

1 **Stable isotope ratio ($^{13}\text{C}/^{12}\text{C}$) mass spectrometry to evaluate**
2 **carbon sources and sinks: changes and trends during the**
3 **decomposition of vegetal debris from eucalyptus clone**
4 **plantations (NW Spain)**

5
6 **New title “Potential biodegradability of eucalyptus litter**
7 **from northwestern Spanish forests planted with a different**
8 **clone: F0 or F1 generation”**

9
10 **I. Fernandez¹, and A. Cabaneiro¹**

11 [1]{Instituto de Investigaciones Agrobiológicas de Galicia, Consejo Superior de
12 Investigaciones Científicas IIAG-CSIC, Santiago de Compostela, Spain}

13 Correspondence to: I. Fernandez (ifernandez@iiag.csic.es)

14
15 **Abstract**

16 Vegetal debris is known to participate in key soil processes such as the formation of soil
17 organic matter (OM), also being a potential source of greenhouse gases to the atmosphere.
18 However, its contribution to the isotopic composition of both the soil OM and the
19 atmospheric carbon dioxide is not clear yet. Hence, the main objective of the present research
20 is to understand the isotopic ^{13}C changes and trends that take place during the successive
21 biodegradative stages of decomposing soil organic inputs. By incubating bulk plant tissues for
22 several months under laboratory controlled conditions, the kinetics of the CO_2 releases and
23 shifts in the ^{13}C natural abundance of the solid residues were investigated using litter samples
24 coming from forest plantations with a different clone (Anselmo: F0, 1st clonal generation
25 attained by morphological selection and Odiel: F1, 2nd clonal generation genetically obtained)
26 of *Eucalyptus globulus* Labill. developed over granitic or schistic bedrocks and located in
27 northwestern Spain. Significant isotopic variations with time were observed, probably due to
28 the isotopically heterogeneous composition of these complex substrates in conjunction with
29 the initial selective consumption of more easily degradable ^{13}C -differentiated compounds
30 during the first stages of the biodegradation, while less available or recalcitrant litter
31 components were decomposed at later stages of biodegradation, generating products that have
32 their own specific isotopic signatures. These results, which significantly differ depending on

1 the type of clone, suggest that caution must be exercised when interpreting carbon isotope
2 studies (at natural abundance levels) since perturbations associated with the quality or
3 chemical composition of the organic debris from different terrestrial ecosystems can have an
4 important effect on the carbon stable isotope dynamics.

5

6 **1 Introduction**

7 Since *Eucalyptus globulus* Labill. is an allochthonous species in Galicia (NW Spain), with
8 characteristics that certainly differs from those of the autochthonous flora, its massive
9 utilization generated a big controversy about its environmental repercussion, especially on the
10 soil quality, that are not still totally elucidated. As a consequence of this concern, some
11 investigations related to this type of plantations within the humid-temperate zone have been
12 carried out (Jones et al., 1999; Álvarez González et al., 2005; Vega-Nieva et al., 2013), some
13 of them focussing on the effect of this type of vegetation on the soil (Calvo de Anta, 1992;
14 Brañas et al., 2000; Álvarez et al., 2002; Camps Arbestain et al., 2004). These studies are of
15 exceptional interest in Galicia, where forest soils are usually acid and sandy although very
16 rich in organic matter (OM), which content and quality practically determines the soil
17 productivity. Given that the success of a sustainable silviculture is mainly based in effective
18 recycling of nutrients, the role of the soil OM that controls cation and water reservoirs is very
19 important for forest plantations with a longer rotation cycle as compared to agricultural soils
20 with annual crops (González-Prieto et al., 1996; Mutabaruka et al., 2002; Matus et al., 2008).
21 Soil OM characteristics in forest ecosystems mainly depend on organic debris (including
22 aboveground and belowground vegetal residues) largely coming from the dominant tree
23 species, and for this reason the study of their biochemical characteristics and mineralization
24 kinetics is essential when we want to know the present and future status of nutrient uptake and
25 cycling in a forest ecosystem.

26 The notable extension of forest surface dedicated to *E. globulus* silviculture, joined to
27 the most recent tendency to use clonal eucalyptus plants oriented to increase wood
28 productivity supports the interest of investigations that point to evaluating the environmental
29 influence of these forest plantations as well as the role of *E. globulus* debris as a C sink/source
30 during its progressive incorporation into the soil.

31 Heavy stable isotopes have been frequently used to trace C-flow through the plant-soil
32 system (Van Dam et al., 1997; Fernandez et al., 2006a, 2006b) since isotopic techniques

1 applied to diverse research fields can provide an integrated and quantitative view of chemical,
2 biological and ecological transformations in nature (Boutton et al., 1998; Griffiths et al.,
3 1999). Due to the precision and efficiency of these techniques many studies use the ^{13}C stable
4 isotope to monitor the C cycle in different biochemical processes, such as photosynthetic
5 fixation of atmospheric CO_2 , decomposition of complex plant debris, etc (Schleser et al.,
6 1999; Fernández and Cadisch, 2003; Fernández et al., 2003, 2004, 2006a, 2006b).

7 Therefore, the purpose of this research is to use stable isotope ratio ($^{13}\text{C}/^{12}\text{C}$) mass
8 spectrometry to obtain direct and updated information about litter decay in *E. globulus* clone
9 plantations from the NW of Spain in order to attain a double objective: i) to elucidate the
10 possible differences between the biodegradability of litter from two different eucalyptus
11 clones and, ii) to evaluate the half-lives or residence times of litter C in this type of
12 ecosystems, whose potential remains unknown for this climatic zone.

13

14 **2 Material and methods**

15

16 **2.1 Study area and experimental design**

17 A laboratory controlled experience about vegetal debris decomposition was carried out using
18 litter samples collected from a total of 9 eucalyptus clone plantations (*E. globulus*) in the SW
19 of Europe (Galicia, NW of Spain), within the temperate-humid climate zone. Because the
20 establishment of eucalyptus clone plantations is still a recent practice in this region, at the
21 present time this type of eucalyptus forests are usually young, so that in our experimental
22 design the age factor was not considered and the following two pre-established selection
23 criteria were used:

24 i) clone type (rooted cuttings):

25 · F0, 1st generation clonal plants (morphologically selected): “Anselmo C-14”, or

26 · F1, 2nd generation clonal plants (genetically obtained): “Odiel”

27 ii) the type of underlying rock on which the soil was developed:

28 · granitic material, or

29 · schistic material

30 Therefore for the present study, 6 clonal plantations were selected over granitic
31 bedrock, half of them (3 stands) planted with the Anselmo clone and the other half (3 stands)
32 using Odiel clonal plants, in order to compare litter biodegradability for both types of clones
33 under similar growing conditions. To highlight the possible effects associated with a

1 particular parent material, other 3 plantations with Anselmo clonal plants developed over
2 schistic bedrock were also selected and compared with the corresponding plantations
3 developed over granitic parent material. Therefore, in this experimental design, a total of 9
4 plots have been chosen, in order to obtain litter samples from 3 forest patches for each soil
5 parent material and clone type. A stand trial of approximately 900 m² (30 m x 30 m) was
6 established in each selected forest plantation.

7

8 **2.2 Field sampling**

9 A representative sample of the litter layer, composed by merging 24 subsamples regularly
10 taken from the whole stand area following a squared pattern (4 rows and 6 columns), was
11 collected from each plot. The litter layer, formed by organic input coming from the vegetal
12 cover was sampled during the winter season, being composed by a mixture of debris at
13 different decomposing stages that were accumulated over time on the soil surface.
14 Simultaneously and following the same pattern, a combined soil sample was also obtained
15 from the upper layer of the A horizon (0-15 cm depth).

16

17 **2.3 Vegetal debris and soil analysis**

18 After the combined soil samples were mixed, the soil pH and the total soil C and N contents
19 were assessed by the methods described by Fernandez et al. (2012). The same methodology
20 was used to determine total C and N contents of litter samples. All results obtained were
21 expressed as means from at least three replicate determinations on oven dry basis (105°C).

22

23 **2.3.1 Litter mineralization under controlled conditions**

24 Long-term aerobic incubations of eucalyptus debris, finely crushed (particle size \approx 400 μ m,
25 Kinematica laboratory grinding mill using sieve with hole \varnothing 2 mm mesh size), were carried
26 out in laboratory incubators (with natural convection) under conditions for optimal microbial
27 activity (from each plot, ten replicates of 2 g were placed into 4 litre hermetic glass containers
28 that were maintained at 28°C and 80% moisture content for 560 days). The flask atmospheres
29 were periodically renewed (every day, every 2 days, every week, etc depending on the flask
30 CO₂ concentration) and the C mineralization during the biodegradative processes was
31 monitored by periodically taking a gas sample from each container and by measuring its CO₂
32 concentration using a multiple infrared gas analyser (7000FM GFC IR ANALYSER, Signal Group

1 Limited). Potential C mineralization was expressed as grams of CO₂-C evolved per kilogram
2 of dry material (g C_{mineralized} kg⁻¹_{d.m.}) and as a percentage of the total litter C (C mineralization
3 coefficient: C_{mineralized}/C_{total}*100).

4

5 **2.3.2 Kinetic modelling**

6 To quantify C mineralization kinetic parameters, the cumulative data on the CO₂ released at
7 different degradation stages along the incubation period were fitted, as described by
8 Cabaneiro et al. 2008, to the double exponential equation proposed by Andr n and Paustian
9 (1987):

$$10 \quad C_t = C_o (1 - e^{-kt}) + (C - C_o)(1 - e^{-ht}) \quad (1)$$

11 where C_t is the cumulative C release after time t (g C kg⁻¹_{d.m.}), C_o is the labile C pool
12 (g C kg⁻¹_{d.m.}), *k* is the instantaneous mineralization rate of the labile C pool (d⁻¹), and *h* is the
13 instantaneous mineralization rate of the recalcitrant C pool (d⁻¹). To avoid parameter
14 estimation errors, Updegraff et al. (1995) convergence criteria were applied.

15 For all debris samples collected from the different eucalyptus clone plantations, the
16 time required for 50% C loss (half-life) and the turnover rate of both labile (L) and
17 recalcitrant (R) organic fractions of the litter was calculated as *Half-life L* = 0.693/*k* or *Half-*
18 *life R* = 0.693/*h* and as *Turnover L* = 1/*k* or *Turnover R* = 1/*h*, respectively.

19

20 **2.3.3 Isotopic analysis (¹³C)**

21 To show possible isotopic changes during the biodegradation process, at the beginning (T₀=0
22 days), during ongoing (T₁=36, T₂=98, T₃=359 days), and at the end (T₄=560 days) of the
23 incubation period, the litter ¹³C/¹²C ratio was determined. Litter samples were oven-dried
24 (40°C), finely ground (<100 μm) and weighed in tin capsules for isotopic analysis. The ¹³C
25 natural abundance was assessed by continuous flow isotope-ratio mass spectrometry using a
26 ThermoFinnigan Delta^{plus} mass spectrometer coupled to a FlashEA 1112 elemental analyzer
27 through a ConFlo II interface. As part of each analytical batch run, a set of international
28 reference materials (NBS 22, IAEA-CH-6, USGS 24) were analysed and, all through the
29 successive isotopic determinations, the precision (standard deviation) for the analysis of δ¹³C
30 of the laboratory standard (acetanilide, n=10) was lower than ± 0.102‰.

1 The $^{13}\text{C}/^{12}\text{C}$ ratio or $\delta^{13}\text{C}$ signature was obtained according to the following equation,
2 which is based on the deviation of the $^{13}\text{C}/^{12}\text{C}$ ratio from the reference standard Vienna Pee
3 Dee Belemnite standard:

$$4 \delta^{13}\text{C} (\text{‰}) = (R_{\text{sample}}/R_{\text{standard}} - 1) \times 10^3 \quad (2)$$

5 where $R = ^{13}\text{C}/^{12}\text{C}$.

7 **2.3.4 Statistical analyses**

8 Data were statistically analysed by one-way ANOVA and least significant difference (LSD)
9 test at the 95% probability level ($P \leq 0.05$) was applied to the results. Pearson correlation
10 coefficients were calculated to examine relationships between the different variables. All
11 statistical analyses were performed using the computer software package IBM[®] SPSS[®]
12 Statistics, version 21.0.0.0 (2012).

14 **3 Results and discussion**

15 All eucalyptus clone plantations selected to carry out this study are developed over acidic
16 soils ($\text{pH}_{\text{H}_2\text{O}}$ ranged from 4.1 to 5.1) with a strong potential acidity (pH_{KCl} ranged from 3.2 to
17 4.1). The soil OM of these forest plantations exhibits values of the C-to-N ratio between 11
18 and 22, with soil C and N contents highly scattered (Table 1). As compared with soils, the
19 litter from the two studied clones of *E. globulus* shows markedly higher values of the total C
20 concentration (with mean values at least five times the total soil C concentration). However,
21 total N concentration in litter samples is closer to total soil N values, resulting in higher
22 eucalyptus debris C-to-N ratios, which present notable differences among plantations,
23 especially in granitic plots (with C-to-N ratios ranging from 45 to 82).

25 **3.1 Biodegradability of eucalyptus debris**

26 Respirometric techniques and incubation procedures for measuring the biological C
27 mineralization are common approaches to assess the potential turnover rate of OM for soils
28 and litter samples (Cabaneiro et al., 2008; Fernandez et al., 2010). Monitoring biodegradation
29 of litter from the two types of eucalyptus clonal plantations studied using long-term aerobic
30 incubations (18 months) under laboratory controlled conditions, allowed not only the
31 continuous determination of the CO_2 released during decomposition but also the tracking of

1 the isotopic ^{13}C dynamics, the quantification of weight losses rates and the progressive
2 changes in the chemical quality of the remaining substrate at different biodegradation stages
3 or incubation times ($T_0=0$, $T_1=36$, $T_2=98$, $T_3=359$, $T_4=560$ days of incubation).

4 When we compare the biodegradability of litter from the two eucalyptus clonal
5 varieties developed over the same bedrock type which in this section trial consisted of granitic
6 parent material, visible weight losses were observed for both clones, with mean decreases of
7 over 20% of the dry material at the end of the incubation (Fig. 1A), these decreases being
8 more pronounced in debris coming from plantations with Odiel clonal plants. As usually
9 reported for plant debris decomposition (Berg et al., 2000), the declines found in our
10 experiment were more pronounced during the first months of incubation, with significant
11 initial differences depending on the type of clone studied (ANOVA, $P<0.014$ and $P<0.021$ at
12 36 and 98 days respectively) that are not longer statistically significant from the first year of
13 incubation. These weight losses, observed during the decomposition of samples collected
14 from the litter layer (which was composed by a mixture of fallen eucalyptus debris with
15 uneven time periods on the atop soil), are lower than those reported bibliographically by other
16 authors during fresh plant material degradation (Lusk et al., 2001, Garcia-Velazquez et al.,
17 2010, Jacob et al., 2010). Besides the previously mentioned weight losses registered during
18 litter degradation, a change in the chemical composition of the substrate was also observed, as
19 reflected by their C-to-N ratio evolution (Fig. 1B). Thus, Odiel clone debris (that had the
20 highest weight loss rate) presented the lowest C-to-N ratios, with values of this parameter that
21 start to differ from the values exhibited by Anselmo litter samples from the first year of
22 incubation (ANOVA, $P<0.034$ and $P<0.043$ for 359 and 560 days of incubation, respectively).
23 The fact that litter samples exhibiting the lowest C-to-N ratios also showed the highest weight
24 losses during the biodegradation seems to agree with the results found by some authors, who
25 associate litter decay to labile C and N availabilities during the initial decomposition phases
26 (Berg et al., 2007; Berg and McClaugherty, 2008), when labile compounds such as
27 carbohydrates, proteins, and other simple compounds are rapidly degraded by fast growing
28 microorganisms requiring high N concentrations (Fioretto et al., 2005).

29 To elucidate the effects that the type of parent material under the eucalyptus
30 plantations can produce on the processes involved on litter biodegradation, the results
31 obtained from the 6 Anselmo clone plantations, 3 of them developed over granitic and 3 over
32 schistic bedrock, were compared. These results seem to indicate that litter decay is slightly
33 more active in eucalyptus stands developed over schistic bedrocks than in plantations

1 developed over granitic parent materials, this being reflected by a more pronounced weight
2 loss (Fig. 1C) and a lower C-to-N ratio (Fig. 1D) in schistic plantations. However, these
3 differences associated to the underlying parent material were not statistically significant for
4 none of these two parameters.

5

6 **3.1.1 Carbon mineralization dynamics**

7 As shown in Table 2, values of the C mineralization coefficient (expressed as % of the total
8 litter C content), obtained at the end of the incubation period for litter from both eucalyptus
9 clones ranged from 20% to barely surpassing 30% in the most active cases. These results are
10 within the range of mineralization coefficients reported by other authors for debris from
11 different tree species (Fernandez et al., 2003). In our study the mean value of the C
12 mineralization activity (expressed as the total quantity of CO₂ evolved per unit weight)
13 positively correlates with both the initial C content of the substrate ($P \leq 0.033$) and the C
14 mineralization coefficient ($P \leq 0.000$). The differences on C mineralization activity between
15 litter samples coming from both eucalyptus clonal plants are clearly illustrated when
16 comparing the C mineralization coefficients presented by debris collected from the
17 plantations developed on granitic rocks exclusively, the values presented by Odiel being
18 significantly higher than those showed by Anselmo plantations (ANOVA, $P \leq 0.040$). A
19 similar behaviour was found when comparing the total quantity of C released as CO₂ during
20 the whole incubation period, that practically in all eucalyptus plantations surpassed 100 g C
21 $\text{kg}^{-1}_{\text{d.m.}}$, debris from Odiel plantations having again significantly higher activity values
22 ($150.3 \pm 14.7 \text{ g C kg}^{-1}_{\text{d.m.}}$) as compared with the other clone ($113.4 \pm 18.3 \text{ g C kg}^{-1}_{\text{d.m.}}$) over the
23 same bedrock type (ANOVA, $P \leq 0.042$).

24

25 **3.1.2 Kinetic modelling and half-life study**

26 Despite the previously mentioned differences on the CO₂ released during the biodegradation
27 of eucalyptus debris collected from Odiel and Anselmo clone plantations, in both cases
28 experimental cumulative data of the CO₂ evolved during the incubation (Fig. 2) significantly
29 fitted to the Eq. (1), first-order double exponential kinetic model proposed by Andrén and
30 Paustian (1987), which supports the hypothesis of two organic pools of different microbial
31 stabilities and mineralization rates, allowing the estimation of the labile and recalcitrant C
32 pools in each substrate (Table 2).

1 By comparing the CO₂ evolved along the incubation of litter samples from the two
2 eucalyptus clonal varieties over the same parent material “granitic bedrock” (Fig. 2A), from
3 the first month of monitoring substrate biodegradation significantly higher CO₂ releases were
4 observed for debris coming from Odiel as compared with Anselmo plantations (P≤0.029), the
5 significance of these differences lasting until the end of the experimental period (T1: P≤0.017,
6 T2: P≤0.021, T3: P≤0.29, T4: P≤0.042). However when comparing the values obtained for all
7 Anselmo clone plantations (Fig. 2B), to elucidate the effects of the underlying parent material
8 (granite/schist), the results indicate that the differences are never statistically significant,
9 although CO₂ released by litter from schistic plots are slightly higher than from granitic ones
10 along the whole incubation period. Determination coefficients (R²) obtained by using the
11 previously mentioned kinetic model were always higher than 0.99 (Table 2) and the estimated
12 values of labile C pool for the different eucalyptus debris ranged from 38.3 g C kg⁻¹_{d.m.} (for a
13 plot with Anselmo clonal plants over granitic bedrock) to 123.3 g C kg⁻¹_{d.m.} (for a plot with
14 Odiel clonal plants over granitic bedrock), representing from 7.6 to 23.9 % of their total C
15 content, respectively. Thus, as compared with debris from Anselmo clone plantations,
16 significantly bigger sizes of the labile C pools were estimated for Odiel litter samples
17 (ANOVA, P≤0.040). Besides, the size of the labile C pool of litter samples collected from the
18 9 studied eucalyptus plantations was found to be positively correlated with both C
19 mineralization indices (P≤0.028 and P≤0.023 with the total C mineralized and the C
20 mineralization coefficient, respectively).

21 As it can be seen in Table 2, the instantaneous mineralization rate of the labile fraction
22 (*k*) is between 50 and 100 times higher than that of the most recalcitrant pool (*h*), without any
23 statistically significant difference between Anselmo and Odiel debris or between granitic and
24 schistic plots. Both *k* and *h* instantaneous mineralization rates show a very strong positive
25 intercorrelation (P≤0.000) and their values point to very different turnover rates for the labile
26 or the recalcitrant fractions (Table 3). Thus, half-life values obtained for the labile fraction of
27 eucalyptus litter ranges from 26 to 77 days, whereas half-life times for the recalcitrant pool
28 vary from approximately 5 to 16 years, which corresponds to turnover rates of up to 111 days
29 for the labile compounds and up to more than 23 years for the more recalcitrant ones, these
30 latter rates being sometimes nearly 100 times higher than the turnover rates of the labile
31 fraction (see Table 3, O3: Odiel clone over granitic bedrock).

32

1 **3.2 Isotopic ¹³C evolution during eucalyptus litter decay**

2 The ¹³C natural abundance values of eucalyptus debris, obtained according Eq. (2), exhibited
3 by both type of clonal plantations (Table 1) are within the range reported for C3 vegetation
4 (Fernandez et al., 2003, 2005, Cabaneiro et al., 2009). The ¹³C isotopic composition was
5 found to be slightly different for the two eucalyptus clones, debris from Odiel clonal
6 plantations being significantly more ¹³C enriched than debris from Anselmo clones (ANOVA,
7 $P \leq 0.046$) with variations that ranged from -30.3‰ (Anselmo plot) to -29.5‰ (Odiel plot).

8 Isotopic differences between Anselmo and Odiel clones on the temporal evolution
9 during debris biodegradation were also revealed when the behaviour of the ¹³C/¹²C ratios of
10 the litter samples collected from both types of eucalyptus plantations over granitic bedrocks
11 (Fig. 3A) were compared along the incubation period, which suggests different ¹³C
12 fractionation dynamics that maybe associated to a dissimilar distribution of some biochemical
13 compounds in both substrates. Thus, during the biodegradation of Odiel debris, visible ¹³C
14 depletion was observed during early decomposition stages, suggesting a selective
15 mineralization of relatively enriched ¹³C compounds, such as proteins, lipids, sugars, starch
16 and other carbohydrates or biomolecules included in the non-ADF fraction described by
17 Fernandez et al. (2003) for decomposing plant materials. On the other hand, organic debris
18 collected from Anselmo clone plantations showed slight and brief initial increases of their ¹³C
19 contents, probably due in this case to the preferential mineralization of some easily
20 degradable biomolecules relatively depleted in this carbon isotope, (e.g. amino acids can be
21 significantly ¹³C depleted according to Borland et al., 1994). As a consequence of these
22 contrasting tendencies, a quick reduction of the initial isotopic differences between litter
23 samples from Odiel and Anselmo plantations was observed, these differences becoming
24 statistically insignificant since the first month of incubation. Once this early stage of high
25 mineralization activity has elapsed, another phase of litter biodegradation with minor ¹³C
26 variations begins and the isotopic composition of the substrate remains practically unaltered
27 until the end of the decomposition experiment. Once more, it can be highlighted that the
28 underlying bedrock did not seem to have a remarkable influence on the isotopic dynamics
29 during litter decay (Fig. 3B), since not significant differences were found between debris
30 collected from Anselmo clone plantations growing either on granitic or on schistic stands,
31 although the litter from the latter is slightly depleted in this carbon isotope as compared to the
32 litter coming from Anselmo plantations over granitic bedrocks.

33

1 **3.3 Factors influencing eucalyptus litter biodegradation**

2 Taken as a whole, after a general scrutiny of all these results, the integrated outcomes of the
3 research including all the different parameters studied during eucalyptus litter decomposition
4 support the hypothesis of the existence of some biochemical heterogeneity between the litter
5 collected from the duff layer of the two studied types of *Eucalyptus globulus* clonal
6 plantations (with F0 or F1 clonal plants) and allows to deduce that, for these kind of forests
7 from northwestern Spain, the biodegradability of the aboveground litter seems to be strongly
8 influenced by the following two key factors:

9 i) The litter decaying stage. The results of eucalyptus litter decomposition illustrates that two
10 differentiated decaying phases during the biodegradative process can be clearly described: a
11 relatively brief more active initial phase (first weeks/months), when the greatest weight
12 losses, CO₂ releases, or isotopic ¹³C shifts occurs, and a later or delayed second less active
13 phase, when all these variables show a posterior progressive stabilization and remain
14 practically constant until the end of the incubation. Two or more decomposing phases during
15 litter decay have already been reported by many authors for different tree species (Melillo et
16 al., 1989; Aber et al., 1990; Guillon et al., 1993; Coûteaux et al., 1995; Rovira and Rovira,
17 2010; Castellanos-Barliza and León, 2011; Patricio et al., 2012).

18 ii) The clonal origin or intrinsic characteristics of the litter. Some dissimilarities in the
19 promptness or slowness of the degradative process can be also distinguished between both
20 types of aboveground residues collected either from the F0 (1st generation) or from the F1 (2nd
21 generation) eucalyptus clonal plantations.

22 More to the point, the influence of these above mentioned two factors appear to be
23 firstly related with the initial chemical composition or quality of the litter (N content, C-to-N
24 ratio, and ¹³C signature) mainly determined by its genetic origin that seems to have a certain
25 influence on its biodegradability and on its C mineralization kinetics. However, although to a
26 lesser extent, it seems to be also moderately affected by the underlying bedrock type.

27

28 **4 Conclusions**

29 The genetic diversity between the two studied clonal plants (Anselmo: F0, first generation
30 clonal variety attained by morphological selection and Odiel: F1, second generation clonal
31 variety genetically obtained) are reflected not only into a different production of vegetal
32 biomass but also into a higher lability of forestal debris coming from Odiel plantations, with

1 lower C-to-N ratios and higher C mineralization coefficients, as compared to litter tissues
2 collected from Anselmo plots. Even if more detailed research that would involve the study of
3 the biochemical and isotopic composition of live leaves as well as of other belowground
4 organic inputs directly entering into the underneath soil layers would be convenient, our
5 findings on the isotopic behaviour during the decomposition of debris coming from the duff
6 layer of Galician forest plantations (NW Spain) with two different *E. globulus* clones seem to
7 reveal the existence of possible differences between both eucalyptus clonal plants at
8 photosynthetic levels, affecting their internal chemistry and therefore the C dynamics of
9 decaying litter. The different isotopic ^{13}C behaviour at early stages of litter decay found for
10 both eucalyptus clonal varieties exhibits evidences of a different proportion of some labile
11 compounds associated to the genotypic characteristics of each type of clone plantation. Thus,
12 microbial fractionation of ^{13}C during detritus decomposition can not be neglected when
13 attempting to evaluate the isotopic aspects of the C cycle comprehension and the
14 quantification of ^{13}C discrimination have to be taken into consideration in order to obtain
15 more reliable estimates of the contribution of decaying vegetal debris to soil OM buildup in
16 each specific ecological context. This has direct implications in studies of soil OM dynamics
17 using isotopic (^{13}C) techniques, particularly to avoid errors in appraising the contribution of
18 eucalyptus litter decay to the global C balance.

19

20 **Acknowledgements**

21 This research was conducted as a part of the project AGL2010-22308-C02-02 financed by the
22 Spanish Government (Ministerio de Ciencia e Innovación). We thank the departments of
23 “Ingeniería Agroforestal” and “Producción Vegetal” of the University of Santiago de
24 Compostela for their invaluable assistance in plot selection. We also thank the Scientific
25 Research Support Services of the University of A Coruña for the isotopic ^{13}C analyses. ENCE
26 is gratefully acknowledged for allowing us to use their clone plantations. Finally, we thank
27 Ana Argibay, Daniel Caride and María García for their technical assistance in the laboratory
28 and fieldwork, as well as SILVANUS, GIT. Forestry Consulting SL, NORFOR and Dirección
29 Xeral de Montes (Consellería do Medio Rural, Xunta de Galicia) for showing interest in this
30 research.

31

1 **References**

- 2 Aber, J., Melillo, J., and McClaugherty, C.: Predicting long-term patterns of mass loss,
3 nitrogen dynamics, and soil organic matter formation from initial fine litter chemistry in
4 temperate forest ecosystems. *Can. J. Bot.*, 68, 2201-2208, 1990.
- 5 Alvarez, E., Monterroso, C., and Fernández Marcos, M. L.: Aluminium fractionation in
6 Galician (NW Spain) forest soils as related to vegetation and parent material. *Forest
7 Ecol. Manag.*, 166, 193-206, 2002.
- 8 Alvarez Gonzalez, J. G., Balboa Murias, M. A., Merino, A., and Rodríguez Soalleiro, R.:
9 Estimación de la biomasa arbórea de *Eucalyptus globulus* y *Pinus pinaster* en Galicia.
10 *Recursos Rurais*, 1, 21-30, 2005.
- 11 Andrén, O. and Paustian, K.: Barley straw decomposition in the field a comparison of models.
12 *Ecology*, 68, 1190-1200, 1987.
- 13 Berg, B. and McClaugherty, C.: *Plant litter. Decomposition, Humus Formation, Carbon
14 Sequestration*, 2nd ed. Ed. Springer Verlag, Heidelberg, Berlin, 2008.
- 15 Berg, B., Meentemeyer, V., and Johansson, M. B.: Litter decomposition in a climatic transect
16 of Norway spruce forests-climate and lignin control of mass-loss rates. *Can. J. Forest Res.*
17 30, 1136-1147, 2000.
- 18 Berg, B., Steffen, K. T., and McClaugherty, C. Litter decomposition rate is dependent on litter
19 Mn concentrations. *Biogeochemistry*, 82, 29-39, 2007.
- 20 Borland, A. M., Griffiths, H., Broadmeadow, M. S. J., Fordham, M. C., and Maxwell C.:
21 Carbon-isotope composition of biochemical fractions and the regulation of carbon balance in
22 leaves of the C₃-crassulacean acid metabolism intermediate *Clusia minor* L. Growing in
23 Trinidad., *Plant Physiol.*, 106, 493-501, 1994.
- 24 Boutton, T. W., Archer, S. R., Midwood, A. J., Zitzer, S. F., and Bol, R.: $\delta^{13}\text{C}$ values of soil
25 organic carbon and their use in documenting vegetation change in a subtropical savanna
26 ecosystem. *Geoderma*, 82, 5-41, 1998.
- 27 Brañas, J., González-Río, F., and Merino, A.: Contenido y distribución de nutrientes en
28 plantaciones de *Eucalyptus globulus* del noroeste de la península ibérica. *Forest systems*, 9(2),
29 317-335, 2000.

- 1 Cabaneiro, A., Fernandez, I., Pérez-Ventura, L., and Carballas, T.: Soil CO₂ Emissions from
2 Northern Andean Páramo Ecosystems: Effects of Fallow Agriculture. *Environ. Sci. Technol.*,
3 42, 1408-1415, 2008.
- 4 Cabaneiro A. and Fernandez I. Testemuño Isotópico (¹³C) do Cambio Global en Galicia, in:
5 Evidencias e Impactos do Cambio Climático en Galicia, edited by Pérez Muñuzuri, V.,
6 Fernandez Cañamero, M., and Gomez Gesteira, J. L., Consellería de Medio Ambiente e
7 Desenvolvemento Sostible, Xunta de Galicia, Santiago de Compostela, 229-245, 2009.
- 8 Calvo de Anta, R.: El Eucalipto en Galicia. Sus relaciones con el medio natural. Ed.
9 Universidad de Santiago de Compostela, 1992.
- 10 Camps Arbestain, M., Mourenza, C., Alvarez, E., and Macías, F.: Influence of parent material
11 and soil type on the root chemistry of forest species grown on acid soils. *Forest Ecol. Manag.*,
12 193, 307-320, 2004.
- 13 Castellanos-Barliza, J. and León, J. D.: Descomposición de hojarasca y liberación de
14 nutrientes en plantaciones de *Accacia mangium* (Mimosaceae) establecidas en suelos
15 degradados de Colombia. *Rev. Biol. Trop. (Int. J. Trop. Biol. ISSN-0034-7744)*, 59, 113-128,
16 2011.
- 17 Coûteaux, M. M., Bottner, P., and Berg, B.: Litter decomposition, climate and litter quality.
18 *Tree*, 10, 63-66, 1995.
- 19 Dewar, R.C. and Cannell, M. G. R.: Carbon sequestration in the trees, products and soils of
20 forest plantations: an analysis using UK examples. *Tree Physiol.* II, 49-71, 1992.
- 21 Ehleringer, J. R., Buchmann, N., and Flanagan, L. B.: Carbon isotope ratios in belowground
22 carbon cycle processes. *Ecol. Appl.*, 10, 412-422, 2000.
- 23 Fernández, I., Cabaneiro, A., and Carballas, T.: Thermal resistance to high temperatures of
24 the different soil organic fractions from soils under pine forests. *Geoderma*, 104, 281-298,
25 2001.
- 26 Fernandez, I. and Cadisch, G.: Discrimination against ¹³C during degradation of simple and
27 complex substrates by two white rot fungi. *Rapid Commun. Mass Spectrom.*, 17, 2614-2620,
28 2003.

- 1 Fernandez, I., Mahieu, N., and Cadisch, G.: Carbon isotopic fractionation during
2 decomposition of plant materials of different quality. *Global Biogeochem. Cy.*, 17, 1075-
3 1085, 2003.
- 4 Fernandez, I., Cabaneiro, A., and González-Prieto, S. J.: Use of ^{13}C to monitor soil organic
5 matter transformations caused by a simulated forest fire. *Rapid Commun. Mass Spectrom.*,
6 18, 435-442, 2004.
- 7 Fernandez, I., González-Prieto, S.J., and Cabaneiro, A.: ^{13}C -isotopic fingerprint of *Pinus*
8 *pinaster* Ait. and *Pinus sylvestris* L. wood related to the quality of standing tree mass in
9 forests from NW Spain. *Rapid Commun. Mass Spectrom.*, 19, 3199-3206, 2005.
- 10 Fernández, I., Cabaneiro, A., and González-Prieto, S. J.: Partitioning CO_2 Effluxes from an
11 Atlantic Pine Forest Soil between Endogenous Soil Organic Matter and Recently Incorporated
12 ^{13}C -Enriched Plant Material. *Environ. Sci. Technol.*, 40, 2552-2558, 2006a.
- 13 Fernandez, I., Pérez-Ventura, L., González-Prieto, S. J., and Cabaneiro, A.: Soil $\delta^{13}\text{C}$ and
14 $\delta^{15}\text{N}$ as a good indicator for predicting the site index of Galician pine forests (*P. pinaster* Ait.
15 and *P. sylvestris* L.). *Forest systems*, 15(1), 3-13, 2006b.
- 16 Fernández, I., Carrasco, B., and Cabaneiro, A.: Comparing the potential carbon mineralization
17 activity of the soil organic matter under tow braoadleaf autochthonous tree species from the
18 NW of Spain (*Quercus robur* L., *Betula alba* L.). *Forestry Ideas*, 16, 258-265, 2010.
- 19 Fernández, I., Carrasco, B., and Cabaneiro, A.: Evolution of soil organic matter composition
20 and edaphic carbon effluxes following oak forest clearing for pasture: climate change
21 implications. *Eur. J. Forest Res.*, 131, 1681-1693, 2012.
- 22 Fioretto, A., Di Nardo, C., Papa, S., and Fuggi, A. Lignin and cellulose degradation and
23 nitrogen dynamics during decomposition of three leaf litter species in a Mediterranean
24 ecosystem. *Soil Biol. Biochem.*, 37, 1083-1091, 2005.
- 25 García-Velásquez, L. M., Ríos-Quintana, A., and Molina-Rico, L.: Structure, plant
26 composition and leaf litter decomposition in soil, at two sites of an andean cloud forest
27 (reforested and in spontaneous succession) in Peñas Blancas, Calarcá (Quindío), Colombia.
28 *Actual Biol.*, 32(93), 147-164, 2010
- 29 Gillon, D., Joffre, R., and Ibrahima, A.: Inital litter properties and decay rate: a microcosm
30 experiment on Mediterranean species. *Can. J. Bot.*, 72, 946-954, 1993..

- 1 González-Prieto, S. J., Cabaneiro, A., Villar, M. C., Carballas, M., and Carballas T.: Effect of
2 soil characteristics on N nimeralization capacity in 112 native and agricultural soils from the
3 northwest of Spain. *Biol. Fertil. Soils*, 22, 252-260, 1996.
- 4 Griffiths, H., Borland, A., Gillon, J., Harwood, K., Maxwell, K., and Wilson, J.: Stable
5 isotopes reveal exchanges between soil, plants and the atmosphere, in: *Physiological Plant
6 Ecology*, edited by: Press, M. C., Scholes, J. D., and Barker, M. G., Blackwell Science,
7 Oxford, 415- 441, 1999.
- 8 Jacob, M., Viedenz, K., Polle, A., and Thomas F. M.: Leaf litter decomposition in temperate
9 deciduous forest stands with a decreasing fraction of beech (*Fagus sylvatica*). *Oecologia*, 164,
10 1083-1094, 2010.
- 11 Jones, H.E., Madeira, M., Herraiez, L., Dighton, J., Fabião, A., González-Rio, F., Fernandez
12 Marcos, M., Gomez, C., Tomé, M., Feith, H., Magalhães, M.C., and Howson, G.: The effect
13 of organic-matter management on the productivity of *Eucalyptus globulus* stands in Spain and
14 Portugal: tree growth and harvest residue decomposition in relation to site and treatment.
15 *Forest Ecol. Manag.*, 122, 73-96, 1999.
- 16 Lusk, C. H., Donoso, C., Jiménez, M., Moya, C., Oyarce, G., Reinoso, R., Saldaña, A.,
17 Villegas, P., and Matus, F.: Descomposición de hojarasca de *Pinus radiata* y tres especies
18 arbóreas nativas. *Revista Chilena de Historia Natural*, 74, 705-710, 2001.
- 19 Matus, F. J., Lusk, C. H., and Maire, C. R.: Effects of soil texture, carbon input rates, and
20 litter quality on free organic matter and nitrogen mineralization in Chilean rain forest and
21 agricultural soils. *Commun. Soil Sci. Plant Anal* , 39, 187-201, 2008.
- 22 Melillo, J. M., Aber, J. D., Linkins, A. E., Ricca, A., Fry, B., and Nadelhoffer, K. J.: Carbon
23 and nitrogen dynamics along the decay continuum: plant litter to soil organic matter. *Plant
24 Soil*, 115, 180-198, 1989.
- 25 Mutabaruka, R., Mutabaruka, C., and Fernandez I.: Diversity of arbuscular micorrhizal fungi
26 associated to tree species in semiarid areas of Machakos, Kenya. *Arid Land Res. Manag.*, 16,
27 385-390, 2002.
- 28 Patricio, M. S., Nunes, L. F., and Pereira, E. L.: Litterfall and litter decomposition in chestnut
29 high forest stands in northern Portugal. *Forest Systems*, 21, 259-271, 2012.

- 1 Rovira, P. and Rovira, R.: Fitting litter decomposition datasets to mathematical curves:
2 Towards a generalised exponential approach. *Geoderma*, 155, 329-343, 2010.
- 3 Schleser, G. H., Frielingsdorf, J., and Blair, A.: Carbon isotope behaviour in wood and
4 cellulose during artificial aging. *Chem. Geol.*, 158, 121-130, 1999.
- 5 Updegraff K., Pastor J., Bridgham S.D., and Johnston. C. A. 1995. Environmental and
6 substrate quality controls over carbon and nitrogen mineralization in northern wetlands. *Ecol.*
7 *Appl.*, 5, 151-163, 1995.
- 8 Van Dam, D., Veldkamp, E., and Van Bremen, N.: Soil organic carbon dynamics: variability
9 with depth in forested and deforested soils under pasture in Costa Rica. *Biogeochemistry*, 39,
10 343-375, 1997.
- 11 Vega-Nieva, D. J., Tomé, M., Tomé J., Fontes, L., Soares, P., Ortiz, L., Basurco, F., and
12 Rodríguez-Soalleiro, R.: Developing a general method for the estimation of the fertility rating
13 parameter of the 3-PG model: application in *Eucalyptus globulus* plantations in northwestern
14 Spain. *Can. J. Forest Res.*, 43, 627-636, 2013.

15

16

17

18

19

20

21

22

23

24

25

26

27

1 **Table 1.** Main chemical characteristics and isotopic ^{13}C composition (mean \pm standard
2 deviation) of both the upper 0-15 cm of soils and litter collected from eucalyptus clone
3 (Anselmo and Odiel) plantations.

		Soil				Litter				
Plot		pH	pH	C	N	C-to-N	C	N	C-to-N	$\delta^{13}\text{C}$ ‰
		H ₂ O	KCl	gC kg ⁻¹ _{d.s.}	gN kg ⁻¹ _{d.s.}		gC kg ⁻¹ _{d.m.}	gN kg ⁻¹ _{d.m.}		
A	Granitic A1	4.5 \pm 0.0	3.8 \pm 0.0	83.1 \pm 0.6	5.5 \pm 0.0	15	463.6 \pm 22.5	10.3 \pm 0.3	45	-30.3 \pm 0.0
N	Granitic A2	4.1 \pm 0.0	3.2 \pm 0.0	137.3 \pm 2.0	7.3 \pm 0.0	19	516.1 \pm 24.3	6.3 \pm 0.2	82	-29.9 \pm 0.0
S	Granitic A3	4.7 \pm 0.0	3.8 \pm 0.0	76.3 \pm 2.5	5.8 \pm 0.3	13	500.7 \pm 19.8	6.4 \pm 0.1	79	-29.8 \pm 0.0
E	Mean value	4.4\pm0.3	3.6\pm0.4	98.9\pm33.4	6.2\pm0.9	16\pm3	493.5\pm27.0	7.6\pm2.3	65\pm21	-30.0\pm0.3
L	Schistic A1	5.1 \pm 0.0	4.1 \pm 0.0	20.2 \pm 0.4	0.9 \pm 0.1	22	523.3 \pm 18.6	9.7 \pm 0.4	54	-30.1 \pm 0.0
M	Schistic A2	4.8 \pm 0.0	4.0 \pm 0.0	57.9 \pm 0.9	3.6 \pm 0.1	16	498.0 \pm 10.6	9.0 \pm 0.0	55	-30.1 \pm 0.0
O	Schistic A3	4.6 \pm 0.0	3.7 \pm 0.0	60.1 \pm 2.7	3.7 \pm 0.3	16	490.8 \pm 18.1	8.1 \pm 0.1	61	-30.2 \pm 0.0
	Mean value	4.8\pm0.3	3.9\pm0.2	46.1\pm22.4	2.8\pm1.6	18\pm4	504.0\pm17.1	8.9\pm0.8	56\pm4	-30.1\pm0.1
O	Granitic O1	4.7 \pm 0.0	3.9 \pm 0.0	47.6 \pm 0.4	3.3 \pm 0.0	15	511.2 \pm 10.4	9.6 \pm 0.0	53	-29.6 \pm 0.0
D	Granitic O2	5.0 \pm 0.0	4.1 \pm 0.0	47.6 \pm 0.9	3.5 \pm 0.1	11	515.3 \pm 19.3	10.4 \pm 0.0	50	-29.8 \pm 0.0
I	Granitic O3	4.6 \pm 0.0	3.7 \pm 0.0	109.6 \pm 1.2	6.5 \pm 0.0	17	517.2 \pm 12.8	6.9 \pm 0.4	75	-29.5 \pm 0.0
E	Mean value	4.8\pm0.2	3.9\pm0.2	68.3\pm35.8	4.4\pm1.8	14\pm3	514.6\pm3.1	9.0\pm1.8	59\pm14	-29.6\pm0.2
L										

1 **Table 2.** Total C mineralized, C mineralization coefficients and kinetic parameters obtained
 2 after 560 days of incubation of litter collected from different eucalyptus clone (Anselmo and
 3 Odiel) plantations by using a first-order kinetic model based on the double exponential
 4 equation proposed by Andrén and Paustian (1987): $C_{\text{mineralized}} = C_o (1-e^{-kt}) + C_r(1-e^{-ht})$.

	Potential C mineralization mean±standard deviation		Kinetic parameters Estimated value±standard asymptotic error				R²
	C mineralized (g kg ⁻¹ _{d.m.})	C mineralization coefficient (%)	C_o (g C kg ⁻¹ _{d.m.})	k (d ⁻¹)	C_r (g C kg ⁻¹ _{d.m.})	h×10⁴ (d ⁻¹)	
ANSELMO CLONE							
Granitic A1	106.3 ±8.4	22.9±1.8	71.8±5.1	0.012±0.001	391.8±27.6	1.8±0.3	0.995
Granitic A2	134.2±19.7	26.0±3.8	107.3±5.3	0.010±0.001	408.8±29.6	1.2±0.3	0.998
Granitic A3	99.6 ±3.3	19.9±0.7	38.3±2.9	0.018±0.002	462.4±22.7	2.7±0.2	0.994
Mean value	113.4	22.9	72.5	0.013	421.0	1.9	
SD	18.3	3.1	34.5	0.004	36.9	0.7	
SCHISTIC CLONE							
Schistic A1	160.9±6.3	30.7±1.2	77.1±3.5	0.027±0.002	446.2±22.1	4.1±0.2	0.995
Schistic A2	123.9±6.7	24.9±1.3	91.0±7.0	0.009±0.001	407.0±17.6	1.6±0.4	0.997
Schistic A3	133.2±0.7	27.2±0.1	87.4±2.4	0.014±0.001	403.4±20.5	2.2±0.1	0.999
Mean value	139.3	27.6	85.2	0.017	418.9	2.6	
SD	19.2	3.0	7.2	0.009	23.7	1.3	
ODIEL CLONE							
Granitic O1	145.9±7.8	28.6±1.5	102.8±3.7	0.015±0.001	408.4±14.1	2.2±0.2	0.998
Granitic O2	166.8±2.7	32.4±0.5	123.3±3.3	0.015±0.001	392.0±22.6	2.2±0.2	0.999
Granitic O3	138.3±8.9	26.7±1.7	107.8±1.8	0.014±0.000	409.4±14.6	1.4±0.1	0.999
Mean value	150.3	29.2	111.3	0.015	403.3	1.9	
SD	14.7	2.9	10.7	0.001	9.8	0.5	

- 5 C_o, carbon of the labile pool
 6 k, instantaneous mineralization rate of the labile carbon pool
 7 C_r, carbon of the recalcitrant pool
 8 h, instantaneous mineralization rate of the recalcitrant carbon pool
 9 R², determination coefficient

10

1 **Table 3.** Values of the time required for 50% mass loss (half-life) and turnover rates of both
 2 labile (L) and recalcitrant (R) organic fractions of litter collected from different eucalyptus
 3 clone (Anselmo and Odriel) plantations estimated by applying a first-order kinetic model
 4 based on the double exponential equation* proposed by Andr en and Paustian (1987) to the
 5 cumulative quantity of C mineralized during 560 days of substrate incubation under
 6 laboratory controlled conditions and calculated as *Half-life L* = 0.693/*k* or *Turnover L* = 1/*k*,
 7 for the labile pool and as *Half-life R* = 0.693/*h* or *Turnover R* = 1/*h* for the recalcitrant
 8 reservoir, respectively.

	Labile fraction		Recalcitrant fraction	
	<i>Half-life L</i> (months.)	<i>Turnover L</i> (months)	<i>Half-life R</i> (months.)	<i>Turnover R</i> (months)
ANSELMO CLONE				
Granitic A1	1.9	2.8	128.3	185.2
Granitic A2	2.3	3.3	192.5	277.8
Granitic A3	1.3	1.9	85.6	123.5
Mean value	1.8	2.7	135.5	195.5
SD	0.5	0.7	57.8	77.7
<hr/>				
Schistic A1	0.9	1.2	56.3	81.3
Schistic A2	2.6	3.7	144.4	208.3
Schistic A3	1.7	2.4	105.0	151.5
Mean value	1.7	2.4	101.9	147.0
SD	0.9	1.3	44.1	63.6
<hr/>				
ODIEL CLONE				
Granitic O1	1.5	2.2	105.0	151.5
Granitic O2	1.5	2.2	105.0	151.5
Granitic O3	1.7	2.4	165.0	238.1
Mean value	1.6	2.3	125.0	180.4
SD	0.1	0.1	34.6	50.0

9 * $C_{\text{mineralized}} = C_o (1 - e^{-kt}) + C_r (1 - e^{-ht})$

10 C_o , carbon of the labile pool

11 C_r , carbon of the recalcitrant pool

12 k , instantaneous mineralization rate of the labile carbon pool

13 h , instantaneous mineralization rate of the recalcitrant carbon pool

14

15

Figure Captions

1
2 Figure 1. Evolution of the weight loss proportion and the C-to-N ratio during the
3 biodegradation of eucalyptus litter collected from two types of clone plantations (Anselmo or
4 Odiel clonal varieties developed over granitic or schistic parent material) as a function of the
5 incubation time (vertical bars are ± 1 standard deviation). Weight loss comparison between
6 both clonal varieties (A). Comparison of the C-to-N ratio between both clones (B). Weight
7 loss of litter for both bedrock types (C). Litter C-to-N ratio for both bedrock types (D).

8
9 Figure 2. Cumulative curves of the C mineralization during incubation of eucalyptus litter
10 collected from from two types of clone plantations (Anselmo or Odiel clonal varieties
11 developed over granitic or schistic bedrock). Comparison between litter from both clonal
12 varieties (A). Comparison between litter from granitic and schistic plantations (B). Vertical
13 bars are ± 1 standard deviation.

14
15 Figure 3. Evolution of the Isotopic ^{13}C composition during incubation of eucalyptus litter
16 collected from from two types of clone plantations (Anselmo or Odiel clonal varieties
17 developed over granitic or schistic bedrock) as a function of the incubation time. Comparison
18 between litter from both clonal varieties (A). Comparison between litter from granitic and
19 schistic plantations (B).

20

21