| 1  | Including high frequency variability in coastal ocean acidification projections                                                                                 |
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| 18 |                                                                                                                                                                 |
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# 24 Abstract

25 Assessing the impacts of anthropogenic ocean acidification requires knowledge of 26 present-day and future environmental conditions. Here, we present a simple model for upwelling 27 margins that projects anthropogenic acidification trajectories by combining high-temporal 28 resolution sensor data, hydrographic surveys for source water characterization, empirical 29 relationships of the CO<sub>2</sub> system, and the atmospheric CO<sub>2</sub> record. This model characterizes CO<sub>2</sub> 30 variability on timescales ranging from hours (e.g. tidal) to months (e.g. seasonal), bridging a 31 critical knowledge gap in ocean acidification research. The amount of anthropogenic carbon in a 32 given water mass is dependent on the age, therefore a density-age relationship was derived for 33 the study region, and was combined with the 2013 Intergovernmental Panel on Climate Change 34 CO<sub>2</sub> emission scenarios to add density-dependent anthropogenic carbon to the sensor time series. 35 The model was applied to time series from four autonomous pH sensors, each deployed in the 36 surf zone, kelp forest, submarine canyon edge, and shelf break in the upper 100 m of the 37 Southern California Bight. All habitats were within 5 km of one another, and exhibited unique, 38 habitat-specific CO<sub>2</sub> variability signatures and acidification trajectories, *demonstrating the* 39 importance of making projections in the context of habitat-specific  $CO_2$  signatures. In general, 40 both the mean and range of  $pCO_2$  increase in the future, with the greatest increases in both 41 magnitude and range occurring in the deeper habitats due to reduced buffering capacity. On the 42 other hand, the saturation state of aragonite  $(\Omega_{Ar})$  decreased in both magnitude and range. This 43 approach can be applied to the entire California Current System, and upwelling margins in 44 general, where sensor and complementary hydrographic data are available.

46 **1 Introduction** 

47 It has become increasingly apparent that upwelling systems, including the California 48 Current System (CCS), are particularly vulnerable to anthropogenic ocean acidification due to 49 their unique physical and chemical traits (Feely et al., 2008, 2010; Gruber et al., 2012; Hauri et 50 al., 2013a, 2013b). Upwelled waters have been isolated from the atmosphere and are naturally 51 elevated in  $CO_2$  from remineralization of organic matter; depending on the age of the upwelled 52 water mass it may also contain anthropogenic carbon (Harris et al., 2013; Sabine et al., 2002). 53 Recent observations estimate the saturation horizon with respect to aragonite (depth at which  $\Omega_{Ar}$ 54 = 1) along the CCS has shoaled by approximately 50 m since preindustrial times, and undersaturated waters ( $\Omega_{Ar}$  < 1) have been observed at the surface near the California-Oregon 55 border during a strong upwelling event (Alin et al., 2012; Feely et al., 2008; Harris et al., 2013). 56 Furthermore, the rate of acidification (i.e.  $\Delta pH yr^{-1}$ ) is expected to be significantly higher along 57 58 upwelling margins than observed in the surface open ocean (Bates et al., 2014; Gruber et al., 59 2012; Hauri et al., 2013b; Leinweber and Gruber, 2013; Rykaczewski and Dunne, 2010) due to 60 the reduced buffering capacity of seawater at higher levels of  $CO_2$  (Frankignoulle, 1994). This 61 effect has caused parts of the CCS to venture beyond the envelope (defined as: mean  $\pm 1$  s.d.) 62 from modeled preindustrial  $\Omega_{Ar}$  conditions (Hauri et al., 2013b). This is a concern because 63 organisms may need to survive outside of the environmental conditions to which they are 64 acclimatized, and the evolutionary potential for key ecological species to adapt to such rapid and 65 unprecedented changes is poorly understood. For example, a significant decrease in calcareous benthic organisms was observed along a natural pH gradient near a cold volcanic CO<sub>2</sub> vent 66 (Hall-Spencer et al., 2008; Kroeker et al., 2011). However, some calcareous organisms such as 67 68 limpets seemed to have adapted to higher  $CO_2$  levels compared to corals and mussels in the same

69 system (Rodolfo-Metalpa et al., 2011). Furthermore for upwelling margins, CO<sub>2</sub> co-varies with 70 other environmental stressors such as temperature and O<sub>2</sub> (Reum et al., 2015), making 71 predictions more difficult due to potential non-linear synergistic effects (Frieder et al., 2014).

72 A critical component in making accurate impact assessments of ocean acidification is the 73 development of robust, ecosystem-specific projections of future CO<sub>2</sub> conditions (Andersson et al., 74 2013; Cai et al., 2011; Feely et al., 2009, 2010; McNeil and Matear, 2008; Sunda and Cai, 2012). 75 Development of surface, open-ocean acidification projections has been relatively straightforward, 76 as they rely on well-defined chemical principles of  $CO_2$  equilibrium at the air-sea interface 77 (Byrne et al., 2010; Lauvset and Gruber, 2014). Models become more complicated when 78 attempting to resolve biological and physical processes that contribute significantly to the natural 79 variability of the system. For example, biologically mediated "enhanced acidification" was 80 identified in the northern Gulf of Mexico, causing significantly faster rates of acidification than 81 the open ocean (Cai et al., 2011). On many tropical coral reefs, seasonal patterns in CO<sub>2</sub> are 82 minimal, whereas the dominant frequency of variability occurs on diel and tidal frequencies 83 (Hofmann et al., 2011). On upwelling margins, both biological and physical processes contribute 84 to the observed natural variability of carbonate conditions (Fassbender et al., 2011).

One approach to develop region-specific ocean acidification projections is to apply an eddy-resolving regional ocean model system (ROMS) coupled with a biogeochemical component, as has been developed for the CCS (Gruber et al., 2011, 2012; Hauri et al., 2013b). Such models have highlighted the importance of capturing physical and biological processes in highly dynamic upwelling systems. The model simulations show complex spatiotemporal variability (Hauri et al., 2013b), and predict that the frequency of "corrosive" upwelling events will intensify in both magnitude and duration by the year 2050 (Hauri et al., 2013a). However, 92 the eddy-resolving ROMS project  $pCO_2$  and saturation state conditions on a 5-km grid, whereas 93 many marine animals experience the environment on the scale of centimeters to meters. In 94 addition, regional models can largely resolve event-scale (weeks) and seasonal features, but 95 cannot capture fluctuations on diel to tidal time scales, which can be the dominant frequency of 96 variability in many near-shore environments (Duarte et al., 2013; Frieder et al., 2012; Hofmann 97 et al., 2011). Due to this discrepancy in both space and time, numerical models tend to 98 underestimate or entirely miss the high-frequency variability that exists for the microclimate of 99 organisms.

100 To transition from region-specific to habitat-specific ocean acidification projections, high 101 temporal resolution data from autonomous chemical sensors deployed across many habitat types 102 can be used to directly quantify the full range of present-day carbonate conditions (Harris et al., 103 2013; Hofmann et al., 2011, 2014; Martz et al., 2014; Sutton et al., 2014). The CCS supports 104 many ecosystems that are of great ecological and economic value. In particular, there is great 105 habitat and species diversity near-shore on the shelf, which includes a large number of 106 commercially important invertebrates and fishes. Habitats in the region include bays and 107 estuaries, rocky and sandy intertidal, eelgrass beds and kelp forests, sub-tidal reefs, canyons and 108 extensive sandy sea floor. There are many endangered and harvested benthic organisms that 109 inhabit only one or a subset of these habitats.

110 Sensor data provide key observations for the mechanistic understanding of the controls 111 on environmental conditions, and are particularly needed for coastal marine environments where 112 complex physical and biological processes influence the observed variability. For example, 113 sensor data from a near-shore kelp forest in the southern CCS revealed that local biological 114 feedbacks and episodic upwelling events were the dominant drivers of CO<sub>2</sub> variability, with

pCO<sub>2</sub> fluctuating by 600 µatm at 17-m water depth (Frieder et al., 2012). This scale of variability associated with near-shore environments is not captured by regional model simulations, but is most relevant for organisms living inside the kelp forest.

118 Here, we present an anthropogenic ocean acidification model to project  $CO_2$  chemistry 119 into the future by combining autonomous chemical sensor data, regional empirical relationships 120 for the CO<sub>2</sub> system (Alin et al., 2012), hydrographic data, and the atmospheric CO<sub>2</sub> record 121 (Keeling et al., 2005). This model was applied to four habitats ranging from the surface to 100-m 122 water depth and all within 5 km of each other in the Southern California Bight (SCB). Each site 123 showed distinct CO<sub>2</sub> variability signatures and acidification trajectories, highlighting the 124 importance of interpreting ocean acidification projections in the context of present and future 125 habitat-specific CO<sub>2</sub> signatures. Implications for future ocean acidification research are discussed.

126 **2** Methods

127 2.1 Study Sites

Moored autonomous sensor packages SeapHOx or SeaFET (Bresnahan et al., 2014) were deployed at four depths (4, 17, 30, and 88 m) within several distinct habitats on the San Diego continental shelf for 1 year starting June 2012 (Figure 1). All sensors were deployed near the seafloor; the three shallowest sensors were deployed within 3 m of the bottom, and the deepest sensor was moored 12 m above the seafloor (Table 1).

A SeaFET was deployed at the Ellen Browning Scripps Pier 2 m above the benthos as a part of the Scripps Ocean Acidification Real-time Monitoring Program. The sensor was located approximately 400 m from the shore in the surf zone. Weekly discrete samples for total alkalinity (TA) and dissolved inorganic carbon (DIC) were taken alongside the sensors for calibration and

quality control following standard protocols (Dickson et al., 2007). The sensor was serviced
every 1-2 months to remove biofouling organisms.

The La Jolla kelp forest is part of the South La Jolla State Marine Reserve and is characterized by a dense population of *Macrocystis pyrifera*. The chemical variability in this ecosystem is strongly influenced by regional physical processes (*e.g.*, upwelling and stratification) and local biological feedbacks (*e.g.*, production and respiration). A SeapHOx was deployed at 17 m in the southern portion of the kelp forest, 3 m above the bottom. The reader is referred to *Frieder et al.*, 2012 for further details on site and deployment description.

145 The La Jolla canyon is a submarine canyon plunging from approximately 20 m to a depth 146 of 1000 m within several km from shore. A SeapHOx was deployed over a sandy bottom at the 147 southern canyon edge at 30 m depth within the Matlahuayl state marine reserve; the  $O_2$  sensor 148 malfunctioned and thus is not included. The water in the La Jolla canyon is characterized by 149 higher salinity, and lower temperature, O<sub>2</sub>, and pH (data not shown). Tidal energy in submarine 150 canyons is significantly amplified (Swart et al., 2011), bringing deep water from the canyon to 151 the canyon edge. Therefore, physical forcings are the dominant drivers for chemical variability at 152 this site (Navarro et al., 2013).

The Del Mar Buoy was first deployed in 2006 at 100 m off of Del Mar in northern San Diego at the shelf break, and has provided continuous time series data (e.g. temperature, salinity, oxygen, and current) at discrete depths (Frieder et al., 2012; Send and Nam, 2012). A SeaFET sensor was deployed on the mooring at 88 m in 2011, and has provided a near-continuous time series of pH since. Co-located sensors include temperature, salinity (SBE 37), and dissolved oxygen (O<sub>2</sub>; Aanderaa Optode). Water at this depth is isolated from the atmosphere and below the euphotic zone, and thus influenced primarily by upwelling and tidal dynamics.

160 2.2 Cruise data

161 Hydrographic data were collected aboard R/V Melville during the student-led San Diego 162 Coastal Expedition cruises in June/July and December of 2012 (Figure 1). The SCB is 163 characterized by relatively weak (compared to the northern CCS), but nearly year-round 164 upwelling. However, there is a clear seasonal cycle based on climatological data, where 165 upwelling intensifies generally between April and August, with the maximum occurring in May 166 (Bograd et al., 2009). The cruises therefore corresponded with upwelling (June/July) and non-167 upwelling (December) seasons. Water samples were collected at stations ranging from >100 km 168 from shore at 1200 m water depth, to within 5 km from shore at 30 m depth. Discrete samples 169 were analyzed for  $O_2$ , pH, and DIC; duplicate samples were collected during every cast.

Discrete samples for  $O_2$  were collected and analyzed by titration using a custom-built system (Martz et al., 2012). The titrant was standardized prior to and after each cruise using KIO<sub>3</sub> standard solutions prepared in house (Fisher, lot 105595); no detectable drift was observed for either cruise. Precision was  $\pm$  0.6 µmol kg<sup>-1</sup> (duplicate n = 62, 1 s.d.), and the accuracy was estimated to be  $\pm$  0.5% because KIO<sub>3</sub> standards were not recrystallized (Emerson, 1999).

175 Samples for DIC and pH were collected in 150-ml or 250-ml Pyrex serum bottles (13-176 mm neck) following standard procedures (Dickson et al., 2007). However, rather than leaving 177 headspace, the bottle was filled completely, and a gray butyl stopper was inserted to prevent gas 178 exchange and samples were analyzed within 4 hours of collection.

DIC samples were analyzed using custom-built system based on an infrared analyzer (LI-COR 7000) similar to systems built by others (Friederich et al., 2002; O'Sullivan and Millero, 181 1998). The DIC measurements were calibrated using Certified Reference Materials provided by 182 the Dickson Lab at SIO, by applying a gain correction (slope) and assuming an offset of zero 183 (intercept). The reference materials were stored in  $CO_2$  impermeable bags (3 L Scholle 184 DuraShield®), and were measured frequently throughout the cruise. The stability of the 185 reference material in the bag was verified by daily measurements of a new bottle; no drift was 186 observed. Precision and accuracy of the DIC measurements were  $\pm 2.5 \ \mu mol \ kg^{-1}$  (duplicate n = 187 67, 1 s.d.).

188 Samples for pH were analyzed spectrophotometrically at 20 °C (Clayton and Byrne, 189 1993) using an automated system (Carter et al., 2013). The pH is reported on the total hydrogen 190 ion concentration scale. The indicator dye (m-cresol purple, ACROS Lot A0264321) was used as 191 received from the manufacturer without further purification. An offset was applied based on 192 measurements in certified Tris buffer provided by the Dickson Lab. The precision and accuracy 193 of the measurements were estimated to be  $\pm 0.0015$  (duplicate n = 86, 1 s.d.) and  $\pm 0.02$  (Liu et 194 al., 2011), respectively. TA and pCO<sub>2</sub> were calculated using CO2SYS (van Heuven et al., 2011) 195 using pH and DIC as inputs, and carbonic acid dissociation constants from Mehrbach et al., 1973 196 refit by Lueker et al., 2000.

197 2.3 Sensor Data

The SeapHOx and SeaFET sensor packages utilize a modified Honeywell Durafet III pH combination electrode for high-frequency pH measurements (Martz et al., 2010). These sensor packages have been successfully deployed in ecosystems worldwide (Frieder et al., 2012; Hofmann et al., 2011; Kroeker et al., 2011; Martz et al., 2014; Price et al., 2012), and have been shown to have excellent stability in seawater for months to years (Bresnahan et al., 2014). The SeapHOx is an integrated sensor package that also consists of an Aanderaa 3835 oxygen optode, and a Seabird SBE 37 Conductivity-Temperature sensor all plumbed into a pumped flow stream; the SeaFET measures pH using a passively flushed cell. Sampling frequencies were 1 hr<sup>-1</sup> or
 greater at all depths.

All pH measurements were calibrated based on discrete TA and DIC samples taken alongside the sensor, at minimum at the beginning and end of each sensor deployment (n > 4 for every site), as recommended by the best practices (Bresnahan et al., 2014). The resolution of the pH measurements is better than 0.0005 pH, stability is estimated to be better than 0.005, and accuracy is estimated to be  $\pm$  0.015. Sensors were removed periodically for maintenance, but all were deployed for > 50 days during both the upwelling and relaxation season.

At the surf zone (surface waters), a constant TA value of 2240 µmol kg<sup>-1</sup> was assumed, 213 214 since discrete TA samples showed low variability (2240  $\pm$  7 (1 s.d.) µmol kg<sup>-1</sup>, n = 57). For the 215 three subsurface sensors, TA was estimated  $(TA^{est})$  using a regional empirical relationship 216 developed for the CCS, with temperature and salinity as inputs (Alin et al., 2012); an offset of +8  $\mu$ mol kg<sup>-1</sup> was applied to TA<sup>est</sup> based on comparisons to discrete samples collected (root mean 217 squared error (RMSE) = 6  $\mu$ mol kg<sup>-1</sup>, n = 25). This offset was a persistent feature over multiple 218 years (2010 - 2012), thus most likely reflecting a regional surface TA influence that is not 219 220 incorporated in the empirical relationship developed for the whole CCS. DIC, pCO<sub>2</sub>, and  $\Omega_{Ar}$ were calculated using CO2SYS (van Heuven et al., 2011) using pH sensor data and TA<sup>est</sup> as 221 222 inputs. Uncertainty for the calculated DIC, pCO<sub>2</sub>, and  $\Omega_{Ar}$  is pH-dependent, but on average is estimated to be  $\pm$  13 µmol kg<sup>-1</sup>,  $\pm$  25 µatm, and  $\pm$  0.04, respectively. Daily range of sensor data 223 224 was calculated by first high-pass filtering the data with a 36 hour window, and then taking the 225 difference between the daily maximum and minimum. The mean daily range was then calculated 226 by averaging the resultant time series.

## 227 2.4 Modelling Future Carbonate Chemistry

## 228 2.4.1 Approach

229 The carbonate conditions were modeled by decreasing or increasing DIC while using TA 230 conditions from 2012. Modeled projections were made for preindustrial times and for year t, where t ranges between 2012 and 2100. The model presented here is based on the  $\Delta C_t^*$  approach 231 232 (Gruber et al., 1996), but instead of using tracers to estimate the age of the water mass (e.g. 233 CFC's), we used the atmospheric CO<sub>2</sub> record as a quasi-age tracer. The age of the water mass 234 ranged between 0 and 50 years in this study region. Although this approach must be used with 235 caution, we demonstrate that our estimates are in good agreement with previously published 236 anthropogenic carbon inventory estimates using age tracer measurements in this region (Feely et 237 al., 2008; Sabine et al., 2002). In this model, it was assumed that ocean acidification is due to 238 anthropogenic  $CO_2$  invasion through the air-sea interface alone. We also assumed that both the 239 path of a particular water mass between the subduction and upwelling site and the rate of 240 remineralization processes remain unchanged. Sensitivity to these assumptions is explored in the Discussion. 241

#### 242

The DIC of the modeled year t  $(DIC_t)$  is calculated by

$$DIC_{\rm t} = DIC_{2012} + \Delta DIC_{\rm anth} \tag{1}$$

where  $DIC_{2012}$  is the DIC observed in 2012, and  $\Delta DIC_{anth}$  is the additional anthropogenic CO<sub>2</sub> that the water mass would have absorbed since 2012. Different formulations for  $\Delta DIC_{anth}$  were used for surface waters (i.e., above the seasonal mixed layer depth, defined here as  $\sigma_{\theta} \leq 25.2$  kg m<sup>-3</sup>) and subsurface waters ( $\sigma_{\theta} > 25.2$  kg m<sup>-3</sup>), and are outlined below.

For surface waters,  $\Delta DIC_{anth}$  was calculated as the difference in surface DIC between year t and 2012. Surface DIC was calculated by assuming atmospheric equilibrium with TA = 249 2240  $\mu$ mol kg<sup>-1</sup> (based on water samples from the Scripps pier) and using pCO<sub>2,atm</sub> projection 250 under the 2013 IPCC RCP6.0 scenario (Hijioka et al., 2008). Although large deviations from 251 equilibrium conditions are often observed in the coastal ocean due to upwelling and biological 252 production (Hales et al., 2005), the mean pCO<sub>2</sub> calculated from sensor data at the surf zone was 253 394 ± 43 (1 s.d.)  $\mu$ atm (Table 2), suggesting that the surface water at the study site was near 254 atmospheric equilibrium.

For subsurface waters,  $\Delta DIC_{anth}$  was quantified as the increase in DIC due to anthropogenic CO<sub>2</sub> when the water parcel was last in contact with the atmosphere. The mass balance of DIC for subsurface waters is:

$$DIC = DIC^{\circ} + \Delta DIC_{\rm bio} \tag{2}$$

where  $DIC^{\circ}$  is the preformed DIC, and  $\Delta DIC_{bio}$  is the DIC added by remineralization processes in the ocean interior.  $DIC^{\circ}$  can be expressed as the sum of DIC if it were in equilibrium with the atmosphere ( $DIC_{eq}$ ) and the degree of air-sea disequilibrium due to slow gas exchange kinetics and biological processes ( $\Delta DIC_{diseq}$ ):

$$DIC^{\circ} = DIC_{eq} + \Delta DIC_{diseq}$$
(3)

Since anthropogenic CO<sub>2</sub> only enters the ocean at the surface, the increase in  $DIC_{eq}$  represents the anthropogenic ocean acidification signal,  $\Delta DIC_{anth}$ , assuming  $\Delta DIC_{diseq}$  is invariant with time. However, in order to use this approach, the age of the water parcel must first be quantified, as this determines the pCO<sub>2,atm</sub> with which it was last in contact.

266 The age of the water parcel was established by combining equations (2) and (3):

$$DIC_{eq}^{2012-age} = DIC - \Delta DIC_{bio} - \Delta DIC_{diseq}$$
(4)

where the superscript denotes the year at which the water parcel was last at the surface (i.e. equal to 2012 - age of the water mass). The age of the water mass was calculated by comparing the atmospheric CO<sub>2</sub> record to the pCO<sub>2,atm</sub> that is necessary to generate  $DIC_{eq}^{2012-age}$ .  $\Delta DIC_{diseq}$  was estimated from published values in the region (Section 2.4.3). Using this information, we calculated  $\Delta DIC_{anth}$  by:

$$\Delta DIC_{\text{anth}} = DIC_{\text{eq}}^{\text{t-age}} - DIC_{\text{eq}}^{2012\text{-age}}$$
(5)

where the superscripts denote the year at which the water parcel was last at the surface, and age is the age of the water parcel. For example, if age = 30 yr and t = 2050, then CO<sub>2</sub> projections for the year 2020, would be used to calculate  $DIC_{eq}^{t-age}$ ;  $DIC_{eq}^{2012-age}$  was calculated from equation (4). The  $\Delta DIC_{anth}$  for subsurface waters was modeled for each projection year as a linear function of  $\sigma_{\theta}$ , and the surface and subsurface  $\Delta DIC_{anth}$  were connected assuming a two end member linear mixing between  $\sigma_{\theta}$  25.2 and 25.5 kg m<sup>-3</sup> to prevent step changes (Figure 2).

## 278 2.4.2 Calculation of $\Delta DIC_{bio}$

#### 279

 $\Delta DIC_{bio}$  was quantified following formulations in Sabine et al., 2002:

$$\Delta DIC_{\rm bio} = r_{\rm C:O}(AOU) - 0.5(TA_{\rm obs} - TA^{\circ} + r_{\rm N:O}(AOU))$$
(6)

where AOU = Apparent Oxygen Utilization = ( $O_{2,sat} - O_{2,obs}$ ),  $TA^{\circ}$  is the preformed alkalinity, and the *r*'s are the elemental remineralization ratios (Anderson and Sarmiento, 1994). The oxygen saturation concentration ( $O_{2,sat}$ ) was calculated using the equations in *Garcia and Gordon*, 1992, and  $TA^{\circ}$  was estimated based on historical near-surface *TA* data in the Pacific (equation 3 in Sabine et al., 2002). Phosphate concentrations necessary to estimate  $TA^{\circ}$  were not directly measured, but were estimated from a regional empirical relationship using historical data 286 (Supplementary Materials); the uncertainty in estimating phosphate using this approach 287 propagates to an error in  $TA^{\circ}$  of 4 µmol kg<sup>-1</sup>.

## 288 2.4.3 Estimation of $\Delta DIC_{diseq}$

289 Making accurate estimates of  $\Delta DIC_{diseq}$  is important because it is a source of large 290 uncertainty for anthropogenic carbon inventory calculations (Matsumoto and Gruber, 2005). 291 Traditionally, age of the water mass is quantified using tracers such as CFCs and then the 292  $\Delta DIC_{diseq}$  is subsequently calculated (Gruber et al., 1996; Sabine and Tanhua, 2010). However, 293 such tracer measurements were not made for this study. Alternatively, we estimated  $\Delta DIC_{diseq}$ 294 based on  $\theta$  & S data to overcome this limitation (Sabine et al., 2002). The mean  $\theta$  and S between  $\sigma_{\theta}$  of 25.5 and 26.5 kg m<sup>-3</sup> were 10.0 °C and 33.9, respectively, resembling water type 1e in 295 Sabine et al. (2010) with a corresponding  $\Delta DIC_{diseq} = -6.24 \ \mu mol \ kg^{-1}$ , the value used in this 296 297 study.

# 298 2.4.4 Calculation of the age of water parcel

299 In order to estimate the age of the water mass, we use equation (4) to calculate  $DIC_{eq}^{2012-age}$ , the DIC of the water parcel was in equilibrium with the atmosphere when it was 300 301 last at the surface (i.e. equal to 2012 - age of the water mass). Therefore the age of the water 302 mass can be calculated by comparing the atmospheric CO<sub>2</sub> record to the pCO<sub>2,atm</sub> that is necessary to generate  $DIC_{eq}^{2012-age}$ . The latter was calculated from the fugacity of CO<sub>2</sub> of the 303 water mass when it was last in contact with the atmosphere at the time of subduction 304  $(fCO_{2.eq}^{2012-age})$ , assuming 100% relative humidity, and barometric pressure of 1 atm [Dickson, 305 2007]. The year that the water parcel subducted was determined by matching the calculated 306 307 CO<sub>2,atm</sub> to the mean annual CO<sub>2,atm</sub> record (Keeling et al., 2005); the age is the difference between 2012 and the calculated year (Figure 3). A relationship between  $\sigma_{\theta}$  and the age was established by fitting a second order polynomial to the subsurface data (n = 186, R<sup>2</sup> = 0.92), and assuming the age of the surface water ( $\sigma_{\theta} < 25.2$ ) is 0 (Figure 3). The non-zero age of the water that appears around  $\sigma_{\theta} = 24.4$  kg m<sup>-3</sup> corresponds to the shallow oxygen maximum layer that formed during the summer. However, since this density range is still shallower than the seasonal mixed layer, its age was considered to be 0. The age of the water ranged between 0 and 50 years between  $\sigma_{\theta}$  of 25.2 and 26.5 kg m<sup>-3</sup>.

# 315 2.4.5 Estimation of preindustrial DIC

316 In order to calculate the preindustrial DIC, equation (3) is written as:

$$DIC^{\circ} = DIC_{eq}^{prein} + DIC_{anth} + \Delta DIC_{diseq}$$
(7)

where  $DIC_{anth}$  represents the anthropogenic carbon present in the water parcel in 2012, and  $DIC_{eq}^{prein}$  is the DIC of the water parcel if it were in equilibrium with pCO<sub>2,atm</sub> = 280 µatm. Combining equations (2) and (7) and rearranging gives

$$DIC_{anth} = DIC - \Delta DIC_{bio} - DIC_{eq}^{prein} + \Delta DIC_{diseq}$$
(8)

320 Calculated *DIC*<sub>anth</sub> as a function of  $\sigma_{\theta}$  is shown in Figure 4. Note that the values calculated 321 here are in good agreement with published values using age tracers (Feely et al., 2008; Sabine et 322 al., 2002) for higher  $\sigma_{\theta}$ , but are significantly higher at lower densities. This is because the 323 literature values were quantified using offshore subsurface waters, whereas our study region is 324 near the coast along an upwelling margin, where subsurface waters are brought near the surface 325 and are thus affected by surface processes. The agreement at higher density where surface 326 influence is minimal demonstrates that the model presented here is capable of making accurate 327 estimates of anthropogenic  $CO_2$ . Furthermore, the comparison illustrates the importance of 328 incorporating surface influence when making acidification projections in shallow, coastal

329 ecosystems. Preindustrial DIC ( $DIC_{prein}$ ) was calculated by subtracting  $DIC_{anth}$  from DIC 330 observed in 2012. Preindustrial pCO<sub>2</sub>,  $\Omega_{Ar}$ , and pH were calculated using  $DIC_{prein}$  and TA 331 conditions from 2012.

**332 3 Results** 

## 333 3.1 Carbonate Chemistry Variability Observed in 2012

334 The results are presented using pH, pCO<sub>2</sub>, or  $\Omega_{Ar}$ , since pH was directly measured, and 335 pCO<sub>2</sub> and  $\Omega_{Ar}$  are commonly used as stress indicators for respiration (Brewer and Peltzer, 2009) 336 and calcification (Langdon et al., 2010), respectively. Across all four sites in 2012, pCO<sub>2</sub> 337 increased with depth. In 2012, the mean  $pCO_2$  in the surf zone was near atmospheric equilibrium 338 (394  $\mu$ atm), while the mean pCO<sub>2</sub> at 88 m was 878  $\mu$ atm (Table 2), and reached a maximum of 339 1270  $\mu$ atm. The variability of pCO<sub>2</sub> also increased with depth (indicated by the s.d. of the time 340 series), which was only 43  $\mu$ atm in the surf zone, but was 149  $\mu$ atm at 88-m depth. The mean (± 341 1 s.d.)  $\Omega_{Ar}$  decreased with depth; the mean  $\Omega_{Ar}$  in the surf zone was 2.4 ± 0.25, where it was 1.05 342  $\pm$  0.18 at 88 m. Undersaturated conditions ( $\Omega_{Ar}$  < 1) were observed 48% of the time at 88 m in 343 2012, but were not observed at other sites. However, unlike pCO<sub>2</sub>, the variability of  $\Omega_{Ar}$ , 344 indicated by the s.d., decreased with depth (0.25 at the surface to 0.18 at 88 m) (Table 2; see 345 Discussion). The mean pH decreased with depth; the mean pH in the surfzone was 8.05, where it 346 was 7.73 at 88 m. The variability in pH increased with depth until 30 m, but decreased at 88 m 347 (Table 2).

Distinct, habitat-specific  $CO_2$  signatures were observed at the four deployment sites (Figure 5, Figure 6, and Figure 7). Here, we define habitat-specific  $CO_2$  signatures as how  $CO_2$ conditions varied in that habitat, regardless of biological or physical origin. In the surf zone, the conditions were near atmospheric equilibrium, with intrusions of higher p $CO_2$  waters through 352 internal tidal bores, a common feature observed in shallow, upwelling environments (Booth et al., 353 2012; Pineda, 1991); temperature and pH were correlated during these events (Supplementary 354 Figure S2). This leads to a high, mean daily range of CO<sub>2</sub> conditions (e.g. 96 µatm, 0.46, and 355 0.085 for pCO<sub>2</sub>,  $\Omega_{Ar}$ , and pH, respectively) (Table 3). However, the signature from the internal 356 bores usually only lasted several hours, and remained at near atmospheric equilibrium for the 357 large majority of the time. Higher occurrence of tidal bores were observed during the spring and 358 summer months relative to winter (Figure 5), consistent with previous observations (Pineda, 359 1991). The mean diel range of  $pCO_2$  at the surf zone was significantly higher than measurements 360 made by a surface mooring located off shore in the SCB (Leinweber et al., 2009).

361 The mean ( $\pm$  s.d.) CO<sub>2</sub> conditions in the kelp forest and the canyon edge were similar, 362 and the s.d. for  $\Omega_{Ar}$  and pH were the highest among the sites (Table 2). However, the timescales 363 of the variability were different, indicating that distinct processes control the CO<sub>2</sub> conditions in 364 these two habitats. For example, the mean daily range for all the variables was significantly 365 higher at the canyon edge compared to the kelp forest (Table 3). Submarine canyons are known 366 to amplify tidal energy (Navarro et al., 2013; Swart et al., 2011), and in fact, periodic variability 367 at the canyon edge occurred on semi-diurnal and diurnal cycles, indicative of tidal forcing. 368 Temeprature and pH were correlated on these shorter timescales (Supplementary Figure 2), 369 further supporting the fact that the variability was dominantly driven by intrusion of cold, deep 370 waters from the canyon.

While tidal forcings and daily biological production are drivers for carbonate chemistry in the La Jolla kelp forest, the largest variability occurred on event time scales (Frieder et al., 2012); event time scales are defined as longer than a day, but shorter than several weeks. For example, pH, pCO<sub>2</sub>, and  $\Omega_{Ar}$  regularly changed by up to 0.3, 250 µatm, and 1.3 on event time 375 scales, more than three times the mean daily range. Variability on event time scales is due to a 376 combination of changing water mass, stratification, and biological respiration (Frieder et al., 377 2012). In addition, a clear seasonal pattern was observed at the canyon edge, where higher  $pCO_2$ 378 and lower pH and  $\Omega_{Ar}$  were observed during the spring and summer months (upwelling season) 379 and lower pCO<sub>2</sub> and higher pH and  $\Omega_{Ar}$  were observed during the fall and winter (relaxation 380 season). Due to incomplete data coverage, a seasonal trend at the kelp forest and pier sites could not be discerned. The largest seasonal change among the four sites for  $\Omega_{Ar}$  (~1) was observed at 381 382 the canyon edge;  $pCO_2$  differed by roughly 200–300 µatm between the two seasons.

383 The shelf break experienced the highest mean CO<sub>2</sub> conditions, and the highest and lowest 384 s.d. for pCO<sub>2</sub> (149  $\mu$ atm) and  $\Omega_{Ar}$  (0.18), respectively (Table 2); the s.d. of pH (0.070) was lower 385 than the kelp forest (0.083) or the canyon edge (0.075). Variability on tidal, event, and seasonal 386 time scales were observed at this site (Figure 5, Figure 6, and Figure 7), as has been previously 387 reported for oxygen (Send and Nam, 2012). In general, upwelling on event time scales led to 388 greater changes in pCO<sub>2</sub> and pH than on tidal frequencies (Figure 5 and Figure 6). The largest 389 variability for all parameters occurred between the seasons, where a change in pCO<sub>2</sub>, pH, and  $\Omega_{Ar}$  were approximately 350 µatm, 0.2, and 0.5, respectively. The close proximity of these four 390 391 sites demonstrates the wide variety of habitat-specific CO<sub>2</sub> signatures that exist over a small 392 spatial scale, especially in near shore environments.

393

## 3.2 Modeled Carbonate Chemistry

Each habitat showed distinct trends in both modeled mean and variability of pCO<sub>2</sub>, pH, and  $\Omega_{Ar}$  owing to increased levels of anthropogenic DIC ( $\Delta DIC_{anth}$ ) (Table 2). For example, the mean pCO<sub>2</sub> at the surf zone (4 m), canyon edge (30 m), and shelf break (88 m) increased by 225, 435, and 738 µatm, respectively, from 2012 to 2100; this drastic difference in increased mean 398  $pCO_2$  is driven by different buffer factors due to depth differences among the sites. The increase 399 in variability (i.e. s.d.) was also larger at 88 m (97 µatm) compared to the surf zone (37 µatm), 400 although the largest increase occurred at the canyon edge at 30 m (126 µatm) (Table 2). Similar 401 trends were observed for mean pH, where the largest mean decrease in pH occurred at 88-m 402 water depth (0.26). However, the s.d. increased into the future for the three shallowest sites, 403 whereas the s.d. decreased at the shelf break (88 m). In contrast, the largest decrease in the mean 404  $\Omega_{Ar}$  occurred at the surface relative to the deeper sites, whereas the decrease in range was 405 equivalent across all depths.

406 The measured and modeled time series for pCO<sub>2</sub> and  $\Omega_{Ar}$  at the shelf break for the year 407 2012 and 2100 are shown in Figure 8 and Figure 9. The variability of pCO<sub>2</sub> increases on both 408 seasonal and tidal time scales; the seasonal amplitude increases from approximately 350 to 650 409 μatm and the mean daily range increases from 110 to 325 μatm by 2100. This greater variability 410 is in addition to an increase in mean  $pCO_2$  of > 700 µatm. On the other hand, the variability of 411  $\Omega_{Ar}$  on both seasonal and shorter time scales decreases. Furthermore, the shelf break is projected 412 to experience undersaturated waters over 90% of the time by 2060, compared to 48% in 2012. 413 Similar patterns were observed in the kelp forest as well, where both the mean conditions and 414 variability of pCO<sub>2</sub> increased, and  $\Omega_{Ar}$  decreased (Figure 10). The largest variability at the kelp 415 forest occurred on timescales of days to weeks, and high frequency (< 1 day) variability was 416 significantly smaller than at the shelf break. Therefore benthic organisms at the kelp forest would 417 experience elevated  $CO_2$  conditions for prolonged periods of time, with only intermittent 418 exposure to near-atmospheric conditions.

419 Preindustrial pCO<sub>2</sub> and  $\Omega_{Ar}$  were compared to conditions observed in 2012 (Table 2). At 420 most sites, the observed pCO<sub>2</sub>, pH, and  $\Omega_{Ar}$  in 2012 were already outside of their preindustrial

421 variability envelopes (defined as mean  $\pm 1$  s.d.), which is consistent with results from a previous 422 ROMS simulation in the CCS (Hauri et al., 2013b). These results suggest that all habitats studied 423 here have left, or are about to leave, the pCO<sub>2</sub>, pH, and  $\Omega_{Ar}$  conditions that were experienced 424 during preindustrial times. This is significant as organisms at these sites are now surviving in 425 conditions that are significantly different than the conditions under which their ancestors evolved.

The modeled habitat-specific pCO<sub>2</sub> and  $\Omega_{Ar}$  conditions for preindustrial, 2012, 2060, and 2100 are shown in Figure 11. The histograms represent the full range of carbonate conditions at each habitat that was captured by the sensors, which includes both the seasonal and highfrequency variability. The shape of each distribution skews towards more "corrosive" conditions at all sites as the model steps forward into the future. This translates to not only increases in mean pCO<sub>2</sub>, but also greater extremes and amount of time spent in extremes.

The projected pCO<sub>2</sub> and  $\Omega_{Ar}$  envelopes (mean ± s.d.) at each habitat throughout the modeled period are shown in Figure 12. An increasing rate of change in pCO<sub>2</sub> ( $\Delta$ pCO<sub>2</sub> yr<sup>-1</sup>) is observed, whereas  $\Omega_{Ar}$  tends to decrease at a relatively constant rate. The rate of increase of pCO<sub>2</sub> is higher than the projected atmospheric CO<sub>2</sub> increase at all subsurface sites. This indicates that as ocean acidification progresses, the effects due to elevated pCO<sub>2</sub> are more likely to become exacerbated with increasing depth. Mean  $\Omega_{Ar}$  is projected to be < 1 at the shelf break by 2020, and leave the 2012 variability envelope around 2070.

439 **4 Discussion** 

## 440 4.1 Changes in the Buffer Factors

441 The general patterns of the acidification trajectories presented here can be explained by 442 changing buffer factors of seawater, as deeper sites are more strongly influenced by  $CO_2$  rich 443 upwelled waters. The buffer factors  $\Pi_{pCO2}$ ,  $\Pi_{pH}$ , and  $\Pi_{CO3}$ , are defined as

$$\Pi_{pCO2} = \frac{\partial pCO_2}{\partial DIC}, \qquad \Pi_{pH} = \frac{\partial pH}{\partial DIC}, \qquad \Pi_{CO3} = \frac{\partial CO_3^{2-}}{\partial DIC}$$
(9)

444 representing the change in each carbonate parameter with respect to a change in DIC 445 (Frankignoulle, 1994). The effect of temperature on  $\Pi$  is small (< 10%) between 0-15 °C for the 446 DIC and TA values observed here, thus subsequent values were calculated assuming a temperature of 10 °C, TA = 2240  $\mu$ mol kg<sup>-1</sup>, and salinity = 33.5 (Figure 13). The ability for 447 448 seawater to buffer changes in pCO<sub>2</sub> diminishes under higher concentrations of DIC. For example, 449  $\Pi_{pCO2}$  increases from 1.6 to 3.3 at the surface between 2012 and 2100 under the RCP6.0 scenario. 450 However, since deeper waters are naturally elevated in DIC, this effect is more pronounced at the 451 shelf break:  $\Pi_{pCO2}$  increases from 6.2 to 12.3 during the same time interval. This explains why 452 the surf zone had the lowest mean increase in pCO<sub>2</sub> (225 µatm) despite having the highest increase in DIC (82 µmol kg<sup>-1</sup>) out of all of the sites. The shelf break on the other hand had the 453 454 highest increase in pCO<sub>2</sub> (737 µatm) while having the smallest increase in mean DIC (77 µmol 455 kg<sup>-1</sup>) during the same time period. Furthermore, the increase in variability with depth can be 456 explained as well, as the same biological and physical forcings on tidal to seasonal cycles cause a 457 larger change in pCO<sub>2</sub>.

458 Changes in  $\Pi_{CO3}$  can explain the patterns for  $\Omega_{Ar}$ , since  $[Ca^{2+}]$  and  $K_{SP}$  remain unchanged. 459 Unlike  $\Pi_{pCO2}$ ,  $|\Pi_{CO3}|$  decreases at higher concentrations of DIC (Figure 13B);  $|\Pi_{CO3}|$  decreases 460 from 0.62 to 0.57 at the surface, and 0.49 to 0.3 at 88 m between 2012 and 2100. This change in 461  $\Pi_{CO3}$  explains both the decrease in rate and range of  $\Omega_{Ar}$  as anthropogenic CO<sub>2</sub> continues to 462 infiltrate the ocean.

463 The  $\Pi_{pH}$  follows a parabolic shape, where there is a maximum decrease in pH per DIC 464 added (Figure 13C). In a pure carbonate solution, this maximum occurs when DIC = TA, but in

seawater it occurs at slightly lower DIC (Frankignoulle, 1994); this maxima occurs at DIC = 465 466 2225 µmol kg<sup>-1</sup> using the parameters listed above. Therefore we would expect to see a similar 467 trend for pH as  $pCO_2$ , where greatest changes occur at depth relative to the surface as long as the 468 mean DIC is lower than this threshold. This condition is only met at the shelf break (88 m) near 469 the end of the century, thus as expected greater decrease in mean pH, and increase in variability 470 (i.e. s.d.) was observed with depth (Table 2). One exception was observed where the s.d. 471 decreased as ocean acidification progressed at the shelf break. This is because an increased 472 proportion of time is spent at greater DIC where  $\Pi_{pH}$  is past its maxima, leading to a smaller 473 variability in pH under the same changes in DIC. It is important to note that the buffer factor of 474  $H^+$  ( $\Pi_{H^+}$ ) follows a similar pattern as pCO<sub>2</sub>, where it continues to increase as DIC increases 475 (Figure 13D). Therefore the rate of increase of [H<sup>+</sup>] will continue to increase as ocean 476 acidification progresses, thus biological responses to  $[H^+]$  may become exacerbated in the future.

# 477 4.2 Observed and Modeled Carbonate Chemistry Variability

478 The carbonate conditions presented here are consistent with previous studies. For example at the shelf break,  $\Omega_{Ar}$  had a strong seasonal cycle, where undersaturated waters were 479 480 observed almost continuously throughout the upwelling season (Nam et al., 2015), and remained 481 supersaturated for the rest of the year. This is in good agreement with previous hydrographic 482 surveys in this region, where aragonite-undersaturated waters have been observed as shallow as 483 60 m during the beginning of the upwelling season (Feely et al., 2008), but were not observed in 484 the upper 100 m at the end of the upwelling season in this region (Bednaršek et al., 2014). 485 Furthermore, estimates based on empirical equations showed a similar seasonal pattern in  $\Omega_{Ar}$  at 486 88 m, where undersaturated waters were observed every upwelling season (Alin et al., 2012). 487 However, undersaturated waters were not observed in the upper 30 m, unlike northern parts of the CCS where undersaturated conditions are repeatedly observed at the surface during the upwelling season (Bednaršek et al., 2014; Feely et al., 2008; Harris et al., 2013). Due to these traits, the southern portion of the CCS is commonly considered less vulnerable to ocean acidification compared to its northern counterpart. However, our results demonstrate that  $\Omega_{Ar}$  as low as 1.3 is routinely observed in the kelp forest (17 m), demonstrating the imminent threat of anthropogenic ocean acidification to the southern CCS.

494 The subsurface habitats characterized in this study routinely experience  $\Omega_{Ar}$  conditions 495 that have been shown to have non-lethal chronic effects on various bivalve larvae between  $\Omega_{Ar}$  of 496 1.2 to 2.0 (Barton et al., 2012; Gaylord et al., 2011; Gazeau et al., 2011; Hettinger et al., 2012; 497 Waldbusser et al., 2015). However, the length of exposure to these unfavorable conditions varies 498 between habitats. For example, the organisms in the kelp forest would be exposed to low  $\Omega_{Ar}$ 499 conditions for days to weeks, whereas large tidal variability at the canyon edge could result in 500 periodic exposure to low  $\Omega_{Ar}$  conditions on the order of hours. Therefore the effects of low  $\Omega_{Ar}$ 501 will largely depend on the reproductive timing and environmental variability that occurs on event 502 to seasonal time scales; the effects of exposure on various timescales are poorly understood. 503 Such events are expected to become more severe in the future (Hauri et al., 2013a), and thus 504 could lead to an increased rate of failed recruitment of bivalves and other keystone organisms 505 (Byrne et al., 2013).

It may be surprising that the mean diel range of pH was the smallest at the kelp forest (Table 3), as one might expect a large diel cycle driven by photosynthesis and respiration in a highly productive kelp forest. This is most likely because the sensor was deployed near the benthos, below the most productive region of the forest. Frieder et al. 2012 observed significantly larger diel pH variability closer to the surface (7 m depth) compared to near the 511 bottom (17 m depth; same as this study), demonstrating that the biologically driven diel cycle 512 diminishes with increasing depth within the canopy. Therefore it is important to keep in mind 513 that the results presented here are not reflecting kelp forest production dynamics, rather, reflect 514 conditions that are experienced by benthic dwelling organisms inside the kelp forest.

515 The trajectories are sensitive to the choice of the emission scenario (Figure 14). Trends 516 are similar at all depths, thus only the mean pCO<sub>2</sub> and  $\Omega_{Ar}$  projections at the shelf break are 517 shown in Figure 14. The highest emission scenario (RCP8.5) diverges from the two intermediate 518 scenarios around 2030, while the lowest emission scenario (RCP2.6) diverges around 2050. The 519 two intermediate scenarios (RCP4.5 and RCP6.0) do not diverge significantly until 2070. The 520 delayed response to different atmospheric CO<sub>2</sub> trajectories occurs because upwelled waters have 521 spent several decades since they were last in contact with the atmosphere (Feely et al., 2008). 522 Therefore the anthropogenic ocean acidification trajectory for the Southern California Bight is 523 already determined for the next several decades, and any mitigation due to changing CO2 524 emissions will be delayed.

The results presented here are site specific, and do not necessarily reflect conditions at all kelp forests, canyon edges, and shelf breaks. However, if sensor pH data and corresponding regional hydrographic surveys are available, then a  $\Delta DIC_{anth}-\sigma_{\theta}$  relationship can be established for that region and applied to the sensor data. For example, this approach can potentially expanded to many regions for the CCS, using the North American Carbon Program West Coast Cruise (Feely et al., 2008) and the  $\Delta DIC_{diseq}$  for the Pacific Ocean (Sabine et al., 2002). If similar data exist, then this approach can be expanded to other upwelling margins as well.

532 The Southern California Bight experiences a steady but weaker degree of upwelling 533 compared to the northern regions of the CCS, where upwelling events are more pronounced

534 (Bograd et al., 2009). These regions could experience more extreme conditions regularly, as well 535 as significantly higher variability of carbonate conditions (Harris et al., 2013). However, such 536 dynamics are poorly understood, and more high-frequency observations of carbonate parameters 537 along this system are needed. Source water properties must be characterized through 538 hydrographic surveys. Alternatively, for regions where such data for source waters are not 539 available, sensor data can be combined with either Global Circulation Model or ROMS outputs. 540 This approach will alleviate the cost associated with characterizing source waters, and to a large 541 degree will incorporate processes such as interannual variability, decadal changes in source 542 water properties, and reduced ventilation. It is critical that inorganic carbon sensors (e.g. pH or 543 pCO<sub>2</sub>) are co-located with basic physical oceanographic measurements (e.g. T and S) to 544 determine source water properties especially for subsurface deployments.

## 545 4.3 Model Assessment

546 The sensitivity of the projected carbonate conditions to the assumptions made in the 547 model is explored here. For example, temperatures observed in 2012 were used to parameterize 548 the model. Sea surface temperature has increased over the past century due to climate change 549 (Smith et al., 2008), and is expected to continue. This will affect the  $CO_2$  equilibrium concentration ( $DIC_{eq}$ ), but the effects are small and will reduce  $DIC_{eq}$  by only several µmol kg<sup>-1</sup>. 550 Both pCO<sub>2</sub> and  $\Omega_{Ar}$  are dependent on in situ temperature; the effects on  $\Omega_{Ar}$  are negligible 551  $(\Delta\Omega/\Delta T < 0.01^{\circ}C^{-1})$ , whereas  $\Delta pCO_2/\Delta T$  increases at higher pCO<sub>2</sub> levels, and can be as large as 552 60  $\mu$ atm °C<sup>-1</sup> at the end of the century, compared to 30  $\mu$ atm °C<sup>-1</sup> at present day at the shelf break. 553 554 These temperature dependencies will affect the mean conditions, but the magnitude of the 555 variability will be relatively unaffected. However, it should be noted that this simple error 556 analysis does not include any biological feedbacks that increased temperature or CO<sub>2</sub> may induce. 557 For example, phase shifts from kelp-dominated to algal turfs might be an outcome of sea surface 558 warming and acidification (Connell and Russell, 2010), with implications for habitat-scale 559 biogeochemical cycling. Likewise, higher temperatures may increase remineralization rates 560 along the path of the subducted water (Rivkin and Legendre, 2001), further enhancing 561 acidification.

562 TA conditions from 2012 were used to calculate pCO<sub>2</sub> and  $\Omega_{Ar}$  for all years. Changes in 563 TA affect the buffer factors of seawater, thus, alterations in TA distribution will either speed up 564 or slow down the progression of ocean acidification. However, trends in TA along the CCS on 565 decadal time scales are unknown due to insufficient data. Reduced ventilation in high latitude 566 seas, altered precipitation patterns, and changes in surface calcification and water-column 567 dissolution rates would all lead to changes in upwelled TA conditions (Fassbender et al., 2011; 568 Lee et al., 2006). Quantifying these processes is difficult and out of the scope of this study. 569 Nevertheless, to demonstrate the magnitude of the uncertainty due to TA, pCO<sub>2</sub> was projected for the year 2100 with a +20  $\mu$ mol kg<sup>-1</sup> bias to TA. The effects were strongly dependent on 570 571 depth: mean pCO<sub>2</sub> was reduced by approximately 240, 130, and 70 µatm at 88 m, 30 m, and the 572 surface, respectively.

Finally, the model presented here projects future carbonate conditions by assuming the dynamics that control the variability at each habitat (e.g. seasonal and episodic upwelling events, internal waves and tides, and biological production and respiration) remain the same as 2012 conditions, and does not account for any variability that occurs on inter-annual to decadal timescales. For example, changes in  $O_2$  and pH on the continental shelf associated with interannual climate events, such as El Niño, have been observed (Nam et al., 2011). However, since 2012 did not correspond with a strong El Niño or La Niña phase, we believe that it was not 580 strongly biased by such events. Furthermore, recent evidence suggests that the proportion of 581 Pacific Equatorial Waters in the California Undercurrent has been increasing over the past 582 several decades, thus modifying the source water properties for upwelled waters onto the 583 continental shelf (Bograd et al., 2015). Since waters of equatorial origin observed between 100-584 500 m are elevated in DIC and lower in  $O_2$  (Bograd et al., 2015), it is expected that the Southern 585 California Bight will experience higher levels of acidification than predicted from this study if 586 this redistribution of water masses of equatorial origin continues. However, at this time, we lack 587 observations with sufficient longevity to predict how climate variability on interannual to 588 decadal time scales might modify the acidification trajectory over the course of the next century. 589 Sustained, high-frequency time series of inorganic carbon parameters are required to elucidate 590 such effects.

# 591 4.4 Implications for Ocean Acidification Research

592 In order to properly assess the impacts of anthropogenic ocean acidification through 593 laboratory manipulation experiments, the control and experimental conditions should accurately 594 reflect the study organism's present-day and future habitat conditions (McElhany and Busch, 595 2013; Reum et al., 2015). The most common control treatment used in ocean acidification 596 experiments for organisms found in the CCS was a pCO<sub>2</sub> value of ~400 µatm, reflecting 597 atmospheric conditions (compiled by Reum et al., 2015). However, our sensor data showed that 598 all subsurface habitats had significantly greater  $pCO_2$  relative to the atmosphere (Table 2). For 599 example, the mean  $pCO_2$  at the kelp forest is about 100 µatm greater than the atmosphere, and 600 routinely experiences conditions of more than 300 µatm above atmospheric. Therefore utilizing 601 atmospheric  $pCO_2$  conditions for control treatments will necessarily underestimate the baseline 602 pCO<sub>2</sub> for organisms collected from subsurface habitats.

603 Recent studies that incorporate natural variability into ocean acidification experiments 604 observed modified responses relative to constant conditions (Dufault et al., 2012; Frieder et al., 605 2014). However, the effect of natural variability on organismal response to ocean acidification, 606 especially through various life stages is still poorly understood. Our model results demonstrate 607 that variability trajectories are also habitat specific. For example, in the kelp forest, the 608 variability, approximated by the s.d., was 93 µatm in 2012, whereas this increased to 202 µatm 609 in 2100 (Table 2). Furthermore, despite having similar mean CO<sub>2</sub> conditions, the largest 610 variability was observed on event time scales in the kelp forest, whereas the dominant variability 611 occurred on tidal and seasonal cycles at the canyon edge. Therefore future ocean acidification 612 studies investigating the effect of natural variability should not only incorporate increasing 613 magnitude into their experimental design, but also consider variability patterns on appropriate 614 time scales.

615 Temperature and O<sub>2</sub> were tightly correlated with carbonate parameters across habitats 616 and various time scales (daily to seasonal) in this study (Figure 15); similar correlation has been 617 documented across the CCS in general (Reum et al., 2014, 2015). These parameters can 618 potentially act as additional stressors (Padilla-Gamiño et al., 2013) or stress reliefs (Gooding et 619 al., 2009) for ocean acidification. However, laboratory experiments incorporating the effects of 620 temperature (Gooding et al., 2009; Padilla-Gamiño et al., 2013) and O<sub>2</sub> (Frieder et al., 2014; 621 Navarro et al. in prep) have just started to be explored for the CCS, and no studies have been 622 conducted that incorporate all three variables in their experimental design. Future studies 623 investigating the synergistic effects of O<sub>2</sub>, temperature, and CO<sub>2</sub> should establish experimental 624 conditions based on environmental data (Figure 15). Although development of systems that can 625 manipulate individual parameters is challenging, important strides have been made to make such

experimental set ups accessible to the community (Bockmon et al., 2013). The development of habitat-specific ocean acidification models provides a link between environment and laboratory to facilitate interpretations of physiological responses to elevated  $CO_2$  in the context of current and future environmental conditions.

630 Discerning habitat-specific CO<sub>2</sub> signatures could lead to the discovery of local 631 populations that are more tolerant of future CO<sub>2</sub> conditions. For example, large high-frequency 632 variability of CO<sub>2</sub> could lead to a greater capacity for physiological and phenotypic plasticity, as 633 organisms are routinely exposed to a wide range of CO<sub>2</sub>. The embryos of Doryteuthis opalescens, 634 an important fishery species in California, can tolerate low pH and O<sub>2</sub>, perhaps due to the fact 635 that they routinely experience a wide range of pH and  $O_2$  (Navarro, 2014). Furthermore, such 636 environmental conditions may be conducive for the existence of high CO<sub>2</sub> tolerant 637 subpopulations, allowing for adaptation to buffer some of the negative effects of ocean 638 acidification (Hofmann and Todgham, 2010). Alternatively, these populations could be living 639 near critical biological thresholds, as has been suggested for the thermal stress of some 640 organisms living in the intertidal (Somero, 2002). A massive failure in an oyster hatchery in 641 Oregon was linked to upwelling of high CO<sub>2</sub> waters during a critical life-stage of oyster larvae 642 (Barton et al., 2012), indicating the existence of  $CO_2$  thresholds for some marine organisms 643 (Bednaršek et al., 2014). However, such thresholds may be dependent on species, life-stage, 644 and/or environmental history. As we begin to realize which populations of species and life stages 645 are living near acidification thresholds versus those that exhibit acidification tolerance, 646 implementation of habitat-specific acidification models can be used as a tool to aid protection, 647 management and remediation efforts of critical marine habitats now and in the future.

648 **5** Conclusions

649 Here we have presented habitat-specific carbonate chemistry projections for four coastal 650 habitats along an upwelling margin. The projections were generated by combining high-651 frequency sensor measurements, a regional empirical relationship for TA, hydrographic survey 652 data to quantify source-water properties of upwelled waters, and the atmospheric CO<sub>2</sub> record. 653 Even though the four habitats were within 5 km of one another, distinct habitat-specific 654 variability signatures and acidification trajectories were observed. These results reveal the 655 existence of highly variable CO<sub>2</sub> signatures within a small geographic area, and the potential for 656 discoveries of habitats that could act as refugia from ocean acidification. Changes in the buffer 657 factors largely explained the observed patterns; however, local biological feedbacks could also 658 produce a large acidification signal. In all habitats studied, carbonate conditions have left, or are 659 leaving preindustrial variability envelopes. Model projections suggest that anthropogenic ocean 660 acidification will continue to progress in the CCS and other upwelling margins over the next 661 several decades regardless of any changes in CO<sub>2</sub> emissions; any impacts from reduced 662 emissions will only be observed mid-century and beyond. This demonstrates the urgency of the 663 situation, and this delayed response must be taken into account when assessing the impacts of 664 ocean acidification and developing mitigation and monitoring strategies.

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## 691 Literature Cited

Alin, S. R., Feely, R. A., Dickson, A. G., Hernández-Ayón, J. M., Juranek, L. W., Ohman, M. D.
and Goericke, R.: Robust empirical relationships for estimating the carbonate system in the
southern California Current System and application to CalCOFI hydrographic cruise data (2005–
2011), J. Geophys. Res., 117(C5), C05033, doi:10.1029/2011JC007511, 2012.

Anderson, L. A. and Sarmiento, J. L.: Redfield ratios of remineralization determined by nutrient
 data analysis, Global Biogeochem. Cycles, 8(1), 65–80, 1994.

Andersson, A. J., Yeakel, K. L., Bates, N. R. and de Putron, S. J.: Partial offsets in ocean acidification from changing coral reef biogeochemistry, Nat. Clim. Chang., 3(11), 1–6, doi:10.1038/nclimate2050, 2013.

Barton, A., Hales, B., Waldbusser, G. G., Langdon, C. and Feely, R. A.: The Pacific oyster,
Crassostrea gigas, shows negative correlation to naturally elevated carbon dioxide levels:
Implications for near-term ocean acidification effects, Limnol. Oceanogr., 57(3), 698–710,
doi:10.4319/lo.2012.57.3.0698, 2012.

Bates, N. R., Astor, Y. M., Church, M. J., Currie, K., Dore, J. E., Gonzalez-Davila, M.,
Lorenzoni, L., Muller-Karger, F., Olafsson, J. and Santana-Casiano, J. M.: A time-series view of
changing surface ocean chemistry due to ocean uptake of anthropogenic CO2 and ocean
acidification, Oceanography, 27(1), 126–141, 2014.

Bednaršek, N., Feely, R. A., Reum, J. C. P., Peterson, B., Menkel, J., Alin, S. R. and Hales, B.:
Limacina helicina shell dissolution as an indicator of declining habitat suitability owing to ocean
acidification in the California Current Ecosystem., Proc. R. Soc. B Biol. Sci., 281(1785),
20140123, doi:10.1098/rspb.2014.0123, 2014.

Bockmon, E. E., Frieder, C. A., Navarro, M. O., White-Kershek, L. A. and Dickson, A. G.:
Technical note: controlled experimental aquarium system for multi-stressor investigation of
carbonate chemistry, oxygen saturation, and temperature, Biogeosciences, 10(9), 5967–5975,
doi:10.5194/bg-10-5967-2013, 2013.

Bograd, S. J., Buil, M. P., Lorenzo, E. Di, Castro, C. G., Schroeder, I. D., Goericke, R.,
Anderson, C. R., Benitez-Nelson, C. and Whitney, F. A.: Changes in source waters to the
Southern California Bight, Deep Sea Res. Part II Top. Stud. Oceanogr., 112, 42–52,
doi:10.1016/j.dsr2.2014.04.009, 2015.

Bograd, S. J., Schroeder, I., Sarkar, N., Qiu, X., Sydeman, W. J. and Schwing, F. B.: Phenology
of coastal upwelling in the California Current, Geophys. Res. Lett., 36(1), 1–5,
doi:10.1029/2008GL035933, 2009.

Booth, J. A. T., McPhee-Shaw, E. E., Chua, P., Kingsley, E., Denny, M., Phillips, R., Bograd, S.
J., Zeidberg, L. D. and Gilly, W. F.: Natural intrusions of hypoxic, low pH water into nearshore

- 726 marine environments on the California coast, Cont. Shelf Res., 45, 108–115, 727 doi:10.1016/j.csr.2012.06.009, 2012.
- Bresnahan, P. J., Martz, T. R., Takeshita, Y., Johnson, K. S. and LaShomb, M.: Best practices for
  autonomous measurement of seawater pH with the Honeywell Durafet, Methods Oceanogr., 9,
  44–60, 2014.
- Brewer, P. G. and Peltzer, E. T.: Limits to marine life, Science (80-.)., 324(5925), 347–348,
  2009.
- Byrne, M., Lamare, M., Winter, D., Dworjanyn, S. A. and Uthicke, S.: The stunting effect of a
  high CO2 ocean on calcification and development in sea urchin larvae, a synthesis from the
  tropics to the poles, Philos. Trans. R. Soc. B Biol. Sci., 368(1627), 1–13,
  doi:10.1098/rstb.2012.0439, 2013.
- Byrne, R. H., Mecking, S., Feely, R. A. and Liu, X.: Direct observations of basin-wide 737 738 the North Pacific Ocean, Geophys. Res. Lett.. acidification of 37(2), 1-5.739 doi:10.1029/2009GL040999, 2010.
- Cai, W.-J., Hu, X., Huang, W.-J., Murrell, M. C., Lehrter, J. C., Lohrenz, S. E., Chou, W.-C.,
  Zhai, W., Hollibaugh, J. T., Wang, Y., Zhao, P., Guo, X., Gundersen, K., Dai, M. and Gong, G.C.: Acidification of subsurface coastal waters enhanced by eutrophication, Nat. Geosci., 4(11),
  766–770, doi:10.1038/ngeo1297, 2011.
- Carter, B. R., Radich, J. A., Doyle, H. L. and Dickson, A. G.: An automated system for
  spectrophotometric seawater pH measurements, Limnol. Oceanogr. Methods, 11, 16–27,
  doi:10.4319/lom.2013.11.16, 2013.
- Clayton, T. D. and Byrne, R. H.: Spectrophotometric seawater pH measurements: total hydrogen
  ion concentration scale calibration of m-cresol purple and at-sea results, Deep Sea Res. Part I
  Oceanogr. Res. Pap., 40(10), 2115–2129, 1993.
- Connell, S. D. and Russell, B. D.: The direct effects of increasing CO2 and temperature on noncalcifying organisms: increasing the potential for phase shifts in kelp forests., Proc. Biol. Sci.,
  277(1686), 1409–1415, doi:10.1098/rspb.2009.2069, 2010.
- Dickson, A. G., Sabine, C. L. and Christian, J. R., Eds.: Guide to best practices for ocean CO2
   measurements, PICES Special Publication 3., 2007.
- Duarte, C. M., Hendriks, I. E., Moore, T. S., Olsen, Y. S., Steckbauer, A., Ramajo, L.,
  Carstensen, J., Trotter, J. A. and McCulloch, M.: Is ocean acidification an open-ocean
  syndrome? Understanding anthropogenic impacts on seawater pH, Estuaries and Coasts, 36(2),
  221–236, doi:10.1007/s12237-013-9594-3, 2013.

- 759 Dufault, A. M., Cumbo, V. R., Fan, T.-Y. and Edmunds, P. J.: Effects of diurnally oscillating
- pCO2 on the calcification and survival of coral recruits, Proc. R. Soc. B Biol. Sci., 279(1740),
  2951–2958, doi:10.1098/rspb.2011.2545, 2012.
- 762 Emerson, S. R.: Accurate measurement of O2, N2, and Ar gases in water and the solubility of N2,
  763 Mar. Chem., 64(4), 337–347, doi:10.1016/S0304-4203(98)00090-5, 1999.
- Fassbender, A. J., Sabine, C. L., Feely, R. A., Langdon, C. and Mordy, C. W.: Inorganic carbon
  dynamics during northern California coastal upwelling, Cont. Shelf Res., 31(11), 1180–1192,
  doi:10.1016/j.csr.2011.04.006, 2011.
- Feely, R. A., Alin, S. R., Newton, J., Sabine, C. L., Warner, M., Devol, A., Krembs, C. and
  Maloy, C.: The combined effects of ocean acidification, mixing, and respiration on pH and
  carbonate saturation in an urbanized estuary, Estuar. Coast. Shelf Sci., 88(4), 442–449,
  doi:10.1016/j.ecss.2010.05.004, 2010.
- Feely, R. A., Doney, S. C. and Cooley, S. R.: Ocean acidification: present conditions and future changes in a high-CO2 world, Oceanography, 22(4), 36–47, 2009.
- Feely, R. A., Sabine, C. L., Hernandez-Ayon, J. M., Ianson, D. and Hales, B.: Evidence for upwelling of corrosive "acidified" water onto the continental shelf., Science, 320(5882), 1490– 1492, doi:10.1126/science.1155676, 2008.
- 775 1472, doi:10.1120/selence.1155070, 2000.
- Frankignoulle, M.: A complete set of buffer factors for acid/base CO2 system in seawater, J. Mar.
  Syst., 5(2), 111–118, 1994.
- Frieder, C. A., Gonzalez, J. P., Bockmon, E. E., Navarro, M. O. and Levin, L. A.: Can variable
  pH and low oxygen moderate ocean acidification outcomes for mussel larvae?, Glob. Chang.
  Biol., 20(3), 754–64, doi:10.1111/gcb.12485, 2014.
- Frieder, C. A., Nam, S., Martz, T. R. and Levin, L. A.: High temporal and spatial variability of
  dissolved oxygen and pH in a nearshore California kelp forest, Biogeosciences, 9(10), 3917–
  3930, doi:10.5194/bg-9-3917-2012, 2012.
- Friederich, G. E., Walz, P. M., Burczynski, M. G. and Chavez, F. P.: Inorganic carbon in the
  central California upwelling system during the 1997 1999 El Niño–La Niña event, Prog.
  Oceanogr., 54(1-4), 185–203, 2002.
- Garcia, H. E. and Gordon, L. I.: Oxygen solubility in seawater: better fitting equations, Limnol.
  Oceanogr., 37(6), 1307–1312, 1992.
- Gaylord, B., Hill, T. M., Sanford, E., Lenz, E. A., Jacobs, L. A., Sato, K. N., Russell, A. D. and
  Hettinger, A.: Functional impacts of ocean acidification in an ecologically critical foundation
  species, J. Exp. Biol., 214(Pt 15), 2586–2594, 2011.

- Gazeau, F., Gattuso, J.-P., Greaves, M., Elderfield, H., Peene, J., Heip, C. H. R. and Middelburg,
- J. J.: Effect of carbonate chemistry alteration on the early embryonic development of the Pacific
- 794 oyster (Crassostrea gigas), PLoS One, 6(8), e23010, doi:10.1371/journal.pone.0023010, 2011.
- Gooding, R. A., Harley, C. D. G. and Tang, E.: Elevated water temperature and carbon dioxide
  concentration increase the growth of a keystone echinoderm, Proc. Natl. Acad. Sci. U. S. A.,
  106(23), 9316–9321, doi:10.1073/pnas.0811143106, 2009.
- Gruber, N., Hauri, C., Lachkar, Z., Loher, D., Frölicher, T. L. and Plattner, G.-K.: Rapid
  progression of ocean acidification in the California Current System, Science (80-.)., 337(6091),
  220–223, doi:10.1126/science.1216773, 2012.
- Gruber, N., Lachkar, Z., Frenzel, H., Marchesiello, P., Münnich, M., McWilliams, J. C., Nagai, T.
   and Plattner, G.-K.: Eddy-induced reduction of biological production in eastern boundary
- 803 upwelling systems, Nat. Geosci., 4(11), 787–792, doi:10.1038/ngeo1273, 2011.
- Gruber, N., Sarmiento, J. L. and Stocker, T. F.: An improved method for detecting anthropogenic
  CO2 in the oceans, Global Biogeochem. Cycles, 10(4), 809–837, 1996.
- Hales, B., Takahashi, T. and Bandstra, L.: Atmospheric CO2 uptake by a coastal upwelling
  system, Global Biogeochem. Cycles, 19(1), 1–11, doi:10.1029/2004GB002295, 2005.
- Hall-Spencer, J. M., Rodolfo-Metalpa, R., Martin, S., Ransome, E., Fine, M., Turner, S. M.,
  Rowley, S. J., Tedesco, D. and Buia, M.-C.: Volcanic carbon dioxide vents show ecosystem
  effects of ocean acidification, Nature, 454(7200), 96–99, 2008.
- Harris, K. E., DeGrandpre, M. D. and Hales, B.: Aragonite saturation state dynamics in a coastal
  upwelling zone, Geophys. Res. Lett., 40(11), 2720–2725, doi:10.1002/grl.50460, 2013.
- Hauri, C., Gruber, N., McDonnell, A. M. P. and Vogt, M.: The intensity, duration, and severity
  of low aragonite saturation state events on the California continental shelf, Geophys. Res. Lett.,
  40(13), 3424–3428, doi:10.1002/grl.50618, 2013a.
- Hauri, C., Gruber, N., Vogt, M., Doney, S. C., Feely, R. A., Lachkar, Z., Leinweber, A.,
  McDonnel, A. M. P., Munnich, M. and Plattner, G.-K.: Spatiotemporal variability and long-term
  trends of ocean acidification in the California Current System, Biogeosciences, 10(1), 193–216,
  2013b.
- Hettinger, A., Sanford, E., Hill, T. M., Russell, A. D., Sato, K. N. S., Hoey, J., Forsch, M., Page,
  H. N. and Gaylord, B.: Persistent carry-over effects of planktonic exposure to ocean acidification
  in the Olympia oyster, Ecology, 93(12), 2758–2768, doi:10.1890/12-0567.1, 2012.
- Van Heuven, S., Pierrot, D., Rae, J. W. B., Lewis, E. and Wallace, D. W. .: MATLAB Program Developed for CO2 System Calculations. ORNL/CDIAC-105b., , doi:10.3334/CDIAC/otg.CO2SXS\_MATLAB\_v1.1\_2011
- 825 doi:10.3334/CDIAC/otg.CO2SYS\_MATLAB\_v1.1, 2011.

- 826 Hijioka, Y., Matsuoka, Y., Nishimoto, H., Masui, M. and Kainuma, M.: Global GHG emissions
- scenarios under GHG concentration stabilization targets, J. Glob. Environ. Eng., 13, 97–108,
- 828 2008.

Hofmann, G. E., Evans, T. G., Kelly, M. W., Padilla-Gamiño, J. L., Blanchette, C. A., Washburn,
L., Chan, F., McManus, M. A., Menge, B. A., Gaylord, B., Hill, T. M., Sanford, E., LaVigne, M.,
Rose, J. M., Kapsenberg, L. and Dutton, J. M.: Exploring local adaptation and the ocean
acidification seascape – studies in the California Current Large Marine Ecosystem,
Biogeosciences, 11(4), 1053–1064, doi:10.5194/bg-11-1053-2014, 2014.

Hofmann, G. E., Smith, J. E., Johnson, K. S., Send, U., Levin, L. A., Micheli, F., Paytan, A.,
Price, N. N., Peterson, B., Takeshita, Y., Matson, P. G., Crook, E. D., Kroeker, K. J., Gambi, M.
C., Rivest, E. B., Frieder, C. A., Yu, P. C. and Martz, T. R.: High-frequency dynamics of ocean
pH: a multi-ecosystem comparison, edited by W.-C. Chin, PLoS One, 6(12), e28983,
doi:10.1371/journal.pone.0028983, 2011.

Hofmann, G. E. and Todgham, A. E.: Living in the now: physiological mechanisms to tolerate a
rapidly changing environment., Annu. Rev. Physiol., 72, 127–45, doi:10.1146/annurev-physiol021909-135900, 2010.

Keeling, C. D., Piper, S. C., Bacastow, R. B., Wahlen, M., Whorf, T. P., Heimann, M. and
Meijer, H. A.: Atmospheric CO2 and 13CO2 exchange with the terrestrial biosphere and oceans
from 1978 to 2000: observations and carbon cycle implications, in A history of atmospheric CO2
and its effects on plants, animals, and ecosystems, edited by J. R. Ehleringer, T. E. Cerling, and
M. D. Dearing, pp. 83–113, Springer Verlag, New York., 2005.

Kroeker, K. J., Micheli, F., Gambi, M. C. and Martz, T. R.: Divergent ecosystem responses
within a benthic marine community to ocean acidification, Proc. Natl. Acad. Sci. U. S. A.,
108(35), 14515–14520, doi:10.1073/pnas.1107789108, 2011.

Langdon, C., Gattuso, J. and Andersson, A.: Measurements of calcification and dissolution of
benthic organisms and communities, in Guide to Best Practices for Ocean Acidificaton Research
and Data Reporting, pp. 213–232, Publications Office of the European Union, Luxembourg.,
2010.

Lauvset, S. K. and Gruber, N.: Long-term trends in surface ocean pH in the North Atlantic, Mar.
Chem., 162, 71–76, doi:10.1016/j.marchem.2014.03.009, 2014.

Lee, K., Tong, L. T., Millero, F. J., Sabine, C. L., Dickson, A. G., Goyet, C., Park, G.-H.,
Wanninkhof, R., Feely, R. A. and Key, R. M.: Global relationships of total alkalinity with
salinity and temperature in surface waters of the world's oceans, Geophys. Res. Lett., 33(19), 1–
5, doi:10.1029/2006GL027207, 2006.

- Leinweber, A. and Gruber, N.: Variability and trends of ocean acidification in the Southern California Current System: a time series from Santa Monica Bay, J. Geophys. Res. Ocean.,
- 862 118(7), 3622–3633, doi:10.1002/jgrc.20259, 2013.

- Leinweber, A., Gruber, N., Frenzel, H., Friederich, G. E. and Chavez, F. P.: Diurnal carbon cycling in the surface ocean and lower atmosphere of Santa Monica Bay, California, Geophys. Res. Lett., 36(8), 3–7, doi:10.1029/2008GL037018, 2009.
- Liu, X., Patsavas, M. C. and Byrne, R. H.: Purification and characterization of meta-cresol
  purple for spectrophotometric seawater pH measurements, Environ. Sci. Technol., 45(11), 4862–
  4868, doi:10.1021/es200665d, 2011.
- Lueker, T. J., Dickson, A. G. and Keeling, C. D.: Ocean pCO2 calculated from dissolved inorganic carbon, alkalinity, and equations for K1 and K2: validation based on laboratory measurements of CO2 in gas and seawater at equilibrium, Mar. Chem., 70(1-3), 105–119, doi:10.1016/S0304-4203(00)00022-0, 2000.
- Martz, T. R., Connery, J. G. and Johnson, K. S.: Testing the Honeywell Durafet for seawater pH
  applications, Limnol. Oceanogr. Methods, 8, 172–184, doi:10.4319/lom.2010.8.172, 2010.
- 875 Martz, T. R., Send, U., Ohman, M. D., Takeshita, Y., Bresnahan, P., Kim, H.-J. and Nam, S.:
- 876 Dynamic variability of biogeochemical ratios in the Southern California Current System,
- 877 Geophys. Res. Lett., 41(7), 2496–2501, doi:10.1002/2014GL059332, 2014.
- Martz, T. R., Takeshita, Y., Rolph, R. and Bresnahan, P.: Tracer monitored titrations:
  measurement of dissolved oxygen., Anal. Chem., 84(1), 290–296, doi:10.1021/ac202537f, 2012.
- 880 Matsumoto, K. and Gruber, N.: How accurate is the estimation of anthropogenic carbon in the 881 ocean? An evaluation of the  $\Delta C^*$  method, Global Biogeochem. Cycles, 19(3), 1–17, 882 doi:10.1029/2004GB002397, 2005.
- McElhany, P. and Busch, D. S.: Appropriate pCO2 treatments in ocean acidification experiments,
  Mar. Biol., 160(8), 1807–1812, doi:10.1007/s00227-012-2052-0, 2013.
- McNeil, B. I. and Matear, R. J.: Southern Ocean acidification: a tipping point at 450-ppm
  atmospheric CO2, Proc. Natl. Acad. Sci. U. S. A., 105(48), 18860–18864,
  doi:10.1073/pnas.0806318105, 2008.
- Mehrbach, C., Culberson, C. H., Hawley, J. E. and Pytkowicz, R. M.: Measurement of the
  apparent dissociation constants of carbonic acid in seawater at atmospheric pressure, Limnol.
  Oceanogr., 18(6), 897–907, doi:10.4319/lo.1973.18.6.0897, 1973.
- Nam, S., Kim, H.-J. and Send, U.: Amplification of hypoxic and acidic events by La Niña
  conditions on the continental shelf off California, Geophys. Res. Lett., 38(22), 1–5,
  doi:10.1029/2011GL049549, 2011.
- Nam, S., Takeshita, Y., Frieder, C. A., Martz, T. and Ballard, J.: Seasonal advection of Pacific
- Equatorial Water alters oxygen and pH in the Southern California Bight, J. Geophys. Res.
  Ocean., 120, n/a–n/a, doi:10.1002/2015JC010859, 2015.

- Navarro, M. O.: Consequences of environmental variability for spawning and embryo
  development of inshore market squid Doryteuthis opalescens, Ph.D. thesis, Unviersity of
  California San Diego. Ann Arbor, USA, ProQuest Dissertations and Theses (Publication No.
  3644287)., 2014.
- Navarro, M. O., Parnell, P. E. and Levin, L. A.: Year-round spawning of the Market Squid,
  Doryteuthis opalescens and associated critical habitat along an upwelling margin, in Coastal and
  Estuarine Research Federation National Conference, San Diego., 2013.
- O'Sullivan, D. W. and Millero, F. J.: Continual measurement of the total inorganic carbon in surface seawater, Mar. Chem., 60(1-2), 75–83, doi:10.1016/S0304-4203(97)00079-0, 1998.
- Padilla-Gamiño, J. L., Kelly, M. W., Evans, T. G. and Hofmann, G. E.: Temperature and CO2
  additively regulate physiology, morphology and genomic responses of larval sea urchins,
  Strongylocentrotus purpuratus, Proc. R. Soc. B Biol. Sci., 280(1759), 20130155,
  doi:10.1098/rspb.2013.0155, 2013.
- Pineda, J.: Predictable upwelling and the shoreward transport of planktonic larvae by internal
  tidal bores, Science (80-.)., 253(5019), 548–551, 1991.
- Price, N. N., Martz, T. R., Brainard, R. E. and Smith, J. E.: Diel variability in seawater pH relates
  to calcification and benthic community structure on coral reefs, PLoS One, 7(8), e43843,
  doi:10.1371/journal.pone.0043843, 2012.
- Reum, J. C. P., Alin, S. R., Feely, R. A., Newton, J., Warner, M. and McElhany, P.: Seasonal
  carbonate chemistry covariation with temperature, oxygen, and salinity in a fjord estuary:
  Implications for the design of ocean acidification experiments, PLoS One, 9(2), e89619,
  doi:10.1371/journal.pone.0089619, 2014.
- Reum, J. C. P., Alin, S. R., Harvey, C. J., Bednarsek, N., Evans, W., Feely, R. A., Hales, B.,
  Lucey, N., Mathis, J. T., McElhany, P., Newton, J., Newton, J. and Sabine, C. L.: Interpretation
  and design of ocean acidification experiments in upwelling systems in the context of carbonate
  chemistry co-variation with temperature and oxygen, ICES J. Mar. Sci., 528–536,
  doi:10.1093/icesjms/fsu231, 2015.
- Rivkin, R. B. and Legendre, L.: Biogenic carbon cycling in the upper ocean: effects of microbial
  respiration., Science, 291(5512), 2398–2400, doi:10.1126/science.291.5512.2398, 2001.
- Rodolfo-Metalpa, R., Houlbrèque, F., Tambutté, É., Boisson, F., Baggini, C., Patti, F. P., Jeffree,
  R., Fine, M., Foggo, a., Gattuso, J.-P. and Hall-Spencer, J. M.: Coral and mollusc resistance to
  ocean acidification adversely affected by warming, Nat. Clim. Chang., 1(6), 308–312,
  doi:10.1038/nclimate1200, 2011.
- 930 Rykaczewski, R. R. and Dunne, J. P.: Enhanced nutrient supply to the California Current
- Ecosystem with global warming and increased stratification in an earth system model, Geophys.
  Res. Lett., 37(21), 1–5, doi:10.1029/2010GL045019, 2010.

- 933 Sabine, C. L., Feely, R. A., Key, R. M., Bullister, J. L., Millero, F. J., Lee, K., Peng, T.-H.,
- Tilbrook, B., Ono, T. and Wong, C. S.: Distribution of anthropogenic CO2 in the Pacific Ocean,
  Global Biogeochem. Cycles, 16(4), 1–17, doi:10.1029/2001GB001639, 2002.
- Sabine, C. L. and Tanhua, T.: Estimation of anthropogenic CO2 inventories in the ocean, Ann.
  Rev. Mar. Sci., 2(1), 175–198, doi:10.1146/annurev-marine-120308-080947, 2010.
- 938 Send, U. and Nam, S.: Relaxation from upwelling: the effect on dissolved oxygen on the 939 continental shelf, J. Geophys. Res., 117(4), 1–9, doi:10.1029/2011JC007517, 2012.
- Smith, T. M., Reynolds, R. W., Peterson, T. C. and Lawrimore, J.: Improvements to NOAA's
  historical merged land–ocean surface temperature analysis (1880–2006), J. Clim., 21(10), 2283–
  2296, doi:10.1175/2007JCLI2100.1, 2008.
- Somero, G. N.: Thermal physiology and vertical zonation of intertidal animals: optima, limits,
  and costs of living, Integr. Comp. Biol., 42(4), 780–789, 2002.
- Sunda, W. G. and Cai, W.-J.: Eutrophication induced CO2-acidification of subsurface coastal
  waters: interactive effects of temperature, salinity, and atmospheric pCO2, Environ. Sci.
  Technol., 46(19), 10651–10659, doi:10.1021/es300626f, 2012.
- Sutton, A. J., Feely, R. A., Sabine, C. L., McPhaden, M. J., Takahashi, T., Chavez, F. P.,
  Friederich, G. E. and Mathis, J. T.: Natural variability and anthropogenic change in equatorial
  Pacific surface ocean pCO2 and pH, Global Biogeochem. Cycles, 28(2), 1–15,
  doi:10.1002/2013GB004679, 2014.
- Swart, N. C., Allen, S. E. and Greenan, B. J. W.: Resonant amplification of subinertial tides in a
  submarine canyon, J. Geophys. Res., 116(9), 1–14, doi:10.1029/2011JC006990, 2011.
- Waldbusser, G. G., Hales, B., Langdon, C. J., Haley, B. A., Schrader, P., Brunner, E. L., Gray, M.
  W., Miller, C. A. and Gimenez, I.: Saturation-state sensitivity of marine bivalve larvae to ocean
  acidification, Nat. Clim. Chang., 5, 273–280, doi:10.1038/NCLIMATE2479, 2015.
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Table 1: Summary of sensor deployments

| Habitat Type             | Deployment Site      | DD <sup>a</sup> | $BD^b$ | Latitude | Longitude | Days <sup>c</sup> |
|--------------------------|----------------------|-----------------|--------|----------|-----------|-------------------|
| Surf Zone                | Scripps Pier         | 4               | 6      | 32.87°N  | 117.26°W  | 122               |
| Kelp Forest              | La Jolla Kelp Forest | 17              | 20     | 32.81°N  | 117.29°W  | 128               |
| Canyon Edge <sup>d</sup> | La Jolla Canyon      | 30              | 30     | 32.86°N  | 117.27°W  | 302               |
| Shelf Break              | Del Mar Buoy         | 88              | 100    | 32.94°N  | 117.32°W  | 335               |

<sup>a</sup> Sensor deployment depth in meters
<sup>b</sup> Bottom depth in meters
<sup>c</sup> Total deployment days between June 2012 and June 2013.
<sup>d</sup> A linear drift correction for salinity was applied for 2 of the 4 deployments. 

|                 | Year    | pCO <sub>2</sub> (µatm) | $\Omega_{ m Ar}$ | pH               |
|-----------------|---------|-------------------------|------------------|------------------|
| Surf Zone       | Preind. | $267\pm26$              | $3.09\pm0.21$    | $8.19\pm0.034$   |
| ( <b>4</b> m)   | 2012    | $394\pm43$              | $2.38\pm0.25$    | $8.05\pm0.038$   |
|                 | 2060    | $473\pm56$              | $2.09\pm0.19$    | $7.98 \pm 0.041$ |
|                 | 2100    | $619\pm80$              | $1.71\pm0.18$    | $7.88\pm0.045$   |
| Kelp Forest     | Preind. | $365 \pm 74$            | $2.28\pm0.42$    | $8.08\pm0.078$   |
| (17 m)          | 2012    | $516\pm108$             | $1.77\pm0.36$    | $7.95\pm0.083$   |
|                 | 2060    | $683 \pm 156$           | $1.43\pm0.33$    | $7.84\pm0.094$   |
|                 | 2100    | $937\pm231$             | $1.11\pm0.29$    | $7.72\pm0.105$   |
| Canyon Edge     | Preind. | $365 \pm 68$            | $2.29\pm0.37$    | $8.08\pm0.068$   |
| ( <b>30</b> m)  | 2012    | $529 \pm 105$           | $1.75\pm0.31$    | $7.94\pm0.075$   |
|                 | 2060    | $702\pm155$             | $1.40\pm0.29$    | $7.83\pm0.085$   |
|                 | 2100    | $964\pm231$             | $1.09\pm0.25$    | $7.70\pm0.095$   |
| Shelf Break     | Preind. | $637 \pm 132$           | $1.38\pm0.27$    | $7.86 \pm 0.083$ |
| ( <b>88 m</b> ) | 2012    | $878 \pm 149$           | $1.05\pm0.18$    | $7.73\pm0.070$   |
|                 | 2060    | $1195\pm200$            | $0.80\pm0.15$    | $7.61\pm0.070$   |
|                 | 2100    | $1639\pm246$            | $0.60\pm0.10$    | $7.47\pm0.065$   |

Table 2. Mean ± s.d of modeled carbonate parameters at in situ conditions for preindustrial, 2012,
2060, and 2100 using the RCP6.0 projection at each habitat.

|                    | pCO <sub>2</sub> (µatm) | $\Omega_{\rm Ar}$ | pН    |
|--------------------|-------------------------|-------------------|-------|
| Surf Zone          | 96                      | 0.46              | 0.085 |
| Kelp Forest        | 68                      | 0.22              | 0.054 |
| <b>Canyon Edge</b> | 167                     | 0.48              | 0.120 |
| Shelf Break        | 110                     | 0.14              | 0.053 |

968 Table 3. Mean daily range of carbonate parameters at in situ conditions for 2012 at each habitat.



Figure 1: Map of study region. Hydrographic stations (black dots) and sensor deployment sites (black squares) are shown. Initials are: CE = canyon edge, SB = shelf break, SZ = surf zone, and 

- KF = kelp forest.



978 Figure 2:  $\Delta DIC_{anth}$  as a function of  $\sigma_{\theta}$  for certain modeled years (indicated above line) using the

- 979 ICPP RCP6.0 projection. The three regimes used in this model, surface, mixing, and subsurface,
- 980 are labeled.
- 981



983Figure 3:  $f CO_{2,eq}^{2012-age}$  calculated from hydrographic data (left) and the calculated age-σθ984relationship (right) is shown. Good agreement (R2 = 0.87) between the data (black circles) and985the fit (age =  $8.852(\sigma\theta - 25.2)2 + 23.132(\sigma\theta - 25.2)$ ; red line) is observed (right). Age of surface986waters ( $\sigma_{\theta} < 25.2$  kg m<sup>-3</sup>) was assumed to be 0.987



Figure 4:  $DIC_{anth}$  as a function of  $\sigma_{\theta}$ . The calculated values and the fit are represented by black

992 circles and a red line, respectively. The blue line shows  $DIC_{anth}$  using the formulations from 993 Feely et al. (2008).



Figure 5: Time series of sensor pH between June 2012 to June. Two lower panels are month-longsnapshots. pH is reported at in situ conditions.



Figure 6: Time series of pCO<sub>2</sub> calculated from sensor pH and *TA*<sup>est</sup> between June 2012 to June
2013. Two lower panels are month-long snapshots. pCO<sub>2</sub> is reported at in situ conditions.



1004 1005 Figure 7: Time series of  $\Omega_{Ar}$  calculated from sensor pH and  $TA^{est}$  between June 2012 to June 1006 2013. Two lower panels are month-long snapshots.  $\Omega_{Ar}$  is reported at in situ conditions. 1007



Figure 8: Observed pCO<sub>2</sub> in 2012 (black) and modeled pCO<sub>2</sub> using the IPCC RCP 6.0 scenario

for the year 2100 (red) at the Del Mar Buoy (88 m) over an annual cycle (top). A close up for the 

month of December is shown on the bottom. Note that the range, but not the absolute values, of the vertical axes for each figure is the same.



1015 Figure 9: Observed  $\Omega_{Ar}$  in 2012 (black) and modeled  $\Omega_{Ar}$  using the IPCC RCP6.0 scenario for

1016 the year 2100 (red) at the shelf break (88 m) over an annual cycle (top). A close up for the month

1017 of December is shown on the bottom. Dashed lines represent  $\Omega_{Ar} = 1$ . Note that the range, but

1018 not the absolute values, of the vertical axes for each figure is the same.



1020 1021 Figure 10: Observed (black) and modelled (red) pCO<sub>2</sub> (top) and  $\Omega_{Ar}$  (bottom) at the La Jolla kelp

1022 forest (17 m). Modelled values correspond to projected values in 2100 using the IPCC RCP6.0 1023 scenario. Note that the range, but not the absolute values, of the vertical axes for each figure is

- 1024 the same.
- 1025





1027Figure 11: Histogram of modeled  $pCO_2$  (left) and  $\Omega_{Ar}$  (right) distribution at the four depths for1028preindustrial (black), 2012 (green), 2060 (blue), and 2100 (red). Atmospheric  $pCO_2$  for the years10292060 and 2100 roughly correspond to 510 and 670 µatm based on the IPCC RCP6.0 scenario.





1032Figure 12: Projected pCO2 (left) and  $\Omega_{Ar}$  (right) between 2013 and 2100 under the IPCC RCP6.01033scenario. The solid line and the shaded region represent the mean and  $\pm 1$  s.d., respectively.1034





Figure 13: Buffer factor  $\Pi$  for pCO<sub>2</sub> (A), CO<sub>3</sub><sup>2-</sup> (B), pH (C), and [H<sup>+</sup>] (D) as a function of DIC. Note the inverted y axis in panels B and C. Model parameters to calculate the  $\Pi$  values were: TA = 2240 µmol kg<sup>-1</sup>, temperature = 10 °C, salinity = 33.5, pressure = 1.013 bar.





1041 Figure 14: Projections of mean pCO<sub>2</sub> (left) and  $\Omega_{Ar}$  (right) at the shelf break (88 m) based on

1042 four projections from the Fifth Assessment of the IPCC.



1045 Figure 15: pCO<sub>2</sub> as a function of O<sub>2</sub> (left) and temperature (right) from the kelp forest and shelf

1046 break. Data observed in 2012 and projected for 2100 are plotted.

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