1 Factors influencing CO₂ and CH₄ emissions from coastal wetlands in the

2 Liaohe Delta, Northeast China

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10 Abstract

11 Many factors are known to influence greenhouse gas emissions from coastal wetlands, but it is still 12 unclear which factors are most important under field conditions when they are all acting 13 simultaneously. The objective of this study was to assess the effects of water table, salinity, soil temperature and vegetation on CH₄ emissions and ecosystem respiration (R_{eco}) from five coastal 14 wetlands in the Liaohe Delta, northeast China: Two Phragmites australis (common reed) wetlands, 15 two Suaeda salsa (sea blite) marshes and a rice (Oryza sativa) paddy. Throughout the growing 16 season, the Suaeda wetlands were net CH₄ sinks whereas the Phragmites wetlands and the rice 17 paddy were net CH₄ sources emitting 1.2-6.1 g CH₄ m⁻² y⁻¹. The *Phragmites* wetlands emitted the 18 19 most CH₄ per unit area and the most CH₄ relative to CO₂. The main controlling factors for the CH₄ 20 emissions were water table, temperature, soil organic carbon and salinity. The CH₄ emission was accelerated at high and constant (or managed) water tables and decreased at water tables below 21 the soil surface. High temperatures enhanced CH₄ emissions, and emission rates were consistently 22 low (<1 mg CH₄ m⁻² h⁻¹) at soil temperatures <18°C. At salinity levels >18 ppt, the CH₄ emission 23 rates were always low (<1 mg $CH_4 m^{-2} h^{-1}$) probably because methanogens were outcompeted by 24 25 sulphate reducing bacteria. Saline *Phragmites* wetlands can, however, emit significant amounts of CH₄ as CH₄ produced in deep soil layers are transported through the air-space tissue of the plants 26 27 to the atmosphere. The CH₄ emission from coastal wetlands can be reduced by creating fluctuating water tables, including water tables below the soil surface, as well as by occasional 28 29 flooding by high-salinity water. The effects of water management schemes on the biological communities in the wetlands must, however, be carefully studied prior to the management in 30 31 order to avoid undesirable effects on the wetland communities.

Keywords: Coastal wetlands, common reed, greenhouse gas emissions, *Phragmites australis,* rice
 paddy, seablite, *Suaeda salsa*

34 1 Introduction

Wetlands play an important role in the global carbon cycling as they function both as carbon sinks, 35 by storing carbon in soils and vegetation, and as carbon sources, by releasing CO₂ and CH₄ into the 36 atmosphere (Brix et al., 2001; Kayranli et al., 2010; Mitsch et al., 2013; Whiting and Chanton, 37 38 2001). Carbon dioxide is fixed by plants and autotrophic microorganisms through photosynthesis 39 and thereby transformed to organic compounds locked away from the atmosphere, a process called carbon sequestration (Kayranli et al., 2010). Wetlands can store organic carbon vectored 40 into the soil for a long time due to the generally slow decomposition rates in anaerobic wetland 41 42 soils (Mitsch et al., 2013). Decomposition of organic matter does however still take place, both through aerobic and anaerobic processes. Aerobic processes are more efficient and mainly form 43 CO_2 as an end-product, whereas anaerobic decomposition is much slower and, along with CO_2 , 44 also produces CH₄. Both gases are known as greenhouse gases, which cause global warming due to 45 46 their ability to absorb solar radiation (IPCC, 2007). The global warming potential (GWP) of CH₄ is 25 times greater than that of CO₂ on a 100 year time scale (IPCC, 2007) and high emissions of CH₄ can 47 therefore have disproportionately adverse effects on the climate. According to Whalen (2005), 48 wetlands contribute to about 24% of global CH₄ emissions from all sources, and are the largest 49 natural source of CH₄. Due to the increasing concern of greenhouse gas emissions and global 50 warming, it is important to gain more knowledge about the factors affecting CO₂ and CH₄ 51 52 emissions in different wetland systems, and understand how the balance might be affected by 53 management actions.

54 Previous work has shown that environmental factors like water table (Altor and Mitsch, 2008; Couwenberg et al., 2011; Hargreaves and Fowler, 1998), soil temperature (Bridgham and 55 Richardson, 1992; Inglett et al., 2012), salinity (Bartlett et al., 1987; Weston et al., 2011) and 56 vegetation biomass and type (Inglett et al., 2012; Kandel et al., 2013) may have strong controlling 57 58 effects on greenhouse gas emissions from wetlands. Decomposition of organic matter in wetland 59 soil is strongly dependent on temperature, and therefore, both CO₂ and CH₄ emissions from decomposition processes tend to increase with increasing soil temperature (Herbst et al., 2011; 60 Inglett et al., 2012). The optimum temperature for methanogenesis is around 20-30 °C, depending 61 on the community of methanogenic archaea (Svensson, 1984). However, methanogens are strictly 62 anaerobic, and for methanogenesis to take place the redox potential must be as low as -200 mV, 63

and other competing terminal electron acceptors must have been reduced (O₂, NO₃ and SO₄) 64 (Mitsch and Gosselink, 2007). The position of the water table is therefore an important controlling 65 factor on CH₄ emissions, as high water tables lead to oxygen depletion and thus low redox 66 potentials, which favors methanogenesis in the wetland soil (Grunfeld and Brix, 1999). 67 Couwenberg et al. (2011) found that CH₄ emissions in peatlands were practically zero when the 68 69 water table was below -20 cm, whereas the emissions varied between near zero and 500 kg CH₄ ha⁻¹ v⁻¹ when the water table was above -20 cm. The more oxidized conditions associated with low 70 water tables favor CH₄ oxidation by aerobic methanotrophic bacteria (Whalen, 2005), as well as 71 72 aerobic decomposition of organic matter, both processes emitting CO₂. It can therefore be difficult to predict gas emissions under field conditions, as both soil temperatures and water tables may be 73 74 subject to large seasonal variations.

The presence of vegetation affects CO₂ fluxes primarily by photosynthesizing and by increasing the 75 total ecosystem respiration (Han et al., 2013; Kandel et al., 2013). However, the vegetation may 76 77 also affect CH₄ emissions. Oxygen released from roots create aerobic microsites in the rhizosphere (Brix, 1994), which favors CH₄ oxidation by aerobic methanotrophs (Grunfeld and Brix, 1999). On 78 79 the other hand, a high primary production also increases the available carbon substrate for 80 methanogens via biomass decomposition and root exudation and can thus lead to higher CH₄ 81 emissions (Van der Nat and Middelburg, 2000; Whiting and Chanton, 1993). In addition, wetland 82 plants with internal air spaces (aerenchyma) provide an additional gas transport pathway, apart 83 from diffusion and ebullition from the sediment, that can enhance CH₄ emissions (Brix et al., 1996; Henneberg et al., 2012; Sorrell and Boon, 1994). Methane produced in the soil can be 84 85 transported through the aerenchyma of the plant tissue and bypass the water column, where it otherwise could have been oxidized by methanotrophs before reaching the atmosphere (Whalen, 86 87 2005). Thus, wetland vegetation can both decrease and enhance CH₄ emissions depending on the specific site conditions and type of vegetation. 88

Acute saltwater intrusion to freshwater wetlands has been reported to increase soil respiration
and lead to elevated CO₂ emissions (Chambers et al., 2011; Weston et al., 2011). However, coastal
wetlands with high salinity usually emit less CH₄ than less saline wetlands (Bartlett et al., 1987;
Poffenbarger et al., 2011). This has been explained by the high concentration of sulphate ions

(SO₄²⁻) in sea water, and the consequent high activity of sulphate reducing bacteria which
outcompete methanogens for organic substrate (Bartlett et al., 1987). Poffenbarger et al. (2011)
analyzed CH₄ and salinity data from a number of coastal wetlands and found a threshold salinity
level of 18 ppt, above which the wetlands emitted significantly less CH₄ than those with a lower
salinity.

98 Although many factors are known to influence CO₂ and CH₄ emissions from coastal wetlands, it is 99 still unclear which factors are most important under field conditions when they are all acting 100 simultaneously. Knowledge of the interactive effects of the factors driving greenhouse gas emissions is a prerequisite to being able to manage wetlands in a way that minimizes greenhouse 101 102 gas emissions, and to predict the effects of future climate change on greenhouse gas emissions 103 from wetlands. The objectives of this study were (i) to quantify the CH₄ emission and ecosystem respiration in the dominant wetland communities in a coastal wetland ecosystem, (ii) to assess the 104 seasonal variation in CH₄ emission and ecosystem respiration in different plant communities, and 105 106 (iii) to determine the main controlling factors for CH₄ emission and ecosystem respiration under field conditions. 107

108 2 Materials and Methods

109 **2.1 Study sites**

The Liaohe Delta is situated in the Liaoning Province in northeast China and comprises a wetland 110 area of around 1,280 km² (Li et al. 2012). About 786 km² of that is marsh vegetated by common 111 reed (Phragmites australis (Cav.) Trin. Ex Steud). The reed marshes in the Liaohe Delta represent 112 113 probably the largest reed fields in the world (Brix et al., 2014). The growing conditions for common reed in the delta marshes have been improved since the 1960s by a freshwater irrigation 114 management practice, that has washed away much of the soil salinity, and as a result, led to an 115 expansion of the reed fields and an increase in productivity (Ji et al., 2009). The reed biomass is 116 117 extensively used for paper production (Ma et al., 1993), and the hydrology is therefore regulated to maximize the biomass yield (Brix et al., 2014). Apart from reed marshes, the main wetland types 118 in the Liaohe Delta are tidal saltmarshes vegetated by Suaeda salsa (L.) Pall., III (seablite), and rice 119 120 paddies planted with Oryza sativa L. (Asian rice). The wetlands of the Liaohe Delta are important

breeding areas for many endangered bird species, and are designated as a Shuangtaizihekou
(Liaohekou) National Nature Reserve since 1986 and also listed as a Ramsar site since 2004 (Li et
al., 2012). However, the wetlands are adversely affected by the polluted water from the Liaohe

River (Zhang et al., 2010) and oil extraction activities, as the Liaohe Delta contains the third largest

125 oil field in China (Zhu et al., 2010).

- 126 Five study sites were selected to embrace the main wetland types of the delta. The five study sites
- included two *Suaeda* marshes, one created and one natural ('Suaeda1' at 40°52'11.09"N;
- 128 121°36′21.72″E and 'Suaeda2' at 40°57′38.62″N; 121°48′20.03″E, respectively), two *Phragmites*
- wetlands for paper production, ('Phrag1' at 40°52'22.34"N; 121°36'08.89"E and 'Phrag2' at
- 130 41°09'33.75"N; 121°47'42.71"E) and a rice paddy ('Rice' at 41°10'38.69"N; 121°41'17.28"E).

131 **2.2 Gas sampling and analysis**

Gas samples for estimation of CO₂ and CH₄ emission were collected monthly from April to 132 133 November 2012, using the static chamber method (Livingston and Hutchinson, 1995). Six 134 quadratic metal frames (0.6 x 0.6 m) were permanently installed in each study site, and wooden 135 boardwalks were built to facilitate access to the frames without disturbing the soil. Small holes were drilled in the sides of the frames just at the ground surface to facilitate water exchange 136 between the inside of the frames and the surrounding wetland between sampling events. These 137 holes were plugged during sampling. At each sampling event, a white plastic chamber (0.55 x 0.55 138 139 x 0.30 m) was placed over the metal frame and an airtight seal was created by water (about 1 cm 140 deep) within a trough inside the frame. The chambers were modified from past designs deployed 141 in shaded forested wetlands (Krauss and Whitbeck, 2012; Yu et al., 2008) by using aluminum foil to 142 cover all inside walls to block out light and prevent photosynthesis completely and to minimize temperature changes. Transparent and opaque chambers have been shown to provide similar CH₄ 143 144 flux estimates (Minke et al., 2014). If the vegetation was taller than the chamber, the plants were 145 bent to fit inside the chamber. At Phrag2, however, the plants grew so tall that they had to be cut 146 in June; we limited what we had to cut as much as possible. A small fan was used to mix the air 147 inside the chamber during sampling, and a PVC tube with the outer end placed in water was used to equilibrate the air pressure inside the chamber with the outside air pressure. Gas samples were 148 149 taken from the chamber through a rubber septum using a 15 mL plastic syringe, and immediately

transferred into pre-evacuated 10 mL glass vials with a thick rubber cap and an aluminum lid. The first sample was taken immediately after placing the chamber onto the frame, and four additional samples were taken with 20 minute intervals. The temperatures at a soil depth of 10 cm and the air temperature in the chamber were recorded at each sampling time. The gas samples were stored at room temperature for a maximum of one week before analysis. For comparison, the CO₂ flux in each chamber was also measured in situ during separate 1 minute incubations on the same day using a portable infrared gas analyzer (LI-COR 8100, Lincoln, NE, USA).

The concentrations of CO₂ and CH₄ in the gas samples were analyzed in 0.6 mL injections on a 157 TRACE Ultra GC-TCD (Thermo Fischer Scientific Inc., Waltham, MA, USA) at Qingdao Institute of 158 Marine Geology and an Agilent 7890A at the Ocean University of China, respectively. Signals from 159 160 the GCs were recorded in GC/MSD ChemStation Software (Agilent Technologies, Inc., Santa Clara, CA, USA) and the peak areas used to calculate the concentrations of CH₄ and CO₂. Gas emissions in 161 mg CH_4 m⁻² h⁻¹ and mg CO_2 m⁻² h⁻¹ (using the weight of the whole molecules of CH_4 and CO_2 , 162 respectively) were determined from the increase in concentration in the chambers over time using 163 linear regression analysis. Regression lines with a coefficient of determination $(R^2) < 0.6$ were not 164 included, except in cases where it was obvious that the low R² value was due to extremely low gas 165 fluxes (zero or near-zero fluxes). In a few cases, extremely deviant data were excluded. Because of 166 technical problems, no data on CO₂ emissions are available from Phrag1 in April and from Suaeda1 167 and Suaeda2 in May, and no data on CO₂ and CH₄ emissions in August from Phrag1. 168

Cumulative CO_2 and CH_4 emissions at each site were calculated as the integral of the mean gas emissions (in mg m⁻² d⁻¹) from the monthly sampling campaigns. As the gas sampling chambers were darkened, CO_2 emissions were assumed to be constant on a daily and nightly basis. And although some studies have found diurnal variations in CH_4 emissions (Käki et al., 2001; Neubauer et al., 2000; Tong et al., 2013), no consistent pattern has been found. Hence, we assumed that the CH_4 emissions were also constant on a daily basis.

175 2.3 Environmental parameters

The water table was measured in a piezometer at each study site, and the soil surface level
differences among the six plots at each site were used to calculate individual water tables for each

plot. Water samples for salinity and pH analyses were taken from the piezometer, and measured
using a Jenco 6010 microcomputer based pH/mV/temperature portable meter (Jenco Electronics,
Ltd., Shanghai, China).

181 The aboveground biomass inside the plots was estimated using a non-destructive method. In the Phragmites wetlands, the heights of all shoots inside the frames were measured, and 25 shoots 182 encompassing the range of heights in the frames were harvested outside the frames. In the 183 Suaeda wetlands, the plant density inside the frames was counted and 20x20 cm plots outside the 184 frame with a similar plant density were harvested. The plants were dried at 60°C and weighed, 185 186 and the biomass inside the plots was calculated from a regression analysis between plant height and dry mass (*Phragmites*) and between plant density and dry mass (*Suaeda*). In the rice paddy, 187 188 five rice plants outside the frames were harvested, dried and weighed, and the biomass within the 189 frames was estimated based on the number of plants.

Soil core samples were taken to 5 cm depth from the topsoil near each frame using a 5 cm
diameter steel cylinder. The samples were dried to constant weight at 60°C for determination of
bulk density and water content. Soil redox potentials (Eh) were measured using platinum
electrodes installed at a depth of 10 cm at least 24 hours before measuring. Redox electrodes
were referenced against a calomel electrode.

195 Two soil core samples were collected to 4 cm depth at each site the following year, mixed and 196 analyzed for selected mineral elements and available nutrients. Total N and TC were analyzed on 197 oven-dried subsamples ground to pass a 2 mm sieve, on a Perkin Elmer 2400 Series II CHNS/O 198 elemental analyzer (Perkin Elmer, Inc., Waltham, MA, USA). For determination of Org-C, another set of subsamples was treated with 4M HCl (Craft, 2007) to remove inorganic carbon before 199 200 analysis on the same instrument. Available nutrients were extracted by the Mehlich-III method (Mehlich, 1984), using an extraction solution prepared from 22.98 mL concentrated CH₃COOH, 201 40.0 g NH₄NO₃, 1.12 g NH₄F, 1.68 mL concentrated HNO₃, 0.58 g EDTA and 1,600 mL deionized 202 water, diluted to 2 L. Air-dried soil subsamples were ground to pass a 1 mm mesh. 2.5 g of the 203 204 ground soil were shaken with 25 mL extraction solution on a reciprocating oscillator for 5 minutes 205 and then centrifuged for 20 minutes. The supernatant was diluted ten times and analyzed for Ca, 206 Cu, Fe, K, Mg, Mn, P and Zn by ICP-OES (Optima 2000 DV, Perkin Elmer, USA).

207 2.4 Statistical analysis

The in situ measurements of CO₂ emissions with the IRGA were used in the statistical analyses. 208 Methane emissions and ecosystem respiration (Reco) were analyzed by Site and Time with Plot as a 209 random factor nested within Site, in a repeated-measures setup using the General Linear Model 210 (GLM) procedure of Statgraphics Centurion XVI (Statpoint Technologies, Inc., Warrenton, Virginia, 211 212 USA). The Bonferroni post-hoc test was used to identify significant differences between different sites at the 5 % significance level. Data of CH₄ emissions and R_{eco} were log-transformed to meet 213 the assumption of equal variances, which was checked using Levene's test (p>0.05). Since the 214 dataset included a few negative gas flux values, a constant was added to the fluxes (CH₄ flux+0.6 215 and Reco+25, respectively) before applying the log-transformations. Data from April, May and 216 August were excluded from the analyses due to missing data at some sites. 217

218 Linear mixed effects models (multiple regressions) using R version 3.0.1 (Team, 2013) were used to assess the relations between the measured environmental factors and CO₂ and CH₄ emissions, 219 220 respectively. The response variables were CO₂ and CH₄ emissions. The fixed effects were plant species (categorical variable), soil temperature (SoilT), water table (WT), aboveground biomass 221 (Biomass) and Salinity (continuous variables). The random effects were Site and Plot. An 222 interaction effect between plant species and aboveground biomass was also included. The effect 223 224 of each variable or interaction was evaluated by removing the variable/interaction from the original model and using a likelihood ratio chi-square test to test for significant differences at the 225 226 5% significance level between the original model and the model excluding the variable/interaction. Data of CO₂ and CH₄ emissions were log-transformed as described before to meet the assumptions 227 228 of normality and equal variances. The original mixed effects model for CO₂ and CH₄ emissions, respectively, was in the form: 229

230
$$\text{Log}_{10}(\text{gas flux})_i = \beta_{1i} \cdot \text{Biomass}_i + \beta_2 \cdot \text{SoilT} + \beta_3 \cdot \text{Salinity} + \beta_4 \cdot \text{WT} + b_1(\text{Site}) + b_2(\text{Plot}) + \epsilon_i$$
 (1)

where β_1 is a coefficient specific for plant species *i*, β_2 , β_3 and β_4 are coefficients for fixed effects common for all plant species, b_1 and b_2 are coefficients for the random effects and ε_i is the residual error for plant species *i*.

234 3 Results

235 **3.1 Environmental parameters**

236 The water tables varied greatly over the season, particularly at Phrag2 where the water table ranged from -35 to +27 cm, and at Suaeda1 where it ranged from -43 to +15 cm (Fig. 1c). At the 237 238 two *Phragmites* wetlands, the water tables were managed to maximize the yield of *Phragmites* 239 biomass. Hence, the water tables at these sites were above the soil surface during most of the growing season. The water tables at the two Sugeda wetlands fluctuated greatly due to tidal 240 241 variations, but the water tables were at the time of sampling usually below the soil surface. At the rice paddy, the water table was fairly stable around +10 cm from June to September due to 242 regulation according to agricultural practice. 243

Soil temperatures at all sites increased from 18-22°C in May to 23-28°C in August, and then
declined to 0-7°C in November (Fig. 1d). We do not have temperature data from the months prior
to our sampling, but usually the soils in the delta are frozen until April, whereafter the
temperature increases over a few weeks.

The amount of aboveground biomass was basically zero during the first sampling campaign in late April. Thereafter, both *Suaeda* and *Phragmites* grew rapidly reaching aboveground biomasses in June of ~800 g dry mass m⁻² for *Suaeda* and ~400 g dry mass m⁻² for *Phragmites* before the cutting in June (Fig. 1e). In the rice paddy, the rice plants were planted in late June. Hence the development of biomass in the rice paddies occurred much later than in the natural *Suaeda* and *Phragmites* wetlands.

The salinity at Suaeda1 was 32-39 ppt during most of the sampling period (Fig. 1f). At Sueda2 the salinity was lower: 10-15 ppt from May to July and then decreasing to 5-6 ppt in August to December. In the *Phragmites* wetlands, the salinities varied between 2 and 19 ppt depending on the water management scheme. The highest salinities were found at Phrag1. At the rice paddy the salinity was constantly low at around 2 ppt.

Soil bulk density varied between 0.93 g cm⁻³ at Phrag2 to 1.50 g cm⁻³ at Suaeda1, and soil water

content between 27% at Suaeda1 and 48% at Phrag2 (Table 1). The mean redox potential was

highest at Suaeda1 (+101 mV) and lowest at Phrag1 (-127 mV). The mean soil water pH was in the

262 interval 7.12 – 7.70 at all sites

Al topsoils consisted largely of fine silt and clay and had a low content of organic matter (Org-C < 2% of the dry matter). However, the contents of organic carbon (12%) and nitrogen (1%) were markedly higher at Phrag2 than at the other sites (Table 1). At Phrag1, the contents of organic carbon (1.8%) and nitrogen (0.17%) were 2-3 times higher than at the Suaeda sites and the rice paddy. Differences in other analyzed mineral elements were less pronounced and probably reflected the predominantly mineral composition of the soils, except for the concentration of P which was higher at Phrag2 and the rice paddy than at the other sites.

270 **3.2 CH₄ emissions**

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271 There were large variations in CH₄ emission rates both among sites and over the season (Fig. 1a) and these differences were statistically significant (Table 2). The highest CH₄ emission rates were 272 found at Phrag2 and at the rice paddy. Peak emissions were 2.5 mg m⁻² h⁻¹ at both sites although 273 the peak values were measured in July at Phrag2 and in August at the rice paddy (Fig. 1a). The 274 highest CH_4 emission rates at Phrag1 (around 0.7 mg m⁻² h⁻¹) were only a fourth of those at 275 Phrag2. At the two *Phragmites* wetlands, the CH₄ emission rates were close to zero in April-May, 276 277 increased rapidly from June to July, and declined again after August. At the rice paddy, the CH₄ emission rates were near zero in June, low in July (0.25 mg $m^{-2} h^{-1}$), increased very sharply from 278 July to August and thereafter declined. At the Suaeda wetlands, the CH4 emission rates were close 279 to zero throughout the sampling period. Means and ranges of CH₄ emission rates over the whole 280 sampling period, and significant differences (p<0.05) among sites, are shown in Table 4. 281

The CH₄ emission rates at sites with significant emissions (Phrag1, Phrag2 and Rice) were positively related to both soil temperature and water table (Table 3; Fig. 3). The CH₄ emission rates were less than 1 mg m⁻² h⁻¹ at temperatures below 18°C and at water tables below the soil surface. The highest CH₄ emission rates were measured at Phrag2 when both the temperature and the water table were high (Fig. 3). The CH₄ emissions decreased significantly (Table 3) with increasing salinity, as CH₄ emission rates were less than 1 mg m⁻² h⁻¹ at salinity levels above 18 ppt (Fig. 4). At

Cumulative CH_4 emissions over the entire growing season in 2012 were highest at Phrag2 with 6.1 g CH_4 m⁻² y⁻¹, corresponding to 154 g CO_2 -equivalents m⁻² y⁻¹ (Fig. 2, Table 4). These emissions were about 1.5 times higher than the cumulative CH_4 emissions from the rice paddy, and about

the highest salinity levels at Suaeda1 (32-38 ppt), CH₄ emission rates were practically zero.

five times higher than the CH₄ emissions from Phrag1. CH₄ emissions from the *Suaeda* wetlands
were negligible.

294 **3.3 Ecosystem respiration (R**eco)

The measured flux of CO₂ in the darkened chamber is the sum of the flux of CO₂ from the soil and 295 296 the respiration of the plant tissue inside the chambers. We here refer to this as the ecosystem 297 respiration (R_{eco}). The ecosystem respiration rates varied significantly both among sites and over time (Fig. 1b, Table 2). The highest ecosystem respiration rates at the rice paddy and at Phrag2 298 (2,400 and 2,300 mg CO₂ m⁻² h⁻¹, respectively) were twice as high as the highest R_{eco} at Phrag1 and 299 three times higher than the Reco at the two Suaeda wetlands. At Phrag2, Reco was highest in June 300 and July, whereas at the rice paddy, the Reco was low at this time of the year and highest in August 301 (Fig. 1b). It should, however, be mentioned that the *Phragmites* stems at Phrag2 were cut in June. 302 Hence, the biomass within the chambers from July and onwards was lower than the biomass in 303 the surrounding reed vegetation. Overall, the ecosystem respiration rates were significantly 304 related to plant biomass, soil temperature and salinity (Table 3) whereas water table had no 305 significant effect on R_{eco} (p>0.05). 306

The cumulative CO₂ emissions, without accounting for photosynthetic CO₂ uptake, varied between 1.7 kg m⁻² y⁻¹ in the *Suaeda* wetlands to 3.0-4.4 kg m⁻² y⁻¹ in the *Phragmites* (Table 4). The cumulative CO₂ emission in the rice paddy was in-between this range (3.3 kg m⁻² y⁻¹).

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311 4 Discussion

312 **4.1 CH₄ emissions**

Over one growing season in 2012, the two *Phragmites* wetlands emitted on average 0.15 and 1.01 313 mg CH₄ m⁻² h⁻¹ (Phrag1 and Phrag2, respectively) and the rice paddy 0.75 mg m⁻² h⁻¹, whereas the 314 emissions from the two Suaeda wetlands were negligible. The large differences in CH₄ emission 315 rates among the five sites can be explained by the differences in soil organic matter, salinity and 316 317 water tables, and, to some extent, vegetation type. For methanogenesis to take place there must be a sufficient amount of labile organic substrate available (Mah et al., 1977), such as dead plant 318 material from the previous growing season and root exudates from the standing vegetation (Mann 319 and Wetzel, 1996; Zhai et al., 2013). Previous studies have reported increasing CH₄ emission rates 320

321 with increasing content of soil organic matter in different types of wetlands (Le Mer and Roger, 2001; Picek et al., 2007; Serrano-Silva et al., 2014; Sha et al., 2011; Tanner et al., 1997). At Phrag2, 322 where CH₄ emission rates were significantly higher than at the other sites, there was a many-fold 323 higher content of organic carbon and nitrogen in the soil compared to the soils at the other sites, 324 325 and the reeds at Phrag2 had a very dense root system in the upper soil layers. Thus, the reason for 326 the high CH₄ emission rates at Phrag2 was most likely the higher content of organic substrate for methanogenesis, originating from dead plant residues and from root exudates. At the rice paddy, 327 where the second highest CH₄ emissions were measured, the organic content of the soil was low, 328 but the soil C:N ratio was lower (8.4) than the ratios at the other sites probably resulting from 329 different plant inputs into the soil. A lower C:N ratio of the organic matter in the soil may increase 330 organic matter lability by decreasing nitrogen limitation for decomposers (Hodgkins et al., 2014). 331 However, the fact that the rice paddy was constantly flooded throughout the growing season 332 333 probably also stimulated methanogenesis and CH₄ emission.

334 Both *P. australis* and rice have well developed aerenchyma in roots, rhizomes and stems, which provides them with a high ability to transport gases between the soil and the atmosphere through 335 336 the plant tissue (Brix et al., 1996; Singh and Singh, 1995). When CH₄ is transported from the soil through the air-space tissues of the plants, it bypasses the aerobic zone in the upper part of the 337 338 soil and the water column, where CH₄ otherwise could have been oxidized by methanotrophic 339 bacteria (Whalen, 2005). Plant-mediated transport has been reported to be the main pathway of 340 CH₄ transport from the soil to the atmosphere and constituting as much as 60-90 % of the CH₄ emissions (Butterbach-Bahl et al., 1997; Huang et al., 2005). In the present study, transport of CH₄ 341 342 through the air-space tissue of the plants may explain the relatively high CH₄ emission rates from the *Phragmites* wetlands and the rice paddy, while the lack of well-developed aerenchyma in S. 343 salsa is consistent with the negligible emission rates from the Suaeda wetlands. The aboveground 344 biomass per se probably had no effect on the plant-mediated CH₄ emissions, as CH₄ has been 345 346 shown to be mainly emitted through micropores in the basal parts of rice plants (Nouchi et al., 1990) and through the basal internodes of *P. australis* (Brix, 1989). Also, Henneberg et al. (2012) 347 showed in a manipulation experiment with Juncus effusus that aboveground biomass was 348 unimportant for the CH₄ transport through the plants, whereas the removal of fine roots and root 349 tips of coarse roots led to significant reductions in plant-mediated CH₄ transport. Thus, it is likely 350

that the extensive root system of the reeds at Phrag2 contributed to the high CH_4 emission rates at this site.

At salinity levels above 18 ppt the CH₄ emission rates were always lower than 1 mg m⁻² h^{-1} across 353 354 all sites (Fig. 4). This is consistent with Poffenbarger et al. (2011) who found a salinity threshold of 18 ppt, above which CH₄ emission rates were significantly lower than at lower salinity levels. The 355 effect of salinity has been explained by the high concentrations of SO₄²⁻ in seawater, which inhibit 356 CH₄ production due to competition from sulphate reducing bacteria (Bartlett et al., 1987; D'Angelo 357 358 and Reddy, 1999). Thus, the lack of CH₄ emissions at the Suaeda sites is most likely an effect of the high salinity, particularly at the Suaeda1 site where salinities were up to 35 ppt. The salinity was, 359 360 however, significantly lower at the Suaeda2 site with salinities of 5-15 ppt, and yet there were no CH₄ emissions as SO₄²⁻ concentrations were still high enough to inhibit methanogenesis. At Phrag2, 361 on the other hand, CH₄ emission rates were high although the water salinity was occasionally as 362 high as 15 ppt. These seemingly contradictory results can be explained by the fact that a high 363 salinity in the water mainly affects the upper soil layers, but not necessarily the deeper layers. 364 Therefore, methanogens may be outcompeted by sulphate reducing bacteria in the upper layers 365 of the soil, but CH_4 can still be produced in the deeper soil layers where all SO_4^{2-} have been 366 reduced. The roots of *P. australis* grow to a soil depth of at least 40-60 cm, and CH₄ can therefore 367 368 be transported from the deeper anoxic zone through the air-space tissue of the plants to the atmosphere. Thus, the relatively high salinity at Phrag2 probably inhibited methanogenesis in the 369 370 upper soil layers, but the CH₄ produced in the deeper soil layers were still transported to the 371 atmosphere through the plants. At the Suaeda wetlands, the generally low and fluctuating water 372 tables indicate that the anaerobic zone where methanogenesis can take place was at a deeper soil depth than at the *Phragmites* wetlands. The roots of *S. salsa* lack aerenchyma and are generally 373 restricted to the upper 20 cm of the soil, and are therefore ineffective conduits for CH₄ from the 374 375 deeper soil layers to the atmosphere. Thus, although salinity levels at Suaeda2 were not always high, any CH₄ that may have been produced in the soil did not reach the atmosphere because of 376 CH₄ oxidation in the upper soil layer. At the rice paddy, the low salinity of around 2 ppt seemingly 377 had no inhibitory effect on the CH₄ production and emission. 378

379 The water table is an important parameter affecting the CH₄ emission rate. The highest CH₄ emissions occurred at the three sites where the water exchange and water table were managed to 380 maximize the reed biomass (Phrag1, Phrag2) and crop yield (Rice) whereas very low CH₄ emission 381 rates were found at the two *Suaeda* wetlands with a natural tidal hydrology. At the rice paddy, the 382 soil was continuously flooded from June until September, and the two *Phragmites* wetlands were 383 384 more or less flooded from June until October, resulting in low redox potentials and relatively high CH₄ emission rates. The soils at the tidally influenced *Suaeda* wetlands were periodically drained 385 and hence partly oxidized inhibiting CH₄ production. When water tables at the *Phragmites* 386 wetlands and the rice paddy were below the soil surface, the CH₄ emission rates were always <1 387 mg $CH_4 m^{-2} h^{-1}$ probably because CH_4 produced in deeper soil layers was oxidized in the upper oxic 388 soil layers, reducing the amount of CH₄ reaching the atmosphere. When the water tables 389 390 approached the soil surface, the CH₄ emission rates increased. This is in agreement with the 391 findings of Zhu et al. (2014), who reported that the seasonal CH₄ emissions from an herbaceous 392 peatland were highly linked to water table fluctuations, and that the water table was the main environmental driver for CH₄ emissions over a single growing season, whereas soil temperature 393 was important on a longer time scale. The important effect of water table on CH₄ emission rates is 394 in agreement with observations in other studies (e.g. Bridgham et al., 2006; Couwenberg et al., 395 2011; Le Mer and Roger, 2001; Serrano-Silva et al., 2014). However, in the present study both soil 396 397 water table and temperature were important drivers.

398 The large seasonal variations in CH₄ emission rates at Phrag1, Phrag2 and Rice were primarily 399 related to the variations in soil temperatures. The highest CH₄ emission rates occurred during the 400 summer months July-September, when temperatures were relatively high. We found an exponential relationship between soil temperature and CH₄ emission rates (Fig. 3) similar to those 401 reported elsewhere (Herbst et al., 2011; Inglett et al., 2012) in accordance with the temperature 402 403 dependency of the methanogenic bacteria. Furthermore, the amount of labile organic carbon 404 substrates from root exudates can be stimulated by high temperatures as Zhai et al. (2013) found significantly higher root exudation rates from *P. australis* roots at 20°C than at 10°C. Also the 405 406 plant-mediated CH₄ transport may be accelerated at higher temperatures as Hosono & Nouchi (1997) reported that the CH₄ transport through rice plants was twice as high at a rhizosphere 407 temperature of 30°C as compared to the transport at 15°C. Thus, the high CH₄ emission rates at 408

409 both Phrag2 and Rice during the warmest months of the year were probably due to the high temperature and its stimulating effect on the activity of the methanogenic bacteria, the root 410 exudation rates and the effectivity of the plant-mediated transport. At soil temperatures below 411 18°C, which occurred before June and after September, CH₄ emission rates were consistently low 412 (<1 mg CH₄ m⁻² h⁻¹). In the spring, the low rates might be associated with a time-lag in the growth 413 of methanogens as the temperature was increasing over a relatively short period. In the autumn 414 415 the low rates might be influenced by low availability of organic carbon, as most carbon might have been 'burned off' during the hot summer months. 416

417 **4.2 Ecosystem respiration (R**eco)

Ecosystem respiration rates were highest in June-July at the Phragmites wetlands, June-August at 418 419 the Suaeda wetlands and August at the rice paddy. The differences among the sites can be explained by the differences in soil organic matter and biomass, whereas the variations over time 420 can be explained mainly by soil temperature and to some extent by differences in biomass. The 421 seasonal pattern of ecosystem respiration was closely related to that of soil temperature at all 422 sites, which suggests that temperature was the main controlling factor for ecosystem respiration. 423 424 This is in agreement with the findings of other studies (Bridgham and Richardson, 1992; Han et al., 425 2013; Happell and Chanton, 1993; Kandel et al., 2013; Krauss et al., 2012; Pulliam, 1993). However, biomass respiration also contributed to the ecosystem respiration rates, particularly late 426 427 in the season when the aboveground biomass was highest. At Phrag1, Suaeda1 and Suaeda2, the 428 seasonal pattern of ecosystem respiration rates correlated to that of the aboveground biomass, 429 indicating that plant respiration may have constituted a large part of the total ecosystem respiration at these sites. This is in agreement with Kandel et al. (2013), who found that plant 430 431 respiration contributed with about 50% of the total ecosystem respiration in a cultivated peatland 432 during the summer months, and Xu et al. (2014), who found ten times higher CO₂ emissions from marshes with plant communities than from those without. Also, the difference in ecosystem 433 respiration rates between the two Suaeda wetlands corresponded to the differences in Suaeda 434 biomass. However, at Phrag2 nearly all CO₂ emissions came from the soil and the belowground 435 biomass, since only short stems were left behind after cutting the reeds in June. At the rice paddy, 436 the ecosystem respiration peaked in August when the aboveground biomass was only about 100 g 437 m⁻². The aboveground rice biomass continued to increase after August, but the ecosystem 438

respiration decreased drastically, indicating that soil respiration constituted the main part ofecosystem respiration at the rice paddy.

441 **4.3 Cumulative emissions**

442 The two Suaeda wetlands were net CH₄ sinks whereas the two Phragmites wetlands and the rice paddy were net CH₄ sources during April to November 2012. Although the peak CH₄ emission rates 443 at the rice paddy were similar to those at Phrag2, the cumulative CH₄ emission rates from Phrag2 444 were 1.5 times higher than those from Rice. The cumulative CO₂ emitted from ecosystem 445 446 respiration followed a similar pattern, with Phrag2 emitting 1.3 times more CO_2 than the rice paddy. Thus, on a yearly basis Phrag2 emitted the highest amounts of both CH₄ and CO₂ per unit 447 area, and also the most CH₄ relative to CO₂. Since CO₂ emissions from vegetated ecosystems are 448 counteracted by photosynthetic CO₂ uptake and possibly carbon sequestration, the CO₂ emissions 449 450 measured as ecosystem respiration does not contribute to the greenhouse effect. However, the CH₄ emissions from wetland ecosystems contribute to the radiative forcing, and therefore CH₄ 451 emission rates should be minimized. It is, however, the balance between carbon sequestrations on 452 the one hand and CH₄ emission on the other hand that determines if a particular wetland can be 453 considered to be a net source or a net sink for radiative greenhouse gases (Mitsch et al., 2013). 454 Based on the present study, it is unfortunately not possible to estimate the carbon sequestration 455 456 of the different wetland communities.

457 4.4 CH₄ emission rates and R_{eco} compared to other studies

The CH₄ emission rates and seasonal pattern at Phrag2 were similar to those measured by Huang 458 et al. (2005) from a reed wetland in the Liaohe delta, where CH₄ emission rates varied from -0.97 459 mg CH₄ m⁻² h⁻¹ in early May to 2.73 mg CH₄ m⁻² h⁻¹ in early September. The average CH₄ emission 460 461 rate at Phrag2 was within the range of CH₄ emission rates from reed wetlands in other parts of China, varying from 0.75 mg m⁻² h⁻¹ (Xu et al., 2014) to 5.13 mg m⁻² h⁻¹ (Tong et al., 2010). The 462 Suaeda wetlands had CH₄ emission rates very similar to those from a Suaeda salsa marsh in the 463 Yellow River delta, China, with rates ranging from -0.74 to 0.42 mg m⁻² h⁻¹ (Sun et al., 2013). The 464 CH₄ emission rates from the rice paddy in the present study were lower than those reported from 465 continuously and intermittently flooded rice paddies in Nanjing, China, which emitted 1-3 mg m⁻² 466 h⁻¹ (Zou et al., 2005). This might be due to temperature differences or differences in soil 467 characteristics at the two sites. 468

The yearly cumulative CH₄ emissions from Phrag2 were similar to those reported by Xu et al. 469 (2014) from a coastal saline grass flat dominated by *P. australis* in southeast China (6.28 g m⁻²). 470 However, markedly higher cumulative CH₄ emissions have been measured from other reed 471 wetlands, such as 39.5 g m⁻² from a tidal reed marsh in southeast China (Tong et al., 2010) and 472 65.9 g m^{-2} from a restored reed fen in northeastern Germany (Koch et al., 2014). The yearly 473 cumulative CH₄ emissions from the rice paddy in our study were about six times higher than the 474 0.54-0.58 g m⁻² measured from rice paddies in eastern China (Zhang et al., 2014) but much lower 475 than the 57 g m⁻² measured over only two months from a rice paddy in the Philippines (Gaihre et 476 al., 2014). The Suaeda wetlands in our study had no net CH₄ emissions over the sampling period, in 477 contrast to a *Suaeda glauca* marsh in southeast China which emitted 0.399 g CH_4 m⁻² y⁻¹ (Xu et al., 478 479 2014).

The average ecosystem respiration rates in this study were in a comparable range to those
recorded from coastal saline wetlands in southeast China by Xu et al. (2014). The average CO₂
emission rates at Phrag1 were somewhat lower than the 569.7 mg m⁻² h⁻¹ from the *Phragmites*wetland in their study, whereas the emissions from Phrag2 were higher. Compared to the *Suaeda glauca* marsh in Xu et al. (2014), which emitted on average 248.6 mg CO₂ m⁻² h⁻¹, Suaeda1 and 2
both had higher average CO₂ emissions.

486 **4.5 Conclusions**

We aimed at determining which factors are most important under field conditions for controlling 487 CH₄ and CO₂ emissions from coastal wetlands in order to be able to predict the effects of future 488 climate change on greenhouse gas emissions from wetlands and potentially to be able to manage 489 490 coastal wetlands in a way that minimizes greenhouse gas emissions. Hence, we quantified the CH₄ 491 emissions and ecosystem respiration from April to November 2012 in five coastal wetlands in the Liaohe Delta, northeast China, and determined the main controlling factors for the seasonal 492 493 variations and the differences among the sites. Over the study period, the two Suaeda wetlands were net CH₄ sinks whereas the *Phragmites* wetlands and the rice paddy were net CH₄ sources. 494 The *Phragmites* wetlands had the highest climatic impact as they emitted the most cumulative CH₄ 495 per unit area and the most CH₄ relative to CO₂ compared to the other wetland types. The main 496 497 controlling factors for the CH₄ emissions were water table, soil organic carbon, temperature and

498 salinity. Methane emissions are accelerated at high and constant (or managed) water tables and decrease at water tables below the soil surface, or fluctuating water tables. Methane emissions 499 are also accelerated at high temperatures and depressed at high salinity levels. Saline wetlands 500 can, however, emit significant amounts of CH₄ as aerenchymatous wetland plants with deep root 501 502 systems can transport CH₄ produced in the deeper soil layers to the atmosphere. The ecosystem 503 respiration of the wetland communities depends largely on temperature and the plant 504 aboveground biomass, but soil organic matter content and belowground biomass are also important. It is, however, necessary to quantify not only the ecosystem respiration, but also the 505 balance between the net CO₂ exchange and the CH₄ emission to determine if a particular wetland 506 can be considered to be a net source or a net sink for radiative greenhouse gases. Our study 507 indicates that the CH₄ emissions from coastal wetlands can be reduced by managing the water in 508 509 the wetland in a way that creates fluctuating water tables, including water tables below the soil 510 surface, as well as by occasional flooding by high-salinity water. However, the effects of potential 511 water management schemes on the biological communities in the wetlands must be carefully studied prior to the implementation of the management in order to avoid negative and 512 undesirable effects on the wetland communities. 513

514 Author contribution

515 S.Y., K.W.K and H.B. designed the study, L.O. and S.Y. performed the field and laboratory

516 measurements, and L.O. prepared the manuscript with contributions from all co-authors.

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- 526 Copenhagen. Any use of trade, product, or firm names is for descriptive purposes only and does
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Table 1. Physical/chemical topsoil characteristics (0-5 cm depth for bulk density, water content and redox

- 693 potential; else 0-4 cm depth) at the five wetland sites (two Suaeda salsa wetlands, two Phragmites australis
 - 694 wetlands and one rice paddy) in the Liaohe Delta, northeast China. Data was collected in 2013 by Siyuan Ye
 - 695 et al. (personal communication).

	Suaeda1	Suaeda2	Phrag1	Phrag2	Rice
Bulk density (g cm ⁻³)	1.50	1.20	1.07	0.93	1.36
Water content (% of FW)	27	37	41	48	30
Redox potential (mV)	101	24	-127	-91	-82
TN (% of DW)	0.08	0.07	0.17	1.02	0.10
TC (% of DW)	0.95	0.83	1.81	12.59	0.88
Org-C (% of DW)	0.53	0.69	1.67	11.81	0.69
C:N ratio	12.4	12.0	9.8	12.3	8.4
Ca (μg g ⁻¹)	6735	4215	3817	2103	2239
Cu (μg g ⁻¹)	9.96	6.78	9.11	7.18	3.44
Fe (μg g ⁻¹)	282	434	396	343	343
Κ (μg g ⁻¹)	849	576	598	892	109
Mg (μg g ⁻¹)	2043	1120	1395	1687	216
Mn (μg g ⁻¹)	291	368	308	104	78
Ρ (μg g ⁻¹)	19.7	27.8	9.9	46.7	37.0
Zn (μg g ⁻¹)	9.6	11.1	17.8	30.8	8.2

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- 699 **Table 2.** Results from repeated-measures ANOVAs with the response variables CH₄-flux and R_{eco},
- respectively, the fixed factors Site and Time and their interaction, and the random factor Plot. Gas fluxes
- 701 were measured during April-November 2012 from six plots at two *Suaeda salsa* wetlands, two *Phragmites*
- 702 *australis* wetlands and one rice paddy in the Liaohe Delta, northeast China. All measurements from April,
- 703 May and August were excluded from the analysis due to missing data from some sites.

Response variable	Factor	df	F-ratio	р
CH ₄ -flux	Site	4	19.9	<0.001
	Time	4	7.5	<0.001
	Site × Time	16	5.9	<0.001
	Plot (random factor)	25	2.0	0.007
R _{eco}	Site	4	23.7	<0.001
	Time	4	379.4	<0.001
	Site × Time	16	55.7	<0.001
	Plot (random factor)	25	1.9	0.010

704 df: degrees of freedom

Table 3. Results from linear mixed effects models, with CH_4 emission rate and ecosystem respiration rate706 (R_{eco}) as response variables, and the fixed effects Plant species, Biomass, Soil temperature, Water table and707Salinity. Shown are the coefficients of the fixed effects to be included in equation 1, standard errors of the708means and p-values.

Response variable	Predictor	Coefficient	SE	р
CH₄ emission rate	Water table	0.0054	0.0014	<0.001
	Soil temperature	0.0017	0.0023	<0.001
	Salinity	-0.0023	0.0030	<0.001
CH ₄ emission rate ^a	Water table	0.0071	0.0019	<0.001
	Soil temperature	0.0074	0.0034	<0.001
R _{eco}	Suaeda*Biomass	-1.93 10 ⁻⁵	3.1 10 ⁻⁴	0.003
	Phrag*Biomass	7.1 10 ⁻⁴	2.5 10 ⁻⁴	0.003
	Rice*Biomass	9.2 10 ⁻⁴	3.0 10 ⁻⁴	0.003
	Soil temperature	0.057	0.0042	<0.001
	Salinity	0.0095	0.0044	0.049

^a Only sites with CH_4 emissions >0 included (Phrag1, Phrag2 and Rice).

- 710 **Table 4.** Mean CH₄ emission and ecosystem respiration rates (R_{eco}) with ranges in parentheses, and
- cumulative CO₂ equivalents from CH₄ and CO₂ emissions, respectively, from two Phragmites australis
- vetlands and one rice paddy during April-November 2012 in the Liaohe Delta, northeast China. CH₄ fluxes
- 713 are converted to CO₂-equivalents using a factor of 25. Superscript letters represent significant differences
- 714 (p<0.05) among sites.

			Cumulative CO ₂ -equivalents		
Site	CH₄ emission rates (mg m ⁻² h ⁻¹)	\mathbf{R}_{eco} (mg CO ₂ m ⁻² h ⁻¹)	CH ₄ (g CO ₂ -eqv m ⁻² y ⁻¹)	CO₂ (g CO ₂ -eqv m ⁻² y ⁻¹)	
Suaeda1	0.01 (-0.31 - 0.44) ^a	278 (-3.6 - 814) ^{ab}	-0.4	1671	
Suaeda2	-0.01 (-0.50 - 0.42) ^a	423 (4.6 - 954) ^b	-1.9	1730	
Phrag1*	0.15 (-0.31 - 1.48) ^{ab}	484 (-14.8 - 1300) ^c	31.1	2963	
Phrag2	1.01 (-0.28 - 6.38) ^c	811 (27.4 - 3357) ^c	153.7	4443	
Rice	0.75 (-0.27 - 4.63) ^b	532 (-0.2 - 3181) ^a	91.6	3337	

715 * No data from August

716 Legends to figures:

Figure 1. Seasonal variation in (a) CH₄ emission rates, (b) ecosystem respiration, (c) water table, (d) soil temperature, (e) aboveground dry biomass and (f) salinity in two *Suaeda salsa* wetlands, two *Phragmites australis* wetlands and one rice paddy during 2012 in the Liaohe Delta, northeast China. Plotted values are the averages for six plots at each site. Data from Phrag2 is missing in August because it was not possible to sample due to extreme flooding. Aboveground biomass data from Suaeda1 is missing in September due to technical issues.

Figure 2. Cumulative CH₄ emissions during the growing season 2012 from two *Suaeda salsa* wetlands, two
 Phragmites australis wetlands and one rice paddy during 2012 in the Liaohe Delta, northeast China. The
 points represent integrals of the monthly mean values from six plots at each site. Measurements are
 missing from Phrag1 in August due to flooding.

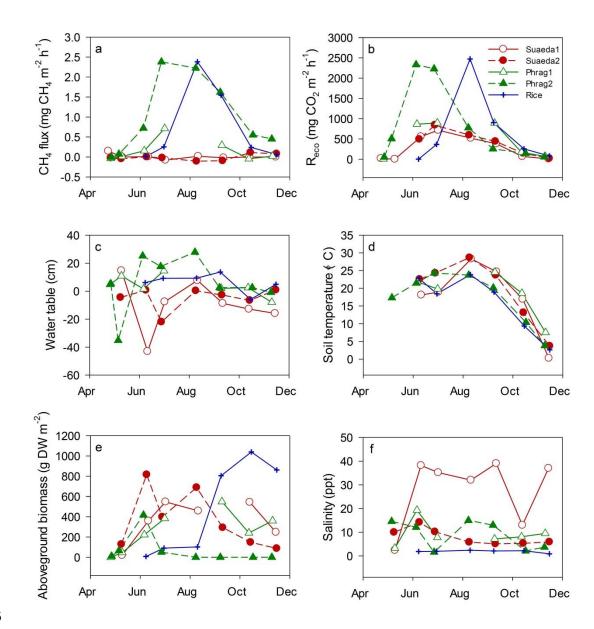
Figure 3. Relationship between CH₄ emission rates and (a) soil temperature, and (b) water table, in two
 Phragmites australis wetlands and a rice paddy in the Liaohe Delta, northeast China. Data points after
 cutting the vegetation at Phrag2 are represented by downward triangles (Phrag2-cut). Measurements were
 done from April to November 2012.

731 Figure 4. Relationship between salinity and CH₄ emission rates in two Suaeda salsa wetlands, two

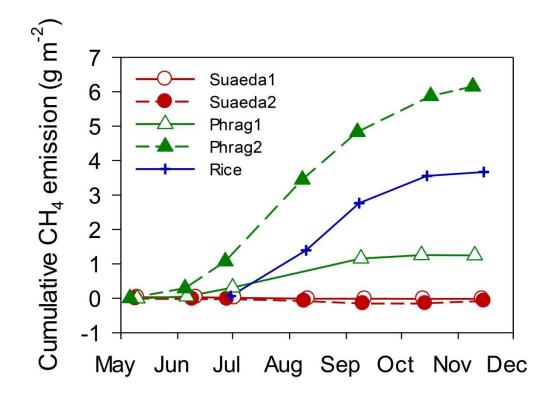
732 Phragmites australis wetlands and one rice paddy during 2012 in the Liaohe Delta, northeast China. Data

points after cutting the vegetation at Phrag2 are represented by downward triangles (Phrag2-cut).

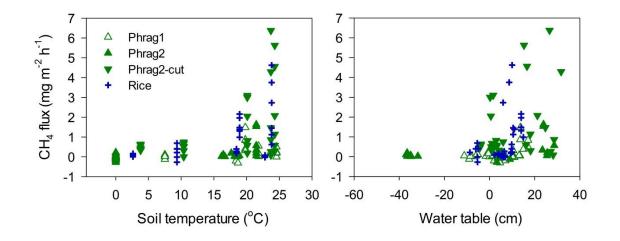
734 Measurements were done from April to November 2012.





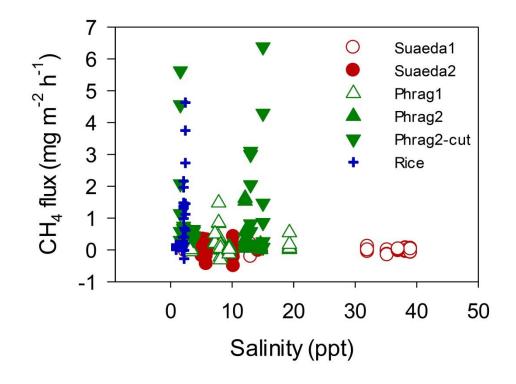












746 Fig. 4