

1 **Nitrogen fixation and the diazotroph community in the**
2 **temperate coastal region of the northwestern North Pacific**

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10
11 **Abstract**

12 Nitrogen fixation in temperate oceans is a potentially important, but poorly understood
13 process that may influence the marine nitrogen budget. This study determined seasonal
14 variations in nitrogen fixation and the diazotroph community within the euphotic zone in the
15 temperate coastal region of the northwestern North Pacific. Nitrogen fixation as high as
16 13.6 nmol N L⁻¹ d⁻¹ was measured from early summer to fall when the surface temperature
17 exceeded 14.2° C (but was lower than 24.3° C) and the surface nitrate concentration was low
18 (≤0.30 μM), although we also detected nitrogen fixation in subsurface layers (42–62 m)

19 where nitrate concentrations were high ($>1 \mu\text{M}$). Clone library analysis results indicated
20 that *nifH* gene sequences were omnipresent throughout the investigation period. During the
21 period when nitrogen fixation was detected (early summer to fall), the genes affiliated with
22 UCYN-A, *Trichodesmium*, and γ -proteobacterial phylotype γ -24774A11 were frequently
23 recovered. In contrast, when nitrogen fixation was undetectable (winter to spring), many
24 sequences affiliated with Cluster III diazotrophs (putative anaerobic bacteria) were recovered.
25 Quantitative PCR analysis revealed that UCYN-A was relatively abundant from early to late
26 summer compared with *Trichodesmium* and γ -24774A11, whereas *Trichodesmium* abundance
27 was the highest among the three groups during fall.

28

29 **1. Introduction**

30 The amount of bioavailable nitrogen introduced into the global ocean via nitrogen fixation is
31 considered to be roughly balanced at the large spatiotemporal scale by nitrogen loss through
32 denitrification, as indicated by the sedimentary nitrogen isotope record during the Holocene
33 epoch (Brandes and Devol, 2002; Deutsch et al., 2004). However, rate measurement data
34 have revealed that denitrification far exceeds nitrogen fixation (Codispoti, 2007). This
35 discrepancy in the nitrogen balance has raised the possibility that the current estimate of
36 marine nitrogen fixation, which is primarily based on data collected in tropical and

37 subtropical oceans where large cyanobacterial diazotrophs (e.g., *Trichodesmium* spp. and
38 *Richelia intracellularis*) are considered to be mainly responsible for nitrogen fixation (e.g.,
39 Capone et al., 1997), might be too low (Codispoti, 2007). This is supported by the results of
40 recent studies using molecular approaches that have increasingly revealed that marine
41 diazotrophs are more diverse and widespread than previously thought (Riemann et al., 2010;
42 Zehr, 2011). Recently discovered marine diazotrophic taxa, including those belonging to
43 unicellular cyanobacteria and heterotrophic bacteria, are abundant in oceanic regions where
44 large cyanobacterial diazotrophs are scarce (Needoba et al., 2007; Moisander et al., 2010;
45 Halm et al., 2012; Bonnet et al., 2013; Rahav et al., 2013; Shiozaki et al., 2014a), suggesting
46 that a failure to account for nitrogen fixation mediated by these diazotrophs might result in
47 underestimation of marine nitrogen fixation.

48 The temperate coastal ocean is one of the regions where nitrogen fixation rates have been
49 understudied and potentially underestimated. Conventionally, nitrogen fixation in temperate
50 oceans has been assumed to be low because of the relatively low temperatures (<~20°C),
51 which generally inhibit the growth of large cyanobacterial diazotrophs (Breitbarth et al.,
52 2007), and development of high dissolved inorganic nitrogen (DIN) concentrations (>1 µM).
53 High DIN concentrations are generally regarded to inhibit nitrogen fixation (Falkowski,
54 1983), especially during mixing periods. However, recent studies have indicated that

55 nitrogen fixation, presumably mediated by unicellular cyanobacteria and heterotrophic
56 bacteria, is detectable even in the relatively cold ($<10^{\circ}\text{C}$) and DIN-rich waters ($>1\ \mu\text{M}$) of the
57 Atlantic coast (Mulholland et al., 2012) and the Baltic Sea estuaries (Bentzon-Tilia et al.,
58 2015). These results highlight the necessity of re-evaluating the extent, variation, and
59 control mechanisms of nitrogen fixation in temperate oceans, with recognition of the
60 widespread occurrence of diverse diazotrophic microbes.

61 This study examined the seasonal variation in nitrogen fixation in the temperate inside bays
62 and open ocean located in the interfrontal zone of the northwestern North Pacific. In this
63 region, physical, chemical, and biological properties vary widely between seasons (Shiozaki
64 et al., 2014b) due to the confluence of three currents: the Kuroshio (warm current), the
65 Tsugaru Warm Current, and Oyashio (cold current). Data on nitrogen fixation rates in the
66 temperate Pacific are limited (Needoba et al., 2007), and to the best of our knowledge, the
67 present study is the first to examine diazotrophy during all seasons in the temperate ocean.

68 This study was conducted as part of a project to monitor the dynamics of the coastal
69 ecosystem and the recovery thereof after the 2011 Tohoku-oki tsunami, which struck the
70 region on 11 March 2011.

71

72 **2. Materials and Methods**

73 The experiments were conducted during six cruises in the temperate coastal region of the
74 western North Pacific. These cruises covered a full seasonal cycle, including spring
75 (KS-14-2_Mar, 14–19 March 2014), early summer (KK-13-1_Jun, 24–29 June 2013),
76 summer (KT-12-20_Aug, 7–12 August 2012), late summer (KK-13-6_Sep, 14–21 September
77 2013), fall (KT-12-27_Oct, 15–22 October 2012), and winter (KT-13-2_Jan, 19–25 January
78 2013). Sampling stations were located along the transect lines OT (39°20'N,
79 141°56'–142°50'E) and ON (38°25'N, 141°29'–142°20'E). Eight stations were located
80 offshore (OT4–6, ON4–8), while two stations were deployed in the Otsuchi (OT1) and
81 Onagawa (ON1) bays (Fig. 1). Just before the KK-13-6_Sep cruise, Typhoon Man-yi
82 passed from southwest to northeast in the study area (Fig. S1).

83 Temperature, salinity, and dissolved oxygen profiles of regions near the bottom floor were
84 measured using a SBE 911-plus conductivity-temperature-pressure (CTD) system (Sea-bird
85 Electronics, Bellevue, WA, USA). Water samples were collected in an acid-cleaned bucket
86 and Niskin-X bottles. At offshore stations, samples for nutrient analysis were collected
87 from 7–15 different depths in the upper 200 m, while at shallower (<200 m) bay stations,
88 samples were collected from 4–9 different depths in the entire water column, except at Stn.
89 OT1 where only surface water samples were collected. Samples for DNA analysis and
90 incubation experiments were collected from the surface at almost every station, and from

91 depths corresponding to 10% and 1% of the surface light intensities at Stns. OT4 and ON5.

92 Light attenuation was determined using a submersible PAR sensor.

93 **2.1. Nutrients**

94 Samples for nutrient analysis were stored in 10-mL polyethylene tubes and kept frozen until

95 onshore analyses. Nitrate, nitrite, ammonium, and phosphate concentrations were

96 determined using an AACSII auto-analyzer (Bran+Luebbe, Norderstedt, Germany). The

97 detection limits of nitrate, nitrite, ammonium, and phosphate ranged from 0.01–0.04 μM ,

98 0.01–0.02 μM , 0.01–0.03 μM , and 0.01–0.02 μM , respectively. The nitracline was defined

99 as the depth where nitrate concentrations increased above 1 μM .

100 **2.2. Nitrogen fixation activity and mannitol enrichment experiment**

101 Nitrogen fixation was determined by the $^{15}\text{N}_2$ gas bubble method (hereafter, the bubble

102 method; Montoya et al., 1996). Samples for incubation were collected in duplicate

103 acid-cleaned 2-L polycarbonate (PC) bottles. The time-zero samples (n=1) were

104 immediately filtered onto precombusted GF/F filters. Two milliliters of $^{15}\text{N}_2$ gas [SI

105 Science Co. Japan, for this gas, contaminations of nitrate, nitrite, and ammonium were

106 determined to be low (< nM level), indicating that the overestimation of nitrogen fixation

107 rates due to the uptake of ^{15}N -labeled contaminants (Dabundo et al. 2014) was minimal

108 (Shiozaki et al., unpublished data)] were injected directly into the incubation bottles through

109 a septum using a gastight syringe. The tracer-added samples were covered with
110 neutral-density screens to adjust the light level and incubated for 24 h in an on-deck incubator
111 filled with flowing surface seawater. After the incubation, the samples were filtered onto
112 precombusted GF/F filters. The isotopic analyses were performed as described previously
113 (Shiozaki et al., 2009). The rate of nitrogen fixation was calculated using the equations of
114 Montoya et al. (1996).

115 To examine the possibility of underestimation of nitrogen fixation as determined by the
116 bubble method (Mohr et al., 2010; Großkopf et al., 2012), we compared the nitrogen fixation
117 rates determined using the $^{15}\text{N}_2$ gas dissolution method (hereafter, the dissolution method;
118 Mohr et al., 2010) with those determined using the bubble method (see above) during the
119 KK-13-6_Sep and KS-14-2_Mar cruises. For the dissolution method, $^{15}\text{N}_2$ -enriched
120 seawater was prepared according to Mohr et al. (2010) and Großkopf et al. (2012). Briefly,
121 filtered seawater was degassed using a Sterapore membrane unit (20M1500A: Mitsubishi
122 Rayon Co., Ltd., Tokyo, Japan) at a flow rate of $\sim 500 \text{ mL min}^{-1}$ (recirculation period, 10 min).
123 Degassed seawater was stored in 1-L Tedlar bags without headspaces and 10 mL $^{15}\text{N}_2$ gas was
124 added. After complete dissolution, the $^{15}\text{N}_2$ -enriched seawater was added to seawater
125 samples contained in 2-L PC bottles, which were incubated and used for isotopic analyses as
126 described above. The $^{15}\text{N}_2$ -enriched seawater was prepared at each station, and was added

127 to the incubation bottles within 1 h after preparation. The nitrogen fixation rate was
128 calculated according to Mohr et al. (2010). For this comparison, triplicate samples were
129 used for both the dissolution and bubble methods.

130 To examine if sugar addition affected nitrogen fixation rates (Bonnet et al., 2013; Rahav et al.,
131 2013; Moisander et al., 2011), we determined nitrogen fixation rates (the $^{15}\text{N}_2$ gas bubble
132 method, see above) for surface seawater samples (stations ON4 and OT6 during the
133 KS-14-2_Mar cruise) with and without addition of mannitol (final conc. $0.8\ \mu\text{M}$) ($n=3$).

134 **2.3. DNA analysis**

135 2.3.1. DNA extraction, sequencing, and phylogenetic analysis

136 Samples (0.38–1 L) for DNA analysis were filtered through 0.2- μm -pore-sized Nuclepore
137 filters and stored in a deep freezer (-80°C) until onshore analysis. Total DNA was extracted
138 using a ChargeSwitch Forensic DNA Purification Kit (Invitrogen, Carlsbad, CA, USA) with
139 slight modification of the manufacturer's protocol (Shiozaki et al., 2014a). Partial *nifH*
140 fragments were amplified using a nested PCR strategy (Zehr and Turner, 2001) from samples
141 collected from surface water at Stns. OT4, ON1, ON5, and ON7 during the KT-12-20_Aug
142 and KT-12-27_Oct cruises, at Stns. OT4, ON1, and ON5 during the KT-13-2_Jan and
143 KS-14-2_Mar cruises, at Stns. OT4, ON1, ON5, and ON8 during the KK-13-1_Jun cruise,
144 and at Stns OT4, ON5, ON7 during the KK-13-6_Sep cruise (Table 1). PCR reagents were

145 applied as described by Shiozaki et al. (2014a). The first and second PCRs were run using
146 the same cycling conditions: 95° C for 30 s followed by 30 cycles of 98° C for 10 s, 52° C for
147 30 s, and 72° C for 30 s; followed by a final extension at 72° C for 7 min. Sterile distilled
148 water was used as the negative control. After PCR analysis, we confirmed there was no
149 band in agarose gel of electrophoresis from the negative control. The PCR products were
150 cloned and sequenced according to Shiozaki et al. (2014a). The present study obtained 197
151 *nifH* sequences in total. The *nifH* sequences were translated into amino acid sequences and
152 searched against the protein database of the National Center for Biotechnology Information
153 using the BLASTp algorithm. Clones with 100% amino acid sequence similarity were
154 defined as the same operational taxonomic unit (OTU) using the CD-HIT suite (Huang et al.,
155 2010). The amino acid sequences were aligned using multiple sequence comparisons by the
156 log-expectation (MUSCLE) module in the MEGA5 package (Tamura et al., 2011). A
157 phylogenetic tree was constructed using the maximum likelihood method employing the
158 Dayhoff matrix-based mode, and 1,000 bootstrap replicates were run. The obtained
159 sequences were assigned to bacterial groups based on known sequences included in a cluster
160 within the phylogenetic tree (Zehr et al., 2003a). The sequences from this study were
161 deposited in the DNA Data Bank of Japan (DDBJ) as accession numbers LC013480 to
162 LC013676.

163 2.3.2. Quantitative PCR (qPCR) analysis

164 The clone library analysis showed that UCYN-A, *Trichodesmium*, and γ -proteobacterial
165 phylotype γ -24774A11 (hereafter γ -24774A11) were likely important diazotrophs from early
166 summer to fall when nitrogen fixation occurred (see below). Therefore, the present study
167 quantified these *nifH* phylotypes by qPCR analysis to examine their relative importance
168 during these seasons. In addition, UCYN-B which is considered to be a major diazotroph in
169 the tropical and subtropical oligotrophic ocean (Moisander et al., 2010), was quantified.
170 TaqMan primer and probe sets previously designed for these four *nifH* phylotypes were used
171 for quantification (Shiozaki et al., 2014a,c). The 20 μ L qPCR reactions contained 10 μ L 2 \times
172 Premix Ex Taq (Probe qPCR; Takara), 5.6 μ L of nuclease-free water, 1 μ L each of the
173 forward and reverse primers, 0.4 μ L of TaqMan probe, and 2 μ L of template DNA. The
174 qPCR assays were performed using LightCycler 480 System (Roche Applied Science,
175 Germany). The qPCR assays were run in triplicate reactions. Linear regression r^2 values
176 for the standard curves were >0.99 for all reactions. The efficiency of the qPCR assays
177 ranged from 90.9 to 98.4%, with an average of 95.1%. As the negative control, sterile
178 distilled water was used, from which no amplification signals were detected. The detection
179 limit was 75 copies L^{-1} .

180

181 **3. RESULTS**

182 **3.1. Comparison of the bubble method and the dissolution method**

183 Nitrogen fixation rates determined by the bubble and dissolution methods were compared
184 during the KK-13-6_Sep and KS-14-2_Mar cruises (Fig. 2). Both methods failed to detect
185 nitrogen fixation in samples collected during the KS-14-2 cruise. During the KK-13-6_Sep
186 cruise, the nitrogen fixation rates determined by the dissolution method were significantly
187 higher (1.5–2.2 fold) than those determined by the bubble method at Stns. OT6 and ON5 (p
188 <0.05). At Stns. OT4 and ON7, the nitrogen fixation rates determined by the two methods
189 did not differ significantly.

190 **3.2. Seasonal variations in nitrogen fixation rates**

191 According to the temperature-salinity (TS) diagram proposed by Hanawa and Mitsudera
192 (1987), both the offshore and bay waters collected during this investigation primarily
193 belonged to either the surface layer water system (SW) or the Tsugaru Warm Current water
194 system (TW) (Fig. 3), with the exception of waters collected from the 1% light depth (119 m)
195 at Stn. ON5 during the KT-13-2_Jan cruise and those collected at the surface of OT5 during
196 the KS-14-2_Mar cruise, which were classified as belonging to the Oyashio water system
197 (OW) and the Coastal Oyashio water system (CO), respectively. These water classifications
198 based on the TS diagram were generally consistent with the geostrophic current field of the

199 investigated region (Fig.S1). Based on these results, it was considered that surface waters
200 collected during the same cruise in a particular season generally belonged to the same water
201 system that was prevalent in the investigated region at the time of our sampling.

202 Sea surface temperatures (SST) (range, 1.5 to 24.3° C) (Figs. 4a and S1) and surface nitrate
203 and phosphate concentrations determined during each cruise were averaged to emphasize the
204 seasonal variability of these parameters (Fig. 4b). In general, surface nitrate and phosphate
205 concentrations were low ($\leq 0.07 \mu\text{M}$ and $\leq 0.20 \mu\text{M}$, respectively) in the warmer seawaters
206 (14.2–24.3° C) sampled in early summer (KK-13-1_Jun), summer (KT-12-20_Aug), and fall
207 (KT-12-27_Oct), whereas they were relatively high ($\geq 0.75 \mu\text{M}$ and $\geq 0.28 \mu\text{M}$, respectively)
208 in the colder seawaters (1.5–9.8° C) sampled during winter (KT-13-2_Jan), and spring
209 (KS-14-2_Mar). During the KK-13-6_Sep cruise (late summer), the nitrate concentrations
210 were relatively high and variable (mean \pm SD; $2.92 \pm 7.90 \mu\text{M}$). This was because the
211 highest nitrate concentration ($22.6 \mu\text{M}$) was determined at the near-shore Stn. OT1 (Fig. S2).

212 Similar to nitrate, surface phosphate concentrations tended to be high during winter
213 (KT-13-2_Jan) and spring (KS-14-2_Mar), while they were low during the warmer seasons.

214 The seasonal variation pattern of the average ammonium concentration at the surface differed
215 from those of nitrate and phosphate concentrations (Fig. 4b), characterized by low
216 concentrations ($\leq \sim 1 \mu\text{M}$) throughout the year. The high variation in surface ammonium

217 concentration during the KK-13-6_Sep cruises were due to relatively high ammonium
218 concentrations at Stn. OT1 (1.41 μM) (Fig. S2).

219 Nitrogen fixation was detected in the surface waters of most samples collected during the
220 four cruises conducted in early summer (KK-13-1_Jun), summer (KT-12-20_Aug), late
221 summer (KK-13-6_Sep), and fall (KT-12-27_Oct), and varied in the range of 0.33–13.6 nmol
222 $\text{N L}^{-1} \text{d}^{-1}$ (Figs. 4c and S2). Relatively high nitrogen fixation rates were determined for
223 samples collected during the KT-12-20_Aug cruise, although the highest value was obtained
224 at Stn. ON7 during the KK-13-6_Sep cruise. Nitrogen fixation was not detected in seawater
225 samples collected during the winter and spring cruises, even after addition of mannitol
226 (KS-14-2_Mar). Furthermore, nitrogen fixation was not detected in DIN-replete water at
227 Stn. OT1 in late summer (KK-13-6_Sep).

228 The rates of nitrogen fixation in samples collected at different depths (0–119 m) were
229 examined at Stns. OT4 and ON5 (Fig. 5). Nitrogen fixation was detectable only during the
230 four cruises conducted in early summer (KK-13-1_Jun), summer (KT-12-20_Aug), late
231 summer (KK-13-6_Sep), and fall (KT-12-27_Oct), the same seasons during which surface
232 nitrogen fixation was observed (Fig. 4). Nitrogen fixation rates tended to be higher at the
233 surface than in the deeper layers during summer (KT-12-20_Aug) and late summer
234 (KK-13-6_Sep (at Stn. OT4)), whereas this vertical trend was less evident during fall

235 (KT-12-27_Oct) and early summer (KK-13-1_Jun). At Stn. OT4, nitrogen fixation was
236 detectable even in deeper layers below the nitracline, where nitrate concentrations were
237 relatively high (KT-12-27_Oct, depth = 62 m; KK-13-1_Jun, depth = 42 m). In this layer,
238 the ammonium concentrations were 0.05 μM (KT-12-27_Oct) and 0.62 μM (KK-13-1_Jun).
239 The nitrogen fixation rate below the nitracline ($1.56 \text{ nmol N L}^{-1} \text{ d}^{-1}$) was higher than that at
240 the surface ($0.87 \text{ nmol N L}^{-1} \text{ d}^{-1}$) during the KK-13-1_Jun cruise. The maximum
241 depth-integrated nitrogen fixation ($294 \mu\text{mol N m}^{-2} \text{ d}^{-1}$) was observed at Stn. OT4 during
242 summer (KT-12-20_Aug).

243 **3.3. Relationship between nitrogen fixation rates and environmental** 244 **variables**

245 Nitrogen fixation rates tended to increase with temperature ($p < 0.01$) (Fig. 6a and Table 2).
246 Nitrogen fixation was detected only when seawater temperatures exceeded 11.7°C , with
247 higher rates ($>6 \text{ nmol N L}^{-1} \text{ d}^{-1}$) noted in waters warmer than 19.5°C . However, there were
248 exceptions to this general relationship between the nitrogen fixation rate and temperature.
249 For example, from the data collected during the KK-13-1_Jun cruise the nitrogen fixation
250 rate was highest at 15.4°C , while it was low (undetectable) at higher temperatures.
251 Nitrogen fixation rates were negatively correlated with nitrate and phosphate concentrations
252 ($p < 0.01$) (Table 2). There was no significant correlation between nitrogen fixation rates

253 and ammonium concentration ($p > 0.05$). We also found no significant correlation between
254 nitrogen fixation rates and the ratio of total inorganic nitrogen (nitrate + nitrite + ammonium)
255 to phosphate (Table 2). A plot of the nitrogen fixation against nitrate concentrations
256 indicated that nitrogen fixation was generally detectable only when nitrate was depleted (Fig.
257 6b), except that relatively high nitrogen fixation rates were determined in the subsurface layer
258 of Stn. OT4 (KT-12-27_Oct and KK-13-1_Jun). Active nitrogen fixation tended to occur at
259 low ammonium concentration $\leq \sim 0.1 \mu\text{M}$. However, seasonal variation in ammonium
260 concentration was small and no statistically significant relationship with nitrogen fixation
261 was observed (Fig. 6c).

262

263 **3.4. Seasonal variation in the diazotroph community**

264 3.4.1. Diazotroph community

265 PCR reagents have been suggested to be a potential source of *nifH* genes during analysis of
266 the diazotroph community (Zehr et al., 2003b). Although we confirmed the absence of any
267 bands from the negative control in agarose gel electrophoresis, sequences with similarity
268 ($>97\%$) at the amino acid level to contaminants in PCR reagents were recovered from
269 samples obtained during the KK-13-6_Sep and KS-14-2_Mar cruises (10 clones in total).
270 We did not include these sequences in our data analysis.

271 The *nifH* gene was recovered from all samples that we collected during this study across
272 different stations and seasons (Table 1). Sixty-one OTUs were grouped from 187 *nifH*
273 clones, based on 100% amino acid sequence similarity. The OTUs were assigned to
274 cyanobacteria, α -, β -, γ -, and δ -proteobacteria, and Cluster III diazotrophs (Zehr et al., 2003a)
275 (Figs. S3 and S4).

276 The detected cyanobacterial sequences belonged to *Trichodesmium*, UCYN-A, and
277 *Leptolyngbya*. The *nifH* sequences of UCYN-B, UCYN-C, and *Richelia intracellularis*
278 were not recovered. The *nifH* sequence of *Trichodesmium* was recovered only during the
279 KT-12-27_Oct cruise (Table 1). UCYN-A was generally observed from early summer to
280 fall, while *nifH* of *Leptolyngbya* was detected during winter. During the KS-14-2_Mar
281 cruise, all recovered sequences were derived from heterotrophic bacteria, and were
282 dominated by Cluster III diazotrophs at Stns. OT4 and ON5. The Cluster III diazotroph
283 *nifH* sequences were recovered on all cruises except the KK-13-1_Jun cruise. Note that 58
284 out of 187 sequences displayed >97% similarity, at the amino acid level, to terrestrial
285 diazotroph sequences derived from soil, mudflats, and lakes (Fig. S3 and S4). These
286 sequences were mainly affiliated with α - and δ -proteobacterial diazotrophs, with 29 of 39
287 α -proteobacterial sequences and 22 of 24 δ -proteobacterial sequences being similar to
288 terrestrial diazotroph sequences.

289 3.4.2. Diazotrophs abundances

290 The *nifH* sequence of *Trichodesmium* was detected by qPCR assay during the KT-12-27_Oct
291 and KK-13-6_Sep cruises (Fig. 7 and 8). During these two cruises, the abundance of
292 *Trichodesmium* ranged from below the detection limit to 8.7×10^4 copies L⁻¹ at all depths.
293 *Trichodesmium* abundance at the surface was higher than those of UCYN-A, UCYN-B, and
294 γ -24774A11 at most stations during the KT-12-27_Oct cruise (Fig. 7 and S5). UCYN-A
295 was detected on all cruises except for the KS-14-2_Mar cruise (Fig. 7 and 8). The
296 maximum abundance of UCYN-A generally occurred at the surface except at Stn. OT4
297 during the KK-13-6_Sep cruise where the peak (1.2×10^3 copies L⁻¹) was observed at 72 m
298 (Fig. 8). The abundance of UCYN-A varied from below the detection limit to 2.6×10^5
299 copies L⁻¹ at all depths. At the surface, UCYN-A was the most abundant among the four
300 groups at most of the stations investigated during the KT-12-20_Aug, KT-13-2_Jan,
301 KK-13-1_Jun, and KK-13-6_Sep cruises (Fig. 7 and S5). UCYN-B was detected only at
302 Stn. ON7 during the KK-13-6_Sep cruise (Fig. 7, 8, and S5). γ -24774A11 was detected
303 during all cruises except for the KS-14-2_Mar cruise (Fig. 7 and 8). The abundance of
304 γ -24774A11 ranged from below the detection limit to 1.8×10^4 copies L⁻¹, with a tendency of
305 a subsurface peak at both stations (Fig. 8).

306

307 4. DISCUSSION

308 4.1. Differences in nitrogen fixation rates between the bubble method and 309 the dissolution method

310 The present study revealed that a significant difference between the bubble and the
311 dissolution methods was not always present. Großkopf et al. (2012) indicated that the
312 difference was smaller when *Trichodesmium* dominated in the diazotroph community than
313 when unicellular cyanobacteria and γ -proteobacteria dominated presumably because
314 *Trichodesmium* can float to the top of the bottle and directly use the added $^{15}\text{N}_2$ in the bubble
315 method. Interestingly, *Trichodesmium* abundance was higher than or similar to those of
316 UCYN-A, UCYN-B, and γ -24774A11 at Stns. OT4 and ON7, at which there was no
317 significant difference detected between the two methods. On the other hand,
318 *Trichodesmium* was not detected and UCYN-A was the most abundant among the four groups
319 at Stns. OT6 and ON5, at which nitrogen fixation determined by the dissolution method was
320 significantly higher than that by the bubble method. These results were consistent with the
321 report by Großkopf et al. (2012). The larger variations in nitrogen fixation at Stns. OT4 and
322 ON7 than at Stns. OT6 and ON5 were probably due to the heterogeneity of *Trichodesmium*
323 abundance (Carpenter et al., 2004). Although nitrogen fixation rates determined by the
324 bubble method in the present study were underestimated on all cruises, the level of

325 underestimation was relatively small during the KT-12-27_Oct cruise when *Trichodesmium*
326 was dominant at most of the stations.

327 **4.2. Seasonal variations in nitrogen fixation rates in the temperate coastal** 328 **ocean**

329 Nitrogen fixation rates were measurable mainly from early summer to fall when nitrate was
330 generally depleted in sample seawaters, although there were some exceptions. Our
331 estimates of the nitrogen fixation rates ($0.33\text{--}13.6 \text{ nmol N l}^{-1} \text{ d}^{-1}$) were significantly ($p < 0.05$)
332 higher than the corresponding values previously reported in the temperate region of the
333 eastern North Pacific ($0.15\text{--}0.31 \text{ nmol N l}^{-1} \text{ d}^{-1}$; Needoba et al., 2007) and the oligotrophic
334 region of the western and central North Pacific ($0.17\text{--}3.62 \text{ nmol N l}^{-1} \text{ d}^{-1}$; Shiozaki et al.,
335 2010), whereas they were comparable to those determined in the Kuroshio ($0.54\text{--}28 \text{ nmol N}$
336 $\text{l}^{-1} \text{ d}^{-1}$; Shiozaki et al., 2010) and the western Atlantic coastal regions ($1.3\text{--}49.8 \text{ nmol N l}^{-1} \text{ d}^{-1}$;
337 Mulholland et al., 2012). Higher nitrogen fixation rates have been determined in other
338 temperate oceans, including the western English Channel (18.9 ± 0.01 and $20.0 \text{ nmol N l}^{-1} \text{ d}^{-1}$;
339 Rees et al., 2009) and the Baltic Sea estuaries ($47\text{--}83 \text{ nmol N l}^{-1} \text{ d}^{-1}$; Bentzon-Tilia et al.,
340 2015).

341 In our study, spatiotemporal variability in nitrogen fixation rates appeared to be partly related
342 to the Tsugaru Warm Current path. This current, which flows from the north (after passage

343 through the Tsugaru Strait) to the study region (Fig. S1), may carry active diazotrophs and
344 therefore enhance nitrogen fixation in our study region. This is supported by the fact that
345 nitrogen fixation rates during individual cruises tended to be higher at Stn. OT4 than at Stn.
346 ON5. These stations were located up- and down-stream of the Tsugaru Warm Current,
347 respectively. In addition, variations in nitrogen fixation rates among stations and seasons
348 might also be related to the extent of vertical mixing in the Tsugaru Warm Current. It has
349 been suggested that vertical mixing may introduce iron-rich subsurface water to the surface
350 of the Tsugaru Strait (Saitoh et al., 2008). Such input of iron may enhance nitrogen fixation
351 rates. Consistent with this notion, our results showed that the nitrogen fixation rate was
352 relatively high at Stn. OT4, where the nitracline was relatively deep.

353 Blais et al. (2012) proposed that nitrogen fixation can proceed even in nutrient-replete waters,
354 if large amounts of iron and organic materials are available for consumption by bacterial
355 diazotrophs. In the present study, this possibility was examined by conducting mannitol
356 addition experiments using surface seawaters collected during spring. These waters,
357 belonging to the Oyashio Current system (Nishioka et al., 2007, 2011; Shiozaki et al., 2014b),
358 were considered to be rich in iron during spring, as indicated by a previous study (iron conc.,
359 0.79–8.46 nM; Nishioka et al. 2007). Despite potentially high iron concentrations, our
360 results showed that nitrogen fixation was undetectable even after the mannitol addition,

361 suggesting that, contrary to the Blais et al. proposition, diazotrophs remained inactive under
362 our experimental settings.

363 Our data showed that nitrogen fixation rates were below the detection limit during winter,
364 spring, and late summer (KK-13-6_Sep), when nitrate concentrations were high. These
365 results were consistent with the results of previous studies in the Pacific Ocean, which
366 indicated that nitrogen fixation rates were low or undetectable in DIN-replete waters
367 (Shiozaki et al., 2010). In contrast, Mulholland et al. (2012) reported that, in temperate
368 regions of the Atlantic Ocean, nitrogen fixation rates were high even in DIN-replete ($>1 \mu\text{M}$)
369 and cold ($<10^\circ\text{C}$) surface seawaters. Their study was conducted downstream of the Gulf
370 Stream, where diazotrophs could be delivered from subtropical oceans where DIN is depleted.
371 Previous studies have suggested that cyanobacterial diazotrophs can travel over long
372 distances ($>1,000 \text{ km}$) in currents, without losing their capacity for N_2 fixation (Shiozaki et
373 al., 2013), and that activity is not lost immediately even after mixing with DIN-replete
374 seawaters (Holl and Montoya, 2005; DeKaemacker and Bonnet, 2011). In our region,
375 because the Tsugaru Warm Current flows from north to south, diazotrophs entrained by the
376 current have little chance of meeting high-DIN water at the surface. DIN-replete water
377 during summer was observed at the inside bay station OT1 (Fig. S2). Concomitantly,
378 low-salinity surface waters spread offshore along the OT transect line (Fig. S6), suggesting

379 that anomalously high DIN concentrations were likely attributable to terrestrial surface
380 discharge enhanced by Typhoon Man-yi, which passed over the region immediately before
381 the cruise. Subramaniam et al. (2008) reported that nitrogen fixation rates near the Amazon
382 River estuary, with low salinity and high nitrate levels, were fairly low. Their results are
383 consistent with ours. Ammonium inhibits nitrogen fixation, especially when ammonium
384 concentrations exceed 1 μM , as demonstrated by *Trichodesmium* (Mulholland et al. 2001).
385 In our study, no negative relationship between nitrogen fixation and ammonium
386 concentration was found. This can likely be explained by relatively low ammonium
387 concentrations ($\leq \sim 1 \mu\text{M}$) throughout the year and across the investigated region.

388 **4.3. Seasonal variation in the diazotroph community in the temperate** 389 **coastal ocean**

390 The qPCR analysis demonstrated that the target groups were quantifiable even at stations at
391 which their sequences were not recovered by the clone library analysis, suggesting that the
392 number of clones was not sufficient to capture the diazotroph community structure on each
393 cruise. Despite this limitation, the sequences more frequently recovered in the clone library
394 generally corresponded to the most abundant group revealed by the qPCR analysis. For
395 example, UCYN-A was frequently recovered in the library during the KT-12-20_Aug,
396 KK-13-1_Jun, and KK-13-6_Sep cruises; for these samples, the qPCR results showed that

397 UCYN-A was the most abundant group among the four examined. Similarly, qPCR data
398 indicated that *Trichodesmium* was the most abundant group during fall, when this group was
399 frequently recovered in the library (during the KT-12-27_Oct cruise). This consistency in
400 the general results obtained by the clone library and qPCR suggests that both of these
401 approaches captured a similar seasonal trend in community composition changes for at least
402 the major diazotroph groups. In the discussion below, we discuss possible factors
403 responsible for seasonal variation in the diazotroph community by focusing on the major
404 diazotroph groups.

405 UCYN-A was detected by qPCR in all seasons except spring (KS-14-2_Mar), suggesting that
406 this group of diazotrophs could be important agents of nitrogen fixation in this region.
407 Especially from early to late summer, the abundance of UCYN-A was generally higher than
408 that of *Trichodesmium*, UCYN-B, and γ -24774A11. UCYN-A has been widely detected in
409 temperate regions, and is considered to be one of the major diazotrophs of these locations
410 (Needoba et al., 2007; Rees et al., 2009; Mulholland et al., 2012; Bentzon-Tilia et al., 2015).
411 UCYN-A is known to be most abundant in relatively warm waters around ~20° C (Needoba
412 et al., 2007; Moisander et al., 2010). UCYN-A was detected by qPCR even during winter at
413 some stations, yet, was not observed during spring. This could be because UCYN-A
414 abundance decreased from fall to winter with decreasing temperatures, eventually

415 disappearing during in spring.

416 *Trichodesmium* was detected from late summer to fall by qPCR analysis, when water
417 temperatures ranged from 19.1 to 23.4° C at the surface. Given that the optimal growth
418 temperature for *Trichodesmium* has been reported to be high (24–30° C) (Breitbarth et al.,
419 2007), *Trichodesmium* detected in the investigated region likely existed under suboptimum
420 conditions. The relatively high abundance of *Trichodesmium* observed during fall, despite
421 the suboptimal temperature conditions, might indicate that *Trichodesmium* was transported
422 from the adjacent subtropical region where seawater temperatures were high (>24° C). In
423 the western North Pacific subtropical region, *Trichodesmium* is abundant from July to
424 September (Marumo and Nagasawa, 1976; Chen et al., 2008). *Trichodesmium* that
425 flourished in the subtropical region during summer could be transported by the Tsugaru
426 Warm Current, displaying peak abundance during fall in the investigated region. This could
427 support the above discussion that waters containing active nitrogen fixation were delivered to
428 this region by the Tsugaru Warm Current.

429 We observed γ -24774A11 by qPCR analysis during all cruises except for the KS-14-2_Mar
430 cruise. This phylotype has not been reported previously in other temperate oceans
431 (Needoba et al., 2007; Rees et al., 2009; Mulholland et al., 2012). The *nifH* sequence of
432 γ -24774A11 was similar to that of *Pseudomonas stutzeri* (94% similarity at the amino acid

433 level), which was observed in waters including temperate regions (Bentzon-Tilia et al., 2015).
434 Bentzon-Tilia et al. (2015) reported that *P. stutzeri*-like *nifH* genes (99% similarity at the
435 nucleotide level) were the most abundant sequences among their samples collected from the
436 temperate Baltic Sea estuary. In the present study, we recovered *P. stutzeri*-like *nifH* genes
437 (>97% similarity at the amino acid level) from Stn. OT4 during the KT-13-2_Jan cruise by
438 the clone library analysis. However, γ -24774A11 was not detected on that occasion by
439 qPCR analysis, suggesting that γ -24774A11 was not quantified as *P. stutzeri* and that *P.*
440 *stutzeri* was not a major diazotroph in this study region. The ecology of γ -24774A11 is still
441 fairly unknown. It remains to be seen, in future studies whether this phylotype contributes
442 to the nitrogen fixation in this region.

443 UCYN-B was not detected by qPCR except at one station. This result is consistent with
444 previous knowledge. UCYN-B becomes abundant with increasing temperature, similar to
445 *Trichodesmium* (Moisander et al., 2010), and is rarely observed in the temperate region
446 (Needoba et al., 2007; Rees et al., 2009; Mulholland et al., 2012; Bentzon-Tilia et al., 2015).
447 Furthermore, UCYN-B abundance is low in shallow nitracline regions (Shiozaki et al.,
448 2014a,c). The nitracline depth in this region (≤ 60 m) was shallower than that of >100-m
449 depths of regions where UCYN-B is abundant (Shiozaki et al., 2014a). Therefore, although
450 UCYN-B might also have been delivered from subtropical region, it could not survive in the

451 shallower nitracline region.

452 In nitrate-rich water during winter and spring, Cluster III diazotrophs were dominant at most
453 of the stations. Furthermore, from early summer to fall, *nifH* sequences of Cluster III
454 diazotrophs were recovered by the clone library analysis in samples from all cruises (except
455 KK-13-1_Jan). Because UCYN-A, *Trichodesmium*, and γ -24774A11 were scarce during
456 winter and spring, Cluster III diazotrophs were likely to be major diazotrophs at these times.

457 Cluster III diazotrophs are putative anaerobes (Hamersley et al., 2011; Farnelid et al., 2013;
458 Bentzon-Tilia et al., 2014), and hence, they are usually dominant in the diazotrophic
459 community of oxygen-depleted waters (Hamersley et al., 2011; Farnelid et al., 2013) or
460 marine sediments (Bertics et al., 2013). In this study, dissolved oxygen was not depleted
461 ($>3.16 \text{ ml L}^{-1}$) in the upper winter maximum mixed layer depth in this region ($\sim 200 \text{ m}$;
462 Shiozaki et al., 2014b) (Fig. S7). Therefore, it is possible that the Cluster III diazotrophs
463 that we detected in the surface layer were derived from resuspensions of coastal marine
464 sediments in which anoxic conditions may prevail because of organic matter decomposition.
465 The Cluster III activity was likely strongly suppressed in the water column because of the
466 high oxygen concentration.

467 Many *nifH* sequences recovered by the clone library analysis were similar to terrestrially
468 derived sequences. These results agree with previous data collected in coastal regions,

469 where terrestrially derived *nifH* sequences were also found (Rees et al., 2009; Mulholland et
470 al., 2012; Blais et al., 2012). We obtained a *Leptolyngbya*-like *nifH* gene during the
471 KT-13-2_Jan cruise. The organism has been found on beaches or coastal land areas (Brito
472 et al. 2012), but not in the open ocean. Because nitrogen fixation was not detected during
473 the KT-13-2_Jan cruise, the organism must have been inactivated after being flushed out
474 from the coastal region.

475

476 **5. CONCLUSION**

477 This study demonstrated that nitrogen fixation can and does proceed at high rates, depending
478 on the season, in the temperate region of the northwestern North Pacific, although we failed
479 to detect nitrogen fixation in DIN-replete cold waters. *nifH* sequences were omnipresent
480 and recovered throughout the year, displaying a marked seasonality in their composition.
481 UCYN-A was a major diazotroph during summer, whereas *Trichodesmium* was abundant
482 during fall, despite low temperatures. It has been suggested that *Trichodesmium* was
483 laterally transported from the adjacent subtropical region, which displays high temperatures.
484 The Cluster III diazotrophs were abundant in surface waters during winter, which was
485 ascribed to their delivery from the anoxic sediments via bottom resuspension. The failure to
486 detect nitrogen fixation when Cluster III was abundant implied that the activity of this

487 diazotroph group was strongly suppressed in oxic water columns.

488

489 **Author Contributions**

490 T.S., T.N., and K.F. designed the experiment and T.S. collected the samples at sea. T.S.

491 determined nitrogen fixation and nutrient concentrations and analyzed satellite datasets. T.S.

492 and M.I. conducted the genetic analyses. T.S. prepared the manuscript with contributions

493 from all co-authors.

494

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509

510 **References**

- 511 Bentzon-Tilia, M., Farnelid, H., Jürgens, K. and Riemann, L.: Cultivation and isolation of
512 N₂-fixing bacteria from suboxic waters in the Baltic Sea, *FEMS Microbiol. Ecol.*, 88,
513 358-371, 2014.
- 514 Bentzon-Tilia, M., Traving, S.J., Mantikci, M., Knudsen-Leerbeck, H., Hansen, J. L. S.,
515 Markager, S., and Riemann, L.: Significant N₂ fixation by heterotrophs, photoheterotrophs
516 and heterocystous cyanobacteria in two temperate estuaries, *ISME J.*, 9, 273-285, 2015.
- 517 Bertics ,V. J., Löscher, C. R., Salonen, I., Dale, A. W., Gier, J., Schmitz, R. A., and Treude,
518 T.: Occurrence of benthic microbial nitrogen fixation coupled to sulfate reduction in the
519 seasonally hypoxic Eckernförde Bay, Baltic Sea, *Biogeosciences*, 10, 1243-1258, 2013.
- 520 Blais, M., Tremblay, J.-E., Jungblut, A. D., Gagnon, J., Martin, J., Thaler, M., and Lovejoy,
521 C.: Nitrogen fixation and identification of potential diazotrophs in the Canadian Arctic, *Glob.*
522 *Biogeochem. Cycles* 26, GB3022, doi:10.1029/2011GB004096, 2012.
- 523 Bonnet, S., Dekaezemacker, J., Turk-Kubo, K. A., Moutin, T., Hamersley, R. M., Grosso, O.,
524 Zehr, J. P., and Capone, D. G.: Aphotic N₂ fixation in the Eastern Tropical South Pacific
525 Ocean, *PLoS one* 8(12), e81265, doi:10.1371/journal.pone.0081265, 2013.
- 526 Brandes, J. A., and Devol, A. H.: A global marine-fixed nitrogen isotopic budget:
527 Implications for Holocene nitrogen cycling, *Glob. Biogeochem. Cycles*. 16, 4, 1120,

528 doi:10.1029/2001GB001856, 2002.

529 Breitbarth, E., Oschlies, A., and LaRoche, J: Physiological constraints on the global
530 distribution of *Trichodesmium* –effect of temperature on diazotrophy, *Biogeosciences*, 4,
531 53-61, 2007.

532 Brito, A., Ramos, V., Seabra, R., Santos, A., Santos, C. L., Lopo, M., Ferreira, S., Martins, A.,
533 Mota, R., Frazão, B., Martins, R., Vasconcelos, V., and Tamagnini, P.: Culture-dependent
534 characterization of cyanobacterial diversity in the intertidal zones of the Portuguese coast: A
535 polyphasic study, *Syst. Appl. Microbiol.* 35, 110-119, 2012.

536 Capone, D. G., Zehr, J. P., Paerl, H. W., Bergman, B., and Carpenter, E. J.: *Trichodesmium*, a
537 globally significant marine cyanobacterium, *Science*, 276, 1221-1229, 1997.

538 Carpenter, E. J., Subramaniam, A., Capone, D. G.: Biomass and primary productivity of the
539 cyanobacterium *Trichodesmium* spp. in the tropical N Atlantic ocean, *Deep-Sea Res.*, 51,
540 173-203, 2004.

541 Church, M. J., Mahaffey, C., Letelier, R. M., Lukas, R., Zehr, J. P., and Karl, D. M.: Physical
542 forcing of nitrogen fixation and diazotroph community structure in the North Pacific
543 subtropical gyre, *Glob. Biogeochem. Cycles.*, 23, GB2020, doi:10.1029/2008GB003418,
544 2009.

545 Chen, Y.-L. L., Chen, H.-Y., Tuo, S.-H., and Ohki, K.: Seasonal dynamics of new production

546 from *Trichodesmium* N₂ fixation and nitrate uptake in the upstream Kuroshio and South
547 China Sea basin, *Limnol. Oceanogr.*, 53(5), 1705-1721, 2008.

548 Codispoti, L. A.: An oceanic fixed nitrogen sink exceeding 400 TgN a⁻¹ vs the concept of
549 homeostasis in the fixed-nitrogen inventory, *Biogeosciences*, 4, 233-253, 2007.

550 Dabundo, R., Lehmann, M. F., Treibergs, L., Tobias, C. R., Altabet, M. A., Moisander, P. H.,
551 and Granger, J.: The contamination of commercial ¹⁵N₂ gas stocks with ¹⁵N-labeled nitrate
552 and ammonium and consequences for nitrogen fixation measurements, *PLoS ONE*, 9(10),
553 e110335, doi:10.1371/journal.pone.0110335, 2014.

554 Dekaezemacker, J., and Bonnet, S.: Sensitivity of N₂ fixation to combined nitrogen forms
555 (NO₃⁻ and NH₄⁺) in two strains of the marine diazotroph *Crocospaera watsonii*
556 (Cyanobacteria), *Mar. Ecol. Progr. Ser.*, 438, 33-46, 2011.

557 Deutsch, C., Sigman, D. M., Thunell, R. C., Meckler, A. N., and Haug, G. H.: Isotopic
558 constraints on glacial/interglacial changes in the oceanic nitrogen budget, *Glob. Biogeochem.*
559 *Cycles.*, 18, GB4012, doi:10.1029/2003GB002189, 2004.

560 Falkowski, P.G.: Enzymology of nitrogen assimilation, in: Carpenter, J., Capone, D. G. (eds)
561 *Nitrogen in the marine environment*, Academic Press, New York, 839-868, 1983.

562 Farnelid, H., Bentzon-Tilia, M., Andersson, A. F., Bertilsson, S., Jost, G., Labrenz, M.,
563 Jürgens, K., and Riemann, L.: Active nitrogen-fixing heterotrophic bacteria at and below the

564 chemocline of the central Baltic Sea, *ISME J.*, 7, 1413-1423, 2013.

565 Großkopf, T., Mohr, W., Baustian, T., Schunck, H., Gill, D., Kuypers, M. M. M., Lavik, G.,
566 Schmitz, R. A., Wallace, D. W. R., and LaRoche, J.: Doubling of marine dinitrogen-fixation
567 rates based on direct measurements, *Nature*, 488, 361-364, 2012.

568 Halm, H., Lam, P., Ferdelman, T. G., Lavik, G., Dittmar, T., LaRoche, J., D'Hondt, S., and
569 Kuypers, M.M.M.: Heterotrophic organisms dominate nitrogen fixation in the South Pacific
570 Gyre, *ISME J.*, 6, 1238-1249, 2012.

571 Hamersley, M. R., Turk, K. A., Leinweber, A., Gruber, N., Zehr, J. P., Gunderson, T., and
572 Capone, D. G.: Nitrogen fixation within the water column associated with two hypoxic basins
573 in the Southern California Bight, *Aquat. Microbial. Ecol.*, 63, 193-205, 2011.

574 Hanawa, K., and Mitsudera, H.: Variation of water system distribution in the Sanriku coastal
575 area, *J. Oceanogr. Soc. Jap.*, 42, 435-446, 1987.

576 Holl, C. M., and Montoya, J. P.: Interactions between nitrate uptake and nitrogen fixation in
577 continuous cultures of the marine diazotroph *Trichodesmium* (Cyanobacteria), *J. Phycol.*, 41,
578 1178-1183, 2005.

579 Huang, Y., Niu, B. F., Gao, Y., Fu, L. M., and Li, W.Z.: CD-HIT Suite: a web server for
580 clustering and comparing biological sequences, *Bioinformatics*, 26, 680-682, 2010.

581 Marumo, R., and Nagasawa S.: Seasonal variation of the standing crop of a pelagic

582 blue-green alga, *Trichodesmium* in the Kuroshio water, Bull. Plankton Soc. Japan, 23(1),
583 19-25, 1976 (in Japanese with English abstract).

584 Moisander, P. H., Beinart, R. A., Hewson, I., White, A. E., Johnson, K. S., Carlson, C. A.,
585 Montoya, J. P., and Zehr, J. P.: Unicellular cyanobacterial distributions broaden the oceanic
586 N₂ fixation domain, Science, 327, 1512-1514, 2010.

587 Moisander, P.H., Zhang, R., Boyle, E.A., Hewson, I., Montoya, J.P., Zehr, J.P.: Analogous
588 nutrient limitations in unicellular diazotrophs and *Prochlorococcus* in the South Pacific
589 Ocean, ISME J., 6, 733-744, 2011.

590 Moisander, P. H., Serros, T., Paerl, R. W., Beinart, R. A., and Zehr, J.P.:
591 Gammaproteobacterial diazotrophs and *nifH* gene expression in surface waters of the South
592 Pacific Ocean, ISME J., 8, 1962-1973, 2014.

593 Mohr, W., Großkopf, T., Wallace, D. W. R., and LaRoche, J.: Methodological
594 underestimation of oceanic nitrogen fixation rates, PLoS ONE, 5(9), e12583,
595 doi:10.1371/journal.pone.0012583, 2010.

596 Montoya, J. P., Voss, M., Kähler, P., and Capone, D. G.: A simple, high-precision,
597 high-sensitivity tracer assay for N₂ fixation, Appl. Environ. Microbiol., 62(3), 986-993, 1996.

598 Mulholland, M. R., Ohki, K., and Capone, D. G.: Nutrient controls on nitrogen uptake and
599 metabolism by natural populations and cultures of *Trichodesmium* (Cyanobacteria), J.

600 *Phycol.*, 37, 1001-1009, 2001.

601 Mulholland, M. R., Bernhardt, P. W., Blanco-Garcia, J. L., Mannino, A., Hyde, K.,
602 Mondragon, E., Turk, K., Moisander, P. H., and Zehr, J. P.: Rates of dinitrogen fixation and
603 the abundance of diazotrophs in North American coastal waters between Cape Hatteras and
604 Georges Bank, *Limnol. Oceanogr.*, 57(4), 1067-1083, 2012.

605 Needoba, J. A., Foster, R. A., Sakamoto, C., Zehr, J. P., and Johnson, K. S.: Nitrogen fixation
606 by unicellular diazotrophic cyanobacteria in the temperate oligotrophic North Pacific Ocean,
607 *Limnol. Oceanogr.*, 52(4), 1317-1327, 2007.

608 Nishioka, J., Ono, T., Saito, H., Nakatsuka, T., Takeda, S., Yoshimura, T., Suzuki, K., Kuma,
609 K., Nakabayashi, S., Tsumune, D., Mitsudera, H., Johnson, W. K., and Tsuda, A.: Iron supply
610 to the western subarctic Pacific: Importance of iron export from the Sea of Okhotsk, *J.*
611 *Geophys. Res.*, 112, C10012, doi:10.1029/2006JC004055, 2007.

612 Nishioka, J., Ono, T., Saito, H., Sakaoka, K., and Yoshimura, T.: Oceanic iron supply
613 mechanisms which support the spring diatom bloom in the Oyashio region, western subarctic
614 Pacific, *J. Geophys. Res.*, 116, C02021, doi:10.1029/2010JC006321, 2011.

615 Rahav, E., Bar-Zeev, E., Ohayon, S., Elifantz, H., Belkin, N., Herut, B., Mulholland, M. R.,
616 and Berman-Frank, I.: Dinitrogen fixation in aphotic oxygenated marine environment, *Front.*
617 *Microbiol.*, 4, 277, doi:10.3389/fmicb.2013.00227, 2013.

618 Riemann, L., Farnelid, H., and Steward, G. F.: Nitrogenase genes in non-cyanobacterial
619 plankton: prevalence, diversity and regulation in marine waters, *Aquat. Microbial Ecol.* 61,
620 235-247, 2010.

621 Rees, A. P., Gilbert, J. A., and Kelly-Gerreyn, B. A.: Nitrogen fixation in the western English
622 Channel (NE Atlantic Ocean), *Mar. Ecol. Progr. Ser.*, 374, 7-12, 2009.

623 Saitoh, Y., Kuma, K., Isoda, Y., Kuroda, H., Matsuura, H., Wagawa, T., Takana, H.,
624 Kobayashi, N., Nagao, S., and Nakatsuka, T.: Processes influencing iron distribution in the
625 coastal waters of the Tsugaru Strait, Japan, *J. Oceanogr.*, 64, 815-830, 2008.

626 Shiozaki, T., Furuya, K., Kodama, T., and Takeda, S.: Contribution of N₂ fixation to new
627 production in the western North Pacific Ocean along 155°E, *Mar. Ecol. Progr. Ser.* 377,
628 19-32, 2009.

629 Shiozaki, T., Furuya, K., Kodama, T., Kitajima, S., Takeda, S., Takemura, T., and Kanda, J.:
630 New estimation of N₂ fixation in the western and central Pacific Ocean and its marginal seas,
631 *Global. Biogeochem. Cycles.*, 24, GB1015, doi:10.1029/2009GB003620, 2010.

632 Shiozaki, T., Kodama, T., Kitajima, S., Sato, M., and Furuya, K.: Advective transport of
633 diazotrophs and importance of their nitrogen fixation on new and primary production in the
634 western Pacific warm pool, *Limnol. Oceanogr.*, 58(1), 49-60, 2013.

635 Shiozaki, T., Ijichi, M., Kodama, T., Takeda, S., and Furuya, K.: Heterotrophic bacteria are

636 major nitrogen fixers in the euphotic zone of the Indian Ocean, *Glob. Biogeochem. Cycles*,
637 28, doi:10.1002/2014GB004886, 2014a.

638 Shiozaki, T., Ito S.-I., Takahashi, K., Saito, H., Nagata, T., and Furuya, K.: Regional
639 variability of factors controlling the onset timing and magnitude of spring algal blooms in the
640 northwestern North Pacific, *J. Geophys. Res.*, 119, 1-13, doi:10.1002/2013JC009187, 2014b.

641 Shiozaki, T., Chen, Y.-L. L., Lin, Y.-H., Taniuchi, Y., Sheu, D.-S., Furuya, K., and Chen,
642 H.-Y.: Seasonal variations of unicellular diazotroph groups A and B, and *Trichodesmium* in
643 the northern South China Sea and neighboring upstream Kuroshio Current, *Cont. Shelf Res.*,
644 80, 20-31, 2014c.

645 Subramaniam, A., Yager, P. L., Carpenter, E. J., Mahaffey, C., Björkman, K., Cooley, S.,
646 Kustka, A. B., Montoya, J. P., Sañudo-Wilhelmy, S. A., Shipe, R., and Capone, D. G.:
647 Amazon River enhances diazotrophy and carbon sequestration in the tropical North Atlantic
648 Ocean, *Proc. Natl. Acad. Sci. U.S.A.*, 108, 2184-2189, 2008.

649 Tamura, K., Peterson, N., Peterson, G., Stecher, M., Nei, M., and Kumar, S.: MEGA5:
650 Molecular Evolutionary Genetics Analysis using maximum likelihood, evolutionary distance,
651 and maximum parsimony methods, *Mol. Biol. Evol.*, 28, 2731-2739, 2011.

652 Zehr, J. P., and Turner, P. J.: Nitrogen fixation: nitrogenase genes and gene expression, *Meth.*
653 *Microbiol.*, 30, 271-285, 2001.

654 Zehr, J. P., Jenkins, B. D., Short, S. M., and Steward, G. F.: Nitrogenase gene diversity and
655 microbial community structure: a cross-system comparison, *Environ. Microbiol.*, 5(7),
656 539-554, 2003a.

657 Zehr, J. P., Crumbliss, L. L., Church, M. J., Omoregie, E. O., Jenkins, B. D.: Nitrogenase
658 genes in PCR and RT-PCR reagents: implications for studies of diversity of functional genes,
659 *BioTechniques*, 35, 996-1005, 2003b.

660 Zehr, J. P.: Nitrogen fixation by marine cyanobacteria, *Trends Microbiol.* 19, 162-173, 2011.

661

662 Table 1. Summary of recovered *nifH* sequences belonging to *Trichodesmium* (Tri), UCYN-A
663 (UA), *Leptolyngbya* (Lep), α -proteobacteria (α -Pro), β -proteobacteria (β -Pro),
664 γ -proteobacteria (γ -Pro), δ -proteobacteria (δ -Pro), and Cluster III (CIII)

Cruise	Station	No. of clones	Cyanobacteria			α -Pro	β -Pro	γ -Pro	δ -Pro	CIII
			Tri	UA	Lep					
KT-12-20	OT4	12		9		3				
	ON1	5		2						3
	ON5	8		8						
	ON7	7		1		6				
Total		32	0	20	0	9	0	0	0	3
KT-12-27	OT4	7	1							6
	ON1	9						4(2)		5(5)
	ON5	6					1			5
	ON7	13	6	1		5(5)		1(1)		
Total		35	7	1	0	5(5)	0	2(1)	4(2)	16(5)
KT-13-2	OT4	11			10			1		
	ON1	1								1
	ON5	14				5(5)			2(2)	7
Total		26	0	0	10	5(5)	0	1	2(2)	8
KK-13-1	OT4	10		2		8(8)				
	ON1	15		3				2	10(10)	
	ON5	11		4		7(7)				
	ON8	1					1			
Total		37	0	9	0	15(15)	1	2	10(10)	0
KK-13-6	OT4	7							4(4)	1
	ON5	11		11						
	ON7	10		2		1		7		
Total		28	0	13	0	1	0	7	4(4)	1
KS-14-2	OT4	10							1(1)	9
	ON1	13				3(3)	3	1(1)	3(3)	
	ON5	15				2(2)				9
Total		38	0	0	0	5(5)	3	1(1)	4(4)	18

665 Numbers in parentheses indicate the number of sequences with >97% similarity at the amino
666 acid level to terrestrial diazotroph sequences.

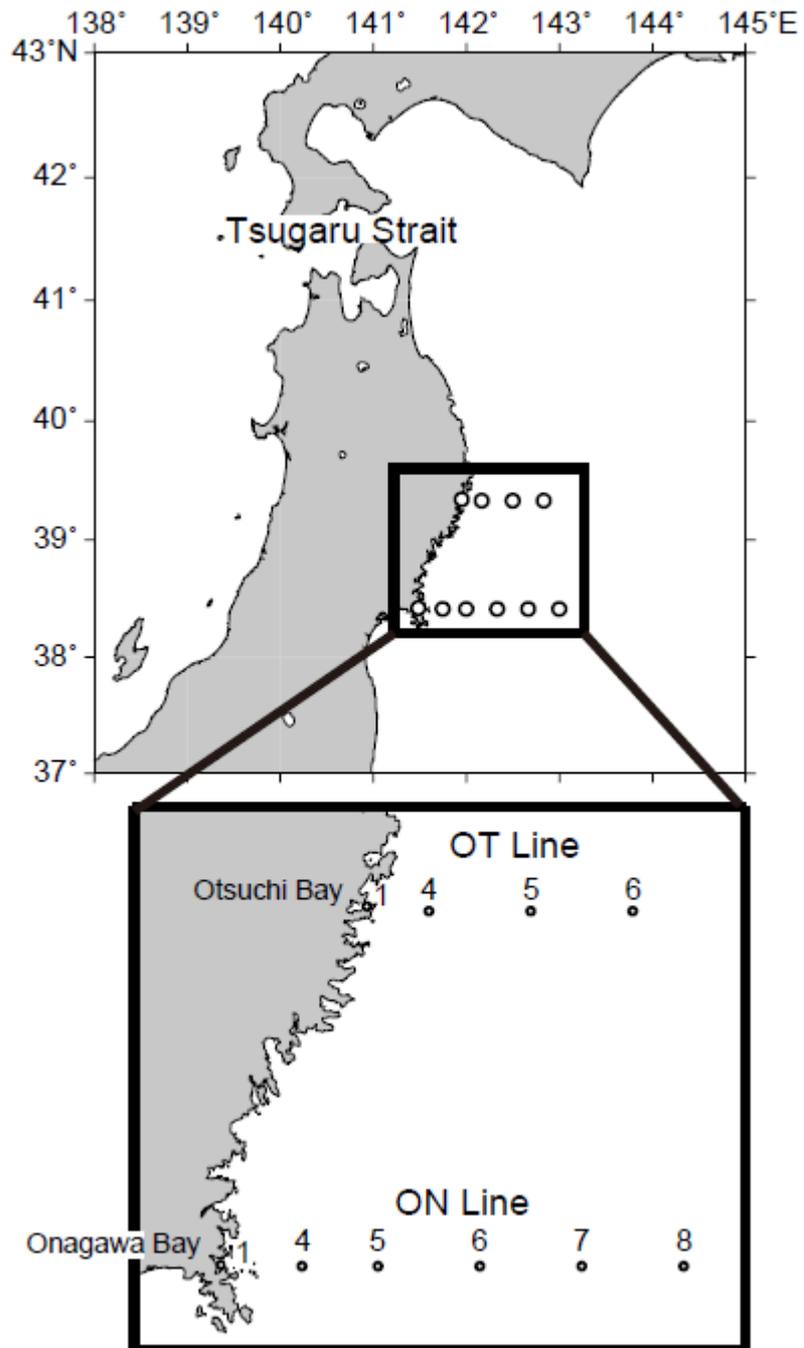
667 Table 2 Pearson's correlation matrix of N₂ fixation rates and water properties at the
 668 surface (n=73).

	Temperature	Nitrate	Ammonium	Phosphate	N/P ratio	N ₂ fixation
Temperature	1					
Nitrate	-0.722**	1				
Ammonium	-0.038	0.440**	1			
Phosphate	-0.863**	0.867**	0.135	1		
N/P ratio	-0.125	0.216	0.171	0.009	1	
N ₂ fixation	0.435**	-0.325**	-0.124	-0.335**	-0.130	1

669 * $p < 0.05$, ** $p < 0.01$

670 N/P ratio denotes the ratio of (nitrate + nitrite + ammonium) to phosphate

671

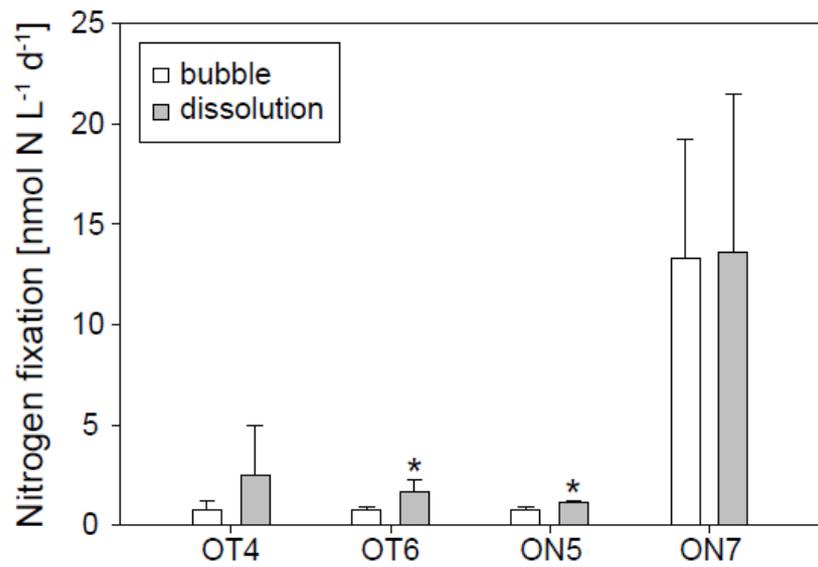


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674 Fig. 1. Sampling locations in the northwestern North Pacific Ocean.

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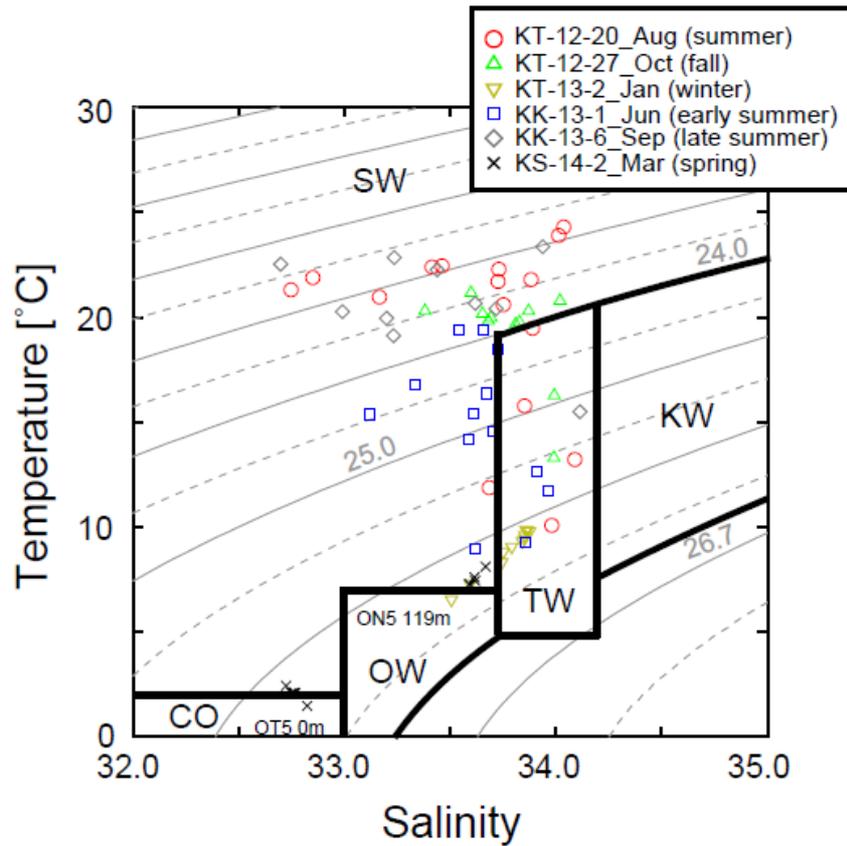


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678 Fig.2. Nitrogen fixation rates estimated simultaneously by the ¹⁵N₂ gas bubble and
 679 dissolution methods during the KK-13-6_Sep cruise. An asterisk indicates a significant
 680 difference between the two methods ($p < 0.05$).

681

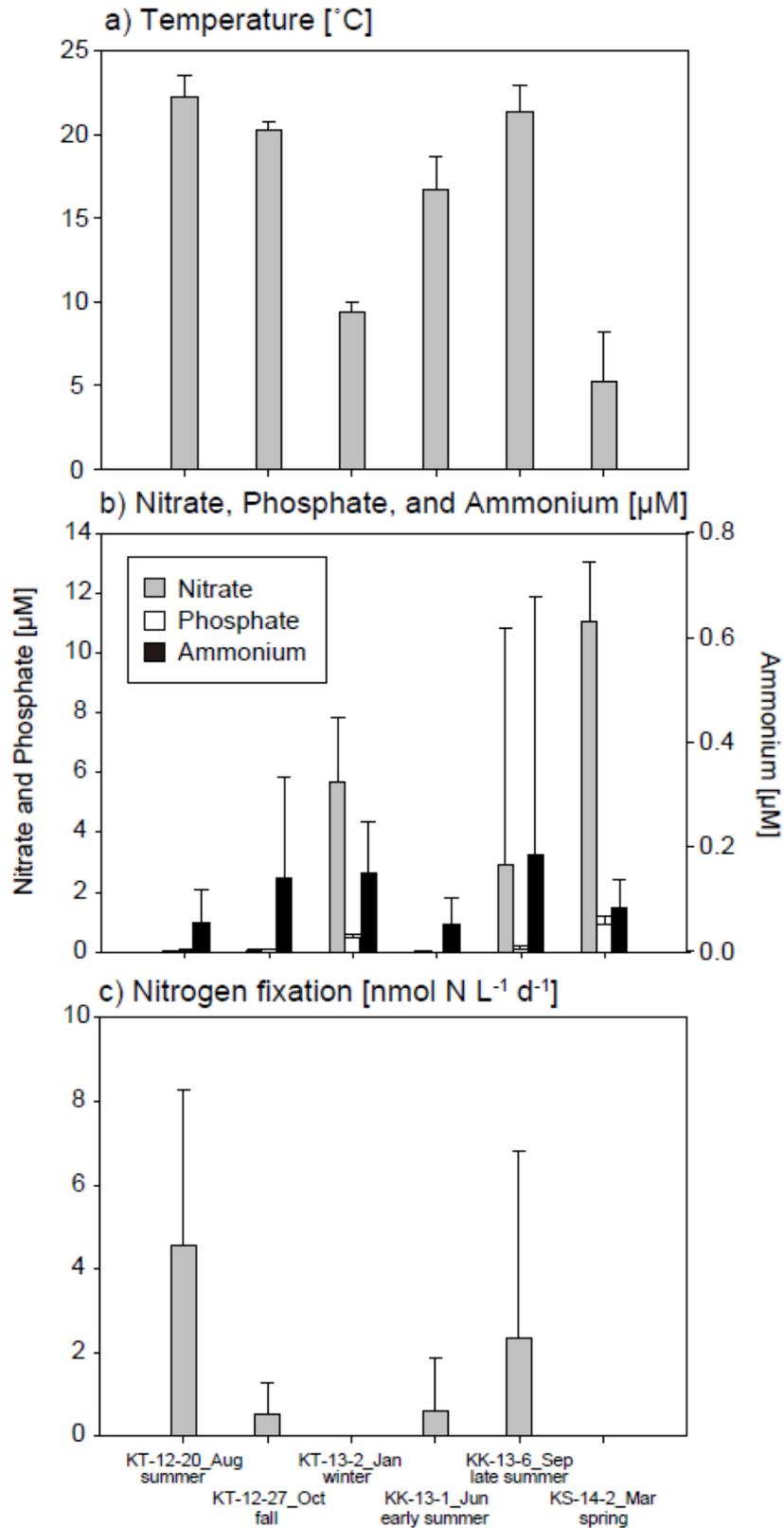


682

683

684 Fig. 3. Temperature-salinity diagram at each sampling point. The water classification was
 685 defined by Hanawa and Mitsudera (1986). SW, KW, TW, OW, and CO denote the surface
 686 layer water system, Kuroshio water system, Tsugaru Warm Current water system, Oyashio
 687 water system, and Coastal Oyashio water system, respectively.

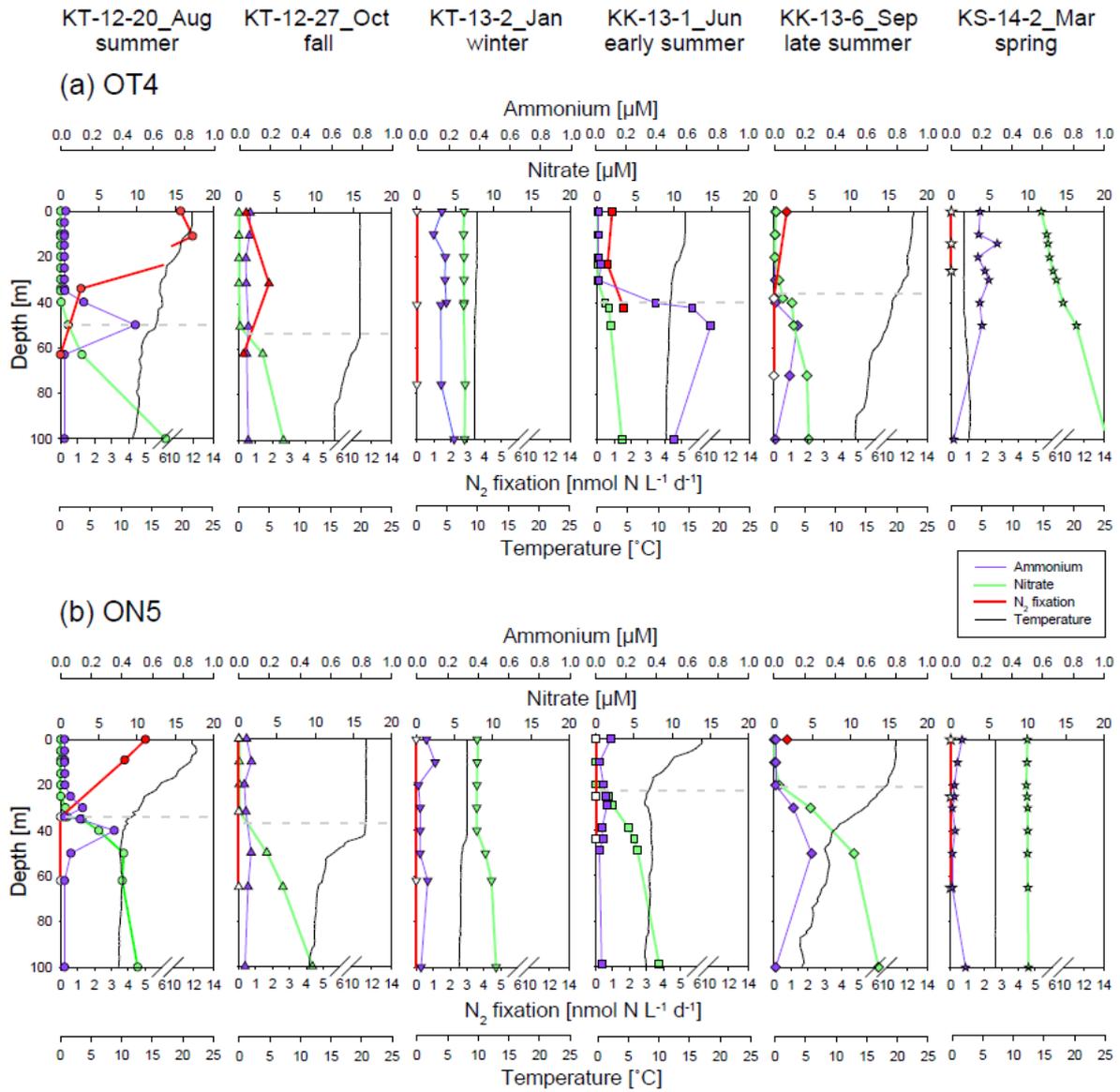
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689

690 Fig. 4. Average (a) temperature [°C], (b) nitrate, phosphate, and ammonium concentrations

691 [μM], and (c) nitrogen fixation [nmol N L⁻¹ d⁻¹] at the surface during each cruise.



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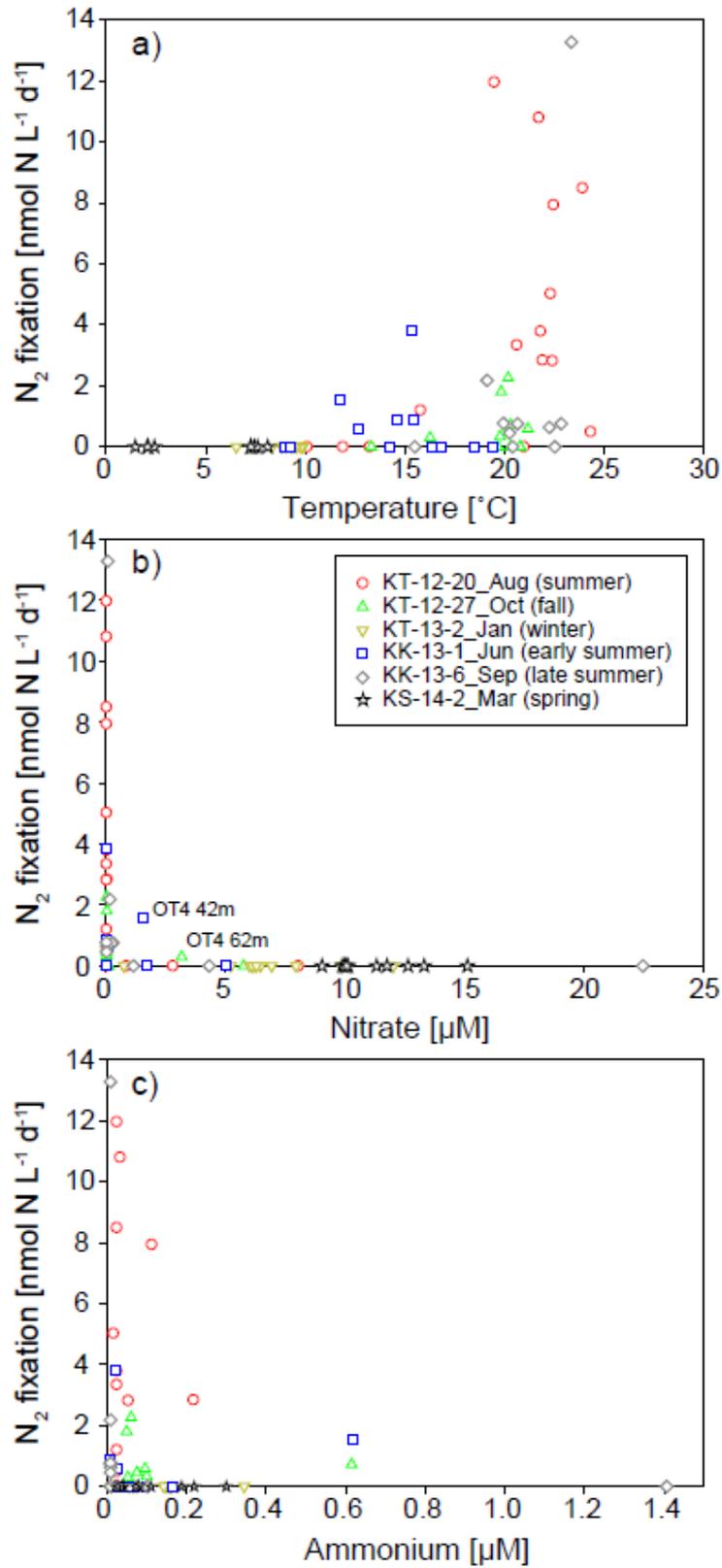
694 Fig. 5. Time-series variations in the vertical profiles of temperature [°C] (black),

695 ammonium (purple) and nitrate (green) concentration [μM], and nitrogen fixation (red) [nmol

696 N L⁻¹ d⁻¹] at Stns (a) OT4 and (b) ON5. Open symbols indicate that nitrogen fixation was

697 not detected. The horizontal dashed line indicates the nitracline depth. The straight lines of

698 temperature and nitrate were ascribable to strong mixing.

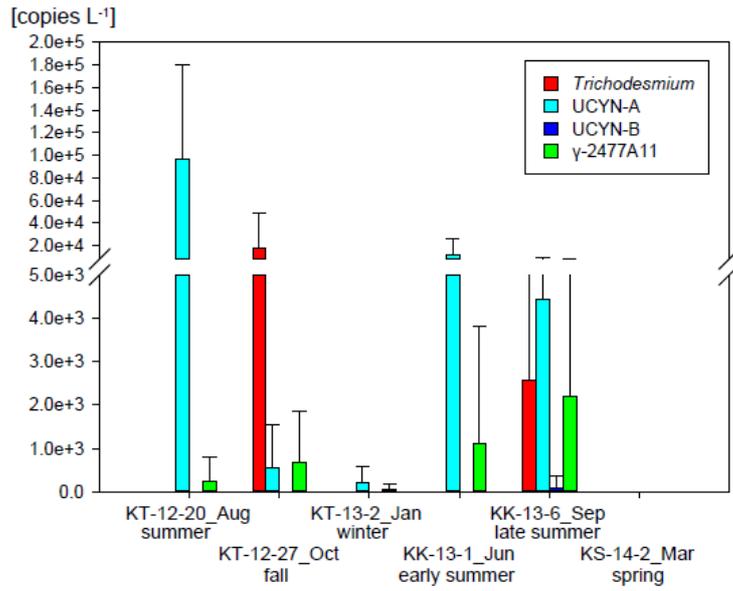


699

700 Fig. 6. Relationship between nitrogen fixation [$\text{nmol N L}^{-1} \text{d}^{-1}$] and (a) temperature [$^{\circ}\text{C}$], (b)

701 nitrate [μM], and (c) ammonium [μM] for all six cruises.

702



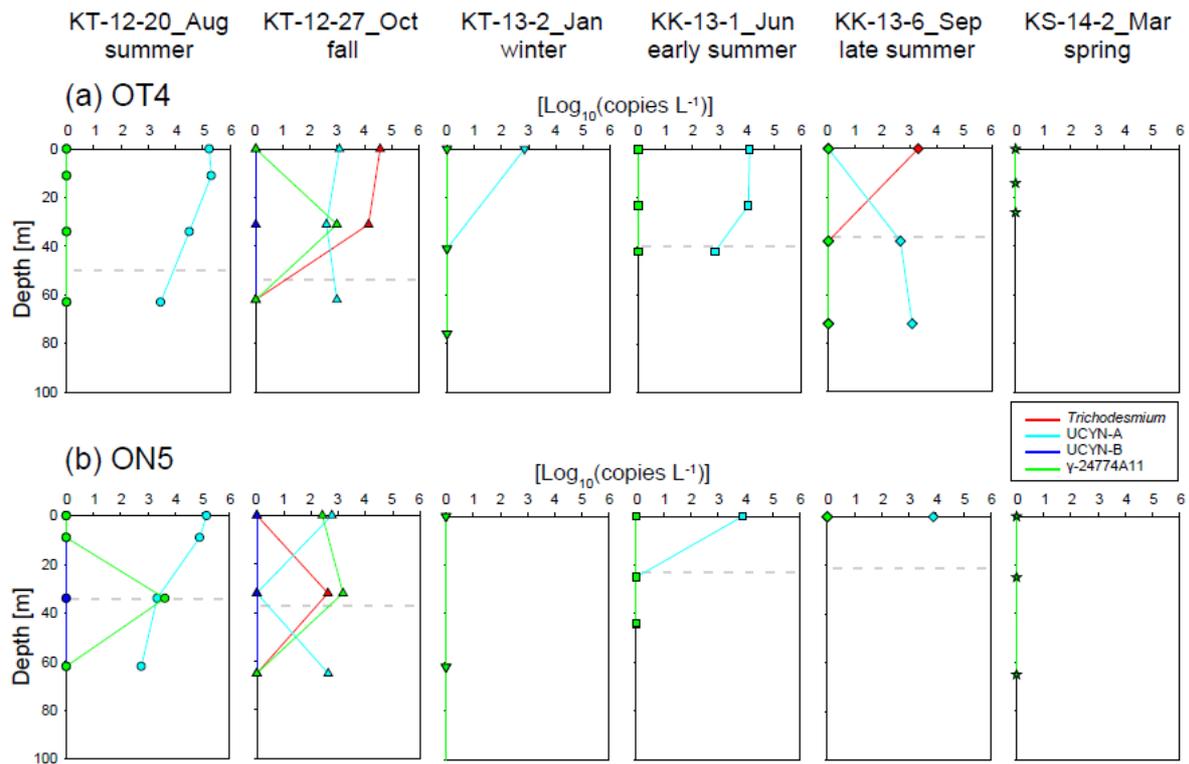
703

704

705 Fig. 7. Average abundances of *Trichodesmium* (red), UCYN-A (light blue), UCYN-B

706 (blue), and γ-24774A11 (green) [copies L⁻¹] at the surface during each cruise.

707



708

709

710 Fig. 8. Time-series variations in the vertical profiles of *Trichodesmium* (red), UCYN-A

711 (light blue), UCYN-B (blue), and γ -24774A11 (green) [copies L⁻¹] at Stns. (a) OT4 and (b)

712 ON5. The horizontal dashed line indicates the nitracline depth.