Thank you very much for taking into consideration the reviewers' comments. I consider that your manuscript has been improved significantly. So we are very pleased to inform you that your manuscript has been accepted for publication in Biogeosciences. Before it is published, please consider my technical corrections below:

L88: a "surface" chl

We have corrected as suggested. (L88)

L89: Did the authors use Aqua/MODIS data? Otherwise, Terra or both?

We used Aqua/MODIS data. We have added the information at L89.

L174–175 plus Supporting Information L10: what does the meaning of 98"+" atom%? Did the authors assume 98 atom% 15N for the calculation of nitrogen fixation rates?

The gas provided from SI Science Co. is labeled 98+ atom%. We assume 98 atom% for the calculation. To avoid confusion, "+" has been deleted from the revised manuscript.

L210–217: What software(s) did the authors use for the nMDS and ANOSIM analyses?

The nMDS and ANOSIM analyses were performed using PRIMER 6 software. We have added this sentence at L218.

L275: phosphate "concentrations" … "were" …

We have corrected as suggested. (L276)

L286: dissolved iron "concentrations" … "were" …

We have corrected as suggested. (L287)

L341: Remove the hyphen between "spp." and "bloom".

We have removed as suggested.

L381: Remove a space between "that" and "in".

We have removed as suggested.

L454: Remove a comma immediately after "plot".

We have removed as suggested.

L564: Start a new line from "Jickells, T.D…".

We have changed as suggested.

Supporting Information L60: Remove a comma immediately after "2015".

We have removed as suggested.

Legend in Fig. S3: KT-07-21

We have corrected as suggested.

Why is *Trichodesmium* abundant in the Kuroshio?

T. Shiozaki[1,2], S. Takeda[1,3], S. Itoh[2], T. Kodama[1,4], X. Liu[1,5], F. Hashihama[6], K. Furuya[1]

[1]{Department of Aquatic Bioscience, Graduate School of Agricultural and Life Sciences, The University of Tokyo, Tokyo, 113-8657, Japan}

[2]{Atmosphere and Ocean Research Institute, The University of Tokyo, Chiba, 277-8564, Japan}

[3]{Faculty of Fisheries, Nagasaki University, Nagasaki, 852-8521, Japan}

[4]{Japan Sea National Fisheries Research Institute, Fisheries Research Agency, Niigata, 951-8121, Japan}

[5]{College of Ocean and Earth Sciences, Xiamen University, Xiamen, 361005, China}

[6]{Department of Ocean Sciences, Tokyo University of Marine Science and Technology, Tokyo, 108-8477, Japan}

Corresponding to: T. Shiozaki (shiozaki@aori.u-tokyo.ac.jp)

Abstract

The genus *Trichodesmium* is recognized as an abundant and major diazotroph in the Kuroshio, but the reason for this remains unclear. The present study investigated the abundance of *Trichodesmium* spp. and nitrogen fixation together with concentrations of dissolved iron and phosphate in the Kuroshio and its marginal seas. We performed the observations near the Miyako Islands, which form part of the Ryukyu Islands, situated along the Kuroshio, since our satellite analysis suggested that material transport could occur from the islands to the Kuroshio. *Trichodesmium* spp. bloomed (>20,000 filaments L^{-1}) near the Miyako Islands, abundance was high in the Kuroshio and the Kuroshio bifurcation region of the East China Sea, but was low in the Philippine Sea. The abundance of *Trichodesmium* spp. was significantly correlated with the total nitrogen fixation activity. The surface concentrations of dissolved iron (0.19–0.89 nM) and phosphate (<3–36 nM) were similar for all of the study areas, indicating that the nutrient distribution could not explain the spatial differences in *Trichodesmium* spp. abundance and nitrogen fixation. Numerical particle-tracking experiments simulated the transportation of water around the Ryukyu Islands to the Kuroshio. Our results indicate that *Trichodesmium* growing around the Ryukyu Islands could be advected into the Kuroshio.

1. Introduction

The Kuroshio is a western boundary current in the North Pacific Ocean that originates in the North Equatorial Current and bifurcates to the east of the Philippines. The main stream of the Kuroshio enters the East China Sea (ECS) northeast of Taiwan, flows out through the Tokara Strait, and runs along the Japanese islands of Shikoku and Honshu. While the Kuroshio and its adjacent waters are characterized by highly oligotrophic conditions, phytoplankton and zooplankton communities in the Kuroshio are distinct compared to those from adjacent waters (McGowan, 1971). McGowan (1971) suggested that some plankton species are delivered by the Kuroshio to the north from the equatorial region.

The abundance of the cyanobacterial genus *Trichodesmium* in the Kuroshio is much higher than that in neighboring seas (Marumo and Asaoka, 1974). Because *Trichodesmium* is a major nitrogen fixer in the Kuroshio, it is believed to be the key genus for understanding the Kuroshio ecosystem (Chen et al., 2008, 2014; Shiozaki et al., 2014a). Nevertheless, the factors controlling the distribution of *Trichodesmium* in this region are poorly understood. Marine nitrogen fixation is thought to be regulated by the supply of iron and phosphorus (Mahaffey et al., 2005), and *Trichodesmium* thrives in iron-rich oligotrophic regions (Moore et al., 2009; Shiozaki et al., 2010,

2014b). A major source of iron in the ocean is atmospheric dust deposition (Jickells et al., 2005; Mahowald et al., 2009). Modeling studies indicate that dust deposition in the western North Pacific decreases exponentially from the continental shelf to the Philippine Sea (Jickells et al., 2005; Mahowald et al., 2009), and hence, deposition is not as high in the Kuroshio as in the adjacent waters. As for phosphorus limitation, iron-enhanced nitrogen fixation causes phosphorus depletion, and the nitrogen fixation is consequently limited by phosphorus (Mather et al., 2008). The phosphate distribution has been examined in this study region using a conventional colorimetric method, and the surface phosphate concentration in the Kuroshio has been reported to be as low as that in the Philippine Sea (Chen, 2008). Therefore, the distinct high abundance of *Trichodesmium* in the Kuroshio is probably not explained by nutrient and trace metal concentrations; however, distributions of dissolved iron and phosphate at the nanomolar level have not been well studied in this region (Obata et al., 1997; Shiozaki et al., 2010; Kodama et al., 2011).

Nitrogen fixation by *Trichodesmium* has recently also been found to be active around oceanic islands; New Caledonia Island, Efate Island, Fiji Island, Tahiti Island, and Northern Mariana Islands (Shiozaki et al., 2010, 2013, 2014c; Lin et al., 2011). Furthermore, these studies demonstrated that abundant *Trichodesmium* is delivered by

72 the current to areas that are remote from the islands. Although this phenomenon was
73 noted in the western Pacific warm pool and western South Pacific, it can also occur in
74 and around the Kuroshio and may contribute to the distribution of *Trichodesmium* in
75 this region.

76 In the present study, we simultaneously determined *Trichodesmium* abundance
77 and bulk water nitrogen fixation together with concentrations of dissolved iron and
78 phosphate at the nanomolar level in the Kuroshio and its marginal seas. In addition,
79 we conducted intensive observations around the Miyako Islands section of the
80 Ryukyu Islands located close to the main stream of the Kuroshio.

81

82 2. Materials and Methods

83 2.1. Oceanographic database

84 Algal blooms in an oligotrophic region may indicate a nitrogen fixation hotspot
85 (Wilson and Qiu, 2008; Shiozaki et al., 2014c). To identify the locations of intensive
86 algal blooms, we used a dataset of chlorophyll (chl) *a* observed by satellite. According
87 to Wilson and Qiu (2008), an algal bloom in an oligotrophic region can be defined as
88 a surface chl *a* value >0.15 mg m^{-3} in summer. In the present study, we used 8-day,
89 moderate-resolution imaging spectroradiometer (MODIS) Aqua level 3 chl *a* with 9

90 km resolution during summer between July 2003 and September 2009. We defined
91 summer as July through September. The bloom frequency for each pixel was
92 calculated from the ratio of counts in which chl *a* was >0.15 mg m^{-3} to the total
93 counts in which chl *a* was detected.
94 To examine the current field, geoelectrokinetograph and ship-mounted acoustic
95 Doppler current profiler (ADCP) data from the uppermost layer for the summers
96 between 1953 and 2008 were obtained from the Japan Oceanographic Data Center
97 (http://www.jodc.go.jp). Regridding, removal of anomalous values, and smoothing of
98 the dataset were performed as described by Isobe (2008).
99

100 ## 2.2. **Cruise observations**

101 Experiments were conducted during summer on-board the R/V *Tansei-maru*
102 (KT-06-21, September 9–17, 2006; KT-07-22, September 5–13, 2007; KT-09-17,
103 September 8–13, 2009; KT-10-19, September 4–12, 2010) and the T/V
104 *Nagasaki-maru* (242, July 19–28, 2007) (Fig. 1a, Table S1). The stations during the
105 KT-06-21, KT-07-22, and *Nagasaki-maru* 242 cruises were divided into three areas
106 based on the temperature-salinity diagram (see Fig.2 of Shiozaki et al., 2011): the
107 ECS, Kuroshio, and Philippine Sea. During the KT-09-17 cruise, we conducted

experiments around the Miyako Islands which were distinguished from the other three areas. During the KT-10-19 cruise, we performed observations in the ECS, the Kuroshio, and around the Miyako Islands (Liu et al., 2013).

2.2.1. **Light intensity, hydrography, nutrients, and chl *a***

Water samples for all of the experiments, with the exception of determination of the dissolved iron concentration, were collected using an acid-cleaned bucket and Niskin-X bottles. The depth profile of light intensity was determined immediately before the water sampling using a light sensor (during the KT-06-22, KT-07-21, KT-09-17, and KT-10-19 cruises) or an empirical equation (during the *Nagasaki-maru* 242 cruise) (Shiozaki et al., 2011). Temperature and salinity profiles to a depth of 200 m were obtained using a conductivity, temperature, and depth (CTD) sensor. Mixed layer depth (MLD) was defined as the depth at which the sigma-t increased by 0.125 from its value at a depth of 10 m. Water samples for nitrate+nitrite (N+N) and phosphate were collected from 0, 10, 30, 40, 50, 60, 70, 80, 90, 100, 125, 150, and 200 m, and from depths at given light intensities. At all of the stations, the N+N and phosphate concentrations were determined at the nanomolar level using a supersensitive colorimetric system consisting of an AutoAnalyzer II (Technicon) and

126 Liquid Waveguide Capillary Cells (World Precision Instruments, USA) (Hashihama et
127 al., 2009). The detection limits of N+N and phosphate were both 3 nM. When the
128 concentration was greater than 0.1 μM, it was determined by conventional methods
129 using a TRAACS 2000 autoanalyzer (Bran:Luebbe, UK). In addition to the
130 observations at the stations, temperature, salinity, and the *in vivo* chl fluorescence of
131 the surface water were monitored continuously during the cruises by a
132 thermosalinograph (Ocean Seven, Idronaut, Italy) and a fluorometer (Minitracka,
133 Chelsea, UK).
134
135 ### 2.2.2. **Dissolved iron**
136 Water was sampled to estimate the dissolved iron concentration from 0.5-m depth
137 during the KT-06-21 and KT-07-22 cruises and from 10-m depth during the KT-09-17
138 cruise using an acid-cleaned Teflon bellows pump (AstiPure PFD2; Saint-Gobain)
139 with Teflon tubing (inner diameter = 12 mm). The water was filtered through an
140 acid-cleaned 0.22 μm pore filter (Millipak100; Millipore) connected to the in-line of
141 the Teflon tubing with a Teflon connector. Filtered seawater was collected in a 125
142 mL low-density polyethylene (LDPE) bottle (Nalgene, Nalge Nunc International),
143 which had been washed using following technique: the sample bottles were

144 sequentially cleaned by soaking in 5% alkali detergent for at least 2 days, in 4 N HCl
145 for at least 1 day, in 0.3 N metal analysis-grade HNO_3 at 60°C overnight, and finally,
146 in Milli-Q water at 60°C overnight. After rinsing with Milli-Q water, the bottles were
147 dried in a laminar flow space and stored in double plastic bags. The filtrate samples
148 were acidified to a pH <1.7 with trace-metal-grade HCl (Tamapure AA-100; Tama
149 Chemicals) in a Class-100 clean-air bench, and stored at room temperature for more
150 than 1 year.

151 The dissolved iron concentration was determined using an automatic Fe(III) flow
152 injection analytical system (Kimoto Electric Co., Ltd.) using a chelating resin
153 pre-concentration and chemiluminescence detection method (Obata et al., 1993). A
154 buffer solution of 10 M formic acid and 2.4 M ammonium formate was added to the
155 samples. The sample pH was adjusted to 3.0 with 20% ammonium hydroxide
156 (NH_4OH; Tamapure AA-10; Tama Chemicals) immediately prior to analysis. The
157 detection limit of this method was 0.05 nM. The SAFe reference standards S1 and D2
158 were measured during the course of sample analysis, and the results were within the
159 range of the published consensus values: S1 = 0.097 ± 0.043 nM and D2 = 0.91 ±
160 0.17 nM (Johnson et al., 2007).

161

2.2.3. **Nitrogen fixation and abundance of *Trichodesmium* spp.**

Samples for the incubation experiments were collected vertically at all of the stations, except at Sts. T0621, GN-3, and T0905, where samples were only collected from the surface. All samples were collected in duplicate in acid-cleaned 4.5-L polycarbonate bottles. During the *Nagasaki-maru* 242 cruise, water samples were collected from four different depths corresponding to 100%, 25%, 10%, and 1% of the surface light intensity. During the other cruises, samples were collected from a depth of 50% surface light intensity. Samples at 100% surface light intensity were collected from 0 m during all of the cruises, except during the KT-10-19 cruise in which the samples were collected from a depth of 5 m. The bulk water nitrogen fixation activity was determined based on primary production using a dual isotopic ($^{15}N_2$ and ^{13}C) technique (Shiozaki et al., 2009). After ^{13}C-labeled sodium bicarbonate (99 atom% ^{13}C; Cambridge Isotope Laboratories) was added to each bottle, 2 mL of $^{15}N_2$ gas (98 ~~±~~ atom% ^{15}N; SI Science Co. Japan) was injected directly into the incubation bottles through a septum using a gastight syringe. The bottles were covered with neutral-density screens to adjust the light level and incubated for 24 h in an on-deck incubator cooled by flowing surface seawater for 24 h. We determined the nitrogen fixation activity using the $^{15}N_2$ gas bubble addition method (Montoya et al., 1996).

This method is believed to underestimate the nitrogen fixation rate relative to the $^{15}N_2$ gas dissolution method (Mohr et al., 2010). The start time of incubation in this study varied at each station (Table S1). Considering daily periodicity of nitrogen fixation in each diazotroph (Zehr, 2011) and the time to reach equilibration of the $^{15}N_2$ gas bubble with seawater (>12 h, Mohr et al., 2010), the level of underestimation could vary at each station. Meanwhile, the level of underestimation is thought to be low in *Trichodesmium* dominant water because *Trichodesmium* can float to the top of the bottle and directly use the added $^{15}N_2$ in the bubble method (Großkopf et al., 2012). Although the bias of underestimation could not be estimated from the results in this study, the actual nitrogen fixation rate could be higher than the obtained rate.

A recent study demonstrated that commercial $^{15}N_2$ gas could be contaminated by ^{15}N-labeled nitrate and ammonium (Dabundo et al., 2014). We tested the contamination in $^{15}N_2$ gas produced by SI Science Co., Ltd., which was used (from different batch numbers) in the present study (see Supporting Information). Briefly, the $^{15}N_2$ gas was dissolved in aged subtropical surface water, and concentrations of nitrate, nitrite, and ammonium at the nanomolar levels were determined using supersensitive colorimetric systems. The results showed that there were no significant differences between the control and samples to which $^{15}N_2$ had been added (Fig. S1),

198 suggesting that the contamination of nitrate, nitrite, and ammonium in the $^{15}N_2$ gas
199 was insignificant (Supporting Information).

200 Water samples were collected for microscopic analysis at all light depths during
201 the *Nagasaki-maru* 242 and KT-07-21 cruises, and only from the surface during the
202 KT-06-22, KT-09-17, and KT-10-19 cruises. The samples were fixed using acidified
203 Lugol's solution. *Trichodesmium* spp. were counted using the Utermöhl method under
204 inverted microscope observation. *Trichodesmium* greater than ca. 300 μm in length
205 were counted as 1 filament and shorter lengths were counted as 0.5 filaments. In
206 addition, phytoplankton other than *Trichodesmium* spp. were identified from the
207 samples obtained during the KT-09-17 cruise.

208

209 ### 2.3. **Statistical analysis of environmental variables**

210 We used non-metric multi-dimensional scaling (nMDS) to investigate the spatial
211 differences in the environmental variables that could influence *Trichodesmium* growth
212 and bulk water nitrogen fixation; temperature, mixed layer depth, nitrate, dissolved
213 iron, and phosphate. The environmental variables were transformed by $\log_{10}(x + 1)$
214 prior to analysis. A dissimilarity/similarity matrix between stations was constructed
215 using the Bray-Curtis index. The nMDS was used to visualize similarities in the

environmental variables among the stations. An Analysis of Similarity (ANOSIM) was used to test the differences in the environmental variables among the stations. The nMDS and ANOSIM analyses were performed using PRIMER 6 software.

2.4. **Numerical experiments**

Numerical particle-tracking experiments were conducted to investigate the transport of water masses at the surface from areas around the Miyako Islands in the summer season from 2003 to 2009. Surface velocity data were derived from the FRA-JCOPE2 reanalysis product (Miyazawa et al., 2009), which is an eddy-resolving (1/12°) ocean model combined with three-dimensional variational data assimilation (satellites, ARGO floats, and shipboard observations), and is one of the most reliable models for the region around Japan for the above time period. The method of tracking particles was basically the same as in Itoh et al. (2009), but we did not include the random walk for simplicity. The release points of particles were selected at the surface of the model grid points around the coastal waters of the Miyako Islands. We assumed that the particles did not increase, die, or sink from the surface during the experiments. To focus on transport during the summer season (July–September), particles were released one month before the summer (June 1) and were tracked until September 30.

234 To examine differences in the output depending on the start time within the same
235 year, we also performed experiments starting on June 1, 11, and 21, and July 1 in
236 2009. The ratio of particles that reached areas downstream of the Tokara Strait
237 (hereafter Area K) (Fig. 7), including the particles' entrainment to the Kuroshio, to
238 total particles released from the Miyako Islands was computed in all experiments. It
239 should be noted that these experiments contained the following two uncertainties.
240 First, the distribution of *Trichodesmium* around the islands, which strongly influences
241 the destinations of particles, was not able to be determined in advance.
242 *Trichodesmium* is known to aggregate and not to occur uniformly in the ocean
243 (Capone et al., 1997). Second, the model cannot reproduce the current very close to
244 the islands. If a water mass very near the islands was delivered to the open ocean by
245 tide and/or river plumes that were not considered in the model, seaward dispersion of
246 particles was likely underestimated.
247

248 ## 3. RESULTS

249 ### 3.1. The Kuroshio path and bloom frequency

250 The average surface current field indicated that the main stream of the Kuroshio
251 flowed along the continental shelf in the ECS, and then passed to the south of the

252 Kyushu and Shikoku Islands (Fig. 1b). In addition, the Kuroshio branch bifurcated
253 northward at 25°N and 30°N at the continental shelf. Hence, all of the stations in the
254 ECS were subject to the influence of the Kuroshio. While the northeastward stream of
255 the Kuroshio was prominent in this region, smaller-scale flows and circulations were
256 observed in the areas around and to the southeast of the Ryukyu Islands. In the west of
257 the main stream of the Kuroshio, because the average chl *a* was over 0.15 mg m^{-3} (Fig.
258 S2), the frequency of chl *a* values >0.15 mg m^{-3} was high (Fig. 1b). In contrast, the
259 bloom frequency in the east of the main stream of the Kuroshio differed from the
260 distribution of the average chl *a*; algal blooms occurred frequently in the Ryukyu
261 Islands. Around the Miyako Islands, water of high bloom frequency was located to the
262 west of the islands, extending to the north.
263
264 ### 3.2. Region-wide environmental conditions, *Trichodesmium* spp., and
265 nitrogen fixation
266 The sea surface temperature (SST) ranged from 25.1–30.5°C at all of the stations
267 (Table S1), and there were no significant differences among the areas (p>0.05,
268 Tukey's honestly significant difference [HSD] test). The MLD varied from 12–60 m
269 at all of the stations, and was relatively deep around the Miyako Islands compared to

the other areas (Table S1). The surface N+N concentration varied between <3 and 42 nM, except around the Miyako Islands (Shiozaki et al., 2010, 2011) (Table S1). The highest surface N+N concentration (374 nM) was observed at St. T0904 where upwelling occurred (see below). No significant difference in the surface N+N was observed among the four areas (p>0.05, Tukey's HSD test). The surface phosphate concentration varied between <3 and 36 nM at all of the stations (Fig. 2a). The phosphate concentrations at the surface and within the MLD ~~was~~ were not significantly different among the four areas (p>0.05, Tukey's HSD test). There was a greater increase in the phosphate concentrations below 40–50 m in the ECS compared to the other areas (Fig. 3a–d). Furthermore, the phosphate concentrations below 40–50 m near the Miyako Islands were higher than those in the Kuroshio and the Philippine Sea, which were depleted down to 100 m, except at St. T1004 located near the continental shelf. The N/P (= N+N/phosphate) ratio at the surface varied from 0.28 to 6.40 except at St. T0904 (N/P = 16.3) (Table S1), and no significant differences were observed among the four areas (p > 0.05, Tukey's HSD test). The surface dissolved iron concentration ranged from 0.19 to 0.89 nM at all of the stations (Fig 2b), with no significant spatial differences among the four areas (p>0.05, Tukey's HSD test). The surface dissolved iron concentrations at Sts. T0622 and T0907 ~~was~~ were elevated to

288 0.83 nM and 0.89 nM, respectively, with lower salinity water than in the adjacent
289 waters (salinity data are shown in Fig. 4a and Kodama et al., 2011). The nMDS
290 showed that the environmental variables at all stations were the same at the >80%
291 similarity level and were >90 % similar excepting station T0904 (Fig. 5). The
292 ANOSIM indicated no significant differences among the stations ($p > 0.05$).
293 The abundance of *Trichodesmium* spp. was highest at the surface at almost all of
294 the stations during the *Nagasaki-maru* 242 and KT-07-21 cruises (Fig. S3). The
295 surface *Trichodesmium* spp. abundances were positively correlated with the
296 depth-integrated abundances ($r^2 = 0.51$, $p < 0.05$) (Fig. 6a). Thus, the surface
297 abundance was used to discuss the geographical distribution of *Trichodesmium* spp.
298 The *Trichodesmium* spp. abundance at the surface varied widely, and there was no
299 significant difference among the four areas ($p > 0.05$, Tukey's HSD test).
300 *Trichodesmium* spp. were observed at all of the stations in the Kuroshio and around
301 the Miyako Islands, whereas they were not always observed in the ECS and the
302 Philippine Sea (Fig. 2c). The average surface abundance in the Philippine Sea was the
303 lowest among all of the areas (Table 1). The highest abundance of *Trichodesmium* spp.
304 (>20000 filaments L^{-1}) was observed near the Miyako Islands at St. T0906, where
305 they bloomed (see below). Tuft-shaped colonies were found at Sts. T0706, T0723,

306 CK-10, and T0906. The nitrogen fixation rate was highest in the upper 25% light
307 depth, and decreased with increasing depth at all of the stations (Fig. 3e–h). The
308 surface rates were positively correlated with the depth-integrated rates ($r^2 = 0.79$, $p <$
309 0.05) (Fig. 6b), suggesting that the distribution of nitrogen fixation was indexed by
310 the surface activity. Surface and depth-integrated nitrogen fixation ranged from 0.54
311 to 62 nmol N L^{-1} d^{-1} and from 29.5 to 753 μmol N m^{-2} d^{-1}, respectively (Fig. 2d and
312 Table S1). Surface nitrogen fixation in the Philippine Sea was significantly lower than
313 that in the Kuroshio ($p < 0.05$, *t*-test).

314 The surface abundance of *Trichodesmium* spp. in the entire study area was
315 positively correlated with the nitrogen fixation rate at the surface ($r^2 = 0.80$; $p < 0.05$
316 [$r^2 = 0.52$; $p < 0.05$ if the datum taken at the *Trichodesmium*-bloom station T0906 is
317 excluded]) (Fig. 6c), suggesting that they significantly contributed to nitrogen fixation
318 in the study region. However, active nitrogen fixation occurred in the ECS where
319 *Trichodesmium* abundance was low, and hence, the other diazotrophs could also be
320 important for nitrogen fixation.

321

322 ### 3.3. **Observation around the Miyako Islands during the KT-09-17 cruise**

323 The SST was lower to the northwest of the Miyako Islands than in adjacent

waters, and chl *a* was enriched in the same location (Fig. 4b,c). Therefore, the enhanced productivity was probably due to nutrient supply by upwelling. This upwelling generally occurs in the lee of islands (Hasegawa et al., 2009), suggesting that there was a northward current during the cruise. The surface salinity was lower east of the Miyako Islands than in the surrounding waters (Fig. 4a). The absence of any large river on the east side of Miyako-jima Island and the separation of low salinity water from the island suggest that the low salinity was caused by rainfall.

St. T0904 was located near the upwelling water; its SST of 29.0°C was lowest and its surface N+N concentration of 374 nM was highest among all of the stations. However, the N+N concentration at St. T0904 at the surface was higher than that at the subsurface (an approximate depth of 50 m; Fig. S4), indicating that St. T0904 was not located in the middle of the upwelling. At St. T0904, the surface phosphate concentration was also highest (23 nM) and the N/P ratio (=16.3) was higher than the Redfield ratio. With the exception of the surface at St. T0904, the phosphate concentration was low (<3–9 nM) in the upper 50 m, with no noticeable variation among the stations (Fig. 2a). The dissolved iron concentration varied between 0.19 and 0.89 nM at the surface (Fig. 2b). The highest dissolved iron concentration was observed at St. T0907.

During the same cruise, we encountered a *Trichodesmium* spp. bloom at St. T0906 (Fig. 2c), which had colored water at the surface. The abundance of *Trichodesmium* spp. at St. T0906 was >20,000 filaments L^{-1}, which was far higher than that at other stations (2–102 filament L^{-1}). The nitrogen fixation rate at the surface (61.9 nmol N L^{-1} d^{-1}) of this station was more than 30-fold that just below the surface, and was the highest among all of the stations (Fig. 3h). The diatom abundance was markedly higher at St. T0904 than that at the other stations. *Cylindrotheca closterium* was the most numerically dominant diatom (59%), followed by *Navicula* spp. (23%) and *Nitzschia* spp. (13%). *C. closterium* was not detected at the other stations, indicating that the high chl *a* induced by the island wake effect mainly consisted of diatoms.

3.4. Numerical simulation

As the Kuroshio generally flows along the continental slope north of the Miyako Islands (Fig. 1b), particles around the Miyako Islands were not transported along the typical path of the Kuroshio to the northeast, especially at their initial stages (Fig. 7a). Some particles migrated around the Miyako Islands, or turned south after they passed the Tokara Strait. Nevertheless, the particles delivered to Area K east of the Tokara

Strait increased as time elapsed, and the ratio of particles delivered to Area K to the total released particles ranged from 13–56% (30 ± 16%) by day 120 in 2003–2009 (Fig. 7b). The year-to-year variations in the ratio are mainly due to influences of mesoscale eddies as partly seen in the particle trajectories in Fig. 7a, and likely occurred over relatively short time scales (shorter than the seasonal time scale). This is supported by another series of experiments in which particles were released on June 1, 11, and 21, and July 1 in 2009, which yielded ratios of 6.2–38% (22 ± 13%) by day 120 (Fig. S5).

4. DISCUSSION

4.1. Distribution of phosphate and dissolved iron concentrations

Phosphate concentrations were consistently low within the MLD in all of the studied areas, and the maximum abundance of *Trichodesmium* spp. and total nitrogen fixation activity generally occurred near the surface, suggesting that the phosphate conditions for surface *Trichodesmium* spp. and other diazotrophs were similar among all of the areas. Furthermore, with the exception of St. T1004 located near the continental shelf, the vertical distribution of phosphate in the Kuroshio was analogous to that in the Philippine Sea. Therefore, at least in the oceanic region of the two areas,

phosphate availability for *Trichodesmium* spp. and the other diazotrophs was similar throughout the water column.

The surface distribution of the dissolved iron concentration demonstrated no significant variation among the areas. The dissolved iron concentration (0.19–0.89 nM) was higher than that in the western North Pacific subtropical region (0.15–0.4 nM) (Brown et al., 2005). Obata et al. (1997) demonstrated that the vertical distribution of the dissolved iron concentration in the ECS showed two peaks (at the surface and in the deep water), suggesting that aerial dust significantly contributes to the high dissolved iron concentration at the surface in all of our study areas. In accordance with our results, previous modeling studies estimated the amount of dust deposition to be similar in all four areas (Jickells et al., 2005; Mahowald et al., 2009). Therefore, iron availability for *Trichodesmium* spp. and the other diazotrophs was also likely similar across all of the study areas. Iron can be supplied from deep water to the surface by mixing processes (Johnson et al., 1999). However, if this were the case, the nitrate concentration would be expected to increase simultaneously at the surface (Johnson et al., 1999), and we observed no noticeable elevation in N+N in any of the areas, except at St. T0904. High concentrations of dissolved iron (>0.8 nM) corresponded with low salinity at Sts. T0622 and T0907, suggesting that wet

deposition was an important process for iron supply. Dry deposition could also be important since the iron-enriched water at Sts. T0601 and T0715 did not correspond with low salinity.

Satellite data analysis indicated that there was a “pipeline” of material transport from the Miyako Islands to the Kuroshio, and this was supported by numerical simulations. According to the hypothesis of Marumo and Asaoka (1974), the growth of *Trichodesmium* in the Kuroshio could be maintained by the supply of iron and phosphorus from the islands situated along the Kuroshio, and the Miyako Islands were considered a possible nutrient source to the Kuroshio. Hence, assuming this hypothesis to be valid, the iron and phosphate concentrations near the Miyako Islands (especially in our observed area) would be expected to be higher than those in the other areas. However, we observed no significant difference in the iron and phosphate concentrations among the four areas. This suggested that there was no detectable washout of iron and phosphorus from the Miyako Islands during our observations, or that diazotrophs and other phytoplankton exhausted the nutrient supply close to the islands.

4.2. Factors controlling the distributions of *Trichodesmium* spp. and

nitrogen fixation

Although there was no statistically significant difference in *Trichodesmium* spp. abundance among the study areas probably because the data were limited and the variation was large, *Trichodesmium* spp. were always observed in the Kuroshio and were abundant at most stations. Furthermore, at St.CK-10 in the East China Sea which is in the Kuroshio branch current, a high abundance of *Trichodesmium* spp. was observed. On the other hand, *Trichodesmium* spp. abundance in the Philippine Sea tended to be lower than in the other areas. Such *Trichodesmium* distribution was also reported in the previous study (Marumo and Asaoka, 1974). The present study also showed lower surface nitrogen fixation in the Philippine Sea compared to that in the Kuroshio ($p < 0.05$, *t*-test). Previous studies demonstrated that *Trichodesmium* spp. flourished in some regions of the subtropical ocean where the iron levels were high (Moore et al., 2009; Shiozaki et al., 2014b), which can be attributed to the high iron requirement of *Trichodesmium* spp. for their growth compared to other diazotrophs and non-diazotrophs (Kustka et al., 2003; Saito et al., 2011). Therefore, the distribution of *Trichodesmium* spp. in the study area was expected to be associated with the dissolved iron concentration at the surface. Furthermore, the iron-enhanced active nitrogen fixation causes phosphorus depletion, and is consequently limited by

phosphorus (Mather et al., 2008). No significant differences in surface iron and phosphate were observed among the study areas, which cannot explain the distribution of *Trichodesmium* spp. and nitrogen fixation in the study region.

Johnson et al. (1999) reported that the iron supply increased around the continental shelf because re-suspension from the bottom to the euphotic zone becomes significant. However, in the continental shelf of the ECS, the abundance of *Trichodesmium* spp. and nitrogen fixation were low (Marumo and Asaoka, 1974; Zhang et al., 2012). Zhang et al. (2012) suggested that the low nitrogen fixation in the continental shelf was attributable to mixing processes and the influence of the Changjiang River. Turbulence near the sea floor influences the surface water in the shallower bottom region (Matsuno et al., 2006), and Zhang et al. (2012) suggested that the physical disturbance reduces diazotrophy since diazotrophs including *Trichodesmium* favor calm seas. Furthermore, the water in the continental shelf of the ECS is strongly influenced by the Changjiang River. The N/P ratio of the Changjiang River plume is significantly higher than the Redfield ratio, which results in phosphorus limitation, and can contribute to the low nitrogen fixation (Zhang et al., 2012). In the present study, despite the fact that the surface phosphate concentration was low throughout the study areas, the N/P ratio was generally lower than the

450 Redfield ratio, suggesting that biological production was limited by the availability of
451 nitrogen compared to phosphate (Moore et al., 2008, 2013). Furthermore, the
452 insignificant difference in MLD among the ECS, the Kuroshio, and the Philippine Sea
453 (p>0.05; Tukey HSD test) indicated similar vertical mixing conditions. Therefore, the
454 environmental variables related to nitrogen fixation only slightly differed as
455 demonstrated by the nMDS plot.

456 In our study, we found a *Trichodesmium* spp. bloom near the Miyako Islands.
457 Recent studies demonstrated that *Trichodesmium* spp. thrived near oceanic islands
458 (Shiozaki et al., 2010, 2014c; Dupouy et al., 2011). Given that some aspect of the
459 environment around the islands increases *Trichodesmium* spp. abundance and that
460 they are transported from the islands to the Kuroshio, this can explain why the
461 *Trichodesmium* distribution was not estimated from environmental variables.
462 Accordingly, the low abundance of *Trichodesmium* spp. in the Philippine Sea was
463 likely due to the low density of islands. Furthermore, higher nitrogen fixation in the
464 Kuroshio than in the Philippine Sea might be explained in the same manner.
465 *Trichodesmium* is a major nitrogen fixer in the Kuroshio (Chen et al., 2008, 2014;
466 Shiozaki et al., 2014a), and our results showed that the bulk water nitrogen fixation
467 was positively correlated with *Trichodesmium* abundance.

468 The numerical simulation demonstrated that released particles from the Miyako
469 Islands were generally transported to the northeast and flowed along the Kuroshio
470 during summer between 2003 and 2009. Thus, if *Trichodesmium* increases and active
471 nitrogen fixation usually occurs around the Miyako Islands, the water would be
472 delivered to the Kuroshio. Furthermore, we performed additional particle tracking
473 experiments whose particle release points were set at major islands in the Ryukyu
474 Islands (Amami Islands, Okinawa Main Island, and the Ishigaki Islands) (Figs. S6 and
475 S7). The results demonstrated that the particles released from the other islands of the
476 Miyako Islands were also delivered to the Kuroshio, with some exceptions. Based on
477 the calculations for 2003–2009, 13–56% (30 ± 16%) of particles released from the
478 islands reached Area K by day 120 (Fig. S7).
479 Studies on nitrogen fixation around islands in the study region are fairly limited
480 (Liu et al., 2013), and the present study is the first report of a *Trichodesmium* bloom
481 around islands in the area. The Miyako Islands are surrounded by reefs, and studies
482 have shown that *Trichodesmium* blooms can be associated with reef environments
483 (Bell et al., 1999; McKinna et al., 2011). However, the factors causing the
484 *Trichodesmium* blooms around islands are not well understood (Shiozaki et al.,
485 2014c). Further studies are required to identify which characteristics of the near island

environment are important for the growth and/or accumulation of *Trichodesmium* and other diazotrophs.

5. CONCLUSIONS

We hypothesize that the high abundance of *Trichodesmium* spp. and active nitrogen fixation in the Kuroshio were ascribable not to the unique nutrient environment, but rather to the supply of *Trichodesmium* spp. and other diazotrophs from the surrounding islands. The Ryukyu Islands would not be the only islands with abundant *Trichodesmium* spp., as *Trichodesmium* spp. also flourish in the upstream Kuroshio near Luzon Island (Chen et al., 2008). Therefore, the abundance of *Trichodesmium* spp. would be generally increased around islands situated along the Kuroshio, and the abundant *Trichodesmium* spp. would likely be transported to the mainstream of the Kuroshio. *Trichodesmium* is a major diazotroph in the Kuroshio (Chen et al., 2008, 2014; Shiozaki et al., 2014a), and diazotrophy in the Kuroshio is considered to influence the nutrient stoichiometry in the North Pacific (Shiozaki et al., 2010). Thus, our results indicate that phenomena around the islands located along the Kuroshio are important for determining the partial nitrogen inventory in the North Pacific.

Author Contributions

T.S., S.T., S.I., and K.F. designed the experiment and T.S., S.T., T.K., X.L., F.H., and K.F. collected the samples at sea. T.S. determined nitrogen fixation and abundance of *Trichodesmium* spp. during the KT-06-21, KT-07-21, KT-09-17, and *Nagasaki-maru* 242 cruises, and X.L. did during the KT-10-19 cruise. T.S. analyzed datasets of satellite and climatological current field. S.T. analyzed concentration of dissolved iron. S.I. performed numerical experiments. T.K. and F.H. determined nutrient concentration. T.S. prepared the manuscript with contributions from all co-authors.

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526

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676

677 Table 1 Summary of *Trichodesmium* at the surface, and depth-integrated nitrogen

678 fixation and its related parameters in the four representative study areas.

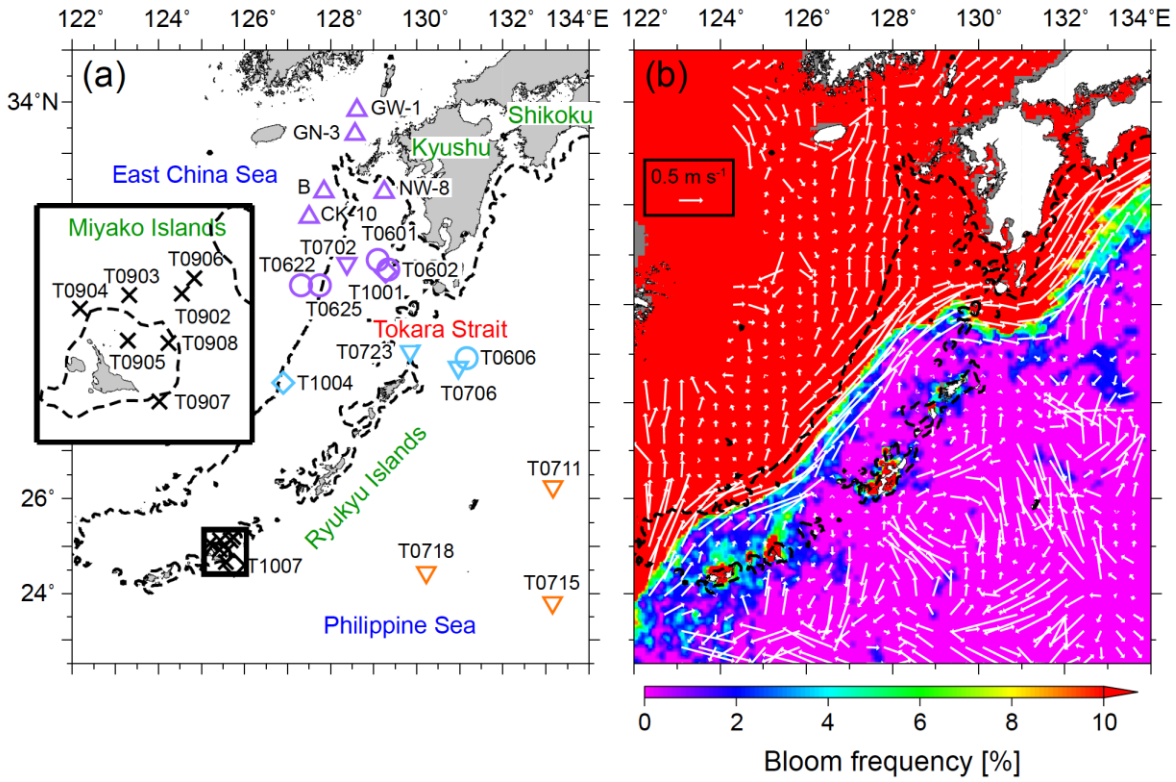
Area	*Trichodesmium*[*] [filaments l^{-1}]	N_2 fixation [μmolN L^{-1} d^{-1}]	Temperature[*] [ºC]	MLD [m]	$NO_3^-+NO_2^{-*,\dagger}$ [nM]	$PO_4^{3-*,\dagger}$ [nM]	DFe[*] [nM]
East China Sea	21±58	170±140	28.5±1.2	24±12	19±11	15±9	0.76±0.18
Kuroshio	43±33	199±142	29.4±0.81	27±8	9±8	15±7	0.45±0.13
Philippine Sea	8±8	58.3±25.1	29.4±0.1	23±3	8±3	14±19	0.51±0.25
Miyako Islands	3019±8478	201±274	29.3±0.3	40±12	61±128	8±7	0.38±0.24

679 [*] values in surface water

680 [†]When the concentration was below the detection limit (3 nM), we assumed a concentration of 3 nM to

681 calculate the mean.

682



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685 Figure 1. (a) Sampling stations during the KT-06-21 (circles), KT-07-22 (inverted
686 triangles), KT-09-17 (crosses), KT-10-19 (diamonds), and 242 (triangles) cruises.
687 Symbols of stations located in the East China Sea, the Kuroshio, the Philippine Sea,
688 and near the Miyako Islands are indicated in purple, light blue, orange, and black,
689 respectively. (b) Climatological surface current fields during summer (1953–2008)
690 from geoelectrokinetograph measurements and ship-mounted ADCP data. The
691 background contour represents the percentage of chlorophyll *a* of >0.15 mg m^{-3}
692 during summer between 2003 and 2009. Dashed lines indicate 200 m isobaths.

693

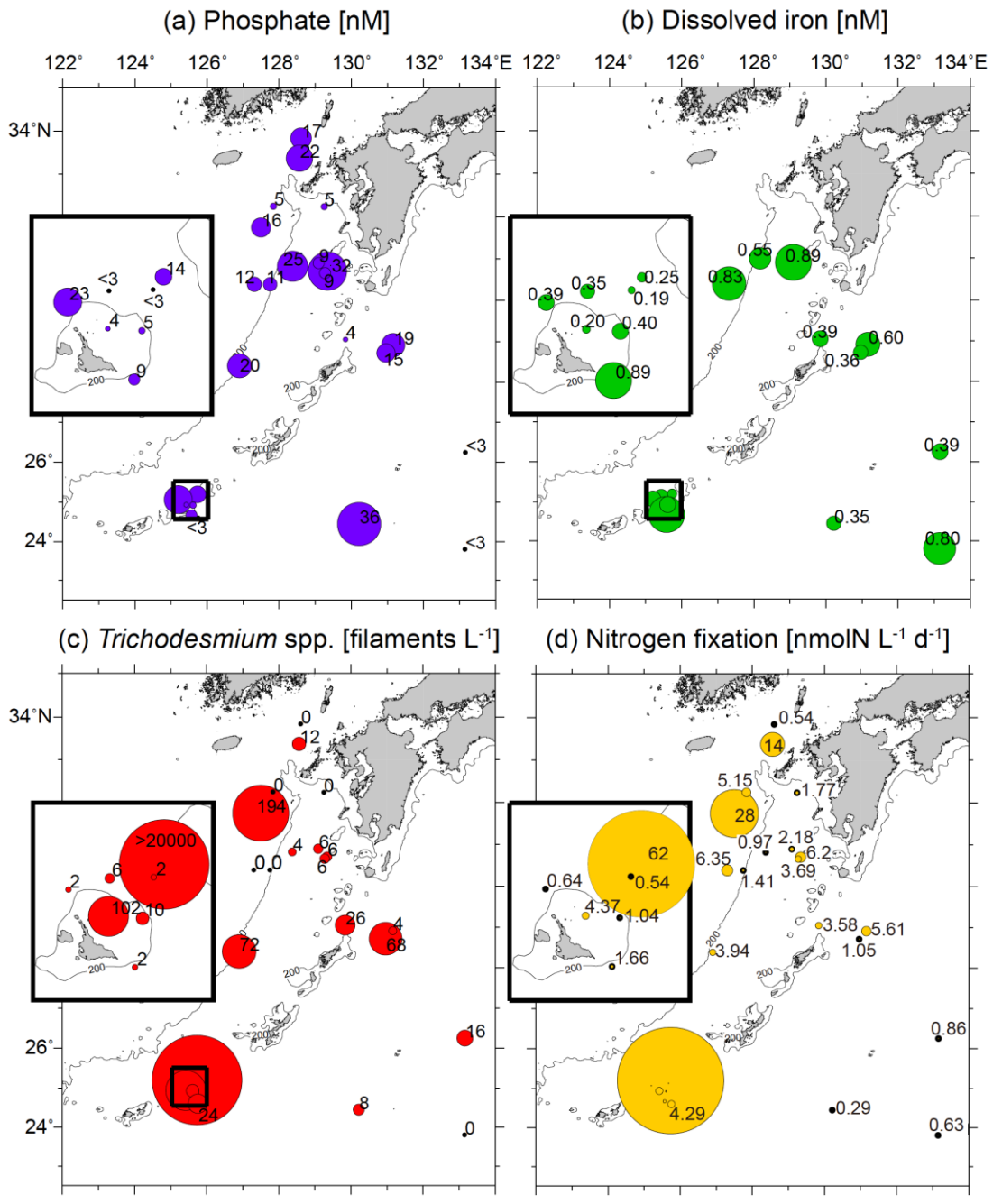


Figure 2. Distribution of (a) phosphate, (b) dissolved iron, (c) *Trichodesmium* spp., and (d) nitrogen fixation at the surface. The parameters in the small boxes indicate results from the KT-09-17 cruise. The areas of the circles are proportional to the concentration, abundance, or activity.

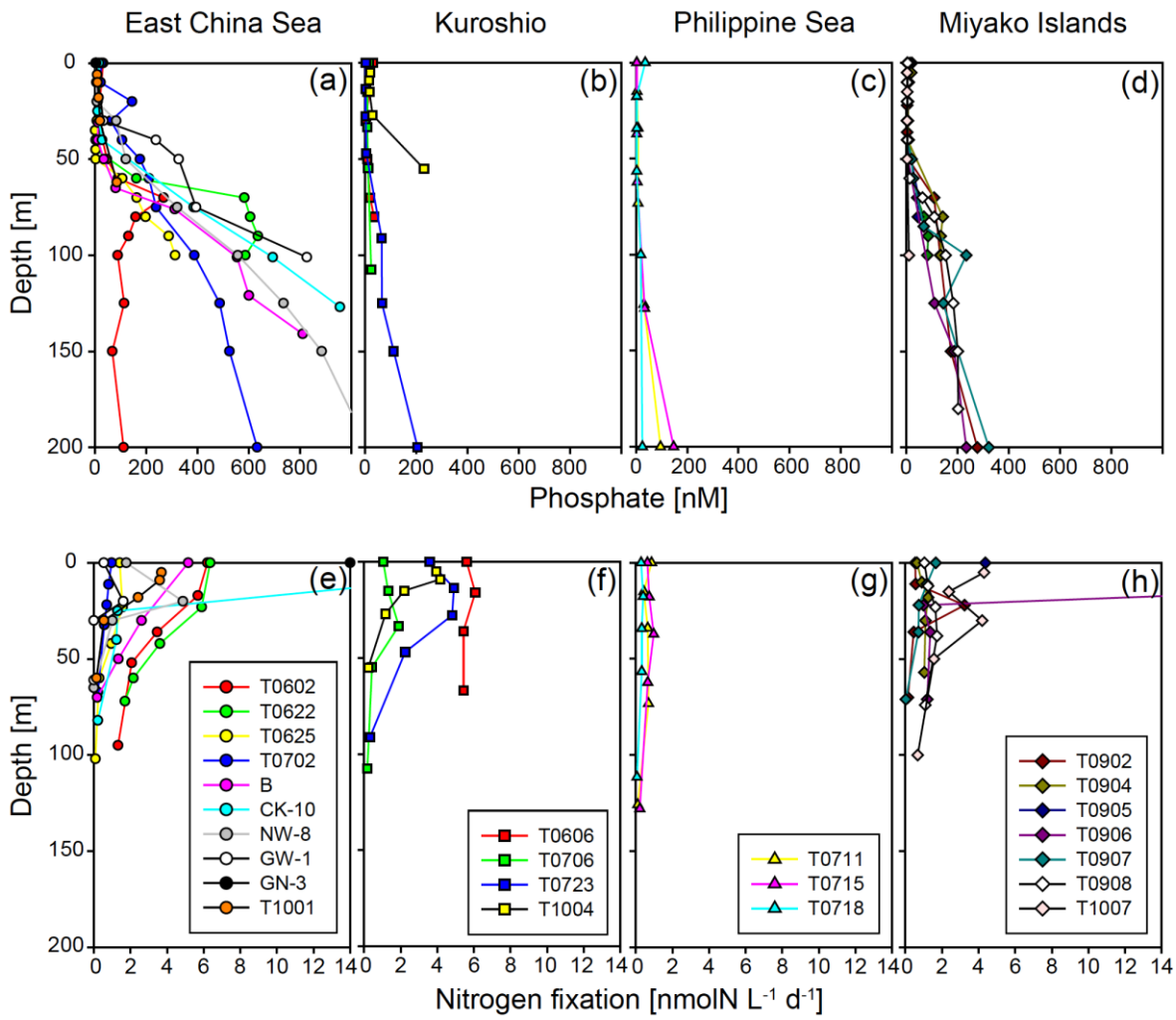
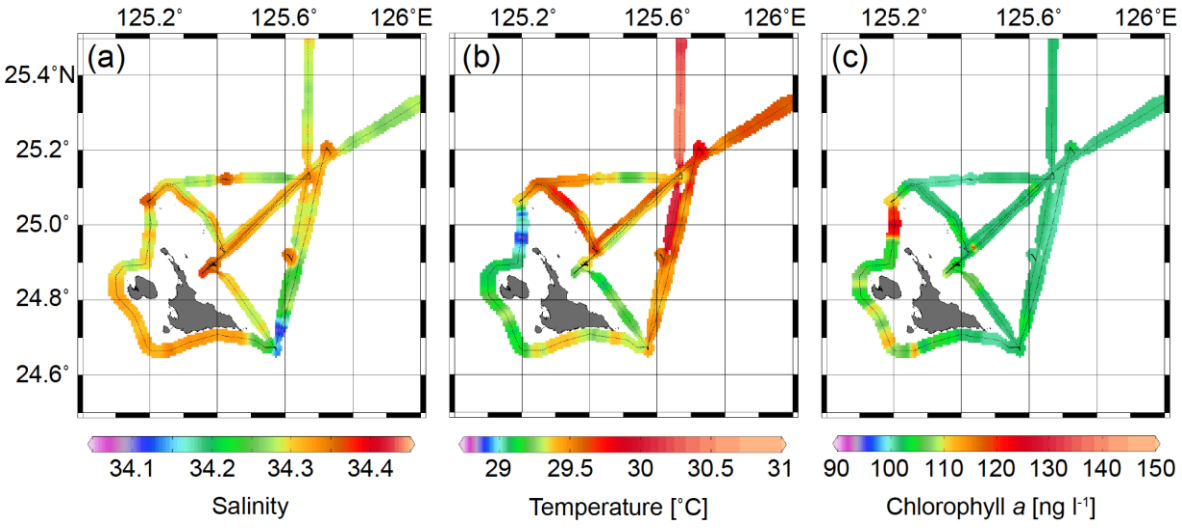


Figure 3. Vertical profiles of phosphate and nitrogen fixation in the East China Sea (a and e), the Kuroshio (b and f), the Philippine Sea (c and g), and the Miyako Islands (d and h).



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708 Figure 4. Surface (a) salinity, (b) temperature, and (c) chlorophyll *a* during the

709 KT-09-17 cruise.

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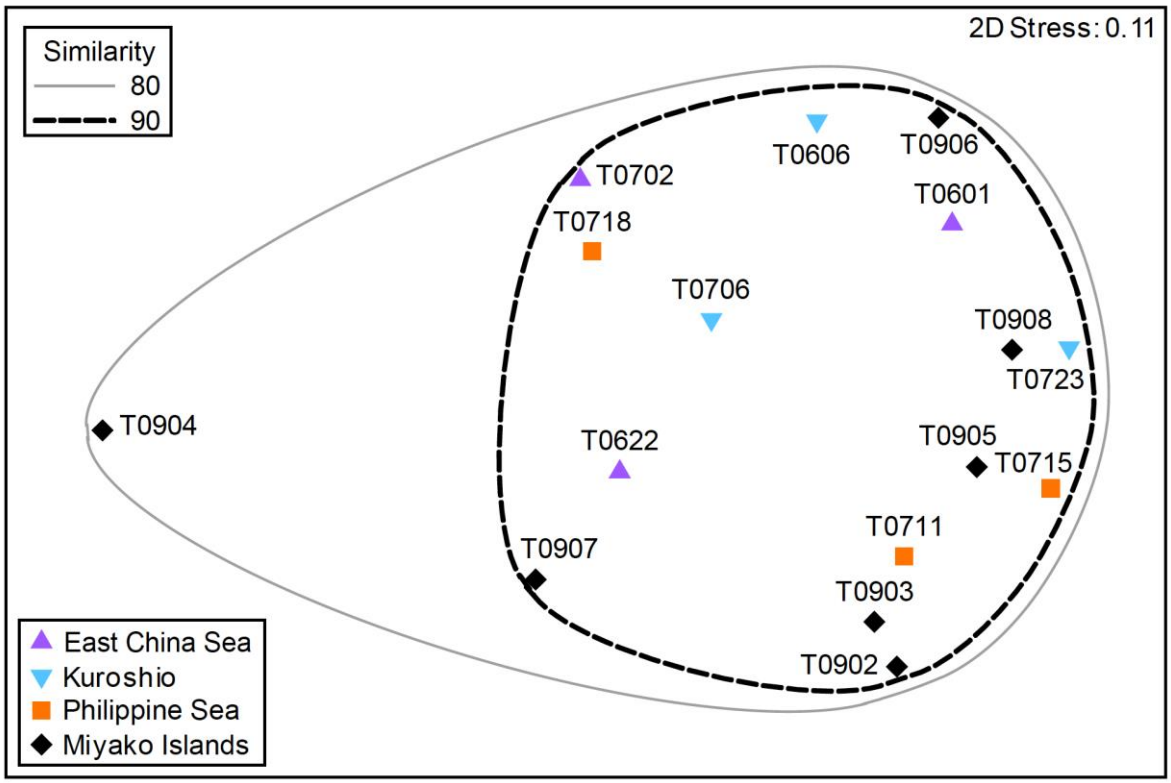

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713 Figure 5. nMDS ordination of sampling stations with environmental variables

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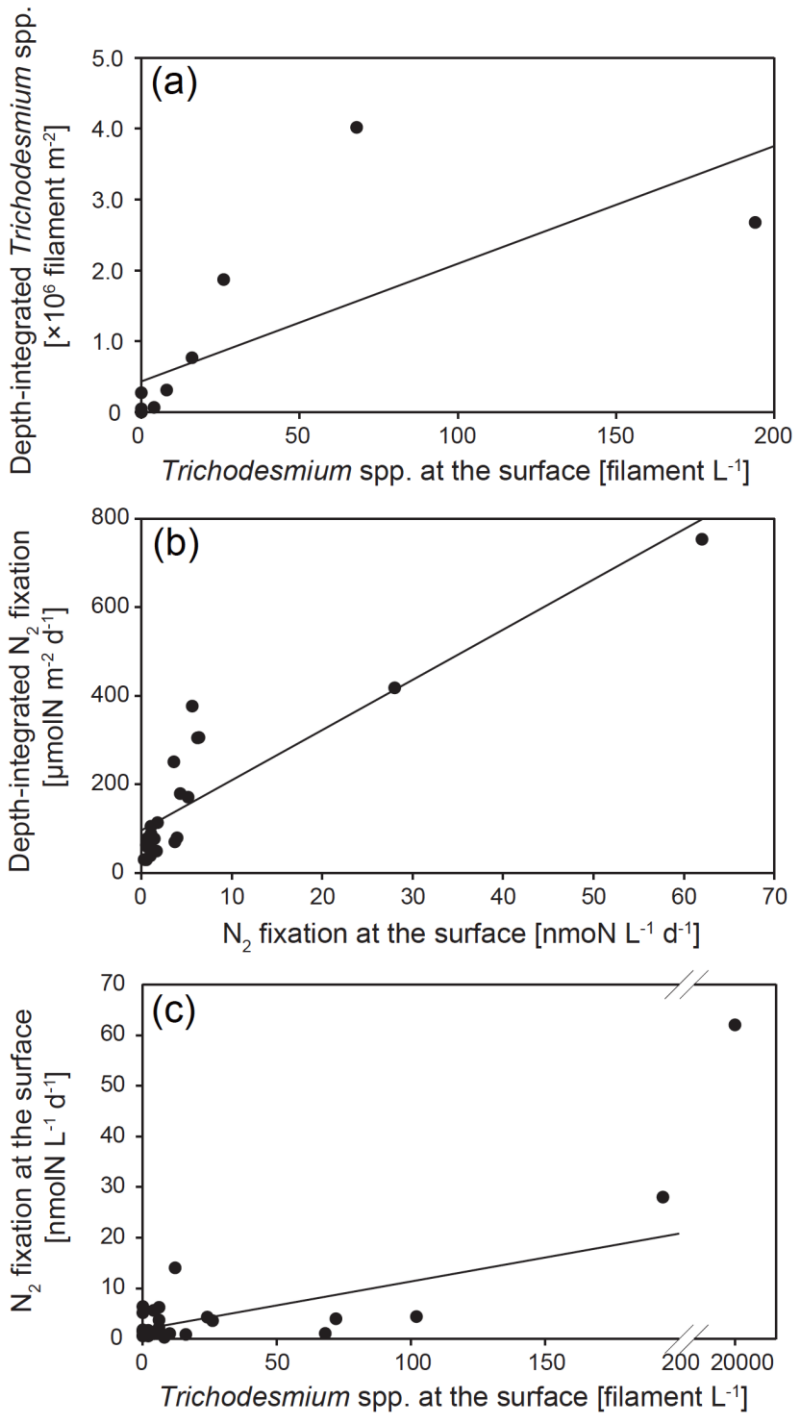

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716

717 Figure 6. Relationships (a) between surface and depth-integrated *Trichodesmium* spp.

718 abundance, (b) between surface and depth-integrated nitrogen fixation rates, and (c)

719 between *Trichodesmium* spp. abundance and nitrogen fixation rate at the surface.

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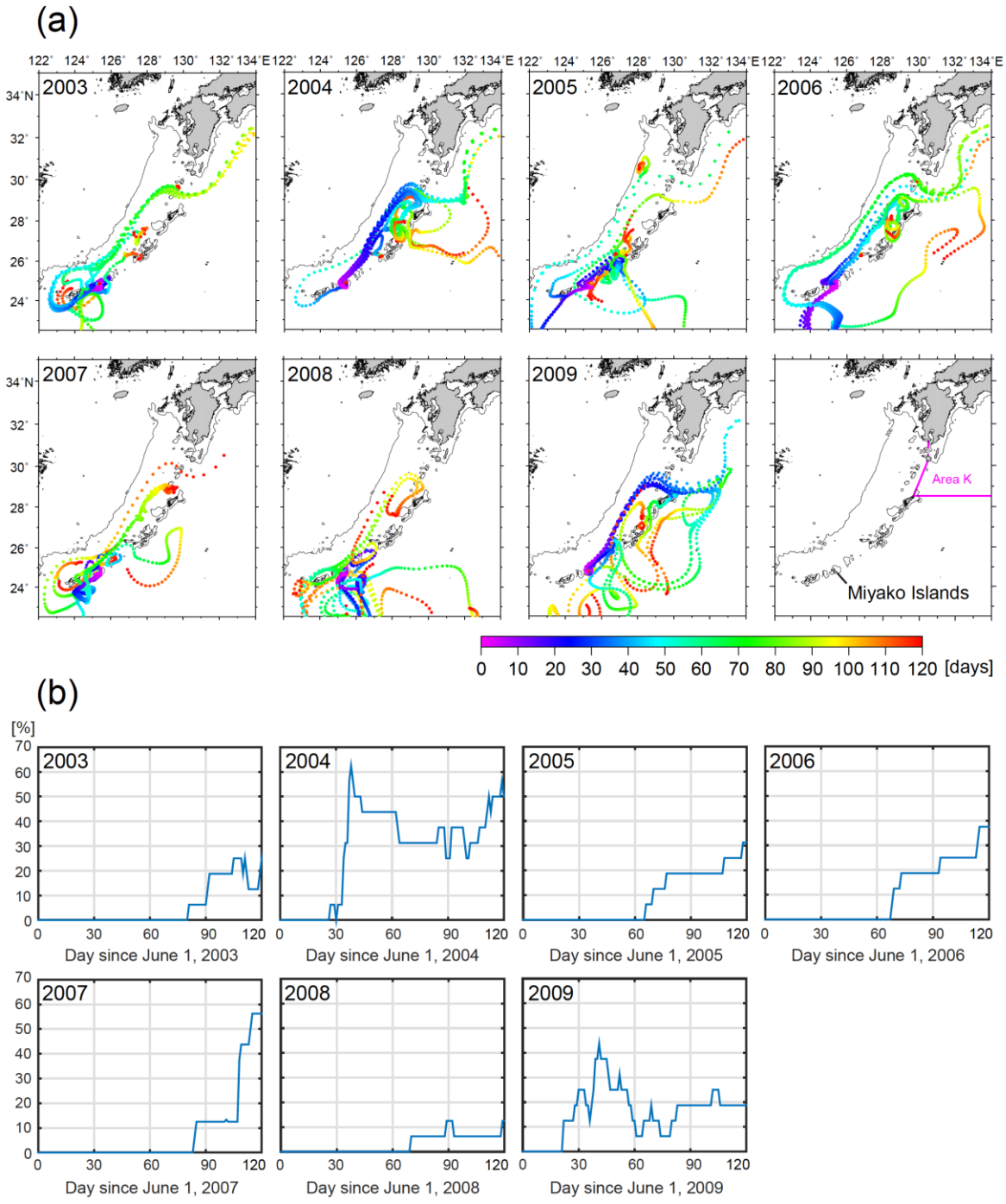

Figure 7. (a) Trajectories of particles released from points around the Miyako Islands on June 1, 2003–2009. (b) The ratio of particles delivered to Area K to the total released particles.