Processes regulating progressive nitrogen limitation under elevated carbon dioxide: A 1 meta-analysis 2 3 4 J. Liang^{1,*}, X. Qi¹, L. Souza^{1,2}, Y. Luo^{1,3,*} 5 ¹Department of Microbiology and Plant Biology, University of Oklahoma, Norman, Oklahoma 6 7 73019, USA ²Oklahoma Biological Survey, University of Oklahoma, Norman, Oklahoma 73019, USA 8 9 ³Center for Earth System Science, Tsinghua University, Beijing 100084, China 10 *Corresponding authors: Junyi Liang (iliang@ou.edu) and Yiqi Luo (yluo@ou.edu). 101 11 David L. Boren Blvd., Norman, Oklahoma 73019, USA. Fax: +1 405 325 7619. Tel: +1 405 325 12 6519. 13 14

Abstract: The nitrogen (N) cycle has the potential to regulate climate change through its influence on carbon (C) sequestration. Although extensive research has explored whether or not progressive N limitation (PNL) occurs under CO₂ enrichment, a comprehensive assessment of the processes that regulate PNL is still lacking. Here, we quantitatively synthesized the responses of all major processes and pools in the terrestrial N cycle with meta-analysis of CO₂ experimental data available in the literature. The results showed that CO₂ enrichment significantly increased N sequestration in the plant and litter pools but not in the soil pool. Thus, the mechanisms that drive PNL occurrence partially exist. However, CO₂ enrichment significantly increased the N influx via biological N fixation and the loss via N2O emission, but decreased the N efflux via leaching. In addition, no general diminished CO₂ fertilization effect on plant growth was observed over time up to the longest experiment of 13 years. Overall, our analyses suggest that the extra N supply by the increased biological N fixation and decreased leaching may potentially alleviate PNL under elevated CO₂ conditions in spite of the increases in plant N sequestration and N₂O emission. Moreover, our syntheses indicate that CO₂ enrichment increases soil ammonium (NH₄⁺) to nitrate (NO₃⁻) ratio. The changed NH₄⁺/NO₃⁻ ratio and subsequent biological processes may result in changes in soil microenvironments, abovebelowground community structures and associated interactions, which could potentially affect the terrestrial biogeochemical cycles. In addition, our data synthesis suggests that more longterm studies, especially in regions other than temperate ones, are needed for comprehensive assessments of the PNL hypothesis.

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1 Introduction

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Fossil-fuel burning and deforestation have led to substantial increase in atmospheric carbon dioxide (CO₂) concentrations, which could stimulate plant growth (IPCC, 2013). The plant growth stimulated by CO₂ fertilization and the resulting terrestrial carbon (C) storage could partially mitigate the further increase in CO₂ concentrations and associated climate warming (IPCC, 2013). However, this effect may be constrained by the availability of nitrogen (N), an essential element for molecular compounds of amino acids, proteins, ribonucleic acids (RNAs) and deoxyribonucleic acids (DNAs) in organisms (Rastetter et al., 1997; Oren et al., 2001; Luo et al., 2004; Reich et al., 2006; Norby et al., 2010; Reich and Hobbie, 2013). A popular hypothesis of the N constraint to the CO₂ fertilization effect is progressive N limitation (PNL) (Luo et al., 2004). Progressive N limitation postulates that the stimulation of plant growth by CO₂ enrichment results in more N sequestered in plant, litter and soil organic matter (SOM) so that, the N availability for plant growth progressively declines in soils over time (Luo et al., 2004). The reduced N availability then in turn constrains the further CO₂ fertilization effect on plant growth on long-term scales. However, whether and to what extent PNL occurs depends on the balance of N demand and supply (Luo et al., 2004; Finzi et al., 2006; Walker et al., 2015). If the N supply meets the N demand, PNL may not occur. Otherwise, the CO₂ fertilization effect on plant growth may diminish over time. The PNL hypothesis has been tested in individual ecosystems during the past decade (e.g., Finzi et al., 2006; Moore et al., 2006; Reich et al., 2006; Norby et al., 2010). Some of the site-level studies support (Reich et al., 2006; Norby et al., 2010), while the others refute PNL (Finzi et al., 2006; Moore et al., 2006). To date, no general pattern of PNL across ecosystems has yet been revealed.

Since the key determining PNL occurrence is that whether N supply meets N demand (Luo et al., 2004), it is important to understand how N supply changes under elevated CO₂. The change in the N supply for plant growth under elevated CO₂ is determined by the responses of multiple N cycling processes, including biological N fixation, mineralization, nitrification, denitrification, and leaching (Chapin III et al., 2011). In addition, the responses of these processes to CO₂ enrichment may be influenced by external N addition, such as N deposition and fertilization (Reay et al., 2008). Thus, synthesizing the responses of processes that regulate PNL to CO₂ enrichment may help reveal the general pattern of PNL in terrestrial ecosystems. In the current study, the main objective was to synthesize data published in the literature on the N limitation to plant growth under enriched CO₂ conditions. Our data synthesis was designed to answer two questions: (i) How do the major processes in the terrestrial N cycle respond to CO₂ enrichment? (ii) Does the CO₂ fertilization effect on plant growth diminish over time? To answer these questions, two sets of data from the literature were collected (Table S1, Table 1). With the first dataset, we quantitatively examined the effects of CO₂ enrichment on all the major processes and pools in the N cycle using meta-analysis. These processes and pools included N sequestered in organic components (i.e., plant tissues, litter and soil organic matter (SOM)), biological N fixation, net mineralization, nitrification, denitrification, leaching, and total inorganic N (TIN), ammonium (NH₄⁺) and nitrate (NO₃⁻) contents in soils. We separated the first dataset according to the experimental durations to explore the responses of the N processes to short- vs. long-term CO₂ treatments. In addition, the responses of the N processes to CO₂ enrichment were compared between without and with N addition conditions. The second dataset was compiled for the plant growth in decadal free air CO₂ enrichment (FACE) experiments.

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82 or not over time.

2 Materials and Methods

2.1 Data collection

For the first dataset, a comprehensive literature search with the terms of "CO₂ enrichment (or CO₂ increase)", "nitrogen" and "terrestrial" was conducted using the online search connection *Web of Science* in Endnote. Then, papers meeting the following two criteria were selected to do the further analyses: (i) including both control and CO₂ enrichment treatments, where the ambient and elevated CO₂ concentrations were around the current and predicted atmospheric CO₂ concentrations by the Intergovernmental Panel on Climate Change (IPCC, 2013), respectively (Fig. S1); (ii) including or from which we could calculate at least one of the major N pools or processes: soil TIN content, soil NH₄⁺ content, soil NO₃⁻ content, aboveground plant N pool (APNP), belowground plant N pool (BPNP), total plant N pool (TPNP), litter N pool (LNP), soil N pool (SNP), N fixation, nodule mass and/or number, net mineralization, nitrification, denitrification, and inorganic N leaching. Overall, there were 175 papers included in the first dataset (Table S1, References S1). For each paper, means, variations (standard deviation (*SD*), standard error (*SE*) or confidence intreval (*CI*) and sample sizes of the variables in both control and CO₂ enrichment treatments were collected.

For those studies that provided SE or CI, SD was computed by

$$SD = SE\sqrt{n}$$
 Eq. (1)

or
$$SD = (CI_u - CI_l)\sqrt{n}/2u_P$$
 Eq. (2)

where n is the sample size, CI_u and CI_l are the upper and lower limits of CI, and u_p is the significant level and equal to 1.96 and 1.645 when $\alpha = 0.05$ and 0.10, respectively. In some studies, if tissue N concentration and biomass were reported, we multiplied the two parts as N pools. When both APNP and BPNP were provided (or calculated), the two were added together

to represent the TPNP. When data from multiple soil layers were provided, they were summed if they were area-based (i.e., m⁻² land), or averaged if they were weight-based (i.e., g⁻¹ soil). In studies where the respective contents of NH₄⁺ and NO₃⁻ were reported, the TIN was calculated by adding the two together. For all the variables, if more than one result were reported during the experiment period, they were averaged by

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$$M = \sum_{i=1}^{j} \frac{M_i}{i}$$
 Eq. (3)

with standard deviation

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$$SD = \sqrt{\frac{\sum_{i=1}^{j} SD_i^2(n_i - 1)n_i}{\left(\sum_{i=1}^{j} n_i - 1\right)\sum_{i=1}^{j} n_i}}$$
Eq. (4)

where j is the number of results, M_i , SD_i and n_i are the mean, SD and sample size of the ith sampling data, respectively (Liang et al., 2013). If additional treatments applied (e.g., N addition), they were treated as independent studies.

Because treatment time and N addition may affect the responses of the N processes to CO_2 enrichment, the dataset was divided into different categories: (i) short-term (≤ 3 years) vs. long-term (> 3 years), and (ii) without N addition vs. with N addition. Moreover, the dataset was also divided into forest, grassland, and cropland to explore possible differences between ecosystem types.

For the second dataset, 15 available time series of plant growth were collected from 7 decadal FACE experiments (Table 1). The ecosystems included 9 forests, 5 grasslands and 1 desert. Because of the limited data, we included variables that can represent plant growth in one way or another, for example, net primary production (NPP), biomass, and leaf production. These data were collected to reveal whether the effect of CO₂ enrichment on plant growth diminishes over treatment time as proposed by the PNL hypothesis (Luo et al., 2004). In the 7 studies, the

treatment lasted from 7 to 13 years, and at least 6 years' production measurements were reported. For each data, the percentage change in NPP (or biomass or leaf production) by CO₂ enrichment was calculated. Then, a linear regression between the percentage change and the treatment year was conducted. A significantly negative slope indicates that the effect of CO₂ enrichment on the plant production diminishes over time. A non-significant slope was treated as 0. After deriving all the slopes, the frequency distribution of the slopes were fitted by a Gaussian function:

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$$y = y0 + ae^{-\frac{(x-\mu)^2}{2\sigma^2}}$$
 Eq. (5)

where x is the mean value of each individual interval, and y is the frequency of each interval. y0 is the base frequency. μ and σ are the mean and SD of the distribution.

138 2.2 Meta-analysis

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- With the first dataset, the effect of CO₂ enrichment for each line of data of the N variables was estimated using the natural logarithm transformed response ratio (*RR*) (Hedges et al., 1999;
- 141 Liang et al., 2013):

$$\log_e RR = \log_e(X_E/X_C) \qquad \text{Eq. (6)}$$

- where X_E and X_C are the variable values under enriched CO_2 and control conditions, respectively.
- The variation of the log RR was

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$$V = \left(\frac{SD_C^2}{n_C X_C^2} + \frac{SD_E^2}{n_E X_E^2}\right)$$
 Eq. (7)

- where SD_C and SD_E are the standard deviation of X_C and X_E , and n_C and n_E are the sample sizes of X_C and X_E .
- Then, the random-effects model was used to calculate the weighted mean. In the random-effects model, the weighted mean was calculated as

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$$M_{weighted} = \frac{\sum_{j=1}^{k} W_{j}^{*} M_{j}}{\sum_{j=1}^{k} W_{j}^{*}}$$
 Eq. (8)

with the variance as

$$V_{weighted} = \frac{1}{\sum_{j=1}^{k} W_j^*}$$
 Eq. (9)

where k is the number of studies, M_j is the Ln(RR) in study j, and W_j^* is the weighting factor which consists of between- and within-study variances (Rosenberg et al., 2000; Liang et al., 2013). The 95% lower and upper limits ($LL_{weighted}$ and $UL_{weighted}$) for the weighted mean were computed as

$$LL_{weighted} = M_{weighted} - 1.96 \times \sqrt{V_{weighted}}$$
 Eq. (10)

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$$UL_{weighted} = M_{weighted} + 1.96 \times \sqrt{V_{weighted}}$$
 Eq. (11)

The weighted mean and corresponding 95% bootstrapping CI (999 iterations) for each variable and category were calculated in MetaWin 2.1 (details are described in the software handbook by Rosenberg et al., 2000). The results were back-transformed and represented as percentage change by $(RR-1) \times 100\%$. The response was considered significant if the 95% CI did not overlap with zero.

3 Results

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The meta-analysis from the first dataset showed that CO₂ enrichment significantly increased N sequestered in plants and litter but not in SOM (Figs. 1A, S2). Whereas CO₂ enrichment had little overall effects on N mineralization, nitrification and denitrification, it significantly increased biological N fixation by 44.3% (with 95% CI from 29.5% to 61.8%). The increased biological N fixation was consistent when using various methods except H₂ evolution (Fig. 2A). In legume species, CO₂ enrichment significantly increased nodule mass and number (Fig. 2B). In addition, CO₂ enrichment increased N₂O emission by 10.7% (with 95% CI from 2.0% to 22.3%), but reduced leaching (i.e., -41.8% with 95% CI from -58.9% to -24.3%) (Fig. 1B). Although CO₂ enrichment did not change the total inorganic N availability in soils, it increased the soil NH_4^+/NO_3^- ratio by 16.9% (with 95% CI from 5.4% to 30.2%) (Fig. 1C). Treatment time had no effect on most of the variables (overlapped 95% CIs for short- and long-term treatments) except nitrification, which was not changed by short-term treatment, but was significantly reduced (-23.4% with 95% CI from -30.4% to -12.1%) by long-term CO₂ enrichment (Fig. 3B). In addition, it seemed that the responses of the NH₄+/NO₃- ratio was strengthened over time, representing a neutral response to short-term CO₂ enrichment, but significantly positive and negative responses to long-term CO₂ enrichment (Fig. 3C). The effects of CO₂ enrichment were influenced by N addition (Fig. 3D – F). For example, nitrification was significantly reduced by CO₂ enrichment without N addition by 19.3% (with 95% CI from -40.5% to -0.65%), but was not changed with N addition. Denitrification and N₂O emission responded to CO₂ enrichment neutrally without N addition, but significantly positively with N addition (Fig. 3E). Additionally, the responses of some variables to CO₂ enrichment were dependent on ecosystem type (Fig. 3G – I). APNP responded to CO₂ enrichment positively in forests and

enrichment in forests or grasslands, while it was significantly increased in croplands (Fig. 3H). Moreover, the change in the TIN was neutral in forests, grassland, but positive, in croplands, respectively (Fig. 3I). In addition, a positive response of the NH₄⁺/NO₃⁻ ratio was only observed in grasslands (Fig. 3I).

The results from the second dataset showed that CO_2 enrichment significantly increased plant growth in most of the decadal FACE experiments (Fig. 4). In addition, the CO_2 fertilization effect on plant growth did not over treatment time change in 11 experiments (P > 0.05), decreased in 2 experiments (slope < 0, P < 0.05), and increased in 2 experiments (slope > 0, P < 0.05), respectively (Table 1, Fig. 4). Overall, the slope of the response of the plant growth vs. treatment time was not significantly different from 0 (i.e., -0.37% year⁻¹ with 95% CI from -1.84% year⁻¹ to 1.09% year⁻¹; Fig. 4).

4 Discussion

In this study, we carried out two syntheses on the responses of the terrestrial N cycle and plant growth to CO₂ enrichment to test whether PNL generally occurs across ecosystems.

4.1 PNL alleviation

According to the PNL hypothesis, a prerequisite for PNL occurrence is that more N is sequestered in plant, litter and SOM (Luo et al., 2004). Our results showed that elevated CO₂ significantly increased N retention in plant tissues and litter, which is consistent with previous meta-analyses (de Graaff et al., 2006; Luo et al., 2006). Thus, the basis of PNL occurrence partially exists. However, the results from the second dataset did not show a general diminished

CO₂ fertilization effect on plant growth on the decadal scale, which disagrees with the expectation of the PNL hypothesis, suggesting that N supply under elevated CO₂ may meet the N demand. In this study, we have identified two processes that increase N supply under elevated CO₂, i.e., biological N fixation and leaching. CO₂ enrichment significantly enhanced the N influx to terrestrial ecosystems through biological N fixation, which reduces dinitrogen (N₂) to NH₄⁺ (Fig. 1B). The enhanced biological N fixation may have resulted from the stimulated activities of symbiotic (Fig. 2B) and free-living heterotrophic N-fixing bacteria (Hoque et al., 2001). In addition, the competition between N₂fixing and non-N₂-fixing species may have contributed to enhance the biological N fixation on the ecosystem level (Poorter and Navas, 2003; Batterman et al., 2013). In addition, the N efflux via leaching was reduced under elevated CO₂ conditions (Fig. 1B). This could be attributed to the decrease in NO₃, which is the primary N form in leaching, (Chapin III et al., 2011), and the increased root growth which may immobilize more inorganic N in soils (Luo et al., 2006; Iversen, 2010). In contrast, gaseous N loss through N₂O emission increased under elevated CO₂, although this increase was only observed when additional N was applied. The net effect of the responses of N processes to CO₂ enrichment resulted in higher N retention in ecosystems, especially within plant tissues and litter (Fig. S2). Because the product of biological N fixation (i.e., NH₄⁺) and the primary form for N leaching loss (i.e., NO₃⁻) can be directly used by plants, the effects of CO₂ enrichment on the two processes directly increase the N availability for plant growth, potentially alleviating PNL (Fig. 5). The increased N in plant tissues can be re-used by plants via resorption (Norby et al., 2000; Norby et al., 2001), and consequently reduce the N demand from soils. This may be another mechanism that alleviates

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235 PNL (Walker et al., 2015). Therefore, the increased N availability from increased N fixation and 236 reduced N leaching could potentially support net accumulation of organic matter in terrestrial ecosystems (Rastetter et al., 1997; Luo and Reynolds, 1999). 237 Since biological N fixation provides at least 30% of the N requirement across natural biomes 238 239 (Asner et al., 2001; Galloway et al., 2004), our results suggest that the positive response of 240 biological N fixation to CO₂ enrichment plays an important role in alleviating PNL. The PNL hypothesis was proposed to characterize long-term dynamics of C-N coupling in response to 241 rising atmospheric CO₂ concentration. Thus, it is critical to understand the long-term response of 242 243 biological N fixation to elevated CO₂. In this paper, we have synthesized 12 studies that lasted 4 - 7 years and binned them in a long-term category (> 3 years). On average of those long-term 244 studies, CO₂ enrichment increased biological N fixation by 26.2%. The increased biological N 245 fixation is supported by evidence at gene level from long-term experiments. For example, Tu et 246 al. (2015) found that the abundance of nifH gene amplicons, which is a widely used marker for 247 analyzing biological N fixation, was significantly enhanced by 12 years of CO₂ enrichment in a 248 249 grassland (BioCON). However, our synthesis showed a relatively wide 95% confidence interval 250 from 2.54% to 59.8%. The wide range can be partially attributed to the relatively small number 251 of studies. In addition, most studies incorporated in the current synthesis were conducted in temperate regions. Thus, longer-term studies, as well as studies in other regions (e.g., boreal and 252 tropical) are critically needed to reveal more general patterns in the future. 253 254 In this study, it is suggested that the general trend of the N cycle changes under elevated CO₂ converges towards increased soil N supply for plant growth, which in theory could alleviate PNL. 255 256 However, the PNL alleviation potential may vary across different ecosystems due to asymmetric 257 distributions of biological N fixation (Cleveland et al., 1999). In addition, PNL alleviation may

also be influenced by other factors. While most of the long-term experiments did not show a diminished CO₂ fertilization effect, it decreased in two sites (i.e., ORNL and Aspen-Birch) (Fig. 4). Plant growth is usually influenced by multiple environmental factors (e.g., nutrients, water, light, ozone). The undiminished CO₂ fertilization effect in most studies indicates that resource limitation (including N) was not aggravated, suggesting that no PNL occurred in these sites. However, in the ORNL and Aspen-Birch (without O₃ treatment) experiments, the diminished CO₂ fertilization effect could be attributed to limitation of N, or other resources, or their combined effect. For example, reduced N availability has been identified as one of the primary factors that lead to the diminished CO₂ fertilization effect on NPP in the ORNL FACE experiment (Norby et al., 2010). In the Aspen-Birch community, however, the deceleration of leaf area increases due to canopy closure was responsible for the diminished CO₂ fertilization effect without O₃ addition (Talhelm et al., 2012). With O₃ addition, O₃ significantly reduced the canopy development, resulting in a relatively open canopy during the experiment period. In addition, the negative effect of O₃ addition increased over time, leading to the apparent increase in the CO₂ fertilization effect (Fig. 4) (Talhelm et al., 2012).

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4.2 Dependence of the responses of N cycling processes upon methodology, treatment duration, N addition and ecosystem types

Methodology may potentially influence findings. Cabrerizo et al. (2001) found that CO₂ enrichment increased the nitrogenase activity measured by acetylene reduction assay (ARA), but not the specific N fixation measured by the H₂ evolution method. In the studies synthesized here, four methods were used to estimate biological N fixation, including isotope, ARA, H₂ evolution and N accumulation. Among them, ARA and H₂ evolution measure nitrogenase activity (Hunt

and Layzell, 1993) whereas isotope and N accumulation methods directly measure biological N fixation. All but the H₂ evolution method showed a significantly positive response to CO₂ enrichment (Fig. 2A). The insignificant response shown by the H₂ evolution method was likely because of the small study numbers (i.e., 3). In addition, the biological N fixation measured by ARA, isotope and N accumulation showed similar response magnitudes (Fig. 2A), suggesting consistency among the three methods. However, further assessment on the H₂ evolution method is needed.

The responses of some N cycling processes that affect N availability are dependent on treatment duration, N addition, and/or ecosystem types (Fig. 3).

N mineralization, in addition to biological N fixation, is a major source of available N in soils. Our meta-analysis showed no change in the net N mineralization in response to CO₂ enrichment, which is consistent with the results by de Graaff et al. (2006). However, the response of net mineralization was dependent upon ecosystem types, showing no change in forests and grasslands, but significant increases in croplands (Fig. 3H). There may be two reasons for the stimulated net mineralization in croplands. First, N fertilization, which is commonly practiced in croplands, can increase the substrate quantity and quality for mineralization (Barrios et al., 1996; Chapin III et al., 2011; Booth et al., 2005; Lu et al., 2011; Reich and Hobbie, 2013). Second, tillage can alter soil conditions (e.g., increasing O₂ content), which can potentially favor the N mineralization under enriched CO₂ (Wienhold and Halvorson, 1999; Bardgett and Wardle, 2010). These findings suggest that CO₂ enrichment can stimulate the N transfer from organic to inorganic forms in managed croplands.

Unlike leaching, the response of nitrification was dependent upon treatment duration (Fig. 3).

Nitrification was not changed by short-term treatment, but was significantly reduced by long-

term CO₂ enrichment (Fig. 3). One possible reason for the reduced nitrification with long-term CO₂ enrichment is the cumulative effect of hydrological changes. CO₂ enrichment generally reduces stomatal conductance and, consequently, water loss via plant transpiration, leading to an increase in soil water content (Niklaus et al., 1998; Tricker et al., 2009; van Groenigen et al., 2011; Keenan et al., 2013). A synthesis by van Groenigen et al. (2011) shows that CO₂ enrichment increases soil water content by 2.6% –10.6%. Increased soil water content may result in less oxygen (O₂) concentration in soils, which could potentially constrain nitrification. In addition, the response of gaseous N loss was dependent on N addition (Fig. 3). The reduced nitrification was only observed under conditions without N addition (Fig. 3E). With N addition, no response of nitrification to CO₂ enrichment was observed (Fig. 3E). Additionally, the response of denitrification to CO₂ enrichment shifted from neutral, without N addition, to significantly positive with N addition (Fig. 3E). One possible reason is that N addition provides more N substrate for nitrifying and denitrifying bacteria (Keller et al., 1988; Stehfest and Bouwman, 2006; Russow et al., 2008). The strengthening trends of both nitrification and denitrification led to a shift of the response of N₂O emission to CO₂ enrichment from neutral without N addition to significantly positive with N addition (Fig. 3E). Our results indicate that CO₂ enrichment significantly increases gaseous N loss when additional N is applied. Our results are consistent with a previous synthesis (van Groenigen et al. 2011). Increased N₂O emissions can partially offset the mitigation of climate change by the stimulated plant CO₂ assimilation as the warming potential of N₂O is 296 times that of CO₂. However, a recent modeling study by Zaehle et al. (2011) found an opposite result showing that CO₂ enrichment reduced radiative forcing of N₂O. In their model, less availability of N substrates for nitrification and denitrification due to the enhanced plant N sequestration attributed to the reduced N2O

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emission. Our synthesis shows that inorganic N does not decrease. Especially with additional N application, enhanced denitrification by CO_2 enrichment results in a greater N_2O emission.

4.3 Changes in soil microenvironment, community structures and above-belowground

The meta-analysis showed that the two major forms of soil available N, NH₄⁺ and NO₃⁻,

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responded to long-term CO₂ enrichment in opposing manners (Fig. 3C). While the enhanced biological N fixation by CO₂ enrichment tended to increase the NH₄⁺ content in soils, the reduced nitrification decreased the NO₃⁻ content in soils, leading to a significant increase in the NH₄⁺/NO₃⁻ ratio (Fig. 3C). Although the total available N did not change under elevated CO₂, the altered proportion of NH₄⁺ over NO₃⁻ in soils may have long-term effects on soil microenvironment and associated aboveground-belowground linkages that control the C cycle (Bardgett and Wardle, 2010). On the one hand, plants would release more hydrogen ion (H⁺) to regulate the charge balance when taking up more NH₄⁺. As a result, the increased NH₄⁺ absorption could acidify the rhizosphere soil (Thomson et al., 1993; Monsant et al., 2008). The lowered pH could have significant effects on soil microbial communities and their associated ecosystem functions. For example, fungal/bacterial ratio increases with the decrease in pH (de Vries et al., 2006; Rousk et al., 2009). The increased fungal/bacterial ratio may result in lower N mineralization because of the higher C/N ratio of fungi and the lower turnover rates of fungal-feeding fauna (de Vries et al., 2006; Rousk and Bååth, 2007). In other words, the increased fungal/bacterial ratio may slow down the N turnover from organic to inorganic forms. On the other hand, the increased NH₄⁺/NO₃⁻ ratio may increase the N use efficiency because it is more energetically expensive for plants to utilize

NO₃⁻ than NH₄⁺ (Chapin III et al., 2011; Odum and Barrett, 2005; Lambers et al., 2008). In addition, since the preferences for plant absorption of different forms of N are different (Chapin III et al., 2011; Odum and Barrett, 2005), the increased NH₄⁺/NO₃⁻ ratio may benefit some plant species while depress others, and consequently alter the community structures over time. These diverse changes in soil microenvironment and microbial and plant community compositions could further affect the terrestrial C cycle on long temporal scales, on which more studies are needed.

5 Summary

This study synthesizes data in the literature on the effects of CO₂ enrichment on the terrestrial N cycle to improve our understanding of the N limitation to plant growth under elevated CO₂. Our results indicate that elevated CO₂ stimulates N influx via biological N fixation but reduces N loss via leaching, leading to increased N supply for plant growth. The additional N supply via the enhanced biological N fixation and the reduced leaching may partially meet the increased N demand under elevated CO₂, potentially alleviating PNL. In addition, increased N₂O emissions can partially offset the mitigation of climate change by stimulated plant CO₂ assimilation.

Moreover, changes in soil microenvironments, ecosystem communities and above-belowground interactions induced by the different responses of NH₄⁺ and NO₃⁻ to CO₂ enrichment may have long-term effects on the terrestrial biogeochemical cycles and climate change.

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References

- Asner, G.P., Townsend, A.R., Riley, W.J., Matson, P.A., Neff, J.C., and Cleveland, C.C.:
- Physical and biogeochemical controls over terrestrial ecosystem responses to nitrogen
- deposition, Biogeochemistry, 54, 1-39, 2001.
- Bardgett, R. D. and Wardle, D. A.: Aboveground-belowground linkages: biotic interactions,
- ecosystem processes, and global change, Oxford University Press, 2010.
- Barrios, E., Buresh, R. J., and Sprent, J. I.: Nitrogen mineralization in density fractions of soil
- organic matter from maize and legume cropping systems, Soil Biology & Biochemistry, 28,
- 385 1459-1465, 1996.
- Batterman, S. A., Hedin, L. O., van Breugel, M., Ransijn, J., Craven, D. J., and Hall, J. S.: Key
- role of symbiotic dinitrogen fixation in tropical forest secondary succession, Nature, 502,
- 388 224-227, 2013.
- Booth, M. S., Stark, J. M., and Rastetter, E.: Controls on nitrogen cycling in terrestrial
- ecosystems: A synthetic analysis of literature data, Ecological Monographs, 75, 139-157,
- 391 2005.
- Cabrerizo, P. M., González, E. M., Aparicio-Tejo, P. M., and Arrese-Igor, C.: Continuous CO₂
- enrichment leads to increased nodule biomass, carbon availability to nodules and activity of
- carbon-metabolising enzymes but does not enhance specific nitrogen fixation in
- pea, Physiologia Plantarum, 113, 33-40, 2001.

- Chapin III, F. S., Matson, P. A., and Vitousek, P.: Principles of terrestrial ecosystem ecology,
- 397 Springer, 2011.
- Cleveland, C. C., Townsend, A. R., Schimel, D. S., Fisher, H., Howarth, R. W., Hedin, L. O.,
- Perakis, S. S., Latty, E. F., Von Fischer, J. C., Elseroad, A., and Wasson, M. F.: Global
- 400 patterns of terrestrial biological nitrogen (N₂) fixation in natural ecosystems, Global
- 401 Biogeochemical Cycles, 13, 623-645, 1999.
- de Graaff, M. A., van Groenigen, K. J., Six, J., Hungate, B., and van Kessel, C.: Interactions
- between plant growth and soil nutrient cycling under elevated CO₂: a meta-analysis, Global
- 404 Change Biology, 12, 2077-2091, 2006.
- de Vries, F. T., Hoffland, E., van Eekeren, N., Brussaard, L., and Bloem, J.: Fungal/bacterial
- ratios in grasslands with contrasting nitrogen management, Soil Biology & Biochemistry, 38,
- 407 2092-2103, 2006.
- 408 Finzi, A. C., Moore, D. J. P., Delucia, E. H., Lichter, J., Hofmockel, K. S., Jackson, R. B., Kim,
- H., Matamala, R., McCarthy, H. R., Oren, R., Pippen, J. S., Schlesinger, W. H.: Progressive
- 410 nitrogen limitation of ecosystem processes under elevated CO₂ in a warm-temperate forest,
- 411 Ecology, 87, 15-25, 2006.
- 412 Galloway, J. N., Dentener, F. J., Capone, D. G., Boyer, E. W., Howarth, R. W., Seitzinger, S. P.,
- Asner, G. P., Cleveland, C. C., Green, P. A., Holland, E. A., and Karl, D. M.: Nitrogen cycles:
- past, present, and future, Biogeochemistry, 70, 53-226, 2004.
- 415 Hedges, L. V., Gurevitch, J., and Curtis, P. S.: The meta-analysis of response ratios in
- experimental ecology, Ecology, 80, 1150-1156, 1999.
- Hoque, M. M., Inubushi, K., Miura, S., Kobayashi, K., Kim, H. Y., Okada, M., and Yabashi, S.:
- Biological dinitrogen fixation and soil microbial biomass carbon as influenced by free-air

- carbon dioxide enrichment (FACE) at three levels of nitrogen fertilization in a paddy field,
- 420 Biology and Fertility of Soils, 34, 453-459, 2001.
- Hunt, S. and Layzell, D.B.: Gas exchange of legume nodules and the regulation of nitrogenase
- activity, Annual Review of Plant Physiology and Plant Molecular Biology, 44, 483-511,
- 423 1993.
- 424 IPCC: Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to
- the Fifth Assessment Report of the Intergovernmental Panel on Climate Change, Cambridge
- 426 University Press, Cambridge, United Kingdom and New York, NY, USA, 2013.
- 427 Iversen, C. M.: Digging deeper: fine-root responses to rising atmospheric CO₂ concentration in
- forested ecosystems, New Phytologist, 186, 346-357, 2010.
- Keenan, T. F., Hollinger, D. Y., Bohrer, G., Dragoni, D., Munger, J. W., Schmid, H. P., and
- Richardson, A. D.: Increase in forest water-use efficiency as atmospheric carbon dioxide
- concentrations rise, Nature, 499, 324-+, 2013.
- Keller, M., Kaplan, W. A., Wofsy, S. C., and Dacosta, J. M.: Emissions of N₂O from Tropical
- Forest Soils: Response to Fertilization with NH₄⁺, NO3⁻, and PO₄³⁻, Journal of Geophysical
- 434 Research-Atmospheres, 93, 1600-1604, 1988.
- Lambers, H., Chapin III, F. S., and Pons, T. L.: Plant Physiological Ecology, Springer, New
- 436 York, 2008.
- Liang, J., Xia, J., Liu, L., and Wan, S.: Global patterns of the responses of leaf-level
- photosynthesis and respiration in terrestrial plants to experimental warming, Journal of Plant
- 439 Ecology, 6, 437-447, 2013.

- 440 Lu, M., Yang, Y. H., Luo, Y. Q., Fang, C. M., Zhou, X. H., Chen, J. K., Yang, X., and Li, B.:
- Responses of ecosystem nitrogen cycle to nitrogen addition: a meta-analysis, New
- 442 Phytologist, 189, 1040-1050, 2011.
- Luo, Y. and Reynolds, J. F.: Validity of extrapolating field CO₂ experiments to predict carbon
- sequestration in natural ecosystems, Ecology, 80, 1568-1583, 1999.
- Luo, Y., Su, B., Currie, W. S., Dukes, J. S., Finzi, A. C., Hartwig, U., Hungate, B., McMurtrie, R.
- E., Oren, R., Parton, W. J., Pataki, D. E., Shaw, M. R., Zak, D. R., and Field, C. B.:
- Progressive nitrogen limitation of ecosystem responses to rising atmospheric carbon dioxide,
- 448 Bioscience, 54, 731-739, 2004.
- Luo, Y. Q., Hui, D. F., and Zhang, D. Q.: Elevated CO₂ stimulates net accumulations of carbon
- and nitrogen in land ecosystems: A meta-analysis, Ecology, 87, 53-63, 2006.
- 451 McCarthy, H. R., Oren, R., Johnsen, K. H., Gallet-Budynek, A., Pritchard, S. G., Cook, C. W.,
- LaDeau, S. L., Jackson, R. B., and Finzi, A. C.: Re-assessment of plant carbon dynamics at
- 453 the Duke free-air CO₂ enrichment site: interactions of atmospheric [CO₂] with nitrogen and
- water availability over stand development, New Phytologist, 185, 514-528, 2010.
- 455 Monsant, A. C., Tang, C., and Baker, A. J. M.: The effect of nitrogen form on rhizosphere soil
- 456 pH and zinc phytoextraction by Thlaspi caerulescens, Chemosphere, 73, 635-642, 2008.
- 457 Moore, D. J. P., Aref, S., Ho, R. M., Pippen, J. S., Hamilton, J. G., de Lucia, E. H.: Annual basal
- area increment and growth duration of *Pinus taeda* in response to eight years of free-air
- 459 carbon dioxide enrichment, Global Change Biology, 12, 1367-1377, 2006.
- Niklaus, P. A., Spinnler, D., and Korner, C.: Soil moisture dynamics of calcareous grassland
- under elevated CO₂, Oecologia, 117, 201-208, 1998.

- Norby, R. J., Cotrufo, M. F., Ineson, P., O'Neill, E. G., and Canadell, J. G.: Elevated CO₂, litter
- chemistry, and decomposition: a synthesis, Oecologia, 127, 153-165, 2001.
- Norby, R. J., Long, T. M., Hartz-Rubin, J. S., and O'Neill, E. G.: Nitrogen resorption in
- senescing tree leaves in a warmer, CO₂-enriched atmosephere, Plant and Soil, 224, 15-29,
- 466 2000.
- Norby, R. J., Warren, J. M., Iversen, C. M., Medlyn, B. E., and McMurtrie, R. E.: CO₂
- 468 enhancement of forest productivity constrained by limited nitrogen availability, Proceedings
- of the National Academy of Sciences of the United States of America, 107, 19368-19373,
- 470 2010.
- Odum, E. P. and Barrett, G. W.: Fundamentals of Ecology, Thomson Brooks/Cole, 2005.
- Oren, R., Ellsworth, D. S., Johnsen, K. H., Phillips, N., Ewers, B. E., Maier, C., Schafer, K. V.
- R., McCarthy, H., Hendrey, G., McNulty, S. G., and Katul, G. G.: Soil fertility limits carbon
- sequestration by forest ecosystems in a CO₂-enriched atmosphere, Nature, 411, 469-472,
- 475 2001.
- Poorter, H. and Navas, M. L.: Plant growth and competition at elevated CO₂: on winners, losers
- and functional groups, New Phytologist, 157, 175-198, 2003.
- 478 Rastetter, E. B., Agren, G. I., and Shaver, G. R.: Responses of N-limited ecosystems to increased
- 479 CO₂: A balanced-nutrition, coupled-element-cycles model, Ecological Applications, 7, 444-
- 480 460, 1997.
- 481 Reay, D. S., Dentener, F., Smith, P., Grace, J., and Feely, R. A.: Global nitrogen deposition and
- carbon sinks, Nature Geoscience, 1, 430-437, 2008.
- Reich, P. B. and Hobbie, S. E.: Decade-long soil nitrogen constraint on the CO₂ fertilization of
- plant biomass, Nature Climate Change, 3, 278-282, 2013.

- Reich, P. B., Hobbie, S. E., Lee, T., Ellsworth, D. S., West, J. B., Tilman, D., Knops, J. M. H.,
- Naeem, S., and Trost, J.: Nitrogen limitation constrains sustainability of ecosystem response
- 487 to CO₂, Nature, 440, 922-925, 2006.
- Reich, P. B., Hobbie, S. E., and Lee, T. D.: Plant growth enhancement by elevated CO₂
- eliminated by joint water and nitrogen limitation, Nature Geoscience, 7, 920-924, 2014.
- 490 Rosenberg, M. S., Adams, D. C., and Gurevitch, J.: MetaWin: statistical software for meta-
- analysis, Sinauer Associates Sunderland, Massachusetts, USA, 2000.
- 492 Ross, D. J., Newton, P. C. D., Tate, K. R., and Luo, D. W.: Impact of a low level of CO₂
- enrichment on soil carbon and nitrogen pools and mineralization rates over ten years in a
- seasonally dry, grazed pasture, Soil Biology & Biochemistry, 58, 265-274, 2013.
- Rousk, J. and Bååth, E.: Fungal biomass production and turnover in soil estimated using the
- acetate-in-ergosterol technique, Soil Biology & Biochemistry, 39, 2173-2177, 2007.
- 497 Rousk, J., Brookes, P. C., and Baath, E.: Contrasting Soil pH Effects on Fungal and Bacterial
- 498 Growth Suggest Functional Redundancy in Carbon Mineralization, Applied and
- Environmental Microbiology, 75, 1589-1596, 2009.
- Russow, R., Spott, O., and Stange, C. F.: Evaluation of nitrate and ammonium as sources of NO
- and N₂O emissions from black earth soils (Haplic Chernozem) based on ¹⁵N field
- experiments, Soil Biology & Biochemistry, 40, 380-391, 2008.
- 503 Schneider, M. K., Luscher, A., Richter, M., Aeschlimann, U., Hartwig, U. A., Blum, H., Frossard,
- E., and Nosberger, J.: Ten years of free-air CO₂ enrichment altered the mobilization of N
- from soil in *Lolium perenne* L. swards, Global Change Biology, 10, 1377-1388, 2004.

- 506 Smith, S. D., Charlet, T. N., Zitzer, S. F., Abella, S. R., Vanier, C. H., and Huxman, T. E.: Long-
- term response of a Mojave Desert winter annual plant community to a whole-ecosystem
- atmospheric CO₂ manipulation (FACE), Global Change Biology, 20, 879-892, 2014.
- 509 Stehfest, E. and Bouwman, L.: N₂O and NO emission from agricultural fields and soils under
- natural vegetation: summarizing available measurement data and modeling of global annual
- emissions, Nutrient Cycling in Agroecosystems, 74, 207-228, 2006.
- Talhelm, A. F., Pregitzer, K. S., and Giardina, C. P.: Long-Term Leaf Production Response to
- Elevated Atmospheric Carbon Dioxide and Tropospheric Ozone, Ecosystems, 15, 71-82,
- 514 2012.
- Thomson, C. J., Marschner, H., and Romheld, V.: Effect of Nitrogen-Fertilizer Form on pH of
- the Bulk Soil and Rhizosphere, and on the Growth, Phosphorus, and Micronutrient Uptake of
- 517 Bean, Journal of Plant Nutrition, 16, 493-506, 1993.
- Tricker, P. J., Pecchiari, M., Bunn, S. M., Vaccari, F. P., Peressotti, A., Miglietta, F., and Taylor,
- G.: Water use of a bioenergy plantation increases in a future high CO₂ world, Biomass &
- 520 Bioenergy, 33, 200-208, 2009.
- Tu, Q., Zhou, X., He, Z., Xue, K., Wu, L., Reich, P., Hobbie, S., and Zhou, J.: The Diversity and
- 522 Co-occurrence Patterns of N₂-Fixing Communities in a CO₂-Enriched Grassland
- Ecosystem, Microbial Ecology, doi:10.1007/s00248-015-0659-7, 2015.
- van Groenigen, K. J., Osenberg, C. W., and Hungate, B. A.: Increased soil emissions of potent
- greenhouse gases under increased atmospheric CO2, Nature, 475, 214-U121, 2011.
- Vitousek, P. M.: Nutrient cycling and limitation: Hawai'i as a model system, Princeton
- 527 University Press, 2004.

Walker, A. P., Zaehle, S., Medlyn, B. E., De Kauwe, M. G., Asao, S., Hickler, T., Parton, W., 528 Ricciuto, D. M., Wang, Y. P., and Wårlind, D.: Predicting long-term carbon sequestration in 529 response to CO₂ enrichment: How and why do current ecosystem models differ?, Global 530 531 Biogeochemical Cycles, 29, 476-495, 2015. 532 Wienhold, B. J. and Halvorson, A. D.: Nitrogen mineralization responses to cropping, tillage, and nitrogen rate in the Northern Great Plains, Soil Science Society of America Journal, 63, 533 192-196, 1999. 534 535 Zaehle, S., Ciais, P., Friend, A.D. and Prieur, V.: Carbon benefits of anthropogenic reactive 536 nitrogen offset by nitrous oxide emissions. Nature Geoscience, 4, 601-605, 2011.

Supporting Information captions 538 Figure S1 Distributions of the experimental duration (A) and the CO₂ concentrations under 539 ambient (B) and elevated (C) treatments and their difference (D) for the 175 collected studies. 540 541 Red dashed lines represent the mean values. 542 Figure S2 Summary of the effect of CO₂ enrichment on ecosystem level N budget. Square boxes 543 are nitrogen pools, ovals are nitrogen processes. Red dashed boxes mean the sum of the pools in 544 the boxes. "+", "-", and "ns" mean the response to CO₂ enrichment are positive, negative, and 545 not significant, respectively. Please see **Figure 1** for abbreviations. 546 547 **Database S1** Database extracted from papers listed in References S1. 548 549 References S1 Papers from which the first dataset was extracted. 550 551

Table 1. Results on the effect of CO_2 enrichment on ecosystem NPP (or biomass or leaf production) in decadal-long free air CO_2 enrichment (FACE) experiments over treatment time. The values of the slope, R^2 and P in the linear regression in **Fig. 4** are shown. The lower and upper n (i.e., n and N) in Refs. Schneider et al., 2004; McCarthy et al., 2010; Reich and Hobbie, 2013 mean without and with N addition, respectively. The lower and upper o (i.e., o and O) in Ref. Talhelm et al., 2012 mean without and with O_3 treatment, respectively.

	Ecosystem	Treatment					
Experiment	type	years	Variable	Slope	R^2	P	Reference
Duke_n	Forest	8	NPP	0.50	0.25	0.21	McCarthy et al., 2010
Duke_N	Forest	8	NPP	-1.39	0.27	0.29	McCarthy et al., 2010
ORNL	Forest	11	NPP	-1.42	0.38	0.04	Norby et al., 2010
BioCON_n	Grassland	13	Biomass	0.42	0.05	0.48	Reich and Hobbie, 2013
BioCON_N	Grassland	13	Biomass	0.23	0.01	0.76	Reich and Hobbie, 2013
NZ	Grassland	10	Biomass	0.95	0.05	0.53	Ross et al., 2013
Swiss_n	Grassland	10	Harvestable biomass	0.30	0.01	0.75	Schneider et al., 2004
Swiss_N	Grassland	10	Harvestable biomass	1.66	0.47	0.03	Schneider et al., 2004
NDFF	Desert	9	Standing biomass	-9.54	0.15	0.40	Smith et al., 2014
Aspen_o	Forest	7	Leaf production	-0.07	0.00	0.97	Talhelm et al., 2012
Aspen_O	Forest	7	Leaf production	0.09	0.00	0.93	Talhelm et al., 2012
AspenBirch_o	Forest	7	Leaf production	-5.27	0.77	0.01	Talhelm et al., 2012
AspenBirch_O	Forest	7	Leaf production	6.48	0.82	0.00	Talhelm et al., 2012
AspenMaple_o	Forest	7	Leaf production	-9.16	0.40	0.13	Talhelm et al., 2012
_AspenMaple_O	Forest	7	Leaf production	1.11	0.11	0.46	Talhelm et al., 2012

558 Figure captions **Figure 1.** Results of a meta-analysis on the responses of nitrogen pools and processes to 559 CO₂ enrichment. In (A), APNP, BPNP, TPNP, LNP, and SNP are the abbreviations for 560 aboveground plant nitrogen pool, belowground plant nitrogen pool, total plant nitrogen 561 pool, litter nitrogen pool, and soil nitrogen pool, respectively. In (C), TIN, NH₄⁺ and 562 563 NO₃ are total inorganic nitrogen, ammonium, and nitrate in soils, respectively. The error bars represent 95% confidence intervals. 564 565 566 Figure 2. Responses of biological N fixation measured by different methods (A) and nodule dry mass and number in legume species (B). ARA: acetylene reduction assay. 567 Mean \pm 95% confidence interval. 568 569 Figure 3. Responses of terrestrial nitrogen pools and processes to CO₂ enrichment (Mean 570 \pm 95% confidence interval) as regulated by experimental durations (A – C; short-term: \leq 571 3 years vs. long-term: > 3 years), nitrogen addition (D – F), and ecosystem types (G – I). 572 Please see Figure 1 for abbreviations. 573 574 **Figure 4.** Time courses of CO₂ effects on ecosystem NPP (or biomass or leaf production) 575 in decadal-long FACE experiments. Please see **Table 1** for details of experiments, 576 577 references and statistical results. Only statistically significant (P < 0.05) regression lines are shown. The panel at the right-low corner shows the distribution of the slopes (-0.37%) 578 year⁻¹ with 95% CI from -1.84% year⁻¹ to 1.09% year⁻¹). 579

Figure 5. Mechanisms that alleviate PNL. PNL hypothesis posits that the stimulated plant growth by CO₂ enrichment leads to more N sequestered in long-lived plant tissues, litter and soil organic matter (SOM) so that, the N availability for plant growth progressively declines over time, and plant growth is downregulated (grey symbols). The current synthesis indicates that the basis of PNL occurrence partially exists (i.e., more N sequestered in plant tissues and litter; black symbols). Despite of the increases in plant N sequestration and N₂O emission, stimulated biological N fixation and reduced N leaching can replenish the N availability, potentially alleviating PNL (blue boxes and arrows). Upward, downward, and horizontal arrows mean increase, decrease, and no change, respectively.

Figure 1.

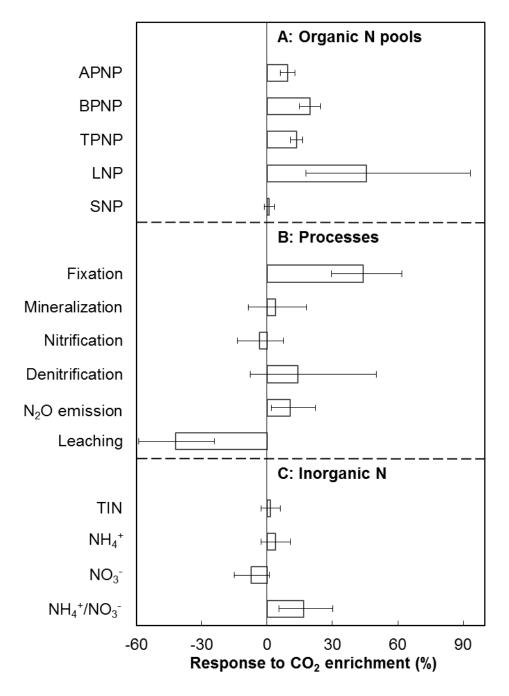


Figure 2.

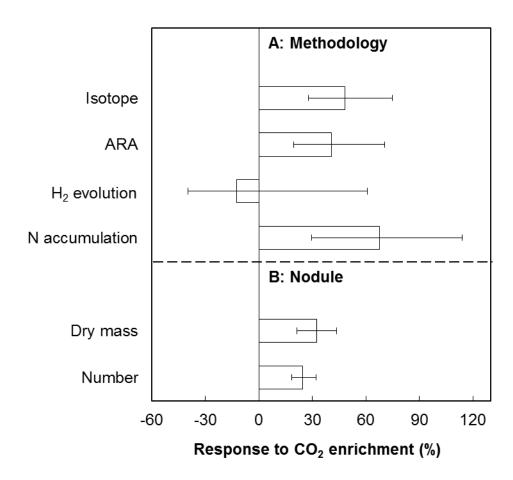


Figure 3.

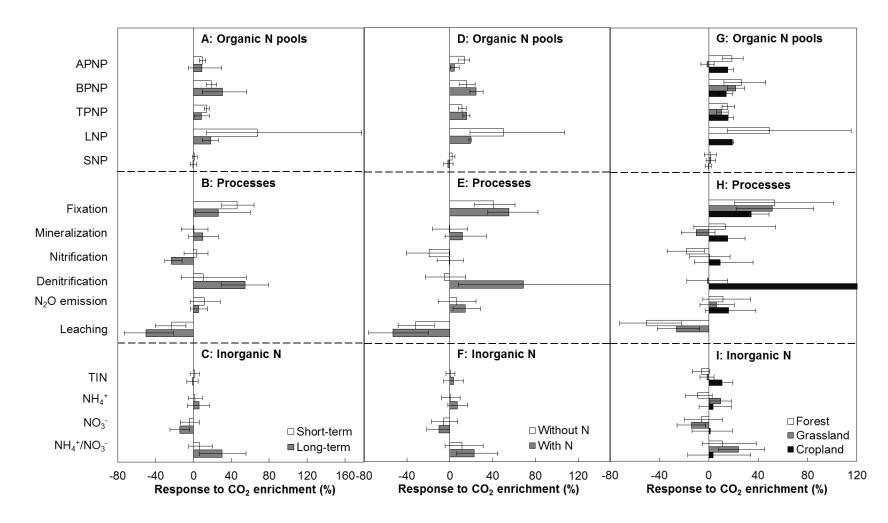


Figure 4.

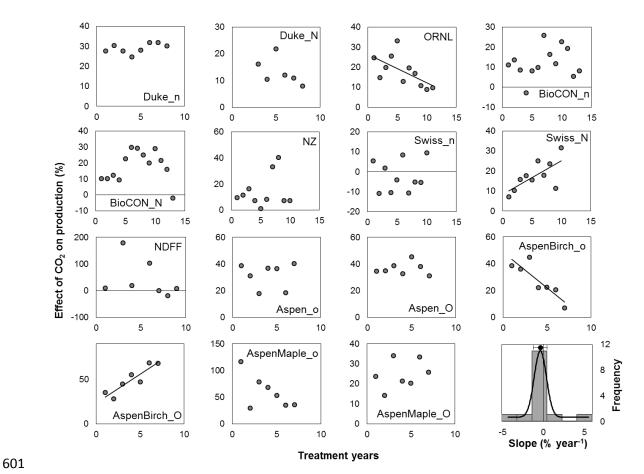


Figure 5.

