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**Title:** Stable isotopes in barnacles as a tool to understand green sea turtle (*Chelonia mydas*) regional movement patterns

**Running page head:** Isotopes in sea turtle barnacles

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21 ABSTRACT: Sea turtles are migratory animals that travel long distances between their feeding and  
22 breeding grounds. Traditional methods for researching sea turtle migratory behavior have  
23 important disadvantages, and the development of alternatives would enhance our ability to  
24 monitor and manage these globally endangered species. Here we report on the isotope signatures  
25 in green sea turtle (*Chelonia mydas*) barnacles (*Platylepas* sp.) and discuss their potential relevance  
26 as tools with which to study green sea turtle migration and habitat use patterns. We analyzed  
27 oxygen ( $\delta^{18}\text{O}$ ) and carbon ( $\delta^{13}\text{C}$ ) isotope ratios in barnacle calcite layers from specimens collected  
28 from green turtles captured at the Palmyra Atoll National Wildlife Refuge (PANWR) in the Central  
29 Pacific. Carbon isotopes were not informative in this study. However, the oxygen isotope results  
30 suggest likely regional movement patterns when mapped onto a predictive oxygen isotope map of  
31 the Pacific. Barnacle proxies could therefore complement other methods in understanding regional  
32 movement patterns, informing more effective conservation policy that takes into account  
33 connectivity between populations.

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36 KEY WORDS: *Chelonia mydas*, *Platylepas* barnacle, epibiont, proxy, oxygen isotope, carbon isotope

## 1. INTRODUCTION

Long distance migratory behavior between breeding and feeding grounds, a key component of sea turtle ecology, creates important research and conservation challenges (Godley et al., 2010). Understanding migratory and habitat use patterns is a critical step in the design of comprehensive conservation and management strategies aimed at protecting all of a species' range, including the corridors connecting distant habitats. For many sea turtle populations we lack detailed spatiotemporal knowledge about migration patterns, as well as fine-scale understanding of habitat use. This dearth of information may hinder conservation efforts, especially in scarcely studied areas such as the Central Pacific (Wallace et al., 2010).

Previous studies on sea turtle movement patterns have been based on mark-recapture, satellite telemetry, or genetic analysis (Godley et al., 2010). Although these methods have provided key insights, they also have important shortcomings. Mark-recapture can have very low return rates (Oosthuizen et al., 2010). Satellite telemetry is a very effective method for tracking turtles across long distances but can be prohibitively expensive, and loss and malfunction of transmitters is common (Hays et al., 2007; Hebblewhite and Haydon, 2010). Genetic studies can be a very effective way of delineating population structure and natal origin, but are uninformative about movements after the sea turtles hatch (Bowen and Karl, 2007). Therefore, additional methods are needed to help us map patterns of movement and habitat use at scales useful for conservation planning (Godley et al., 2010).

Because of their intimate connections, species that are associates of particular hosts have been used as proxies for the study of host ecology, demography, and evolutionary history (Nieberding and Olivieri, 2007). Recent research has shown that studying associate species such as parasites and commensals, can be a cost-effective alternative to ecological research on the host themselves (Byers et al., 2011; Hechinger et al., 2007). Several barnacle species are commonly found in sea turtles,

attached to the skin and shell. Barnacles are found in the majority of green turtles observed in a long-term study of marine turtles at Palmyra Atoll National Wildlife Refuge (Gómez, A., personal observation), and they have been reported widely from sea turtle populations from across the world (Casale et al., 2004; Frick et al., 2010; Rawson et al., 2003; Schwartz, 1960; Torres-Pratts et al., 2009; Zardus and Balazs, 2007). As obligate commensals, these barnacles form close, long-lasting associations with their hosts, and may thus provide useful information about turtle ecology.

Previous studies have shown that isotopes in barnacle calcite can be used to reconstruct migratory patterns and habitat use in California gray whales (Killingley, 1980) and loggerhead turtles (Killingley and Lutcavage, 1983). Isotope ratios in calcite layers can be used to approximate the water temperature throughout the life of individual barnacles because warmer waters have reduced oxygen ratios (Killingley and Lutcavage, 1983), where the oxygen isotopes in the barnacle calcite fractionate or change in relative proportion during calcite formation depending on the oxygen ratios in the surrounding water (Kendall and Caldwell, 1998). Therefore, oxygen isotope ratios obtained from barnacles can be informative about turtle movements at large scales, as long as those movements occurred along water temperature gradients (Killingley and Lutcavage, 1983). These movements can be traced by comparing barnacle oxygen isotope ratios to mapped prediction for these values. Temporal reconstruction could potentially also be added as our understanding of the pace at which successive barnacle calcite layers are laid down improves. Carbon isotope ratios can be expected to vary as microhabitats differ in the concentration of dissolved carbon, and can therefore provide information about habitat occupancy across sites, with lagoons and the pelagic zone assumed to have low and high carbon conditions respectively (Killingley and Lutcavage, 1983). Here we report on oxygen ( $\delta^{18}\text{O}$ ) and carbon ( $\delta^{13}\text{C}$ ) isotopes in the barnacle *Platylepas hexastylus*, an epibiont of turtles, collected from green sea turtles (*Chelonia mydas*) at Palmyra Atoll National Wildlife Refuge in the Central Pacific and discuss the potential of this method as a tool with which to study sea turtle movements.

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87 **2. MATERIALS AND METHODS**

88 The barnacle specimens used in the experiment were collected at Palmyra Atoll National Wildlife  
89 Refuge (PANWR; 05 ° 52' N, 162 ° 05' W), central Pacific Ocean. The atoll has a wide shallow reef,  
90 extensive reef terraces at both the eastern and western ends, and three lagoons (Collen et al., 2009).  
91 The islets and 12 nautical miles of the surrounding ocean have been designated a marine protected  
92 area by the U.S. Fish and Wildlife Service since 2001. In 2005, the Center for Biodiversity and  
93 Conservation of the American Museum of Natural History initiated a research and conservation  
94 program for sea turtles at PANWR. The program includes research into the turtles' distribution and  
95 abundance, connectivity, feeding ecology, health, and threats (McFadden et al., 2014; Sterling et al.,  
96 2013). The sea turtle population at this site has been studied using mark-recapture, satellite  
97 telemetry, and genetic analysis (Sterling et al., 2013).

98 *Platylepas* sp. barnacles were collected from adult green sea turtles caught in PANWR during the  
99 summer of 2011. These barnacles were found embedded in the turtles' soft tissue (A. Gómez,  
100 personal observation). Barnacles were removed from the turtle's skin and stored in vials with 90%  
101 ethanol until analysis. We analyzed a total of 12 barnacles. In order to assess the consistency of  
102 recorded isotope ratios of different barnacles from a given turtle we sampled three barnacles per  
103 turtle. The barnacles were dissected and milled along their axis of growth using a Merchantek  
104 MicroMill (Electro Scientific Industries, Inc., Portland, United States) to take calcite samples. The  
105 mill was programmed to make passes on the outer facing surface of the paries perpendicular to the  
106 axis of growth in distances 0.3-0.4 mm apart. For each sample, a record was kept of the distance  
107 along the growth axis from barnacles' base to where each pass had been made. Samples were taken  
108 from the outermost part of the paries to exclude any calcite deposits that might have been the  
109 result of ageing and thickening of the individual plates. It should be noted that nothing is known

about growth rates in this species of barnacle. The calcite samples were sent to the Keck Paleoenvironmental & Environmental Stable Isotope Laboratory at the University of Kansas, where they were analyzed for oxygen ( $\delta^{18}\text{O}$ ) and carbon ( $\delta^{13}\text{C}$ ) stable isotope ratios. A Kiel Carbonate Device III and a Finnigan MAT253 isotope ratio mass spectrometer (Finnigan MAT, Bremen, Germany) were used to perform the laboratory analyses.

Oxygen isotope ratios in barnacle calcite can be expected to vary predictively as a function of the water's oxygen isotope ratios and temperature and can be solved for using a conversion formula (Epstein et al., 1953) with a required modification for barnacle calcite (Killingley and Newman, 1982). We reversed the formula by rearranging variables for the water's oxygen isotope ratio, which accounts for variations in salinity, and temperature to create a map of predicted barnacle oxygen isotope ratios. We used annual average sea surface temperature data from NOAA's World Ocean Database (NOAA, 2005) for the temperature variable in the equation, and published water oxygen isotope figures from 2006 (LeGrande and Schmidt, 2006) as inputs in the equation. The resulting map allowed us to put the oxygen isotope results from the barnacles into geographic context. We used this map to create an isoscape, thereby defining the largest possible area from which the isotope values measured from the calcite could have accumulated across the life of the barnacles sampled. A detailed methodology is included as an electronic supplement.

### 3. RESULTS

Because some of the calcite samples were not sufficiently large to be analyzed with precision in the mass spectrometer, we obtained a complete set of results for barnacles from two of the four sea turtles sampled and only partial results from one other. We included nine barnacles from three turtles in our analysis, as results from the fourth were too incomplete. The selected barnacles on the respective turtles had the following sizes measured from the base to the aperture: (i) 1.6 mm,

1.3 mm and 1.6 mm on GD42, (ii) 1.6 mm, 2.2 mm, and 2.5 mm on GI41, and (iii) 2.0 mm, 2.1 mm and 1.6 mm on GI43. A summary of the stable isotope ratios are reported in Table 1. The youngest part of the barnacle is that closest to the basal margin or bottom, as the barnacle grows outward. These isotope ratios represent the values across the growth axis of the barnacle shell going from the youngest to the oldest part of the barnacle. The carbon and oxygen isotope ratios are reported versus the Vienna Pee Dee Belemnite (VPDB) scale (Coplen, 1995), which is used as benchmark value. The maps predicting calcite oxygen isotope ratios in the Central Pacific showed uniform ratios along the equator and steep gradients towards northern and southern latitudes.

Oxygen isotope ratios in our calcite samples did not show major fluctuations throughout the life of the barnacle, while the carbon isotope ratios of the barnacles spanned three orders of magnitude. The highest measured oxygen isotope ratio in the collected barnacles was  $-0.951 \delta^{18}\text{O}$ . We used this value as a contour to create an envelope in which we would expect our sea turtles to have stayed throughout the lifetime of the barnacle (Fig. 1). The resulting isoscape included PANWR. We also created a more conservative isoscape that corrected for the fact that the original isoscape maps might be overestimating the isotope ratios. The first step was to identify the expected oxygen isotope ratio at PANWR on the map, as the isotopes in the barnacles' youngest layer would be expected to coincide with it. The map predicted a calcite oxygen isotope ratio of  $-1.075 \delta^{18}\text{O}$ , while the average youngest layers of the barnacles collected were  $-1.337 \delta^{18}\text{O}$ , giving a difference of  $0.262 \delta^{18}\text{O}$ . Adding this difference to the original isoscape value of  $-0.951 \delta^{18}\text{O}$  gave a corrected calcite oxygen ratio of  $-0.688 \delta^{18}\text{O}$ . This ratio was then used to produce a larger standardized isoscape delineating the sea turtles movements during the barnacles' lifetime (Fig. 2).

#### 4. DISCUSSION

Our study found that oxygen isotopes in barnacles' calcite could be used to broadly delineate the area in which the sampled sea turtles moved during the life of the barnacles, allowing us to exclude visitation of major breeding grounds in the Pacific. Carbon isotopes were not informative in this study and assessing their utility as proxies with which to explore sea turtle habitat use requires further study. Oxygen isotope values observed in the barnacles in this study indicated that the calcite ratios conform to sea temperatures of 28°C and 30°C. Assuming that average temperatures above 28°C are found in the warmest waters of the Central Pacific that are in proximity to the equator, then our data suggest that turtles did not venture beyond these waters during the lifespan of the barnacles collected. This is consistent with observations from the field, which suggest that turtles spend extended periods of time in PANWR (Sterling et al., 2013).

To obtain a more concrete picture of the sea turtles' movements, we used the predicted calcite oxygen isotope map estimating the area within which the sea turtles may have moved. The contour delineating the isoscape of possible movements was large (Fig. 1 and 2) as water temperatures in the Central Pacific are relatively uniform. However, some major known green turtle grounds that are within in the potential migratory range of green turtles from PANWR were not within this isoscape (STC, 2012). These include Ogasawara Island (Japan), NW Australia and Hawaii, which also remain outside of the boundary when using the more conservative adjusted oxygen isoscape. Importantly, recent research shows that the natal origin of sea turtles in PANWR can almost exclusively be found to the West and South of the Central Pacific (Naro-Maciel et al., 2014). Therefore, the boundaries we delineate in this study: 1) include PANWR, 2) are consistent with ecological observation, and 3) are consistent with new genetic evidence about the population structure of green sea turtles at PANWR.



181 Because we cannot exclude the possibility that our isoscapes simply reflect residency at Palmyra,  
182 we are unable to quantify the method's utility as an indicator of large scale movements. However,  
183 our data suggest that it can be used to delineate envelopes of likely residency across the Pacific  
184 basin. Therefore we suggest that this method has the potential to provide valuable data to inform  
185 comprehensive management strategies, by helping identify specific ecological and political areas  
186 within or outside a given population's range.

187 A wide range in the barnacles carbon isotopes may indicate that turtles made use of a variety of  
188 microhabitats around the atoll, possibly moving between areas like the lagoon and the pelagic zone,  
189 which are assumed to have low and high carbon conditions respectively (Killingley and Lutcavage,  
190 1983). An alternative explanation is that the turtles are frequenting ecologically heterogeneous  
191 areas beyond PANWR. However, any conclusions drawn from these results need to be viewed  
192 conservatively, as a heterogeneous environment does not necessarily explain the lack of  
193 consistency in our data, which have marked dissimilarities in carbon isotope ratios between  
194 barnacles on the same turtle. There could be differences in uptake or expression of carbon isotopes  
195 in each barnacle possibly limiting the use of the carbon isotope data in this study system. Previous  
196 studies used a larger barnacle species than the ones found on the green turtles at PANWR  
197 (Killingley and Lutcavage, 1983). *Platylepas* sp. specimens that we collected had sizes ranging  
198 between 1.3 and 2.5 mm, which is a magnitude smaller than the *Chelonibia testudinaria* recovered  
199 from loggerhead turtles in previous studies (Killingley, 1980; Killingley and Lutcavage, 1983). This  
200 resulted in fewer data points and limited statistical analysis of the results.

201 In summary, this limited dataset suggests that inferences about green sea turtle spatial ecology  
202 obtained from isotope analysis are broadly consistent with field observations and genetic analyses.  
203 Isotope analysis may provide low-resolution information about sea turtle connectivity, potentially  
204 defining areas of interest for research and management. Therefore, we suggest that this method can

only complement but not replace other tools to investigate turtle migration and habitat use patterns. One advantage of the method is its low cost. The total cost of analyzing three barnacles on one sea turtle was below 170 USD (56 USD per barnacle in 2011). This makes using barnacle proxies an option that could be explored further in the study of spatial ecology and could be improved in future applications.

Future research can add critical information with which to improve this method. We lack basic information about the natural history of many turtle epibionts. Because of the dearth of data on baseline growth rates for *Platylepas* sp., the time span between successive calcite layers is unknown, and therefore the system cannot be attached to an absolute temporal scale. We also lack benchmarks for isotope ratios in barnacles. Therefore, it is difficult to draw conclusions about the significance of fluctuations that we observed, especially for the variation in carbon isotope ratios. The utility of barnacles as proxies of sea turtle movement at study sites such as PANWR might not be fully realized until these key knowledge gaps are addressed.

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## References

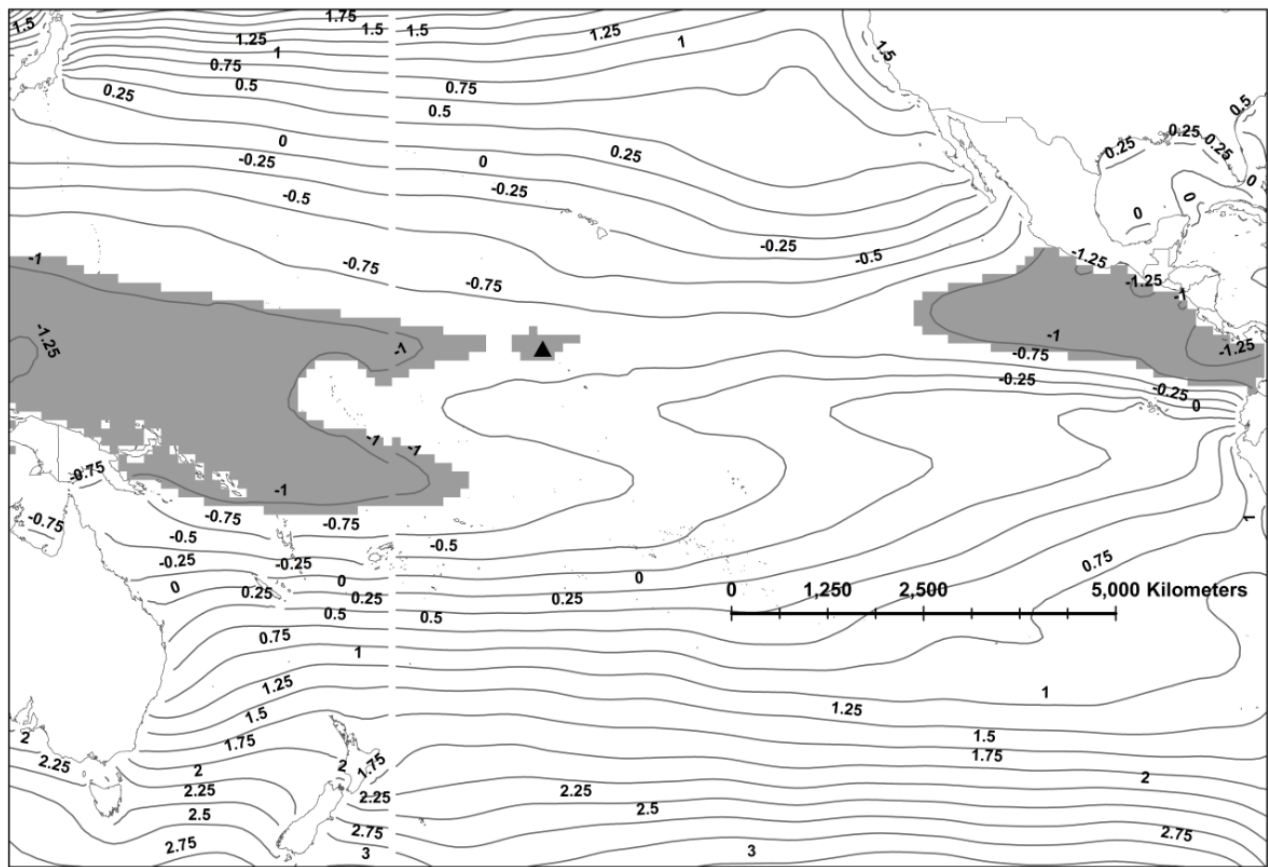
- Bowen, B. W. and Karl, S. A.: Population genetics and phylogeography of sea turtles, *Molecular Ecology*, 16, 4886-4907, 2007.
- Byers, J. E., Altman, I., Grosse, A. M., Huspeni, T. C., and Maerz, J. C.: Using parasitic trematode larvae to quantify an elusive vertebrate host, *Conservation Biology*, 25, 85-93, 2011.
- Casale, P., Freggi, D., Basso, R., and Argano, R.: Epibiotic barnacles and crabs as indicators of *Caretta caretta* distribution and movements in the Mediterranean Sea, *Journal of the Marine Biological Association of the United Kingdom*, 84, 1005-1006, 2004.
- Collen, J. D., Garton, D. W., and Gardner, J. P. A.: Shoreline changes and sediment redistribution at Palmyra Atoll (Equatorial Pacific Ocean): 1874–Present, *Journal of Coastal Research*, doi: 10.2112/08-1007.1, 2009. 711-722, 2009.
- Coplen, T.: Reporting of stable hydrogen, carbon, and oxygen isotopic abundances, *Geothermics*, 24, 707-712, 1995.
- Epstein, S., Buchsbaum, R., Lowenstam, H. A., and Urey, H. C.: Revised carbonate-water isotopic temperature scale, *Geological Society of America Bulletin*, 64, 1315-1326, 1953.
- Frick, M. G., Zardus, J. D., and Lazo-Wasem, E. A.: A new *Stomatolepas* barnacle species (Cirripedia: Balanomorpha: Coronuloidea) from leatherback sea turtles, *Bulletin of the Peabody Museum of Natural History*, 51, 123-136, 2010.
- Godley, B. J., Barbosa, C., Bruford, M., Broderick, A. C., Catry, P., Coyne, M. S., Formia, A., Hays, G. C., and Witt, M. J.: Unravelling migratory connectivity in marine turtles using multiple methods, *J. Appl. Ecol.*, 47, 769-778, 2010.
- Hays, G. C., Bradshaw, C. J. A., James, M. C., Lovell, P., and Sims, D. W.: Why do Argos satellite tags deployed on marine animals stop transmitting?, *Journal of Experimental Marine Biology and Ecology*, 349, 52-60, 2007.
- Hebblewhite, M. and Haydon, D. T.: Distinguishing technology from biology: a critical review of the use of GPS telemetry data in ecology, *Philosophical Transactions of the Royal Society B: Biological Sciences*, 365, 2303-2312, 2010.
- Hechinger, R. F., Lafferty, K. D., Huspeni, T. C., Brooks, A. J., and Kuris, A. M.: Can parasites be indicators of free-living diversity? Relationships between species richness and the abundance of larval trematodes and of local benthos and fishes, *Oecologia*, 151, 82-92, 2007.
- Kendall, C. and Caldwell, E. A.: Fundamentals of isotope geochemistry, *Isotope tracers in catchment hydrology*, 1998. 51-86, 1998.
- Killingley, J. and Newman, W.:  $^{18}\text{O}$  fractionation in barnacle calcite: a barnacle paleotemperature equation, *Journal of Marine Research*, 40, 893-902, 1982.
- Killingley, J. S.: Migrations of California gray whales tracked by Oxygen-18 variations in their epizoic barnacles, *Science*, 207, 759-760, 1980.
- Killingley, J. S. and Lutcavage, M.: Loggerhead turtle movements reconstructed from  $^{18}\text{O}$  and  $^{13}\text{C}$  profiles from commensal barnacle shells, *Estuarine, Coastal and Shelf Science*, 16, 345-349, 1983.
- LeGrande, A. N. and Schmidt, G. A.: Global gridded data set of the oxygen isotopic composition in seawater, *Geophysical Research Letters*, 33, L12604, 2006.
- McFadden, K. W., Gómez, A., Sterling, E. J., and Naro-Maciel, E.: Potential impacts of historical disturbance on green turtle health in the unique & protected marine ecosystem of Palmyra Atoll (Central Pacific), *Marine pollution bulletin*, 89, 160-167, 2014.
- Naro-Maciel, E., Gaughran, S. J., Putman, N. F., Amato, G., Arengo, F., Dutton, P. H., McFadden, K. W., Vintinner, E. C., and Sterling, E. J.: Predicting connectivity of green turtles at Palmyra Atoll, central Pacific: a focus on mtDNA and dispersal modelling, *Journal of The Royal Society Interface*, 11, 20130888, 2014.

- Nieberding, C. M. and Olivieri, I.: Parasites: proxies for host genealogy and ecology?, *Trends in Ecology & Evolution*, 22, 156-165, 2007.
- NOAA: <http://www.nodc.noaa.gov/OC5/indprod.html> last access: 03/04 2012.
- Oosthuizen, W. C., De Bruyn, P., Bester, M. N., and Girondot, M.: Cohort and tag-site-specific tag-loss rates in mark-recapture studies: A southern elephant seal cautionary case, *Marine Mammal Science*, 26, 350-369, 2010.
- Rawson, P. D., Macnamee, R., Frick, M. G., and Williams, K. L.: Phylogeography of the coronulid barnacle, *Chelonibia testudinaria*, from loggerhead sea turtles, *Caretta caretta*, *Molecular Ecology*, 12, 2697, 2003.
- Schwartz, F.: The barnacle *Platylepas hexastylus* encrusting a green turtle, *Chelonia mydas mydas*, from Chincoteague Bay, Maryland, *Chesapeake Science*, 1, 116-117, 1960.
- STC: <http://www.conserveturtles.org/seaturtlenestingmap.php>, last access: 05/15 2012.
- Sterling, E. J., McFadden, K., Holmes, K., Vintinner, E., Arengo, F., and Naro-Maciel, E.: Ecology and conservation of marine turtles in a foraging ground in the Central Pacific, *Chelonian Conservation Biology*, 12, 2-16, 2013.
- Torres-Pratts, H., Scharer, M. T., and Schizas, N. V.: Genetic diversity of *Chelonibia caretta*, commensal barnacles of the endangered hawksbill sea turtle *Eretmochelys imbricata* from the Caribbean (Puerto Rico), *Journal of the Marine Biological Association of the United Kingdom*, 89, 719-725, 2009.
- Wallace, B. P., DiMatteo, A. D., Hurley, B. J., Finkbeiner, E. M., Bolten, A. B., Chaloupka, M. Y., Hutchinson, B. J., Abreu-Grobois, F. A., Amorcho, D., Bjorndal, K. A., Bourjea, J., Bowen, B. W., Duenas, R. B., Casale, P., Choudhury, B. C., Costa, A., Dutton, P. H., Fallabrino, A., Girard, A., Girondot, M., Godfrey, M. H., Hamann, M., Lopez-Mendilaharsu, M., Marcovaldi, M. A., Mortimer, J. A., Musick, J. A., Nel, R., Pilcher, N. J., Seminoff, J. A., Troeng, S., Witherington, B., and Mast, R. B.: Regional Management Units for Marine Turtles: A novel framework for prioritizing conservation and research across multiple scales, *PLoS One*, 5, 2010.
- Zardus, J. D. and Balazs, G. H.: Two previously unreported barnacles commensal with the green sea turtle, *Chelonia mydas* (Linnaeus, 1758), in Hawaii and a comparison of their attachment modes, *Crustaceana*, 80, 1303-1315, 2007.

Table 1. Distance from paries' base, oxygen isotope ratio and carbon isotope ratio in *Platylepas* sp. barnacles collected from three green sea turtles (GD42, GI41, and GI43) in Palmyra Atoll National Wildlife Refuge. Rows show the average of three barnacles per turtle sampled. Distance is given in millimeters and isotope ratios are reported versus the VPDB scale.

Distance from Base			$\delta^{18}\text{O}$ Concentration			$\delta^{13}\text{C}$ Concentration		
<i>GD42</i>	<i>GI41</i>	<i>GI43</i>	<i>GD42</i>	<i>GI41</i>	<i>GI43</i>	<i>GD42</i>	<i>GI41</i>	<i>GI43</i>
0.350	0.350	0.350	-1.359	-1.343	-1.310	0.729	-0.451	-0.299
0.719	0.727	0.743	-1.283	-1.220	-1.431	0.798	-0.398	-0.619
1.052	1.135	1.107	-1.414	-1.168	-1.200	0.624	-0.124	-0.914
1.403	1.559	1.451	-1.500	-1.097	-1.160	1.090	0.009	-0.481
1.550	1.937	1.725	-1.503	-1.004	-1.476	1.430	-0.002	0.096
n/a	2.354	2.067	n/a	-1.321	-1.379	n/a	0.227	-0.811

**Fig. 1.** Oxygen isoscape (shaded in gray) showing the area in which we would expect our sea turtles to have resided throughout the life of the barnacles tested. This isoscape was calculated using an oxygen isotope ratio of  $-0.951 \delta^{18}\text{O}_c$ . PANWR is located within this area and depicted by the black triangle. Solid lines are contours of predicted oxygen isotope ratios in barnacle calcite ( $\delta^{18}\text{O}_c$ ).



353 **Fig. 2.** Adjusted oxygen isoscape (shaded in gray). PANWR is depicted by the black triangle. Solid  
354 lines are contours of predicted oxygen isotope ratios in barnacle calcite ( $\delta^{18}\text{O}_\text{c}$ ).

