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Net soil–atmosphere fluxes mask patterns in gross production and consumption of nitrous oxide and methane in a managed ecosystem

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Received: 3 November 2015 – Accepted: 25 November 2015 – Published: 4 December 2015

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Published by Copernicus Publications on behalf of the European Geosciences Union.

BGD

12, 19167–19197, 2015

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

Abstract

Nitrous oxide (N₂O) and methane (CH₄) are potent greenhouse gases that are both produced and consumed in soil. Production and consumption of these gases are driven by different processes, making it difficult to infer their controls when measuring only net fluxes. We used the trace gas pool dilution technique to simultaneously measure gross fluxes of N₂O and CH₄ throughout the growing season in a cornfield in northern California, USA. Net N₂O fluxes ranged from 0–4.5 mg N m⁻² d⁻¹ with the N₂O yield averaging 0.68 ± 0.02. Gross N₂O production was best predicted by net nitrogen (N) mineralization, soil moisture, and soil temperature ($R^2 = 0.60$, $n = 39$, $p < 0.001$). Gross N₂O reduction was correlated with the combination of gross N₂O production rates, net N mineralization rates, and CO₂ emissions ($R^2 = 0.74$, $n = 39$, $p < 0.001$). Overall, net CH₄ fluxes averaged -0.03 ± 0.02 mg C m⁻² d⁻¹. The methanogenic fraction of carbon mineralization ranged from 0 to 0.27% and explained 40% of the variability in gross CH₄ production rates ($n = 37$, $p < 0.001$). Gross CH₄ oxidation exhibited a strong positive relationship with gross CH₄ production rates ($R^2 = 0.67$, $n = 37$, $p < 0.001$), which reached as high as 5.4 mg C m⁻² d⁻¹. Our study is the first to demonstrate the simultaneous in situ measurement of gross N₂O and CH₄ fluxes, and results highlight that net soil–atmosphere fluxes can mask significant gross production and consumption of these trace gases.

1 Introduction

Greenhouse gas emissions from soils are major contributors to climate change (Ciais et al., 2013). While carbon dioxide (CO₂) is the most abundant greenhouse gas in the atmosphere, both nitrous oxide (N₂O) and methane (CH₄) are more potent with 34 and 298 times the global warming potential of CO₂ on a 100-year time scale, respectively (Myhre et al., 2013). Both N₂O and CH₄ are produced and consumed in soils by microbially-mediated redox-sensitive processes. However, most studies only measure

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



net soil–atmosphere exchange of N_2O and CH_4 . This approach cannot differentiate between production and consumption of these trace gases and thus limits our ability to infer controls on these processes and to diagnose model inaccuracies in predicting net N_2O and CH_4 fluxes. This hinders predictions of how soil–atmosphere N_2O and CH_4 fluxes will respond to future changes in land use practices or climate change.

Nitrous oxide consumption in soils by denitrifying bacteria leads to the production of dinitrogen gas (N_2), completing the N cycling. Nitrous oxide consumption is not generally considered to be an important process in upland soils because it is an anaerobic process. Rates of N_2O reduction to N_2 decrease as O_2 and NO_3^- availability increase (Weier et al., 1993; Firestone et al., 1980). Theoretically, this results in a high N_2O yield ($\frac{\text{N}_2\text{O}}{\text{N}_2\text{O}+\text{N}_2}$) in unsaturated soil where diffusive resupply of O_2 and the production of NO_3^- from nitrification would inhibit N_2O reduction. However, N_2O yields measured in oxic, upland soils span the entire range from 0 to 1 (Schlesinger, 2009; Stevens and Laughlin, 1998). This high variability in part reflects the difficulty in measuring rates of N_2O reduction to N_2 , particularly under field conditions (Groffman et al., 2006). The large range in N_2O yields also suggests that N_2O reduction to N_2 could play an important role in mitigating soil N_2O emissions to the atmosphere in some upland ecosystems.

Upland soils globally consume atmospheric CH_4 at a rate similar to the accumulation of CH_4 in the atmosphere (Ciais et al., 2013), and thus changes in the CH_4 sink strength of soils could influence atmospheric CH_4 concentrations. The inhibition of CH_4 oxidation associated with fertilizer application of NO_3^- (Aronson and Helliker, 2010), urea (Mosier et al., 1991), and NH_4^+ (Bedard and Knowles, 1989) is thought to cause lower net rates of CH_4 uptake in agricultural systems compared to natural ecosystems (Nesbit and Breitenbeck, 1992; Bender and Conrad, 1994; Koschorreck and Conrad, 1993; Dutaur and Verchot, 2007; Mosier et al., 1991). Inhibition by NH_4^+ has been attributed to enzymatic substrate competition due to the similarities between the CH_4 monooxygenase and NH_4^+ monooxygenase enzymes (Gulledge and Schimel, 1998) and to toxicity effects from nitrite produced during NH_4^+ oxidation (King and Schnell, 1994). However, the effect of N on CH_4 oxidation varies by soil (Gulledge et al., 1997),

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[⏪](#)[⏩](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



and at least some of this effect is due to inhibition by salts included in the fertilizer applications (Adamsen and King, 1993; Dunfield et al., 1993; Gulledge and Schimel, 1998; Nesbit and Breitenbeck, 1992). In addition, the response of CH_4 oxidation to NH_4^+ and NO_3^- may depend on the methanotrophic community; for example the high affinity Type II methane-oxidizing bacteria that dominate under low (< 1000 ppm) CH_4 conditions (Bender and Conrad, 1992) may be less sensitive to mineral N availability (Jang et al., 2011; Reay and Nedwell, 2004; Wang and Ineson, 2003). Thus, there remains uncertainty surrounding N inhibition of CH_4 oxidation as the mechanism leading to low net rates of CH_4 uptake in agricultural soils.

A major confounding factor in studies assessing controls on CH_4 oxidation is the simultaneous occurrence of methanogenesis and CH_4 oxidation. Net changes in CH_4 concentrations under oxic soil conditions are assumed to reflect only CH_4 oxidation (e.g., Nesbit and Breitenbeck, 1992) because methanogenesis occurs only under highly reducing conditions (Conrad, 1996). However, von Fischer and Hedin (2002) demonstrated that CH_4 production occurred in a wide range of dry, oxic soils with water-filled pore space as low as 20%. Similarly, Teh et al. (2005) documented the occurrence of methanogenesis under well-aerated conditions in an upland tropical forest soil. Macroaggregates can support net CH_4 efflux in unsaturated soil (Jackel et al., 2001; Sey et al., 2008), likely because O_2 consumption in the centers of the aggregates exceeds diffusive re-supply of O_2 to create reducing conditions (Sexstone et al., 1985). Microsites of methanogenesis could also occur in the rhizosphere where high rates of O_2 consumption from rhizosphere priming could create reducing conditions (Cheng et al., 2003). Because the controls on methanogenesis and CH_4 oxidation are likely very different, the co-occurrence of these processes means that we must measure gross rates of both processes simultaneously to elucidate the mechanisms driving patterns in net soil–atmosphere CH_4 fluxes.

We used the stable isotope trace gas pool dilution technique to measure gross N_2O and CH_4 fluxes in cornfield soils throughout the growing season. Fertilized agroecosystems are typically large net N_2O sources and small net CH_4 sinks (Haile-Mariam et al.,

2008; Kessavalou et al., 1998; Gelfand et al., 2013; Nangia et al., 2013; Robertson et al., 2000). However, little is known about the rates of gross production and consumption of these gases in upland soils, or their controlling factors. Different controls on production and consumption processes may result in complex responses of net soil–atmosphere gas fluxes to climate or management. Thus, the objectives of this study were to quantify field rates of gross N₂O and CH₄ production and consumption, and explore environmental and plant-mediated controls on these rates.

2 Materials and Methods

2.1 Study Site

The study site was a cornfield planted on a drained peatland located on Twitchell Island (38.11° N, 121.65° W) in the Sacramento–San Joaquin River Delta region of northern California. The region is very productive agriculturally, producing USD 500 million in crops in 1993 (Ingebritsen and Ikehara, 1999). The climate is Mediterranean with a winter wet season and summer dry season. The mean annual temperature is 20.5 °C, and annual precipitation ranges 375–625 mm (Atwater, 1980). The soils consist of mucky clay over buried peat and are classified as fine, mixed, superactive, thermic Cumulic Endoaquolls (Drexler et al., 2009). The field was fertilized once, at seeding, at a rate of 118 kg N ha⁻¹ with UAN 32, which consists of 45 % ammonium nitrate, 35 % urea, and 20 % water. The water table was maintained around 50 cm soil depth throughout the growing season via subsurface irrigation.

2.2 Study Design

We measured gross and net fluxes of CO₂, CH₄, and N₂O at five time points during the growing season from May to November 2012 on the following days after seeding (DAS): 11 (germination stage), 24 (seedling stage), 59 (peak growth stage), 94 (flowering stage), and 171 (senesced stage). The corn began senescing around DAS 104

19171

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



and was harvested on DAS 178. We performed measurements in row and inter-row locations with the assumption that plant effects, if any, would be greater in the rows where the corn was growing (Cai et al., 2012; Haile-Mariam et al., 2008; Kessavalou et al., 1998). We established three parallel transects spaced 50 m apart. We measured gross production and consumption of CH₄ and N₂O as well as net fluxes of CO₂, CH₄, and N₂O along the northern most transect and measured only net fluxes in the other two transects. In each transect, we used paired measurements in the bed (in-between corn rows) and furrow (in-row) with replicate pairs spaced 10 m apart ($n = 4$ pairs per transect). After each gas flux measurement was completed, we measured air, chamber headspace, and soil temperature at the surface flux chamber location. We also used an auger to sample the soil from the chamber footprint in 10 cm increments to 50 cm depth for the gross flux transect and only 0–10 cm depth in the net flux transects. The soils were processed the next day for determination of gravimetric soil moisture and net rates of nitrogen (N) mineralization and nitrification as described below.

2.3 Laboratory Assays

We determined net rates of N mineralization and nitrification from six-day laboratory incubations. We mixed each soil core by hand and subsampled 15 g for extraction in 75 mL of 2 M KCl, 10 g for determination of gravimetric soil moisture, and 50 g for incubation in Mason jars kept in the dark at ambient temperature. The jars were covered in perforated plastic wrap to minimize evaporation during the incubation. After six days, the soils in the jars were mixed and 15 g of soil was subsampled for KCl extractions. The KCl extracts were analyzed colorimetrically for NH₄⁺ and NO₃⁻ concentrations on a Lachat Quick Chem flow injection auto-analyzer (Lachat Instruments, Milwaukee, WI, USA). We calculated net N mineralization rates from the change in NH₄⁺ plus NO₃⁻ concentrations over the incubation period and net nitrification rates from the change in NO₃⁻ concentrations over the incubation period.

The remaining soil not utilized in the net rates incubation was air-dried for archival. Air-dried samples from the May sampling date were ground in a Spex Mill (Metuchen,

NJ, USA) for total C and N analyses on a Vario Micro Cube elemental analyzer (Elementar, Hanau, Germany).

2.4 Gas Flux Measurements

We used the stable isotope trace gas pool dilution technique to measure field rates of gross N_2O and CH_4 production and consumption (von Fischer and Hedin, 2002; Yang et al., 2011). We injected 10 mL of isotopically enriched spiking gas into the headspace of a 28 L surface flux chamber inserted 6 cm into the soil surface. The spiking gas consisted of 70 ppm N_2O at 98 atom % ^{15}N enrichment, 280 ppm CH_4 at 99 atom % ^{13}C enrichment, and 28 ppm SF_6 to achieve a ^{15}N - N_2O enrichment of 5.42 atom % and ^{13}C - CH_4 enrichment of 5.61 atom %. This spiking gas injection increased the chamber headspace gas composition by 25 ppb N_2O , 100 ppb CH_4 , and 10 ppb SF_6 . We sampled the chamber headspace at 5, 15, 30, 45, and 60 min after spiking gas injection. We analyzed samples on a Shimadzu GC-14A gas chromatograph (Columbia, MD, USA) equipped with a thermal conductivity detector, flame ionization detector, and electron capture detector for determination of CO_2 , CH_4 , N_2O , and SF_6 concentrations. We analyzed separate samples for ^{15}N - N_2O and ^{13}C - CH_4 on an IsoPrime 100 continuous flow isotope ratio mass spectrometer interfaced with a trace gas pre-concentration unit (Isoprime Ltd, Cheadle Hulme, UK) and Gilson GX271 autosampler (Middleton, WI). The trace gas analyzer was equipped with a combustion furnace using palladium to catalyze the conversion of CH_4 to CO_2 for isotopic analysis after CO and CO_2 were scrubbed from the sample (Fisher et al., 2006). One out of the 40 gross N_2O flux measurements and three out of the 40 gross CH_4 flux measurements were lost due to autosampler needle clogs that occurred during isotopic analysis.

Gross N_2O and CH_4 production and consumption rates were estimated using the pool dilution model as described by Yang et al. (2011) and von Fischer and Hedin (2002). The iterative model solves for gross production rates based on the isotopic dilution of the isotopically enriched chamber headspace pool of N_2O or CH_4 by natural

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

abundance N_2O or CH_4 emitted by the soil. Gross consumption rates were estimated from the empirical loss of the $^{15}\text{N}_2\text{O}$ or $^{13}\text{CH}_4$ tracer, using the loss of the SF_6 tracer to account for physical losses such as diffusion. We assumed that the isotopic composition of produced N_2O was 0.3431 atom % ^{15}N and the fractionation factor associated with N_2O reduction to N_2 was 0.9924. The justification for these assumptions is discussed by Yang et al. (2011). We assumed that the isotopic composition of produced CH_4 was 1.0473 atom %, based on measurements of the ^{13}C isotopic composition of soil CH_4 in a nearby study site (Y. Teh, personal communication, 2011). We assumed that the fractionation factor associated with CH_4 oxidation was 0.98 as justified by von Fischer and Hedin (2002). Sensitivity analyses performed by both Yang et al. (2011) and von Fischer and Hedin (2002) showed that the pool dilution model output is not sensitive to these assumed values at the high isotopic enrichments used. Net fluxes of CO_2 , N_2O , and CH_4 were determined from the change in concentration over time using an iterative model that fits an exponential curve to the data (Matthias et al., 1978). Fluxes were considered to be zero when the relationship between trace gas concentration and time was not significant at $p = 0.05$.

2.5 Statistical Analyses

We used SYSTAT Version 13 (SPSS Inc., Evanston, IL, USA) to perform statistical analyses and Microsoft Excel 2007 (Microsoft Corporation, Redmond, WA, USA) to run the iterative pool dilution model. We log-transformed the data to meet the normality assumptions of ANOVAs; soil moisture, soil temperature, soil C and N concentrations, and soil C : N ratios did not require transformation. We analyzed net and gross fluxes of CO_2 , N_2O , and CH_4 using sampling date as the within-subjects factor and location (i.e., bed vs. furrow) as the between-subject factor in repeated measures ANOVAs. We also analyzed net N mineralization and nitrification rates using sampling date as the within-subjects factor, and soil depth and location as the between-subjects factors in repeated measures ANOVAs. We explored relationships between trace gas fluxes and potential drivers (e.g., soil moisture, air and soil temperatures, soil NH_4^+ and NO_3^-

and $12.9 \pm 2.0 \mu\text{gNg}^{-1}$ in inter-rows. Soil NO_3^- concentrations were lower on DAS 11 than all other sampling dates (Table 3), averaging $53.5 \pm 7.2 \mu\text{gNg}^{-1}$ on DAS 11 and $215 \pm 33 \mu\text{gNg}^{-1}$ across all other sampling dates at 0–10 cm depth. Soil NO_3^- concentrations decreased with depth (Tables 2 and 3). On DAS 59 and 94 only, soil NO_3^- -concentrations were higher in rows ($387 \pm 117 \mu\text{gNg}^{-1}$) than in inter-rows ($156 \pm 23 \mu\text{gNg}^{-1}$) (Table 3).

Across the entire data set ($n = 216$), net N mineralization rates averaged $3.3 \pm 0.5 \mu\text{gNg}^{-1} \text{d}^{-1}$ and net nitrification rates averaged $2.7 \pm 0.6 \mu\text{gNg}^{-1} \text{d}^{-1}$. Net N mineralization and nitrification rates did not differ significantly among soil depths, sampling locations, or sampling dates (Table 3), although rates trended higher at 0–10 cm depth across all sampling dates and locations (Table 2). Across all sampling dates and soil depths, 96 % of the variability in net nitrification rates was explained by net N mineralization rates ($p < 0.001$, $n = 215$, Table 4).

Total C and N concentrations for soils sampled on DAS 11 differed between row and inter-row sampling locations (soil C, $F_{1,30} = 5.295$, $p = 0.03$; soil N, $F_{1,30} = 4.546$, $p = 0.04$) but not among soil depths (Table 2). Both soil C and N concentrations were higher in rows than in inter-rows, averaging $16.1 \pm 0.8\%$ C and $0.99 \pm 0.03\%$ N in rows and $13.7 \pm 0.5\%$ C and $0.89 \pm 0.02\%$ N in inter-rows. Soil C : N ratios averaged 15.8 ± 0.2 overall ($n = 40$), and did not differ significantly between sampling locations or among soil depths.

3.2 Gross and net N_2O fluxes

Across the entire data set, net N_2O fluxes ranged from 0–4.5 $\text{mgNm}^{-2} \text{d}^{-1}$ and averaged $1.6 \pm 0.2 \text{mgNm}^{-2} \text{d}^{-1}$ ($n = 112$). Net N_2O fluxes differed significantly among sampling dates ($F_{4,56} = 3.0$, $p = 0.03$) but not between sampling locations (Fig. 1a). Net N_2O fluxes were best predicted by net N mineralization, soil moisture, and soil CO_2 emissions together ($R^2 = 0.49$, Table 4).

BGD

12, 19167–19197, 2015

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

Gross N₂O production ranged from 0.09–6.6 mg N m⁻² d⁻¹ and gross N₂O reduction rates ranged from 0.00–0.95 mg N m⁻² d⁻¹. The N₂O yield averaged 0.68 ± 0.02 (*n* = 40). Both gross N₂O production and consumption rates differed significantly among sampling dates ($F_{4,20} = 4.5$, $p = 0.009$ and $F_{4,20} = 4.4$, $p = 0.01$, respectively) but not between sampling locations (Fig. 2a). The highest gross production and consumption rates occurred on DAS 59 and 171. Overall, gross N₂O production rates were best predicted by net N mineralization, soil moisture, soil temperature, and soil CO₂ emissions ($R^2 = 0.60$, Table 4). At peak growth (DAS 59 and 94), these variables explained 89% of the variability in gross N₂O production rates (*n* = 15, $p < 0.001$). When the corn was not actively growing, gross N₂O production was most strongly correlated with CO₂ emissions alone ($R^2 = 0.68$, *n* = 24, $p < 0.001$).

Gross N₂O reduction rates increased with gross N₂O production rates ($R^2 = 0.60$, *n* = 39, $p < 0.001$, Fig. 3a). Rates were also positively correlated with soil CO₂ emissions ($R^2 = 0.36$, *n* = 39, $p < 0.001$); this relationship was stronger when the corn was not actively growing (DAS 11, 24, and 171), with 80% of the variability in gross N₂O reduction rates explained by CO₂ emissions on these dates (*n* = 24, $p < 0.001$, Fig. 3b). Gross N₂O reduction was most strongly correlated with the combination of gross N₂O production rates, net N mineralization rates, and CO₂ emissions ($R^2 = 0.74$, *n* = 39, $p < 0.001$, Table 4).

3.3 Gross and net CH₄ fluxes

Net CH₄ fluxes ranged from -1.3 to 0.44 mg C m⁻² d⁻¹ but net fluxes were not detectable for 94 out of 112 measurements. Overall net CH₄ fluxes averaged -0.03 ± 0.02 mg C m⁻² d⁻¹. Using the trace gas pool dilution technique, we detected gross CH₄ production in 36 out of 37 measurements. Gross CH₄ production reached as high as 5.4 mg C m⁻² d⁻¹ with rates trending higher throughout the growing season (Fig. 2b). However, rates were only significantly different between DAS 11 and 94 ($F_{4,12} = 4.1$, $p = 0.03$). Gross CH₄ production rates were marginally significantly higher in rows than

BGD

12, 19167–19197, 2015

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



in inter-rows ($F_{1,3} = 5.8$, $p = 0.10$). Overall, gross CH_4 production rates were weakly correlated to soil CO_2 emissions ($R^2 = 0.17$, Table 4) but exhibited a stronger positive correlation with the methanogenic fraction of C mineralization ($R^2 = 0.40$, $n = 37$, $p < 0.001$, Fig. 4a), which ranged from 0 to 0.27% and averaged $0.06 \pm 0.01\%$. The strength of the relationship increased to $R^2 = 0.60$ ($n = 23$, $p < 0.001$) when considering only dates when the corn was not actively growing (Fig. 4a). When only peak growth sampling dates were considered (DAS 59 and 94), 57% of the variability in gross CH_4 production rates was predicted by the combination of CO_2 emissions, net N mineralization, and net nitrification ($n = 14$, $p = 0.03$).

Gross CH_4 oxidation did not differ significantly among sampling dates (Fig. 2b), averaging $1.1 \pm 0.2 \text{ mg C m}^{-2} \text{ d}^{-1}$ across all measurements ($n = 37$). Rates were marginally significantly higher in rows than in inter-rows ($F_{1,3} = 6.1$, $p = 0.09$). Gross CH_4 oxidation showed a strong positive relationship with gross CH_4 production ($R^2 = 0.67$, $n = 37$, $p < 0.001$). When gross CH_4 production rates exceeded $0.22 \text{ mg C m}^{-2} \text{ d}^{-1}$, gross CH_4 oxidation rates exhibited a tight 1 : 1 relationship with gross CH_4 production rates (slope = 1.06 ± 0.05 ; $R^2 = 0.95$, $n = 27$, $p < 0.001$, Fig. 4b). Below this threshold of gross CH_4 production, gross CH_4 oxidation was not correlated to gross CH_4 production rates alone, but was strongly correlated to the combination of gross CH_4 production and soil temperature ($R^2 = 0.67$, $n = 10$, $p = 0.02$); oxidation rates exhibited a negative relationship with soil temperature ($R^2 = 0.40$, $n = 10$, $p = 0.05$). Overall, gross oxidation rates were best predicted by the combination of gross CH_4 production rates, soil temperature, and CO_2 emissions ($R^2 = 0.79$, Table 4).

3.4 CO_2 emissions

Carbon dioxide emissions ranged from $0.6\text{--}10.5 \text{ g C m}^{-2} \text{ d}^{-1}$ across the entire data set. Emissions trended higher in the rows than in the inter-rows after the corn germinated, but repeated measures ANOVA showed that CO_2 emissions differed significantly among sampling dates ($F_{4,56} = 80.1$, $p < 0.001$) but not between row and inter-

BGD

12, 19167–19197, 2015

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



row locations (Fig. 1b). The highest CO₂ emissions occurred on DAS 59 and 94, at the height of the growing season, averaging $6.7 \pm 0.2 \text{ g C m}^{-2} \text{ d}^{-1}$; the lowest emissions occurred on DAS 11 and 24 at the beginning of the growing season, averaging $2.6 \pm 0.2 \text{ g C m}^{-2} \text{ d}^{-1}$. The variability in CO₂ emissions was poorly explained by environmental and soil variables with soil moisture and soil temperature together as the best, yet weak, predictors ($R^2 = 0.15$, Table 4).

4 Discussion

4.1 N₂O Dynamics

Net N₂O fluxes at our study site were comparable to those reported for other fertilized crop fields (Gelfand et al., 2013; Smith et al., 2011; Stevens and Laughlin, 1998; Nangia et al., 2013; Robertson et al., 2000), averaging $1.5 \pm 0.2 \text{ mg N m}^{-2} \text{ d}^{-1}$ across the growing season. Prior field estimates of N₂O yield using ¹⁵NH₄ or ¹⁵NO₃ addition at application rates of 200–300 kg N ha⁻¹ span a wide range from 0.06 to 0.7 (Mosier et al., 1986; Rolston et al., 1976, 1978, 1982). In contrast, the N₂O yield varied little throughout the growing season at our site, averaging 0.68 ± 0.02 , despite significant differences in both net and gross N₂O fluxes among sampling dates. This is similar to a field estimate of the N₂O yield for a nearby pasture on the same soil type (0.70 ± 0.04 ; Yang et al., 2011). Soil NO₃⁻ concentrations in surface soils (0–10 cm depth) were 1–2 orders of magnitude greater in the cornfield than in the pasture, so it is surprising that the N₂O yields were similar. Soil NO₃⁻ concentration was the strongest predictor of N₂O yield in a US Midwest cornfield soil incubated in the laboratory (Woli et al., 2010). Other factors such as soil pH, labile C availability, or soil aggregation may have played a more important role in controlling the N₂O yield in our cornfield (Sey et al., 2008).

The best predictors of gross N₂O production and consumption changed over the growing season, likely reflecting the influence of plant-microbial competition for N on N₂O dynamics. When the corn was actively growing, 89% of the variability in gross

BGD

12, 19167–19197, 2015

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

N_2O production was explained by soil moisture, soil temperature, net N mineralization, and CO_2 emissions together. In contrast, when the corn was not actively growing, both gross N_2O production and reduction were best predicted by soil CO_2 emissions alone. This may reflect the role of CO_2 emissions as proxy for the availability of labile C as an electron donor for denitrification; during the growing season, the contribution of autotrophic respiration to soil CO_2 emissions obscured this role. Net N mineralization was an explanatory variable for gross N_2O production only during the growing season when plant uptake of N could have limited N_2O production.

Overall, gross N_2O reduction rates were strongly correlated to gross N_2O production rates. This relationship was also observed in a managed grassland with high soil mineral N concentrations and net soil N_2O emissions (Yang et al., 2011), but not in a salt marsh with low mineral N availability where net N_2O uptake by soil occurred (Yang and Silver, 2015). The strong relationship between N_2O production and reduction may have driven the well-constrained N_2O yields in both this study and the managed grassland study because N_2O reduction increased proportionally to N_2O production rates. Additional studies using the trace gas pool dilution technique in the field could elucidate whether or not this relationship holds only in soils with high mineral N concentrations to drive high rates of N_2O production.

4.2 CH_4 dynamics

The small and zero net CH_4 fluxes we observed, which are typical of cornfields (Mosier et al., 2006), masked gross CH_4 fluxes which were two orders of magnitude greater. Net CH_4 fluxes were generally undetectable because CH_4 oxidation was tightly coupled to methanogenesis, especially at high gross CH_4 production rates. The ability of methanotrophs to adjust activity to match but not exceed rates of methanogenesis could reflect oxidation of soil-derived CH_4 at high concentrations near methanogenic microsites but not atmospheric CH_4 at low concentrations in the bulk soil. There are a few mechanisms that could drive a stimulatory effect of high CH_4 concentrations on CH_4 oxidation without increasing oxidation rates at atmospheric concentrations (Ben-

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

stead and King, 1997). First, high microsite CH_4 concentrations can increase the number of methanotrophs as well as shift the methanotrophic community composition from high affinity Type II methanotrophs, who consume CH_4 at low concentrations, to low affinity Type I methanotrophs, who consume CH_4 only at high concentrations, in or near the methanogenic microsites (Bender and Conrad, 1992, 1995). Second, the enzyme affinity of Type II methanotrophs can change from high affinity in the presence of atmospheric CH_4 concentrations to low affinity at high CH_4 concentrations, thereby reducing their capability to oxidize CH_4 at low concentrations (Dunfield et al., 1999). Third, high CH_4 availability may be needed to stimulate enzyme synthesis (Bender and Conrad, 1992, 1995; Nesbit and Breitenbeck, 1992), and thus methanotrophic activity may be induced only near methanogenic microsites and not in the bulk soil. Additional studies investigating gross CH_4 dynamics in soil aggregates or through the soil profile could provide insight into the mechanisms coupling CH_4 production and consumption. Regardless of the mechanisms, our observations suggest that using in situ methods that preserve spatial variability in soil CH_4 concentrations and allow for the occurrence of both CH_4 production and oxidation, such as the trace gas pool dilution technique, is important for accurately characterizing CH_4 dynamics in soil.

Gross CH_4 production rates were strongly positively correlated with the methanogenic fraction of C mineralization, an index of anaerobic soil microsites where electron acceptors are depleted relative to C supply (von Fischer and Hedin, 2007). Von Fischer et al. (2007) found that the methanogenic fraction was constrained below 0.04 % and gross CH_4 production rates below $1 \text{ mg C m}^{-2} \text{ d}^{-1}$ in tropical and temperate forest soils with less than 60 % water-filled pore space. Though the slope of the relationship between gross CH_4 production rates and the methanogenic fraction observed here was similar to that reported by von Fischer et al. (2007), the maximum methanogenic fraction observed here was nearly 7 times greater. The maximum gross CH_4 production rate was also an order of magnitude greater than the maximum rate of $0.5 \text{ mg C m}^{-2} \text{ d}^{-1}$ reported by von Fischer and Hedin (2002) for a range of unsaturated upland soils in which net CH_4 fluxes were near zero (-0.2 to $0.2 \text{ mg C m}^{-2} \text{ d}^{-1}$). This

suggests a higher potential for the development of methanogenic microsites in these drained peatland soils, which are rich in C.

The near zero net CH₄ fluxes measured in our cornfield are consistent with other studies in agricultural systems, but the relatively high gross CH₄ oxidation rates we documented challenge the paradigm that agricultural soils have low potential for CH₄ oxidation compared to unsaturated soils in natural ecosystems (Bender and Conrad, 1994; Koschorreck and Conrad, 1993; Mosier et al., 1991; Nesbit and Breitenbeck, 1992; Zhuang et al., 2013). Our soils had high NH₄⁺ and NO₃⁻ concentrations, which did not limit the ability of methanotrophs to completely consume soil-derived CH₄. Undisturbed soils in which CH₄ production and consumption occur simultaneously could behave differently than manipulated soils incubated in the laboratory under conditions to isolate CH₄ oxidation from CH₄ production, and vice versa. Application of the trace gas pool dilution technique to other agricultural fields could reveal whether or not the tight coupling of CH₄ production and consumption rather than low rates of CH₄ production and oxidation could be responsible for the general observation of small and near zero net CH₄ fluxes in agricultural ecosystems. A greater understanding of limitations on gross CH₄ oxidation under field conditions is needed to accurately predict how land use change will alter soil–atmosphere CH₄ exchange and to better manage agricultural soils to be atmospheric CH₄ sinks.

Our data provide circumstantial evidence that plants could mediate gross CH₄ dynamics in upland soil. First, we observed a steady, though not statistically significant, increase in gross CH₄ fluxes over the course of the growing season. Both gross CH₄ production and oxidation rates were approximately 2.5 times greater at DAS 171 compared to DAS 11. This trend in gross CH₄ fluxes cannot be explained by changes in environmental variables such as soil temperature, which peaked in the middle of the growing season, and soil moisture, which decreased over the growing season. An increase in plant C inputs to the soil over the growing season may have driven the increase in rates of methanogenesis. Second, both gross CH₄ production and oxidation rates were higher in rows than in inter-rows. Kessavalou et al. (1998) found that net

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



CH₄ fluxes did not differ between row and inter-row, but our results demonstrate that net fluxes may mask patterns in gross CH₄ dynamics. Greater methanogenesis in rows could reflect a greater number of anaerobic microsites caused by rhizosphere priming fueling biological O₂ demand for C mineralization (Zhu et al., 2014).

5 Conclusions

Our study demonstrates that the anaerobic processes of N₂O reduction to N₂ and methanogenesis can play important roles in mediating soil-atmosphere greenhouse gas fluxes in upland crop field soils where these processes have previously been discounted. Moreover, despite high soil NO₃⁻ and NH₄⁺ concentrations that theoretically inhibit N₂O reduction to N₂ as well as CH₄ oxidation, gross N₂O reduction rates were approximately half that of gross N₂O production rates and CH₄ oxidation kept pace with methanogenesis that reached relatively high rates for unsaturated soil. Our field measurements of gross N₂O and CH₄ fluxes thus challenge our current understanding of the controls on the production and consumption of N₂O and CH₄ in upland soils. The strong correlations that gross N₂O and CH₄ fluxes exhibited with soil characteristics and soil N cycling process rates can help guide controlled studies to investigate the controls on the processes that lead to the production and consumption of N₂O and CH₄. A better understanding of the controls on these processes can help refine modeling efforts to characterize the effects of anoxic microsites in unsaturated soil on greenhouse gas emissions (Riley et al., 2011) and also inform land management decisions to mitigate soil greenhouse gas emissions from crop fields.

Acknowledgements. We appreciate field and lab assistance from Heather Dang, Andrew McDowell, Gavin McNicol, Julia Cosgrove, Rebecca Ryals, Zoe Statman-Weil, Jonathan Treffkorn, Jonathan Lee, Taichi Nataka, Ryan Salladay, and Kristina Solheim. Funding was provided by the California Department of Water Resources, by a US National Science Foundation grant (DEB-0543558) to WLS and by the USDA National Institute of Food and Agriculture, McIntire Stennis project (CA-B-ECO-7673-MS) to WLS.

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- 30

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)

[⏪](#)

[⏩](#)

[◀](#)

[▶](#)

[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)

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Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)

[⏪](#)

[⏩](#)

[◀](#)

[▶](#)

[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)

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Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

[Title Page](#)

[Abstract](#)

[Introduction](#)

[Conclusions](#)

[References](#)

[Tables](#)

[Figures](#)

[⏪](#)

[⏩](#)

[◀](#)

[▶](#)

[Back](#)

[Close](#)

[Full Screen / Esc](#)

[Printer-friendly Version](#)

[Interactive Discussion](#)

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**Gross methane and
nitrous oxide fluxes
in a managed
ecosystem**W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

I ◀

▶ I

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



Zhu, B., Gutknecht, J. L. M., Herman, D. J., Keck, D. C., Firestone, M. K., and Cheng, W. X.: Rhizosphere priming effects on soil carbon and nitrogen mineralization, *Soil Biol. Biochem.*, 76, 183–192, doi:10.1016/j.soilbio.2014.04.033, 2014.

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Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Table 1. Environmental and soil (0–10 cm depth) variables by sampling date (mean \pm SE).

Variable	<i>F</i> statistic	Sampling date				
		DAS 11 (<i>N</i> = 8)	DAS 24 (<i>N</i> = 24)	DAS 59 (<i>N</i> = 24)	DAS 94 (<i>N</i> = 16)	DAS 171 (<i>N</i> = 24)
Air temperature (° C)	$F_{4,56} = \mathbf{3.4}$	27.6 \pm 0.9 a	26.8 \pm 1.2 a	25.8 \pm 0.5 ab	28.2 \pm 0.7 a	24.5 \pm 0.7 b
Soil temperature (° C)	$F_{4,56} = \mathbf{245}$	18.4 \pm 0.2 a	22.1 \pm 0.3 b	24.2 \pm 0.3 c	22.8 \pm 0.2 b	14.8 \pm 0.1 d
Soil moisture (g H ₂ O g ⁻¹ soil)	$F_{4,24} = \mathbf{34}^*$	0.38 \pm 0.02	0.34 \pm 0.02	0.35 \pm 0.01	0.24 \pm 0.01	0.32 \pm 0.01

Degrees of freedom are shown in subscripts, and statistically significant *F* statistics at $P < 0.05$ are indicated by bold text. Letters indicate statistically significant differences among sampling dates.

* One transect was excluded from the repeated measures ANOVA because data are missing for one sampling date.

[Title Page](#)
[Abstract](#)
[Introduction](#)
[Conclusions](#)
[References](#)
[Tables](#)
[Figures](#)
[◀](#)
[▶](#)
[◀](#)
[▶](#)
[Back](#)
[Close](#)
[Full Screen / Esc](#)
[Printer-friendly Version](#)
[Interactive Discussion](#)

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Table 2. Soil characteristics and N cycling rates across all sampling dates by soil depth in the gross flux transect (mean \pm SE).

Variable	N	P value	0–10 cm	10–20 cm	20–30 cm	30–40 cm	40–50 cm
Soil moisture ($\text{g H}_2\text{O g}^{-1}$)	40	< 0.001	0.34 \pm 0.01 a	0.35 \pm 0.01 a	0.37 \pm 0.01 ab	0.40 \pm 0.01 b	0.46 \pm 0.02 c
NH_4^+ concentration ($\mu\text{g N g}^{-1}$)	40	< 0.001	23.3 \pm 9.6 a	15.2 \pm 6.3 b	7.0 \pm 1.8 b	5.0 \pm 0.7 b	5.7 \pm 0.8 b
NO_3^- concentration ($\mu\text{g N g}^{-1}$)	40	< 0.001	183 \pm 28 a	110 \pm 22 b	58.9 \pm 8.4 c	41.9 \pm 6.1 c	29.5 \pm 3.2 c
Net mineralization ($\mu\text{g N g}^{-1} \text{d}^{-1}$)	40		5.9 \pm 2.6	1.0 \pm 0.9	1.5 \pm 0.5	1.8 \pm 0.7	3.3 \pm 0.5
Net nitrification ($\mu\text{g N g}^{-1} \text{d}^{-1}$)	40		6.7 \pm 2.4	1.7 \pm 0.8	2.1 \pm 0.5	2.3 \pm 0.7	3.8 \pm 0.5
Soil C concentration (%)	8*		14.1 \pm 0.5	15.4 \pm 1.6	14.8 \pm 0.9	15.2 \pm 1.6	15.0 \pm 0.9
Soil N concentration (%)	8*		0.93 \pm 0.02	0.96 \pm 0.07	0.98 \pm 0.05	0.91 \pm 0.06	0.93 \pm 0.05

Letters indicate statistically significant differences among soil depths.

* Data from DAS 11 only.

[Title Page](#)
[Abstract](#)
[Introduction](#)
[Conclusions](#)
[References](#)
[Tables](#)
[Figures](#)
[⏪](#)
[⏩](#)
[◀](#)
[▶](#)
[Back](#)
[Close](#)
[Full Screen / Esc](#)
[Printer-friendly Version](#)
[Interactive Discussion](#)

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Table 3. Results from repeated measures ANOVAs with sampling date, the interaction of sampling date and soil depth, and the interaction of sampling date and sampling location as the within subjects, and soil depth and sampling location as the between subjects factors.

	Sampling date	Soil depth	Sampling location location	Sampling date * Soil depth	Sampling date * Sampling location
Soil moisture ($\text{g H}_2\text{O g}^{-1}$)	$F_{4,120} = \mathbf{135}$	$F_{4,30} = \mathbf{31}$	$F_{1,30} = \mathbf{5.1}$	$F_{16\ 120} = \mathbf{1.9}$	$F_{4,120} = \mathbf{4.6}$
NH_4^+ concentration ($\mu\text{g N g}^{-1} \text{d}^{-1}$)	$F_{4,120} = \mathbf{7.9}$	$F_{4,30} = \mathbf{7.7}$	$F_{1,30} = \mathbf{4.0}$	$F_{16\ 120} = 0.90$	$F_{4,120} = 1.9$
NO_3^- concentration ($\mu\text{g N g}^{-1} \text{d}^{-1}$)	$F_{4,120} = \mathbf{17}$	$F_{4,30} = \mathbf{36}$	$F_{1,30} = \mathbf{18}$	$F_{16\ 120} = 1.0$	$F_{4,120} = \mathbf{7.1}$
Net mineralization ($\mu\text{g N g}^{-1} \text{d}^{-1}$)	$F_{4,120} = 1.5$	$F_{4,30} = 1.5$	$F_{1,30} = 1.5$	$F_{16\ 120} = 1.1$	$F_{4,120} = 1.7$
Net nitrification ($\mu\text{g N g}^{-1} \text{d}^{-1}$)	$F_{4,120} = 1.4$	$F_{4,30} = 0.22$	$F_{1,30} = 1.9$	$F_{16\ 120} = 0.31$	$F_{4,120} = 1.8$

Degrees of freedom are shown in subscripts, and statistically significant F statistics at $P < 0.05$ are indicated by bold text.

[Title Page](#)
[Abstract](#)
[Introduction](#)
[Conclusions](#)
[References](#)
[Tables](#)
[Figures](#)
[Back](#)
[Close](#)
[Full Screen / Esc](#)
[Printer-friendly Version](#)
[Interactive Discussion](#)


Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Table 4. Coefficients for multiple linear regressions predicting trace gas fluxes using soil variables.

Dependent variable	N	R ²	Effect	Coefficient	SE	P value
Log(Net nitrification, $\mu\text{g N g}^{-1} \text{d}^{-1}$)	215	0.96	Constant	0.162	0.020	< 0.001
			Log(Net N mineralization, $\mu\text{g N g}^{-1} \text{d}^{-1}$)	0.906	0.012	< 0.001
Log(CO ₂ emissions, $\text{g C m}^{-2} \text{d}^{-1}$)	96	0.15	Constant	0.466	0.195	0.02
			Soil moisture ($\text{g H}_2\text{O g}^{-1}$)	-1.019	0.416	0.02
			Soil temperature ($^{\circ}\text{C}$)	0.022	0.007	0.002
Log(Net N ₂ O flux, $\text{mg N m}^{-2} \text{d}^{-1}$)	56	0.49	Constant	-1.671	0.743	0.03
			Log(Net N mineralization)	1.342	0.274	< 0.001
			Soil moisture	4.356	1.539	0.007
			Log(CO ₂ emissions)	1.404	0.288	< 0.001
Log(Gross N ₂ O production, $\text{mg N m}^{-2} \text{d}^{-1}$)	39	0.60	Constant	1.743	0.453	0.001
			Log(Net N mineralization)	0.516	0.139	0.001
			Soil moisture	2.226	0.839	0.01
			Soil temperature	-0.043	0.013	0.002
			Log(CO ₂ emissions)	1.056	0.180	< 0.001
Log(Gross N ₂ O reduction)	39	0.74	Constant	-2.525	0.556	< 0.001
			Log(Net N mineralization)	0.680	0.258	0.01
			Log(Gross N ₂ O production)	0.983	0.226	< 0.001
			Log(CO ₂ emissions)	1.199	0.292	< 0.001
Log(Gross CH ₄ production, $\text{mg C m}^{-2} \text{d}^{-1}$)	37	0.17	Constant	2.264	0.199	< 0.001
			Log(CO ₂ emissions)	0.921	0.348	0.01
Log(Gross CH ₄ oxidation)	37	0.79	Constant	0.794	0.621	0.21
			Log(Gross CH ₄ production)	1.090	0.142	0.002
			Soil temperature	-0.086	0.024	0.001
			Log(CO ₂ emissions)	1.096	0.335	< 0.001

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

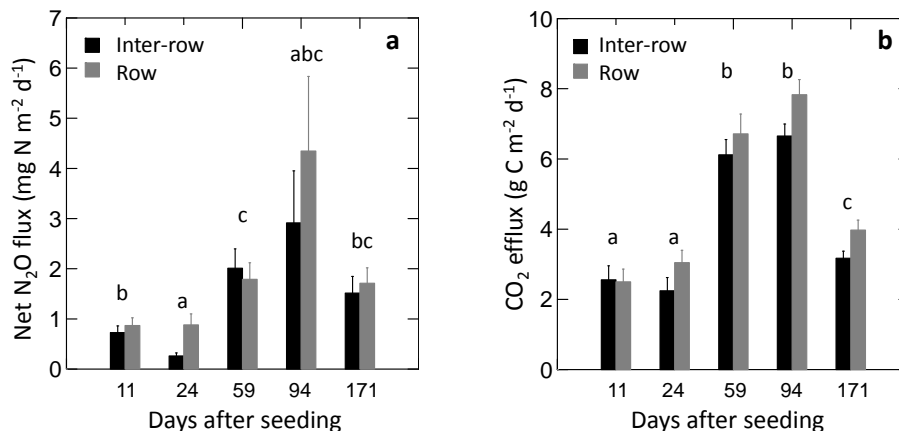


Figure 1. Mean (a) net N₂O flux and (b) CO₂ efflux for all three transects ($n = 24$ per sampling date except $n = 16$ on DAS 94) in inter-rows (black bars) and rows (grey bars). Error bars represent standard errors, and different letters indicate statistically significant differences among sampling dates.

[Title Page](#)
[Abstract](#)
[Introduction](#)
[Conclusions](#)
[References](#)
[Tables](#)
[Figures](#)
[⏪](#)
[⏩](#)
[◀](#)
[▶](#)
[Back](#)
[Close](#)
[Full Screen / Esc](#)
[Printer-friendly Version](#)
[Interactive Discussion](#)

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

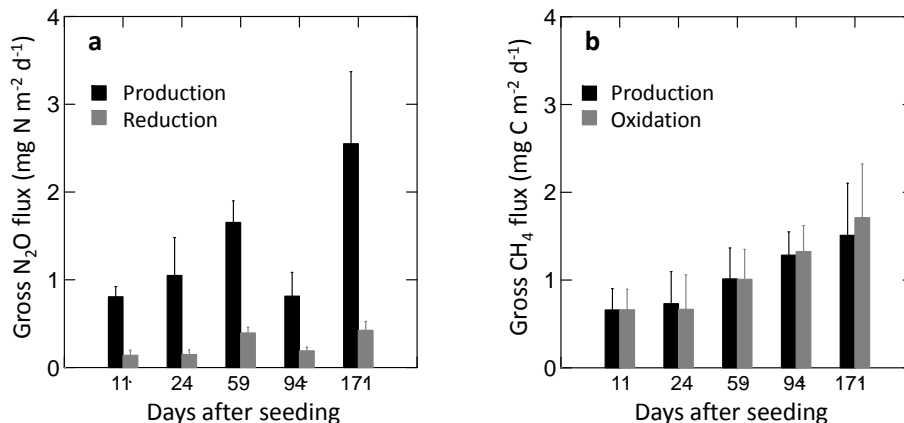


Figure 2. Mean (a) gross N₂O production rates (black bars) and reduction rates (grey bars) and (b) gross CH₄ production rates (black bars) and oxidation rates (grey bars). Error bars represent standard errors ($n = 8$ per sampling date).

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

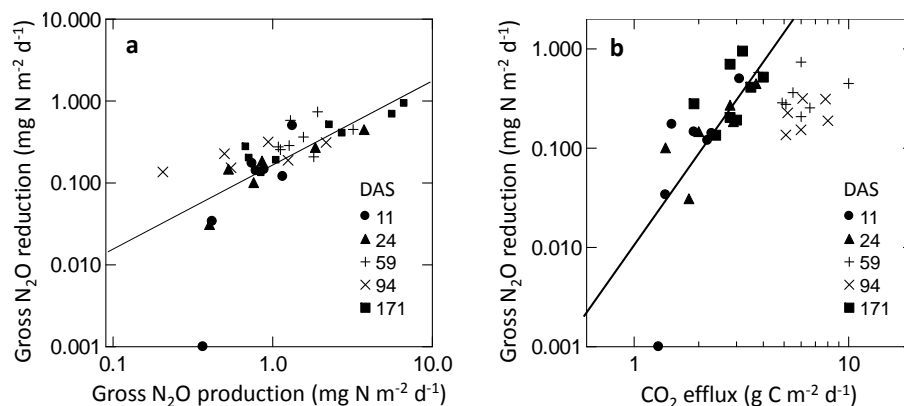


Figure 3. Gross N_2O reduction rates vs. **(a)** gross N_2O production and **(b)** CO_2 efflux. Symbols represent sampling on different days after seeding (DAS): circles are DAS 11, triangles are DAS 24, pluses are DAS 59, crosses are DAS 94, and squares are DAS 171. The line represents the regression line for sampling dates when the corn was not at peak growth (DAS 11, 24, and 171), $\log_{10}(y) = [3.663 \times \log_{10}(x)] + 0.822$ ($R^2 = 0.80$, $n = 24$, $p < 0.001$).

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

Gross methane and nitrous oxide fluxes in a managed ecosystem

W. H. Yang and
W. L. Silver

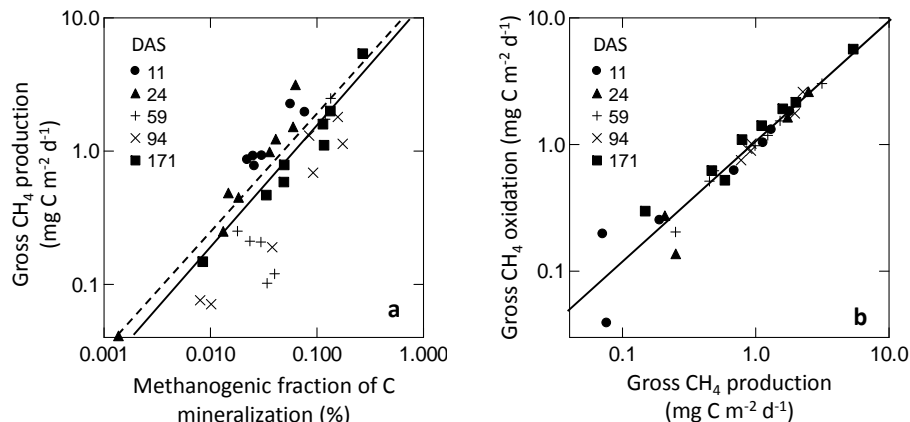


Figure 4. (a) Gross CH₄ production rates vs. methanogenic fraction of C mineralization, and (b) gross CH₄ oxidation rates vs. gross CH₄ production rates. Symbols represent sampling on different days after seeding (DAS): circles are DAS 11, triangles are DAS 24, pluses are DAS 59, crosses are DAS 94, and squares are DAS 171. The solid lines represent the regression line for all sampling dates together, (a) $\log_{10}(y) = [18.953 \times \log_{10}(x)] + 2.245$ ($R^2 = 0.40$, $n = 37$, $p < 0.001$) and (b) $\log_{10}(y) = [1.308 \times \log_{10}(x)] - 1.028$ ($R^2 = 0.67$, $n = 37$, $p < 0.001$). The dashed line represents the regression line for DAS 11, 24, and 171 only, $\log_{10}(y) = [22.681 \times \log_{10}(x)] + 1.904$ ($R^2 = 0.60$, $n = 23$, $p < 0.001$).