

1 **Survival and settling of larval *Macoma balthica* in a large-scale mesocosm experiment at different**
2 ***f*CO₂ levels**

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13

14 **Abstract**

15 Anthropogenic carbon dioxide (CO₂) emissions are causing severe changes in the global inorganic
16 carbon balance of the oceans. Associated ocean acidification is expected to impose a major threat to
17 marine ecosystems worldwide, and it is also expected to be amplified in the Baltic Sea where the system
18 is already exposed to relatively large natural seasonal and diel pH fluctuations. We studied the responses
19 of larvae of the benthic key-species *Macoma balthica* to a range of future CO₂-scenarios using six ~ 55
20 m³ mesocosms encompassing the entire pelagic community. The mesocosms were deployed in the
21 northern Baltic Sea in June 2012. We focused on the survival, growth and subsequent settlement process
22 of *Macoma balthica* when exposed to different levels of future CO₂. The size and time to settlement of
23 *M. balthica* increased along the CO₂ gradient, suggesting a developmental delay. With on-going climate
24 change, both the frequency and extent of regularly occurring high CO₂ conditions is likely to increase,
25 and a permanent pH decrease will likely occur. The strong impact of increasing CO₂ levels on early-
26 stage bivalves is alarming as these stages are crucial for sustaining viable populations, and a failure in
27 their recruitment would ultimately lead to negative effects on the population.

28

29 **1 Introduction**

30 Anthropogenic CO₂-emissions are causing severe changes in the oceans (Feely et al., 2004). Future
31 ocean acidification (OA), which includes changes in the inorganic carbon balance of the seawater
32 coupled with a decrease in pH, is occurring at a rate faster than experienced in the geological past
33 (Hönisch et al., 2012), and is expected to impose a major threat to marine ecosystems worldwide (Orr et
34 al., 2005; Fabry et al., 2008). The sea surface pH is estimated to decrease by 0.4 units in the global open
35 oceans by the year 2100 (Caldeira and Wickett, 2003), whereas many coastal areas already experience
36 large pH fluctuations reaching to considerably lower pH levels than predicted for the near future
37 (Blackford and Gilbert, 2007; Johnson et al., 2013). The multiple environmental stressors impacting
38 coastal areas and the local processes that impact watersheds make the precise modelling of future pH
39 levels exceedingly challenging for these areas (Borges and Gypens, 2010; Duarte et al., 2013).

40

41 The majority of studies investigating the biological effects of future CO₂ levels have focused on its
42 impacts on calcifying species and on pelagic primary producers. Pelagic calcifiers such as bivalve early
43 life-stages are generally considered susceptible to increasing CO₂ levels (Kurihara, 2008; Dupont and
44 Thorndyke, 2009), with a range of observed (mostly negative) impacts on development, survival and
45 growth of larval stages as consequences of the CO₂ increase (Gazeau et al., 2013). Also the settling and
46 survival of post-larvae are impacted by the changes in the water chemistry (Green et al., 2004, 2009;
47 Clements and Hunt, 2014). The response of organisms to future CO₂ levels has traditionally been
48 studied in experiments focusing on single species, and the community-wide responses are still not well
49 known. In mesocosms, the natural community can be maintained to a high degree, and organismal
50 performance can be measured in near-natural surroundings (Riebesell et al., 2010). Mesocosm studies
51 have the additional advantages of allowing experimental manipulation of environmental factors such as
52 CO₂, possibility for replication, and repeated sampling of the closed study systems over long
53 experimental duration.

54

55 In the Baltic Sea a drop in pH of 0.5 units is estimated for the surface waters within this century
56 (Hjalmarsson et al., 2008; Omstedt et al., 2012). Similar to coastal and estuarine areas (Duarte et al.,

2013), however, the natural pH variability in the Baltic Sea is large and regularly exceeds the estimates made for the near-future (Omstedt et al., 2009; Melzner et al., 2012; Jansson et al., 2013). For example, during the summer season pH changes of nearly one unit per day driven by changes in primary production and respiration are common in the shallow coastal areas of the northern Baltic Proper (pers. obs.). Yet, ocean acidification is likely to increase the pH fluctuations, making the occasionally experienced extreme pH levels even more pronounced, further expanding the pH range which the Baltic species are exposed to (Thomas and Schneider, 1999; Omstedt et al., 2010; Melzner et al., 2012). A key species in the Baltic Sea soft-bottom communities, the bivalve *Macoma balthica* (L.), is experiencing variable conditions throughout its life-cycle. During the larval phase, it is exposed to large pelagic diel pH-fluctuations (Jansson et al., 2013; Almén et al., 2014) followed by the harsh reducing conditions of the sedimentary system when settling into the benthic environment (Woodin et al., 1998). The tolerance of *M. balthica* to low pH conditions has so far been studied in aquarium experiments of different types and durations (van Colen et al., 2012; Jansson et al., 2013), which have shown negative effects on the early-stage bivalves. In such experiments, however, the potential impact of future environmental changes on e.g. the settlement process is challenging to study.

The aim of the whole large-scale pelagic mesocosm experiment was to study the responses of the Baltic Sea pelagic community to different future $f\text{CO}_2$ -scenarios. In this specific study we wanted to explicitly shed light on 1) growth and survival of *M. balthica* larvae and 2) the subsequent settling of the post-larvae, when exposed to different levels of future CO_2 in their natural surroundings. Based on the results of our previous experiments (Jansson et al., 2013; van Colen et al., in prep.), we predicted the growth of the larvae to decrease along the increasing $f\text{CO}_2$ gradient and the survival and settling to be negatively impacted by the $f\text{CO}_2$ increase.

2 Material and methods

2.1 The study species

The infaunal bivalve *M. balthica* is abundant throughout the Baltic Sea, often dominating biomass in soft sediments from organic mud to sandy bottoms from the very shallow down to 190 m depth

85 (Segerstråle, 1960; Elmgren et al., 1986; Bonsdorff, 2006). The spawning of *M. balthica* occurs when
86 water temperature has reached approximately 7°C (Caddy, 1967). The planktonic life stage (ca. 6
87 weeks) ends when the individual has reached a sufficient size and developmental stage (including
88 increased mobility of the foot) to metamorphose and settle to the seafloor (Caddy, 1969). A majority of
89 the very newly settled bivalves encountered in the Baltic Sea have a size of 250–300 µm (Ankar, 1980;
90 Elmgren et al., 1986; Olafsson, 1989). Peak settling in the northern parts of the Baltic Sea typically
91 occurs in July. During the pelagic larval phase, abundances of up to 12 000 larvae m⁻³ are measured in
92 the Baltic Sea, with a settling population of around 30 000 m⁻² each year, at peak settling even up to
93 300 000 m⁻² (Ankar, 1980; Elmgren et al., 1986; Bonsdorff et al., 1995). *M. balthica* is an important
94 prey organism, and has a central role in sediment reworking and bioturbation, contributing to the overall
95 health and functioning of the benthic ecosystem (Michaud et al., 2006). In the species-poor northern
96 Baltic Sea, this species is essential to the functioning of the benthic ecosystem through these key
97 processes (Villnäs et al., 2012; Norkko et al., 2013).

98

99 **2.2 Experimental set-up**

100 Six pelagic mesocosms (KOSMOS, Riebesell et al. 2013a) of ~55 m³ were deployed in the western Gulf
101 of Finland (59° 51.5' N, 23° 15.5' E) on 12 June 2012 to study responses of the Baltic Sea plankton
102 community to increased fugacity of carbon dioxide (*f*CO₂). The mesocosm bags were lowered down to a
103 depth of 17 m to enclose the natural plankton community, excluding organisms larger than 3 mm by a
104 mesh installed at the top and bottom of the cylindrical bags. With the bags fully submerged below the
105 sea surface, water and organisms inside the bags could exchange with the surrounding water mass for
106 five days before closing the mesocosms on 17 June (day -5, 5 days before CO₂ manipulation). To seal
107 the bottom of each mesocosm, a two meter long sediment trap funnel collecting settling particles and
108 organisms was installed by divers to replace the 3 mm mesh. The top end of the bags was
109 simultaneously pulled above the sea surface to fully isolate the enclosed water bodies. Bubbling the
110 systems with compressed air for three and a half minutes right after closure destroyed the halocline
111 present inside the bags. The mesocosms were manipulated with filtered (50 µm), CO₂-saturated
112 seawater as described by Riebesell et al. (2013a) on four consecutive days (day 0–3) to establish a range

113 of four $f\text{CO}_2$ target treatments (600–1650 μatm) and two ambient blind manipulated mesocosms (Table
114 1). On day 15 $f\text{CO}_2$ was readjusted inside the treated mesocosms to counteract outgassing of CO_2 . For a
115 more detailed description of the experimental set-up, manipulations and maintenance of the mesocosms
116 please see Paul et al. (2015).

117

118 **2.3 Sampling the mesocosms**

119 **2.3.1 Water parameters**

120 CTD profiles were measured daily with a handheld self-logging CTD60M probe (Sea and Sun
121 Technology) from 0.3 down to 18 m (mesocosms) and to 30 m (surrounding bay) with sensors for
122 salinity, temperature, dissolved oxygen, PAR (photosynthetic active radiation) and pH. Details on the
123 sensors and their accuracy are described in Schulz and Riebesell (2013). Depth-integrated water samples
124 (IWS, HYDRO-BIOS Kiel) were collected regularly (daily to every other day, see Paul et al., 2015)
125 from all mesocosms and the surrounding water body to measure e.g. total pH (pHT), total alkalinity
126 (TA) and dissolved inorganic carbon (DIC) for determining the inorganic carbon components, and
127 chlorophyll *a* to follow the development of the phytoplankton bloom. pHT was determined by analyzing
128 samples with a Cary 100 (Varian) spectrophotometer (Dickson et al., 2007). The details of the procedure
129 ($f\text{CO}_2$ was calculated from measured DIC and pHT) are described in Paul et al. (2015). CTD pH
130 measurements were corrected to pH on the total scale by linear correlations of mean water column
131 potentiometric pH measurements to spectrophotometric pHT measurements. Exact details of all
132 sampling procedures, equipment used and sample analyses are described in Riebesell et al. (2013a),
133 Schulz et al. (2013) and Paul et al. (2015).

134

135 **2.3.2 Water column: Mesozooplankton sampling and quantification of *M. balthica* larvae**

136 Mesozooplankton samples from the six mesocosms were taken with an Apstein net of 17 cm diameter
137 and 100 μm mesh size by pulling the net vertically from 17 m depth to the sea surface. Net hauls were
138 taken from the mesocosms on eleven sampling days: prior to the first CO_2 addition (days -3, -2, -1), on
139 the day of the first CO_2 addition (day 0), and after the first CO_2 addition in a seven day rhythm (days 3,
140 10, 17, 24, 31, 38, 45). Mesozooplankton samples were preserved in 70% ethanol. The larvae of *M.*

141 *balthica* were counted in the whole sample under a stereo microscope (WILD M3B). For size range
142 determination, on average 70 individuals were measured from each mesocosm on days 0 and 10. The
143 individuals were photographed using a dissecting microscope connected to a Nikon DS-Fi2 camera
144 system, and sizes were determined by measuring shell lengths using the Nikon DS camera interface.
145 Zooplankton abundance was calculated as individuals per cubic meter, assuming 100% filtering
146 efficiency of the net. For more details on mesozooplankton sampling and processing see Lischka et al.
147 (2015).

148

149 **2.3.3 Sediment traps: collection of material, subsampling and quantification of settling *M. balthica***

150 The sediment traps were emptied every second day using a gentle vacuum to pump the samples through
151 a silicon tube into sampling flasks at the sea surface (for more details see Boxhammer et al., 2015).
152 Subsamples of 20 mL were taken with a pipette of the homogeneously mixed samples (on average
153 2.5 L) and preserved in 4% buffered formalin for quantification and size determination of settling
154 bivalves. Abundance and size range determinations of settled bivalves were made on 3 replicates of 1
155 mL subsamples. *M. balthica* collected in the sediment traps included settled individuals as well as
156 individuals that had died in the water column or in the sediment trap after settling. However, the gaping
157 shells of individuals that were dead at the time of sampling were identified in the preserved samples and
158 such individuals were not counted. Individuals that were assessed to be living at the time of sampling
159 were counted and photographed using a dissecting microscope connected to a Nikon DS-Fi2 camera
160 system. During the main settling period (days 11, 13, 15 and 17) on average 35 individuals were
161 measured from each mesocosm. Sizes were determined by measuring shell lengths using the Nikon DS
162 camera interface.

163

164 **2.4 Numerical analysis**

165 The abundance of bivalve larvae in the water column of each mesocosm over time was compared by
166 calculating a rate of change between each sampling day and comparing the timing of decreasing
167 abundances. This was done by calculating Spearman correlation ranks for each time point. To analyse
168 the differences in post-larval settling between the mesocosms, we performed a chi-square test to

169 compare the cumulative abundances of settled individuals on days 9, 11, 13, 15, 17 and 19. Graphical
170 post-hoc tests were performed to identify differences between mesocosms.

171

172 The sizes of both the larvae in the water column and the post-larvae in the settling traps in the different
173 $f\text{CO}_2$ levels were compared by a linear regression model. To standardize the comparisons, they were
174 conducted on average sizes of a batch of individuals measured in each mesocosm. The residuals of the
175 regressions adhered to the assumption of normality. All analyses were performed in the software R
176 (version 3.0.2; R Development Core Team, 2012). The differences were considered significant at $p <$
177 0.05 for all tests. The data for the carbonate system parameters are shown as averages until day 17 (the
178 settling period of *M. balthica*). The graphs are based on actual $f\text{CO}_2$ values (presented in table 1). Data
179 are presented as means \pm SE.

180

181 **3 Results**

182 **3.1 Abiotic conditions in the mesocosms**

183 Water temperature varied from 8°C to 16°C during the experiment, following the natural conditions in
184 the bay. Salinity was on average 5.7 and total alkalinity on average $1550 \text{ mmol kg}^{-1}$ at the closing of the
185 mesocosms. Both parameters remained fairly constant during the experiment in all mesocosms (Paul et
186 al. 2015, this issue). Initial pHT after closing of the mesocosms and before the CO_2 -manipulations was
187 ca. 8.2 in the mesocosms and the bay. Average pHT levels and other parameters of each mesocosm over
188 the course of the experiment are shown in table 1.

189

190 **3.2 Larval abundance**

191 After the closing of the mesocosms (day -3 to -2), some unexplained variation was found in the
192 abundance of bivalve larvae (Fig. 1). On day 0, however, the abundances in the water column were
193 relatively similar within the mesocosms ($5522\text{-}5936 \text{ ind. m}^{-3}$), except in the $319 \mu\text{atm}$ ambient
194 mesocosm. This is likely due to a sampling issue or an artifact caused by a mesocosm maintenance
195 method (bubbling to destroy the halocline on day -3). During the first week after the CO_2 -manipulation,
196 by day 10, the larval abundance had decreased strongest in the two ambient mesocosms, with $> 80\%$

197 decrease in abundance in comparison to the 35-50 % decrease in the two highest $f\text{CO}_2$ mesocosms
198 ($>1000 \mu\text{atm}$) (Spearman $r = -0.83$, $p < 0.05$). Consequently, on day 10 the highest abundance was
199 measured in the highest $f\text{CO}_2$ mesocosm (3194 ind. m^{-3}) and the lowest abundances in both ambient
200 mesocosms ($319\text{-}321 \mu\text{atm}$) ($545 \text{ resp. } 1064 \text{ ind. m}^{-3}$). A strong decrease in abundance ($> 85 \%$)
201 occurred a week later (day 10 to 17) in all the high, $>400 f\text{CO}_2$, mesocosms, with up to a 93% decrease
202 found in the $1347 \mu\text{atm}$ mesocosm (Spearman $r = 0.94$, $p < 0.05$). From day 17 onwards, the abundances
203 were low in all of the mesocosms (Fig. 1).

204

205 **3.3 The abundance of settled individuals**

206 The abundances of settled individuals differed significantly between mesocosms and sampling days of
207 the main settling period (days 9-17, chi-square $\chi^2 = 1168.588$, $df = 25$, $p < 0.001$). The graphical post-
208 hoc tests showed three distinct settling peaks of *M. balthica*. In the ambient and near-ambient (<500
209 μatm) $f\text{CO}_2$ mesocosms a large increase in the abundance of settled individuals was found between days
210 9-13, with 71 %, 74 % and 54 % of all the individuals having settled by day 13. In comparison, only 39
211 % and 47 % of the individuals had settled during that time period in the two highest ($1072\text{-}1347 \mu\text{atm}$)
212 $f\text{CO}_2$ mesocosms (Fig. 2a and b). In the 857 and $1072 \mu\text{atm}$ $f\text{CO}_2$ mesocosms, a smaller settling event
213 occurred on days 11-15 and in the highest $f\text{CO}_2$ mesocosm the settling peaked on day 17, where after the
214 settling soon ceased in all mesocosms. On average 6130 ± 240 individuals settled in the mesocosms
215 during the course of the experiment, with the exception of $1072 \mu\text{atm}$ $f\text{CO}_2$ mesocosm where only ca.
216 4850 individuals settled (Fig. 2b).

217

218 **3.4 Larval sizes in the water column**

219 On day 0, larval size in the water column was on average $287 \pm 23 \mu\text{m}$ with no difference found
220 between the mesocosms. After 10 days of exposure to different $f\text{CO}_2$ levels, the average size of the
221 larvae in the water column (0-17 m) varied from $286 \mu\text{m}$ to $313 \mu\text{m}$, increasing significantly along the
222 increasing $f\text{CO}_2$ gradient ($R^2 = 0.78$, $F = 14.47$, $p = 0.019$, Fig 3) with ca. 10 % larger larvae still in the
223 water column in the two highest $f\text{CO}_2$ mesocosms (1072 and $1347 \mu\text{atm}$).

224

225 3.5 The sizes of settled individuals

226 On average > 80% of the individuals settled in the mesocosms during days 11 to 17. No significant
227 differences were found in the sizes of the settled individuals in the different $f\text{CO}_2$ levels at any of these
228 investigated time points (Fig. 4). On days 11 and 13 the average size within the mesocosms varied
229 between 285 μm to 303 μm , and on days 15 and 17 the average size varied between 293 μm to 317 μm .

230

231 4 Discussion

232 In this study we investigated the effects of different future CO_2 scenarios on the larval survival, growth
233 and settling of a Baltic Sea benthic key species *M. balthica* in a large-scale mesocosm setting. We found
234 that *M. balthica* settled later along the increasing $f\text{CO}_2$ gradient of the mesocosms. Moreover, an
235 indication that *M. balthica* larvae settled at a larger size in the high $f\text{CO}_2$ treatments was also observed,
236 possibly indicating that at increasing $f\text{CO}_2$ a sufficient mass for settling is not reached until a larger shell
237 length has been attained.

238

239 During the week after first CO_2 manipulation (day 3 to day 10) settling of *M. balthica* occurred faster in
240 the ambient and middle $f\text{CO}_2$ mesocosms (319 to 469 μatm) than in the higher $f\text{CO}_2$ mesocosms.
241 Consequently, the main settling peak occurred ca. 6 days earlier in these mesocosms (<500 μatm).
242 When comparing the sizes of the larvae, we found that the ones remaining in the water column on day
243 10 had an average size of 290 μm in both ambient mesocosms, whereas in the other mesocosms ($f\text{CO}_2$
244 >400 μatm), the sizes of the remaining larvae were 300-315 μm . We hypothesise that in the ambient
245 $f\text{CO}_2$ the bivalves settled at the expected size (<300 μm), and thus only the smaller larvae remained in
246 the upper water column when the settling was reaching its peak. In the high $f\text{CO}_2$ treatments the
247 development of the *M. balthica* larvae might have been compromised and/or delayed as on day 10,
248 despite being relatively large (>300 μm), a large part of the bivalves remained in the upper water
249 column without initiating settlement.

250

251 The observed inconsistency between the growth and settling of the early-stage bivalves can be explained
252 by proximate factors that regulate settling. For successful metamorphosis and settling from the

253 planktonic phase to the benthos, the individuals need to reach a sufficient size or weight and
254 developmental stage, including increased mobility/appearance of the foot (Caddy, 1969; Drent, 2002).
255 Shell growth alone, the growth measure used in our experiment as in many other studies, does not
256 automatically reflect the overall biomass production and developmental stage of the organism (Lewis
257 and Cerrato, 1997; Wood et al., 2008). In undersaturated conditions, calcification of the shell might be
258 compromised so that even though shell length reaches its typical size for settling, shell thickness is
259 reduced. This could be a factor that restricts the gaining of necessary mass to settle to the sea floor
260 (Waldbusser et al., 2010). During the entire experiment, undersaturation with respect to aragonite
261 occurred in all mesocosms apart from the two ambient mesocosms, and the three highest $f\text{CO}_2$
262 treatments were also undersaturated with respect to calcite (Table 1). It is also likely that at decreased
263 pHT levels shell growth was occurring at the cost of tissue development and biomass increase.
264 Unfortunately we were not able to measure soft tissue weight of collected larvae due to the very small
265 size. Larvae that stay longer in the water column, e.g. due to slower growth or delayed development,
266 face a higher risk of predation. The population dynamics of a bivalve species is largely dependent on
267 successful settlement and recruitment of the post-larvae, and dispersal of larval and post-larval stages
268 (Pedersen et al., 2008; Pineda et al., 2009; Valanko et al., 2010). As larval mortality of planktonic
269 invertebrates is also generally high (yet variable; estimates range from 3–23% daily), mainly due to
270 predation and environmental factors (Pineda et al., 2009), a reduced survival of the early-life stages, as
271 found in the present study, is alarming. As the key species of the soft-bottom ecosystems of the Baltic
272 Sea, *M. balthica* is an essential contributor to the overall health and functioning of the benthic
273 ecosystem. Future CO_2 -mediated changes to this species' population size might thus affect the diversity
274 and ecosystem functioning of the area.

275
276 Some other important factors that impact the settlement process, but cannot be mimicked in this
277 mesocosm setup include, e.g., sediment type and quality, cues from adult conspecifics and water
278 movements that can prevent or facilitate the settlement process (Woodin et al., 1986). Some limits to
279 ecosystem realism also arise from the exclusion of factors such as currents and large predators, which
280 impact the natural succession and dispersion patterns of the species. To understand complex, system-

281 wide responses that take into account ecological processes such as competition, predation and the effect
282 of/on different trophic levels, several species interactions need to be tested simultaneously. The
283 interactions between factors such as increasing CO₂ and predation is a topic for future studies, but it is
284 likely that individuals stressed by high CO₂ also would suffer higher predation rates.

285

286 In a previous aquarium experiment conducted with newly hatched larvae (ca. 150 µm) from the same
287 bay (Jansson et al., 2013), both the growth and survival of the larvae were found to be negatively
288 impacted by decreasing pH. In this mesocosm experiment, however, survival was not found to be
289 affected, and it was not possible to study growth in the same level of detail as in a laboratory
290 experiment. Other typical consequences of pH decrease found in early-stage bivalves are e.g. delayed
291 and/or abnormal development (Kurihara et al., 2008; Talmage and Gobler, 2010; Crim et al., 2011),
292 reduced calcification (Miller et al., 2009) and higher mortality (Talmage and Gobler, 2009; Crim et al.,
293 2011; van Colen et al., 2012). The settling of post-larvae to the seafloor may be impacted by the changes
294 in the water chemistry created by CO₂ increase (Green et al., 2004; Cigliano et al., 2010; Clements and
295 Hunt, 2014). The major part of ocean acidification research has been conducted by studying the
296 response of single species, with a few studies focusing on the interactions between a small number of
297 species, whereas studies on intact communities have so far only rarely been conducted (but see e.g.
298 work done at CO₂ vents by Hall-Spencer et al., 2008 or Kroeker et al., 2011 and previous/other
299 mesocosm studies by Christen et al., 2013; Riebesell et al., 2013b). For species such as *M. balthica*, a
300 mesocosm setting provides an excellent platform to study the development and succession of pelagic
301 early-life stages resulting in recruitment into the benthic system, which cannot be studied in a simple,
302 small-scale aquarium experiment. The direct and indirect factors that essentially impact the early life
303 success of a bivalve, e.g. natural food quality and quantity, can be incorporated in a mesocosm setting in
304 a more comprehensive way. In the case of future ocean acidification, potential changes in phytoplankton
305 dynamics due to increased CO₂ levels are likely to have consequences for the other trophic levels. The
306 growth of nanoplankton and diatom species (< 20 µm), which are the main food particles of larval
307 bivalves (Bos et al., 2006), has been shown to benefit from changing CO₂ conditions (e.g. Engel et al.,
308 2008; Feng et al., 2009; Meakin and Wyman, 2011; but see also e.g. Tortell et al., 2002), potentially

309 impacting the capacity of the larvae to survive in a changing environment via consequences in their
310 energy balance. In this study, no significant changes were detected in the phytoplankton abundance or
311 the total chlorophyll *a* concentration within the mesocosms during the main occurrence of *M. balthica*
312 larvae in the water column (until days 10 and 17). An increase in the abundance of phytoplankton and
313 Chl *a* concentration in the highest $f\text{CO}_2$ mesocosms was, however, found later on during the experiment
314 (day 16 onwards; Crawford et al., 2015; Paul et al., 2015). By the time the differences in phytoplankton
315 abundance started emerging, most of the *M. balthica* larvae had already settled from the water column.

316

317 The Baltic Sea is a unique system to study future ocean acidification. Large pH fluctuations that already
318 occur seasonally in the northern Baltic Sea in the shallow coastal areas, primarily due to changes in
319 productivity (Thomas and Schneider, 1999; Schneider et al., 2003), result in high pH values of up to 8.4
320 during daytime and low pH values such as 7.4 during respiration at night (pers. obs.). For areas such as
321 this, accurate modelling of the future pH change is generally challenging. Yet, future ocean acidification
322 is predicted to permanently decrease the pH and thus shift the pH range the organisms are exposed to
323 towards lower values (Omstedt et al., 2010). In our study we found negative effects of increasing CO_2
324 levels on the settling and early development of *M. balthica*. The impact on the success of these early-
325 stage bivalves is alarming as these stages are crucial for sustaining viable populations. A failure in their
326 recruitment would ultimately lead to negative effects on the population, and considering the key role *M.*
327 *balthica* has in the Baltic Sea, also for the functioning and resilience of the benthic ecosystem.

328

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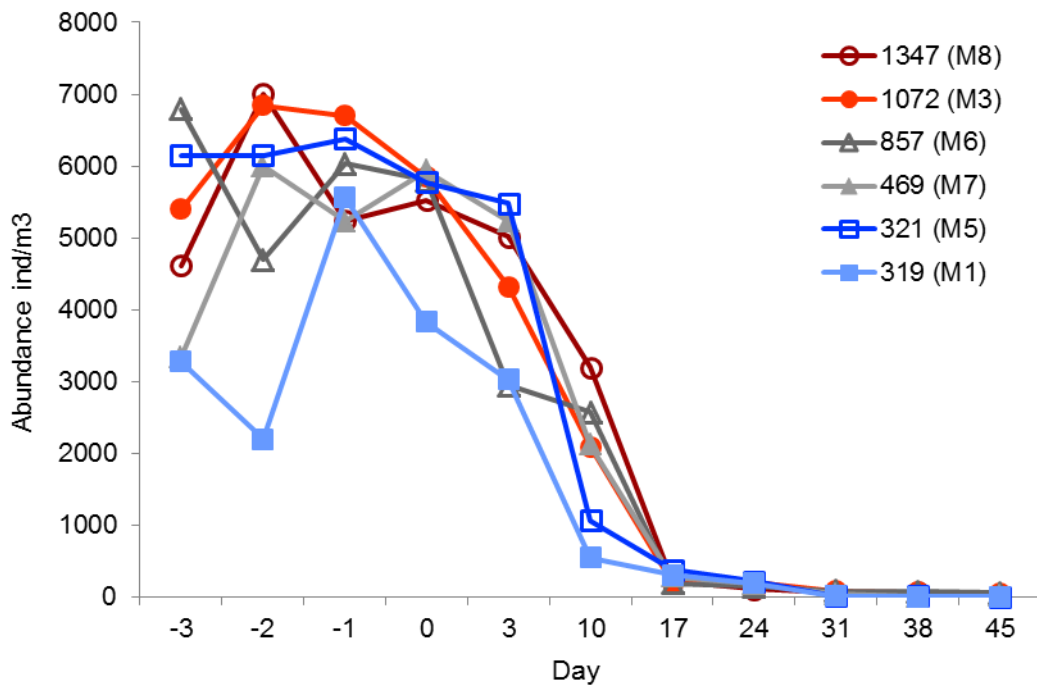
595

596 Table 1. Carbonate system parameters in the mesocosms during the experiment (average values on days
 597 0-17, the main settling period of *M. balthica*).

	M1	M5	M7	M6	M3	M8	Bay
Target $f\text{CO}_2$ (μatm)	ambient/control	ambient/control	600	950	1300	1650	ambient
$f\text{CO}_2$ (μatm)	319	321	469	857	1072	1347	282
pHT	7.94	7.94	7.80	7.59	7.51	7.43	7.99
Ω aragonite	1.07	1.06	0.77	0.47	0.39	0.33	1.19
Ω calcite	1.92	1.91	1.39	0.84	0.71	0.59	2.14

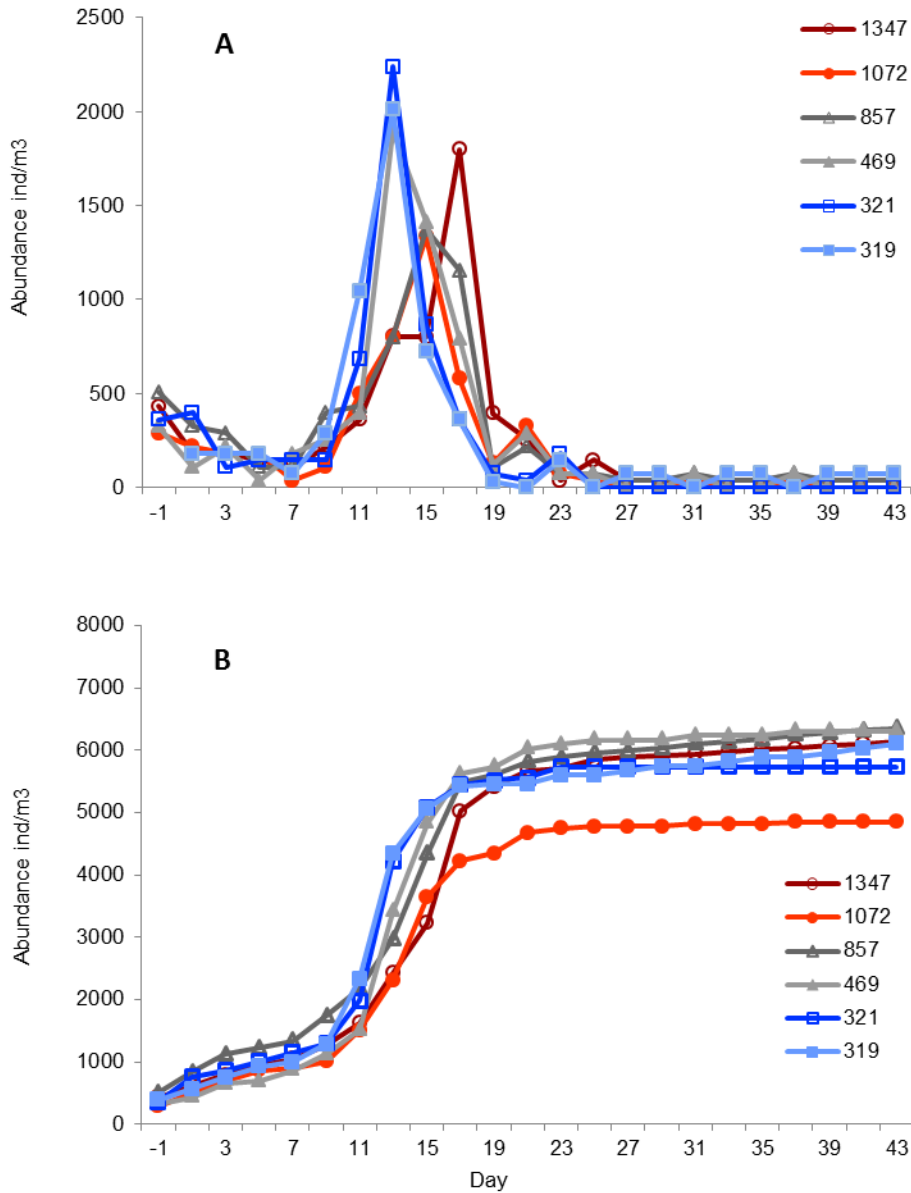
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601 Figure1. Larval abundance in the water column of the individual mesocosms over time.

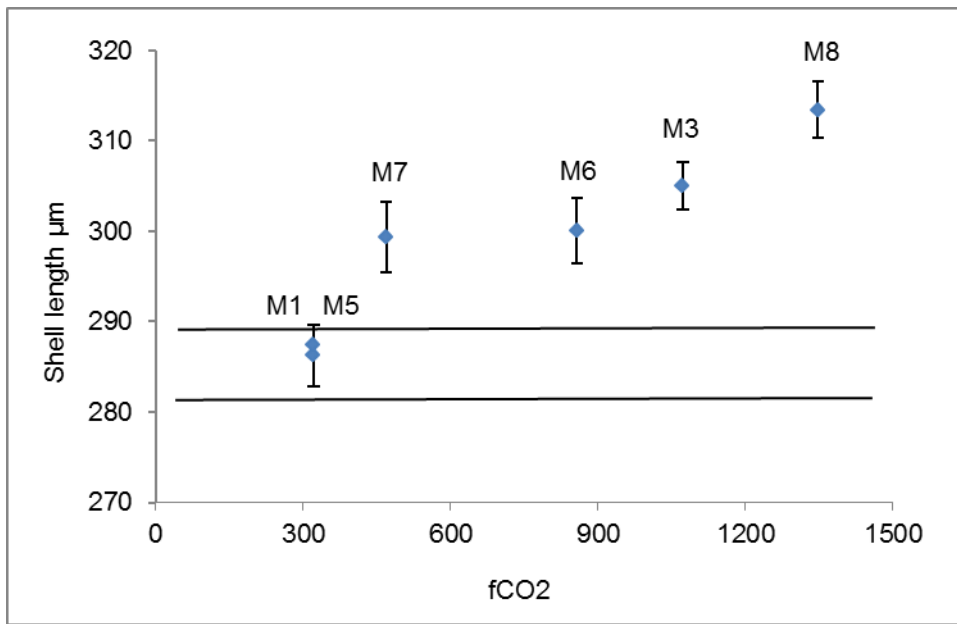


602

603 Figure 2. A. The abundance of settled individuals per cubic meter water mass enclosed in the different

604 mesocosms over the course of the experiment. B. The cumulative abundance of settled *M. balthica* per

605 cubic meter of individual mesocosm volume.

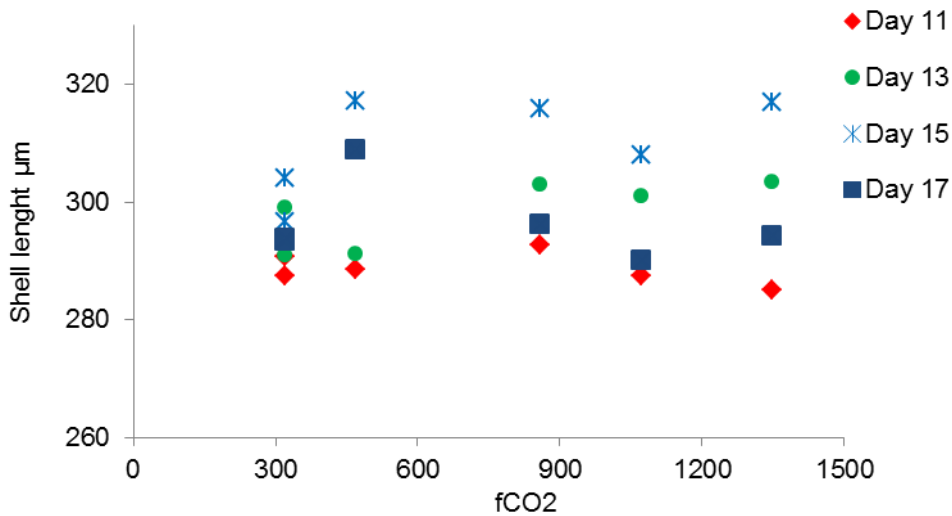


606

607 Figure 3. Larval sizes in different $f\text{CO}_2$ levels at day 10. Data is presented as means \pm SE, $n = \text{ca. } 70$

608 individuals. The horizontal lines indicate the range of average larval sizes on day 0.

609



610

611 Figure 4. Sizes of the settled individuals exposed to different $f\text{CO}_2$ levels on days 11, 13, 15 and 17.

612 Data is presented as means, $n = \text{ca. } 35$ at each data point. For clarity, SE are not shown.

613