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Soil redistribution and weathering controlling the fate of geochemical and physical carbon
stabilization mechanisms in soils of an eroding landscape

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Abstract

The role of eroding landscapes in organic carbon stabilization operating as C sinks or sources have been frequently discussed, but the underlying mechanisms are not fully understood. Our

analysis aims to clarify the effects of soil redistribution on physical and biogeochemical soil organic carbon (SOC) stabilization mechanisms along a hillslope transect. The observed mineralogical differences seem partly responsible for the effectiveness of geochemical and physical SOC stabilization mechanisms as the mineral environment along the transect is highly variable and dynamic. The abundance of primary and secondary minerals and the weathering status of the investigated soils differ drastically along this transect. Extractable iron and aluminum components are largely abundant in aggregates, but show no strong correlation to SOC, indicating their importance for aggregate stability but not for SOC retention. We further show that pyrophosphate extractable soil components, especially manganese, play a role in stabilizing SOC within non-aggregated mineral fractions. The abundance of microbial residues and measured ^{14}C ages for aggregated and non-aggregated SOC fractions demonstrate the importance of the combined effect of geochemical and physical protection to stabilize SOC after burial at the depositional site. Mineral alteration and the breakdown of aggregates limit the protection of C by minerals and within aggregates temporally. The ^{14}C ages of buried soil indicate that C in aggregated fractions seem to be preserved more efficiently while C in non-aggregated fractions is released, allowing a re-sequestration of younger C with this fraction. Old ^{14}C ages and at the same time high contents of microbial residues in aggregates suggest that microorganisms either feed on old carbon to build up microbial biomass, or that these environments consisting of considerable amounts of old C are proper habitats for microorganisms and preserve their residues. Due to continuous soil weathering and, hence, weakening of protection mechanisms, a potential C sink through soil burial is finally temporally limited.

1. Introduction

Soil organic carbon (SOC) is one of the most important terrestrial C pools. Carbon in soils can be protected against decomposition by three key mechanisms: (i) inherent biochemical recalcitrance; (ii) organo-mineral associations; by interaction of organic molecules with mineral surfaces, and (iii) physical protection; making the SOC inaccessible to degraders/consumers within soil aggregates (Sollins et al., 1996; Six et al., 2002). These mechanisms are interactive (e.g. aggregation of organo-mineral associations could already include biochemically recalcitrant SOC) and their contribution to SOC stabilization is strongly influenced by soil environmental conditions and landform (Salomé et al., 2010; Berhe et al., 2012; Dungait et al., 2012; Wang et al., 2014a). Distinguishing the role of a single mechanism for stabilizing C is, hence, a difficult task and a matter of ongoing debate and research (Berhe et al., 2012). For example, some authors argue that the biochemical recalcitrance of organic molecules does not exist *per se* and has always to be seen in an environmental context (Kleber et al., 2010), while others argue that “*recalcitrant SOM can be defined by intrinsic molecular properties, but these properties may be fairly irrelevant under specific environmental conditions*” (von Luetzow and Koegel-Knabner, 2010).

Studying SOC dynamics in landscapes with lateral soil fluxes requires the combined study of geomorphological, climatic, biogeochemical, and microbial parameters (Park et al., 2014). In most studies on SOC dynamics, the landscapes in which C exchange takes place are stable surfaces with no or only little lateral fluxes of soil or C. Hence, SOC fluxes in complex pool models are limited to a vertical exchange between soil and atmosphere. Connecting the different controls on SOC dynamics across soil depths and topographic positions has only recently received attention in landscape scale studies on SOC dynamics (Berhe et al., 2012; Doetterl et al., 2012). Bringing the detailed knowledge that we possess about SOC dynamics at plot and micro-scales into perspective at the landscape scale remains largely neglected.

1 Recently, soil redistribution in cropland and grassland systems has been shown to lead to
2 distinct qualitative and quantitative SOC modifications along geomorphic gradients compared
3 to soils in stable landforms (Yoo et al., 2006; Dlugoss et al., 2011; Berhe et al., 2012). Soils at
4 eroding sites are usually C depleted while soils in depositional settings can store more SOC
5 due to burial of topsoil with eroded sediment, potentially storing C for centuries (Van Oost et
6 al., 2012; Hoffmann et al., 2013; Johnson 2014). The removal of weathered topsoil material
7 from eroding positions, the replacement of eroded SOC and its burial at depositional sites can
8 potentially lead to a net sink for atmospheric C depending on the fate of the eroded SOC
9 (Harden et al., 1999; Doetterl et al., 2012; Wiaux et al., 2014a). SOC at the depositional site is
10 often regarded to be more stable with longer turnover times, depending on microbial activity,
11 environmental conditions (Wang et al., 2013) and biogeochemical characteristics of the
12 transported C fractions. However, areas (or landscapes) with a fast burial can lead to the
13 accumulation (storage) of labile SOC which is still vulnerable to decomposition if the
14 conditions at the site of burial change (Wiaux et al., 2014b). Thus, there is an ongoing
15 discussion about depositional sites of highly dynamic landscapes as C sink or source.

16 Soil properties with relation to C stability, such as aggregate stability, availability of reactive
17 mineral surfaces or soil water saturation, differ strongly between stable and dynamic
18 landscapes, i.e. landscapes where high rates of soil redistribution take place. This may have
19 consequences for SOC sequestration and stabilization in soils and requires the use of proxies
20 to investigate the stability of specific fractions. For example, the abundance of amino sugars
21 (AS) and the ¹⁴C derived age of a fraction can indicate the potential of a specific mechanism
22 to protect SOC against further degradation. The large majority (>99%) of AS found in soils
23 are considered to be of microbial origin and constitute an important building block in cell
24 walls and extracellular polysaccharides (Glaser, 2004; 2005; Simpson, 2004). In soils, only
25 glucosamine (GluN), galactosamine (GalN), and muramic acid are found in quantifiable

1 concentrations. Amino sugars provide a generally readily available energy source for
2 microorganisms and are easy to decompose compared to more recalcitrant organic matter
3 such as lignin or lipids (Amelung et al., 2001, 2008; von Luetzow et al., 2006; Roberts et al.,
4 2007; Schmidt et al., 2011). Amino sugar turnover is, therefore, fast compared to other
5 stabilized SOC fractions. Turnover times of AS, however, are strongly affected by
6 interactions with the soil mineral matrix and the soil environment (Bodé, 2013) and this can
7 lead to an accumulation of AS after cell death in soils (Guggenberger et al., 1999; Glaser et
8 al., 2004). This fact can be used to investigate the effectiveness of different stabilization
9 mechanisms. During the transport of sediment to and accumulation and burial at the site of
10 deposition, easily available SOC fractions have been decomposed. However, at the
11 depositional site, subsoil C is at least partly derived from buried topsoil C. Hence, at the
12 depositional site changes in the abundance of AS and the ^{14}C derived age of a fraction can
13 indicate the potential of a specific mechanism to protect SOC against further degradation
14 (Wang et al., 2014b).

15 Our work is a conceptual approach to analyze and illustrate the mechanisms of soil
16 redistribution effects on C dynamics. For our study, we hypothesize that the soil redistribution
17 history of a soil profile influences the present weathering status of the soil material, and is
18 therefore a primary control on the abundance and composition of the reactive soil mineral
19 phase, stabilizing SOC through geochemical (association of organic molecules with minerals)
20 and physical (aggregation) mechanisms. To complement our analysis on the importance of
21 different mechanisms to stabilize C along a geomorphic transect, we measured the ^{14}C and the
22 abundance of microbial residues in the form of AS in the different soil fractions. We further
23 hypothesize that with ongoing soil redistribution, the reactive soil mineral phase will be
24 highly variable, i.e. horizontally in space along the slope transect, vertically with soil depth,
25 and in time due to removal or burial of soil. We use the total reserve in base cations (TRB in

cmol_c kg⁻¹) to characterize the weathering status of the soils along the geomorphic transect and complement this with a qualitative and quantitative analysis of clay minerals. If changes in the mineralogical composition of the investigated soils due to soil redistribution act at a faster pace than weathering related changes, distinct mineralogical differences between profiles along the slope should be identifiable.

2. Material and methods

2.1 Study site and sampling

We performed our analysis on nine bulk soil samples and the dominating soil C fractions that were collected in the study of Doetterl et al. (2012) (Table 1) from a geomorphic transect on cropland situated in central Belgium (50°45'41.50''N; 4°44'07.36''E) with a south/southeast facing slope. The study area is characterized by a smooth rolling topography with plateaus, slopes (up to 20 percent slope) and dry valley bottoms. The climate of the region is a temperate oceanic climate with mild winters and cool summers (Köppen climate Cfb, 821 mm yr⁻¹; 9.7°C on 30-year average) (IRM, 2011). The geological substrate is a several meters thick Pleistocene aeolian deposit of calcareous loess, in which Luvisols have developed and are overlaying Tertiary sands (Wouters and Vandenberghe, 1994). Soils are well drained and show no evidence of long lasting hydromorphic conditions. First traces of agriculture in the region date back to the Late Bronze/Iron age and both sites have been under continuous agricultural land use since at least 1770 (Rommens et al., 2005; Lannoo, 2009). No detailed information about the long-term (i.e. several centuries) crop rotation or SOC input is available. However, this study focuses on the relative differences between geomorphic positions which were managed in the same way. Soil samples were taken from a stable, non-eroding profile at the hilltop plateau, an eroding profile from the hillslope shoulder (200cm soil loss) and a depositional profile in the colluvial valley bottom (350cm soil gain). In order

to quantify the vertical and horizontal distribution of SOC fractions, the soil cores were cut into the following depth intervals: the topsoil (0-15cm), the shallow subsoil (35-50cm), and the deeper subsoil (55-70cm).

2.2 SOC fractionation

The SOC content was measured in duplicate on 1g grinded soil subsamples using a VarioMax CN dry combustion Analyzer (Elementar GmbH, Germany) with a measuring range of 0.2-400mg C per g soil (absolute C in sample) and a reproducibility of <0.5% (relative deviation) on powdered soil samples. To derive functional SOC fractions, Doetterl et al. (2012) used a method based on the conceptual SOC fraction model proposed by Six et al. (1998; 2002) (Fig. 1). For details on the gathered fractions see the original study Doetterl et al. (2012) and the concepts of Six et al. (1998; 2002). In short, the scheme consists of a series of chemical and physical fractionation techniques applied to isolate functional SOC fractions, differentiated by stabilization mechanisms (chemical, biochemical and physical) which can also be associated with different (potential) turnover times (see also von Luetzow et al., 2006). Background information to the key soil and geomorphological properties as well as the abundance and composition of identified SOC fractions as described by Doetterl et al. (2012) and Wang et al. (2014b) for the investigated soils can be found in table 1. For our analysis on the microbial composition and ¹⁴C age of the fractions we analyzed the environment in which C is stabilized, hence analyzing macro- and microaggregates as a whole. For the mineralogical analysis we are focusing on comparable fractions for the mineralogical analysis, hence comparing aggregated and non-aggregated silt and clay fractions.

2.3 Soil physico-chemical characterization

Soil pH values were determined as the mean of two measurements per sample after a response time of 30 min and 24 hours respectively in a 1 : 2.5 soil/solution ratio in 25 ml 0.01 M CaCl₂

1 potentiometrically with a glass electrode using a portable multiparameter Meter HI9828
2 (Hanna Instruments US Inc., USA). All samples have shown no reaction when treated with
3 10% HCl and are considered free of carbonates.

4 The soil weathering degree in the different soil depth layers was estimated by measuring the
5 Total Reserve in Bases (TRB, the sum of total content in Ca, Na, K, Mg, in $\text{cmol}_c\text{kg}^{-1}$)
6 following Herbillon et al. (1986). The TRB can be used to compare soil horizons relative to
7 the parent material to evaluate the weathering degree of soil material by assessing the relative
8 loss of Ca, Na, K and Mg cations during weathering. Total elemental content was determined
9 by Inductively Coupled Plasma-Atomic Emission Spectrometry (ICP-AES) on the bulk soil of
10 the three profiles and three depths and the parent loess material after borate fusion (Chao and
11 Sanzalone, 1992).

12 A three-step sequential extraction scheme of pedogenic organo-mineral associations and oxy-
13 hydroxides (Stucki et al., 1988) was carried out in duplicate in the following order: sodium-
14 pyrophosphate at pH 10 (Bascomb, 1968), ammonium oxalate-oxalic acid at pH 3 (Dahlgren,
15 1994) and dithionite-citrate-bicarbonate (DCB) at pH 8 (Mehra and Jackson, 1960). This
16 allows assessing the amount of Mn, Mg, Fe and Al-bearing phases in the different fractions
17 and their correlation with organic C in the different SOC fractions. The specific extraction
18 was performed on the bulk soil of each sampling position, the (micro)aggregated (s+cm) and
19 non-aggregated (s+c) silt and clay associated SOC fraction separately. These latter mentioned
20 fractions were identified as the two key fractions to change in their abundance along the
21 hillslope between eroding and depositional (sub-)soils (Doetterl et al., 2012) and are the
22 building blocks for larger aggregates. Each extract was analyzed for its Al, Fe, Mg and Mn
23 content by ICP-AES.

24 In our sequential extraction, pyrophosphate (p) extractable components are interpreted as
25 predominantly organically complexed metals. Oxalate (o) extractable components reflect the
26 amorphous secondary Fe and Mn oxides and poorly crystalline aluminosilicates (imogolite-

type materials, ITM). Dithionite (d) extractable components included predominantly crystalline oxy-hydroxides of Mn, Mg, Fe and Al. Several authors could show that pyrophosphate extractable Al may not be attributable only to Al bound to organo-metallic complexes since the alkaline extractant could also extract Al from Al hydroxide phases and from poorly crystalline aluminosilicates (i.e. Schuppli et al., 1983; Kaiser and Zech, 1996). The results of the pyrophosphate extraction must, therefore, be treated with caution due to uncertainty about the origin of the extracted minerals. Hence, we limit our analysis to interpreting the abundance and spatial patterns of the various extractable components in the bulk soil and the SOC fractions and discuss the pedological implications of the observed carbon/mineral correlations.

2.4 Soil mineralogy

The mineralogy of the clay-sized fraction was determined by X-ray diffraction (XRD, Cu K α , D8, Bruker Advance) after K⁺ and Mg²⁺ saturation, ethylene glycol solvation and thermal treatments at 300 and 550 °C (Robert and Tessier, 1974). Clay minerals were classified and peak-identified according to Brindley and Brown (1980). Quantitative analyses were performed using the Rietveld method through the software package Siroquant V4.0. Refinement of Rietveld parameters was carried out following the instructions provided in the “Siroquant V4.0 Technical and Clay Manuals” until obtaining a Chi-Square value < 3. Key primary minerals that can interfere with the spectra (quartz) and act as sources for clay formation such as amphiboles (hornblende), feldspars (albite, orthoclase, plagioclase), mica (muscovite, phengite, biotite) as well as olivine and pyroxene have been quantified in order to analyze their relative abundance in relation to the secondary aluminosilicates. For the latter, we focused on the identification of expandable 2:1 layered minerals such as smectite and vermiculite, the non-expandable 2:1 layered illite and chlorite and the non-expandable 1:1

layered kaolinite. Note that no distinction has been made between primary and secondary chlorite. In a final step, the quantified primary and secondary minerals were then compared along the geomorphic gradient, i.e. soil layers from the same depth across different positions or soil layers from different depths but the same profile, to find relevant trends.

2.5 Compound specific analysis of amino sugars

Amino sugar concentrations were determined using liquid chromatography for the bulk soil, macroaggregates ($>250\text{ }\mu\text{m}$), microaggregates ($53\text{--}250\text{ }\mu\text{m}$) as well as for the non-aggregated silt and clay fractions ($<53\text{ }\mu\text{m}$). Each sample was extracted in duplicate. Amino sugar extraction and analysis was based on the procedure described by Bodé et al. (2009). Shortly, samples corresponding to 0.3 mg of N were successively hydrolysed with 6M HCl (20 ml per gram of sample) at $105\text{ }^{\circ}\text{C}$ for 8 hours after which AS were purified on a cationic exchange resin (AG50W-X8, 100-200 Mesh, Hydrogen form, Bio-Rad lab.) and eluted by protons prior to LC-IRMS analysis. The chromatographic separation was performed using a LC pump (Surveyor MS-Pump Plus, Thermo Scientific, Bremen, Germany) mounted with a PA20 CarboPac analytical anion-exchange column ($3\times 150\text{mm}$, $6.5\mu\text{m}$) and a PA20 guard column (Thermo Scientific, Bremen, Germany). Basic AS (GluN and GalN) were eluted with 2mM NaOH and a column temperature of 15°C . An analysis of the repeatability between replicates shows deviations of ca. 11% between the replicates for GalN with contents of $>100\text{ }\mu\text{g GalN}$ per g soil. For smaller values, deviations were higher (ca. 31%). For GluN with contents of $>250\text{ }\mu\text{g GluN}$ per g soil deviations of c. 12% were observed, whereas smaller values differed about 40% between replicates. Hence, we limit our analysis of this data to observations that exceed the latter mentioned uncertainty and to a description of trends within the dataset.

2.6 Isotopic composition and ¹⁴C age

Using ¹⁴C radiocarbon dating we estimate the age of the organic material associated with the macroaggregates, microaggregates and the non-aggregated silt and clay fractions at the depositional site in order to see relative differences between different fractions and burial depths. We use these relative differences to evaluate the effectiveness of different fractions in stabilizing C against decomposition after burial. In short, roughly 1 mg C was sealed into an evacuated Pyrex tube and reduced to graphite (Xu et al., 2007). Sample preparation backgrounds have been subtracted, based on measurements of ¹⁴C-free coal. The radiocarbon signature of the graphite was measured with accelerator mass spectrometry (NEC 0.5MV 1.5SDH-2 AMS system) at the Keck-Carbon Cycle AMS facility at UC Irvine (CA, USA). Results have been corrected for isotopic fractionation according to the conventions of Stuiver and Polach (1977), with $\delta^{13}\text{C}$ values measured on the prepared graphite using the AMS spectrometer. Radiocarbon concentrations are given as fractions of a modern oxalic acid standard and conventional radiocarbon age following the conventions of Stuiver and Polach (1977).

2.7 Implementation and statistical analyses

All statistical analysis was realized using SAS 9.3 (SAS Institute Inc., Cary, NC) and R 2.11.1 (R Development Core Team, 2010). Differences between the means of classes have been performed using multi-group ANOVA Bonferroni corrections and Tamhane's T2. We also performed linear regressions to test correlations between the abundance of the reactive soil phases and SOC in the bulk soil and for the isolated fractions. All statistical tests were evaluated using $p < 0.1$ as the level of significance.

3. Results

3.1 Total reserve in bases and degree of soil weathering along the hillslope

The TRB for the loess parent material was $139 \text{ cmol}_c \text{ kg}^{-1}$. For the different soil layers, TRB ranged between 101 and $118 \text{ cmol}_c \text{ kg}^{-1}$, which represents a TRB loss of 15 to 27%. These values are in the same range as reported for other Luvisol profiles in the region (Brahya et al., 2001). Losses in TRB are mostly related to decalcification (Ca losses compared to parent material: $80 \pm 2\%$) (Table 2). The strongest losses of cations (lowest TRB in Table 2), and hence the most advanced weathering stage, are observed for the deposited soils at the foothill (relative difference of TRB at the depositional site compared to the loess parent material: 25 - 27%), while the layers at the eroding hillslope have been facing the lowest losses of cations (highest TRB in Table 2), and hence the weakest degree of weathering (relative difference of TRB at the depositional site compared to the loess parent material 15 -20%). Nevertheless, these differences are small compared to differences between soil types in the same region (Brahya et al., 2001) and indicate a rather small effect of soil redistribution on TRB in comparison. At the stable plateau and the eroding hillslope, a significant difference ($p < 0.1$) with higher TRB at greater depth can be observed, but with weaker increases of TRB with soil depth than reported from similar soils (Brahya et al., 2001). In contrast, no effect of depth on TRB was observed in the depositional profile.

3.2 Reactive soil phases

Pyrophosphate extractable components are largely abundant in microaggregates, especially as Fe(p) ($1118 \pm 293 \text{ mg kg}^{-1} \text{ soil}$) and Al(p) ($1647 \pm 385 \text{ mg kg}^{-1} \text{ soil}$) (Fig. 2). The ratio of non-aggregated to aggregated pyrophosphate extracts in silt and clay is generally between 1:3 – 1:7 and shows a clear association of these components with soil microaggregates. In contrast, oxalate and DCB extractable phases do not show such a pattern between aggregated and non-aggregated soil samples. No significant differences in the amount of oxalate and DCB

1 extractable components in aggregated vs. non-aggregated soil fractions were detected for Fe,
2 Mg and Mn. Al(o) showed about 40% lower concentrations in non-aggregated silt and clay
3 (s+c) compared to aggregated silt and clay (s+cm). Approximately the same amount of
4 oxalate extractable Al, Fe, Mg and Mn is present in aggregated ($1690 \pm 378 \text{ mg kg}^{-1}$ soil) and
5 non-aggregated ($2421 \pm 648 \text{ mg kg}^{-1}$ soil) soil samples with ca. five times more DCB
6 extractable elements than oxalate in both soil compartments.

7 Despite the small contribution of Mn(p) ($40 \pm 36 \text{ mg kg}^{-1}$ soil) to the total pyrophosphate
8 extractable phase of the bulk soil compared to Al(p) ($122 \pm 64 \text{ mg kg}^{-1}$ soil) and Fe(p)
9 ($222 \pm 115 \text{ mg kg}^{-1}$ soil) and Mg(p) ($101 \pm 33 \text{ mg kg}^{-1}$ soil), it is the only phase with a
10 significant correlation to SOC in the bulk soil across the whole slope transect (Fig. 3). Table 3
11 shows that the positive correlation of Mn(p) with SOC along the slope for the bulk soil is
12 driven by the non-aggregated silt and clay fractions. In contrast to Mn(p), a correlation of
13 Fe(p) and Al(p) with SOC could only be identified at the depositional site. Only poor or
14 negative correlations were found for all pyrophosphate extractable phases with SOC in the
15 microaggregate silt and clay (s+cm) fraction. Depth patterns of Al(p), Fe(p) and Mg(p) are not
16 consistent for eroding and stable profiles, but a significant decrease with depth can be
17 recognized at the depositional site. Mn(p), on the other hand, decreases significantly ($p < 0.1$)
18 with depth at all slope positions except for s+c at the depositional site and for s+cm at the
19 eroding and depositional site.

20 No strong correlation of oxalate extractable phases in the bulk soil, s+c or s+cm with SOC
21 could be identified. Oxalate and DCB and extractable elements are generally not significant or
22 negatively correlated to SOC, except for a significantly positive correlation of Fe(o) to SOC
23 in the bulk soil and non- aggregated silt and clay fraction.

3.3 Clay-sized fraction mineralogy

The abundance of secondary and primary minerals in the clay fraction differs largely along the transect and for different depths (Fig. 4), while the concentration of quartz remains fairly constant across the different transect profiles and soil depths ($15\pm1.6\%$, data not shown). Generally, the eroding slope profile has the highest number of investigated primary minerals (26 - 45%), while the lowest values are measured at the stable plateau (31 - 36%) and the deepest layer of the depositional site ($26\pm0.6\%$). The opposite trend can be observed for the abundance of pedogenic secondary minerals with the highest contents measured at the stable plateau (40 - 62%) and the depositional site (38 - 52%) while the erosional profile shows the lowest values (35 - 42%). A closer analysis of the secondary mineral fraction of all investigated soils shows the presence of both expandable (vermiculite; smectite) and non-expandable 2:1 layered clay minerals (illite; chlorite). Also larger amounts of 1:1 layered clays such as kaolinite were found, partly inherited from the parent material and as to be expected in Luvisols of this region (Van Ranst et al., 1982) (Fig. 4). Within the clay fraction, the expandable 2:1 layered mineral range between 6 – 26% relative abundance with highest values at the depositional topsoil ($26.1\pm0.5\%$) and the lowest values at the eroding slope topsoil ($5.9\pm0.4\%$). The abundance of illite is highest at the erosional subsoils ($32\pm0.9\%$) with no clear depth or slope related pattern for the remaining samples (10 - 24%), while the highest relative abundance of chlorite has been measured at the depositional deeper (55 - 70cm) subsoil ($49\pm0.7\%$). Kaolinite contents are generally increasing with depth at the stable plateau (22-62%) and decreasing with depth at the eroding profile (46-22%), while no pattern was observed for the depositional profile (35-64%).

3.4 Amino sugar abundance

The contribution of AS *per g SOC* (Fig. 5) at the depositional site is several times higher than at other positions, especially for glucosamine. Generally, AS concentrations per unit C

decrease with the size of the fraction and are especially low in the non-aggregated silt and clay fraction. In addition, the non-aggregated silt and clay fraction shows generally the highest differences between top- and subsoil abundance of AS per unit C at all slope positions; this is especially pronounced at the stable plateau and eroding slope position. Moreover, the bulk soil AS values are similar for the topsoil along the slope (GalN-C: $22.5 \pm 1.9 \text{ mg g}^{-1} \text{ SOC}$; GluN-C: $34.7 \pm 22.2 \text{ mg g}^{-1} \text{ SOC}$) with highest values at the depositional site. In contrast, subsoils of the stable plateau (GalN-C: $7.2 \pm 0.6 \text{ mg g}^{-1} \text{ SOC}$; GluN-C: $16.3 \pm 4.6 \text{ mg g}^{-1} \text{ SOC}$) and the eroding slope (GalN-C: $6.3 \pm 1.0 \text{ mg g}^{-1} \text{ SOC}$; GluN-C: $15.7 \pm 3.5 \text{ mg g}^{-1} \text{ SOC}$) profile show significantly ($p > 0.1$) lower values compared to subsoils at the depositional site (GalN-C: $25.1 \pm 6.1 \text{ mg g}^{-1} \text{ SOC}$; GluN-C: $54.2 \pm 6.9 \text{ mg g}^{-1} \text{ SOC}$). All fractions contain a large portion of AS per unit ~~SOC~~ in the topsoil (GalN-C: $19.1 \pm 3.5 \text{ mg g}^{-1} \text{ SOC}$; GluN-C: $42.0 \pm 9.7 \text{ mg g}^{-1} \text{ SOC}$), while in subsoils the contribution of AS to total C is higher in the macroaggregates (GalN-C: $16.3 \pm 2.5 \text{ mg g}^{-1} \text{ SOC}$; GluN-C: $36.5 \pm 7.3 \text{ mg g}^{-1} \text{ SOC}$) than in the microaggregate (GalN-C: $12.7 \pm 6.0 \text{ mg g}^{-1} \text{ SOC}$; GluN-C: $31.1 \pm 13.0 \text{ mg g}^{-1} \text{ SOC}$) and the non-aggregated silt and clay fraction (GalN-C: $6.5 \pm 5.7 \text{ mg g}^{-1} \text{ SOC}$; GluN-C: $12.2 \pm 12.2 \text{ mg g}^{-1} \text{ SOC}$). Interestingly, at the depositional site, the amount of AS per unit C in the microaggregate and non-aggregated silt and clay fraction decreases with depth compared to the macroaggregate fraction, where no such trend could be observed ([Fig. 5](#)).

The abundance of the two investigated ~~amino-sugars~~AS is closely correlated to SOC in the bulk soil and across all fractions (Table 4), whereas the correlation of GalN to SOC is higher ($r = 0.97^*$) than the correlation of GluN to SOC ($r = 0.71^*$). The lowest changes in the abundance of ~~amino-sugars~~AS and SOC *per g soil* ([Fig. 5](#)) are observed between top- and subsoil in the macroaggregate fractions (Ratio top- vs subsoil: 1.7-3.6), while changes in the microaggregate fractions ((Ratio top- vs subsoil: 3.1-11.5) and especially in the non-aggregated silt and clay fractions are much higher (Ratio top- vs subsoil: 4.9-32.8). Generally, the highest abundance of AS was found in topsoils, while lowest in deeper subsoil layers. But

1 this trend is much weaker in the depositional profile than in the non-eroding plateau and
2 eroding slope profile.

3 4 5 3.5 Radiocarbon ages for fractions at the depositional site

6 The measured ^{14}C ages for the investigated fractions at the depositional site differ largely for
7 different soil depths (Table 65). Generally, the ^{14}C ages increase with depth and range
8 between 635 ± 15 and 3655 ± 15 years. Consistently, the macroaggregate fraction has the oldest
9 ^{14}C age at all depths of all fractions (975 ± 15 - 3090 ± 20 years), representing distinctively older
10 C than associated with the respective microaggregate (680 ± 15 - 2670 ± 20 years) or non-
11 aggregated silt and clay fraction (635 ± 15 - 2200 ± 15 yrs). Furthermore, the data shows a shift
12 in the relative age of microaggregate vs. non-aggregated C. In the topsoil and shallow subsoil
13 the measured ^{14}C ages for these fractions are rather similar (difference of approx. 50-150yrs).
14 In the deepest subsoil a clear distinction can be observed with microaggregate associated C
15 being about approx. 500 years older than C associated with non-aggregated silt and clay.
16 Hence, ^{14}C ages for the non-aggregated silt and clay fraction in subsoils indicate a relative
17 young age of this fraction compared to the aggregated fractions.

18 19 4. Discussion

20 4.1 Organo-mineral interactions along the toposequence

21 The reactive mineral phases, predominantly manganese, that are related to the abundance of
22 SOC seem to be largely extracted with pyrophosphate (Fig. 3, Table 3). Hence, we suggest
23 that manganese influences significantly the dynamics of SOC in the investigated soil profiles
24 by promoting the formation of organo-mineral complexes and by the redistribution of these
25 complexes along the toposequence. The measured C concentrations in the pyrophosphate
26 extract (Panel I, Fig. 2) represented about 81 % of the total bulk C in these soils. This points

1 to a minor importance of well-crystallized minerals to stabilize C, as extracted with DCB, and
2 confirms earlier findings. Kodama and Schnitzer (1980) could show that the presence of
3 organic ligands in the soil solution can prevent the formation of crystalline Al and Fe oxy-
4 hydroxides e.g. due to strong complexation gradients of fulvic acids with metals which
5 prevent their complete hydroxylation. However, the absence of correlations between SOC and
6 poorly-crystalline minerals extracted with oxalate is contradictory to former studies (Kodama
7 and Schnitzer, 1980; Kaiser and Zech, 1996; Mikutta et al., 2009). Oxalate extractable soil
8 phases are generally assumed the most reactive group in many types of soils and presumably
9 strong candidates for stabilizing SOC with minerals by adsorption on mineral surfaces (Huang
10 et al., 1977; Kaiser and Guggenberger, 2003; Kleber et al., 2005; Eusterhues et al., 2005). We
11 interpret our result in terms of (i) the applied extraction scheme, (ii) the specific soil
12 conditions in which interactions between the reactive mineral phase and SOC take place and
13 (iii) to the varying mobility of the extracted phases in those soils.

14 First, in contrast to many other studies, our extraction is sequential, with pyrophosphate,
15 oxalate and DCB applied in this order on the same sample. Other studies that do not use
16 pyrophosphate extract with oxalate a potentially large part of organo-mineral associations.
17 These associations are in our design already partly extracted with pyrophosphate. Second,
18 even though concentrations of pyrophosphate extracted iron and aluminum were high in
19 microaggregates, no strong positive correlations with C were found for these elements (Table
20 3, Fig. 2). The fact that pyrophosphate extractable elements are largely abundant in aggregates
21 raises questions about the importance of the isolated elements for supporting the physical
22 protection of C against decomposition. One explanation could be that pyrophosphate
23 extractable Fe and Al are important for aggregate stability as ligands between clay minerals,
24 even without involving SOC, but not for SOC retention itself. Fe(p) and Al(p) could act as
25 proxies for the formation of aggregates through the formation of organo-metallic complexes,
26 but also through the formation of Fe, Al oxy-hydroxides (Kaiser and Zech, 1996).

Pyrophosphate in that case might simply act as a strong reagent to disaggregate soil (Muneer and Oades, 1989) and therefore the abundance of pyrophosphate extractable elements in aggregates does not point at an identifiable group of organo-mineral associations (Kaiser and Zech, 1996). Third, the pH-values in our soils are near neutral pH (Table 1), which is a pH buffer zone where Mn is highly mobile as Mn^{2+} , potentially forming organo-mineral complexes, while the mobility of Fe and Al is strongly limited at $pH > 6$ (Lindsay, 1979). However, the data indicates a potential impact of oxalate extractable Fe oxides to stabilize SOC where physical protection of SOC by aggregation is weak, namely at the eroding site (Table 3). There, the aggregation potential is small (Doetterl et al., 2012) due to the continuous removal of aggregate-rich topsoil layers.

4.2 Importance of clay-sized mineral distribution

The presented data for the abundance of different clay-sized minerals (Fig. 4) clearly indicates a large variety of clay minerals along the slope transect. In combination with the high TRB values at the eroding site, the large abundance of primary minerals and illite (= non-expandable 2:1 layered clay) and the relatively low content of pedogenic clay minerals (1:1 and 2:1 layered clay minerals) indicate a lower level of weathering at the eroding compared to the plateau and depositional site. This data is in accordance with the general assumption that i.e. kaolinite is one of the end products in the weathering sequence in soils and that the eroding profile is less weathered than the stable and depositional profile (see section 3.1). The deepest layer of the eroding profile is the least weathered part of soil along the sequence. Kaolinite there is mostly derived from the parent material, hence lowest in comparison to other samples undergoing more advanced weathering.

The applied quantitative approach allows us to document a relative depletion of expandable 2:1 clay minerals (smectite and vermiculite) in the topsoil of the eroding site ($3.2 \pm 0.4\%$) and their enrichment in the topsoil of the depositional site ($15.4 \pm 1.7\%$). This suggests the

1 preferential mobilization of smectite and vermiculite at the eroding site and its deposition at
2 the foothill as soil gets mobilized and eroded mostly as aggregates (Wang et al., 2010).
3 Aggregates dominated by expandable clay minerals have been identified to be less stable due
4 to shrinking and swelling than kaolinite or illite-rich aggregates (Fan et al., 2008) or
5 aggregates rich in amorphous and crystalline Al and Fe oxide (Goldberg, 1989). The lower
6 stability of smectite or vermiculite-dominated aggregates then leads to a preferential
7 breakdown and mobilization of these aggregates resulting in smaller particles more
8 susceptible for erosion, leading to transport to and burial at depositional sites. Our data (Fig. 4
9 and Doetterl et al. 2012) is consistent with the observation of the preferential export of small
10 particle sizes out of the catchment, which are often dominated by smectite colloids (Gibbs,
11 1967). At the depositional site, weathering continues and alters clay minerals which are
12 involved in the formation of aggregates and SOC stabilization, for example by transforming
13 expandable clay into non-expandable forms such as chloride (Fig. 4; depositional footslope
14 55-70cm). Both mechanisms together (weathering and transportation) can hence explain the
15 low amount of aggregates at eroding sites, and the loss of aggregates with depth at the
16 depositional site (Table 1) as weathering continues. However, we cannot elaborate whether
17 the breakdown of aggregates at the depositional site is induced first by decomposition of C, or
18 mineral weathering. Nevertheless, no strong and significant correlations between specific clay
19 minerals and C content in different fractions or the clay content and SOC concentration could
20 be identified along the slope (data not shown). Even though a direct quantification of the
21 impact of clay minerals on stabilizing SOC is not possible with our analysis, the data indicates
22 that in these soils with rather low bulk SOC concentrations of 1.2-0.2%, clay surfaces are
23 likely not the limiting factor for C stabilization, even if the amount of highly reactive clay
24 minerals such as vermiculite or smectite are small. This is consistent with observations of
25 Duemig et al. (2012) where higher SOC loadings of clay minerals were observed in

consequence of a shortage of reactive surface area in clay depleted soils compared to more clay-rich soils.

4.3 Radiocarbon ages in relation to microbial residues, respiration and particulate organic matter

The fact that the AS content per unit C in the macroaggregate and, to a weaker extent, in the microaggregate and non-aggregated silt and clay fractions at the depositional site does not decrease significantly after burial, in comparison to very low subsoil AS concentrations at the eroding site (Fig. 5), leads to the conclusion that burial of soil can lead to the storage of an otherwise relatively easy decomposable part of SOC in soils for decades and centuries. At the same time C in macroaggregates has shown the oldest ^{14}C ages of all isolated fractions (Table 6).

X. Wang et al. (2014a) report for a cropland slope from the same region with similar topographic setting, a re-aggregation of deposited and buried C within macroaggregates. Our data showing old ^{14}C ages of macroaggregate associated C is supportive of this, indicating indeed a preferential accumulation of old carbon in macroaggregates at the depositional site (Table 65). The measured old ^{14}C ages and the high potential respiration rates measured for these soils rich in macroaggregates (i.e. Doetterl et al., 2012) indicate furthermore the presence of C sources with differing availability to decomposers within the macroaggregate fraction. On the one hand, these aggregates represent a proper habitat for microorganisms to thrive on C (Denef et al., 2001). Respiration data (presented in our former work (Doetterl et al. 2012) clearly shows that soils in this sequence with large amounts of macroaggregate C also respire at a higher rates than soils low on macroaggregate C. On the other hand, the presence of high amounts of microbial residues (Fig. 5) can enhance the formation of stable aggregates (Bossuyt et al., 2001; Cotrufo et al., 2012). Hence, macroaggregates represent hotspots of microbial life and activity (Kolb et al., 2009) with at the same time old ^{14}C ages,

1 indicating higher stability. A conversion of SOC from non-microbial to microbial carbon
2 within stable aggregates would then not alter the ^{14}C signature of the fraction, but the
3 potential stability of this fraction would still change. This is supported by recent studies that
4 highlight that the genetic variety of microbial communities are maintained in buried surface
5 soils, and that large amounts of microbial biomass can be found in deeper layers of burial
6 sites, driven by the abundance of organic matter as a nutrient source (Helgason et al., 2014).
7 Hence, we argue that ^{14}C ages and high AS contents in macroaggregates indicate that
8 microorganisms either feed on old carbon to build up microbial biomass, if conditions are
9 suitable, or that these environments contain considerable amounts of old C and are proper
10 habitats for microorganisms where their residues are preserved.

12 4.4 Soil redistribution and the depth related changes of carbon at the depositional sites

13 Our results show the ongoing rejuvenation of the soil at the eroding hillslope by removing soil
14 material and exposing less weathered material to the surface: The lower loss of cations
15 compared to the parent material (Table 2) indicates that the continuous removal of soil at the
16 hillslope is acting at a faster rate than the natural soil weathering and this keeps the eroding
17 profile TRB higher, and hence less weathered, than at the stable plateau site. This is consistent
18 with the absence of a depth related trend in the TRB at the depositional site (Table 2) as this
19 material is buried former topsoil, similar in TRB. Changes in C stocks with depth at the
20 depositional sites can shed light on the underlying mechanistic relationships as the buried
21 topsoil should have similar C content and composition as the present topsoil if no changes in
22 respect to the composition of C fractions occur, given the low C contents in subsoils of soils
23 not affected by burial processes (Table 1). Hence, decreasing amounts of only certain
24 fractions must be related to the decomposition of C within these fractions. Furthermore,
25 comparing this decrease to C contents associated with other fractions allows assessing
26 information on the effectiveness of protection through a specific set of stabilization

[mechanisms after burial](#). A recently published study on SOC stabilization along a 500 year chronosequence from wetland to cropland in China illustrates the importance of understanding the interaction of stabilization mechanisms for SOC by pointing out that “*beyond providing a physical barrier between microbes and carbon substrates, soil aggregation could contribute to SOC stabilization by bringing organic matter and soil particles together and promoting organo-mineral interaction*” (Cui et al., 2014).

In contradiction to the findings of [X. Wang et al. \(2014a\)](#) our data indicates the strengthening effect of physical protection/aggregation on the stabilization of C against decomposition. While the relative contribution of C associated with the non-aggregated silt and clay fraction is increasing with depth (Table 1), its age compared to C associated with aggregates is becoming younger (Table [5](#)). We conclude from this that non- aggregated C is not as stable against decomposition as often assumed (Yu et al., 2011). Within aggregates on the other hand, mineral surfaces are “locked away” from new C input in a similar way as aggregated C is not as easily accessible for decomposers as non-aggregated C and can hence result in a slower turnover of this C. Instead, mineral surfaces of the non- aggregated silt and clay C can act very efficiently in stabilizing younger C (Eusterhues et al., 2005; 2008; Duemig et al., 2011; 2012) resulting in younger radiocarbon ages and at the same time similar C loading as for microaggregates (Table 5). However, the amount of macro- and microaggregates is steadily decreasing with depth, indicating that (i) the ongoing weathering of soils and hence alterations of minerals ([Fig. 6](#)) that build up the aggregates and (ii) the slow but steady decomposition of C leads to a deterioration of aggregates and, hence, this mechanism of protecting C through physical isolation.

5. Conclusions

Our findings ([Fig. 6](#)) highlight 1) the control of soil redistribution on the distribution of functional SOC fractions; 2) the abundance and weathering of clay minerals and pedogenic

oxyhydroxides is partly responsible for the effectiveness of geochemical and physical stabilization mechanism of SOC along the slope and with soil depth; 3) mineral alteration and the breakdown of aggregates limit the protection of C by minerals and within aggregates temporally; 4) pyrophosphate extractable iron and aluminum are important for aggregate stability but not for SOC retention in near neutral pH soils 5) the combined effect of geochemical and physical protection of C after burial is prevalent at the depositional sites but changes through time, and 6) microorganisms feed on old carbon to build up microbial biomass within aggregates or aggregates are environments containing considerable amounts of old C where microorganisms thrive and their residues are preserved. The question remains, however, what is the importance of these findings for our understanding of C sequestration and potential sink and source functions in dynamic landscapes?

In conclusion, a considerable amount of C has been stabilized with the mineral phase. These minerals play a crucial role in supporting the dynamic replacement (Harden et al., 1999) of eroded C with new C input while at the same time burying the eroded C at depositional sites can provide a potential sink for atmospheric C. However, this potential sink function is largely temporally and spatially limited. Due to the continues weathering and hence weakening of different protection mechanisms, the buried C at depositional sites is not indefinitely stable and will be turned over and, slowly but steadily, be released back to the atmosphere with a delay of decades to centuries.

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performed research and analyzed the data and all authors interpreted the data and wrote the paper.

7. References

Amelung, W., Miltner, A., Zhang, X., and Zech, W.: Fate of microbial residues during litter decomposition as affected by minerals, *Soil Sci.*, 166, 598–606, 2001.

Amelung, W., Brodowski, S., Sandhage-Hofmann, A., and Bol, R.: Combining biomarker with stable isotope analyses for assessing the transformation and turnover of soil organic matter, *Adv. Agron.*, 100, 155–250, 2008.

Bascomb, C. L.: Distribution of pyrophosphate-extractable iron and organic carbon in soils of various groups, *J. Soil Sci.*, 19, 251–268, 1968.

Berhe, A. A., Harden, J. W., Torn, M. S., Kleber, M., Burton, S. D., and Harte, J.: Persistence of soil organic matter in eroding vs. depositional landform positions, *J. Geophys. Res.*, 117, G02019, doi:10.1029/2011JG001790, 2012.

Bodé, S.: Development of a Compound Specific Stable Isotope Method for Amino Sugars as Tracers of Microbial Dynamics in Terrestrial Ecosystems, PhD thesis, Ghent University, Belgium, 206 pp., 2013.

Bodé, S., Denef, K., and Boeckx, P.: Development and evaluation of a high-performance liquid chromatography/isotope ratio mass spectrometry methodology for delta C-13 analyses of amino sugars in soil, *Rapid Commun. Mass Sp.*, 23, 2519–2526, 2009.

Bossuyt, H., Denef, K., Six, J., Frey, S. D., Merckx, R., and Paustian, K.: Influence of microbial populations and residue quality on aggregate stability, *Appl. Soil Ecol.*, 16, 195–208, 2001.

Brahy, V. and Delvaux, B.: Cation exchange resin and test vermiculite to study soil processes in situ in a toposequence of Luvisol and Cambisol on loess, *Eur. J. Soil Sci.*, 52, 397–408, 2001.

Brindley, G. W. and Brown, G.: Crystal Structures of Clay Minerals and Their Identification, Mineralogical Society Monograph no. 5, Mineralogical Society, London, 495 pp., 1980.

Chao, T. T. and Sanzolone, R. F.: Decomposition techniques, *J. Geochem. Explor.*, 44, 65–106, 1992.

Cotrufo, M. F., Wallenstein, M. D., Boot, C., Denef, K., and Paul, E.: The Microbial Efficiency- Matrix Stabilization (MEMS) framework integrates plant litter decomposition with soil organic matter stabilization: do labile plant inputs form stable soil organic matter?, *Glob. Change Biol.*, 19, 988–995, 2012.

Cui, J., Li, Z., Liu, Z., Ge, B., Fang, C., Zhou, C., and Tang, B.: Physical and chemical stabilization of soil organic carbon along a 500 year cultivated soil chronosequence originating

- from estuarine wetlands: temporal patterns and land use effects, *Agr. Ecosyst. Environ.*, 196, 10–20, 2014.
- Dahlgren, R. A.: Quantification of allophane and imogolite, in: *Quantitative methods in soil mineralogy*, edited by: Zelazny, L. W. and Amonette, J. E., Soil Science Society of America, Madison, 430–451, 1994.
- Denef, K., Six, J., Bossuyt, H., Frey, S. D., Elliott, E. T., Merckx, R., and Pasutian, K.: Influence of dry-wet cycles on the interrelationship between aggregate, particulate organic matter, and microbial community dynamics, *Soil Biol. Biochem.*, 33, 1599–1611, 2001.
- Dlugoss, V.: Impacts of Soil Redistribution Processes on Soil Organic Carbon Stocks and Fluxes in a Small Agricultural Catchment, PhD thesis, Geographisches Institut der Universität zu Köln, Köln, 143 pp., 2011.
- Doetterl, S., Six, J., VanWesemael, B., and Van Oost, K.: Carbon cycling in eroding landscapes, geomorphic controls on soil organic C pool composition and C stabilization, *Glob. Change Biol.*, 18, 2218–2232, 2012.
- Duemig, A., Smittenberg, R., and Koegel-Knabner, I.: Concurrent evolution of organic and mineral components during initial soil development after retreat of the Damma glacier, 5 Switzerland, *Geoderma*, 163, 83–94, 2011.
- Duemig, A., Haeusler, W., Steffens, M., and Koegel-Knabner, I.: Clay fractions from a soil chronosequence after glacier retreat reveal the initial evolution of organo-mineral associations, *Geochim. Cosmochim. Ac.*, 85, 1–18, 2012.
- Dungait, J. A. J., Hopkins, D. W., Gregory, A. S., and Whitmore, A. P.: Soil organic matter turnover is governed by accessibility not recalcitrance, *Glob. Change Biol.*, 18, 1781–1796, 2012.
- Eckmeier, E., Gerlach, R., Skjemstad, J. O., Ehrmann, O., and Schmidt, M.W. I.: Minor changes in soil organic carbon and charcoal concentrations detected in a temperate deciduous forest a year after an experimental slash-and-burn, *Biogeosciences*, 4, 377–383, doi:10.5194/bg-4-377-2007, 2007.
- Eusterhues, K., Rumpel, C., and Koegel-Knabner, I.: Organo-mineral associations in sandy acid forest soils, importance of specific surface area, iron oxides and micropores, *Eur. J. Soil Sci.*, 56, 753–763, 2005.
- Eusterhues, K., Wagner, F. E., Haeusler, W., Hanzlik, M., Knicker, H., Totsche, K. U., Koegel-Knabner, I., and Schwertmann, U.: Characterization of ferrihydrite-soil organic matter coprecipitates by X-ray diffraction and Mössbauer spectroscopy, *Environ. Sci. Technol.*, 42, 7891–7897, 2008.
- Fan, Y., Lei, T., and Cai, Q.: Effects of wetting rate on soil surface strength and aggregate stability, *Transactions of the Chinese Society of Agricultural Engineering*, 24, 46–50, 2008.
- Fontaine, S., Barot, S., Barre, P., Bdioui, N., Mary, B., and Rumpel, C.: Stability of organic carbon in deep soil layers controlled by fresh carbon supply, *Nature*, 450, 277–280, 2007.

- 1 Gibbs, R. J.: The geochemistry of the Amazon River system. Part I. The factors that control
2 the salinity and the composition and concentration of the suspended solids, *Bull. Geol. Soc.*
3 *Am.*, 78, 1203–1232, 1967.
- 4
- 5 Glaser, B. and Gross, S.: Compound-specific delta C-13 analysis of individual amino sugars –
6 a tool to quantify timing and amount of soil microbial residue stabilization, *Rapid Commun.*
7 *Mass Sp.*, 19, 1409–1416, 2005.
- 8
- 9 Glaser, B., Turrion, M. B., and Alef, K.: Amino sugars and muramic acid – biomarkers for
10 soil microbial community structure analysis, *Soil Biol. Biochem.*, 36, 399–407, 2004.
- 11
- 12 Goldberg, S.: Interaction of aluminium and iron oxides and clay minerals and their effect on
13 soil physical properties: a review, *Commun. Soil Sci. Plan.*, 20, 1181–1207, 1989.
- 14
- 15 Guggenberger, G., Frey, S. D., Six, J., Paustian, K., and Elliott, E. T.: Bacterial and fungal
16 cell-wall residues in conventional and no-tillage agroecosystems, *Soil Sci. Soc. Am. J.*, 63,
17 1188–1198, 1999.
- 18
- 19 Harden, J. W., Sharpe, J. M., Parton, W. J., Ojima, D. S., Fries, T. L., Huntington, T. G., and
20 Dabney, S. M.: Dynamic replacement and loss of soil carbon on eroding cropland, *Global*
21 *Biogeochem. Cy.*, 13, 885–901, 1999.
- 22
- 23 Helgason, B. I., Konschu, H. J., Bedard-Haughn, A., and VandenBygaart, A. J.: Microbial
24 distribution in an eroded landscape: buried A horizons support abundant and unique
25 communities, *Agr. Ecosyst. Environ.*, 196, 94–102, 2014.
- 26
- 27 Herbillon, A. J.: Chemical estimation of weatherable minerals present in the diagnostic
28 horizons of low activity clay soils, in: *Proceedings of the eighth international soil*
29 *classification workshop – Classification, Characterization and Utilization of Oxisols. Part 1 –*
30 *Papers*, edited by: Beinroth, F. H., Camargo, M. N., and Eswaran, H., Rio de Janeiro, 12 to 23
31 May 1986, 39–48, 1988.
- 32
- 33 Huang, P. M., Wang, T. S. C., Wang, M. K., Wu, M. H., and Hsu, N. W.: Retention of
34 phenolic acids by noncrystalline hydroxy-aluminum and -iron compounds and clay minerals
35 of soils, *Soil Sci.*, 123, 213–219, 1977.
- 36
- 37 Hoffmann, T., Schlummer, M., Notebaert, B., Verstraeten, G., and Korup, O.: Carbon burial
38 in soil sediments from Holocene agricultural erosion, Central Europe, *Global Biogeochem.*
39 *Cy.*, 27, 828–835, doi:10.1002/gbc.20071, 2013.
- 40
- 41 Kaiser, K. and Guggenberger, G.: Mineral surfaces and soil organic matter, *Eur. J. Soil Sci.*,
42 54, 219–236, 2003.
- 43
- 44 Kaiser, K. and Zech, W.: Defects in estimation of aluminum in humus complexes of podzolic
45 soils by pyrophosphate extraction, *Soil Sci.*, 161, 452–458, 1996.
- 46
- 47 Kleber, M.: What is recalcitrant soil organic matter?, *Environ. Chem.*, 7, 320–332, 2010.
- 48
- 49 Kleber, M., Mikutta, R., Torn, M. S., and Jahn, R.: Poorly crystalline mineral phases protect
50 organic matter in acid subsoil horizons, *Eur. J. Soil Sci.*, 56, 717–725, 2005.
- 51

- 1 Kodama, A. and Schnitzer, M.: Effects of fulvic acids on the crystallization of aluminum
2 hydroxides, *Geoderma*, 24, 195–205, 1980.
- 3
- 4 IRM: Climatogram for the Period 1961–1990, Station 06447 (Uccle; Belgique), retrieved 28
5 July 2011, available at: <http://www.recordmeteo.com/weather-hi-low/weather-station-ducclen-6447.html> (last access: 25 November 2014), 2011.
- 6
- 7
- 8 Johnson, W. C.: Carbon cycle: sequestration in buried soils, *Nat. Geosci.*, 7, 398–399, 2014.
- 9 Kolb, S. E., Fermanisch, K. J., and Dornbush, M. E.: Effect of charcoal 5 quantity on
10 microbial biomass and activity in temperate soils, *Soil Sci. Soc. Am. J.*, 7, 1173–1181, 2009.
- 11
- 12 Lannoo: *Le grand Atlas de Ferraris*, Lannoo, 2009.
- 13
- 14 Lindsay, W. L. (ed): *Chemical equilibria in soils*, The Blackburn Press, West Caldwell, USA,
15 450 pp., 1979.
- 16
- 17 Marin-Spiotta, E., Chapricha, N. T., Plante, A. F., Diefendorf, A. F., Mueller, C. W., Grandy,
18 A. S., and Mason, J. A.: Long-term stabilization of deep soil carbon by fire and burial during
19 early Holocene climate change, *Nat. Geosci.*, 7, 428–432, 2014.
- 20
- 21 Mehra, O. P. and Jackson, M. L.: Iron oxide removal from soils and clays by a dithionite –
22 citrate system buffered with sodium bicarbonate, *Clay. Miner.*, 5, 317–327, 1960.
- 23
- 24 Mikutta, R., Schaumann, G. E., Gildemeister, D., Bonneville, S., Kramer, M. G., Chadwick,
25 O. A., and Guggenberger, G.: Biogeochemistry of mineral-organic associations across a long-
26 term mineralogical soil gradient (0.3–4100 kyr), Hawaiian Islands, *Geochim. Cosmochim.*
27 *Ac.*, 73, 234–260, 2009.
- 28
- 29 Muneer, M. and Oades, J. M.: The role of Ca-organic interactions in soil aggregate stability.
30 III. Mechanisms and models, *Aust. J. Soil Res.*, 27, 411–423, 1989.
- 31
- 32 Park, J.-H., Meusburger, K., Jang, I., Hojeong, K., and Alewell, C.: Erosion-induced changes
33 in soil biogeochemical and microbiological properties in Swiss Alpine grasslands, *Soil Biol.*
34 *Biochem.*, 69, 382–392, 2014.
- 35
- 36 R Development Core Team: *R, a Language and Environment for Statistical Computing*, R
37 Foundation for Statistical Computing, available at: <http://www.r-project.org>, last access: July
38 2014, 2010.
- 39
- 40 Robert, M. and Tessier, D.: Méthode de préparation des argiles des sols pour des études
41 minéralogiques, *Annals Agronomiques*, 25, 859–882, 1974 (in French).
- 42
- 43 Roberts, P., Bol, R., and Jones, D. L.: Free amino sugar reactions in soil in relation to soil
44 carbon and nitrogen cycling, *Soil Biol. Biochem.*, 39, 3081–3092, 2007.
- 45
- 46 Rommens, T., Verstraeten, G., Poesen, J., Govers, G., Van Rompaey, A., Peeters, I., and
47 Lang, A.: Soil erosion and sediment deposition in the Belgian loess belt during the Holocene,
48 establishing a sediment budget for a small agricultural catchment, *Holocene*, 15, 1032–1043,
49 2005.
- 50

- 1 Salomé, C., Nunan, N., Pouteau, V., Lerch, T. Z., and Chenu, C.: Carbon dynamics in topsoil
2 and in subsoil may be controlled by different regulatory mechanisms, *Glob. Change Biol.*, 16,
3 416–426, 2010.
- 4
- 5 Schmidt, M. W. I., Torn, M. S., Abiven, S., Dittmar, T., Guggenberger, G., Janssens, I. A.,
6 Kleber, M., Koegel-Knabner, I., Lehmann, J., Manning, D. A. C., Nannipieri, P., Rasse, D. P.,
7 Weiner, S., and Trumbore, S. E.: Persistence of soil organic matter as an ecosystem property,
8 *Nature*, 478, 49–56, 2011.
- 9
- 10 Schnitzer, M. and Kodama, H.: Interactions between organic and inorganic components in
11 particle-size fractions separated from four soils, *Soil Sci. Soc. Am. J.*, 56, 1099–1105, 1992.
- 12
- 13 Schuppli, P. A., Ross, G. J., and McKeague, J. A.: The effective removal of suspended
14 materials from pyrophosphate extracts of soils from tropical and temperate regions, *Soil Sci.*
15 *Soc. Am. J.*, 47, 1026–1032, 1983.
- 16
- 17 Simpson, R. T., Frey, S. D., Six, J., and Thiet, R. K.: Preferential accumulation of microbial
18 carbon in aggregate structures of no-tillage soils, *Soil Sci. Soc. Am. J.*, 68, 1249–1255,
19 2004.
- 20
- 21 Six, J., Elliott, E. T., Paustian, K., and Doran, J. W.: Aggregation and soil organic matter
22 accumulation in cultivated and native grassland soils, *Soil Sci. Soc. Am. J.*, 62, 1367–1377,
23 1998.
- 24
- 25 Six, J., Conant, R. T., Paul, E. A., and Paustian, K.: Stabilization mechanisms of soil organic
26 matter, implications for C-saturation of soils, *Plant Soil*, 241, 155–176, 2002.
- 27
- 28 Sollins, P., Homann, P., and Caidwell, B. A.: Stabilization and destabilization of soil organic
29 matter: mechanisms and controls, *Geoderma*, 74, 65–105, 1996.
- 30
- 31 Stucki, J. W., Goodman, B. A., and Schwertmann, U.: *Iron in Soils and Clay Minerals*,
32 Springer, Dordrecht, 893 pp., 1988.
- 33
- 34 Stuiver, M. and Polach, H. A.: Discussion – reporting of ¹⁴C data, *Radiocarbon*, 19, 355–363,
35 1977.
- 36
- 37 Süsser, P. and Schwertmann, U.: Proton buffering in mineral horizons of some acid forest
38 soils, *Geoderma*, 49, 63–76, 1991.
- 39
- 40 Van Oost, K., Verstraeten, G., Doetterl, S., Notebaert, B., Wiaux, F., Broothaerts, N., and Six,
41 J.: Legacy of human-induced C erosion and burial on soil-atmosphere C exchange, *P. Natl.*
42 *Acad. Sci. USA*, 109, 19492–19497, doi:10.1073/pnas.1211162109, 2012.
- 43
- 44 Van Ranst, E., De Coninck, F., Tavernier, R., and Langohr, R.: Mineralogy in silty to loamy
45 soils of central and high Belgium in respect to autochthonous and allochthonous materials,
46 *Bulletin de la Société Belge de Géologie*, 91, 27–44, 1982.
- 47
- 48 von Luetzow, M. and Koegel-Knabner, I.: Response to the concept paper: “What is
49 recalcitrant soil organic matter?” by Markus Kleber, *Environ. Chem.*, 5 7, 333–335, 2010.
- 50

- 1 von Luetzow, M., Koegel-Knabner, I., Ekschmitt, K., Matzner, E., Guggenberger, G.,
2 Marschner, B., and Flessa, H.: Stabilization of organic matter in temperate soils, mechanisms
3 and their relevance under different soil conditions – a review, *Eur. J. Soil Sci.*, 57, 426–445,
4 2006.
- 5
- 6 Wang, X., Cammeraat, L. H., Wang, Z., Zhou, J., Govers, G., and Kalbitz, K.: Stability of
7 organic matter in soils of the Belgian Loess Belt upon erosion and deposition, *Eur. J. Soil*
8 *Sci.*, 64, 219–228, 2013.
- 9
- 10 Wang, X., Cammeraat, E. L. H., Cerli, C., and Kalbitz, K.: Soil aggregation and the
11 stabilization of organic carbon as affected by erosion and deposition, *Soil Biol. Biochem.*, 72,
12 55–65, 2014.
- 13
- 14 Wang, Z., Govers, G., Steegen, A., Clymans, W., Van den Putte, A., Langhans, C., Merckx,
15 R., and Van Oost, K.: Catchment-scale carbon redistribution and delivery by water erosion in
16 an intensively cultivated area, *Geomorphology*, 124, 65–74, 2010.
- 17
- 18 Wang, Z., Van Oost, K., Lang, A., Quine, T., Clymans, W., Merckx, R., Notebaert, B., and
19 Govers, G.: The fate of buried organic carbon in colluvial soils: a long-term perspective,
20 *Biogeosciences*, 11, 873–883, doi:10.5194/bg-11-873-2014, 2014.
- 21
- 22 Wiaux, F., Cornelis, J.-T., Cao, W., Vanclooster, M., and Van Oost, K.: Combined effect of
23 geomorphic and pedogenic processes on the distribution of soil organic carbon quality along
24 an eroding hillslope on loess soil, *Geoderma*, 214, 36–47, 2014a.
- 25
- 26 Wiaux, F., Vanclooster, M., Cornelis, J.-T., and Van Oost, K.: Factors controlling soil organic
27 carbon persistence along an eroding hillslope in the loess belt, *Soil Biol. Biochem.*, 77, 187–
28 196, 2014b.
- 29
- 30 Wouters, L. and Vandenberghe, N.: *Géologie de la Campine, Essai de synthèse*, ONDRAF,
31 Brussels, 1994.
- 32
- 33 Xu, X., Trumbore, S. E., Zheng, S., Southon, J. R., McDuffee, K. E., Luttgen, M., and Liu, J.
34 C.: Modifying a sealed tube zinc reduction method for preparation of AMS graphite targets:
35 reducing background and attaining high precision, *Nucl. Instrum. Meth. B*, 259, 320–329,
36 2007.
- 37
- 38 Yu, H., Ding, W., Luo, J., Geng, R., Ghani, A., and Cai, Z.: Effects of long-term compost and
39 fertilizer application on stability of aggregate-associated organic carbon in an intensively
40 cultivated sandy loam soil, *Biol. Fert. Soils*, 48, 325–336, doi:10.1007/s00374-011-0629-2,
41 2011.
- 42
- 43 Yoo, K., Amundson, R., Heimsath, A. M., and Dietrich, W. E.: Spatial patterns of soil organic
44 carbon on hillslopes: integrating geomorphic processes and the biological C cycle, *Geoderma*,
45 130, 47–65, 2006.
- 46
- 47
- 48

8. Tables

Table 1: Key soil and geomorphological properties and identified SOC fractions as described by Doetterl et al. (2012) and Wang et al. (2014b) for the investigated soils at different slope positions and soil depths.

Parameter		Units	Stable plateau			Eroding slope			Depositional footslope		
Slope		[%]	3			10			4		
Total Erosion(-) & Deposition (+)		[cm]	0			-200			350		
Current Erosion & Deposition rate		[mm yr ⁻¹]	0			-1.32			1.16		
Depth		[cm]	0-15	35-50	55-70	0-15	35-50	55-70	0-15	35-50	55-70
Bulk density		[g cm ⁻³]	1.58	1.56	1.59	1.52	1.63	1.51	1.55	1.64	1.58
pH [CaCl ₂]			7.0	7.1	7.0	7.2	7.3	7.1	6.8	6.8	6.9
SOC (g kg ⁻¹)	Bulk soil		9.6	2.6	2.4	9.3	2.1	2.1	11.6	6.7	3.9
	Macroaggregate		14.3	3.0	4.1	12.1	3.0	5.2	14.3	7.8	6.3
	Microaggregate		7.3	2.6	2.2	7.3	2.8	2.4	9.3	5.9	3.5
	Non-aggregated silt+ clay		8.4	2.3	2.0	7.5	1.7	1.6	9.1	6.7	3.1
Relative C (% bulk SOC)	Macroaggregate		48	19	24	46	19	27	58	37	35
	Microaggregate		35	48	47	24	34	24	21	37	21
	Non-aggregated silt+ clay		17	33	29	30	47	49	21	26	44
C stock (gC m ⁻² cm ⁻¹ depth)	Bulk soil		152	41	38	141	34	32	180	110	62
	Macroaggregate		72	8	9	63	7	9	105	40	22
	Microaggregate		54	19	18	33	12	8	37	40	13
	Non-aggregated silt+ clay		26	13	11	41	16	16	39	29	27

- 1 Table 2: Total content in Ca, K, Mg, Na and Total Reserve in Bases (TRB) values for the
- 2 different soil profiles and depths and the parent material.

Parameter	Unit	Stable plateau			Eroding slope			Depositional footslope			Parent material (Loess)
Depth	[cm]	0-15	35-50	55-70	0-15	35-50	55-70	0-15	35-50	55-70	>100cm
Ca	[cmol _c kg ⁻¹]	14	13	13	15	12	14	15	13	13	70
K		46	46	48	47	50	46	42	41	43	32
Mg		15	20	19	17	19	19	14	14	16	18
Na		31	34	32	32	33	38	32	32	32	18
Total TRB		107	113	112	111	115	118	103	101	103	139
<i>TRB relative. to parent material</i>	(%)	77	81	81	80	83	85	75	73	74	100

1 Table 3: Correlations (Pearson r) between pyrophosphate, oxalate and DCB extractable solid soil phases and SOC or soil depth (* = p <0.1).

SOC fraction	Position	Parameter	Pyrophosphate extractable				Oxalate extractable				DCB extractable			
			Al _(p)	Fe _(p)	Mg _(p)	Mn _(p)	Al _(o)	Fe _(o)	Mg _(o)	Mn _(o)	Al _(d)	Fe _(d)	Mg _(d)	Mn _(d)
Bulk soil	Non-eroding	Pearson r Phase vs depth	0.34	-0.45	-0.27	-0.95*	0.84*	0.09	-0.73*	0.37	0.74*	0.84*	0.60	0.61*
		Pearson r Phase vs. SOC	-0.37	0.4	0.06	0.99*	-0.87*	0.02	0.84*	-0.09	-0.57	-0.68*	-0.33	-0.36
		Range mg kg ⁻¹	70-223	118-410	62-119	6-73	647-1076	1512-1928	38-75	255-379	788-1280	6401-12161	81-132	43-60
	Eroding slope	Pearson r Phase vs depth	0.17	-0.12	0.1	-0.9*	0.09	-0.85*	-0.41	0.83	0.18	0.57	0.53	-0.53
		Pearson r Phase vs. SOC	0.04	0.34	0.13	0.99*	-0.38	0.75*	0.41	-0.95	-0.35	-0.72*	-0.54	0.55
		Range mg kg ⁻¹	53-298	91-498	58-207	6-84	554-865	829-1531	42-66	191-282	783-1027	6277-9593	73-128	44-79
	Depositional footslope	Pearson r Phase vs depth	-0.23	-0.67*	-0.68	-0.99*	0.83*	0.05	-0.43	0.99*	0.5	0.64*	0.29	0.25
		Pearson r Phase vs. SOC	0.23	0.67*	0.69	0.99*	-0.83*	-0.05	0.43	-0.99*	-0.49	-0.64*	-0.29	-0.25
		Range mg kg ⁻¹	58-180	106-320	69-133	18-112	432-636	930-1157	40-65	181-265	569-1234	5319-10324	67-146	41-60
Non-aggregated s+c	Non-eroding	Pearson r Phase vs depth	0.61*	0.05	0.24	-0.88*	-0.17	-0.33	0.19	-0.42	0.8	0.97*	0.12	0.56
		Pearson r Phase vs. SOC	-0.47	0.2	-0.02	0.95*	-0.08	0.57	0.18	0.46	-0.92*	-0.99*	0.17	-0.39
		Range mg kg ⁻¹	105-403	67-252	42-93	10-107	792-1842	1557-1858	38-75	152-301	1003-1626	7188-10760	100-137	53-57
	Eroding slope	Pearson r Phase vs depth	0.44	0.18	0.29	-0.74*	0.16	-0.92*	0.29	0.64*	0.21	0.59*	0.78*	0.54
		Pearson r Phase vs. SOC	-0.27	-0.06	-0.12	0.72*	-0.49	0.81*	-0.36	-0.77*	-0.32	-0.81*	-0.82*	-0.46
		Range mg kg ⁻¹	87-568	46-362	33-152	4-153	746-1035	686-1360	42-63	113-168	1016-1294	8288-10783	115-172	50-56
	Depositional footslope	Pearson r Phase vs depth	-0.76*	-0.48	-0.5	-0.5	0.54	0.55	0.57	0.29	0.71*	0.76*	0.07	0.85*
		Pearson r Phase vs. SOC	0.76*	0.48	0.5	0.5	-0.46	-0.51	-0.63*	-0.2	-0.71*	-0.76*	-0.07	-0.85*
		Range mg kg ⁻¹	107-425	73-271	31-125	13-168	513-871	802-1085	44-62	68-175	855-1087	6612-8795	117-167	45-69
Aggregated s+cm	Non-eroding	Pearson r Phase vs depth	0.80*	0.58	0.5	-0.69*	0.75*	0.62*	0.42	0.52	0.33	0.54	0.21	-0.4
		Pearson r Phase vs. SOC	-0.96*	-0.76*	-0.64*	0.81*	-0.98*	-0.77*	0.07	-0.82*	-0.82*	-0.94*	-0.54	-0.21
		Range mg kg ⁻¹	1290-1947	934-1189	165-219	62-169	460-760	1113-1476	36-44	85-166	647-1004	5797-9048	73-103	32-45
	Eroding	Pearson r Phase vs depth	0.68*	0.39	0.32	-0.31	0.72*	-0.41	0.52	0.93*	0.49	0.46	0.8*	0.65*
		Pearson r Phase vs. SOC	0.57	0.71*	0.58	0.38	-0.81*	0.39	-0.55	-0.98*	-0.61	-0.62*	-0.8*	-0.72*
		Range mg kg ⁻¹	1117-1954	681-1599	142-291	89-216	463-688	812-1050	35-56	69-124	753-1064	7027-10764	83-136	34-40
	Depositional footslope	Pearson r Phase vs depth	0.2	0.37	0.34	0.43	0.27	-0.38	0.22	0.52	0.2	0.52	0.61	-0.32
		Pearson r Phase vs. SOC	-0.27	-0.25	-0.09	0	-0.22	0.4	0.16	-0.54	-0.24	-0.49	-0.21	0.37
		Range mg kg ⁻¹	1137-2685	849-2021	132-356	58-221	286-529	640-945	32-50	39-66	557-789	5017-7972	78-114	29-33

1 Table 4: Correlations (Pearson r) between SOC and the extracted [AS](#) Galactosamine (GalN)
2 and Glucosamine (GlcN) from different fractions. (* = p < 0.1).

	Bulk soil	Macroaggregate	Microaggregate	Non-aggregated s+c
Pearson r GalN-C	0.97*	0.87*	0.94*	0.92*
Pearson r GlcN-C	0.71*	0.76*	0.96*	0.87*
N Observations GalN-C	31	16	19	18
N Observations GlcN-C	31	14	16	20
Range GalN-C $\mu\text{g g}^{-1}$ soil	12-325	33-360	4-248	2-206
Range GlcN-C $\mu\text{g g}^{-1}$ soil	26-860	60-887	23-546	4-403
Mean GalN-C $\mu\text{g g}^{-1}$ soil	296±280	172±93	100±82	69±65
Mean GlcN-C $\mu\text{g g}^{-1}$ soil	555±679	396±224	228±183	138±130

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1 | Table 5: Conventional radiocarbon ages for investigated fractions for different depths at the
2 | depositional site.

Fraction	Depth (cm)	¹⁴ C age (years BP)
Macroaggregates (>250µm)	0-15	975±15
	35-50	2275±20
	55-70	3090±20
Microaggregates (<250-53µm)	0-15	680±15
	35-50	1680±15
	55-70	2670±20
Non-aggregated silt and clay (<53µm)	0-15	635±15
	35-50	1835±15
	55-70	2200±15

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9. Figures

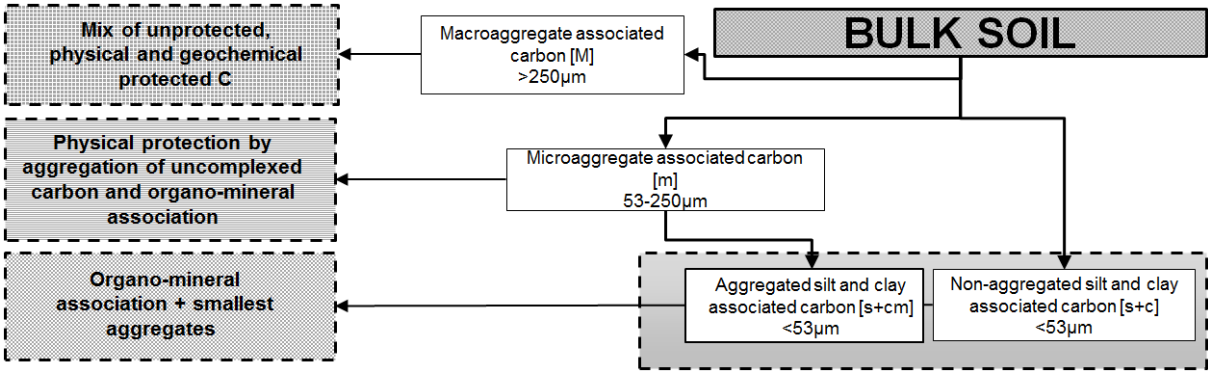


Figure 1: The resulting fractions of the applied fractionation scheme and interpretation of the present carbon stabilization mechanisms in each fraction.

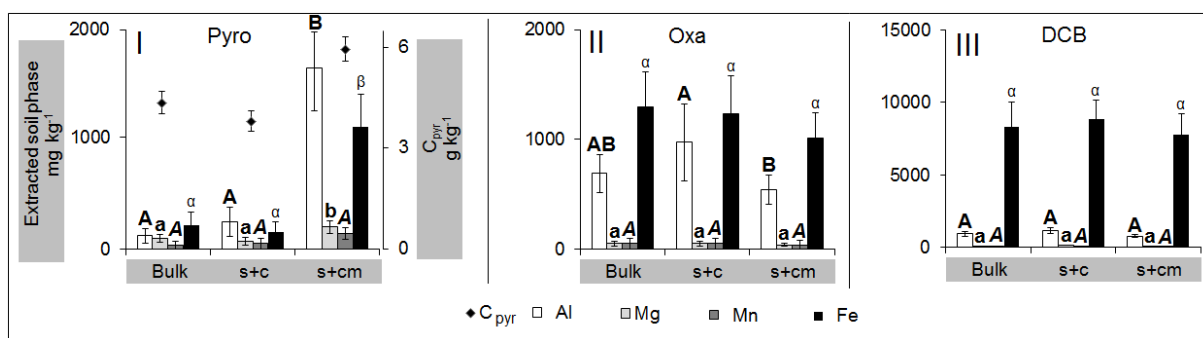


Figure 2: The distribution of pyrophosphate (I), oxalate (II) and dithionite (III) extractable Al, Mn and Fe between the Bulk soil and its compartments non-aggregated silt+clay (s+c) and aggregated silt+clay (s+cm) for all depths and positions combined (bars). Dots represent the the C extracted by pyrophosphate for the bulk soil and the fractions (Panel I only). Different letter above bars indicate a significant difference (p<0.1, tested for extracted element in the different SOC fractions and treatments separately as indicated by differences in font types).

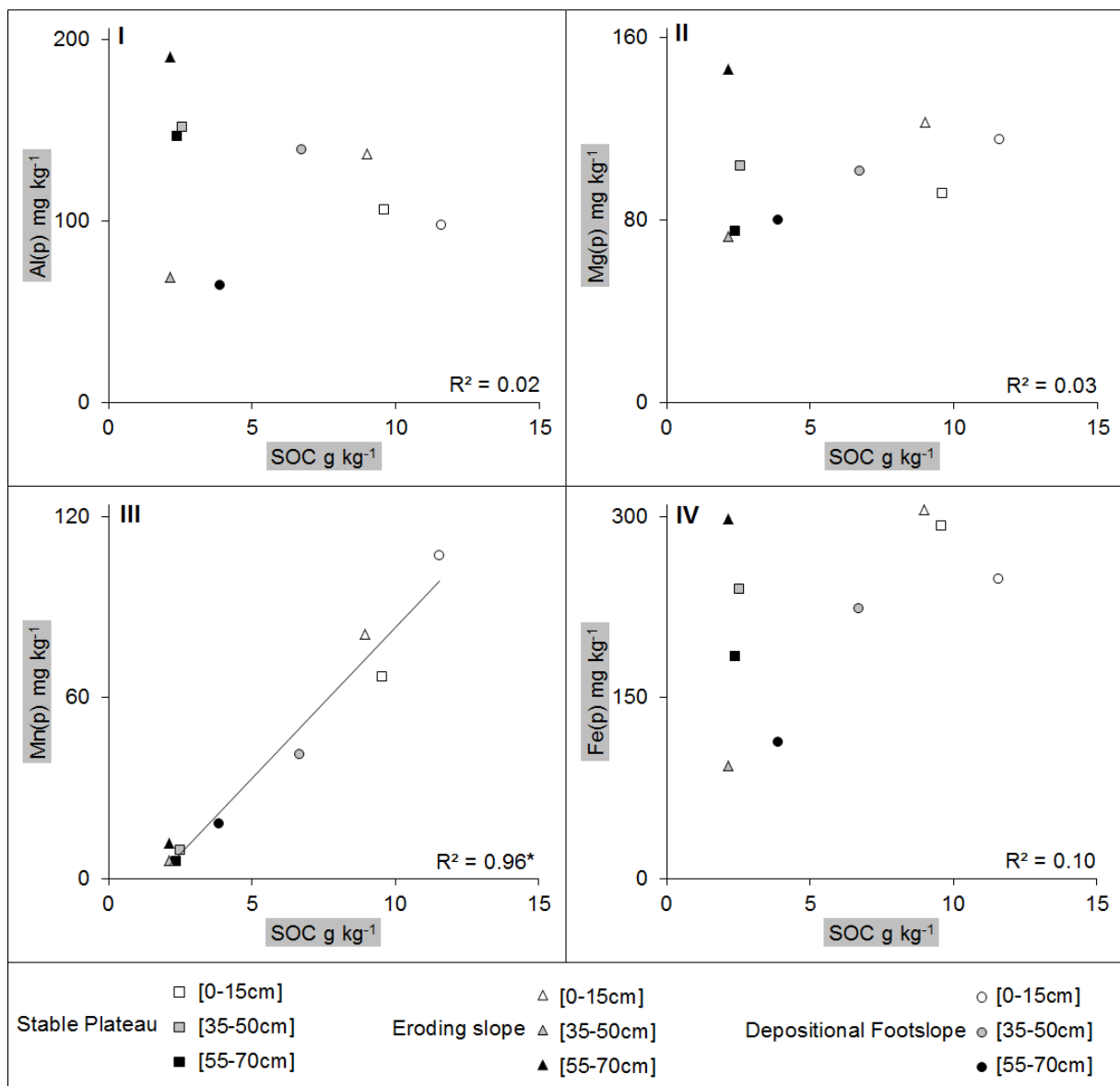


Figure 3: Mean SOC (g per kg soil) and pyrophosphate extractable Al (I), Mg (II), Mn (III) and Fe (IV).

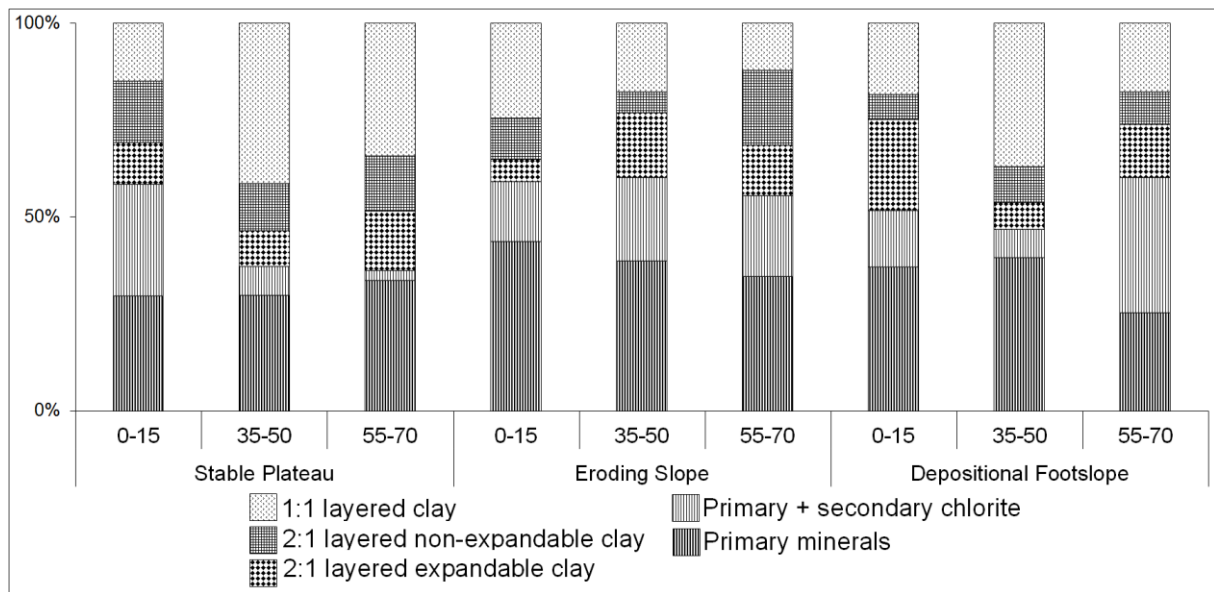


Figure 4: Composition of the mineral phase and quantification of primary vs secondary minerals for the different slope positions and soil depths of the clay sized fraction (no distinction between primary and secondary chlorite).

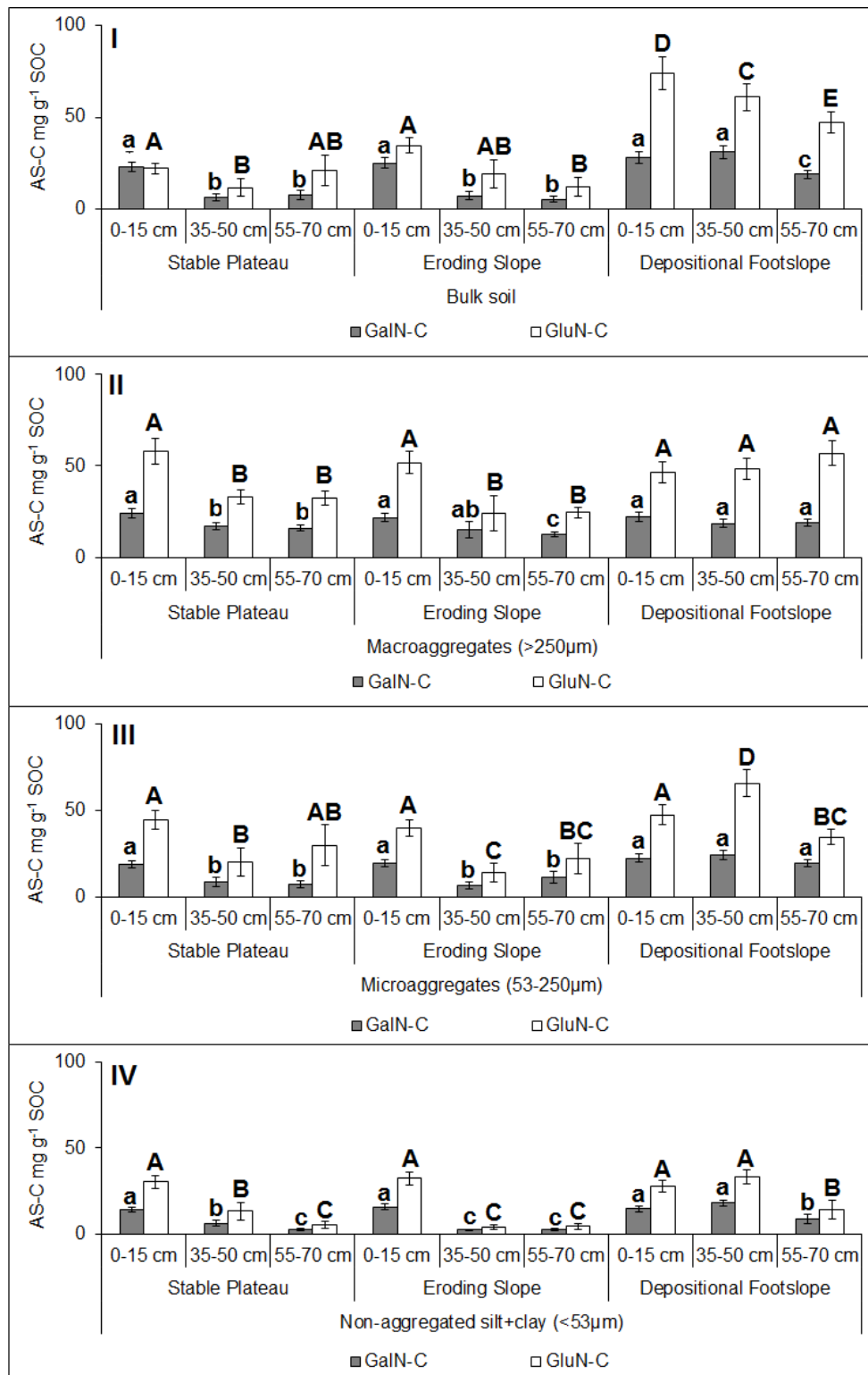


Figure 5: Amino sugars per unit SOC in the bulk soil (I) and the fractions (II-IV) along the slope and for different depths. Different letters above bars indicate a significant difference ($p < 0.1$, for the two different AS and the bulk soil and three fractions separately as indicated by differences in font types).

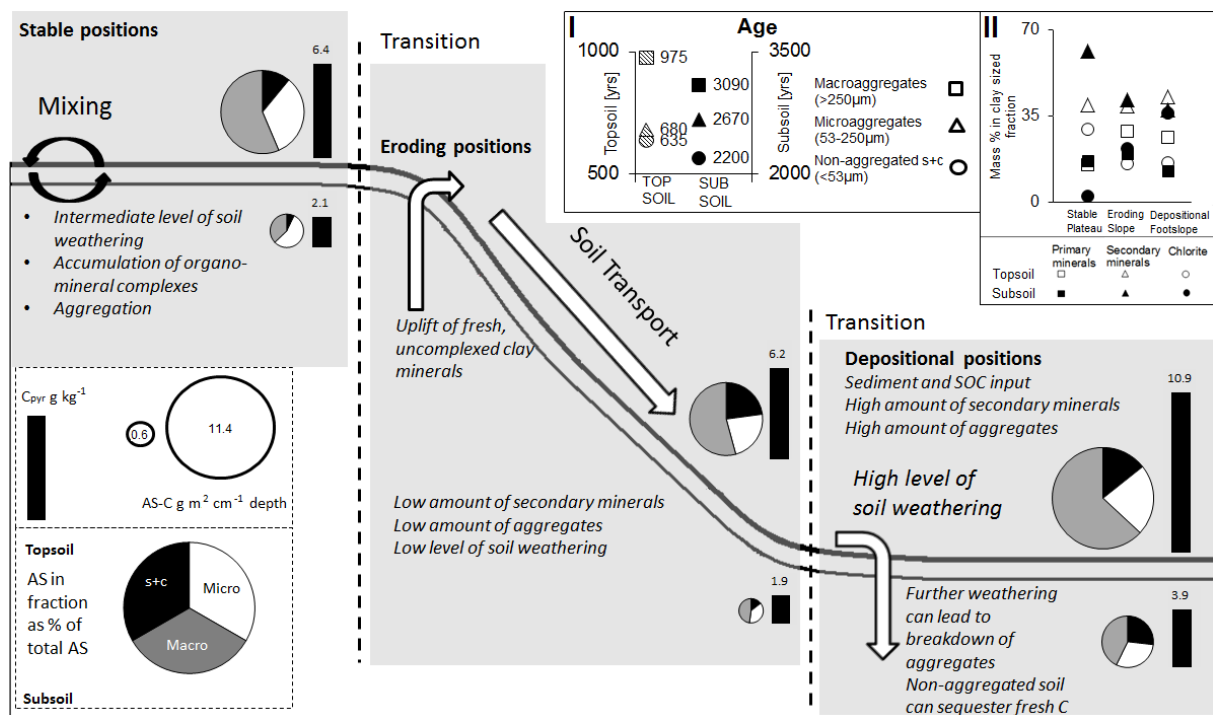


Figure 6: Conceptual figure showing the mineralogical changes along the slope in relation to the abundance of AS in non-aggregated silt and clay (s+c), microaggregates (Micro) and Macroaggregates (Macro). Statement on the level of soil weathering along the transect refers to both top and subsoil. Pie charts show the relative contribution of AS in each fraction to total AS by multiplying AS per g C in a fraction with C stock estimates of the respective fraction given in table 1. The size of the pie chart indicates the absolute AS content of these fractions combined in g per m² and per cm depth of a profile. The amount of C extracted with pyrophosphate in the bulk soil is given as a column next to each AS pie char. Figures on the top right show the distribution of primary and secondary minerals along the slope in different depths (I) (see also table 4) and the relative ¹⁴C derived ages of these fractions in top and subsoil for the depositional site (II), indicating differences between the different fractions (see also table 5).