

1 **Use of near-infrared spectroscopy to assess phosphorus**
2 **fractions of different plant availability in forest soils**

3 J. Niederberger^{1*}, B. Todt^{1*}, A. Boča², R. Nitschke¹, M. Kohler¹, P. Kühn³ and J. Bauhus¹

4

5 1 Chair of Silviculture, Institute of Forest Science, University of Freiburg, Freiburg,
6 Germany.

7 2 Wildland Resources Department, Utah State University, Logan, UT, United States.

8 3 Department of Geosciences, Physical Geography and Soil Science, University of Tübingen,
9 Tübingen, Germany.

10 * These authors contributed equally to this work

11

12 Correspondence to: J. Niederberger (joerg.niederberger@waldbau.uni-freiburg.de)

13

14 **Abstract**

15 The analysis of soil phosphorus (P) in fractions of different plant availability is a common
16 approach to characterize the P status of forest soils. However, quantification of organic and
17 inorganic P fractions in different extracts is labor-intensive and therefore rarely applied for
18 large sample numbers. Therefore, we examined whether different P fractions can be predicted
19 using near-infrared spectroscopy (NIRS).

20 We used the Hedley sequential extraction method (modified by Tiessen & Moir, 2008) with
21 increasingly strong extractants to determine P in fractions of different plant availability and
22 measured NIR spectra for soil samples from sites of the German forest soil inventory and
23 from a nature reserve in south-eastern China.

24 The R^2 of NIRS calibrations to predict P in individual Hedley fractions ranged between 0.08
25 and 0.85. When these fractions were combined into labile, moderately labile and stable P
26 pools, R^2 of calibration models was between 0.38 and 0.88 (all significant). Model prediction
27 quality was higher for organic than for inorganic P fractions and increased with the
28 homogeneity of soil properties in soil sample sets. Usable models were obtained for samples

1 originating from one soil type in subtropical China, whereas prediction models for sample sets
2 from a range of soil types in Germany were only moderately useable or not useable.

3 Our results indicate that prediction of Hedley P fractions with NIRS can be a promising
4 approach to replace conventional analysis, if models are developed for sets of soil samples
5 with similar physical and chemical properties, e.g. from the same soil type or study site.

6

7 **Keywords:** Soil P, Hedley fractions, P pools, model calibration, model validation

8

9 **1 Introduction**

10 Phosphorus (P) is limiting plant growth and ecosystem productivity in many parts of the
11 world (e.g. Elser et al., 2007, Vitousek et al., 2010). Unlike nitrogen, the world's P stores for
12 the production of fertilizer are finite and therefore will decrease in the future (i.a. Cordell et
13 al., 2009, Edixhoven et al. 2013). It is therefore of paramount importance to use P as
14 efficiently as possible in agricultural and forestry production systems. In several parts of
15 Europe and other parts of the world, concerns have been expressed that a substantial
16 proportion of forest stands may suffer from P limitation (Fox et al., 2011, Khanna et al., 2007,
17 Lorenz et al., 2003). A number of hypotheses, to explain this phenomenon of wide-spread
18 phosphorus limitation have been put forward (e.g. nitrate uptake suppressing P uptake, soil
19 acidification reduces rate of organic P mineralization and increases inorganic P sorption
20 capacity) (Mohren et al., 1986, Paré and Bernier, 1989, Gillespie and Pope, 1991). In long-
21 term development of forest ecosystems following disturbance, P (in contrast to N) can only
22 diminish through processes such as erosion and timber harvest and therefore becomes limiting
23 to biological activity. Consequently, increasing P deficiency is also a natural process of
24 ecosystem retrogression (Walker et al., 1976, Wardle, 2009), which seems to be accelerated
25 by anthropogenic activities. The increasing demand for forest biomass, such as for renewable
26 energy, might exacerbate the P nutrition problem in many forests (Vanguelova et al., 2010).
27 Therefore, suitable methods for monitoring the current status and the medium to long-term
28 trends of P nutrition in forest ecosystems are urgently needed.

29

30 For forest soils, often solely total P contents are measured. But total P is not a meaningful
31 variable as it is not very sensitive to environmental or management influences over time and

1 is not indicative of the amount of plant-available P in forest soils, where P can occur in many
2 organic and inorganic forms of different availability (Khanna et al., 2007). Especially for
3 forest soils, it is important to differentiate between organic and inorganic P forms, since
4 organic P, although not directly available for plant uptake, plays a major role in tree nutrition
5 (Turner et al., 2005, Shen et al., 2011, Rennenberg and Herschbach, 2013). Analytical
6 approaches, such as the Hedley fractionation (Hedley et al. 1982), have been developed to
7 quantify organic and inorganic P in fractions of different plant availability. These fractions
8 have been successfully applied to characterize the dynamic nature of P availability in forest
9 ecosystems (e.g. Richter et al., 2006). Several authors discussed the usefulness of this
10 commonly used sequential extraction method to determine P in fractions of distinctly different
11 solubility (Cross and Schlesinger, 1995, Condon and Newman, 2011, Yang and Post, 2011).
12 The ecological relevance of chemically derived Hedley fractions has been critically discussed
13 in some cases (Turner et al., 2005) and it is still unclear which fractions are relevant for plant
14 growth (Rennenberg and Herschbach, 2013). However, the application of this sequential
15 extraction method in forest soils has proven to be useful. In contrast to agricultural soils, the
16 slowly cycling P pool in forest soils contributes substantially to plant nutrition, whereas labile
17 P pools stay relatively constant, indicating that higher amounts of P from more slowly cycling
18 pools “feed” the labile P pool (Compton and Cole, 1998, Richter et al., 2006). However, the
19 Hedley fractionation method is very time consuming, labor intensive and therefore costly,
20 which renders it unsuitable for the analysis of large numbers of samples (Cécillon et al.,
21 2009). Therefore, it would be useful, if Hedley P fractions could be determined by means of
22 an indirect and less expensive technique. For this purpose, the use of near-infrared
23 spectroscopy (NIRS) may be a promising approach.

24 NIRS is a rapid and non-destructive analytical method routinely used in a wide range of
25 fields, mostly in quality control in the chemical, forage or food industry. In the field of soil
26 sciences, NIRS has been successfully applied to predict soil chemical and soil biological
27 parameters like carbon (in the form of total C, SOC, microbial C, and mineralizable C) or
28 nitrogen (in the form of total N, microbial N, and mineralizable N) contents (Chang et al.,
29 2001, Ludwig et al. 2002, Zornoza et al., 2008) and was also applied to agricultural soils to
30 predict available P (Olsen P, Mehlich III P) (eg Maleki et al., 2006, Dhawale et al., 2013). An
31 overview of available reflectance spectra for different soil properties is available at the global
32 soil vis-NIR spectral library (World Agroforestry Centre – ICRAF and ISRIC-World soil
33 Information, 2010).

1 The mineral fractions of soil, which commonly constitute the major portion of soils, are
2 usually hardly visible for NIRS (Malley et al., 2004). Despite the complexity and
3 heterogeneity of chemical and physical soil properties, the spectral information of soils is
4 surprisingly uniform. For example, 30 soil types of the USDA soil classification system could
5 be grouped into only 5 different spectra classes (Stoner and Baumgardner, 1981). Owing to
6 the low dipole moment between P and oxygen, phosphates cannot be excited by infrared
7 radiation and therefore not be detected directly with NIRS. However, P can be detected
8 indirectly, if it is bound organically or through correlation with the above stated other soil
9 properties such as C and N content that are detectable by NIRS.

10

11 To develop robust NIRS models, it is essential to produce a reliable quality in spectral data
12 sets. Several publications on NIRS deal with the influence of the quality of soil samples on
13 spectral information. Stenberg et al. (2010) and Brunet et al. (2007) discussed the influence of
14 ground soil samples in comparison with sieved samples (<2 mm) and found increasing model
15 quality for more homogeneous ground samples. The quality of the data with regard to the
16 heterogeneity of samples was addressed by Abrams et al. (1987) and Shenk and Westerhaus
17 (1993). Global models used for the prediction of forage quality in hay, where “global” means
18 that the model is based on the largest sample population comprising a wide variety of hay
19 samples and hay composition from many sites, produced predictions that were as good as
20 local models derived from one single site. However, in other studies using plant material (pine
21 needles), both, improvements and reductions in model quality for more heterogeneous data
22 sets (addressed as global) when compared with homogeneous (local) data, were observed
23 (Gillon et al., 1999). Reductions in model quality were observed for the prediction of C
24 content in sample sets with high variation in chemical composition associated with high
25 spectral variation. Since high variation in chemical composition and spectral properties is true
26 for forest soil samples, it might be a challenging process to build robust models. The problem
27 of detecting soil characteristics using NIRS methods is not new (Stoner and Baumgardner,
28 1981, Ben-Dor and Banin, 1995, Udelhofer et al., 2003, Cécillon et al., 2009, Genot et al.
29 2011). To our knowledge, up to now only few publications (Guerrero et al., 2010, Sankey et
30 al., 2008) have addressed the problem of NIRS model calibration and prediction quality for
31 forest soil samples and none of these has focused on soil P.

1 Currently, there are only few NIRS models available to predict soil-P parameters. These
2 includes NIRS-prediction models for Mehlich-III extractable P (Chang et al., 2001), for total
3 P and exchangeable P (Malley et al., 2004). Hitherto, there have been no studies focusing on
4 the use of NIRS to predict organic and inorganic Hedley P fractions in forest soils. This study
5 aimed to examine the general ability of NIRS methods to assess these P fractions. Specifically
6 we addressed the following hypotheses:

- 7 1. Hedley fractions of different P availability can be sufficiently well predicted through
8 NIRS models. The criteria used to quantify the quality of NIRS models will be
9 introduced in the Material and Methods section.
- 10 2. The quality of predictions through NIRS models is higher for organic than for
11 inorganic P fractions.
- 12 3. The quality of predictions of P fractions depends on the variability in Phosphorus
13 content and soil properties of soil sample data sets.

15 **2 Material and methods**

16 **2.1 Soil samples**

17 Two different sets of mineral soil samples were used for this study. Approximately half of the
18 soil samples originated from the German nationwide forest soil inventory (BZE). This data set
19 covers 282 archived soil samples from 145 sites belonging to the BZE inventory grid.
20 Including 70 sites in Baden-Württemberg (70 samples 0-5 cm, 69 samples 10-30 cm), 23 sites
21 in Hessen (23 samples from both depths), 33 sites in Lower Saxony (32 samples 0-5 cm, 28
22 samples 10-30 cm) and 19 sites in Saxony Anhalt (18 samples 0-5 cm, 19 samples 10-30 cm).
23 The area covered by the BZE samples was a region of approximately 200 km width and 700
24 km length. The BZE plots were part of German Forest Soil inventory net, which is realized as
25 a grid with a size of 8 x 8 km. From each site, if available, two samples (0-5 cm and 10-30 cm
26 depth) were analyzed. BZE samples are composite samples from 8 replicates (BZE 2006).
27 The samples included in this data set were selected to cover the majority of forest soil types in
28 Germany and a large variation in total phosphorous concentration. Development of robust
29 NIRS-models requires sample populations that cover the whole calibration range with an
30 approximately even distribution of samples across the range of the variable to be predicted. In
31 contrast, sample populations with normally distributed samples tend to overestimate low

1 values and underestimate high values in model calibration (Williams, 2001). The BZE soil
2 samples were grouped by soil type in eleven major groups: Regosols (n = 20), Gleysols (n =
3 6), Luvisols (n = 59), Vertisols (n = 2), Podzols (n = 10), Chernozem (n = 2), Stagnosols (n =
4 38), chromic Cambisols (n = 10), Anthrosols (n = 4), cambic Umbrisol (n = 78), and
5 Cambisol (n = 53) (IUSS Working Group WRB 2006). The total P content covered by the
6 BZE samples ranged from 58 to nearly 2800 mg kg⁻¹ (Tab. 1).

7 The other half of the soil samples originated from the international research project
8 “Biodiversity and Ecosystem Functioning in China (BEF China)” (Bruehlheide et al., 2011).
9 The 294 soil samples from the BEF China project were collected from 27 so-called Comparative
10 Study Plots (= CSP, 30 x 30 m) in the Gutianshan National Nature Reserve, which has a size of
11 81.07 km² (mean distance between all CSPs = 3.4 km; Min = 0.04 km; Max = 8.98 km). At all
12 27 CSPs the soil type was a Cambisol (reference soil group after WRB, 2006) comparable to
13 the “typical Brown Earth” after the German classification. Thus the 294 BEF samples
14 represented a more homogeneous sample set than the BZE samples. At each CSP, nine soil
15 cores that were evenly distributed across the plot were collected. One bulk sample was then
16 combined from the nine cores, for each of the three depth increments (0-5 cm, 5-10 cm and
17 10-20 cm), i.e. 81 samples in total. At twelve CSPs, an additional 37 samples were collected
18 from the three topmost diagnostic soil horizons, predominantly Ah and Bw horizons (at one
19 plot, the four topmost horizons were sampled) of representative soil profiles of the CSPs
20 (ranging in depth between 47 and 120 cm). In addition, 176 samples (0-20 cm) were collected
21 in four replicates from 44 so-called Neighborhood Diversity Tree Clusters, which were
22 located in the proximity of some of the permanent Comparative Study Plots. Each of the four
23 samples of tree clusters, were also composite samples from three cores each. Each composite
24 sample represents different conditions within the cluster; they were collected at the base of
25 individual trees belonging to different or the same tree species and in the center of a triangle
26 between these trees. We cannot rule out a spatial correlation between these samples. Total P
27 content of all BEF samples ranged from 26 to 330 mg kg⁻¹ (Tab. 1).

28 Both sample sets, BZE and BEF, contained samples that originated from different depth but
29 the same location (soil cores). Since these samples might not have been spatially independent,
30 the autocorrelation among samples could have influenced the quality of NIRS models. For
31 example, the validation may not be truly independent, if calibration and validation sample sets
32 contain samples that are systematically correlated with each other. To address this issue, we

1 calculated the Durbin-Watson coefficient (d) as a measure of autocorrelation between
2 measured and predicted values and compared this coefficient between sample sets from all
3 soil depths (containing samples from the same location) and from one soil depth only
4 (spatially independent). This test did not indicate autocorrelation within our sample sets (d
5 values between 1.75 and 2.23). In addition, we found no differences for the Durban-Watson
6 coefficient for all BZE samples (depth: 0-5 cm and 10-30 cm) and BZE samples sets from one
7 depth only. Since there was no indication of autocorrelation between samples of different
8 depths, we included all samples in our calibration and validation steps.

9 The data for C and N content as well as the pH values of the BZE samples were provided by
10 the Northwest German Research Station and the Forest Research Institute Baden-
11 Wuerttemberg. Samples were prepared and measured according to the German Forest science
12 standard (BMVEL, 2009). Total soil C and N contents of the horizon-wise samples and the
13 three depth increments of the BEF samples were determined after dry combustion (1150°C)
14 with an element analyzer (Elementar Vario EL III) (DIN ISO 10964). Since the material is
15 non calcareous, total soil C equals soil organic C. For these samples, pH values were derived
16 in H₂O (2.5 ml : 1 g water solution to soil) with a Sentix 81 electrode and pH meter (WTW,
17 Germany). C and N contents of the 176 neighbourhood diversity samples of the BEF dataset
18 were determined with a TruSpec Elemental Analyzer (Leco, USA) and pH values were
19 measured in H₂O (2.5 ml : 1g water solution to soil) with a blueLine electrode and pH meter
20 (Schott, Germany).

21 All soil samples were dried at 40°C for 24 h and sieved (< 2 mm). For C and N analyses,
22 samples were finely ground. The intensity of sieving respectively grinding has an impact on
23 the reproducibility and the distribution of extractable P over the fractionation process (Tiessen
24 and Moir 2008) and the NIR spectra (Brunet et al., 2007). Due to the more “natural”
25 extractability of P compared to ground samples and the ecological importance of soil
26 aggregates, we used < 2 mm sieved soil samples for the determination of P fractions.

27 Before measurement of the NIR spectra, the samples were ground to powder by a mixer mill
28 MM400 (Retsch, Germany) and dried over night at 40°C.

29

1 2.2 Phosphorus fractionation

2 Soil samples were analyzed by the sequential extraction method according to Hedley (Hedley
3 et al., 1982b). We used 24 samples in one batch consisting of 23 individual samples including
4 six replicate samples (as random quality check) and one in-house soil standard (as standard
5 quality check). In the protocol applied to the samples, 0.5 g soil was extracted by different
6 extractants with increasing strength (Fig. 1), starting with distilled water containing an anion
7 exchange resin - Dowex 1x8; 20-50 mesh; Sigma-Aldrich, Taufkirchen, Germany (Resin),
8 followed by sodium bicarbonate, 0.5 M (NaHCO_3), sodium hydroxide, 0.1 M (NaOH),
9 sodium hydroxide, 0.1 M with ultrasonic treatment - ultrasonic bath RK510H, 35kHz; 23W/l;
10 Bandelin, Berlin, Germany; (S-NaOH), hydrochloric acid, 1M (1M HCl) and a final acid
11 digestion (Residual). Tiessen and Moir (2008) introduced an additional extraction step with
12 hot concentrated hydrochloric acid (HCl conc.) before the final digestion. Our fractionation
13 scheme considered all of these 7 fractionation steps, but used 25 ml solution for the
14 extractions and 2.2 ml concentrated nitric acid for the final acid digestion. The whole
15 fractionation process is described in detail by Tiessen and Moir (2008).

16 Total P content as index value for the sum of all fractions was measured separately after nitric
17 acid digestion, which is a standard method recommended by the German forest soil survey
18 (BMVEL, 2009).

19 Phosphorus was determined colorimetrically with the spectrophotometer UV mini-1240 by
20 Shimadzu through molybdate reactive P method described by Murphy and Riley (1962)
21 modified by John (1970), but keeping the ammonium molybdenum concentration at 10 g l^{-1} .
22 Every extract needs to be adjusted for pH, as development and stability of the blue
23 phosphomolybdic complex is pH dependent (Drummond and Maher, 1995, Huang and Zhang,
24 2008, Tiessen and Moir, 2008). The P extracted within every step is only partly detectable as
25 free phosphate. To determine the total phosphate, the NaHCO_3 , NaOH and the HCl conc.
26 extracts were oxidized with ammonium persulfate in an autoclave. The difference between
27 total P after digestion (TP) and the free phosphate in the extract (P_i) provides the organically
28 bound P (P_o) (Rowland and Haygarth, 1997, Tiessen and Moir, 2008). The amount of
29 organically bound P in the resin and the 1 M HCl extracts were considered negligible (Tiessen
30 and Moir, 2008) and therefore not analyzed. We summed up the values P_i of the NaOH and P_i
31 of the S-NaOH fraction as well as P_o of the NaOH and P_o of the S-NaOH fraction because
32 the ultrasonic treated NaOH step is chemically hardly different from the main NaOH fraction

1 and a differentiation between these two fractions with NIRS did not seem meaningful. Similar
2 considerations applied to the distinction between Pi and Po in the HCl conc. extract, since
3 concentrated HCl is a strong oxidant and an additional oxidation with persulfate is chemically
4 comparable. In our analysis, the precision of the Pi value of the HCl conc. extract lacked
5 reproducibility within replicates compared to the TP value and this might be an indicator for
6 an incomplete reaction during the boiling procedure. Also several organic phosphates are
7 unstable under these acid conditions (Turner et al., 2005) and the results of Pi and Po of the
8 HCl conc. fraction lacked also reproducibility within our standard. The total P content of the
9 HCl conc. fractions showed good reproducibility and therefore was used for calibration.

10 In summary, we selected the following fractions to perform calibrations with NIRS: Pi Resin,
11 Pi NaHCO₃, Po NaHCO₃, Pi NaOH, Po NaOH, Pi 1M HCl, P HCl conc., P residual. For the
12 purpose of subsequent statistical analyses, the Hedley fractions were grouped by their
13 solubility in labile (Pi Resin, Pi and Po NaHCO₃), moderately labile (1M HCl, Pi and Po
14 NaOH) and stable P pools (P HCl conc. and P residual) as commonly found in literature
15 (Cross and Schlesinger, 1994, Stevenson and Cole, 1999, De Schrijver et al., 2011, Yang and
16 Post, 2011). Several tests were conducted to ensure that Hedley fractions were reproducible
17 and accurately determined. One indicator of the reliability of the Hedley fractionation method
18 is the recovery rate of total P in the sum of fractions, which was very good for the soils in this
19 study (Fig. 2). Only for samples with high P contents, the sum of fractions may either
20 systematically exceed the total P content or the amount of measured total P may be
21 underestimated. Errors for samples with high P content could occur either due to necessary
22 dilution steps for measuring fractions with very high P concentration, where small differences
23 during colorimetric measurements lead to relatively high errors for the final value, or to
24 incomplete digestion of P with nitric acid. To avoid these sources of error, we performed
25 repeated measurements of these samples and we reduced the sample weight to 400 mg and
26 increased the amount of acid to 3 ml in these samples. In addition, this problem occurred only
27 in a very low proportion of samples (10 out of 576) and removing these samples from the data
28 set did not change the calculated recovery rate.

29

1 **2.3 Near-infrared spectroscopy**

2 Molecules with dipole moments can be excited to stretching, bending, and rotating
3 oscillations by absorbing infrared radiation. While an excitation of fundamental oscillations
4 occurs in the mid infrared range of the light, the near-infrared spectrum of a substance shows
5 overtone peaks and combination bands of molecular vibrations, mainly from bonds like O-H,
6 C-H and N-H (Burns and Ciurczak, 2001, Malley et al., 2004, Cécillon et al., 2009). This
7 makes NIRS especially suitable for organic compounds (Weyer, 1985, Miller, 2001).
8 Phosphates and other inorganic P compounds are hardly detectable by NIRS due to the weak
9 P-O dipole moment (Malley et al., 2004). The detection of nutrients such as in P compounds
10 seems to be possible, if they are correlated with other NIRS-detectable soil properties such as
11 soil texture, water content, organic compounds, and mineralogy, in particularly soil organic
12 matter or organic carbon (Stenberg et al., 2010).

13 Each type of molecule has characteristic absorption bands for near-infrared radiation, but
14 there is much interference and overlap between the overtone and combination oscillations.
15 Therefore molecules cannot be identified by single peaks. The whole spectral information or
16 at least several regions of the spectrum are needed to determine a molecule (Burns and
17 Ciurczak, 2001, Günzler and Gremlich, 2003, Roberts et al., 2004).

18 The spectral region of NIR radiation ranges from 800 nm to 2500 nm wavelength, following
19 directly the visible light region. In infrared spectroscopy, commonly the wavenumber ν in cm^{-1}
20 is used, which is the reciprocal of the wavelength λ .

$$21 \quad \nu = \lambda^{-1}$$

22 Since the O-H bond has a strong influence on near-infrared absorption (Schmitt, 2000, Malley
23 et al., 2004) ground samples were dried at 40°C and measured under constant humidity
24 conditions. For each soil sample, five spectra were taken with a Fourier Transform Mid- and
25 Near Infrared combination instrument (Tensor 37, Brukeroptics, Ettlingen, Germany). Each
26 spectrum is a mean of 64 individual scans over the range of 12000 to 4000 cm^{-1} wave
27 numbers with a resolution of 16 cm^{-1} . Model development was done with the OPUS 6.5
28 spectroscopy software (Brukeroptics, Ettlingen, Germany). Before statistical analyses, a
29 number of mathematical data pre-processing options were tested. The pre-processing options
30 providing the best results were first derivative, vector normalization, or a combination of
31 these two. These were used as data treatment before partial least square regressions were
32 conducted and applied to a variety of frequency ranges to find the best fit with the value of the

1 respective component; spacing was 1; several smoothing point values (5, 9, 13, 17, 21, 25) for
2 the derivatives were tested, too. Calibration was performed with cross validation, a common
3 approach for small data sets (Williams, 2001, Conzen, 2005). Here, a defined number of
4 samples, in our case one sample, were stepwise excluded from the calibration process. The
5 rest of the samples were used to predict the excluded samples. This procedure was performed
6 until all samples were excluded once, and the best models to predict all samples were found
7 (Conzen, 2005). For each dataset, only 80% of the samples were used for this procedure and
8 the development of models, the remaining 20% were left out for later independent validation
9 of the models to test their prediction quality. This distribution of samples was carried out with
10 an automatic function in the Opus software (Conzen, 2005, Bruker Optik User Manual, 2006)
11 selecting samples for each individual model equally over the whole range of each dataset.
12 The min and max values were always assigned to the calibration set and the samples second
13 to min and max were always assigned to the validation set. To avoid confusion with the final
14 validation process of independent samples, we refer to the cross-validation process as
15 calibration.

16 Our initial intent was to generate a global prediction model for P-concentration that integrated
17 the whole variety of forest soil types. The aim was to test, if such a global model could be
18 used to estimate P-concentration of all BZE samples and for future inventories. Since this
19 approach proved to be difficult, we also tried to develop NIRS models for a sub-set of the
20 total BZE sample pool with reduced variation in soil properties including P. Therefore we
21 selected the BZE samples belonging to the German soil classification group “brown earth”.
22 This was the only sub-set of soil types (using soil type as an indication of similar soil
23 properties) in the BZE data set with sufficient number of samples for NIRS model
24 development. Additionally, we used soil samples from the above mentioned BEF China
25 project, which represented a data set from a small geographic region and similar soil
26 properties.

27 To address the hypothesis that the predictive quality of NIRS models increases with the
28 homogeneity of sample sets, we finally established calibrations for sample populations of
29 varying heterogeneity (Fig. 3):

- 30 1) a full dataset containing all soil samples with the greatest variability (n = 576)
- 31 2) the complete BZE dataset with almost similar variability comprising a wide range of
32 different soil types (n = 282)

- 1 3) the less variable BZE sub-set of “Brown Earths” soil samples (n = 84)
- 2 4) the BEF-China sample set comprising soils from one soil type and from one small
- 3 region (n = 294).

4

5 To assess whether the prediction of P by NIRS depended on direct detection of soil P -
6 regardless of organic or inorganic form - or rather indirectly on properties of soil organic
7 matter, we used regression analyses to see whether the quality of NIRS models to predict P
8 could be explained by the relationships between P and soil C and N. As mentioned above, it
9 was presumed that P bound to C or N could be modelled with better quality.

10

11 The BZE Brown Earths dataset (Set 3) with 82 samples was too small for a robust validation
12 with external samples that were not used in the calibration. Still, we included it in our study,
13 because it was the only meaningful and less variable subset of samples that could be separated
14 from the whole BZE data set (Set 2). The sizes of the sample Sets 1 and 2 were comparable
15 with those used in other studies to develop “global” models (Abrams et al., 1987; Shenk and
16 Westerhaus, 1993, Gillon et al., 1999). Sample Set 4 could be regarded as an extensive but
17 local data set. A principal component analysis (Fig. 3) (OPUS 6.5 software) was carried out to
18 assess spectral properties of the selected subsamples. The general spectral properties of the
19 soil samples indicated that the BEF China sample set (Set 4) was more homogenous than the
20 BZE sample sets and clearly separated from the BZE sample set as a whole as well as from
21 the subset BZE-“Brown-Earth”. The spectral properties of the two subsets of BZE samples
22 were not different

23 To evaluate the prediction quality of NIRS models, the statistical parameters R^2 and the ratio
24 of performance to deviation (RPD = ratio of standard deviation to standard error of
25 prediction) were used. The usefulness of these statistical parameters to characterize the quality
26 of NIRS models was discussed controversially (Chang et al., 2001, Williams, 2001, Malley et
27 al., 2004, Zornoza et al., 2008). Especially for soil analysis, high values for RPD and R^2 are
28 hardly achieved (Malley et al., 2004). To judge our RPD and R^2 values, we followed the
29 classification suggested by Malley et al. (2004), who provided an overview and summary on
30 this topic. We applied the following 4 levels to classify model quality:

- 31 • Level A: $R^2 > 0.95$ and $RPD > 4$, excellent calibrations, usable for all applications

- 1 • Level B: R^2 between 0.90 and 0.95, RPD between 3 and 4, successful calibrations
2 usable for most applications some with caution, e.g. quality assurance
- 3 • Level C: R^2 between 0.80 and 0.90, RPD between 2.25 and 3, moderately successful
4 calibrations, usable with caution, e.g. research
- 5 • Level D: R^2 between 0.70 and 0.80, RPD between 1.75 and 2.25, moderately useful
6 calibrations, usable for screening of samples

7 Calibrations with lower values might still be useful for rough screening purposes (Chang et
8 al., 2001).

9 All statistical analyses shown in this study are performed with SPSS Version 20.

10

11 **3 Results**

12 **3.1 Soil phosphorus contents**

13 Total P contents were higher and showed a wider range in the BZE samples than in the BEF-
14 China samples for all pools of P availability and individual fractions (Tab. 1). The sub group
15 “Brown Earths” was characterized by a large variation in total P content (130 to 2200 mg kg⁻¹
16 ¹, as the sum of all Hedley fractions), which had a smaller range of values compared to the
17 pool of all BZE samples but, as indicated by the standard deviation, a similar variation in total
18 P. The same applied to P in the labile, moderately labile, and stable pools. The mean
19 percentage of organic P, here calculated as the sum of all Po fractions without the HCl conc.
20 fraction, was higher in the BEF-China samples (57%) than in the BZE and BZE “Brown
21 Earths” samples (39% and 43%). However, there was no difference in the range of organic P
22 (10-70% of the total) for the three datasets. The organic P fraction with the highest
23 concentrations in all three datasets was the one extracted with NaOH (mean values for BZE
24 155 mg kg⁻¹, equivalent to 28% of P_t, BZE “Brown Earth” 196 mg kg⁻¹, equivalent to 31% of
25 P_t, and BEF-China 35 mg kg⁻¹, 37% of P_t). The P concentrations in the NaHCO₃-extractable
26 and the 1M HCl-extractable P_i fraction in the BEF samples were so low as to fall below the
27 detection limit of the Murphy & Riley (1962) method.

28

1 3.2. NIRS models for P fractions

2 Initially, NIRS models were calibrated for both, the complete sample set as well as for three
3 different subsets (BZE, BZE Brown Earths, and BEF-China). The global model for all
4 samples produced calibrations that were only below D level quality ($R^2 = 0.08\text{--}0.68$; RPD =
5 $1.04\text{--}1.74$). The BZE model, which was developed for samples from different soil types,
6 showed only for the P HCl conc. fraction a moderately useful calibration, while all other
7 fractions could not be calibrated sufficiently well (Tab. 3). For all fractions except the
8 inorganic NaOH, better calibrations were achieved with the BZE “Brown Earths” subset than
9 with the complete BZE dataset. However, calibration models achieved only D level quality
10 for two of the fractions (Po NaOH, P HCl conc.). The more homogenous BEF-China dataset
11 yielded the best calibrations. Only the inorganic NaHCO_3 and the inorganic 1M HCl fraction
12 were, owing to the very low contents, not calibrated successfully. Calibrations to predict the
13 Pi Resin, Po NaHCO_3 , Pi NaOH and P Residual fraction produced models of level D quality,
14 while calibrations for the organic NaOH and the P HCl conc. fractions yielded models of level
15 C quality. Overall, best calibration models were achieved for the BEF-China dataset
16 especially for the organic P fractions extracted with NaHCO_3 , NaOH, and the P HCl conc.
17 fraction (Tab. 3).

18 Models providing level D calibrations were subsequently tested with a validation dataset
19 comprising samples not included in the calibration. The validation of the level D calibration
20 model of the BZE set for the HCl conc. P fraction was not successful. Validation of this
21 fraction in samples of the more homogenous subset BZE “Brown Earths” yielded a model
22 quality of level C and validation models for the P HCl conc. fraction in BEF soil samples
23 achieved a level B quality (Fig. 4).

24 The validation of models for the Po NaHCO_3 ($R^2=0.72$, RPD=1.87), Pi NaOH ($R^2=0.68$,
25 RPD=1.77) and P Residual ($R^2=0.75$, RPD=1.98) fractions of the BEF China dataset reached
26 level D quality, while the model for the Pi Resin fraction could be validated at level C quality
27 ($R^2=0.83$, RPD=2.39). The validation of the model for the Po NaOH fraction reached level B
28 ($R^2=0.91$, RPD=3.23).

29

1 **3.3 NIRS models for P pools**

2 Grouping of the Hedley fractions into labile, moderately labile and stable P fractions did result
3 in good models for the BEF-China dataset, while only the stable fractions of the other three
4 datasets (BZE+BEF, BZE, BZE Brown Earth) could well be predicted with NIRS models (Fig.
5 5). Validation of the best calibration models for the stable P pool in the three datasets
6 BZE+BEF, BZE and BZE “Brown Earths” yielded a level D model ($R^2=0.79$, $RPD=2.16$) for
7 the complete set (BZE+BEF) as well as the BZE dataset ($R^2=0.73$, $RPD=1.91$) and a level C
8 model ($R^2=0.88$, $RPD=2.83$) for the BZE “Brown Earths” dataset.

9 Validations for the BEF China datasets achieved a level C quality for the labile and the
10 moderately labile P pool, and level D quality for the stable P pool (Fig. 6).

11
12 To assess whether P content in Hedley fractions was determined indirectly through
13 relationships with soil C and N, we calculated relationships between NIRS model quality and
14 coefficient of determination for the relationship between P in Hedley fractions and soil total C
15 and total N (Fig. 7). For organic P fractions high coefficients of determination (r^2) between P in
16 Hedley fractions and total soil C or total N coincided with high r^2 values of NIRS models. In
17 contrast, the P HCl conc. fraction showed the best model quality for all three data sets (r^2 0.72-
18 0.85) but very poor r^2 for the relationship with total C and total N. For inorganic P fractions for
19 the BZE samples, coefficients of determination (r^2) were poor for both, the relationships
20 between soil C and N and P fractions and NIRS models (Fig. 7).

21

22 **4 Discussion**

23 **4.1 NIRS models for P fractions and pools**

24 Calibration and validation models of individual fractions achieved good to satisfactory results
25 for some but not all individual P fractions. Generally, the accuracy and reproducibility of the
26 reference method (here Hedley fractionation) is of crucial importance for the quality of
27 predictions with NIRS models (Williams 2001). To increase reproducibility in our study, all
28 samples were measured according to the same protocol in the same laboratory at the
29 University of Freiburg. In addition, the strong agreement between the recovery of total soil P
30 and the sum of all individual fractions indicated a high reproducibility of the method. This

1 reproducibility, however, does not provide information about the distribution of P_i and P_o
2 within individual fractions. For the individual fractions, it is widely acknowledged that
3 significant potential errors may occur owing to variation in alkalinity or acidity in the extracts
4 leading to acid hydrolysis and precipitation of inorganic P. This can influence the accurate
5 distinction between organic and inorganic forms of P and may thus lead to over- or
6 underestimation of these forms within individual fractions (Magid et al., 1996, Condrón et al.,
7 2005, Turner et al., 2005, Tiessen and Moir, 2008, Condrón and Newman, 2011). However,
8 we achieved good reproducibility for P_i and P_o in repeatedly analyzed samples for NaHCO_3
9 and NaOH fractionation steps. For example, the standard errors of 16 repeated measurements
10 of the same sample were 5.0% for P_i NaOH, 8.3% for P_o NaOH, and 7.5% for P_{total} NaOH .

11 Other than these methodological limitations of the Hedley fractionation method, the level of P
12 content of each fraction is also of importance. The P content in the P_i NaHCO_3 and 1M HCl
13 fractions of the soil samples from subtropical China were close to the detection limit.
14 Therefore the reference data of these two fractions could not be regarded as reliable and hence
15 no meaningful calibration models with NIRS could be produced.

16

17 Using all Hedley fractions, the description of soil P availability becomes easily confusing.
18 Some fractions could not be calibrated owing to very low P concentrations or only with very
19 low prediction quality. In case of the BEF China data set, grouping the fractions into labile,
20 moderately labile, and stable pools showed a slight increase for the quality parameters R^2 and
21 RPD, but the quality of models remained at the same level. These pools incorporated the
22 inorganic NaHCO_3 and 1 M HCl fractions, which could not be predicted individually. As part
23 of the labile respectively moderately labile P pool, a reasonable prediction of the total P
24 content for the BEF China samples could still be possible. While pooling these fractions does
25 not reduce the work involved in reference analysis of the Hedley fractionation, it is still a
26 significant reduction in the modelling process, reducing the number of models from 8 to 3 for
27 each dataset.

28

29 **4.2 Calibration of organic and inorganic P fractions**

30 Whether P is in organic or inorganic form seemed to be of importance for the quality of
31 predictions of Hedley fractions with NIRS. Regardless of the datasets and sample origin,

1 models predicting the organic P fractions performed better than models predicting the
2 inorganic P fractions (Table 3). Similar results have been obtained for N (Ludwig et al.,
3 2002). The superior quality of models for organic P fractions is related to the underlying
4 mechanisms generating NIR spectra, because organic compounds can be more easily excited
5 by the irradiation than inorganic ones. Spectral variation in relation to soil P content is often
6 associated with organic matter or crystal water (Ben-Dor and Banin, 1995, Chang et al., 2001,
7 Malley et al., 2005, Stenberg et al., 2010). Therefore, we also calibrated NIRS models for the
8 sum of P in all organic fractions (data not shown). However, we could not observe any
9 improvement for prediction of the sum of assigned organic P in comparison with single
10 organic fractions or the pools. This might be caused by the large diversity in organic P
11 compounds (Turner, 2008). In contrast to organic P fractions, the relationships between
12 inorganic P fractions and other NIRS-detectable soil properties were obviously not strong
13 enough to aid the prediction of inorganic P using NIRS.

14 In our study, we were not able to identify spectral regions to be specific for a P signal as was
15 found in other studies (Malley et al., 2004). Therefore we had also assessed, if focusing on
16 typical NIR spectral regions for C-H, N-H and O-H bonds could influence NIRS model
17 quality. The organic residual which is connected with the phosphate molecule could be
18 dominated by CH, NH, OH bonds or a mixture of them. For this purpose we compared NIRS
19 models based on optimized spectral regions (automated procedure by OPUS software), on the
20 whole spectral range and on specific spectral regions, which are known to represent C-H, N-H
21 and O-H bonds (Conzen, 2005). We found that in all cases, the OPUS-software optimized
22 spectral selection yielded superior models followed by models covering the whole spectral
23 area. Models for selected bonds were in all cases of substantially lower quality, and were thus
24 not presented in detail. The best results based on r^2 and RPD were obtained for O-H bonds for
25 the $P_0\text{-HCO}_3$ and P-HCL conc. fractions. This was followed by models focusing on C-H
26 bonds and. The lowest quality models were obtained for models focusing on N-H bonds.

27 There was no general pattern for the relationship between NIRS model quality (r^2) for
28 different P fractions and the coefficient of determination of relationships between soil C or N
29 and P in these fractions. This indicated that NIRS models were based mostly on original
30 properties of organic P compounds and the predicted P content was not simply a function of
31 soil organic matter content. This corroborates the underlying assumptions of the Hedley
32 fractionation that P in the individual organic fractions is of different availability owing to the

1 different properties of the organic matter dominating the respective fraction. In turn, this
2 indicates that total organic soil C and N can be used only to a limited extent to predict organic
3 P in different fractions.

4 **4.3 Homogeneity of datasets**

5 The quality of predictions of P fractions depended on the homogeneity of individual datasets.
6 The question of homogeneity has also been discussed in the use of global and local calibration
7 models by Abrams et al. (1987) and Shenk & Westerhaus (1993), who found that global
8 models, with a higher number of samples per dataset, were potentially as accurate as more
9 local calibrations for mixtures of hay consisting of different plant species and origins.
10 Variable results were obtained for calibrations of the total C, N and P between global and
11 local models of heterogeneous plant material (Gillon et al., 1999). The global model, which
12 combined all individual datasets of pine needle categories (needles on trees, fallen needles
13 and litter), was more accurate or similar than the “local” individual datasets for each category
14 of needles (Gillon et al., 1999). However, for global models predicting carbon, a decline in
15 model quality was observed. Also, a comparison of NIRS models for soil organic C, inorganic
16 C and clay content showed a decreasing accuracy from local to global calibration models
17 (Sankey et al., 2008). In contrast Brunet et al. (2007) found for local NIRS models higher
18 model quality predicting total C or total N for local models compared with global models.
19 This increasing effect was stronger for ground samples (< 0.2mm) than for sieved samples
20 (<2mm). For our soil data sets, a similar incline in model quality with increasing homogeneity
21 of sample origin could be observed. The general observation that an increase in the range of
22 values leads to an increase in the robustness of NIRS models (Abrams et al., 1987, Shenk and
23 Westerhaus, 1993) did not apply in our study. Our results indicated that regardless of the
24 organic or inorganic origin or the availability of P, the quality of NIRS models to predict P
25 was better for more homogenous than heterogeneous sets of soil samples. For example,
26 reducing the soil samples from the nationwide forest soil monitoring program to the subset
27 BZE “Brown Earths”, we produced a more homogenous subset that improved model
28 calibrations for all but the inorganic NaOH fraction. Although an upgrade in model category
29 (into category D) was only observed for the organic NaOH fraction. The most homogenous
30 BEF China sample set produced even better calibration models, including improvements of
31 model category (into D as well as from D to C), than the BZE subset.

1 Since P is not “recognized” directly by near-infrared radiation but produces signals in the
2 spectra that are related to P compounds, it appears that the signals for the same group of P
3 compounds that relate to a specific Hedley fraction vary greatly among different soil types.
4 For organic P, these groups of compounds are probably present in more than one fraction and
5 the various extracts are unlikely to be either exhaustive or unique with respect to the target
6 compounds (Turner et al., 2005). To our knowledge, the chemical nature of the organic
7 phosphorus within the operationally defined fractions is still poorly understood (Magid et al.,
8 1996, Turner et al., 2005). In contrast, the inorganic P forms represented in the distinct P
9 fractions are more specific in their chemical nature and well known (Stevenson and Cole,
10 1999, Tiessen and Moir, 2008). This variation of P forms within Hedley fractions may be
11 greatly reduced through restricting the development of NIRS models to comparable soil types.

12

13 **5 Conclusions**

14 The approach to predict soil P Hedley fractions of different availability with NIRS can be
15 regarded successful only when certain limitations are considered. NIRS models for prediction
16 of individual inorganic P-fractions were at best moderately useful but unsuitable for soil
17 inventories. We found that the quality of NIRS models for prediction of P, for single fractions
18 as well as for pools, depended on the homogeneity of sample sets. The homogeneity may not
19 only be related to the same soil types (improved models after grouping of soil types in the
20 BZE subset), but also to the same geology that has a large influence on soil properties (better
21 calibration models of the BEF China sample set - with the same geology - than the BZE
22 subset originating from different geologies). The approach may therefore be well suited for
23 large experiments that require a great number of spatial and temporal replication of samples
24 within well-defined soil types. For large-scale, grid based soil inventories such as those
25 conducted in the context of environmental monitoring programs for entire regions, like
26 nation-wide soil surveys, it appears that NIRS-models to predict P fractions need to be
27 developed for individual soil groups or other classes of samples according to specific soil
28 properties. While the development of NIRS models requests a considerable initial investment,
29 these efforts are likely to pay off only for large sample numbers and in repeated inventories.

30

31 **Acknowledgments**

1 B. Todt was financed by a DFG grant (BEF China, FOR 891, BA 2821/10-1) to Jürgen
2 Bausch. J. Niederberger was financed through a grant provided by the Federal Ministry of
3 Food, Agriculture and Consumer Protection, administered through the Von-Thünen Institute.
4 We wish to thank K. Winschu for the technical support, the Forest Research Institute Baden-
5 Wuerttemberg and the Northwest German Forest Research Station for providing the BZE
6 samples and C. Geißler and S. Flaiz of the University of Tübingen for preparing and
7 providing a share of the BEF China samples. We greatly appreciate the comments of Dr.
8 Partap K. Khanna and three anonymous reviewers on an earlier version of the manuscript.

9

10 **References**

- 11 Abrams, S. M., Shenk, J. S., Westerhaus, M. O., and Barton F. E. II: Determination of forage
12 quality by near infrared reflectance spectroscopy efficacy of broad-based calibration
13 equations, *J. Dairy Sci.*, 70, 806-813, 1987.
- 14 Attiwill, P. M. and Adams, M. A.: Nutrient cycling in forests. *New Phytol.*, 124, 561-582,
15 1993.
- 16 Ben-Dor, E. and Banin A.: Near infrared analysis (NIRA) as a method to simultaneously
17 evaluate spectral featureless constitutions in soil, *Soil Sci.*, 159, 259-270, 1995.
- 18 BMELV: Handbuch Forstliche Analytik, HFA-Teil A, Boden und Humusuntersuchungen,
19 2009. [http://www.bmelv.de/SharedDocs/Downloads/Landwirtschaft/Wald-
20 Jagd/Bodenzustandserhebung/Handbuch/A-Boden-und-
21 Humusuntersuchungen.pdf?__blob=publicationFile/](http://www.bmelv.de/SharedDocs/Downloads/Landwirtschaft/Wald-Jagd/Bodenzustandserhebung/Handbuch/A-Boden-und-Humusuntersuchungen.pdf?__blob=publicationFile/) , Accessed 1. April 2014
- 22 Bruelheide, H., Böhnke, M., Both, S., Fang, T., Assmann, T., Baruffol, M., Bausch, J.,
23 Buscot, F., Chen, X-Y., Ding, B., Durka, W., Erfmeier, A., Fischer, M., Geißler, C., Guo, D.,
24 Guo, L-D., Härdtle, W., He, J-S., Hector, A., Kröber, W., Kühn, P., Lang, A., Nadrowski, K.,
25 Pei, K., Scherer-Lorenzen, M., Shi, X., Scholten, T., Schuldt, A., Trogisch, S., von Oheimb,
26 G., Welk, E., Wirth, C., Wu, Y-T., Yang, X., Zeng, X., Zhang, S., Zhou, H., Ma, K. and
27 Schmid, B.: Community assembly during secondary forest succession in a Chinese
28 subtropical forest, *Ecol. Monogr.*, 81, 25–41, 2011.
- 29 BRUKER OPTIK GmbH: User Manual Quant, Opus Spectroscopy Software, Version 6,
30 Ettlingen, 2006.

1 Brunet, D., Barthès, B. G., Chotte, J. L. and Feller, Ch.: Determination of carbon and nitrogen
2 contents in Alfisols, Oxisols and Ultisols from Africa and Brazil using NIRS analysis: Effects
3 of sample grinding and set heterogeneity, *Geoderma*, 139, 106-117, 2007.

4 Burns, D. A. and Ciurczak, E. W. (eds): *Handbook of Near-Infrared Analysis*, Second
5 Edition, in: *Practical Spectroscopy a Series*, Vol 27, Marcel Dekker Inc New York, 2001

6 Cécillon, L., Barthès, B. G., Gomez, C., Ertlen, D., Genot, V., Hedde, M., Stevens, A. and
7 Brun, J. J.: Assessment and monitoring of soil quality using near-infrared reflectance
8 spectroscopy (NIRS), *Eur. J. Soil Sci.*, 60(5), 770-784, 2009.

9 Chang, C.-W., Laird, D. A., Mausbach, M. J and Hurburgh, C. R.: Near-Infrared Reflectance
10 Spectroscopy - Principal Components Regression Analyses of Soil Properties Journal Paper
11 no. J-18766 of the Iowa Agric. and Home Econ. Exp. Stn., Ames, IA. *Soil. Sci. Soc. Am. J.*
12 65, 480-490, 2001.

13 Compton, J. E. and Cole, D. W.: Phosphorus cycling and soil P fractions in Douglas-fir and
14 red alder stands, *For. Ecol. Manag.*, 110, 101-112, 1998.

15 Condon, L. M., Turner, B. L. and Cade-Menun, B. J.: Chemistry and dynamics of soil
16 organic phosphorus. In: Sims, T. and Sharpley, A. N. (eds): *Phosphorus: Agriculture and the*
17 *Environment*, Am. Soc. Agron., Madison, Wisconsin, USA, 87-121, 2005.

18 Condon, L. M. and Newman, S.: Revisiting the fundamentals of phosphorous fractionation of
19 sediments and soil, *J. Soil Sediment*, 11, 830-840, 2011.

20 Conzen, H.-P.: *Multivariate Kalibration, Ein praktischer Leitfaden zur*
21 *Methodenentwicklung in der analytischen Analytik*, Bruker Optik GmbH, 2005.

22 Cordell, D., Drangert, J.-O. and White, S.: The story of phosphorus: Global food security and
23 food for thought, *Glob. Environ. Chang.*, 19, 292-305, 2009.

24 Cross, F. and Schlesinger, W. H.: A literature review and evaluation of the Hedley
25 fractionation: Applications to the biogeochemical cycle of phosphorous in natural ecosystems,
26 *Geoderma*, 64, 197-214, 1995.

27 De Schrijver, A., Vesterdal, L., Hansen, K., De Frenne, P., Augusto, L., Achat, D. L.,
28 Staelens, J., Baeten, L., De Keersmaeker, L., De Neve, S. and Verheyen, K.: Four decades of
29 post-agricultural forest development have caused major redistributions of soil phosphorus
30 fractions, *Oecologia*, 167, 221-234, 2012.

1 Dhawale, N. M., Adamchuck, V. I., Viscarra Rossel, R. A., Prascher, S. O., Whalen, J. K. and
2 Ismail, A. A.: Predicting extractable soil phosphorus using visible / near-infrared
3 hyperspectral soil reflectance measurements, The Canadian Society for Bioengineering 2013
4 Annual Meeting, Saskatoon, Saskatchewan, 7-10 July 2013, Paper No. CSBE13-047.

5 Drummond, L. and Maher, W.: Determination of phosphorus in aqueous solution via
6 formation of the phosphoantimonymolybdenum blue complex. Re-examination of optimum
7 conditions for the analysis of phosphate, *Anal. Chim. Acta.*, 302(1), 69-74, 1995.

8 Duckworth, J.: Mathematical Data Preprocessing, in: Roberts, C. A., Workmann, J. J. and
9 Reeves J. B. III (eds): *Near-Infrared Spectroscopy in Agriculture*, in: *Agronomy Series*, Vol
10 44, Am. Soc. Agron. Inc, Crop. Sci. Soc. Am. Inc, Soil Sci. Soc. Am. Inc, Madison: 729 –
11 784, 2004.

12 Edixhoven, J. D., Gupta, J., and Savenije, H. H. G.: Recent revisions of phosphate rock
13 reserves and resources: reassuring or misleading? An in-depth literature review of global
14 estimates of phosphate rock reserves and resources, *Earth Syst. Dynam. Discuss.*, 4, 1005-
15 1034, doi:10.5194/esdd-4-1005-2013, 2013.

16 Elser, J. J., Bracken, M. E. S., Cleland, E. E., Gruner, D. S., Harpole, W. S., Hillebrand, H.,
17 Ngai, J. T., Seabloom, E. W., Shurin, J. B. and Smith, J. E.: Global analysis of nitrogen and
18 phosphorus limitation of primary producers in freshwater, marine and terrestrial ecosystems,
19 *Ecol. Lett.*, 10, 1135-1142, 2007.

20 Fox, T. R., Miller, B. W., Rubilar, R., Stape, J. L. and Albaugh, T. J.: Phosphorus Nutrition of
21 Forest Plantations: The Role of Inorganic and Organic Phosphorus, in: *Soil Biology* 26,
22 Chapter 13, 317-338, Springer-Verlag, Berlin/Heidelberg, Germany, 2011.

23 Genot, V., Colinet, G., Bock, L., Vanvyve, D., Reusen, Y. and Dardenne, P.: Near infrared
24 reflectance spectroscopy for estimating soil characteristics valuable in the diagnosis of soil
25 fertility, *J. Near Infrared Spectrosc.*, 119, 117-138, 2011.

26 Gillespie, A. R. and Pope, P. E.: Consequences of rhizosphere acidification on delivery and
27 uptake kinetics of soil phosphorus, *Tree Physiol.*, 8, 195-204, 1991.

28 Gillon, D., Houssard, C. and Joffre R.: Using near-infrared reflectance spectroscopy to predict
29 carbon, nitrogen and phosphorus content in heterogeneous plant material, *Oecologia*, 118,
30 173-182, 1999.

- 1 Gruselle, M.-C. and Bauhus, J.: Assessment of the species composition of forest floor
2 horizons in mixed spruce-beech stands by Near Infrared Reflectance Spectroscopy (NIRS).
3 *Soil Biol. Biochem.*, 42, 1347-1354, 2010.
- 4 Günzler, H. and Gremlich H.-U.: IR Spektroskopie eine Einführung, Wiley-VCH Verlag,
5 Weinheim, 2003
- 6 Hättenschwiler, S., Aeschlimann, B., Coûteaux, M.-M., Roy, J. and, Bonal, D.: High variation
7 in foliage and leaf litter chemistry among 45 tree species of a neotropical rainforest
8 community, *New Phytol.*, 179, 165-175, 2008.
- 9 Hedley, M. J., Stewart, J. W. B. and Chauhan, B. S.: Changes in Inorganic and Organic Soil
10 Phosphorus Fractions Induced by Cultivation Practices and by Laboratory Incubations, *Soil*
11 *Sci. Soc. Am. J.*, 46, 970-976, 1982a.
- 12 Hedley, M. J., White, R. E. and Nye, P. H.: Plant-Induced changes in the rhizosphere of rape
13 (*Brassica Napus* Var. Emerald) seedlings, III. Changes in L value, soil phosphate fractions
14 and phosphatase activity, *New Phytol.*, 91, 45-56, 1982b.
- 15 Huang, X. L. and Zhang, J. Z.: Rate of phosphoantimonymolybdenum blue complex
16 formation in acidic persulfate digested sample matrix for total dissolved phosphorus
17 determination: Importance of post-digestion pH adjustment, *Talanta*, 77(1), 340-345, 2008.
- 18 John, M. K.: Colorimetric Determination of Phosphorus in Soil and Plant Materials with
19 Ascorbic Acid, *Soil Sci.*, 109, 214-220, 1970.
- 20 Khanna, P. K., Bauhus, J., Meiwes, K. J., Kohler, M., Rumpf, S. and Schönfelder, E.:
21 Assessment of changes in the Phosphorus status of forest ecosystems in Germany – Literature
22 review and analysis of existing data. Report to the German Federal Ministry of Food,
23 Agriculture and Consumer Protection, 2007.
- 24 Lei, P. and Bauhus, J.: Use of near-infrared reflectance spectroscopy to predict species
25 composition in tree fine-root mixtures, *Plant Soil*, 333, 93-103, 2010.
- 26 Lorenz, M., Mues, V., Becher, G., Müller-Edzards, C., Luysaert, S., Raitio, H., Fürst, A. and
27 Langouche, D.: Forest Condition in Europe - Results of the 2002 Large-scale Survey. UNECE
28 and EC, Geneva and Brussels, 2003.

- 1 Ludwig, B., Khanna, P. K., Bauhus, J. and Hopmans, P.: Near infrared spectroscopy of forest
2 soils to determine chemical and biological properties related to soil sustainability. *For. Ecol.*
3 *Manag.*, 171, 121-132, 2002.
- 4 Magid, J., Tiessen, H. and Condon, L. M.: Dynamics of organic phosphorus in soils under
5 natural and agricultural ecosystems, in: Piccolo, A. (ed): *Humic Substances in Terrestrial*
6 *Ecosystems*, Elsevier Sci., Oxford, 429-466, 1996.
- 7 Maleki, M. R., Van Holm, L., Ramon, H., Merckx, R., De Baerdemaeker, J. and Mouazen, A.
8 M.: Phosphorus Sensing for Fresh Soils using Visible and Near Infrared Spectroscopy,
9 *Biosystems Engineering*, 95, 425-436, 2006.
- 10 Malley, D. F., Martin, P. D. and Ben-Dor, E.: Application in Analysis of Soils, in: Roberts, C.
11 A., Workmann, J. J. and Reeves, J. B. III (eds): *Near-Infrared Spectroscopy in Agriculture*, in:
12 *Agronomy Series*, Vol 44, Am. Soc. Agron. Inc, Crop Sci. Soc. Am. Inc, Soil Sci. Soc. Am.
13 Inc, Madison, 729 – 784, 2004.
- 14 Miller, C. E.: Chemical principles of near infrared technology, in Williams, P. and Norris, K.:
15 *Near-infrared technology in the agricultural and food industries*, Am Association Cereal
16 Chemist. Inc, 19-37, 2001.
- 17 Mohren, G. M. J., Van den Burg, J. and Burger, F. W.: Phosphorus deficiency induced by
18 nitrogen input in Douglas fir in the Netherlands, *Plant Soil*, 95, 191-200, 1986.
- 19 Murphy, J. and Riley, J. P.: A modified single solution method for the determination of
20 phosphate in natural waters, *Anal. Chim. Acta.*, 27, 31-36, 1962.
- 21 Paré, D. and Bernier, B.: Origin of the phosphorus deficiency observed in declining sugar
22 maple stands in the Quebec Appalachians, *Can. J. For. Res.*, 19, 24-34, 1989.
- 23 Rennenberg, H. and Herschbach, C.: Phosphorus nutrition of woody plants: many questions –
24 few answers, *Plant Biol.*, 15, 785–788, 2013.
- 25 Richter, D., Allen, H. L., Li, J., Markewitz, D. and Raikes, J.: Bioavailability of slowly
26 cycling soil phosphorus: major restructuring of soil P fractions over four decades in an
27 aggrading forest, *Oecologia*, 150, 259-271, 2006.
- 28 Roberts, C. A., Workmann, J. J. and Reeves, J. B. III (eds): *Near-Infrared Spectroscopy in*
29 *Agriculture*, in: *Agronomy Series*, Vol 44, Am. Soc. Agron. Inc, Crop Sci. Soc. Am. Inc, Soil
30 Sci. Soc. Am. Inc, Madison, 729–784, 2004.

- 1 Rowland, A. P. and Haygarth, P. M.: Determination of total dissolved phosphorus in soil
2 solutions, *J. Environ. Qual.*, 26(2), 410-415, 1997.
- 3 Sankey, J. B., Brown, D. J., Bernard, M. L. and Lawrence, R. L.: Comparing local vs global
4 visible and near-infrared (Vis-NIR) diffuse reflectance spectroscopy (DRS) calibrations for
5 the prediction of soil clay, organic C and inorganic C, *Geoderma*, 148, 149–158, 2008.
- 6 Schmidt, W.: *Optische Spektroskopie eine Einführung*, Wiley-VCH Verlag, Weinheim, 2000.
- 7 Shen, J., Yuan, L., Zhang, J., Li, H., Bai, Z., Chen, X., Zhang, W. and Zhang, F.: Phosphorus
8 Dynamics: From Soil to Plant, *Plant Physiol.*, 156, 997-1005, 2011.
- 9 Shenk, J. S. and Westerhaus, M. O.: Near infrared reflectance analysis with single- and
10 multiproduct calibrations, *Crop Sci.*, 33, 582-584, 1993.
- 11 Stenberg, B., Viscarra-Rossel, R. A., Mouazen, A. M. and Wetterlind, J.: Visible and Near
12 Infrared Spectroscopy in Soil Science, in: Sparks, D. L. (ed): *Adv. Agron.*, 107, Burlington:
13 Academic Press, 163-215, 2010.
- 14 Stevenson, F. and Cole, M.: Phosphorus, in: *Cycles of soil: Carbon, nitrogen, phosphorus,*
15 *sulfur, and micronutrients*, John Wiley & Sons, Inc., New York, 2nd edition, 279-329, 1999.
- 16 Stoner, E. R. and Baumgardner, M. F.: Characteristic variations in reflectance of surface soils,
17 *Soil Sci. Soc. Am. J.*, 45, 1161-1165, 1981.
- 18 Tiessen, H. and Moir, J. O.: Characterization of available P by sequential extraction, in:
19 Carter, M. R. and Gregorich, E. G. (eds): *Soil sampling and methods of analysis*, chapter 25,
20 293-306, *Can. Soc. Soil Sci.*, Boca Raton Fl, USA, 2nd edition, 2008.
- 21 Turner, B. L., Mahieu, N. and Condon, L. M.: The phosphorus composition of temperate
22 grassland soils determined by NaOH-EDTA extraction and solution ³¹P NMR spectroscopy,
23 *Org. Geochem.*, 34, 1199-1210, 2003.
- 24 Turner, B. L., Cade-Menun, B. J., Condon, L. M. and Newman, S.: Extraction of soil organic
25 phosphorus, *Talanta*, 66, 294-306, 2005.
- 26 Turner, B. L., Newman, S. and Reddy, K. R.: Overestimation of organic phosphorus in
27 wetland soils by alkaline extraction and molybdate colorimetry, *Environ. Sci. Technol.*, 40,
28 3349-3354, 2006.

- 1 Turner, B. L. Resource partitioning for soil phosphorus: a hypothesis, *J. Ecol.*, 96, 698-702,
2 2008.
- 3 Udelhofer, T., Emmerling, Ch. and Jarmer, T.: Quantitative analysis of soil chemical
4 properties with diffuse reflectance spectrometry and partial least-square regression: feasibility
5 study, *Plant Soil*, 251, 319-329, 2003.
- 6 Vanguelova, E., Pitman, R., Luro, J. and Helmisaari, H.-S.: Long term effects of whole tree
7 harvesting on soil carbon and nutrient sustainability in the UK, *Biogeochem.*, 101, 43-59,
8 2010.
- 9 Vitousek, P. M., Porder, S., Houlton, B. Z. and Chadwick, O. A.: Terrestrial phosphorus
10 limitation: mechanisms, implications, and nitrogen-phosphorus interactions, *Ecol. Appl.*, 20,
11 5-15, 2010.
- 12 Walker, T. W. and Syers, J. K.: The fate of phosphorus during pedogenesis, *Geoderma*, 15: 1-
13 19, 1976.
- 14 Wardle, D.: Aboveground and belowground consequences of long-term forest retrogression in
15 the timeframe of millennia and beyond, in: Wirth, C., Gleixner, G. and Heimann, M. (eds):
16 Old-growth forests: Function, fate and value, *Ecol. Stud.*, 207, 193-209, Springer Verlag
17 Berlin Heidelberg, 2009.
- 18 Weyer, L. G.: Near-Infrared Spectroscopy of Organic Substances. *Appl. Spectrosc. Rev.*,
19 21(1), 1-43, 1985.
- 20 Williams, P. C.: Implementation of near-infrared technology. in: Williams, P. C. and Norris,
21 K. H. (eds): *Near-Infrared Technology in the Agricultural and Food Industries*, Am. Assoc.
22 Cereal Chemist, St. Paul, Minnesota, USA, 145–169, 2001.
- 23 Workman, J. J.: NIR Spectroscopy Calibration Basis, in: Burns, D. A. and Ciurczak, E. W.
24 (eds): *Handbook of Near-Infrared Analysis, Second Edition*, in: *Practical Spectroscopy a*
25 *Series*, Vol 27, Marcel Dekker Inc New York, 2001
- 26 World Agroforestry Centre (ICRAF) and ISRIC-World Soil Information: ICRAF–ISRIC Soil
27 vis–NIR spectral Library, World Agroforestry Centre (ICRAF), Nairobi, Kenya, 2010.
- 28 WRB: IUSS Working Group World Reference Base for Soil Resources 2006, *World Soil*
29 *Resources Report No. 103*, FAO, Rome, 2006.

- 1 Yang, X. and Post, W. M.: Phosphorus transformations as a function of pedogenesis: A
2 synthesis of soil phosphorus data using Hedley fractionation method, *Biogeosciences*, 8,
3 2907-2916, 2011.
- 4 Zornoza, R., Guerrero, C., Mataix-Solera, J., Scow, K. M., Arcenegui, V. and Mataix-
5 Beneyto, J.: Near infrared spectroscopy for determination of various physical, chemical and
6 biochemical properties in Mediterranean soils, *Soil Biol. Biochem.*, 40, 1923-1930, 2008.
7

1 **Table 1.** Descriptive statistical parameters of soil P content (mg kg^{-1}) as total sum of Hedley
 2 P fractions and in pools of different plant availability, grouped by data sets used for NIRS
 3 modelling.

Data Set	P-Pool	n	Min	Mean	Max	SD	Skewness	Kurtosis
BZE all	total sum	282	57.8	564.0	2796.1	413.4	1.9	6.2
	labile		6.0	123.6	850.0	110.7	2.4	8.9
	moderately labile		8.1	296.7	2307.7	266.2	3.2	18.5
	stable		6.7	141.1	805.2	127.4	1.	4.1
BZE “Brown Earths”	total sum	84	128.1	657.7	2178.0	440.59	1.2	1.7
	labile		11.0	166.3	850.0	146.89	2.1	5.6
	moderately labile		46.9	357.3	1227.0	256.9	1.2	1.4
	stable		6.7	134.3	534.3	118.4	1.4	1.9
BEF China	total sum	294	25.7	94.8	328.6	47.9	1.7	4.5
	labile		1.2	17.0	78.9	10.6	1.8	6.5
	moderately labile		10.8	43.7	161.2	21.8	1.7	5.6
	stable		10.8	34.1	174.8	23.1	3.0	11.8

4

5

1 **Table 2.** Descriptive statistical parameters of carbon (g kg⁻¹), nitrogen (g kg⁻¹) and pH (H₂O)
 2 in soil, grouped by data sets used for NIRS modelling.

		n	Min	Mean	Max	SD	Skewness	Kurtosis
BZE	carbon		0.2	42.9	314.7	39.7	2.0	7.6
	nitrogen	282	0.1	2.7	14.2	2.3	1.6	3.3
	pH		3.2	4.5	8.2	0.8	1.9	4.9
BZE	carbon		3.6	56.2	178.0	45.2	0.8	-0.3
„Brown Earths“	nitrogen	84	0.2	3.4	10.0	2.5	0.8	-0.2
	pH		3.4	4.4	7.8	0.7	2.7	11.3
BEF-China	carbon		3.4	30.9	118.1	17.2	1.4	3.0
	nitrogen	294	0.3	1.6	5.9	0.8	1.5	4.5
	pH		3.5	4.5	5.1	0.2	-0.7	1.7

3

4

1 **Table 3.** Quality parameters for NIRS model calibration (R^2 , RPD) for all Hedley fractions in
 2 the three data sets: BZE, BZE “Brown Earths”, and BEF-China.

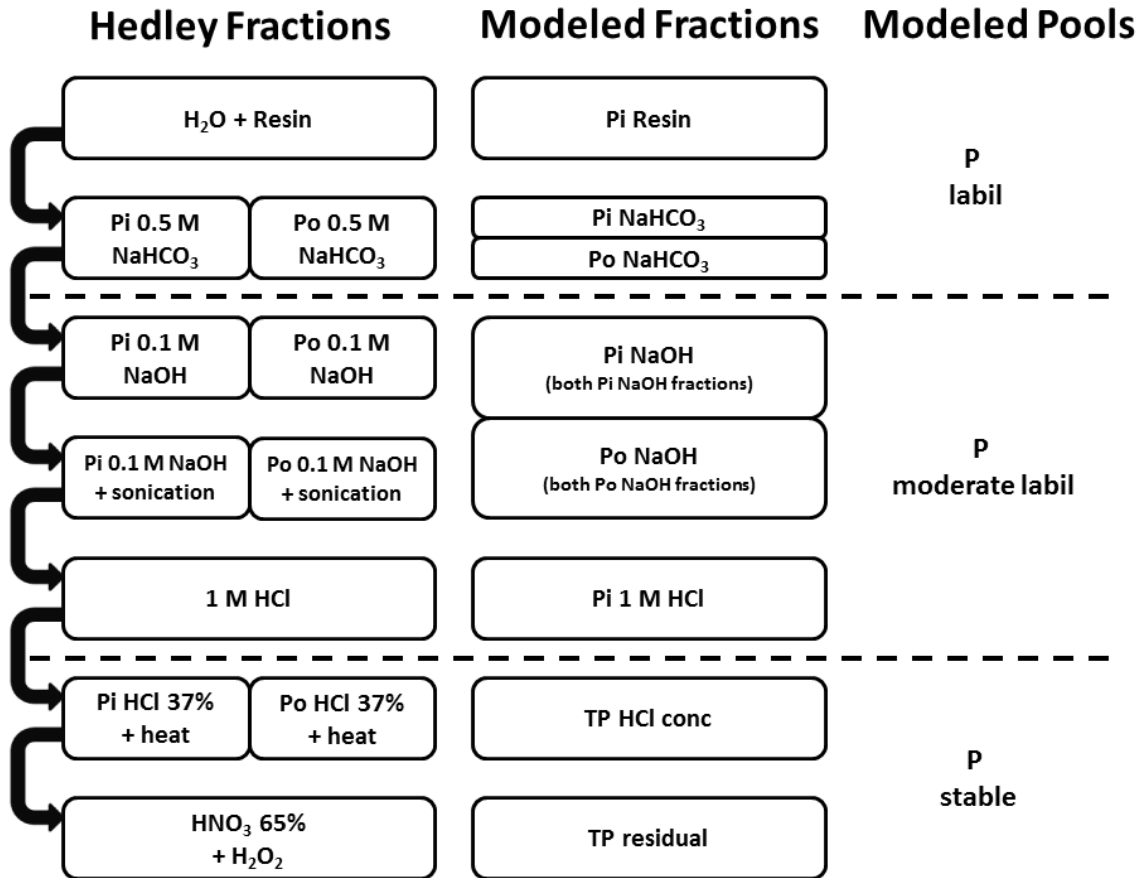
Hedley Fraction	BZE		BZE „Brown Earths“		BEF-China	
	R^2	RPD	R^2	RPD	R^2	RPD
Pi Resin	0.26	1.16	0.51	1.44	0.77	2.08 ^d
Pi NaHCO₃	0.08	1.04	0.19	1.10	0.48	1.39
Po NaHCO₃	0.61	1.60	0.63	1.64	0.78	2.13 ^d
Pi NaOH	0.23	1.14	0.15	1.09	0.72	1.88 ^d
Po NaOH	0.61	1.60	0.75	1.98 ^d	0.82	2.35 ^c
Pi 1M HCl	0.30	1.19	0.38	1.26	0.34	1.23
P HCl conc.	0.72	1.88 ^d	0.76	2.05 ^d	0.85	2.57 ^c
P Residual	0.56	1.50	0.57	1.52	0.69	1.79 ^d

3 ^d) model quality level D; ^c) model quality level C;

4

5

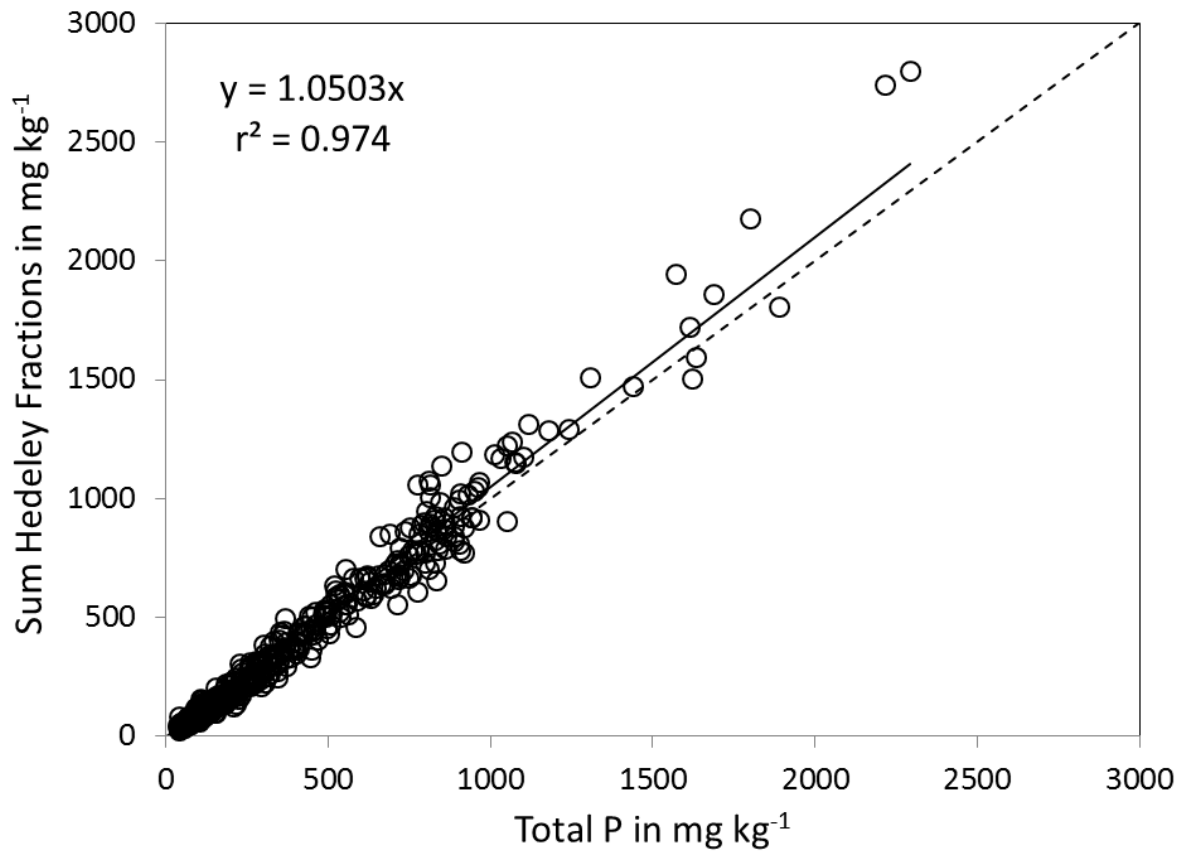
1 **Figure 1.** Sequential extraction scheme of Hedley fractionation modified by Tiessen and
 2 Moir, grouped by pools of availability; NaHCO₃, NaOH and HCl conc. steps provide both
 3 organic and inorganic compounds for this fraction step.



4

5

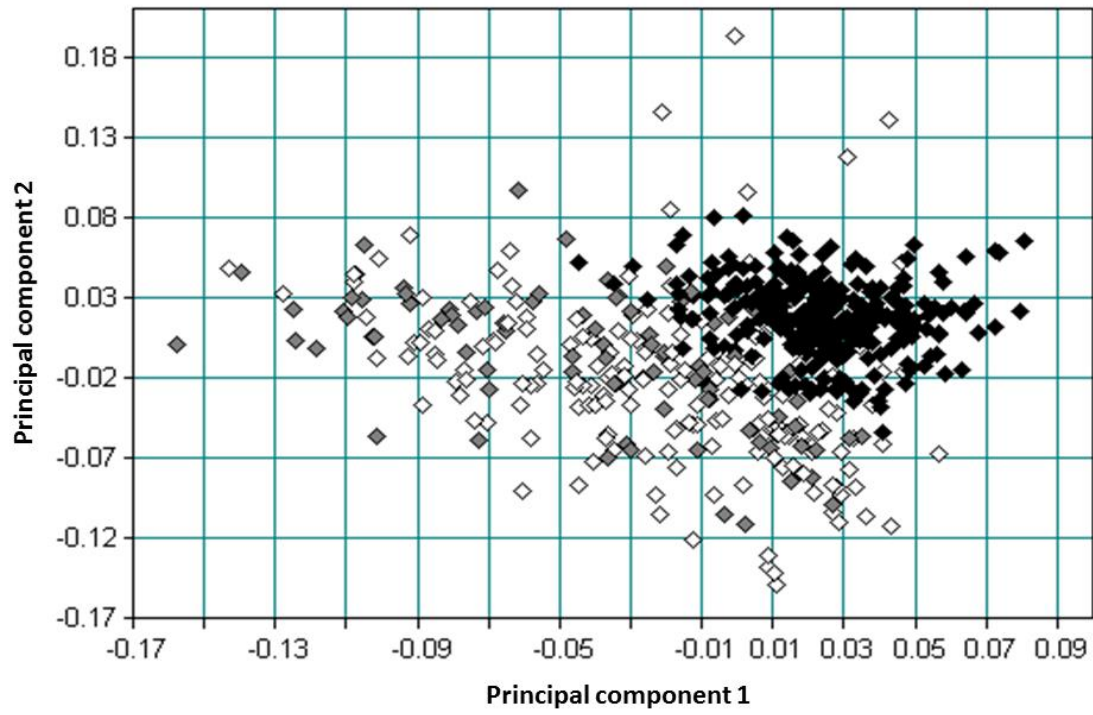
- 1 **Figure 2.** Relationship between the sum of recovered Hedley P fractions and independently
- 2 determined total P (mg kg^{-1}) in soil samples.



3

4

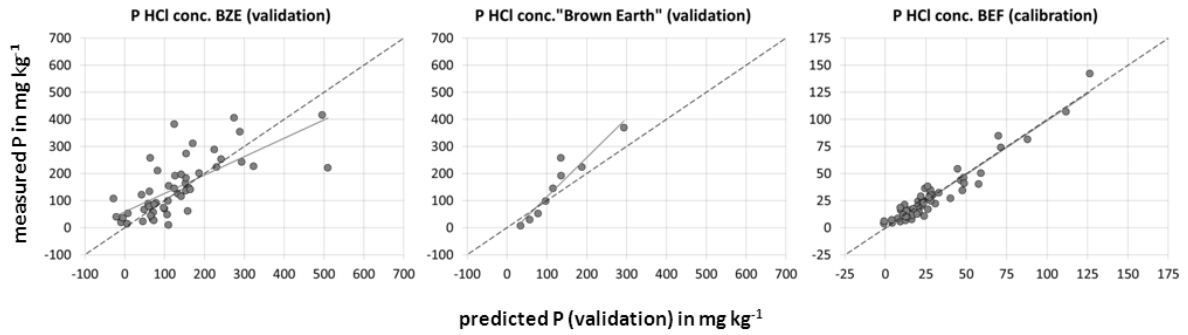
- 1 **Figure 3.** Score plot of principal component analysis (total of five components derived;
- 2 principal component 1 vs. principal component 2) for the three datasets BZE (white, set 2),
- 3 BZE “Brown Earths” (grey, set 3) and BEF China (black, set 4).



4

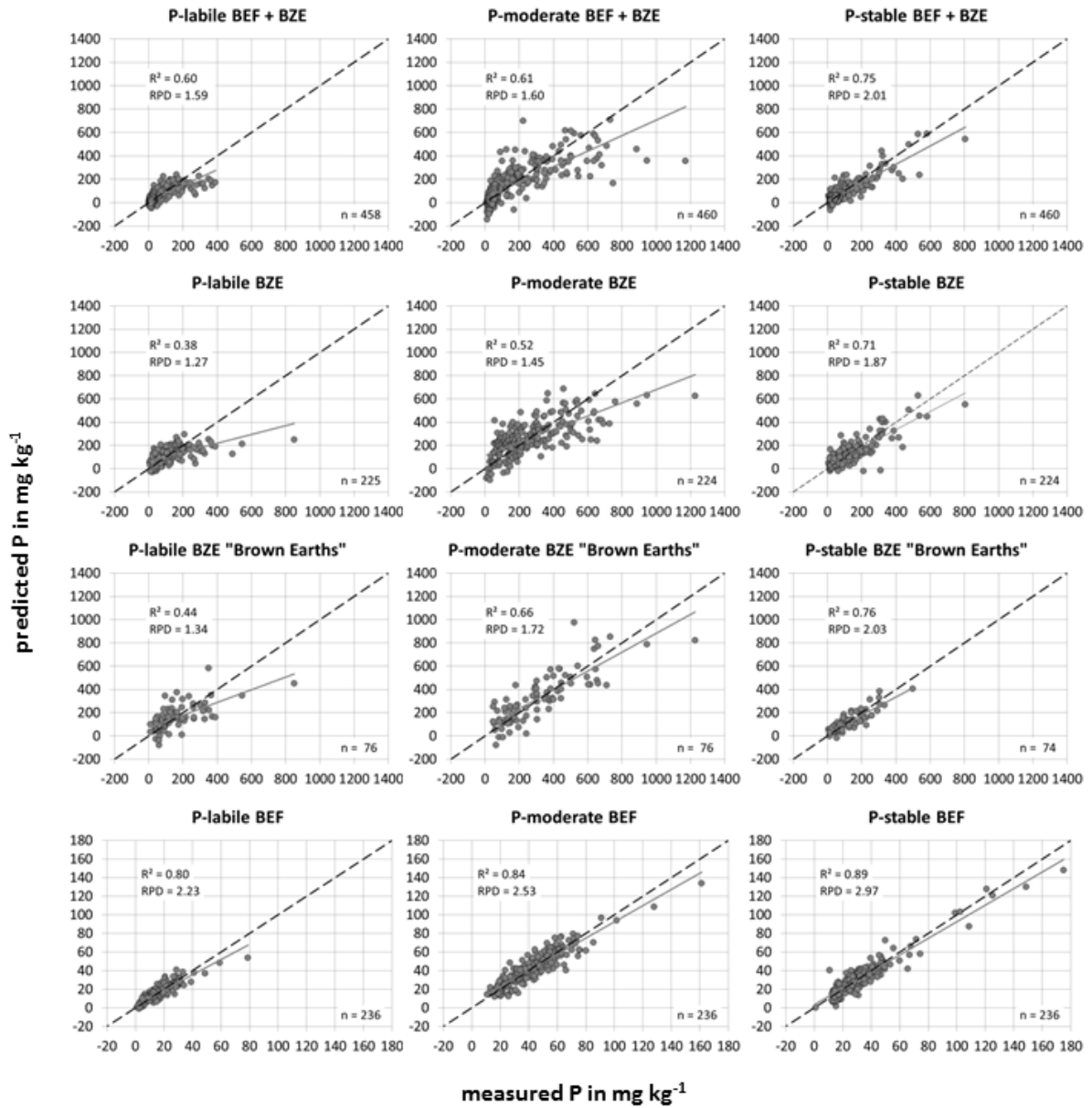
5

- 1 **Figure 4.** Validation for P HCl conc. fraction of the three datasets BZE, BZE “Brown Earths”
- 2 and BEF China (mg kg^{-1}), x-axis measured, y-axis predicted values, please note the difference
- 3 in scale among graphs.



- 4
- 5

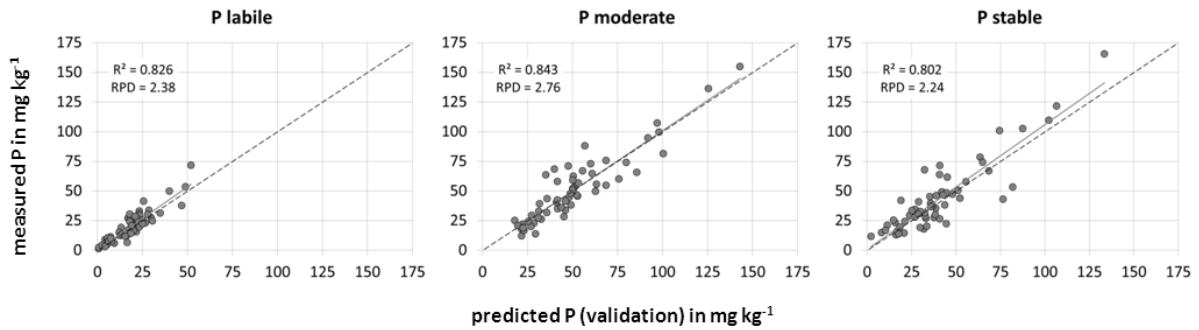
1 **Figure 5.** Calibration models for the four datasets BEF China+BZE, BZE, BZE “Brown
 2 Earths” and BEF China, pooled for labile, moderately labile (mod. labile) and stable
 3 phosphorus (mg kg^{-1}), x-axis measured, y-axis predicted values, please note the difference in
 4 scales.



5

6

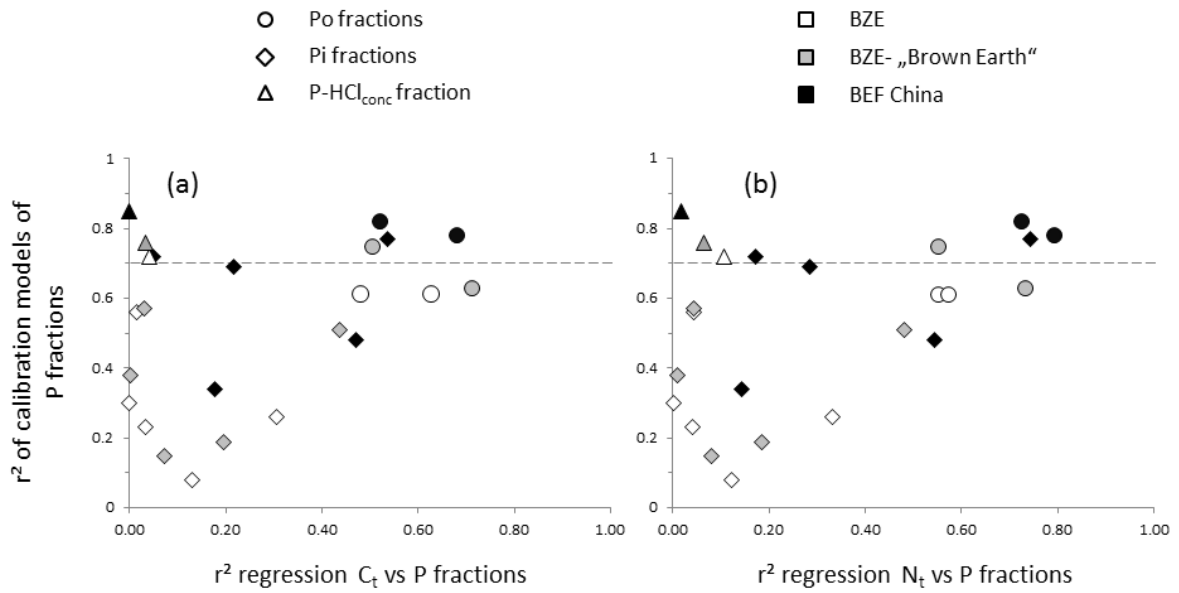
1 **Figure 6.** Validation for BEF China calibration models with external datasets pooled for
2 labile moderately labile (mod. labile) and stable phosphorus (mg kg^{-1}), x-axis measured, y-
3 axis predicted values.



4

5

1 **Figure 7.** Goodness of fit of calibration models (r^2) for all individual P fractions and
 2 coefficient of determination (r^2) of regression analysis of total organic carbon **(a)**, total
 3 nitrogen **(b)** and the P fractions (circles=organic fractions; squares=inorganic fractions;
 4 triangles= HCl_{conc} fraction) in soils for all three datasets (black=BEF China; grey=BZE
 5 “Brown Earths”; white=BZE), dashed line indicates the minimum model quality of $r^2 = 0.7$.



6