detection limit (Fig. 1d) and represents only a tiny fraction (<0.01%) of the total Hg
9 associated with particles.

10 Despite Hg concentrations lower than typically amenable to X-ray techniques, we were able to
11 detect and map Hg on surfaces of these particles by XRF on individual diatoms and mineral
12 particles (Figs. 2 and 3). Four individual diatoms with different morphological properties and
13 Hg-loadings (with or without Hg additions) were studied along with mineral particulate clusters
14 (Fig. 4). We analyzed Hg on diatoms and mineral particles by conducting XRF microscopy
15 measurements and signal collection both above and below the Hg L_{III} edge to confirm that the
16 observed Hg signal is not an artifact due to spectral leakage from other elements. Hg appears to
17 be co-localized with Fe, Mn, S, Zn, P, Cl, and Ca in most cases but not with Si on both diatoms
18 and mineral particles (Fig. 3, details are shown on individual sub-figures of the lower left in 3a
19 and of the upper left in 3b). The correlations between Hg and P, Cl, S, and Ca are consistent with
20 the association of Hg with biomass either as cellular components or preferred ligands (in the case
21 of S) (Glasauer et al., 2007). This is also shown in the electron backscattering analysis of these
22 elements (Supplement 2). Other elements were not shown due to their relatively low
23 concentrations or insensitivity to XRF microscopy. Provided in Table 1 is the average elemental

1 compositions on the entire diatom and selected mineral particles determined with XRF in Figs.
2 2–4. On a micron scale (at 100-nm resolution), quantification of Hg concentrations on diatoms
3 and minerals revealed highly heterogeneous distribution, particularly on mineral particles with
4 localized areas of high Hg concentrations. For diatoms, with the exception of locations in the
5 lower part of the diatoms in Fig. 3, Hg is predominantly associated with cell walls or outer
6 membranes. This observation suggests that surface adsorption of Hg(II) is the main process for
7 Hg uptake by diatoms since > 99% of the Hg in EFPC water is in the form of inorganic Hg(II)
8 (Fig. 1d) (Brooks and Southworth, 2011). It agrees with previous studies, which showed that
9 passive adsorption is the principal mechanism for the membrane-bound Hg on diatoms or
10 phytoplankton since both living and heat-killed cells have similar VCFs for inorganic Hg (Mason
11 et al., 1996; Pickhardt and Fisher, 2007; Watras and Bloom, 1992). The result is also similar to
12 the adsorption of such metal ions as Zn^{2+} , which is predominately sorbed onto diatom
13 membranes, rather than Si frustule (Gelabert et al., 2007), and thus explains the co-localization
14 of Hg and Zn since both of them are soft metals. As for MeHg, the VCFs were 1.5–5 times
15 higher in living cells than dead, thus an active uptake by phytoplankton cells has been suggested
16 to be the main mechanism (Pickhardt and Fisher, 2007).

17 The elevated concentrations of Hg noted on the diatoms in Fig. 3 likely result from mineral
18 particulates such as iron oxides laying on or attached to cells, as evidenced by elevated
19 concentrations of Fe, S, Ca, and Mn in those areas in the image. In general, when normalized to
20 the surface area, average Hg loadings on the mineral particles are higher than the average Hg
21 loadings on the entire diatom (Table 1). Additional analyses of several selected mineral particles
22 (Fig. 4a) indicate a highly heterogeneous distribution of Hg. On many particles (Fig. 4a, upper
23 two-thirds of the image) Hg was not detected, whereas on others (lower center region), up to

1 about 0.14 $\mu\text{g}/\text{cm}^2$ Hg was measured (Table 1). Similar to observations on diatoms, Hg on
2 mineral particles was co-localized with Fe, Mn, S, and Zn, but not with Si. These correlations are
3 not completely surprising and can be attributed to the presence of Fe- and Mn-oxide minerals
4 and the associated NOM coatings and microbial biomass, which are known to strongly sorb Hg
5 (Dong et al., 2011; Hu et al., 2013a; Hu et al., 2013b)).

6 Significant sorption of NOM on Fe- and Mn-oxide minerals is expected because of their strong
7 binding affinities, as shown in previous studies (Fu and Quan, 2006; Gu et al., 1994; 1995;
8 Parfitt et al., 1977; Schwertmann et al., 1986). Strong association between NOM and naturally-
9 occurring Fe- or Mn-oxyhydroxides is commonly observed in water (Balogh et al., 2008; Gu et
10 al., 1994; Hintelmann and Harris, 2004; Quemerais et al., 1998). The dissolved organic matter
11 concentration in EFPC water is ~ 3 mg/L (Dong et al., 2010; Miller et al., 2009). Using the
12 electron backscattering elemental analysis we can clearly distinguish the organic carbon (C)
13 coating on mineral particles (< 3 μm) (Fig. 4b) and, more importantly, the distribution of C in the
14 image appears well correlated with those of Fe, Al, and O. Additionally, the FTIR spectrum of
15 mineral particles showed strong absorbance bands in the region between 1000 and 1700 cm^{-1}
16 (Fig. 4c), indicating the presence of organic matter with carboxylic and phenolic carbon moieties
17 (at ~ 1640 cm^{-1}) (Fu and Quan, 2006; Gu et al., 1994; 1995; Tejedor-Tejedor and Anderson,
18 1990). The broad band between 900 cm^{-1} and 1200 cm^{-1} also suggests the presence of organic
19 matter such as polysaccharide C–O functional groups that are likely associated with minerals (Fu
20 and Quan, 2006; Gu et al., 1994; 1995; Parfitt et al., 1977). The absorbance peaks at about 1030,
21 800, 750, and 700 cm^{-1} could be attributed to the presence of minerals such as lepidocrocite and
22 goethite (Parfitt et al., 1992; Schwertmann and Wolska, 1990).

1 These observations and analyses (Fig. 4b,c) suggest that NOM-coated Fe-oxyhydroxide minerals
2 may serve as strong sorption sites for Hg through the formation of Hg-NOM complexes. In other
3 words, these organic materials may have acted as a bridging agent by forming the Hg-(S-NOM-
4 O)-Fe-oxide ternary complexes because the complexation between Fe-oxides and NOM involves
5 primarily the carboxyl and hydroxyl functional groups (Fu and Quan, 2006; Gu et al., 1994;
6 1995; Tejedor-Tejedor et al., 1992), whereas Hg binding with NOM occurs through reduced
7 sulfur functional groups on NOM (Dong et al., 2011; Gu et al., 2011; Miller et al., 2009; Nagy et
8 al., 2011; Skyllberg et al., 2006). As a result, it led to greatly increased Hg loadings on these
9 particles (Table 1).

10

11 **4 Conclusions**

12 XRF microprobe imaging is shown to provide a useful tool for directly visualizing and
13 quantifying the localization of Hg and its correlations to elemental distributions on suspended
14 particles of both living organisms (e.g., diatoms) and inorganic minerals. Relatively high levels
15 of localized Hg on particles (e.g., > 50 $\mu\text{g/g}$) may be needed for XRF analysis, and this level of
16 Hg could be found in contaminated water bodies such as the EFPC water with dissolved Hg
17 concentrations of >100 ng/L and the San Francisco Bay Delta waters (Pickhardt and Fisher,
18 2007). The technique is particularly useful to identify the presence or absence of certain minerals
19 and particles such as cinnabar or metacinnabar based on the clustering of Hg with S or other
20 elements (Bernaus et al., 2006; Terzano et al., 2010). It is important to realize the ubiquitous
21 presence of NOM and its association with iron oxyhydroxides, which can enhance Hg uptake by
22 suspended particles in freshwater ecosystems. These organic materials sorb onto Fe and Al oxide
23 surfaces via carboxyl or hydroxyl functional groups, whereas sulfhydryl functional groups in
24 NOM form complexes with Hg(II). Such intimate association between Hg and NOM and

1 suspended particles could potentially control the partitioning and reactivity of Hg and may thus
2 have important implications in determining the geochemical cycling and bioavailability of Hg in
3 natural aquatic environments.

4 **Supplementary material related to this article is available online at**

5 <http://www.biogeosciences.net/>

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8

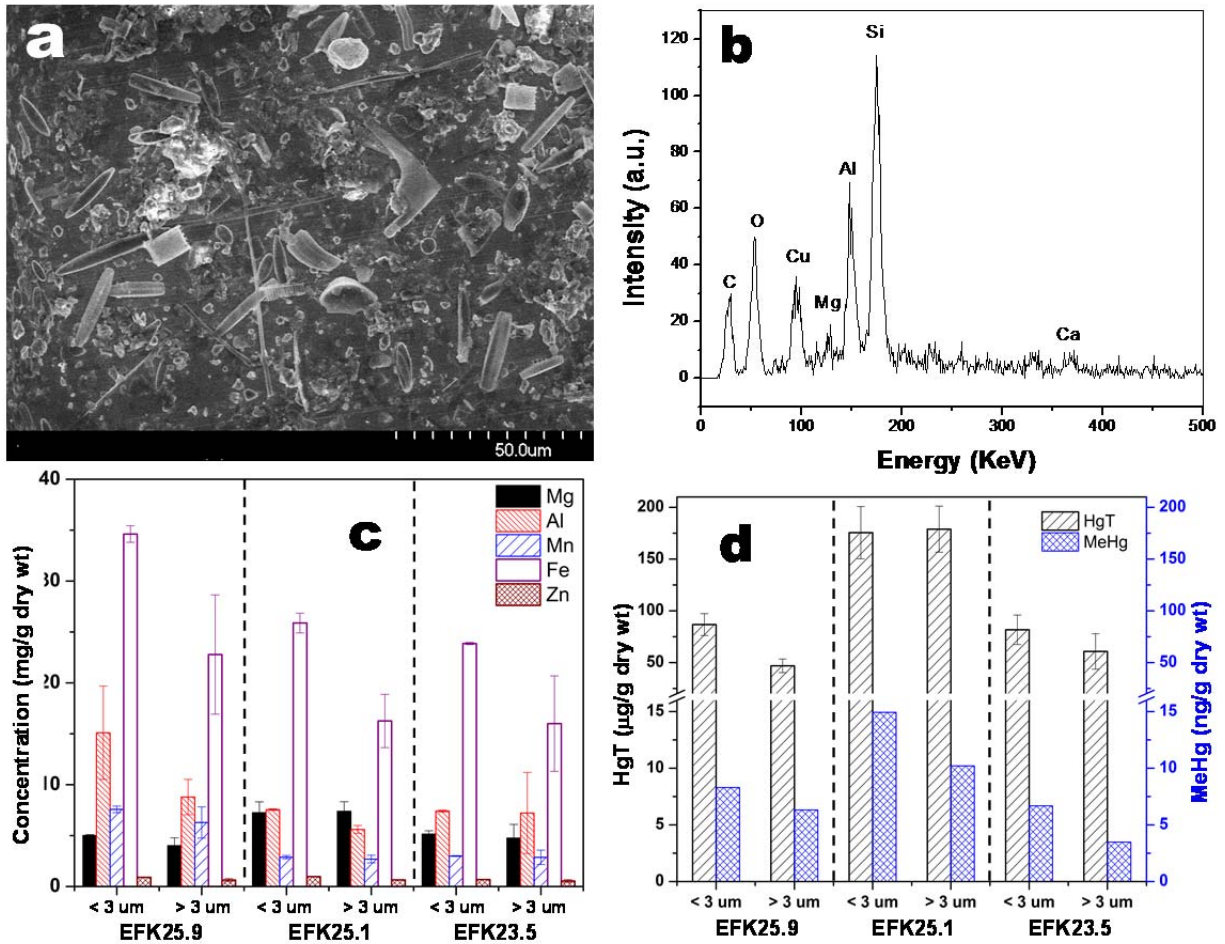
1 Table 1. The average elemental compositions ($\mu\text{g}/\text{cm}^2$) on the entire diatom and selected
 2 mineral particles determined by XRF in Figures 2–4.
 3

Elements	Figure 2a diatom	Figure 2b diatom	Figure 3a diatom	Figure 3b diatom	Figure 3a mineral (lower left)	Figure 3b mineral (top left)	Figure 4a mineral particles
Si	32	34	32.245	24	5.256	6.0	4.8
P	0.506	0.637	0.126	0.168	0.355	0.220	2.112
S	0.508	0.824	0.149	0.217	0.380	0.387	1.968
Cl	0.998	1.559	0.643	0.651	0.662	0.728	1.352
Ca	1.926	2.152	0.900	1.060	2.138	2.040	3.9
Mn	0.145	0.0569	0.098	0.315	1.560	0.991	1.206
Fe	0.479	0.388	0.800	2.923	9.243	5.434	12.591
Zn	0.220	0.0588	0.190	0.799	0.259	0.147	0.29
Hg	0.006	0.005	0.032	0.092	0.154	0.142	0.138

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1 **Figure 1.**

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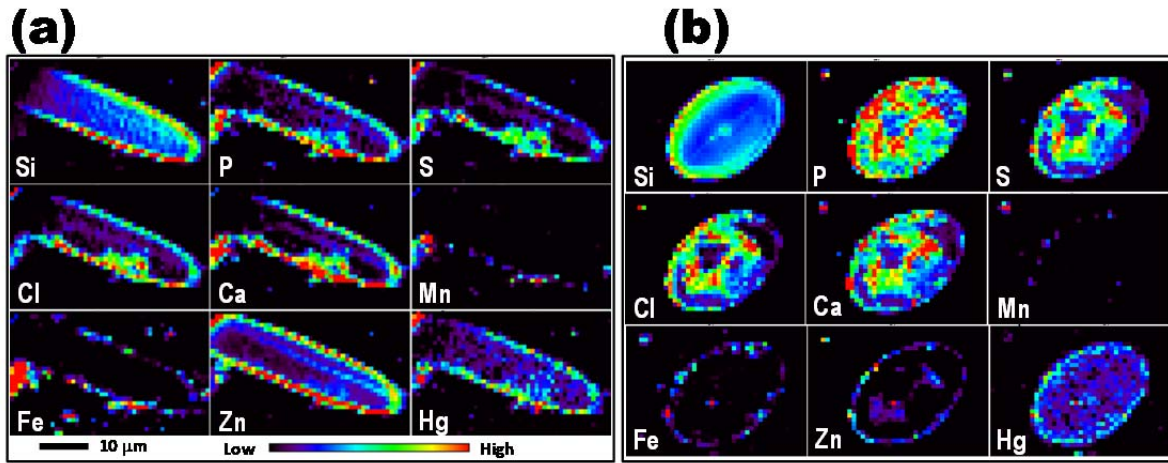
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6 Figure 1. (a) SEM image of suspended particulates containing both diatoms and mineral particles
7 from EFPC in Oak Ridge, TN; (b) Energy dispersive X-ray (EDX) analysis of particulates in (a),
8 showing dominant elemental composition of Si, Al, O, and C (Note: Cu signal is from the use of
9 copper grid sample holders); (c) Bulk analysis of major cations on EFPC particles collected from
10 three locations (EFK25.9, EFK25.1, EFK23.5) and two size fractions by ICP-MS following
11 digestion in 1:1 concentrated HCl:HNO₃; (d) Distribution of total Hg (HgT) and MeHg
12 associated with EFPC particles of two different size fractions.

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Figure 2.



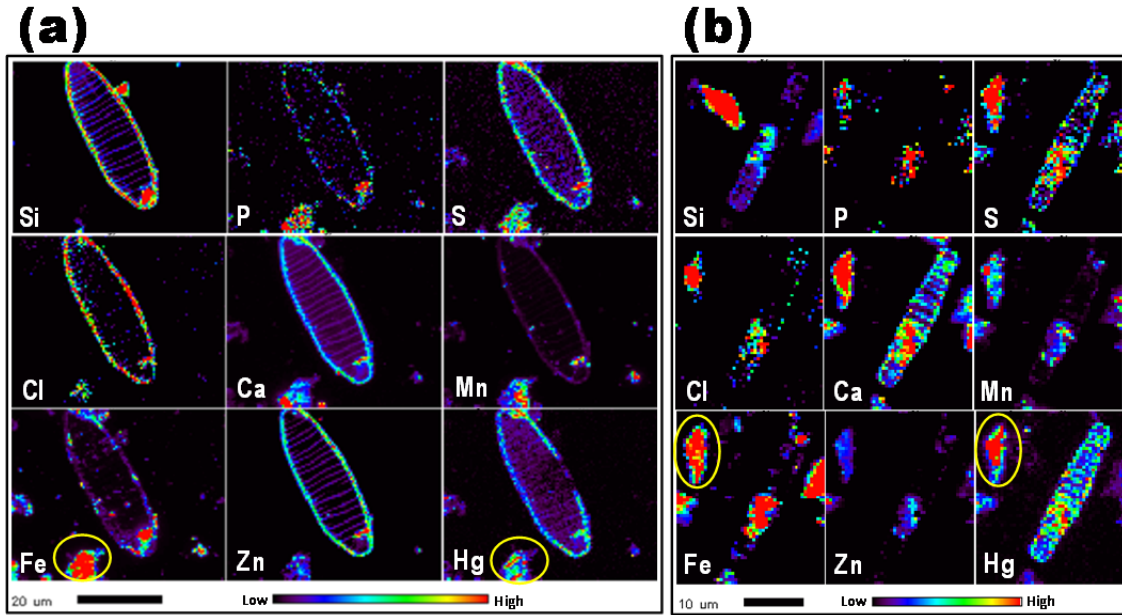
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Figure 2. Direct X-ray microprobe fluorescence imaging and elemental analysis of two individual diatoms collected from a Hg-contaminated EFPC creek (EFK23.5) without any treatment. Only selected elements of Hg, Zn, Fe, Mn, Ca, Cl, S, P, and Si were presented due to their relatively high concentrations. The average total Hg content is about 70 μg/g particles (dry wt).

1 **Figure 3.**

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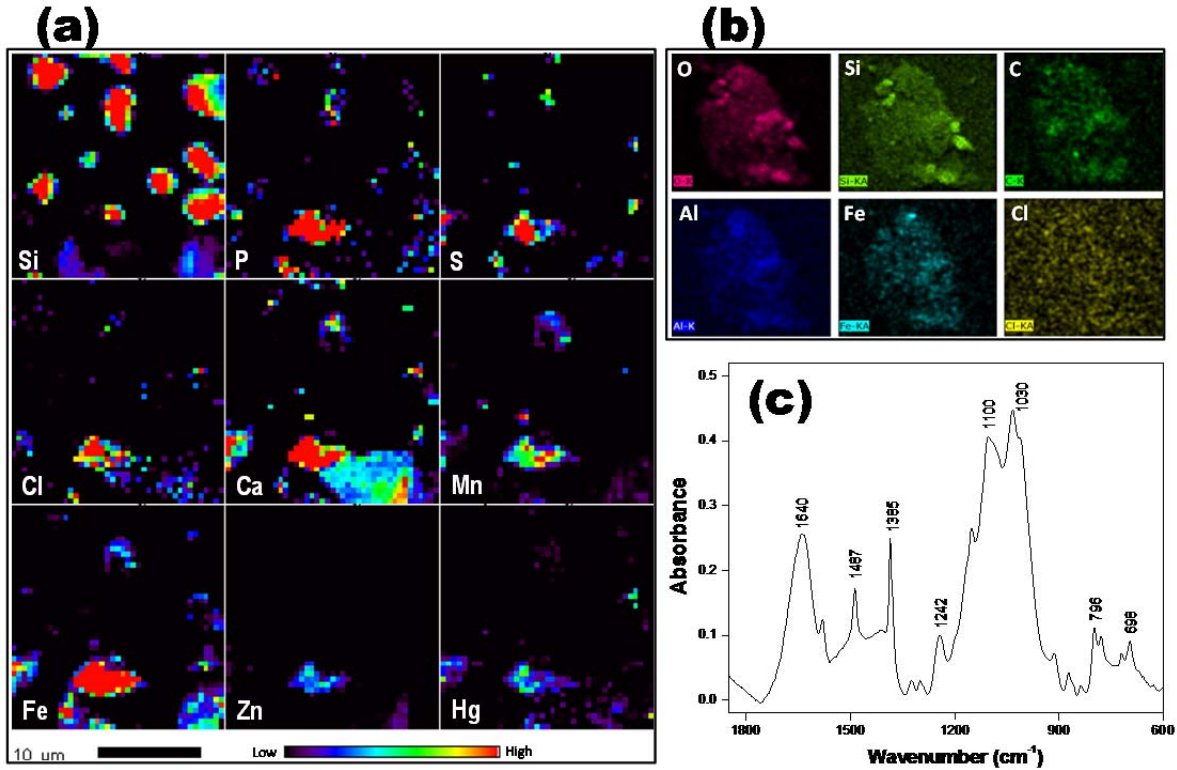
6

7 Figure 3. X-ray microprobe fluorescence imaging and elemental analysis of two individual
8 diatoms and mineral particles with different morphologies collected from a Hg-contaminated
9 EFPC creek (EFK23.5). Both samples were spiked with Hg to a final sorbed Hg concentration of
10 617 μg/g (dry wt) on average (see text for additional details). Iron-rich mineral particles can be
11 seen at the lower left of the image (a) and upper left of the image (b) (yellow circles).

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1 **Figure 4.**

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Figure 4. Experiments used to support the hypothesis that organic matter (carbon) is associated with iron oxide minerals and acts as a bridge for Hg sorption. (a) X-ray microprobe fluorescence imaging and elemental analysis (e.g., Hg, Fe, etc.) of selected mineral particles (from EFK23.5) spiked with Hg at 617 μg/g (dry wt); (b) scanning electron microscope (SEM) backscattering elemental analysis of a selected cluster of mineral particles showing the association between carbon and Fe, and (c) FTIR spectroscopic analysis of natural organic matter associated with mineral particles.