

Mechanisms of microbial carbon sequestration in the ocean

—future research directions

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1 **Abstract**

2 This paper reviews progress on understanding biological carbon sequestration in the ocean with special
3 reference to the microbial formation and transformation of recalcitrant dissolved organic carbon (RDOC),
4 the microbial carbon pump (MCP). We propose that RDOC is a concept with a wide continuum of
5 recalcitrance. Most RDOC compounds maintain their levels of recalcitrance only in a specific environmental
6 context (RDOC_i). The ocean RDOC pool also contains compounds that may be inaccessible to microbes due
7 to their extremely low concentration (RDOC_c). This differentiation allows us to appreciate the linkage
8 between microbial source and RDOC composition on a range of temporal and spatial scales.

9 Analyses of biomarkers and isotopic records show intensive MCP processes in the Proterozoic oceans
10 when the MCP could have played a significant role in regulating climate. Understanding the dynamics of the
11 MCP in conjunction with the better constrained biological pump (BP) over geological timescales could help
12 to predict future climate trends. Integration of the MCP and the BP will require new research approaches and
13 opportunities. Major goals include understanding the interactions between particulate organic carbon (POC)
14 and RDOC that contribute to sequestration efficiency, and the concurrent determination of the chemical
15 composition of organic carbon, microbial community composition and enzymatic activity. Molecular
16 biomarkers and isotopic tracers should be employed to link water column processes to sediment records, as
17 well as to link present-day observations to paleo-evolution. Ecosystem models need to be developed based
18 on empirical relationships derived from bioassay experiments and field investigations in order to predict the
19 dynamics of carbon cycling along the stability continuum of POC and RDOC under potential global change
20 scenarios. We propose that inorganic nutrient input to coastal waters may reduce the capacity for carbon
21 sequestration as RDOC. The nutrient regime enabling maximum carbon storage from combined POC flux
22 and RDOC formation should therefore be sought.

23
24 **Key words:** Biological pump (BP), microbial carbon pump (MCP), particulate organic carbon (POC),
25 recalcitrant dissolved organic carbon (RDOC), anthropogenic perturbation, climate change

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1. Introduction

The ocean absorbs approximately 30% of anthropogenic CO₂ (IPCC, 2013), mitigating global warming in a profound way. However the biological mechanisms for long-term carbon sequestration in the ocean are not fully understood. The biological pump (BP) is the collective term for a suite of processes by which carbon dioxide that is fixed by phytoplankton photosynthesis in the euphotic zone is exported to the deep ocean. These processes include the passive flux of sinking organic particles (dead cells, faecal pellets, etc.), the active flux of dissolved and particulate organic material mediated by vertical migration of zooplankton, and the vertical transport of dissolved organic material by physical processes. Around 50% of the photosynthetically produced particulate organic carbon (POC) is transformed through mechanisms including excretion, zooplankton grazing, viral lysis and the action of microbial ectohydrolases into dissolved organic carbon (DOC) (Anderson and Tang, 2010). The production rate and chemical composition of this dissolved organic matter (DOM) is influenced by the nutrient status and community composition of the microbial foodweb. Operationally DOC is defined as all compounds less than 0.2 μm in size (Carlson et al., 2002), and thus will include microparticulates (e.g., cell wall fragments, membranes, viruses etc.) and metabolites leaked/released by photo-autotrophs, defecated by phago-heterotrophs and associated with viral lysis of host cells. Marine microbes readily utilise most of this DOC, producing CO₂ and in turn transforming the composition of the DOM. However, an estimated ~5-7 % of the microbially produced DOC is recalcitrant (RDOC) and resists rapid remineralisation (Ogawa et al., 2001; Gruber et al., 2006; Koch et al., 2014), which enables the DOC to be exported below the seasonal thermocline and sequestered in the oceans' interior.

The microbial carbon pump (MCP) (Jiao et al., 2010a) describes the ecological processes and chemical mechanisms that produce RDOC throughout the water column. The resilience of RDOC to degradation by marine microbes is an important mediator of the global carbon cycle and the marine carbon pool. Since the current reservoir of RDOC is comparable to the inventory of atmospheric CO₂ (Hansell et al., 2009), trade off between the two carbon pools would influence climate change. Hence the relative rates of POC export, production of RDOC and respiration of POC and DOC regulate the timescale over which carbon is stored in the ocean's interior, and small changes to these rates would have a major, potentially detrimental, impact on atmospheric CO₂.

While numerous experiments have assessed the sensitivity of POC export to changes in stratification, mixing and remineralization depth (Kriest et al., 2010; Romanou et al., 2014), little attention has been paid to the environmental factors and anthropogenic perturbations, such as ocean acidification (OA) and

1 eutrophication, which might control the rates of RDOC production and transformation. Given the vast
2 abundance and diversity of microbes (ranging from auto- and hetero- trophic prokaryotes through to
3 photoauto-, mixo- and phagohetero- trophic protists), the complexity of microbial ecosystems and the
4 sensitivity of microbes to environmental change, small shifts in microbial metabolic efficiency potentially
5 cause large changes to carbon sequestration (Mitra et al., 2014). Without fully understanding the microbial
6 processes, we risk overlooking a crucial feedback of the overall system that is caused by a seemingly minor
7 perturbation of an individual process.

8 The production and transformation of RDOC is intricately linked with the production and
9 transformation of POC; thus it is timely to investigate these interactions. It is not known whether the cycling
10 of POC and DOC would interact to enhance or decrease their individual effects, but it is possible that
11 perturbations such as eutrophication or increasing temperature could cause a shift in the balance of carbon
12 sequestration via dissolved versus particulate forms. In addition, their combined response to environmental
13 conditions may be regulated differently under different conditions, for example, in coastal eutrophic waters
14 compared to oceanic oligotrophic waters.

15 A multidisciplinary effort is required to address these challenges. To this end, the international
16 IGBP/SCOR programme Integrated Marine Biogeochemistry and Ecosystem Research (IMBER), convened
17 a workshop entitled “The impact of anthropogenic perturbations on open ocean carbon sequestration via the
18 dissolved and particulate phases of the biological carbon pump”, at the IMBIZO III conference in Goa, India
19 in January 2013. Microbial ecologists, marine biogeochemists, organic chemists, climatologists, fisheries
20 scientists and economists presented recent research on the biological and microbial carbon pumps, discussed
21 future natural and social science research needs to integrate POC and DOC research, and brainstormed to
22 better understand the microbial carbon storage mechanisms of the ocean.

23 The objective of this paper is to identify the challenges of and devise strategies for the integration of
24 observations and models of ocean POC and DOC cycling and sequestration, with reference to: the chemical
25 composition of marine DOC, the microbial processing of DOC, the environmental controls on the
26 composition and processing of DOC, interactions between POC and DOC cycling, impacts of anthropogenic
27 perturbations on the BP and MCP in different environments, and approaches for ecosystem sustainability
28 and management.

29 30 **2. The nature and controls of DOC in the ocean**

1 Most of the DOC produced by photosynthesis is labile and can be remineralized or assimilated by
2 microbes within minutes to a few days (Fuhrman, 1987). The remaining DOC can be gradually degraded
3 and transformed by microbes and abiotic processes to a huge variety of new compounds with residence
4 times from days to months, decades, hundreds and even thousands of years (Sherr, 1988; Marchant and Scott,
5 1993). The oceanic DOC pool has been classified into two major classes: labile DOC (LDOC) which does
6 not accumulate in the ocean due to rapid microbial turnover and recalcitrant DOC (RDOC) which serves as
7 a reservoir until its eventual mineralization or removal. In some studies, RDOC is subdivided into fractions
8 defined by their lifetimes: semi-labile (~1.5 years), semi-refractory (~20 years), refractory (~16 000 years)
9 and ultra-refractory (~40 000 years) (Hansell et al., 2012). Obviously there are major gaps between the
10 timescales describing the recalcitrance of RDOC, to say nothing of the difference in definitions between
11 scientific disciplines. While geochemists can define RDOC according to the ^{14}C age on time scales of
12 thousands of years and modelers use the turnover rates as the basis of their definition for the different types
13 of RDOC, microbiologists may identify RDOC according to the absence of the genes that encode the
14 enzymes required to metabolize the specific RDOC compound with no link to the time scale required. Such
15 differences must be identified and addressed to ensure the interdisciplinary collaborations needed for a
16 comprehensive understanding of the interactions between microbes and their geochemical environment and
17 the consequences of microbial processing of carbon on outgassing of CO_2 and carbon sequestration.

18 While age can be used to identify RDOC, not all old DOC is RDOC. For example, petroleum
19 components can be very old but when exposed to microbial action they can be rapidly decomposed. In
20 addition, different microbes have different decomposition capabilities under different environmental
21 conditions. Thus, recalcitrance can vary between different species, different functional groups and different
22 environments (Carlson et al., 2011; Jiao et al., 2011; Kujawinski, 2011). The classification of RDOC as
23 microbial species-, functional group- or environmental context-specific recalcitrant could diminish
24 confusion between biological and geochemical descriptions. Therefore we propose the term $\text{RDOC}_{\text{context}}$
25 (RDOC_t) (Fig. 1).

26 For a better understanding of the nature and behavior of RDOC, effort needs to be directed to isolation
27 of different DOC molecules and subsequent chemical analyses of the principal components of the DOC pool.
28 One promising approach is ultrahigh resolution mass spectrometry (Fourier transform ion cyclotron
29 resonance mass spectrometry; FT-ICR-MS), which is based on the analysis of exact molecular masses from
30 which the molecular elemental composition of marine DOM can be deduced (Koch et al., 2005; Hertkorn et
31 al., 2006). To date several thousand molecular formulae have been identified and many of them may act as

1 potential indicators for microbial sources and transformation processes (e.g. Kujawinski et al., 2004;
2 Gonsior et al., 2009). Solid-phase extracted DOM is composed of highly oxygenated molecules (average
3 oxygen to carbon ratio ~0.45), which implies that they should be utilizable by prokaryotes. This high oxygen
4 content, which primarily exists in carboxylic functions, reflects a high degree of polarity and may therefore
5 need a highly specific and energy-efficient uptake system by distinct microbes (Kattner et al., 2011). A low
6 average proportion of hydrogen (average hydrogen to carbon ratio ~1.25) reflects a substantial proportion of
7 stable aromatic backbones, structures known to be difficult for prokaryotes to degrade. Approximately
8 one-third of the detectable formulae are present in all marine samples and most likely represent a common
9 refractory background in DOM (Koch et al., 2005; Kattner et al., 2011). Within limits (on a molecular
10 formula level), FT-ICR-MS allows us to distinguish between labile and refractory marine DOM generated
11 within the MCP. D-Glucose incubations show that microbially-derived marine DOM resembles labile
12 material and that longer incubations are required to reach refractory element compositions (Koch et al.,
13 2014). Recently, the chemical composition of DOM has been related to the degradation state and age of
14 DOM (Flerus et al., 2012; Lechtenfeld et al., 2014). These studies reveal that the most persistent compounds
15 encompass a very narrow range of average molecular elemental ratios H/C and O/C and show a continuum
16 of residence times of refractory DOM in the ocean, the longest of which substantially exceeded the average
17 age of marine DOC of ~5000 years (Bauer et al., 1992).

18 Besides describing the molecular composition of RDOC, understanding the microbial inaccessibility of
19 RDOC is essential to determining why the ocean holds such a huge DOC pool in the presence of such an
20 abundance of microbes. Appropriate (meta-) genomic and (meta-) transcriptomic methods are now available
21 to examine the microbial genetic and enzymatic repertoire for cleaving and decomposing as well as taking
22 up and transforming DOC compounds (Kujawinski, 2011). For example, the average genome of marine
23 bacteria contains 3,000-5,000 proteins, which according to comparative genomics analysis share similarities
24 for primary metabolic pathways but differences for specific substrate assimilation.

25 Genes associated with cross-membrane transport, extracellular hydrolysis, motility and chemotaxis are
26 critical for accessing the breadth of DOC molecules available for microbial assimilation. *Roseobacter* strains
27 tend to assimilate carbohydrate-rich DOC while SAR11 bacteria prefer nitrogen-containing DOM because
28 they have suites of high-affinity carbohydrate and amino acid ABC transporter systems, respectively (Jiao
29 and Zheng, 2011). *Bacteroides* and *Gammaproteobacteria* are able to consume a diverse array of DOC
30 because they have TonB-dependent transporter genes (Tang et al., 2012). *Bacteroides* can take up and
31 assimilate N-acetyl glucosamine while SAR11 cannot, due to a lack of N-acetyl glucosamine transporter and

1 its deacetylase. Genes associated with motility and chemotaxis vary from ~0.5% to ~1.2% in the
2 metagenomes of common marine environments. These genes provide a mechanism by which
3 microorganisms can respond to microscale DOC gradients and access nutrient-enriched patches (Stocker,
4 2012).

5 High-molecular-weight (HMW) compounds must be cleaved into smaller chemical units by
6 extracellular enzymes before microbial uptake (Arnosti, 2011). It is hard for cell wall materials such as
7 peptidoglycan (accounting for 2% of the cell biomass) (Park and Uehara, 2008) to be decomposed
8 completely when they are released as fragments into the environment during viral lysis or grazing processes.
9 Usually heterotrophic prokaryotes need at least seven combined enzymatic transformations to cleave and
10 decompose peptidoglycan for reutilization (Jiang et al., 2010). Even if peptidoglycan is cleaved, the
11 fragments containing certain components such as N-acetylglucosamine - N-acetylmuramic acid and
12 anhydro-N-acetylmuramic acid can remain inaccessible to the uptake and assimilation systems of some
13 microbes; however, many heterotrophic prokaryotes can take up and metabolize N-acetylglucosamine
14 (Riemann and Azam, 2002). Although D- amino acids can be transformed into L-amino acids by racemases
15 inside the cell (Jørgensen and Middelboe, 2006), the transformation can not be carried out extracellularly,
16 and if no membrane transporter is available then this leads to the accumulation of D-amino acids in the
17 water column as RDOC. Structural RDOC molecules such as many D-amino acids (D-cysteine,
18 D-tryptophan, D-tyrosine), are intrinsically very resistant to microbial utilization. Theoretically, an organic
19 molecule/compound can be intrinsically recalcitrant to a specific microbial species or functional group if the
20 microbes do not have the gene that encodes the corresponding enzyme to take up or decompose the
21 molecule/compound. However, in the natural environment of diverse microbes and variable conditions, all
22 RDOC molecules/compounds are in a transitional stage, subject to further cleaving or decomposition, and
23 their recalcitrance is a continuum dependent upon microbial community structure and environmental
24 conditions. Throughout the water column, microbial processing alters the nature of DOC through
25 decomposition, assimilation and regeneration. Microbes can produce complex structures, such as biofilms,
26 and low molecular weight (LMW) molecules such as antibiotics, toxins and virulence factors. In addition, in
27 the surface water, photochemical reactions alter the composition of DOC, produce LMW organic
28 compounds (Kieber et al., 1990) and influence the availability of DOC to microbes. The successive and
29 repetitive processing of DOC compounds by the diverse prokaryote community could generate smaller and
30 smaller fragments forming a LMW DOC pool. Although the total concentration of this LMW DOC pool is
31 not low (~40 μM , about half of the surface ocean DOC concentration), since it is composed of billions of

1 different molecular species (Baldock et al., 2004; Koch et al., 2005), the concentration of most individual
2 LMW DOC constituents would be extremely low. This, rather than their recalcitrance could prevent
3 energy-efficient microbial uptake (Kattner et al., 2011; Stocker, 2012). Although microbes are able to
4 exploit substrates at very low concentrations, a low threshold exists (Jannasch, 1995) for energetically
5 profitable substrate utilization (Barber, 1968; Kattner et al., 2011). These LMW DOC molecules would stay
6 inaccessible to microbial uptake until they accumulate to a threshold level.

7 Based on the above considerations, RDOC can be classified into two categories, environmental
8 context-dependent RDOC_t and concentration-constrained RDOC (RDOC_c). RDOC_t would be recalcitrant in
9 a given biogeochemical context but could become accessible to microbial degradation in a different context,
10 RDOC_c would be composed of molecules at extremely low individual concentrations, which are below the
11 corresponding microbial uptake thresholds. Organic carbon ages as it is transformed gradually and
12 successively from labile to recalcitrant, from young HMW compounds in the upper ocean to old LMW
13 compounds in the deep ocean, thus creating the continuum between microbial and geochemical processing
14 of RDOC in the ocean (Fig. 1).

16 **3. RDOC processing in current and ancient oceans**

17 The activity of marine microorganisms leaves fingerprints in the geological records that are traceable
18 using organic biomarkers and stable carbon isotopes. Thus integration of microbial identity and function
19 with stable carbon isotopes on a geological time scale can improve our understanding of the mechanisms
20 and processes of DOC production, accumulation and transformation in the modern ocean as well as their
21 relationships with climate variability.

22 In addition to its present day role, the MCP may have been crucial in the formation of a huge RDOC
23 reservoir in the Pre-Cambrian Ocean. A MCP mediated by sulfate-reducing or iron-reducing microbes under
24 hypoxic or anoxic conditions may have facilitated the accumulation of authigenic carbonate (i.e. derived
25 from DOC) in sediments or bottom water, which may have played an important role in the global carbon
26 budget through earth's history (Canfield and Kump, 2013; Schrag et al., 2013).

27 Geochemical records indicate an intensive prokaryote driven MCP, and production of a large RDOC
28 reservoir in ancient oceans. Logan et al. (1995) found that the *n*-alkyl lipids preserved in Proterozoic rocks
29 are generally isotopically heavier than coexisting isoprenoidal lipids, while the opposite is observed for most
30 modern and Phanerozoic sediments. This suggests that in the Proterozoic ocean, the *n*-alkyl lipids received

1 stronger heterotrophic reworking than recalcitrant isoprenoidal lipids and that the MCP was therefore
2 stronger in the Proterozoic oceans relative to that in modern and Phanerozoic oceans. A strong negative shift
3 (down to -15‰) in the C-isotopic composition of sedimentary carbonates alongside a generally unchanged
4 C-isotopic composition of coexisting organic matter (Fike et al., 2006; Swanson-Hysell et al., 2010;
5 Grotzinger et al., 2011) during the Neoproterozoic (~0.85 to 0.54 Ga) has been proposed to indicate the
6 presence of an unusually large RDOC reservoir at this time (Rothman et al., 2003).

7 The development of this large RDOC reservoir coincided with a series of extreme “Snowball Earth”
8 glaciations (Swanson-Hysell et al., 2010), culminating in the birth of the earliest animals on Earth (Fike et
9 al., 2006; McFadden et al., 2008). It is likely that the great glaciations set up a lateral gradient of oxidants in
10 the postglacial oceans (Li et al., 2010), which not only favored an intensive anaerobic MCP in shallow
11 subsurface waters, but also created an extremely reduced deep ocean for storage of the resulting RDOC;
12 these favorable geochemical conditions allowed the accumulation of the largest ocean RDOC reservoir
13 known in the Earth’s history (at least 10^2 - 10^3 times larger than the modern RDOC reservoir in size and 10^4
14 years longer in turnover time (Rothman et al., 2003)). Further work is required on the biogeochemical
15 mechanisms and effects of the unusual accumulation of RDOC in the Neoproterozoic deep ocean, as
16 understanding the processes involved in the MCP in the deep past is important to improve our predictive
17 capability for a future ocean where anoxia is likely to increase (and thus potentially increase the ocean
18 carbon storage capacity via the RDOC reservoir).

19 20 **4. Interactions between POC and DOC sequestration**

21 Differentiation of carbon sequestration to either the particulate or dissolved phase depends on the size
22 threshold used to divide POC from DOC, and the lifetime of the various size fractions. Organic matter
23 produced by the marine foodweb covers a size range of almost 10 orders of magnitude, from the smallest
24 organic molecules (e.g. glucose, 0.7 nm) to baleen whales (up to 30 m). For convenience, researchers divide
25 this size range into DOC and POC (the threshold depending on the filter used to retain particles), but
26 actually the size distribution is almost continuous. The lifetime of any substance is defined, assuming
27 exponential decay, as the time over which its concentration decreases to $1/e$ of its initial value, where e is the
28 Napierian constant 2.71828; this corresponds to the “ e -folding lifetime”, which is different from the related
29 concept of “half-life” where $1/2$ is used instead of $1/e$ (Hansell, 2013). Organic matter has lifetimes that range
30 from less than a day to tens of thousands of years. The rates of production of the DOC fractions defined by

1 Hansell (2013) are inversely related to their average lifetimes, i.e. organic compounds with a long lifetime
2 are produced at small rates according to the following equation (based on values given in Table 1 of Hansell,
3 2013):

$$\log_{10}(\text{production within a DOC fraction}) = 0.29 - 0.40 \log_{10}(\text{average lifetime of the fraction}) \quad (1)$$

4 with $r^2 = 0.96$. In addition, the intrinsic lifetimes of organic compounds may be significantly
5 lengthened by their storage in geochemical reservoirs. For example, organic matter that sinks into deep
6 water may be either remineralised (respired) to CO_2 , or buried in sediments. In the first case, the CO_2 that is
7 dissolved in deep waters will return to the atmosphere on average about 1000 years later. In the latter case,
8 the organic carbon may be incorporated in sediments, carried by the ocean floor until it is subducted in
9 trenches (the age of the ocean floor is generally <125 million years), and released to the atmosphere by
10 volcanoes, or incorporated into continental rocks until ultimately released to the atmosphere by weathering.
11

12 The Intergovernmental Panel on Climate Change (IPCC) (2013) defines carbon sequestration as the
13 addition of carbon containing substances to a reservoir, e.g. the ocean, which has the capacity to store,
14 accumulate or release carbon. Economists are interested in the timescale of carbon sequestration because
15 companies or countries can earn carbon credits by artificially capturing and securing the storage of carbon
16 that would otherwise be emitted to or remain in the atmosphere. Within the context of ocean fertilisation,
17 Lampitt et al. (2008) proposed that sequestration requires carbon which persists at least 100 years.
18 According to this 100-year timescale (e.g. Legendre and Le Fèvre, 1991; Legendre and Le Fèvre, 1995;
19 Legendre and Rassoulzadegan, 1996; Passow and Carlson, 2012), POC that reaches deep ocean waters is
20 sequestered, as are the most refractory fractions of RDOC. Carbon sequestration is also achieved by the
21 solubility and the carbonate pumps (Volk and Hoffert, 1985).

22 The rate of POC sequestration can be estimated from the POC sinking flux measured in sediment traps
23 at 2000 m. Estimates of this flux range from $0.43 \text{ PgC year}^{-1}$ (Honjo et al., 2008) to $0.66 \text{ PgC year}^{-1}$ (Henson
24 et al., 2012). The POC flux to the sediment is estimated to be $0.1\sim 0.16 \text{ PgC year}^{-1}$ (Hedges and Keil, 1995;
25 Prentice et al., 2001). In contrast, there are few estimates of the rate of DOC sequestration. Using the
26 100-year sequestration criterion, a minimum estimate would be the combined production rates of refractory
27 and ultra-refractory DOC (average lifetimes of 16 000 and 40 000 years, production of 0.043 and
28 $0.000012 \text{ Pg C year}^{-1}$) (Hansell, 2013). In addition, at least part of the production of semi-refractory DOC
29 should be included, since its average lifetime is 20 years (production of $0.34 \text{ Pg C year}^{-1}$) (Hansell, 2013)
30 and that of the next fraction, refractory DOC, is 16 000 years. Hence the combined production rates of these

1 three fractions of RDOC would be $0.38 \text{ Pg C/year}^{-1}$, roughly consistent to earlier estimates of $0.5\text{-}0.6 \text{ Pg C}$
2 year^{-1} (Brophy and Carlson, 1989). It seems that sequestration from the RDOC-based MCP and the
3 POC-based BP are of the same order of magnitude.

4 Sequestration would be high when there is rapid downward transport of POC or substantial
5 transformation of organic matter to RDOC. The interactions between POC flux and RDOC production are
6 numerous. For example, the attenuation of POC flux is accompanied by DOC generation throughout the
7 water column, while the microbial transformation of DOC can also be accompanied by the formation of
8 particles large enough to sink (Fig. 2). About 10% of marine DOC exists in the form of gels which harbor
9 heterotrophic prokaryotes (Azam and Malfatti, 2007) and can accelerate carbon transformation (Ziervogel et
10 al., 2011), while POC or aggregates attract copiotrophs such as *Bacteroides* (Arnosti et al., 2012) that have
11 motility and chemotaxis genes and can potentially follow DOC gradients (Stocker, 2012). Many marine
12 heterotrophic prokaryotes produce polysaccharides, which help them attach to biotic and abiotic surfaces to
13 form aggregates. The matrix of the aggregate, known as extracellular polymeric substances (EPS) or
14 transparent exopolymer particles (TEP), is composed of polysaccharides, proteins, nucleic acids, and lipids.
15 These cohesive, three-dimensional polymers interconnect cells, forming aggregates which then contribute to
16 POC flux. Microbes colonize sinking aggregates, and can grow by means of exoenzymatic decomposition of
17 the aggregated organic particles, which in turn could lead to a DOC plume following the sinking aggregate.
18 In fact, this plume may account for a significant fraction of the microbial production and remineralization
19 (Kiorboe and Jackson, 2001). Thus the balance between the rate at which aggregates form and sink on the
20 one hand and the rate at which they are remineralized and secrete DOC on the other hand has a major impact
21 on ocean carbon flux. It has been hypothesized that the strategy of heterotrophic prokaryotes in oligotrophic
22 environments is to grow in biofilms on surfaces where nutrients are locally available, and to persist in
23 nutrient- deprived zones such as floating biofilms with the capacity to return to optimum growth when
24 nutrients again become available (Costerton et al., 1995). This is supported by evidence suggesting that
25 carbon-limited deep-ocean prokaryotes show a preferential particle-related life strategy (DeLong et al., 2006;
26 Arístegui et al., 2009; Baltar et al., 2009; Baltar et al., 2010b).

27 Aggregating and scavenging processes are common in the water column. Sinking POC particles could
28 be nuclei for RDOC molecules to attach to or aggregate with (Druffel and Williams, 1990; Hwang and
29 Druffel, 2003; Roland et al., 2008). If the POC particles scavenge enough RDOC molecules, they could
30 become coated with RDOC (Fig. 2). As POC is relatively labile, it is considered to be an important food
31 source for deep ocean microbes. In the case of "RDOC-coated POC", since RDOC is resistant to microbial

utilization, if the POC particle is coated with enough RDOC molecules, it is theoretically no longer subject to microbial attack and can safely reach the seabed where it can be buried for millions of years. In a simplified scenario that a spherical particle with diameter of d is sinking directly from the bottom of the euphotic zone to the sea floor, the carbon content of this particle is:

$$POC = \frac{4}{3} \pi \cdot \left(\frac{d}{2}\right)^3 \cdot \rho \quad (2)$$

where ρ is the particle's carbon density. Assuming a weight density of the particle of 3 mg mm^{-3} (Carder et al., 1982) and the molar ratio of 106:16:1:106 for C:N:P:O of the particle, we can estimate $\rho=0.1 \text{ mmol C mm}^{-3}$. If the depth from the bottom of the euphotic zone to the seabed is z , the path of the sinking particle forms a water column with diameter d and length z , in which all the RDOC molecules ($RDOC_{ect}$) are encountered by the particle:

$$RDOC_{ect} = \pi \cdot \left(\frac{d}{2}\right)^2 \cdot z \cdot [RDOC] \quad (3)$$

We can estimate $[RDOC]$ to be $40 \text{ }\mu\text{M}$ and z to be $4,000 \text{ m}$ for the typical open ocean. If the probability that a RDOC molecule is scavenged by the particle is p , the carbon ratio of the RDOC and the original POC in this particle after reaching the sea floor is:

$$r = \frac{POC}{RDOC_{ect} \cdot p} = \frac{2}{3} \cdot \frac{\rho}{z \cdot [RDOC]} \cdot \frac{d}{p} \quad (4)$$

By using the parameter values we estimated above, Eq. (4) simplifies to

$$r = 2400 \cdot \frac{p}{d} \text{ mm}^{-1} \quad (5)$$

This equation indicates that the ratio r depends on the particle size and the scavenge probability. For a small particle with diameter of order 0.01 mm , a very low scavenge probability $p=4 \times 10^{-6}$ will roughly give $r \approx 1$, i.e. about half of the carbon in the particle reaching the seabed is from RDOC. For a larger particle with diameter of $\sim 1 \text{ mm}$, the same scavenge probability would predict that $\sim 1\%$ of the carbon of the particle reaching the seabed is from RDOC.

This calculation is a simplification as it does not limit the maximum number of RDOC molecules that can be scavenged by a particle and does not consider eddies and currents generated by the sinking particles. In addition, if a particle does not sink directly to the sea floor but also moves horizontally or even upwards,

1 its path can be even longer and so it would encounter more RDOC molecules.

2 Another effect of the “RDOC coating process” is bias in ^{14}C dating (Fig.2). As indicated in the above
3 calculation, assuming the average age of RDOC is 5000 yr, a one year old POC particle landing at the
4 seabed could be falsely dated as 50-2500 yr old depending on its size (0.01-1mm) using our assumed
5 scavenge probability. In fact, one of the loss processes of RDOC in the water column is aggregation that
6 ultimately leads to transfer of aged organic carbon as POC to the sediment (Engel et al., 2004b; Jiao et al.,
7 2010a).

8 Physical processes such as stratification, mixing and ocean currents influence carbon sequestration in
9 the ocean. Increasing stratification restricts nutrient supply from deep water to the euphotic zone, and
10 therefore primary production which will in turn impact the export of POC (Doney, 2006; Capotondi et al.,
11 2012; Passow and Carlson, 2012). Episodic vertical water movement such as solitary waves could enhance
12 POC flux and these are thought to be responsible for the unexpected presence of *Prochlorococcus* in aphotic
13 waters (300-1000m) in the western Pacific marginal seas (Jiao et al., 2014). Mesoscale eddies are ubiquitous
14 features in the ocean (Cheney and Richardson, 1976; Arístegui et al., 1997; van Haren et al., 2006), and
15 could play a major role in the generation, accumulation and downward transport of biogenic production in
16 the ocean. Cyclonic eddies enhance nutrient inputs to the surface ocean increasing new production
17 (Falkowski et al., 1991; Harris et al., 1997; Moran et al., 2001) and chlorophyll concentrations (Arístegui et
18 al., 1997; McGillicuddy Jr. et al., 1998; Tarran et al., 2001). The presence of eddies has also been related to
19 increased bacterial abundance (Arístegui and Montero, 2005) and production (Bode et al., 2001; Baltar et al.,
20 2007), even in the mesopelagic zone (Baltar et al., 2010a). However, the contribution of eddies to particle
21 flux is still poorly constrained. Mesoscale eddies were shown to enhance POC export by a factor of 2-4
22 (Alonso-González et al., 2010) in the Canary Island region, whereas eddies in Hawaii did not increase the
23 efficiency of POC export to mesopelagic waters as most of the particle production was rapidly remineralized
24 in the upper 150 m (Maiti et al., 2008).

25 A recent study of cyclonic eddies in the western South China Sea (Jiao et al., 2014) suggested that the
26 intensity, timing and duration of nutrient input influenced the plankton community structure which affected
27 whether eddy induced upwelling was associated with an increase (diatom dominated) or decrease
28 (dominance of cyanobacteria) in POC flux. Legendre and Le Fèvre (1995) previously stressed the significant
29 role of the microbial food web in carbon export. Along a nutrient gradient from eutrophy to oligotrophy,
30 POC export decreases as there is a transition in the structure of the microbial food web from phytoplankton

1 prey-microzooplankton predators to picoplanktonic cyanobacteria, heterotrophic bacteria, and archaea prey
2 and mixotrophic protist predators (Zubkov and Tarran, 2008; Hartmann et al. 2012). The ratio of DOC
3 production to total primary production increases with increasing oligotrophy (Teira et al., 2001), with some
4 of this DOC likely converted to RDOC. Thus, the contribution of the MCP to carbon storage could be
5 expected to be relatively high in the oligotrophic ocean. A similar transition from dominance of the BP to
6 dominance of the MCP might be expected along a latitudinal gradient from polar regions to the tropics and
7 from surface waters to the mesopelagic (Fig. 3).

9 **5. Impact of anthropogenic perturbations on carbon sequestration**

10 **5.1 Relevance to society**

11 The BP and MCP operate in concert to keep a large reservoir of carbon out of the atmosphere by storing
12 POC, DOC and dissolved inorganic carbon (DIC) in the ocean. Without marine biological carbon
13 sequestration, it has been estimated that the atmospheric CO₂ concentration would be 50% (200 ppmv)
14 higher than the current value (Parekh et al., 2006). This storage of carbon thus has intrinsic value as an
15 ‘ecosystem service’ (e.g. Luisetti et al., 2011). The term “Blue Carbon” has previously been applied to
16 coastal ecosystems which have the capacity to store carbon year-on-year, with the intention of valuation and
17 possible subsequent carbon trading (Ullman et al., 2013) but that definition has not previously been
18 extended to continental shelf sediments or the open ocean (Grimsditch et al., 2013).

19 The deep ocean natural carbon store is relatively secure on short (decadal to centennial) timescales due
20 to the long residence times of DIC (10³ years) and DOC (10⁴ years) in the deep ocean. More relevant to
21 society and how our activities may impact this system is the balance between input and output terms of the
22 ocean carbon inventory. Any action we can take to increase the efficiency of the BP and MCP, or reduce the
23 rate of the return pathway(s) will lead to net accumulation of carbon in the deep ocean. Assuming the net
24 biological pump (BP+MCP) is ~1-10 Pg C year⁻¹ (IPCC, 2013), this represents roughly 1-10% of net global
25 primary production and between 10 and 100% of global anthropogenic CO₂ emissions. Thus, for example, a
26 10% increase in the annual input term to the ocean carbon store could lead to significant additional annual
27 carbon sequestration. Conversely a decrease of the input term or increase to the output could lead to
28 significant additional emissions. Thus understanding the interactions between our actions (and their
29 subsequent effects) and the efficiency of the BP and MCP are of particular importance, both for
30 understanding the likely response to future global change and in informing whether or how marine

1 management options might be employed to enhance (or reduce degradation of) pump efficiency.

2 Many of the interactions over which we may be able to exercise management options take place in
3 shelf seas, which are active areas for DOM cycling (Prowe et al., 2009; Johnson et al., 2013) and carbon
4 export (Tsunogai and Noriki, 1991; Thomas et al., 2004). Although covering only 8% of the ocean's surface
5 area, they account for 20% of the ocean's capacity to absorb CO₂ (Thomas et al., 2004). Shelf seas are also
6 the regions subject to strongest human pressures (Emeis et al., 2014), thus they represent a strong 'pressure
7 point' for controlling BP and MCP efficacy. These human pressures include nutrient input, hypoxia and
8 trawling amongst others, and we do not yet know how these pressures or combinations of pressures will
9 affect carbon storage. In the following section we consider the potential effects of two important
10 anthropogenic forcings: i) nutrient input to the oceans (a largely shelf sea pressure) and ii) ocean
11 acidification (a global pressure).

12 **5.2 Nutrient supply**

13 Generally, an increase in nutrient supply to coastal waters is expected to lead to an increase in primary
14 production, POC, and consequently an increase of the BP. However high nutrient concentrations could have
15 a negative impact on the MCP (Jiao et al., 2010b). Primary production, bacterial respiration and bacterial
16 growth efficiency would respond differently to increasing nutrients (Fig. 4). As phytoplankton populations
17 increase with increasing nutrients, a maximum will be reached when light shading becomes important and
18 primary production starts to decrease. In contrast, providing DOC was in adequate supply, heterotrophic
19 bacterial growth would be less influenced by the light field. The phytoplankton bloom would provide a
20 steady supply of labile DOC for bacterial growth, together with riverine derived semi-labile DOC (SL-DOC)
21 (Fig. 4). Oxygen consumption due to this bacterial respiration can eventually lead to hypoxia. Under
22 hypoxic and anoxic condition, anaerobic bacteria would degrade the remaining organic matter, generating
23 gasses such as methane and H₂S. The former is a potent greenhouse gas and the latter is a potential source of
24 acidic rain (Fig. 5). Such scenarios could have occurred during geological events in the history of the earth.
25 In terms of carbon preservation, even if more carbon is fixed, it does not necessarily lead to increased carbon
26 storage. . This is verified by a systematic field survey which indicated an inverse correlation between nitrate
27 and organic carbon in all terrestrial and marine environments (Taylor and Townsend, 2010). Thus excess
28 nutrients can lead to lower organic carbon storage.

29 On the other hand, if nutrient input were reduced, although the BP is apparently decreased, the organic
30 matter that is produced would have relatively high C/N or C/P ratios, and thus be of poor food quality for

1 zooplankton resulting in high ingestion and gut transit rates and low digestion. Such organic matter would
2 also be relatively resistant to microbial utilization, enhancing the MCP. Microbial carbon accumulation is
3 known to occur where/when nutrients are limiting (Carlson et al., 2002; Gasol et al., 2009; Lauro et al., 2009;
4 Thomas et al., 1999; Jiao et al., 2010b). For example, in postbloom, nutrient depleted conditions in
5 temperate systems, net CO₂ fixation continues with the carbon likely being stored in high C/N, or high C/P
6 DOM, while the POC/PON still approximates Redfield values (Craig et al., 2013). In support of this,
7 oceanic DOC concentrations tend to be highest in the low nutrient oligotrophic gyres (Hansell et al., 2009).
8 Even in eutrophic waters, as long as the C/N ratio reaches a threshold value, microbial cells will start to
9 store carbon and produce more recalcitrant compounds or polymers (Bhaskar and Bhosle, 2005; Kadouri et
10 al., 2005; Jiao et al., 2010b). In a 14-day *in situ* nutrient enrichment experiment undertaken in the Western
11 Pacific oligotrophic gyre, more than 30% of the ambient organic carbon was respired in the incubation with
12 addition of inorganic nutrients compared to the control (Liu et al., 2014).

13 **5.3 Ocean acidification**

14 The absorption of CO₂ by the ocean results in an increase in the partial pressure of CO₂ (pCO₂),
15 bicarbonate ion [HCO₃⁻] and hydrogen ion [H⁺] concentration, and a decrease in carbonate ion concentration
16 [CO₃²⁻] – so-called ocean acidification (OA) (Doney et al., 2009). Enhanced photosynthesis and nitrogen
17 fixation have been shown to occur under higher pCO₂ conditions in laboratory and field experiments (Fu et
18 al., 2007; Riebesell and Tortell, 2011). Phytoplankton production of TEP is stimulated by higher pCO₂
19 treatments at both species and community levels (Engel, 2002; Engel et al., 2004a; Mari, 2008; Pedrotti et
20 al., 2012), which contributes to the formation of aggregates and so the vertical flux of POC and DOC. OA
21 inhibits phytoplankton calcification which will decrease the ballast effect of calcium carbonate, so
22 decreasing the vertical transport of POC (Barker et al., 2003). In addition, the efficient degradation of TEP
23 by marine microbes (Koch et al., 2014) may be enhanced at lower pH (Piontek et al., 2010). Shifts in
24 phytoplankton community structure have occurred in high CO₂ treatments that could impact the
25 composition and bioavailability of the DOC produced (Tortell et al., 2002; Paulino et al., 2008; Brussaard et
26 al., 2013). Although there is no clear impact of OA on bacterial abundance (Rochelle-Newall et al., 2004;
27 Grossart et al., 2006; Allgaier et al., 2008; Paulino et al., 2008; Newbold et al., 2012; Brussaard et al., 2013),
28 gross and cell specific bacterial production are usually stimulated by high pCO₂ treatments (Grossart et al.,
29 2006; Allgaier et al., 2008; Motegi et al., 2013; Piontek et al., 2013). The activity of bacterial protease,
30 glucosidase and leucine-aminopeptidase is also stimulated by higher pCO₂ (Grossart et al., 2006; Piontek et

1 al., 2010; Piontek et al., 2013). Changes in the community structure of bacterioplankton were observed when
2 pCO₂ was artificially adjusted (Allgaier et al., 2008; Newbold et al., 2012; Zhang et al., 2013). However, it
3 is possible that the observed responses of bacterioplankton to OA are due to tight coupling of phytoplankton
4 and heterotrophs in experiments with whole water samples. Nevertheless, higher bacterial activities in high
5 pCO₂ conditions may reduce carbon sequestration by POC flux but enhance the efficiency of the MCP by
6 producing more RDOC (Piontek et al., 2010; Piontek et al., 2013). Indeed, Kim et al. (2011) found an
7 enhanced DOC: POC production ratio in higher pCO₂ treatments in a mesocosm study. Still, we have a very
8 limited understanding of ecological processes (such as viral lysis and grazing) involved in both POC and
9 DOC cycling under high CO₂ conditions.

11 **6. Strategies for future research to maximize carbon storage in the ocean**

12 **6.1. Monitoring**

13 Satellite remote sensing of the surface ocean's optical properties (ocean color) has been fundamental in
14 developing the prevailing view of global ocean phytoplankton production and the BP. However, standard
15 band-ratio chlorophyll products, designed for "Class I" open ocean waters, have limitations when dealing
16 with other optically active constituents related to DOC transformation. The premise underpinning the "Class
17 I" water classification is that all optical properties co-vary with phytoplankton. However, increasing
18 evidence that aged DOC occurs at significant concentrations in the open ocean (Hansell et al., 2009),
19 independent of phytoplankton dynamics, may require this assumption to be re-evaluated. A range of
20 products have been developed that give an indication of the surface distributions of pools of organic carbon
21 constituents in the ocean, including colored (or chromophoric) dissolved organic material (CDOM; Siegel et
22 al., 2002; Maritorena and Siegel, 2005; Siegel et al., 2005; Morel and Gentili, 2009), DOC (Mannino et al.,
23 2008), POC (Stramski et al., 1999; Loisel et al., 2002; Gardner et al., 2006; Sathyendranath et al., 2009;
24 Stramska, 2009), phytoplankton size classes (PSC; Ciotti and Bricaud, 2006; Hirata et al., 2008a; Brewin et
25 al., 2010a; Brewin et al., 2010b; Uitz et al., 2010; Devred et al., 2011; Hirata et al., 2011) and particle size
26 distribution (PSD; Hirata et al., 2008b; Kostadinov et al., 2009; Kostadinov et al., 2010). Key uncertainties
27 relate to the relationship between the absorption of light by CDOM and concentrations of DOC and to the
28 contribution of very small particles (e.g. viruses) to particle backscattering signals used in the derivation of
29 POC and PSD products (Stramski et al., 2008; Dall'Olmo et al., 2009).

30 The major limitation of using satellite observations for investigation of the MCP is that they are

1 restricted to surface layers of the ocean. Nonetheless, there may be some regions and seasons where this
2 limitation can be partially overcome, for example, in upwelling regions or regions of deep seasonal
3 overturning, where deeper DOC-rich waters are mixed to the surface. Recent advances in sub-surface remote
4 sensing, by the addition of biogeochemical and optical sensors to profiling floats (such as those deployed in
5 the Argo array), provide a new opportunity to investigate distributions of CDOM and POC throughout the
6 water column, extending our knowledge of surface bio-optical distributions into the interior ocean and
7 connecting them with environmental gradients in nutrients, oxygen and pH (Johnson et al., 2009). Future
8 incorporation on the floats of novel sensors for rapid characterization of genetic material *in situ* would
9 significantly advance our ability to describe prokaryote diversity throughout the water column alongside a
10 better characterization of DOM.

11 Marine biogeochemical time-series data sets, such as the Hawaii Ocean Time-Series (HOT), the
12 Bermuda Atlantic Time-Series Study (BATS) and the Porcupine Abyssal Plain (PAP) Observatory are vital
13 to the study of inter-annual variability in the linkage between phyto- and bacterioplankton community
14 structure and the BP and MCP (Carlson et al., 2004; Bidigare et al., 2009). These single point data sets are
15 complemented with transect time series datasets such as the Atlantic Meridional Transect (Robinson et al.,
16 2006) and multi-decadal biological datasets such as those collected with the Continuous Plankton Recorder
17 (CPR; Hinder et al., 2012). Data from the CPR survey show the link between climate variability and the
18 dominant phytoplankton functional group (PFG) (Hays et al., 2005), and a global collation of sediment trap
19 data demonstrate the relationship between dominant PFG and POC export efficiency and transfer efficiency
20 (Henson et al., 2012). Inclusion of measurements of DOC quantity, quality and reactivity alongside
21 microbial community structure in these monitoring programmes would improve our understanding of
22 linkages between climate-derived changes in plankton community structure and oceanic storage of organic
23 carbon.

24 **6.2 Environmental context**

25 The sequestration of carbon in the ocean is indispensably linked to the cycling of nitrogen, phosphorus,
26 sulfur, and iron. Bacterial and archaeal activities contribute to the regeneration of N and P by consuming
27 DOC (White et al., 2012). Complex interactions between prokaryotes and eukaryotic microbes, such as
28 cooperation or competition for nutrients, exist in the marine environment. For example, Mitra et al. (2014)
29 have proposed a new paradigm where, in oligotrophic waters, the mixotrophic protists through production of
30 DOM effectively engage in “bacterial farming” to ensure ample provision of food. Thus, in different

1 biogeochemical environments, the MCP could be expected to operate with different efficiencies for carbon
2 storage.

3 An example of the changing efficiency of DOC-derived carbon sequestration is that which occurs along
4 an estuarine gradient. Due to high terrigenous input of nutrients and organic matter, estuarine ecosystems
5 usually experience intense heterotrophic respiration processes that rapidly consume dissolved oxygen,
6 potentially producing extensive hypoxic and anoxic zones in the water column. The lowered availability of
7 dissolved oxygen and the increased load of nutrients such as nitrate from river input prompt enhanced
8 anaerobic respiration processes. Thus, most of the nutrients may be consumed by anaerobically respiring
9 heterotrophic microorganisms instead of being utilized by phytoplankton for POC and DOC production (Fig.
10 6). Anthropogenic eutrophication in estuarine and coastal areas may thus reduce the efficiency of the MCP
11 (Dang and Jiao, 2014). This reduced efficiency may be exacerbated by the potential ‘priming’ effect of labile
12 organic matter addition stimulating the respiration of RDOC, as recently seen in soil environments (Wieder
13 et al., 2013).

14 **6.3 Bioassay and perturbation experiments**

15 In order to investigate mechanistic relationships between changing environmental parameters such as
16 temperature, OA and nutrients, and microbial organic carbon cycling, bioassay or manipulation experiments
17 are required. Mesocosm experiments have become the preferred approach for these manipulations due to
18 their ability to: a) study community dynamics of three or more trophic levels for a prolonged period of time,
19 b) measure the pools and fluxes of bio-active compounds and to perform mass balance estimates, and c)
20 study the interactions of ecosystem dynamics and biogeochemical processes under defined experimental
21 conditions (Riebesell et al., 2013).

22 Mesocosm experiments have been instrumental in observing the influence of OA on DOC
23 concentration (Schulz et al., 2008), TEP production (Engel et al., 2004a) and community respiration (Egge et
24 al., 2009) , the impact of nutrient supply on production, partitioning and the elemental composition of
25 dissolved and particulate material (DOM, POM), and the impact of increasing temperatures on the
26 accumulation and stoichiometry of DOM and POM (Wohlers-Zöllner et al., 2012), the coupling of
27 phytoplankton and bacterial processes (Hoppe et al., 2008) and the balance between autotrophic and
28 heterotrophic metabolism (Müren et al., 2005).

29 Most data on the effect of climate change on organic matter dynamics were obtained in perturbation

1 experiments studying the response to a single factor. Recent data highlights the need to study the interactions
2 between multiple drivers (e.g. temperature, nutrients, light, OA). For example, the contradictory responses
3 of phytoplankton TEP production to OA (Engel et al., 2004a; Schulz et al., 2008; Egge et al., 2009) indicates
4 that additional factors, such as total alkalinity (Mari, 2008; Passow, 2012) or nutrient stoichiometry (Corzo
5 et al., 2000; Staats et al., 2000; Passow, 2002; Beauvais et al., 2006) should be considered in future
6 experiments that investigate TEP aggregation (Passow and Carlson, 2012). Synergistic effects of increased
7 temperature and OA on microbial community composition (Lindh et al., 2012), and OA and increased
8 inorganic nutrients on bacterial production (Baltar et al., 2013) have also been found. It is therefore crucial
9 to move towards a multiple-factor approach in the design of mesocosm experiments to better constrain the
10 effect of multiple environmental drivers on the MCP and the BP. However, studying the impact of multiple
11 factors demands a more complex experimental design and statistical approach to distinguish between subtle
12 and interacting effects (Breitburg et al., 1998). Oceanographers would benefit from the experience of
13 multiple stressor studies undertaken in freshwater and terrestrial environments by ecotoxicologists to
14 develop hypotheses and concepts linking global, regional and local anthropogenic drivers and their
15 combined effects on ocean biota (Calow, 1989; Boyd and Hutchins, 2012).

16 *In situ* mesoscale addition experiments where a single water mass tagged with the inert tracer sulphur
17 hexafluoride along with a potentially limiting nutrient (e.g. iron) or nutrients (e.g. iron and phosphate) (Boyd
18 et al., 2007) could also be adapted for the study of the BP and MCP. The unequivocal lagrangian sampling
19 mode which this allows could be used to study the effect of changing DOC concentration and composition
20 associated with for example whale or zooplankton excretion, melting sea-ice, increased river flow or the
21 offshore movement of upwelled water (Loucaides et al., 2012).

22 **6.4 Improved chemical analytical and genomic approaches**

23 Lipid biomarkers and their carbon isotopes can be powerful tools for identification of the
24 microorganisms participating in POM and DOM cycling (White et al., 1979; Zhang et al., 2002), which may
25 also help link biogeochemical processes in the water column and sediments. The concentrations of
26 ester-linked phospholipid fatty acids (PLFA) and intact polar archaeal lipids (IPAL) indicate the biomasses of
27 extant bacteria and archaea, respectively, in complex ecosystems (White et al., 1979; Zhang et al., 2002; Sturt
28 et al., 2004; Lipp et al., 2008; Liu et al., 2011). Furthermore, certain lipids can be used as biomarkers because
29 they are characteristic of, or unique to, certain microbes. Such lipid biomarkers or their combinations can
30 reflect community structure, physiological and nutritional status, and the dynamic biogeochemical processes

1 carried out by the microbes (White et al., 1998; Suzumura, 2005). In addition, the carbon isotopes of lipid
2 biomarkers can be used as tracers for molecular level flow of carbon and thus serve to evaluate the
3 efficiency of the MCP and quantify its relationship with the BP because the products of BP-based organisms
4 may serve as the substrates of MCP-based organisms.

5 Radiocarbon is another powerful approach for quantification of MCP or BP activities in the ocean.
6 From POC to DIC to DOC, the ^{14}C values decrease sequentially. If the BP plays a dominant role in the
7 ocean, a more ^{14}C positive signature would be seen in the water column and sediment organic carbon; on the
8 other hand, if MCP dominates, the reworking of POC in the water column may shift organic carbon toward
9 older DOC with depleted ^{14}C (McNichol and Aluwihare, 2007). The ^{14}C of lipid biomarkers can help to
10 evaluate pathways of carbon metabolism by deep ocean microbes. For example, the ^{14}C values of glycerol
11 dialkyl glycerol tetraethers from deep sea ammonia-oxidizing archaea are closer to the ^{14}C value of DIC,
12 indicating that these organisms fix CO_2 in the deep ocean (Ingalls et al., 2006). Such an approach would
13 help evaluate the autotrophic versus heterotrophic capabilities of meso- and bathypelagic prokaryotes.
14 Studying the ^{14}C signature of DNA collected from mesopelagic Pacific waters, Hansman et al. (2009)
15 concluded that both DIC and fresh DOC (presumably released from sinking POC) are utilized, while
16 ambient DOC is not a major substrate.

17 What is more challenging is linking the taxonomic composition of microbial communities with their
18 possible functions in the carbon cycle. In particular, it is not clear if the biological formation of RDOC is
19 carried out by all microbes or by a subset of the microbial community. New sequencing technology has been
20 instrumental in the proliferation of genomic approaches for exploring potential metabolisms and
21 biogeochemical roles of microbes in the oceans. These approaches include metagenomic and
22 metatranscriptomic methods, which are based on the direct sequencing of genomes and transcripts from all
23 organisms in a particular size class (usually the one dominated by bacteria and archaea) without isolation or
24 separation of individual taxa (Moran, 2008; Kirchman, 2012). These methods give insights into the potential
25 function of microbes and have suggested new pathways, such as light-harvesting by proteorhodopsin (Béjà
26 et al., 2000) in the open oceans. Data from genomic sequencing of single cells isolated by flow cytometry
27 have suggested metabolisms, such as chemolithotrophy based on sulfur oxidation in mesopelagic waters
28 (Swan et al., 2011).

29 These “omic” approaches have provided insights into processes involving labile DOC (Poretsky et al.,
30 2010), but work is needed to get a complete picture of how microbes interact with all DOC components in

1 the oceans. Microbial oceanographers face several challenges using omic approaches to explore DOC use
2 and formation. One challenge is that analysis of sequence data heavily depends on databases of sequences
3 from laboratory-grown organisms with known metabolic functions. These laboratory-grown organisms may
4 not be representative of uncultivated oceanic microbes. Another problem is that not all of the genes in even
5 heavily-studied organisms, such as *Escherichia coli*, are completely known, and typically 10-20% of a
6 prokaryotic genome will not be similar to genes from any organism (Koonin and Wolf, 2008). Even when
7 identified by its similarity to known genes, more detailed enzymatic analysis may not show the predicted
8 function (Cottrell et al., 2005). Consequently, there are problems in using genomic information to examine
9 even known functions. It is even more difficult to use genomic data to gain insights into processes such as
10 RDOC formation when the basic biogeochemical mechanisms are unknown.

11 It is unlikely that a single gene or gene cluster will provide the answer to RDOC formation, just as few
12 diseases can be traced to defects in one or two genes. However, genomes represent the evolutionary record
13 of how microbes have interacted with DOC over millennia. If we can read that record, we are likely to learn
14 much about DOC-microbe interactions. Coupling omic studies with geochemical studies on all DOC
15 components is likely to give new insights into the MCP.

16 **6.5 Ecosystem modeling**

17 Biogeochemical models have been used to study DOC cycling and its interactions with heterotrophic
18 prokaryotes in the surface ocean, indicating that oligotrophic conditions can lead to the accumulation and
19 export of semi-labile DOC (SLDOC, a fraction of DOC, which resides mainly in the upper layer but
20 becomes labile when transported to deep water) (Polimene et al., 2006; Luo et al., 2010). To better
21 understand carbon sequestration, further modeling studies are required to quantify the relative importance of
22 the MCP and BP, identify processes which contribute the greatest uncertainties in their quantification, and
23 guide the priorities for future field and laboratory work.

24 The key processes to be represented in a MCP model are those that describe the production and
25 removal of RDOC in different vertical regimes. An initial step could be to set up a conceptual model
26 representing the ocean as simply 3 vertical boxes: the epipelagic (surface 200 m), mesopelagic (200 to 1,000
27 m) and bathypelagic zones (below 1,000 m) (**Fig. 7**). In the epipelagic zone, such a model would focus on
28 quantifying the production of SLDOC. The model of Flynn et al. (2008) provides a basis for such work,
29 describing the production of different types of DOM as a function of the growth and nutrient status of
30 primary producers. Similar models are needed for DOM production by zooplankton, and for the

1 consumption of all fractions of DOM by bacteria and other microbes. The proposal made by Mitra et al.
2 (2014) places an additional reason to develop these simulations, with mixotrophs simultaneously producing
3 DOM through phototrophy and phagotrophy, while consuming bacteria whose growth is supported by the
4 same DOM.

5 In the mesopelagic zone, the model would need to address the lability continuum of RDOC. A
6 mechanism would be needed to determine a flexible lability continuum according to the nutrient conditions
7 in the overlying epipelagic layer: the continuum would move toward recalcitrance in oligotrophic oceans
8 while being more labile in eutrophic regions (Jiao and Zheng, 2011). Such a model could also include a
9 description of processes in the mesopelagic layer allowing the modification of the accessibility of RDOC to
10 heterotrophic bacteria. This requires the development of a model which describes the ability of bacteria to
11 discriminate between different types of DOM with changes in stoichiometry (C:N:P), and which can
12 reflect changes in growth rates, growth efficiency and respiration. While such models exist (e.g., Flynn,
13 2005; Luo et al., 2010), other than for use in conceptualized scenarios, all these models remain largely
14 empirical because of the practical challenges surrounding the characterisation and quantification of different
15 DOM fractions.

16 Accepting that limitation, assuming that the DOC concentration in the bathypelagic zone does not
17 change substantially with depth (Hansell, 2013), the model can address the suggestion that RDOC is not
18 directly transported to local bathypelagic zones, but moves with the thermohaline circulation and is
19 degraded slowly within all 3 vertical layers. This initial model simulates CO₂ production as the result of
20 degrading RDOC in different vertical layers. Together with an estimation of deep water ventilation and
21 exchange rates of water masses between the 3 vertical boxes, it is expected to quantify the role of the MCP
22 in carbon sequestration on various time scales (Fig. 7).

23 Models of these processes within coastal ocean scenarios will help in evaluating anthropogenic impacts
24 on carbon sequestration by both the BP and the MCP. *In silico* experiments could be set up with different
25 rates and stoichiometries of riverine nutrient inputs. Carbon sequestration resulting from the BP in coastal
26 oceans also needs to consider the remineralization of POC in subsurface layers, as well as the resuspension
27 of benthic POC; however, while models may clearly differentiate between POC and DOC, in real world
28 sampling the so-called DOC fraction is contaminated by micro-particulates (i.e., POC) that compounds the
29 already challenging definition of labile, semi-labile and refractory fractions. Here, perhaps more than for
30 ocean systems, it is necessary to describe the variable stoichiometry of the DOM fractions. To quantify

1 carbon sequestration by the MCP, parameters which control both DOC lability and its interactions with
2 bacteria should be described in the model. To further quantify the fate of DOC after use by bacteria, variable
3 stoichiometry for bacterial cellular composition is required. The model ultimately needs to calculate the net
4 rate of supply of RDOC to the adjacent open ocean and thus give estimates for the net carbon sequestration
5 rate by the MCP.

6 These model experiments could be used to determine a relationship between carbon sequestration (from
7 the BP and the MCP) and the nutrient input from rivers, in order to estimate an optimal rate and
8 stoichiometry of riverine nutrient input for maximum carbon sequestration. Reverse modeling methods
9 (Friedrichs et al., 2007; Luo et al., 2010), may be applied to improve these estimates, and to unveil the
10 uncertainties associated with model processes and parameters.

11 Modeling of ocean ecosystems has relied heavily on the concept of functional groups, with more
12 complex models having multiple functional types within each modelled group. For example, the singular
13 phytoplankton box of early nutrient-phytoplankton-zooplankton-detritus (NPZD) models has been extended
14 to take account of size (e.g. pico, nano and micro size classes), biogeochemical function (e.g. silicifiers,
15 calcifiers, dimethylsulphide producers, Le Quere et al., 2005) or taxonomy (e.g. diatoms, dinoflagellates,
16 Blackford et al., 2004). While understanding of microbial heterotrophic communities in the ocean is
17 advancing rapidly, there is no clear differentiation of a set of bacterial and archaeal functional types. As with
18 other marine ecosystem modelling approaches, the resolution and descriptions used will vary according to
19 the question posed. For example, taxonomically it may be desirable to separate archaea from bacteria,
20 whereas in the context of understanding biogeochemical transformations of POC and RDOC it may be more
21 opportune to model the affinities of organisms to different substrates (particles, gels or DOC) or their
22 evolutionary strategies for responding to different resource environments. In this regard, partitioning into
23 copiotrophs that use chemotaxis, motility and fast uptake kinetics to exploit microscale gradients of high
24 nutrient concentration, versus non-motile oligotrophs that are adapted to life in nutrient poor environments
25 (Stocker, 2012) may provide a good starting point.

26 Modeling studies would eventually need to extend to the global scale, aiming to reproduce the
27 distribution of DOC and predict how the MCP and BP will change with changing anthropogenic influences.
28 Overall, two-way interaction should be maintained between the MCP-BP modeling and observational
29 scientific communities: the results from experiments will help parameterize the model, and model results
30 will then guide further experimentation. First and foremost, however, is the need to rationalize the chemical,

1 biological and modelling descriptions of different types of DOM.

2 **6.6 Strategies to enhance carbon sequestration**

3 Based on the discussion above (5.2, Fig. 4) and a statistical study which shows that concentrations of
4 organic carbon and nitrate in natural environments ranging from soils and rivers, to coastal and oceanic
5 waters are inversely correlated (Taylor and Townsend, 2010), it seems that if we want to store more organic
6 carbon in the environment we must avoid high concentrations of nutrients. Therefore, reduced nutrient levels
7 in otherwise eutrophic coastal waters could result in a greater proportion of the fixed carbon becoming
8 RDOC (or RDOC_t) (Jiao et al., 2010b). Then we may be able to enhance carbon sequestration in the coastal
9 zone by controlling the discharge of nutrients from land. This can be achieved through the integrative
10 management of the land-ocean system, for example by using methods of fertilization which avoid loss of
11 nutrients to rivers and reducing sewage discharge to coastal waters. Such an integrative management
12 approach would have the additional advantage of reducing eutrophication. Furthermore, the carbon storage
13 capacity of coastal regions could be enhanced by optimizing both the BP and the MCP (Fig. 4). If the key to
14 the efficiency of carbon storage in an ecosystem is the nutrient status, then it is critical to perform
15 field-based research to fully understand the tipping point of the nutrient concentration or stoichiometry that
16 leads to different types of carbon metabolism. An “optimal nutrient concentration/stoichiometry” could be
17 obtained for a given ecosystem through theoretical calculation and field experimentation.

19 **7. Summary**

20 This synthesis of current research and gaps in our knowledge leads to the following conclusions and
21 suggestions for future research:

22 RDOC can be classified as RDOC_t and RDOC_c depending on the composition and concentration of the
23 RDOC molecules as well as the prevailing environmental conditions. State-of-the-art analytical chemical
24 and genomic methods should be used to determine the microbial source and composition of RDOC and
25 assess the environmental conditions which influence the recalcitrance of RDOC.

26 Analyses of biomarkers and isotopic records show intensive MCP processes in the Proterozoic oceans
27 when the MCP could have played a more significant role in regulating climate. Understanding MCP
28 dynamics in the past will aid in predicting how carbon storage could change with changing climatic
29 conditions in the future.

1 Future research programs should integrate the study of POC flux and DOC transformation in order to
2 elucidate the interactions between POC and DOC cycling and the environmental controls on these
3 interactions. This includes an investigation of the occurrence and lability of RDOC-coated POC.

4 Bioassay and field experiments should be undertaken to assess the combined effects of multiple
5 environmental drivers (temperature, OA, nutrient supply) on marine carbon storage. Ecosystem models need
6 to be developed and tested with mechanistic relationships derived from these experiments in order to predict
7 the dynamics of and interactions between the BP and the MCP under global change scenarios including
8 changing mixing and circulation, changing nutrient and oxygen distributions, and increasing temperature
9 and decreasing pH.

10 We hypothesize that increased nutrient supply to an ecosystem above a tipping point will decrease the
11 efficiency of carbon storage by the MCP and in the long term by the BP as well. If supported, this could
12 have important implications for management strategies for carbon sequestration and coastal ecosystem
13 health including reduced eutrophication and hypoxia.

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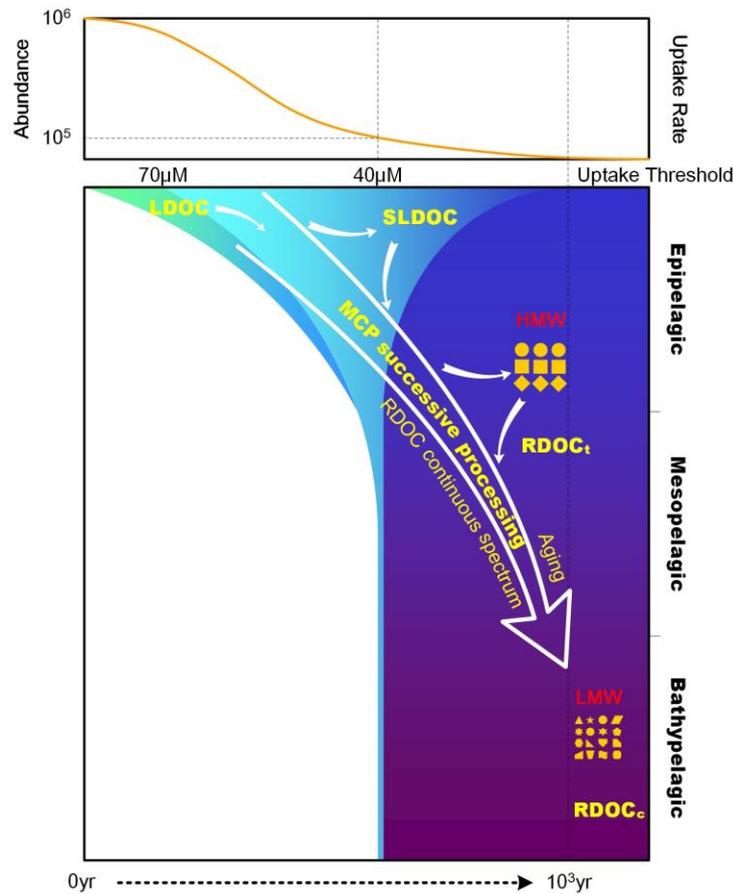
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 3 **Fig. 1. Linking RDOC at multiple dimensions: Temporal (age) and spatial (depth) transformations**
 4 **of RDOC. Lower panel:** Successive microbial processing of organic carbon results in the generation
 5 of RDOC of different recalcitrance and different potential residence time: MCP --- microbial carbon
 6 pump; RDOC_t --- RDOC compounds that are resistant to microbial consumption in certain environments,
 7 but subject to further cleaving and decomposition when the situation changes. RDOC_c --- composed of
 8 diverse small molecules which are inaccessible to microbial uptake due to their low concentration. **Upper**
 9 **panel:** Microbial response (in terms of abundance or uptake rate) to DOC availability as a reference to
 10 conceptualize the microbial uptake threshold for RDOC_c, microbial abundance corresponding to DOC
 11 concentrations of 40 μM in the deep and 70 μM in the surface oceans. LDOC --- labile DOC, a fraction of
 12 DOC, which is immediately accessible to microbial utilization. SLDOC --- semi-labile DOC, a fraction
 13 of DOC, which resides mainly in the upper layer but which becomes labile when transported to deep
 14 water.

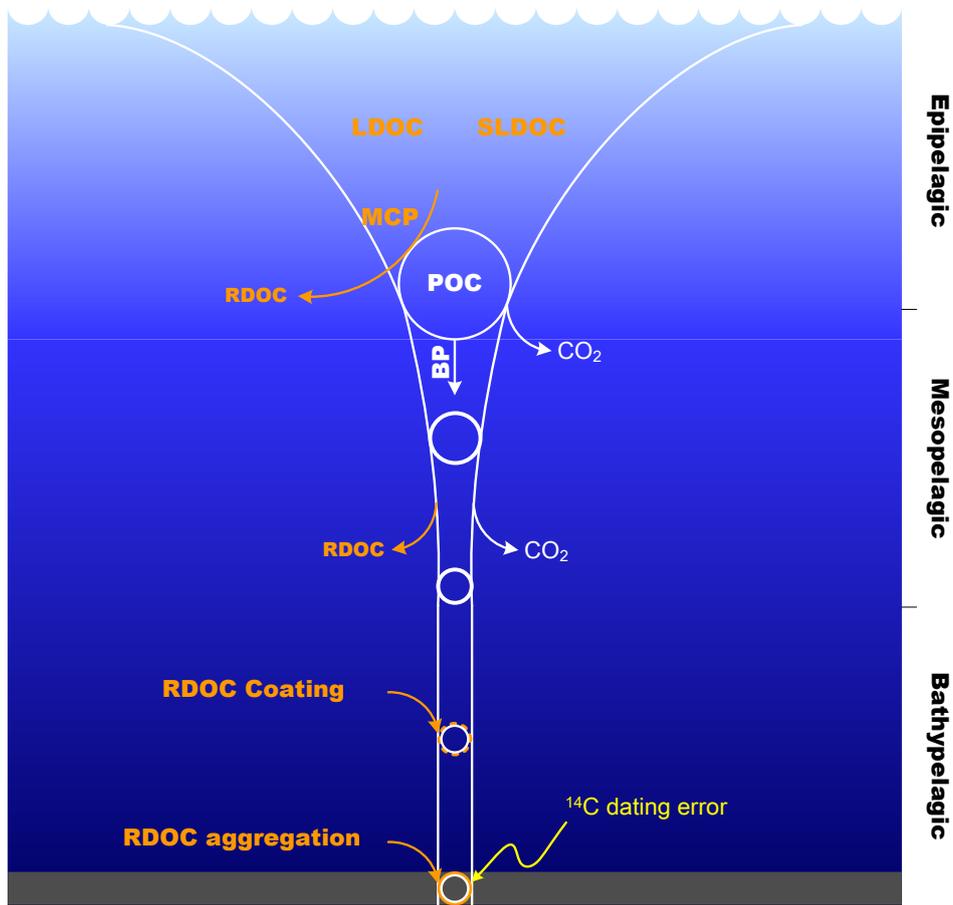
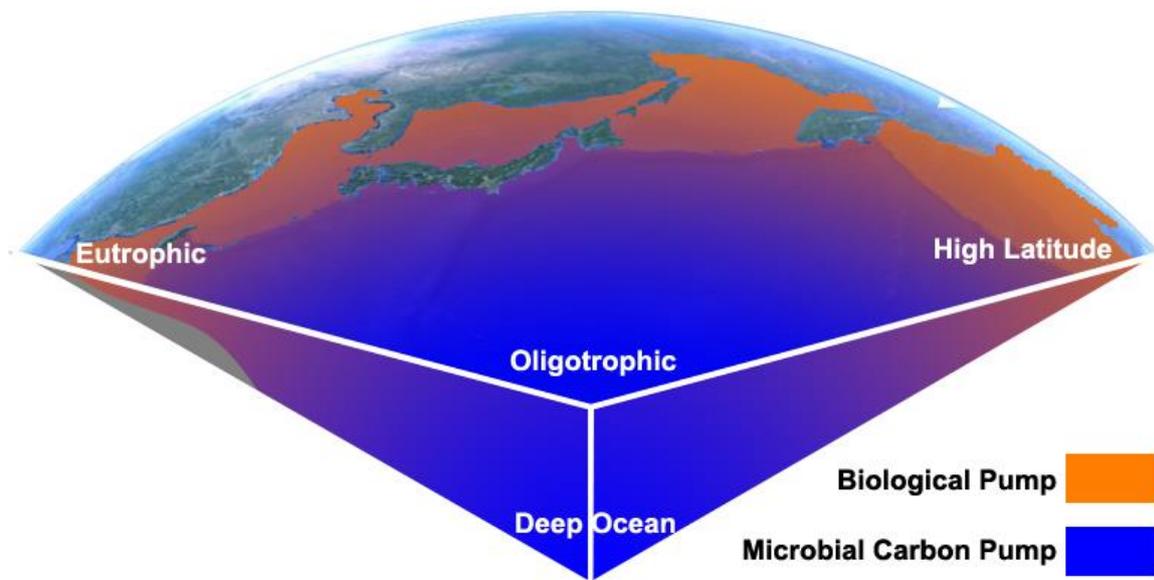


Fig. 2. Transformation of DOC and POC through decomposition and scavenging processes that could influence carbon sequestration and ¹⁴C dating (see text for details). MCP --- microbial carbon pump, BP --- biological pump; POC --- particulate organic carbon, DOC --- dissolved organic carbon, LDOC --- labile DOC, SLDOC --- semi-labile DOC, RDOC --- refractory DOC, RDOC coating --- the process of RDOC attaching to the exterior of or being incorporated into a particle, RDOC aggregation --- the process of RDOC molecules accumulating and combining

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3 **Fig. 3.** A demonstration of trends in the relative dominance of the BP and the MCP along environmental
4 gradients.

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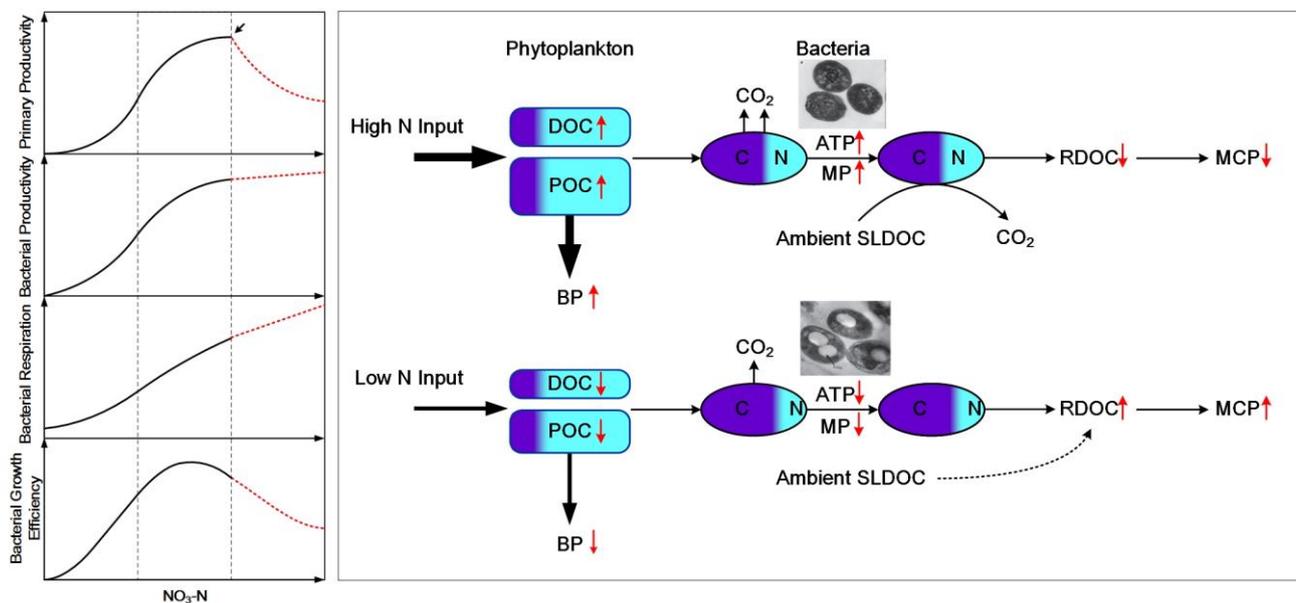
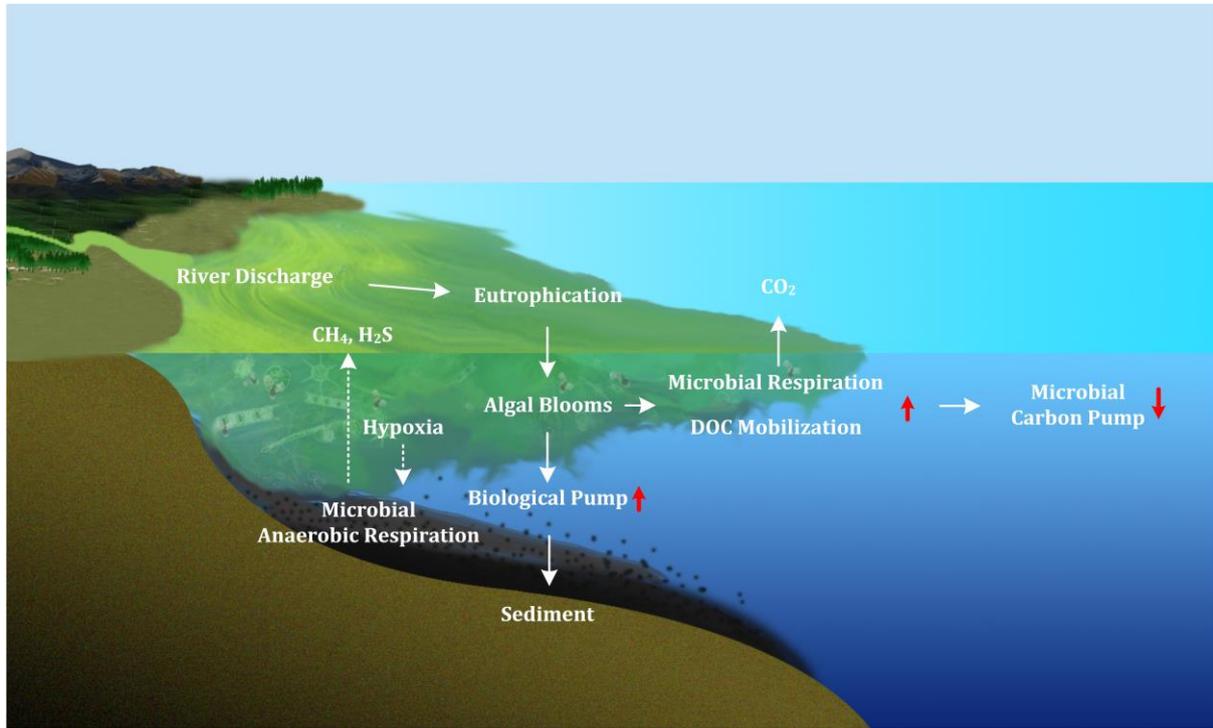


Fig. 4 The impact of nutrient supply on the sequestration of carbon via the BP and the MCP

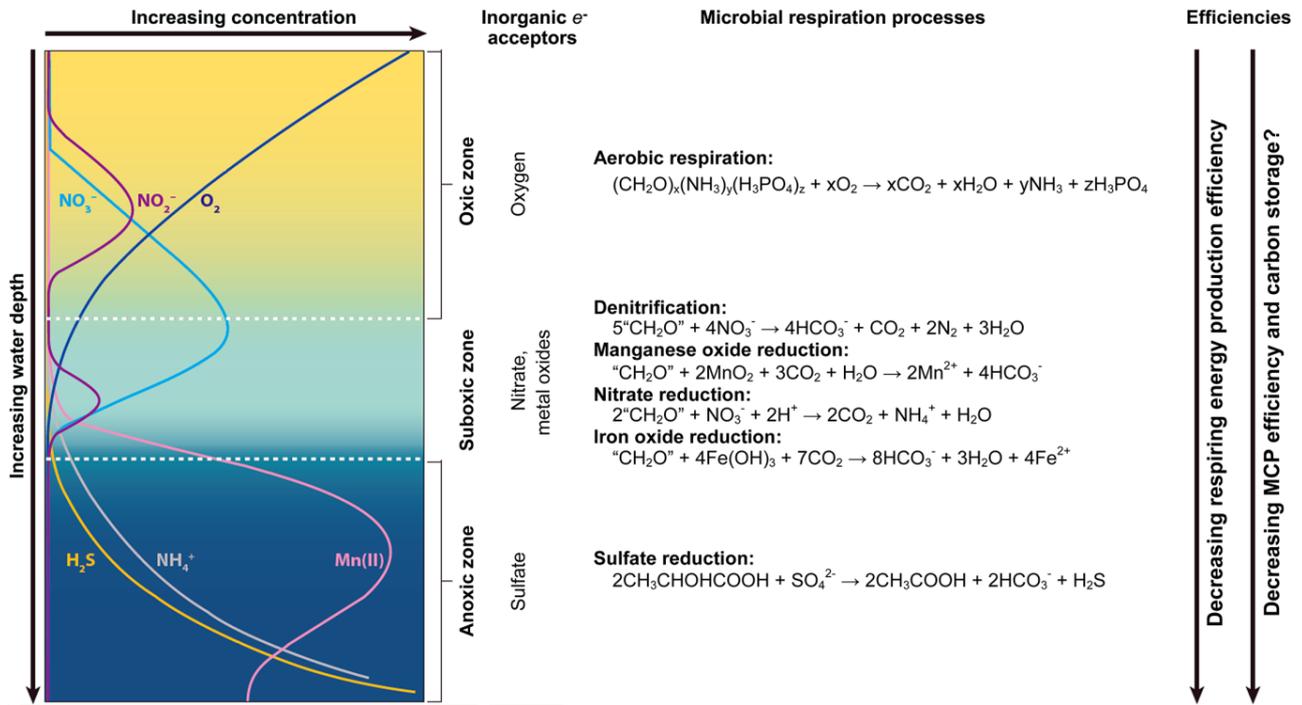
Left panel: Primary production, bacterial respiration and bacterial growth efficiency as functions of nutrients. The arrow in the top graph shows a tipping point of nutrient concentration beyond which primary production could drop down due to constraints of limiting factors other than nutrients, such as light availability and environmental carrying capacity. The red lines after the tipping point emphasize the differences between phytoplankton and bacteria in their response to high nutrients. The appropriate nutrient concentrations for a healthy ecosystem would range between the vertical dash lines where the ecosystem could remain sustainable while running at a high level of biological efficiency.

Right panel: Responses of the BP and the MCP to nutrient inputs. (MP – membrane potential; ATP – adenosine triphosphate). With high nutrient input, although the BP could be enhanced, the MCP could be reduced, because microbial respiration can also be stimulated by nutrients. Meanwhile ambient semi-labile DOC could be remobilized for microbial utilization, especially with the priming effects of the labile DOC generated by enhanced primary production. In contrast, if nutrient input is appropriate, the BP could remain moderate, semi-labile DOC could remain persistent, the MCP could be enhanced, and the storage capacity of the combined BP and MCP could be maximized.



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Fig. 5. Hypothesized consequences of excess nutrients in coastal waters. Excess nutrients from river discharge cause eutrophication, harmful algal blooms and hypoxia, which in turn influence overall carbon sequestration efficiency.



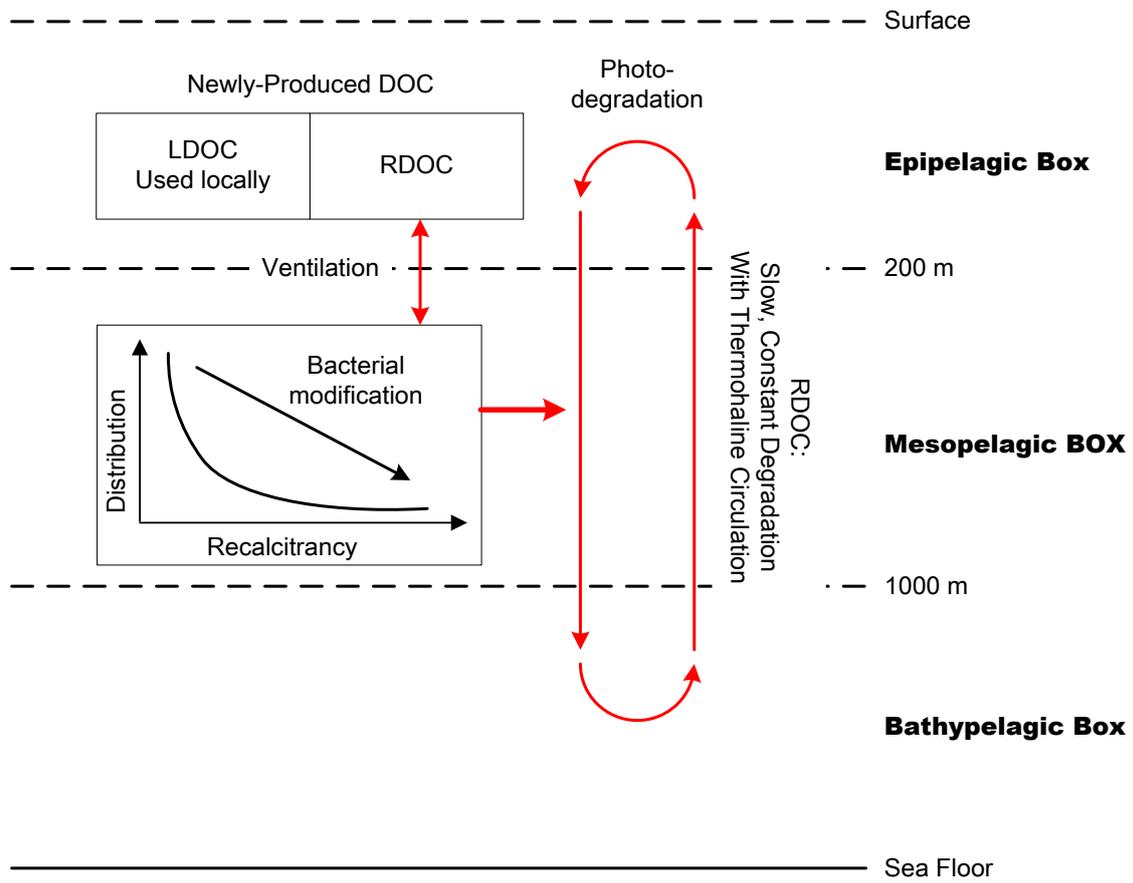
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3 **Fig. 6** The influence of microbial respiration processes on the efficiency of the MCP. Left panel after
 4 Moore et al., 2009.

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2 **Fig. 7. A conceptual framework for ecosystem modeling.**

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