

## Answers to the interactive comment by Anonymous Referee #1

Before we address the concerns of referee #1, we would like to thank the referee for the time she/he invested to write her/his thoughtful and constructive comments. For clarity we have copied the review in italics and address the concerns of the referee, using normal fonts

*GENERAL COMMENTS Oil palm is one of the most rapidly expanding and financially important cropping systems in the tropics, yet little is known or understood about the biogeochemistry of these ecosystems, or their ability to release or sequester carbon. The work presented here is therefore interesting, important and novel because it provides much-needed, high quality empirical data on the fluxes of carbon from representative systems in the Southeast Asia region, over an annual cycle. This manuscript distinguishes itself from many of the other papers that have preceded it, because the sampling is more spatially extensive and over a longer time period than other work, enabling the investigators to make more confident assertions about the annual fluxes of CO<sub>2</sub> and CH<sub>4</sub> from these systems.*

*The findings from this work will provide an excellent basis for understanding the mechanistic controls on CO<sub>2</sub> and CH<sub>4</sub> fluxes from oil palm systems planted on mineral soils, with the potential for future modelling or up-scaling. The focus on smallholder systems is also noteworthy, because smallholder cultivation typically accounts for 40-60 % of all plantations in Southeast Asia, and therefore represents a sizeable proportion of land cover under this cropping system. Studies on smallholder systems are also less common, and therefore this study serves as a useful point of comparison compared to measurements from large, corporate plantations. However, one of the obvious scientific challenges for this study is the fact that smallholder management tends to be more heterogeneous than that of large agribusinesses (e.g. see page 7 line 22 – page 8 line 9). Yet despite this constraint, I believe that this dataset is still easily interpretable, given the selection of adequate controls (i.e. forest, jungle rubber) and the concomitant measurement of continuous independent variables (e.g. available N, P, soil texture, temperature, moisture, etc.). In addition, the use of small-scale manipulative field experiments, such as the simulated fertiliser addition study, helped to more unambiguously establish the mechanistic links between the fluxes and measured environmental variables.*

*My only other comment/minor concern is with respect to the content and organization of the Discussion; while the authors do a good job of comparing their findings against the existing literature, I felt that the paper could be made more impactful if the authors made it clearer in the Discussion which of their findings was novel or interesting. As it is written, it is sometimes difficult for the reader to identify the most exciting results from this study. The Discussion could also be slightly streamlined in terms of length, or slightly re-packaged/revised to better highlight the most important findings.*

We have addressed this concern by making the following changes: throughout our discussion we now start each paragraph with an introductory sentence in which the most important and novel interpretation discussed in this paragraph is presented, followed by the reasoning behind this. In the previous version, in some parts of the discussion this was already the case, but in other parts, the most exciting findings were 'hidden' in the text.

In the previous discussion, we were not able to cite a source showing reliably that under oil palm the soil carbon stocks strongly decrease with time. The source that we referred to (Allen et al., 2015) did not show a decrease, which was caused by the large spatial variability of soil carbon stocks among

our sites. However, we were now able to 'streamline' the discussion by citing a study that was recently published by our group, which show strong reductions of soil C stocks in oil palm plantations (van Straaten et al., 2015). This study, which was also conducted in Jambi province, had many more site replicates, and showed that the observed decreases in soil C stocks were highly significant.

We think that by making these changes we have adequately addressed this concern of reviewer #1.

*Specific comments on individual portions of the text are provided in the section below.*

*SPECIFIC COMMENTS 1. Page 9, line 14: Were any of the CO<sub>2</sub> or CH<sub>4</sub> data nonlinear? If so, how were these data dealt with?*

In page 10 L3-L12, we addressed this question in the revised manuscript. Only in very few measurements of CO<sub>2</sub> fluxes, the last concentration measurement at the last sampling time (at 31 minutes after chamber closure) was not linear compared to the first 3 sampling times (at 1, 11 and 21 minutes after chamber closure). In such few cases, we excluded the last data point and calculated the fluxes based on the linear increase in concentrations during the first 3 sampling times. For the large majority of CH<sub>4</sub> flux measurements, the measured CH<sub>4</sub> concentrations were strongly linear with time (during 31 minutes after chamber closure). Only in a few cases when CH<sub>4</sub> uptake was low, CH<sub>4</sub> concentration change with time of chamber closure showed low R<sup>2</sup> in linear regression. In these few cases, however, the corresponding CO<sub>2</sub> concentrations (from the same gas sample as the gas sample was analyzed consecutively for CH<sub>4</sub> and CO<sub>2</sub>) were linear, indicating no mistake in sampling. Thus, we still estimated fluxes from these low changes in CH<sub>4</sub> concentration with time using linear regression because, even if there was low linearity exhibited, this flux was a real manifestation of the balance between CH<sub>4</sub> uptake in and emission from the soil. All CH<sub>4</sub> flux measurements were included in all statistical analysis.

*2. Page 13, lines 25: Please revise this sentence, as the structure is a bit awkward and the sentence does not read smoothly.*

We changed this sentence into (page 11 L27):

'We extended the LME model to include either 1) a variance function that allows different variances of the fixed effect, and/or 2) a first-order temporal autoregressive process, which assumes that the correlation between measurements decreases with increasing time difference, if this improved the relative goodness of the model fit based on the Akaike information criterion.'

*3. Page 22, lines 19-29: As the authors allude, one cause for the reduced soil respiration fluxes in oil palm system may be because of lower root respiration in the oil palm system relative to the forest due to lower overall root biomass. Do the authors know if the oil palm systems had lower root biomass than their other ecosystems? Could they use data from spatially explicit sampling (e.g. sampling in gradients away from palms/trees; see page 13 lines 19-28) to estimate root versus heterotrophic (saprotrophic) respiration? I realise that this would be a back-of-the-envelope calculation, but it may be a useful point of discussion, given that the spaces between palms in many plantations have very sparse plant cover (and therefore could be used to estimate the root-free rate of soil respiration).*

As we point out in the discussion, fine and coarse root production was lower in the oil palm plantation compared to the forest (Kotowska et al., 2015). We did not conduct spatially explicit sampling of the root biomass and unfortunately cannot make the back-of-the-envelope calculation as suggested by reviewer #1.

## References

Allen, K., Corre, M. D., Tjoa, A., and Veldkamp, E.: Soil nitrogen-cycling responses to conversion of lowland forests to oil palm and rubber plantations in Sumatra, Indonesia, *PloS one*, 10, e0133325, 2015.

Kotowska, M. M., Leuschner, C., Triadiati, T., Meriem, S., and Hertel, D.: Quantifying above- and belowground biomass carbon loss with forest conversion in tropical lowlands of Sumatra (Indonesia), *Glob Chang Biol*, doi: 10.1111/gcb.12979, 2015. 2015.

van Straaten, O., Corre, M. D., Wolf, K., Tchienkoua, M., Cuellar, E., Matthews, R. B., and Veldkamp, E.: Conversion of lowland tropical forests to tree cash crop plantations loses up to one-half of stored soil organic carbon, *Proc Natl Acad Sci U S A*, 112, 9956-9960, 2015.

## Answers to the interactive comment by Anonymous Referee #2

Before we address the concerns of referee #2, we would like to thank the referee for the time she/he invested to write her/his thoughtful and constructive comments. For clarity we have copied the review in italics and address the concerns of the referee, using normal fonts

*The presented study aims to document the magnitude and investigate the drivers of soil-atmosphere carbon dioxide and methane exchange in Sumatra, Indonesia. To this end, two land-use change gradients on soils of differing texture were used to investigate the influence of soil type and forest conversion. Given considerable economic pressure for forest conversion, the reported data represents a timely and comprehensive contribution to the relatively sparse literature addressing the dynamics of soil-atmosphere greenhouse gas exchange in the region. The experimental design reflects considerable dedication in the field and manuscript is generally well written and reasoned. With this in mind I have a few queries and suggestions that I feel should be considered to improve the clarity of the work for future readers.*

*1) Very minor but I'd quite like to see annual fluxes reported in the abstract to provide a point of reference for the key findings described.*

Following this good suggestion we have now included the mean CO<sub>2</sub> and CH<sub>4</sub> fluxes (or range of fluxes) in the abstract. We chose to report the mean fluxes instead of the annual fluxes, since the mean fluxes are purely based on measurements, while the annual fluxes are based on trapezoidal extrapolation.

*2) I think the methods section would be clearer if the text (Page 9174, line 15 – Page 9175, line 7) describing the experimental set-up of the fertilisation manipulation is moved from '2.2. CO<sub>2</sub> and CH<sub>4</sub> flux measurement' and included in the previous section '2.1. Study area and experimental design'.*

We agree with this suggestion and moved the description of the fertilization experiment to section 2.1

*3) It is stated that carbon dioxide fluxes were calculated using a linear model fit, however, the text does not make it clear if this approach was also applied to methane fluxes (Page 9174, lines 5 –9). Evidence of non-linearity in the change of headspace concentration (a common observation in static chamber data that is acknowledged by the authors on Page 9173, line 1) might imply that a different approach to flux calculation e.g. Pedersen et al., 2010 may be more appropriate. Clarification of the approach used is required.*

In page 10 L3-L12, we addressed this question in the revised manuscript. Only in very few measurements of CO<sub>2</sub> fluxes, the last concentration measurement at the last sampling time (at 31 minutes after chamber closure) was not linear compared to the first 3 sampling times (at 1, 11 and 21 minutes after chamber closure). In such few cases, we excluded the last data point and calculated the fluxes based on the linear increase in concentrations during the first 3 sampling times. For the large majority of CH<sub>4</sub> flux measurements, the measured CH<sub>4</sub> concentrations were strongly linear with time (during 31 minutes after chamber closure). Only in a few cases when CH<sub>4</sub> uptake was low, CH<sub>4</sub> concentration change with time of chamber closure showed low R<sup>2</sup> in linear regression. In these few cases, however, the corresponding CO<sub>2</sub> concentrations (from the same gas sample as the gas sample was analyzed consecutively for CH<sub>4</sub> and CO<sub>2</sub>) were linear, indicating no mistake in sampling. Thus, we still estimated fluxes from these low changes in CH<sub>4</sub> concentration with time using linear regression

because, even if there was low linearity exhibited, this flux was a real manifestation of the balance between CH<sub>4</sub> uptake in and emission from the soil. All CH<sub>4</sub> flux measurements were included in all statistical analysis. This approach was also followed in the study by (Verchot et al., 2000), that was referred to by reviewer #2.

*4) Similarly, the authors indicate that net zero methane fluxes were retained in their dataset (Page 9174, line 10) but do not state the criteria used to define measurements as such e.g. Verchot et al., 2000, Pedersen et al., 2010, Parkin et al., 2012. I think it is important to indicate the lines along which measurements are defined as zero or omitted from the dataset (i.e. non-significant fits resulting from small flux rates vs. those caused by errors in sampling, storage or analysis) and the number of 'zeros' retained. The treatment of zero fluxes in the literature can be somewhat patchy, despite the fact their inclusion / exclusion inherently introduces biases, so this sort of information is very useful to future readers when comparing reported flux rates across studies. Again, clarification is required.*

Apparently our description how we handle methane fluxes that were not significantly different from zero was not clear. We recognized that some of the fluxes were not significantly different from zero; however these fluxes were not omitted from the dataset. Therefore, we did not use any criteria to exclude zero fluxes and accordingly we do not report these criteria. We have now changed the description in the methods section as follows (page 10, L8):

‘There were a few measurements when CH<sub>4</sub> concentrations changed only minutely with time of chamber closure, mostly when net CH<sub>4</sub> uptake was low; in such cases, the calculated CH<sub>4</sub> flux using linear regression was not significantly different from zero. These fluxes were however retained in the statistical analyses to avoid bias by excluding low CH<sub>4</sub> fluxes or by assuming that these fluxes were zero.’

*5) Which variables were transformed and where these transformations were subsequently used in statistical tests should be indicated (Page 9176, line 13). Full assessment of the reported relationships is not really possible in the absence of this information.*

We now specified (page 11, L 17-18) which transformations were used for the measured parameters, whenever necessary for the comparisons among land-use types within each landscape or between landscapes within each land-use type.

‘. . . and if necessary a logarithmic (for CO<sub>2</sub>, CH<sub>4</sub>, and mineral N) or square root (for WFPS) transformation was used.’

*6) Mixed effect models are used to account for spatial and temporal structures in the experimental design when testing the effect of categorical variables (i.e. landscape and land-use) on mean fluxes at the level of plot/palm and sampling date. However, this approach is not extended to testing relationships between fluxes and other continuous variables. Instead the dataset is reduced and Pearson's or Spearman's correlation tests are applied to investigate temporal and spatial variability. This seems fine to me but I think it might be useful for the authors to (v. briefly) explain the reasoning behind their approach (Page 9176, line 14 – Page 9177, line 15).*

The statistical results are essentially the same either using LME with mean centering of the data set or using the mean values of replicate plots for each treatment (land use) on each sampling day and conducting Pearson correlation across the measurement period (i.e. in our study n=12 months) in order to assess the temporal controls on soil CO<sub>2</sub> and CH<sub>4</sub> fluxes (section 3.3). We straightforwardly

used Pearson correlation as this is more commonly reported; also, as the correlation coefficients using Pearson or LME with mean centering of the data are the same anyway. The reason why we used the mean values of the 4 replicate plots for each land use type on each sampling day is in order to represent the center of the spatial structure per land use and focus the analysis mainly on the temporal pattern for such land use. We added this reason in page 12 L6-7.

For the spatial controls on soil CO<sub>2</sub> and CH<sub>4</sub> fluxes (section 3.4), we used the annual fluxes from each replicate plot within each landscape (n=16 for 4 land-use types x 4 plots) and the average soil physical and biochemical characteristics per replicate plot (Table A1), which were only measured once in 2013 since these soil variables are not going to vary within a year. In the same analogy as that with the temporal control, we used the annual values to represent the temporal pattern across the monthly measurement and focus this analysis on spatial pattern. Since we used the individual plot value (not the average of the 4 plots per land use), we used the Spearman rank correlation so as not to conduct any data transformation of the soil variables which have non-normal distribution. We added this reason in page 12 L13-14.

These are the approaches that we have typically employed in many of our earlier works on soil trace gas flux measurements (e.g. Koehler et al., 2009; Veldkamp et al., 2013; Corre et al., 2014) in order to present the data analysis correctly yet in a less confusing manner.

*7) It's not always clear which statistical tests have been applied throughout the results section. Maybe include a reminder in sections 3.3 and 3.4 that the reported correlation coefficients are Pearson's  $r$  and Spearman's  $\rho$ , respectively.*

We have partly followed this suggestion. Since all our figures and tables clearly state our statistical methods (including Table 3 where the Pearson's correlations are reported) we did not add this information in section 3.3. However, the first time we report the Spearman's  $\rho$  correlation coefficients in section 3.4, we specify this test. Furthermore, we changed the symbol that we used for Spearman's correlation to  $\rho$  ( $p$ ) throughout the text in order to make this clearer.

*8) The results of the fertilisation manipulation seem to receive relatively limited discussion beyond consideration of spatial footprints. For example, smaller carbon dioxide fluxes were identified at the furthest chamber location from a palm (Page 9179, line 7). It's not clear whether this is a result of the application of the fertilizer or whether it results from a more general pattern related to distance from palms e.g. driven by differences in root biomass and respiration. Is a relationship present between flux and distance from palms in the main dataset (i.e. when considering chambers positioned 1.8 – 5 m from palms and measured monthly throughout the year; Page 9179, line 13)? Or did the flux return to pre-fertilization levels as reported for methane (Page 9179, line 27- 28)?*

We did not put a lot of emphasis on the fertilization manipulation since the temporal and spatial effects that we measured were relatively insignificant, as we also stated in the Results (page 14, L1-5, L19-21). There was no systematic change (i.e. significant correlation) in CO<sub>2</sub> and CH<sub>4</sub> fluxes with distance to palms when we conducted Spearman correlation using annual fluxes from all individual chambers. Since the subplots where measurements were conducted were selected randomly and the chamber bases were also randomly located in the subplot, such absence of systematic change with distance to palms indicated that the subplot measurements represented the plot. Moreover, none of the chamber bases in this dataset were closer than 1.8 m from the palm tree, whereas the strongest effects from fertilization was within 1 m from the palm trees. The effects that we reported for CO<sub>2</sub>

fluxes in the fertilization experiment were the result of a general pattern, probably related to differences in root biomass and respiration. They did not disappear or change with time as was the case with CH<sub>4</sub> fluxes.

*9) Support for the main thrust of the manuscript in relating carbon dioxide and methane fluxes to soil fertility is heavily reliant on the results reported in section '3.4 Spatial controls of annual CO<sub>2</sub> and CH<sub>4</sub> fluxes across land-use types within each landscape' i.e. the relationships presented in the abstract relating carbon dioxide flux positively with SOC and negatively with 15N, extractable P and base saturation (Page 9165, line 14 – 16) and methane uptake negatively with N availability and positively with Al availability (Page 9165, line 20 – 23) are presented here. However, I find this section a little hard to follow as a correlation matrix (like Table 3 for temporal relationships) isn't shown. I realise there are a large number of variable pairs and multiple scales considered but I think some sort of table could be very useful given how central these results are to the manuscript. Particularly, I'm unclear as to whether relationships between annual means of fluxes and environmental variables were considered as drivers of spatial variability. Given the highlighted importance of WFPS in driving temporal variability in methane flux (Table 3) from these soils I would like to see the possibility that variability in WFPS could be driving spatial variability addressed. Indeed, a lack of a relationship here, as similarities in bulk density (Table A1) and unclear patterns in WFPS between land-uses (Figure 1) might suggest, would serve to strengthen the argument made for fertility as the key driver across this system.*

The main reason why we chose not to present the correlation matrices in section 3.4 is that there are simply too many variables, and instead we chose to present only the correlations of annual soil CO<sub>2</sub> and CH<sub>4</sub> fluxes with soil variables that showed significant correlations. So the correlations mentioned in the text are the only ones that were significant and hence it makes no sense to present all soil variables in a correlation matrix table. As stated in Statistical analysis (page 12, L2-7): we first conducted a Spearman correlation of annual CO<sub>2</sub> and CH<sub>4</sub> fluxes with the soil physical and biochemical characteristics (Appendix Table A1) for the reference land uses across the two studied landscapes (correlation matrix 1), and second across land-use types for each landscape separately (correlation matrices 2 & 3). We think that such correlation matrices are too much to present, even for an Appendix because many soil variables are auto-correlated with each other and only few soil variables (the ones mentioned in the text) showed significant correlations. This last point was maybe not clear and we have now added the following sentence to paragraph 3.4 (page 15, L28-29):

'Apart from the correlations reported here, there were no other significant correlations with any of the tested soil physical and biochemical characteristics.'

There was no significant correlation between annual CH<sub>4</sub> fluxes and WFPS, since we had to average the WFPS across the monthly measurements to correlate with one annual value. In essence, if one has to look at the spatial pattern of annual soil CH<sub>4</sub> fluxes across plots in a landscape, WFPS (which displayed quite significant seasonal variability; Fig. 1a, b) is not a good predictor variable but probably annual rainfall. WFPS is more appropriate variable to correlation with temporal pattern of soil CH<sub>4</sub> fluxes (as we have reported in Table 3 and section 3.3).

## References

Corre, M. D., Sueta, J. P., and Veldkamp, E.: Nitrogen-oxide emissions from tropical forest soils exposed to elevated nitrogen input strongly interact with rainfall quantity and seasonality, *Biogeochemistry*, 118, 103-120, 2014.

Koehler, B., Corre, M. D., Veldkamp, E., Wullaert, H., and Wright, S. J.: Immediate and long-term nitrogen oxide emissions from tropical forest soils exposed to elevated nitrogen input, *Global Change Biology*, 15, 2049-2066, 2009.

Veldkamp, E., Koehler, B., and Corre, M. D.: Indications of nitrogen-limited methane uptake in tropical forest soils, *Biogeosciences*, 10, 5367-5379, 2013.

Verchot, L. V., Davidson, E. A., Cattanio, J. H., and Ackerman, I. L.: Land-use change and biogeochemical controls of methane fluxes in soils of eastern Amazonia, *Ecosystems*, 3, 41-56, 2000.



Marked-up manuscript version in the next pages.



# Soil fertility controls soil-atmosphere carbon dioxide and methane fluxes in a tropical landscape converted from lowland forest to rubber and oil palm plantations

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## Abstract

Expansion of palm oil and rubber production, for which global demand is increasing, causes rapid deforestation in Sumatra, Indonesia and is expected to continue in the next decades. Our study aimed to 1) quantify changes in soil CO<sub>2</sub> and CH<sub>4</sub> fluxes with land-use change, and 2) determine their controlling factors. In Jambi Province, Sumatra, we selected two landscapes on heavily weathered soils that differ mainly in texture: loam and clay Acrisol soils. At each landscape, we investigated the reference land uses: forest and secondary forest with regenerating rubber, and the converted land uses: rubber (7-17 years old) and oil palm plantations (9-16 years old). We measured soil CO<sub>2</sub> and CH<sub>4</sub> fluxes monthly from December 2012 to December 2013. Annual soil CO<sub>2</sub> fluxes from the reference land uses were correlated with soil fertility: low extractable phosphorus (P) coincided with high annual CO<sub>2</sub> fluxes from the loam Acrisol soil that had lower fertility than the clay Acrisol soil (P<0.05). Soil CO<sub>2</sub>

fluxes from the oil palm (107.2 to 115.7 mg C m<sup>-1</sup> h<sup>-1</sup>) decreased compared to the other land uses (between 178.7 and 195.9 mg C m<sup>-1</sup> h<sup>-1</sup>; P<0.01). Across land uses, annual CO<sub>2</sub> fluxes were positively correlated with soil organic carbon (C) and negatively correlated with <sup>15</sup>N signatures, extractable P and base saturation. This suggests that the reduced soil CO<sub>2</sub> fluxes from oil palm was a result of strongly decomposed soil organic matter and reduced soil C stocks due to reduced litter input, and possible reduction in C allocation to roots due to improved soil fertility from liming and P fertilization in these plantations. Soil CH<sub>4</sub> uptake in the reference land uses was negatively correlated with net nitrogen (N) mineralization and soil mineral N, suggesting N limitation of CH<sub>4</sub> uptake, and positively correlated with exchangeable aluminum (Al), indicating decrease in methanotrophic activity at high Al saturation. Reduction in soil CH<sub>4</sub> uptake in the converted land uses (ranging from -3.0 to -14.9 μg C m<sup>-2</sup> h<sup>-1</sup>) compared to the reference land uses (ranging from -20.8 to -40.3 μg C m<sup>-2</sup> h<sup>-1</sup>; P<0.01) was due to decrease in soil N availability in the converted land uses. Our study shows for the first time that differences in soil fertility control soil-atmosphere exchange of CO<sub>2</sub> and CH<sub>4</sub> in a tropical landscape, a mechanism that we were able to detect by conducting this study at the landscape scale.

## 1 Introduction

Oil palm (*Elaeis guineensis*) and rubber (*Hevea brasiliensis*) are two of the fastest expanding tree cash crops in the tropics (Clay, 2013). Global oil palm production has quintupled from 1990 to 2013 and is currently grown on an estimated area of 17 million hectare (Mha) (Food and Agricultural Organization, 2014). Indonesia contributes nearly half of global palm oil production (Food and Agricultural Organization, 2014), and is planning to double its production in the coming decade (Carlson et al., 2013). Similarly, rubber is grown on around 10 Mha globally, whereby Indonesia is the second largest rubber producer (Food and Agricultural Organization, 2014). Most Indonesian oil palm and rubber production are located in Sumatra (Indonesian Ministry of Agriculture, 2014), where conversion of lowland rainforest to plantations has been widespread (Laumonier et al., 2010). It has been estimated that plantation establishment has caused a loss of 7.5 Mha of Sumatran natural forest in the last two decades (1990-2010) (Margono et al., 2012), and future expansion will probably be at the expense of large areas of tropical forest, unless a properly planned and spatially explicit development strategy will be implemented (Koh and Ghazoul, 2010).

1 Although the majority of remaining lowland tropical forests are located on nutrient poor,  
2 heavily weathered soils, these ecosystems are among the most productive worldwide and  
3 contain globally significant above- and belowground carbon stocks. The high ecosystem  
4 productivity is possible despite the nutrient poor soils because of efficient cycling of rock-  
5 derived nutrients (phosphorus (P) and base cations) between vegetation and soil, and also high  
6 soil nitrogen (N) availability caused by biological N fixation (Hedin et al., 2009). Conversion  
7 of tropical forest to agricultural land-use systems does not only decrease biodiversity and  
8 contribute to climate change (Danielsen et al., 2009) but also alters soil fertility and soil  
9 physical properties in the newly established land-use systems (Dechert et al., 2004; Klinge et  
10 al., 2004). Burning of slashed vegetation is typically part of forest conversion, releasing large  
11 amounts of nutrients previously bound in the vegetation. A considerable part of these  
12 nutrients ends up in the soil but is susceptible to losses (through leaching and gaseous  
13 emission), which are especially high in the earlier years of crop establishment and decrease  
14 with time (Klinge et al., 2004). Furthermore, forest conversion is often associated with  
15 increases in soil bulk density. These dynamic changes in soil fertility and soil bulk density  
16 following forest conversion do not only affect agricultural production but also the soil-  
17 atmosphere exchange of trace gases like carbon dioxide (CO<sub>2</sub>) and methane (CH<sub>4</sub>) since their  
18 production, consumption and exchange are directly related to soil fertility and soil bulk  
19 density (Keller et al., 1993; Veldkamp et al., 2008).

20 Globally, soils are the largest natural source of CO<sub>2</sub> (IPCC, 2007), which is released during  
21 respiration processes of microbial communities and roots (Raich and Schlesinger, 1992).  
22 While the important proximal controllers of soil CO<sub>2</sub> fluxes are soil temperature and moisture,  
23 it has been demonstrated that other distal regulators such as vegetation type and soil physical  
24 and biochemical properties (e.g. bulk density, texture, pH, carbon stocks) also affect soil CO<sub>2</sub>  
25 fluxes (Raich and Schlesinger, 1992). Soils also play a dominant role in the production and  
26 consumption of CH<sub>4</sub>, a greenhouse gas with a global warming potential of 23 times that of  
27 CO<sub>2</sub> over a 100-year time horizon (IPCC, 2007). In soils, CH<sub>4</sub> can be produced during  
28 anaerobic decomposition by methanogenic archaea, while CH<sub>4</sub> can also be consumed by  
29 methanotrophic bacteria which are able to utilize CH<sub>4</sub> as an energy source. Whether net  
30 consumption or net emission of CH<sub>4</sub> occurs at the soil surface depends on the balance  
31 between production and consumption in the soil. For soil CH<sub>4</sub> fluxes, the proximal controllers  
32 are soil moisture, gas diffusivity and temperature, while other distal regulators include

1 microbial activity, N availability and aluminum toxicity (Verchot et al., 2000; Tamai et al.,  
2 2003; Bodelier and Laanbroek, 2004; Veldkamp et al., 2013).

3 In tropical lowland forest landscapes with heavily weathered soils, much of the spatial  
4 variability in trace gas fluxes appears to be related to soil texture (Sotta et al., 2006). In the  
5 Brazilian Amazon, lowland forests on Acrisol and Ferralsol soils display high soil CO<sub>2</sub>  
6 emissions with large variations among sites that relate to soil texture: soils with sandy loam to  
7 sandy clay loam texture had 21 - 36% higher CO<sub>2</sub> emissions than soils with clay texture  
8 (Keller et al., 2005; Sotta et al., 2006). Moreover, although well-drained soils in tropical  
9 lowland forests act generally as a sink for CH<sub>4</sub> (Keller and Reiners, 1994; Verchot et al.,  
10 2000; Veldkamp et al., 2013), their differences in CH<sub>4</sub> uptake are explicable by their  
11 differences in soil texture. In a review of 16 tropical lowland forests, the only factor  
12 correlating annual CH<sub>4</sub> fluxes with site characteristics was a significant positive correlation  
13 with clay contents, indicating that the higher the clay content the lower is the CH<sub>4</sub> uptake  
14 (Veldkamp et al., 2013).

15 Since much of the original forest in our study area have been converted to oil palm and rubber  
16 plantations, the management practices in these land uses added important factors that  
17 influence soil CO<sub>2</sub> and CH<sub>4</sub> fluxes from these converted landscapes. Earlier studies have  
18 shown that forest conversion to agricultural land uses in the tropics lead to considerable  
19 changes in soil CO<sub>2</sub> fluxes, which were related to changes in belowground C allocation  
20 (Davidson et al., 2000; Salimon et al., 2004), carbon quality (Werner et al., 2006), living fine  
21 root biomass and litter input (Ishizuka et al., 2002; Sheng et al., 2010). Conversion of tropical  
22 forest to agricultural land uses causes a reduction in soil CH<sub>4</sub> uptake or even turns the soil into  
23 a source of CH<sub>4</sub>. Often this trend is explained by soil compaction, which leads to reduced gas  
24 diffusivity and accordingly limits aerobic CH<sub>4</sub> oxidation while enhancing anaerobic CH<sub>4</sub>  
25 production (Keller et al., 1993; Veldkamp et al., 2008). Changes in N availability may also  
26 play a role since CH<sub>4</sub> uptake may be N limited (Bodelier and Laanbroek, 2004; Veldkamp et  
27 al., 2013) and high concentrations of ammonium (NH<sub>4</sub><sup>+</sup>, e.g. from fertilization) can inhibit  
28 CH<sub>4</sub> oxidation (Veldkamp et al., 2001; Werner et al., 2006). Finally, termites are known to  
29 produce CH<sub>4</sub> and their presence may also affect the balance between production and  
30 consumption of CH<sub>4</sub> (Seiler et al., 1984).

31 Although Sumatra, Indonesia represents a hot spot of land-use change, especially for the  
32 establishment of rubber and oil palm plantations, how this affects soil CO<sub>2</sub> and CH<sub>4</sub> fluxes

remains highly uncertain for the following reasons: (1) most studies relating land-use change to trace gas emissions have been conducted in South and Central America (Keller and Reiners, 1994; Davidson et al., 2000; Verchot et al., 2000; Veldkamp et al., 2001; Salimon et al., 2004) and only few studies were conducted in Southeast Asia (Ishizuka et al., 2002; Veldkamp et al., 2008); (2) most studies have focused on forest conversion to traditional land uses such as maize, pastures, slash-and-burn agriculture, cacao and coffee, and less on the rapidly expanding tree cash crops such as rubber and oil palm; (3) the few studies that reported CO<sub>2</sub> and CH<sub>4</sub> fluxes from oil palm plantations were conducted on peat soils (Melling et al., 2005a, b) whereas the studies conducted on mineral soils, where most of the rubber and oil palm plantations are located, were either conducted without spatial replication, covered only short periods of measurements (Ishizuka et al., 2002; Adachi et al., 2005; Werner et al., 2006) or measured only once (Ishizuka et al., 2005). It is imperative that better information becomes available on trace gas fluxes from these economically-important and rapidly-expanding rubber and oil palm plantations. Whether palm oil-based biofuel indeed has environmental advantages compared to fossil fuel depend, among many facets, on the greenhouse gas balance during oil palm fruit production.

In the present study, our aims were to 1) quantify changes in soil-atmosphere fluxes of CO<sub>2</sub> and CH<sub>4</sub> with land-use change, and 2) determine their controlling factors in a converted lowland landscape in Sumatra, Indonesia. Soil-atmosphere fluxes of CO<sub>2</sub> and CH<sub>4</sub> were measured in forest and secondary forest with regenerating rubber (hereafter called jungle rubber, which is a more traditional rubber agroforestry system (Gouyon et al., 1993)) as reference land uses and the converted land uses of monoculture rubber and oil palm plantations. Our study was designed to cover these four land-use types in each of the two landscapes on highly weathered soils that differed mainly in texture: clay and loam Acrisol soils. We tested the following hypotheses: 1) soil CO<sub>2</sub> emissions and CH<sub>4</sub> uptake will be higher in loam than in clay Acrisol soils, and 2) soil CO<sub>2</sub> fluxes and CH<sub>4</sub> uptake rates will be higher in the reference land uses (forest and jungle rubber) than in the converted land uses (rubber and oil palm plantations). Here, we present the first spatially replicated study with a full year of measurements that investigates soil CO<sub>2</sub> and CH<sub>4</sub> fluxes from conversion of forest or jungle rubber to rubber and oil palm plantations on mineral soils. We also evaluate the effect of management intensity since we compare rubber plantations without fertilizer inputs with fertilized oil palm plantations. Our results will be a critical contribution to trace gases life-cycle assessment of rubber and palm oil at the production stage.

## 2 Material and Methods

### 2.1 Study area and experimental design

The study area is located in the lowlands (35-95 m above sea level) of Jambi province, Sumatra, Indonesia. In the past two decades, forest cover in Jambi province decreased by 1.14 Mha, which was about 40% of the forest cover in 1990 (Margono et al., 2012). The climate is humid tropical with a mean annual air temperature of  $26.7 \pm 0.1$  °C and a mean annual precipitation of  $2235 \pm 385$  mm (1991-2011; data from Jambi-Sultan-Thaha airport of the Indonesian Meteorological, Climatological and Geophysical Agency). The dry season is usually from May to September and the rainy season occurs from October to April. In 2013, during our study period, the wet season lasted slightly longer, while a drier period was detected between mid-June and end-October. During this dry period, rainfall was reduced by 35 - 57% compared to the wetter months during which rainfall was 333 - 362 mm per month.

We selected two landscapes on heavily weathered soils that mainly differed in texture: loam Acrisol soil ( $36 \pm 6$  % sand,  $32 \pm 4$  % silt and  $32 \pm 2$  % clay in the top 0.5 m) and clay Acrisol soil ( $26 \pm 6$  % sand,  $29 \pm 3$  % silt and  $45 \pm 4$  % clay in the top 0.5 m). This textural difference led to differences in soil fertility: forest sites in the clay Acrisol soil had higher base saturation, Bray-extractable P and lower Al saturation compared to those in the loam Acrisol soil ( $P \leq 0.01$  to 0.04; Appendix Table A1; Allen et al., 2015). Detailed soil physical and biochemical characteristics from our study sites were measured by Allen et al. (2015) and are summarized in Appendix Table A1. Acrisol soils cover about 50% of the land area in Sumatra and about one third of Indonesia (FAO et al., 2009). The clay Acrisol landscape was located about 160 km southwest of Jambi City between 01.94° S, 102.58° E and 02.14° S, 102.85° E. Forest sites in this landscape were established within the Bukit Duabelas National Park (administered by the Ministry of Forestry, PHKA). The loam Acrisol landscape was located about 80 km southwest of Jambi City between 01.79° S, 103.24° E and 2.19° S, 103.36° E. The forest sites in this landscape were established within the Harapan Forest Reserve and had been partially logged in the past (administered by the Restoration Ecosystem Indonesia Harapan, PT REKI).

In each landscape, we studied four land-use types: lowland forest, jungle rubber, and smallholder monoculture plantations of rubber and oil palm. In Jambi province, the



smallholder rubber and oil palm plantations were established after clearing and burning either the forest (often partially logged) or jungle rubber (based on interviews conducted by Euler et al., unpublished data). Thus, in our study the lowland forest and jungle rubber served as the reference land uses, representing the baseline conditions with which we compared the rubber and oil palm plantations.

For each of the four land-use types within each landscape, we selected four replicate plots (50 m x 50 m each with a minimum distance of 200 m between plots), totalling to 32 plots that were all located on relatively flat, well drained positions in the landscape. Additional information on tree species composition, tree density, tree height, basal area, and plantation age of these plots are reported in Appendix Table A2. Within each plot, we established a 10 x 10 grid which was used to select four randomly nested subplots (5 m x 5 m each) that were at least 5 m from the plot's border. In each subplot, we randomly deployed one permanent chamber base to measure soil trace gas fluxes.

This approach of comparing soil CO<sub>2</sub> and CH<sub>4</sub> fluxes from the converted land uses to the reference land use in order to assess the effects of land-cover change has the implicit assumption that before land-use conversion soil characteristics were comparable. We tested this assumption by comparing the land-use independent soil characteristics, i.e. clay content in 0.50-2.00 m depth, among land uses within each landscape. Since there were no significant differences in clay contents between the reference and converted land uses at these depths (Appendix Table A1; Allen et al., 2015), we deduced that the sites within each landscape had previously similar soil characteristics and that differences in trace gas fluxes can be attributed to the changes in land uses and its associated management practices.

Since all the plantations were managed by smallholders, management practices of rubber and oil palm were diverse. The following information on management practices were based from our interviews of the smallholders. In 2013, oil palm and rubber plantations were weeded once or twice a year either manually or using herbicide (most commonly 2-5 L Gramaxone® or Roundup® ha<sup>-1</sup> year<sup>-1</sup>). Oil palm plantations were fertilized whereas rubber plantations were not. Oil palm plantations in the clay Acrisol landscape were fertilized only once during the rainy season, whereas those in the loam Acrisol landscape had a second fertilizer application in the dry season. The most commonly used fertilizers were NPK complete fertilizer (i.e. Phonska, Mahkota), potassium chloride (KCl) and urea. Plantations fertilized once received about 300 kg NPK-fertilizer ha<sup>-1</sup> year<sup>-1</sup> and plantations fertilized twice received

about 550 kg NPK-fertilizer ha<sup>-1</sup> year<sup>-1</sup>. In terms of added nutrient element, these rates were equivalent to 48-88 kg N ha<sup>-1</sup> year<sup>-1</sup>, 21-38 kg P ha<sup>-1</sup> year<sup>-1</sup> and 40-73 kg K ha<sup>-1</sup> year<sup>-1</sup>. Additionally, three of the smallholders applied 157 kg K-KCl ha<sup>-1</sup> year<sup>-1</sup> and 143 kg KCl-K ha<sup>-1</sup> year<sup>-1</sup> and two of the smallholders applied 138 kg urea-N ha<sup>-1</sup> year<sup>-1</sup>. One of the smallholders also applied lime in 2013 at about 200 kg dolomite ha<sup>-1</sup> year<sup>-1</sup>. Smallholders typically applied the fertilizer around the oil palm tree at about 0.8 – 1 m distance from the tree base. Additionally, senescing fronds were regularly cut and piled on the inter-rows of an oil palm plantation, typically in the middle of the 9 m distance between rows of oil palms. Oil palm fruits were harvested every two weeks, whereas the latex of the rubber and jungle rubber was collected weekly.

In addition to the monthly flux measurements conducted at the smallholder farms described above, we simulated fertilizations and conducted more frequent measurements (6 to 11 times) during 3 to 8 weeks following fertilization as the monthly sampling may have missed the short-term effect of fertilization on soil CO<sub>2</sub> and CH<sub>4</sub> fluxes. We chose 3 plots of oil palm plantations in each of the 2 landscapes, and in each plot we selected 3 trees separated by an inter-row distance of 9 m (in total, 18 oil palm trees). At 0.8-m distance from the base of each tree, we applied manually the fertilizer within a width of 0.2 m around the tree using the same rate that smallholders applied to these oil palm plantations (i.e. equivalent to 2 kg fertilizer per tree, based from 300 kg NPK-fertilizer ha<sup>-1</sup> divided by 134-140 trees ha<sup>-1</sup>; Appendix Table A2). We used the same fertilizer forms that smallholders applied, i.e. NPK complete fertilizer in the clay Acrisol landscape and a combination of KCl, ammonium sulfate and NPK complete fertilizer in the loam Acrisol landscape. One chamber base was placed at 0.3 m distance from the tree base (chamber location a); another chamber base was placed at 0.8 m distance from the tree wherein the fertilizer was applied (chamber location b); and a third chamber was placed at 4 - 4.5 m distance from the tree that served as a reference chamber without direct fertilizer application (chamber location c). In the clay Acrisol landscape, measurements in the 3 oil palm plots were done from mid-October to mid-December 2013, mid-February to mid-March 2014, and mid-February to mid-April 2014. In the loam Acrisol landscape, measurements were done from the end of October 2013 to mid-December 2014, mid-January to mid-March 2014, and mid-March to the start of April 2014. Shorter intervals of sampling days (Appendix Fig. B1) were conducted right after the fertilizer application.

## 2.2 CO<sub>2</sub> and CH<sub>4</sub> flux measurement

Soil CO<sub>2</sub> and CH<sub>4</sub> fluxes were measured monthly from December 2012 to December 2013, using static vented chambers. We are aware that the use of static vented chambers for CO<sub>2</sub> and CH<sub>4</sub> flux measurements could have possible sources of error due to changes in diffusion gradients during chamber closure. We have however adapted the design of our chambers and the flux calculation to minimize, if not avoid, these possible errors (see below). A clear advantage of the static vented chamber method compared to other methods is that it remains the only operational method that can be used to measure trace gas fluxes at a large number of plots (in our case 32) spread over a large area with regular measurements within a year. Furthermore, it also yields information on short-distance spatial variability of trace gas fluxes, which cannot be quantified by more integrative tower-based methods. Our chamber bases were made of polyvinyl chloride (0.05 m<sup>2</sup> area) and inserted ~0.03 m into the soil. In each of the four subplots per replicate plot, we randomly placed a permanent chamber base one month before the first measurement started. Since the area occupied by piled fronds or applied with fertilizer in oil palm plantations was relatively small, none of these randomly placed chamber bases (range of distance to the tree base was 1.8 – 5 m) happened to be located on such area. During sampling, the chamber bases were covered with polyethylene hoods (0.27 m total chamber height, and 12 L total volume) equipped with a Luer-lock sampling port and a vent for pressure equilibrium. Four gas samples (30 ml each) were removed at 1, 11, 21 and 31 minutes after chamber closure using a plastic syringe connected to the Luer-lock port. Immediately after sampling, gas samples were stored with overpressure into pre-evacuated 12 mL Labco Exetainers<sup>®</sup> (Labco Limited, Lampeter, UK) with rubber septa that were only used once. Our group has tested these exetainers for extended period of sample storage (e.g. up to 6 months) and airfreight transport by storing and transporting standard gases of known concentrations in overpressure, and these exetainers were proven to be leak proof. Gas samples in exetainers were transported to Germany by airfreight every 3-4 months and were analyzed upon arrival using a gas chromatograph (GC 6000 Vega Series 2, Carlo Erba Instruments, Milan, Italy with an ASPEC autosampler, Gilson SAS, Villiers, Le Bel, France), equipped with a flame ionization detector and an electron capture detector. Three standard gases were used for calibration with concentrations from 350 to 5000 ppm for CO<sub>2</sub> and 1000 to 5000 ppb for CH<sub>4</sub> (Deuste Steininger GmbH, Mühlhausen, Germany). The software Probe66 Version 1.3 (Messwert GmbH, Göttingen, Germany) was used to determine CH<sub>4</sub> and CO<sub>2</sub> sample concentrations by comparing integrated peak areas with those of the standard

gases. Fluxes were calculated from the concentration change over time of chamber closure, and adjusted with actual air temperature and pressure measured at the time of sampling. Linearity of increase of CO<sub>2</sub> concentrations with time of chamber closure ( $R^2 \geq 0.98$ ) was checked for each chamber measurement and in a few cases where concentration curved at the last sampling time we excluded the last data point and calculated the fluxes based on the linear increase in concentrations during the first 3 samplings. The majority of the measurements showed linear change in CH<sub>4</sub> concentrations with time of chamber closure. There were a few measurements when changes in CH<sub>4</sub> concentrations with time of chamber closure were small, mostly when net CH<sub>4</sub> uptake was low; in such cases, the calculated CH<sub>4</sub> flux using linear regression was not significantly different from zero. These fluxes were however retained in the statistical analyses to avoid bias by excluding low CH<sub>4</sub> fluxes or by assuming that these fluxes were zero. Assuming constant flux rates per day, annual soil CO<sub>2</sub> and CH<sub>4</sub> fluxes from the monthly sampling and total fluxes during a fertilization event (see below) were calculated by adopting the trapezoidal rule on time intervals between measured flux rates, similar to the method we employed in our earlier studies (e.g. Koehler et al., 2009; Veldkamp et al., 2013)

### 2.3 Auxiliary measurements

Soil temperature, moisture and mineral N content were measured with each trace gas measurement. Soil temperature was determined in the top 0.05 m depth using a GMH 1170 digital thermometer (Greisinger electronic GmbH, Regenstauf, Germany). Soil samples for mineral N content were also taken from the top 0.05 m at approximately 1 m distance from each chamber with 4 soil samples per plot. These 4 soil samples were pooled to represent each replicate plot on each day of measurement, and roots, leaves and twigs were manually removed. For the fertilization experiment, soil samples taken near each chamber location (a, b and c) were processed separately. Since even short storage of disturbed soil samples can affect mineral N concentrations (Arnold et al., 2008), extraction was done in the field immediately following soil sampling. A soil sample was added to a prepared 250 mL plastic bottle containing 150 mL of 0.5 mol L<sup>-1</sup> K<sub>2</sub>SO<sub>4</sub> (approximately 1:3 ratio of fresh soil to extractant volume) and transported to the field station. At the field station, samples were shaken for 1 h, filtered through pre-washed (with 0.5 mol L<sup>-1</sup> K<sub>2</sub>SO<sub>4</sub>) filter papers (Whatman, GE Healthcare Life Sciences, 4 µm nominal pore size) and the filtrate were immediately stored in a freezer. The remaining field-moist soil samples were stored in plastic bags and gravimetric moisture

content was determined at the field station, for which 50 - 100 g of fresh soil was dried at 105° C for at least 1 day. The gravimetric moisture content was used to calculate the dry mass of the soil extracted for mineral N. Frozen extracts were transported by airfreight to Germany to ensure that they stayed frozen throughout the transport until analysis. At our laboratory in the University of Göttingen, Germany,  $\text{NH}_4^+$  and  $\text{NO}_3^-$  concentrations in the extracts were analyzed using continuous flow injection colorimetry (SEAL Analytical AA3, SEAL Analytical GmbH, Norderstedt, Germany).  $\text{NH}_4^+$  was determined by salicylate and dicloro-isocyanuric acid reaction (Autoanalyzer Method G-102-93), and  $\text{NO}_3^-$  by cadmium reduction method with  $\text{NH}_4\text{Cl}$  buffer (Autoanalyzer Method G-254-02). Soil water content was expressed as water-filled pore space (WFPS), calculated using a particle density of 2.65 g  $\text{cm}^{-3}$  for mineral soil and the measured bulk densities in our study sites (Appendix Table A1).

## 2.4 Statistical Analysis

All statistical analyses of the monthly measurements of soil  $\text{CO}_2$  and  $\text{CH}_4$  fluxes were conducted using the means of the four chambers (or subplots) that represent each replicate plot on a given sampling day. Data were checked for normal distribution (using Shapiro-Wilk's tests), and if necessary a logarithmic (for  $\text{CO}_2$ ,  $\text{CH}_4$ , and mineral N) or square root (for WFPS) transformation was used. For our first objective, we conducted comparisons of the reference land uses between the two landscapes in order to test the first hypothesis. Then we carried out comparisons among land-use types within each landscape to test our second hypotheses. Linear mixed effect models (LME) were applied (Crawley, 2009) with either landscape (i.e. comparing landscapes for each reference land use) or land use (i.e. comparing land-use types within each landscape) as the fixed effect and replicate plots and sampling days as the random effects. For the fertilization experiment, we tested differences in soil  $\text{CO}_2$  and  $\text{CH}_4$  fluxes between chamber locations within each oil palm plantation plot, using LME with chamber location as the fixed effect and palm trees and sampling days as the random effects. We extended the LME model to include either 1) a variance function that allows different variances of the fixed effect, and/or 2) a first-order temporal autoregressive process, which assumes that correlation between measurements decreases with increasing time difference, if this improved the relative goodness of the model fit based on the Akaike information criterion. Fixed effect was considered significant based on analysis of variance at  $P \leq 0.05$ , and differences between landscapes or land-use types (or chamber locations for the

fertilization experiment) were assessed using Fisher's least significant difference test  $P \leq 0.05$ . For our second objective, we assess how soil factors influence the seasonal variations of soil CO<sub>2</sub> and CH<sub>4</sub> fluxes, using Pearson's correlation tests with soil temperature, WFPS, NO<sub>3</sub><sup>-</sup>, NH<sub>4</sub><sup>+</sup> and total mineral N. This assessment of seasonal controls of trace gas fluxes was conducted for each land use within each landscape using the means of the four replicate plots (as spatial representation) on each of the 12 monthly measurements in order to focus the analysis on temporal variation. Lastly, we assessed the influence of soil physical and biochemical characteristics (Appendix Table A1) on the spatial variations of soil annual CO<sub>2</sub> and CH<sub>4</sub> fluxes first on the reference land uses across landscapes (16 plots) and second across land-use types within each landscape (16 plots), using Spearman's rank correlation test. The first was to assess the spatial controls of trace gas fluxes from the reference land uses, and the second was to evaluate which soil factors drive the spatial variation of trace gas fluxes across land uses. We used the annual fluxes as temporal representation in order to focus this analysis on the spatial variation. For all correlation tests, statistical significance was taken at  $P \leq 0.05$ , except in a few cases for which marginal significance at  $P \leq 0.09$  was considered because our experimental design encompassed the inherent spatial variability in the studied landscapes. All statistical analyses were conducted using R 3.0.2 (R Development Core Team, 2013).

### 3 Results

#### 3.1 Reference land uses (forest and jungle rubber): comparison between landscapes

WFPS in the jungle rubber was higher in the clay than loam Acrisol soils ( $P < 0.01$ ), but in the forest WFPS did not differ between landscapes ( $P = 0.56$ ; Fig. 1a, b). In both landscapes, we detected a distinctly lower WFPS during the drier period (mean WFPS ranged 38-80 % between mid-June and October) compared to the wetter period (mean WFPS ranged 50-96 %;  $P < 0.01$ ; Fig. 1a, b). Soil temperatures in the forest were lower in the clay than loam Acrisol soils ( $P = 0.02$ ; Fig. 1c, d), which was probably due to the difference in the time of the day when measurements were conducted. Soil temperatures in the jungle rubber did not differ between landscapes ( $P = 0.17$ ).

In both landscapes, NH<sub>4</sub><sup>+</sup> was the dominant form of mineral N (Table 1). Soil NH<sub>4</sub><sup>+</sup> contents in the jungle rubber were higher in the clay than loam Acrisol soils ( $P = 0.02$ ), but in the

forest soil  $\text{NH}_4^+$  contents did not differ between landscapes ( $P = 0.90$ ; Table 1). Soil  $\text{NO}_3^-$  contents in the forest were higher in the clay than loam Acrisol soils ( $P < 0.01$ ), whereas soil  $\text{NO}_3^-$  contents in the jungle rubber was higher in the loam than clay Acrisol soils ( $P = 0.02$ ; Table 1). Total mineral N contents in both reference land uses did not differ between landscapes ( $P = 0.11 - 0.19$ ; Table 1).

We detected no differences in soil  $\text{CO}_2$  fluxes between landscapes for the reference land-use types ( $P = 0.63 - 0.69$ ; Table 2; Fig. 2a, b). Similarly, soil  $\text{CH}_4$  fluxes from both reference land uses were also comparable between the two landscapes ( $P = 0.26 - 0.27$ ; Table 2; Fig. 2c, d). However, in the loam Acrisol soil, two of the four forest sites displayed net  $\text{CH}_4$  emissions in 26 % of the measurements, which resulted in its largest variation among plots (as indicated by its largest standard error around the mean; Table 2).

### **3.2 Converted land uses (rubber and oil palm plantations): comparisons to the reference land uses within each landscape**

While in both landscapes WFPS did not differ among land-use types ( $P = 0.12 - 0.26$ ; Fig 1a, b), soil temperatures were slightly higher in the plantations ( $27.2 \pm 0.1$  °C) compared to the reference land uses ( $25.9 \pm 0.1$  °C) in each landscape (both  $P < 0.01$ ; Fig. 1c, d). Soil  $\text{NH}_4^+$  contents in rubber were lower than in all other land uses in the clay Acrisol soil ( $P = 0.05$ ), and soil  $\text{NH}_4^+$  contents in both rubber and oil palm were also lower than in the reference land uses in the loam Acrisol soil ( $P = 0.03$ ; Table 1). In the clay Acrisol soil,  $\text{NO}_3^-$  contents in rubber and oil palm were lower than in forest ( $P < 0.01$ ), and in the loam Acrisol soil  $\text{NO}_3^-$  contents in rubber were lowest whereas these were intermediary in oil palm ( $P < 0.01$ ; Table 1). The latter was related to a fertilizer application by the owner of one of the oil palm plots just a month prior to our measurement. Total mineral N showed similar differences among land-use types in each landscape as those with soil  $\text{NH}_4^+$  and  $\text{NO}_3^-$  contents (both  $P < 0.01$ ; Table 1).

Soil  $\text{CO}_2$  fluxes from oil palm were 45% lower in the clay Acrisol soil and 38% lower in the loam Acrisol soil compared to the forest (both  $P < 0.01$ ; Table 2; Fig. 2a, b). From the fertilization experiment, soil  $\text{CO}_2$  fluxes from within 1 m distance to the oil palm base (chamber locations a and b) were on average  $2.3 \pm 0.2$  times higher than those at 4 - 4.5 m from the tree base (chamber location c) in both landscapes (all  $P < 0.01$ ; Appendix Table A3).

1 However, this area within 1 m distance to the tree base is only 3 m<sup>2</sup> per tree or 4 % on a  
2 hectare basis, and so even if we would weight with area coverage the annual soil CO<sub>2</sub> fluxes  
3 (Table 2), which were measured from chambers placed randomly between 1.8 - 5 m from the  
4 oil palm base, such high fluxes within 1 m distance to the tree base would still account less  
5 than the standard errors (7 - 9 %) of the mean annual fluxes.

6 Soil CH<sub>4</sub> uptake in the plantations were 84% (oil palm) and 93% (rubber) lower compared to  
7 the forest and 69% (oil palm) and 86% (rubber) lower compared to the jungle rubber in the  
8 clay Acrisol soil ( $P < 0.01$ ; Table 2; Fig. 2c, d). Also in the loam Acrisol soil, CH<sub>4</sub> uptake in  
9 the plantations was lower (reduction of 64% for rubber and 44% for oil palm) compared to  
10 jungle rubber ( $P = 0.02$ ; Table 2; Fig. 2c, d). However, CH<sub>4</sub> uptake in the forest on loam  
11 Acrisol soils deviated from the differences detected in the clay Acrisol soils because of the  
12 two forest sites that displayed net CH<sub>4</sub> emissions (see above; Table 2; Fig. 2d). From the  
13 fertilization experiment, soil CH<sub>4</sub> uptake from the area of fertilizer application (chamber  
14 location b) were  $2.6 \pm 0.2$  times lower than the unfertilized chamber locations a and c  
15 (respectively at 0.3 m and 4 – 4.5 m distance from the oil palm base) ( $P \leq 0.01 - 0.05$ ;  
16 Appendix Table A3), with the exception of plot 3 in the clay Acrisol soil ( $P = 0.45$ ; Appendix  
17 Table A3). In most cases, CH<sub>4</sub> uptake in chamber location b was reduced immediately  
18 following fertilizer application and was restored to pre-fertilization values after about 6 weeks  
19 (Appendix Fig. B1). Thus, even if we would weight with the area coverage (~1.3% of the area  
20 in a hectare) and time duration (12% of the time in a year) the annual soil CH<sub>4</sub> fluxes, the  
21 effect of this fertilized location would be negligible.

### 23 3.3 Seasonal controls of CO<sub>2</sub> and CH<sub>4</sub> fluxes from each land-use type

24 In the clay Acrisol soil, CH<sub>4</sub> fluxes were positively correlated with WFPS (Table 3) in each of  
25 the four land-use types, signifying the higher CH<sub>4</sub> uptake in the dry than wet season ( $P \leq 0.01$   
26 - 0.03; Fig. 2c). Soil CH<sub>4</sub> fluxes correlated negatively with NO<sub>3</sub><sup>-</sup> contents in forest and with  
27 soil CO<sub>2</sub> fluxes in rubber (Table 3). Across all land-use types, soil CH<sub>4</sub> uptake was negatively  
28 correlated with total mineral N content ( $R = -0.47$ ,  $P \leq 0.01$ ,  $n = 41$ ; Fig. 3a) and NO<sub>3</sub><sup>-</sup> content  
29 ( $R = -0.73$ ,  $P \leq 0.01$ ,  $n = 41$ ). Some correlations in Table 3 were possibly spurious: in oil  
30 palm, soil CO<sub>2</sub> fluxes were positively correlated with soil temperatures (Table 3) even if the  
31 temperate range was small (25.5 - 28.8 °C) such that this correlation was likely caused by the



differences in sampling time during the day rather by seasonal temperature pattern. In jungle rubber, plotting soil CO<sub>2</sub> fluxes against soil NO<sub>3</sub><sup>-</sup> contents showed that their correlation (Table 3) was apparently caused by a group of many low NO<sub>3</sub><sup>-</sup> contents against one high NO<sub>3</sub><sup>-</sup> value and this correlation became insignificant when the one high value was removed. Also, in rubber, the marginal negative correlation between soil CH<sub>4</sub> fluxes with NO<sub>3</sub><sup>-</sup> contents was because of the correlation between WFPS and NO<sub>3</sub><sup>-</sup> contents.

In the loam Acrisol soil, seasonal variations of soil CO<sub>2</sub> fluxes were positively correlated with WFPS in jungle rubber and negatively correlated with WFPS in rubber (Table 3). Some correlations in Table 3 were also spurious caused by differences in sampling time during the day rather than by seasonal pattern: correlation between soil CO<sub>2</sub> fluxes and soil temperature in forest and oil palm despite narrow temperate ranges (24.8 - 27.2 °C in forest and 25.8 - 29.4 °C in oil palm). The negative correlation between soil CO<sub>2</sub> fluxes and NO<sub>3</sub><sup>-</sup> contents in jungle rubber was driven by the negative correlation between WFPS and NO<sub>3</sub><sup>-</sup> (Table 3). As was observed in the clay Acrisol soil, seasonal variation in soil CH<sub>4</sub> fluxes from the loam Acrisol soil was positively correlated with WFPS in each of the three land-use types (Table 3) with the exception of the forest which was caused by one extreme CH<sub>4</sub> emission from a single chamber (656.47 µg C m<sup>-2</sup> h<sup>-1</sup>). When this one value was excluded, a positive correlation between soil CH<sub>4</sub> fluxes and WFPS was also detected for forest ( $R = 0.60$ ,  $P \leq 0.01$ ,  $n = 12$ ). Soil CH<sub>4</sub> fluxes correlated also positively with soil CO<sub>2</sub> fluxes in jungle rubber whereas this correlation was negative in rubber (Table 3). In the jungle rubber, soil CH<sub>4</sub> fluxes correlated negatively with soil NO<sub>3</sub><sup>-</sup> contents (Table 3). Across land-use types, we also observed negative correlations of soil CH<sub>4</sub> uptake with total mineral N content ( $R = -0.52$ ,  $P \leq 0.01$ ,  $n = 38$ ; Fig. 3b) and NO<sub>3</sub><sup>-</sup> content ( $R = -0.75$ ,  $P \leq 0.01$ ,  $n = 38$ ).

### **3.4 Spatial controls of annual CO<sub>2</sub> and CH<sub>4</sub> fluxes across land-use types within each landscape**

For these correlation analyses, we used all soil physical and biochemical characteristics, which are reported in Appendix Table A1. Apart from the correlations reported here, there were no other significant correlations with any of the tested soil physical and biochemical characteristics. First, analyzing both reference land uses (forest and jungle rubber) across landscapes, the only significant correlation between annual soil CO<sub>2</sub> emissions and soil

parameters was with sand content (*Spearman's*  $\rho = -0.51$ ,  $P = 0.08$ ,  $n = 16$ ). However, analyzing for each landscape separately, annual soil CO<sub>2</sub> fluxes from the reference land uses in the loam Acrisol soil correlated with Bray-extractable soil P ( $\rho = -0.74$ ,  $P = 0.04$ ,  $n = 8$ ). Furthermore, annual soil CH<sub>4</sub> fluxes from both reference land uses across landscapes were correlated with net N mineralization rates ( $\rho = -0.75$ ,  $P < 0.01$ ,  $n = 16$ ) and, for each landscape separately, with exchangeable Al ( $\rho = 0.74$ ,  $P = 0.04$ ,  $n = 8$  in the clay Acrisol soil, and  $\rho = 0.69$ ,  $P = 0.06$ ,  $n = 8$  in the loam Acrisol soil).

Second, analyzing across four land-use types within each landscape, annual soil CO<sub>2</sub> fluxes correlated only with soil <sup>15</sup>N natural abundance signatures in the clay Acrisol soil ( $\rho = -0.49$ ,  $P = 0.05$ ,  $n = 16$ ). In the loam Acrisol soil, annual soil CO<sub>2</sub> fluxes correlated with soil organic C ( $\rho = 0.49$ ,  $P = 0.06$ ,  $n = 16$ ), base saturation ( $\rho = -0.53$ ,  $P = 0.04$ ,  $n = 16$ ), Bray-extractable P ( $\rho = -0.71$ ,  $P < 0.01$ ,  $n = 16$ ) and soil <sup>15</sup>N natural abundance signatures ( $\rho = -0.60$ ,  $P = 0.02$ ,  $n = 16$ ). Annual soil CH<sub>4</sub> fluxes across all land uses in the clay Acrisol soil correlated with net N mineralization rates ( $\rho = -0.52$ ,  $P = 0.04$ ,  $n = 16$ ), whereas in the loam Acrisol soil this correlation only showed up after exclusion of one plot in rubber that had an unusually high net N mineralization ( $\rho = -0.51$ ,  $P = 0.07$ ,  $n = 15$ ). Net N mineralization significantly decreased in rubber that had no fertilization and intermediate in oil palm that had fertilization, particularly in the clay Acrisol soil (Appendix Table A1).

## 4 Discussion

### 4.1 CO<sub>2</sub> and CH<sub>4</sub> fluxes from the reference land uses

Mean soil CO<sub>2</sub> fluxes from our forest sites (Table 2) were within the range of reported fluxes (123 - 228 mg C m<sup>-2</sup> h<sup>-1</sup>) from tropical rainforests in Asia (Adachi et al., 2005; Ohashi et al., 2008) and Latin America (Davidson et al., 2000; Schwendenmann et al., 2003; Keller et al., 2005; Sotta et al., 2006; Koehler et al., 2009). Compared to measurements conducted in Indonesia, our lowland forests had higher soil CO<sub>2</sub> fluxes than a montane forest in Sulawesi at 1000 m elevation with similar spatially replicated and temporally intensive measurements (127 mg C m<sup>-2</sup> h<sup>-1</sup>; van Straaten et al., 2011) and higher than the seven partially logged forest sites in Jambi with only one-time measurement (162 mg C m<sup>-2</sup> h<sup>-1</sup>; Ishizuka et al., 2005). While the difference with this last study may be caused by their one-time sampling, the only

other study that measured CO<sub>2</sub> fluxes from the same region (that conducted nine measurements spread over one year at three plots) reported values that were as low as 33% to 50% of our measured soil CO<sub>2</sub> fluxes (63 – 94 mg C m<sup>-2</sup> h<sup>-1</sup>; Ishizuka et al., 2002). Such values are hard to reconcile with our and other measurements in tropical lowland forests, including the measurements by Ishizuka et al. (2005).

Seasonal variation of soil CO<sub>2</sub> fluxes from the reference land uses was driven by changes in soil water content, as suggested by the positive correlation with WFPS in jungle rubber on the loam Acrisol soil (Table 3). Other studies conducted in tropical rainforests have shown that seasonal changes in soil CO<sub>2</sub> fluxes are often caused by changes in soil water content (e.g. Sotta et al., 2007; Koehler et al., 2009; van Straaten et al., 2011), and sometimes in combination with reduction in solar irradiation caused by clouds during the wet season (Schwendenmann et al., 2003). In tropical forest soils, the relationship of soil CO<sub>2</sub> flux with soil water content is curvilinear with the highest fluxes typically at field capacity (pF ~2 or WFPS between 50 – 55 %; Sotta et al., 2007; Koehler et al., 2009; van Straaten et al., 2011), which explains why WFPS did not show correlation in forests in both landscapes where WFPS (mostly ≥60 – 80%; Fig. 1a, b) fluctuated at the top curve of this curvilinear relationship.

In contrast to our first hypothesis, soil texture was not the proximal factor controlling annual soil CO<sub>2</sub> fluxes, but instead sand content indirectly affected soil fertility (e.g. Bray-extractable P) which, in turn, influenced soil CO<sub>2</sub> fluxes. In the reference land uses, the negative correlation of annual soil CO<sub>2</sub> fluxes with the sand contents contrasted with results in the Amazon Basin where sandy Ferralsol soil had higher soil CO<sub>2</sub> fluxes than the clay Ferralsol soil (Sotta et al., 2006). In the study by Sotta et al. (2006), annual CO<sub>2</sub> emissions were negatively correlated with total soil P content. In our loam Acrisol soil, which had lower soil fertility (i.e. lower Bray-extractable P and base saturation and higher Al saturation) than the clay Acrisol soil (Appendix Table A1; Allen et al., 2015), there may be strong competition for P such that trees have to allocate more C to their root or root-mycorrhizal system to obtain this nutrient. From the same study sites, there was also lower P concentration in fine roots in the top 0.2-m soil depth of the reference land uses in the loam than clay Acrisol soils (Sahner et al., 2015). This strategy of high below-ground C investment was reflected in the negative correlation of annual soil CO<sub>2</sub> fluxes from the reference land uses with Bray-extractable P contents in the loam Acrisol soil.

Mean soil CH<sub>4</sub> fluxes from our forest sites (Table 2) fall within the range for tropical lowland forests reported by other studies (-6.3 to -55.9 µg CH<sub>4</sub>-C m<sup>-2</sup> h<sup>-1</sup>; summarized by Veldkamp et al., 2013); however, our measured CH<sub>4</sub> uptake rates were at the upper end (towards more negative values) of these reported rates and were also higher than the CH<sub>4</sub> uptake rates reported for old-growth forests in **Jambi, Indonesia** (-21.3 to +4.2 µg CH<sub>4</sub>-C m<sup>-2</sup> h<sup>-1</sup>; Ishizuka et al., 2002).

Seasonal variation of soil CH<sub>4</sub> fluxes was strongly controlled by soil water content with higher uptake in the dry season (Fig. 1a, b), as shown by the strong positive correlations with WFPS in all land uses in both landscapes (Table 3). Such seasonal changes reflect diffusional limitation on the supply of CH<sub>4</sub> to methanotrophs at high WFPS (Keller and Reiners, 1994) and the possible occurrence of anaerobic decomposition, producing CH<sub>4</sub>, which may partially offsets CH<sub>4</sub> consumption (Keller and Reiners, 1994; Verchot et al., 2000). Since we measured occasional net CH<sub>4</sub> emissions from some reference land uses (Fig. 2d), we cannot exclude this anaerobic CH<sub>4</sub> production. High microbial and root activity consume oxygen in the soil, which may contribute to the creation of anaerobic microsites where CH<sub>4</sub> can be produced. This may have occurred in the jungle rubber on the loam Acrisol soil, where we detected a positive correlation of soil CO<sub>2</sub> fluxes with soil CH<sub>4</sub> fluxes (Table 3). Positive correlations of soil CO<sub>2</sub> fluxes and CH<sub>4</sub> fluxes have been reported also for tropical forests (Verchot et al., 2000). In addition to WFPS, soil mineral N dynamics also influenced the seasonal variation of soil CH<sub>4</sub> fluxes. The negative correlations of soil CH<sub>4</sub> fluxes with soil NO<sub>3</sub><sup>-</sup> contents in the forest on the clay Acrisol soil and in the jungle rubber on the loam Acrisol soil (Table 3) imply that some of the observed seasonal variability may have been caused by temporal N limitation of CH<sub>4</sub> oxidation (Bodelier and Laanbroek, 2004; Veldkamp et al. 2013).

**We found strong indications that CH<sub>4</sub> uptake in this converted tropical lowland was both N-limited and affected by high, potentially toxic, exchangeable Al concentrations in the soil. We interpreted** negative correlations of annual soil CH<sub>4</sub> fluxes from the reference land uses with net N mineralization rates (see 3.4) across landscapes **as evidence for N-limited CH<sub>4</sub> uptake**. Indications of N-limited CH<sub>4</sub> uptake have been reported for tropical forests in Panama (Veldkamp et al., 2013) and Ecuador (Wolf et al., 2012), but this is the first time that it was observed on a landscape scale in the tropics. Furthermore, the positive correlations of annual soil CH<sub>4</sub> fluxes from the reference land uses with exchangeable Al within each landscape signified the lower CH<sub>4</sub> uptake measured in sites with more exchangeable Al in the soil. Soil

Al saturation in our reference land uses was high (mean values ranged from 61% to 80%; Appendix Table A1). High  $\text{Al}^{3+}$  concentrations in the soil solution and higher exchangeable Al in the soil are known to be toxic for plants which root growth may be inhibited (Ma et al., 2001). Dissolved  $\text{Al}^{3+}$  can also be toxic for soil microorganisms and it has been shown that high dissolved Al concentrations in the soil inhibited  $\text{CH}_4$  uptake in a temperate forest soil in Japan (Tamai et al., 2003). We are not aware of any study reporting such a relationship for tropical ecosystems, which is not surprising since in most trace gas studies exchangeable Al in the soil is either not measured or does not reach such high levels as in our sites.

In summary, seasonal variation of soil  $\text{CO}_2$  fluxes from the reference land uses were related to soil water content (i.e. jungle rubber in loam Acrisol soil), while spatial control of annual soil  $\text{CO}_2$  fluxes across landscapes were related to soil fertility: low Bray-extractable P concentrations coincided with high annual soil  $\text{CO}_2$  fluxes from the loam Acrisol soil, which had lower soil fertility than the clay Acrisol soil. Seasonal variation of  $\text{CH}_4$  fluxes from the reference land uses were mainly explained by soil water content, although we found indications that also temporal N limitation may have played a role. Spatial controls of annual soil  $\text{CH}_4$  fluxes across landscapes were also related to soil fertility, as shown by their negative correlation with soil N availability, suggesting N limitation on  $\text{CH}_4$  uptake, and positive correlation with exchangeable Al, suggesting Al toxicity on methanotrophs, which has not yet been reported for tropical ecosystems. These results are in contrast to our first hypothesis - soil texture was not the proximal factor controlling soil  $\text{CO}_2$  and  $\text{CH}_4$  fluxes but only indirectly through its influence on soil fertility that, in turn, controlled the spatial variations of these trace gases across our studied landscapes.

## **4.2 Effects of land-use change on $\text{CO}_2$ and $\text{CH}_4$ fluxes**

Mean soil  $\text{CO}_2$  fluxes from our rubber plantations (Table 2) were in the same order of magnitude as those reported for seven rubber plantations in Jambi (Indonesia) measured once (171  $\text{mg C m}^{-2} \text{ h}^{-1}$ ; Ishizuka et al., 2005), while soil  $\text{CO}_2$  fluxes from a rubber plantation in a sandy clay loam Nitisol soil in Malaysia with one measurement were lower (123  $\text{mg C m}^{-2} \text{ h}^{-1}$ ; Adachi et al., 2005). Some other studies reported soil  $\text{CO}_2$  fluxes that are much lower than our measured fluxes: a rubber plantation on a heavily weathered silty clay soil in China (35  $\text{mg C m}^{-2} \text{ h}^{-1}$ ; Werner et al., 2006) and a rubber plantation in Jambi (Indonesia) with nine

1 measurements ( $75 \text{ mg C m}^{-2} \text{ h}^{-1}$ ; Ishizuka et al., 2002). Since this last study also reported 33 -  
2 50% lower soil  $\text{CO}_2$  fluxes from forests (see 4.1) as well as 50% lower soil  $\text{CO}_2$  fluxes from  
3 oil palm ( $51 \text{ mg C m}^{-2} \text{ h}^{-1}$ ; Ishizuka et al., 2002) than our measured fluxes from the same  
4 region (Table 2), we suspect some methodological issues in this study. Mean soil  $\text{CO}_2$  fluxes  
5 from our oil palm sites were comparable with other reported fluxes from five oil palm  
6 plantations in Jambi (Indonesia) that were measured once ( $98 \text{ mg C m}^{-2} \text{ h}^{-1}$ ; Ishizuka et al.,  
7 2005). Lastly, soil  $\text{CO}_2$  fluxes from an oil palm plantation that were more than double of our  
8 measured fluxes were reported from a one-time measurement in a sandy clay loam Nitisol soil  
9 in Malaysia ( $222 \text{ mg C m}^{-2} \text{ h}^{-1}$ ; Adachi et al., 2005).

10 Seasonal variation of soil  $\text{CO}_2$  fluxes from oil palm was not as pronounced as that from  
11 rubber (Fig. 2a, b). In rubber plantations in loam Acrisol soil, where WFPS were all above 55  
12 % (Fig. 1b), the seasonal variation of soil  $\text{CO}_2$  fluxes reflected the curvilinear relationship of  
13 soil  $\text{CO}_2$  fluxes with WFPS, whereby soil  $\text{CO}_2$  fluxes typically decline at WFPS larger than  
14 about 55 % (e.g. Sotta et al., 2007; Koehler et al., 2009), and hence the negative correlation  
15 with WFPS (Table 3).

16 The spatial variation of annual soil  $\text{CO}_2$  fluxes across land uses reflected the changes in soil  
17 organic matter quality and quantity with changes in land use, as indicated by the negative  
18 correlations with soil  $^{15}\text{N}$  natural abundance signatures (see 3.4) and the positive correlation  
19 with soil organic C content. Soil  $^{15}\text{N}$  signatures in our studied oil palm plantations were  
20 significantly higher than the reference land uses (i.e. loam Acrisol soil; Appendix Table A1;  
21 Allen et al., 2015), which we interpreted as an indication of the degree of decomposition of  
22 soil organic matter. The more decomposed the soil organic matter, the higher is the soil  $^{15}\text{N}$   
23 signature, as illustrated by increasing soil  $^{15}\text{N}$  signatures with increasing depth of tropical  
24 forest soils (Sotta et al., 2008; Baldos et al., 2015). The more decomposed soil organic matter  
25 in the oil palm plantations was probably due to their lower inputs from litterfall and fine and  
26 coarse root production than those in the forests (measured from the same sites by Kotowska et  
27 al., 2015). The low litter input and root production influenced not only the degree of  
28 decomposition of the soil organic matter but also lead to strong reductions in soil organic C  
29 stocks of oil palm and rubber plantations (measured in the same study region by van Straaten  
30 et al. 2015). This may have contributed to the low  $\text{CO}_2$  emissions, since we also detected a  
31 positive correlation of soil  $\text{CO}_2$  emissions with soil organic C content. Similar findings were  
32 reported from forest conversion to tree plantations on Acrisol soil in subtropical southern

China where decreases in soil CO<sub>2</sub> fluxes were also explained by decreases in annual litterfall and root biomass (Sheng et al., 2010). In addition to changes in soil organic matter quality, spatial variation of annual soil CO<sub>2</sub> fluxes across land uses in the loam Acrisol soil was also controlled by changes in soil fertility with land-use change, as shown by their negative correlations with base saturation and Bray-extractable P (see 3.4). Conversion of forest or jungle rubber to rubber and oil palm plantations was accompanied by burning of slashed vegetation, whereby considerable amounts of bases and P could be released from the plant biomass to ashes (Klinge et al., 2004). Input of these nutrients to the soil from the ashes, combined with P fertilization and liming (particularly in the oil palm plantations), significantly increased soil pH in both rubber and oil palm as well as base saturation and Bray-extractable P in oil palm (Appendix Table A1; Allen et al., 2015). The negative correlations of annual soil CO<sub>2</sub> fluxes with base saturation and Bray-extractable P across land uses suggest that C allocation to its root-mycorrhizal system may have decreased with increased base cations and P availability, contributing to the observed decrease in soil CO<sub>2</sub> fluxes from oil palm compared to the other land uses (Table 2). In contrast, the speculation by Ishizuka et al. (2005) that low soil CO<sub>2</sub> fluxes from oil palm plantations could be explained by higher soil bulk densities related to intensive management practices we could not support since soil bulk densities in these converted land uses were comparable to the reference land uses (Appendix Table A1; Allen et al., 2015).

Mean soil CH<sub>4</sub> fluxes from rubber plantations (Table 2) were comparable with those reported for a rubber plantation in southwest China (-5.7 µg CH<sub>4</sub>-C m<sup>-2</sup> h<sup>-1</sup>; Werner et al., 2006) and for seven rubber plantations in Jambi (Indonesia) measured only once (-5.8 µg CH<sub>4</sub>-C m<sup>-2</sup> h<sup>-1</sup>; Ishizuka et al., 2005). From the oil palm plantations, mean soil CH<sub>4</sub> fluxes (Table 2) were comparable with those reported for five oil palm plantations in Jambi (Indonesia) measured only once (-20.1 µg CH<sub>4</sub>-C m<sup>-2</sup> h<sup>-1</sup>; Ishizuka et al., 2005) but larger (or more CH<sub>4</sub> uptake rate) than that reported for an oil palm plantation in Jambi with one measurement (-4.2 µg CH<sub>4</sub>-C m<sup>-2</sup> h<sup>-1</sup>; Ishizuka et al., 2002).

As was the case for the reference land uses, seasonal variation of soil CH<sub>4</sub> fluxes from the converted land uses were also controlled by WFPS (Table 3), and the possible mechanisms were the same (see 4.1). Moreover, strong negative correlations of soil CH<sub>4</sub> uptake with total mineral N (Fig. 3) and NO<sub>3</sub><sup>-</sup> contents across all land uses (see 3.3), of which total mineral N was lowest in the converted land uses (Table 1), also suggest temporal N limitation on



1 methanotrophic activity (Veldkamp et al. 2013) that may have contributed to the decrease in  
2 CH<sub>4</sub> uptake in the converted land uses (Fig. 2c, d; Table 2).

3 The negative correlations of annual soil CH<sub>4</sub> fluxes with net N mineralization rates across  
4 land uses further suggest N limitation of CH<sub>4</sub> uptake, as indicated by the lowest CH<sub>4</sub> uptake in  
5 the converted land uses (Table 2) that had the lowest (i.e. rubber with no N fertilization) to  
6 intermediate (i.e. oil palm with N fertilization) net N mineralization rates (see 3.4). The  
7 results from the fertilization experiment in the oil palm sites that showed inhibition of CH<sub>4</sub>  
8 uptake in the fertilized spot (chamber location b; Appendix Table A3) within 6 weeks  
9 following fertilizer application (Appendix Fig. B1) was probably caused by salt effect, as has  
10 been observed in a fertilization experiment in tropical pastures (Veldkamp et al., 2001).  
11 However, this CH<sub>4</sub> inhibition following fertilizer application did not influence our annual flux  
12 estimates because of the negligible area coverage of the fertilized spots (~1.3% of the area in  
13 a hectare) and its short-term effect (less than 6 weeks or 12% of the time in a year).

14 In summary, soil CO<sub>2</sub> fluxes decreased only in oil palm and not in rubber, which partly  
15 supports our second hypothesis. These converted land uses showed decrease in soil CH<sub>4</sub>  
16 uptake, which supports our second hypothesis. Seasonality of soil CO<sub>2</sub> and CH<sub>4</sub> fluxes in the  
17 converted land uses appeared to be controlled by the same factors as those in the reference  
18 land uses. The strong decrease in soil CO<sub>2</sub> fluxes from the oil palm was probably caused by a  
19 combination of strongly decomposed soil organic matter and low soil carbon stocks (caused  
20 by the low input of litterfall and low fine root production), and possibly low C allocation to  
21 root or root-mycorrhizal system (due to the improved base cations and P availability from  
22 liming and P fertilization). Reduction in annual CH<sub>4</sub> uptake in the converted land uses were  
23 primarily caused by the decrease in soil N availability in these converted land uses.

## 25 **5 Consequences of land-use change on soil trace gas fluxes**

26 Our study shows that land-use change had a profound effect on the soil-atmosphere fluxes of  
27 the trace gases CO<sub>2</sub> and CH<sub>4</sub>, with reduced soil CO<sub>2</sub> fluxes from oil palm plantations and  
28 reduced soil CH<sub>4</sub> uptake in both rubber and oil palm plantations. The reduced soil CO<sub>2</sub> fluxes  
29 in the oil palm should not be interpreted as reduced net ecosystem emissions because we did  
30 not measure the net CO<sub>2</sub> uptake by the vegetation and the changes in soil and vegetation  
31 carbon stocks. Rather the strong decrease in soil CO<sub>2</sub> fluxes from oil palm is a reflection of  
32 the present belowground carbon dynamics in this land use. Due to decreases in litterfall and



fine root production (Kotowska et al., 2015) as well as frond management practice (stacking them on inter-rows) that reduced fresh litter input on the whole area, **soil organic C stocks in these oil plantations decrease over time (van Straaten et al., 2015), reflecting the** reductions in soil CO<sub>2</sub> emissions.

Our estimate of decrease in CH<sub>4</sub> uptake from conversion of forest or jungle rubber to rubber and oil palm in these landscapes was about 2 kg CH<sub>4</sub>-C ha<sup>-1</sup> year<sup>-1</sup> (based on average of values in Table 2). If we multiply this with 0.52 Mha, the increase in areal coverage of oil palm and rubber plantations in Jambi from 1996 to 2011 (BPS, 2012), this suggests that the capacity of the province of Jambi to remove this potent greenhouse gas from the atmosphere has decreased by about 1040 Mg CH<sub>4</sub>-C year<sup>-1</sup> as a result of this land-use conversion. This calculation does not take into account land-use changes that occurred in the peatlands. Finally, we detected important soil fertility controls on trace gas exchange in this converted tropical landscape on highly weathered Acrisol soils, including the controls of base cation and P availability on annual soil CO<sub>2</sub> fluxes and N availability and Al toxicity on annual soil CH<sub>4</sub> fluxes. Such controls at the landscape scale have not yet been reported, and thus we stress the importance of conducting landscape-scale studies as field studies on a few small plots or laboratory-based studies may not be able to detect such important controls.

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Table 1. Mean ( $\pm$ SE,  $n = 4$ ) soil extractable mineral nitrogen in the top 0.05-m depth for different land-use types within each soil landscape in Jambi, Sumatra, Indonesia, measured monthly from December 2012 to December 2013. Means followed by different lowercase letter indicate significant differences among land-use types within a soil landscape and different capital letter indicate significant differences between soil landscapes within a land-use type (Linear mixed effects models with Fisher's LSD test at  $P \leq 0.05$ ).

Land-use type	NH <sub>4</sub> <sup>+</sup> (mg N kg <sup>-1</sup> )	NO <sub>3</sub> <sup>-</sup> (mg N kg <sup>-1</sup> )	mineral N (mg N kg <sup>-1</sup> )
clay Acrisol soil			
Forest	6.99 $\pm$ 1.03 <sup>a,A</sup>	2.15 $\pm$ 0.36 <sup>a,A</sup>	9.14 $\pm$ 1.34 <sup>a,A</sup>
Jungle Rubber	7.33 $\pm$ 0.21 <sup>a,A</sup>	0.23 $\pm$ 0.06 <sup>b,B</sup>	7.56 $\pm$ 0.26 <sup>b,A</sup>
Rubber	4.25 $\pm$ 0.23 <sup>b,A</sup>	0.05 $\pm$ 0.01 <sup>b,B</sup>	4.30 $\pm$ 0.23 <sup>c,A</sup>
Oil Palm	5.80 $\pm$ 0.64 <sup>a,A</sup>	0.81 $\pm$ 0.49 <sup>b,A</sup>	6.60 $\pm$ 0.42 <sup>b,A</sup>
loam Acrisol soil			
Forest	5.94 $\pm$ 0.40 <sup>a,A</sup>	0.61 $\pm$ 0.15 <sup>ab,B</sup>	6.55 $\pm$ 0.28 <sup>a,A</sup>
Jungle Rubber	5.64 $\pm$ 0.28 <sup>a,B</sup>	1.25 $\pm$ 0.63 <sup>a,A</sup>	6.89 $\pm$ 0.59 <sup>a,A</sup>
Rubber	4.14 $\pm$ 0.57 <sup>b,A</sup>	0.12 $\pm$ 0.02 <sup>b,A</sup>	4.26 $\pm$ 0.58 <sup>b,A</sup>
Oil Palm	4.20 $\pm$ 1.10 <sup>b,B</sup>	0.60 $\pm$ 0.36 <sup>ab,B</sup>	4.81 $\pm$ 1.44 <sup>b,B</sup>



Table 2. Mean ( $\pm$ SE,  $n = 4$ ) soil CO<sub>2</sub> and CH<sub>4</sub> fluxes and annual soil CO<sub>2</sub> and CH<sub>4</sub> fluxes from different land-use types within each soil landscape in Jambi, Sumatra, Indonesia, measured monthly from December 2012 to December 2013. Means followed by different lowercase letter indicate significant differences among land-use types within a soil landscape and different capital letter indicate significant differences between soil landscapes within a land-use type (Linear mixed effects models with Fisher's LSD test at  $P \leq 0.05$ ). Annual soil CO<sub>2</sub> and CH<sub>4</sub> fluxes were not statistically tested for differences between landscapes or land-use types since these annual values are trapezoidal extrapolations. For the loam Acrisol soil, CH<sub>4</sub> fluxes in parenthesis included only the two forest sites that had dominantly net CH<sub>4</sub> uptake, and comparison among land-use types was conducted between jungle rubber, rubber and oil palm that all showed net CH<sub>4</sub> uptake.

Land-use type	CO <sub>2</sub> fluxes (mg C m <sup>-2</sup> h <sup>-1</sup> )	Annual CO <sub>2</sub> fluxes (Mg C ha <sup>-1</sup> year <sup>-1</sup> )	CH <sub>4</sub> fluxes (μg C m <sup>-2</sup> h <sup>-1</sup> )	Annual CH <sub>4</sub> fluxes (kg C ha <sup>-1</sup> year <sup>-1</sup> )
clay Acrisol soil				
Forest	195.93 $\pm$ 13.51 <sup>a,A</sup>	16.93 $\pm$ 1.19	-40.3 $\pm$ 10.25 <sup>c,A</sup>	-3.63 $\pm$ 0.89
Jungle Rubber	185.25 $\pm$ 9.40 <sup>a,A</sup>	16.11 $\pm$ 0.72	-20.79 $\pm$ 7.24 <sup>b,A</sup>	-1.85 $\pm$ 0.59
Rubber	182.77 $\pm$ 16.22 <sup>a,A</sup>	16.09 $\pm$ 1.40	-3.00 $\pm$ 1.26 <sup>a,A</sup>	-0.29 $\pm$ 0.12
Oil Palm	107.24 $\pm$ 7.23 <sup>b,A</sup>	9.22 $\pm$ 0.61	-6.37 $\pm$ 3.06 <sup>a,A</sup>	-0.52 $\pm$ 0.26
loam Acrisol soil				
Forest	186.64 $\pm$ 13.72 <sup>a,A</sup>	16.21 $\pm$ 1.17	-1.56 $\pm$ 17.07 <sup>A</sup> (- 29.45 $\pm$ 11.92)	-0.18 $\pm$ 1.55
Jungle Rubber	178.69 $\pm$ 11.17 <sup>a,A</sup>	15.55 $\pm$ 0.94	-26.92 $\pm$ 3.85 <sup>b,A</sup>	-2.42 $\pm$ 0.34
Rubber	182.86 $\pm$ 14.47 <sup>a,A</sup>	16.52 $\pm$ 1.32	-9.73 $\pm$ 3.79 <sup>a,A</sup>	-0.93 $\pm$ 0.35
Oil Palm	115.74 $\pm$ 10.99 <sup>b,A</sup>	10.29 $\pm$ 0.88	-14.94 $\pm$ 3.14 <sup>a,A</sup>	-1.38 $\pm$ 0.31

1 Table 3. *Pearson correlation coefficients ( $n = 12$ ) between soil CO<sub>2</sub> flux (mg C m<sup>-2</sup> h<sup>-1</sup>), soil*  
2 *CH<sub>4</sub> flux (μg C m<sup>-2</sup> h<sup>-1</sup>), soil temperature (°C, top 0.05-m depth), water-filled pore space*  
3 *(WFPS) (% , top 0.05-m depth) and extractable mineral nitrogen (mg N kg<sup>-1</sup>, top 0.05-m*  
4 *depth), using the means of the four replicate plots per land-use type on monthly measurement*  
5 *between December 2012 – December 2013.*

Land use	Variable	Soil CH <sub>4</sub> flux	Soil temp.	WFPS	NH <sub>4</sub> <sup>+</sup>	NO <sub>3</sub> <sup>-</sup>	Min. N
clay Acrisol soil							
Forest	Soil CO <sub>2</sub> flux	0.19	0.42	0.49	-0.17	0.37	-0.01
	Soil CH <sub>4</sub> flux		0.25	0.68 <sup>b</sup>	0.18	-0.59 <sup>b</sup>	-0.09
	Soil temperature			0.34	0.63 <sup>b</sup>	-0.32	0.54 <sup>a</sup>
	WFPS				0.25	-0.18	0.18
Jungle Rubber	Soil CO <sub>2</sub> flux	-0.03	0.38	0.21	-0.39	0.61 <sup>b</sup>	0.27
	Soil CH <sub>4</sub> flux		0.49	0.74 <sup>c</sup>	0.33	-0.19	0.34
	Soil temperature			0.78 <sup>c</sup>	0.34	0.19	0.39
	WFPS				0.25	0.07	0.28
Rubber	Soil CO <sub>2</sub> flux	-0.51 <sup>a</sup>	0.49	-0.39	0.05	0.14	0.06
	Soil CH <sub>4</sub> flux		-0.14	0.84 <sup>c</sup>	-0.06	-0.52 <sup>a</sup>	-0.1
	Soil temperature			-0.24	0.3	0.16	0.31
	WFPS				-0.06	-0.53 <sup>a</sup>	-0.1

Oil Palm	Soil CO <sub>2</sub> flux	-0.29	0.82 <sup>c</sup>	-0.37	0.31	0.24	0.41
	Soil CH <sub>4</sub> flux		-0.09	0.69 <sup>c</sup>	0.19	0.13	0.25
	Soil temperature			-0.19	0.32	0.32	0.52 <sup>a</sup>
	WFPS				0.16	0.08	0.16
loam Acrisol soil							
Forest	Soil CO <sub>2</sub> flux	0.12	0.58 <sup>b</sup>	0.05	-0.12	0.23	-0.01
	Soil CH <sub>4</sub> flux		0.19	0.32	0.09	-0.24	-0.24
	Soil temperature			0.42	0.41	-0.03	0.37
	WFPS				0.4	-0.33	0.23
Jungle Rubber	Soil CO <sub>2</sub> flux	0.74 <sup>c</sup>	0.21	0.59 <sup>b</sup>	-0.05	-0.60 <sup>b</sup>	-0.41
	Soil CH <sub>4</sub> flux		0.35	0.74 <sup>c</sup>	0.35	-0.58 <sup>b</sup>	0.11
	Soil temperature			0.42	0.47	-0.22	0.38
	WFPS				0.32	-0.67 <sup>b</sup>	0.05
Rubber	Soil CO <sub>2</sub> flux	-0.74 <sup>c</sup>	0.16	-0.54 <sup>a</sup>	0.06	-0.07	0.05
	Soil CH <sub>4</sub> flux		-0.07	0.84 <sup>c</sup>	0.33	-0.11	0.32
	Soil temperature			0.07	0.57 <sup>b</sup>	-0.42	0.54 <sup>a</sup>
	WFPS				0.23	-0.24	0.2

Oil Palm	Soil CO <sub>2</sub> flux	-0.05	0.57 <sup>a</sup>	-0.29	0.25	0.36	-0.05
	Soil CH <sub>4</sub> flux		0.16	0.86 <sup>c</sup>	0.06	0.17	0.1
	Soil temperature			0.08	0.13	-0.19	0.16
	WFPS				-0.08	-0.05	-0.07

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1 <sup>a</sup>  $P \leq 0.09$ , <sup>b</sup>  $P \leq 0.05$ , <sup>c</sup>  $P \leq 0.01$

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3

## List of Figures

Figure 1. Mean ( $\pm$ SE,  $n = 4$ ) soil water-filled pore space (WFPS) and soil temperature in the top 0.05-m depth under forest ( $\blacklozenge$ ), jungle rubber ( $\diamond$ ), rubber ( $\blacktriangle$ ) and oil palm ( $\triangle$ ) on the clay Acrisol soil (**a** and **c**) and the loam Acrisol soil (**b** and **d**) in Jambi, Sumatra, Indonesia, measured monthly from December 2012 to December 2013. Grey shadings mark the dry season

Figure 2. Mean ( $\pm$ SE,  $n = 4$ ) soil CO<sub>2</sub> fluxes and soil CH<sub>4</sub> fluxes from forest ( $\blacklozenge$ ), jungle rubber ( $\diamond$ ), rubber ( $\blacktriangle$ ) and oil palm ( $\triangle$ ) on the clay Acrisol soil (**a** and **c**) and the loam Acrisol soil (**b** and **d**) in Jambi, Sumatra, Indonesia, measured monthly from December 2012 to December 2013. Grey shadings mark the dry season.

Figure 3. Relationship between soil CH<sub>4</sub> uptake and soil mineral N content, using the means of four replicate plots per land-use type on monthly measurement between December 2012 – December 2013: forest ( $\blacklozenge$ ), jungle rubber ( $\diamond$ ), rubber ( $\blacktriangle$ ) and oil palm ( $\triangle$ ) on the clay Acrisol soil (Pearson correlation:  $R = -0.47$ ,  $P = 0.01$ ,  $n = 41$ ) (**a**) and the loam Acrisol soil (Pearson correlation:  $R = -0.52$ ,  $P < 0.01$ ,  $n = 38$ ) (**b**). Correlations exclude net CH<sub>4</sub> emissions (fluxes above 0) in both landscapes and an outlier plot of oil palm on the loam Acrisol soil (shaded grey).

Table A1. *Mean ( $\pm$ SE,  $n = 4$ ) soil physical and biochemical characteristics in the top 0.10-m depth (except for the clay content, 0-2-m depth with  $n = 3$ ) for different land-use types within each soil landscape in Jambi, Sumatra, Indonesia. Means followed by different lowercase letter indicate significant differences among land-use types within each soil landscape and different capital letter indicate significant differences between soil landscapes within a land-use type (Linear mixed effects models with Fisher's LSD test at  $P \leq 0.05$  and marginally significant at  $^{\dagger}P \leq 0.09$ ). Soil characteristics were measured by Allen et al. (2015). Soil sampling and analysis are described in Appendix C.*

Soil characteristics	Land use			
	Forest	Jungle Rubber	Rubber	Oil Palm
clay Acrisol soil				
Clay (0-0.5 m) (%)	31.4 $\pm$ 5.4 <sup>a</sup>	47.2 $\pm$ 12.40 <sup>a</sup>	42.4 $\pm$ 3.1 <sup>a</sup>	59.7 $\pm$ 5.2 <sup>a,A</sup>
Clay (0.5-1.0 m) (%)	34.9 $\pm$ 9.0 <sup>b†</sup>	51.4 $\pm$ 12.6 <sup>ab†</sup>	36.8 $\pm$ 8.00 <sup>b†</sup>	69.7 $\pm$ 4.8 <sup>a†A</sup>
Clay (1.0-1.5 m) (%)	39.0 $\pm$ 13.0 <sup>a</sup>	62.8 $\pm$ 12.6 <sup>a</sup>	40.8 $\pm$ 10.3 <sup>a</sup>	62.8 $\pm$ 3.6 <sup>a,A</sup>
Clay (1.5-2.0 m) (%)	41.3 $\pm$ 11.2 <sup>a</sup>	46.6 $\pm$ 16.2 <sup>a</sup>	36.5 $\pm$ 10.8 <sup>a</sup>	63.3 $\pm$ 6.1 <sup>a,A</sup>
Sand (0-0.10 m) (%)	36 $\pm$ 11 <sup>a</sup>	27 $\pm$ 20 <sup>a</sup>	35 $\pm$ 7 <sup>a</sup>	11 $\pm$ 2 <sup>a,B†</sup>
Bulk density (g cm <sup>-3</sup> )	1.0 $\pm$ 0.1 <sup>a</sup>	0.8 $\pm$ 0.1 <sup>a</sup>	0.9 $\pm$ 0.1 <sup>a</sup>	0.9 $\pm$ 0.1 <sup>a,B</sup>
pH (1:4 H <sub>2</sub> O)	4.2 $\pm$ 0.4 <sup>b</sup>	4.5 $\pm$ 0.0 <sup>a,A</sup>	4.5 $\pm$ 0.1 <sup>a</sup>	4.4 $\pm$ 0.1 <sup>a</sup>
Soil organic C (kg C m <sup>-2</sup> )	3.3 $\pm$ 0.5 <sup>a</sup>	4.3 $\pm$ 0.4 <sup>a,A</sup>	2.8 $\pm$ 0.4 <sup>a</sup>	3.5 $\pm$ 0.2 <sup>a,A</sup>
Total N (g N m <sup>-2</sup> )	263.4 $\pm$ 67.1 <sup>a</sup>	331.4 $\pm$ 34.1 <sup>a,A</sup>	198.4 $\pm$ 32.5 <sup>a</sup>	260.2 $\pm$ 22.6 <sup>a,A</sup>
C:N ratio	13.1 $\pm$ 1.3 <sup>a</sup>	13.0 $\pm$ 0.3 <sup>a</sup>	14.3 $\pm$ 0.6 <sup>a,A</sup>	13.5 $\pm$ 0.2 <sup>a</sup>
Effective cation exchange capacity (cmol <sub>c</sub> kg <sup>-1</sup> )	9.4 $\pm$ 4.1 <sup>a</sup>	12.4 $\pm$ 2.6 <sup>a,A</sup>	7.1 $\pm$ 2.2 <sup>a</sup>	7.8 $\pm$ 0.8 <sup>a,A</sup>
Base saturation (%)	23 $\pm$ 6 <sup>a,A</sup>	23 $\pm$ 6 <sup>a</sup>	20 $\pm$ 3 <sup>a</sup>	38 $\pm$ 7 <sup>a</sup>
Aluminum saturation (%)	61 $\pm$ 3 <sup>ab,B</sup>	71 $\pm$ 6 <sup>a</sup>	73 $\pm$ 4 <sup>a</sup>	53 $\pm$ 7 <sup>b</sup>
Bray-extractable	1.4 $\pm$ 0.1 <sup>ab,A</sup>	0.8 $\pm$ 0.1 <sup>bc</sup>	0.4 $\pm$ 0.0 <sup>c</sup>	4.7 $\pm$ 1.5 <sup>a,A†</sup>

phosphorus (g P m <sup>-2</sup> )				
<sup>15</sup> N natural abundance (‰)	4.5 ± 0.0 <sup>a</sup>	4.0 ± 0.3 <sup>a</sup>	4.6 ± 0.4 <sup>a</sup>	5.2 ± 0.4 <sup>a</sup>
Net N mineralization (mg N kg <sup>-1</sup> d <sup>-1</sup> )	1.2 ± 0.3 <sup>a</sup>	0.5 ± 0.0 <sup>b</sup>	0.5 ± 0.2 <sup>b</sup>	0.9 ± 0.2 <sup>ab</sup>
loam Acrisol soil				
Clay (0-0.5 m) (%)	26.0 ± 2.6 <sup>a</sup>	30.6 ± 4.6 <sup>a</sup>	37.3 ± 10.3 <sup>a</sup>	33.4 ± 2.2 <sup>a,B</sup>
Clay (0.5-1.0 m) (%)	28.7 ± 4.8 <sup>a</sup>	38.8 ± 9.0 <sup>a</sup>	45.1 ± 11.3 <sup>a</sup>	41.0 ± 3.1 <sup>a,B</sup>
Clay (1.0-1.5 m) (%)	33.3 ± 7.56 <sup>a</sup>	42.4 ± 9.9 <sup>a</sup>	46.1 ± 9.9 <sup>a</sup>	43.3 ± 2.8 <sup>a,B</sup>
Clay (1.5-2.0 m) (%)	37.3 ± 8.7 <sup>a</sup>	44.5 ± 10.0 <sup>a</sup>	43.4 ± 6.5 <sup>a</sup>	47.6 ± 4.5 <sup>a,B</sup>
Sand (0-0.10 m) (%)	39 ± 8 <sup>a</sup>	42 ± 19 <sup>a</sup>	26 ± 13 <sup>a</sup>	43 ± 14 <sup>a,A†</sup>
Bulk density (g cm <sup>-3</sup> )	1.0 ± 0.0 <sup>ab</sup>	0.9 ± 0.0 <sup>b</sup>	1.1 ± 0.1 <sup>a</sup>	1.1 ± 0.1 <sup>a,A</sup>
pH (1:4 H <sub>2</sub> O)	4.3 ± 0.0 <sup>b†</sup>	4.3 ± 0.0 <sup>b†,B</sup>	4.5 ± 0.1 <sup>ab†</sup>	4.5 ± 0.1 <sup>a†</sup>
Soil organic C (kg C m <sup>-2</sup> )	2.6 ± 0.2 <sup>a</sup>	2.7 ± 0.3 <sup>a,B</sup>	2.0 ± 0.3 <sup>a</sup>	1.8 ± 0.2 <sup>a,B</sup>
Total N (g m <sup>-2</sup> )	182.9 ± 10.8 <sup>a</sup>	186.19 ± 11.0 <sup>a,B</sup>	172.6 ± 23.8 <sup>a</sup>	145.0 ± 13.5 <sup>a,B</sup>
C:N ratio	14.3 ± 0.2 <sup>a</sup>	13.7 ± 0.8 <sup>a</sup>	11.7 ± 0.7 <sup>b,B</sup>	12.5 ± 0.5 <sup>ab</sup>
Effective cation exchange capacity (mmol <sub>c</sub> kg <sup>-1</sup> )	4.5 ± 0.5 <sup>a</sup>	4.1 ± 0.8 <sup>a,B</sup>	4.6 ± 0.5 <sup>a</sup>	4.0 ± 0.8 <sup>a,B</sup>
Base saturation (%)	11 ± 1 <sup>b†,B</sup>	16 ± 2 <sup>ab†</sup>	21 ± 8 <sup>ab†</sup>	28 ± 5 <sup>a†</sup>
Aluminum saturation (%)	80 ± 1 <sup>a,A</sup>	78 ± 2 <sup>a</sup>	73 ± 8 <sup>a</sup>	67 ± 5 <sup>a</sup>
Bray-extractable phosphorus (g P m <sup>-2</sup> )	0.5 ± 0.1 <sup>a,B</sup>	0.7 ± 0.1 <sup>a</sup>	0.5 ± 0.1 <sup>a</sup>	0.8 ± 0.1 <sup>a,B†</sup>
<sup>15</sup> N natural abundance (‰)	4.3 ± 0.2 <sup>b</sup>	4.5 ± 0.1 <sup>b</sup>	5.0 ± 0.4 <sup>ab</sup>	5.4 ± 0.3 <sup>a</sup>
Net N mineralization (mg N kg <sup>-1</sup> d <sup>-1</sup> )	0.8 ± 0.2 <sup>a</sup>	0.7 ± 0.1 <sup>a</sup>	0.7 ± 0.3 <sup>a</sup>	0.5 ± 0.2 <sup>a</sup>

1 Table A2. *Plantation age and mean ( $\pm$ SE,  $n = 4$ ) tree density, tree height, basal area, diameter at breast height (DBH) of trees with  $\geq 0.10$  m*  
 2 *BDH and the most common tree species for different land-use types within each soil landscape in Jambi, Sumatra, Indonesia.*

Land-use type	Age (years)	range	Tree density (n ha <sup>-1</sup> ) <sup>a</sup>	Tree height (m) <sup>a</sup>	Basal area (m <sup>2</sup> ha <sup>-1</sup> ) <sup>a</sup>	DBH (cm) <sup>a</sup>	Most common tree species <sup>b</sup>
clay Acrisol soil							
Forest	not mined (ND)	deter-	471 $\pm$ 31	17.0 $\pm$ 0.5	29.4 $\pm$ 1.7	23.0 $\pm$ 0.4	<i>Archidendron sp.</i> , <i>Baccaurea spp.</i> , <i>Ochanostachys sp.</i>
Jungle rubber	ND		685 $\pm$ 72	15.2 $\pm$ 0.3	21.1 $\pm$ 1.4	17.3 $\pm$ 0.6	<i>Artocarpus spp.</i> , <i>Endospermum sp.</i> , <i>Hevea sp.</i> , <i>Macaranga spp.</i>
Rubber	7-16		497 $\pm$ 15	13.4 $\pm$ 0.1	10.0 $\pm$ 1.4	15.2 $\pm$ 0.7	<i>Hevea brasiliensis</i>
Oil Palm	9-13		134 $\pm$ 6	4.0 $\pm$ 0.3	not applicable (NA)	NA	<i>Elaeis guineensis</i>



loam Acrisol soil						
Forest	ND	658 ± 26	20.0 ± 0.6	30.7 ± 1.0	21.0 ± 0.5	<i>Aporosa spp.</i> , <i>Burseraceae spp.</i> , <i>Dipterocarpaceae spp.</i> , <i>Fabaceae spp.</i> , <i>Gironniera spp.</i> , <i>Myrtaceae spp.</i> , <i>Plaquium spp.</i> , <i>Porterandia sp.</i> , <i>Shorea spp.</i>
Jungle Rubber	ND	525 ± 60	14.0 ± 0.2	16.6 ± 0.4	16.8 ± 0.5	<i>Alstonia spp.</i> , <i>Artocarpus spp.</i> , <i>Fabaceae sp.</i> , <i>Hevea sp.</i> , <i>Macaranga spp.</i> , <i>Porterandia sp.</i> , <i>Sloetia sp.</i>
Rubber	14-17	440 ± 81	13.4 ± 0.5	12.2 ± 1.6	17.8 ± 1.2	<i>Hevea brasiliensis</i>
Oil Palm	12-16	140 ± 4	4.9 ± 0.6	NA	NA	<i>Elaeis guineensis</i>

1 <sup>a</sup> Kotowska et al. (2015).

2 <sup>b</sup> Rembold et al. (unpublished data), based on trees found in five subplots (5 m x 5 m) of each replicate plot (50 m x 50 m) which had ≥ 20  
3 individuals, except *Fabaceae spp.* which had ≤ 20 individuals.

Table A3. Mean ( $\pm$ SE,  $n = 3$  oil palm trees) soil CO<sub>2</sub> and CH<sub>4</sub> fluxes from three different chamber locations during a fertilization in three oil palm plantations within each soil landscape, measured 6 to 11 times during 3-8.5 weeks following fertilization. Means followed by different letter indicate significant differences among chamber locations within each oil palm plantation site (Linear mixed effects models with Fisher's LSD test at  $P \leq 0.05$ ). Chamber locations a, b and c were placed at 0.3 m, 0.8 m, and 4-4.5 m, respectively, from each of the three trees in each oil palm plantation site. Smallholders fertilized around the base of each tree at about 0.8 – 1 m from the tree base, and thus chamber location b was on this fertilized area and chamber location c serves as the reference chamber not receiving any fertilizer. The same fertilization rate and form were used as the smallholders applied in these studied oil palm plantations, described in 2.2 CO<sub>2</sub> and CH<sub>4</sub> flux measurement.

Oil palm plantation site	Chamber location	CO <sub>2</sub> fluxes (mg C m <sup>-2</sup> h <sup>-1</sup> )	CH <sub>4</sub> fluxes (μg C m <sup>-2</sup> h <sup>-1</sup> )
clay Acrisol soil			
1	a	272.83 $\pm$ 36.68 <sup>a</sup>	-23.66 $\pm$ 2.56 <sup>b</sup>
	b	218.25 $\pm$ 25.91 <sup>b</sup>	-12.61 $\pm$ 5.12 <sup>a</sup>
	c	103.56 $\pm$ 11.72 <sup>c</sup>	-16.66 $\pm$ 8.68 <sup>ab</sup>
2	a	226.16 $\pm$ 38.17 <sup>a</sup>	-28.44 $\pm$ 1.48 <sup>b</sup>
	b	246.39 $\pm$ 42.80 <sup>a</sup>	-6.64 $\pm$ 2.07 <sup>a</sup>
	c	86.04 $\pm$ 7.83 <sup>b</sup>	-10.60 $\pm$ 5.29 <sup>a</sup>
3	a	222.56 $\pm$ 72.49 <sup>b</sup>	-8.13 $\pm$ 4.77 <sup>a</sup>
	b	311.63 $\pm$ 89.87 <sup>a</sup>	-10.38 $\pm$ 3.61 <sup>a</sup>
	c	105.49 $\pm$ 12.06 <sup>c</sup>	-14.49 $\pm$ 2.03 <sup>a</sup>
loam Acrisol soil			
1	a	334.67 $\pm$ 32.12 <sup>a</sup>	-14.00 $\pm$ 3.31 <sup>b</sup>
	b	378.47 $\pm$ 50.97 <sup>a</sup>	-4.12 $\pm$ 2.24 <sup>a</sup>

	c	$160.35 \pm 20.48^b$	$-18.53 \pm 1.32^b$
2	a	$271.35 \pm 17.31^a$	$-13.18 \pm 1.22^a$
	b	$233.27 \pm 18.98^b$	$-8.63 \pm 0.52^a$
	c	$127.66 \pm 17.13^c$	$-19.47 \pm 5.08^b$
3	a	$240.81 \pm 23.12^a$	$-28.13 \pm 3.40^b$
	b	$243.92 \pm 24.23^a$	$-10.34 \pm 2.70^a$
	c	$136.55 \pm 19.08^b$	$-29.41 \pm 2.39^b$

1  
2

1 Appendix Fig. B1. Mean ( $\pm$ SE,  $n = 3$  oil palm trees) soil CH<sub>4</sub> fluxes during a fertilization in  
2 one oil palm plantation site in the clay Acrisol soil (▲) and loam Acrisol soil (●). Smallholders  
3 fertilized around the base of each tree at about 0.8 – 1 m from the tree base, and these fluxes  
4 were measured on this fertilized location (chamber location b) with the same rate and form  
5 that smallholders applied in these oil palm plantations (described in 2.2 *CO<sub>2</sub> and CH<sub>4</sub> flux*  
6 *measurement*).

## Appendix: soil sampling and analysis

Soil samples were taken from randomly selected ten subplots per plot that were at least 5 m distance from the plot's border. Soil characteristics for each replicate plot were the average of the ten subplots. Soil sampling was conducted between June 2013 and December 2013. Soil samples were taken at various depth intervals down to 2 m, and we report here the values from the top depth interval (0-0.1 m), except for soil texture, which we report for the entire 2 m. Soil texture was analyzed using the wet sieving and pipette methods. Soil bulk density was measured using the core method. Soil pH (H<sub>2</sub>O) was analyzed in a 1:4 soil-to-water ratio. Soil organic C and total N concentrations were analyzed from air-dried, sieved (2 mm) and ground samples using a CN analyzer (Vario EL Cube, Elementar Analysis Systems GmbH, Hanau, Germany). Air-dried and sieved soils were used to determine effective cation exchange capacity (ECEC) by percolating with unbuffered 1 mol L<sup>-1</sup> NH<sub>4</sub>Cl and cations (Ca, Mg, K, Na, Al, Fe, and Mn) were measured in percolate using an inductively coupled plasma-atomic emission spectrometer (iCAP 6300 Duo VIEW ICP Spectrometer, Thermo Fischer Scientific GmbH, Dreieich, Germany). Base and aluminum saturation were calculated as percent exchangeable base cations and aluminum of the ECEC. Extractable P was determined using the Bray 2 method, which is typically used for acidic tropical soils. For soil <sup>15</sup>N natural abundance signatures, ground soil samples were analyzed using isotope ratio mass spectrometry (IRMS; Delta Plus, Finnigan MAT, Bremen, Germany). Net N mineralization rate was measured in two subplots per plot that were at least 10 m from the plot's border, using the buried bag method on intact soil cores incubated in situ for 7 days. This was conducted between January 2013 and May 2013 during the rainy season. The same field extraction of the soil with 0.5 M K<sub>2</sub>SO<sub>4</sub>, analysis of NH<sub>4</sub><sup>+</sup> and NO<sub>3</sub><sup>-</sup> concentrations, and calculation of rate are used as described in our earlier work (Arnold et al., 2008). Net N mineralization rate for each plot was the average of two subplots.