1	Wet-season spatial variability of N ₂ O emissions from a tea field in subtropical central China
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17 Abstract

Tea fields emit large amounts of nitrous oxide (N_2O) to the atmosphere. Obtaining accurate 18 19 estimations of N₂O emissions from tea-planted soils is challenging due to strong spatial variability. We examined the spatial variability of N₂O emissions from a red-soil tea field in 20 Hunan province, China, on 22 April 2012 (in a wet season) using 147 static mini chambers 21 approximately regular gridded in a 4.0 ha tea field. The N₂O fluxes for a 30-min snapshot 22 (10–10.30 a.m.) ranged from -1.73 to 1659.11 g N ha⁻¹ d⁻¹ and were positively skewed with 23 an average flux of 102.24 g N ha⁻¹ d⁻¹. The N₂O flux data were transformed to a normal 24 distribution by using a logit function. The geostatistical analyses of our data indicated that the 25 logit-transformed N₂O fluxes (FLUX30t) exhibited strong spatial autocorrelation, which was 26 characterized by an exponential semivariogram model with an effective range of 25.2 m. As 27 observed in the wet season, the logit-transformed soil ammonium-N (NH4Nt), soil nitrate-N 28 29 (NO3Nt), soil organic carbon (SOCt), total soil nitrogen (TSNt) were all found to be significantly correlated with FLUX30t (r = 0.57-0.71, p < 0.001). Three spatial interpolation 30 methods (ordinary kriging, regression kriging and cokriging) were applied to estimate the 31 32 spatial distribution of N₂O emissions over the study area. Cokriging with NH4Nt and NO3Nt as covariables (r = 0.74 and RMSE = 1.18) outperformed ordinary kriging (r = 0.18 and 33 RMSE = 1.74), regression kriging with the sample position as a predictor (r = 0.49 and 34 RMSE = 1.55) and cokriging with SOCt as a covariable (r = 0.58 and RMSE = 1.44). The 35 predictions of the three kriging interpolation methods for the total N₂O emissions of 4.0 ha 36 tea field ranged from 148.2 to 208.1 g N d⁻¹, based on the 30 min snapshots obtained during 37

38	the wet season. Our findings suggested that to accurately estimate the total N_2O emissions
39	over a region, the environmental variables (e.g., soil properties) and the current land use
40	pattern (e.g., tea row transects in the present study) must be included in spatial interpolation.
41	Additionally, compared with other kriging approaches, the cokriging prediction approach
42	showed great advantages in being easily deployed, and more importantly providing accurate
43	regional estimation of N ₂ O emissions from tea-planted soils.

45 Introduction

According to the latest data, which show rapid increases in their atmospheric concentrations 46 (IPCC, 2013), nitrous oxide (N₂O), carbon dioxide (CO₂) and methane (CH₄) are three major 47 greenhouse gases in the atmosphere that significantly contribute to global warming. Among 48 these major greenhouse gases, N₂O has a very high radiative forcing per unit mass (265-fold 49 stronger than CO₂ on a 100 year horizon) and plays an important role in ozone depletion in 50 the stratosphere (Ravishankara et al., 2009). The primary sources of N₂O are from agriculture 51 development and the subsequent increased use of chemical N fertilizers (Ambus and 52 53 Christensen, 1994; Mosier et al., 1996, 1998; Yanai et al., 2003; Tokuda and Hayatsu, 2004; Akiyama et al., 2006; Ravishankara et al., 2009). Agricultural soils produce 2.8 (1.7-4.8) Tg 54 of N₂O-N yr⁻¹ (IPCC, 2013). The N₂O is emitted from soils via the microbial processes of 55 nitrification under aerobic conditions and denitrification under anaerobic conditions 56 (Firestone and Davidson, 1989; Wrage et al., 2004). The magnitude of soil N₂O emissions is 57 highly variable and strongly influenced by changes in environmental conditions. 58

59	Among the different agricultural soils, tea-planted soils are important sources of N ₂ O that
60	are rapidly attracting attention due to recent large increases in the number of tea plantations
61	and large N fertilizer inputs (Akiyama et al., 2006; Lin and Han, 2009; Fu et al., 2010, 2012;
62	Hirono and Nonaka, 2012; Han et al., 2013; Li et al., 2013). In China, the total tea-planted
63	area was approximately 2.10 million ha (mostly distributed in Fujian, Anhui, Zhejiang and
64	Hunan) in 2013 (NBSC, 2014). Compared with other agricultural soils, tea-planted soils
65	provide optimal conditions (e.g., low soil pH, high temperature and ample moisture) for
66	microbes to emit significant amounts of N ₂ O (Hayatsu, 1993; Venterea and Rolston, 2000; Li
67	et al., 2013). However, because few measurements of N_2O emissions from tea-planted soils
68	have been reported in China (Fu et al., 2012; Li et al., 2013; Han et al., 2013), it is difficult to
69	conduct precise spatial and temporal evaluations of N_2O emissions from tea-planted soils. To
70	estimate the N_2O emissions from tea-planted soils accurately and to understand the roles that
71	tea plantations play in global warming, it is necessary to investigate the spatial and temporal
72	patterns and related mechanisms of N ₂ O emissions from tea fields. This information will lead
73	to the development of effective land management options for mitigating N_2O emissions from
74	a significant source, tea plantation.
75	The N ₂ O fluxes have large spatial variability in agricultural soils (Konda et al., 2008,

2010; Meda et al., 2012; Li et al., 2013). Many previous studies in tea fields have found
pronounced seasonal fluctuations in N₂O fluxes, with higher N₂O emissions during the wet
season than during the dry season (Fu et al., 2012; Han et al., 2013). The seasonal and spatial
variability of N₂O emissions significantly contributes to the uncertainty when estimating the

80	contributions of subtropical tea-planted ecosystems to N_2O flux. Moreover, most of our
81	knowledge regarding seasonal changes and the spatial variability of N_2O fluxes is based on a
82	small number of measurements taken from tea-planted soils. Li et al. (2013) investigated the
83	spatial structure of N_2O fluxes for tea-planted soils during the dry season in October 2010
84	and found that the spatial distribution of the N_2O fluxes was primarily associated with field
85	elevation ($r = -0.42$, $p < 0.001$). The other soil properties (e.g., soil organic carbon, soil water
86	and soil mineral nitrogen) were not significantly related to N_2O flux. To obtain a more
87	accurate evaluation of the interannual variability of N2O emissions from tea-planted soils, a
88	study on the spatial structure and distribution of N_2O emissions during a wet season (in
89	contrast to the dry season) is necessary.
90	To understand the structure of the spatially distributed data and to predict the N_2O fluxes
91	at the unsampled locations, geostatistical analyses can be useful (Goovaerts, 1997; Webster
92	and Oliver, 2001). Geostatistics provide statistical tools for describing the quantitative spatial
93	variability of field observations for the accurate mapping and planning of rational sampling
94	schemes that efficiently utilize the available labor (Webster, 1985). Several geostatistical
95	methods are used to examine the spatial variability of N_2O fluxes, including simple kriging
96	(SK), ordinary kriging (OK), regression kriging (RK) and cokriging (CK). The most
97	commonly used method is OK (Clemens et al., 1999; Röver et al., 1999; Mathieu et al., 2006;
98	Konda et al., 2008, 2010), which uses the derived theoretical semivariogram models to
99	interpolate the spatial distribution of N ₂ O fluxes. However, research has demonstrated that
100	RK and CK approaches, which use related auxiliary variables, improve the prediction

101	accuracy (Goovaerts, 1997; Webster and Oliver, 2001; Hengl et al., 2004). The RK method
102	combines multiple regressions, including linear regressions, generalized linear models,
103	generalized added models and regression tree models, with the auxiliary variables used for
104	kriging (Odeh et al., 1994). In the RK method, linear regressions are commonly used. The
105	CK approach uses correlations that may exist between the predicted variables and other more
106	easily measured variables. These variables can be measured at the same points as the
107	predicted variable, at other points, or at both. Compared with the RK approach, the CK
108	approach is commonly applied when the measurement of a covariable is less expensive than
109	the cost of a predicted variable (Stein et al., 1988; Odeh et al., 1995). In addition to the
110	feature correlation as a criterion for selecting covariables, the CK approach also requires that
111	both of the predicted variable and covariables have similar spatial structures (Odeh et al.,
112	1994). In this study, we used three interpolation methods (OK, RK and CK) to estimate the
113	spatial distribution of N_2O fluxes in a tea field.
114	In contrast with the dry season, the spatial variability of the N_2O emissions was
115	investigated during the wet season in April 2012 from the same tea-planted catchment that
116	was studied by Li et al. (2013). The catchment consisted of a completely independent
117	hydrological system. Thus, the spatial distribution of the N_2O emissions within the catchment
118	was expected to have intrinsic characteristics. The objectives of this study were to (i) evaluate
119	the spatial variability of N_2O emissions from soils planted with tea in subtropical central
120	China during the wet season, (ii) determine the key environmental factors controlling N_2O
121	emissions, and (iii) assess the prediction efficiency of three kriging interpolation methods.

123 **2. Materials and Methods**

124 **2.1 Site description**

The field experiment was conducted in a small catchment (4.0 ha) in Jinjing, Changsha, in 125 Hunan province, China (28°32′50″N and 113°19′58″ E and elevation 90 to 111 m) (Fig. 1). 126 The region has a subtropical monsoon climate with a mean annual air temperature of 17.5°C 127 128 and a mean annual precipitation of 1400 mm (average from 1979 to 2012). The site had four distinct seasons: spring (February to April), summer (May to July), autumn (August to 129 November), and winter (December to January). On average, 70% of the annual precipitation 130 occurred in April, May and June. The daily air temperature and precipitation for 2012 were 131 recorded by an automatic weather station (Intelimet A, IMET-ADV2, Dynamax, USA) 132 located next to the studied catchment (Fig. 2). The soil of the catchment was a Haplic Alisol 133 (FAO/UNESCO soil taxonomy) that was derived from a granitic parental material. Tea 134 (Camellia sinensis L., cv. Baihaozao) was contour-planted 10 years ago using an inter-row 135 spacing of 0.5 m in the catchment. 136

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<Insert Fig. 1 & Fig. 2 near here>

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140 **2.2 Sampling positions**

In the 4.0 ha tea-planted catchment, 1964 evenly-distributed points with plane coordinates
and elevation values and 456 centerlines of tea tree row were recorded by locally calibrated
differential Geographic Positioning System (DGPS) receiver (Sanding Southern Survey Co.,

144	China), and then were used to develop the local DEM and land use data (at a spatial
145	resolution of 0.1 m, respectively, as shown in Fig. 1c and d). The land use data showed the
146	four positions where the chambers were placed, including the inter-row, fertilization point,
147	under tea tree and in tea tree row, as described in Li. et al. (2013). The spatial positions of the
148	gas sampling points in a 15 m \times 15 m regular grid over the catchment were originally
149	determined using a DGPS receiver on 20 April 2012. Some of the chamber positions were
150	slightly adjusted (because of a lack of space in the tea tree rows or to avoid roads and
151	trenches). Thus, the chambers were placed in one of four locations mentioned above (Fig. 1d)
152	Overall, 147 sampling points were determined, and the Euclidean distances between each
153	point and its nearest neighbors ranged from 14.6 m to 16.7 m. The x-y coordinates, the gas
154	sampling position information (the inter-row, fertilization point, under tea tree and in tea tree
155	row along tea row transects), and the elevations at the sampling points were recorded.

157 **2.3 Gas and soil properties measurements**

Gas and soil samples were collected at each grid point on 22 April 2012 using a closed mini chamber technique. A mini chamber set was composed of PVC and had two parts (base and chamber). The base was 0.15 m in diameter and 0.05 m high. The chamber was 0.15 m in diameter and 0.15 m high, and was equipped with rubber septa on the top for gas sampling. In the field operation, the base was gently inserted vertically into the soil on 20 April 2012, and the chamber was clipped on the base with the sponge seals in between to stop gas leaking before gas sampling on 22 April 2012. Therefore, the effective static chambers volume was

165	equal to the chamber volume of 0.002651 m^3 . Gas samples were collected from the
166	headspace between 10 and 10.30 a.m. For simultaneous sampling, 25 skilled gas sampling
167	persons helped to accomplish the field sampling. Each person only took care of one column
168	containing 4 to 8 sampling positions (see Fig. 1), and started sampling at the same time of 10
169	a.m. At each point, three gas sample replicates were collected from the headspace into
170	pre-evacuated 12 mL vials (Exetainers, Labco, UK) at 0 and 30 min after the chamber body
171	was clipped. After collecting the gas samples, the air temperature in each chamber was
172	measured for subsequent correction of the flux calculation, and then three replicate soil cores,
173	0.05 m in diameter and 0.20 m in depth, were collected from the soils inside the mini
174	chambers. Soil samples were put straight into clean zip-lock bags for avoiding soil moisture
175	loss, and quickly transported back to the laboratory in thermal insulation boxes and stored in
176	a refrigeration room at 4 °C for preventing any microbial activity (such as mineralization,
177	nitrification and denitrification). The N_2O concentrations of the gas samples were analyzed
178	using a gas chromatograph (Agilent 7890A, Agilent, USA) that was fit with a ⁶³ Ni-electron
179	capture detector and an automatic sample injector system. The N_2O fluxes (FLUX30, g N ha ⁻¹
180	d ⁻¹) were calculated as described by Li et al. (2013). The soil physical/chemical properties
181	determined by using fresh soil, e.g., the soil ammonium content (NH4N), soil nitrate content
182	(NO3N), soil dissolved organic carbon content (DOC), soil volumetric water content (SWC),
183	and soil bulk density (BD), were measured within three days after sampling, while those
184	using air-dried soil, e.g., total soil nitrogen content (TSN), soil organic carbon content (SOC)
185	and soil clay/silt/sand content (CLAY, SILT and SAND), were determined within two weeks

186 after the field work.

188	2.4	Data	anal	lyses
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- 189 The descriptive statistical and geostatistical analyses were performed using R (\mathbf{R}
- 190 Development Core Team, 2014) with the gstat package (DGUU, 2010).
- 191 Descriptive statistical analyses were used to determine the mean, median, minimum and
- 192 maximum values, SD, coefficient of variation (CV), and skewness of the original and
- 193 logit-transformed data. These analyses were based on the four chamber placement positions.
- Because the FLUX30, NH4N, NO3N, SOC, TSN and SWC data were highly skewed, these
- values were transformed by using a logit function (Hengl et al., 2004). The transformed
- 196 variables were named FLUX30t, NH4Nt, NO3Nt, SOCt, TSNt and SWCt. Using a Pearson's
- 197 correlation, the relationships between FLUX30t, NH4Nt, NO3Nt, SOCt, TSNt, SWCt, DOC,
- BD, SAND, SILT, and CLAY were tested. The significance of the differences in the FLUX30t
- and environmental factors (NH4Nt, NO3Nt, SOCt, TSNt and DOC) between any two of the
- 200 different chamber positions along the entire tea-tree row transect were evaluated using the
- 201 Tukey's Honest Significant Difference method.
- In the geostatistical analyses, an experimental semivariogram of FLUX30t was
- calculated, and the theoretical semivariogram models were fit. The ratio of the partial sill to
- the total sill was used as an index of spatial dependence. Armstrong (1998) stated that a
- variable with a higher ratio of partial sill to sill and a longer semivariogram range were more
- structured. The spatial distribution of FLUX30t across the catchment was predicted using

207	three kriging interpolation methods (OK, RK and CK). These data were transformed back to
208	the original scale of FLUX30 for mapping. The Leave-One-Out cross-validation method was
209	used to evaluate the accuracy of interpolating FLUX30t using the three different kriging
210	methods.
211	
212	3. Results
213	3.1 Exploratory data analyses
214	In the 4.0 ha tea-planted catchment, the N_2O fluxes during the 30-min one-time
215	measurements performed on 22 April 2012 ranged from -1.73 to 1,659.11 g N ha ⁻¹ d ⁻¹ , with a
216	median value of 27.56 g N ha ⁻¹ d ⁻¹ and a CV of 234.7 % (Table 1). The N ₂ O flux data were
217	positively skewed (Table 1 and Fig. 3a), and their logit-transformations were approximately
218	normally distributed (Table 1 and Fig. 3b). From Table 3, the logit-transformed N_2O fluxes
219	(FLUX30t) were the highest in the fertilization points, and the differences in the FLUX30t
220	values among the chamber placement positions were statistically significant ($p < 0.001$).
221	
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224	The ELEVATION, BD, DOC, SWC, SAND, SILT, and CLAY were approximately
225	normally distributed, with skewness values of less than 1 (Table 1). Additionally, DOC
226	displayed a moderate CV of 34.6 %, and the other variables had lower CVs (4.1–23.8 %). The
227	NH4N, NO3N, SOC and TSN were positively skewed, and the logit-transformations (NH4Nt,

228	NO3Nt, SOCt and TSNt) had approximately normal distributions (Table1). The NH4N and
229	NO3N had very high CVs (190.8 % and 141.6 %, respectively), and the SOC and TSN had
230	moderate CVs (50.1 % and 38.3 %, respectively).
231	The NH4Nt, NO3Nt, SOCt, TSNt and SWC were significantly correlated with the N_2O
232	fluxes (Fig. 5), and the NH4Nt, NO3Nt and TSNt had strong positive relationships with N_2O
233	($r = 0.71$, 0.70 and 0.57, respectively, $p < 0.001$). The N ₂ O emissions and some soil
234	properties (NH4N, NO3N, SOC, TSN and SWC) in the fertilization points were significantly
235	different ($p < 0.001$) from the other three chamber placement positions (Fig. 6). These
236	variables were used as auxiliary covariables for the CK approach.
237	
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239	
240	3.2 Spatial variability of N_2O emissions and related environmental factors
241	Because most of the soil properties were significantly correlated with the chamber placement
242	positions, two types of semivariogram models were calculated for the N_2O and soil
243	parameters (correlated with N_2O fluxes) in the wet season (Table 2). The FLUX30t exhibited
244	strong spatial autocorrelation and was characterized by an exponential semivariogram model,
245	a theoretical distance parameter of 8.40 m (equivalent to an effective range of 25.2 m) and a
246	zero nugget. The NH4Nt, SWCt, SAND and SILT showed almost no spatial dependency,
247	while NO3Nt and TSNt demonstrated weak spatial dependency with a range parameter of

248 91.9 and 58.0 m, respectively (equivalent to an effective range of 163.7 and 102.6 m,

249	respectively). The SOCt exhibited a moderate spatial dependency within 93.0 m. By
250	detrending the influence of the chamber placement position, large changes in the
251	semivariogram models occurred regarding the above variables. Although the semivariograms
252	of the regression residuals of FLUX30t, NH4Nt, NO3Nt and SOCt were best-fit with the
253	same semivariogram model (exponential) with a similar range of 17.4 m (equivalent to an
254	effective range of 52.1 m), the spatial dependencies of those variables were different (Table
255	2). Of the soil properties, only SOCt had a similar spatial structure to FLUX30t when the
256	influence of the chamber placement position was detrended (Table 2). Based on these
257	correlation analyses and spatial variability analyses, the covariables for the CK method were
258	determined.
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260	<insert 2="" here="" near="" table=""></insert>
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262 **3.3 Spatial interpolation of N₂O emissions by three methods**

Three spatial interpolation methods were used in this study to predict the spatial distribution of N_2O emissions from tea soils in the catchment. In the first method, the derived theoretical semivariogram model for FLUX30t that is presented in Table 2 was used for the OK prediction. In the second method, RK was used and the chamber placement position was identified as the auxiliary regression predictor. Thus, the semivariogram of the regression residuals of FLUX30t were calculated and best-fit with the theoretical semivariogram model shown in Fig. 6. In the third method, CK involved two groups of covariables. As described

270	previously, because SOCt (detrending the influence of chamber placement position) showed a
271	similar spatial structure to FLUX30t (detrending the influence of chamber placing position), a
272	CK process was performed using SOCt as the covariable. Firstly, the direct and
273	cross-semivariograms of FLUX30t and SOCt (detrending the influence of the chamber
274	placement position) were calculated and best-fit with a linear model for co-regionalization
275	(LMC). Next, the fitted LMC was used to predict the spatial surface of N_2O emissions.
276	Because NH4Nt and NO3Nt were significantly correlated with FLUX30t (Fig. 5), a second
277	CK with NH4Nt and NO3Nt as the covariables was processed, similarly to that of the CK
278	with SOCt. However, these covariables had different spatial structures (Table 2). As reflected
279	by the lower root mean squared error (RMSE) and higher r values (Table 4), the CK method
280	performed better than the other spatial interpolation methods. Furthermore, the CK with
281	NH4Nt and NO3Nt as two covariables outperformed the CK with SOCt as the covariable.
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283	<insert figs.6-9="" here="" near=""></insert>
284	
285	As shown in Fig. 9, the surface map for the spatial distribution of N_2O emissions
286	interpolated by OK was rougher than the maps obtained from the other interpolation
287	approaches. The kriging standard deviation maps were showed in Fig. 10, and clearly
288	indicated that the RK and CK methods with lower kriging standard deviations outperformed
289	the OK method with higher kriging standard deviations. The four kriging interpolations of
290	OK, RK, CK with SOCt as the covariable and CK with NH4Nt and NO3Nt as the covariables

291	were able to predict that the total amount of N_2O emissions in the tea fields during the wet
292	season were 208.1 g N d ⁻¹ , 148.2 g N d ⁻¹ , 149.7 g N d ⁻¹ and 150.5 g N d ⁻¹ , respectively. From
293	the performance evaluations of the four spatial interpolations, the total N_2O emissions from
294	the tea field on 22 April 2012 during the wet season were approximately 150 g N d ⁻¹ .
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296	<insert 10="" fig.="" here="" near=""></insert>
297	
298	Discussion
299	4.1 Seasonal differences of N_2O fluxes in the red soil planted with tea
300	The N_2O emissions from soils have obvious seasonal fluctuations, with emissions that are
301	significantly higher during the wet season than during the dry season (Konda et al., 2010). To
302	understand the seasonal changes in the spatial structures of N_2O fluxes, we compared the
303	N_2O emissions between the wet (this study) and dry (Li et al., 2013) seasons. In general, the
304	mean, SD and CV (102.24, 239.96 g N ha ⁻¹ d ⁻¹ and 234.7 %, respectively) of the N ₂ O fluxes
305	in the wet season were all higher than those (2.88, 8.94 g N ha ⁻¹ d ⁻¹ and 152.0 %, respectively)
306	during the dry season (Table 3). Furthermore, in contrast with the dry season, the N_2O fluxes
307	during the wet season were significantly different among the four chamber placement
308	positions, with the highest fluxes occurring at the fertilization points and the inter-row
309	positions (Table 3). During the wet season, the high N_2O fluxes at the fertilization points and
310	the inter-row positions resulted from the high soil moisture, due to more rainfall, and from
311	the fertilization that occurred on 19 February 2012 (Fig. 2). The soil N and the soil organic C
312	availability are directly increased by the application of chemical and organic N fertilizers.

313	The additional in the available C and N supplied by fertilization resulted in increased soil
314	microbial activity, which stimulated the nitrification and denitrification processes that
315	contribute to soil N ₂ O emissions (Davidson et al., 1993; Kiese et al., 2003; Werner et al.,
316	2007).
317	
318	<insert 3="" here="" near="" table=""></insert>
319	
320	4.2 Spatial structure of N_2O emissions from red soils planted with tea
321	Soil type, topography and land management (fertilization, tillage and irrigation) are the
322	primary factors that affect the spatial structures of N2O emissions (Folorunso and Rolston,
323	1984; Clemens et al., 1990; Velthof et al., 1996; Konda et al., 2008). During the wet season,
324	the N_2O fluxes showed a strong spatial dependence (with a range of approximately 25.3 m)
325	that was similar to the dry season range of approximately 28.0 m in the tea-planted fields (Li
326	et al., 2013). These results indicated that the spatial dependence of N_2O fluxes at the current
327	spatial sampling scale was comparable between seasons. Our findings for a fertilized tea field
328	were similar to those of Konda et al. (2010) for a tropical forest. However, these results
329	contrasted those of many previous investigations for agricultural fields, including winter
330	wheat (Ball et al., 1997; Clemens et al., 1999; Röver et al., 1999; Mathieu et al., 2006),
331	summer maize (Clements et al., 1999), onion (Yanai et al., 2003), and grassland (Ambus and
332	Christensen, 1994; Velthof et al., 1996; van den Pol-van Dasselaar et al., 1998; Turner et al.,
333	2008) fields, in which the N_2O flux presented no, weak or moderate spatial dependence. This

334	discrepancy primarily occurred because of the unique geographical characterization and land
335	management of the tea plantation. Compared with other agricultural fields in flat areas, tea
336	fields are always distributed in hills or mountains. Therefore, the contributions of the
337	topography to the spatial dependence of the N_2O flux were strong (Li et al., 2013).
338	Additionally, tea is a perennial plant. Thus, apart from fertilization and weeding, the soil
339	disturbance in tea fields is always very low.
340	During the dry season, the topography (elevation) had a significant effect on the spatial
341	pattern of N_2O fluxes in the tea-planted fields (Li et al., 2013). Similar spatial patterns of N_2O
342	fluxes with topography were also observed in forest soils (Van Kessel et al., 1993; Konda et
343	al., 2010). Theoretically, the SWC varies with the topography and affects the spatial pattern
344	of N ₂ O fluxes by controlling the conditions for soil nitrification and denitrification (Firestone
345	and Davidson, 1989; Wrage et al., 2004). Although the SWC had no relationships with N_2O
346	and elevation during the dry season (Li et al., 2013), a correlation existed in the present study
347	(Fig. 5). The microstructures of the tea tree-row transect and the land management practices
348	of tea production were the primary influences on the spatial pattern of soil water in the
349	tea-planted fields (Li et al., 2013). During the wet season, fertilization contributed to the
350	spatial pattern of N_2O fluxes in the tea-planted fields, with the highest averaged fluxes at the
351	fertilization sites (198.81 g N ha ⁻¹ d ⁻¹) (Table 3). Fertilization resulted in similar spatial
352	patterns of N ₂ O fluxes in other agricultural soils (Ball et al., 1997; Clements et al., 1999;
353	Röver et al., 1999; Mathieu et al., 2006; Yanai et al., 2003).

In view of the analysis of the primary factors that affected the spatial pattern of N₂O 354 fluxes, we detrended the influences of the environmental factors when the N₂O flux 355 356 semivariograms were calculated to more deeply explore the spatial structures of the N₂O emissions in the tea-planted fields. For example, during the dry and wet seasons, the spatial 357 influences of elevation (Li et al., 2013) and chamber placement position, respectively, were 358 detrended when computing the N₂O flux semivariograms. Because the relationship between 359 chamber placement position and N₂O flux was more relevant than the relationship between 360 elevation and N₂O flux, the effect of detrending the influence of chamber placement position 361 during the wet season was more obvious than that of detrending the influence of elevation 362 during the dry season (Li et al., 2013). This effect was also reflected in the evaluation of the 363 performance of the RK method for the wet and dry seasons (Table 4). 364

365

4.3 Spatial interpolations of N₂O emissions by three methods

The three interpolation methods (OK, RK and CK) were used to predict the spatial 367 distributions of N₂O emissions from the red soils planted with tea during dry (Li et al., 2013) 368 and wet seasons (this study). However, these three methods resulted in significantly different 369 performances between the dry and wet seasons (Table 4). We conducted comparative 370 analyses for the performance of the three interpolation methods using two aspects: different 371 seasons and different methods. Firstly, the OK method performed better when predicting the 372 spatial distribution of N₂O fluxes for the dry season relative to the wet season. Because the 373 OK method directly used the fitted theoretical semivariogram model of the target variable to 374

375	predict the spatial distribution, its performance reflected the predictive ability of the original
376	data (Goovaerts, 1997). During the wet season, more factors (e.g., NH4N, NO3N, SOC, TSN
377	and SWC) influenced the spatial distributions of the N_2O fluxes than the dry season (Table 2
378	and Fig. 5). The values of the original data were concealed. Thus, other sophisticated kriging
379	methods, such as RK and CK, which reconcile the relationships between N_2O fluxes and
380	environmental factors, could be useful. The RK method performed better when elevation was
381	used as an auxiliary regression predictor during the dry season than when the chamber
382	placement position was used during the wet season (Table 4). This finding primarily occurred
383	because the chamber placement position was a categorical variable with a lower regression
384	fitting ability than elevation, which was a continuous variable (Goovaerts, 1997). The
385	performances of the CK with two groups of covariables during the wet season were better
386	than those of the CK with three groups of covariables during the dry season (Table 4).
387	Particularly, the CK with strongly correlated covariables of NO3N and NH4N ($r = 0.70-0.71$
388	and $p < 0.001$) (Fig. 5) performed the best ($r = 0.74$ and RMSE = 1.04) (Table 4).
389	Secondly, by comparing the performances of the three interpolation methods, the RK and
390	CK methods, which are more sophisticated kriging technologies, performed better than the
391	OK method for the dry and wet seasons. Similar results were obtained by previous
392	researchers (Stein et al., 1988; Odeh et al., 1995; Goovaerts, 1997; Hengl et al., 2004).When
393	comparing the performances of RK and CK, no differences were observed for the dry season.
394	However, during the wet season, the CK significantly outperformed the RK (Table 4). Overall,
395	few attempts have been made to provide a good method for selecting interpolation methods

396	between RK and CK (Kontters et al., 1995; Odeh et al. 1995). Li et al. (2013) suggested that
397	RK was a good choice because of the performance of the two interpolation methods and the
398	difficulties encountered when applying CK. However, in this study, the CK method was
399	better than the RK method because of its high predictive performance (Table 4), its readily
400	available required covariables (e.g., NH4N, NO3N and SOC) at co-locations, and because
401	expensive surface data were not needed (e.g., DEM and land use data, which are required by
402	RK) (Goovaerts, 1997; Webster and Oliver, 2001). Our conclusions were similar to those of
403	many previous studies that found that CK was the most versatile and rigorous statistical
404	technique for estimating spatial points (Stein et al., 1988; Odeh et al., 1995; Webster and
405	Oliver, 2001). For the application of CK, the covariables must show a correlation with the
406	target variable and present a similar spatial structure as the target variable (Odeh et al., 1995;
407	Goovaerts, 1997; Webster and Oliver, 2001). Therefore, we further compared the effects of
408	the two groups of covariables for CK in this study. We found that CK method with NH4Nt
409	and NO3Nt (showed significant correlations with FLUX30t) as covariables outperformed the
410	CK method with SOCt (presented a similar spatial structure to FLUX30t) as a covariable,
411	indicating that the feature correlation was more important than the similarity of the spatial
412	structure when selecting CK covariables. This finding can be regarded as a prerequisite for
413	selecting covariables for CK application.
414	

<Insert Table 4 near here>

417	The three spatial interpolation methods predicted similar total N_2O emissions from the
418	tea-planted red soils in the 4.0 ha catchment on 30 October 2010 (in the dry season) and on
419	22 April 2012 (in the wet season), ranging from 21.2 to 22.1 g N d^{-1} and from 148.2 to 208.1
420	g N d^{-1} (Table 4), respectively. The predicted errors during the wet season were higher than
421	those of the dry season (Table 4). This result mainly occurred because fertilization was a
422	major factor that affected the N_2O emissions from the tea fields during the wet season.
423	Following fertilization, the horizontal and vertical movement of NH4N and NO3N in the
424	topsoil of the tea fields potentially produced the strong spatial heterogeneity of N_2O
425	emissions. In addition, it is possible that the variations in the availability of oxygen in the
426	soils was regulated by soil moisture, which determined the spatio-temporal heterogeneity of
427	N ₂ O emissions by inducing different degrees of soil nitrification and denitrification
428	(Davidson et al., 2000; Konda et al., 2010). Thus, spatial interpolation methods must be
429	chosen carefully to accurately estimate the spatial distribution of N_2O emissions when the
430	emissions are high and have strong spatial variability in the fields.

432 **5** Conclusions

433 During the wet season of 2012, a 30-min one-time measurement of N_2O emissions from a 4.0

ha red-soil tea field in the subtropical region of central China were determined at 147 points.

435 The N_2O fluxes significantly varied with space. In addition, the N_2O fluxes were significantly

436 correlated with the NH4N, NO3N, SOC and TSN contents (r > 0.27 and p < 0.001). The

437 logit-transformed N₂O fluxes demonstrated a strong spatial dependency and were

438	characterized by an exponential semivariogram model with an effective range of 25.2 m.
439	Three spatial interpolation methods (OK, RK and CK) were used to predict the spatial
440	distribution of N_2O emissions. The RK and CK methods were relatively accurate for
441	predicting results. Although the N_2O emissions were much higher during the wet season than
442	in the dry season, the N_2O emissions exhibited similar spatial structure during both seasons.
443	Such a phenomenon was mainly attributed to the low soil disturbance (e.g., only fertilizing in
444	a very small proportion of area and weeding) in the tea field.
445	To effectively mitigate high N_2O emissions from the tea field soils, the biological and
446	chemical mechanisms of N_2O emissions must be deeply explored. In addition, the responsive
447	land management practices, such as biochar application, deep fertilization (under 20 cm), the
448	use of controlled-release fertilizers and ecological engineering, must be recommended and
449	deployed, especially during the wet season.
450	

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454 **References**

- 455 Akiyama, H., Yan, X. Y., and Yagi, K.: Estimations of emission factors for fertilizer-induced
- direct N₂O emissions from agricultural soils in Japan: Summary of available data, Soil Sci.
- 457 Plant Nutr., 52, 774-787, 2006.
- Ambus, P., and Christensen, S.: Measurement of N₂O emission from a fertilized grassland: an
 analysis of spatial variability, J. Geophys. Res., 99, 16557-16567, 1994.
- 460 Armstrong, M.: Basic linear Geostatistics, Springer Verlag, Berlin, 153 pp., 1998.
- Ball, B. C., Horgan, G. W., Clayton, H., and Parker, J. P.: Spatial variability of nitrous oxide
- fluxes and controlling soil and topographic properties, J. Environ. Qual., 26, 1399-1409,
 1997.
- Clemens, J. Schillinger, M. P., Goldbach, H., and Huwe, B.: Spatial variability of N₂O
 emissions and soil parameters of an arable silt loam a field study, Bio. Fert. Soils, 28,
 403-406, 1999.
- 467 Davidson, E. A., Matson, P. A., Vitousek, P. M., Riley, R., Dunkin, K., García-Méndez, G.,
- and Maass, J. M.: Processes regulating soil emissions of NO and N₂O in a seasonally dry
 tropical forest, Ecology, 74, 130-139, 1993.
- Davidson, E. A., Keller, M., Erickson, H. E., Verchot, L. V., and Veldkamp, E.: Testing a
 conceptual model of soil emissions of nitrous and nitric oxides, Bioscience, 50, 667-680,
- 472 2000.
- 473 DGUU (Department of Geography, Utrecht University): Introduction for Gstat, available at:
- 474 http://www.gstat.org/index.html (last access: 15 December 2010), 2010.

- Firestone, M., and Davidson, E.: Microbial basis of NO and N₂O production and
 consumption, in: Exchange of Trace Gases Between Ecosystems and the Atmosphere,
 edited by: Andreae, M.O. and Schimel, D.S., John Wiley, Chichester, 7-21, 1989.
- Folorunso, O. A., and Rolston, D. E.: Spatial variability of field measured denitrification gas
 fluxes, Soil Sci. Soc. Am. J., 48, 1214-1219, 1984.
- 480 Fu, X., Li, Y., Xiao, R., Tong, C., and Wu, J.: N₂O emissions from a tea field in subtropical
- 481 China. In: Proceedings of the 19th World Congress of Soil Science, Soil Solutions for a
- 482 Changing World, 1–6 August 2010, Brisbane (published on CDROM), 161-163, 2010.
- 483 Fu, X., Li, Y., Su, W., Shen, J., Xiao, R., Tong, C., and Wu, J.: Annual dynamics of N2O
- 484 emissions from a tea field in southern subtropical China, Plant Soil Environ., 58,
 485 373-378, 2012.
- Goovaerts, P.: Geostatistics for Natural Resources Evaluation, Oxford University Press, New
 York, 483 pp., 1997.
- Gorres, J. H., Dichiaro, M. J., and Lyons, J. A.: Spatial and temporal patterns of soil
 biological activity in a forest and an old field, Soil Biol. Biochem., 30, 219-230, 1998.
- Han, W., Xu, J., Wei, K., Shi, W., and Ma, L.: Estimation of N₂O emission from tea garden
- soils, their adjacent vegetable garden and forest soils in eastern China, Environ. Earth Sci.,
- 492 70, 2495-2500, 2013.
- Hayatsu, M.: The lowest limit of pH for nitrification in tea soil and isolation of an acidophilic
 ammonia oxidizing bacterium, Soil. Sci. Plant Nutr., 39, 219-226, 1993.

495	Hengl, T., Heuvelink, G. B. M., and Stein, A.: A generic framework for spatial prediction of
496	soil variables based on regression-kriging. Geoderma, 120, 75-93, 2004.

- 497 Hirono, Y., and Nonaka, K.: Nitrous oxide emissions from green tea fields in Japan:
- 498 contribution of emissions from soil between rows and soil under the canopy of tea plants,
- 499 Soil. Sci. Plant Nutr., 58, 384-392, 2012.
- 500 IPCC: Climate change 2013: the physical science basis. Contribution of working group I, in:
- 501 Fourth assessment report of the intergovernmental panel on climate change, edited by:
- 502 Solomon S., Qin D., Manning, M., Chen Z., Marquis, M., Averyt, K.B., Tignor, M.,
- 503 Miller, H.L., Cambridge University Press, Cambridge, 996 pp., 2013.
- ISM (Institute for Statistics and Mathematics): The R Project for Statistical Computing,
 available at: http://www.r-project.org/ (last access: 15 December 2010), 2010.
- 506 Kiese, R., Hewett, B., Graham, A., and Butterbach-Bahl, K.: Seasonal variability of N₂O
- 507 emissions and CH₄ uptake by tropical rainforest soils of Queensland, Australia. Global
 508 Biogeochem. Cy., 17, 1043, doi:10. 1029/2002GB002014, 2003.
- Konda, R., Ohta, S., Ishizuka, S., Arai, S., Ansori, S., Tanaka, N., and Hardjono, A.: Spatial
 structures of N₂O, CO₂, and CH₄ fluxes from Acacia mangium plantation soils during a
- relatively dry season in Indonesia, Soil Biol. Biochem., 40, 3021-3030, 2008.
- 512 Konda, R., Ohta, S., Ishizuka, S., Heriyanto, J., and Wicaksono, A.: Seasonal changes in the
- spatial structures of N_2O , CO_2 and CH_4 fluxes from Acacia mangium plantation soils in
- 514 Indonesia, Soil Biol. Biochem., 42, 1512-1522, 2010.

515	Li, Y., Fu, X., Liu, X., Shen, J., Luo, Q., Xiao, R., Li, Y., Tong, C., and Wu, J.: Spatial
516	variability and distribution of N_2O emissions from a tea field during the dry season in
517	subtropical central China, Geoderma, 193, 1-12, 2013.
518	Lin, Y., and Han, W.: N ₂ O emissions from different soils, Chinese Journal of Tea Science, 29,
519	456-464, 2009.

- Mathieu, O., Lévêque, J., Hénault, C., Milloux, M. J., Bizouard, F., and Andreux, F.:
 Emissions and spatial variability of N₂O, N₂ and nitrous oxide mole fraction at the field
 scale, revealed with ¹⁵N isotopic techniques, Soil Biol. Biochem., 38, 941-951, 2006.
- Meda, B., Flechard, C. R., Germain, K., Robin, P., Walter, C., and Hassouna, M.:
 Greenhouse gas emissions from the grassy outdoor run of organic broilers,
 Biogeosciences, 9, 1493-1508, doi:10.5194/bg-9-1493-2012, 2012.
- Mosier, A. R., Duxbury, J. M., Freney, J. R., Heinemeyer, O., and Minami, K.: Nitrous oxide
 emissions from agricultural fields: assessment, measurement and mitigation. Plant Soil,
 181, 95-108, 1996.
- 529 Mosier, A. R., Kroeze, C., Nevison, C., Oenema, O., Seitzinger, S., and van Cleamput, O.:
- $Closing the global N_2O budget:$ nitrous oxide emissions through the agricultural nitrogen
- 531 cycle, Nutr. Cycl. Agroecosys., 52, 225-248, 1998.
- NBSC (a): China Statistical Yearbook, annual publication, National Bureau of Statistics of
 China, Beijing, 2014.

534	Odeh, I. O. A., McBratney, A. B., and Chittleborough, D. J.: Spatial prediction of soil
535	properties from landform attributes derived from a digital elevation model, Geoderma, 63,
536	197-214, 1994.

- Odeh, I. O. A., McBratney, A. B., and Chittleborough, D. J.: Further results on prediction of
 soil properties from terrain attributes: heterotopic cokriging and regression kriging,
 Geoderma, 67, 215-226, 1995.
- 540 R Development Core Team: R: a language and environment for statistical computing. R
- 541 Foundation for Statistical Computing, 2014.
- Ravishankara, A. R., Daniel, J. S., and Portmann, R. W.: Nitrous oxide (N₂O): the dominant
 ozone-depleting substance emitted in the 21st century, Science, 326, 123-125, 2009.
- Röver, M., Heinemeyer, O., Munch, J. C., and Kaiser, E. A.: Spatial heterogeneity within the
- plough layer: high variability of N₂O emission rates, Soil Biol. Biochem., 31, 167-173,
 1999.
- Stein, A., van Dooremolen, W., Bouma, J., and Bregt, A. K.: Cokriging point data on
 moisture deficit. Soil Sci. Soc. Am. J., 52, 1418-1423, 1988.
- Tokuda, S. I., and Hayatsu, M.: Nitrous oxide flux from a large amount of nitrogen fertilizer
 and soil environmental factors controlling the flux, Soil. Sci. Plant Nutr., 50, 365-374,
 2004.
- Turner, D. A., Chen, D., Gellbally, I. E., Li, Y., Edis, R. B., Leuning, R., Kelly, K., and
 Phillips, F.: Spatial variability of nitrous oxide emissions from an Australian irrigated
 dairy pasture, Plant soil, 309, 77-88, 2008.

555	Van den Pol-van Dasselaar, A., Corré, W. J., Klemedtsson, A., Weslien, P., Stein, A.,
556	Klemedtsson, L., and Oenema, O.: Spatial variability of methane, nitrous oxide, and
557	carbon dioxide emissions from drained grasslands, Soil Sci. Soc. Am. J., 62, 810-817,
558	1998.

- Van Kessel, C., Pennock, D.J., and Farrell, R.E.: Seasonal-variations in denitrification and
 nitrous oxide evolution at the landscape scale, Soil Sci. Soc. Am. J., 57, 988-995, 1993.
- Velthof, G. L., Jarvis, S. C., Stein, A., Allen, A. G., and Oenema, O.: Spatial variability of
- nitrous oxide fluxes in mown and grazed grasslands on a poorly drained clay soil, Soil
- 563 Biol. Biochem., 28, 1215-1225, 1996.
- Venterea, R. T., and Rolston, D. E.: Mechanisms and kinetics of nitric and nitrous oxide
 production during nitrification in agricultural soil, Glob. Change Biol., 6, 303-316, 2000.
- 566 Webster, R.: Quantitative spatial analysis of soil in the field, in: Advances in Soil Science,
- edited by: Stewart, B.A., Springer, New York, 1-70, 1985.
- Webster, R., and Oliver, M. A.: Geostatistics for Environmental Scientists, John Wiley &
 Sons, Chichester, 2001.
- 570 Werner, C., Kiese, R., and Butterbach-Bahl, K.: Soil-atmosphere exchange of N₂O, CH₄, and
- CO_2 and controlling environmental factors for tropical rain forest sites in western Kenya,
- 572 J. Geophys Res., 112, D03308, doi:10.1029/2006JD007388, 2007.
- Wrage, N., Velthof, G. L., Laanbroek, H. J., and Oenema, O.: Nitrous oxide production in
 grassland soils: assessing the contribution of nitrifier denitrification, Soil Biol. Biochem.,
 36, 229-236, 2004.

576	Yanai, J., Lee, C. K., Umeda, M., and Kosaki, T.: Spatial variability of soil chemical
577	properties in a paddy field. Soil Sci. Plant Nutr., 46, 473-482, 2000.
578	Yanai, J., Sawamoto, T., Oe, T., Kusa, K., Yamakawa, K., Sakamoto, K., Naganawa, T.,
579	Inubushi, K., Hatano, R., and Kosaki, T.: Atmospheric pollutants and trace gases: spatial
580	variability of nitrous oxide emissions and their soil-related determining factors in an
581	agricultural field, J. Environ. Qual., 32, 1965-1977, 2003.

Variable ^a	Mean	Minimum	Maximum	CV	Skewness of the	Skewness of the	
				(%)	original	logit-transformed data	
					data		
FLUX30	102.24 ^b	-1.73	1,659.11	234.7	4.37	0.6	
ELEVATION	80.64	74.25	87.96	4.1	0.04	-	
BD	1.26	0.90	1.56	10.1	-0.28	-	
DOC	185.56	43.70	424.14	34.6	0.75	-	
NH4N	62.33	1.89	842.55	190.8	3.28	0.17	
NO3N	21.54	0.48	135.29	141.6	1.85	0.28	
SOC	13.33	5.11	52.52	50.1	2.27	-0.44	
TSN	1.52	0.81	4.12	38.3	1.73	-0.01	
SWC	0.33	0.19	0.47	16.6	0.07	-	
SAND	39.73	16.98	63.79	23.8	0.02	-	
SILT	47.15	26.78	64.17	16.1	-0.29	-	
CLAY	13.12	8.68	21.68	21.5	1.00	-	

Table 1 Descriptive statistics of the N_2O fluxes and environmental factors.

^aFLUX30 is the N₂O flux (g N ha⁻¹ d⁻¹); ELEVATION is the elevation (m); and BD, DOC,
NH4N, NO3N, SOC, TSN, SWC, SAND, SILT and CLAY are the soil bulk density (Mg m⁻³),
soil dissolved organic carbon (mg C kg⁻¹ soil), soil ammonium (mg N kg⁻¹ soil), soil nitrate
(mg N kg⁻¹ soil), soil organic carbon (g C kg⁻¹ soil), soil total nitrogen (g N kg⁻¹ soil),

- gravimetric soil water (g H₂O g⁻¹ soil), soil sand particle (%), soil silt particle (%) and soil
- clay particle (%) content, respectively, of the 0-20 cm of topsoil.
- ^bThe median and standard deviation of the FLUX30 were 27.56 and 239.96 g N ha⁻¹ d⁻¹,
- 591 respectively.

Variable	Model	Nugget	Partial	Sill(nugget+ Distance		Effective	Partial
			sill	partial sill)	Parameter (m)	range (m)	sill/sill
FLUX30t ^a	Exp	0	3.7186	3.7186	8.40	25.2	1.00
NH4Nt ^a	ND ^c	ND ^c	ND ^c	ND ^c	ND ^c	ND ^c	ND ^c
NO3Nt ^a	Ste	4.0794	0.6113	4.6907	91.92	163.7	0.13
SOCt ^a	Sph	1.1198	0.7744	1.8942	92.96	93.0	0.41
TSN t ^a	Ste	1.0422	0.2816	1.3238	57.97	102.6	0.21
SWCt ^a	ND ^c	ND ^c	ND ^c	ND ^c	ND ^c	ND ^c	ND ^c
SAND ^a	ND ^c	ND ^c	ND ^c	ND ^c	ND ^c	ND ^c	ND ^c
SILT ^a	ND ^c	ND ^c	ND ^c	ND ^c	ND ^c	ND ^c	ND ^c
FLUX30t ^b	Exp	1.1911	2.0560	3.2471	17.36	52.1	0.63
NH4Nt ^b	Exp	2.0473	0.7185	2.7658	17.36	52.1	0.26
NO3Nt ^b	Exp	1.6241	1.1188	2.7429	17.36	52.1	0.41
SOCt ^b	Exp	0.6043	1.0777	1.6820	17.36	52.1	0.64
TSNt ^b	Ste	0.9347	0.3114	1.2461	59.53	105.4	0.25
SWCt ^b	ND ^c	ND ^c	ND ^c	ND ^c	ND ^c	ND ^c	ND ^c
SAND ^b	ND	ND	ND	ND	ND	ND	ND
SILT ^b	ND	ND	ND	ND	ND	ND	ND

Table 2 Semivariogram models for N_2O fluxes and the environmental factors.

593 ND, not determined.

^aSemivariogram models for the OK method.

- ^bSemivariogram models for the RK method using the chamber placement position as the
- 596 auxiliary regression predictor.
- ^cSpatial structures were not apparent.

Sample position	Mean	SD	Median	Max.	Min.	CV (%)
Dry season						
Inter-row (58)	5.15	4.95	4.09	22.43	-2.83	96.1
Fertilization point (50)	7.19	12.04	4.34	79.56	-6.42	167.4
Under tree (28)	3.58	2.91	2.36	10.28	0.68	81.3
In tree row (11)	5.95	10.38	3.98	52.17	-5.69	174.5
Wet season						
Inter-row (45)	101.69	287.23	27.56	1,659.11	-0.81	282.5
Fertilization point (45)	198.81	295.70	73.42	1,404.32	0.85	148.7
Under tree (22)	16.74	17.00	10.64	61.24	-1.73	101.6
In tree row (33)	28.30	38.34	14.72	177.08	0.19	135.5

599 Table 3 Statistics for N_2O fluxes during the dry and wet seasons.

600 The numbers in the parentheses represent the sample numbers for each chamber placement

601 position.

$\label{eq:constraint} {\mbox{Table 4 Cross-validations of the three different kriging interpolations for N_2O fluxes during}$

the dry and wet seasons.

Method of spatial	Auxiliary	ME	RMSE	r	Predicted total
interpolation	variable	(no dimension)	(no dimension)		N ₂ O emissions
					$(g N d^{-1})$
Dry season					
ОК	-	0.0002	0.102	0.52	22.1 ^a
RK	ELEVATION	0.0008	0.098	0.57	21.1 ^a
СК	SOCt	0.0006	0.103	0.51	22.0 ^a
СК	ELEV	0.0008	0.099	0.57	21.5 ^a
СК	SOCt and	0.0009	0.098	0.57	21.2 ^a
	ELEV				
Wet season					
ОК	-	-0.0005	1.739	0.18	208.1
RK	POSITION	-0.0006	1.549	0.49	148.2
СК	SOCt	0.0020	1.439	0.58	149.5
	(POSITION)				
СК	NH4Nt	0.0001	1.185	0.74	150.5
	(POSITION)				
	and NO3Nt				
	(POSITION)				

605	OK, RK and CK correspond to ordinary kriging, regression kriging and cokriging,
606	respectively; For the dry season campaign, ELEVATION, SOCt and ELEV are the normalized
607	elevation, the normalized soil organic carbon content and the inverse of the normalized
608	elevation, respectively. For the wet season campaign, SOCt, NH4Nt and NO3Nt are the
609	logit-transformations of soil organic carbon, soil ammonium and soil nitrate concentrations,
610	respectively. "POSITION" (in the parentheses) indicates the process of detrending the
611	influence of chamber placement position. The ME, RMSE, and r are the mean prediction
612	error, the root mean squared error (the mean squared deviation ratio of the prediction
613	residuals to the kriging standard errors), and the Pearson's correlation coefficient between the
614	observations and the predictions, respectively.
615	$^{\mathrm{a}}\text{The}$ predicted total $N_{2}O$ emissions during the dry season were recalculated because the study
616	area changed from 4.8 ha to 4.0 ha for the wet season.

618	Figure captions
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2.
) the logit-transformed
JX30t, NH4Nt, NO3Nt,
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ts (<i>r</i>) of the N ₂ O fluxes natrix are normally 'ION is the elevation (m); 'LAY are the soil bulk

soil), soil nitrate (mg N kg⁻¹ soil), soil organic carbon (g C kg⁻¹ soil), total soil nitrogen (g N
kg⁻¹ soil), gravimetric soil water (g H₂O g⁻¹ soil), soil sand particle (%), soil silt particle (%)
and soil clay particle (%) contents of the top 0-20 cm of the soil, respectively. Furthermore, *,
** and *** represent the statistical significance at probability levels of 0.05, 0.01 and 0.001,
respectively. The lowercase letter t represents the logit transformation.
Figure 6. Semivariograms (open circles) and best-fitted models (solid lines) of the normal

646 logit-transformed N₂O fluxes (FLUX30t) (no dimension) for ordinary kriging (\mathbf{a}) and the 647 regression residuals of FLUX30t (no dimension) with chamber placement position as the 648 predictor for regression kriging (\mathbf{b}).

649

Figure 7. Direct and cross-semivariograms (open circles, detrending the influence of chamber
placement position for cokriging) and the best-fitted linear model of the co-regionalization
(solid lines) of the normal logit-transformed N₂O fluxes (FLUX30t) (no dimension) and the
normal SOC (SOCt, no dimension). The linear model of co-regionalization was characterized
by using the same range and different sills for its component models.

655

Figure 8. Direct and cross-semivariograms (open circles, detrending the influence of chamber
placement position for cokriging) and the best-fit linear model of co-regionalization (solid
lines) for the normal logit-transformed N₂O fluxes (FLUX30t) (no dimension), NH4N

659 (NH4Nt, no dimension) and NO3N (NO3Nt, no dimension). The linear model of

660 co-regionalization was characterized by the same range and different sills for its component661 models.

662

663	Figure 9. Spatial distributions of the N_2O fluxes as predicted by (a) OK, (b) RK with
664	chamber placement position as the regression predictor, (c) CK with SOCt (with the influence
665	of chamber placement position detrended) as the covariable, and (d) CK with NH4Nt (with
666	the influence of chamber placement position detrended) and NO3Nt (with the influence of
667	chamber placement position detrended) as two covariables. Here, SOCt, NH4Nt and NO3Nt
668	represent the logit-transformed soil organic carbon, soil ammonium and soil nitrate content,
669	respectively.
670	
671	Figure 10. Spatial distributions of kriging standard deviations of the predicted N_2O fluxes by
672	(a) OK, (b) RK, (c) CK with SOCt as the covariable, and (d) CK with NH4Nt and NO3Nt as

673 two covariables.



675 Figure 1





679 Figure 3



		100 105 110 115		100 200 300 400		-10 -5 0 5		-6 -4 -2 0 2		20 40 60		10 15 20	
	FLUX30t					÷.			- *				ο . φ
75 80 85	0.05	ELEVATION											
	-0.05	-0.18*	BD	×.	-				.				1.0 1.4
100 300	0.11	-0.03	-0.25**	DOC									
	0.71***	0.11	0.04	-0.04	NH4Nt						-1. • • • • • • • • • • • • • • • • • • •		-10 -5 0
-10 0 5	0.70***	0.04	0.03	0.01	0.8***	NO3Nt							
	0.36***	-0.12	-0.05	0.31***	0.34***	0.27***	SOCt	- And - Contraction				-	-5 0 5
	0.57***	0.08	-0.10	0.28***	0.64***	0.56***	0.77***	TSNt	-	-		*** ***	
	0.27***	0.05	0.29***	0.27**	0.35***	0.40***	0.17*	0.35***	SWCt	nini -	3		-202
20 40 60	0.21*	0.34***	-0.05	-0.02	0.15	0.04	0.29***	0.24**	-0.02	SAND	A DESCRIPTION OF THE OWNER		
	-0.21*	-0.32***	0.06	-0.02	-0.15	-0.04	-0.27***	-0.23**	0	-0.97***	SILT		30 40 50 60
10 15 20	-0.14	-0.28***	0	0.12	-0.08	0	-0.25**	-0.17*	0.08	-0.74***	0.55***	CLAY	

683 Figure 5



686 Figure 6







691 Figure 8









0 25 50