Dear Bronwen Currie,

Thanks for the thorough editing and commenting on our manuscript. Major concerns of the reviewers were that a general focus in the draft manuscript is missing and that several parts are not well connected. We now tried to resolve this by significant shortening and rewriting large parts of the text and starting discussions on topics that are now at the beginning to gain attention. Of course, it is difficult to provide a review on such a topic that addresses all aspects and all relevant literature fully adequately. By now focusing on potential feedback coupling and control mechanisms in the OMZ, we restricted the topic and could hopefully address this problem. We further put a stronger emphasis on the quantitative aspects and added a table on major primary production, N cyle and export production parameters.

While it has been suggested to cut the section on marine zooplankton and on viruses, we think they indeed impact largely on biogeochemistry of the water column and therefore now put more emphasis on this in order to make it clear, why these parts should not be cut.

The direct suggestions (such as missing references and changes concerning the wording in the abstract) were addressed.

Best regards,

Carolin Löscher

- 1 Water column biogeochemistry of oxygen minimum zones in the eastern
- 2 tropical North Atlantic and eastern tropical South Pacific Oceans

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1 Abstract

2 Recent modeling results suggest that oceanic oxygen levels will decrease significantly over the next decades to centuries in response to climate change and altered ocean circulation. 3 4 Hence the future ocean may experience major shifts in nutrient cycling triggered by the expansion and intensification of tropical oxygen minimum zones (OMZs), which are 5 connected to the most productive upwelling systems in the Ocean. There are numerous 6 feedbacks among oxygen concentrations, nutrient cycling and biological productivity; 7 however, existing knowledge is insufficient to understand physical, chemical and biological 8 9 interactions in order to adequately assess past and potential future changes.

In the following, one decade of research performed in the framework of the collaborative 10 research center SFB 754 focusing on Climate- Biogeochemistry interactions in tropical OMZs 11 is summarized. The influence of low environmental oxygen conditions on biogeochemical 12 cycles organic matter formation and remineralization, greenhouse gas production and the 13 ecology in OMZ regions of the eastern tropical South Pacific compared to the weaker OMZ of 14 15 the eastern tropical North Atlantic were investigated. The impact of sulfidic events on water column biogeochemistry, as well as their specific microbial community capable of highly 16 17 efficient carbon fixation, nitrogen turnover and N₂O production is discussed. Based on our findings, a coupling of primary production and organic matter export via the nitrogen cycle is 18 19 proposed, which may, however, be impacted by several additional factors, e.g. micronutrients, particles acting as microniches, vertical and horizontal transport of organic material and the 20 21 role of zooplankton and viruses therein.

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1 1 Introduction

2 Eastern boundary upwelling systems are ocean areas where cold and nutrient-rich waters are upwelled to the sea surface, where they fuel high biomass production (Carr, 2002; Chavez and 3 4 Messie, 2009). While only covering 0.2% of the ocean, those upwelling areas account for 5 about 50-58% of global fish catch (Pauly and Christensen, 1995). Two eastern boundary 6 upwelling systems were subject to this study: the upwelling off Mauritania, located in the 7 eastern tropical North Atlantic (ETNA) and the upwelling system off Peru located in the eastern tropical South Pacific (ETSP). While both systems are characterized by intense 8 primary production, their biogeochemical properties differ strongly (Karstensen et al., 2008), 9 which partially results from a combination of different water mass ages and characteristics 10 (Körtzinger et al., 2004), topography and atmospheric impacts (e.g. Duce et al., 2008). A 11 major difference between the ETNA and the ETSP is the intensity of the oxygen minimum 12 zone (OMZ) associated with those upwelling regions (Capone and Hutchins, 2013): The 13 ETNA OMZ has O_2 concentrations typically above 40 µmol kg⁻¹, whereas the large and 14 persistent OMZ in the ETSP located off Peru and Chile has O₂ concentrations below the 15 detection limit based on conventional methods (~2 μ mol kg⁻¹, Figure 1) with sometimes even 16 17 sulfidic conditions on the shallower shelf (Schunck et al., 2013).

Besides the age of the water mass and other physical constraints, biological remineralization 18 and respiration processes consume O_2 below the highly productive surface waters and 19 contribute therefore to the development and maintenance of OMZ waters (Walsh, 1981, 20 21 Quinones et al., 2010). The intensity of the OMZ may therefore be determined by a positive feedback, with increased primary production leading to enhanced organic matter export back 22 23 to underlying O_2 -depleted waters (Dale et al.2015). As a consequence of enhanced organic matter export, respiration processes may increase. Stronger anoxia on the other hand would 24 25 promote O₂ sensitive N loss processes, therefore creating an nitrogen (N) deficit in upwelled waters. This would then stimulate N_2 fixation at the sea surface and enhance again primary 26 production. In how far a feedback between primary production, organic matter 27 remineralization and the N cycle is a valid model in OMZ waters and what role sulphidic 28 29 conditions take in there is critical to understand. Modeling results (Bopp et al., 2013; Cocco et al., 2013), predict that O₂ levels will decrease 30 significantly over the next decades in response to climate change and eutrophication. Hence, 31

- 32 the future ocean may experience major shifts in nutrient cycling triggered by the possible
- expansion and intensification of tropical OMZs (Codispoti, 2010). Currently, the estimated
- 34 volume of OMZs with O₂ concentrations $<20 \mu$ mol kg⁻¹ is about 1% of the global ocean

volume (Lam and Kuypers, 2011). Approximately 0.05% of the global ocean volume has O_2 1 levels below 5 μ mol kg⁻¹. The effects of O₂-sensitive nutrient cycling processes occurring in 2 these relatively small regions (Codispoti, 2010) are conveyed to the rest of the ocean (see e.g. 3 Deutsch et al. 2007). Hence comparatively "small" volumes of OMZs can significantly impact 4 5 nutrient budgets, biological productivity and the overall potential for CO₂ fixation of the ocean. An important factor is further that deoxygenation of OMZs has been proposed to 6 increase the production of the greenhouse gas nitrous oxide (N₂O) (Codispoti, 2010). 7 Therefore, understanding the present biogeochemistry of those systems and exploring the 8 9 potential to respond to climate change is critical. The following review of the major biogeochemical processes in OMZ waters is based on 10 studies of the Collaborative Research Centre 754 (SFB754) "Climate-Biogeochemistry 11 Interactions in the Tropical Ocean" (www.sfb754.de). The comparison between the ETNA 12 13 and ETSP upwelling systems, their OMZs and differences in remineralization processes and associated marine sources and sinks of important nutrient elements are discussed in order to 14

understand potential controls on the intensity of those OMZs, as well as their future
 development.

17

18 2 Primary production in the ETSP and ETNA

In eastern boundary upwelling systems, phytoplankton blooms are stimulated by nutrient 19 supply from upwelled waters and provide the basis for vibrant ecological systems. Both, the 20 ETNA and the ETSP, are major sites of primary production (Longhurst, 1995); with the 21 ETNA exceeding primary production of the ETSP by a factor of ~2 depending on the applied 22 method (see Table 1 for an overview of major primary production-related parameters). This 23 difference may be explained, e.g. by the stoichiometry of the macronutrients N and phosphate 24 (P), with the deficit of N based on a Redfieldian equilibrium of N:P = 16:1 being significantly 25 stronger in the ETSP compared to the ETNA (Deutsch et al., 2007). This difference may be 26 due to intensified N loss mirrored by a strongly positive d^{15} N-nitrate signal (Ryabenko et al., 27 2012) in the more O_2 depleted water column and sediments of the ETSP. On the other hand, 28 enhanced P release from the sediments at decreasing O_2 (Ingall and Jahnke, 1994), or a 29 difference between N and P remineralization from organic material (Jilbert et al., 2011) may 30 impact decreased N:P ratios. A stronger N deficit in the water column may influence primary 31 production in different ways: It may either stimulate N₂ fixation in order to replenish the N 32

1 deficit, or it may stimulate non-Redfield primary production. A major aspect to detangle these

2 potential responses is therefore to understand the community composition.

Franz et al. (2012a) reported in situ observations along an east-west transect in the ETSP at 3 4 10°S stretching from the upwelling region above the narrow continental shelf to the wellstratified oceanic section of the eastern boundary regime. The study showed that new 5 6 production in the coastal upwelling was driven by large-sized phytoplankton (e.g. diatoms) 7 with generally low N:P ratios (<16:1), thus speaking for non-Redfield surface water primary 8 production. A deep chlorophyll *a* maximum consisting of nano- (*Synechococcus*, flagellates) and microphytoplankton occurred within a pronounced thermocline in subsurface waters 9 10 above the shelf break. Here, intermediate particulate N:P ratios were close to Redfield proportions. High PON:POP (>20:1) ratios were observed in a stratified open ocean section, 11 12 coinciding with a high abundance of the pico-cyanobacterium *Prochlorococcus*. Excess P was present along the entire transect but did not appear to stimulate growth of N₂ fixing 13 cyanobacteria, as pigment fingerprinting and phylogenetic studies did not indicate the 14 presence of diazotrophic cyanobacteria at most of our sampling stations (Franz et al., 2012a; 15 Löscher et al., 2014). These findings are mostly in accordance with other studies from this 16 area (Bonnet et al., 2013; Fernandez et al., 2011; Turk-Kubo et al., 2014). Therefore, the 17 excess P generated within the OMZ seemed to be rather consumed by non-Redfield processes, 18 19 i.e. primary production by large phytoplankton found in shelf surface waters, instead of stimulating surface N₂ fixation. A possible explanation can be deducted from the optimality-20 based model of N₂ fixation by Pahlow et al. (2013). The model is based on the assumption that 21 22 natural selection should tend to produce organisms optimally adapted to their environment. The competitive advantage of diazotrophs is most pronounced under conditions of low 23 dissolved inorganic N and increased dissolved inorganic P (DIN, DIP) availability (Houlton et 24 al., 2008). The ability to compete for DIP is therefore less important at high DIP. Thus, high P 25 concentrations above the ETSP OMZ might actually reduce the selective advantage of 26 diazotrophs compared to non-Redfield primary producers. This could partially explain why 27 cyanobacterial N2 fixers were apparently not stimulated by excess phosphate in surface waters 28 29 of the abovementioned transect.

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2.1 The impact of changing N:P ratios as a result of ocean deoxygenation: What can
 we learn from mesocosm experiments and bioassay studies?

A series of on board mesocosm experiments and bioassay incubations were performed in 1 order to identify nutrient limitations in both areas and to specifically address the impact of 2 stoichiometry on primary production. Despite the fundamental differences between the 3 ETNA and ETSP with regard to the N deficit, the results of short-term mesocosm experiments 4 implied N limitation of surface plankton communities in both areas (Franz et al., 2012a; Franz 5 et al., 2012b). Further, the partitioning and elemental composition of dissolved and particulate 6 7 organic matter was investigated. Maximum accumulation of POC and PON was observed under high N supply, indicating that primary production was controlled by N availability. Part 8 9 of the excess P was consumed by non-Redfield production, predominantly by diatoms, as also observed from direct monitoring as described above. While particulate N:P of the 10 11 accumulated biomass generally exceeded the supply ratio (Franz et al., 2012b), excess P of the dissolved nutrient pool was channeled into release of dissolved organic phosphorus (DOP) 12 13 by phytoplankton. These results demonstrated that excess P upwelled into the surface ocean overlying O₂-deficient waters represents a net source for DOP and motivated further 14 15 dedicated mesocosm experiments in the ETNA to elucidate the fate of DOP. While the direct monitoring and the results of mesocosm studies were strongly speaking for a shift to non-16 17 Redfield primary production due to changes in N:P, a general stimulating effect of DOP on N₂ fixation has been observed (Meyer et al., 2015). This is in line with a recent modeling study 18 based on large-scale surface data sets of global DON and Atlantic Ocean DOP. Here, the 19 model suggests an important role of DOP for stimulating growth of N₂ fixing organisms 20 (Somes and Oschlies, 2015). This model indicates that the marine N- budget is sensitive to 21 DOP, provided that access to the relatively labile DOP pool expands the ecological niche for 22 diazotrophs. Taken together, changes in N:P may lead to a combination of both, non-Redfield 23 primary production and enhanced N₂ fixation via DOP. 24

Besides a direct effects of N:P ratios, primary production and N₂ fixation, due to the 25 26 compareably high Fe requirements of the diazotrophs (Gruber, 1997) is largely influenced by 27 trace metal availability (Mills et al., 2004). Comparing the ETNA and ETSP regions, an obvious difference with regard to potentially limiting nutrients is related to the iron (Fe) 28 source: In the ETNA, Saharan dust input contributes 71-87% of dissolved Fe to the water 29 (Conway, 2014). Several studies highlighted the importance of atmospheric Fe supply to the 30 ETNA (Voss et al., 2002; Mills et al., 2004) as a major factor of primary production. A 31 comparable atmospheric Fe source is however missing in the ETSP (Baker et al., 2016). 32 Previous studies (Scholz et al., 2014) identified the ETSP Fe supply as benthic, however, the 33

question on how much Fe is transported from the sediments to the sea surface is so far not
 fully clarified.

Results of bioassay incubations and correlation studies demonstrated that primary production and N_2 fixation in this region respond significantly to Fe additions (Dekaezemacker et al., 2013). N_2 fixation could further be directly limited by inorganic nutrient availability, or indirectly through the stimulation of primary production and the subsequent excretion of dissolved organic matter and/or the formation of micro-environments favorable for heterotrophic N_2 fixation (Dekaezemacker et al., 2013).

9

10 2.2 What is the role of N_2 fixation for primary production in the ETNA and ETSP?

Several studies (Voss et al., 2002; Mills et al., 2004; Langlois et al, 2005 & 2007) 11 demonstrated the important role of N2 fixation for surface primary production in ETNA 12 waters. Voss et al. (2002) estimated an average N_2 fixation of 24-140 µmol m⁻² d⁻¹ translating 13 into a contribution of 0.1-0.7% to primary production assuming Redfield stoichiometry on the 14 15 basis of the rates given in Table 1. This is below the global average of 5.3% (Duce et al., 2008), potentially due to the relatively high deposition of reactive N via Saharan dust input. 16 17 For the ETSP, N₂ fixation was higher compared to the ETNA with rates of 25-657 μ mol m⁻² d⁻¹ (Dekaezamacker et al., 2013; Löscher et al., 2014); while C fixation was rather lower 18 (Table 1). Here, theoretically, N₂ fixation contributes 0.2-7.1% C fixation (Table 1). However, 19 while N loss does not play a role in the ETNA water column (see, e.g., Bange et al. (2010)) 20 with the exception of O₂ depleted mesoscale eddies (Löscher et al., 2015), high N loss 21 removes between one and three orders of magnitude more N (Kalvelage et al., 2013; Table 1) 22 than is made available by N_2 fixation. This would decrease the contribution of N_2 fixation to 23 C fixation to zero. When comparing N₂ fixation to N loss, it must be considered that first N 24 loss has only been detected on and close to the shelf, while N_2 fixation rates were detectable 25 throughout the OMZ water column (Fig. 2). Second, while N_2 fixation is measured via direct 26 27 N₂ incorporation and herefore mirroring in situ rates, N loss is likely being overestimated as 28 measured following addition of the substrates, which may artificially stimulate the respective process (up to 2-3 orders of magnitude, as discussed in Kalvelage et al., 2013). An entirely 29 correct budget of N₂ fixation vs. N loss based on rate measurements is therefore difficult to 30 31 obtain.

In both areas, N₂ fixation may however be considered important for the productivity of the 1 respective system. Still, given the previously described observations of non-Redfield primary 2 production, the contribution to C fixation remains to be fully established. 3 From the comparison of the N_2 fixation in the ETNA and ETSP, the question raises why there 4 is such a strong difference between those systems. A possible explanation may be found in the 5 character of the diazotrophic communities: While the classical view of oceanic N₂ fixation 6 7 mainly attributed to phototrophic cyanobacteria, such as Trichodesmium or Crocosphaera, (Capone et al., 1997; Zehr and Turner, 2001) may be mostly true for the ETNA (e.g. Langlois 8 et al., 2005, 2007; Großkopf et al., 2012), a different community of diazotrophs is present in 9 the ETSP. 10 A growing number of different *nifH* sequences (the key functional gene of N_2 fixation, 11 12 encoding the α subunit of nitrogenase) detected within the Peruvian OMZ (Bonnet et al., 2013; Dekaezemacker et al., 2013; Fernandez et al., 2011; Löscher et al., 2014; Turk-Kubo et 13 al., 2014), did not belong to common oxygenic phototrophs, but to some unknown 14 diazotrophic microorganisms that might be specifically adapted to O_2 deficient conditions. 15 These diazotrophs, as well as the extension of their habitat to deeper waters might be one 16 reason for the possible underestimation of N gain compared to N loss in the ocean (Codispoti, 17 18 2007). In combination with a novel method for N_2 fixation rate measurements (Mohr et al., 2010) (Großkopf et al., 2012) revealed up to 6-fold higher N_2 fixation rates when considering 19 N_2 fixation below the euphotic zone. When extrapolated to all ocean basins this resulted in a 20 N₂ fixation rate of 177 ± 8 Tg N yr⁻¹, which-depending on the assumed budget may balance 21 22 50-100% of oceanic N loss (Codispoti, 2007; Gruber and Sarmiento, 1997). 23

24 2.3 In how far is N_2 fixation in the ETSP OMZ coupled to N loss?

Model studies (Deutsch et al., 2007), assuming that a N-deficit resulting from N loss or enhanced P release (Ingall and Jahnke, 1994) provides a niche for diazotrophs and that a coupling of N loss in OMZs and N₂ fixation in overlying surface waters might restore the N:P ratio towards Redfield proportions.

In the ETSP OMZ, N is indeed continuously removed by the anaerobic oxidation of ammonium (anammox) (Francis et al., 2007; Kuypers et al., 2005; Kuypers et al., 2003; Thamdrup and Dalsgaard, 2002), which has been shown to be the dominating N loss process in this region (Kalvelage et al., 2013, Lam et al., 2009), as well as in other OMZ waters (off Namibia (Kuypers et al., 2005), Peru (Hamersley et al., 2007) and Chile (Thamdrup et al., 2006)). Moreover, N is (i) lost by denitrification (the 4-step reduction of NO₃⁻ to N₂ (Devol, 2008)), e.g. in the Arabian Sea OMZ (Ward et al., 2009), where denitrification has been
identified the dominant N loss process. However, off Peru, denitrification was only detectable
in connection to sulfidic events (Kalvelage et al, 2013, Schunck et al., 2013).

4 The prevalence of novel nifH genes and active N₂ fixation, derived from samples collected directly in the OMZ waters off Peru, where anammox bacteria were abundant and active 5 6 (Kalvelage et al., 2013; Löscher et al., 2014), supports the view of a positive feedback 7 between N loss and N gain communities (Figure 3). Evidence for co-occurrence of denitrification and N₂ fixation has previously been documented only for an anoxic lake (Halm 8 et al., 2009), and for cyanobacterial aggregates in the Baltic Sea (Klawonn et al., 2015). 9 Recent investigations from Baltic Sea sediments on N2 fixation and diazotrophic abundance in 10 sediments show, however, that a very close spatial link between N loss and N₂ fixation might 11 exist (Bertics et al., 2013). Still, too little is currently known about the interactions among the 12 stoichiometry of inorganic nutrient supply, primary production, N_2 fixation, and 13 remineralization under anoxic conditions, to allow a definite characterization of the conditions 14 15 leading to fixed-nitrogen exhaustion in the OMZs.

This coupling which seems to exist in OMZ waters may in fact have far reaching 16 consequences: While N loss may provide a niche for N₂ fixation, model studies, suggest that 17 denitrification of N₂ fixation-derived organic matter may lead to a net N loss that further 18 stimulates N₂ fixation, because 120 moles of nitrate per mole of phosphorus are used to 19 remineralize Redfield organic matter via denitrification (Landolfi et al., 2013). In contrast, N2 20 21 fixation fixes only 16 moles N (per mole P). Because of those stoichiometric constraints, any addition of fixed N to the surface ocean only exacerbates the problem (Canfield, 2006) unless 22 23 the corresponding primary production is prevented from being remineralized in the underlying OMZ (Landolfi et al., 2013). Lipschultz et al. (1990) indeed stated that N loss in the ETSP 24 25 OMZ is high enough to respire all produced organic material. Only by spatial or temporal decoupling of N₂ Fe limitation or dissolved organic matter cycling, the N inventory may 26 27 stabilize, otherwise the OMZ would become completely void of fixed inorganic N. Whether these stoichiometric constraints are valid for anammox as dominant N loss process instead of 28 29 denitrification is however not clear.

Concerning the stoichiometric aspects of ultimate N loss from OMZ waters, a to date largely disregarded aspect should be taken into consideration: As shown for the Gotland basin (Jilbert et al., 2011) enhanced preferential P release from organic matter remineralization was quantitatively important for creating a N deficit. This preferential P release was present in the water column and was further increased under O_2 depleted, reduced conditions. Although the quantitative contribution to the N deficit in the ETSP is not entirely clear, yet, it may act as a factor decoupling the 'vicious' cycle between N_2 fixation and N loss Landolfi et al., 2013), as shifting the abovementioned stoichiometric constraints.

5 2.4 Factors determining N loss

6 The net rate of N loss in OMZs is determined by the balance of remineralization of sinking 7 particulate organic carbon (POC) and O_2 supply to the OMZ. Interestingly, recent studies 8 attributed the fact whether anammox or denitrification dominates in a certain environment to 9 organic matter composition and availability (Babbin et al., 2014). While the supply of O_2 is 10 mostly determined by physical transport, the rate of N loss depends on the activity of the 11 bacteria responsible for denitrification and anammox as well as the POC export and sinking 12 velocity.

The intensity of this feedback may be overestimated in current biogeochemical models, owing 13 to spurious nutrient trapping (Dietze and Loeptien, 2013). The extent of the coupling between 14 15 primary production at the surface and denitrification in the OMZ, and hence the strength of the positive feedback, is a strong function of the elemental (C:N:P) stoichiometry of the 16 17 exported primary production. Phytoplankton C:N:P stoichiometry in turn is influenced by the stoichiometry of inorganic nutrients (Franz et al., 2012a; Franz et al., 2012b). Recently 18 developed process models of primary production and N_2 fixation (Pahlow et al., 2013; Pahlow 19 and Oschlies, 2013) specifically address the response of phytoplankton elemental 20 21 stoichiometry to ambient nutrient concentrations and light.

It is generally assumed that both zooplankton and heterotrophic bacteria vary much less in 22 their elemental stoichiometry than phytoplankton (e.g., Touratier et al. (2001)). In both cases, 23 the heterotrophs appear to respond to variable nitrogen content in their food by regulating 24 25 their gross growth efficiency for carbon (Anderson and Williams, 1998; Kiørboe, 1989). In 26 OMZ regions, this implies that strong nutrient limitation in the surface ocean, which is 27 associated with high C:N ratios in primary producers (e.g., data used in Pahlow et al. (2013)), 28 should intensify denitrification in the OMZ relative to the export flux from the surface. Higher surface nutrient concentrations would then be expected to reduce C:N ratios in the export flux 29 30 and hence have a somewhat mitigating effect. Since denitrification and anammox in the OMZ cause lower nitrate concentrations in upwelled waters, the variable stoichiometry of 31 phytoplankton could add to the positive feedback between denitrification and N_2 fixation by 32 increasing C:N ratios in response to decreasing surface nitrate concentrations. 33

Combined ¹⁵N-incubation experiments and functional gene expression analyses indicate that 1 anammox in the Peruvian OMZ benefits from other N-cycling processes for reactive 2 substrates (Kalvelage et al., 2011). Excretion of ammonium and other reduced N-compounds 3 by diel vertical migrators was also proposed (Bianchi et al., 2014), but recent experiments 4 indicate that ammonium excretion of diel vertical migrators is strongly reduced at anoxia 5 (Kiko et al. 2015 a, Kiko et al. 2015 b). Additionally, anammox activity has been described to 6 7 depend on export of organic matter (Kalvelage et al., 2013), potentially resulting from the availability of ammonium recycled from particulate organic N (Ganesh et al., 2015). In the 8 absence of significant denitrification, these results indicate that anammox relies on NH_4^+ 9 oxidation and NO_3^- reduction as NO_2^- source. Further, NH_4^+ may be derived from 10 remineralization of organic matter via NO₃⁻ reduction with a possibly important role of 11 12 microaerobic respiration (Kalvelage et al., 2015). The overlap between aerobic and anaerobic 13 N-cycling processes in particular in the coastal shelf waters and the upper part of the OMZ is supportive of microaerobic activity in the OMZ. As DNRA was insignificant in the water 14 15 column during our studies in the ETSP, sedimentary fluxes could be an important ammonium source, particularly for the inner shelf sediments (Bohlen et al., 2011; Kalvelage et al., 2013). 16 17 However, it has been suggested that sulfate reduction is more widespread in OMZ waters than previously believed and could be responsible for substantial NH_4^+ production (Canfield et al., 18 2010) and sulfate reducers have been detected in the Peruvian OMZ (Schunck et al., 2013). 19 Direct evidence for the actual link between sulfate reduction and NH₄⁺ production is, 20 however, still missing. 21

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3 What part is lost from the upwelling region: Organic matter export and

24 remineralization in the ETSP OMZ

25 3.1 Sinking of particles

Knowledge about particle fluxes in areas of tropical OMZs is scarce and predominantly 26 derived from deep moored traps (Honjo et al., 2008) or models (Dale et al., 2015, Table 1). 27 Only few studies have addressed upper ocean export fluxes and mesopelagic flux attenuation 28 in tropical OMZs (Martin et al. (1987), Devol and Hartnett (2001) and VanMooy et al. (2002) 29 for the Eastern Tropical Pacific by means of surface tethered sediment traps, Buesseler et al. 30 (1998) for the Arabian Sea by means of ²³⁴Th, and Iversen et al. (2010) at the northern edge of 31 32 the ETNA OMZ by means of particle camera profiling). In the eastern tropical north Pacific (ETNP; Martin et al. (1987); Van Mooy et al. (2002); Devol and Hartnett (2001)), as well as 33

in the ETSP (Martin et al. 1987; Dale et al. (2015)) mesopelagic POC fluxes were less 1 attenuated with depth (Martin curve exponent 'b' of 0.32-0.81) compared with the widely 2 used "open ocean composite" of b=0.86 (Martin et al. 1987). Those studies indicate that a 3 greater proportion of the sinking OM escapes degradation while sinking through the eastern 4 tropical Pacific OMZ. On the other hand, it has been shown that microbial degradation of 5 organic N and proteins under suboxia is not strongly affected (Pantoja et al., 2009; Pantoja et 6 7 al., 2004; Van Mooy et al., 2002). In addition, organic matter degradation seems not to be significantly affected by decreased O₂ (Dale et al., 2015). 8

9 Still, little is known about the microbial controls on the decomposition of organic matter under lower O₂ concentrations. Microorganisms are generally considered responsible for most 10 of the remineralization in the ocean. This view is probably justified with respect to carbon, 11 given the high rates of microbial respiration (del Giorgio and Cole, 1998). Owing to the 12 relatively low N and phosphorous (P) content of dissolved organic matter, however, bacteria 13 may be less important for the remineralization of N and P and in fact often compete with 14 15 phytoplankton for inorganic nutrients in the surface ocean (Anderson and Williams, 1998; Pahlow and Vézina, 2003). Remineralization of N and P may thus be largely due to 16 17 zooplankton activity (Caron et al., 1988; Garber, 1984; Pahlow et al., 2008).

Classically, the most abundant organisms detected in OMZs belong to the Proteobacteria, 18 Bacteroidetes, Thaumarchaeota of the marine group A, Actinobacteria and Planctomycetes 19 (Schunck et al., 2013; Wright et al., 2012). Several candidate clusters have previously been 20 identified among which are the SAR11, SAR324 and SUP05 clusters (Schunck et al., 2013; 21 Wright et al., 2012). Most investigations of the microbial phylogenetic and functional 22 diversity resort to observing and correlating changes in oxygen concentrations to changes in 23 the microbial phylogenetic diversity. Indeed, several studies, including our own datasets 24 corroborate this idea: A combined statistical analysis of our metagenomic data of the ETSP 25 26 OMZ (Kalvelage et al., 2015) and datasets from the Chilean OMZ (Canfield et al., 2010; Stewart et al., 2011) has resulted in a partitioning of the OMZ in 5 different habitats, namely 27 surface, subsurface (defined as below the mixed layer and above waters with $O_2 > 20 \mu \text{mol kg}^-$ 28 ¹), oxyclines, OMZ core ($O_2 < 5 \mu mol kg^{-1}$) and sulfidic waters (Figure 4). High-resolution 29 sampling in the eastern tropical North Pacific OMZ has shown that the microbial richness is 30 highest at the base of the euphotic zone and the upper oxycline (Beman and Carolan, 2013), 31 often along with high organic flux, low O₂ concentrations and dynamic cycling of C, N, and 32 sulfur (S). This may be interpreted in a way that the upper oxycline is of higher importance 33

34 for remineralization than the OMZ.

1

3.3 The impact of zooplankton for organic matter export and remineralization 2 An important consideration for explaining the lowered flux attenuation in the OMZ could be 3 4 deducted from the diminished abundance of metazoans in the core of the OMZ. If particles 5 are not repackaged, fed upon, or destroyed, they might sink at greater speeds through the OMZ, which would result in decreased degradation. 6 Zooplankton and nekton organisms are essential components of the biological pump as they 7 8 egest packaged organic matter as rapidly sinking fecal pellets. Many zooplankton and nekton species also feed in surface waters during the night and migrate to midwater depth at daybreak 9 to avoid predation (Lampert, 1989), and to conserve energy (McLaren, 1963). This behavior 10 11 is known as diel vertical migration (DVM) and also contributes to the activity of the 12 biological pump as it enhances the export of organic matter from the photic zone by continued respiration, excretion and egestion in mid-water layers (Burd et al., 2010; Hannides et al., 13 14 2009; Robinson et al., 2010; Steinberg et al., 2000). In addition to changes in temperature with depth, DVM organisms experience low O_2 concentrations during the daytime in OMZ 15 regions (Brewer and Peltzer, 2009; Paulmier et al., 2011), and O₂ concentrations below a 16 certain threshold level can restrict DVM of most zooplankton and nekton (e.g. Hauss et al 17 Biogeosciences Discussions). On a regional scale, the upper boundary of the oxycline is the 18 single most critical factor structuring the habitat of most zooplankton organisms in the 19 Peruvian upwelling system (Escribano et al., 2009). Nevertheless, some specifically adapted 20 species are able to downregulate their metabolic activity at low oxygen levels and can remain 21 22 at OMZ depth (non-migrators) or actively migrate into suboxic or anoxic OMZs (Seibel 2011, Kiko et al. 2015 a, b). 23 Abundance and biomass of metazoans living permanently at extremely low oxygen 24 concentrations is rather low (Auel and Verheye, 2007; Escribano et al., 2009; Fernández-25 Álamo and Färber-Lorda, 2006; Saltzmann and Wishner, 1997; Wishner et al., 1998) although 26 27 animals have evolved physiological (such as metabolic suppression) and/or morphological adaptations (such as increased gill surface area) allowing them to cope temporarily or 28 29 permanently with O₂ depleted conditions (e.g. copepods, e.g. *Eucalanus inermis*, (Flint et al., 30 1991), euphausiids, e.g. Euphausia mucronata (Antezana, 2009), decapods (Pearcy et al., 31 1977), cephalopods, e.g. Dosidicus gigas (Rosa and Seibel, 2010), and teleosts (Friedman et al., 2012; Luo et al., 2000)). According to Seibel (2011), adaptations to low oxygen levels are 32 needed below approximately 40 μ mol O₂ kg⁻¹. Strong physiological adaptations seem thus 33

necessary to thrive in the ETSP OMZ, but not in the ETNA OMZ where O₂ concentrations are 1 normally greater than 40 μ mol kg⁻¹ (Teuber et al., 2013). 2 Estimates of zooplankton and nekton mediated carbon fluxes in OMZ regions are rare. For the 3 4 northern Chilean upwelling in the ETSP, Escribano (2009) found that migrations of only two 5 key species (*Eucalanus inermis* and *Euphausia mucronata*) contribute approximately 7.2 g C m⁻² d⁻¹ to the OMZ through respiration, mortality, and production of fecal pellets within the 6 7 OMZ. However, these estimates are probably too high, as the reduction of respiration at low oxygen levels (Kiko et al. 2015, Kiko et al. 2015b) was not accounted for in the calculations. 8 9 As stated above, a particular role of DVMs for the N cycle could result from the secretion of 10 ammonium: Ammonium is an important nutrient in the anammox reaction which represents 11 nearly 30-50% of N-loss activity in the OMZ (Codispoti et al., 2001; Emery et al., 1955; Gruber, 2004). Bianchi et al. (2014) suggested that DVMs could supply as much as 30% of 12 13 the ammonium for the anammox reaction, assuming no reduction of the rate ammonium excretion under OMZ conditions. This assumption is unlikely to hold, however, as 14 15 ammonium excretion is e.g. reduced four-fold in the squat lobster Pleuroncodes monodon (Kiko et al., 2015a; Kiko et al., 2015b) and six-fold in the euphausiid Euphausia mucronata 16 17 (Kiko et al., 2015a; Kiko et al., 2015b) upon exposure to anoxia at OMZ temperatures. Thus, the significance of excretion by zooplankton as a source of ammonium for the anammox 18 19 reaction remains to be established. 20 3.4 In how far could viruses impact primary production and organic matter feedback? 21 A recent model study quantifying the effect of viruses on ecosystem function in the ocean 22 demonstrated that viruses affect biological productivity and remineralization (Weitz et al., 23 2015). In line with field studies (Breitbart, 2012), this model showed enhanced organic matter 24 cycling, e.g. by cell lysis. Viruses lyse $\sim 10-40\%$ of the present prokaryotes every day (Suttle, 25 26 2005), which may-besides generally supplying nutrients to the surrounding waters- impact stoichiometry on smaller scales. Specifically, (cyano)phages in the ETSP have been shown to 27 release micronutrients such as Fe into surrounding waters at an estimated flux of 10 pmol L^{-1} 28 d⁻¹ (Poorvin et al., 2004). Likewise, virus-induced bacterial lysis was calculated to contribute 29 \sim 1-6 Gt N a⁻¹ to bacterial primary production, which would significantly support 30 phytoplankton production (Shelford et al., 2012). The transfer of nutrients from living 31

- organisms into the dissolved phase is called the 'viral shunt' (Breitbart, 2012). Besides the
 'viral shunt', the model showed a reduced transfer of organic material to higher trophic levels,
- 34 which was interpreted to stabilize primary production. Quantitatively, net primary production

1	was found increased by $\sim 11\%$ in the presence of viruses. This strongly speaks for a viral
2	impact on the efficiency of the biological pump (Azam, 1998). On the other side, viruses were
3	shown to influence particle formation and disaggregation through discharging adhesive cell
4	components (Peduzzi and Weinbauer, 1993) and cell lysis (Weinbauer et al., 2011),
5	respectively.
6	In OMZ waters, highly specific viral communities have been discovered which show
7	unusually low diversity and a low viral-to-microbial ratio (VMR) (Cassmann et al., 2012).
8	Specific viruses appear to be only present in OMZ waters as exemplarily shown by genomic
9	studies of uncultivated SUP05 bacteria isolated from the ETSP OMZ (Roux et al., 2014).
10	Interestingly, various genes involved in the cycling of nitrogen and sulfur have also been
11	found in viromes of ETSP waters (see Tables S3 and S4 in Cassman et al. (2012), Roux et al.,
12	2014)).
13	Recent studies analyzing samples from the weaker ETNA OMZ indicated that one of the most
14	abundant archaeal nitrifiers in the ETNA OMZ (Thaumarchaeota, "Cand. Nitrosopelagicus
15	brevis") contains several viral genes in its genome arguing that this archaeon is infected by an
16	OMZ-specific hitherto uncharacterized virus (Neulinger and Schmitz, unpublished results),
17	thus confirming earlier studies from the global ocean dataset (Santoro et al., 2015).
18	Considering that Cand. Nitrosopelagicus brevis is most likely the most important producer of
19	the greenhouse gas nitrous oxide in the ETNA and ETSP OMZs (Löscher et al., 2012), these
20	findings add a potential role for greenhouse gas production to the current picture of viruses in
21	the ocean.
22	

23 3.2 Physical fluxes of DOM

Besides particle fluxes and organic matter export via DVM, DOM transport is due exclusively 24 to physical horizontal and vertical transport processes, induced by mesoscale (horizontal 25 26 scales of 10 - 100 km) and sub-mesoscale (100 m - 10 km) motion and vertical fluxes due to diapycnal mixing. As an example of lateral eddy transport, elevated DOM concentrations 27 have been detected $(+11 \mu mol C L^{-1})$ in the Canada Basin within an eddy originating from the 28 shelf region (Mathis et al., 2007). Lasternas et al. (2013) suggested a mechanism for DOM 29 accumulation within anticyclonic eddies, where nutrient downwelling causes a progressive 30 oligotrophication, enhanced cell mortality and lysis, which results in additional DOM release. 31 Numerical model simulations of the Peruvian upwelling regime show that mesoscale 32 dynamics increase the downward and offshore export of nutrients and biomass out of the 33 coastal surface ocean (Lathuiliere et al., 2010). For the understanding of remineralization 34

processes and feedbacks in upwelling systems, a quantification of the material that is lost to 1 the open ocean is critical as it may directly impact the system's productivity. Gruber et al. 2 (2011) found that mesoscale eddy activity in upwelling regimes results in a net reduction of 3 biological productivity. Additionally, sub-mesoscale upwelling filaments can enhance the off-4 shelf flux of labile DOM (Alvarez-Salgado et al., 2001). Vertical velocities are higher at sub-5 mesoscale density fronts (Klein and Lapeyre, 2009; Levy et al., 2012; Thomas et al., 2008), 6 7 which are prominent features in eastern boundary upwelling systems (Durski and Allen, 2005). These vertical velocities often extend to below the mixed layer (Klein et al., 2008), 8 9 where they can drive sizeable vertical fluxes of solutes. Mahadevan (2014) proposes the subduction of organic matter-rich surface water into the subsurface layers within 10 11 submesoscale cold filaments as a new export mechanism, which differs strongly from export via particle sinking. In filaments the organic matter is subducted together with large amounts 12 of O₂, which then can directly be used for decomposition of organic matter. Vertical mixing 13 of DOM from the euphotic into to the upper mesopelagic zone is another important transport 14 15 mechanism in (sub)tropical waters (Hansell, 2002). The Bermuda Atlantic Time-Series Study provides a well-documented example of this process (Carlson et al., 1994). The efficiency of 16 17 the downward DOM transport depends on the concentration gradient of DOM between the surface layer and the OMZ, and on the activity of the microbial population along this gradient. 18 Produced by high primary production in upwelling regions, DOM can accumulate in the 19 euphotic zone with maximum concentrations of 100-300 µmol C L⁻¹ off Peru (Franz et al., 20 2012a; Romankevich and Ljutsarev, 1990). Due to the vicinity of the DOM-rich surface layer 21 above and the O₂-depleted waters below the shallow and sharp oxycline of the Peruvian 22 OMZ, physical vertical transport may bring large amounts of labile organic matter to the 23 OMZ, where it may be utilized by heterotrophic communities (Hoppe et al., 2000; Hoppe and 24 Ullrich, 1999; Pantoja et al., 2009). DOM supply via (sub-) mesoscale vertical transport 25 26 processes and diapycnal mixing may therefore contribute importantly to sustaining microbial 27 activity in the Peruvian OMZ and may thus largely impact biogeochemical cycles.

28

29 4 What role do sulfidic events play in the ETSP?

30 Oceanic sulfidic events are cases of extreme anoxia following periods of enhanced primary

31 production and organic matter export. They are understood to mostly originate from sulfide

32 production in sediments (Figure 5) and have been documented sporadically since the 19th

century for the ETSP OMZ (Burtt, 1852; Dugdale et al., 1977). To date sulfidic events have

34 been reported in three of the five OMZs by only a handful of studies and hence our current

understanding of their regulation, initiation and termination is still limited. Possible analogs 1 for oceanic events are permanently sulfidic areas in enclosed basins of the Baltic Sea (Brettar 2 et al., 2006; Brettar and Rheinheimer, 1991; Glaubitz et al., 2009), the Black Sea (Glaubitz et 3 al., 2010; Jørgensen et al., 1991; Sorokin et al., 1995), the Cariaco basin off Venezuela 4 (Hayes et al., 2006; Taylor et al., 2001; Zhang and Millero, 1993) and Saanich Inlet in Canada 5 (Tebo and Emerson, 1986; Walsh et al., 2009). Here, sulfide accumulates to milli-molar 6 concentrations under O₂ and nitrate-free conditions and is released by a diffusive flux into the 7 overlying pelagic water column where it reaches low micro-molar concentrations (Lavik et 8 al., 2009; Schunck et al., 2013). These events are then terminated or detoxified in the pelagic 9 water column by a community of sulfide-oxidizing bacteria. This occurs when sulfide and 10 11 nitrate are both present thus stimulating sulfide-oxidizing nitrate-reducing bacteria (soNRB). soNRB re-oxidize sulfide back to sulfate or elemental sulfur while reducing nitrate to either 12 N_2 via autotrophic denitrification or NH_4^+ via dissimilatory nitrate reduction to ammonium 13 (Lam and Kuypers, 2011). If nitrate is limiting, sulfur is the more likely end product of sulfide 14 15 oxidation, which occurs in the following reaction stoichiometry for the denitrification pathway, $2NO_3^{-} + 5HS^{-} + 7H^{+} \rightarrow N_2 + 5S^{0} + 6H_2O$. A steady state is reached when the 16 diffusive fluxes (mmol $m^{-2} d^{-1}$) of nitrate and sulfide are in a 1:2.5 ratio. If the sulfide flux 17 exceeds the nitrate flux by more than a factor of 2.5, then sulfide will diffuse into the oxic 18 layer (Lam and Kuypers, 2011). Importantly, the activity of soNRB help to detoxify sulfide to 19 sulfur, preventing it from reaching overlying productive surface waters, hence most sulfidic 20 events likely go unnoticed (Lavik et al., 2009). However, with the increase in eutrophication 21 and the expansion of OMZs in both the Atlantic and Pacific (Stramma et al., 2008), sulfidic 22 events are expected to become more frequent., as already demonstrated for a time series 23 station in the Baltic Sea (Lennartz et al., 2014). 24

The first quantitative measurements and detailed profiles of a sulfidic event in the Peruvian 25 26 upwelling came from Schunck et al. (2013). During RV Meteor cruise M77/3 in January 2009 sulfidic waters covered $>5500 \text{ km}^2$ and contained approximately 2.2 x 10^4 tons of sulfide, 27 28 making it one of the largest plumes recorded. A total of 9 stations were taken along the coastal transect from Lima to Pisco which showed a ~80 m thick sulfide-rich layer extending 29 at times just below the oxycline. At this interface oxygen (< 1 μ mol kg⁻¹), nitrate (<1 μ mol kg⁻¹) 30 ¹) and nitrite (2 μ mol kg⁻¹) profiles overlapped with detectable sulfide concentrations. Stable 31 isotope rate measurements and targeted gene assays using quantitative PCR indicated that 32 various oxidants could have been used by the microbial community to oxidize sulfide at the 33 34 time of sampling. The most abundant sulfide oxidizers identified from the 16S rRNA

diversity belonged to the phylum proteobacteria within the subphylum gamma-, including the 1 2 SUP05/ARCTIC96BD-19-clade, Candidatus Ruthia magnifica, and Candidatus Vesicomyosocius okutanii, but also epsilon- such as Sulfurovum spp. Metagenomics 3 confirmed that all were capable of sulfide or sulfur oxidation, either with nitrate and oxygen 4 (facultative soNRB) or exclusively with oxygen. Indeed, both subphyla appear to be 5 ubiquitous in other seasonally oxic/anoxic waters and OMZs, (Canfield et al., 2010; Lavik et 6 al., 2009; Stevens and Ulloa, 2008; Stewart et al., 2011; Stewart et al., 2012; Swan et al., 7 2011; Walsh et al., 2009). Both gamma- and epsilon- proteobacteria members are known 8 chemolithoautotrophs, which assimilate carbon dioxide as the carbon source without the use 9 of sunlight. Subsurface C- assimilation rates were between 0.9 to 1.4 μ mol C L⁻¹ d⁻¹ during 10 this sulfidic event. In this study, "dark" primary production had contributed up to 25% of the 11 total CO_2 fixation in the Peruvian upwelling region at the time of sampling, which is 12 13 comparable to values observed in the Baltic and Black Seas (Schunck et al. (2013) and references therein). Paradoxically, some of these studies showed that measured rates of CO₂ 14 15 assimilation exceed rates possible by chemolithoautotrophic processes alone. Thus, while chemolithoautotrophic CO₂-fixation is considered a significant process, the specific activity 16 17 and main contributors of CO₂-fixation during sulfidic events (down to the genus-level) still remain unknown. 18

Different from our current knowledge of OMZ sulfur cycling is whether the production of 19 sulfide can originate as well from pelagic waters itself. Simultaneous reduction of different 20 electron acceptors (like NO_3^{-} , SO_4^{2-} and CO_2) can occur in defined niches where particle 21 aggregates have formed and are sinking through the water column (Wright et al., 2012). 22 These aggregates, more commonly known as marine snow, contain micro-scale redoxclines 23 under anoxic conditions (Alldredge and Cohen, 1987; Karl and Tilbrook, 1994; Woebken et 24 al., 2007). Moreover, aggregate communities appear to be distinct from bulk water collected 25 26 samples (Fuchsman et al., 2011). These communities were suggested to have active 27 manganese reduction, sulfate reduction and sulfide oxidation at the interior of the aggregates. 28 How much sulfide is generated in the water column during a sulfidic event is not well resolved. Nevertheless, in situ incubation experiments done in the Chilean upwelling have 29 shown the capacity for sulfate reduction in the offshore OMZ occurring under 30 thermodynamically unfavorable nitrate-rich conditions. In separate incubations measured 31 rates of potential sulfide oxidation were larger than rates of sulfate reduction indicating that 32 any produced sulfide is immediately re-oxidized (Canfield et al., 2010). The authors 33 34 intriguingly suggested an active but cryptic sulfur cycle linked to nitrogen cycling in the pelagic OMZ. From a biogeochemical perspective large-scale sulfate-reduction coupled to
 organic matter remineralization releasing inorganic nitrogen could represent a significant
 supply of ammonium for anammox bacteria.

4

5 5 The impact of extreme anoxia: Trace gases in the ETSP

The upper 1000 m of the ocean (incl. the euphotic zone) are the key regions where the 6 7 production of climate-relevant trace gases such as carbon dioxide (CO_2), nitrous oxide (N_2O), methane (CH₄) and dimethyl sulfide (DMS) occurs (see, e.g., Liss and Johnson (2014)). 8 While the pathways of CO₂ and DMS are dominated by phytoplankton in the oxic euphotic 9 zone, N₂O and CH₄ pathways are dominated by microbial processes at midwater depth (i.e. in 10 the OMZ). This is especially important since some OMZs are connected to coastal upwelling 11 regions where OMZ waters — enriched in both nutrients and trace gases such as CO₂, N₂O 12 13 and CH₄ — are brought to the surface fuelling phytoplankton blooms and releasing trace gases to the atmosphere (see, e.g., Capone and Hutchins (2013)). Thus, although they are usually 14 15 not in direct contact with the atmosphere, OMZs play an important role for oceanic emissions of climate-relevant trace gases (see e.g. Arévalo-Martinez et al. (2015)). 16

17

18 5.1 Nitrous oxide (N₂O) in OMZ

A comprehensive overview of both nitrous oxide (N₂O) distributions and pathways in OMZ
has been published in (Naqvi et al., 2010). Therefore, we concentrate here on recent findings
from the ETNA and ETSP.

N₂O production in the ocean is dominated by microbial nitrification and denitrification 22 processes. It is formed as a by-product during nitrification and as an intermediate during 23 denitrification. The paradigm that N₂O is exclusively produced by bacteria has been 24 challenged by the discovery of nitrifying (i.e. NH_4^+ oxidising) archaea (e.g., *Cand.* 25 26 Nitrosopelagicus brevis, see above) dominating N₂O production in the ETSP and ETNA (Löscher et al., 2012), which is supported by results of a culture study (Löscher et al., 2012) 27 28 and a marine microbial enrichment experiment (Santoro et al., 2011). The production of N_2O by archaea (and bacteria) depends on dissolved O₂ and is increasing with decreasing O₂ 29 concentrations (Frame and Casciotti, 2010; Löscher et al., 2012). Denitrifying bacteria do not 30 produce N₂O in the presence of O₂ (> 10 μ mol kg⁻¹); however, when O₂ concentrations are 31 approaching 0 µmol kg⁻¹, N₂O is consumed during denitrification. There is no N₂O production 32 under anoxic conditions. The significance of N₂O production during anammox (Kartal et al., 33 34 2007) and DNRA (Giblin et al., 2013) in OMZs (see Section 5) remains to be proven.

The detailed investigation of $\Delta N_2 O/AOU$ (= excess N₂O/apparent oxygen utilization) and 1 $\Delta N_2 O/\; \Delta^{15} NO_3^{-1}$ relationships from the ETNA and ETSP revealed two facts (Ryabenko et al., 2 2012): (i) The lower O_2 concentrations found in the core of the OMZ of the ETSP (< 5 µmol 3 kg^{-1}) favour N₂O consumption by denitrification which is not observed in the ETNA because 4 of its comparably high O₂ concentrations and (ii) the maximum observed N₂O concentrations 5 were higher in the ETSP than in the ETNA. This is in line with the results of two model 6 7 studies of N₂O in the ETSP by Zamora et al. (2012) and Cornejo and Farias (2012), which suggested that the switching point between N₂O production and N₂O consumption occurs at 8 higher O_2 concentration (~ 8-10 µmol kg⁻¹) than previously thought. 9

In contrast to the open ocean, OMZs in coastal (i.e. shelf) regions show a higher spatial and 10 temporal variability: Seasonally occurring suboxic or even anoxic/sulfidic OMZs have been 11 12 observed in coastal regions worldwide (see e.g. Diaz and Rosenberg (2008)). One of the most prominent areas where widespread sulfidic conditions have been recently observed is the shelf 13 off Peru (Schunck et al., 2013) (Section 5). Figure 6 shows the distribution of N₂O, water 14 temperature, nutrients and H₂S during the sulfidic event described by Schunck et al. (2013) on 15 16 the shelf off Peru during December 2008/January 2009. Here, extreme N₂O concentrations are found at the boundary to the H₂S containing bottom waters. No N₂O is found in the core 17 sulfidic layer. This suggests again that there is a narrow range of low O₂ concentrations which 18 is associated with exceptionally high N₂O production. As soon as the O₂ concentrations are 19 close to zero (anoxic/sulfidic conditions) N₂O production turns into N₂O consumption. 20 21 Similar N₂O distributions during anoxic/sulfidic events were found off the west coast of India, 22 in the Gotland Deep (central Baltic Sea) and in Saanich Inlet (Brettar and Rheinheimer, 1991; Cohen, 1978; Naqvi et al., 2000). Brettar and Rheinheimer (1991) suggested a close coupling 23 between H_2S oxidation and NO_3^- reduction in a narrow layer where NO_3^- and H_2S coexist. 24 This is in line with recent findings from the anoxic event off Peru by Schunck et al. (2013) 25 and similar to the suggestion of a cryptic sulfur cycle where sulfate reduction is coupled to 26 27 rapid H_2S oxidation by NO₃⁻ proposed for the OMZ off Chile by Canfield et al. (2010).

28

29 5.2 The role of OMZs in trace gas emissions

In OMZs with O_2 concentrations below 20 µmol kg⁻¹, N₂O production does not take place in the core of the OMZ. Instead, N₂O production is found at the oxycline. Exceptionally high N₂O concentrations have so far only been found in temporarily occurring anoxic/sulfidic regions off Peru/Chile and West India (Farías et al., 2015; Naqvi et al., 2010). Stagnant

sulfidic systems such as in the Baltic and Black Seas as well as the Cariaco Basin, have 1 2 shown only slightly enhanced N₂O concentrations at the oxic/anoxic interfaces (Bange et al., 2010, and references therein). This implies that significant pulses of N_2O emissions to the 3 atmosphere occur only when a shallow coastal system rapidly shifts from oxic to 4 anoxic/sulfidic conditions and vice versa (Bange et al., 2010). This can be explained by a lag 5 of N₂O reduction by denitrifiers, when they switch from oxygen to nitrogen respiration 6 7 (Codispoti, 2010) or N₂O production during the reestablishment of nitrification after O₂ 8 ventilation (Schweiger et al., 2007).

9 CH₄ production is also tightly connected to OMZs (see overview in Naqvi et al., 2010).
10 Similar to N₂O, upwelling areas are considerable hotspots for CH₄ emissions, albeit organic
11 material-enriched shallow coastal zones such as estuaries and mangroves or shallow
12 sediments with geological CH₄ sources show higher emissions (Bakker et al., 2014).

13 Since DMS is produced by phytoplankton in the euphotic zone, an accumulation of DMS in OMZs appears unlikely. However, measurements at the Candolim Time-Series Station 14 15 (CaTS) on the shelf off Goa (India) revealed an unprecedented 40-fold increase in DMS concentrations in the sulfidic layers during an anoxic event (Shenoy et al., 2012). These high 16 17 concentrations could not be explained by any known pathways and may imply an unknown most likely microbial — DMS production pathway under anoxic conditions either in the water 18 column or in the underlying sediments (Shenoy et al., 2012). Only recently it has been shown 19 that phytoplankton communities exposed to anoxic conditions increase their DMS production 20 significantly (Omori et al., 2015). This implies a potential accumulation of DMS at 21 22 oxic/anoxic boundaries of coastal OMZs which, in turn, might result in high DMS emissions from shallow coastal zones during anoxic/sulfidic events. 23

5.3 Trace gas production in OMZ and environmental changes

Trace gas production in OMZs is expected to be influenced primarily by deoxygenation 25 (Naqvi et al., 2010; Stramma et al., 2012). It is also well-known that eutrophication, warming 26 27 and supply of limiting nutrients (e.g. iron) will increase subsurface respiration of organic material, which leads to deoxygenation in open ocean and coastal OMZs (Bijma et al., 2013; 28 29 Gruber, 2011). Acidification of the upper ocean may result in a decrease of calcium carbonate (produced by calcifying organisms), which can act as ballast material for sinking organic 30 matter. Less ballast means a reduction in the sinking speed of organic particles, which could 31 increase the residence time of organic material and cause higher respiration rates (Riebesell et 32 33 al., 2009). Therefore, on-going environmental changes such as deoxygenation, eutrophication, warming and acidification have both direct and indirect effects on trace gas production in 34

OMZs. In general, we might expect enhanced production of N₂O, CH₄ and DMS in OMZs
 because of the on-going loss of O₂.

Deoxygenation in open ocean and coastal environments may lead, on the one hand, to 3 enhanced N_2O production when approaching the N_2O production/consumption switching 4 point (see above), but on the other hand, when O_2 concentrations fall below the switching 5 point this may lead to a consumption of N₂O (Zamora et al., 2012). Moreover, we do not 6 7 know whether the frequency of coastal anoxic events will continue to increase and how this 8 may affect the coastal net N₂O production/consumption. A recent modelling study on the 9 influence of anthropogenic nitrogen aerosol deposition and its effect on N₂O production anammhave revealed that the effect is small on a global scale but that the OMZ of the Arabian 10 11 Sea is especially sensitive to atmospheric nitrogen deposition resulting in an enhanced N₂O production (Suntharalingam et al., 2012). 12

13

14 6 Conclusions

15 While there is a growing amount of data on primary production and the pelagic N cycle in and associated to OMZ waters, quantitative estimates of microbial production and respiration, 16 17 particularly at ultra-low O_2 levels, are still not fully explored. This translates into an 18 uncertainty concerning the origin of the N deficit. While it had been clearly demonstrated that 19 N loss processes respond sensitively to minimal changes in O_2 (Dalsgaard et al., 2014), a potential uncertainty may result from additional processes, such as preferential P release 20 21 directly in the water column may be important to create the N deficit in the water column. In this context, the character and size were shown important (DeVries et al., 2014), therefore 22 linking P release from particles to the character of N loss (Babbin et al., 2014). The character 23 of the N loss/ N deficit term is, however, highly important, as it determines the extent of N 24 depletion of the entire OMZ due to the above explained stoichiometric discrepancy between N 25 26 loss and N₂ fixation. Interestingly, a strong impact of decadal climate variations on respiration rates, primary production and the intensity of N loss has been described for the South Pacific 27 (Deutsch et al., 2011). This may directly link to the character of N loss derived from our and 28 other measurements and has to be taken into consideration for future studies. 29 A coupling via the proposed primary production chain may indeed act in OMZs associated to 30

31 upwelling systems (an overview of major processes in the ETNA and ETSP is depicted in Fig.

32 7), the important term of organic matter export, either horizontally or vertically need to be

33 more dedicated investigations: To date, a quantification of DOM supply via (sub-) mesoscale

vertical transport processes out of the OMZ area and diapycnal mixing sustaining microbial 1 activity in the Peruvian OMZ is missing. Further, for POM supply to the OMZ, DVM seems 2 to play a key role, despite some quantitative uncertainties. Although some organisms 3 performing DVM have certain strategies to cope with anoxic conditions, mostly by down-4 5 regulating the aerobic metabolism, there are limits for zooplankton and nekton. Thus, a reduction of OM export by DVM may result with a further expansion and deoxygenation of 6 7 OMZs. Deutsch et al (2014) describe in this context that a decrease of the habitat caused by global warming and ocean deoxygenation increases competition among species and may even 8 9 result in a loss of metabolic functionality by 20%. 10 A quantification of DOM and POM import and export rates to and from the ETNA is currently not available, an extensive discussion of POC dynamics from the ETSP OMZs is 11 12 provided in this issue (Dale et al., 2015). Information on the character of microbial processes responsible for POM degradation within 13 the OMZ is however missing. Here, first studies (e.g. Ganesh et al., 2014, 2015) indicate a 14 key role of particulate organic matter acting as microniches for microbes and thus host certain 15 processes such as microaerobic respiration in OMZ waters. By containing strong redox 16 17 gradients in relatively narrow vicinity, and by providing nutrients and trace metals, particles 18 might strongly influence biogeochemical cycles. It is well-known that in the core of OMZs 19 adjacent to coastal upwelling regions, such as those found off Peru, Mauritania and the 20 Arabian Sea, a pronounced POM/particle-enriched turbid layer (a so-called intermediate nepheloid layer) exists (see, e.g., Stramma et al. (2013); Naqvi et al. (1993); Fischer et al. 21 (2009)).22 With regard to sulfidic events, which make the lower limit of anoxia, the positive feedback 23 24 coupling could be thought to stabilize itself: While a direct toxic effect of H2S on primary 25 production is mostly mitigated by the respective detoxifying community, decoupling of the 26 supply of benthic nutrients to the sea surface might decrease primary production. This may be of particular importance in areas, such as the ETSP where the benthic supply of e.g. trace 27 metals is dominant. Resulting decreased respiration in the OMZ could be thought to 28 29 subsequently lead to regeneration to non-sulphidic, less pronounced anoxia, which may stabilize the OMZ to a certain extent. Whether this hypothesis is valid has, however, to be 30 resolved. 31 32 A critical consequence of ocean deoxygenation is visible from the comparison of the ETSP and ETNA regions: Massive supersaturation of N₂O, connected to sulfidic plumes, has been 33

detected repeatedly, in the ETSP. OMZs are important sites of enhanced production of climate 1 relevant trace gases such as N_2O , CH_4 , and DMS. N_2O production is significantly enhanced at 2 oxic/anoxic boundaries of OMZs and we suggest that it mainly results from habitat 3 4 compression, where in extreme cases (such as sulfidic events, sharpening gradients) 5 nitrification and denitrification can occur simultaneously. Maximum N₂O concentrations and subsequent emissions to the atmosphere have been observed in dynamic coastal systems that 6 7 rapidly shift from oxic to anoxic conditions and vice versa. Although OMZs are usually not in direct contact with the atmosphere, their vicinity to coastal upwelling systems plays an 8 9 important role for oceanic emissions of climate-relevant trace gases such as N_2O , CH₄, and 10 DMS with potential feedbacks on global warming, which then may again impact on ocean 11 deoxygenation. Our studies from the ETSP (Arevalo-Martinez, 2015, 2016, Kock et al., 2016) confirm intense production of N_2O in the coastal upwelling. This is in line with an increase of 12 13 N2O production from OMZ areas as concluded from forced climate models. However, the same model describes a global decrease of N_2O formation by 4-12%, mostly linked to the 14 15 western basins of the Pacific and Atlantic oceans (Martinez-Rev et al., 2015). 16 Marine ecosystems and biogeochemical cycles are increasingly impacted by a growing 17 number of stress factors, some of which act locally, such as eutrophication and pollution,

18 others globally. Global stressors are associated with anthropogenic carbon dioxide (CO₂)

emissions and affect the ocean either directly through CO₂-induced acidification or indirectly

20 through climate change-induced ocean warming and deoxygenation (Ciais et al., 2013). How

21 these stressors will impact marine ecosystems and biogeochemistry, individually or in

22 combination, is still largely unknown.

23 Ocean warming, acidification and deoxygenation occur globally and simultaneously,

24 although with distinct regional differences. Through increased stratification and decreased

25 nutrient supply to the surface layer, ocean warming is expected to decrease the biological

26 production in the already stratified low to mid latitudes.

27 While research on ocean warming is relatively advanced, far less is known about the impacts

28 of ocean acidification and deoxygenation on marine organisms and ecosystems. Because the

29 three stressors have mostly been studied in isolation, knowledge on the combined effects of

30 two or more of them is scarce. In principle, additive, synergistic (more than additive) and

antagonistic (less than additive, i.e. compensatory) interactions of effects are possible, but *a*

32 *priori* it is impossible to judge what the combined effects will be. One example for a

33 synergistic effect is that of ocean acidification narrowing the thermal tolerance window of

34 some organisms, amplifying the impact of warming (Pörtner and Farrell, 2008). However,

1 we consider interactions among stressors in marine communities largely understudied.

2 7 Outlook

Major issues remaining unresolved, in addition to those highlighted above, concerning (1) a mechanistic understanding of organic matter degradation and nutrient cycling at low or variable oxygen concentrations in the water column and the role of DVM for organic matter supply to the OMZ, (2) the sensitivities of heterotrophic microbes and their sensitivity to low oxygen conditions, and (3) biogeochemical feedback processes in oxygen minimum zones and their impacts on local to global scales.

9 Future studies in the framework of the SFB 754 will therefore combine measurements of 10 particle flux, zooplankton abundance, microbial activities and O_2 concentrations in order to 11 answer the following key questions:

- I. What is the effect of low oxygen conditions (below 20 μmol kg⁻¹) on organic matter
 degradation? And what is the partitioning between DOM and POM in OMZ waters?
- II. How do the rates of nutrient cycling and loss in OMZs relate to particles and associated microniches?
- III. What are the rates of oxygen supply and consumption in the upper OMZ? And whatis regulating respiration rates?
- IV. Do small-scale processes (e.g. viral lysis) affect fluxes on larger scales? And how can
 models represent these important processes?

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1 Figures



2

Figure 1. Global distribution of O_2 at σ_{Θ} = 26.4 kg m⁻³ (~ 400 m depth): The major regions of low oxygen in the world ocean are all located in the tropical oceans, at shallow to intermediate depths. The area off Peru represents one of the most pronounced OMZs. The investigated areas in the eastern tropical South Pacific and the eastern tropical North Atlantic Oceans are marked with black boxes; examples of the O₂ distribution are given along two sections from the coast to the open ocean at 10°S in the OMZ off Peru and at 18°N in the eastern tropical North Atlantic; O₂ concentrations are indicated by the color code.

10





Figure 2. Co-occurrence of anammox as determined by rate measurements and the key functional marker gene for N2 fixation, nifH, in the ETSP OMZ (modified from Kalvelage et al., 2013 and Löscher et al., 2014).



Figure 3. The marine nitrogen (N) cycle with the major onshore and offshore processes in the ETSP OMZ, modified from Kalvelage et al. (2013). Numbers indicate fluxes of N [Tg y⁻¹].



1

Figure 4. Redundancy analysis ordination model of microbial taxa (vectors) identified from 2 pyrosequencing reads of multiple samples (points) in the ETSP. Spherical k-means clustering 3 revealed a fivefold partitioning that reflects distinct OMZ habitats (see legend). Each point is 4 5 colored according to the cluster that dominated the microbial population in the respective 6 sample.









minimum zone, e.g. in the ETSP. The sulfide and nitrate fluxes are shown in steady state. 11

- 1 Sulfate-reducing bacteria produce sulfide from the sediment while the complementary
- 2 detoxification process occurs in the water column at overlapping profiles.



3

Figure 6. Distributions of N_2O , NH_4^- , NO_2^- , NO_3^- , H_2S , and water temperature during December 2008/January 2009 (R/V Meteor cruise M77/3) on the shelf along the coast of Peru. Max. N_2O concentrations have been detected right above the sulfidic zone, where a sharp oxycline is present and ammonium and nitrate are available.

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- 1 Figure 7. Scheme of the (A) ETNA and (B) ETSP OMZs with major processes identified. The
- O_2 background is taken from the SOPRAN cruise P399, along 18°N in the ETNA and from
- 3 the SFB754 cruise M77/3, along 10° S in the ETSP.

- Table 1: A comparison of the O₂ minimum, excess nitrogen (N*), primary production, organic 1
- 2 C export, N₂ fixation and N loss in the ETNA and ETSP upwelling regions.
- 3

	ETSP-shelf	ETSP- offshore	ETNA-shelf	ETNA- offshore
O ₂ min. [µmol kg ⁻¹]	0 (sulfidic)	0	25	40
N* [mol m ⁻²]	-1,9- (-5,98) ^a			
Primary Production [mmol C m ⁻² d ⁻¹]	101-122 ^b 68,5 ^c 61,4 ^d	73-94 ^b	137 ^c 167 ^d	
Organic C export [mmol C m ⁻² d ⁻¹]	10,6-75,3 ^b	2,6-11,1 ^b	6,4- 9,3 ^e	0,67-2,6 ^e
N_2 fixation [µmol N m ⁻² d ⁻¹]	25-657 ^f		24-140 ^g	
% contribution of N ₂ fixation to primary production*	0,2-4 ^b 0,2-6,4 ^c 0,3-7,1 ^d		0,2-0,7 ^c 0,1-0,6 ^d	
N loss [mmol N m ⁻² d ⁻¹]				
anammox denitrification	1-10 up to 70 (in presence of H ₂ S)	0 0	000	

* Calculated based on the Redfield ratio of C:N = 106:16 using the primary production and N_2 fixation rates

given in this table in consistency with the % contribution given in Duce et al. (2008). ^a Kalvelage et al., 2013, ^b Dale et al., 2015, ^c Behrenfeld and Falkowski, 1997, ^d Longhurst et al., 1995, ^e Iversen et al., 2010, ^f Dekaezemacker et al., 2013 and Löscher et al., 2014, ^g Voss et al., 2002.

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