Responses to specific comments of Editor

I went through the manuscript again. The reviewer's comments and edits have been incorporated. I did realize however, that you use PSU as salinity unit. In a strict sense salinity is defined as a ratio and hence without a unit. Please delete 'PSU' throughout the text. Also, it is the rule in Biogeosciences that DOC concentrations are given in micromol L-1. Hence, please convert your mg/l to micromol/l throughout the text, tables and in the figures. Subsequently, please upload the revised manuscript again. **Response:** Thank you for your suggestion. We have deleted 'PSU' through the test in the revised manuscript. The DOC concentrations have converted from 'mg L⁻¹' to ' μ mol L⁻¹' in the revised manuscript.

1	Seasonal characterization of CDOM for lakes in semi-arid regions of
2	Northeast China using excitation-emission matrices fluorescence and
3	parallel factor analysis (EEM-PARAFAC)
4	Ying Zhao ¹ , Kaishan Song ^{1*} , Zhidan Wen ¹ , Lin Li ² , Shuying Zang ³ , Tiantian Shao ¹ ,
5	Sijia Li^1 , Jia Du^1
6 7	¹ Northeast Institute of Geography and Agroecology, Chinese Academy of Sciences, Changchun, Jilin, 130102, China
8 9	² Department of Earth Sciences, Indiana University-Purdue University, Indianapolis, IN, USA
10	³ College of Geographical Science, Harbin Normal University, Harbin, China
11	*corresponding author E-mail: <u>songks@iga.ac.cn</u> ; Tel: 86-0431-85542364
12	
13	Abstract. The seasonal characteristics of fluorescent components in CDOM for lakes
14	in the semi-arid region of Northeast China were examined by excitation-emission
15	matrix (EEM) spectra and parallel factor analysis (PARAFAC). Two humic-like (C1
16	and C2) and the protein-like (C3 and C4) components were identified using
17	PARAFAC. The average fluorescence intensity of the four components differed under
18	seasonal variation from June and August 2013 to February and April 2014.
19	Components 1 and 2 exhibited a strong linear correlation ($R^2 = 0.633$). Significantly
20	positive linear relationships between CDOM absorption coefficients $a(254)$ (R ² =0.72)
21	0.46, $p < 0.01$), a(280) (R ² = 0.77, 0.47, $p < 0.01$), a(350) (R ² = 0.76, 0.78, $p < 0.01$)
22	and F_{max} for two humic-like components (C1 and C2) were exhibited, respectively. A

23	significant relationship $(R^2 = 0.931)$ was found between salinity and DOC. However,
24	almost no obvious correlation was found between salinity and EEM-PARAFAC
25	extracted components except for C3 ($R^2 = 0.469$). Results from this investigation
26	demonstrate that the EEM-PARAFAC technique can be used to evaluate the seasonal
27	dynamics of CDOM fluorescent components for inland waters in the semi-arid
28	regions of Northeast China, and to quantify CDOM components for other waters with
29	similar environmental conditions.

- 30 Keywords: CDOM, fluorescent components, EEMs, PARAFAC, DOC, Salinity
- 31 32

33 **1 Introduction**

Dissolved organic matter (DOM), a heterogeneous mixture of humic acids, 34 35 proteins and carbohydrates, plays important roles in aquatic ecosystems (Zhang et al., 2010). Chromophoric dissolved organic matter (CDOM), the colored fraction of 36 DOM, absorbs light energy in the ultraviolet (UV) and visible region of the spectrum 37 and inhibits the propagation of UV radiation. CDOM in waters also affects the 38 transport and bio-availability of materials such as trace metals and other pollutants 39 (Song et al., 2013), so it can be used as a proxy of water quality. In natural water 40 bodies, CDOM originates from the degradation of plant materials and other organisms 41 and terrestrially imported substances, which varies in time and space and is controlled 42 by its structure and composition (Stedmon et al., 2003). CDOM is compositionally 43 complex, making it difficult to isolate hydrophobic from hydrophilic acids using XAD 44 ion-exchange resins (Aiken et al., 1992; Spencer et al., 2010). Nonetheless, some 45

optically active components of CDOM can emit fluorescence after absorbing light at certain wavelengths (Zhang et al., 2010) so that the fluorescence spectroscopic techniques can be used to provide detailed information about the source and concentration of CDOM. The traditional fluorescence techniques including fluorescence emission spectrometry and synchronous fluorescence scanning applied to examine CDOM components have the drawback that the output was restricted to a linear scan (Hudson et al., 2007).

53 Recently, excitation-emission matrix fluorescence spectroscopy (EEM) has been 54 applied to identify CDOM components because of its ability to produce synchronous scan spectra in the form of contours (Stedmon et al., 2003; Zhang et al., 2010). The 55 EEM spectroscopy is considered the most effective technique for studying the 56 57 composition of fluorophores given its high selectivity and sensitivity to CDOM in water columns (Zhang et al., 2010). In recent years, EEM spectroscopy has been 58 widely used to investigate the dynamics of marine, freshwater and ice-water 59 ecosystems as well as snow melting water (Barker et al., 2006, 2009, 2010, 2013; 60 Coble, 2007; Fellmanet al., 2010; Guo et al., 2010; Hudson et al., 2007; Stedmon et 61 al., 2007). Moreover, the EEM spectroscopy can also be used to distinguish 62 allochthonous and autochthonous CDOM sources in aquatic environments (Coble et 63 al., 1998; Mayer et al., 1999; Yamashita et al., 2008, 2010; Zhang et al., 2013). Based 64 on the peak positions in EEMs, two main fluorescent components, i.e., humic-like and 65 protein-like substances, have been identified and investigated (DelCastillo et al., 1999; 66 Jaffe' et al., 2004). However, overlapped fluorophores of CDOM EEMs could make 67

this traditional 'peak-picking' method unreliable to evaluate CDOM dynamics in 68 aquatic ecosystems (Coble, 1996; Stedmon et al., 2003). Recently, the combined 69 70 EEMs-PARAFAC (parallel factor analysis) technique has been shown to effectively decompose EEM of CDOM into independent fluorescent components and assess the 71 72 source of CDOM and relationships with other water quality parameters. A number of investigators have used EEMs-PARAFAC to characterize DOM in freshwater and 73 marine aquatic environments (Broisover et al., 2009; Cory et al., 2005; Guo et al., 74 2010; Stedmon et al., 2003; Stedmon and Markager, 2005; Yamashita, 2008; Zhang et 75 76 al., 2010, 2011, 2013). Stedmon et al. (2003) introduced PARAFAC and identified five distinct DOM components for a Danish estuary and its catchment. In coastal 77 environments, Yamashita et al. (2008) reported on seven components using the 78 79 combined EEMs-PARAFAC technique and assessed the dynamic of individual fluorophores and their relationship with salinity in Ise Bay. Zhang et al. (2011) also 80 found three different components by PARAFAC modeling and analyzed the 81 82 correlations between the fluorescent components and absorption coefficients of CDOM for Lake Tianmu and its catchment. 83

The Songnen Plain is a fluvial plain with semi-arid climate, in which many fresh and brackish waters are distributed according to its geomorphological characteristics (Song et al., 2013). Dissolved organic carbon (DOC) characterisitics of these fresh and brackish waters across the Songnen Plain have been studied by Song et al. (2013); the results indicated that a huge amount of DOC was stored in these waters. In particular, brackish waters would exhibit high average DOC concentrations and

90 significantly contributed the carbon budget to inland waters (Duarte et al., 2008; Song et al., 2013; Tranvik et al., 2009). However, little study has been made on the detailed 91 92 information of DOC sources for these waters in the Songnen Plain. Therefore, it motivated us to investigate the components in CDOM for both fresh and brackish 93 94 waters in the semi-arid region. In the present study, the absorption and fluorescence of CDOM were determined for the water samples collected from seven lakes in the 95 western part of Jilin province, which varied at different seasons. The specific 96 objectives of this study are to: 1) characterize CDOM components contained in these 97 98 lakes using EEMs and their origins through the EEM-PARAFAC method; 2) assess 99 the dynamic of individual fluorescent components of CDOM under seasonal variation; and most importantly 3) link CDOM fluorescence intensities, absorption coefficients, 100 101 DOC concentrations and salinity to each other, in order to establish proxies for CDOM bioavailability and photoreactivity in waters. 102

103

104 **2 Materials and Methods**

105 **2.1 Lakes and water sampling**

The water bodies investigated in this study were located in the western part of Jilin Province, which belongs to the semi-arid part of the Songnen Plain (Song et al., 2013). Two groups of lakes were investigated, i.e., the Chagan lake group and the Yuelianghu lake group. The Chagan lake group is made up of Lake Chagan (CGL), Xinmiaopao (XMP), Xindianpao (XDP) and Kulipao (KLP). The Yuelianghu lake group mainly includes Lake Yueliang (YLL), Talahong (TLH) and Xinhuangpao (XHP) (Fig. 1).

The two groups are about 60 km away from each other, of which each includes both 112 fresh and brackish waters. The primary economic value for these lakes is fisheries, 113 114 agricultural irrigation and recreation. The average annual precipitation is about 391 mm, but the average evaporation is up to 1790 mm, resulting in water scarcity. Due to 115 the area dominated by saline-alkali soil, the rainfall flush and agricultural catchment 116 land use can result in an increase of lake salinities. These seven lakes are endowed 117 with similar geological, hydrological and climatic settings, thus we presume that 118 similar processes may control the CDOM components. In order to characterize the 119 120 CDOM fluorescent components under seasonal variation using EEMs-PARAFAC, 67 water samples were collected from the surface of the seven lakes in 1-liter 121 acid-cleaned plastic bottles during four field campaigns in June and August 2013 as 122 123 well as in February and April 2014, respectively. These samples were collected during the ice covering period using an ice drilling auger. The under-ice surface water was 124 coming up when a hole was drilled in the ice layer by the auger. The ice shavings 125 were collected in plastic bags and the under-ice surface water was collected in plastic 126 bottles. The collected samples were held on ice and immediately transported to the 127 laboratory in the Changchun City of Jilin province within 3-5 hours. In the laboratory, 128 these samples were filtered within 24 h and then kept at 4 $^{\circ}$ C until analysis within two 129 days. Latitude and longitude of each sample location were recorded in situ using a 130 Trimble Global Positioning System (GPS). 131

132

Figure 1. Locations of the water sampling sites for 7 lakes in the western part of Jilin
province, Northeast China. a) Yueliang lake group: YLL, Yueliang Lake; XHP;

135 Xinhuangpao; TLH; Talahong; b) Chagan lake group: CGL, Chagan Lake; XDP,
136 Xindianpao; XMP, Xinmiaopao; KLP, Kulipao.

137

138 **2.2 Analytical procedures**

To characterize the basic parameters of water quality, salinity was measured through a 139 DDS-307 electrical conductivity (EC) meter in the laboratory. Salinity was expressed 140 141 in the basis of the UNESCO practical salinity unit (PSU 1978). The pH was measured using a PHS-3C pH meter at room temperature $(20 \pm 2 \degree C)$ in the laboratory. Water 142 turbidity was determined using the Shimadzu UV-2600PC UV-Vis dual beam 143 spectrophotometer with matching 3 cm quartz cells at room temperature (20 \pm 2 °C) 144 with Milli-Q water the reference (UV talk letter 145 as vol. 10, https://shimadzu.com.au/uv-talk-letter-volume-10). To determine DOC concentrations, 146 147 water samples were filtered through 0.45 µm filters and then measured using a Shimadzu TOC-5000 Analyzer and a 1.2 % Pt on silica catalyst at 680 °C. Potassium 148 hydrogen phthalate was used as a standard. The reproducibility of the analytical 149 150 procedure was within 2-3 % for the current study (APHA, 1998; Song et al., 2011).

151

152 **2.3 Absorption measurement**

In the laboratory, all the samples were filtered at low pressure, first through a pre-combusted Whatman GF/F filter (0.7 μ m), and then through a pre-rinsed 25 mm Millipore membrane cellulose filter (0.22 μ m) into glass bottles. Absorption spectra of the samples were measured between 200 and 800 nm at 1 nm increments using the Shimadzu UV-2600PC UV-Vis dual beam spectrophotometer with a 1 cm quartz 158 cuvette and Milli-Q water as reference. The absorption coefficient a_{CDOM} was 159 calculated from the measured optical density (OD) of the sample using Eq. (1):

160
$$a_{CDOM}(\lambda) = 2.303 \left[OD_{(\lambda)} - OD_{(null)} \right] / \gamma \quad (1)$$

where γ is the cuvette path length (0.01 m) and the factor 2.303 converts from base 10 to base natural logarithm transformation. Some fine particles possibly remained in the filtered solution (Babin et al., 2003; Bricaud et al., 1995), therefore it was necessary to correct for scattering by fine particles and in this case, $OD_{(null)}$ is the average optical density over 740-750 nm where the absorbance of CDOM can be assumed to be zero.

166 A CDOM absorption spectrum $(a_{CDOM}(\lambda))$ can be expressed as an exponential 167 function (Babin et al., 2003; Bricaud et al., 1995):

168
$$a_{CDOM}(\lambda_i) = a_{CDOM}(\lambda_r) \exp[-S(\lambda_i - \lambda_r)]$$
(2)

where $a_{CDOM}(\lambda_i)$ is the CDOM absorption at a given wavelength λ_i , $a_{CDOM}(\lambda_r)$ is the absorption estimate at the reference wavelength λ_r (440 nm), and *S* is the spectral slope of the CDOM absorption. According to Helms et al. (2008), *S* is calculated by fitting a linear model to the data over a wavelength range of 275 to 295 nm (*S1*) or 350 to 400 nm (*S2*). To eliminate the inter-laboratory variability, the slope ratio S_R = *S1/S2* is defined to indicate the molecular weight and photo-bleaching of CDOM (Helms et al., 2008; Zhang et al., 2010).

176

177 **2.4 Three-dimensional fluorescence measurement**

The EEMs analysis of CDOM were conducted using a Hitachi F-7000 fluorescence
spectrometer (Hitachi High-Technologies, Tokyo, Japan) with a 700-voltage xenon

180 lamp. The scanning ranges were 200–450 nm for excitation, and 250–500 nm for 181 emission. Readings were collected in the ratio mode at 5 nm intervals for excitation, 182 and at 1 nm intervals for emission, using a scanning speed of 2400 nm min⁻¹. The 183 band-passes were 5 nm for both excitation and emission. A Milli-Q water blank of the 184 EEMs was subtracted to eliminate the water Raman scatter peaks (McKnight et al., 185 2001; Stemdon et al., 2003; Zhang et al., 2010, 2011).

The inner-filter effect, which results from reabsorption and excitation of the fluorescence itself, can reduce the fluorescence intensity by 5% (Larsson et al., 2007; McKnight et al., 2001). In order to eliminate the inner-filter effect, the EEMs were corrected for absorbance by multiplying each value in the EEMs with a correction factor based on the assumption that the average path length of absorption of the excitation and emission light is one-half length of the cuvette (McKnight et al., 2001; Zhang et al., 2010). The correction function is expressed as follows:

193

$$F_{corr} = F_{obs} \times 10^{(A_{ex} + A_{em})/2}$$
(3)

194

where F_{Corr} and F_{obs} are the corrected and uncorrected fluorescence intensities and A_{ex} and A_{em} are the absorbance values at the respective excitation and emission wavelengths.

The measured fluorescence intensity is dependent on the concentration of the dissolved fluorophores in water bodies. Finally, the fluorescence intensities of all samples' EEMs were normalized to the area under the Milli-Q water Raman peak $(\lambda ex=350 \text{ nm}, \lambda em=371-428 \text{ nm})$ measured daily (Lawaetz and Stedmon, 2009). The 202 contour figures of the EEMs were plotted using the Matlab 10.0 software package203 (Math Works, Natick Massachusetts, America).

204

205 2.5 The PARAFAC modeling

206 PARAFAC, a three-way method, is applied to decompose the CDOM fluorescence into separate fluorescent signals (Andersen and Bro, 2003; Stedmon and Bro, 2008). 207 According to Stedmon and Bro (2008), a similar PARAFAC analysis is carried out in 208 the present study using the DOMFluor toolbox in MATLAB with the "N-way toolbox 209 for MATLAB" (Andersson et al., 2000). Before PARAFAC modeling, the excitation 210 wavelengths from 200 to 220 nm and the emission wavelengths from 250 to 300 nm 211 were deleted because of their poor quality. In order to remove the effect of Rayleigh 212 213 scatter on PARAFAC modeling, the missing values (NaN-Not a number) were inserted in the regions (Ex-20 \leq Em \leq Ex+20 and 2Ex-20 \leq Em \leq 2Ex+20; unit: nm) 214 which are significantly influenced by the first and second order scattering from the 215 216 measured spectroscopic data (Hua et al., 2007; Stedmon and Bro, 2008).

To determine the appropriate number of PARAFAC components, the split-half validation procedure was executed to verify whether the model was valid by comparing the emission and excitation loadings from each half (Stedmon and Bro, 2008). Split-half analysis is the most effective method for implementing the PARAFAC models, in which the EEMs are randomly divided into four groups of equal size, and then analyzed for two half splits (1-2 and 3-4 half) respectively. If the correct number of components is chosen, the excitation and emission loadings from the two groups should show the same shape and size (Bro, 1997, 1999). The fluorescence intensity of every component was represented by F_{max} (Raman unit: nm⁻¹) (Stedmon and Markager, 2005).

227

228 **2.6 Statistical analysis**

Statistical analysis was conducted using the SPSS 16.0 software package (Statistical Program for Social Sciences). Regression and correlation analysis was used to describe the relationship between CDOM absorption coefficient, DOC concentration, salinity and F_{max} . A model II-ANOVA was performed to determine seasonal variability is higher than between-lake variability. The difference is considered to be statistically significant when *p*-values are less than or equal to 0.05.

235

3 Results and discussion

237 **3.1 Water quality conditions**

The water quality parameters, i.e., pH, salinity, turbidity for the 67 water samples 238 collected from June 2013 to April 2014 in the western part of Jilin province are 239 displayed in Table 1. When the set of samples from various field trips was pooled 240 together, the waters had high pH values and high salt contents. The highest salinity 241 was present when the lakes were frozen in February 2014, whereas relatively constant 242 values (around 0.40 PSU) were exhibited in the other three seasons. Also the water 243 bodies were highly turbid. The highest turbidity was present in June 2013, and then 244 reduced in August 2013, and the lowest value was recorded in February 2014. 245

Compared with February 2014, the turbidity had almost no change in April 2014(Table 1).

248

Table 1. Mean value of water quality parameters from June 2013 to April 2014.

- 250
- **3.2 EEMs characterization of CDOM**

Based on the EEMs 'peak picking' technique, the key fluorescence peaks can be 252 observed in 67 water samples: two humic-like and two protein-like substances (Coble, 253 1996; Stedmon et al., 2003). The humic-like components are the mixture of aromatic 254 and aliphatic compounds-humic-like acids from terrestrial substances, and aquatic 255 humic-like substances of phytoplankton origin. With respect to the protein-like 256 components (i.e., tyrosine-like and tryptophan-like substance), they mainly consist of 257 dissolved amino acids. As an example, Fig. S1 displays the EEMs of samples from 258 259 lake Xindianpao at different seasons. The peaks comprise two humic-like fluorescence peaks: one in the ultraviolet range (Ex/Em = 220-240/410-430 nm) and 260 the other in the visible range (Ex/Em = 300-340/410-450 nm) and the protein-like 261 fluorescence peaks: tyrosine-like (Ex/Em = 210-230, 270-280/310-330 nm) and 262 tryptophan-like (Ex/Em = 220-230, 280-300/350-370 nm). 263

264

In our study, four separate fluorescent components (Fig. 2a-d) and the excitation and emission loadings (e-h) of the four components identified by EEM-PARAFAC are summarized in Fig. 2 and Table 2. The first fluorescent

268	component (C1) was a biological degradation humic-like component comparable to
269	humic-like peaks (M and N) in marine and in phytoplankton degradation experiments
270	for inland waters (Coble, 1996; Zhang et al., 2009). Component 2 was consistent with
271	the humic-like peaks (A and C) defined by Coble (1996). Component 3 resembles the
272	tryptophan-like (T) component as found by Baker et al. (2004) and Hudson et al.
273	(2007). For component 4, it is likely related to tyrosine-like component (B) (Hudson
274	et al., 2007). Components 3 and 4 represent autochthonous semi-labile CDOM
275	associated with bacteria activity and phytoplankton degradation (Borisover et al.,
276	2009; Stedmon et al., 2003). Particularly, there was a shoulder at the excitation
277	wavelength 310-330 nm in component 3 and 330-340 nm in component 4, which may
278	be due to the residual Raman peaks in some water samples (Fig. 2c-d). In this study,
279	not all of the four components were present in all of the samples.

Figure 2. The PARAFAC modeling output shows the contour plots of the four PARAFAC fluorescent components (a-d) and excitation (black) and emission (red) loadings (e-h) of each component. Fluorescence is in Raman units: nm⁻¹.

284

Table 2. Positions of the fluorescence maximum peaks of the four components identified by
PARAFAC modeling in the present study compared with those previously identified.
Secondary excitation maxim is given in brackets.

288

289 **3.3 Temporal distribution of PARAFAC components**

290 These fresh and brackish water in Jilin province in northeast China are endowed with 291 similar geological, hydrological and climatic settings, thus it is presumed that similar

292	processes may control the CDOM components. When a model II-ANOVA using
293	season and lake as random effect factors was performed, it shows that the seasonal
294	variability ($F > F_{crit}$, $p < 0.05$) is higher than between-lake variability. Therefore, the
295	water samples from different lakes for every season were pooled together in order to
296	study the seasonal variation of the fluorescent components. As shown in Fig. 3a, the
297	average fluorescence intensity of the four components had seasonal variation. When
298	all the water samples at different seasons were pooled together, the average value of
299	total fluorescence intensity was 2.05 \pm 0.93 nm ⁻¹ , corresponding to the intensities of
300	0.71 ± 0.32 (C1), 0.33 ± 0.11 (C2), 0.50 ± 0.24 (C3), and 0.51 ± 0.26 (C4) nm ⁻¹ for
301	different components. These results can demonstrate that the fluorescence intensity
302	was dominated by C1, implying most of the CDOM for the seven inland lakes
303	originated from the degradation of phytoplankton and microorganisms. The
303 304	originated from the degradation of phytoplankton and microorganisms. The protein-like components (C3 and C4), related to bioavailability and microbial activity
304	protein-like components (C3 and C4), related to bioavailability and microbial activity
304 305	protein-like components (C3 and C4), related to bioavailability and microbial activity of CDOM, had almost the same magnitude. At all four seasons, the fluorescent
304 305 306	protein-like components (C3 and C4), related to bioavailability and microbial activity of CDOM, had almost the same magnitude. At all four seasons, the fluorescent component C2, which was terrestrially imported to water bodies, contributed less to
304305306307	protein-like components (C3 and C4), related to bioavailability and microbial activity of CDOM, had almost the same magnitude. At all four seasons, the fluorescent component C2, which was terrestrially imported to water bodies, contributed less to total fluorescence than the other three. The total fluorescence intensity differed under
 304 305 306 307 308 	protein-like components (C3 and C4), related to bioavailability and microbial activity of CDOM, had almost the same magnitude. At all four seasons, the fluorescent component C2, which was terrestrially imported to water bodies, contributed less to total fluorescence than the other three. The total fluorescence intensity differed under seasonal variation, varying from 2.54 ± 0.68 nm ⁻¹ in June to 1.93 ± 0.70 nm ⁻¹ in
 304 305 306 307 308 309 	protein-like components (C3 and C4), related to bioavailability and microbial activity of CDOM, had almost the same magnitude. At all four seasons, the fluorescent component C2, which was terrestrially imported to water bodies, contributed less to total fluorescence than the other three. The total fluorescence intensity differed under seasonal variation, varying from $2.54 \pm 0.68 \text{ nm}^{-1}$ in June to $1.93 \pm 0.70 \text{ nm}^{-1}$ in August 2013, and then increased to $2.34 \pm 0.92 \text{ nm}^{-1}$ in February and reduced to the
 304 305 306 307 308 309 310 	protein-like components (C3 and C4), related to bioavailability and microbial activity of CDOM, had almost the same magnitude. At all four seasons, the fluorescent component C2, which was terrestrially imported to water bodies, contributed less to total fluorescence than the other three. The total fluorescence intensity differed under seasonal variation, varying from $2.54 \pm 0.68 \text{ nm}^{-1}$ in June to $1.93 \pm 0.70 \text{ nm}^{-1}$ in August 2013, and then increased to $2.34 \pm 0.92 \text{ nm}^{-1}$ in February and reduced to the lowest $1.57 \pm 0.55 \text{ nm}^{-1}$ in April 2014 (Fig. 3c). The intensities of four fluorescent

314	average except C2 ($0.32 \pm 0.06 \text{ nm}^{-1}$). This can be explained by enhanced activities
315	from plant degradation and microbial activities, but less terrestrial substances were
316	imported to the water bodies in June and therefore the fluorescence intensity of C2
317	was lower than the seasonal average. Compared to the fluorescence intensity in June,
318	the three fluorescence intensities $(0.65 \pm 0.14 \text{ (C1)}, 0.33 \pm 0.16 \text{ (C3)}, 0.52 \pm 0.36 \text{ (C4)})$
319	nm ⁻¹) from the samples collected in August 2013 were reduced, but an increased value
320	was recorded for C2 (0.42 \pm 0.05 nm ⁻¹) (Fig. 3d). Especially, the fluorescence
321	intensities of two protein-like components showed an obvious difference. This can be
322	attributed to substantially increased precipitation up to 180 mm in July from June to
323	August 2013 (Fig. 3b) so that floods occurred when rainfall continued to increase in
324	August. Gradually, DOM contained in terrestrial CDOM was flushed by rainfall to the
325	lakes so that the C2 (0.42 \pm 0.05 nm ⁻¹) fluorescence intensity became higher. In
326	accordance with Cheng et al. 2010, the rainwater CDOM for this study was largely
327	characterized by protein-like components (Cheng et al., 2010). The fluorescence
328	intensity of the rainwater CDOM was very weak, and also the rainwater CDOM
329	contained much lower humic-like concentration (Fig. S2b). The intensities of the
330	other three components decreased because of dilution resulting from heavy rain and
331	relatively weak microbial decomposition of plants.

The highest C1 ($1.02 \pm 0.38 \text{ nm}^{-1}$) presented in February 2014, and the C2 ($0.39 \pm 0.12 \text{ nm}^{-1}$) intensity remained almost the same as that in August 2013. However, the protein-like components indicated that the C3 ($0.57 \pm 0.25 \text{ nm}^{-1}$) intensity was higher than the C4 ($0.35 \pm 0.17 \text{ nm}^{-1}$) intensity, which was opposite to the results from other

months (Fig. 3d). In cold winter, the surface waters formed a thick layer of ice 336 covering the lake waters. Because the ice cover reduced light penetration and 337 restricted gas exchange between the underlying water and atmosphere, vigorous 338 biological activity in the lakes would be reduced at low temperature and low light 339 level (Thomas K., 1983; Uusikiv et al., 2010; Wharton, et al., 1993). Although the 340 biological activity was very weak, there could still be a bit of production of C1 and 341 C3 in lake water. Also, dissolved materials were left in the underlying surface waters 342 and little terrestrial matter was imported to the lakes once covered by ice (Stedmon et 343 344 al., 2007). Therefore, the C1 and C3 in the water of the lakes beneath the ice layers would be produced and accumulated simultaneously, whereas, the C2 remained the 345 same. Obviously, the fluorescence intensity of component 1 reached the highest value 346 for the winter samples. As shown in Fig. S2a, another striking feature for the winter 347 samples was that the fluorescence of CDOM in the ice was dominated by the 348 tyrosine-like C4 component, which is consistent with the findings of Barker et al. 349 (2009, 2013) and Stedmon et al. (2007). It showed that the C4 component was left in 350 the ice-cover when the lakes were frozen. Therefore, it is not surprising that the 351 intensity of component C4 for water beneath ice layers was reduced and the 352 concentrated C3 showed a much higher fluorescence intensity. In April 2014, the 353 intensities of four fluorescent components (0.47 \pm 0.17 (C1), 0.25 \pm 0.08 (C2), 0.40 \pm 354 0.16 (C3), and 0.45 \pm 0.13 (C4) nm⁻¹) (Fig. 3d) exhibited similar seasonal trends 355 though these values were much lower than the average. Our interpretation is that the 356 ice CDOM was characterized by tyrosine-like component (C4) (Fig. S2a), and the 357

fluorescence intensity of C4 contributed by the ice-melt water was very weak. However, the underlying lake CDOM included both humic-like (C1 and C2) and protein-like (C3 or C4) components. When the ice in the lakes melted into water with warming weather and biological degradation and human activity was weak, the lake CDOM was diluted by the ice-melted water and the fluorescence intensity would reach to the lowest value in early spring.

364

Figure 3. a) Seasonal average of F_{max} for EEM-PARAFAC components (C1, C2, C3 and C4) for lakes in the western part of Jilin province; b) Monthly variation of rainfall for the lakes in western part of Jilin province from April 2013 to February 2014; c) Seasonal variation of the total fluorescence intensity at different seasons; d) Seasonal variation of the four EEM-PARAFAC components at different seasons. The error bars represent one standard deviation.

371

372 **3.4 CDOM versus EEM-PARAFAC extracted components**

The concentration of DOC, CDOM absorption coefficients and the slope ratio S_R are 373 shown in Table 3. The DOC concentrations ranged from $\frac{10.03}{10.03}$ to $\frac{56.60 \text{ mg L}^{-1}}{10.03}$ with 374 an average value of $\frac{37.60 \pm 18.05 \text{ mg } \text{L}^{-\frac{1}{2}}}{\text{during the period from June 2013 to April$ 375 2014, demonstrating a seasonal dynamics that can be attributed to hydrological, 376 climatic and landscape variations (Song et al., 2013). The highest average DOC 377 concentration $(55.04 \pm 20.00 \text{ mg L}^{-1})$ was present in February 2014 (ice-covered 378 period); whereas, relatively constant values of approximate 30 mg L⁻¹ were observed 379 in the ice-free season. The relative high DOC concentration in ice-free season was 380 caused by evapo-condensed effect due to the prolonged sunshine duration for the 381

lakes in the Songnen Plain. With respect to the higher DOC concentration in winter, it 382 can be attributed to the accumulated DOC left in the liquid phase when ice formation 383 384 took place, resulting in the higher DOC concentration in the underlying water (unpublished material). Generally, the absorption coefficient a(350) is used as a proxy 385 for characterizing CDOM concentration (Guo et al., 2010; Zhang et al., 2011), a(280) 386 is related to DOC biodegradation (McDowell et al., 2006), and a(254) can be used to 387 characterize the optical properties of DOC aromaticity (Jaffe' et al., 2004; Weishaar et 388 al., 2003). The highest averaged CDOM absorption coefficients a(350), a(280), a(254) 389 390 were also present in February 2014, corresponding to the highest DOC concentration. The S_R values of the two wavelength ranges (275-295 nm over 350-400 nm) were 391 used to represent DOM molecular weight (Helms et al., 2008). The lowest mean of S_R 392 393 was present in August 2013 suggesting the relatively weak microbial decomposition of plants and lots of terrestrially imported substances through rainwash resulted in the 394 higher average molecular weight of DOC. 395

396

397Table 3. Mean values of DOC concentration and CDOM absorption coefficients groups at398different seasons. S_R : the slope ratio of $S_{275-295nm}$: $S_{350-400nm}$.

399

When the whole data set (N = 67) was pooled together, there were significantly positive linear relationships between a(254), a(280), a(350) and F_{max} for two humic-like components (C1 and C2), respectively, but mostly such correlations were not observed for the protein-like components (Fig. 4a and b, Table 3). These results were in accordance with previous investigations (Zhang et al., 2010, 2011).

Components 1 and 2 were strongly linearly correlated with each other ($R^2 = 0.633$) 405 (Fig. 4c), indicating that the concentrations of the two humic-like components were 406 controlled by common sources (Baker and Spencer, 2004). There was a weak 407 relationship ($R^2 = 0.051$) between the protein-like components (C3 and C4) possibly 408 409 because of a complex origin of CDOM such as rainfall in summer, ice in winter and organic pollutants derived from domestic, agricultural and industrial sewerage, which 410 represent the complex origins of CDOM. However, there was almost no correlation 411 between the humic-like and protein-like components. The linkage of a fluorescence 412 413 signal to DOC was very complicated because of the seasonal impacts, i.e., increased rainfall, algal blooms and ice-cover, which affect the DOC concentration. Due to both 414 steady and labile CDOM fluorescent components in DOC, the fluorescent signal 415 416 would change with the ratio of fluorescent and non-fluorescent CDOM components (Henderson et al., 2009). A weak relationship $(R^2 = 0.42)$ (Fig. 4d) was found between 417 DOC and component 3 likely from the decay of plants through microbial activity or 418 419 the pollution from human and animal wastes.

Different from the findings by Yamashita et al. (2008) for ocean water, this study did not find obvious correlation between salinity and EEM-PARAFAC extracted components with the exception of C3 ($R^2 = 0.469$) (Table 4 and Fig. 4f). The most important finding for the water samples collected at different seasons from the Songnen Plain is a significant relationship ($R^2 = 0.934$) between salinity and DOC (Fig. 4e). This is because DOC is evapo-condensed from spring to autumn and freeze-accumulated in winter in the semi-arid region. A prolonged sunshine duration 427 can result in an evapo-condensed DOC concentration in ice-free season. On the other
428 hand, the DOC is accumulated when the lakes freeze in winter leaving DOC in the
429 liquid phase.

430

Table 4. Correlation coefficients (R) and significance levels (p) of the linear relationships
between CDOM absorption, DOC, salinity and fluorescent components.

433

Figure 4. Relationships between CDOM absorption coefficient a(350) with a) $F_{max}(C1)$, b) with $F_{max}(C2)$, c) peak $F_{max}(C1)$ versus $F_{max}(C2)$, d) peak $F_{max}(C3)$ versus DOC, e) Salinity versus DOC, f) Salinity versus $F_{max}(C3)$.

437

438 **4 Conclusions**

A model II-ANOVA using season and lake as random effect factors shows that the 439 seasonal variability ($F > F_{crit}$, p < 0.05) is higher than between-lake variability. In this 440 study, the application of EEM-PARAFAC to characterize four fluorescent 441 components under seasonal variation in CDOM was presented with 67 water samples 442 443 collected from June 2013 to April 2014 in the semi-arid region of the Songnen Plain. Two humic-like and the protein-like components were identified using the PARAFAC 444 445 modeling. The average fluorescence intensity of the four components differed under 446 seasonal variation from June 2013 to April 2014. The highest C1 1.02 nm⁻¹ was presented in February 2014 probably due to the condensed CDOM caused by ice 447 formation in winter. Especially in summer when quantities of rainfall take place and 448 in winter when water is frozen, the fluorescence intensity is dominated by 449 tyrosine-like components in rain and ice-melt water. Component 1 and 2 exhibited a 450

strong linear correlation ($R^2 = 0.633$). There were significantly positive linear 451 relationships between F_{max} and CDOM absorption coefficient a(254) (R² = 0.72, 0.46, 452 p < 0.01), a(280) (R² = 0.77, 0.47, p < 0.01), a(350) (R² = 0.76, 0.78, p < 0.01) for two 453 humic-like components (C1 and C2), respectively. A weak relationship ($R^2 = 0.42$) 454 was found between DOC and component 3 from the decay of plants through 455 microbial activity or the pollution from human and animal wastes. However, almost 456 no obvious correlation was found between salinity and EEM-PARAFAC extracted 457 components except C3 ($R^2 = 0.469$), though the correlation was not as strong as with 458 DOC concentration. Most importantly, a significant relationship $(R^2 = 0.931)$ was 459 found between salinity and DOC. In order to understand the biogeochemical effects 460 on the aquatic ecosystem, further study should be required to identify CDOM source 461 462 and assess physical/chemical, bioavailable and photoreactive transformation in various lakes with larger saline gradients in the semi-arid region, Northeast China. 463

464

465 Acknowledgements

The research was jointly supported by the "One Hundred Talents" program from Chinese Academy of Sciences and the National Natural Science Foundation of China (No. 41471290). The authors thank Zhi Ding, Ying Guan, Lei Liu and Ming Wang for their persistent assistance with both field sampling and laboratory analysis.

470

471 **References**

472	APHA/AWWA/WE F .: Standard Methods for the Examination of Water and
473	Wastewater, Washington, DC: American Public Health Associstion, 1998.
474	Aiken, G. R., McKnight, D. M., Thorn, K. A., and Thurman, E. M.: Isolation of
475	hydrophobic organic-acids from water using nonionic macro porous resins, Org.
476	Geochem., 18, 567-573, 1992.
477	Andersen, C. M., and Bro, R.: Practical aspects of PARAFAC modeling of
478	fluorescence excitation-emission data, J. Chemom., 17, 200-215, 2003.
479	Andersso, C. A., and Bro. R.: The N-way Toolbox for MATLAB, Chemom. Intell.
480	Lab. Syst., 52, 1–4, 2000.
481	Babin, M., Stramski, D., Ferrari, G. M., Claustre, H., Bricaud, A., Obolensky, G.,
482	and Hoepffner, N.: Variations in the light absorption coefficients of
483	phytoplankton, nonalgal particles, and dissolved organic matter in coastal waters
484	around Europe, J. Geophys. Res., 108, 3211-3230, 2003.
485	Baker, A., Ward, D., Lieten, Shakti H., Periera, R., Simpson, Ellie C., and Slater, M.:
486	Measurement of protein-like fluorescence in river and waste water using a
487	handheld spectrophotometer, Water Res., 38, 2934-2938, 2004.
488	Baker, A., and Spencer, R. G. M.: Characterization of dissolved organic matter from
489	source to sea using fluorescence and absorbance spectroscopy, Sci. Total
490	Environ., 333, 217-232, 2004.

491 Barker, J. D., Dubnick, A., Lyons, W. B., and Chin, Y. P.: Changes in Dissolved
492 Organic Matter (DOM) Fluorescence in Proglacial Antarctic Streams, Arct.

Antarct. Alp. Res., 45, 305-317, 2013.

- Barker, J. D., Sharp, M. J., Fitzsimons, S. J., and Turner, R. J.: Abundance and
 dynamics of dissolved organic carbon in glacier systems, Arct. Antarct. Alp. Res.,
 38, 163–172, 2006.
- Barker, J. D., Sharp, M. J., and Turner, R. J.: Using synchronous fluorescence
 spectroscopy and principal components analysis to monitor dissolved organic
 matter dynamics in a glacier system, Hydrol. Processes, 23, 1487–1500, 2009.
- 500 Barker, J. D., Klassen, J. L., Sharp, M. J., Fitzsimons, S. J., and Turner, R. J.:
- 501 Detecting biogeochemical activity in basal ice using fluorescence spectroscopy,
 502 Ann. Glaciol., 51, 47–55, 2010.
- Borisover, M., Laor, Y., Parparov, A., Bukhanovsky, N., and Lado, M.: Spatial and
 seasonal patterns of fluorescent organic matter in Lake Kinneret (Sea of Galilee)
 and its catchment basin, Water Res., 43, 3104-3116, 2009.
- Bricaud, A., Babin, M., Morel, A., and Claustre, H.: Variability in the
 chlorophyll-specific absorption coefficients of natural phytoplankton: Analysis
 and parameterization, J. Geophys. Res., 100, 13321–13332, 1995.
- Bro, R.: PARAFAC tutorial and applications, Chemom. Intell. Lab. Syst., 38, 149-171,
 1997.
- 511 Bro, R.: Exploratory study of sugar production using fluorescence spectroscopy and 512 multi-way analysis, Chemom. Intell. Lab. Syst., 46,133-147, 1999.
- 513 Cheng, Y. Y., Guo, W. D., Long, A. M., and Chen, S. Y.: Study on optical

- characteristic of chromophoric dissolved organic matter in rainwater by
 fluorescence excitation-emission matrix and absorbance spectroscope (Article in
 Chinese), Spectrosc. Spect. Anal., 30, 2413-2416, 2010.
- 517 Coble, P. G.: Characterization of marine and terrestrial DOM in seawater using 518 excitation-emission matrix spectroscopy, Mar. Chem., 51, 325–346, 1996.
- Coble, P. G.: Marine optical biogeochemistry: the chemistry of ocean color, Chem.
 Rev., 107, 402-418, 2007.
- Coble, P. G., Del Castillo, C. E., and Avril, B.: Distribution and optical of CDOM in
 the Arabian Sea during the 1995 Southwest Monsoon, Deep-Sea Res. part II, 45,
 2195–2223, 1998.
- 524 Cory, R. M., and McKnight, D. M.: Fluorescence spectroscopy reveals ubiquitous 525 presence of oxidized and reduced quinines in dissolved organic matter, Environ.
- 526 Sci. Technol., 39, 8142-8149, 2005.
- 527 DelCastillo, C. E., Coble, P. G., Morell, J. M., Lopez, J. M., and Corredor, J. E.:
- Analysis of the optical properties of the Orinoco River plume by absorption and
 fluorescence spectroscopy, Mar. Chem., 66, 35–51, 1999.
- 530 Duarte, C. M., Montes, C., Cole, J. J., Striegl, R. G., Melackand, J., and Downing, J.
- A.: CO2 emissions from saline lakes: Aglobal estimate of asurprisingly large flux,
 J. Geophys. Res. Biogeosci., 113, G04041, 2008.
- 533 Fellman, J. B., Hood, E., and Spencer, R. G. M.: Fluorescence spectroscopy opens
- new windows into dissolved organic matter dynamics in freshwater ecosystems:

A review, Limnol. Oceanogr., 55, 2452-2462, 2010.

536	Guo, W. D., Xu, J., Wang, J. P., Wen, Y. G., Zhou, J. F., and Yan, Y. C.:
537	Characterization of dissolved organic matter in urban sewage using excitation
538	emission matrix fluorescence spectroscopy and parallel factor analysis, J.
539	Environ. Sci., 22, 1728-1734, 2010.
540	Henderson, R. K., Baker, A., Murphy, K. R., Hambly, A., Stuetz, R. M., and Khan, S.
541	J.: Fluoresence as a potential monitoring tool for recycled water system: A
542	review, Water Res., 43, 863-881, 2009.
543	Helms, J. R., Stubbins, A., Ritchie, J. D., Minor, E. C., Kieber, D. J., and Mopper, K.:
544	Absorption spectral slopes and slope ratios as indicators of molecular weight,
545	source, and photo bleaching of chromophoric dissolved organic matter, Limnol.

546 Oceanogr., 53, 955–969, 2008.

- Hua, B., Dolan, F., Mcghee, C., Clevenger, ThomasE., and Deng, B. L.: Water-source
 characterization and classification with fluorescence EEM spectroscopy:
 PARAFAC analysis, Int. J. Environ. Anal. Chem., 87, 135-147, 2007.
- Hudson, N., Baker, A., and Reynolds, D.: Fluorescence analysis of dissolved organic
 matter in natural, waste and polluted waters a review, River Res. Appl., 23,
 631–649, 2007.
- Jaffe', R., Boyer, J. N., Lu, X., Maie, N., Yang, C., Scully, N. M., and Mock, S.:
 Source characterization of dissolved organic matter in subtropical
 mangrove-dominated estuary by fluorescence analysis, Mar. Chem., 84, 195–210,

556 2004.

574

557	Larsson, T., Wedborg, M., and Turner, D.: Correction of inner-filter effect in
558	fluorescence excitation-emission matrix spectrometry using Raman scatter, Anal.
559	Chim. Acta., 583, 357-363, 2007.
560	Lawaetz, A. J., and Stedmon, C. A.: Fluorescence Intensity Calibration Using the
561	Raman Scatter Peak of Water, Appl. Spectrosc., 63, 936-940, 2009.
562	Mayer, L. M., Schick, L. L., and Loder, T. C.: Dissolved protein fluorescence in two
563	Maine estuaries, Mar. Chem., 64, 171–179, 1999.
564	McDowell, W. H., Zsolnay, A., Aikenhead-Peterson, J. A., Gregorich, E. G., Jones, D.
565	L., Jo"dermann, D., Kalbitz, K., Marschner, B., and Schwesig, D.: A
566	comparison of methods to determine the biodegradable dissolved organic
567	carbon from different terrestrial sources, Soil Biol. Biochem., 38, 1933-1942,
568	2006.
569	Mcknight, D. M., Boyer, E. W., Westerhoff, P. K., Doran, P. T., Kulbe, T., and
570	Andersen, D. T.: Spectrofluorometriccharacterization of dissolved organic
571	matter for indication of precursor organic material and aromaticity, Limnol.
572	Oceanogr., 46, 38–48, 2001.
573	Song, C. C., Wang, L. L., Guo, Y. D., Song, Y. Y., Yang, G. S., and Li, Y. C.: Impacts

- 575 Plain, Northeastern China, J. Hydrol., 398, 26-32, 2011.
- 576 Song, K. S., Zang, S. Y., Zhao, Y., Du, J., Lin, L., Zhang, N. N., Wang, X. D., Shao, T.

of natural wetland degradation on dissolved carbon dynamics in the Sanjiang

577	T., Guan, Y., and Liu, L.: Spatiotemporal characterization of dissolved carbon
578	for inland waters in semi-humid/semiarid region, China, Hydrol. Earth Syst.
579	Sci., 17, 4269-4281, 2013.
580	Spencer, R. G. M., Hernes, P. J., Ruf, R., Baker, A., Dyda, R. Y., Stubbins, A., and
581	Six, J.: Temporal controls on dissolved organic matter and lignin
582	biogeochemistry in apristine tropical river, J. Geophys. Res. Biogeosci., 115,
583	G03013, 2010.
584	Stedmon, C. A., and Bro, R.: Characterizing dissolved organic matter fluorescence
585	with parallel factor analysis: a tutorial, Limnol. Oceanogr. Methods, 6, 572-579,
586	2008.
587	Stedmon, C. A., and Markager, S.: Tracing the production and degradation of
588	autochthonous fractions of dissolved organic matter by fluorescence analysis,
589	Limnol. Oceanogr., 50, 1415-1426, 2005.
590	Stedmon, C. A., Markager, S., and Bro, R.: Tracing dissolved organic matter in

- aquatic environments using a new approach to fluorescence spectroscopy, Mar.
 Chem., 82, 239-254, 2003.
- Stedmon, C. A., Thomas, D. N., Granskog, M., Kaartokallio, H., Papadimitriou, S.,
 and Kuosa, H.: Characteristics of dissolved organic matter in Baltic coastal sea
 ice: allochthonous or autochthonous origins? Environ. Sci. Technol., 41,
 7273–7279, 2007.
- 597 Thomas K. B.: Under Landfast ice, Arctic, 36, 328-340, 1983.

598	Tranvik, L. J., Downing, J. A., Cotner, J. B., Loiselle, S. A., Striegl, R. G., Ballatore, T.
599	J., Dillon, P., Finlay, K., Fortino, K., Knoll, L. B., Kortelainen, P. L., Kutser, T.,
600	Larsen, S., Laurion, I., Leech, D. M., McCallister, S. L., McKnight, D. M.,
601	Melack, J. M., Overholt, E., Porter, J. A., Prairie, Y., Renwick, W. H., Roland, F.,
602	Sherman, B. S., Schindler, D. W., Sobek, S., Tremblay, A., Vanni, M. J.,
603	Verschoor, A. M., Wachenfeldt, E. V., and Weyhenmeyer, G. A.: Lakes and
604	reservoirs as regulators of carbon cycling and climate, Limnol. Oceanogr., 54,
605	2298–2314, 2009.
606	Uusikiv, J., Vahatal, A.V., Granskog, M.A., Sommaruga, R., 2010. Contribution of
607	mycosporine-like amino acids and colored dissolved and particulate matter to
608	sea ice optical properties and ultraviolet attenuation, Limnol. Oceanogr., 55(2),
609	703–713.
610	UV talk letter vol. 10, 2013. https://shimadzu.com.au/uv-talk-letter-volume-10
611	Weishaar, J. L., Aiken, G. R., Bergamaschi, B. A., Farm, M. S., Fujii, R., and Mopper,
612	K.: Evaluation of specific ultraviolet absorbance as an indicator of the chemical
613	composition and reactivity of dissolved organic carbon, Environ. Sci. Technol.,
614	37, 4702–4708, 2003.
615	Wharton, R. A., Jr., McKay, C. P., Clow, G. D., and Andersen, D. T.: Perennial ice
616	covers and their influence on Antarctic lake ecosystems, Antarct. Res. Ser., 59,
617	53–70, 1993.
618	Yamashita, Y.: Assessing the dynamics of dissolved organic matter (DOM) in coastal
619	environments by excitation emission matrix fluorescence and parallel factor

analysis (EEM-PARAFAC), Limnol. Oceanogr., 53, 1900-1908, 2008.

621	Yamashita, Y., Cory, R. M., Nishioka, J., Kuma, K., Tanoue, E., and Jaffe', R.,:
622	Fluorescence characteristics of dissolved organic matter in the deep waters of the
623	Okhotsk Sea and the northwestern North Pacific Ocean, Deep Sea Res. Part II,
624	57, 1478–1485, 2010.
625	Zhang, Y. L., Liu, X. H., Osburn, C. L., Wang, M. Z., Qin, B. Q., and Zhou, Y. Q.:
626	Photo bleaching response of different Source of Chromophoric Dissolved
627	Organic Matter Exposed to Natural Solar Radiation Using Absorption and
628	Excitation-Emission Matrix Spectra, Plos one, 8, e77515, 2013.
629	Zhang, Y. L., Yin, Y., Feng, L. Q., Zhu, G. W., Shi, Z. Q., Liu, X. H., and Zhang, Y. Z.:
630	Characterizing chromophoric dissolved organic matter in Lake Tianmuhu and its
631	catchment basin using excitation-emission matrix fluorescence and parallel
632	factor analysis, Water Res., 45, 5110-5122, 2011.
633	Zhang, Y. L., Zhang, E. L., Yin, Y., VanDijk, M. A., Feng, L. Q., Shi, Z. Q., Liu, M. L.,
634	and Qin, B. Q.: Characteristics and sources of chromophoric dissolved organic
635	matter in lakes of the Yungui Plateau, China, differing in trophic state and
636	altitude, Limnol. Oceanogr., 55, 2645-2659, 2010.
637	Zhang, Y. L., VanDijk, M. A., Liu, M. L., Zhu, G. W., and Qin, B. Q.: The
638	contribution of phytoplankton degradation to chromophoric dissolved organic
639	matter (CDOM) in eutrophic shallow lakes: Field and experimental evidence.
640	Water Res., 43, 4685–4697, 2009.

641

Table 1. Mean value of water quality parameters from June 2013 to April 2014. Turbdenotes water turbidity; N denotes sampling numbers.

Sampling season	pН	Salinity (PSU)	Turb (NTU)	Ν
Jun.2013	8.54	0.40	166.20 ± 108.73	15
Aug.2013	8.63	0.37	63.13±31.21	13
Feb.2014	8.35	0.70	21.33 ± 15.87	17
Apr.2014	8.67	0.43	22.24 ± 16.42	22
All	8.55	0.48	62.18±79.07	67

Table 2. Positions of the fluorescence maximum peaks of the four components
identified by PARAFAC modeling in the present study compared with those
previously identified. Secondary excitation maxima is given in brackets.

	Component	$Ex_{max}(nm)$	Em _{max} (nm)	Description and source	Components	Components
	No				(Coble) and	(Stedmon and Markager)
	C1	230 (300)	425	Marine humic-like (phytoplankton degradation)	(Zhang) M	6
	C2	255 (350)	460	Terrstrial humic-like	A and C	1 and 4
	C3	225 (290)	360	Autochthonous tryptophan-like	Т	
	C4	220 (275)	320	Autochthonous tyrosine-like	В	8
672	Fluoresence	peaks were na	amed as Comp) by Coble et al. (1996, 1998) and Zhang et al.
673	(2010, 2011),	, while as Cor	nponets (Stedi	mon and Markager) by Stee	dmon and Markage	er (2005).
674						
675						
676						
677						
678						
679						
680						
681						
682						
683						
684						
685						
686						
687						
(00						
688						

691	Table 3. N	Mean values of	f DOC concentration	ration and CI	DOM absorp	ption coeffici	ents
692	groups at d	lifferent seasons	S_R : the slope ra	tio of S275-295m	m: S350-400nm.		
693							
	Sampling	a(254) m ⁻¹	a(280) m ⁻¹	a(350) m ⁻¹	\mathbf{S}_R	DOC mg L⁻¹	N
	season						
	Jun.2013	38.39±9.23	25.98±6.38	5.73±1.68	1.29±0.16	31.84±14.67	15
	Aug.2013	29.71±4.73	19.36±2.91	5.82±0.81	0.96±0.22	32.83±14.78	13
	Feb.2014	52.88±18.13	34.62±11.54	6.36±2.17	1.18±0.11	55.04±20.00	17

 22.45 ± 7.36

25.73±9.58

4.17±1.49

5.40±1.84

 $1.32{\pm}0.13$

1.21±0.20

30.86±10.91

37.60±18.05

nts

6	9	4
	_	_

Apr.2014

All

 $34.43{\pm}11.38$

39.08±14.73

Table 4. Correlation coefficients (R) and significance levels (*p*) of the linear
relationships between CDOM absorption, DOC, salinity and fluorescent components.

/15										
		a(254)	a(280)	a(350)	DOC	Salinity	C1	C2	C3	C4
	DOC	0.711**	0.646**	0.294*	1.000**					
	Salinity	0.650**	0.579**	0.159	0.965**	1.000**				
	C1	0.850**	0.875**	0.873**	0.496**	0.383**	1.000**			
	C2	0.677**	0.686**	0.885**	0.414**	0.270^{*}	0.796**	1.000**		
	C3	0.452**	0.417**	0.134	0.648**	0.685**	0.267^{*}	0.103	1.000**	
	C4	-0.040	-0.016	0.078	-0.101	0.135	0.084	0.069	0.225	1.000**
716	**p<0.0	1 level ; * <i>p</i> <	<0.05 level.							
717										
718										
710										
719										
720										
720										
721										
, = 1										
722										
723										
724										
, <u> </u>										

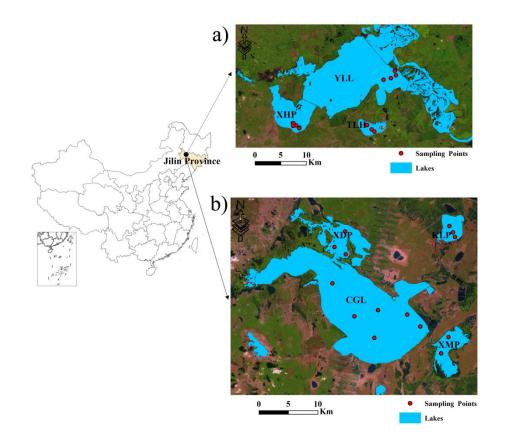


Figure 1. Locations of the water sampling sites for 7 lakes in the western part of Jilin
province, Northeast China. a) Yueliang lake group: YLL, Yueliang Lake; XHP;
Xinhuangpao; TLH; Talahong; b) Chagan lake group: CGL, Chagan Lake; XDP,
Xindianpao; XMP, Xinmiaopao; KLP, Kulipao.

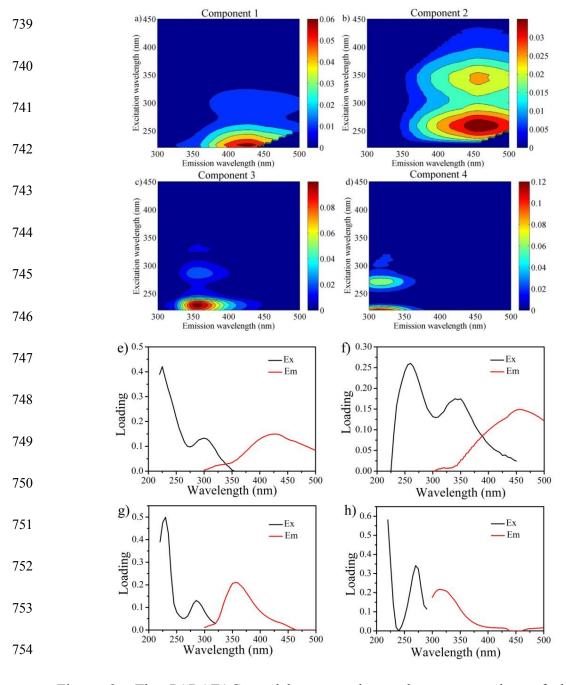


Figure 2. The PARAFAC model output shows the contour plots of the four
PARAFAC fluorescent components (a-d) and excitation (black) and emission (red)
loadings (e-h) of each component. Fluorescence is in Raman units: nm⁻¹.

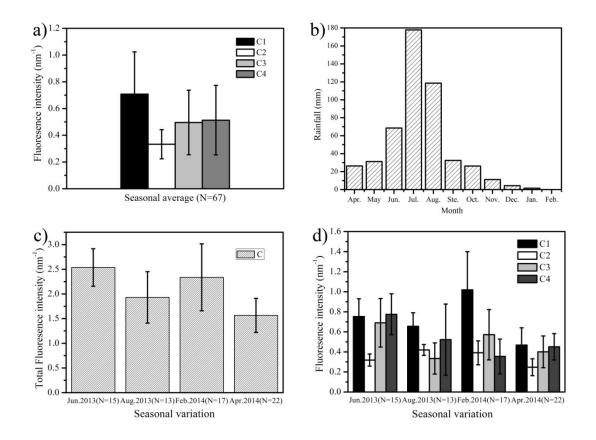




Figure 3. a) Seasonal average of F_{max} for EEM-PARAFAC components (C1, C2, C3 and C4) for lakes in the western part of Jilin province; b) Monthly variation of rainfall for the lakes in western part of Jilin province from April 2013 to February 2014; c) Seasonal variation of the total fluorescence intensity at different seasons; d) Seasonal variation of the four EEM-PARAFAC components at different seasons. The error bars represent one standard deviation.

- 766
- 767

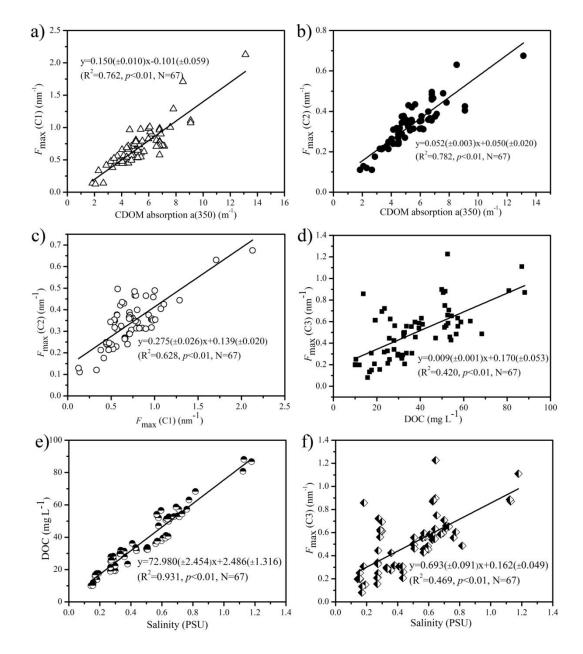




Figure 4. Relationships between CDOM absorption coefficient a(350) with a) $F_{max}(C1)$, b) with $F_{max}(C2)$, c) $F_{max}(C1)$ versus $F_{max}(C2)$, d) $F_{max}(C3)$ versus DOC, e) Salinity versus DOC, f) Salinity versus $F_{max}(C3)$.

- 773
- 774
- 775
- -
- 776

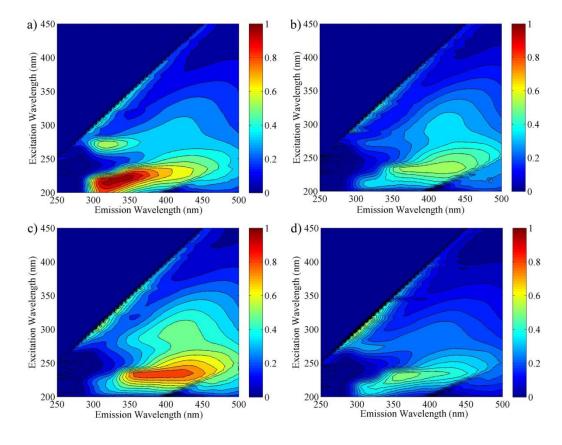
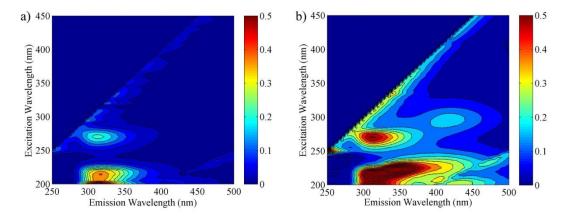


Figure S1. Examples of EEMs for one water sample from Xindianpao Lake in the
western part of Jilin province at different seasons a) June 2013; b) August 2013; c)
February 2014; d) April 2014 (Fluorescence is in Raman unit: nm⁻¹).



790

Figure S2. Representative examples of EEMs for a) lake ice-melt water sample, and b)

rainwater CDOM in the western part of Jilin province (Raman: nm⁻¹).