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The impact of sedimentary alkalinity release on the water column CO₂ system in the North Sea

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Abstract

Recently, it has been proposed that alkalinity release from sediments can play an important role in the carbonate dynamics on continental shelves, lowering the pCO_2 of seawater and hence increasing the CO_2 uptake from the atmosphere. To test this hypothesis, sedimentary alkalinity generation was quantified within permeable and muddy sediments across the North Sea during two cruises in September 2011 (basinwide) and June 2012 (Dutch coastal zone). Benthic fluxes of alkalinity (A_T) and dissolved inorganic carbon (DIC) were determined using shipboard closed sediment incubations. These results show that sediments can be an important source for alkalinity, particularly in the shallow southern North Sea, where high A_T and DIC fluxes were recorded in near shore sediments of the Belgian, Dutch and German coastal zone. In contrast, fluxes of A_T and DIC are substantially lower in the deeper, seasonally stratified, northern part of the North Sea. Overall, our results show that sedimentary alkalinity generation should be considered an important factor in the CO₂ dynamics of shallow coastal systems.

1 Introduction

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Coastal seas play a crucial role in the global carbon cycle by connecting the terrestrial, oceanic and atmospheric reservoirs (Regnier et al., 2013). Although continental shelves cover only 7 % of the total global ocean surface, they are estimated to account for up to 30 % of the oceanic primary production (Gattuso et al., 1998) and between 10 and 25 % of the current day oceanic CO_2 uptake (Regnier et al., 2013; Liu et al., 2010). In general, most open shelves in the temperate and high-latitude regions seem undersaturated with respect to atmospheric CO_2 , while the low-latitude shelves gener-

ally tend to be supersaturated (Chen and Borges, 2009). However, this rule is far from absolute. Although temperate coastal areas are generally acting as a CO₂ sink, considerable variability has been observed in the CO₂ uptake within and between systems,





which may be explained by a dominance of different drivers of CO_2 uptake. High nutrient inputs from land fuel intense primary production and hence stimulate atmospheric CO_2 uptake, while respiration of organic matter exported from terrestrial ecosystems stimulates the release of CO_2 in coastal areas (Bozec et al., 2006; Jönsson et al., 2011;

Schiettecatte et al., 2007). In addition to these biological sinks and sources, the effect of temperature on the CO₂ solubility controls the magnitude and direction of the CO₂ exchange between coastal waters and the atmosphere (Kitidis et al., 2012; Borges and Frankignoulle, 2003). Therefore the question remains as to what particular drivers are governing the CO₂ dynamics in a given coastal system (Borges and Frankignoulle, 2003).

In addition to water column processes, sediments have also been suggested to play a role, as the shallowness of coastal seas permits a close interaction between the sediment, the water body and the atmosphere. Coastal water bodies are characterized by either a permanently or seasonally mixed water column, which hence establishes

- ¹⁵ a direct link between the sediment and the atmosphere, acting on a time scale of days to months. In contrast, in the open ocean, the sediment and the atmosphere can only interact over much longer time scales (\approx 1000 yr of global oceanic circulation). However, the extent to which sediment geochemistry plays a role in the CO₂ uptake of coastal system remains largely unresolved.
- It has been proposed that alkalinity generation caused by anaerobic organic matter degradation in shallow coastal sediments can increase the CO_2 buffer capacity of coastal waters and therefore increase atmospheric CO_2 uptake. When coining this hypothesis, Thomas et al. (2009) estimated that alkalinity generation in Wadden Sea sediments could be responsible for 20–25% of the total CO_2 uptake of the entire North
- Sea. In the East China Sea it was already suggested by Chen and Wang (1999) that alkalinity generation due to anaerobic degradation processes in sediments adds considerably to the total alkalinity budget in this area. Furthermore, Chen (2002) proposed that shelf-generated alkalinity release from benthic anaerobic processes could be almost as important as alkalinity generated by dissolution of carbonates in the open





ocean. Finally, based on modeled nitrogen and carbon budgets for the northwestern North Atlantic continental shelf, Fennel (2010) suggested that shelf sediments must be an important source of alkalinity. However, Hu and Cai (2011a) obtained a much smaller alkalinity flux from global coastal sediments based on the upscaling of local

⁵ denitrification and sulfate reduction rates. They concluded that sedimentary alkalinity fluxes are too low to significantly affect the alkalinity budget of the global ocean, but also stated that sedimentary anaerobic processes could be important on regional scales as an alkalinity release mechanism. From these results, it is clear that currently, there is uncertainty as to what extent coastal sediments are an important source of alkalinity to coastal oceans.

In this study, we aim to quantify sedimentary alkalinity generation within various sediment-types in the North Sea. During two cruises samples were collected in September 2011 (basin-wide) and June 2012 (Dutch coastal zone). Benthic fluxes of alkalinity, dissolved inorganic carbon and oxygen were determined using closed sedi-¹⁵ ment incubations and microsensor profiling. Subsequently, we analysed the sources of the observed sedimentary alkalinity release, and after implementing an alkalinity budget for both water column and sediment, we constrained the *net* CO₂ uptake from the atmosphere associated with the sedimentary alkalinity release.

2 Materials and methods

20 2.1 North Sea system

In this paper we divide the North Sea into three different hydrogeographical zones: the southern North Sea (SNS), the northern North Sea (NNS) and the Skagerrak and the Norwegian Trench (SKNT). The SNS encompasses the Southern Bight (between Belgium/Netherlands and the UK), the shallow Wadden Sea (running along the Dutch and Corman apart up to Fabiara in Danmark) the Corman Bight and the control part

²⁵ and German coast up to Esbjerg in Denmark), the German Bight, and the central part of the North Sea (Fig. 1). The major difference between the SNS and the NNS is the





stratification regime. Whereas the shallow SNS has a fully mixed water column throughout the year, the NNS is thermally stratified in summer, but fully mixed during winter, due to strong wind forcing and surface cooling (Elliott et al., 1991). As the operational border between the SNS and NNS, we use the 100 m depth bathymetric isoline, as employed in previous studies (Pätsch and Kühn, 2008) . The third hydrogeographical zone constitutes the Skagerrak and the Norwegian Trench (SKNT), which forms one of the major sediment depositional areas of the North Sea (de Haas and van Weering, 1997). The Skagerrak is part of the transition area that connects the North Sea with the Baltic Sea. It is a rather small strait (200 by 100 km) between the southeast coast of

- Norway, the southwest coast of Sweden, and the Jutland peninsula of Denmark. The average depth of the Skagerrak is about 210 m. The Skagerrak is strongly stratified in summer, but also features a weak stratification in winter driven by Baltic freshwater inputs (Gustafsson and Stigebrandt, 1996; Rodhe, 1987). The Skagerrak is connected to the Norwegian Sea through the Norwegian trench with a sill depth of 270 m. The Norwegian trench with a sill depth of 270 m. The Skagerrak is about 210 m.
- ¹⁵ Norwegian Trench itself is a deep sedimentary basin (250–700 m) that reaches from the Oslofjord in the southeast to the Stad peninsula in the northwest up (Rodhe, 1996). Like the Skagerrak, the Norwegian Trench is characterized by haline stratified water masses (Reid and Edwards, 2001).

The water transport in the NNS is dominated by the large open boundary with the North Atlantic in the north. Water entering through the Shetland Channel and the Faire Island Channel turns eastwards and leaves the North Sea via the Norwegian Trench in the east. The residence time of the water is about one year. Generally, water entering the NNS does not influence the SNS, as just 5% of the North Atlantic water entering the northern boundary reaches the SNS (Lenhart and Pohlmann, 1997). The water

transport in the SNS is mainly determined by inflow of Atlantic Ocean through the English Channel, which mixes with low salinity water coming from rivers and moves along the eastern coastlines towards the north-east. Baltic Sea water entering through the Kattegat and coastal run-off are important in maintaining the Norwegian Coastal Current that initiates in the Skagerrak. This northwards directed current is the only outflow



of the North Sea and thus balances all incoming water inputs as described above. Additionally, the shallow SNS is influenced by a strong tidally induced currents and mixing (Dauwe, 1999). The tides in the SNS are diurnal, whereas maximum surface currents at spring tide occur on the western and southern parts of the SNS. More the the north and into the German Bight, the tidal current velocities decrease (Van der Molen, 2002).

The seafloor of the North Sea predominantly consists of permeable sediments. Medium and fine sand is the main sediment type and occupies the largest part of the North Sea basin. Coarse sand is found at confined locations throughout the entire North Sea basin with larger areas of coarse grained sediments present along the English coast and in front of the German and Danish coasts. Mud and sandy mud are

mainly found in the deep trenches along the Norwegian coast, off the coast of Scotland, in particular locations along the Belgian coast and in smaller areas north and east of the German Bight (Lüders, 1955; Schlüter and Jerosch, 2009; Braeckman et al., 2014).

2.2 Sediment sampling

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- ¹⁵ During a cruise onboard RV *Pelagia* in September 2011 we sampled a total of 19 stations across the whole North Sea basin (Table 1). On a second cruise in June 2012, we sampled 7 stations along a transect perpendicular to the Dutch coast from the Wadden island of Terschelling up to the Oyster Grounds in the central North Sea (1b–7b in Fig. 1). Sediment cores were collected at each site using a Reineck box corer. Poly-
- ²⁰ methyl methacrylate (PMMA) core barrels (19 cm inner diameter) were subsequently inserted into the sediment of the box core to a depth of 10 to 15 cm enclosing 15 to 20 cm of overlying water. The cores were excavated from the box core, closed off with a lid at the bottom and immediately transferred into a water-filled reservoir in a thermo-controlled container that was kept at in-situ bottom water temperature. These sediment cores were subsequently used for closed core flux incubations as described below.

During the September 2011 cruise, small sediment cores were retrieved for solid phase analysis and microsensor profiling and these were taken from the same box core as the flux cores. For solid phase analysis, acrylic core barrels (5 cm i.d.) were





inserted into the sediment of the box core. The upper 10 cm of each core was sliced in 1 cm intervals and sediment samples were analyzed for porosity and grain size distribution. Porosity was determined by weight loss after freeze-drying, accounting for salt precipitation in the saline pore water. Grain size distribution was determined using 5 a Malvern Mastersizer 2000 particle analyzer.

For O₂ and pH microsensor profiling, acrylic core liners (3 cm i.d.) were inserted into the sediment of the box core, and afterwards, the sediment was brought level to the rim of the core liner. Cores were subsequently placed in an aquarium containing bottom water at in-situ temperature, which was constantly bubbled with ambient air. Bottom water to top up sediment cores for flux measurements was retrieved by casts with 25 L Niskin bottles (Ocean Test Equipment/Fort Lauderdale, USA) retrieving water at approximately 1 m above the sediment surface. A conductivity, temperature, depth (CTD) was mounted on a standard rosette frame together with 24 Niskin bottles. The CTD was equipped with a SBE3+ thermometer, a SBE4 conductivity meter and a SBE43 dissolved oxygen sensor (Seabird, USA).

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2.3 Solute flux measurements

Flux chamber incubations are potentially susceptible to various methodological artifacts resulting from sediment enclosure, which relate to the sensitivity of benthic fluxes to changes in ambient hydrodynamics and altered benthic faunal activity (Santschi et al., 1991; Tengberg et al., 2005; Lehrter et al., 2011). In permeable sediments, 20 fluxes are particularly susceptible to the imposed stirring regime and the associated local pressure gradients that are generated in the flux chamber, which drive the advective pore water exchange between sediment and overlying water (Huettel and Gust, 1992; Janssen et al., 2005). The flux chamber type employed here was based on the design as in Huettel and Gust (1992), which was specifically developed for flux studies 25 in permeable sediments. The sediment cores retrieved were closed off with a PMMA top lid that was equipped with a large central stirring disc (diameter: 14 cm). The ro-





gradient, which drives pore water exchange in the chamber. A major challenge for flux chamber studies in permeable sediments is the selection of the appropriate stirring regime. Without knowledge of the local in-situ hydrodynamics, it is impossible to predict a priori which stirring regime is appropriate for a given site. Here we used two different stirring rates at each station (40 and 80 RPM), to mimic a range of interfacial

- pressure gradients and solute exchange conditions (Huettel and Gust, 1992; Janssen et al., 2005; Rao et al., 2012). By taking this approach, we are able to discern how sensitive the fluxes at each station are to advective exchange, and thus, we get an idea of the uncertainty on our flux estimates for the permeable sites that we visited.
- ¹⁰ Prior to the start of the flux measurements, the overlying water in each core was replaced with ambient bottom water to ensure that the chemical composition of the overlying water closely resembled in-situ conditions. Each set of flux measurements began by securing gas-tight lids equipped with O-ring seals on each core. Core lids contained two sampling ports on opposite sides for subsampling during the incubation
- and these ports were carefully purged with bottom water prior to the start of each incubation to remove any air bubbles trapped. Fiber-optical oxygen sensors (FireSting OXF1100) were inserted into a third opening in the top lid. The chambers were closely inspected to ensure that no gas bubbles remained inside the chamber. Two or three replicate chamber incubations were made per station.
- The temporal evolution of the oxygen concentration in the overlying water of the flux chambers was continuously monitored using oxygen optodes at a sample interval of 1 min. Optodes were pre-calibrated on the same day using a two-point calibration with ambient seawater at 0 % (saturated with sodium sulphite) and 100 % O₂ saturation (bubbled with air). Optode results were verified with measurements of the O₂ concentration by Winkler titration in discrete water samples before and after incubation

(Grasshoff et al., 2009).

A water subsample (~ 50 mL) was withdrawn from the chambers at 4–6 h intervals for solute analysis as described below. When a water sample was extracted via one sampling port, an equal amount of ambient bottom water entered through the replace-





ment tube connected to the other sampling port. Samples were collected in plastic syringes for alkalinity (A_T) and in glass syringes for dissolved inorganic carbon (DIC) analysis. Water samples for A_T (~ 10 mL) were filtered (0.45 µm Millex-HA syringe filter) and stored in the dark at 4 °C. DIC water samples (~ 12 mL) were not filtered, but poisoned with 10 µL HgCl₂ and stored submerged at 4 °C in a fridge.

The Total Oxygen Uptake (TOU) rate and the flux of DIC and A_T were determined from a linear regression of overlying water concentrations vs. incubation time (Eq. 1).

$$J = \frac{V_{\rm ow}}{A} \frac{{\rm d}C_{\rm ow}}{{\rm d}t}$$

At the end of the incubation, the top lid was removed, and the height of the overlying water ($H = V_{ow}/A$) was measured at four points along the side of the core using a ruler and the mean height was calculated. Glud (2008) suggested that an oxygen decrease by more than 10–15% from the initial conditions can already stimulate processes that cause a non-linear decrease in oxygen concentrations, which also might affect other solute fluxes across the sediment–water interface. Here we found that the linear re-15 gressions used to calculate the oxygen uptake were insensitive to an oxygen decrease of 30% or more (for examples see Fig. 3).

2.4 Analytical methods

Total A_{T} was determined via an open-cell titration procedure, using a Metrohm Titrando 888 system with a combined Metrohm glass electrode (Unitrode) following the proce-²⁰ dure SOP3a as described in Dickson et al. (2007). Samples (10 mL) were placed in a temperature regulated open cell (25 °C) and titrated with a solution of hydrochloric acid (0.1 N) in a two-stage process. First the sample (10 mL) was acidified to a pH close to 3.5 and then titrated in small steps down to a pH of 3.0. Subsequently, A_{T} was calculated using a non-linear regression approach based on SOP3a from Dickson

et al. (2007). Two replicate measurements were carried out for each sample analyzed. Titrations (n = 10) of Certified Reference Materials (CRM Batch 116 provided by A. G.



(1)



Dickson) were on average within $4\,\mu\text{mol}\,\text{kg}^{-1}$ of the nominal value with a precision of $5\,\mu\text{mol}\,\text{kg}^{-1}.$

DIC was determined using an AS-C3 DIC analyzer (Apollo SciTEch, USA), in which the sample (10 mL) was acidified and the released CO_2 was detected using a solid state infra-red CO_2 detector (LI-7000, LI-COR Biosciences, USA). Two replicate measurements were carried out for each sample analyzed. Quality assurance of the DIC analysis was also based on CRM (Batch 116 – accuracy and precision: $3 \mu mol kg^{-1}$).

2.5 O₂ and pH microprofiling

Microsensor profiling was performed using commercial Clark-type O₂ and potentiometric pH microsensors operated with a motorized micromanipulator (Unisense A.S., Denmark). Vertical depth profiles of O₂ were recorded using an electrode with a tip size of 100 µm at 250 µm steps, beginning at 2 mm above the sediment–water interface until either anoxia or 20 mm depth. The O₂ microsensors were calibrated with a 2-point calibration made in air-saturated seawater (100 % saturation) and at depth in anoxic sediment (0 % saturation). Depth profiles of pH were measured using microsensors with a tip size of 200 µm in 1 mm steps, beginning 4 mm above the sediment surface until 35 mm depth. Measurements were always started within 1 h after sampling. The pH microsensors were calibrated using NBS buffers (pH 4 and 7) and TRIS buffer (Delvalls and Dickson, 1998), and the pH is reported on the total scale.

²⁰ The diffusive oxygen uptake (DOU) of the sediment was calculated from the O₂ depth profiles as

$$DOU = -D_{O_2} \frac{d[O_2]}{dz}$$
(2)

where *z* is the depth and $[O_2]$ denotes O_2 concentration. Profiles were measured under diffusive conditions and the slope $d[O_2]/dz$ was determined from the gradient in the diffusive boundary layer (Glud, 2008). The molecular diffusion coefficient of O_2 in





seawater (D_{O_2}) was calculated as a function of the bottom water salinity and temperature using the *CRAN:marelac* extension package in the open-source programming language R (Soetaert et al., 2010).

2.6 Statistical analyses

⁵ Results are reported as the mean ± 1 standard deviation (SD) of *n* replicate measurements. Nonparametric statistics were used in the interpretation of results, including the Mann–Whitney *U* test (*p*) for comparison of the mean of two independent groups of measurements, and the Spearman's rank correlation coefficient (*p*) as a measure of the statistical dependence between two variables. Statistical analyses were conducted in R using the *CRAN:stats* package.

3 Results

3.1 Bottom water characteristics

Bottom water salinity, temperature and oxygen concentration as obtained by CTD profiling are listed in Table 1, and characteristic temperature depth profiles are displayed ¹⁵ in Fig. 2. Bottom water salinity and temperature of stations from the basin wide North Sea cruise ranged from 35.39 PSU (NNS) to 29.35 PSU (SNS) and 17.41 °C (SNS) to 6.02 °C (NNS). The bottom water in the NNS and SKNT was generally colder and more saline than bottom water in the SNS (Mann–Whitney; Temperature: p = 0.001; Salinity: p < 0.001). A thermocline was formed between between 20–50 m water depth in all

²⁰ stations of the NNS and SKNT, but just in two stations of the SNS (Fig. 2 and Table 1). The bottom water oxygen concentrations did however not exhibit significant differences between the SNS and other parts of the North Sea (Mann–Whitney; p > 0.5).





3.2 Sediment properties

The median grain size for all stations visited during the basin wide North Sea campaign in 2011 ranged from 21 to 499 μ m (mean median grain size: 215 μ m). In the SNS the median grain size ranged from 138–499 μ m (mean median grain size: 215 μ m),

⁵ and according to the Wentworth scale, the SNS sediments can be classified as fine to medium sand (Table 1). Sediments in the NNS (median grain size range 119–314 µm; mean 227 µm) were not significantly different from the SNS (Mann–Whitney; p = 0.8), hence also classifying as fine to medium sand. In contrast, sediments from the Skagerrak and Norwegian Trench (SKNT) were considerably finer (Mann–Whitney; p < 0.01), with the median grain size ranging from 22–62 µm. Thus, sediments of SKNT classified as fine to coarse silt on the Wentworth scale (Table 1).

Porosity for all stations in the North Sea basin varied from 0.32 to 0.74. Between sediments of the SNS (mean: 0.34) and NNS (mean: 0.37) no significant differences in porosity were found (Mann–Whitney; p = 0.31). However, sediments of the SKNT displayed a significantly higher porosity than sediments from the SNS or NNS (Mann–Whitney; p < 0.05).

3.3 Benthic flux chamber incubations

The temporal evolution of the solute concentrations (O₂, DIC and A_T) in the overlying water from flux chamber incubations at representative stations from each of the three zones (SNS, NNS, SKNT) is presented in Fig. 3. The oxygen data display a linear decrease, while the DIC and A_T data display a increasing trend, albeit with greater variability due to the limited number of subsamples. Fluxes were only considered if their calculation is based on a linear regression of at least 4 points combined with an r^2 value greater than 0.8. Furthermore, fluxes were set to zero in case the slope of the linear regression was not significantly different from zero.

During the basin-wide campaign in September 2011, TOU rates ranged from 3.1 to $28.7 \text{ mmol m}^{-2} \text{d}^{-1}$ for the SNS, 0.7 to $6.2 \text{ mmol m}^{-2} \text{d}^{-1}$ for the NNS and 2.9 to



5.7 mmol m⁻² d⁻¹ for the SKNT. During the Dutch transect cruise in June 2012, TOU rates ranged from 6.5 to 25.1 mmol m⁻² d⁻¹ (Table 2). Thereby, the highest oxygen uptake rates were measured in the SNS, followed by the SKNT, while the lowest TOU rates were measured in the NNS (mean value SNS: 10.0 mmol m⁻² d⁻¹ in 2011 and 13.5 mmol m⁻² d⁻¹ in 2012; SKNT: 3.9 mmol m⁻² d⁻¹; NNS: 3.4 mmol m⁻² d⁻¹). In general, TOU rates in the SNS are significantly higher than TOU rates of the NNS and the SKNT (Mann–Whitney; *NNS* : $p < 10^{-5}$; SKNT: p < 0.001). Between TOU rates from the NNS and TOU rates from the SKNT no statistical difference could be identified (Mann–Whitney; p = 0.6). Furthermore, TOU rates measured in the SNS in June 2012 are not significantly different from TOU rates measured in September 2011 (Mann–

Whitney; p > 0.05).

Within the permeable sediments of the SNS and NNS, flux chamber incubations were performed at two different stirring speeds (40 and 80 RPM). In general, the TOU rates were about 80 % higher at the higher stirring speed (Fig. 4a), indicating an advective dominated transport within the sediments (Spearman's; p < 0.01). On the other

hand, a correlation between median grain size of the sediments and TOU rates could not be confirmed (Fig. 4b).

In general, A_T and DIC concentrations increased linearly with time in the overlying water of the flux incubations for most stations. A_T fluxes for both campaigns ranged

²⁰ from 0 to 21.4, 0 to 3.0 and 1.4 to 9.9 mmolm⁻² d⁻¹ for the SNS, NNS and SKNT respectively. The highest A_T fluxes were observed in the SNS (mean: 6.5 mmolm⁻² d⁻¹ in 2011 and 5.7 mmolm⁻² d⁻¹ in 2012) and in the SKNT (4.3 mmolm⁻² d⁻¹). The lowest mean A_T flux was calculated for the NNS (1.8 mmolm⁻² d⁻¹). Only the A_T fluxes of the SNS are significantly higher than A_T fluxes of the NNS (Mann–Whitney; SNS: p < 0.001; SKNT: p = 0.2). A_T fluxes measured during the basin wide cruise in 2011 are

similar to those measured along the Dutch transect in 2012 (Mann–Whitney; p = 0.3).

DIC fluxes varied between 1.5–29.1, 0–6.2 and 4.2–7.2 mmol m⁻² d⁻¹ for the SNS, NNS and SKNT respectively. The trends in DIC flux were similar as those obtained for the TOU rates and A_{T} fluxes. The highest DIC fluxes were measured in the SNS,





followed by the SKNT and the NNS (mean: SNS: 11.5 mmol m⁻² d⁻¹ in 2011 and 12.3 mmol m⁻² d⁻¹ in 2012; SKNT: 6.1 mmol m⁻² d⁻¹; NNS: 0.9 mmol m⁻² d⁻¹). DIC fluxes were significantly higher in the SNS than in the NNS, but showed no differences to the DIC fluxes measured in the SKNT (Mann–Whitney; NNS: p < 0.01; SKNT:

p = 0.3), while also no significant difference was found between DIC fluxes of the SKNT and the NNS (Mann–Whitney; p = 0.08). As found for TOU rates and A_T fluxes, the DIC fluxes in the SNS were similar between both campaigns (Mann–Whitney; p = 0.7). This allows us to examine all solute fluxes recorded in the SNS as one single group in the discussion section.

10 3.4 O₂ and pH microprofiling

Representative examples of O_2 and pH depth profiles in stations from all three zones of the North Sea are presented in Fig. 5. In some stations pore water profiles could not be measured due to the coarse grain size and the presence of carbonate shell fragments, which induces a high risk of damaging the microsensors. In general, pore water

O₂ declines as a result of O₂ consumption associated with organic matter degradation. From the O₂ depth profiles the DOU rates of the sediments were calculated as detailed in Sect. 2.5. In the SNS, DOU rates ranged from 0.20 to 6.95 mmol m⁻² d⁻¹, while in the NNS and SKNT, a range of 1.47 to 4.33 mmol m⁻² d⁻¹ and 0.83 to 2.49 mmol m⁻² d⁻¹ were found (Table 3). The lowest mean DOU rate was measured in the SKNT, followed by the DOU rates of the NNS and SNS (SKNT: 1.77 mmol m⁻² d⁻¹, NNS: 2.20 mmol m⁻² d⁻¹; SNS: 2.39 mmol m⁻² d⁻¹).

The oxygen penetration depth (OPD) is defined as the thickness of the oxic zone in marine sediments (Cai and Sayles, 1996), and was operationally defined as the depth below which the O_2 concentration drops below 1 µmol kg⁻¹. In some permeable sediments, the oxygen did not fully deplete over the measured depth profiles (first 2 cm), and so the OPD could not be determined. In the remaining cores, the OPD





was as shallow as 5.3 mm for the SNS, 3.20 mm for the SKNT and 7.10 mm for the NNS.

Table 3 compares the TOU with the corresponding DOU for the basin-wide campaign in September 2011. The TOU/DOU ratio ranged from 50.1 (SNS) to 1.1 (NNS), and DOU rates were significantly lower than TOU rates (Spearman's; p < 0.01), indicating that physical or biological driven advective transport strongly enhanced the sedimentary oxygen uptake.

Depth pH profiles of representative cores are presented in Fig. 5. Depth profiles generally show a decline of pH in all profiled cores, which can be attributed to the release of CO_2 resulting from organic matter degradation. Furthermore, the cores from the NNS and SKNT were typically characterized by a sub-surface minimum in pH, while the depths of these pH minima matched the corresponding OPD. Hence, we interpret these sub-surface pH minima as resulting from the aerobic oxidation of reduced substances transported upwards from deeper sediment layers. Generally, the pH increased again at depth in the cores, most likely due to A_T generation associated with the anoxic degradation of organic matter (see discussion below).

4 Discussion

4.1 Benthic mineralization in the North Sea

Several studies have previously addressed benthic oxygen consumption in the North

- ²⁰ Sea, although most studies are restricted to the southern North Sea (Table 4). The TOU rates found in the present study for the North Sea (range $3.1-28.7 \text{ mmol O}_2 \text{ m}^{-2} \text{ d}^{-1}$) fall within the range of previously published TOU rates (range $0-57.1 \text{ mmol O}_2 \text{ m}^{-2} \text{ d}^{-1}$), which is however large. Due to temporal variability (seasonality), spatial variability (e.g. substrates ranging from cohesive mud to highly permeable sand), but also differences
- ²⁵ in methodology (see discussion below on the impact of stirring rate), it remains difficult to intercompare the oxygen consumption rates that have been obtained in different





studies. So despite this growing database of TOU rates, a more accurate assessment of the spatial and temporal variation of the oxygen consumption in the North Sea remains an outstanding challenge.

- In our basin wide campaign in 2011, we obtained the highest mean TOU rate in the SNS (10.0 mmol m⁻² d⁻¹), followed by the SKNT (3.9 mmol m⁻² d⁻¹), while the lowest TOU rates were measured in the NNS (3.4 mmol m⁻² d⁻¹). One important environmental factor in controlling the TOU rates is the amount of primary production in these different regions. Part of this organic matter produced during photosynthesis is respired in the water column, while the remaining part sinks down to the sediments and undergoes respiration there. As oxygen serves as the ultimate electron acceptor for almost all respired organic carbon (Thamdrup and Canfield, 2000), high primary production rates tend to relate to high TOU rates in the sediments. Note, that besides primary production rates, also the water depth and thereby the amount of organic carbon which
- reaches the sediments is of great importance. Because of the shallow water depth, the fraction of organic carbon which reaches the sediments in the southern North Sea is considerably higher than in the northern North Sea, hence further increasing the TOU rates in this region (Kühn et al., 2010). Based on measured surface chlorophyll concentration in the southern North Sea, Joint and Pomroy (1993) estimated primary production rates for a period from August 1988 to October 1989, and found clear
- ²⁰ gional differences in primary production in the SNS ranging from 18 mmol C m⁻² d⁻¹ along the British Coast to 59 mmol m⁻² d⁻¹ in the German Bight. A model study by Moll (1998) covering the whole North Sea confirmed the large regional variation in depth-integrated annual primary production rates ranging from 21 mmol C m⁻² d⁻¹ in the NNS to 79 mmol C m⁻² d⁻¹ in the German Bight of the SNS. TOU rates observed are con-²⁵ gruent with these previous observational and modelling studies, with high values in
- the southern North Sea, and in particular in the German Bight, to lower values in the northern North Sea.

A second important environmental factor controlling the north–south gradient in TOU rates could be summer bottom water temperature. In a recent seasonal flux study of





shallow coastal sediments from the North Sea, a positive correlation was found between benthic solute fluxes and water temperature (Rao et al., 2014). These authors proposed that higher bottom water temperatures in summer are a main driver for higher benthic fluxes as increased temperatures enhance the metabolic activity in the sedi-

- ⁵ ment. In September 2011, the thermocline was formed between 20 and 50 m (Fig. 2 and Sect. 3.1), and the whole NNS and SKNT were stratified, while in contrast, the SNS was fully mixed (apart from the stations 38 and 45 on the border with the NNS). As a result, the bottom water in the NNS and SKNT was substantially colder (≈ 11°C difference between SNS and NNS/SKNT Table 1). Adopting a temperature Q10 co-
- efficient of ≈ 2 for benthic respiration (Denman and Pena, 2002; Schrum et al., 2006), one hence would expect a doubling of the TOU between the NNS and SNS due to bottom water temperature, all other environmental factors being the same. Accordingly, lower bottom water temperatures in the NNS and SKNT could indeed partially explain the lower TOU values recorded compared to the SNS.
- ¹⁵ To further identify the drivers of the observed spatial variability across the North Sea, we examined the correlation of TOU rates with water depth, porosity and grain size (Fig. 6). We found significant correlations between TOU values and water depth (Spearman's; p < 0.01), suggesting that increased water depth reduced the benthic pelagic coupling in the NNS and SKNT. As more organic matter is remineralized upon
- ²⁰ the longer transit through the water column, less detritus reaches the seafloor, and this hence decreases the contribution of benthic mineralization in the overall respiration. Finally, we also found both a negative correlation between TOU and porosity (Spearman's; p < 0.02) and a significant negative correlation between TOU and median grain size (Spearman's; p < 0.01). This opposes the classical picture of sedimentary diage-
- nesis, where high oxygen consumption rates and intense biogeochemical cycling are typically linked to fine-grained organic rich sediments. Instead, we found the highest TOU values in the permeable sediments of mostly the SNS (low porosity, high median grain size), which supports the more recent ideas of permeable sediments as active bio-catalytic filters (Huettel and Rusch, 2000), which actively trap suspended detritus





by means of advective currents through the upper sediment layers and rapidly mineralize this trapped organic matter. As a result, these permeable sediments display low standing stocks of organic matter, but high TOU values.

The importance of advective transport for the benthic oxygen dynamics is emphasized by the deep oxygenation of the surface sediment at permeable sites (Fig. 5a), the impact of benthic chamber stirring speed on the TOU (Fig. 4), and by the strong difference between TOU values as measured by benthic incubations and the corresponding DOU values as obtained by microprofiling (Table 3). We found that DOU values were always smaller than TOU values for all stations throughout the North Sea. In general, the total oxygen uptake can be decomposed as TOU = DOU + BMU + AMU (Glud, 2008), where BMU represents benthos mediated O_2 uptake resulting from burrow irrigation and the respiration of infauna, and AMU represents the enhancement of the sedimen-

tary O₂ uptake by purely physical transport processes, such as pore water advection induced by currents over bottom topography (Huettel and Rusch, 2000; Meysman et al., 2007) and oscillatory pore water mixing induced by waves (Shum, 1992).

A second signature of physical and or biological pore water irrigation is the deep oxygenation of the surface sediment. As already noted above, in some highly permeable sites, the surface sediment remained completely oxygenated over the whole surface layer that was examined by O₂ microsensor profiling (first 20 mm), and so no oxygen penetration depth (OPD) could be determined. For those stations that did allow to determine the OPD (i.e., the pore water O₂ signal decreased to zero), we employed the analytical model of Cai and Sayles (1996), which provides an inverse relation between OPD and DOU (Fig. 7a) (Eq. 3).

$$L = 2\phi D_s \frac{[O_2]_{bw}}{F_{O_2}^0},$$

1

²⁵ In this expression, *L* is the OPD of the sediment, ϕ is the porosity, and D_s is the effective diffusivity of O₂ in the pore water (i.e., corrected for tortuosity), and $[O_2]_{bw}$. The above relation was used to estimate the theoretical OPD from the measured DOU as shown in





(3)

Fig. 7b. The actually measured OPD is systematically larger than the predicted OPD for all stations. As the model 3 holds for sediments that only experience diffusive transport, the discrepancy suggests that non-diffusive transport (i.e. bio-irrigation or physical advection) increases the oxygen availability and penetration in the sediment (Archer and Devol, 1992).

4.2 Benthic DIC release in the North Sea

5

10

The benthic DIC release in the North Sea follows the same spatial pattern as the benthic oxygen uptake: the highest DIC effluxes were recorded in the SNS (11.5 mmol m⁻² d⁻¹ in 2011 and 12.3 mmol m⁻² d⁻¹ in 2012), followed by the SKNT (6.1 mmol m⁻² d⁻¹) and the NNS (0.9 mmol m⁻² d⁻¹). However, while TOU values were comparable in NNS and SKNT, the DIC flux in the SKNT was significantly higher than in the NNS. As for the TOU, we found a significant positive correlation between the DIC efflux and water depth (Spearman's; p = 0.02), and a weak negative correlation between the DIC efflux and the porosity (Spearman's; p = 0.07) and a significant neg-

ative correlation between the DIC efflux and median grain size (Spearman's; p < 0.01; Fig. 6). Overall, this suggests that the same environmental factors (pelagic primary production, temperature, water depth, sediment permeability) that are driving the TOU are also controlling the spatial pattern of the sedimentary DIC release in the North Sea.

Overall, we found the DIC to be strongly positively correlated with the TOU (Fig. 8;

- ²⁰ Spearman's; p < 0.0001). The ratio between the DIC efflux and the TOU at a given site represents the respiratory quotient (RQ), and we found that the mean RQ value for the SNS (0.95), NNS (0.94) and SKNT (1.00) to be all similar. No significant differences were detected between the three zones (Mann–Whitney; p > 0.05). In general, the RQ can be used as an indicator for the degree of reoxidation of reduced compounds as-
- ²⁵ sociated with anaerobic remineralization (Therkildsen and Lomstein, 1993). Assuming that (1) no carbonate dissolution occurs in the sediment, (2) that the stoichiometry of organic matter follows the Redfield ratio (C : N : P = 106 : 16 : 1; Redfield, 1958), and (3) that organic matter is oxidized using oxygen as the sole electron acceptor, one



would obtain an RQ of 0.77 (solid line in Fig. 8; Paulmier et al., 2009). The slope of the regression in the DIC efflux vs. TOU plot equals 0.71 (Fig. 8a), which is close to Redfield ratio.

In principle, the RQ value is expected to increase with increasing importance of suboxic and anoxic respiration pathways, and subsequent accumulation of reduced compounds such as pyrite, as respiration along these pathways is releasing DIC without consuming oxygen. Alternatively, dissolution of carbonates may also lead to an increased RQ due to the release of two moles DIC per mole CaCO₃ dissolved. Equally, the RQ value is expected to decrease when a stock of reduced compounds is being oxidized, i.e., the annihilation of a previously accumulated *oxygen debt*.

4.3 Benthic alkalinity release in the North Sea

Our results show that North Sea sediments can be a substantial source of A_7 with sediment effluxes ranging from 0 to 28.7 mmol Eqm⁻² d⁻¹ (Fig. 2). The A_7 fluxes reported here were obtained by monitoring the temporal evolution of A_7 in the overlying water of enclosed sediment incubations. In the same sediment incubations, Burt et al. (2014) measured the efflux of the short-lived Radium isotopes (²²⁴Ra and ²²³Ra) and subsequently estimated A_7 fluxes. To this end, these authors used the Ra-isotope data to estimate the overall water exchange rate between the pore water and the overlying water column. This water exchange rate was then multiplied by the excess concentration

- ²⁰ of A_{T} in pore water, which was estimated from pore water analysis. This indirect estimate procedure resulted in benthic A_{T} fluxes between 4.7 and 22.1 mmol Eqm⁻² d⁻¹, and is in very good agreement with the A_{T} flux values obtained here (Fig. 9). The disagreement between observed and calculated A_{T} flux may result from applying the same A_{T} pore–water concentration for all stations in calculating the A_{T} fluxes based
- on Radium-isotope data. This assumption might be wrong and could lead to an overestimation of the A_T flux in station 38. For a more detailed discussion on the correlation between both methods see Burt et al. (2014).





The benthic A_{τ} release in the North Sea follows the same spatial pattern as the benthic oxygen uptake and the DIC efflux, suggesting that organic matter mineralization is the primary driver for the A_{τ} release from sediments. We observed the highest A_{τ} release in the SNS (6.5 mmol m⁻² d⁻¹ in 2011 and 5.7 mmol m⁻² d⁻¹ in 2012), closely followed by the SKNT (4.3 mmol m⁻² d⁻¹). The lowest A_{τ} fluxes were obtained in the NNS (1.8 mmol m⁻² d⁻¹). As for the TOU and DIC efflux, we found a weak positive correlation between A_{τ} efflux and water depth (Spearman's; $\rho = 0.06$), though no significant correlation between A_{τ} efflux and porosity (Spearman's; $\rho = 0.4$) and a significant negative correlation between the A_{τ} efflux and median grain size (Spearman's; $\rho < 0.01$;

- Fig. 6). The lack of negative correlation with porosity is mainly due to SKNT stations, which showed a high A_T efflux at high porosity. Furthermore, the increase of A_T fluxes with increasing TOU rates (Fig. 8) is an indication for metabolic driven dissolution of CaCO₃, as it is fueled by CO₂ released during organic matter mineralization and by the oxidation of reduced compounds produced during anaerobic respiration. Metabolic
- ¹⁵ dissolution would induce a A_T /DIC flux ratio of 1, which is indeed confirmed in some stations, though not for all (Fig. 10). A positive correlation is obtained between A_T and DIC fluxes (Spearman's; p = 0.02), but there are marked differences in the A_T /DIC flux ratio between different stations, which suggests that a different type of biogeochemistry is acting at different locations. Below we will discuss in more detail how different ²⁰ biogeochemical processes in the sediment are releasing A_T and DIC in distinct ratios.

To our knowledge, the A_{T} fluxes presented here are the first field observations of such fluxes in the North Sea area. Thomas et al. (2009) estimated sedimentary A_{T} fluxes in the southeastern bight of the North Sea and the Wadden Sea during autumn based on water column data. In this approach, the sedimentary A_{T} release appeared as the unknown closure term in an A_{T} budget based on water column data, which provided A_{T} flux estimates of about 9.6 mmol m⁻² d⁻¹. These values are similar to the benthic A_{T} fluxes reported here for the SNS.

 A_T release from sediments has also been the subject to a number of studies on other coastal systems than the North Sea. Based on basin-wide budget calculations,

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Gustafsson et al. (2014) estimated that sediments of the Baltic Sea generate A_{T} with a mean rate of 2.4 mmol Eq m⁻² d⁻¹, which is in the same order as the A_{T} input by rivers in that basin. As main sources for A_{T} these authors propose denitrification together with sulfate reduction and/or silicate weathering. Chen and Wang (1999) estimated that sediments in the East China sea generate between 2.9 and 4.9 mmol Eq m⁻² d⁻¹

that sediments in the East China sea generate between 2.9 and 4.9 mmol Eqm ² d A_T . Over 80% of this A_T flux was thereby attributed to iron and sulfate reduction, with no contribution from carbonate dissolution.

Krumins et al. (2013) used a one-dimensional reactive transport model to estimate A_T fluxes from coastal sediments. For that study, the authors divided the global shelf into four different environments, each with distinctive particular organic and inorganic carbon fluxes. Ultimately, they identified non-carbonate shelves (e.g. the North Sea) as an A_T source of 2.7 mmol Eq m⁻² d⁻¹.

On a global scale, Krumins et al. (2013) estimated an A_T flux for the coastal ocean of 29 Tmolyr⁻¹. Whereas this estimate agrees well with an independent estimation by

- ¹⁵ Chen (2002) (16–31 Tmol yr⁻¹), Hu and Cai (2011a) obtained a much smaller flux of 4– 6 Tmol yr⁻¹. Note, that all these three studies are based on different assumption on the underlying processes that are generating A_T . Whereas the A_T flux estimated by Chen (2002) is mainly generated by sulfate reduction with zero contribution of carbonate dissolution, Krumins et al. (2013) acknowledged carbonate dissolution as a benthic A_T
- ²⁰ source. Hu and Cai (2011a) is based on anaerobic A_T generation alone, hence possible contributions by carbonate dissolution are not taken into account. Furthermore, these latter authors treat the coastal sediment–water column system as a single system, while the other two papers only consider the sediment.

To put these different assumptions into perspective, and verify their consequences, we now develop an alkalinity budget for the SNS, in which we first provide a sediment budget, and subsequently, we extend this argument to arrive at a combined sediment and water column alkalinity budget. We restrict this discussion to the SNS, as we have shown in Sect. 3.3 that A_{T} fluxes in the SNS significantly exceed those of the other two zones. High primary production, riverine input of terrestrial organic matter, and





the shallow water depth favor a high respiration rate in the SNS. Furthermore, in the context of atmospheric CO_2 uptake, a direct link between alkalinity release from the sediments and the atmosphere is needed, which requires a non-stratified water column as encountered in most parts of the SNS.

5 4.4 Sources of alkalinity in sediments of the SNS

The high benthic A_{τ} effluxes observed in the SNS invoke the question as to which processes are generating A_{τ} in the sediment. In the following we will discuss how different biogeochemical pathways contribute to the overall A_{τ} release from sediments, and in this way, we will try to assemble a closed A_{τ} budget for the SNS sea floor. To this end, we developed a simplified biogeochemical model of the SNS sediment, which accounts

- ¹⁰ We developed a simplified biogeochemical model of the SNS sediment, which accounts for carbonate dissolution (CD), aerobic respiration (AR), nitrification (NI), denitrification (DN), iron reduction (IR), sulfate reduction (SR), free sulfide oxidation (SO), and pyrite formation (PF) as biogeochemical pathways. The input parameters, diagenetic relations, and output variables are summarized in Table 5. The A_T /DIC flux ratio for each
- of these reactions is indicated as a straight line in Fig. 10. In the next paragraphs, we now discuss how the rates of the individual reactions can be constrained based on our flux measurements and literature data.

In theory, the dissolution of one mole of $CaCO_3$ releases two moles of A_7 , assuming no other processes are acting.

²⁰ CaCO₃
$$\rightarrow$$
 CO₃²⁻ + Ca²⁺

25

The undersaturation of the pore water with respect to the phases of calcium carbonate present (e.g. high Mg calcite, aragonite or calcite) determines the rate at which carbonate will dissolve. In general the undersaturation of the pore water can have two causes (Boudreau et al., 2010): (1) undersaturation of the overlying water and (2) additional undersaturation of the pore water due to metabolic respiration. In the North Sea, the overlying water is always oversaturated with respect to all common carbonate phases (Frankignoulle et al., 1998), and so, dissolution of CaCO₃ in the sediment must



(4)



be exclusively metabolically driven. Both the CO_2 production during organic matter mineralization as well as the oxidation of reduced compounds produced during anaerobic respiration processes can make the pore water undersaturated with respect to carbonates, and therefore, fuel metabolic driven dissolution (Jahnke et al., 1994). The

⁵ increase of the A_{T} fluxes with increasing TOU rates (Fig. 8) forms indeed an indication for metabolic driven dissolution of CaCO₃ (Jahnke and Jahnke, 2004).

From the flux dataset available here, it is not possible to constrain the actual CaCO₃ dissolution rate in each station. However, it is still possible to provide an upper limit for the CaCO₃ dissolution rate on a larger geographic scale. Gazeau et al. (personal communication) recently estimated carbonate production in the southern North Sea to be 2.7 mmol Cm⁻² d⁻¹. Net carbon burial does not occur in the North Sea, except for small amounts in the Skagerrak and the Norwegian Channel (de Haas et al., 2002; Thomas et al., 2005; Bozec et al., 2006). Therefore, we consider carbonate production and dissolution to be in steady state. Accordingly, carbonate dissolution in sediments can release up to 5.4 mmol Eq m⁻² d⁻¹ of A_T .

In addition to carbonate dissolution, A_T generation can be linked to various organic matter degradation pathways and secondary re-oxidation reactions. In the presence of oxygen, aerobic respiration is used to break down organic matter:

 $OM + 15H^{+} + 106O_{2} \rightarrow 106CO_{2} + 16NH_{4}^{+} + H_{2}PO_{4}^{-} + 106H_{2}O$

²⁰ where we assume organic matter (OM) to be of Redfield elemental composition (Redfield, 1958). This reaction essentially generates A_T by the consumption of protons linked to ammonium release, and to a lesser extent, it consumes A_T by the release of phosphate. Thamdrup and Canfield (2000) have estimated that aerobic respiration accounts for between 5–25% of the total benthic mineralization in shelf and coastal sediments. If we adopt a value of the benthic mineralization rate $R_{min} = 10.0 \text{ mmol Cm}^{-2} \text{ d}^{-1}$, which is the mean TOU value recorded for the SNS in September 2011 (Table 2; see also discussion below), we can estimate the aerobic respiration rate to range between 0.50 and 2.50 mmol Cm $^{-2} \text{ d}^{-1}$ for the SNS (mean:



(5)



1.50 mmol C m⁻² d⁻¹). Accounting for stoichiometry (A_T/O_2 ratio = 15/106), the corresponding A_T release ranges from 0.07 to 0.35 mmol Eq m⁻² d⁻¹, with a mean of 0.21 mmol Eq m⁻² d⁻¹.

Nitrification of ammonium released during mineralization consumes A_T and can be represented by the reaction equation:

 $NH_4^+ + 2O_2 \rightarrow NO_3^- + H_2O + 2H^+$

10

Per mole of ammonium that is released during mineralization, exactly 2 mole of A_{τ} is consumed. If we use the TOU as a proxy for R_{\min} , the rate of ammonification (i.e. 16/106 R_{\min}) becomes 1.51 mmolNm⁻²d⁻¹ for the SNS. If all the ammonium is re-oxidized, and hence no ammonium escapes the sediment, the nitrification rate should match the ammonification rate, and so the associated A_{τ} consumption becomes 3.02 mmolEq m⁻²d⁻¹.

Denitrification is a second mineralization pathway via which organic matter degradation proceeds in the sediment, and can be represented as:

¹⁵ OM + 84.8NO₃⁻ + 99.8H⁺
$$\rightarrow$$
 106CO₂ + 42.4N₂ + 16NH₄⁺ + H₂PO₄⁻ + 106H₂O (7)

Per mole of organic matter denitrified, the A_T increases by 0.94 = 99.8/106 mole. To discuss the impact of denitrification on sedimentary A_T release, we need to consider the origin of the nitrate that is used for denitrification. A portion of the nitrate is internally generated in the sediment through coupled nitrification/denitrification, while an-

²⁰ other part of the nitrate is derived externally (i.e., from the overlying water column). It has been estimated that these coupled nitrification/denitrification reactions account for 80 % of the total denitrification rates in coastal environments (Middelburg, 1996; Seitzinger et al., 2006), which thus implies that the total denitrification rate should scale as 1/0.8 times the nitrification rate. This way, we obtain a denitrification rate of 1.89 mmol C m⁻² d⁻¹, with an associated A_7 release of 1.78 mmol Eq m⁻² d⁻¹. In the North Sea, a tight coupling of nitrification and denitrification has indeed been reported



(6)

by Lohse et al. (1993) and Pätsch and Kühn (2008), and observed benthic denitrification rates in the SNS range from 1.6 to 4.6 mmol C m⁻² d⁻¹ (Lohse et al., 1996; Hydes et al., 1999), a range that centrally embraces our estimate here. Note that the coupling of ammonification, nitrification and subsequently denitrification does not lead to net ⁵ production of A_{τ} . Only if nitrate is derived from the overlying water column, one obtains a net production of A_{τ} in the pore water (Hu and Cai, 2011a).

Once nitrate and oxygen are fully consumed, dissimilatory iron reduction and sulfate reduction are the prevailing respiration pathways (in the SNS we assume manganese oxides to be a minor electron acceptor, and so we ignore this pathway).

¹⁰ OM +
$$53SO_4^{2-}$$
 + $121H^+ \rightarrow 53H_2S + 106CO_2 + 16NH_4^+ + H_2PO_4^- + 106H_2O$ (8)

 $OM + 424FeOOH + 863H^+ \rightarrow 424Fe^{2+} + 106CO_2 + 16NH_4^+ + H_2PO_4^- + 106H_2O_4^-$ (9)

Both reactions are consuming protons, and hence producing A_{T} . Per mole organic carbon oxidized, sulfate- and iron reduction are releasing 1.14 and 8.14 moles of A_{τ} respectively. However, to assess the *net* A_{T} generation in the sediment, we need to take the fate of the reduced species into account. The free sulphide that is generated 15 by sulfate reduction can follow two major pathways. Some of the free sulphide (fraction p) will react with the reduced iron liberated by iron reduction to form pyrite. Due to its thermodynamic stability, pyrite is considered to be the primary sink for both iron and sulfur on the time scales of early diagenesis (Hu and Cai, 2011a). The overall process of pyrite formation can be represented by the reaction equation:

$$\frac{1}{2}Fe^{2+} + \frac{1}{4}O_2 + H_2S \rightarrow \frac{1}{2}FeS_2 + H^+ + \frac{1}{2}H_2O$$
(10)

20

The remaining part of the free sulphide generated by sulfate reduction (fraction 1 - p) is transported upwards towards the oxic zone and re-oxidized with oxygen

 $H_2S + 2O_2 \rightarrow SO_4^{2-} + 2H^+$ (11)



Both pyrite formation and free sulfide reoxidation consume alkalinity. However, when combined with the alkalinity impact of sulfate reduction, pyrite formation results in a *net* generation of alkalinity, while the alkalinity consumed in free sulfide reoxidation exactly compensates the alkalinity generated during sulfate reduction. Thamdrup et al. (1994)

- calculated for sediments of the Aarhus Bay that the burial of reduced sulfur accounts for only 20% of the total sulfate reduction rate, and similar values are reported in other studies (Jørgensen, 1977; Berner and Westrich, 1985; Jørgensen et al., 1990). The sediments of SNS are generally coarser and more permeable than those of Aarhus Bay, which leads to more advection and reoxygenation, which favors sulfide reoxidation
 as opposed to pyrite formation. Therefore, here we adopt a reduced degree of pyrite
 - formation (p = 0.1; Table 1).

Adopting steady state, our simplified diagenetic model provides a set of linear relations (Table 5), which generate a particular division of the total organic matter mineralization into aerobic respiration (15%), denitrification (24%), iron reduction (0.4%)

- and sulfate reduction (61 %). These estimates are comparable to the model analysis of Krumins et al. (2013), who estimated that sulfate reduction is the main respiratory pathway on continental shelves, accounting for 77–85 % of the organic matter mineralization, followed by aerobic respiration (16 %), denitrification (2–4 %) and iron-reduction (0.3–0.7 %). Similarly, Nedwell et al. (1993) estimated that sulfate reduction accounted
- for 10–53% of the total organic matter mineralization in the SNS, while a coupled benthic–pelagic model for the North Sea suggests that up to 30% of the organic matter is mineralized using sulfate as the electron acceptor (Luff and Moll, 2004). The TOU rate predicted by the model (10.1 mmol $O_2 m^{-2} d^{-1}$) agrees well with the mean TOU observed in the SNS in September 2011 (10.0 mmol $O_2 m^{-2} d^{-1}$), but the predicted res-
- ²⁵ piratory coefficient (RQ = 1.26) is slightly higher than the observed one (RQ = 1.15). This higher simulated RQ could be due to an overestimate of carbonate dissolution rate, which must be reduced to 1.6 mmol C m⁻² d⁻¹ to match the observed RQ = 1.15. However, such a reduction of the carbonate dissolution rate would at the same time



lead to a substantial underprediction of the sedimentary alkalinity release, and so we stick to the carbonate dissolution rate $CD = 2.7 \text{ mmol C m}^{-2} \text{ d}^{-1}$.

Accounting for the contributions of all biogeochemical reactions, the total alkalinity generation in the sediment R_{sed} becomes (Fig. 11):

$$R_{\text{sed}} = 2r_{\text{CD}} + \frac{16-1}{106}r_{\text{AR}} + \frac{84.8+16-1}{106}r_{\text{DNF}} + \frac{848+16-1}{106}r_{\text{IR}} + \frac{106+16-1}{106}r_{\text{SR}}$$
(12)
$$-2r_{\text{NIT}} - 2r_{\text{SO}} - r_{\text{PF}},$$

As we assume no alkalinity flux to deeper sediment layers, the total alkalinity generation in the sediment should match the efflux of alkalinity across the sediment—water interface. The model predicts a total alkalinity production $R_{sed} = 5.96 \text{ mmol Eqm}^{-2} \text{ d}^{-1}$, which is indeed in good agreement with the mean A_T flux of 6.50 mmol Eqm $^{-2} \text{ d}^{-1}$ as measured in our flux incubations within the SNS. Positive contributions to the A_T efflux are due to carbonate dissolution, aerobic respiration, denitrification, sulfate reduction and iron reduction, whereas A_T is consumed during nitrification, sulfur oxidation and pyrite formation. The most dominant A_T producing reaction is carbonate dissolution, followed by sulfate reduction and denitrification. The most dominant A_T consuming reactions are sulfide oxidation and nitrification. The A_T contribution of iron reduction is small and balances that of pyrite formation.

4.5 System-wide alkalinity budget

Above we have documented how the sediments of the SNS can be a source of A_{T} for the water column of the SNS. However, before we can estimate the resulting effect of this benthic A_{T} release on the pCO_{2} dynamics of the southern North Sea, we first need to assess the A_{T} balance of the water column. An efflux of A_{T} from the sediment does necessarily result in an increase of A_{T} whole system, as in the water column, some biogeochemical processes are opposing the A_{T} generation in the sediment. Here, we identify carbonate formation (CF), primary production (PP), aerobic respiration (AR)





and nitrogen fixation (NF) as processes that potentially produce or consume alkalinity in the water column.

As shown above, the dissolution of carbonates in the sediment (Eq.) forms a strong source of A_T , but the production of carbonates in the water column has the opposite ⁵ effect on A_T .

$$Ca^{2+} + 2HCO_3^- \rightarrow CaCO_3 + CO_2 + H_2O$$

If the annual production of carbonate in the water column matches the carbonate dissolution in the sediments, there will be no net generation of A_T on a system wide scale over a whole seasonal cycle. Note however that when there is a temporal shift in production vs. dissolution of carbonates, these processes may still have an impact on the coastal A_T balance on shorter time scales. Here we assume that no net burial or export of solid carbonates occurs in the SNS on an annual scale (de Haas et al., 2002; Thomas et al., 2005; Bozec et al., 2006). Accordingly, the annual benthic carbonate dissolution must be fully compensated by carbonate production in the water column, hence carbonate production in the water column will consume 5.4 mmol Eq m⁻² d⁻¹ of A_T .

A second process which consumes alkalinity in the water column is primary production. This occurs mostly pelagic, but also benthic in some very shallow waters of the SNS. If we assume that nitrate is the main nitrogen source, primary production can be described by the reaction equation:

$$106CO_2 + 16NO_3^- + H_2PO_4^- + 122H_2O + 17H^+ \to OM + 138O_2$$
(14)

20

Joint and Pomroy (1993) estimated primary production in the southern North Sea to be $199 \text{gCm}^{-2} \text{yr}^{-1}$, or equally, $45.45 \text{ mmol} \text{Cm}^{-2} \text{d}^{-1}$, thus producing 7.28 mmol Eq m⁻² d⁻¹ of A_T .

²⁵ Aerobic respiration in the water column can be described by the opposite reaction equation of primary production:

$$OM + 138O_2 \rightarrow 106CO_2 + 16NO_3^- + H_2PO_4^- + 122H_2O + 17H^+$$

12423



(13)

(15)

To estimate the total respiration rate we need to account for two different sources of the organic matter that is being respired. The majority of the organic matter is derived from local primary production, while a smaller part originates from riverine input of terrestrial compounds. It has been estimated that about 80 % of the locally produced organic mat-

- ⁵ ter respires in the water column, whereas the remaining 20% sinks down to the sediments (Radach and Moll, 1993; Moll, 1998). The value of $R_{min} = 10 \text{ mmol } m^{-2}d^{-1}$ as derived in the previous section, compare well with this assessment, as it comes down to 22% of the PP value of 45 mmol C m⁻² d⁻¹ as estimated above. The pelagic mineralization rate derived from local net primary production hence becomes 35 mmol C m⁻² d⁻¹.
- ¹⁰ Additionally, organic matter input from land also fuels respiration, either in the water column or in the benthic compartment. The total riverine input of organic matter in the SNS was estimated by Kühn et al. (2010) to be in the range of 1.0 mmol Cm⁻² d⁻¹, which is hence small compared to the local net primary production. Accordingly, the total respiration rate in the water column must be 36.45 mmol Cm⁻² d⁻¹, thus consuming 5.85 mmol Eq m⁻² d⁻¹ of A_T .

Finally, the loss of fixed nitrogen due to denitrification in the sediment can be compensated by nitrogen fixation in the water column. However, the salinity of the North Sea is too high for diazotrophic cyanobacteria and too cold for open ocean cyanobacteria (Stal, 2009). Thus, we consider N_2 fixation in the North Sea to be negligible.

Similar as was done for the sediment, we can define the total alkalinity generation in the water column R_{wc} as:

$$R_{\rm wc} = \frac{17}{106} r_{\rm PP} - \frac{17}{106} r_{\rm AR} - 2r_{\rm CF} - r_{\rm NF},$$

20

which hence leads to a total consumption of 3.96 mmol Eqm⁻² d⁻¹ of A_{T} .



(16)

Based on the analysis above, we can now write an alkalinity balance for combined the sediment and water column of the SNS:

$$V_{\text{sed}} \frac{dA_T^{\text{sed}}}{dt} = -F_{\text{sed}} + R_{\text{sed}}$$
(17)
$$V_{\text{wc}} \frac{dA_T^{\text{wc}}}{dt} = F_{\text{river}} + F_{\text{sed}} - F_{\text{out}} + R_{\text{wc}}$$
(18)

⁵ In steady state, the net generation of alkalinity amounts to $R_{net} = R_{sed} + R_{wc} = 5.96 - 3.96 = 2.00 \text{ mmol Eq m}^{-2} \text{d}^{-1}$ or 139 Gmol Eq yr⁻¹. As shown in Table 5, 33% can be attributed to sulfur and iron cycling (i.e., anoxic mineralization taking place in the sediment) and the remaining 68% is linked to nitrogen cycling (this part is dominated by denitrification in the sediment). Furthermore, the riverine input of alkalinity was estimated by Pätsch and Kühn (2008) as $F_{river} = 1.5 \text{ mmol Eq m}^{-2} \text{d}^{-1}$, based on a river load compilation for the years 1977 to 2002. Accordingly, the A_T export flux from the SNS to the rest of the North Sea (SKNT and NNS) is $F_{out} = F_{river} + R_{sed} + R_{wc} = 1.5 + 5.96 - 3.96 = 3.50 \text{ mmol m}^{-2} \text{d}^{-1}$ or 266 Gmol yr⁻¹.

As shown in Table 5, the different terms that contribute to net generation of alkalinity in the SNS have been attributed to four different types of elemental cycling (carbon, nitrogen, sulphur + iron, phosphate). This analysis reveals that no alkalinity is associated with carbon and phosphate cycling, but that 33 % can be attributed to sulfur and iron cycling (i.e., anoxic mineralization taking place in the sediment) and the remaining 68 % is linked to nitrogen cycling (this part is dominated by denitrification in the sediment). Sedimentary denitrification is hence the most prominent driver of net alkalinity generation in the SNS. Applying our estimated mean denitrification rate (1.9 mmol N m⁻² d⁻¹) to the whole SNS, this implies a removal of 133 Gmol N yr⁻¹ of bioavailable nitrogen, which closely matches the denitrification estimate of Pätsch and Kühn (2008) of 119 Gmol N yr⁻¹ for the whole North Sea. Accounting that the sediments

²⁵ of the SNS are the prime locations for denitrification, our estimates of denitrification







rates and the associated alkalinity generation rate in the SNS seems to be in line with previous studies.

4.6 Impact on water column pCO₂ dynamics of the SNS

Figure 12 shows the associated DIC and A_{τ} budget of the water column in SNS. The ⁵ direction of a CO₂ flux between the surface water and the atmosphere is determined by the pCO_2 gradient between water and atmosphere, which is ultimately governed by the ratio of internal DIC over internal A_{τ} release (Frankignoulle, 1994; Egleston et al., 2010; Hu and Cai, 2011b). Assuming mean values of the SNS for the time of sampling (salinity: 34 PSU, temperature: 16.3 °C, A_7 : 2270 µmol kg⁻¹) and an atmospheric CO_2 concentration of 392 ppm, one mole A_7 can take up 0.85 mole atmospheric CO_2 . As shown above, the net generation of alkalinity amounts to $2.0 \text{ mmol Eq m}^{-2} \text{ d}^{-1}$, and hence, 1.70 mmol $Cm^{-2}d^{-1}$ of CO_2 can be taken up. However, as noted above, riverine input of organic matter and subsequent respiration of this organic matter releases 1.0 mmol $Cm^{-2}d^{-1}$ of DIC. Therefore, the total atmospheric CO₂ uptake due to internal A_T generation in the SNS amounts to $1.70-1.0 = 0.70 \text{ mmol Eg m}^{-2} \text{ d}^{-1}$ or 15 48 Gmol Cyr⁻¹, which is around 6% of the total CO₂ uptake of the entire North Sea (Thomas et al., 2004; Egleston et al., 2010). This value is lower than the estimate by Thomas et al. (2009), who calculated that sedimentary A_{τ} can potentially facilitate up to 25% of the total CO₂ uptake of the North Sea. This latter estimate was not based on direct flux measurements, but derived from an A_{T} budget for the North Sea, where 20 A_{T} generation in Wadden Sea sediments was introduced as a closure term.

The idea that internal alkalinity generation in the SNS can drive atmospheric CO_2 uptake is further supported by previous CO_2 studies, although significant seasonality has been observed in the atmospheric CO_2 uptake. Thomas et al. (2005) re-

²⁵ ported an uptake of CO₂ for the months February to August ($-0.52 \text{ mmol Cm}^{-2} \text{d}^{-1}$), while the SNS was reported to be a source for CO₂ between September and January ($+0.60 \text{ mmol Cm}^{-2} \text{d}^{-1}$). Bozec et al. (2005) described the SNS as a strong source for CO₂ with fluxes of 0.8 to 1.7 mmol Cm⁻² d⁻¹ in late summer, while on an annual scale, the SNS was characterized as a sink of CO₂ with an air–sea flux of -0.55 mmol Cm⁻² d⁻¹. Due to the temporal variability of CO₂ fluxes, and the seasonal cycles of primary production and respiration, it is hard to compare these numbers with our calculated CO₂ air–sea flux of -0.7 mmol Cm⁻² d⁻¹, but the estimated impact on the *p*CO₂ dynamics of the SNS by benthic A_7 release appears to be in the same order of magnitude as determined in previous studies of the *p*CO₂ dynamics of the SNS. Overall, the A_7 release from sediments, driven by denitrification and anoxic respiration of organic matter, seems to play an important role in the atmospheric CO₂ uptake of the North Sea.

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¹⁵ data of Fig. 9 and further input regarding to sediment—water exchange rates. This manuscript has benefited significantly from fruitful discussions with Mathilde Hagens. This work was supported by the ZKO program of the Netherlands Organisation for Scientific Research (NWO) and ERC grant 306933 to F. J. R. Meysman. U. Braeckman was financially supported by FWO project nr G.0033.11.

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sedimentary alkalinity release on the water column CO₂ system

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Table 1. Sampled stations from both cruises in 2011 and 2012 with coordinates and maximum water depth, as well as with sediment characteristics and bottom water characteristics as measured by the CTD.

Station	Latitude	Longitude	Water depth	Temperature	Salinity	Oxygen Rottom water	Porosity	Median	Sediment classification
			(m)		DOLLOIN WALEI	(umolka ⁻¹)	(vol/vol)	(um)	(Montworth)
			(11)	(0)	(F30)	(µmorky)	(00/001)	(μπ)	(wentworth)
2011	North Sea	basin-wide							
2	51.52533	1.96633	40.66	17.41	34.94	153.49	-	499	medium sand
7	52.60000	3.50117	29.84	17.11	35.06	153.80	0.32	292	medium sand
11	53.19983	2.50083	30.53	16.33	34.51	157.29	0.32	287	medium sand
17	53.80033	4.00050	29.95	17.03	34.14	150.84	0.35	237	fine sand
20	54.40033	8.10050	18.23	17.16	29.35	151.10	0.32	209	fine sand
30	55.00000	4.99950	39.40	15.29	34.76	155.67	0.37	150	fine sand
32	55.00000	1.99983	22.99	14.25	34.63	154.34	0.32	215	fine sand
38	1.99983	1.99983	81.09	7.32	34.45	140.10	0.42	138	fine sand
42	56.00167	7.50033	20.19	16.29	33.87	154.17	0.31	219	fine sand
45	57.00000	5.25100	53.96	7.02	35.01	146.19	0.33	287	medium sand
52	57.50050	7.50150	209.65	6.18	35.18	165.70	0.63	62	coarse silt
56	58.00000	0.49933	109.03	9.29	35.37	143.73	0.46	119	fine sand
59	58.00050	4.25100	95.23	7.66	35.21	151.43	0.31	314	medium sand
62	58.00000	9.50083	304.24	6.03	35.15	169.51	0.64	36	coarse silt
65	58.49983	9.49983	539.12	6.02	35.18	172.03	0.74	28	medium silt
71	59.00033	2.50000	115.11	6.87	35.27	148.17	0.40	149	fine sand
80	60.00017	0.50000	116.46	8.90	35.39	150.42	0.35	283	medium sand
88	61.00000	3.49950	350.27	6.29	35.39	150.42	0.35	283	medium sand
91	60.99950	0.50033	141.16	7.60	35.33	146.49	0.44	269	medium sand
2012	Dutch coastal transect								
1b	54.43190	3.40820	43.92	8.68	34.64	267.63			
2b	54.24760	4.02550	44.65	9.51	34.73	260.38			
3b	54.08090	4.20060	47.58	11.21	34.67	253.80			
4b	53.55040	4.36280	35.55	12.21	34.74	264.95			
5b	53.82750	4.52470	41.00						
6b	53.41270	4.28640	36.33	12.43	34.55	272.63			
7b	53.42550	4.48820	36.66						



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		September 2011		June	2012	Literature	
		Mean	Range	Mean	Range	Mean	Range
TOU	SNS	10.0 ± 2.4	3.1–28.7	13.5 ± 4.5	6.5–25.1	14.9 ^a	5.2–28.4 ^a
	NNS	3.4 ± 0.4	0.7–6.2	-	-	_	-
	SKNT	3.9 ± 0.4	2.9–5.7	_	-	_	_
A_{T}	SNS	6.5 ± 5.2	0–21.4	5.7 ± 3.7	0.5–18.7	9.6 ^b	-
flux	NNS	1.8 ± 2.4	0–3	_	_	_	_
	SKNT	4.3 ± 3.3	1.4–9.9	_	_	_	_
DIC	SNS	11.5 ± 5.4	1.5-29.1	12.3 ± 2.6		_	_

Table 2. Fluxes at the sediment–water interface for O_2 (TOU), A_T and DIC.

^a Hydes et al. (1999).

NNS

SKNT

 0.9 ± 2.1

 6.14 ± 2.6

flux

^b Thomas et al. (2009). Units mmol Cm⁻² d⁻¹ (DIC flux); mmol Eqm⁻² d⁻¹ (A_7 flux); mmol O₂ m⁻² d⁻¹ (TOU rate).

0-6.2

4.2-7.4

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Table 3. Comparison between TOU rates as measured by FireSting optodes and diffusive oxygen uptake rates (DOU) obtained from O_2 microprofiles.

Station	TOU	DOU	OPD
11	10.02	0.20	NA
20	22.40	2.07	5.30
30	11.33	6.95	1.10
38	3.73	1.06	13.30
45	7.32	1.67	NA
52	5.49	0.83	17.00
56	3.76	1.47	NA
59	4.94	4.33	3.20
62	4.80	2.49	NA
65	3.01	1.88	15.10
71	3.67	1.76	7.10
80	4.62	1.25	8.80
88	2.25	1.87	14.80

Unit: $mmolO_2 m^{-2} d^{-1}$; oxygen penetration depth (OPD) for cores used for microprofiling in mm. NA indicates that oxygen does not deplete over the measured depth.

Table 4. TOU rates $(mmol O_2 m^2 d^{-1})$ measured in the North Sea and other coastal systems taken from literature compared to this study.

Literature	Location	Month and year	TOU
North Sea			
Wilde et al. (1984)	OG	5, 8, 9 (1980, 1981)	3.6-14.4
Cramer (1990)	FF	5, 6 (1986), 8, 9 (1987)	23.3-51.8
Raaphorst et al. (1990)	DB	7, 8 (1988)	4–20
	BF	1, 4, 5, 8, 11 (1989)	2-22
van Duyl et al. (1992)	FF	1, 4, 5, 8, 11 (1989)	15-40
Upton (1993)	U1	9, 10 (1988), 2, 4, 6, 8, 9 (1989)	5-16
	FF	9, 10 (1988), 2, 4, 6, 8, 9 (1989)	5-28
	U3	9, 10 (1988), 2, 4, 6, 8, 9 (1989)	7–25
	U4	9, 10 (1988), 2, 4, 6, 8, 9 (1989)	10-11
	U5	9, 10 (1988), 2, 4, 6, 8, 9 (1989)	6-10
	U6	9, 10 (1988), 2, 4, 6, 8, 9 (1989)	7–18
Lohse et al. (1996)	OG	7 (1994)	5.6-6.1
Osinga et al. (1996)	BF	2 (1993), 7, 10 (1994)	< 24
	OG	2 (1993), 7, 10 (1994)	< 63
Boon et al. (1998)	BF	2, 3, 4, 6, 8, 11 (1993)	< 19.2
	FF	2, 3, 4, 6, 8, 11 (1993)	< 48
Trimmer et al. (2000)	Thms	7, 10 (1996), 4, 7 (1997)	11.4-5.8
Trimmer et al. (2005)	USP	10 (2001), 7 (2002)	16.1-57.1
	Thms	10 (2001), 7 (2002)	14-43.9
Weston et al. (2008)	OG	9 (2003)	12.6-30.6
Franco et al. (2010)	F115bis	2, 4, 10 (2003)	5.5-18.8
	F330	2, 4, 10 (2003)	1.2-8.7
Provoost et al. (2013)	F115	9, 10, 11, 12 (2002), 1, 2, 3, 4, 5, 7, 8, 9, 10 (2003)	4.5-32.9
Braeckman et al. (2014)	BM	2, 3, 4, 5, 6, 7, 8, 9 (2011)	0-41.92
	BFS	2, 3, 4, 5, 6, 7, 8, 9 (2011)	< 56.31
	BFS	10 (2011)	43.88
	BS	2, 3, 4, 5, 6, 7, 8, 9 (2011)	< 22.94
	BS	10 (2011)	6.63
Other coastal systems			
Lansard et al. (2008)	MS	06 (2001 and 2002)	3.9-25.6
Nordi et al. (2013)	FS	04 05 06 07 08 (2011) 02 06 07 (2012)	3.3-6.8
Archer et al. (1992)	WS	06 (1988)	1.0-18.3
Grebmeier and McRoy (1989)	BC	07 08 09 (1984–1986)	0.3-16.9
Smith Jr (1974)	SD	10 (1973)	0.4-3.9
Boetius and Damm (1998)	AC	08 09 (1993)	0.2-2.3
Witte and Pfannkuche (2000)	AS	10 (1995)	0.9-6.3
Devol and Christensen (1993)	WS	06 07 (1988) 06 (1991)	2.9-18.5
Berelson et al. (2003)	MB	06 (1991)-10 (1995)	5.1-13.5
Friedl et al. (1998)	BS	Summer (1995)	0.0-33.0
Hartnett et al. (2008)	AP	03 06 10 (2000) 02 (2001)	1.5-2.1
Larsen et al. (2013)	CS	07 (2008	5.8-9.0
This study	SNS	9 (2011)	3.12-28.65
	SNS	4 (2012)	6.50-25.11
	NNS	9 (2011)	0.74-6.20

OG: Oyster Ground, FF: Frisian Front, DB: Dogger Bank, BF: Broad Fourteens, U1–U3: (near) English Channel, U4–U6: Central North Sea, F115bis, F330, BM, BFS, BS: Belgian Coast, Thms: Thames, USP: Outer Silver Pit, MS: Mediterranean Sea, FS: Faroe Shelf, WS: Washington Shelf, BC: northern Bering and Chukchi Seas, SD: San Diego Trough, AC: Arctic Continental Slope, AS: Arabian Sea, MB: Monterey Bay, BS: Black Sea, AP: western Antarctic Peninsula, CS: Celtic Sea, SNS: southern North Sea, NNS: northern North Sea.

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Table 5. Input parameters and rate expressions for all diagentic and pelagic processes including in our model. Additionally, the impact of the different elemental cycles are expressed in percentages.

Input parameter	Expression	Value	Units	A_T release
Mineralization rate	Rmin	10.0	mmol C m ^{-2} d ^{-1}	-
Carbonate dissolution rate	CD	2.70	$mmol C m^{-2} d^{-1}$	-
Aerobic respiration fraction	a	0.15	-	-
Fraction of DNF supported by NI	b	0.8	-	-
Pyrite formation fraction	p	0.2	-	
Pelagic processes				
Primary production	PP	45.40	mmol C m $^{-2}$ d $^{-1}$	+7.28
Aerobic respiration	AR	36.45	$mmol C m^{-2} d^{-1}$	-5.85
Carbonate formation	CF	2.70	mmol C m ⁻² d ⁻¹	-5.40
Nitrogen fixation	NF	0	$mmol C m^{-2} d^{-1}$	0
Alkalinity generation	See text		$mmol Eq m^{-2} d^{-1}$	-3.96
Diagenetic processes				
Carbonate dissolution	CD	2.70	mmol C m ⁻² d ⁻¹	+5.40
Aerobic respiration	$AR = a \cdot Rmin$	1.50	$mmol C m^{-2} d^{-1}$	+0.21
Nitrification	NIT = $\frac{16}{400}$ · Rmin	1.51	mmol C m ⁻² d ⁻¹	-3.02
Denitrification	$DNF = \frac{1}{20} \cdot \frac{1}{5} \cdot NIT$	1.89	$mmol C m^{-2} d^{-1}$	+1.78
Sulfate reduction	$SR = \frac{Bmin-AR-DNF}{1+1}$	6.57	$mmol C m^{-2} d^{-1}$	+7.50
Pyrite formation	$PF = 0.5 \cdot p \cdot SR$	0.33	mmol C m $^{-2}$ d $^{-1}$	-0.33
Sulfide oxidation	$SO = 0.5(1 - p) \cdot SR$	2.96	$mmol C m^{-2} d^{-1}$	-5.91
Iron reduction	$IR = \frac{1}{16} \cdot p \cdot SR$	0.04	$mmolCm^{-2}d^{-1}$	+0.33
Alkalinity generation	see text		mmol Eq m ⁻² d ⁻¹	+5.96
Total oxvgen uptake	$TOU = AR + 2 \cdot NIT + 2 \cdot SO + 0.25PF$	10.1	$mmol O_2 m^{-2} d^{-1}$	
Respiratory quotient	$RQ = \frac{Rmin+CD}{TOU}$	1.26	2	
Linked to				
Carbon cycle	2.CD - 2.CF	0	%	-
Nitrogen cycle	84.8/106 · DNF + 16/106 · (AR + DNF + IR + SR) – 2 · NIT + (16/106) · (PP – WAR)	68	%	-
Sulphur and iron cycle	8 · IR + SR – 2 · SO – PF	33	%	-
Phosphor cycle	–1/106 · (AR + DNF + IR + SR) + (1/106) · (PP – WAR)	0	%	-

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Figure 1. Map of sampled stations. Red symbols: southern North Sea, black symbols: Skagerrak, blue symbols: northern North Sea, circles: sampled in September 2011, squares: sampled in June 2012. Border of the Skagerrak as defined by the International Hydrographic Organization, border between SNS and NNS roughly represents the 100 m depth isoline.





Figure 2. Temperature profiles as recorded by a CTD cast for: **(a)** Station 11 (SNS); **(b)** Station 45 (SNS); **(c)** Station 65 (SKNT); Station 80 (NNS).







Figure 3. Representative concentrations of A_7 , DIC and O₂ saturation over time for all three regions of the North Sea. Red: Station 11 (SNS); blue: Station 71 (NNS); black: Station 65 (SKNT).







Figure 4. (a) Total oxygen uptake rates measured in benthic incubation chambers at two different stirring rates (RPM = rounds min⁻¹); **(b)** TOU rates in correlation to the median grain size. Dashed line: 1 : 1 line; solid line: linear regression; red symbols: SNS; blue symbols: NNS. All rates in mmol $O_2 m^{-2} d^{-1}$.







Figure 5. Examples of microsensor depth profiles from different regions of the North Sea. **(a)** O₂; **(b)** pH; red: Station 11 (SNS); blue: Station 65 (SKNT); black: Station 80 (NNS); solid line: sediment–water interface.





Figure 6. Correlation plots between DIC-, A_T -fluxes, TOU rates and water depth, porosity and grain size. Red: SNS, black: SKNT, blue: NNS; Spearman's p value given in each plot. Units mmol C m⁻² d⁻¹ (DIC); mmol Eq m⁻² d⁻¹ (A_7); mmol O₂ m⁻² d⁻¹ (TOU).



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Figure 7. (a) Oxygen Penetration Depth (OPD) plotted vs. Diffusive Oxygen Uptake (DOU) rate. Filled circles: measured values; empty circles: theoretical values **(b)** measured OPD vs. theoretical OPD. Dashed line: 1 : 1 line; red symbols: SNS; blue symbols: NNS; black: SKNT. For calculation and details see text.







Figure 8. (a) Correlation between A_7 fluxes and TOU rates. **(b)** Correlation between DIC fluxes and TOU rates. Dashed line: line of RQ = 0.77; solid line: linear regression; Spearman's p value given in each plot. TA and DIC fluxes in mmol C m⁻² d⁻¹. TOU rates in mmol O₂ m⁻² d⁻¹.













Figure 10. Correlation between A_{τ} and DIC fluxes for sites of the SNS from both campaigns. Different lines are representing different reaction stoichiometry. For more explanation and reaction equations see text. Short dashed line: aerobic respiration; dotted line: denitrification; dotted/dashed line: sulfate reduction coupled to pyrite formation and burial; long dashed line: carbonate dissolution; p: Spearman's rank correlation coefficient. All fluxes in mmol C m⁻² d⁻¹.







Figure 11. Contribution of different processes to the AT budget in the SNS. For more details on different reaction rates see text. NI: nitrification; SO: sulfide oxidation; PF: pyrite formation; CD: carbonate dissolution; DN: denitrification; SR: sulfate reduction; IR: dissimilatory iron reduction; AR: aerobic respiration. Fluxes in mmol $Cm^{-2}d^{-1}$.







Figure 12. DIC and A_T budgets of the water column in the SNS. Bold arrows: fluxes; dashed arrows: reaction rates. F_{sed} : net flux from the sediments; F_{river} : riverine input; F_{out} : flux into the NNS/SKNT; F_{air} : atmospheric DIC uptake; R_{pp} : primary production rate; R_{resp} : aerobic respiration rate; R_{cf} : carbonate production rate. Unit: mmol Cm⁻² d⁻¹.

