

1 Author's response to anonymous reviewer #2

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3 We really appreciate thoroughly giving comments and helpful suggestion to our study by anonymous
4 reviewer #2. And I am grateful for careful proofreading on our manuscript for publication in
5 Beogeosciences. We have answered all the comments made by the referees #2. RC, AR and RS stand
6 for Reviewers comment, Authors response and Revised sentence, respectively. We hope that our
7 explanations and revise plan are acceptable and satisfactory.

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9 [RC-1-1] P1L19: "copepods" instead of "copepod"

10 [AR-1-1] I replaced "copepod" "copepods".

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12 [RC-1-2] P2L5: "initiate" instead of "cause"

13 [AR-1-2] I replaced "cause" "initiate".

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15 [RC-1-3] P2L6: "early" instead of "earlier"

16 [AR-1-3] I replaced "earlier" "early".

17

18 [RC-1-4] P2L11: "has" instead of "appears to have"

19 [AR-1-4] I replaced "appears to have" "has".

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21 [RC-1-5] P2L14: remove "very" before "shallow"

22 [AR-1-5] I removed "very" before "shallow".

23

24 [RC-1-6] P2L14: add "extensive" before "shallow"

25 [AR-1-6] I added "extensive" before "shallow".

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27 [RC-1-7] P2L16: Add a reference to back this statement. Otherwise remove it.

28 [AR-1-7] Thank you for helpful comment. I removed it.

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[RC-1-8] P2L23: “matter” instead of “material”

[AR-1-8] I replaced “material” “matter”.

[RC-1-9] P2L31: “merge” instead of “merged”

[AR-1-9] I replaced “merged” “merge”.

[RC-1-10] P3L19: replace “because” “as”

[AR-1-10] I replaced “because” “as”.

[RC-1-11] P3L26: NORPAC is not explained here. I would remove it and keep the definition to the M&M

[AR-1-11] I removed it from this section. And I added “North Pacific Standard (” before the explanation of NORPAC net.

[RS-1-11] P4L9-12: Zooplankton samples were collected during the day or at night using vertical tows with a North Pacific Standard (NORPAC) net (mouth diameter 45 cm, mesh size 335 μm) from 5 m above the bottom to the surface (the depths of most stations were approximately 50 m).

[RC-1-12] P3L32: Is there a scientific question that can be formulated from what has been described before?

[AR-1-12] Yes, I have scientific question that there is the difference of response for environmental change between species. The other reviewer also indicated about it. So, I changed the last paragraph of introduction as [RC-12].

[RS-1-12] P3L24–P4L4: The distribution patterns of both Pacific and Arctic copepods in the Arctic seas have been reported in these previous studies. However, recent and future drastic climate changes potentially trigger the shifts in the distributions of copepod species or change of their habitat. This phenomenon has already been reported for some species (e.g., Eisner et al., 2014; Ershova et al., 2015). In order to comprehend the response of each copepod group to the environmental changes in the Arctic, a statistical understanding of the relationship between environmental factors and the group’s abundance is required. Since Pacific and Arctic copepods have different life-cycles, suitable habitat,

1 and reproductive characteristics, their response to the environmental changes are expected to differ.
2 Therefore in the present study, we aim to construct an adequate model to illustrate the suitable
3 environmental characteristics for each Pacific and Arctic copepods group that will help us predict the
4 risks they might face in the future. Here, we propose the use of generalized additive models (GAMs)
5 to determine the factors affecting the spatial pattern of copepod abundances based on data collected by
6 net-sampling during the summers of 2007, 2008, and 2013.

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8 [RC-1-13] Use scientific writing of dates: 30 July and not July 30. July 30–August 24, June 30–July
9 13 and July 4–17 should be changed 30 July–24 August, 30 June–13 July and 4–17 July, respectively.

10 [AR-1-13] I modified them. Please check [RS-13].

11 [RS-1-13] P3L7–9: We sampled copepods and water onboard of T/S *Oshoro-maru* (Hokkaido
12 University) during 30 July–24 August 2007 (31 stations), 30 June–13 July 2008 (26 stations), and 4–
13 17 July 2013 (31 stations; Fig. 1).

14

15 [RC-1-14] P4L6: replace “NORPAC” “North Pacific Standard (NORPAC)” before “net”

16 [AR-1-14] I added “North Pacific Standard (” before “NORPAC”.

17

18 [RC-1-15] P4L10: remove “the” before “land” and “identification”

19 [AR-1-15] I removed remove “the” before “land” and “identification”.

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21 [RC-1-15] P4L12: remove “the” before “identification”

22 [AR-1-15] I removed remove “the” before “identification”.

23

24 [RC-1-16] P4L13: replace “listed” “list”

25 [AR-1-16] I replaced “listed” “list”.

26

27 [RC-1-17] P4L20: replace “measured” “made” and remove “a” before “conductivity-temperature-
28 depth”

29 [AR-1-17] I replaced “measured” “made” and remove “a” before “conductivity-temperature-depth”.

1
2 [RC-1-18] move “(<100 mmHg)” to before “filtered”
3 [RC-1-18] I removed “(<100 mmHg)” at the end of sentence and add it before “filtered”. Please check
4 [RS-1-18].
5 [RS-1-18] P5L4-5: Water samples were gently filtered (<100 mmHg) onto GF/F filters.
6
7 [RC-1-19] P4L28: “nsicde” instead of “nsicde”
8 [AR-1-19] I’m sorry. It’s my misspelling. I modified it.
9
10 [RC-1-20] P4L30: “relationship” instead of “relationships”
11 [AR-1-20] I replaced “relationships” “relationship”.
12
13 [RC-1-21] P5L2: “complex” instead of “complicated”, and remove “quantitatively” before on
14 [AR-1-21] I replaced “complicated” “complex” and removed “quantitatively.”
15
16 [RC-1-22] P5L4: “group” instead of “groups”
17 [AR-1-22] I replaced “groups” “group”.
18
19 [RC-1-23] P5L5: “variables” instead of “variable”.
20 [AR-1-23] I replaced “variable” “variables”.
21
22 [RC-1-24] P5L13: add “the” before “shallow”
23 [AR-1-24] I added “the” before “shallow”.
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25 [RC-1-25] P7L27: add “in” before Fig. 2b
26 [AR-1-25] I added “in” before Fig. 2b.
27

1 [RC-1-26] P7L30–P8L1: remove “: this” and replace “could be” “, possibly”
2 [AR-1-26] I removed “: this” and replace “could be” “, possibly”.
3
4 [RC-1-27] P8L21: What's the purpose of only mentioning Pseudocalanus taxa as part of CopS_{arc} since
5 this group is also composed of other taxa?
6 [AR-1-27] Because Pseudocalanus taxa numerically dominate CopS_{arc}, we mentioned habitat of them.
7
8 [RC-1-28] P10L3: remove “invasion” before “northward” and replace “of” ”by”
9 [AR-1-28] I removed “invasion” before “northward” and replaced “of” ”by”.
10
11 [RC-1-29] P10L8: add “the” before “Arctic”
12 [AR-1-29] I added “the” before “Arctic”.
13
14 [RC-1-30] P10L8: I don't understand this sentence. What is a mean developmental stage?
15 [AR-1-30] It means that the stage of Copepodite I–V. I added the explanation of this term in revised
16 manuscript.
17 [RS-1-30] P10L20–21: There is a strong relationship between the mean developmental stage
18 (Copepodite stage I–V) of *C. glacialis* and surface temperature (Ershova et al., 2015).
19
20 [RC-1-31] P10L16: “leads to” instead of “cause”
21 [AR-1-31] I replaced “cause” “leads to”.
22
23 [RC-1-32] P10L17: replace “their” “an”
24 [AR-1-32] I replaced “their” “an”.
25
26 [RC-1-33] P10L18: replace “more” “a large supply of” and remove “their” before “high”
27 [AR-1-33] I replaced “more” “a large supply of” and removed “their” before “high”.
28

1 [RC-1-34] P10L18–19: Not sure I got this sentence right. But I suggest this change to “relatively
2 warm ice-free waters, or even cold when close to the melt period”.

3 [AR-1-34] Thank you for giving helpful suggestion. As you suggested, I rewrote the sentence.

4 [RS-1-34] P10L29-P11L2: For copepods, the spring bloom resulting from early sea ice retreat is an
5 important energy source, because a large supply of food can be utilized while maintaining high
6 activity in relatively warm ice-free waters or even cold, when close to the melt period. Thus, earlier
7 sea ice retreat should have positive effects on the growth and reproduction of copepods that do not rely
8 on sea ice production in the northern Bering and Chukchi Seas.

9

10 [RC-1-35] P10L20: “that do not rely on sea-ice production” instead of “without using sea-ice”

11 [AR-1-35] I replaced “without using sea-ice” “that do not rely on sea-ice production”.

12

13 [RC-1-36] P10L29: “cluster” instead of “clustere”

14 [AR-1-36] I replaced “clustere” “cluster”.

15

16 [RC-1-37] P11L4: remove “water with” before “cold/lower”, and add “water mass” after “IMW”

17 [AR-1-37] I removed “water with” and added “water mass” after “IMW”.

18

19 [RC-1-38] P11L6: add “is” after “masses” and use “correlated with” instead of “affects”

20 [AR-1-38] I added “is” after “masses” and replaced “affects” “correlated with”.

21

22 [RC-1-39] P11L7: add “, represented solely by *Calanus glacialis* in the study area.” after “(Fig. 5)”
23 and replace “*Calanus glacialis*, which represents CopLarc in this study,” “This species”

24 [AR-1-39] I added “, represented solely by *Calanus glacialis* in the study area.” after “(Fig. 5)” and
25 replaced “*Calanus glacialis*, which represents CopLarc in this study,” “This species”

26

27 [RC-1-40] P11L8: “Huntly” instead of “Huutly” and “Ashujian” instead of “Ashjian”, and “of” instead
28 of “on”

29 [RC-1-40] I modified them “Huntly”, “Ashjian” and “of”, respectively.

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2 [RC-1-41] P11L9: replace “is” “has been shown to be”, add “in the study area” before (Ershova et al.,
3 2015) and replace “reflected” “backs”

4 [AR-1-41] I replaced “is” “has been shown to be”, added “in the study area” before (Ershova et al.,
5 2015) and replaced “reflected” “backs”.

6

7 [RC-1-42] P11L19: “throughout” instead of “through”

8 [AR-1-42] I replaced “through” “throughout”.

9

10 [RC-1-43] P11L28: *P. mimus* and *P. newmani* are considered more Pacific whereas *P. acuspes* and *P.*
11 *minutus* would Arctic (Questel et al. 2016).

12

13 Questel, J.M., Blanco-Bercial, L., Hopcroft, R.R., Bucklin, A., 2016. Phylogeography and
14 connectivity of the *Pseudocalanus* (Copepoda: Calanoida) species complex in the eastern North
15 Pacific and the Pacific Arctic Region. J. Plankton Res.

16

17 It is probably worth discussing the effect of different origins for habitat preference of this taxonomic
18 complex.

19 [AR-1-43] Thank you for helpful comments and suggestion. According to Questel et al., (2016), *P.*
20 *mimus* and *P. newmani*, summarized into CopS_{arc} in our study, are considered more Pacific. The
21 criterion of identification on Arctic/Pacific species is producible/unproducible in Arctic, so that *P.*
22 *mimus* and *P. newmani* are identified as CopS_{arc}. Unfortunately, we didn’t analyze the genetic type of
23 copepods individually, so that we couldn’t identify the detail of their origins. However, as you pointed
24 out, the difference of origins might lead to the difference of response for environmental variables. So,
25 I would like to add the explanation about it. Please check [RS-1-43].

26 [RS-1-43] P12L8–21: In this study, CopS_{arc} were dominated by *Pseudocalanus*, including
27 *Pseudocalanus acuspes*, *P. mimus*, *P. minutus*, *P. newmani*, and undefined *Pseudocalanus* spp. (mean
28 72 % of CopS_{arc} abundance). *Pseudocalanus* occurs in the entire of Bering Sea shelf and in the Arctic
29 area (Frost, 1989). This distribution is thought to result from *Pseudocalanus* being initially abundant
30 in the warm water originating from the Bering Sea. According to Questel et al., (2016), *P. mimus* and
31 *P. newmani*, summarized into CopS_{arc} in our study, are considered more Pacific in origin.

1 Arctic/Pacific species are identified as such based on whether or not they are reproducible in Arctic
2 region; thus, *P. mimus* and *P. newmani* are identified as CopS_{arc}. Unfortunately, we did not analyze the
3 genetic type of copepods individually, so we could not determine their origins. However, *P. mimus*
4 and *P. newmani* might be transported to the Arctic by the Pacific inflow. Therefore CopS_{arc} are
5 significantly abundant in the warm-water masses such as ACW and BSW. The abundance of CopL_{arc}
6 could be associated with cold-water masses in which CopS_{arc} are less abundant.

7

8 [RC-1-44] P11L29: “Pseudocalanus” instead of “Puseudocalanus”

9 [AR-1-44] I replaced “Puseudocalanus” “Pseudocalanus”.

10

11 [RC-1-45] P12L17: “dominant” instead of “dominat”

12 [AR-1-45] I replaced “dominat” “dominant”.

13

14 [RC-1-46] P12L18: “is” instead of “was”

15 [AR-1-46] I replaced “was” “is”.

16

17 [RC-1-47] P12L29: replace “Although there is” “besides” and remove “copepods at”

18 [AR-1-47] I replaced “Although there is” “besides” and removed “copepods at”.

19

20 [RC-1-48] P12L30: replace “by using” “with”

21 [AR-1-48] I replaced “by using” “with”.

22

23 [RC-1-48] P13L1: But then if the match is before the sampling period, the zooplankton abundances
24 would have integrated the new recruitment. I think one important effect for the lack of correlation
25 between phyto and zooplankton values is the different temporal scales in population growth. A
26 relationship may have showed using the cumulative phytoplankton production from ice break-up to
27 sampling time, which is difficult to obtain.

28 [AR-1-48] Thank you for helpful comments and suggestion. As you indicated, the reason why there is
29 the lack of correlation between phyto and zooplankton values is the different temporal scales in
30 population growth. In addition, it is difficult to obtain the relationship between the cumulative

1 phytoplankton production from ice break-up to sampling time, as you pointed out, too. So, I rewrote
2 this section in line with your comments.

3 [RS-1-48] P13L24–L32: Consequently, we also expected a positive effect of the bottom chlorophyll *a*
4 concentration (Chl.*a*_{BOT}) on the abundance of all copepod groups. However, clear positive effects were
5 not observed (Fig. 5). In addition, another important explanation for the non-correlation between
6 phyto- and zooplankton values is the different temporal scales in population growth. A relationship
7 may have been shown using the cumulative phytoplankton production from the ice break-up to the
8 sampling time, which is difficult to obtain. Therefore, it is difficult to link the chlorophyll *a*
9 concentration to the copepod abundance using the time lag between the blooms of phytoplankton and
10 copepods.

11

12 [RC-1-49] P13L22: remove “quantitatively” after “abundance”

13 [AR-1-49] I removed “quantitatively” after “abundance”.

14

15 [RC-1-50] P13L25: “already” instead of “well”

16 [AR-1-50] I replaced “well” “already”.

17

18 [RC-1-51] P13L29: “increase” instead of “warming”

19 [AR-1-51] I replaced “warming” “increase”.

20

21 [RC-1-52] P13L29: replace “increase of fresh water content” “enhanced sea water freshening”

22 [AR-1-52] I replaced “increase of fresh water content” “enhanced sea water freshening”.

23

24 [RC-1-53] P20L4–5 Figure 1: remove “The color scale indicates bottom water depth (m).”

25 [AR-1-53] I removed “The color scale indicates bottom water depth (m).”.

26

27 [RC-1-54] P20L8 Figure 2: “indicates” instead of “indicated”

28 [AR-1-54] I replaced “indicated” “indicates”.

29

1 [RC-1-55] P20L8–13: remove “Water mass designations are Alaskan coastal water (ACW;
2 temperature 2.0–13.0 °C and salinity < 31.8), Bering Shelf Water (BSW; 0.0–10.0 °C and 31.8–32.5),
3 Anadyr Water (AW; -1.0–1.5 °C and 32.3–33.3), Bering Shelf Anadyr water (BSAW; BSW and AW
4 combined), ice melt water (IMW; < 2.0 °C and < 30.0) and dense water (DW; < -1 °C and 31.0–
5 33.0).”, because I Find a way to show the water masses on the graphs or describe in the Results section.

6 [AR-1-55] As you indicated, I removed the explanations of water masses.

7

8 [RC-1-56] P20L14 Figure 3: remove “The” before “distribution”.

9 [AR-1-56] I removed “The” before “distribution”.

10

11 [RC-1-57] P20L15 Figure 3: “indicates” instead of “indicated the”.

12 [AR-1-57] I replace “indicated the” “indicates”.

13

14 [RC-1-58] P20L1617 Figure 4: remove “The” before “distribution” and “Colored circles indicted the
15 abundance of copepods”

16 [AR-1-58] I removed “The” before “distribution” and “Colored circles indicted the abundance of
17 copepods”.

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1 Author's response to anonymous reviewer #3

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3 We really appreciate thoroughly giving comments and helpful suggestion to our study by anonymous
4 reviewer #3. And I am grateful for careful proofreading on our manuscript for publication in
5 Beogeosciences. We have answered all the comments made by the referees #3. RC, AR and RS stand
6 for Reviewers comment, Authors response and Revised sentence, respectively. We hope that our
7 explanations and revise plan are acceptable and satisfactory.

8

9 [RC-2-1] This paper deals with the distribution of copepods in the Arctic Ocean in relation to the
10 environmental variables. The analysis was based in an intensive sampling for 3 summers and the
11 phenomenon seemed to be well reproduced with the model.

12 What I feel discontent in this paper is found in the introduction, lacking the question or clear
13 hypothesis which should be revealed in the paper. Because of this defect, it is unclear to locate the new
14 scientific findings in this paper, which should be added our previous understanding in terms of Arctic
15 copepod ecology. As authors already mentioned in the introduction, it has been well known that the
16 distribution of copepods in this region is well corresponds to properties of water masses. Therefore I
17 require the authors to rewrite the introduction to emphasize the importance to construct the adequate
18 model to predict the copepod community structure in relation to the climate change so that the
19 significance of the paper will be highlighted.

20 [AR-2-1] We appreciate for giving us helpful comments to revise introduction. As you pointed out,
21 there is no question or clear hypothesis which should be revealed in the paper. The most importance of
22 constructing the adequate model is to evaluate the effects of environmental variables and adapted to
23 predict the risks faced by organisms, in future. Because Pacific, small and large Arctic copepods have
24 different life cycle and reproductive characteristics, the response for environmental change caused by
25 various phenomena such as climate change can be different. So, I rewrote the last paragraph as follows.

26 [RS-2-1] P3L24–P4L4: The distribution patterns of both Pacific and Arctic copepods in the Arctic
27 seas have been reported in these previous studies. However, recent and future drastic climate changes
28 potentially trigger the shifts in the distributions of copepod species or change of their habitat. This
29 phenomenon has already been reported for some species (e.g., Eisner et al., 2014; Ershova et al., 2015).
30 In order to comprehend the response of each copepod group to the environmental changes in the
31 Arctic, a statistical understanding of the relationship between environmental factors and the group's
32 abundance is required. Since Pacific and Arctic copepods have different life-cycles, suitable habitat,
33 and reproductive characteristics, their response to the environmental changes are expected to differ.

1 Therefore in the present study, we aim to construct an adequate model to illustrate the suitable
2 environmental characteristics for each Pacific and Arctic copepods group that will help us predict the
3 risks they might face in the future. Here, we propose the use of generalized additive models (GAMs)
4 to determine the factors affecting the spatial pattern of copepod abundances based on data collected by
5 net-sampling during the summers of 2007, 2008, and 2013.

6

7 [RC-2-2] Another minor comment is about the grouping of ‘cyclopoid copepods’. Although the
8 authors did not mentioned anything about the ‘cyclopoid copepods’ in the text, some species such as
9 *Oithona similis* or *Triconia* are known to be abundant and could be ubiquitous in any water mass even
10 if they use the large mesh size. In this light, the authors need to describe the genus of ‘cyclopoid
11 copepod’ clearly (it should not be so difficult) as well as the reason why they designated the
12 ‘cyclopoid copepod’ as the ‘small Arctic copepods’

13 [AR-2-2] Thank you for helpful comments. In our study, we identified copepods that they can
14 reproduce in Arctic as Arctic species. And, the definition of size (small or large) is not depending on
15 the actual body length of copepod, but the generation length and the number of times of reproduction.
16 Depend on these definitions, ‘cyclopoid copepod’ is identified as small Arctic species. However, we
17 are sorry that we didn’t identify the genus of ‘cyclopoid copepod’ clearly because the rate of their
18 abundance was very small and we cannot do quantitative evaluation on the abundance of them
19 sampled by coarse net. Therefore, we are sorry that we cannot describe the genus of ‘cyclopoid
20 copepod’. We add the reason why we designated the samples into three groups on revised manuscript.
21 Please check [RS2-2]

22 [RS-2-2] P4L16–P4L32: For the dominant taxa (calanoid copepods), identification was made at the
23 species level. In addition to calanoid copepods, cyclopoid copepods such as *Oithona similis* also
24 widely appear in this study area (Llinás et al., 2009). However, we summarized all species as
25 cyclopoid copepods, because we did not perform identifications at the species level. The species were
26 separated into Pacific and Arctic species based on their dominant reproducing grounds. The applied
27 definition of size (small or large) did not depend on the actual body length of the copepod specimen,
28 but on the generation length and the number of times of reproduction. Falk-Petersen et al. (2009) and
29 Dvoretzky and Dvoretzky (2009) list the copepod characteristic of distribution, generation length and
30 reproduction. The life-cycles of large Arctic copepods includes one or fewer generations per year,
31 whereas small Arctic copepods have multiple generations in the Arctic (e.g., Dvoretzky and Dvoretzky,
32 2009; Falk-Petersen et al., 2009). Following these two sources, we summarized the copepod species
33 into three groups (Table 2): large Arctic (CopL_{arc}: reproducible in the Arctic, and generation length is
34 greater than one year, and reproduction occurs once), small Arctic (CopS_{arc}: reproducible in Arctic,

1 generation length less than one year, and reproduction occurs multiple times a year), and Pacific
2 copepods (Cop_{pac} : not reproducible in the Arctic, generation length is greater than one year, and
3 reproduction occurs once).

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1 **Distribution of Arctic and Pacific copepods and their**
2 **habitat in the northern Bering and Chukchi Seas**

3

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15

16 **Abstract**

17 The advection of warm Pacific water and the reduction in sea ice in the western Arctic Ocean
18 may influence the abundance and distribution of copepods, a key component of food webs. To
19 quantify the factors affecting the abundance of copepods in the northern Bering and Chukchi
20 Seas, we constructed habitat models explaining the spatial patterns of large and small Arctic
21 and Pacific copepods, separately. Copepods were sampled using NORPAC nets. The
22 structures of water masses indexed by using principle component analysis scores, satellite-
23 derived timing of sea ice retreat, bottom depth, and chlorophyll *a* concentration were
24 integrated into generalized additive models as explanatory variables. The adequate models for
25 all copepods exhibited clear continuous relationships between the abundance of copepods and
26 the indexed water masses. Large Arctic copepods were abundant at stations where the bottom
27 layer was saline; however they were scarce at stations where warm fresh water formed the
28 upper layer. Small Arctic copepods were abundant at stations where the upper layer was
29 warm and saline and the bottom layer was cold and highly saline. In contrast, Pacific

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削除: copepod

1 copepods were abundant at stations where the Pacific-origin water mass was predominant (i.e.
2 a warm, saline upper layer and saline and a highly saline bottom layer). All copepod groups
3 showed a positive relationship with early sea ice retreat. Early sea ice retreat has been
4 reported to initiate spring blooms in open water, allowing copepods to utilize more food while
5 maintaining their high activity in warm water without sea ice and cold water. This finding
6 indicates that early sea ice retreat has positive effects on the abundance of all copepod groups
7 in the northern Bering and Chukchi Seas, suggesting a change from a pelagic–benthic-type
8 ecosystem to a pelagic–pelagic type.

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削除: cause

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削除: earlier

9

10 1 Introduction

11 Over the last decade, seasonal sea ice coverage has changed dramatically in the northern
12 Bering and Chukchi Seas (Comiso et al., 2008; Parkinson and Comiso, 2012), possibly
13 because of an increase in the inflow of Pacific water from the Bering Sea through the Bering
14 Strait (Shimada et al., 2006). The Bering Strait is shallow (<30 m) and has a gentle shelf
15 extending to the Arctic Shelf break through the Chukchi Sea. On this extensive shallow shelf,
16 the food webs are short and efficient, and even small changes in production pathways can
17 affect organisms at higher trophic levels (Grebmeier et al., 2006). The recent change in the
18 sea ice melt timing contributes to stratification, nutrient trapping at the surface, and lower
19 primary production with insufficient sunlight (Clement, 2004). In contrast, it has been
20 suggested that the timing of the phytoplankton bloom has also altered (Kahru et al., 2011) and
21 that its annual primary production has increased (Arrigo et al., 2008). Changes in the timing
22 and location of primary production and associated grazing by zooplankton have a direct
23 influence on the energy and matter transfer to the benthic community (Grebmeier et al., 2010).

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24 In the Bering and Chukchi Seas, several water masses have been identified based on
25 their basis of salinity and temperature (Table 1). The water masses include the relatively
26 warm/low-salinity Alaskan coastal water (ACW; temperature 2.0–13.0 °C and salinity <31.8)
27 that originates from the eastern Bering Sea; the warm/saline Bering shelf water (BSW; 0.0–
28 10.0 °C and 31.8–33.0) from the middle Bering shelf; and the cold/higher-salinity Anadyr
29 water (AW; –1.0–1.5 °C and 32.3–33.3) originating from the Gulf of Anadyr at depth along
30 the continental shelf of the Bering Sea. The BSW and AW merge to form the Bering Sea
31 Anadyr water (BSAW; Coachman et al., 1975; Springer et al., 1989). In addition, cold/lower-
32 salinity ice-melt water (IMW; <2.0 °C and <30.0) originates from sea ice, and colder/high-

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1 salinity dense water (DW; less than $-1.0\text{ }^{\circ}\text{C}$ and $32.0\text{--}33.0$) forms in the previous winter
2 during freezing of both the Bering and Chukchi Seas (Weingartner et al., 2013). These water
3 masses often show vertical consistency both geographically and seasonally (Iken et al., 2010;
4 Eisner et al., 2013; Weingartner et al., 2013).

5 In the northern Bering and Chukchi Seas, copepods are primary consumers of
6 phytoplankton and are the main prey of foraging fish (e.g., polar cod *Boreogadus saida*,
7 Nakano et al., 2015), seabirds (e.g., phalaropes, shearwaters and crested auklets *Aethia*
8 *crisatella*; Piatt and Springer, 2003; Hunt et al., 2013), and baleen whales (e.g., bowhead
9 whale *Balaena mysticetus*; Lowry et al., 2004). Therefore, copepods are a key components of
10 the Arctic marine food webs (Lowry et al., 2004). In this region, large Arctic copepods
11 (*Calanus glacialis*) and small Arctic copepods (e.g., *Acartia hudsonica*, *Centropages*
12 *abdominalis*, *Eurytemora herdmani* and *Pseudocalanus acuspes*) are abundant (Springer et al.,
13 1996). In addition, Pacific copepods (*C. marshallae*, *Eucalanus bungii*, *Metridia pacifica*,
14 *Neocalanus cristatus*, *N. flemingeri*, and *N. plumchrus*) are often transported from the Bering
15 Sea (Lane et al., 2008; Hopcroft et al., 2010). Copepod communities are associated with the
16 distribution of water masses (e.g., Springer et al., 1989; Hopcroft et al., 2010; Eisner et al.,
17 2013): *Pseudocalanus* species are abundant in the ACW and Pacific species are abundant in
18 the AW, as they are transported from the Bering Sea. Pacific copepod species (e.g., *E. bungii*)
19 expanded their distribution into the Chukchi Sea in 2007 (Matsuno et al., 2011). *C. glacialis*
20 is abundant in Arctic waters, and it is considered to be a native species to the Arctic shelves
21 (Canover and Huntley, 1991; Ashjian et al., 2003). Therefore, the distribution of copepod
22 communities in this region appears to be affected by both the inflow of Pacific water and the
23 water from sea ice melting.

24 The distribution patterns of both Pacific and Arctic copepods in the Arctic seas have
25 been reported in these previous studies. However, recent and future drastic climate changes
26 potentially trigger the shifts in the distributions of copepod species or change of their habitat.
27 This phenomenon has already been reported for some species (e.g., Eisner et al., 2014;
28 Ershova et al., 2015). In order to comprehend the response of each copepod group to the
29 environmental changes in the Arctic, a statistical understanding of the relationship
30 between environmental factors and the group's abundance is required. Since Pacific and
31 Arctic copepods have different life-cycles, suitable habitat, and reproductive characteristics,
32 their response to the environmental changes are expected to differ. Therefore in the present

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削除: The objective of this study was to determine the factors affecting the spatial pattern of copepod abundance based on the data collected by the NORPAC net sampling conducted by T/S *Oshoro-maru* in the summers of 2007, 2008 and 2013. We categorized copepods into three groups; large Arctic, small Arctic, and Pacific copepods. The life cycles of large Arctic copepods are one or fewer generation

1 study, we aim to construct an adequate model to illustrate the suitable environmental
2 characteristics for each Pacific and Arctic copepods group that will help us predict the risks
3 they might face in the future. Here, we propose the use of generalized additive models
4 (GAMs) to determine the factors affecting the spatial pattern of copepod abundances based on
5 data collected by net-sampling during the summers of 2007, 2008, and 2013.

6 2 Materials and methods

7 2.1 Field sampling

8 We sampled copepods and water onboard of T/S *Oshoro-maru* (Hokkaido University) during
9 30 July–24 August 2007 (31 stations), 30 June–13 July 2008 (26 stations), and 4–17 July
10 2013 (31 stations; Fig. 1). Zooplankton samples were collected during the day or at night
11 using vertical tows with a North Pacific Standard (NORPAC) net (mouth diameter 45 cm,
12 mesh size 335 μm) from 5 m above the bottom to the surface (the depths of most stations
13 were approximately 50 m). The volume of water filtered through the net was estimated using
14 a flow-meter mounted on the mouth of the net. Zooplankton samples were immediately
15 preserved with 5 % v/v borax-buffered formalin. In a laboratory on land, identification and
16 enumeration of taxa were performed on the zooplankton samples under a stereomicroscope.
17 For the dominant taxa (calanoid copepods), identification was made at the species level. In
18 addition to calanoid copepods, cyclopoid copepods such as *Oithona similis* also widely appear
19 in this study area (Llinás et al., 2009). However, we summarized all species as cyclopoid
20 copepods, because we did not perform identifications at the species level. The species were
21 separated into Pacific and Arctic species based on their dominant reproducing grounds. The
22 applied definition of size (small or large) did not depend on the actual body length of the
23 copepod specimen, but on the generation length and the number of times of reproduction.
24 Falk-Petersen et al. (2009) and Dvoretzky and Dvoretzky (2009) list the copepod
25 characteristic of distribution, generation length and reproduction. The life-cycles of large
26 Arctic copepods includes one or fewer generations per year, whereas small Arctic copepods
27 have multiple generations in the Arctic (e.g., Dvoretzky and Dvoretzky, 2009; Falk-Petersen
28 et al., 2009). Following these two sources, we summarized the copepod species into three
29 groups (Table 2): large Arctic (CopL_{arc}: reproducible in the Arctic, and generation length is
30 greater than one year, and reproduction occurs once), small Arctic (CopS_{arc}: reproducible in
31 Arctic, generation length less than one year, and reproduction occurs multiple times a year).

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下へ移動 [2]: per year, whereas small Arctic copepods have multiple generations in the Arctic (e.g., Dvoretzky and Dvoretzky, 2009; Falk-Petersen et al., 2009).

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1 and Pacific copepods (Cop_{pac} : not reproducible in the Arctic, generation length js greater than
2 one year, and reproduction occurs once).
3 _____ At the zooplankton sampling stations, vertical profiles of temperature and salinity
4 were made using conductivity-temperature-depth (CTD: Sea-Bird Electronics Inc., SBE 911
5 Plus) casts. Water samples for chlorophyll *a* were obtained with Niskin bottles on the CTD
6 rosette from the bottom (21–56 m) to the surface. Water samples were gently filtered (<100
7 mmHg) onto GF/F filters. Phytoplankton pigments on the filters were extracted with *N,N*-
8 dimethylformamide (Suzuki and Ishimaru, 1990), and chlorophyll *a* concentrations were
9 determined by the fluorometric method using a Turner Designs 10-AU fluorometer
10 (Welschmeyer, 1994). In order to investigate the relationships between the abundance of
11 copepods and the sea ice condition, we used SSM/I Daily Polar Gridded Sea Ice
12 Concentration (SIC) data obtained from the National Snow and Ice Data Center
13 (<http://nsidc.org/>).

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14 2.2 Data analysis

15 The relationship between the abundance of copepods and traditionally defined water masses
16 has been reported (Hopcroft and Kosobokova, 2010; Eisner et al., 2013). In these studies, the
17 surface and bottom water masses were identified based on the basis of temperature and
18 salinity. However, the quantitative evaluation of the effects of complex water properties on
19 the copepod abundance is difficult. In order to quantify the factors affecting the spatial pattern
20 of abundance of each copepod group using GAMS (See Section 2.3), explanatory variables
21 that are correlated with other variables must be removed to avoid the problem of
22 multicollinearity. This procedure may hinder the recovery of important oceanographic
23 features such as the combination of water masses in the upper and bottom layers, because
24 water temperature and salinity in both layers are often strongly correlated. In this study, to
25 delineate the combination of water masses in the upper and bottom layers, we summarized the
26 water-mass properties in these layers as scores using principal component analysis (PCA).
27 These scores can be used as continuous explanatory variables in GAMS.

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28 As the vertical structure of the water mass in our focused region basically forms a one-
29 or two-layered structure because of the shallow bathymetry, we can divide the water column
30 into a maximum of two layers (i.e., the layers above and below the pycnocline are defined as
31 the upper and bottom layers, respectively). The density (ρ) was calculated from the

1 | temperature and the salinity measured by CTD profiles with a vertical data resolution of 1 m.
2 | We calculated the vertical density gradient ($\frac{d\rho}{dD}$) at a specific depth using 2 m-mean densities
3 | immediately above and below the specific depth. $\frac{d\rho}{dD}$ was calculated for all depths except for
4 | the two uppermost and the two lowermost depth levels. The depth of the maximum density
5 | gradient ($\frac{d\rho}{dD_{max}}$) was defined as the pycnocline of each sampled site. Then environmental
6 | variables (temperature, salinity, and log-transformed chlorophyll *a*) were vertically averaged
7 | within the upper and bottom layers and defined as T_{UPP}, T_{BOT}, S_{UPP}, S_{BOT}, Chl.*a*_{UPP} and
8 | Chl.*a*_{BOT}, respectively (see Table 3 and Figures A1–A4 in Supplementary Materials). PCA
9 | was applied to determine the water-mass structure using $\frac{d\rho}{dD_{max}}$, T_{UPP}, T_{BOT}, S_{UPP} and S_{BOT} at
10 | all 88 stations. As the principal water masses in the Bering and Chukchi Seas are
11 | characterized by the temperature and salinity of the water column (Coachman et al., 1975),
12 | Chl.*a*_{UPP}, Chl.*a*_{BOT} and SIC were not used in the PCA to determine the water-mass structure.
13 | These five parameters ($\frac{d\rho}{dD_{max}}$, T_{UPP}, T_{BOT}, S_{UPP} and S_{BOT}) were standardized prior to the PCA
14 | to reduce the biases between the units of the variables. Several principal components and their
15 | factor loadings (correlations of factors to the derived principal components) were presented.
16 | The PCA scores were used as covariates of the water-mass structures in the habitat models. In
17 | addition, we used the anomaly of timing of sea ice retreat (aTSR) at each sampling station as
18 | an index of sea ice condition. The values of aTSR were calculated using satellite-derived sea
19 | ice images for 1991–2013. Although sea ice concentration images had been projected using
20 | polar stereographic coordinates with 25km spatial resolution, we interpolated them using the
21 | nearest-neighbor method and resampled them into 9km spatial resolution. Considering the
22 | missing values and land contamination, we defined SIC <50 % as non-ice-covered pixels, and
23 | aTSR was defined as the anomalous last date when the SIC fell below 50 % prior to the date
24 | of the annual sea ice minimum in the Arctic Ocean.

25 | 2.3 Statistical analysis

26 | Before producing the habitat models, we examined the multicollinearity between the
27 | explanatory variables by correlation analysis. To examine the relationships between the
28 | copepod abundance (CopL_{arc}, CopS_{arc}, and Cop_{pac}) and the environmental variables, we
29 | constructed habitat models using GAMs. GAMs are a non-parametric extension of

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1 | generalized linear models (GLMs) such as multiple-regression models (Eq. (1)), with the only
2 underlying assumption that the functions are additive and that the components are smooth (Eq.
3 (2)). The basic concept is the replacement of the parametric GLM structure:

$$4 \quad g(\mu) = \alpha + \beta_1 x_1 + \beta_2 x_2 + \beta_3 x_3 + \dots + \beta_i x_i \quad (1)$$

5 with the additive smoothing function structure:

$$6 \quad g(\mu) = \varepsilon + s_1(x_1) + s_2(x_2) + s_3(x_3) + \dots + s_i(x_i) \quad (2)$$

7 where α and ε are the intercepts and β_i and s_i are the coefficients and the smooth functions of
8 the covariates, respectively (Wood, 2006). To select the most adequate model in our approach,
9 we used Akaike's Information Criterion. Model validation was applied to the optimal models
10 to verify our assumptions and reproducibility of the results. Specifically, we plotted the
11 original values versus the fitted values and judged the adequacy of our optimal models based
12 on R^2 . The deviance explained (Eq. (3)) indicates the percentage of the variance that can be
13 explained by the most adequate model, and it is calculated as follows:

$$14 \quad \text{Deviance explained (\%)} = (1 - \text{Residual Deviance/Null Deviance}) \times 100 \quad (3)$$

15 where the residual deviance denotes the deviance produced by the model that includes
16 explanatory variables and the null deviance is the deviance produced by the model without
17 explanatory variables. All statistical analyses were undertaken using R (version 2.15.0
18 <http://www.r-project.org>).

19

20 **3 Results**

21 **3.1 Principal component analysis and water mass**

22 The first principal component (PC1) explained 47.1 % of the total variability. In the PC1
23 score, the loading coefficient was positive for $\frac{d\rho}{dT}_{\max}$, indicating that the magnitude of
24 stratification increased with an increase in PC1. In contrast, PC1 was strongly negative for
25 T_{UPP} and T_{BOT} , indicating that lower temperatures in the whole water mass resulted in smaller
26 PC1 (Table 4). Additionally, PC1 was negative for S_{UPP} , indicating a low-salinity water mass
27 in the surface layer with higher PC1, but weakly positive for S_{BOT} . According to Fig. 2a,
28 which shows the T-S diagram colored according to the PC1 score, a higher PC1 value (>1)

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1 value indicated a combination of the cold/lower salinity IMW₂ in the upper layer₂ and the
2 colder/high-salinity DW₂ in the bottom layer. In contrast, a low PC1 value₂ denoted a warm
3 water mass in both layers and/or low-salinity surface water (Table 4). From Fig. 2a, a lower
4 PC1 value (<-1.5) indicated a combination of warmer/low-salinity ACW₂ in the upper layer₂,
5 and warm/saline BSW or cold/higher-salinity AW or BSAW₂ in the bottom layer. A low-
6 medium PC1 score (-1.5-0.5) indicated a combined water mass with both BSW and
7 AW/BSAW (Fig. 2a). PC1 was higher at the stations north of 69°N as compared to ones to the
8 south in 2008 and 2013 and low for all stations in 2007 (Fig. 3), suggesting that the
9 combination of IMW and DW was dominant in the northern stations in 2008 and 2013, and
10 ACW was dominant at almost all stations in 2007.

11 The second principal component (PC2) explained 34.8 % of the total variability. In the
12 PC2 score, the loading coefficient was negative for $\frac{dp}{dD_{max}}$ and temperature and positive for
13 salinity in both the upper and bottom layers (Table 4). These results indicated that there is
14 highly saline water in both layers that tended to decrease the magnitude of stratification and
15 form a single layered structure with higher PC2. As illustrated in Fig. 2b, medium-high PC2
16 values (>0.5) indicated waters with a single-layered structure, BSW, AW₂ or BSAW. Low-
17 medium PC2 value (<0.5) denoted waters with a two-layered structure, with warmer-
18 temperature and lower-salinity water in the upper layer compared to the bottom layer,
19 possibly IMW in the upper layer and DW in the bottom layer, or ACW in the upper layer and
20 BSW/AW/BSAW in the bottom layer. PC2 was high at stations <69°N in all years and low at
21 stations east of the survey area in 2007 (Fig. 4), implying that a single-layered structure with
22 BSW/AW/BSAW was dominant in the Bering Strait. However, a combination of ACW with
23 BSW/AW/BSAW was observed northeast of the survey area in 2007.

24 The third principal component (PC3) explained 14.2 % of the total variability. The
25 PC3 score was correlated positively with all physical variables (Table 4), especially with T_{UPP}
26 and S_{BOT}. According to the T-S diagram colored according to the PC3 values (Fig. 2c),
27 relatively high PC3 values (>0.5) with relatively warm T_{UPP} (>4.0°C) and/or high S_{BOT}
28 (>32.0) suggested that the water columns were composed of ACW in the upper layer and/or
29 high-salinity BSW/AW at the bottom. PC3 was higher in 2007 than in 2008 and 2013,
30 particularly at the stations in the north of the Bering Strait (Fig. 3), indicating that relatively
31 warm BSW/ACW made up the upper layer and/or higher salinity AW/ BSAW/DW the
32 bottom layer.

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1 3.2 Copepod abundance

2 The **recorded** abundance of copepods at each station ranged between 150 and 146,323 inds.
3 m⁻² (median: 14,488). CopL_{arc} included only *Calanus glacialis* (Table 2), which represented
4 0.00 %–48.2 % of the total abundance and was found over almost **the entire** study area.
5 CopL_{arc} were more abundant in 2013 than in 2007 and 2008 (Fig. 4). CopS_{arc} made up
6 1.47 %–55.6 % **of the total copepod abundance** at each station and included *Pseudocalanus*
7 spp, *P. minutus*, *P. mimus*, *P. newmani*, and *P. acuspes* (Table 2). CopS_{arc} were dominant
8 throughout the study area in all study seasons (Fig. 4). Cop_{pac} included *C. marshallae*, *N.*
9 *cristatus*, *N. flemingeri*, *N. plumchrus*, *E. bungii*, and *M. pacifica*. Cop_{pac} were more abundant
10 in the south (<69°N) than in the north during all studied time intervals (Fig. 4).

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11 3.3 Copepod habitats

12 We constructed habitat models using aTSR, the quantitative index of the water masses (PC1,
13 PC2, and PC3), bottom depth (Bdepth), and averaged log-transformed chlorophyll *a* in the
14 upper layer (Chl.a_{UPP}) and **in the** bottom layer (Chl.a_{BOT}) as potential explanatory variables.
15 Averaged physical factors in the upper layer and bottom layers were excluded from potential
16 explanatory variables, as these were already included in the quantitative index of the water
17 masses.

18 The **model most adequately** explaining the abundance of CopL_{arc} included all
19 explanatory variables (Table 5). CopL_{arc} were abundant at stations with lower aTSR (<0 days)
20 and with deeper Bdepth, especially in the **areas** with bottom depths greater than 45 m (Fig. 5).
21 CopL_{arc} appeared to be abundant at stations with medium–higher PC1 (**≥** -0.5), low–high PC2
22 (-1 to 1), and low–medium PC3 (-1 to 0). The abundance of CopL_{arc} was relatively high in
23 **waters** with low (less than -0.5) and high (0.2–0.5) Chl.a_{UPP}. **However**, the effects of Chl.a_{UPP}
24 and Chl.a_{BOT} on CopL_{arc} were not clear.

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25 The **model which explains** the abundance of CopS_{arc} **most adequately**, included all
26 explanatory variables except PC2 (Table 5). CopS_{arc} were abundant at stations with lower
27 aTSR (< 5days) and with deeper Bdepth, especially in the **areas where** the sea depth was
28 greater than 40 m (Fig. 5). The abundance of CopS_{arc} was high for low–high PC1 (between
29 -1.5 and 2) and medium PC3 (0–1.2), and **for** medium–high Chl.a_{UPP} (>0; Fig. 5). The effect
30 of Chl.a_{BOT} was unclear.

1 | The abundance of Cop_{pac} was most adequately explained by the model with all
2 | explanatory variables except Chl.a_{UPP} (Table 5). Cop_{pac} were abundant at stations with low
3 | aTSR (<0 days), deeper Bdepth with a clear positive effect in waters deeper than 35 m, low-
4 | medium PC1 (-2 to 0.5) and PC3 (-0.5 to 1) and PC2 (<-0.5); it is less abundant at stations
5 | with medium-high PC2 (>-0.5) and high PC1 (>0.5; Fig. 5). The abundance of Cop_{pac} was
6 | high in the waters with low (<-0.2) and high (>0.5) Chl.a_{BOT}; however, the effect of Chl.a_{BOT}
7 | on Cop_{pac} was not clear.

9 | 4 Discussion

10 | 4.1 Effect of sea ice on copepod abundance

11 | The models most adequate to explain the abundance of copepods included aTSR as an
12 | explanatory variable (Table 5). As shown in the GAM plot, earlier sea ice retreat had positive
13 | effects on the abundance of all copepod groups (Fig. 5); in particular, the effect of early sea
14 | ice retreat was more obvious for Cop_{arc} than for the other two groups. The Cop_{pac} typified by
15 | *C. marshallae* and *N. cristatus*, are often transported from the Bering Sea through the Bering
16 | Strait (Lane et al., 2008; Hopcroft et al., 2010; Matsuno et al., 2011). Sea ice reduction is
17 | strongly related to an increase in the inflow of Pacific water from the Bering Sea through the
18 | Bering Strait (Shimada et al., 2006). Increasing water-mass transportation into the Chukchi
19 | Sea (Woodgate et al., 2012) and sea ice retreat enhances the northward invasion by larger
20 | Pacific water species. Our results reflect that future increases in advection from the Bering
21 | Sea will carry more Pacific zooplankton through the Bering Strait with even further
22 | penetration into the Arctic.

23 | Temperature and food are important for the growth of Cop_{Larc} and Cop_{Sarc} that
24 | reproduce in the Arctic. There is a strong relationship between the mean developmental stage
25 | (Copepodite stage I-V) of *C. glacialis* and surface temperature (Ershova et al., 2015). Early
26 | sea ice retreat leads to a longer ice-free period and warmer surface temperature. In our study,
27 | aTSR is negatively correlated with T_{UPP} and T_{BOT} ($\rho = -0.59$ and -0.69 , respectively;
28 | Spearman's correlation test $p < 0.001$), i.e., the sampling stations with early sea ice retreat
29 | have relatively high temperature and favorable conditions for copepod growth. The spring
30 | bloom inevitably forms at the ice edge and its timing is controlled by the timing of the sea ice
31 | retreat in the northern Bering Sea (Brown and Arrigo, 2013). In the shelf regions of the

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1 Bering and Chukchi Seas, early sea ice retreat leads to spring blooms in open water (Fujiwara
2 et al., 2016). For copepods, the spring bloom resulting from early sea ice retreat is an
3 important energy source, because a large supply of food can be utilized while maintaining
4 high activity in relatively warm ice-free waters or even cold, when close to the melt period.
5 Thus, earlier sea ice retreat should have positive effects on the growth and reproduction of
6 copepods that do not rely on sea ice production in the northern Bering and Chukchi Seas.

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7 4.2 Effects of water mass on copepod abundance

8 The abundance of all copepods was variably related to the combination of water masses in the
9 northern Bering and Chukchi Seas. In these seas, it has been well documented that the
10 community structure and abundance of zooplankton species differ in the different water
11 masses (e.g., Lane et al., 2008; Hopcroft et al., 2010; Matsuno et al., 2011), including the six
12 major water masses: ACW, IMW, DW, BSW, AW, and BSAW (e.g., Coachman et al., 1975;
13 Springer et al., 1989). These water masses and their combinations have mostly been described
14 by cluster analysis using temperature and salinity (e.g., Norcross et al., 2010; Eisner et al.,
15 2013; Ershova et al., 2015). In the present study, we quantitatively characterized these water
16 masses using PCA incorporating the combined water masses, the number of layers (single- or
17 double-layered masses), and the occurrence of high-salinity water in the bottom layer and/or
18 warm water in the upper layer (Fig. 2).

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19 CopL_{arc} were relatively abundant in the northern part of the Chukchi Sea (>69°N),
20 which is dominated by the cold/lower-salinity IMW water mass in the upper layer and the
21 colder/high-salinity DW in the bottom layer ($PC1 > 1$, $-1 < PC2 < -0.8$, and $-1 < PC3 < 0$;
22 Figs. 3, 4). This combination of water masses is positively correlated with the abundance of
23 CopL_{arc} (Fig. 5), represented solely by *Calanus glacialis*, in the study area. This species is
24 considered to be native to Arctic shelves (Conover and Huntley, 1991; Ashjian et al. 2003).
25 The Arctic population of *C. glacialis* appears in winter water in the study area (Ershova et al.,
26 2015). Our results back these CopL_{arc} habitats. Previous findings have reported that *C.*
27 *glacialis* were also abundant in water masses with ACW in the upper layer and BSAW in the
28 bottom layer (Eisner et al., 2013). In the present study, CopL_{arc} were relatively abundant in
29 the Bering Strait, in areas dominated by cold/high to higher-salinity BSAW and AW in both
30 layers ($-1.5 < PC1 < 1$, $-0.8 < PC2 < 1.2$, and $PC3 < -1$) in 2013. However, CopL_{arc} in this
31 study are less abundant in the water off Point Hope (southern part of the Chukchi Sea); this

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1 area was characterized by ACW in the upper layer and BSAW in the bottom layer ($-2.5 <$
2 $PC1 < -1.5$ and $PC3 > 0$; Fig. 5) during the summer of 2007. Our results slightly contradict
3 those of the above previous study; however, the presence of BSAW/AW is important for
4 CopL_{arc}.

5 In contrast to CopL_{arc}, CopS_{arc} were common in the entire study area. This copepod
6 group was abundant in waters with medium PC1 and PC3, indicating that these taxa were
7 distributed in waters with a wide range of temperature and salinity, i.e., warm/saline BSW.
8 However, CopS_{arc} were less abundant in waters with higher PC1, i.e., colder/low-salinity
9 IMW in the upper layer and cold/high-salinity DW in the bottom layer. These support the
10 previous findings that small Arctic copepods (e.g., *Pseudocalanus* spp., *A. hudsonica* and *A.*
11 *longiremis*) were abundant in warm BSW and relatively warm ACW in the upper and/or
12 bottom layers (Eisner et al., 2013; Ershova et al., 2015). In this study, CopS_{arc} were dominated
13 by *Pseudocalanus*, including *Pseudocalanus acuspes*, *P. mimus*, *P. minutus*, *P. newmani*, and
14 undefined *Pseudocalanus* spp. (mean 72 % of CopS_{arc} abundance). *Pseudocalanus* occurs in
15 the entire of Bering Sea shelf and in the Arctic area (Frost, 1989). This distribution is thought
16 to result from *Pseudocalanus* being initially abundant in the warm water originating from the
17 Bering Sea. According to Questel et al., (2016), *P. mimus* and *P. newmani*, summarized into
18 CopS_{arc} in our study, are considered more Pacific in origin. Arctic/Pacific species are
19 identified as such based on whether or not they are reproducible in Arctic region; thus, *P.*
20 *mimus* and *P. newmani* are identified as CopS_{arc}. Unfortunately, we did not analyze the
21 genetic type of copepods individually, so we could not determine their origins. However, *P.*
22 *mimus* and *P. newmani* might be transported to the Arctic by the Pacific inflow. Therefore
23 CopS_{arc} are significantly abundant in the warm-water masses such as ACW and BSW. The
24 abundance of CopL_{arc} could be associated with cold-water masses in which CopS_{arc} are less
25 abundant.

26 Pacific zooplankton are advected into the western Arctic Ocean through the Bering
27 Strait (Springer et al., 1989). Previous studies demonstrated that Pacific zooplankton
28 communities occurred in high-salinity water (BSW/AW) in the northern Bering and Chukchi
29 Seas (Springer et al., 1989; Lane et al., 2008; Hopcroft et al., 2010; Matsuno et al., 2011;
30 Eisner et al., 2013). In this study, Pacific copepods (Cop_{pac}) were abundant in the Bering
31 Strait and the Chukchi Sea south of Point Hope, areas which have low-medium PC1 and PC2,
32 associated with warmer/low-salinity ACW in the upper layer and cold/higher-salinity AW and

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1 warm/saline BSW or BSAW in the bottom layer, or single-layered AW, BSW, and BSAW.
2 These results support the previous observations. Our study further confirms the effects of the
3 interannual water-mass variability on copepod abundance. During the summer of 2007,
4 Pacific water masses (ACW, BSW and BSAW) extended to the north of 69°N (Fig. 3) and
5 transported Cop_{pac} into the Chukchi Sea (Matsuno et al., 2011). In contrast, in the summers of
6 2008 and 2013, when IMW and colder/high-salinity DW were dominant, few Cop_{pac} were
7 collected in the northern part of the Chukchi Sea (Fig. 4).

8 The combinations and distributions of water masses are known to be affected by the
9 Pacific inflow (Weingartner et al., 2005) and related to the sea ice retreat (Coachman et al.,
10 1975; Day et al., 2010). The inflow of warmer Pacific ACW was dominant in 2007
11 (Woodgate et al., 2010), and this strong inflow is believed to have triggered the sea ice retreat
12 in the western Arctic Ocean (Woodgate et al., 2012). Thus, the variability of the water masses
13 and their combinations as illustrated by PCA were in good agreement with the conventional
14 description of the dynamics of water masses. Our index can be used for the quantitative
15 evaluation of the effects of water-mass combinations with multiple components of water
16 properties and so may be useful for predicting copepod distributions with climate changes.

17 4.3 Effects of phytoplankton and bottom depth

18 The species categorized as Cop_{Sarc} (e.g., *Pseudocalanus* spp.) graze phytoplankton and
19 reproduce in the surface layer during day and night in the summer (Norrbín et al., 1996;
20 Plourde et al., 2002; Harvey et al., 2009). We therefore expected positive effects of Chl.*a*_{Upp}
21 on the Cop_{Sarc} abundance. However, the models did not yield obvious relationships between
22 the abundance of any copepods and Chl.*a*_{Upp}. Besides, there is possibility that young
23 copepodite stages could not be sampled with a coarse net (> 300 μm) such as the NORPAC
24 net used for our sampling. Moreover, another plausible explanation is that the sampling
25 period (June–August) did not coincide with the high-grazing and reproduction season when
26 copepods require a large amount of food intake. Cop_{Larc} reproduce during the spring
27 phytoplankton bloom (e.g., Falk-Petersen et al., 2009); thus our sampling period was not the
28 time of their reproduction. Phytoplankton cells sinking to the bottom water layers are
29 important food for copepods (Sameoto et al., 1986). Consequently, we also expected a
30 positive effect of the bottom chlorophyll *a* concentration (Chl.*a*_{Bot}) on the abundance of all
31 copepod groups. However, clear positive effects were not observed (Fig. 5). In addition,

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1 | another important explanation for the non-correlation between phyto- and zooplankton values
2 | is the different temporal scales in population growth. A relationship may have been shown
3 | using the cumulative phytoplankton production from the ice break-up to the sampling time,
4 | which is difficult to obtain. Therefore, it is difficult to link the chlorophyll *a* concentration to
5 | the copepod abundance using the time lag between the blooms of phytoplankton and
6 | copepods.

7 | A few previous studies have reported associations between the copepod abundance
8 | and the bottom depth of the shelf in the northern Bering and Chukchi Seas (e.g., Ashjian et al.,
9 | 2003). The reason for copepod groups being less abundant in waters shallower than 32 m
10 | bottom depth was unclear. In this survey, because the shallower area is correlated with the
11 | longitude ($\rho = -0.73$; Spearman's rank correlation test of longitude (°E) vs. Bdepth, $p <$
12 | 0.001), the result indicates that copepods are less abundant near the land. As shown in Figure
13 | 5, the smallest number of copepods was recorded at sampling stations of 25 m Bdepth. Except
14 | for these two stations, Cop_{L-arc} are not obviously related to Bdepth, whereas Cop_{pac} and
15 | Cop_{S-arc} gradually increase with depth.

16 | The associations between environmental factors and the abundance of copepods have
17 | been well documented (e.g., Springer et al., 1989; Lane et al., 2008; Matsuno et al., 2011).
18 | Recently these relationships were analyzed using clustered water masses (Eisner et al., 2013;
19 | Ershova et al., 2015). In the present study, we indexed the water masses and then
20 | quantitatively modeled the relationships between the water-mass characteristics and the
21 | spatial patterns of copepod abundance. Our evaluation of the effect of changes in the timing
22 | of sea ice retreat on copepod abundance confirms that suitable environments for copepods are
23 | formed by early sea ice retreat. The influence of the changes in sea ice on the Arctic
24 | ecosystem has been already documented; however, to the best of our knowledge, this is the
25 | first quantitative study to describe the relationships between the early sea ice retreat and
26 | copepod abundance. Quantitative analyses using the habitat models are useful for
27 | understanding various phenomena and risks faced by organisms (e.g., sea ice loss,
28 | temperature increase, and enhanced sea water freshening). Furthermore, this type of analysis
29 | can be adapted to predict ecosystem changes in the future by incorporating climate and
30 | predicted environmental data, and can also be used to understand the responses of organisms
31 | to environmental change in the northern Bering and Chukchi Seas.

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1 **Author contributions**

2 T.K. designed and coordinated this research project. K.M. and A.Y. collected the zooplankton
3 samples, performed species identification and enumeration of the zooplankton samples in the
4 land laboratory. A.F. operated and calculated sea-ice concentration data. H.U. and M.O.
5 calculated the stratification index by using CTD profiles. H.S. and Y.W. wrote the manuscript
6 with contributions from all co-authors.

7

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1 **Figure captions**

2 **Figure 1.** Study area and sampling stations in the northern Bering and Chukchi Seas during
3 the summers of 2007, 2008 and 2013. The symbols denote the sampling stations
4 where NORPAC net and CTD water samplings were conducted. Modified from
5 figure presented in Spall et al. (2014) and Grebmeier et al. (2015).

6 **Figure 2.** T-S diagrams of principal component scores (a) PC1, (b) PC2 and PC3 (c). Colored
7 circle indicated the magnitude of each PC.

8 **Figure 3.** Distribution of main principal component score (PC1–3) in 2007, 2008 and 2013.
9 Colored circles indicates magnitude of PC.

10 **Figure 4.** Distribution of copepods abundance in 2007, 2008 and 2013, large Arctic (CopL_{arc}),
11 small Arctic (CopS_{arc}) and Pacific (Cop_{pac}) copepods.

12 **Figure 5.** GAM plot of the best model in each copepod groups: large Arctic (CopL_{arc}), small
13 Arctic (CopS_{arc}) and Pacific (Cop_{pac}) copepods. The horizontal axes show the
14 explanatory variable: the anomaly of the timing of sea-ice retreat (aTSR), principal
15 component score (PC1–3) averaged log-transformed chlorophyll *a* concentration
16 within the layer above and below pycnocline, (Chl *a*_{UPP} and Chl *a*_{BOT}) and bottom
17 depth (Bdepth). Shade area represents 95% confidence intervals. The vertical axes
18 indicate the estimate smoother for the abundance of copepods. The estimated
19 smoother converts the explanatory variable to fit the models, so it shows positive
20 effects for response variables and the magnitude of its effects when estimated
21 smoother is positive, and vice versa. Short vertical lines located on the *x* axes of
22 each plot indicate the values at which observations were made.

23 **Supplementary materials**

24 **Figure A1.** Maximum density gradient (10⁻³ kg m⁻¹) at each sampling station.

25 **Figure A2.** Horizontal distributions of temperature (°C) averaged within the upper (T_{UPP}, top
26 panels) and the bottom (T_{BOT}, bottom panels) layers at each sampling station in
27 2007 (left panels), 2008 (middle panels) and 2013 (right panels).

28 **Figure A3.** Same as figure A2 but for salinity (S_{UPP} and S_{BOT}).

29 **Figure A4.** Same as figure A2 but for Chlorophyll-*a* concentration (Chl*a*_{UPP} and Chl*a*_{BOT}).

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削除: The color scale indicates bottom water depth (m).

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削除: Water mass designations are Alaskan coastal water (ACW; temperature 2.0–13.0 °C and salinity < 31.8), Bering Shelf Water (BSW; 0.0–10.0 °C and 31.8–32.5), Anadyr Water (AW; -1.0–1.5 °C and 32.3–33.3), Bering Shelf Anadyr water (BSAW; BSW and AW combined), ice melt water (IMW; < 2.0 °C and < 30.0) and dense water (DW; < -1 °C and 31.0–33.0).

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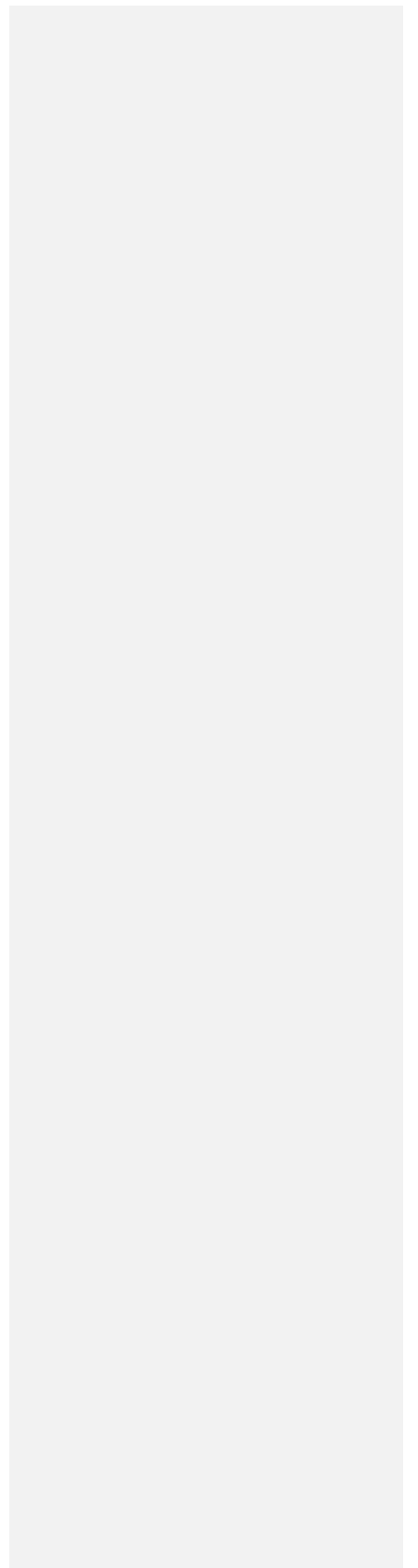
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削除: Colored circles indicted the abundance of copepods:



1 **Table 1.** Water mass properties in the northern Bering and Chukchi Seas.

Water mass	Temperature	Salinity	Reference
Alaskan coastal water (ACW)	relatively warm (2.0–13.0 °C)	low (< 31.8)	Coachman et al. (1975)
Bering Shelf Water (BSW)	warm (0.0–10.0 °C)	saline (31.8–32.5)	Coachman et al. (1987) Grebmeier et al. (1988) Springer et al. (1989)
Anadyr water (AW)	cold (-1.0–1.5 °C)	high (32.5–33.3)	Coachman et al. (1987) Grebmeier et al. (1988) Springer et al. (1989)
Bering Shelf Anadyr water (BSAW)	cold (-1.0–2.0 °C)	high (31.8–33.0)	Grebmeier et al. (1989) Eisner et al. (2013)
ice melt water (IMW)	cold (< 2.0 °C)	low (< 30.0)	Weingartner et al. (2005)
dense water (DW)	cold (< -1.0 °C)	high (32.0–33.0)	Coachman et al. (1975) Feder et al. (1994)

2

1 **Table 2.** The copepods species included in each copepod groups: large Arctic (CopL_{arc}), small
 2 Arctic (CopS_{arc}) and Pacific (Cop_{pac}) copepods.

Response Variables	Description	Species
CopL _{arc}	large Arctic copepods	<i>Calanus glacialis</i>
CopS _{arc}	small Arctic copepods	<i>Acartia hudsonica</i>
		<i>Acartia longiremis</i>
		<i>Acartia tumida</i>
		<i>Centropages abdominalis</i>
		<i>Eurytemora herdmani</i>
		<i>Epilabidocera amphitrites</i>
		<i>Microcalanus pygmaeus</i>
		<i>Pseudocalanus acuspes</i>
		<i>Pseudocalanus mimus</i>
		<i>Pseudocalanus minutus</i>
		<i>Pseudocalanus newmani</i>
		<i>Pseudocalanus</i> spp.
		<i>Scolecithricella minor</i>
		<i>Tortanus discaudatus</i>
		Cyclopoid copepods
Cop _{pac}	Pacific copepods	<i>Calanus marshallae</i>
		<i>Eucalanus bungii</i>
		<i>Metridia pacifica</i>
		<i>Neocalanus cristatus</i>
		<i>Neocalanus flemingeri</i>
		<i>Neocalanus plumchrus</i>

3

1 **Table 3.** The covariates for principal component analysis and explanatory variables for
 2 Generalize Additive Models (GAMs).

Explanatory variables in GAMs	Environmental Variables	Description	Unit
The principal components (PC1, PC2 and PC3)	$\frac{d\rho}{dD_{\max}}$	Magnitude of the maximum potential density gradient	10^{-3} g m^{-1}
	T_{UPP}	Vertical averaged temperature above the depth of the maximum potential density gradient	$^{\circ}\text{C}$
	T_{BOT}	Vertical averaged temperature under the depth of the maximum potential density gradient	$^{\circ}\text{C}$
	S_{UPP}	Vertical averaged salinity above the depth of the maximum potential density gradient	
	S_{BOT}	Vertical averaged salinity under the depth of the maximum potential density gradient	
BDepth	Depth	Bottom depth	m
Chl. a_{UPP}	Chl. a_{UPP}	Vertical averaged log-transformed Chlorophyll- a concentration above the depth of the maximum potential density gradient	
Chl. a_{BOT}	Chl. a_{BOT}	Vertical averaged log-transformed Chlorophyll- a concentration under the depth of the maximum potential density gradient	
aTSR	aTSR	Temporal difference from the Timing of Sea ice Retreat (TSR) anomaly to TSR between 1991 and 2013	days

3

1 **Table 4.** Eigenvalue and factor loadings of principle component analysis. The variances and
 2 eigenvalue of each principal component (PC) are also given. Descriptions of
 3 elements are same as Table 3 (See Table 3).

Elements	Eigenvector (Factor loadings)									
	PC1		PC2		PC3		PCA4		PCA5	
$\frac{dp}{dD_{max}}$	0.36	(0.55)	-0.55	(-0.73)	0.45	(0.38)	-0.27	(-0.10)	0.54	(0.15)
T _{UPP}	-0.51	(-0.78)	-0.38	(-0.50)	0.38	(0.32)	-0.38	(-0.13)	-0.56	(-0.15)
S _{UPP}	-0.43	(-0.66)	0.54	(0.71)	0.11	(0.09)	-0.54	(-0.19)	0.47	(0.13)
T _{BOT}	-0.60	(-0.92)	-0.18	(-0.24)	0.21	(0.18)	0.65	(0.23)	0.37	(0.10)
S _{BOT}	0.27	(0.41)	0.48	(0.63)	0.77	(0.65)	0.24	(0.08)	-0.21	(-0.06)
Eigenvalue	2.66		1.74		0.71		0.12		0.07	
Standard deviation	1.54		1.32		0.84		0.35		0.27	
Proportion of variance (%)	47.13		34.79		14.17		2.43		1.49	
Cumulative proportion (%)	47.13		81.92		96.08		98.51		100.00	

4

1 **Table 5.** Best models of each copepod groups: large Arctic (CopL_{arc}), small Arctic (CopS_{arc})
 2 and Pacific (Cop_{pac}) copepods.

Response variables	Best models	Deviance Explained (%)	Observed vs. Fitted
			R ²
CopL _{arc}	s(aTSR)+s(PC1)+s(PC2)+s(PC3)+s(Chl.a _{UPP})+s(Chl.a _{BOT})+s(Bdepth)+ ε	92.4	0.94
CopS _{arc}	s(aTSR)+ s(PC1)+s(PC3)+s(Chl.a _{UPP})+s(Chl.a _{BOT})+s(Bdepth)+ ε	89.9	0.88
Cop _{pac}	s(aTSR)+ s(PC1)+s(PC2)+s(PC3)+s(Chl.a _{BOT})+s(Bdepth)+ ε	75.3	0.38

3