

Revision of Global soil nitrous oxide emissions in a dynamic carbon-nitrogen model

Dear Soenke,

Thank you very much for your competent handling of our manuscript and the opportunity to publish our work in Biogeosciences.

We have taken care of the suggested corrections, as follows

p6 l28 Te-> The

Done

p7 line 29: Please change Zaehle & Friend 2010 to Smith et al. 2014 BGS. I actually do not use a transpiration relationship in a transient sense.

Done

p10 l 6: generate -> generates

Corrected

p23 l17ff. If is unclear here whether you talk about the LM3V modelled soil moisture or the "data-derived" models. please be very explicit here.

We changed the sentence as follows:

“However, despite using data-derived soil moisture, it appears that the prediction of soil moisture is an impediment towards validating N₂O emissions at field scale *for both LM3V-N simulated and reanalysed soil moisture.*”

p25 l8 include references here

We added the following references: Xu-Ri et al., 2012; Zaehle et al., 2011

p25 l13: provide reference or explain briefly why the BNF in models requires improvement (please no exhaustive discussion!)

We changed this part of the paragraph as follows

The representation of BNF in models requires improvement: currently, simple empirical relationships are used, yet BNF is the largest source of new N in terrestrial systems, and therefore is critical in the determination of N availability to nitrifiers and denitrifiers. Here we showed that different implementations of BNF (prescribed vs. responsive to plant N demand) are globally important for N₂O emissions.

p27 l23 opposite -> of opposite sign?

Corrected

p27123ff “,while for climate it is the combination of temperature and interaction”. Sorry I don’t understand this.

We rephrased to:

“On the other hand, simulations with O-CN (Zaehle et al., 2011) showed the effects of CO₂ and climate to be approximately equal and of opposite sign for historic simulations covering the past 300 years that also include land-cover changes. They evaluated the effect of climate change as the difference between a simulation that consider both CO₂ and climate and a simulation that does not consider climate change. Thus their climate effect contains both the single effect of climate and the interaction of climate with CO₂. The temperature (i.e. climate) response on top of CO₂ can in our simulation be calculated as the temperature effect plus the interaction effect (Fig 8). Analyzed in this fashion, LM3V-N’s results are congruent with those of Zaehle et al. (2011), albeit we found a slightly weaker temperature effect compared to CO₂.”

p29 l 4 “distinct and new results ... is treated differently”. Please be more precise!

We are now more explicit:

“Our sensitivity analysis shows that processes of the larger plant-soil N cycle affect fast N cycle processes as evidenced by the response to the fire and BNF modification. This sensitivity can lead to differences in N₂O across models (e.g. in the response to CO₂ and climate) even if existing nitrification-denitrification schemes are identical.”

p29 l 5. a response of N₂O, or the N cycle in general. And - what does this imply...?

We added the central implication of the rarity of tropical manipulative experiments.

“Further, our work suggests a much stronger response to warming and CO₂ in tropical forests compared to extratropical forest, thus extrapolation of mostly extra-tropical field studies to the globe warrants caution.”

check spelling and formatting (N₂O etc).

We did further editorial changes to eliminate spelling, grammatical, and formatting error. Please see the attached marked-up version.

Best regards,
Stefan Gerber and Yuanyuan Huang

1 Global soil nitrous oxide emissions in a dynamic carbon- 2 nitrogen model

3 Y. Y. Huang¹ and S. Gerber¹

4 [1] {Soil and Water Science Department, Institute of Food and Agricultural Sciences,
5 University of Florida, Gainesville, Florida 32611}

6 Correspondence to: S. Gerber (sgerber@ufl.edu)

7 8 Abstract

9 Nitrous oxide (N₂O) is an important greenhouse gas that also contributes to the depletion of
10 stratospheric ozone. Due to its high temporal and spatial heterogeneity, a quantitative
11 understanding of terrestrial N₂O emission, its variabilities and responses to climate change is
12 challenging. We added a soil N₂O emission module to the dynamic global land model LM3V-
13 N, and tested its sensitivity to mechanisms that affect the level of mineral [nitrogen \(N\)](#) in soil
14 such as plant N uptake, biological N fixation, amount of volatilized N redeposited after fire, and
15 nitrification-denitrification. We further tested the relationship between N₂O emission and soil
16 moisture, and assessed responses to elevated CO₂ and temperature. Results extracted from the
17 corresponding gridcell (without site-specific forcing data) were comparable with the average
18 of cross-site observed annual mean emissions, although differences remained across individual
19 sites if stand-level measurements were representative of gridcell emissions. Processes, such as
20 plant N uptake and N loss through fire volatilization that regulate N availability for nitrification-
21 denitrification have strong controls on N₂O fluxes in addition to the parameterization of N₂O
22 loss through nitrification and denitrification. Modelled N₂O fluxes were highly sensitive to
23 water filled pore space (WFPS), with a global sensitivity of approximately 0.25 TgN per year
24 per 0.01 change in WFPS. We found that the global response of N₂O emission to CO₂
25 fertilization was largely determined by the response of tropical emissions with reduced N₂O
26 fluxes in the first few decades and increases afterwards. The initial reduction was linked to N
27 limitation under higher CO₂ level, and was alleviated through feedbacks such as biological N
28 fixation. The extratropical response was weaker and generally positive, highlighting the need
29 to expand field studies in tropical ecosystems. We did not find synergistic effects
30 ~~between~~[between](#) warming and CO₂ increase as reported in analyses with different models.

1 Warming generally enhanced N₂O efflux and the enhancement was greatly dampened when
2 combined with elevated CO₂, although CO₂ alone had a small effect. The differential response
3 in the tropics compared to extratropics with respect to magnitude and sign suggests caution
4 when extrapolation from current field CO₂ enrichment and warming studies to the [global](#)
5 [scale](#).

6

7 **1 Introduction**

8 Nitrous oxide (N₂O) is a major reactant in depleting stratospheric ozone as well as an important
9 greenhouse gas (Ravishankara et al., 2009; Butterbach-Bahl et al., 2013; Ciais et al., 2013). With
10 a global warming potential of 298 times more (per unit mass) than that of carbon dioxide (CO₂)
11 over a 100-year period (Forster et al., 2007), the contributions of N₂O emissions to global
12 radiative forcing and climate change are of critical concern (Zaehle and Dalmonech, 2011). The
13 concentration of atmospheric N₂O has been increasing considerably since the industrial
14 revolution with a linear rate of 0.73 ± 0.03 ppb yr⁻¹ over the last three decades (Ciais et al., 2013).
15 Although applications of synthetic fertilizer and manure during agriculture intensification have
16 been identified as the major causes of this increase (Davidson, 2009; Zaehle and Dalmonech,
17 2011; Zaehle et al., 2011), nonagricultural (natural) soil is still an important N₂O source (Ciais
18 et al., 2013; Syakila and Kroeze, 2011). N₂O fluxes from nonagricultural soils are highly
19 heterogeneous, which limits our ability to estimate and predict global scale [budgets](#), and
20 quantify ~~its~~ [the](#) response [of natural N₂O fluxes](#) to global environmental changes (Butterbach-
21 Bahl et al., 2013; Ciais et al., 2013).

22 Most of the N₂O fluxes from soil are produced by microbial nitrification and denitrification
23 (Braker and Conrad, 2011; Syakila and Kroeze, 2011). Nitrification is an aerobic process that
24 oxidizes ammonium (NH₄⁺) to nitrate (NO₃⁻), during which some N is lost as N₂O.
25 Denitrification reduces nitrate or nitrite to gaseous N (i.e. NO_x, N₂O and N₂), a process that is
26 fostered under anaerobic conditions. During denitrification N₂O is generated in intermediary
27 steps where a small portion can escape from [the](#) soil before further reduction to N₂ takes place.
28 Soil texture, soil NH₄⁺, soil water filled pore space (WFPS), mineralization rate, soil pH, and
29 soil temperature are well-known regulators of nitrification N₂O fluxes (Parton et al., 1996; Li et
30 al., 2000; Parton et al., 2001). Denitrification and associated N₂O emissions depend primarily
31 on carbon supply, the redox potential and soil NO₃⁻ (Firestone and Davidson, 1989; Parton et

1 al., 1996). Soil moisture has a particularly strong impact (Galloway et al., 2003;Schlesinger,
2 2009) as it influences nitrification and denitrification rates through its regulations on substrate
3 availability and soil redox potential (as oxygen diffusion proceeds at much slower rate in water
4 filled than in air filled pore space), thereby also controlling the partitioning among various
5 denitrification products (i.e. NO_x, N₂O and N₂) (Firestone and Davidson, 1989;Parton et al.,
6 2001). Although emissions are known to be sensitive to soil moisture, quantitative
7 understanding of its role in terrestrial N₂O fluxes and variability is limited (Ciais et al., 2013).

8 At regional to global scale, the application of the “hole-in-pipe” concept (Firestone and
9 Davidson, 1989) in the CASA biosphere model pioneered one of the earliest process-based
10 estimation of natural soil N₂O fluxes. The model calculated the sum of NO, N₂O and N₂ fluxes
11 as a constant portion of gross mineralized N, and the relative ratios of N trace gases
12 (NO_x:N₂O:N₂) as a function of soil moisture (Potter et al., 1996). While the early models of
13 nitrification and denitrification are primarily conceptual driven, recent global N₂O models
14 combine advancements in global dynamic land models with more detailed processes, including
15 microbial dynamics. Xu-Ri and Prentice (2008) simplified nitrification and denitrification
16 modules from DNDC (i.e., DeNitrification-DeComposition) (Li et al., 1992;Li et al., 2000) in
17 their global scale dynamic N scheme (DyN) and incorporated DyN into the LPJ dynamic global
18 vegetation model. In the DNDC approach, nitrification and denitrification were allowed to
19 occur simultaneously in aerobic and anaerobic microsites. Zaehle et al. (2011) incorporated a
20 nitrification-denitrification scheme into the O-CN land model following largely the LPJ-DyN
21 with minor modifications and additions of the effects of soil pH and chemo-denitrification that
22 originated from DNDC (Li et al., 2000). Stocker et al. (2013) embedded the LPJ-DyN approach
23 into an Earth System Model and investigated the feedbacks of N₂O emissions, together with
24 CO₂ and CH₄, to climate. Compared to LPJ-DyN approach, Saikawa et al. (2013) retained the
25 explicit simulation of nitrifying and denitrifying bacteria from DNDC in their CLMCN-N₂O
26 module based on CLM V3.5 land model. Simulations with O-CN demonstrated a positive
27 response of N₂O emissions to historical warming and a negative response to historical CO₂
28 increase, globally. While CO₂ and interaction with climate change resulted in an increase in
29 historical and future N₂O emissions in LPJ-DyN (Xu-Ri et al., 2012) and its application in LPX-
30 Bern (Stocker et al., 2013), respectively, historical CO₂ change alone, i.e. single factor of Xu-
31 Ri et al. (2012), caused a slight decrease in historical N₂O emissions. The negative CO₂
32 response seems to be in disagreement with one meta-analysis of manipulative field experiments

1 showing an increase in N₂O emissions at elevated levels of CO₂ (Zaehle et al., 2011; Xu-Ri et
2 al., 2012; van Groenigen et al., 2011). The discrepancy in response to global change factors
3 needs to be addressed both in models and in the interpretation of manipulative field experiments.
4

5 Here we add a N₂O gas emission module to LM3V-N, a land model developed at the Geophysical
6 Fluid Dynamics Laboratory (GFDL). In this paper, we will first briefly introduce LM3V-N and
7 describe the added N₂O emission module. We then subject the model to historic changes in CO₂,
8 N deposition, and recent climate change to infer natural N₂O emissions in the past few decades.
9 We test the model's sensitivity to soil water regime, by addressing the parameterization of soil
10 WFPS, and by replacing the model soil moisture with two different soil moisture reanalysis
11 products. We also conduct sensitivity tests with regard to the general N cycling and
12 parameterization of N₂O emissions. We then subject the model to step changes in atmospheric
13 CO₂ and temperature to understand modelled responses to CO₂ fertilization ⁴ and climate change.
14 Since we build largely on existing parameterization of nitrification-denitrification processes,
15 we will briefly discuss implications from transferring process formulations to LM3V-N where
16 other aspects of the N cycle are treated differently.

17 **2 Methods**

18 **2.1 Model description**

19 LM3V is capable of simulating ecosystem dynamics and exchange of CO₂, water and energy
20 between land and atmosphere with the fastest time step of 30 minutes (Shevliakova et al., 2009).
21 LM3V-N expands the LM3V land model with a prognostic N cycle (Gerber et al., 2010), and
22 includes five plant functional types (PFTs): C3 and C4 grasses, tropical, temperate deciduous
23 and cold evergreen trees. Each PFT has five vegetation C pools (leaf, fine root, sapwood, labile,
24 and wood), two litter and two soil organic C pools and their corresponding N pools based on
25 the specific C:N ratios. Photosynthesis is coupled with stomatal conductance on the basis of the
26 Collatz et al.'s (1991, 1992) simplification of the Farquhar scheme (Farquhar et al., 1980). Soil
27 hydrology in LM3V-N follows partly on Land Dynamics (LaD) with further improvements
28 (Shevliakova et al., 2009; Milly and Shmakin, 2002; Milly et al., 2014). N enters the ecosystem
29 through atmospheric N deposition and biological N fixation (BNF), losses via fire and leaching
30 of dissolved organic N (DON) as well as mineral N. We briefly describe the major

1 characteristics of LM3V-N in the next subsection (2.1.1), and details are available in Gerber et
2 al. (2010).

3 **2.1.1 Main characteristic of LM3V-N**

4 **2.1.1.1 C-N coupling in vegetation**

5 We briefly describe the larger plant-soil N cycle and how it links to mineral N (ammonium and
6 nitrate). Plants adjust their uptake of C and N to maintain their tissue specific C:N ratios, which
7 are PFT-dependent constants. Instead of varying C:N ratios in tissues, short-term asynchronies
8 in C and N assimilations or temporary imbalances in stoichiometry are buffered by [an](#) additional
9 N storage pool (S) in which N is allowed to accumulate once plant N demand is satisfied. The
10 optimum storage size S_{target} is based on tissue turnover $Q_{N,liv}$,

$$11 \quad S_{target} = t_h Q_{N,liv} \quad (1)$$

12 where t_h is the time span that buffer plant N losses (currently set as 1 year). Plant N status (x)
13 is defined as the fraction of the actual N storage compared to the target storage: $x = S/S_{target}$.
14 Consequently, N constraints on photosynthesis and soil N assimilation are based on plant N
15 status:

$$16 \quad A_{g,N} = A_{g,pot} (1 - e^{-x\varphi}) \quad (2)$$

$$17 \quad U_{N,P} = U_{N,P,pot} * \begin{cases} 1 & \text{if } S < S_{target} \\ 0 & \text{else} \end{cases} \quad (3)$$

18 where $A_{g,N}$ indicates N constrained rate of gross photosynthesis ($\mu\text{molC m}^{-2} \text{s}^{-1}$) and $A_{g,pot}$
19 corresponds to the potential photosynthetic rate without N limitation. The parameter φ mimics
20 the metabolic deficiency as plant N decreases. $U_{N,P,pot}$ is the potential inorganic N uptake rate
21 from soil available ammonium and nitrate pools. The actual inorganic N uptake rate ($U_{N,P}$)
22 operates at its potential if plants are N limited and drops to zero when N storage (S) reaches its
23 target size. Overall this set-up intends to overcome short-term asynchronies between C and N
24 supply.

25 **2.1.1.2 Soil C-N interactions in organic matter decomposition**

26 Organic matter decomposition is based on a modified CENTURY approach (Bolker et al.,
27 1998), and amended with formulations of N dependent C and N mineralization rates. Here, we
28 use a 3 pool model where the pools broadly represent labile and structural litter, and processed

1 soil organic matter. Decomposition is the main source of available N for nitrification and
 2 denitrification. In turn, NO_3^- and NH_4^+ can both trigger the decomposition of “light” organic
 3 matter and stabilize C in “heavy” organic matter in LM3V-N. Formation of a slow
 4 decomposable organic matter pool leads to immobilization of ammonium and nitrate to satisfy
 5 the fixed carbon to nitrogen ratio of this pool.

6 **2.1.1.3 Competing sinks of available N**

7 The fate of soil mineral N (i.e. ammonium and nitrate) depends on the relative strength of the
 8 competing sinks, with the broad hierarchy of sorption > soil immobilization > plant uptake >
 9 leaching/denitrification. This creates a tight N cycle, since internal (plant and soil) sinks
 10 dominate over N losses. Denitrification thus far has been lumped with leaching losses and
 11 summed into a generic N loss term. Sorption/desorption buffers available N and is assumed to
 12 have the highest priority and be at steady state in each model time step. N immobilization into
 13 organic matter occurs during transfers among litter and soil organic matter pools. Leaching
 14 losses of available N are simulated on the basis of drainage rate. Plant uptake of mineral N is a
 15 combination of both active and passive processes. The active uptake is modeled as a Monod
 16 function, and the passive transport is a function of available N and plant transpiration:

$$17 \quad U_{N,P,pot,i} = \frac{v_{max}C_r N_{i,av}}{h_s(k_{p,1/2} + [N_{av}])} + [N_{i,av}]Q_{W,T} \quad (4)$$

18 where v_{max} ($\text{yr}^{-1} \text{kgC}^{-1}$) stands for the maximum uptake rate per unit root mass C_r , h_s is soil depth,
 19 $k_{p,1/2}$ is the half saturation constant, and $Q_{W,T}$ represents the transpiration flux of water. ~~The~~
 20 subscript j refers to either ammonium or nitrate, while $[N_{av}]$ is the concentration of the combined
 21 dissolved ammonium nitrate pool. Potential uptake and thus effective removal of available N
 22 occurs if plants are N limited (see Equation 3).

23 **2.1.1.4 N losses from organic pools**

24 With the implementation of high ecosystem N retention under limiting condition where internal
 25 N sinks outcompeting losses from the ammonium/nitrate pools, losses via organic pathways
 26 become important (Gerber et al., 2010; Thomas et al., 2015). Over the long term, N losses via
 27 fire and DON are thus critical factors limiting ecosystem N accumulation and maintaining N
 28 limitation in LM3V-N. N volatilized via fire is approximated as a function of CO_2 produced in
 29 a fire, stoichiometric ratio of burned tissues but reduced by a global retention factor representing
 30 the fraction of N that is retained as ash (*ash_fraction*, currently set as 0.45). DON leaching is

Formatted: Font: Italic

1 linked to hydrologic losses of dissolved organic matter (L_{DOM}) and its C:N ratio. In turn L_{DOM}
2 is based on drainage rate ($Q_{w,D}$) and a buffer or sorption parameter b_{DOM} (currently set as 20).

$$3 \quad L_{DOM} = \frac{Q_{w,D}}{h_s b_{DOM}} DOM \quad (5)$$

4 where DOM is the amount of dissolve organic matter in the soil column. Soil depth (h_s) is used
5 to convert DOM unit to concentration (in unit of kgC m^{-3}). Production of DOM (in unit of kgC
6 m^{-2}) is assumed to be proportional to the decomposition flux of the structural litter and soil
7 water content. Both, losses via fire and via DOM are losses from a plant-unavailable pool
8 (Thomas et al., 2015), and have the potential to increase or maintain N limitation over longer
9 timescales, and consequently reduce N availability for N_2O production through sustained and
10 strong plant N uptake.

11 **2.1.1.5 Biological nitrogen fixation (BNF)**

12 BNF in LM3V-N is dynamically simulated on the basis of plant N availability, N demand, and
13 light condition. BNF increases if plant N requirements are not met by uptake. The rate of up-
14 regulation is swift for tropical trees but constrained by light penetrating the canopy for other
15 PFTs, mimicking the higher light requirements for new recruits that possibly can convert
16 atmospheric N_2 into plant available forms. In turn, sufficient N uptake reduces BNF. The BNF
17 parameterization thus creates a negative feedback, where high plant available N and thus the
18 potential for denitrification is counteracted with reduction of N input into the plant-soil system.
19 This explicit negative feedback is different to other models where BNF is parameterized based
20 on NPP (Thornton et al., 2007), or transpiration (~~Zachle and Friend, 2010~~; [Smith et al., 2014](#)).
21 The inclusion of BNF as a negative feedback contributes to a rather tight cycling within LM3V-
22 N, with low overall rates of BNF under unperturbed conditions (Gerber et al., 2013).

23 **2.1.2 Soil N_2O emission**

24 LM3V-N assumes that nitrification is linearly scaled to ammonium content, and modified by
25 soil temperature and soil moisture. Gaseous losses so far were not differentiated from
26 hydrological leaching. We add a soil nitrification-denitrification module which accounts for N
27 gaseous losses from NH_3 volatilization, nitrification and denitrification. The nitrification-
28 denitrification scheme implemented here combines features from both the DNDC model (Li et
29 al., 1992; Li et al., 2000) and the CENTURY/DAYCENT [model](#) (Parton et al., 1996; Parton et
30 al., 2001; Del Grosso et al., 2000). In this subsection, we provide details on the nitrification-

1 denitrification module which explicitly simulates N ~~gaseous gas~~ losses ~~from~~ via nitrification and
 2 denitrification, as well as other process modifications compared to the original LM3V-N.

3 2.1.2.1 Nitrification-Denitrification

4 Transformation among mineral N species (ammonium and nitrate) occurs mainly through two
 5 microbial pathways: nitrification and denitrification. Although ongoing debate exists in whether
 6 nitrification rates may be well described by bulk soil ammonium concentration or soil N
 7 turnover rate (Parton et al., 1996; Zaehle and Dalmonech, 2011), we adopt the donor controlled
 8 scheme (ammonium concentration). In addition to substrate, soil texture, soil water filled pore
 9 space (WFPS, the fraction of soil pore space filled with water), and soil temperature are all well
 10 known regulators of nitrification. As a first order approximation, nitrification rate (N , in unit,
 11 $\text{kgN m}^{-2} \text{ year}^{-1}$) is simulated as a function of soil temperature, NH_4^+ availability and WFPS,

$$12 \quad N = k_n f_n(T) f_n(WFPS) \frac{N_{\text{NH}_4^+}}{b_{N,\text{NH}_4^+}} \quad (6)$$

13 where k_n is the base nitrification rate (11000 year^{-1} , the same as in LM3V-N) (Gerber et al.,
 14 2010); $N_{\text{NH}_4^+}$ is ammonium content (in unit, kgN m^{-2}); b_{N,NH_4^+} is the buffer or sorption
 15 parameter for NH_4^+ (unitless, 10 in LM3V-N) (Gerber et al., 2010); $f_n(T)$ is the temperature
 16 response function following Li et al. (2000), with an optimum temperature for nitrification at
 17 35°C ; and $f_n(WFPS)$ is the soil water response function. The effect of WFPS on nitrification is
 18 texture dependent, with most of the reported optimum value around 0.6 (Parton et al., 1996; Linn
 19 and Doran, 1984). We adopt the empirical WFPS response function from Parton et al. (1996)
 20 with medium soil texture.

$$21 \quad f_n(T) = \left(\frac{60 - T_{\text{soil}}}{25.78}\right)^{3.503} \times e^{\frac{3.503 \times (T_{\text{soil}} - 34.22)}{25.78}} \quad (7)$$

$$22 \quad f_n(WFPS) = \left(\frac{WFPS - 1.27}{-0.67}\right)^{\frac{1.9028}{0.59988}} \times \left(\frac{WFPS - 0.0012}{0.59988}\right)^{2.84} \quad (8)$$

23 where T_{soil} is the soil temperature in degree Celsius.

24 Denitrification is controlled by substrate NO_3^- (electron acceptor), labile C availability (electron
 25 donor), soil moisture and temperature. Labile C availability is estimated by soil heterotrophic
 26 respiration (HR). Following LPJ-DyN (Xu-Ri and Prentice, 2008), denitrification is assumed
 27 to have a Q_{10} value of 2 when the soil temperature is between 15 and 25°C . The soil moisture
 28 response function is adopted from Parton et al. (1996). Soil pH is reported to be an important

1 indicator of chemodenitrification which occurs predominantly in acidic soils (pH<5) under
 2 conditions of high nitrite concentration (Li et al., 2000). However, its role for N₂O production
 3 is not well studied (Li et al., 2000) and we do not model chemodenitrification explicitly.

$$4 \quad D = k_d f_d(T) f_d(WFPS) f_g NO_3^- \quad (9)$$

$$5 \quad \text{And } f_g = \frac{HR}{HR + K_c} \frac{NO_3^-}{NO_3^- + K_n} \quad (10)$$

$$6 \quad NO_3^- = \frac{N_{NO_3^-}}{b_{NO_3^-}} \quad (11)$$

7 where D is the denitrification rate (in unit, kgN m⁻² year⁻¹); k_d is the base denitrification rate
 8 (8750 year⁻¹); f_g mimics the impact of labile C availability and substrate (nitrate) on the growth
 9 of denitrifiers, adapted from Li et al. (2000); K_c and K_n are half-saturation constants taken from
 10 Li et al. (2000) (0.0017 and 0.0083 kgN m⁻² respectively, assuming an effective soil depth of
 11 0.1m); $b_{NO_3^-}$ is the buffer or sorption parameter for NO₃⁻ (unitless, 1 in LM3V-N) (Gerber et
 12 al., 2010); $N_{NO_3^-}$ and NO_3^- are nitrate content before and after being buffered (in unit, kgN m⁻²),
 13 respectively; and $f_d(T)$ and $f_d(WFPS)$ are empirical soil temperature and water reponse
 14 function for denitrification, adopted from Xu-Ri and Prentice (2008) and Parton et al. (1996),
 15 respectively.

$$16 \quad f_d(T) = e^{308.56 \times \left(\frac{1}{68.02} - \frac{1}{T_{soil} + 46.02} \right)} \quad (12)$$

$$17 \quad f_d(WFPS) = \frac{1.56}{12.0 \left(\frac{16.0}{12.0(2.01 \times WFPS)} \right)} \quad (13)$$

18 2.1.2.2 Gaseous partitions from nitrification-denitrification

19 N₂O is released as a byproduct during both nitrification and denitrification. The fraction of N₂O
 20 lost during net nitrification is uncertain (Li et al., 2000; Xu-Ri and Prentice, 2008). Here we set
 21 this fraction to be 0.4%, which is higher than Goodroad and Keeney (1984), but at the low end
 22 provided by Khalil et al. (2004). Nitrification also ~~generate~~generates NO_x gas, in addition to
 23 N₂O. N losses as NO_x emissions during nitrification are scaled to the -N₂O release using a
 24 variable NO_x:N₂O ratio ($R_{NO_x:N_2O}$). $R_{NO_x:N_2O}$ varies with relative gas diffusivity (D_r , the relative
 25 gas diffusivity in soil compared to air) (Parton et al., 2001), which is calculated from air filled
 26 porosity ($AFPS$, i.e., the portion of soil pore space that is filled by air) (Davidson and Trumbore,
 27 1995)

Formatted: Subscript

$$1 \quad R_{NOx:N2O} = 15.2 + \frac{35.5 \times ATAN(0.68 \times \pi \times (10 \times D_r - 1.68))}{\pi} \quad (14)$$

$$2 \quad D_r = 0.209 \times AFPS^{\frac{4}{3}} \quad (15)$$

3 where ATAN stands for the trigonometric arctangent function; $AFPS$ is the air filled porosity
4 (1-WFPS), and π is the mathematical constant, approximately 3.14159.

5 During denitrification, the gaseous ratio between N_2 and N_2O ($R_{N2:N2O}$) is calculated following
6 the empirical function derived by Del Grosso et al. (2000), which combines the effects of
7 substrate (NO_3^-) to electron donor (HR , the proxy for labile C) ratio and WFPS. $R_{N2:N2O}$ is
8 updated at every time step and for each grid cell.

$$9 \quad R_{N2:N2O} = Fr\left(\frac{NO_3^-}{HR}\right) \cdot Fr(WFPS) \quad (16)$$

10 With

$$11 \quad Fr\left(\frac{NO_3^-}{HR}\right) = \max(0.16 \times k, k \times e^{(-0.8 \times \frac{NO_3^-}{HR})}) \quad (17)$$

$$12 \quad Fr(WFPS) = \max(0.1, 0.015 \times WFPS - 0.32) \quad (18)$$

13 where k is a texture dependent parameter (Table 1) estimated from Del Grosso et al. (2000). k
14 controls the maximum value of the function $Fr\left(\frac{NO_3^-}{HR}\right)$.

15 **2.1.2.3 Other modified processes**

16 To complete the N loss scheme in LM3V-N, we also added NH_3 volatilization into LM3V-N.
17 NH_3 volatilization in soil results from the difference between the equilibrium NH_3 partial
18 pressure in soil solution and that in the air. Dissolved NH_3 is regulated by ammonium
19 concentration and pH. The net flux of NH_3 from soil to the atmosphere varies with soil NH_3 ,
20 moisture, temperature, therefore

$$21 \quad NH_3 = k_{nh} f(pH) f_{NH_3}(T) (1 - WFPS) \frac{N_{NH_4^+}}{b_{N,NH_4^+}} \quad (19)$$

22 where NH_3 is the net ammonia volatilization flux (in unit, $kgN \ m^{-2} \ year^{-1}$); k_{nh} is the base
23 ammonia volatilization rate ($365 \ year^{-1}$); $f(pH)$ is the pH factor and $f(T)$ is the temperature factor
24 which are given by the following two equations:

$$25 \quad f(pH) = e^{2 \times (pH_{soil} - 10)} \quad (20)$$

$$26 \quad f_{NH_3}(T) = \min\left(1, e^{308.56 \times \left(\frac{1}{71.02} - \frac{1}{T_{soil} + 46.02}\right)}\right) \quad (21)$$

1 where pH_{soil} is the soil pH which is prescribed instead of simulated dynamically. $f(pH)$ and $f(T)$
2 follow largely on the NH_3 volatilization scheme implemented in the dynamic global vegetation
3 model LPJ-DyN (Xu-Ri and Prentice, 2008).

4 **2.2 Model experiments**

5 **2.2.1 Global hindcast with potential vegetation**

6 To understand the model performance and compare with other models and observations, we
7 conducted a hindcast simulation with potential vegetation. The model resolution was set to 3.75
8 degrees longitude by 2.5 degrees latitude. We forced the model with 3 hourly reanalysis weather
9 data based on Sheffield et al. (2006). We used a 17 year recycled climate of 1948-1964 for the
10 spin-up and simulation years prior to 1948. Atmospheric CO_2 concentration was prescribed
11 with 284 ppm for model spin-up and based on ice core and atmospheric measurements for
12 transient simulations (Keeling et al., 2009). N deposition was set as natural background for
13 simulations before 1850 (Dentener and Crutzen, 1994), and interpolated linearly between the
14 natural background and a snapshot of contemporary (1995) deposition (Dentener et al., 2006)
15 for simulations after 1850. Soil pH was prescribed and derived from the Harmonized World
16 Soil Database (HWSD) version 1.1, the same as NACP model driver data (Wei et al., 2014).

17 The model was spun up from bare ground without C-N interactions for the first 68 years and
18 with C-N interactions for the following 1200 years to develop and equilibrate C and N stocks.
19 To accelerate the spin-up process, slow litter and soil C and N pools were set to the equilibrium
20 values based on litterfall inputs and decomposition/leaching rates every 17 years. We
21 determined the model to reach a quasi-equilibrium state by confirming the drift to be less than
22 0.03 PgC yr^{-1} for global C storage and 0.2 TgN yr^{-1} for global N storage. From this quasi
23 equilibrium state, we initialized the global hindcast experiment starting from 1850 using the
24 corresponding climatic forcings, CO_2 and N deposition data. In the following analysis, we will
25 focus mostly on the last three decades (1970-2005).

26 **2.2.2 Sensitivity to soil water filled pore space (WFPS)**

27 While LM3V-N carries a simplified hydrology, we bracketed effects of soil moisture by
28 exploring the parameterization of WFPS and by substituting the predicted soil moisture with 3-
29 hourly re-analysis data. Levels of soil water (in unit kg m^{-2}) therefore stem from: (1) the

1 simulated water content based on LM3V-N soil water module, hereafter LM3V-SM (2) the
 2 Global Land Data Assimilation System Version 2 with the land surface model NOAH 3.3
 3 (Rodell et al., 2004), hereafter NOAH-SM, and (3) the ERA Interim reanalysis dataset from
 4 European Center for Medium range Weather Forecasting (ECMWF) (Dee et al., 2011),
 5 hereafter ERA-SM. The latter two datasets integrate satellite and ground based observations with
 6 land surface models. When overriding soil moisture, we linearly interpolated the 3 hourly data
 7 onto the 30 minutes model time step. In these simulations, we allowed soil C and N dynamics
 8 to vary according to different soil moisture datasets, but kept the model prediction of soil water
 9 to use for plant productivity and evapotranspiration.

10 Parameterization of the soil moisture effect on nitrification and denitrification are based on
 11 WFPS. LM3V-N uses the concept of plant available water, where water that is available to
 12 plants varies between the wilting point and field capacity. Water content above the available
 13 water capacity (i.e., the difference between field capacity and wilting point) leaves the soil
 14 immediately (Milly and Shmakin, 2002), and thus WFPS does not attain high values typically
 15 observed during denitrification. To explore the effect of WFPS – soil moisture relationship on
 16 N₂O emissions, we calculated WFPS using three methods. Method 1 assumes WFPS is the ratio
 17 of available water and the available water capacity in the rooting zone. In Method 2 we assumed,
 18 WFPS is the ratio of the water filled porosity and total porosity which is derived from bulk
 19 density (BD, in unit g cm⁻³). BD was obtained from the Harmonized World Soil Database
 20 (HWSD) version 1.1 (Wei et al., 2014). The calculation is given by

$$21 \quad WFPS = \frac{\theta}{1 - \frac{\rho h_r}{PD}} \quad (22)$$

22 where θ (kg m⁻²) is the root zone soil water; h_r (m) is the effective rooting depth of vegetation;
 23 ρ is the density of water (1000 kg m⁻³); and PD is the particle density of soil (2650 kg m⁻³).
 24 Method 1 generally leads to an overestimation of WFPS because the available water capacity
 25 is smaller than total pore space. In contrast, the use of Method 2 with LM3V-SM creates an
 26 underestimation since water is not allowed to accumulate beyond field capacity and misses high
 27 WFPS to which nitrification and denitrification are sensitive. Meanwhile, for NOAH-SM and
 28 ERA-SM data, Methods 2 is more close to the “real” WFPS and is the default method when
 29 using these data sets. The third approach, which is also the default method with LM3V-SM that
 30 is applied in the global hindcast experiment, the subsequent elevated CO₂ and temperature

1 responses experiment, and sensitivity tests with regard to N cycling, calculates WFPS as the
2 average of the previous two methods.

3 For each soil moisture dataset (3 in total, 2 replacements and 1 simulated by LM3V-N), we
4 calculated WFPS using three methods mentioned above. We conducted transient simulations
5 with the nine different WFPSs (3 datasets × 3 methods) starting from the near equilibrium state
6 obtained in the global hindcast experiment in 2.2.1. The use of less realistic Method for WFPS
7 for each soil moisture driver (LM3V-SM, NOAH-SM and ERA-SM) offers insights of the
8 sensitivity of N₂O emissions to soil moisture. The simulation procedure was the same as that in
9 global hindcast experiment except for the WFPS. ERA-SM is only available starting from 1979,
10 prior to which simulations were conducted with model default soil moisture (LM3V-SM).
11 Results from ERA-SM were analyzed starting from 1982, leaving a short period for adjustment.

12 **2.2.3 Sensitivity to N cycling processes and parameterization**

13 N₂O emission is constrained by ecosystem availability of mineral N, which is linked to different
14 N cycling processes in addition to nitrification and denitrification processes. To test the
15 sensitivity of modelled N₂O emission to the larger plant-soil N cycle, we conducted the
16 following sensitivity analyses, in form of a one at a time perturbation. We replaced the dynamic
17 BNF scheme with empirically reconstructed preindustrial fixation rates (Cleveland et al., 1999;
18 Green et al., 2004), removing the negative feedback between BNF and plant N availability. We
19 further shut off N loss pathways through DON leaching and fire volatilization (with
20 *ash_fraction* =1). We expect that these three modifications alleviate N limitation: Prescribed
21 BNF may continuously add N beyond plant N demand. Further eliminating fire and DOM N
22 losses leave loss pathways that have to pass the available N pool thereby opening the possibility
23 of increasing gaseous losses. Further, removing these plant-unavailable pathways (Thomas et
24 al., 2015) increases N retention and opens the possibility of alleviating N limitation. In addition,
25 we modified key parameters related to general N cycling and N₂O emissions one-at-a-time. We
26 multiplied several parameters that directly affect ammonium and nitrate concentration or N₂O
27 fluxes by 10 (x10) or 0.1 (x0.1), while kept other parameters as defaults. Those parameters
28 control the active root N uptake rates (v_{max}), nitrification rate (k_n), denitrification rate (k_d, K_c, K_n)
29 and the fraction of net nitrification lost as N₂O (*frac*),

1 2.2.4 Responses to elevated CO₂ and temperature

2 Responses of N₂O emissions to atmospheric CO₂ and global warming have been reported at field
3 scale (Dijkstra et al., 2012; van Groenigen et al., 2011). Here, we evaluate the model's response
4 to step changes in form of a doubling of preindustrial CO₂ level (284 ppm to 568 ppm) and a
5 2K increase in atmospheric temperature. Starting from the same quasi-equilibrium state with
6 potential vegetation as in the global hindcast experiment in 2.2.1, we conducted four transient
7 model runs: (1) the CONTROL run with the same drivers as spin-up; (2) the CO₂_FERT run
8 with the same drivers as the CONTROL except a doubling of atmospheric CO₂ level; (3) the
9 TEMP run with the same drivers as the CONTROL except a 2K rise in atmospheric temperature;
10 and (4) the CO₂_FERT×TEMP run with both the doubling of CO₂ and 2K rise in temperature.
11 For each experiment, we ran the model for 100 years and evaluated the corresponding results.

12 2.3 Comparisons with observations

13 We compared our model results for annual N₂O gas loss with field data: We compiled annual
14 N₂O emissions from peer-reviewed literature (see Appendix A for more information). To
15 increase the representativeness of the measurements, we included only sites with more than 3
16 months or 100 days experimental span. ~~We limited our datasets where there was no reference~~
17 ~~to a disturbance of any kind.~~ Only locations with at least 50 years non-disturbance history for
18 forests and 10 years for vegetation other than forests were included. The compiled 61
19 measurements cover a variety of spatial ranges with vegetation types including tropical
20 rainforest, temperate forest, boreal forest, tundra, savanna, perennial grass, steppe, alpine grass
21 and desert vegetation. Multiple measurements falling into the same model grid cell were
22 averaged. If the authors had indicated the dominant vegetation or soil type, we used the values
23 reported for the dominant type instead of the averaged. For multiyear measurements, even if
24 the authors gave the individual year's data, we averaged the data to avoid overweighting of long
25 term studies. If the location was between borders of different model grid cells, we averaged
26 across the neighboring grid cells.

27 We also compared monthly N₂O fluxes at a group of sites: (a) the Tapajós National Forest in
28 Amazonia (3°S, 55°W), taken from Davidson et al. (2008); (b) the Hubbard Brook
29 Experimental Forest in New Hampshire, USA (44°N, 72°W), as described in Groffman et al.
30 (2006); (c) the cedar forest from Oita, Japan (33°N, 131°E), as described in Morishita et al.

1 (2007); (d) the *Leymus chinensis* (LC) and *Stipa grandis* (SG) steppe in Inner Mongolia, China
2 (44°N, 117°E), taken from Xu-Ri et al. (2003); (e) the cedar forest in Fukushima, Japan (37°N,
3 140°E), taken from Morishita et al. (2007); and (f) the primary (P1 and P2) and secondary (L1
4 and L2) forests located at the Pasir Mayang Research Site (1°S, 102°E), Indonesia, taken from
5 Ishizuka et al. (2002). In addition, daily measurements of soil temperature, soil moisture and
6 N₂O emissions were compared at four German forest sites located in the same grid cell (50°N,
7 8°E), as described in Schmidt et al. (1988).

8 **3 Results**

9 **3.1 Global budget, seasonal and inter-annual variability**

10 Our modelled global soil N₂O flux is 6.69±0.32 TgN yr⁻¹ (1970-2005 mean and standard
11 deviation among different years) (Fig.1) with LM3V-SM (Method 3, default method for
12 LM3V-N calculated soil moisture), 5.61±0.32 TgN yr⁻¹ with NOAH-SM (Method 2) and
13 7.47±0.30 TgN yr⁻¹ with ERA-SM (1982-2005, Method 2) which is within the range of reported
14 values: The central estimate of N₂O emission from soils under natural vegetation is 6.6 TgN yr⁻¹
15 based on the Intergovernmental Panel on Climate Change (IPCC) AR5 (Ciais et al., 2013)
16 (range, 3.3–9.0 TgN yr⁻¹) for the mid-1990s. Mean estimation for the period of 1975-2000
17 ranged from 7.4 to 10.6 TgN yr⁻¹ with different precipitation forcing data (Saikawa et al., 2013).
18 Xu-Ri et al. (2012) reported the decadal-average to be 8.3-10.3 TgN yr⁻¹ for the 20th century.
19 Potter and Klooster (1998) reported a global mean emission rate of 9.7 TgN yr⁻¹ over 1983-
20 1988, which is higher than the earlier version of their model (6.1 TgN yr⁻¹) (Potter et al., 1996).
21 Other estimates include 6-7 TgN yr⁻¹ (Syakila and Kroeze, 2011), 6.8 TgN yr⁻¹ based on the O-
22 CN model (Zaehle et al., 2011), 3.9-6.5 TgN yr⁻¹ for preindustrial periods from a top-down
23 inversion study (Hirsch et al., 2006), 1.96-4.56 TgN yr⁻¹ in 2000 extrapolated from field
24 measurements by an artificial neural network approach (Zhuang et al., 2012), 6.6-7.0 TgN yr⁻¹
25 for 1990 (Bouwman et al., 1995), and 7-16 TgN yr⁻¹ (Bowden, 1986) as well as 3-25 TgN yr⁻¹
26 (Banin, 1986) from two earlier studies.

27 Following Thompson et al. (2014), El Niño years are set to the years with the annual
28 multivariate ENSO index (MEI) greater than 0.6. 1972, 1977, 1982, 1983, 1987, 1991, 1992,
29 1993, 1994, 1997 and 1998 were chosen as El Niño years. We detected reduced emissions

1 during El Niño years (Fig. 1), in line with the global atmospheric inversion study of Thompson
2 et al. (2014) and the process based modelling study from Saikawa et al. (2013).

3 Figure 2 shows the simulated global natural soil N₂O emissions in 4 seasons averaged over the
4 period of 1970-2005 based on LM3V-SM (Method 3). The northern hemisphere displays a large
5 seasonal variability, with the highest emissions in the northern summer (JJA, June to August)
6 and lowest in winter (DJF, December to February). Globally, northern spring (MAM, March to
7 May) has the highest emission rate (2.07 TgN) followed by summer (1.89 TgN). The smaller
8 emissions in summer compared to spring stems from a reduced contribution of the southern
9 hemisphere during northern summer.

10 As expected, a large portion (more than 60%) of the soil N₂O fluxes have tropical origin (23.5
11 S to 23.5N), while emissions from cooler regions are limited by temperature and arid/semi-arid
12 regions by soil water. Our modelling results suggest year-round high emission rates from humid
13 zones of Amazonia, east central Africa, and throughout the islands of Southeast Asia, with small
14 seasonal variations (Fig. 2). Emissions from tropical savannah are highly variable, with
15 locations of both high fluxes (seasonal mean > 30 mgN m⁻² month⁻¹ or 3.6 kg ha⁻¹ yr⁻¹) and low
16 fluxes (seasonal mean < 1.3 mgN m⁻² month⁻¹ or 0.16 kg ha⁻¹ yr⁻¹). The simulated average
17 tropical emission rate is 0.78 kgN ha⁻¹ yr⁻¹ (1970-2005), within the range of estimates (0.2-1.4
18 kgN ha⁻¹ yr⁻¹) based on site-level observations from the database of Stehfest and Bouwman
19 (2006), but smaller than a more detailed simulation study (1.2 kgN ha⁻¹ yr⁻¹) carried out by
20 Werner et al. (2007). Our analysis here excluded land cover, land use changes and human
21 management impacts, while most of the observation-based or regional modelling studies did
22 not factor out those impacts. Our modelling result in natural tropics is comparable with another
23 global modelling study (average emission rate, 0.7 kgN ha⁻¹ yr⁻¹) (Zaehle et al., 2010), in which
24 the authors claimed they may underestimate the tropical N₂O sources compared to the inversion
25 estimates from the atmospheric transport model TM3 (Hirsch et al., 2006).

26 **3.2 Sensitivity to WFPS**

27 The different parameterization of WFPS and the use of different soil moisture modeling and
28 data allows to test the sensitivity of soil N₂O emissions to variable WFPS. Globally, emissions
29 generally increase with WFPS (Fig. 3). WFPS derived from Method 1 is higher than that based
30 on Method 2. Data-derived soil moisture datasets combined with different calculation methods
31 together produced a range of 0.15-0.72 for the global mean WFPS (1982-2005). While mean

1 values greater than 0.6 (approximately field capacity) are less realistic, these high WFPS values
2 provide the opportunity to test the model's response to the soil moisture-based parameterization
3 of redox conditions in soils. Global soil N₂O emissions are highly sensitive to WFPS, with
4 approximately 0.25 TgN per year per 0.01 change in global mean WFPS in the range 0 to 0.6.
5 The spatial and temporal characteristic of WFPS also matters. Emission rate from LM3V-SM
6 (Fig. 3 green cycle) is 1.13 TgN yr⁻¹ higher than that from NOAH-SM (Fig. 3 blue triangle),
7 while both model configuration have the same mean WFPS (*ca.* 0.21), highlighting effects of
8 regional and temporal differences between the soil moisture products.

9 **3.3 Model-observation comparisons**

10 Modelled N₂O emissions capture the average of cross-site observed annual mean emissions
11 (0.54 vs. 0.53 kgN ha⁻¹ yr⁻¹ based on LM3V-SM) reasonably (Appendix A and Fig. 4a), but
12 spread considerably along the 1:1 line. The points deviating the most are from tropical forests,
13 with overestimations from montane tropical forest and underestimations from lowland tropical
14 forests if those measurements are representative of gridcell emissions. These patterns are
15 similar as results from NOAH-SM (Appendix A and Fig. 4b) and ERA-SM (Appendix A and
16 Fig. 4c), except that the application of WFPS from NOAH-SM slightly underestimates the
17 observed global mean (0.54 vs. 0.47 kgN ha⁻¹ yr⁻¹ from NOAH-SM with WFPS based on
18 Method 2).

19 At the Tapajós National Forest, results from LM3V-SM capture some of the variations in N₂O
20 fluxes, but the model is not able to reproduce the high emissions observed during spring (Panel
21 (a), Fig. 5), which might be caused by the underestimation of WFPS from models. We used a
22 total porosity of 0.62 (Davidson et al., 2004) to estimate root zone WFPS based on the reported
23 soil water content (Davidson et al., 2008). The average WFPS from observation is estimated to
24 be 0.49, which is higher than the modelled average of root zone WFPS for all 3 model
25 configurations (LM3V-SM 0.27, NOAH-SM 0.30, and ERA-SM 0.37). WFPS varies between
26 < 0.05 and 0.45 in LM3V-SM (range from 0.20 to 0.36 in NOAH-SM and 0.30 to 0.41 in ERA-
27 SM), and contrasts with observation that show seasonal variations with WFPS in the range of
28 0.37 to 0.58. At the Hubbard Brook Experimental Forest, the correlations between model results
29 and observations are 0.51 (LM3V-SM), 0.56 (NOAH-SM) and 0.62 (ERA-SM) for yellow
30 birch, 0.66 (LM3V-SM), 0.68 (NOAH-SM) and 0.70 (ERA-SM) for sugar maple. However,
31 the model is less robust in reproducing the magnitude of emission peaks. Groffman et al. (2006)

1 suggested high emissions of N₂O in winter were associated with soil freezing. However, the
2 model assumes little emissions when soil temperature is under 0 °C. In addition, observations
3 suggested N₂O uptake (negative values in Panel (b), Fig. 5) while the model does not
4 incorporate mechanisms to represent N₂O uptake. At the Oita cedar forest, LM3V-N reproduces
5 the seasonality of N₂O emissions accurately (Panel (c), Fig. 5). ERA-SM overestimates the
6 magnitude of N₂O fluxes from Inner Mongolia grassland, while the magnitudes produced from
7 LM3V-SM and NOAH-SM are comparable with observations. However, the timing of the
8 emission peaks are one or two month in advance in the model compared to observations (Panel
9 (d), Fig. 5). WFPS at a nearby meteorological station fluctuated between 0 and 0.5 for 0-20cm
10 depth (Xu-Ri et al., 2003) which agrees with our values based on LM3V-SM and ERA-SM, but
11 the range is lower for NOAH-SM (0.05 to 0.35). At the specific plots, Xu-Ri et al. (2003)
12 reported a mean WFPS of 0.32 in one plot (LC) and 0.20 from in the other plot (SG) for the 0
13 to 0.1 m depth interval which are close to ERA-SM and NOAH-SM (LM3V-SM 0.14, NOAH-
14 SM 0.19, ERA-SM 0.30), however, no temporal information was provided for the specific sites.
15 At the Fukushima cedar forest, similar as at the Oita cedar forest, models are less robust at
16 capturing the magnitude of high peaks of N₂O emissions although the seasonality produced by
17 the model are good (Panel (e), Fig. 5). Emissions in the primary and secondary tropical
18 rainforest at the Pasir Mayang Research Site are highly variable, which makes the comparison
19 difficult (Panel (f), Fig. 5). LM3V-SM (but not ERA-SM and NOAH-SM) reproduces the low
20 emissions in September-November 1997 and the increase of emissions from secondary forests
21 in December, 1997. Overall, modeled variability is smaller compared to observation across
22 these sites.

23 The strong variability of measured N₂O emissions is further illustrated in Fig. 6. Differences in
24 measured N₂O fluxes between different forest sites are large, reflecting heterogeneity that is not
25 captured within one model grid cell. In addition, the error bars, which represent the standard
26 deviation of measured N₂O fluxes at three different plots of the same forest, are large. The
27 standard deviation is as high as 49.27 μgN m⁻²h⁻¹, indicating the strong variability of measured
28 N₂O fluxes at the plot scale. Modeled N₂O fluxes are generally within the range of measured
29 N₂O emissions. Model outputs slightly underestimate N₂O emissions largely due to the
30 underestimation of soil water content (Panel (b) Fig. 6).

1 3.4 Sensitivity to N cycling processes and parameterization

2 Disallowing N losses through DON and fire volatilization enhance ecosystem N accumulation
3 and availability to plants and microbes, and therefore increases N₂O emissions (Panel (a), Fig.7).
4 The gain in N₂O emissions from disallowing DON loss is small (0.12 TgN yr⁻¹). However, N₂O
5 emission is on average (1950-2005) increased by 3.63 TgN yr⁻¹ in the absence of fire
6 volatilization N loss (we note, that fires do occur, but N is retained as ash in the litter). The gain
7 is most evident in tropical regions (not shown), indicating the importance of fire in regulating
8 ecosystem N status. Simulated preindustrial BNF is smaller than the empirical reconstructed
9 BNF (72 in LM3V-N vs. 108 TgN yr⁻¹ from empirical based data Green et al., 2004). However,
10 BNF in LM3V-N increases with time under historical varying climate, increasing atmospheric
11 CO₂ level and N deposition. The global average BNF ~~during simulated for the period~~ 1950-2005
12 is 100 TgN yr⁻¹, close to the empirical value. Nevertheless, substitution of BNF in LM3V-N by
13 empirical preindustrial value increased N₂O flux by 1.2 TgN yr⁻¹ (Panel (a), Fig.7).

14 Among the specific parameters tested, N₂O emission is most sensitive to the 10 times change
15 (x10) of the fraction of net nitrification lost as N₂O gas. The relative magnitude of N₂O flux on
16 average (1950-2005) reaches 6.5 times of the default (Panel (b), Fig.7). Reduction (x0.1) of
17 maximum active plant N uptake strength (v_{max}) strongly increases N₂O emissions (*ca.* by 3 times
18 of the default). Meanwhile, enhancement of v_{max} also increases N₂O fluxes, reflecting the non-
19 linear response of N₂O emissions to v_{max} . x10 ~~in k_n~~ the maximum nitrification rate k_n and
20 denitrification rate k_d increase N₂O emissions, while x0.1 decrease N₂O flux. N₂O increases
21 more with increasing k_d than with increasing k_n , whereas reduction of k_n (x0.1) produces a
22 stronger response than reduction of k_d . The half-saturation constant that represents the
23 regulation of labile carbon availability on denitrification rate, K_c , is the least sensitive parameter.
24 Meanwhile, reduction (x0.1) of the half-saturation constant K_n that represents the regulation of
25 substrate availability on denitrification rate on average increased N₂O fluxes by 4.5 TgN yr⁻¹
26 (Panel (b), Fig.7).

27 3.5 CO₂ and temperature responses

28 Globally, N₂O emissions respond to a step CO₂ increase first with a decline to ultimately
29 increased levels after approximately 40 years (Fig. 8a, black line). The simulated global
30 response follows largely the behaviour as simulated for tropical forests (Fig. 8a, yellow line).
31 The shift from a negative to a positive response indicates possible competing mechanisms

Formatted: Font: Not Italic

1 operating on different time scales. Field level experiments revealed the highly variable effects
2 of CO₂ fertilization on N₂O emissions. Based on a meta-analysis, van Groenigen et al. (2011)
3 suggested that elevated CO₂ significantly increased N₂O emission by 18.8%, while Dijkstra et
4 al. (2012) argued for a non-significant response in non-N-fertilized studies. In contrast to
5 observation studies, the global C-N cycle model analyses from O-CN suggested negative CO₂
6 fertilization effects on N₂O emissions (Zaehle et al., 2011). The negative impacts (reduced N₂O
7 flux), which are also reported in manipulative experiments, are likely from increased plant N
8 and immobilization demand under CO₂ fertilization, reducing N availability for nitrifiers and
9 denitrifiers (Dijkstra et al., 2012). CO₂ fertilization on average (over 100 years) increased the
10 global mean plant nitrogen uptake rate by 10.02 kgN ha⁻¹ yr⁻¹, as shown in Fig. 9 (Panel (b)).
11 Modelled soil inorganic N content (ammonium and nitrate) is reduced at first, but the reduction
12 is not sustained. One mechanism to alleviate CO₂ fertilization induced N limitation is through
13 BNF, which is on average (over 100 years) more than doubled (Fig. 9 Panel (e)). Similar to
14 manipulative field experiments (Dijkstra et al., 2012), positive effects (increase N₂O fluxes)
15 can result from the impacts of elevated CO₂ level to increase litter production (Fig. 9 Panel (a))
16 and consequently C sources for denitrifiers, and to increase soil moisture (Fig. 9 Panel (d)) from
17 reduced stomatal conductance and leaf transpiration (Fig. 9 Panel (c)). With both positive and
18 negative mechanisms embedded in our model, the net effects depend on the relative strength of
19 the opposing forces.

20 Temperate deciduous forests, where most of the forest CO₂ fertilization experiments are
21 conducted, respond positively to elevated CO₂ ~~level~~ levels (Fig. 8a, green line). The slight
22 increase in modelled N₂O emission are comparable with the mean response of field data
23 compiled for temperate forests (*ca.* 0.01-0.03 kgN yr⁻¹ ha⁻¹) (Dijkstra et al., 2012). A similar
24 positive response was detected for cold evergreen forests (Fig. 8a, pink line) with stronger
25 magnitude compared to temperate deciduous forests. For grasslands, Dijkstra et al. (2012)
26 reported small negative mean response from northern mixed prairie (Δ N₂O, *ca.* -0.01 to -0.03
27 kgN yr⁻¹ ha⁻¹), zero mean response from shortgrass steppe and positive mean response from
28 annual grassland (*ca.* 0.03-0.06 kgN yr⁻¹ ha⁻¹). Our model shows a small negative mean response
29 from C4 grassland (Fig. 8a, cyan line) with the similar magnitude of that reported for the
30 Northern mixed prairie, where the composition of C4 grass varies (Dijkstra et al., 2012). A CO₂
31 increase in C3 grassland initially reduces N₂O emission (Fig. 8a, blue line). However, this slight
32 negative response turns into a small positive within one decade.

1 Elevated temperature generally increases N₂O emissions except for the slight negative effect in
2 C4 grass (Fig. 8b). Overall the response to a 2 degree warming is bigger than that of doubling
3 of CO₂. The simulated temperature effects are more pronounced in the first decade and decrease
4 over time in tropical forests (Fig. 8b, yellow line), while for the temperate deciduous forests
5 (Fig. 8b, green line) and boreal forests (Fig.8b pink line), the temperature effects become more
6 pronounced over time. Simulated temperate forest response (in the first decade) is close to that
7 of observed mean (*ca.* 0.2-0.5 kgN yr⁻¹ ha⁻¹) (Dijkstra et al., 2012). Our modelled slight negative
8 response in C4 grass and positive in C3 grass are in alignment with data compiled by Dijkstra
9 et al. (2012) who reported both positive and negative responses in grasslands.

10 The results of combining CO₂ and temperature are similar to the CO₂ effect alone (Fig. 8c),
11 despite the fact, that the individual effect of temperature is much stronger than that of CO₂. This
12 antagonistic interaction (i.e. the combined enhancement in N₂O flux from elevated CO₂ and
13 temperature are smaller than the summary of their individual effects) is also evident for C3
14 grass (first 50 years), temperate deciduous tree and cold evergreen forests (Fig. 8d).

15 **4 Discussion**

16 Our model combines two of the most widely applied biogeochemical models (DNDC and
17 CENTURY) with current advancements in field level studies. The model was capable of
18 reproducing the global mean natural N₂O emissions in other modeling and inverse methods,
19 and the average of observed cross-site annual mean behavior. By focusing on the role of soil
20 moisture in N₂O emissions, we found on a global scale a high dependence of simulated N₂O
21 emissions on soil moisture (WFPS), mainly driven by emissions from tropical regions. The
22 model broadly reproduced the magnitude and direction of responses to elevated CO₂ and
23 temperature from manipulative field experiments where data is available. Both the global total
24 emission as well as the global response to temperature and CO₂ increase followed largely the
25 response of tropical forests, where field experiments are rare and no evaluation of CO₂
26 responses exist.

27 Soil moisture is a key variable in climate system but difficult to derive or measure at the global
28 scale (Seneviratne et al., 2010). Our modelled fluxes were highly sensitive to WFPS, which is
29 in agreement with observation and model synthesis studies (Heinen, 2006; Butterbach-Bahl et
30 al., 2013). ~~The large range when calculating~~Calculating WFPS from different methods resulted
31 in a difference of more than 5 TgN yr⁻¹ in global soil N₂O fluxes. Saikawa et al. (2013) found

1 an up to 3.5 TgN yr⁻¹ gap induced by different precipitation forcing data from CLMCN-N2O.
2 It is difficult to single out the difference caused by soil moisture alone from their results.
3 Nevertheless, ~~those two studies did suggest~~our and previous results highlight the importance
4 of improving the dynamics of soil water and representation of WFPS for the purpose of
5 predicting soil N₂O emission and climate feedbacks.

6 The root zone soil water in LM3V-N is based on a single layer bucket model. This simplified
7 treatment of soil water dynamics may increase the difficulty in reproducing the temporal and
8 spatial dynamics of WFPS. As a first step, we used the average between the original analog in
9 LM3V-N and a formulation that was derived from soil total porosity to account for actual soil
10 moisture and the possibility of soil water above field capacity. Meanwhile, overriding soil
11 moisture with data-derived products (NOAH-SM and ERA-SM) suggests that the most realistic
12 average (1970-2005) soil N₂O emission is in the range of 5.61-7.47 TgN yr⁻¹. However, despite
13 using data-derived soil moisture, it appears that the prediction of soil moisture is an impediment
14 towards validating N₂O emissions at field scale- for both LM3V-N simulated and reanalysed
15 soil moisture. If evaluated against field data, the model was capable of representing the mean
16 across sites and to a certain degree also compared adequately with site-specific time series.
17 However, across the models there are differences between sites model and data (Fig. 4) ~~and~~
18 ~~also~~particularly peak emissions were poorly represented in the model (Fig. 5), and they can at
19 least partly be attributed to mismatches in WFPS. Overall, comparison against field data
20 revealed that the model's variability is smaller compared to observation for both across field
21 sites (Fig. 4) and at different sites (Figs. 5 and 6). One of the reason for this shortcoming may
22 be that fast transitions, such as freeze-thaw cycle (Groffman et al., 2006) and pulsing (Yienger
23 and Levy, 1995) are not sufficiently captured.

24 Perhaps equally important to address in future analysis, is the tremendous variability of N₂O
25 emissions from site to site within the same region (see Fig. 6) This field-scale variability
26 highlights the complexity of microscale interactions for N₂O production, which creates
27 notorious large spatial and temporal variabilities and are undoubtedly difficult to constrain even
28 at the stand level (Butterbach-Bahl et al., 2013). The homogeneous representation of
29 environmental drivers within model grid cells casts doubt on site-specific model-observation
30 comparison in global simulations. For example, N₂O emissions vary with topography which
31 are not treated explicitly in most of the global C-N models. 3.8 times difference was detected
32 in a montane forest (Central Sulawesi, Indonesia) moving from 1190 m to 1800m (Purbopuspito

1 et al., 2006), and 4.3 times difference was found from a tropical moist forest (Brazilian Atlantic
2 Forest) with the altitude changing from 100m to 1000m (Sousa Neto et al., 2011).

3 Globally, N₂O emissions from nitrification-denitrification were similar to O-CN and LPJ-DyN
4 as they are all derived from DNDC's formulation- (Xu-Ri et al., 2012; Zaehle et al., 2011).
5 Embedding an established N₂O emission module into LM3V-N enables evaluation of the
6 response of N₂O emissions under different assumptions across models with respect to the
7 dynamics of the larger plant-soil N cycle. Generally higher inputs from BNF or restriction of
8 losses through organic N (fire, DON) enhance N₂O emissions. The representation of BNF in
9 models requires improvement ~~but~~; currently, simple empirical relationships are used, yet BNF
10 is the largest source of new N in terrestrial systems, and therefore is critical in the determination
11 of N availability to nitrifiers and denitrifiers. Here we showed ~~here~~ that different
12 implementations of BNF (prescribed vs. responsive to plant N demand) are globally important
13 for N₂O emissions. Similarly, the magnitude of N lost through fire impacted N₂O emissions in
14 fire prone regions, while N emission factors are poorly constrained globally (Andreae and
15 Merlet, 2001). The strength of plant uptake of N posed a strong constraint on the availability of
16 N for nitrification-denitrification losses as it can draw down N substantially (Gerber and
17 Brookshire, 2014). A reduction of plant uptake strength allows for relatively more N allocated
18 for denitrification. More surprising was the positive effect of a stronger plant uptake capacity
19 on N₂O emissions: Enhanced plant uptake allow increased vegetation production and N
20 throughput through litterfall and mineralization in the long run, which ultimately may allow
21 higher N₂O losses. In addition to those N cycling processes, N₂O emissions were highly
22 sensitive to the fraction of N lost as N₂O during net nitrification. The fraction of N₂O lost during
23 net nitrification is uncertain. Goodroad and Keeney (1984) suggested a value of 0.1-0.2% ,
24 while Khalil et al. (2004) reported a range of 0.16%-1.48% depending on the O₂ concentration.
25 We applied a global constant of 0.4% in our default simulation, bearing in mind the large
26 uncertainties associated with this parameter.

27 Our results showed that tropical forests play a major role in both rates of emission and responses
28 to perturbations. Tropical forests contributed with more than 60% to the global soil N₂O fluxes.
29 El Niño events triggered reduced soil N₂O emissions that are in our simulations similar to earlier
30 estimates (Saikawa et al., 2013; Thompson et al., 2014). El Niño events are known to have
31 induced several of the most well known large scale droughts and altered soil moisture dynamics
32 (Schwalm et al., 2011) in the tropics. Tropical forest N₂O emissions were highly correlated with

1 root zone soil water content and contributed strongly to the global-scale fluxes of N₂O in our
2 model. Similarly, global responses to elevated CO₂ and temperature were dominated by the
3 tropical response. In contrast to temperate and boreal forests, tropical forests responded
4 negatively to elevated CO₂ in the first few decades. The initial negative response of N₂O
5 emissions to CO₂ fertilization in tropical forests produced by LM3V-N stemmed largely from
6 increased demand and uptake of mineral N due to enhanced vegetation growth under elevated
7 atmospheric CO₂ level. Consequently, less N is available for gaseous losses as the N cycle
8 tightens. If gross mineralization is used as an indicator of the rate of N flow in the “hole-in-the-
9 pipe” concept and gaseous losses are proportional to mineralization, the initial negative response
10 is unlikely to be detected. We found increased mineralization rate with increased litterfall under
11 elevated CO₂, while N availability is reduced from LM3V-N. The mineralization based
12 approach is likely to predict an increase of losses regardless of N limitation.

13 The marked decrease in our simulation for the tropical forests also contrasts somewhat findings
14 from manipulative field experiments where CO₂ enrichment caused decrease, ~~unchanged~~
15 ~~no~~ ~~change~~ or increase across extratropical ecosystems (Dijkstra et al., 2012; van Groenigen et al.,
16 2011), whereas no empirical evidence is available in tropical forests. Overall, the marked
17 differences between tropics and extratropics in the response to environmental forcing, and the
18 large contribution of tropical forests to global N₂O emissions suggests caution when
19 extrapolating field studies mostly carried out in extratropical regions to the globe.

20 Based on single factor analysis with LM3V-N, the initial response of N₂O emission to a
21 temperature increase was much larger than the response to ~~increase~~ ~~increased~~ atmospheric CO₂
22 (Fig. 8). However, we found large interactions between warming and CO₂ fertilization, in that
23 the combined effect much more resembled the CO₂ effect alone. This interaction is the result
24 of two antagonistic responses where a warming lead to increased N mineralization and potential
25 N surplus, whereas a CO₂ increase fostered plant N demand that competed with microbial N₂O
26 production. While these mechanisms are part of most models, both comparison against different
27 models show notable differences when analyzing these two opposing effects. For example,
28 Stocker et al. (2013) found that under future climate change scenarios, CO₂ and climate effects
29 are amplifying each other, in accordance with earlier model experiments (Xu-Ri et al., 2012).
30 In LM3V-N we find that these interactions are negative. On the other hand, simulations with
31 O-CN (Zaehle et al., 2011) showed the ~~marginal~~ effects of CO₂ and climate to be approximately
32 equal and ~~of opposite~~. ~~The marginal effects in~~ ~~sign for historic simulations covering the~~

1 ~~modeling setup of Zaehle et al. (2011) compare best with our past 300 years that also include~~
2 ~~land-cover changes. They evaluated the effect of climate change as the difference between a~~
3 ~~simulation that consider both CO₂ and climate and a simulation that does not consider climate~~
4 ~~change. Thus their climate effect contains both the single effect for CO₂, while for of climate,~~
5 ~~it is and the combination of interaction of climate with CO₂. The temperature and interaction~~
6 ~~(i.e. climate) response on top of CO₂ can in our simulation be calculated as the temperature~~
7 ~~effect plus the interaction effect (Fig- 8). Analyzed in ~~their~~this fashion, LM3V-N's results are~~
8 congruent with those of Zaehle et al. (2011), albeit we found a slightly weaker temperature
9 effect compared to CO₂. ~~This~~Over time, the initial response ~~then~~ transitions into a much larger
10 CO₂ effect, while the response to temperature diminishes. This long-term response of a positive
11 CO₂ effect can be expected in a model that strongly retains N under limiting conditions such as
12 LM3V-N. Retention ultimately allows build-up of N stocks, thereby alleviating limitation and
13 increasing the substrate for nitrifiers and denitrifiers. This transition into a positive CO₂
14 response was likely facilitated by up-regulation of BNF (Figure 9), which acts to reduce
15 ecosystem N deficits and plant N demand in medium- to long-term. Up-regulation is expected
16 to be much weaker or absent in models where BNF is parameterized based on
17 evapotranspiration (Thomas et al., 2015). We realize that strong interactions as found here and
18 elsewhere (Xu-Ri et al., 2012; Stocker et al., 2013) may also play out when other factors are
19 considered (Brown et al., 2012), including N deposition, precipitation and land ~~use~~cover change
20 ~~(disturbance)~~. Therefore some of the discrepancy with other models may be caused by
21 differences in the modeling setup. In addition, step changes in atmospheric CO₂ and
22 temperature compared to gradual and sustained increases may also lead to differences. Yet
23 applying step changes is useful to test our conceptual understanding and may help explain the
24 discrepancy between the previous modeling study and meta-analysis of manipulative field
25 experiments with regard to CO₂ fertilization responses (Zaehle et al., 2011; van Groenigen et
26 al., 2011)

27 **5 Conclusions**

28 We present estimates of terrestrial soil N₂O fluxes under natural vegetation (1970 to 2005)
29 based on existing N₂O emission formulations embedded into the global C-N cycle model
30 LM3V-N. To determine the sensitivity of the modelling result to soil water (WFPS), we
31 replaced the root zone soil water with two other derived datasets and altered the way in which
32 WFPS is calculated. Our best estimate of modelled global soil N₂O flux is 5.61-7.47 TgN yr⁻¹

1 (1970-2005), within the range of current understanding of soil N₂O emissions, but highly
2 sensitive to WFPS, general N cycling and parameterization of N₂O losses through nitrification
3 and denitrification. Comparison against field experiments suggests that LM3V-N was able to
4 capture mean values, although site-to-site and temporal mismatches remained. Given the
5 sensitivity of N₂O emissions to WFPS, improvements in soil hydrology are likely to improve
6 soil N₂O emission estimates. As expected, we found that processes in the model that alleviate
7 ecosystem N limitation, such as reduced N losses through fire volatilization and increased N
8 inputs through higher biological nitrogen fixation (BNF) rate, enhance N₂O emissions.
9 Responses to CO₂ and temperature perturbations showed differences compared to other models.
10 In particular elevated CO₂ curbs N₂O emissions sharply initially, but this negative response is
11 alleviated after a few decades, likely in conjunction with fast N replenishment from up-
12 regulated BNF. Our sensitivity analysis ~~and the comparison with other models showed~~ shows
13 that existing parameterizations of processes of the larger plant-soil N cycle affect fast N cycle
14 processes such as as evidenced by the response to the fire and BNF modification. This
15 sensitivity can lead to differences in N₂O across models (e.g. in the response to CO₂ and climate)
16 even if existing nitrification-denitrification lead to distinct and new results if the larger plant-
17 soil N cycle is treated differently. More importantly schemes are identical. Further, our work
18 suggests a ~~strong~~ much stronger response to warming and CO₂ in tropical forests, ~~where few~~
19 manipulative compared to extratropical forest, thus extrapolation of mostly extra-tropical field
20 studies have been carried out to the globe warrants caution.

21

22

1 Appendix A: Observed annual N₂O fluxes data

2 Annual N₂O fluxes data were compiled from peer-reviewed literature. We applied simple
 3 selection criteria (see the main text) to reduce the mismatches between model outputs and field
 4 measurements, bearing in mind the gaps between complex field conditions and idealized model
 5 forcings. Latitudes (Lat) and longitudes (Lon) in Table A1 are based on model grids.

6 Table A1 Observed annual N₂O emission data for model comparison

No	Country	Lon	Lat	Location	Veg Type	N ₂ O kgN ha ⁻¹ yr ⁻¹			ERA	Reference
						OBS	LM3V-N	NOAH		
1	Australia	133.1	-12.3	Douglas Daly region	Savanna	0.02	0.15	0.25		Grover et al. (2012)
2	Australia	148.1	-37.3	Moe	Temperate forest	0.11	0.58	0.74	0.72	Khalil et al. (1990)
3	Australia	151.9	-27.3	South-east Queensland	Tropical forest	0.52	0.01	0.03		Rowlings et al. (2012)
4	Austria	16.9	47.8	Klausenleopoldsdorf	Temperate forest	0.62	0.64	0.52	0.53	Kesik et al. (2005)
5	Austria	9.4	47.8	Achenkirch	Temperate forest	0.35	0.54	0.48	0.47	Kesik et al. (2005)
6	Austria	13.1	47.8	Innsbruck	Temperate forest	0.08	0.42	0.36	0.31	Henrich and Haselwandter (1997)
7	Austria	16.3	48.2	Schottenwald and Klausenleopoldsdorf	Temperate forest	0.76	0.61	0.54	0.53	Kitzler et al. (2006)
8	Brazil	-61.9	-2.3	Manaus	Tropical rain forest	1.9	1.6	1.68	1.56	Luizao et al. (1989)
9	Brazil	-61.9	-2.3	Manaus	Tropical rain forest	1.930	1.71	1.74	1.55	Keller et al. (1986)
10	Brazil	-54.4	-4.8	East-central Amazonia	Tropical rain forest	2.1	1.34	2.19	1.57	Davidson et al. (2008)
11	Brazil	-46.9	-2.3	Paragominas	Rainforest	2.430	1.22	1.19	1.11	Verchot et al. (1999)
12	Burkina Faso	-1.9	10.3	Ioba	Savanna	0.6	0.03	1.32		Bruemmer et al. (2008)
13	Canada	-80.6	50.3	Ontario	Boreal forest	0.04	0.11	0.14	0.12	Schiller and Hastie (1996)
14	Canada	-106.9	52.8	Saskatchewan	Boreal forest	0.28	0.01	0.01	0.01	Simpson et al. (1997)
15	Canada	-103.1	52.8	Saskatchewan	Boreal forest	0.07	0.21	0.17		Matson et al. (2009)
16	Canada	-106.9	52.8	Saskatchewan	Boreal forest	0.09	0.01	0.01		Matson et al. (2009)
17	Canada	-73.1	45.3	Mont St. Hilaire	Temperate forest	0.42	0.54	0.46		Ullah and Moore (2011)
18	China	91.9	35.3	Tibet	Alpine grassland	0.07	0	0	0	Pei (2003)
19	China	125.6	40.3	Changbai mountain	Alpine tundra, temperate forest	0.56	0.73	0.64	0.45	Chen et al. (2000)
20	China	114.4	42.8	Inner mongolia	Temperate forest	0.73	0.1	0.14	0.71	Du et al. (2006)
22	China	133.1	47.8	Sanjiang Experimental Station	Freshwater marshes	0.21	0.34	0.35	0.34	Yu et al. (2007)
23	Denmark	13.1	55.3	Solo	Temperate forest	0.29	0.27	0.42	0.06	Kesik et al. (2005)

24	Denmark	13.1	55.3	Denmark	Temperate forest	0.52	0.28	0.37	0.05	Struwe and Kjoller (1989)
25	Ecuador	-80.6	-4.8	Bombuscaro	Tropical forest	0.3	1.02	0		Wolf et al. (2011)
26	Finland	24.4	60.3	Southern	Boreal forest	0.78	0.62	0.35	0.17	Maljanen et al. (2006)
27	Germany	9.4	50.3	Average	Temperate forest	0.57	0.6	0.53	0.5	Templer et al. (2012)
28	Germany	9.4	52.8	Kiel	Temperate forest	0.4	0.48	0.53	0.52	Mogge et al. (1998)
29	Germany	9.4	47.8	Southwest	Temperate forest	0.93	0.56	0.51	0.49	Jungkunst et al. (2004)
30	Germany	13.1	47.8	Höglwald	Temperate forest	0.41	0.47	0.4	0.39	Luo et al. (2012)
31	Germany	9.4	52.8	Average	Temperate forest	0.66	0.44	0.5	0.5	Brumme et al. (1999)
32	Germany	9.4	52.8	Harz mountains	Mire	0.25	0.48	0.56	0.52	Tauchnitz et al. (2008)
34	Indonesia	103.1	-2.3	Jambi	Lowland tropical rainforest	0.260	0.44			Ishizuka et al. (2002)
35	Indonesia	121.9	-2.3	Central Sulawesi	Tropical seasonal rain forest	0.800	1.73	2.31	1.7	Purbopuspito et al. (2006)
36	Indonesia	114.4	-2.3	Central Kalimantan	Tropical forest	2.51	2	2.45	1.73	Takakai et al. (2006)
37	Italy	9.4	45.3	P.Ticino BoscoNegri	Temperate forest	0.18	1.38	2.8	1.82	Kesik et al. (2005)
38	Malaysia	110.6	-2.3	Sarawak	Mixed peat swamp forest	0.7	0.66	0.65	0.57	Melling et al. (2007)
39	New Zealand	170.6	-44.8	New Zealand	Temperate forest	0.01	1.24	2.84	1.24	Price et al. (2004)
40	Norway	9.4	60.3	Norway	Temperate forest	0.73	0.52	0.52	0.38	Sitaula et al. (1995)
41	Panama	-80.6	7.8	Gigante Peninsula	Tropical forests	1.6	0.2	0.39	0.39	Koehler et al. (2009)
42	Sweden	13.1	57.8	Southwestern	Temperate forest	0.07	1.86	1.67		Klemetsson et al. (1997)
43	Sweden	13.1	57.8	Asa experimental forest	Undrained bog	0.65	0.36	0.45	0.36	von Arnold et al. (2005)
44	UK	-1.9	55.3	Northumberland	Grassland	0.3	0.4	0.5	0.41	Ball et al. (2007)
45	USA	-73.1	42.8	Harvard forest	Mixed hardwood	0.04	0.56	0.54	0.48	Bowden et al. (1990)
46	USA	-73.1	40.3	New York	Temperate forest	0.9	0.4	0.49	0.41	Duxbury et al. (1982)
47	USA	-80.6	25.3	Florida	Marsh	1	0.45	0		Duxbury et al. (1982)
48	USA	-73.1	42.8	New Hampshire	Temperate forest	0.070	0.64	2.15		Groffman et al. (2006)
49	USA	-106.9	35.3	New mexico	Temperate forest	0.06	0.41	0.51	0.43	Matson et al. (1992)
50	USA	-118.1	45.3	Washington	Temperate shrub-steppe	0.15	0.02	0.02	0.02	Mummey et al. (1997)
51	USA	-114.4	37.8	Mojave desert	Perennial grasses	0.11	0.02	0.02	0.02	Billings et al. (2002)
52	USA	-106.9	40.3	Wyoming	Sagebrush steppe	0.21	0.01	0.02	0.03	Matson et al. (1991)
53	USA	-73.1	45.3	Northeastern	Temperate forest	0.18	0.05	0.04	0.05	Castro et al. (1992)
54	USA	-69.4	45.3	Northeastern	Temperate forest	0.03	0.53	0.46	0.44	Castro et al. (1992)
55	USA	-103.1	40.3	Colorado	Temperate steppe	0.14	0.37	0.53	0.4	Mosier et al. (1996)
56	USA	-88.1	42.8	Wisconsin	Grass	0.040	0.03	0.05	0.05	Cates and Keeney (1987)
57	USA	-114.4	37.8	Nevada	Mojave desert	0.11	0.45	0.45		Billings et al. (2002)

58	USA	-110.6	32.8	Arizona	Sonoran desert	0.4	0.04	0.04	0.05	Guilbault and Matthias (1998)
59	USA	-118.1	45.3	Ft. Collins, Colorado	Temperate grassland	0.12	0.01	0.03	0.03	Parton et al. (1988)
60	Venezuela	-61.9	10.3	Venezuela	Savana	0.73	0.06	0.07	0.07	Simona et al. (2004)
61	Zimbabwe	31.9	-17.3	Harare	Miombo woodland savanna	0.51	0.83	1.61	0.57	Rees et al. (2005)

1

2 References cited in the Appendix

- 3 Ball, T., Smith, K. A., and Moncrieff, J. B.: Effect of stand age on greenhouse gas fluxes from
4 a Sitka spruce [*Picea sitchensis* (Bong.) Carr.] chronosequence on a peaty gley soil, *Global*
5 *Change Biol.*, 13, 2128-2142, doi:10.1111/j.1365-2486.2007.01427.x, 2007.
- 6 Billings, S. A., Schaeffer, S. M., and Evans, R. D.: Trace N gas losses and N mineralization in
7 Mojave desert soils exposed to elevated CO₂, *Soil Biol. Biochem.*, 34, 1777-1784,
8 doi:10.1016/s0038-0717(02)00166-9, 2002.
- 9 Bowden, R. D., Steudler, P. A., Melillo, J. M., and Aber, J. D.: Annual nitrous-oxide fluxes
10 from temperate forest soils in the northeastern United-States, *J. Geophys. Res.-Atmos.*, 95,
11 13997-14005, doi:10.1029/JD095iD09p13997, 1990.
- 12 Bruemmer, C., Brueggemann, N., Butterbach-Bahl, K., Falk, U., Szarzynski, J., Vielhauer, K.,
13 Wassmann, R., and Papen, H.: Soil-atmosphere exchange of N₂O and NO in near-natural
14 savanna and agricultural land in Burkina Faso (W. Africa), *Ecosystems*, 11, 582-600,
15 doi:10.1007/s10021-008-9144-1, 2008.
- 16 Brumme, R., Borken, W., and Finke, S.: Hierarchical control on nitrous oxide emission in forest
17 ecosystems, *Glob. Biogeochem. Cycles*, 13, 1137-1148, doi:10.1029/1999gb900017, 1999.
- 18 Castro, M. S., Steudler, P. A., Melillo, J. M., Aber, J. D., and Millham, S.: Exchange of N₂O
19 and CH₄ between the atmosphere and soils in spruce-fir forests in the northeastern United-
20 States, *Biogeochemistry*, 18, 119-135, doi:10.1007/bf00003273, 1992.
- 21 Cates, R. L., Jr., and Keeney, D. R.: Nitrous oxide emission from native and reestablished
22 prairies in southern Wisconsin, *American Midland Naturalist*, 117, 35-42, 1987.
- 23 Chen, G. X., Huang, B., Xu, H., Zhang, Y., Huang, G. H., Yu, K. W., Hou, A. X., Du, R., Han,
24 S. J., and VanCleemput, O.: Nitrous oxide emissions from terrestrial ecosystems in China,
25 *Chemosphere: Global Change Science*, 2, 373-378, 2000.
- 26 Davidson, E. A., Nepstad, D. C., Ishida, F. Y., and Brando, P. M.: Effects of an experimental
27 drought and recovery on soil emissions of carbon dioxide, methane, nitrous oxide, and nitric
28 oxide in a moist tropical forest, *Global Change Biol.*, 14, 2582-2590, doi:10.1111/j.1365-
29 2486.2008.01694.x, 2008.
- 30 Du, R., Lu, D., and Wang, G.: Diurnal, seasonal, and inter-annual variations of N₂O fluxes from
31 native semi-arid grassland soils of inner mongolia, *Soil Biol. Biochem.*, 38, 3474-3482,
32 doi:10.1016/j.soilbio.2006.06.012, 2006.
- 33 Duxbury, J. M., Bouldin, D. R., Terry, R. E., and Tate, R. L.: Emissions of nitrous-oxide from
34 soils, *Nature*, 298, 462-464, doi:10.1038/298462a0, 1982.

- 1 Groffman, P. M., Hardy, J. P., Driscoll, C. T., and Fahey, T. J.: Snow depth, soil freezing, and
2 fluxes of carbon dioxide, nitrous oxide and methane in a northern hardwood forest, *Global*
3 *Change Biol.*, 12, 1748-1760, doi:10.1111/j.1365-2486.2006.01194.x, 2006.
- 4 Grover, S. P. P., Livesley, S. J., Hutley, L. B., Jamali, H., Fest, B., Beringer, J., Butterbach-
5 Bahl, K., and Arndt, S. K.: Land use change and the impact on greenhouse gas exchange in
6 north Australian savanna soils, *Biogeosciences*, 9, 423-437, doi:10.5194/bg-9-423-2012, 2012.
- 7 Guilbault, M. R., and Matthias, A. D.: Emissions of N₂O from Sonoran Desert and effluent-
8 irrigated grass ecosystems, *J. Arid Environ.*, 38, 87-98, doi:10.1006/jare.1997.0300, 1998.
- 9 Henrich, M., and Haselwandter, K.: Denitrification and gaseous nitrogen losses from an acid
10 spruce forest soil, *Soil Biol. Biochem.*, 29, 1529-1537, doi:10.1016/s0038-0717(97)00010-2,
11 1997.
- 12 Ishizuka, S., Tsuruta, H., and Murdiyarto, D.: An intensive field study on CO₂, CH₄, and N₂O
13 emissions from soils at four land-use types in Sumatra, Indonesia, *Global Biogeochem. Cycles*,
14 16, GB1049, doi:10.1029/2001gb001614, 2002.
- 15 Jungkunst, H. F., Fiedler, S., and Stahr, K.: N₂O emissions of a mature Norway spruce (*Picea*
16 *abies*) stand in the black forest (southwest Germany) as differentiated by the soil pattern, *J.*
17 *Geophys. Res.-Atmos.*, 109, D07302, doi:10.1029/2003jd004344, 2004.
- 18 Keller, M., Kaplan, W. A., and Wofsy, S. C.: Emissions of N₂O, CH₄ and CO₂ from tropical
19 forest soils, *J. Geophys. Res.-Atmos.*, 91, 1791-1802, doi:10.1029/JD091iD11p11791, 1986.
- 20 Kesik, M., Ambus, P., Baritz, R., Brüggemann, N. B., Butterbach-Bahl, K., Damm, M., Duyzer,
21 J., Horvath, L., Kiese, R., Kitzler, B., Leip, A., Li, C., Pihlatie, M., Pilegaard, K., Seufert, G.,
22 Simpson, D., Skiba, U., Smiatek, G., Vesala, T., and Zechmeister-Boltenstern, S.: Inventories
23 of N₂O and NO emissions from European forest soils, *Biogeosciences*, 2, 353-375, 2005.
- 24 Khalil, M. A. K., Rasmussen, R. A., French, J. R. J., and Holt, J. A.: The influence of termites
25 on atmospheric trace gases: CH₄, CO₂, CHCl₃, N₂O, CO, H₂, and light-hydrocarbons, *J.*
26 *Geophys. Res.-Atmos.*, 95, 3619-3634, doi:10.1029/JD095iD04p03619, 1990.
- 27 Kitzler, B., Zechmeister-Boltenstern, S., Holtermann, C., Skiba, U., and Butterbach-Bahl, K.:
28 Nitrogen oxides emission from two beech forests subjected to different nitrogen loads,
29 *Biogeosciences*, 3, 293-310, 2006.
- 30 Klemetsson, L., Klemetsson, A. K., Moldan, F., and Weslien, P.: Nitrous oxide emission
31 from Swedish forest soils in relation to liming and simulated increased N-deposition, *Biol. Fert.*
32 *Soil.*, 25, 290-295, doi:10.1007/s003740050317, 1997.
- 33 Koehler, B., Corre, M. D., Veldkamp, E., Wullaert, H., and Wright, S. J.: Immediate and long-
34 term nitrogen oxide emissions from tropical forest soils exposed to elevated nitrogen input,
35 *Global Change Biol.*, 15, 2049-2066, doi:10.1111/j.1365-2486.2008.01826.x, 2009.
- 36 Luizao, F., Matson, P., Livingston, G., Luizao, R., and Vitousek, P.: Nitrous oxide flux
37 following tropical land clearing, *Glob. Biogeochem. Cycles*, 3, 281-285,
38 doi:10.1029/GB003i003p00281, 1989.
- 39 Luo, G. J., Brüggemann, N., Wolf, B., Gasche, R., Grote, R., and Butterbach-Bahl, K.: Decadal
40 variability of soil CO₂, NO, N₂O, and CH₄ fluxes at the Hölglwald Forest, Germany,
41 *Biogeosciences*, 9, 1741-1763, doi:10.5194/bg-9-1741-2012, 2012.

- 1 Maljanen, M., Jokinen, H., Saari, A., Strommer, R., and Martikainen, P. J.: Methane and nitrous
2 oxide fluxes, and carbon dioxide production in boreal forest soil fertilized with wood ash and
3 nitrogen, *Soil Use Manage.*, 22, 151-157, doi:10.1111/j.1475-2743.2006.00029.x, 2006.
- 4 Matson, A., Pennock, D., and Bedard-Haughn, A.: Methane and nitrous oxide emissions from
5 mature forest stands in the boreal forest, Saskatchewan, Canada, *Forest Ecol. Manag.*, 258,
6 1073-1083, doi:10.1016/j.foreco.2009.05.034, 2009.
- 7 Matson, P. A., Volkman, C., Coppinger, K., and Reiners, W. A.: Annual nitrous-oxide flux
8 and soil-nitrogen characteristics in sagebrush steppe ecosystems, *Biogeochemistry*, 14, 1-12,
9 1991.
- 10 Matson, P. A., Gower, S. T., Volkman, C., Billow, C., and Grier, C. C.: Soil nitrogen cycling
11 and nitrous oxide flux in a Rocky Mountain Douglas-fir forest: effects of fertilization, irrigation
12 and carbon addition, *Biogeochemistry*, 18, 101-117, doi:10.1007/bf00002705, 1992.
- 13 Melling, L., Hatano, R., and Goh, K. J.: Nitrous oxide emissions from three ecosystems in
14 tropical peatland of Sarawak, Malaysia, *Soil Sci. Plant Nutr.*, 53, 792-805, doi:10.1111/j.1747-
15 0765.2007.00196.x, 2007.
- 16 Mogge, B., Kaiser, E. A., and Munch, J. C.: Nitrous oxide emissions and denitrification N-
17 losses from forest soils in the Bornhöved Lake region (Northern Germany), *Soil Biol.*
18 *Biochem.*, 30, 703-710, doi:10.1016/s0038-0717(97)00205-8, 1998.
- 19 Mosier, A. R., Parton, W. J., Valentine, D. W., Ojima, D. S., Schimel, D. S., and Delgado, J.
20 A.: CH₄ and N₂O fluxes in the Colorado shortgrass steppe .1. Impact of landscape and nitrogen
21 addition, *Glob. Biogeochem. Cycles*, 10, 387-399, doi:10.1029/96gb01454, 1996.
- 22 Mummey, D. L., Smith, J. L., and Bolton, H.: Small-scale spatial and temporal variability of
23 N₂O flux from a shrub-steppe ecosystem, *Soil Biol. Biochem.*, 29, 1699-1706,
24 doi:10.1016/s0038-0717(97)00077-1, 1997.
- 25 Parton, W. J., Mosier, A. R., and Schimel, D. S.: Rates and pathways of nitrous-oxide
26 production in a shortgrass steppe, *Biogeochemistry*, 6, 45-58, 1988.
- 27 Pei, Z. Y.: Carbon dynamics in the alpine grassland ecosystem on the tibetan plateau—a case
28 study of wudaoliang, qinghai province, PhD thesis, Institute of Geographic Sciences and
29 Natural Resources Research, Beijing, China, 2003.
- 30 Price, S. J., Sherlock, R. R., Kelliher, F. M., McSeveny, T. M., Tate, K. R., and Condon, L.
31 M.: Pristine new zealand forest soil is a strong methane sink, *Global Change Biol.*, 10, 16-26,
32 doi:10.1046/j.1529-8817.2003.00710x, 2004.
- 33 Purbopuspito, J., Veldkamp, E., Brumme, R., and Murdiyarso, D.: Trace gas fluxes and
34 nitrogen cycling along an elevation sequence of tropical montane forests in Central Sulawesi,
35 Indonesia, *Glob. Biogeochem. Cycles*, 20, GB3010, doi:10.1029/2005gb002516, 2006.
- 36 Rees, R. M., Wuta, M., Furley, P. A., and Li, C. S.: Nitrous oxide fluxes from savanna
37 (miombo) woodlands in Zimbabwe, *J. Biogeogr.*, 33, 424-437, 2005.
- 38 Rowlings, D. W., Grace, P. R., Kiese, R., and Weier, K. L.: Environmental factors controlling
39 temporal and spatial variability in the soil-atmosphere exchange of CO₂, CH₄ and N₂O from an
40 Australian subtropical rainforest, *Global Change Biol.*, 18, 726-738, doi:10.1111/j.1365-
41 2486.2011.02563.x, 2012.

1 Schiller, C. L., and Hastie, D. R.: Nitrous oxide and methane fluxes from perturbed and
2 unperturbed boreal forest sites in northern Ontario, *J. Geophys. Res.-Atmos.*, 101, 22767-
3 22774, doi:10.1029/96jd01620, 1996.

4 Simona, C., Ariangelo, D. P. R., John, G., Nina, N., Ruben, M., and Jose, S. J.: Nitrous oxide
5 and methane fluxes from soils of the Orinoco savanna under different land uses, *Global Change*
6 *Biol.*, 10, 1947-1960, doi:10.1111/j.1365-2486.2004.00871.x, 2004.

7 Simpson, I. J., Edwards, G. C., Thurtell, G. W., den Hartog, G., Neumann, H. H., and Staebler,
8 R. M.: Micrometeorological measurements of methane and nitrous oxide exchange above a
9 boreal aspen forest, *J. Geophys. Res.-Atmos.*, 102, 29331-29341, doi:10.1029/97jd03181,
10 1997.

11 Sitaula, B. K., Bakken, L. R., and Abrahamsen, G.: N-fertilization and soil acidification effects
12 on N₂O and CO₂ emission from temperate pine forest soil, *Soil Biol. Biochem.*, 27, 1401-1408,
13 doi:10.1016/0038-0717(95)00078-s, 1995.

14 Struwe, S., and Kjoller, A.: Field determination of denitrification in water-logged forest soils,
15 *FEMS Microbiol. Ecol.*, 62, 71-78, 1989.

16 Takakai, F., Morishita, T., Hashidoko, Y., Darung, U., Kuramochi, K., Dohong, S., Limin, S.
17 H., and Hatano, R.: Effects of agricultural land-use change and forest fire on N₂O emission
18 from tropical peatlands, Central Kalimantan, Indonesia, *Soil Sci. Plant Nutr.*, 52, 662-674,
19 doi:10.1111/j.1747-0765.2006.00084.x, 2006.

20 Tauchnitz, N., Brumme, R., Bernsdorf, S., and Meissner, R.: Nitrous oxide and methane fluxes
21 of a pristine slope mire in the German National Park Harz Mountains, *Plant and Soil*, 303, 131-
22 138, doi:10.1007/s11104-007-9493-0, 2008.

23 Templer, P. H., Mack, M. C., Chapin, F. S., III, Christenson, L. M., Compton, J. E., Crook, H.
24 D., Currie, W. S., Curtis, C. J., Dail, D. B., D'Antonio, C. M., Emmett, B. A., Epstein, H. E.,
25 Goodale, C. L., Gundersen, P., Hobbie, S. E., Holland, K., Hooper, D. U., Hungate, B. A.,
26 Lamontagne, S., Nadelhoffer, K. J., Osenberg, C. W., Perakis, S. S., Schleppei, P., Schimel, J.,
27 Schmidt, I. K., Sommerkorn, M., Spoelstra, J., Tietema, A., Wessel, W. W., and Zak, D. R.:
28 Sinks for nitrogen inputs in terrestrial ecosystems: A meta-analysis of ¹⁵N tracer field studies,
29 *Ecology*, 93, 1816-1829, 2012.

30 Ullah, S., and Moore, T. R.: Biogeochemical controls on methane, nitrous oxide, and carbon
31 dioxide fluxes from deciduous forest soils in eastern Canada, *J. Geophys. Res.-Biogeo.*, 116,
32 GB3010, doi:10.1029/2010jg001525, 2011.

33 Verchot, L. V., Davidson, E. A., Cattanio, J. H., Ackerman, I. L., Erickson, H. E., and Keller,
34 M.: Land use change and biogeochemical controls of nitrogen oxide emissions from soils in
35 eastern Amazonia, *Glob. Biogeochem. Cycles*, 13, 31-46, doi:10.1029/1998gb900019, 1999.

36 von Arnold, K., Nilsson, M., Hanell, B., Weslien, P., and Klemedtsson, L.: Fluxes of CO₂, CH₄
37 and N₂O from drained organic soils in deciduous forests, *Soil Biol. Biochem.*, 37, 1059-1071,
38 doi:10.1016/j.soilbio.2004.11.004, 2005.

39 Wolf, K., Veldkamp, E., Homeier, J., and Martinson, G. O.: Nitrogen availability links forest
40 productivity, soil nitrous oxide and nitric oxide fluxes of a tropical montane forest in southern
41 Ecuador, *Glob. Biogeochem. Cycles*, 25, doi:10.1029/2010gb003876, 2011.

1 Yu, J., Liu, J., Wang, J., Sun, W., Patrick, W. H., Jr., and Meixner, F. X.: Nitrous oxide emission
2 from *deyeuxia angustifolia* freshwater marsh in northeast China, *Environ. Manage.*, 40, 613-
3 622, doi:10.1007/s00267-006-0349-9, 2007.
4

1 **Acknowledgements**

2 The soil moisture data used in this study were acquired as part of the mission of NASA's Earth
3 Science Division and archived and distributed by the Goddard Earth Sciences (GES) Data and
4 Information Services Center (DISC). We thank the European Centre for Medium-Range
5 Weather Forecasts for providing the reanalysed soil moisture dataset and the Oak Ridge
6 National Laboratory (ORNL) Distributed Active Archive Center (DAAC) for sharing N₂O
7 observation and soil property dataset. We would like to thank Matthew J. Cohen, Patrick Inglett
8 and Jeremy W. Lichstein for their constructive comments throughout the study. We would also
9 like to thank Lex Bouwman, Benjamin Stocker and an anonymous reviewer for constructive
10 comments and suggestions.

11 **References**

- 12 Andreae, M. O., and Merlet, P.: Emission of trace gases and aerosols from biomass burning,
13 *Glob. Biogeochem. Cycles*, 15, 955-966, doi:10.1029/2000gb001382, 2001.
- 14 Banin, A.: Global budget of N₂O: The role of soils and their change, *Sci. Total Environ.*, 55,
15 27-38, doi:10.1016/0048-9697(86)90166-x, 1986.
- 16 Bolker, B. M., Pacala, S. W., and Parton, W. J.: Linear analysis of soil decomposition: Insights
17 from the century model, *Ecol. Appl.*, 8, 425-439, doi:10.2307/2641082, 1998.
- 18 Bouwman, A. F., Vanderhoek, K. W., and Olivier, J. G. J.: Uncertainties in the global source
19 distribution of nitrous oxide, *J. Geophys. Res.-Atmos.*, 100, 2785-2800, doi:10.1029/94jd02946,
20 1995.
- 21 Bowden, W. B.: Gaseous nitrogen emissions from undisturbed terrestrial ecosystems: An
22 assessment of their impacts on local and global nitrogen budgets, *Biogeochemistry*, 2, 249-279,
23 doi:10.1007/bf02180161, 1986.
- 24 Braker, G., and Conrad, R.: Diversity, structure, and size of N₂O-producing microbial
25 communities in soils-what matters for their functioning?, in: *Advances in Applied*
26 *Microbiology*, Vol 75, edited by: Laskin, A. I., Sariaslani, S., and Gadd, G. M., *Advances in*
27 *Applied Microbiology*, 33-70, 2011.
- 28 Brown, J. R., Blankinship, J. C., Niboyet, A., van Groenigen, K. J., Dijkstra, P., Le Roux, X.,
29 Leadley, P. W., and Hungate, B. A.: Effects of multiple global change treatments on soil N₂O
30 fluxes, *Biogeochemistry*, 109, 85-100, doi:10.1007/s10533-011-9655-2, 2012.
- 31 Butterbach-Bahl, K., Baggs, E. M., Dannenmann, M., Kiese, R., and Zechmeister-Boltenstern,
32 S.: Nitrous oxide emissions from soils: how well do we understand the processes and their
33 controls?, *Philosophical Transactions of the Royal Society B-Biological Sciences*, 368,
34 20130122–20130122, doi:10.1098/rstb.2013.0122, 2013.
- 35 Ciais, P., Sabine, C., Bala, G., Bopp, L., Brovkin, V., Canadell, J., Chhabra, A., DeFries, R.,
36 Galloway, J., Heimann, M., Jones, C., Quéré, C. L., Myneni, R. B., Piao, S., and Thornton, P.:
37 Carbon and Other Biogeochemical Cycles, in: *Climate Change 2013: The Physical Science*
38 *Basis. Contribution of Working Group I to the Fifth Assessment Report of the*

1 Intergovernmental Panel on Climate Change, edited by: Stocker, T. F., Qin, D., Plattner, G.-K.,
2 Tignor, M., Allen, S. K., Boschung, J., Nauels, A., Xia, Y., Bex, V., and Midgley, P. M.,
3 Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, 2013.

4 Cleveland, C. C., Townsend, A. R., Schimel, D. S., Fisher, H., Howarth, R. W., Hedin, L. O.,
5 Perakis, S. S., Latty, E. F., Von Fischer, J. C., Elseroad, A., and Wasson, M. F.: Global patterns
6 of terrestrial biological nitrogen (N₂) fixation in natural ecosystems, *Global Biogeochemical*
7 *Cycles*, 13, 623-645, 10.1029/1999gb900014, 1999.

8 Collatz, G. J., Ball, J. T., Grivet, C., and Berry, J. A.: Physiological and environmental
9 regulation of stomatal conductance, photosynthesis and transpiration: a model that includes a
10 laminar boundary layer, *Agr. Forest Meteorol.*, 54, 107-136, doi:10.1016/0168-
11 1923(91)90002-8, 1991.

12 Collatz, G. J., Ribas-Carbo, M., and Berry, J. A.: Coupled photosynthesis-stomatal conductance
13 model for leaves of C4 plants, *Aust. J. Plant Physiol.*, 19, 519-538, 1992.

14 Davidson, E. A., and Trumbore, S. E.: Gas diffusivity and production of CO₂ in deep soils of
15 the eastern Amazon, *Tellus Series B-Chemical and Physical Meteorology*, 47, 550-565,
16 doi:10.1034/j.1600-0889.47.issue5.3.x, 1995.

17 Davidson, E. A., Ishida, F. Y., and Nepstad, D. C.: Effects of an experimental drought on soil
18 emissions of carbon dioxide, methane, nitrous oxide, and nitric oxide in a moist tropical forest,
19 *Global Change Biology*, 10, 718-730, doi:10.1111/j.1365-2486.2004.00762.x, 2004.

20 Davidson, E. A., Nepstad, D. C., Ishida, F. Y., and Brando, P. M.: Effects of an experimental
21 drought and recovery on soil emissions of carbon dioxide, methane, nitrous oxide, and nitric
22 oxide in a moist tropical forest, *Global Change Biology*, 14, 2582-2590, 10.1111/j.1365-
23 2486.2008.01694.x, 2008.

24 Davidson, E. A.: The contribution of manure and fertilizer nitrogen to atmospheric nitrous oxide
25 since 1860, *Nature Geoscience*, 2, 659-662, doi:10.1038/ngeo608, 2009.

26 Dee, D. P., Uppala, S. M., Simmons, A. J., Berrisford, P., Poli, P., Kobayashi, S., Andrae, U.,
27 Balmaseda, M. A., Balsamo, G., Bauer, P., Bechtold, P., Beljaars, A. C. M., van de Berg, L.,
28 Bidlot, J., Bormann, N., Delsol, C., Dragani, R., Fuentes, M., Geer, A. J., Haimberger, L., Healy,
29 S. B., Hersbach, H., Holm, E. V., Isaksen, I., Kallberg, P., Koehler, M., Matricardi, M.,
30 McNally, A. P., Monge-Sanz, B. M., Morcrette, J. J., Park, B. K., Peubey, C., de Rosnay, P.,
31 Tavolato, C., Thepaut, J. N., and Vitart, F.: The ERA-Interim reanalysis: configuration and
32 performance of the data assimilation system, *Quarterly Journal of the Royal Meteorological*
33 *Society*, 137, 553-597, doi:10.1002/qj.828, 2011.

34 Del Grosso, S. J., Parton, W. J., Mosier, A. R., Ojima, D. S., Kulmala, A. E., and Phongpan, S.:
35 General model for N₂O and N₂ gas emissions from soils due to denitrification, *Glob.*
36 *Biogeochem. Cycles*, 14, 1045-1060, 2000.

37 Dentener, F., Drevet, J., Lamarque, J. F., Bey, I., Eickhout, B., Fiore, A. M., Hauglustaine, D.,
38 Horowitz, L. W., Krol, M., Kulshrestha, U. C., Lawrence, M., Galy-Lacaux, C., Rast, S.,
39 Shindell, D., Stevenson, D., Van Noije, T., Atherton, C., Bell, N., Bergman, D., Butler, T.,
40 Cofala, J., Collins, B., Doherty, R., Ellingsen, K., Galloway, J., Gauss, M., Montanaro, V.,
41 Mueller, J. F., Pitari, G., Rodriguez, J., Sanderson, M., Solmon, F., Strahan, S., Schultz, M.,
42 Sudo, K., Szopa, S., and Wild, O.: Nitrogen and sulfur deposition on regional and global scales:
43 A multimodel evaluation, *Global Biogeochemical Cycles*, 20,
44 doi:10.1029/2005gb002672, 2006.

1 Dentener, F. J., and Crutzen, P. J.: A three-dimensional model of the global ammonia cycle, *J.*
2 *Atmos. Chem.*, 19, 331-369, doi:10.1007/bf00694492, 1994.

3 Dijkstra, F. A., Prior, S. A., Runion, G. B., Torbert, H. A., Tian, H., Lu, C., and Venterea, R.
4 T.: Effects of elevated carbon dioxide and increased temperature on methane and nitrous oxide
5 fluxes: evidence from field experiments, *Frontiers in Ecology and the Environment*, 10, 520-
6 527, doi:10.1890/120059, 2012.

7 Farquhar, G. D., Caemmerer, S. V., and Berry, J. A.: A biochemical model of photosynthetic
8 CO₂ assimilation in leaves of C₃ species, *Planta*, 149, 78-90, doi:10.1007/bf00386231, 1980.

9 Firestone, M. K., and Davidson, E. A.: Microbiological basis of NO and N₂O production and
10 consumption in soil, *Exchange of trace gases between terrestrial ecosystems and the*
11 *atmosphere.*, edited by: Andreae, M. O., and Schimel, D. S., 7-21 pp., 1989.

12 Forster, P., Ramaswamy, V., Artaxo, P., Berntsen, T., Betts, R., Fahey, D. W., Haywood, J.,
13 Lean, J., Lowe, D. C., Myhre, G., Nganga, J., Prinn, R., Raga, G., Schulz, M., and Dorland, R.
14 V.: Changes in Atmospheric Constituents and in Radiative Forcing, in: *Climate Change 2007:*
15 *The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report*
16 *of the Intergovernmental Panel on Climate Change*, edited by: Solomon, S., Qin, D., Manning,
17 M., Chen, Z., Marquis, M., Averyt, K. B., M.Tignor, and Miller, H. L., Cambridge University
18 Press, Cambridge, United Kingdom and New York, NY, USA, 2007.

19 Galloway, J. N., Aber, J. D., Erisman, J. W., Seitzinger, S. P., Howarth, R. W., Cowling, E. B.,
20 and Cosby, B. J.: The nitrogen cascade, *Bioscience*, 53, 341-356, doi:10.1641/0006-
21 3568(2003)053[0341:tnc]2.0.co;2, 2003.

22 Gerber, S., and Brookshire, E. N. J.: Scaling of physical constraints at the root-soil interface to
23 macroscopic patterns of nutrient retention in ecosystems, *Am. Nat.*, 183, 418-430,
24 doi:10.1086/674907, 2014.

25 Gerber, S., Hedin, L. O., Keel, S. G., Pacala, S. W., and Shevliakova, E.: Land use change and
26 nitrogen feedbacks constrain the trajectory of the land carbon sink, *Geophysical Research*
27 *Letters*, 40, 5218-5222, doi:10.1002/grl.50957, 2013.

28 Gerber, S., Hedin, L. O., Oppenheimer, M., Pacala, S. W., and Shevliakova, E.: Nitrogen
29 cycling and feedbacks in a global dynamic land model, *Glob. Biogeochem. Cycles*, 24,
30 doi:10.1029/2008gb003336, 2010.

31 Goodroad, L. L., and Keeney, D. R.: Nitrous oxide emission from forest, marsh, and prairie
32 ecosystems, *J. Environ. Qual.*, 13, 448-452, 1984.

33 Green, P. A., Vorosmarty, C. J., Meybeck, M., Galloway, J. N., Peterson, B. J., and Boyer, E.
34 W.: Pre-industrial and contemporary fluxes of nitrogen through rivers: a global assessment
35 based on typology, *Biogeochemistry*, 68, 71-105, doi:10.1023/b:biog.0000025742.82155.92,
36 2004.

37 Groffman, P. M., Hardy, J. P., Driscoll, C. T., and Fahey, T. J.: Snow depth, soil freezing, and
38 fluxes of carbon dioxide, nitrous oxide and methane in a northern hardwood forest, *Global*
39 *Change Biology*, 12, 1748-1760, 10.1111/j.1365-2486.2006.01194.x, 2006.

40 Heinen, M.: Simplified denitrification models: Overview and properties, *Geoderma*, 133, 444-
41 463, doi:10.1016/j.geoderma.2005.06.010, 2006.

1 Hirsch, A. I., Michalak, A. M., Bruhwiler, L. M., Peters, W., Dlugokencky, E. J., and Tans, P.
2 P.: Inverse modeling estimates of the global nitrous oxide surface flux from 1998-2001, *Glob.*
3 *Biogeochem. Cycles*, 20, doi:10.1029/2004gb002443, 2006.

4 Ishizuka, S., Tsuruta, H., and Murdiyarso, D.: An intensive field study on CO₂, CH₄, and N₂O
5 emissions from soils at four land-use types in Sumatra, Indonesia, *Global Biogeochemical*
6 *Cycles*, 16, 10.1029/2001gb001614, 2002.

7 Keeling, R. F., Piper, S. C., Bollenbacher, A. F., and Walker, J. S.: Atmospheric CO₂ records
8 from sites in the SIO air sampling network, in *Trends: A compendium of data on global change.*
9 Carbon Dioxide Information Analysis Center, Oak Ridge National Laboratory, U.S.
10 Department of Energy, Oak Ridge, Tenn., U.S.A [online] Available from:
11 <http://cdiac.ornl.gov/trends/co2/sio-mlo.html> (Accessed 13 August 2010), 2009.

12 Khalil, K., Mary, B., and Renault, P.: Nitrous oxide production by nitrification and
13 denitrification in soil aggregates as affected by O₂ concentration, *Soil Biol. Biochem.*, 36, 687-
14 699, doi:10.1016/j.soilbio.2004.01.004, 2004.

15 Li, C. S., Frolking, S., and Frolking, T. A.: A model of nitrous-oxide evolution from soil driven
16 by rainfall events .1. model structure and sensitivity, *J. Geophys. Res.-Atmos.*, 97, 9759-9776,
17 1992.

18 Li, C. S., Aber, J., Stange, F., Butterbach-Bahl, K., and Papen, H.: A process-oriented model of
19 N₂O and NO emissions from forest soils: 1. Model development, *J. Geophys. Res.-Atmos.*, 105,
20 4369-4384, doi:10.1029/1999jd900949, 2000.

21 Linn, D. M., and Doran, J. W.: Effect of water-filled pore space on carbon dioxide and nitrous
22 oxide production in tilled and nontilled soils, *Soil Sci. Soc. Am. J.*, 48, 1267-1272, 1984.

23 Milly, P. C. D., and Shmakin, A. B.: Global modeling of land water and energy balances. Part
24 I: the land dynamics (LaD) model, *J. Hydrometeorol.*, 3, 283-299, doi:10.1175/1525-
25 7541(2002)003<0283:gmolwa>2.0.co;2, 2002.

26 Milly, P. C. D., Malyshev, S. L., Shevliakova, E., Dunne, K. A., Findell, K. L., Gleeson, T.,
27 Liang, Z., Philipps, P., Stouffer, R. J., and Swenson, S.: An enhanced model of land water and
28 energy for global hydrologic and Earth-System studies, *J. Hydrometeorol.*, 15, 1739-1761,
29 2014.

30 Morishita, T., Sakata, T., Takahashi, M., Ishizuka, S., Mizoguchi, T., Inagaki, Y., Terazawa,
31 K., Sawata, S., Igarashi, M., Yasuda, H., Koyama, Y., Suzuki, Y., Toyota, N., Muro, M., Kinjo,
32 M., Yamamoto, H., Ashiya, D., Kanazawa, Y., Hashimoto, T., and Umata, H.: Methane uptake
33 and nitrous oxide emission in Japanese forest soils and their relationship to soil and vegetation
34 types, *Soil Science and Plant Nutrition*, 53, 678-691, 10.1111/j.1747-0765.2007.00181.x, 2007.

35 Parton, W. J., Mosier, A. R., Ojima, D. S., Valentine, D. W., Schimel, D. S., Weier, K., and
36 Kulmala, A. E.: Generalized model for N₂ and N₂O production from nitrification and
37 denitrification, *Glob. Biogeochem. Cycles*, 10, 401-412, doi:10.1029/96gb01455, 1996.

38 Parton, W. J., Holland, E. A., Del Grosso, S. J., Hartman, M. D., Martin, R. E., Mosier, A. R.,
39 Ojima, D. S., and Schimel, D. S.: Generalized model for NO_x and N₂O emissions from soils, *J.*
40 *Geophys. Res.-Atmos.*, 106, 17403-17419, doi:10.1029/2001jd900101, 2001.

41 Potter, C. S., Matson, P. A., Vitousek, P. M., and Davidson, E. A.: Process modeling of controls
42 on nitrogen trace gas emissions from soils worldwide, *J. Geophys. Res.-Atmos.*, 101, 1361-
43 1377, doi:10.1029/95jd02028, 1996.

1 Potter, C. S., and Klooster, S. A.: Interannual variability in soil trace gas (CO₂, N₂O, NO) fluxes
2 and analysis of controllers on regional to global scales, *Glob. Biogeochem. Cycles*, 12, 621-
3 635, doi:10.1029/98gb02425, 1998.

4 Purbopuspito, J., Veldkamp, E., Brumme, R., and Murdiyarso, D.: Trace gas fluxes and
5 nitrogen cycling along an elevation sequence of tropical montane forests in Central Sulawesi,
6 Indonesia, *Glob. Biogeochem. Cycles*, 20, doi:10.1029/2005gb002516, 2006.

7 Ravishankara, A. R., Daniel, J. S., and Portmann, R. W.: Nitrous oxide (N₂O): The dominant
8 ozone-depleting substance emitted in the 21st century, *Science*, 326, 123-125,
9 doi:10.1126/science.1176985, 2009.

10 Rodell, M., Houser, P. R., Jambor, U., Gottschalck, J., Mitchell, K., Meng, C. J., Arsenault, K.,
11 Cosgrove, B., Radakovich, J., Bosilovich, M., Entin, J. K., Walker, J. P., Lohmann, D., and
12 Toll, D.: The global land data assimilation system, *Bulletin of the American Meteorological*
13 *Society*, 85, 381-394, doi:10.1175/bams-85-3-381, 2004.

14 Saikawa, E., Schlosser, C. A., and Prinn, R. G.: Global modeling of soil nitrous oxide emissions
15 from natural processes, *Glob. Biogeochem. Cycles*, 27, 972-989, doi:10.1002/gbc.20087, 2013.

16 Schlesinger, W. H.: On the fate of anthropogenic nitrogen, *Proceedings of the National*
17 *Academy of Sciences of the United States of America*, 106, 203-208,
18 doi:10.1073/pnas.0810193105, 2009.

19 Schmidt, J., Seiler, W., and Conrad, R.: Emission of nitrous oxide from temperate forest soils
20 into the atmosphere, *Journal of Atmospheric Chemistry*, 6, 95-115, 10.1007/bf00048334, 1988.

21 Schwalm, C. R., Williams, C. A., Schaefer, K., Baker, I., Collatz, G. J., and Roedenbeck, C.:
22 Does terrestrial drought explain global CO₂ flux anomalies induced by El Niño?,
23 *Biogeosciences*, 8, 2493-2506, doi:10.5194/bg-8-2493-2011, 2011.

24 Seneviratne, S. I., Corti, T., Davin, E. L., Hirschi, M., Jaeger, E. B., Lehner, I., Orlowsky, B.,
25 and Teuling, A. J.: Investigating soil moisture-climate interactions in a changing climate: A
26 review, *Earth-Sci. Rev.*, 99, 125-161, doi:10.1016/j.earscirev.2010.02.004, 2010.

27 Sheffield, J., Goteti, G., and Wood, E. F.: Development of a 50-year high-resolution global
28 dataset of meteorological forcings for land surface modeling, *J. Clim.*, 19, 3088-3111,
29 doi:10.1175/jcli3790.1, 2006.

30 Shevliakova, E., Pacala, S. W., Malyshev, S., Hurtt, G. C., Milly, P. C. D., Caspersen, J. P.,
31 Sentman, L. T., Fisk, J. P., Wirth, C., and Crevoisier, C.: Carbon cycling under 300 years of
32 land use change: Importance of the secondary vegetation sink, *Glob. Biogeochem. Cycles*, 23,
33 doi:10.1029/2007gb003176, 2009.

34 [Smith, B., Warlind, D., Arneeth, A., Hickler, T., Leadley, P., Siltberg, J., and Zaehle, S.:](#)
35 [Implications of incorporating N cycling and N limitations on primary production in an](#)
36 [individual-based dynamic vegetation model, *Biogeosciences*, 11, 2027-2054, doi:10.5194/bg-](#)
37 [11-2027-2014, 2014.](#)

38 Sousa Neto, E., Carmo, J. B., Keller, M., Martins, S. C., Alves, L. F., Vieira, S. A., Piccolo, M.
39 C., Camargo, P., Couto, H. T. Z., Joly, C. A., and Martinelli, L. A.: Soil-atmosphere exchange
40 of nitrous oxide, methane and carbon dioxide in a gradient of elevation in the coastal Brazilian
41 Atlantic forest, *Biogeosciences*, 8, 733-742, doi:10.5194/bg-8-733-2011, 2011.

42 Stehfest, E., and Bouwman, L.: N₂O and NO emission from agricultural fields and soils under
43 natural vegetation: summarizing available measurement data and modeling of global annual

1 emissions, *Nutrient Cycling in Agroecosystems*, 74, 207-228, doi:10.1007/s10705-006-9000-
2 7, 2006.

3 Stocker, B. D., Roth, R., Joos, F., Spahni, R., Steinacher, M., Zaehle, S., Bouwman, L., Xu-Ri,
4 and Prentice, I. C.: Multiple greenhouse-gas feedbacks from the land biosphere under future
5 climate change scenarios, *Nature Climate Change*, 3, 666-672, doi:10.1038/nclimate1864, 2013.

6 Syakila, A., and Kroeze, C.: The global nitrous oxide budget revisited, *Greenhouse Gas
7 Measurement and Management*, 1, 17-26, doi:10.3763/ghgmm.2010.0007, 2011.

8 Thomas, R. Q., Brookshire, E. N. J., and Gerber, S.: Nitrogen limitation on land: how can it
9 occur in Earth system models?, *Global Change Biol.*, 21, 1777-1793, doi:10.1111/gcb.12813,
10 2015.

11 Thompson, R. L., Chevallier, F., Crotwell, A. M., Dutton, G., Langenfelds, R. L., Prinn, R. G.,
12 Weiss, R. F., Tohjima, Y., Nakazawa, T., Krummel, P. B., Steele, L. P., Fraser, P., O'Doherty,
13 S., Ishijima, K., and Aoki, S.: Nitrous oxide emissions 1999 to 2009 from a global atmospheric
14 inversion, *Atmospheric Chemistry and Physics*, 14, 1801-1817, doi:10.5194/acp-14-1801-2014,
15 2014.

16 Thornton, P. E., Lamarque, J.-F., Rosenbloom, N. A., and Mahowald, N. M.: Influence of
17 carbon-nitrogen cycle coupling on land model response to CO₂ fertilization and climate
18 variability, *Glob. Biogeochem. Cycles*, 21, doi:10.1029/2006gb002868, 2007.

19 van Groenigen, K. J., Osenberg, C. W., and Hungate, B. A.: Increased soil emissions of potent
20 greenhouse gases under increased atmospheric CO₂, *Nature*, 475, 214-U121,
21 doi:10.1038/nature10176, 2011.

22 Wei, Y., Liu, S., Huntzinger, D.N., Michalak, A.M., Viovy, N., Post, W.M., Schwalm, C.R.,
23 Schaefer, K., Jacobson, A.R., Lu, C., Tian, H., Ricciuto, D.M., Cook, R.B., Mao, J., and Shi,
24 X. : NACP MsTMIP: Global and North American Driver Data for Multi-Model
25 Intercomparison, Data set. Available on-line [<http://daac.ornl.gov>] from Oak Ridge National
26 Laboratory Distributed Active Archive Center, Oak Ridge, Tennessee, USA. Available at:
27 <http://dx.doi.org/10.3334/ORNLDAAC/1220>, 2014

28 Werner, C., Butterbach-Bahl, K., Haas, E., Hickler, T., and Kiese, R.: A global inventory of
29 N₂O emissions from tropical rainforest soils using a detailed biogeochemical model, *Glob.
30 Biogeochem. Cycles*, 21, GB3010, doi:10.1029/2006gb002909, 2007.

31 Xu-Ri, Wang, Y. S., Zheng, X. H., Ji, B. M., and Wang, M. X.: A comparison between
32 measured and modeled N₂O emissions from Inner Mongolian semi-arid grassland, *Plant and
33 Soil*, 255, 513-528, 2003.

34 Xu-Ri and Prentice, I. C.: Terrestrial nitrogen cycle simulation with a dynamic global
35 vegetation model, *Global Change Biology*, 14, 1745-1764, doi:10.1111/j.1365-
36 2486.2008.01625.x, 2008.

37 Xu-Ri, Prentice, I. C., Spahni, R., and Niu, H. S.: Modelling terrestrial nitrous oxide emissions
38 and implications for climate feedback, *New Phytologist*, 196, 472-488, doi:10.1111/j.1469-
39 8137.2012.04269.x, 2012.

40 Yienger, J. J., and Levy, H.: Empirical model of global soil-biogenic NO_x emissions, *J. Geophys.
41 Res.-Atmos.*, 100, 11447-11464, doi:10.1029/95jd00370, 1995.

1 Zaehle, S., and Friend, A. D.: Carbon and nitrogen cycle dynamics in the O-CN land surface
2 model: 1. Model description, site-scale evaluation, and sensitivity to parameter estimates, *Glob.*
3 *Biogeochem. Cycles*, 24, doi:10.1029/2009gb003521, 2010.

4 Zaehle, S., Friend, A. D., Friedlingstein, P., Dentener, F., Peylin, P., and Schulz, M.: Carbon
5 and nitrogen cycle dynamics in the O-CN land surface model: 2. Role of the nitrogen cycle in
6 the historical terrestrial carbon balance, *Global Biogeochemical Cycles*, 24,
7 doi:10.1029/2009gb003522, 2010.

8 Zaehle, S., Ciais, P., Friend, A. D., and Prieur, V.: Carbon benefits of anthropogenic reactive
9 nitrogen offset by nitrous oxide emissions, *Nat. Geosci.*, 4, 601-605, doi:10.1038/ngeo1207,
10 2011.

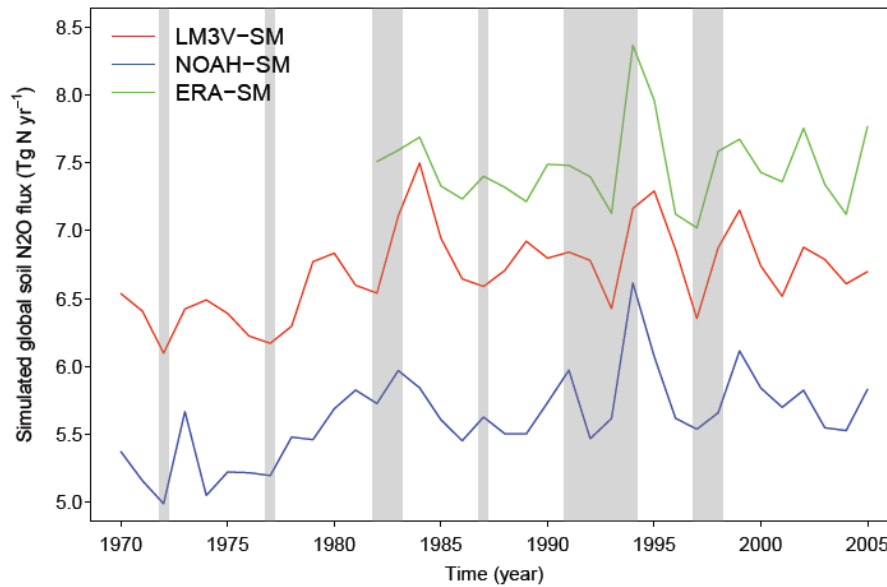
11 Zaehle, S., and Dalmonech, D.: Carbon-nitrogen interactions on land at global scales: current
12 understanding in modelling climate biosphere feedbacks, *Current Opinion in Environmental*
13 *Sustainability*, 3, 311-320, doi:10.1016/j.cosust.2011.08.008, 2011.

14 Zhuang, Q., Lu, Y., and Chen, M.: An inventory of global N₂O emissions from the soils of
15 natural terrestrial ecosystems, *Atmos. Environ.*, 47, 66-75,
16 doi:10.1016/j.atmosenv.2011.11.036, 2012.

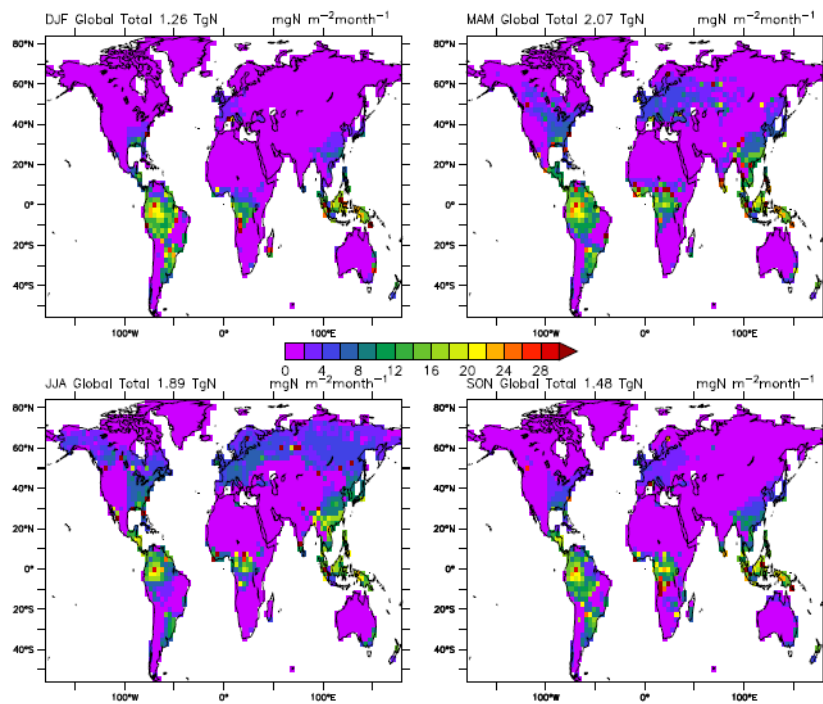
17

18

1 **Figures and Tables**

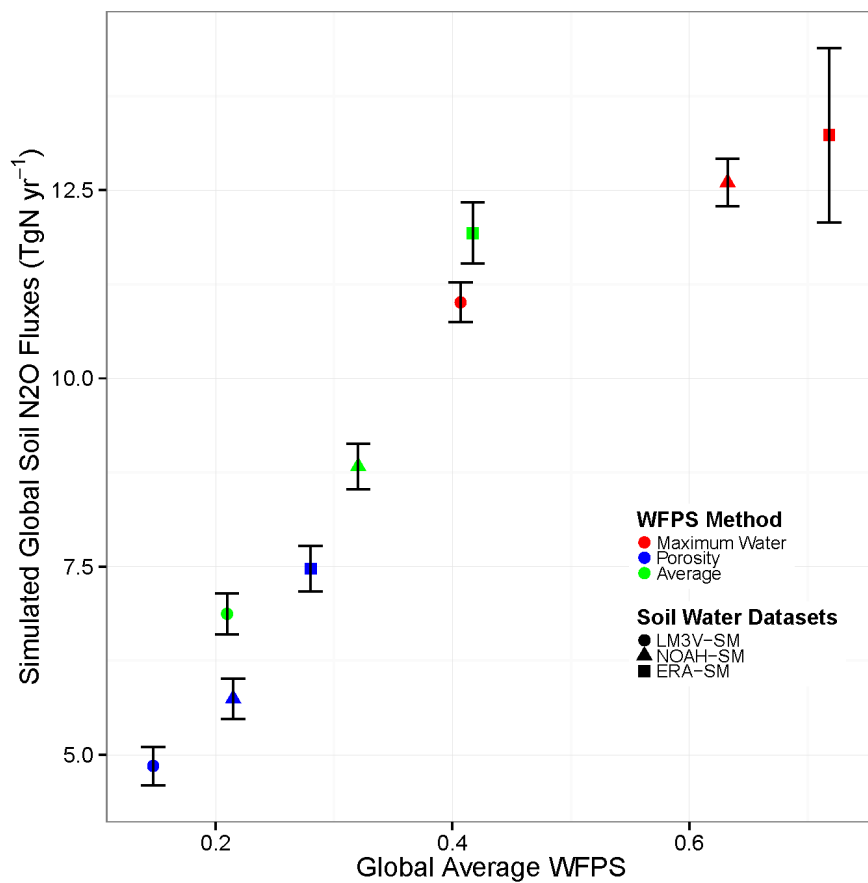


2
3 Figure 1. Simulated annual global soil N₂O emissions based on potential vegetation (1970-
4 2005). Shaded grey area indicates El Niño years with the annual multivariate ENSO index (MEI)
5 greater than 0.6. Colours refer to different soil moisture dataset used in the estimation: red for
6 LM3V-SM (with WFPS calculated by Method 3); blue for NOAH-SM (Method 2) and green
7 for ERA-SM (Method 2). Details for these soil moisture dataset and WFPS calculating methods
8 is available in the main text.
9

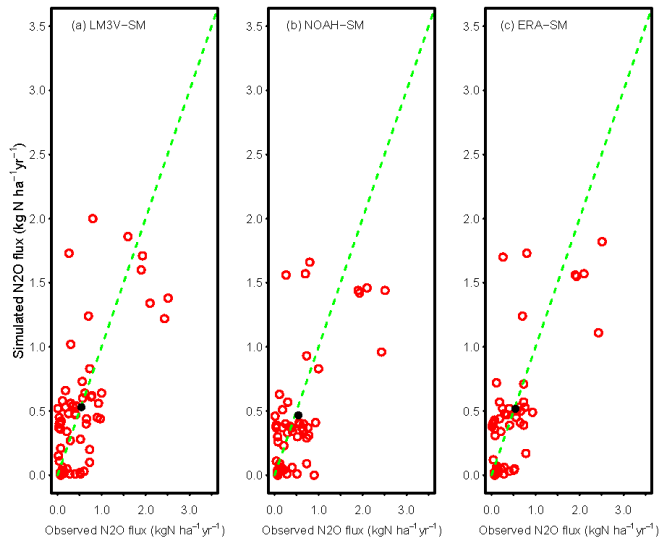


1
 2 Figure 2. Global seasonal mean soil N₂O emissions (with potential vegetation) averaged over
 3 the years 1970-2005. DJF (December, January and February), stands for Northern
 4 Hemisphere Winter; MAM (March, April and May) for Spring; JJA (June, July and August)
 5 for Summer; and SON (September, October and November) for Autumn.

6

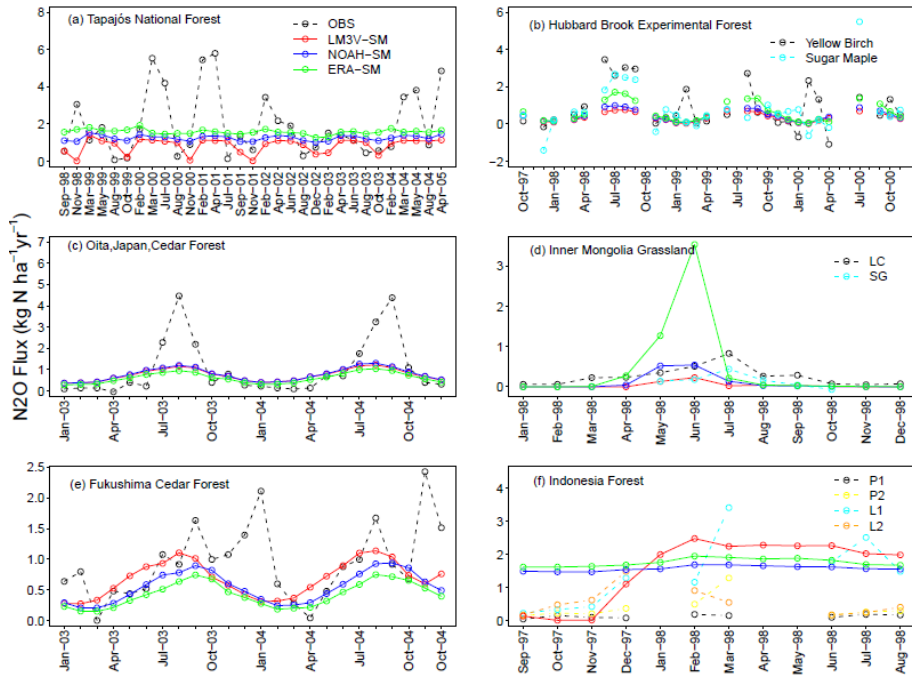


1
2 Figure 3. Sensitivity of simulated global soil N₂O emissions (with potential vegetation) to
3 water filled pore space (WFPS). The x-axis is the WFPS averaged globally over 1982-2005;
4 the y-axis represents the corresponding global total N₂O fluxes. A total of nine sets of WFPS
5 are obtained through either different soil water datasets (colours) or varied calculation
6 methods (symbols). Maximum water, porosity and average correspond to method 1, method 2
7 and method 3 in the main text, respectively. Coloured symbols represent means and error bars
8 indicate interannual standard deviations.



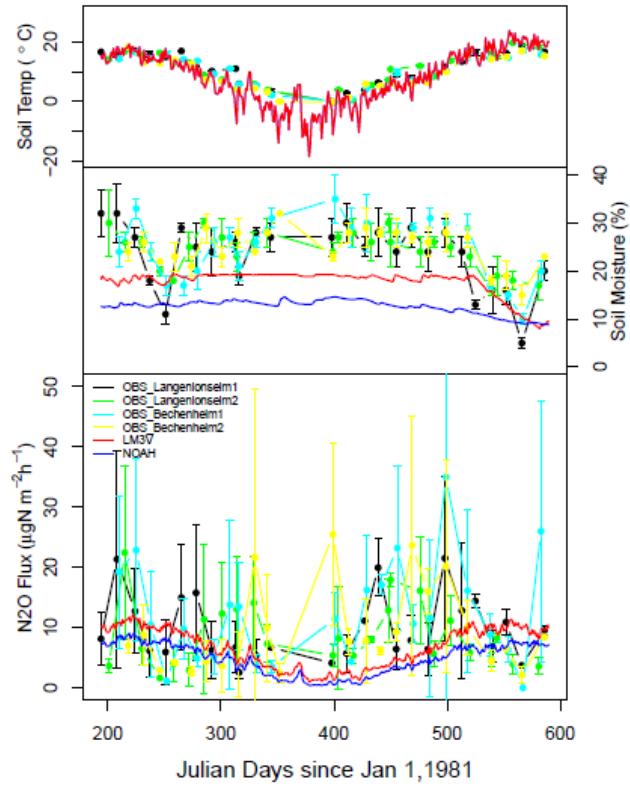
1
 2 Figure 4. Observed vs. simulated annual N₂O emissions from natural soils. Dashed green lines
 3 are the 1:1 lines. The solid circles represent the overall means. Different panels represent
 4 simulations with different soil moisture data: (a) LM3V-SM (simulated by LM3V-N); (b)
 5 NOAH-SM (based on land surface model NOAH 3.3 in Global Land Data Assimilation System
 6 Version 2); and (c) ERA-SM (reanalysis data from ECMWF). Water filled pore space (WFPS)
 7 is calculated using the average of the one based on available water capacity and the one based
 8 on the total porosity (Method 3, see the main text for detailed description) for panel (a); and
 9 using the total porosity (Method 2) for panel (b) and (c).

10



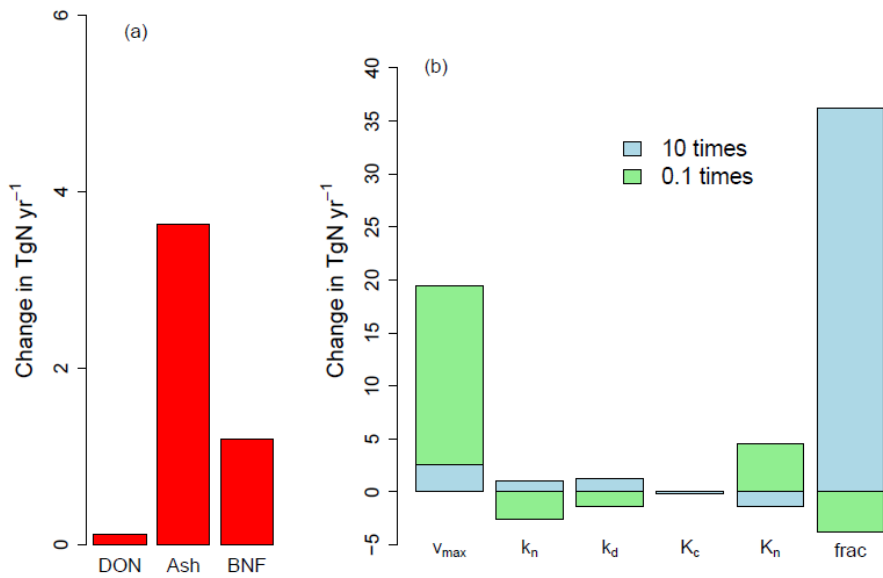
1
 2 Figure 5. Observed vs. simulated monthly N_2O emissions at (a), the Tapajós National Forest in
 3 east-central Amazonia ($3^{\circ}S$, $55^{\circ}W$), taken from Davidson et al. (2008); (b), the Hubbard Brook
 4 Experimental Forest in New Hampshire, USA ($44^{\circ}N$, $72^{\circ}W$), taken from Groffman et al. (2006);
 5 (c), a cedar forest at Oita, Japan ($33^{\circ}N$, $131^{\circ}E$), taken from Morishita et al. (2007) ; (d), the
 6 *Leymus chinensis* (LC) and *Stipa grandis* (SG) steppe in Inner Mongolia, China ($44^{\circ}N$, $117^{\circ}E$),
 7 taken from Xu-Ri et al. (2003); (e), a cedar forest in Fukushima, Japan ($37^{\circ}N$, $140^{\circ}E$), taken
 8 from Morishita et al. (2007); and (f), the primary (P1 and P2) and secondary (L1 and L2) forests
 9 located at the Pasir Mayang Research Site, Indonesia, taken from Ishizuka et al. (2002) ($1^{\circ}S$,
 10 $102^{\circ}E$). Shown are modeled results from three WFPS schemes (LM3V-SM, NOAH-SM and
 11 ERA-SM) the same as in Figure 4.

12
 13
 14



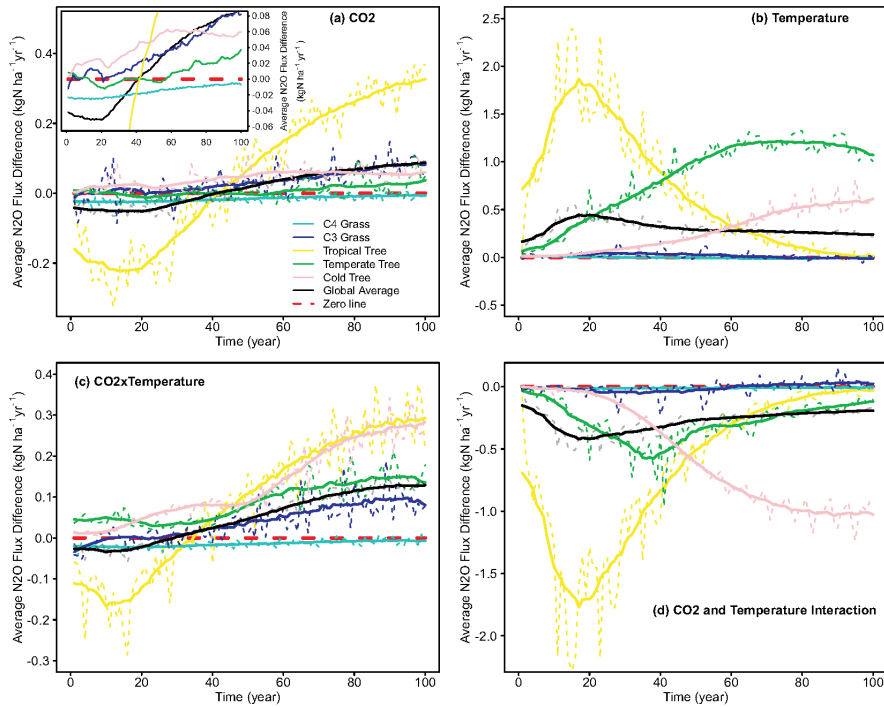
1
 2 Figure 6. Comparison of (a) soil temperature (2cm from observation and 1 cm from model)
 3 in °C; (b) soil moisture (2cm from observation and root zone from model) in % and (c) soil
 4 N₂O emissions in µgN m⁻² h⁻¹ from observations and model outputs at four forest sites from
 5 Germany (50°N, 8°E), taken from Schmidt et al. (1988). Shown are modeled results from two
 6 WFPS schemes (LM3V-SM and NOAH-SM) similar as in Figure 4.

7
 8

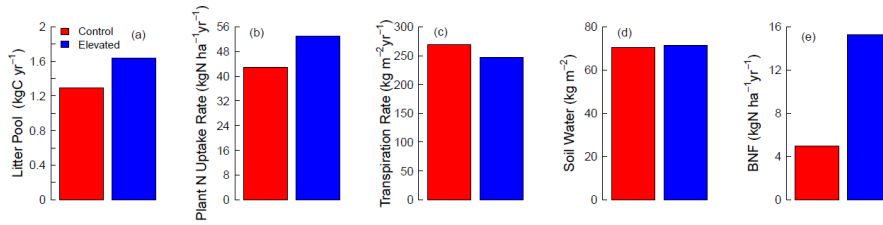


1
 2 Figure 7. Changes in simulated global average N₂O (1950-2005) emissions from modifying
 3 general N cycling processes (a) and model parameters one-at-a-time (b). Altered processes
 4 include disallowing N losses through dissolved organic matter (DON in (a)) and fire
 5 volatilization (Ash in (a)), and replacing simulated biological N fixation with preindustrial N
 6 fixation rate (BNF in (a)). Parameters include: v_{max} , the maximum active N uptake rate per unit
 7 root biomass; k_n , the optimum nitrification rate; k_d , the optimum denitrification rate; K_c and K_n ,
 8 the half saturation constants for labile C availability and nitrate respectively; and $frac$ is the
 9 fraction of net nitrification lost as N₂O. Parameters are either increased by multiplying 10
 10 (lightblue) or reduced by multiplying 0.1 (lightgreen) relative to the defaults .

11
 12
 13



1
 2 Figure 8. Soil N₂O emissions in response to step increases in atmospheric CO₂ and temperature.
 3 Panel (a) is the response to CO₂ fertilization alone, expressed as the difference between CO₂
 4 increased run and the control run (CO₂_FERT - CONTROL), the inset zooms into the y axis
 5 (flux difference) around zero; Panel (b) is the response to temperature increase alone (TEMP-
 6 CONTROL); Panel (c) is the combined response to both CO₂ enrichment and temperature rise
 7 (CO₂_FERT×TEMP-CONTROL); and Panel (d) is the interactive effect of CO₂ and
 8 temperature responses, which is the difference between the combined (results from Panel (c))
 9 and minus the individual responses (results from Panel (a) and (b)). Results are shown as annual
 10 values (thin dashed lines) and as running average with a moving window of 17 years (period of
 11 recycled climate forcing, thick solid lines). The black lines represent the global average
 12 response. Coloured lines indicate responses for biome as represented by each plant functional
 13 type (PFT) considered in LM3V-N: C4 grass (cyan), C3 grass (blue), tropical forest (yellow),
 14 temperate deciduous forest (green) and cold evergreen forest (pink). Dashed red line represents
 15 the zero line.



1
 2 Figure 9. CO₂ fertilization effects (no temperature change) on litter pool size (Panel (a)), plant
 3 nitrogen uptake rate (Panel (b)), canopy transpiration rate (Panel (c)), soil water content in the
 4 root zone (Panel (d)) and biological nitrogen fixation (BNF) rate (Panel (e)). Shown are the
 5 100-year average of global means (spatial) for control (284 ppm, red) and with elevated CO₂
 6 (568 ppm, blue).

7
 8 Table 1 Texture dependent parameter *k*, which partitions N₂O/N₂ gas fractions during
 9 denitrification, estimated from Del Grosso et al. (2000)

Soil Texture	Coarse	Medium	Fine	Coarse medium	Coarse/ fine	Medium/ fine	Coarse/ medium/ fine	Organic
<i>k</i>	2	10	22	6	12	16	11	2

10
 11