

Dear Editor(s),

28 October 2015

Please find attached our revised manuscript, including a marked version here with revisions marked in track changes mode. The revisions in our manuscript were done in response to comments provided by reviewers one and two (our point by point responses documenting these revisions have already been uploaded, and we copied them below here as well). In addition, we made changes described below in response to the final comments by Associate Editor Warwick F. Vincent on 12 Oct 2015.

Comments from W.F. Vincent: Please keep in mind that the concerns raised by the referees may also be those of future readers. It will be useful to transfer some of your very helpful response comments (in condensed form) to your revised manuscript. For example, one point that seems a bit contradictory is in response to referee 2 who said: "Could the authors thereby combine in situ spectral UV absorption and residence time (treated separately in much of the Discussion) into a single equation = cumulative UV exposure or absorption. In this way 'light-limitation' could occur through a) low surface irradiance and light limitation at all depths; b) complete absorption of irradiance in the surface waters and light limitation at depth; or c) insufficient time (cumulative exposure) for complete photochemical breakdown. You responded: "This amount of "UV light exposure" and "DOC degradation" (from equation 3) is then multiplied by the residence time (which is what it seems the reviewer is requesting), and the results are presented in Fig. 9." But then you say: "Beyond what is presented in a figure, it might be convenient to have a single equation highlighting light exposure and residence time – this equation would multiply equation 3 by  $R_t$ , residence time. However, the situation as we present it in the manuscript is more complicated...." This seems to contradict your previous statement. One way to handle this (but you may have a better way) would be to condense your comments into something in the Discussion like 'Equation 3 was multiplied by residence time ( $R_t$ ) to generate the results in Fig. 9. However, a precise estimate of  $R_t$  is difficult to achieve in practice given that there are inputs of water and fresh CDOM as a parcel of water moves downstream.'

Author response: We took your suggestion and added condensed versions of our responses to reviewer comments to the discussion on (1) the effects of residence time on DOM photo-degradation (including the challenges associated with this assessment), and (2) the counter-intuitive result that protection of the bottom water from UV light under stratified conditions actually leads to more DOM photo-degradation under these conditions. These revisions are on pages 20-21 in the MARKED (track changes) manuscript (attached in this document).

Sincerely,



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### **Author responses to Anonymous Referee #1**

Reviewer: - Introduction, the introduction is well written and provides a solid background and rational for the presented study - Methods, the methods used in this study are robust and thoroughly presented - Results and discussion, the results in the text are consistent with the data presented in the tables and figures. The discussion walks the reader through the conceptual model clearly and thoroughly and is well supported with literature. - The word “significant” is used throughout the results and discussion with no p values (or alternative measures of significance) presented. The authors should add in tests of significance or consider revising the text to reflect that their differences are measured, but not tested statistically for significance.

Reviewer: Technical Corrections: - Figures 3 and 4 are tough to read with such a small font. Would it be possible for the authors to increase the size? - Page 9818, line 7, “attention” should be revised to read “attenuation”.

*Author Response: Thank you for the review of our manuscript.*

*We revised Tables 2 and 3 to more clearly indicate when measures of DOM composition were statistically, significantly different. For example, for Table 2, we used t-tests to evaluate whether means for a water type were significantly different between years (2011 vs. 2012) at  $p < 0.05$ . For the data shown in Table 3, we did an ANOVA comparing mean values among the three water types (soil water, pool bottom water, and pool surface water), across both years ( $p < 0.01$ ). As now indicated in the footnote of Table 3, all water types were statistically different from one another for all variables.*

*In contrast to Tables 2 and 3 which evaluate mean water or DOM chemistry by water type over all dates, the data shown in Figure 5 compare DOM concentration and chemistry between surface and bottom water in each pool on two dates, with mean  $\pm$  SE (of replicates from the same sample bottle). Because a single discrete sample was analyzed from each depth in each pool, no t-tests could be conducted on surface vs. bottom water values from each pool in this Figure. Nonetheless, the results in Figure 5 show that the concentration and composition of DOM in surface waters generally did not overlap with the values of DOM in the bottom water (analytical replicates). Thus, the data in Figure 5 provide an example of one date to show why surface waters as a group were significantly different than bottom waters when pools were stratified (as quantified by t-tests or ANOVA in Tables 2 and 3, respectively).*

*We increased the font and can request that the journal re-size Figures 3 and 4 (new versions uploaded).*

*We replaced the typo “attention” with attenuation on line 9818.*

### **Author Responses to Anonymous Referee #2**

*Author Response: Thank you for the review of our manuscript.*

Reviewer: P9795, Line 26: “We suggest that degradation, and thus export, of DOM in CDOM-rich streams or ponds similar to Imnavait is typically light-limited under most flow conditions.”

This does not sound logically correct to me: if degradation reduces DOM export, then a factor that limits degradation should favor rather than limit DOM export.

*Author response: We revised the text in the abstract to clarify that because DOM degradation is light-limited in Innavait Creek, export of DOM from this stream will be less under conditions that increase the light available for DOM photo-degradation (i.e., low flows, sunny days).*

Reviewer: P9081, Line 2: Optical measurements were at what time of day (zenith angle, used later as an important correction for a) and under what sky conditions (OVC, scattered, clear skies)?

*Author response: In-situ attenuation coefficients were measured between 10 -11 am Alaska Standard Time on 28 June 2011 and between 11 am - 2:30pm on 23 June 2012. Conditions were mostly sunny (28 June 2011) or sunny (23 June 2012). Specifically, the cloudiness factor for these dates was 0.7 and 1, respectively on these dates, as calculated in Cory et al. 2014 to estimate the amount of direct vs. indirect light reaching the water surface export of DOM. The cloudiness factor was calculated as the ratio of mean measured to mean modeled (SMARTS) solar irradiance at Toolik Field Station, and was found to range from ~ 0.3 on cloudy days typical in August to ~ 1 for the clear skies characteristic of May-June at the field station near Innavait Creek (Fig. S6 in Cory et al. 2014). As described in Cory et al. 2014, in-situ  $K_{d,\lambda}$  values were compared to  $a_{CDOM,\lambda}$  values obtained from the spectrophotometer after correcting for the solar zenith angle corresponding to the time of day the  $K_{d,\lambda}$  values were measured.*

*We cited the Cory et al. paper for these methods on  $K_{d,\lambda}$  to account for the time and sky conditions when  $K_{d,\lambda}$  was measured, but did not revise the text to add more of this background information to the manuscript given that it is published in the Cory et al. 2014 paper.*

Reviewer: The term UV exposure is used but not fully defined (e.g. P9797, Line 25). It would be interesting to distinguish exposure rate (spectrally integrated mol photons absorbed by CDOM  $m^{-2} s^{-1}$  and cumulative exposure or dose (exposure rate integrated over the time for a parcel of water to travel through a defined reach). Could the authors thereby combine in situ spectral UV absorption and residence time (treated separately in much of the Discussion) into a single equation = cumulative UV exposure or absorption. In this way 'light-limitation' could occur through a) low surface irradiance and light limitation at all depths; b) complete absorption of irradiance in the surface waters and light limitation at depth; or c) insufficient time (cumulative exposure) for complete photochemical breakdown.

*Author response: We revised the text in the methods to clarify that UV exposure means the amount of light reaching the water surface that is available to be absorbed by CDOM ( " $Q_{ds0, \lambda}$ " in Equation 3), where UV exposure varies by time of day, day of year, and by sunny vs. cloudy days.*

*We also revised the text in the discussion on residence time and light-limitation of photo-degradation in response to AE Vincent's request to condense our initial response to this reviewer*

*here (that is, our discussion below in response to this reviewer's comment was condensed and added to the discussion):*

*The reviewer is correct that in a general sense the overall “light limitation” is a function of “exposure rate” and the “length of exposure” or residence time. This is what we are showing in our conceptual diagram in Fig. 8, and we have quantified this in our Fig. 9. Our equation 3 (a standard equation in photochemical studies) describes how much light is available at the water surface (UV exposure), how much light is absorbed by CDOM, and then per light absorbed how much DOM is degraded (for example, the mol of CO<sub>2</sub> produced per mol of light absorbed, which is the apparent quantum yield). This amount of “UV light exposure” and “DOC degradation” (from equation 3) is then multiplied by the residence time (which is what it seems the reviewer is requesting), and the results are presented in Fig. 9. In this figure the DOC loss is shown as a function of residence time; thus, at any given residence time the various lines (for different conditions of light availability, CDOM amount, and apparent quantum yields) give the cumulative loss of DOC. It appears that this is what the reviewer is asking for, an independent treatment of in-situ absorption and degradation (the three lines of different slopes in Fig. 9) and of residence time (the Y-axis in Fig. 9). We experimented with adding a third axis to our conceptual diagram to illustrate the relationships between (a) light at the surface, (b) light absorption and limitation at depth, and (c) residence time (as mentioned by the reviewer), but found that the figure became very complicated and lost its simplicity at conveying our ideas. Also, we would need a fourth axis to convey the factor of DOM lability (represented in the apparent quantum yield) that we show is important as well – therefore, we believe that the most simple and effective way to present this information is the combination of Figs. 8 and 9.*

*Beyond what is presented in a figure, it might be convenient to have a single equation highlighting light exposure and residence time – this equation would multiply equation 3 by  $Rt$ , residence time. However, the situation as we present it in the manuscript is more complicated. For example,  $Rt$  as a concept is simple (the stock divided by the input or output rate at steady state), but it can be extremely difficult to calculate in practice and a discussion of those calculations is beyond this manuscript. Furthermore, both our current approach and the approach suggested by the reviewer do not consider inputs of water (and CDOM) as a parcel of water moves downstream. Considering the input of water containing “fresh” (labile) CDOM is needed to quantify the total, integrated amount of light absorbed by CDOM as a parcel of water moves downstream over time. However, this as well is beyond the scope of the current study.*

Reviewer: Fig. 2. It is interesting that  $K_d$  and the absorption coefficients (zenith angle adjusted) were so close – so no optical scattering in this environment? Or masked by the effects of having such high  $a_{CDOM}$ . What was the range of suspended sediment concentrations and POC in this water?

*Author response:* We modified the text in the results to clarify that your interpretation is correct, there was very little optical scattering and absorption by POC in this very high CDOM water. Strong agreement between  $K_{d,\lambda}$  and  $a_{CDOM,\lambda}$  values here are consistent with agreement reported for a wider range of surface waters in the Arctic by Cory et al. 2014 and Gareis et al. 2010, for example (as noted in the manuscript on pg 9811, first paragraph). We did not collect samples

*for POC analysis from Imnavait Creek for this study because POC is on average ~ 20-fold less than DOC concentrations in streams like Imnavait Creek near Toolik Lake (Kling et al. 2000). Relatively low POC concentrations (relative to DOC) are consistent with the strong agreement between  $K_{d,\lambda}$  and  $a_{CDOM,\lambda}$  in Imnavait Creek suggesting there was little scattering in the UV by organic or mineral particles. There are nearby rivers that are strongly dominated by glacial inputs of suspended sediments, and in this situation the relationship between  $K_{d,\lambda}$  and  $a_{CDOM,\lambda}$  weakens (see Cory et al. 2014).*

Reviewer: P9806, Line 18 onwards: There is a long speculative section to discuss the outlier points in this graph – this could be contracted, transferred to the discussion or deleted. Also, could these outliers simply be the result of absorption and scattering by naturally suspended sediments on those dates?

*Author response: We shortened this section, which was included to speculate on the reasons other than scattering by sediment (see response to above comment), to explain the poor agreement between  $K_{d,\lambda}$  and  $a_{CDOM,\lambda}$  for a few samples. We removed interpretations from the text other than the most likely reason for these outliers, i.e., the measurement error associated with measuring  $K_{d,\lambda}$  values in-situ in streams containing high concentrations of CDOM where the UV light is rapidly attenuated.*

Reviewer: P9811, Line 10: ‘Because UV and PAR account for approximately 51% of the energy within the shortwave radiation portion of the spectrum (300–2500 nm), absorption of sunlight by CDOM contributes to the frequency and extent of stratification by restricting warming to the surface layers (Merck and Neilson, 2012).’ See also: Caplanne S & Laurion I (2008) Effect of chromophoric dissolved organic matter on epilimnetic stratification in lakes. *Aquatic Sciences* 70 (2), 123-133.

*Author response: We added a citation to the work of Caplanne and Laurion (2008) on the role of CDOM in lake stratification, which showed that as CDOM absorption coefficient increased, UVA and especially visible light played an increasing role in stratification compared to low CDOM lake.*

Reviewer: Figs 3 and 4: The axis labels need to be checked-  $\_C$ , m3/s. Also PxVA or ‘vertical array’ is not a variable – the pond labels could be, for example for Pond 5: P5T ( $\_C$ ). For these figures, it would be helpful to know what the measurement depths were for each pond. If this would make the caption too long, each probe depth for each pond could be spelled out in the Methods (at the moment it is only given as a broad range).

*Author response: We revised the figure to fix the axes as suggested. We added pool depth to the legend for each pool, and added text in the figure captions for Figs 3 and 4 to indicate that the sensors were placed in each pool starting 5 to 15 cm from the bottom of the pool and then at intervals ranging from 5 to 50 cm (which is also in the methods section on in-situ monitoring).*

Reviewer: P9817, Line 14: ‘but this stratification serves to protect DOM from UV light by isolating water masses in pool bottoms (e.g., Table 3, Fig. 5).’ How does the river flow during these conditions – is water flowing across the surface of the stratified pond? In

which case, stratification is reducing the 'light limitation' of the flowing water because its mean depth is shallower than under conditions of full water column mixing? What is the volume of the water sequestered at the bottom of a typical pool relative to the river flow – i.e. equivalent to how many seconds, minutes hours, days of average flow? Is it a major or minor component?

*Author response: We took the associate editor Vincent's suggestion to add condensed versions of our responses to this reviewer's comments (below) to the discussion.*

*Original responses (condensed versions added to discussion): You are correct that under stratified conditions water moves across the surface of the ponds. These conditions increase the residence time of the pool bottom water because these waters are stagnant until the next mixing event. The effect of stratification on the light exposure of DOM in the bottom waters is low because, as stated on p.9807 lines 20-26, the depth of the UVB and UVA light penetration is always less than the diel mixing depth when stratified, meaning that DOM in the bottom of the pools is protected from UV light under stratified conditions.*

*It is also important to note that the depth of light penetration into the ponds does not differ between stratified (low flow) or mixed (high flow) conditions as shown by the limited differences in  $a_{CDOM\lambda}$  values at 305 nm between these conditions in Innavait Creek (comparing pool surface  $a_{CDOM\lambda}$  values in 2011 vs. 2012, Table 3). Thus, the amount of CDOM exposed to light, or the rate of light absorption, does not differ between stratified vs. mixed conditions (for a given amount of sunlight under given sky conditions). The only difference is the amount of time for the photo-degradation to occur (greater photo-degradation under longer residence times associated with low-flow, stratified conditions; Figure 9).*

*Of course it is more complicated than that because during peak radiation nearly the entire water column can become stratified, thus increasing the residence time in the surface waters until there is cooling at night and downward mixing (although not all the way to the bottom). Such complex dynamics require a model to characterize and understand, and while interesting it is beyond the scope of the current paper.*

*The volume of water sequestered in the pool bottoms (below the mixing depth) under stratified conditions varies, but on average was about 70% of the total pool volume. Pool volumes ranged from  $< 1 \text{ m}^3$  (for pool 4), to  $\sim 100 \text{ m}^3$  (pool 7), and active volumes (defined as the volume of the pool undergoing mixing), were around 30% of pool volume under stratified conditions (compared to 100% of the pool volume mixing under mixed conditions). This means that under stratified conditions, the majority of the pool volume was sequestered in the bottom, below the depth of UV light penetration (surface mixing layer depth was on average 50 cm, compared to depth of UVB and UVA light penetration of 8 – 45 cm as discussed on pg. 9807 lines 20-30).*

*Although most water was sequestered in the pool bottoms under stratified conditions, more DOM is actually lost due to photo-degradation under these conditions. This finding may seem counter-intuitive at first, considering that most of the DOC was protected from photo-degradation when it was sequestered in pool bottoms under stratified conditions. However, as we demonstrated in*

*this paper, there was enough light-absorbing DOM that is labile to photo-degradation even in the pool surface waters under all conditions that DOM photo-degradation was never limited by substrate (DOM supply). This means that the amount of water and DOM sequestered in the bottom waters does not influence the amount of DOM that can be degraded by light in this system.*

1 **Controls on dissolved organic matter (DOM)**  
2 **degradation in a headwater stream: the influence of**  
3 **photochemical and hydrological conditions in**  
4 **determining light-limitation or substrate-limitation of**  
5 **photo-degradation**

6

7 **R. M. Cory,<sup>1</sup> K.H. Harrold,<sup>1</sup> B. T. Neilson<sup>2</sup>, G.W. Kling<sup>3</sup>**

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14

15 **Abstract**

16 We investigated how absorption of sunlight by chromophoric dissolved organic matter (CDOM)  
17 controls the degradation and export of DOM from Imnavait Creek, a beaded stream in the  
18 Alaskan Arctic. We measured concentrations of dissolved organic carbon (DOC), as well as  
19 concentrations and characteristics of CDOM and fluorescent dissolved organic matter (FDOM),  
20 during ice-free periods of 2011-2012 in the pools of Imnavait Creek and in soil waters draining  
21 to the creek. Spatial and temporal patterns in CDOM and FDOM in Imnavait Creek were  
22 analyzed in conjunction with measures of DOM degradation by sunlight and bacteria and  
23 assessments of hydrologic residence times and in-situ UV exposure. CDOM was the dominant  
24 light attenuating constituent in the UV and visible portion of the solar spectrum, with high  
25 attenuation coefficients ranging from  $86 \pm 12 \text{ m}^{-1}$  at 305 nm to  $3 \pm 1 \text{ m}^{-1}$  in the  
26 photosynthetically active region (PAR). High rates of light absorption and thus light attenuation  
27 by CDOM contributed to thermal stratification in the majority of pools in Imnavait Creek under

1 low-flow conditions. In turn, thermal stratification increased the residence time of water and  
2 DOM, and resulted in a separation of water masses distinguished by contrasting UV exposure  
3 (i.e., UV attenuation by CDOM with depth resulted in bottom waters receiving less UV than  
4 surface waters). When the pools in Imnavait Creek were stratified, DOM in the pool bottom  
5 water closely resembled soil water DOM in character, while the concentration and character of  
6 DOM in surface water was reproduced by experimental photo-degradation of bottom water.  
7 These results, in combination with water column rates of DOM degradation by sunlight and  
8 bacteria, suggest that photo-degradation is the dominant process controlling DOM fate and  
9 export in Imnavait Creek. A conceptual model is presented showing how CDOM amount and  
10 lability interact with incident UV light and water residence time to determine whether photo-  
11 degradation is “light-limited” or “substrate-limited”. ~~We suggest that degradation, and thus~~  
12 ~~export, of DOM in CDOM-rich streams or ponds similar to Imnavait is typically light-limited~~  
13 ~~under most flow conditions.~~ We suggest that degradation of DOM in CDOM-rich streams or  
14 ponds similar to Imnavait is typically light-limited under most flow conditions. Thus, export of  
15 DOM from this stream will be less under conditions that increase the light available for DOM  
16 photo-degradation (i.e., low flows, sunny days).

17

## 18 **1 Introduction**

19 The decomposition of dissolved organic matter (DOM) to CO<sub>2</sub> and its subsequent  
20 transport to and release from surface waters is an important process in the carbon cycling of  
21 inland waters (e.g., Cole et al., 1994, 2007; Kling et al., 1991). This decomposition has been  
22 mainly attributed to bacterial respiration in the water column and sediments (e.g., Battin et al.,  
23 2009; Cole et al., 2007; Wetzel, 2001). Exposure to ultraviolet (UV) light is also a key control  
24 on the photochemical conversion of DOM to CO<sub>2</sub> in surface waters (e.g., Cory et al., 2007;  
25 Moran et al., 2000; Vähätalo and Wetzel, 2008), and coupled photochemical and microbial  
26 processing can enhance DOM degradation beyond the effect of bacteria or light alone (Cory et  
27 al., 2013; Judd et al., 2007; Tranvik and Bertilsson, 2001).

28 Recent work demonstrated that in shallow arctic lakes and streams the photo-degradation  
29 of DOM can greatly exceed bacterial respiration, accounting for up to 94% of the total DOM  
30 processed in the water column (Cory et al., 2014). The water column rate of DOM photo-

1 degradation to CO<sub>2</sub> (photo-mineralization) or to partially degraded DOM (e.g., photo-stimulated  
2 bacterial respiration; Cory et al., 2013) depends on (1) the amount of UV radiation from sunlight  
3 reaching the water surface, (2) the absorption of UV light by chromophoric DOM (CDOM), and  
4 (3) the apparent quantum yield, a term quantifying the lability of DOM as moles of product  
5 formed per moles of photons absorbed by DOM. Water column rates of DOM photo-  
6 degradation increase linearly with increasing UV light reaching the water surface, or with  
7 increasing photo-lability of DOM. However, the rate of DOM photo-degradation in the water  
8 column depends non-linearly on CDOM concentrations and depth due to attenuation of light  
9 mainly by CDOM with depth in the water column (Hu et al., 2002; Miller, 1998).

10 As CDOM concentrations increase, the depth of UV light penetration decreases to  
11 shallower depths, but the average rate of light absorption by CDOM increases in the water  
12 column (Hu et al., 2002). Thus, while the depth of UV light penetration is low, on the order of  
13 10 to 100 cm in the streams and small ponds in the Arctic characterized by high concentrations  
14 of CDOM (Cory et al., 2007, 2014; Gareis et al., 2010; Prairie et al., 2009; Watanabe et al.,  
15 2011), the rate of light absorption by CDOM may be high. If the light absorption rate is high  
16 enough, photo-degradation rates of DOM reach an asymptote such that increasing CDOM has no  
17 effect on photo-degradation integrated through the water column (Hu et al., 2002). At this point,  
18 where photo-degradation is insensitive to changing CDOM concentrations, the system is 'light  
19 limited' – in a light-limited system, as the amount of incident UV light increases so does the rate  
20 of photo-degradation. In contrast, in very clear waters light attenuation by CDOM is low and  
21 rates of DOM photo-degradation are limited by insufficient CDOM to absorb the available light  
22 – in these 'substrate-limited' systems, increasing the incident UV light has no effect but higher  
23 CDOM concentrations increase rates of photo-degradation. Thus, depending on the incident  
24 light available, the CDOM concentrations, and the depth of the water column, the rates of DOM  
25 degradation in surface waters may be either light-limited, substrate-limited, or co-limited by light  
26 and substrate. To our knowledge, the range of conditions and the interactions of controls on  
27 photo-degradation across the continuum of light- versus substrate-limitation have not been  
28 described or characterized.

29 At the scale of a stream reach, lake, or catchment, DOM degradation is related to both  
30 photochemical processing and the influence of hydrology on light exposure. Water residence  
31 times in a stream or river are generally a function of watershed and channel characteristics, but

1 may also be influenced by surface and subsurface transient storage (e.g., (Chapra and Runkel,  
2 1999; Neilson et al., 2010; Zarnetske et al., 2011) and thermal stratification that can isolate water  
3 masses (e.g., Merck and Neilson, 2012). While the influence of these factors on biogeochemical  
4 processes and solute concentrations has been studied (e.g., Boano and Harvey, 2014; Miller et  
5 al., 2009), the relative importance of CDOM concentration and lability, UV exposure, and water  
6 residence times on the degradation of DOM is unknown for stream or lake ecosystems.

7         Running waters, and especially lower-order streams, may be expected to have relatively  
8 low DOM degradation in the water column due to their high flow rates and short water residence  
9 times. Although small streams are often shallow and if unshaded by riparian vegetation may  
10 have light penetration to the bottom, the water residence time in any given reach is short and  
11 therefore there is little time for substantial photo-degradation of DOM. However, in areas of low  
12 relief, the headwater streams have longer residence times and greater light exposure through a  
13 shallow water column. In the Arctic, residence times within low gradient, first-order beaded  
14 streams are controlled by thermal stratification of the beads (pools) (Merck et al. 2012, Merck  
15 and Neilson 2012). Strong thermal stratification (up to 10 °C temperature difference within 0.5  
16 m depth) observed in Innavait Creek on the North Slope of Alaska was attributed to a  
17 combination of high concentrations of CDOM, low wind stress at the stream surface, underlying  
18 frozen soils, and low in-stream discharge (Merck et al. 2012). Because sunlight is rapidly  
19 attenuated in high-CDOM waters, warming by solar radiation is restricted to surface layers and  
20 can cause strong thermal stratification and density gradients (Fee et al., 1996; Houser, 2006;  
21 Kling, 1988). Merck et al. (2012) found that this stratification isolated the pool surface water  
22 from the bottom water and increased the water residence times in a reach from minutes under  
23 mixed conditions to hours or weeks when the pools were stratified. At the same time, there were  
24 distinct gradients in the concentrations of chromophoric and fluorescent fractions of dissolved  
25 organic matter (CDOM and FDOM, respectively) between pool surface and bottom waters  
26 (Merck et al. 2012). The authors suggested that these gradients in CDOM and FDOM were due  
27 to photo-degradation of DOM in the surface waters, and that stratification regulated the residence  
28 times of water and DOM and thus controlled the extent of DOM degradation in this stream.

29         To quantify the role of photo-degradation in producing observed DOM gradients in  
30 stratified stream pools, and to generally determine the influence of in-stream stratification, water  
31 residence times, and UV exposure on DOM degradation, we measured the lability and rates of

1 DOM degradation by sunlight and bacteria along with changes in CDOM and FDOM within the  
2 pools of Imnavait Creek in two summers with differing discharge and stratification patterns. We  
3 demonstrate that photo-degradation is the dominant process altering DOM chemistry and  
4 producing CO<sub>2</sub> in the water column of this headwater stream under all conditions, and we show  
5 how rates of photo-degradation are governed by the amount and lability of DOM (CDOM), light  
6 attenuation, patterns of stratification, and residence time. We suggest that in relatively shallow,  
7 high CDOM headwater streams, DOM photo-degradation is limited by available light instead of  
8 by available substrate (DOM) under a wide range of hydrological conditions.

## 9 **2 Methods**

### 10 **2.1 Site description**

11 Imnavait Creek is a headwater, beaded stream located on the North Slope of Alaska in a  
12 glacial valley formed during the Sagavanirktok glaciation in the Kuparuk River basin (68.616  
13 °N, 149.318 °W; (Detterman et al., 1958; Hamilton, 1986). The creek primarily lies in the  
14 organic soil layer and only occasionally cuts through to the mineral soil (McNamara et al., 1998).  
15 The connected pools, or beads, were formed by the erosion and melting of large ice deposits that  
16 had underlain the creek (McNamara et al., 1998; Walker et al., 1989) .

17 Previous studies of Imnavait Creek found that spring snowmelt accounts for 23 to 75 % of  
18 the watershed's annual water flux(Kane et al., 2004; McNamara et al., 2008) compared to 6 to 9  
19 % produced by the largest, single summer storm events (McNamara et al., 2008). Subsurface  
20 water paths are limited to the thawed active layer as the region is underlain with up to several  
21 hundred meters of permafrost, which effectively separates the active layer from any deep ground  
22 water (Osterkamp and Payne, 1981). Typical seasonally-thawed active layer depths at Imnavait  
23 ranged from 25 to 40 cm, occasionally extending to 100 cm (Hinzman et al., 1991). Water  
24 inputs from the riparian zone occur through both surface and diffuse subsurface flow(Kane et al.,  
25 2000). In addition to surface chutes that connect the stream pools, water travels between pools  
26 through the riparian zone with both subsurface flow through the active layer and surface flow  
27 during significant precipitation events (Merck and Neilson, 2012).

28 We studied a ~120 m reach of the creek consisting of a series of seven pools connected by  
29 short chutes (Fig. 1). Pools were named starting with pool 1 and proceeding downstream  
30 sequentially to pool 7. Across these 7 pools, surface areas ranged from 2 to 129 m<sup>2</sup>, volumes

1 ranged from 0.2 to 102 m<sup>3</sup>, and pool depths were between ~0.21 – 2 m. Along the reach of  
2 creek studied, we collected soil water from a water track that drains from the adjacent eastern  
3 hillslope into the pools. Seventeen sites were sampled along the water track from the hill top to  
4 the valley bottom along the water track with distances between sites ranging from 30 to 190 m.  
5 Soil water was also collected from an array of 55 sites within a 150 m by 90 m grid in a riparian  
6 zone on the eastern hillslope adjacent to the study pools in Imnavait Creek (Fig. 1).

## 7 **2.2 Water sample collection**

8 Water samples were collected from the surface and bottom of the seven pools monthly  
9 from 23 June through 4 August 2011, and weekly from 27 June through 18 August 2012. Pool  
10 water was collected from the surface and bottom of each pool through MasterFlex<sup>®</sup> tubing (Cole-  
11 Parmer, Vernon Hills, IL) using a peristaltic pump (GeoPump Inc., Medina, NY). Soil water  
12 was collected from the 17 sites in the water track flowing into the pools (Fig. 1), once in June  
13 and twice each in July and August in both 2011 and 2012, and from the riparian zone adjacent to  
14 the study pools monthly from June through August 2011. From the water track or riparian area,  
15 soil water was withdrawn using stainless-steel soil needles inserted into the soil, through  
16 MasterFlex<sup>®</sup> tubing, into plastic syringes. Temperature, conductivity, and pH were measured  
17 from pool and soil water at the time of collection using WTW meters (models 3210; Xylem,  
18 White Plains, NY). All pool and soil water samples were filtered in the field into high-density  
19 polyethylene bottles and kept cool and dark until analysis. Aliquots for analysis of DOM  
20 quantity and quality were filtered through pre-combusted Whatman GF/F glass fiber filters  
21 (Whatman, Clifton, NJ).

## 22 ***Sunlight attenuation***

23 Light attenuation with depth was measured in pools 1, 2, 3, and 6 on 27 June 2011 and in  
24 pools 1, 2, 3, 6 and 7 on 23 June 2012 using a compact optical profiling system for UV light in  
25 natural waters (UV C-OPS; Biospherical Instruments Inc., San Diego, CA) as previously  
26 described (Cory et al., 2013, 2014). The C-OPS measured downwelling cosine-corrected  
27 irradiance at 7 wavebands (305, 313, 320, 340, 380, 395, and 412 nm) and photosynthetically  
28 active radiation (PAR, 400-700 nm). Attenuation coefficients ( $K_{d,\lambda}$ ) were calculated from the  
29 downwelling irradiance ( $E_{\lambda}$ ) as a function of depth ( $z$ ) at each waveband:

1  $E_{\lambda,z} = E_{\lambda,0}e^{-K_d\lambda z}$  (1)

2 From multiple casts in each pool (n = 2 to 5), the coefficient of variation of  $K_{d,\lambda}$  ranged from 1 to  
3 3 % in the UV and 9 % for PAR. Means  $\pm$  standard error (SE) of  $K_d$  are reported.

#### 4 **2.3 In-situ monitoring**

5 Temperature sensor arrays (HOBO<sup>®</sup> Water Temp Pro v2; Onset Computer Corporation,  
6 Inc., Bourne, MA) were deployed vertically in each pool (n = 1 to 5 per pool in 2011 and n=3-25  
7 per pool in 2012) from late-June through mid-August, measuring at 5 minute intervals. The  
8 probes were wrapped with aluminum foil to prevent radiation-caused heating (Neilson et al.,  
9 2010) and placed starting 5 to 15 cm from the bottom of the pool and then at intervals ranging  
10 from 5 to 50 cm. Additional monitoring of pool 2 was conducted for one week in July 2011 and  
11 for most of July and part of August in 2012 where two sondes were deployed near the surface  
12 and bottom of the pool with oxygen, pH, specific conductance, and temperature probes (YSI  
13 6920 V2 sonde with ROX<sup>™</sup> optical dissolved oxygen, 6561 pH, 6560 conductivity, and 6560  
14 temperature sensors; YSI Inc., Yellow Springs, OH) measuring at 15 minute intervals. Finally,  
15 discharge data were collected at a weir further downstream to compare and contrast the flow  
16 variability between summer 2011 and 2012 (Kane and Hinzman, 2011; Kane, 2015).

#### 17 **2.4 Meteorological measurements**

18 Air temperature 1 m above the ground and precipitation were measured hourly at a  
19 meteorological station on the west-facing ridge of the Imnavait Creek basin approximately 1 km  
20 upstream of the study site using a temperature probe (model HMP45C; Campbell<sup>®</sup> Scientific,  
21 Logan, UT) and tipping bucket rain gauge, respectively (Kane and Hinzman, 2011). Global  
22 solar, UVA and UVB radiation were each measured at five minute intervals at Toolik Field  
23 Station (TFS, 11 km West of Imnavait at 68.616 °N, 149.318 °W) with pyranometers from Kipp  
24 & Zonen (CMP-6) and Yankee Environmental System, Inc. (UVB-1 and UVA-1), respectively.  
25 The global solar pyranometer measured a spectral range of 310 to 2800 nm, while UVB and  
26 UVA pyranometers measured 280-320 nm and 320-400 nm, respectively. Photosynthetically  
27 active radiation (PAR; 400-700 nm) was measured hourly using a quantum sensor by Li-Cor (LI-  
28 190S) at the same location.

## 1 2.5 DOM quantity and quality

2 Samples for dissolved organic carbon (DOC) concentration were acidified with trace-metal  
3 grade hydrochloric acid to approximately pH 3 after filtration through Whatman GF/F filters and  
4 stored in the dark at 4 °C until analysis using a high-temperature platinum-catalyzed combustion  
5 followed by infrared detection of CO<sub>2</sub> (Shimadzu TOC-V; Shimadzu, Columbia, MD).

6 The chromophoric and fluorescent fractions of DOM (CDOM and FDOM, respectively)  
7 were analyzed within hours to at most several days of collection. Samples were stored in the  
8 dark at 4 °C until warmed to room temperature (20 to 25 °C) just prior to analysis. UV-Vis  
9 absorbance spectra of CDOM were collected using 1-cm path length quartz cuvettes with a  
10 spectrophotometer (USB 2000+UV-VIS; Ocean Optics, Inc., Dunedin, FL or Aqualog; Horiba  
11 Scientific). Sample absorption was measured against laboratory-grade deionized (DI) water  
12 blanks (Barnstead E-Pure and B-Pure; Barnstead Thermolyne, Dubuque, IA). The spectral slope  
13 ratio ( $S_R$ ) was calculated from the absorbance spectrum of each sample as the ratio of the slope  
14 from 275 to 295 nm to the slope from 350 to 400 nm (Helms et al., 2008). CDOM absorption  
15 coefficients ( $a_{CDOM,\lambda}$ ) were calculated as follows:

$$16 \quad a_{CDOM,\lambda} = \frac{A_\lambda}{l} 2.303 \quad (2)$$

17 where  $A$  is the absorbance reading at wavelength  $\lambda$  and  $l$  is the pathlength in meters.  
18  $SUVA_{254}$  was calculated as absorbance at 254 nm divided by the cuvette pathlength (m) and then  
19 divided by the DOC concentration (mg C L<sup>-1</sup>; Weishaar et al., 2003).

20 Excitation-emission matrices (EEMs) were measured on all water samples with a  
21 Fluoromax-4 fluorometer or an Aqualog (Horiba Scientific, Edison, NJ) following previously  
22 described procedures (Cory et al., 2010). An aliquot of sample was placed in the 1-cm quartz  
23 cuvette for each EEM and diluted with DI if necessary to bring  $A_{254} < 0.6$ . EEMs were corrected  
24 for inner-filter effects and for instrument-specific excitation and emission corrections in Matlab  
25 (version 7.7) following Cory et al. (Cory et al., 2010). The fluorescence index (McKnight et al.,  
26 2001) was calculated from each corrected EEM as the ratio of emission intensity at 470 nm over  
27 the emission intensity at 520 nm at an excitation wavelength of 370 nm (Cory et al., 2010).  
28 Emission intensity at FDOM peaks A, C, and T was evaluated at excitation/emission pairs  
29 250/450, 350/450, 275/340 (nm/nm), respectively, in Raman Units (RU; Stedmon et al., 2003).

## 1 2.6 DOM degradation

2 Water collected from Innavait Creek in amber HDPE bottles in the field was used for  
3 photochemical and bacterial degradation experiments, as described in Cory et al. (2014).  
4 Briefly, dark bacterial respiration was measured from whole water samples incubated for five to  
5 seven days in the dark at 6-7 °C alongside killed controls (1% HgCl<sub>2</sub>) in air-tight, pre-combusted  
6 12-mL borosilicate exetainer vials (Labco, Inc). For DOM photo-degradation, GF/F filtered  
7 water was placed in air-tight, pre-combusted 12-mL borosilicate exetainer vials and exposed to  
8 natural sunlight at Toolik Field Station for ~12 hours alongside foil-wrapped dark controls at  
9 temperatures ranging from 10-16 °C. Bacterial re-growth experiments for photo-stimulated  
10 bacterial respiration were conducted as described in Cory et al. (2013, 2014). There were four  
11 independent replicates from each water sample for every analysis type and treatment. Membrane  
12 inlet mass spectrometry (MIMS) was used to measure bacterial or photochemical oxygen  
13 consumption relative to killed or dark controls, respectively. Bacterial or photochemical  
14 production of CO<sub>2</sub> during complete oxidation (mineralization) of DOM was quantified as  
15 production of dissolved inorganic carbon (DIC) relative to killed or dark controls, respectively,  
16 using a DIC analyzer (model AS-C3, Apollo SciTech, Inc.). After exposure to sunlight or  
17 bacteria, subsamples were analyzed for CDOM and FDOM as described above. Changes in  
18 DOM quality are reported as mean ± standard error (SE) of the four replications of each  
19 treatment.

20 We used previously reported lability and rates of DOM degradation by bacteria and  
21 sunlight in Innavait Creek, measured from experiments described above, during our study  
22 periods in 2011 and 2012 to determine the sensitivity of DOM degradation in Innavait Creek to  
23 photochemical and hydrological factors. The conversion of experimental measures of DOM  
24 degradation to water column rates of degradation is described for Innavait Creek and other  
25 waters in the Arctic in Cory et al. (2013, 2014). In the current study, we quantified how rates of  
26 DOM photo-degradation in the water column varied with available light, CDOM concentrations,  
27 and lability of DOM measured in Innavait Creek during the 2011-2012 summer seasons.

28 The integrated, water-column rate of DOM photo-degradation is:

29 
$$\text{Photo-degradation (mol C m}^{-2} \text{ d}^{-1}) = \int_{\lambda_{\min}}^{\lambda_{\max}} \phi_{\lambda} Q_{dso,\lambda} (1 - e^{-K_d \lambda z}) \frac{a_{CDOM,\lambda}}{a_{tot,\lambda}} d\lambda \quad (3),$$

1 where  $\lambda_{\min}$  and  $\lambda_{\max}$  are the minimum and maximum wavelengths of light contributing to the  
2 photo-degradation of DOC (280 and 700 nm, respectively).  $\Phi_{\lambda}$  is the apparent quantum yield  
3 spectrum for photo-degradation of DOM (mol product mol<sup>-1</sup> photons absorbed nm<sup>-1</sup>; a measure  
4 of DOM lability to photo-degradation which decreases exponentially with increasing  
5 wavelength). We used previously reported spectra of DOM lability ( $\Phi_{\lambda}$ ) in Innavait Creek for  
6 photo-mineralization DOC to CO<sub>2</sub> and photo-stimulated bacterial respiration (Cory et al. 2014).  
7  $Q_{dso,\lambda}$  is the spectrum of the UV from sunlight that reaches the water surface (accounting for  
8 reflection; Cory et al. 2014).  $Q_{dso,\lambda}$  varies by location (latitude/longitude), time of day, date, and  
9 sunny vs. cloudy sky conditions, as described in Cory et al. 2014.  $K_{d,\lambda}$  is the attenuation  
10 coefficient with depth (Eqn. 1).  $Q_{dso,\lambda}$  is the spectrum of UV light just below the water surface  
11 (accounting for reflection; Cory et al. 2014), and  $K_{d,\lambda}$  is the attenuation coefficient with depth  
12 (Eqn. 1).  $a_{CDOM,\lambda}$  is the concentration of CDOM measured as described above (Eqn. 2).  
13  $a_{CDOM,\lambda}/a_{tot,\lambda}$  is the spectrum of the ratio of absorption by CDOM to the total absorption (where  
14  $a_{tot,\lambda}$  is the total absorption in the water column due to CDOM, particles, and water). The ratio  
15 of  $a_{CDOM,\lambda}/a_{tot,\lambda}$  was assumed to be 1 at all wavelengths (i.e., CDOM was the main UV-  
16 absorbing constituent in Innavait Creek; Cory et al. 2014).

17 From Eqn. 3, it follows that as DOM lability to photo-degradation ( $\Phi_{\lambda}$ ) or incoming UV  
18 light ( $Q_{dso,\lambda}$ ) increase, the depth-integrated or areal water-column rate of DOM photo-  
19 degradation increases. In addition, photo-degradation of DOM throughout the water column  
20 increases with increasing  $K_{d,\lambda}$  up to a point where at high  $K_{d,\lambda}$  the relationship is asymptotic (Fig.  
21 S1). This is because while increasing CDOM provides more DOM to absorb light and photo-  
22 degrade, the absorption of UV light by CDOM also controls light attenuation (i.e.,  $a_{CDOM,\lambda} \approx K_{d,\lambda}$   
23 ; presented in the *Results* section below). Thus as light attenuation ( $K_{d,\lambda}$ ) increases, at some  
24 point light becomes limiting and adding more CDOM (increasing  $K_d$ ) results in no change in the  
25 integrated water column rate of photo-degradation (Fig. S1). In this study, we used the range of  
26 terms in Eqn. 3 observed in Innavait Creek (Fig. S2) to develop a conceptual model of controls  
27 on DOM photo-degradation in this and similar systems. For example, we used the range of  
28  $Q_{dso,\lambda}$  spectra for Innavait Creek representing the average, minimum and maximum UV light  
29 reaching the surface of Innavait Creek over the course of the day during the study period, as  
30 well as the average, minimum and maximum of  $a_{CDOM,\lambda}$  observed in Innavait Creek.

### 31 **3 Results**

### 1 3.1 Meteorological Conditions

2 There was no difference in mean air temperatures between the mid-June through mid-  
3 August study periods in 2011 vs. 2012 at Imnavait Creek. However, June-August was generally  
4 sunnier and drier in 2011 compared to 2012 (Table 1). The total global solar radiation (310-2800  
5 nm) and total UV and visible photon flux (280-700 nm) were 17 % and 24 % higher,  
6 respectively, in 2011 compared to 2012 (Table 1, Fig. S3). Precipitation was three times greater  
7 during the summer 2012 compared with 2011, and the lower precipitation in 2011 resulted in  
8 significantly lower volume of water passing the weir in 2011 versus 2012 (Table 1, Fig. S1).

### 9 3.2 Pool stratification and separation of water masses

10 Light attenuation coefficients ( $K_{d,\lambda}$ ) in Imnavait Creek decreased exponentially with  
11 increasing wavelength from  $88 \pm 12 \text{ m}^{-1}$  at 305 nm to  $17 \pm 3 \text{ m}^{-1}$  at 412 nm to  $3 \pm 1 \text{ m}^{-1}$  in the  
12 photosynthetically active region (PAR). In most pools, there was no significant difference  
13 between in-situ  $K_{d,\lambda}$  values and CDOM absorption coefficients ( $a_{\text{CDOM},\lambda}$ ) collected from filtered  
14 water from the same pool at the time  $K_{d,\lambda}$  was measured (as shown for 320 nm and 412 nm in  
15 Fig. 2). These results demonstrate that in most pools CDOM was the dominant light absorbing  
16 constituent in the water column (Fig. 2), consistent with low particulate matter concentrations in  
17 this stream and previous work showing that CDOM dominates UV light absorption in these  
18 streams (Cory et al. 2014). However, in two pools sampled in June 2011, the in-situ  $K_{d,\lambda}$  values  
19 at 320 nm were significantly higher than the corresponding CDOM absorption coefficients  
20 measured from filtered water (Fig. 2). This was likely due to the inherent challenges deploying  
21 an instrument to quantify  $K_{d,\lambda}$  in the UVB range in high CDOM waters where 99 % of light at  
22 320 nm is attenuated by ~ 8 cm (based on the mean  $K_{d,\lambda}$  or CDOM coefficients in Fig. 2). This  
23 could be due to inadvertent mixing of sediment from the bottom during the instrument casts to  
24 measure light attenuation, or to general difficulties quantifying  $K_{d,\lambda}$  in the UVB range in high  
25 CDOM waters, given that 99 % of light at 320 nm is attenuated by ~ 8 cm (based on the mean  
26  $K_{d,\lambda}$  or CDOM coefficients in Fig. 2). In addition, co-precipitation of iron and DOM during  
27 filtration may have caused these differences between in situ  $K_{d,\lambda}$  vs. CDOM coefficients given  
28 the relatively high concentrations of iron in Imnavait Creek, ranging from 4 to 21  $\mu\text{M}$  total iron  
29 in the surface waters with most iron present as ferrous iron (Page et al., 2013, 2014).

1           Five of the seven pools (2, 3, 5, 6, 7) were repeatedly thermally stratified with a nearly 10  
2 °C temperature difference between top and bottom waters during sunny and dry (low-flow)  
3 conditions in Imnavait Creek (Figs. 3 and 4). In contrast, pools 1 and 4 did not exhibit  
4 stratification in 2011 or 2012. During the sunnier and drier summer of 2011, pools 2, 3, 5, 6, 7  
5 were stratified on 43 to 46 out of 50 days measured (Fig. 3), while during the wetter summer of  
6 2012, these pools stratified only 11 out of 49 days measured (Fig. 4). Within each summer, the  
7 roles of solar radiation and precipitation were evident in the frequency and extent of stratification  
8 in each pool. For example, pools 2, 3, 5, 6, and 7 showed the greatest extent of thermal  
9 stratification once the discharge from snowmelt had receded, coinciding with the period when  
10 solar irradiance was highest (e.g., late June in 2011 and 2012; Figs. 3, 4). In addition, during  
11 both summers a portion of each stratified pool mixed nightly due to surface heat loss, followed  
12 by re-stratification with increasing solar irradiance during the day (Merck and Neilson, 2012).  
13 Following substantial precipitation events, stream flow increased and caused stratified pools to  
14 mix completely within hours of precipitation, as demonstrated for example after a rain even on  
15 17 July 2011 (Fig. 3). After mixing, pools stratified again within four to five days (Fig. 3).

16           Under stratified conditions, water and DOM in the pools experienced contrasting UV  
17 exposure. For example, Pool 3 had a depth of 1.4 m from the water surface to the sediment, and  
18 on average the depth of the surface mixing layer was 50 cm (ranging from 20 – 50 cm below the  
19 water surface; Fig. 3). It follows that DOM below 50 cm was in the “bottom water”, defined as  
20 water trapped below the diel mixing depth during stratified conditions. The depth of UVB and  
21 UVA light penetration was always less than 50 cm in Imnavait Creek. For example, using the  
22 mean CDOM absorption coefficients at 320 nm and 412 nm (Fig. 2), 99 % of all incoming UVB  
23 and UVA light was absorbed within  $8 \pm 1$  cm and  $35 \pm 3$  cm, respectively (average  $\pm$  SE;  
24 maximum depth of UVA penetration observed was 45 cm based on attenuation coefficients at  
25 412 nm in Fig. 2). Therefore, DOM in the surface of the pools experienced UV light exposure  
26 each day while DOM trapped in pool bottom waters was protected from UV light during  
27 stratified conditions.

### 28 **3.3 Stream and soil water chemistry**

29           Both the pool water and the soil water draining into Imnavait Creek had low pH, low  
30 conductivity, and high concentrations of DOC, CDOM, and FDOM (Table 2). The pH ranged

1 from  $5.2 \pm 0.1$  to  $5.7 \pm 0.1$  and the conductivity ranged from  $25 \pm 10$  to  $12 \pm 2 \mu\text{S cm}^{-1}$  in soil  
2 and pool water, respectively. DOC concentrations ranged from  $1412 \pm 78$  and  $785 \pm 13 \mu\text{M C}$  in  
3 pool and soil water, respectively, during summer of 2012 (Table 2). CDOM and FDOM proxies  
4 for the chemical composition of soil and pool water DOM were consistent with a terrestrial  
5 source of DOM, i.e., high molecular weight, aromatic compounds derived from degradation of  
6 plant and soil organic matter. For example, in the soil waters draining to Innavait Creek, the  
7 spectral  $S_R$ , a proxy for average molecular weight of DOM (Helms et al. 2008) was  $0.75 \pm 0.08$ ,  
8 and the specific UV absorbance at 254 nm ( $\text{SUVA}_{254}$ ), strongly correlated with aromatic C  
9 content, was  $4.4 \pm 0.1 \text{ L mg C}^{-1} \text{ m}^{-1}$ . The fluorescence index, a proxy for DOM source and  
10 aromatic C content, was  $1.59 \pm 0.07$ . Although there were some significant differences in mean  
11 values between soil and pool water DOC, CDOM and FDOM (discussed below), similar ranges  
12 of DOC, CDOM and FDOM were observed for the pool bottom waters in Innavait Creek and  
13 for the soil water draining into the pool bottoms (Table 2).

14 There were no significant differences in average pH, conductivity, DOC, CDOM, or  
15 FDOM concentrations in the soil waters between 2011 and 2012 (Table 2). There was no  
16 difference in average optical character of soil water DOM between summers except for  
17  $\text{SUVA}_{254}$ , which was significantly lower in the soil waters in 2012 compared to 2011 (Table 2).  
18 In contrast, there were significant differences in DOM quality between 2011 and 2012 when  
19 comparing pool bottom or surface water across years (Table 2), likely due to differences in the  
20 extent of stratification between years.

21 When the pools in Innavait Creek were stratified, there were significant differences in  
22 water chemistry between pool surface and bottom water. In-situ data collected under stratified  
23 conditions in Pool 2 from 8 - 15 July 2011 showed significantly higher dissolved oxygen in the  
24 surface compared to the bottom pool water (Fig. 3). Discrete water samples collected when  
25 pools were stratified in 2011 and 2012 showed significantly higher concentrations of DOC,  
26 CDOM, and FDOM in bottom waters compared to surface waters (as shown in Fig. 5 for pools  
27 sampled on 14 July 2011). The surface waters of most pools sampled under stratified conditions  
28 had different DOM quality compared to bottom waters, as indicated by significantly lower  
29  $\text{SUVA}_{254}$ , higher  $S_R$ , and lower FI (Table 3; Fig. 5). When the stream pools stratified, DOM in  
30 pool bottom water was not significantly different than soil water for most CDOM and FDOM  
31 measures or DOM quantity and quality (Table 3). In contrast, when the pools were mixed, there

1 were no significant differences in DOC, CDOM, and FDOM between pool surface and bottom  
2 waters (e.g., as shown in Fig. 5 for pools sampled on 21 July 2012).

### 3 **3.4 Photochemical degradation of DOM**

4 Previous work showed that photo-mineralization of DOM to CO<sub>2</sub> accounted for the  
5 majority of DOM degradation in Imnavait Creek ( $24.69 \pm 18.28$  mmol C m<sup>-2</sup> d<sup>-1</sup>, mean  $\pm$  SE;  
6 Cory et al. 2014). Here we show that in addition to mineralization of DOM, exposure of  
7 Imnavait Creek DOM to ~12 hours of sunlight altered the chemical quality of the remaining  
8 DOM, likely due to preferential mineralization of the aromatic fraction (Cory et al., 2007;  
9 Stubbins et al., 2010) and to partial photo-oxidation of the DOM (Cory et al., 2013, 2014).  
10 These photochemical alterations of DOM resulted in a significant loss of CDOM and FDOM at  
11 each wavelength compared to dark controls ( $12.1 \pm 1.4$  % to  $27.2 \pm 2.1$  % less CDOM or FDOM  
12 compared to the dark control depending on CDOM wavelength or FDOM peak, Table 4).  
13 Because there was greater loss of CDOM at long wavelengths compared to shorter wavelengths,  
14 photo-degradation significantly increased the S<sub>R</sub> by  $15.8 \pm 1.3$  % and decreased the FI by  $-11.7 \pm$   
15  $0.8$  % on average. There was an increase in peak T intensity by  $4.1 \pm 1.5$  % for photo-exposed  
16 DOM compared to dark controls (Table 4).

17 Photo-degradation of DOM enhanced the respiration of bacteria fed the photo-exposed  
18 DOM (compared to DOM kept in the dark); the water column rate of photo-stimulated bacterial  
19 respiration was  $3.04 \pm 1.31$  mmol C m<sup>-2</sup> d<sup>-1</sup> (Cory et al. 2014). Relative to the initial photo-  
20 exposed DOM, bacterial incubation generally increased CDOM and FDOM (from  $3 \pm 1$  to  $15 \pm 1$   
21 %, Table 4). The exception was that following bacterial degradation of the photo-exposed  
22 DOM, there was a  $12 \pm 1$  % loss of peak T fluorescence (Table 4). Coupled photo and bacterial  
23 degradation increased the fluorescence index by  $5 \pm 1$  % (Table 4).

### 24 **3.5 Bacterial degradation of DOM**

25 The average areal rate of dark bacterial respiration of DOM integrated over the mean  
26 depth (0.5 m) in Imnavait Creek was  $2.35 \pm 0.34$  mmol O<sub>2</sub> m<sup>-2</sup> d<sup>-1</sup>. Bacterial degradation  
27 resulted in significant loss of CDOM and FDOM compared to the killed control over the six day  
28 incubation period at 6 - 7 °C ( $-2.9 \pm 1.6$  % to  $-7.6 \pm 2.4$  %, Table 4). There was no detectable

1 change in the  $S_R$  or the FI after dark bacterial degradation of DOM compared to the killed  
2 control (Table 4).

## 3 **4 Discussion**

### 4 **4.1 Stratification in beaded streams**

5 Stratification is likely widespread during the summer in beaded pools and small ponds  
6 across the Arctic. Stratification in tundra ponds is well documented (Boano and Harvey, 2014;  
7 Hobbie, 1980), as are the factors conducive to stratification including high light attenuation by  
8 CDOM (Cory et al., 2007, 2014; Gareis et al., 2010; Watanabe et al., 2011), adequate solar  
9 radiation, and in streams low wind stress at the surface and low enough discharge coupled with  
10 permafrost below the stream (Merck and Neilson, 2012; Merck et al., 2012; this study). For  
11 example, nearly one third of both lower-order streams and coastal plain lakes sampled in the  
12 Alaskan Arctic had average CDOM absorption coefficients at 305 nm greater than or equal to the  
13 absorption coefficients observed in the surface of Imnavait Creek (Cory et al., 2014), consistent  
14 with high CDOM absorption coefficients reported in streams and small ponds and lakes across  
15 the Arctic (Gareis et al., 2010; Watanabe et al., 2011). While fewer studies have reported in-situ  
16  $K_{d,\lambda}$  values compared to reports of CDOM absorption coefficients in arctic freshwaters, strong  
17 agreement between  $K_{d,\lambda}$  and  $a_{CDOM,\lambda}$  in this and other studies (Fig. 2; Cory et al., 2014; Gareis et  
18 al., 2010; MORRIS et al., 1995) demonstrate that CDOM was the main UV and visible (PAR)  
19 light absorbing constituent in surface waters across the Arctic. Because UV and PAR account  
20 for approximately 51 % of the energy within the shortwave radiation portion of the spectrum  
21 (300 - 2500 nm), absorption of sunlight by CDOM contributes to the frequency and extent of  
22 stratification by restricting warming to the surface layers (Merck and Neilson, 2012).

23 Given that there was no significant difference in the average CDOM absorption  
24 coefficients in pool surface waters between 2011 and 2012 (Table 3), or between pool bottom  
25 water temperatures or in wind stress (Table 1), differences in the extent and frequency of  
26 stratification in the pools between years were most likely due to differences in discharge (Table  
27 1; Fig. S1). Low in-stream discharges common in 2011 were due to significantly less  
28 precipitation than in 2012. Low discharge led to low turbulence, allowing for more common  
29 thermal stratification in the pools throughout a large fraction of the open water season (Table 1,  
30 Fig. 3). The higher flows during summer 2012 resulted in approximately five-fold greater stream

1 volume compared to 2011, which increased the turbulence that induced frequent mixing  
2 throughout the water column in the pools (Table 1; Fig. 4).

### 3 **4.2 DOM composition and photo-degradation**

4 There are three lines of evidence in support of photo-degradation as the main control on  
5 the observed differences in DOM quantity and composition between pool surface and bottom  
6 waters in Imnavait Creek under stratified conditions: (1) water column rates of photo-  
7 mineralization of DOM and photo-stimulated bacteria respiration each exceeded dark bacterial  
8 respiration (Cory et al. 2014; Table 4), (2) experimental photo-degradation of Imnavait DOM  
9 closely reproduced the depth differences in DOM concentration and composition (Table 4), and  
10 (3) depth differences in DOM concentration and composition were consistent with effects of  
11 sunlight on DOM observed in other studies.

12 The mean water column rate of photo-mineralization of DOM to CO<sub>2</sub> was about ten times  
13 faster than photo-stimulated bacterial respiration or dark bacterial respiration in Imnavait Creek  
14 (Cory et al. 2014, Table 4). Thus, although 99 % of all UVB and UVA light causing photo-  
15 mineralization of DOM was attenuated within 8 to 35 cm below the pool surface, respectively  
16 (Fig. 2), thereby confining photo-degradation of DOM to the top ~ 35 cm of the pool, photo-  
17 mineralization in this surface layer was fast enough to exceed rates of bacterial respiration  
18 occurring throughout the UV-exposed and UV-protected portions of the water column (Table 4).

19 The second line of evidence in support of photo-degradation as the dominant process  
20 creating differences in DOM character between surface and bottom waters was that photo-  
21 degradation experiments reproduced the magnitude and direction of depth differences in DOC,  
22 CDOM, and FDOM observed under stratified conditions (Tables 2 and 4, Fig. 5). Compared to  
23 pool bottom waters, surface waters had significantly lower CDOM and FDOM concentrations,  
24 higher S<sub>R</sub>, and lower FI, by 9 to 55 % (Tables 2 and 4, Fig. 5). Exposure of bottom water to ~12  
25 hours of natural sunlight resulted in a comparable loss of CDOM and FDOM, and similarly  
26 higher S<sub>R</sub>, and lower FI (11 to 27 % loss or change in CDOM or FDOM compared to dark  
27 controls; Table 4). Thus, DOM in pool surface waters and in photo-exposed bottom waters had  
28 lower concentrations of aromatic DOM (i.e., CDOM and FDOM) with lower average molecular  
29 weight compared to DOM protected from UV in the bottom waters. Many studies have  
30 demonstrated that photo-degradation of DOM results in a loss of CDOM and FDOM Granéli et

1 al., 1996), a decrease in aromaticity (Brooks et al., 2007; Stubbins et al., 2010), and a decrease in  
2 average molecular weight (e.g.,  $S_R$ , Cory et al., 2007; Helms et al., 2008; Spencer et al., 2010),  
3 just as we observed.

4 It is unlikely that bacterial processing of DOM could produce similar changes to those  
5 observed between surface and bottom waters or to the results of photo-degradation experiments.  
6 Bacteria degrade stream DOM and decrease CDOM and FDOM (Cory and Kaplan, 2012), but, at  
7 the same time some bacterial processes can regenerate CDOM and FDOM (e.g., Amado et al.,  
8 2006; Moran et al., 2000), leading to a net balance between degradation and regeneration. In our  
9 experiments the dark bacterial degradation of DOM resulted in a net loss of CDOM and FDOM,  
10 but the loss was much lower compared to photo-degradation (e.g., ~ 2 to 8 % decrease over six  
11 days, Table 4). Furthermore, bacterial degradation had no detectable effect on the  $S_R$  or FI, and  
12 thus cannot explain the significant differences in  $S_R$  and FI between surface and bottom waters  
13 in Imnavait Creek (Tables 2, 3 and 4, Fig. 5).

14 In an earlier study of Imnavait Creek, Merck et al. (2012) found that soil water FDOM  
15 was chemically similar to pool bottom water FDOM. Cold soil water from subsurface lateral  
16 flows plunged into the pool bottom during stratified conditions, bringing in FDOM that remained  
17 relatively unchanged in composition from the FDOM in soil waters. In our study we show that,  
18 similarly, FDOM but also pH, conductivity, DOC, and CDOM are comparable in quantity or  
19 quality between soil waters and pool bottom waters (Tables 2 and 3). These results suggest that  
20 either DOM in soil waters draining into isolated pool bottoms experiences little degradation in  
21 the pool, as would be expected based on relatively slow rates of bacterial respiration in cold,  
22 acidic, low nutrient, and often anoxic water (Table 4, Fig. 3-4), or that the pathways, rates, and  
23 fate of DOM degradation in both soils and bottom waters are similar.

#### 24 **4.3 Synthesis of factors controlling DOM degradation**

25 Given that photochemical processes dominate the degradation and alteration of DOM in  
26 Imnavait Creek, estimates of DOM degradation or export at a stream reach or whole catchment  
27 scale must be based on an integration of the photochemical and hydrological controls on DOM  
28 degradation. These controls include (1) the amount of surface UV available to be absorbed by  
29 CDOM ( $Q_{dso,\lambda}$ , Eqn. 3), (2) the amount of CDOM to absorb and attenuate UV light in the water  
30 column ( $a_{CDOM,\lambda}$  and  $K_d$ ; Eqn. 3), (3) the lability of DOM to photo-degradation, quantified as

1 apparent quantum yields ( $\Phi_\lambda$ ; *Eqn. 3*), and (4) the residence time of DOM in the water column or  
2 in a stream reach as affected by flow rates and stratification that in turn control the total UV light  
3 exposure and amount of DOM photo-degradation. These main controls and their feedbacks are  
4 summarized in Fig. 6, which shows that as light exposure increases, DOM photo-degradation  
5 increases. Similarly, there are different pathways through which increased CDOM can affect  
6 light exposure. First, higher CDOM increases light attenuation and thus helps facilitate thermal  
7 stratification. Stratification in turn results in increased water residence times that can increase  
8 the opportunity for light exposure and photo-degradation. For example, in pool surface waters  
9 where light exposure is greatest, during the day under low-flow conditions nearly the entire  
10 upper layer is stratified (Fig. 3), which increases residence time and light exposure (right side of  
11 Fig. 6). Second, although increasing CDOM increases light attenuation, the effect on light  
12 exposure can vary (left side of Fig. 6). In general, if  $K_d$  is relatively low and light penetrates to  
13 the bottom of the water column, increasing CDOM will result in greater total light exposure of  
14 DOM in the system, and thus greater total light absorption by CDOM to drive photochemical  
15 reactions. However, if  $K_d$  is relatively high and light is extinguished well before it reaches the  
16 bottom of the water column, the system is light-limited and adding more CDOM will not  
17 increase the overall light exposure of DOM (or light absorption by CDOM). The relative  
18 importance of these scenarios can be estimated by first examining the sensitivity of various  
19 components in the photo-degradation model (*Eqn. 3*).

20 To investigate the sensitivity of DOM photo-degradation to the amount of surface UV,  
21 CDOM, or  $\Phi_\lambda$ , we varied each term independently in the equation for the water column rate of  
22 DOM photo-mineralization (*Eqn. 3*), using the average, minimum, and maximum values  
23 observed in Imnavait Creek (Fig. S2). Holding surface UV and  $\Phi_\lambda$  constant (using the average  
24 observed values) and varying CDOM across the range observed in the pool surface waters of  
25 Imnavait Creek (39 to 63  $\text{m}^{-1}$  at 305 nm, mean of  $53 \pm 2$ ; Table 2), there was little variation in  
26 water column rates of photo-mineralization (Fig. 7). This result indicates that photo-degradation  
27 of DOM in Imnavait Creek is represented by higher  $K_d$  and falls in the asymptotic range shown  
28 in Fig. S1, meaning it is limited by insufficient UV light. It follows that increasing UV light  
29 (while holding CDOM and  $\Phi_\lambda$  constant), should significantly increase the rate of DOM photo-  
30 mineralization, as shown in Fig. 7. That is, the daily total surface UV light varied by nearly 10-  
31 fold over the course of the summer season due to differences in solar zenith angle or cloud cover

1 (as shown for 2012 in Fig. S3), and thus there was a nearly 10-fold higher rate of photo-  
2 mineralization when using the maximum vs. minimum surface UV light available in *Eqn. 3* (Fig.  
3 7). The greatest effect on the rate of photo-degradation occurred when holding the UV light and  
4 CDOM to their average values, and varying the lability of DOM to photo-mineralization (i.e.,  
5 varying  $\Phi_\lambda$ ). In this case, with a 6 to 20-fold range in  $\Phi_\lambda$  (depending on wavelength; Fig. S2),  
6 DOM was converted to CO<sub>2</sub> by sunlight ~ 13-fold faster using the maximum vs. the minimum  
7 observed  $\Phi_\lambda$  (Fig. 7).

8         The second step in understanding photo-degradation for any system is to consider the  
9 integrated effects of light attenuation with flow rates, stratification, and anticipated residence  
10 time distributions. Integrating the photochemical and hydrological factors produces a continuum  
11 of conditions that can represent or classify any particular system (Fig. 8a,b). For example, as  
12 light exposure and residence times increase, the amount of DOM lost through photo-  
13 mineralization increases (Fig. 8a). However, the quantity and photo-lability (*quality*,  $\Phi_\lambda$ ) of  
14 CDOM both determine the rates and the total amount of DOM photo-degradation. In systems  
15 with high CDOM concentrations or low light exposure, photochemical processes can be light  
16 limited rather than substrate (CDOM) limited. In such systems waters are rarely photo-bleached  
17 clear before inputs from soil waters or sediments replenish the CDOM lost to photo-degradation.  
18 This is the situation in Imnavait Creek (e.g., Table 3; Fig. 8a, left side;), which applies to most  
19 arctic headwater catchments or any system that receives high inputs of organic matter (Koehler  
20 et al., 2014). Similarly, if the photo-lability (and thus the  $\Phi_\lambda$ ) of the DOM is low (the lower,  
21 dashed line on Fig. 8a), then more UV is required for photo-degradation, and the system would  
22 be again considered light-limited. This is likely the case in the lower-CDOM Kuparuk River  
23 (spectral characteristics described in Cory et al. 2014), a fourth-order stream where it is joined by  
24 Imnavait Creek, although under some situations this river may be co-limited by light and  
25 substrate. Most of the waters in the Alaskan Arctic would fall in between these examples on this  
26 conceptual diagram, based on CDOM concentrations and  $\Phi_\lambda$  values (Cory et al. 2014). In these  
27 cases where DOM photo-degradation is light-limited, the amount of time the DOM is exposed to  
28 UV becomes more important than the mass of DOM exposed. On the other hand, if photo-  
29 lability is very high and CDOM concentrations are very low, then the system is substrate-limited  
30 and the total mass of DOM exposed is more important than the amount of time the DOM spends  
31 exposed to UV (Fig. 8a, right side). This is because when the system is substrate limited, even a

1 short exposure to UV will result in rapid and substantial photo-degradation, and exposing greater  
2 amounts of DOM even over short residence times will increase the overall photochemical  
3 processing in the system. Similarly, as one moves further right on Fig. 8a, the DOM loss as a  
4 percentage of initial amount declines once the system has switched from light- to substrate-  
5 limitation.

6 Finally, the nature and controls on DOM photo-degradation of a river reach (or whole  
7 system) can be expressed as a function of light attenuation and residence times (Fig. 8b). When  
8 water is flowing quickly through a stream the residence times are very short and the water  
9 column is well mixed, DOM spends less time exposed to UV light, and even at medium to low  
10 values of CDOM and  $K_d$  the system is light-limited (Fig. 8a,b left) and DOM loss to photo-  
11 mineralization is low (Fig. 8a left). At the other extreme, low flow conditions create long  
12 residence times, and even at medium values of CDOM the system is substrate-limited (Fig. 8b,  
13 right) and again DOM photo-mineralization may become low (Fig. 8a, right).

14 ~~The relationship between residence time and photochemical controls is plotted for~~  
15 ~~measured values in Imnavait Creek in Fig. 9. Combinations of minimum and maximum values~~  
16 ~~of incident UV light, CDOM, and  $\Phi_\lambda$  were used to create a 'lowest case' and 'highest case'~~  
17 ~~scenario of photo-degradation over a range of residence time from hours to 20 days. Because it~~  
18 ~~was shown in Fig. 7 (left set of bars) that the natural variability in CDOM concentrations has no~~  
19 ~~effect on water column rates of photo-degradation, the scenarios were created by varying~~  
20 ~~incoming UV light and DOM quality ( $\Phi_\lambda$ ). For example, the minimum UV light and minimum~~  
21  ~~$\Phi_\lambda$  values observed resulted in low rates of photo-mineralization; over a 20 day residence time~~  
22 ~~less than 5% of the DOM in surface waters could be converted to  $\text{CO}_2$  by photo-mineralization.~~  
23 ~~Conversely, combining the maximum UV light and maximum  $\Phi_\lambda$  values shows that 100% of the~~  
24 ~~DOM could be converted to  $\text{CO}_2$  by photo-mineralization at the end of about one week (Fig. 9).~~  
25 The relationship between residence time and DOM photo-degradation in Imnavait Creek was  
26 explored by multiplying Eqn. 3 (representing the water column rate of DOM photodegradation as  
27 the product of UV light, CDOM, and  $\Phi_\lambda$ ) by residence time to generate the results in Fig. 9.  
28 Combinations of minimum and maximum values of incident UV light, CDOM, and  $\Phi_\lambda$  were  
29 used to create a 'lowest case' and 'highest case' scenario of photo-degradation over a range of  
30 residence time from hours to 20 days. Thus, Fig. 9 shows cumulative DOM loss as a function of  
31 residence for a range of conditions (i.e., UV light availability, CDOM, and  $\Phi_\lambda$ ) generated using a

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1 'lowest case' and 'highest case' scenario of photo-degradation. Because it was shown in Fig. 7  
2 (left set of bars) that the natural variability in CDOM concentrations has no effect on water  
3 column rates of photo-degradation, the scenarios were created by varying incoming UV light and  
4 DOM quality ( $\Phi_\lambda$ ). For example, the minimum UV light and minimum  $\Phi_\lambda$  values observed  
5 resulted in low rates of photo-mineralization; over a 20 day residence time less than 5% of the  
6 DOM in surface waters could be converted to  $\text{CO}_2$  by photo-mineralization. Conversely,  
7 combining the maximum UV light and maximum  $\Phi_\lambda$  values shows that 100% of the DOM could  
8 be converted to  $\text{CO}_2$  by photo-mineralization at the end of about one week (Fig. 9). However, a  
9 precise estimate of residence time is difficult to achieve in practice given that there are inputs of  
10 water and "fresh" (labile) CDOM as a parcel of water moves downstream; accounting for these  
11 inputs is needed to quantify the total, integrated amount of DOM broken down by light as a  
12 function of residence time in a stream.

13 In all surface waters there is a variety of combinations of photochemical and hydrological  
14 controls that vary in space and time, and that define the "range" of DOM photo-degradation  
15 rates. For example, in Imnavait Creek longer residence times occur during times of low flow and  
16 stratification, but this stratification serves to protect DOM from UV light by isolating water  
17 masses in pool bottoms (e.g., Table 3, Fig. 5). Although exposure of surface waters to UV light  
18 did create chemical differences in DOM between surface and bottom waters, overall there was a  
19 relatively narrow range of CDOM concentrations at all locations and times in Imnavait Creek. In  
20 all surface waters there is a variety of combinations of photochemical and hydrological controls  
21 that vary in space and time, and that define the "range" of DOM photo-degradation rates. For  
22 example, in Imnavait Creek longer residence times occur during times of low flow and  
23 stratification, and this stratification serves to protect DOM from UV light by isolating water  
24 masses in pool bottoms (e.g., Table 3, Fig. 5). The volume of water sequestered in the pool  
25 bottoms (below the mixing depth) under stratified conditions was on average about 70% of the  
26 total pool volume (Figs. 3, 4; Merck et al. 2012). Thus, under stratified conditions, the majority  
27 of the pool volume was sequestered in the bottom, below the depth of UV light penetration (8 –  
28 45 cm see results section 3.2). However, the depth of light penetration into the ponds does not  
29 differ between stratified (low flow) or mixed (high flow) conditions as shown by the limited  
30 differences in  $a_{\text{CDOM}\lambda}$  values at 305 nm between these conditions in Imnavait Creek (comparing  
31 pool surface  $a_{\text{CDOM}\lambda}$  values in 2011 vs. 2012, Table 3). Thus, the amount of CDOM exposed to

1 light, or the rate of light absorption, does not differ between stratified vs. mixed conditions (for a  
2 given amount of sunlight under given sky conditions). The only difference is the amount of time  
3 for the photo-degradation to occur (greater photo-degradation under longer residence times  
4 associated with low-flow, stratified conditions; Fig. 9). Therefore, although most water was  
5 sequestered in the pool bottoms under stratified conditions, more DOM is lost due to photo-  
6 degradation under these conditions. This is because there is enough light-absorbing DOM that is  
7 labile to photo-degradation even in the pool surface waters under all conditions that DOM photo-  
8 degradation is not limited by substrate (DOM supply). The amount of water and DOM  
9 sequestered in the bottom waters does not influence the amount of DOM that can be degraded by  
10 light in this system.

11 Rates of photo-mineralization varied little over this narrow range of CDOM (Fig. 7, left),  
12 and because CDOM was very high photochemical reactions were light-limited (Fig. 8b). In  
13 addition, the consistently high CDOM concentrations observed across space (pool to pool) and  
14 averaged over time (2011 to 2012) in Innavait Creek (Tables 2 and 3, Fig. 5) suggests that  
15 CDOM lost to photo-mineralization under any photochemical or hydrological conditions is  
16 rapidly replenished from riparian soil waters over relatively short time periods (see also Merck et  
17 al. 2012). ~~In other words, it~~ is likely that stream reaches with high CDOM concentrations  
18 (substrate rich) and residence times in the range observed in Innavait Creek are similarly always  
19 light-limited (Fig. 8). Increased residence times, or lower CDOM concentrations such as those  
20 observed in the Kuparuk River, will move a system from light-limitation toward co-limitation by  
21 light and substrate (Fig. 8), again depending on the combination of photochemical and  
22 hydrological properties and their variability in space and time.

## 23 **5 Conclusions**

24 Results from this study demonstrate that in Innavait Creek photo-degradation dominates  
25 over bacterial degradation of DOM, and photo-degradation can create substantial differences in  
26 DOM chemistry between water masses isolated during stratification. The amount and lability of  
27 CDOM and the light attenuation by CDOM form a critical control point in DOM degradation –  
28 higher CDOM attenuates light faster with depth but results in no change or an increase in the  
29 overall rate of light absorption in the water column. With increasing CDOM and thus increasing  
30 rates of light-absorption, photo-degradation rates in the water column are more likely to be light-

1 limited and rates will increase with incident UV light or residence time. Given that higher light  
2 attenuation by CDOM traps heat in surface waters and creates stratification, which lengthens  
3 residence times and thus the time-integrated light exposure of DOM, low-flow conditions in  
4 Imnavait Creek likely maximize the conditions for photo-degradation of DOM.

5 On the other hand, if CDOM concentrations are very low, then the system is substrate-  
6 limited and the total mass of DOM exposed is more important than the amount of time the DOM  
7 spends exposed to UV (Fig. 8a, right side). This is because when the system is substrate limited,  
8 even a short exposure to UV will result in rapid and substantial photo-degradation, and exposing  
9 greater amounts of DOM even over short residence times will increase the overall photochemical  
10 processing in the system. In addition, in our conceptual model the lability of DOM to photo-  
11 degradation acts as a control on processing rates independent of whether a system is light- or  
12 substrate limited (Fig. 8a). Finally, at the scale of a stream reach or catchment, the balance  
13 between light- vs. substrate-limitation of DOM degradation varies with changes in water  
14 residence times, the incident UV light, and photo-lability of DOM. Our analyses indicate that  
15 the hydrological and photochemical conditions in Imnavait Creek create light-limitation for  
16 DOM photo-degradation, and we suggest that photo-degradation in most streams and ponds with  
17 high CDOM is similarly light-limited.

#### 18 **Author contributions**

19 GWK, BTN and RMC designed the field sampling plan and all authors contributed to the field  
20 work and data analysis. RMC, GWK and BTN prepared the manuscript with contributions  
21 KHH.

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#### 27 **References**

28 Amado, A. M., Farjalla, V. F., Esteves, F. D. A., Bozelli, R. L., Roland, F. and Enrich-Prast, A.:  
29 Complementary pathways of dissolved organic carbon removal pathways in clear-water

1 Amazonian ecosystems: Photochemical degradation and bacterial uptake, *FEMS Microbiol.*  
2 *Ecol.*, 56(1), 8–17, doi:10.1111/j.1574-6941.2006.00028.x, 2006.

3 Battin, T. J., Kaplan, L. a., Findlay, S., Hopkinson, C. S., Marti, E., Packman, A. I., Newbold, J.  
4 D. and Sabater, F.: Biophysical controls on organic carbon fluxes in fluvial networks, *Nat.*  
5 *Geosci.*, 2(8), 595–595, doi:10.1038/ngeo602, 2009.

6 Boano, F. and Harvey, J.: Hyporheic flow and transport processes: Mechanisms, models, and  
7 biogeochemical implications, *Rev. Geophys.*, 603–679, doi:10.1002/2012RG000417.Received,  
8 2014.

9 Brooks, M. L., Meyer, J. S. and McKnight, D. M.: Photooxidation of wetland and riverine  
10 dissolved organic matter: Altered copper complexation and organic composition, *Hydrobiologia*,  
11 579(1), 95–113, doi:10.1007/s10750-006-0387-6, 2007.

12 [Caplanne, S. and Laurion, I.: Effect of chromophoric dissolved organic matter on epilimnetic](#)  
13 [stratification in lakes, \*Aquat. Sci.\*, 70\(2\), 123–133, doi:10.1007/s00027-007-7006-0, 2008.](#)

14 Chapra, S. C. and Runkel, R. L.: Modeling Impact of Storage Zones on Stream Dissolved  
15 Oxygen, *J. Environ. Eng.*, 125(5), 415–419, doi:10.1061/(ASCE)0733-9372(1999)125:5(415),  
16 1999.

17 Cole, J. J., Caraco, N. F., Kling, G. W. and Kratz, T. K.: Carbon dioxide supersaturation in the  
18 surface waters of lakes., *Science*, 265(5178), 1568–1570, doi:10.1126/science.265.5178.1568,  
19 1994.

20 Cole, J. J., Prairie, Y. T., Caraco, N. F., McDowell, W. H., Tranvik, L. J., Striegl, R. G., Duarte,  
21 C. M., Kortelainen, P., Downing, J. A., Middelburg, J. J. and Melack, J.: Plumbing the global  
22 carbon cycle: Integrating inland waters into the terrestrial carbon budget, *Ecosystems*, 10(1),  
23 171–184, doi:10.1007/s10021-006-9013-8, 2007.

24 Cory, R. M. and Kaplan, L. A.: Biological lability of streamwater fluorescent dissolved organic  
25 matter, *Limnol. Oceanogr.*, 57(5), 1347–1360, doi:10.4319/lo.2012.57.5.1347, 2012.

26 Cory, R. M., McKnight, D. M., Chin, Y. P., Miller, P. and Jaros, C. L.: Chemical characteristics  
27 of fulvic acids from Arctic surface waters: Microbial contributions and photochemical  
28 transformations, *J. Geophys. Res. Biogeosciences*, 112(4), doi:10.1029/2006JG000343, 2007.

1 Cory, R. M., Miller, M. P., McKnight, D. M., Guerard, J. J. and Miller, P. L.: Effect of  
2 instrument-specific response on the analysis of fulvic acid fluorescence spectra, *Limnol.*  
3 *Oceanogr. Methods*, 8, 67–78, doi:10.4319/lom.2010.8.0067, 2010.

4 Cory, R. M., Crump, B. C., Dobkowski, J. a and Kling, G. W.: Surface exposure to sunlight  
5 stimulates CO<sub>2</sub> release from permafrost soil carbon in the Arctic., *Proc. Natl. Acad. Sci. U. S.*  
6 *A.*, 110(9), 3429–34, doi:10.1073/pnas.1214104110, 2013.

7 Cory, R. M., Ward, C. P., Crump, B. C. and Kling, G. W.: Sunlight controls water column  
8 processing of carbon in arctic fresh waters, *Science* (80-. ), 345(6199), 925–928,  
9 doi:10.1126/science.1253119, 2014.

10 Detterman, R. L., Bowsher, A. L. and Dutro Jr., J. T.: Glaciation on the Arctic Slope of the  
11 Brooks Range, Northern Alaska, *Arctic*, 11(1), 43–61, 1958.

12 Fee, E. J., Hecky, R. E., Kasian, S. E. M. and Cruikshank, D. R.: Effects of lake size, water  
13 clarity, and climatic variability on mixing depths in Canadian Shield lakes, *Limnol. Oceanogr.*,  
14 41(5), 912–920, doi:10.4319/lo.1996.41.5.0912, 1996.

15 Gareis, J. a L., Lesack, L. F. W. and Bothwell, M. L.: Attenuation of in situ UV radiation in  
16 Mackenzie Delta lakes with varying dissolved organic matter compositions, *Water Resour. Res.*,  
17 46(9), 1–14, doi:10.1029/2009WR008747, 2010.

18 Graneli, W., Lindell, M. and Tranvik, L. J.: Photo-oxidative production of dissolved inorganic  
19 carbon in lakes of different humic content, *Limnol. Oceanogr.*, 41(4), 698–706,  
20 doi:10.4319/lo.1996.41.4.0698, 1996.

21 Hamilton, T. D.: Late Cenozoic glaciation of the central Brooks Range, in *Glaciation in Alaska-*  
22 *The geologic record*, edited by T. D. Hamilton, K. M. Reed, and R. M. Thorson, pp. 9–50,  
23 Alaska Geological Society., 1986.

24 Helms, J. R., Stubbins, A., Ritchie, J. D., Minor, E. C., Kieber, D. J. and Mopper, K.: Absorption  
25 spectral slopes and slope ratios as indicators of molecular weight, source, and photobleaching of  
26 chromophoric dissolved organic matter, *Limnol. Oceanogr.*, 53(3), 955–969,  
27 doi:10.4319/lo.2008.53.3.0955, 2008.

28 Hinzman, L. D., Kane, D. L., Gieck, R. E. and Everett, K. R.: Hydrologic and thermal properties  
29 of the active layer in the Alaskan Arctic, *Cold Reg. Sci. Technol.*, 19, 95–110, 1991.

1 Hobbie, J. E.: Limnology of Tundra Ponds, Barrows, Alaska, US/IBP Synth. Ser. n.13 Dowden,  
2 Hutchinson Ross, Inc. Stroudsbeng, PA 514 pp., 0(0), NULL, 1980.

3 Houser, J. N.: Water color affects the stratification, surface temperature, heat content, and mean  
4 epilimnetic irradiance of small lakes, *Can. J. Fish. Aquat. Sci.*, 63(11), 2447–2455,  
5 doi:10.1139/f06-131, 2006.

6 Hu, C. M., Muller-Karger, F. E. and Zepp, R. G.: Absorbance, absorption coefficient, and  
7 apparent quantum yield: A comment on common ambiguity in the use of these optical concepts,  
8 *Limnol. Oceanogr.*, 47(4), 1261–1267, 2002.

9 Judd, K., Crump, B. and Kling, G.: Bacterial responses in activity and community composition  
10 to photo-oxidation of dissolved organic matter from soil and surface waters, *Aquat. Sci. - Res.*  
11 *Across Boundaries*, 69, 96–107, doi:10.1007/s00027-006-0908-4, 2007.

12 Kane, D. L.: No Title, [online] Available from:  
13 <http://ine.uaf.edu/werc/projects/NorthSlope/northslope.html>, 2015.

14 Kane, D. L. and Hinzman, L. D.: Climate data from the North Slope Hydrology Research  
15 project, , accessed Feb. 2013 [online] Available from:  
16 <http://ine.uaf.edu/werc/projects/NorthSlope>, 2011.

17 Kane, D. L., Hinzman, L. D., McNamara, J. P., Zhang, Z. and Benson, C. S.: An overview of a  
18 nested watershed study in Arctic Alaska, *Nord. Hydrol.*, 31(4-5), 245–266 [online] Available  
19 from: <Go to ISI>://WOS:000167153700002, 2000.

20 Kane, D. L., Gieck, R. E., Kitover, D. C., Hinzman, L. D., McNamara, J. P. and Yang, D.:  
21 Hydrological cycle on the North Slope of Alaska, in *Northern Research Basins Water Balance*,  
22 edited by D. L. Kane and D. Yang, pp. 224–236, IAHS Press, Wallingford., 2004.

23 Kling, G. W.: Comparative transparency, depth of mixing, and stability of stratification in lakes  
24 of Cameroon, West Africa, *Limnol. Oceanogr.*, 33(1), 27–40, doi:10.4319/lo.1988.33.1.0027,  
25 1988.

26 Kling, G. W., Kipphut, G. W. and Miller, M. C.: Arctic lakes and streams as gas conduits to the  
27 atmosphere: implications for tundra carbon budgets., *Science*, 251(4991), 298–301,  
28 doi:10.1126/science.251.4991.298, 1991.

- 1 Koehler, B., Landelius, T., Weyhenmeyer, G. a., Machida, N. and Tranvik, L. J.: Sunlight-  
2 induced carbon dioxide emissions from inland waters, *Global Biogeochem. Cycles*, 28(7), 696–  
3 711, doi:10.1002/2014GB004850, 2014.
- 4 McKnight, D. M., Boyer, E. W., Westerhoff, P. K., Doran, P. T., Kulbe, T. and Andersen, D. T.:  
5 Spectrofluorometric characterization of dissolved organic matter for indication of precursor  
6 organic material and aromaticity, *Limnol. Oceanogr.*, 46(1), 38–48,  
7 doi:10.4319/lo.2001.46.1.0038, 2001.
- 8 McNamara, J. P., Kane, D. L. and Hinzman, L. D.: An analysis of streamflow hydrology in the  
9 Kuparuk River Basin, Arctic Alaska: a nested watershed approach, *J. Hydrol.*, 206(1-2), 39–57,  
10 doi:10.1016/S0022-1694(98)00083-3, 1998.
- 11 McNamara, J. P., Kane, D. L., Hobbie, J. E. and Kling, G. W.: Hydrologic and biogeochemical  
12 controls on the spatial and temporal patterns of nitrogen and phosphorus in the Kuparuk River,  
13 arctic Alaska, *Hydrol. Process.*, 22, 3294–3309, doi:10.1002/hyp.6902, 2008.
- 14 Merck, M. F. and Neilson, B. T.: Modelling in-pool temperature variability in a beaded arctic  
15 stream, *Hydrol. Process.*, 26(25), 3921–3933, doi:10.1002/hyp.8419, 2012.
- 16 Merck, M. F., Neilson, B. T., Cory, R. M. and Kling, G. W.: Variability of in-stream and riparian  
17 storage in a beaded arctic stream, *Hydrol. Process.*, 26(19), 2938–2950, doi:10.1002/hyp.8323,  
18 2012.
- 19 Miller, M. P., McKnight, D. M., Chapra, S. C. and Williams, M. W.: A model of degradation and  
20 production of three pools of dissolved organic matter in an alpine lake, *Limnol. Oceanogr.*,  
21 54(6), 2213–2227, doi:10.4319/lo.2009.54.6.2213, 2009.
- 22 Miller, W. L.: Effects of UV-radiation on aquatic humus: photochemical principles and  
23 experimental considerations, in *Aquatic Humic Substances - Ecology and Biogeochemistry*,  
24 edited by D. O. Hesson and L. J. . Tranvik, pp. pp. 126–143, Springer Berlin Heidelberg., 1998.
- 25 Moran, M. A., Sheldon, W. M. and Zepp, R. G.: Carbon loss and optical property changes during  
26 long-term photochemical and biological degradation of estuarine dissolved organic matter,  
27 *Limnol. Oceanogr.*, 45(6), 1254–1264, doi:10.4319/lo.2000.45.6.1254, 2000.
- 28 Morris, D. P., Zagarese, H., Williamson, C., BALSEIRO, E. G., HARGREAVES, B. R.,  
29 MODENUTTI, B., MOELLER, R. and QUEIMALINOS, C.: The attenuation of solar UV

1 radiation in lakes and the role of dissolved organic carbon, *Limnol. Oceanogr.*, 40(8), 1381–  
2 1391, doi:10.4319/lo.1995.40.8.1381, 1995.

3 Neilson, B. T., Hatch, C. E., Ban, H. and Tyler, S. W.: Solar radiative heating of fiber-optic  
4 cables used to monitor temperatures in water, *Water Resour. Res.*, 46(8), W08540,  
5 doi:10.1029/2009WR008354, 2010.

6 Osterkamp, T. E. and Payne, M. W.: Estimates of permafrost thickness from well logs in  
7 northern Alaska, *Cold Reg. Sci. Technol.*, 5(1), 13–27, doi:10.1016/0165-232X(81)90037-9,  
8 1981.

9 Page, S. E., Kling, G. W., Sander, M., Harrold, K. H., Logan, J. R., McNeill, K. and Cory, R. M.:  
10 Dark formation of hydroxyl radical in arctic soil and surface waters, *Environ. Sci. Technol.*,  
11 47(22), 12860–12867, doi:10.1021/es4033265, 2013.

12 Page, S. E., Logan, J. R., Cory, R. M. and McNeill, K.: Evidence for dissolved organic matter as  
13 the primary source and sink of photochemically produced hydroxyl radical in arctic surface  
14 waters., *Environ. Sci. Process. Impacts*, 16(4), 807–22, doi:10.1039/c3em00596h, 2014.

15 Prairie, Y., Breton, J., Vallières, C. and Laurion, I.: Limnological properties of permafrost thaw  
16 ponds in northeastern Canada, *Can. J. Fish. Aquat. Sci.*, 66(10), 1635–1648, doi:10.1139/F09-  
17 108, 2009.

18 Spencer, R. G. M., Hernes, P. J., Ruf, R., Baker, A., Dyda, R. Y., Stubbins, A. and Six, J.:  
19 Temporal controls on dissolved organic matter and lignin biogeochemistry in a pristine tropical  
20 river, Democratic Republic of Congo, *J. Geophys. Res. Biogeosciences*, 115(3),  
21 doi:10.1029/2009JG001180, 2010.

22 Stedmon, C. A., Markager, S. and Bro, R.: Tracing dissolved organic matter in aquatic  
23 environments using a new approach to fluorescence spectroscopy, *Mar. Chem.*, 82(3-4), 239–  
24 254, doi:10.1016/S0304-4203(03)00072-0, 2003.

25 Stubbins, A., Spencer, R. G. M., Chen, H., Hatcher, P. G., Mopper, K., Hernes, P. J., Mwamba,  
26 V. L., Mangangu, A. M., Wabakanghanzi, J. N. and Six, J.: Illuminated darkness: Molecular  
27 signatures of Congo River dissolved organic matter and its photochemical alteration as revealed  
28 by ultrahigh precision mass spectrometry, *Limnol. Oceanogr.*, 55(4), 1467–1477,  
29 doi:10.4319/lo.2010.55.4.1467, 2010.

- 1 Tranvik, L. J. and Bertilsson, S.: Contrasting effects of solar UV radiation on dissolved organic  
2 sources for bacterial growth, *Ecol. Lett.*, 4(5), 458–463, doi:10.1046/j.1461-0248.2001.00245.x,  
3 2001.
- 4 Vähätalo, A. V. and Wetzel, R. G.: Long-term photochemical and microbial decomposition of  
5 wetland-derived dissolved organic matter with alteration of  $^{13}\text{C}:^{12}\text{C}$  mass ratio, *Limnol.*  
6 *Oceanogr.*, 53(4), 1387–1392, doi:10.4319/lo.2008.53.4.1387, 2008.
- 7 Walker, D. A., Binnian, E., Evans, B. M., Lederer, N. D., Nordstrand, E. and Webber, P. J.:  
8 Terrain, vegetation and landscape evolution of the R4D research site, Brooks-Range-foothills,  
9 Alaska, *Holarct. Ecol.*, 12(3), 238–261, 1989.
- 10 Watanabe, S., Laurion, I., Chokmani, K., Pienitz, R. and Vincent, W. F.: Optical diversity of  
11 thaw ponds in discontinuous permafrost: A model system for water color analysis, *J. Geophys.*  
12 *Res. Biogeosciences*, 116(2), doi:10.1029/2010JG001380, 2011.
- 13 Weishaar, J. L., Aiken, G. R., Bergamaschi, B. A., Fram, M. S., Fujii, R. and Mopper, K.:  
14 Evaluation of specific ultraviolet absorbance as an indicator of the chemical composition and  
15 reactivity of dissolved organic carbon, *Environ. Sci. Technol.*, 37(20), 4702–4708,  
16 doi:10.1021/es030360x, 2003.
- 17 Wetzel, R. G.: *Limnology: Lake and River Ecosystems.*, 2001.
- 18 Zarnetske, J. P., Haggerty, R., Wondzell, S. M. and Baker, M. A.: Dynamics of nitrate  
19 production and removal as a function of residence time in the hyporheic zone, *J. Geophys. Res.*  
20 *Biogeosciences*, 116(1), doi:10.1029/2010JG001356, 2011.

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**Table 1.** Meteorological conditions at Toolik Field Station during study periods

Period	<sup>a</sup> Global Solar Radiation	<sup>a</sup> UV+Visible Photon Flux	<sup>b</sup> Air Temperature (1 m)	<sup>a</sup> Precipitation	<sup>c</sup> Total discharge at weir
	kW m <sup>-2</sup>	mol photons m <sup>-2</sup>	°C	mm	m <sup>3</sup>
2011	12	2111	9	63	41x10 <sup>3</sup>
2012	10	1600	10	189	233x10 <sup>3</sup>

<sup>a</sup>Sum of daily average values from 23-June through 18-August in each year (sampling periods in 2011 and 2012).

<sup>b</sup>Daily average value from 23-June through 18-August. <sup>c</sup> Sum discharge passing the weir at Imnavait from 28-Jun to 18-August.

**Table 2.** Annual average hydrologic and chemical characteristics of soil and stream water samples from Imnavait Creek.

		<u>Soil Water</u>		<u>Pool Bottom Water</u>		<u>Pool Surface Water</u>	
		<u>2011</u>	<u>2012</u>	<u>2011</u>	<u>2012</u>	<u>2011</u>	<u>2012</u>
Water Temp	°C			11 ± <1	11 ± <1	14 ± 1	11 ± 1
pH		5.2 ± 0.4	5.2 ± 0.1	5.5 ± 0.1	5.6 ± 0.1	5.7 ± 0.1	5.7 ± 0.1
Conductivity	µs cm <sup>-1</sup>	26 ± 1	25 ± 2	42 ± 10	17 ± 3	13 ± <1	12 ± <1
DOC	µM C	1357 ± 110	1412 ± 78	1252 ± 85	1100 ± 17	785 ± 13	1028 ± 21
a <sub>305</sub>	m <sup>-1</sup>	96 ± 10	87 ± 7	118 ± 19	61 ± 2	53 ± 2	53 ± 1
S <sub>R</sub>		0.75 ± 0.01	0.74 ± 0.01	0.70 ± 0.02	0.75 ± 0.01	0.78 ± 0.02	0.76 ± <0.01
SUVA <sub>254</sub>	(L mg C <sup>-1</sup> m <sup>-1</sup> )	4.4 ± 0.1 <sup>A</sup>	4.1 ± 0.1 <sup>B</sup>	5.2 ± 0.5	3.9 ± 0.1	4.5 ± 0.1	3.7 ± <0.1
Peak A	RU	3.3 ± 0.2	2.9 ± 0.2	3.5 ± 0.1	2.8 ± <0.1	2.5 ± <0.1	2.7 ± 0.1
FI		1.59 ± 0.01	1.60 ± 0.01	1.55 ± 0.01	1.57 ± 0.01	1.51 ± 0.01	1.55 ± <0.01

All values are seasonal averages across all dates in 2011 or 2012 shown with standard error in parentheses.

**Table 2.** Annual average hydrologic and chemical characteristics of soil and stream water samples from Imnavait Creek.

		<u>Soil Water</u>		<u>Pool Bottom Water</u>		<u>Pool Surface Water</u>	
		<u>2011</u>	<u>2012</u>	<u>2011</u>	<u>2012</u>	<u>2011</u>	<u>2012</u>
<u>Water Temp</u>	<u>°C</u>			<u>11 ± &lt;1</u>	<u>11 ± &lt;1</u>	<u>14 ± 1<sup>A</sup></u>	<u>11 ± 1<sup>B</sup></u>
<u>pH</u>		<u>5.2 ± 0.4</u>	<u>5.2 ± 0.1</u>	<u>5.5 ± 0.1</u>	<u>5.6 ± 0.1</u>	<u>5.7 ± 0.1</u>	<u>5.7 ± 0.1</u>
<u>Conductivity</u>	<u>µs cm<sup>-1</sup></u>	<u>26 ± 1</u>	<u>25 ± 2</u>	<u>42 ± 10<sup>A</sup></u>	<u>17 ± 3<sup>B</sup></u>	<u>13 ± &lt;1</u>	<u>12 ± &lt;1</u>
<u>DOC</u>	<u>µM C</u>	<u>1357 ± 110</u>	<u>1412 ± 78</u>	<u>1252 ± 85<sup>A</sup></u>	<u>1100 ± 17<sup>B</sup></u>	<u>785 ± 13<sup>A</sup></u>	<u>1028 ± 21<sup>B</sup></u>
<u>a<sub>305</sub></u>	<u>m<sup>-1</sup></u>	<u>96 ± 10</u>	<u>87 ± 7</u>	<u>118 ± 19<sup>A</sup></u>	<u>61 ± 2<sup>B</sup></u>	<u>53 ± 2</u>	<u>53 ± 1</u>
<u>S<sub>R</sub></u>		<u>0.75 ± 0.01</u>	<u>0.74 ± 0.01</u>	<u>0.70 ± 0.02<sup>A</sup></u>	<u>0.75 ± 0.01<sup>B</sup></u>	<u>0.78 ± 0.02</u>	<u>0.76 ± &lt;0.01</u>

<b>SUVA<sub>254</sub></b>	(L mg C <sup>-1</sup> m <sup>-1</sup> )	4.4 ± 0.1 <sup>A</sup>	4.1 ± 0.1 <sup>B</sup>	5.2 ± 0.5 <sup>A</sup>	3.9 ± 0.1 <sup>B</sup>	4.5 ± 0.1 <sup>A</sup>	3.7 ± <0.1 <sup>B</sup>
<b>Peak A</b>	RU	3.3 ± 0.2	2.9 ± 0.2	3.5 ± 0.1 <sup>A</sup>	2.8 ± <0.1 <sup>B</sup>	2.5 ± <0.1 <sup>A</sup>	2.7 ± 0.1 <sup>B</sup>
<b>FI</b>		1.59 ± 0.01	1.60 ± 0.01	1.55 ± 0.01 <sup>A</sup>	1.57 ± 0.01 <sup>B</sup>	1.51 ± 0.01 <sup>A</sup>	1.55 ± <0.01 <sup>B</sup>

All values are seasonal averages ± standard error across all dates in 2011 or 2012. Letters indicate significant difference (p < 0.05) in mean values between years (2011 vs. 2012) for each water type (soil waters, pool bottom waters, and pool surface waters).

**Table 3. DOM in soil waters compared to pool water in Innavaik Creek under stratified conditions**

		<b>Soil Water</b>	<b>Pool Bottom Water</b>	<b>Pool Surface Water</b>
DOC	µM C	1382 ± 69	1188 ± 56	815 ± 12
α <sub>305</sub>	m <sup>-1</sup>	92 ± 6	100 ± 13	50 ± 4
S <sub>R</sub>		0.75 ± 0.01	0.71 ± 0.01	0.77 ± 0.01
SUVA <sub>254</sub>	(L mg C <sup>-1</sup> m <sup>-1</sup> )	4.4 ± 0.1	4.9 ± 0.3	4.2 ± 0.1
Peak A	RU	3.1 ± 0.2	3.2 ± 0.1	2.38 ± 0.05
FI		1.60 ± 0.01	1.57 ± 0.01	1.52 ± 0.01

All values shown as average ± standard error; calculated in the pool surface or bottom across all dates in 2011 and 2012 when pools were stratified (2011: 27 Jun, 29 Jun, 14 Jul, 4 Aug; 2012: 23 Jun, 30 Jun). Soil water values were calculated as average over both years (2011-2012).

**Table 3.** DOM in soil waters compared to pool water in Innavaik Creek under stratified conditions

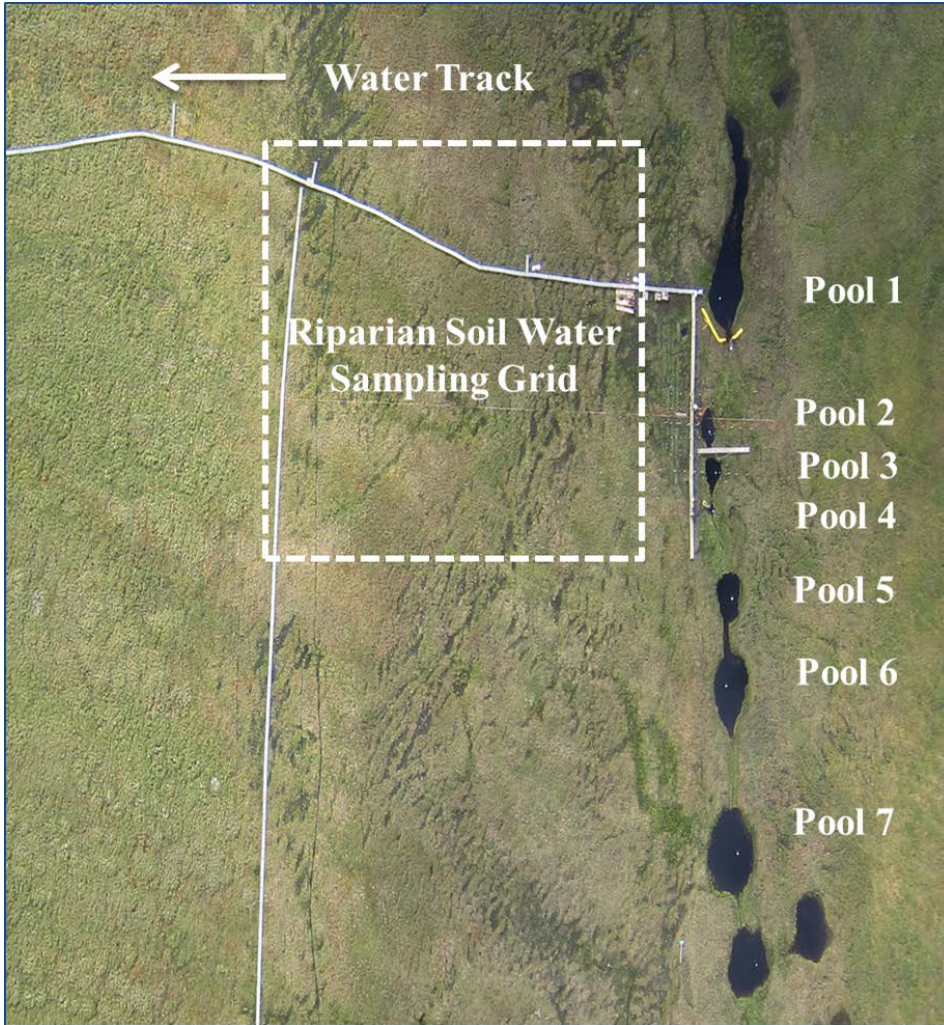
		<u>Soil Water</u>	<u>Pool Bottom Water</u>	<u>Pool Surface Water</u>
<u>DOC</u>	<u>μM C</u>	<u>1382 ± 69</u>	<u>1188 ± 56</u>	<u>815 ± 12</u>
<u>a<sub>305</sub></u>	<u>m<sup>-1</sup></u>	<u>92 ± 6</u>	<u>100 ± 13</u>	<u>50 ± 1</u>
<u>S<sub>R</sub></u>		<u>0.75 ± 0.01</u>	<u>0.71 ± 0.01</u>	<u>0.77 ± 0.01</u>
<u>SUVA<sub>254</sub></u>	<u>(L mg C<sup>-1</sup> m<sup>-1</sup>)</u>	<u>4.4 ± 0.1</u>	<u>4.9 ± 0.3</u>	<u>4.2 ± 0.1</u>
<u>Peak A</u>	<u>RU</u>	<u>3.1 ± 0.2</u>	<u>3.2 ± 0.1</u>	<u>2.38 ± 0.05</u>
<u>FI</u>		<u>1.60 ± 0.01</u>	<u>1.57 ± 0.01</u>	<u>1.52 ± 0.01</u>

All values shown as average ± standard error; calculated in the pool surface or bottom across all dates in 2011 and 2012 when pools were stratified (2011: 27-Jun, 29-Jun, 14-Jul, 4-Aug; 2012: 23-Jun, 30-Jun). Soil water means were calculated from over both years (2011-2012). All water types were statistically different from one another for all variables (ANOVA  $p < 0.01$ ).

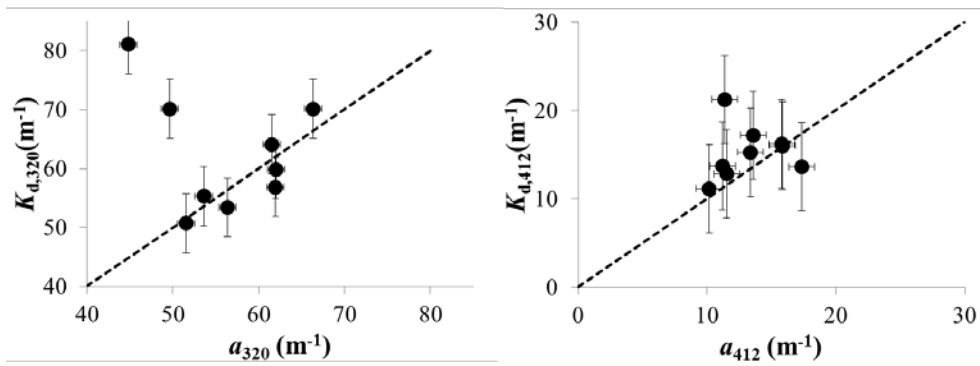
**Table 4.** Effect of sunlight and bacteria on CDOM, FDOM and DOM mineralization.

Process	$a_{305}$ $m^{-1}$	$S_R$	$SUVA_{254}$ $L\ mg\ C^{-1}\ m^{-1}$	FI	Peak A $RU$	Peak C $RU$	Peak T $RU$	Mineralization rate ( $mmol\ C\ m^{-2}\ d^{-1}$ )
<sup>a</sup> Photo-degradation	$-12.4 \pm 1.8$	$15.8 \pm 1.3$	$-4.9 \pm 0.01$	$-11.7 \pm 0.8$	$-12.1 \pm 1.4$	$-27.2 \pm 2.1$	$4.1 \pm 1.5$	$24.7 \pm 18.3$
<sup>b</sup> Bacterial degradation	$-2.9 \pm 1.6$	ND	$5.0 \pm 0.03$	ND	$-7.6 \pm 2.4$	$-3.6 \pm 1.5$	$-2.5 \pm 8.2$	$2.35 \pm 0.34$
<sup>c</sup> Photo + bacterial degradation	$3 \pm 1$	$1 \pm 1$	NM	$5 \pm 1$	$7 \pm 1$	$15 \pm 1$	$-12 \pm 1$	$3.04 \pm 1.31$

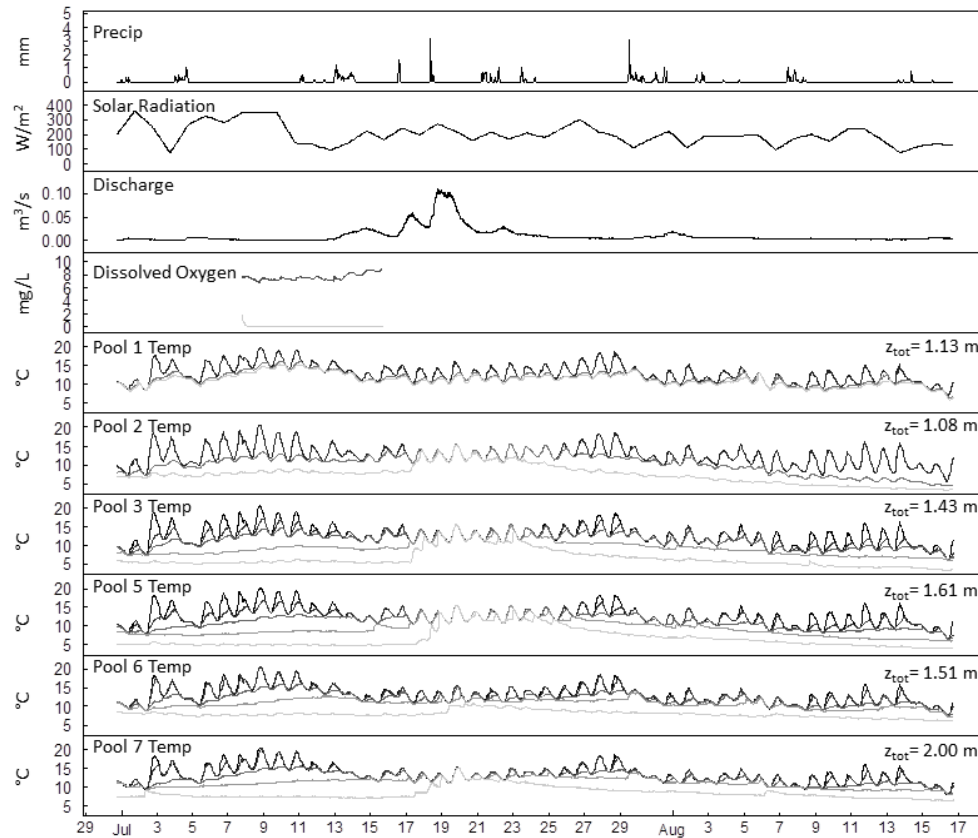
DOM degradation quantified as percent change in CDOM or FDOM of Imnavait Creek waters after <sup>a</sup>exposure to 12 h sunlight compared to dark control (photo-degradation), <sup>b</sup> after seven days of incubation of whole waters at 6-7 °C compared to killed control (bacterial degradation), or <sup>c</sup> after 12 h sunlight followed by seven days of incubation with natural bacterial community in stream water at 6-7 °C relative to killed controls (here the percent change is relative to the initial photo-exposed water). Data shown as average  $\pm$  standard error of three photochemical experiments, two bacterial incubations, and one coupled photo-bacterial incubation. Within each of the bacterial or photo + bacterial experiments there were four replicates per treatment (light, dark, live or killed controls). ND = none detected based on no significant difference compared to killed controls. NM = not measured.



**Fig. 1.** Study area showing the 120 m reach of Innavait Creek containing seven consecutive pools, and the locations of soil water collection (riparian zone and water track; only the bottom portion of the water track is shown in this image). Also visible in this areal image as white solid lines are boardwalks installed to access sampling sites.

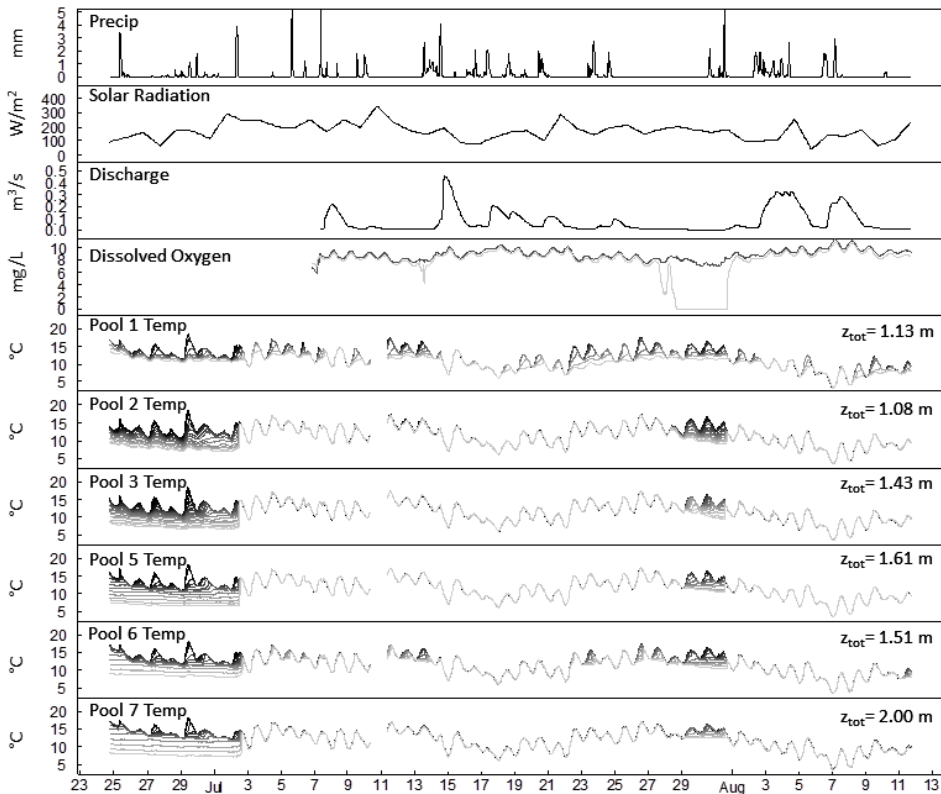


**Fig. 2.**  $K_{d,\lambda}$  vs.  $a_{\text{CDOM},\lambda}$  at 320 nm (left) and 412 nm (right) plotted vs. 1:1 line (dotted line).  $a_{\text{CDOM},\lambda}$  measured on the laboratory spectrophotometer was corrected for the average cosine of downwelling radiation for the time of day (i.e., zenith angle) that the in-situ  $K_{d,\lambda}$  values were measured in Innavait Creek. Thus  $a_{\text{CDOM},\lambda}$  values in this figure are not directly comparable to the values presented in Tables 2 and 3.



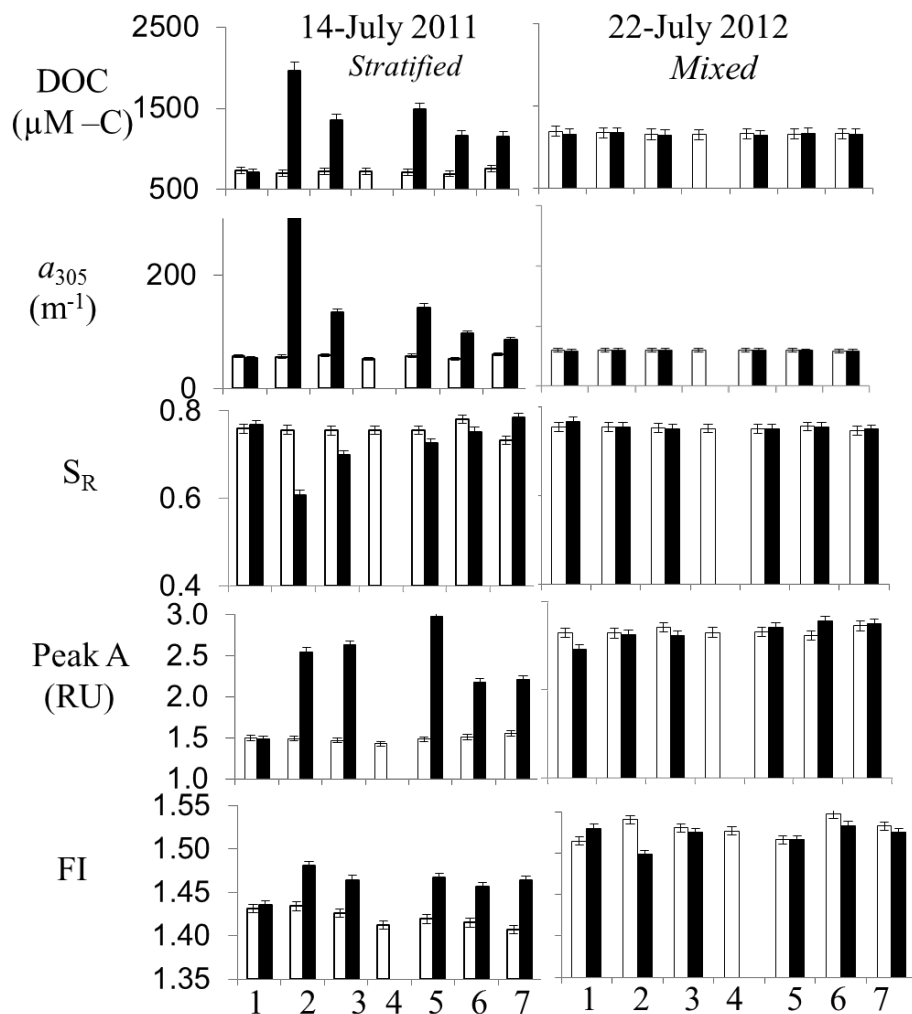
**Fig. 3.** Innavaic Creek precipitation, solar radiation, discharge (at location downstream of study reach), dissolved oxygen in pool 2 top and bottom, and vertical arrays (VA) of temperature sensors within each study pool (P) in summer 2011. The darkest lines represent the sensor at the top of the water column; subsequent lines become lighter with depth of each sensor. The sensors were placed in each pool starting 5 to 15 cm from the bottom of the pool and then at intervals ranging from 5 to 50 cm over the depth of each pool ( $z_{tot}$ ) as indicated in the figure.

**Comment [RMC1]:** Revised Figure replaces former Fig 3.



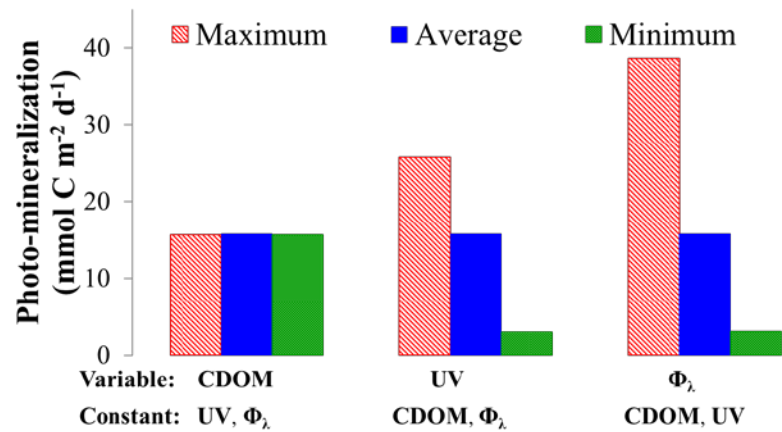
**Fig. 4.** Innavaik Creek precipitation, solar radiation, discharge (at location downstream of study reach), dissolved oxygen in pool 2 top and bottom, and vertical arrays (VA) of temperature sensors within each study pool (P) in summer 2012. The darkest lines represent the sensor at the top of the water column; subsequent lines become lighter with depth of each sensor. The sensors were placed in each pool starting 5 to 15 cm from the bottom of the pool and then at intervals ranging from 5 to 50 cm over the depth of each pool ( $z_{tot}$ ) as indicated in the figure.

Comment [RMC2]: Revised figure replaces former Fig 4.

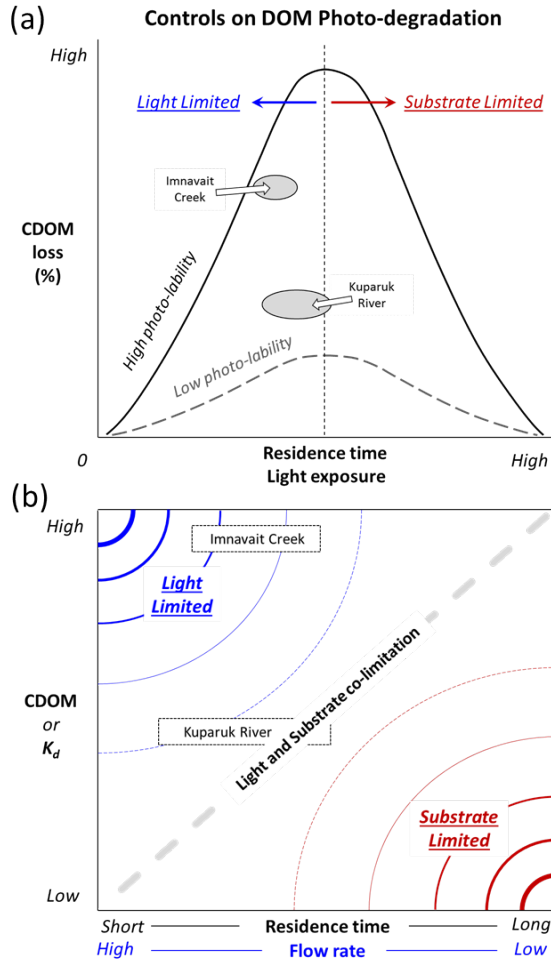


**Fig. 5.** Concentration and quality of DOM in the surface (open bars) and bottom waters (filled bars) in pools 1-7 in Innavaik Creek under stratified (left) and mixed (right) conditions in 2011 and 2012, respectively. CDOM and FDOM concentrations shown as absorption coefficients at 305 nm ( $a_{305}$ ) and emission intensities at Peak A (Raman Units; RU), respectively. DOM quality shown as slope ratio ( $S_R$ ) and fluorescence index (FI); CDOM and FDOM proxies for DOM described in text.

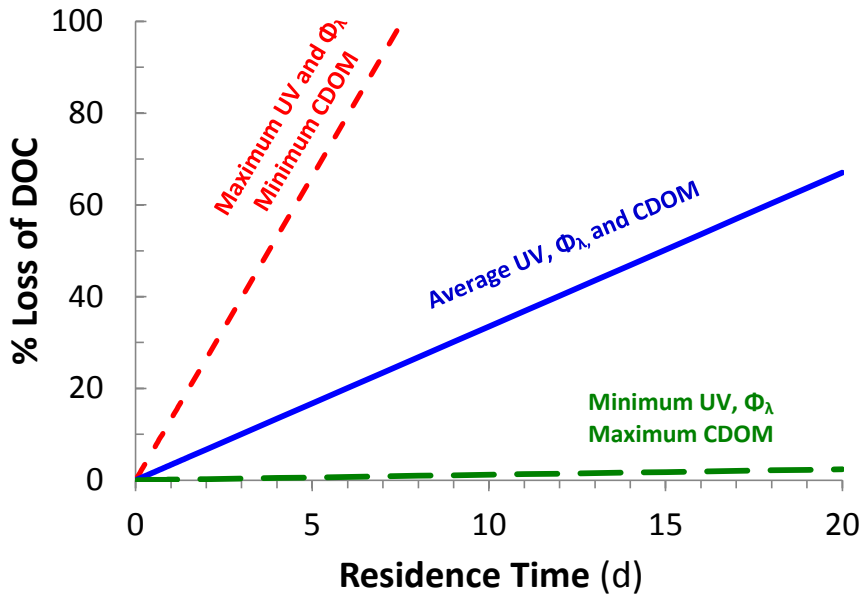




**Fig. 7.** Effect of CDOM, incident UV, and  $\Phi_\lambda$  (apparent quantum yield) on water column rates of photo-mineralization of DOM to  $\text{CO}_2$  in Innavait Creek. For each scenario, two variables from *Eqn. 3* were held constant, and one was varied using the average, minimum, and maximum values observed over the study period at Innavait Creek (2011-2012). UV = daily total UV reaching water surface at Innavait Creek (dependent on solar zenith angle and cloud cover).

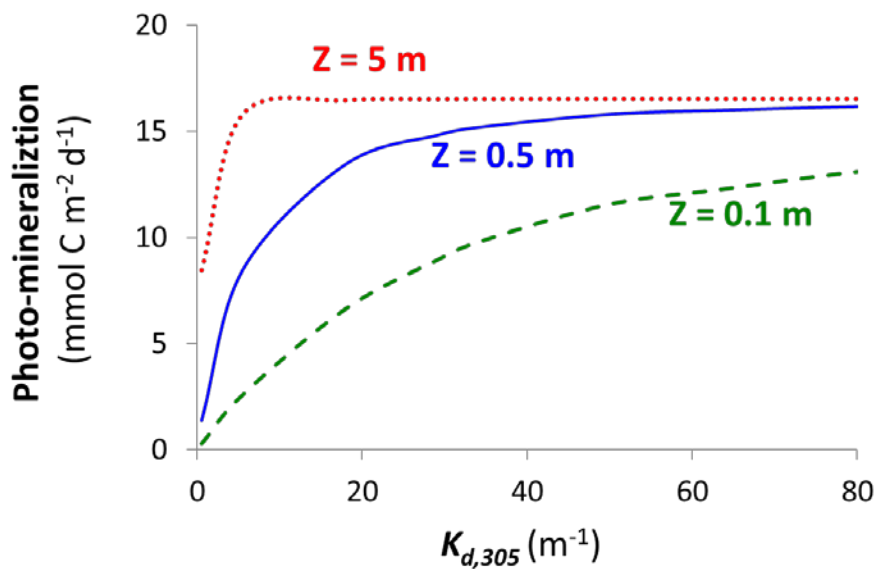


**Fig. 8.** Controls on DOC photo-degradation. **(a)** As residence time or light exposure increase, so does DOC photo-degradation (% loss of the initial pool of CDOM, without replacement). The CDOM loss for any given light exposure is greater for higher photo-labile DOC (solid line) than it is for lower photo-labile DOC (dashed line). At low light exposure levels photo-degradation is “light-limited”, but after sufficient CDOM is lost the process switches to be “substrate-limited” (insufficient CDOM). **(b)** This shows the light- vs. substrate- limitation in terms of CDOM concentrations or light attenuation ( $K_d$ ), Y-axis, and the water residence time or flow rate (X-axis). At high CDOM and short residence time there is insufficient light available for photo-degradation (upper left), while at low CDOM concentrations and long residence times there is abundant light yet insufficient CDOM. The range of conditions for DOC photo-degradation in Imnavait Creek is likely always light-limited, while in the Kuparuk River conditions may be substrate-limited at times.

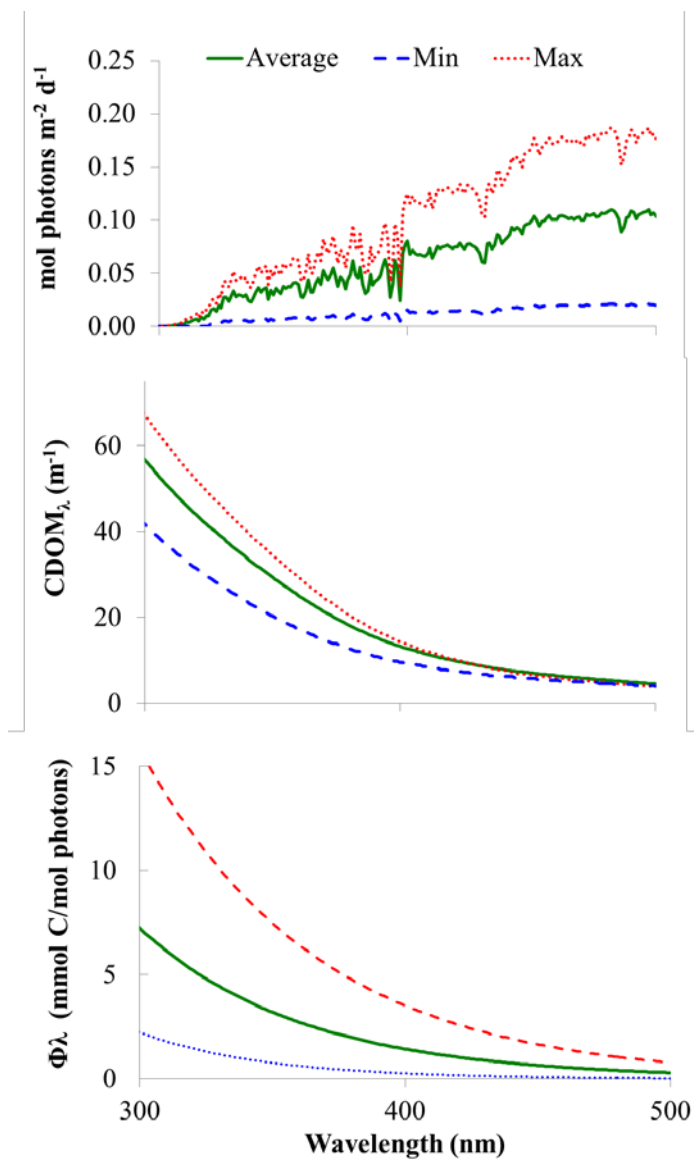


**Fig. 9.** Cumulative percent of DOC loss in Innavait Creek lost through photo-mineralization as calculated from Eqn. 3 (removal of DOC as  $\text{mol C m}^{-2} \text{d}^{-1}$  over the mean depth in Innavait Creek; 0.5 m) in Innavait Creek using combinations of the range of surface UV light exposure, CDOM concentrations, and apparent quantum yields ( $\Phi_\lambda$ ) measured in this study for up to a 20 day residence time. For each scenario, the initial DOC concentration was set to  $943 \mu\text{M C}$ , the average surface water DOC concentration over both 2011 and 2012 (Table 2). Calculations do not include (1) the effect of DOC loss on changing light attenuation ( $K_d$ ) over the residence time (i.e., CDOM and thus  $K_{d,\lambda}$  remain constant over the residence time for each scenario), or (2) the effect of UV light exposure on  $\Phi_\lambda$  over time ( $\Phi_\lambda$ , or DOM lability, remained constant over the residence time for each scenario).

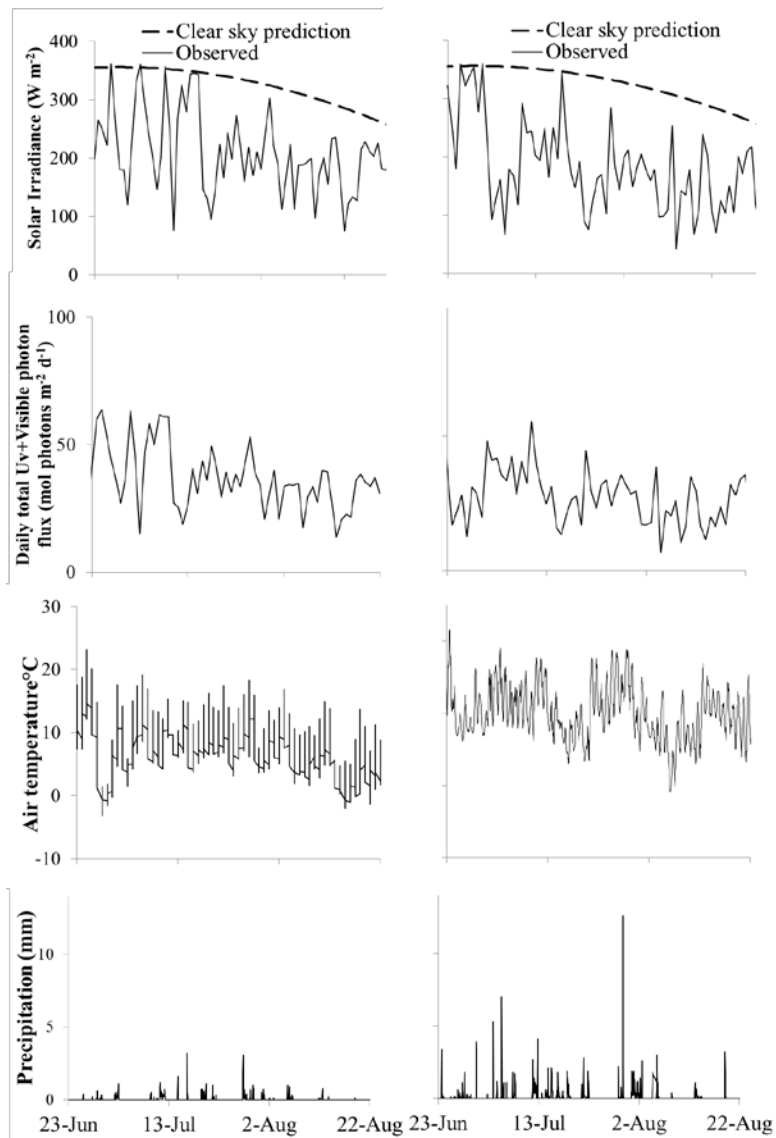
## Supporting Information – Figures and Data



**Fig. S1.** The dependence of water column rates of photo-mineralization as a function of  $K_d$ , which increases with increasing CDOM. Three situations with varying depth ( $z$ ) of the water column are shown. In all but the shallowest water columns, the photo-mineralization rate asymptotes at relatively low  $K_d$  values – the range of  $K_d$  at 305 nm measured in Innavait Creek was 39 to 63 m<sup>-1</sup>.



**Fig. S2.** Average, minimum, and maximum spectra of incident UV light (daily photon flux), CDOM, and apparent quantum yield of photo-mineralization measured in Innavait Creek.



**Fig. S3.** UV and climate data at Innavait Creek for 2011 (left) and 2012 (right) during the study periods. (A) Predicted and observed mean daily global solar irradiance. (B) Daily total sum of UV+ Visible photon flux reaching water surface. (C) Air temperature measured 1m above ground at Innavait Creek. (D) Precipitation.