Interactive comment on "Mesozooplankton structure and functioning during the onset of the Kerguelen phytoplankton bloom during the Keops2 survey" by F. Carlotti et al.

Anonymous Referee #3
Received and published: 17 March 2015

Journal: BG

Title: Mesozooplankton structure and functioning during the onset of the Kerguelen Bloom during Keops2 survey

Author(s): F. Carlotti et al. MS No.: bg-2014- 598

Referee:
General Comments Carlotti et al. present and extensive and intensive overview of the zooplankton abundance, biomass, taxonomic composition, and stable isotope composition observed around the Kerguelen Island survey during the spring of 2011. They particularly investigate an undulation of the Polar Front east of the region, and the effect of time over their 6 week survey (a positive effect with time approaching early summer), the effect of day-night (little effect), and the influence of HNLC waters and Fe enrichment over the plateau. The zooplankton is sampled with a bongo net and 333 um mesh; it is significant that all the samples are analysed with Zooscan which is an achievement in itself.
In some ways this paper is actually 2 papers in one.
The separation and identification of specific taxa for stable isotope analysis is impressive; Figures 5, 7 and 8 are very revealing.

Answers: We thank the reviewer for his/her thorough review and highly appreciate the comments and suggestions, which significantly contributed to improving the quality of the publication. Please find below a detailed response to the each of the comments.

My concerns are:

Referee:
1) It is a rich data set and the conclusions mostly sound, but from an external perspective of this paper for a special Keops issue it seems rather colloquial. I realise the readership will be from the Keops2 group, but to others it may seem rich with jargon on the station names and "T-groups" and it is hard to glean the major findings. At some points the paper seems like a technical report.

Answers:
We agree about the heaviness of the names of stations and group of stations.
We rewrote the paragraph describing the cruise strategy and the different stations. The names and terminology are common between all papers dedicated to the KEOPS2, and we maintain them. We tried to make it simpler and clearer to the reader, guiding him in the Figure1, and quoting other key papers. The figure caption of figure 1 has been reworked with more details.

Concerning the results about isotopic ratios, the mention of IS groups is now suppressed in the sections 'Data analysis', 'Results' and 'Discussion'. We kept only the groups of stations (T-groups) individualised by Trull et al (2015) based on the chemometric characteristics of phytoplankton.

Referee:

2) Could the analyses be made more general rather than cruise specific, by relating the conditions of zooplankton to water mass and bathymetry rather than latitude, longitude and voyage track?

Answers:

We rewrote the introduction to better specified the scientific objectives and explain the sampling strategy in relation with these objectives. The paragraph 2.1 Study site and sampling strategy has been rewritten to better link the different group of stations and hydrodynamical features. Other papers of the KEOPS2 special issue have been quoted in a way to better guide the reader for complementary information.

Moreover, we add in the result a paragraph "3.1. Hydrology and trophic conditions" to better explain the hydrological and trophic environmental conditions met by the sampled mesozooplankton at each stations.

Referee:

3) More importantly there is no discrete question on why this survey was done. The main objective is to compare the zooplankton with Keops1 (which was not explicitly possible with OPC vs. zooscan?) and "its responses to primary production" – presumably to Chl-a biomass (as primary production was not measured).

Answers:

When rewriting the introduction, webetter specified the scientific objectives both of the whole KEOPS 2 cruise and the specific objectives of the present paper.

At the end of the introduction, the last paragraph sum up these objectives

"The main objective of the KEOPS2 study was to investigate the early phase (October–November 2011) of the seasonal marine productivity in this Kerguelen region in order to gain new insights on the biogeochemistry and ecosystem response to iron fertilization. The study was conducted in contrasted environments differently impacted by iron availability, i.e. on the plateau waters, in areas common with KEOPS1, and in productive oceanic deep waters with strong mesoscale activity to the east of the Kerguelen Islands. The focus of the present paper is to document the responses of zooplankton in terms of species diversity, density and biomass in the mosaic of blooms observed during the survey, and to characterize the trophic pathways from primary production to large mesozooplanktonic organisms.

Referee:

1) The stable isotope analysis lacks an ecosystem analysis, to compare composition of phytoplankton (?) (the source) with the other members of the zooplankton community. There are many elegant methods (some Bayesian) in the public domain to quantitatively compare the predator-prey relationships. Most copepods are omnivorous, and the degree herbivory reflects the availability of alternative prey.

Answers:

Stable isotope values of zooplankton were compared to those of phytoplankton recorded by Trull et al. (2015) in the same stations. This information was synthesized in the new figure 10 and added in the Discussion section. We discussed the link between phyto- and zooplankton in the different groups of stations and calculated the mean trophic fractionation between these two broad trophic levels.

Zooplankton size fractions were composed of organisms with different feeding regimes (herbivores, omnivores and carnivores in varying proportions, as indicated in the discussion). Thus, it would be incorrect to use mixing models (Bayesian SIAR for example) for inferring

predator-prey relationships between zooplanktonic fractions. However, we calculated the mean trophic fractionation between phytoplankton and zooplankton as a whole. The low fractionation values observed (+ 0.40 ‰ for $\delta^{13}C$ and + 2.69 ‰ for $\delta^{15}N$) indicated a dominance of herbivory in zooplankton, and confirmed the conclusions based on zooplankton composition.
The Discussion section on stable isotope results was rewritten and this information added to the text (p 21-22).

Referee:
In summary, the Introduction needs to better justify why this study was made, and where the knowledge gaps are that need to be filled.
Answers:
Introduction has been rewritten consequently.

Referee:
In the Methods section (p. 2386) are many papers of 2014 about the fate of phytoplankton, but not much about how this paper fits in. These papers should be cited more in the Introduction.
Answers:
In the paragraph "3.1. Hydrology and trophic conditions", we better explain the hydrological and trophic environmental conditions met by the sampled mesozooplankton at each stations, and we quote a restricted number of relevant papers of KEOPS 2 needed to discuss our results. In the discussion part, we gave more explanations about the linkages between our results and those of KEOPS2 companion papers dedicated to the fate of phytoplankton.

Specific Comments

Referee:
The mesh size does affect the size data from sieves, so that the smaller sizes (as they acknowledge) are not quantitatively sampled, but merely indicative because of occasional, sporadic clogging. The species composition is useful for long-term ocean observing, but it does not contribute to their specific questions (how does the biodiversity compare with Keops1?).
Answers:
Indeed, during the cruise we used both 120 µm and 330 µm mesh size nets on the Bongo frame. The results of the 120 µm were not presented in the first version, but we added them in the present version. In many stations, the 120 µm size net was often clogged with phytoplankton cells and aggregates, and the cod-end contents could not be used for dry weight and ZOOSCAN process. Abundances from taxonomic counting obtained with the 120 µm size net are now presented

Results corresponding to the taxonomic determination with the 120 µm mesh size net are presented in the Table 1 and on the figure 6, and in the paragraph 3.3 Metazooplankton community composition and distribution.
Another paper is in preparation to discuss in more detail about biodiversity patterns during the KEOPS2 cruise (from bacteria to mesozooplankton) and in the present paper, we only mention in paragraph 4.2: "The taxonomic composition did not show major differences between shelf and oceanic waters, except that the contribution of copepods to the whole mesozooplankton was higher in oceanic waters than on the shelf, and these taxonomic patterns were quite similar between the KEOPS 1 (see Fig. 7 in Carlotti et al. 2008) and KEOPS2 survey (Fig. 6)."

Referee:
They could take their ECD data, or sieve data, and compare it with the Keops1 OPCdata series by amalgamating size classes.

Answers:
We were not quite sure about the comment understanding. Indeed, we defined the same size fractions for the abundance and biomass results of KEOPS1 and KEOPS2 from the ESD data. In the part "**4.2 Comparison with previous results",** we explain why the results are comparable even using different laboratory technologies (Lab OPC during KEOPS1 and ZOOSCAN during KEOPS2) and we give more details about the comparison of results.
A new table 4 synthesizes the data of abundance and biommas size fractions:
"Table 4: Seasonal variations of zooplankton abundance and biomass from KEOPS2 (15 October – 20 November 2011) and KEOPS1 (January 19- February 13, 2005) surveys with contribution of different size fractions (<500 µm, 500-1000 µm; 1000-2000 µm; > 2000 µm). The reference stations were A3 (shelf waters) and C11 (oceanic waters) for KEOPS1 (see Carlotti et al., 2008, their Figs. 3 and 5) , and A3 (shelf waters) and TNS6-TNS5 and E4E-E5 (oceanic waters) for KEOPS2."

Referee:
Can Tables 1 and 2 be put into an appendix or supplementary information (it is very useful data) but can they be graphed in some way?

Answers:
Information of Table 1 is used in Figure 6, and Table 2 data are graphed in Figures 9 and 10.

Referee:
Line 5, p. 238, 330 micron (not mm)

Answers: OK, we changed it

Referee:
Line 6 – how did the bongo nets to 250 m depth compare with the thermocline depth?

Answers:
As written before, we add in the result a paragraph "3.1. Hydrology and trophic conditions" to better explain the hydrological and trophic environmental conditions met by the sampled mesozooplankton at each stations. Particularly we add more information about the MLD and quote papers which have deeper description about the vertical physical structure of the water column at the different stations (Trulls et al, their table 4a, Jouandet et al., 2014). Our bongo nets to 250 m depth always included the mixed layer.

Referee:
Line 21, p 2390. You may have compared 13C to VPDB and 15N to atmospheric N, but there is the internal laboratory (working) standard of acetanilide. This is not a simple comparison. How was this compared; did the working standard overlap the observed values for zooplankton? A two point calibration is needed, see Paul D, Skrzypek G, Forizs I (2007) Normalization of Measured Stable Isotopic Compositions to Isotope Reference Scales - a Review. Rapid Communications in Mass Spectrometry 21:3006-3014); and Coplen TB, Brand WA, Gehre M, Groning M, Meijer HAJ, Toman B, Verkouteren RM (2006). New guidelines for delta c-13 measurements. Analytical Chemistry 78:2439-2441.

Answers:
Stable isotope values were properly corrected following routine standard procedures in the laboratory where the analyses were done (UMR LIENSs, University of La Rochelle). Calibrations to VPDB and N2 are performed regularly using certified reference materials

(USGS-24, IAEA-CH6, -600 for carbon; IAEA-N2, -NO-3, -600 for nitrogen), as well as intercalibration between several facilities. The replicated measurement of internal standards each 10 analyses are used to determine the accuracy of the values and to detect any analytical drift. Acetanilide is used as internal standard. It has values in the range of the analyzed samples: -27.0 ‰ for $\delta^{13}C$, +1 ‰ for $\delta^{15}N$. These precisions were added to the § 2.6 on stable isotope analyses in the Materials and methods section.

As detailed before, we do make calibrations regularly but we do not realize two point calibrations while running each batch of samples. This procedure is carried out regularly but it appears that it does not give a better precision.

Referee:
Line 20, p. 2392. The ANOVA tables would be useful, at least as supplementary information.
Answers:
The tables are presented below

ANOVA tables for linear regression of abundances versus time

Source	SS	DF	MS	F
Treatments	25909,18	1	25909,18	24,62164
Error	36830,26	35	1052,29	
Total (corrected)	62739,44	36		

ANOVA table for linear regression of biomasses versus time

Source	SS	DF	MS	F
Treatments	10,01	1	10,01	6,491218
Error	53,96	35	1,54179	
Total (corrected)	63,97	36		

Referee:
Fig. 6. Pie charts are very hard to quantitatively compare – can these be presented as bar graphs?
Answers:
We maintained the pie charts which allow to present the distributions for several stations in a synthetic way. Moreover we had pie charts for two size class from the 120 *m mesh size net. To help the reader, we increased the size of the police on the pie charts, and the figure caption of the figure 6 has been more detailed:
"Figure 6. Distributions of main taxa abundances at stations A3-1, A3-2, E3 and E5 from binocular observation. Distributions are presented for four size fractions (small, medium, large, and very large) for the organisms observed in the 330 µm mesh size net samples (four upper bands on the figure), and distributions are presented for the two lower size fractions (small and medium) for the 120 µm mesh size net samples (two lower bands on the figure). Distributions are average values between day and night samples. For each size fraction (the

four pie charts on the same horizontal band), the color labels for the different taxa are similar.”

Referee:

Fig. 7. The 80% similarity for grouping your samples is arbitrary, and the discrimination of groups is tenuous considering that there are branching just above and below 80%.

What was the stress statistic for the associate MDS plot?

Answers:

The value stress statistic for the associate MDS plot is 0,12. We added the associated MDS plot in the figure 7

1 **Revised version of the manuscript (below)**

Mesozooplankton structure and functioning during the onset of the Kerguelen Phytoplankton Bloom during the Keops2 survey

François Carlotti
Mediterranean Institute of Oceanography (MIO)
Aix Marseille Université, CNRS, Université de Toulon, IRD, MIO UM 110,
13288, Marseille, France
Email: francois.carlotti@mio.osupytheas.fr

Marie-Paule Jouandet
Mediterranean Institute of Oceanography (MIO)
Aix Marseille Université, CNRS, Université de Toulon, IRD, MIO UM 110,
13288, Marseille, France
Email: marie-paule.jouandet@mio.osupytheas.fr

Antoine Nowaczyk
Mediterranean Institute of Oceanography (MIO)
Aix Marseille Université, CNRS, Université de Toulon, IRD, MIO UM 110,
13288, Marseille, France
Email: antoine.nowaczyk@u-bordeaux.fr

Mireille Harmelin-Vivien
Mediterranean Institute of Oceanography (MIO)
Aix Marseille Université, CNRS, Université de Toulon, IRD, MIO UM 110,
13288, Marseille, France
Email: Mireille.Harmelin-vivien@mio.osupytheas.fr

Dominique Lefèvre

Mediterranean Institute of Oceanography (MIO)
Aix Marseille Université, CNRS, Université de Toulon, IRD, MIO UM 110,
13288, Marseille, France

Email: dominique.lefèvre@mio.osupytheas.fr

Pierre Richard

Littoral Environnement et Sociétés, UMR 7266 CNRS-Université de La Rochelle, 2 Rue Olympe de Gouges, 17000 La Rochelle, France.

Email : pierre.richard@univ-lr.fr

Yiwu Zhu

University of Massachusetts Boston, Boston, MA 02125

Mediterranean Institute of Oceanography (MIO)
Aix Marseille Université, CNRS, Université de Toulon, IRD, MIO UM 110,
13288, Marseille, France

Email: Yiwu.Zhu@umb.edu

Meng Zhou

University of Massachusetts Boston, Boston, MA 02125

Mediterranean Institute of Oceanography (MIO)
Aix Marseille Université, CNRS, Université de Toulon, IRD, MIO UM 110,
13288, Marseille, France

Email: Meng.Zhou@umb.edu

Abstract.

This paper presents results on the spatial and temporal distribution patterns of mesozooplankton in the naturally fertilized region to the east of the Kerguelen Islands (Southern Ocean) visited at early bloom stage during the KEOPS2 survey (15 October – 20 November 2011). The aim of this study is to compare the zooplankton response in contrasted environments localised over the Kerguelen Plateau in waters of the east shelf and shelf edge and in productive oceanic deep waters characterized by conditions of complex circulation and rapidly changing phytoplankton biomass.

The mesozooplankton community responded to the growing phytoplankton blooms earlier on the plateau than in the oceanic waters, where complex mesoscale circulation stimulated initial more or less ephemeral blooms before a broader bloom extension. The mesozooplankton species composition showed a high degree of similarity across the whole region, and the populations initially responded to spring bloom with a large production of larval forms increasing abundances, without biomass changes. Taxonomic composition and stable isotope ratios of size-fractionated zooplankton indicated the strong domination of herbivores, and the total zooplankton biomass values over the survey presented a significant correlation with the integrated chlorophyll concentrations in the mixed layer depth.

The biomass stocks observed at the beginning of the KEOPS2 cruise were around 1.7 g C m^{-2} above the plateau and 1.2 g C m^{-2} in oceanic waters. Zooplankton biomass in oceanic waters remained on average below 2 g C m^{-2} over the study period, except for one station in the Polar Front Zone (FL), whereas zooplankton biomasses were around 4 g C m^{-2} on the plateau at the end of the survey. The most remarkable feature during the sampling period was the stronger increase in abundance in the oceanic waters (25 10^3 to 160 10^3 ind.m^{-2}) than on the plateau (25 10^3 to 90 10^3 ind. m^{-2}). The size structure and taxonomic distribution patterns revealed a cumulative contribution of various larval stages of dominant copepods and euphausiids particularly in the oceanic waters, with clearly identifiable stages of progress during a Lagrangian survey. The reproduction and early stage development of dominant species were sustained by mesoscale-related initial ephemeral blooms in oceanic waters but individual growth was still food-limited and zooplankton biomass stagnated. By contrast, zooplankton abundance and biomass on the shelf were both in a growing phase, at slightly different rates, due to individual growth under sub-optimal conditions. Combined with our observations during the KEOPS1 survey (January-February 2015), the present results deliver a consistent understanding of patterns in mesozooplankton abundance and biomass from early spring to summer in the poorly documented oceanic region east of the Kerguelen Islands.

Introduction

The eastern part of the Kerguelen Plateau sustains one of the most important local foraging areas for land-based marine predators (birds, penguins, seals and elephant seals) and for whales (Hindell et al, 2011; Blain et al, 2013). Satellite-based chlorophyll images of this region highlight the intensive seasonal Kerguelen bloom and its southeast extension off the archipelago (Schlitzer, R., 2002; Thomalla et al., 2011; Blain et al, 2013; Trull et al, 2015). During the KEOPS1 survey (KErguelen Ocean and Plateau compared Study), the origin and fate of the elevated phytoplankton biomass over the Kerguelen plateau were addressed (Blain et al., 2008), with a focus on the mechanisms supplying the surface waters with iron. The Kerguelen Plateau, oriented along the 70º E meridian, forms a large north-west/south-east topographical barrier of the Antarctic Circumpolar Current, forcing the Polar Front (PF) to pass above the plateau south of the Kerguelen Islands in a meandering course (Figure 1). The PF flow on the shelf induces entrainment and mixing of Fe enriched shelf waters from plateau sediments in the oceanic upper layer in the eastern area of Kerguelen and drives relatively high phytoplankton bloom concentrations, with a strong increase from October to December (Blain et al., 2007; Blain et al, 2013), initially dominated by diatoms of high growth rates (Quéguiner, 2013) contrasting with the generally high-nutrient low-chlorophyll (HNLC) surface oceanic waters of the Southern Ocean. This enhanced biological productivity in the eastern area of the Kerguelen Islands fuels the trophic level of zooplankton and micronekton, which are potential prey of fish and squid forage required to meet the demand of top predators. During the KEOPS1 cruise (January–February 2005), the mesozooplankton populations, mainly copepods, were already well established without significant spatial and temporal changes in species composition and biomass, around 10.6 g C m^{-2} above the plateau and around 5 g C m^{-2} in HNLC oceanic waters (Carlotti et al., 2008). The KEOPS1 survey occurred during the decline phase of a natural long-term spring bloom initiated in November.

How the zooplankton populations increase from overwinter stocks by exploiting new primary production in early spring is still poorly documented because descriptions of seasonal variations of mesozooplankton standing stocks in oceanic Antarctic regions are scarce. The implementation of the Southern Ocean CPR survey delivers consistent information regarding the seasonal succession of zooplanktonic communities in the Southern Ocean south of Australia (Hosie et al, 2003; Hunt and Hosie, 2006 a and b). In the PF zone, a relatively strong increase in zooplankton abundance occurs in spring, from October-November (see Hosie et al, 2003, their Fig. 3), mainly due to changes in density of all common taxa from average winter levels still maintained until October (Hunt and Hosie, 2006 b). The largest copepods of the region (*Rhincalanus gigas*, *Calanoides acutus*, *Ctenocalanus citer*) are

seasonal migrators which arrive in the surface layer from winter diapause depths when Chl*a* concentrations increase (October - November). Overwintering females may spawn reserves even before the full bloom, whereas overwintering stages other than adult stages resume their growth in surface water up to mature adults which produce new cohorts during the full bloom period (Atkinson, 1998; Hunt and Hosie, 2006 b). Other smaller species (*Calanus similimus*, *Oithona* sp., etc.) resume their population development from survivors from the previous year and start reproduction earlier in spring (Atkinson, 1998). There is no historical CPR data around the Kerguelen Islands, but some pieces of the puzzle suggest similar patterns. Zooplankton distribution patterns observed by Semelkina (1993) during the SKALP cruises around the Kerguelen Islands (46–52°S, 64–73°E) from February 1997 to February 1998 showed a change in biomass (4-fold higher) from winter (July-August) to mid-summer (February), but did not describe this early spring period. Despite the particular environmental conditions above the plateau, it is worth noting that the seasonal zooplankton abundances recorded from February 1992 to January 1995 at the KERFIX station, located around 60 miles southwest of the Kerguelen Islands in 1700 m of water, show a major increase in copepod densities from September to January (Razouls et al., 1998).

The main objective of the KEOPS2 study was to investigate the early phase (October–November 2011) of the seasonal marine productivity in this Kerguelen region in order to gain new insights on the biogeochemistry and ecosystem response to iron fertilization. The study was conducted in contrasted environments differently impacted by iron availability, i.e. on the plateau waters, in areas common with KEOPS1, and in productive oceanic deep waters with strong mesoscale activity to the east of the Kerguelen Islands. The focus of the present paper is to document the responses of zooplankton in terms of species diversity, density and biomass in the mosaic of blooms observed during the survey, and to characterize the trophic pathways from primary production to large mesozooplanktonic organisms.

2 Material and methods

2.1 Study site and sampling strategy

The KEOPS2 survey was performed east of the Kerguelen Islands in the Indian sector of the Southern Ocean, on board R.V. Marion Dufresne, between the 15th of October and the 20th of November 2011. It firstly consisted of predefined stations along two transects (Fig. 1) the first oriented north-south between 46°50 S and 49°08 S, and subsequently referred to as TNS transect (Stations TNS1 to TNS10, blue dots in Fig. 1), and the second oriented east-west between 69°50 E and 74°60 E, referred to as TEW transect (Stations TEW1 to TEW8, green dots on Fig. 1). Along these two transects, zooplankton samples were collected once at

each station. The TEW transect crossed the Polar front twice, first between TEW3 and TEW4 where the southern branch of the PF flows northwards along the shelf-break, and secondly between TEW6 and TEW7, where the PF is directed southwards after a semicircle trajectory maintaining a large stationary meander in this area. The most westerly stations were located over the inner (TEW1) and outer (TEW2) parts of the Kerguelen shelf. The most easterly stations (TEW7 and TEW8) were situated in Sub-Antarctic Mode Water whereas the central section (TEW4 to TEW6) within the stationary meander was covered by mixed Antarctic surface water (Farias et al., 2015; Trull et al., 2015).

In addition, intensive sampling (24 hours) was performed at 9 strategic stations (Fig. 1) located in the eastern bloom in the polar frontal zone (F-L station), in the north-eastern bloom (set of E stations), in the south-eastern bloom above the Kerguelen plateau (A3 station) and in the deep waters south west of the Kerguelen Islands considered as a HNLC reference station (R station). Station A3 (common with KEOPS1) was sampled twice during KEOPS2: at pre-winter (A3-1) and spring stage (A3-2). The patterns of change over time of the Northeastern bloom, located in a complex recirculation area inside the stationary meander of the Polar front (Park et al., 2014; Zhou et al., 2014), was studied by a quasi-lagrangian survey including 5 stations (E1-E2-E3-E4E-E5).

Real time satellite images (chlorophyll and altimetry) in combination with trajectories of 50 drifters released during the first part of the cruise were used to carefully decide the positions of these 5 stations (Trull et al, 2015, their Fig. 2). In addition, we visited a productive station (E4W, red dot in Fig. 1) located in the plume of chlorophyll observed downstream of the plateau and close to the jet induced by the PF.

2.2 Mesozooplankton sampling

Zooplankton collection was conducted at 27 stations with a double Bongo (60 cm mouth diameter) with one 330-µm mesh net and a 120-µm mesh net mounted with filtering cod ends. Hauls were done from 250 m depth to the surface at 0.5 ms^{-1}. The stations of the TNS transect (stations TNS 1 to TNS10) and the stations of the TEW transect (stations TEW1 to TEW8) were sampled once each. During the long-term stations study (A3 visited twice, R2, F-L, and the set of stations E), zooplankton samples were taken twice daily, by day and by night(stations were named R2-d and R2-n, for instance).

For each sampling station, two successive net tows at each station were done: the first net tow was taken for the ZOOSCAN processing, taxonomy study and dry weight, and a second net tow was taken for isotopes. The cod-end contents of the first tow was kept fresh and split into two parts with a Motoda box (Motoda, 1959). The first part was preserved in 4%

borax-buffered formalin seawater for further laboratory study of zooplankton community structure (taxonomy, abundance and size structure) and biomass estimates from organism biovolume (see below). The second half of the sample was preserved for dry weight measurements. As many of the 120 µm size nets were clogged with phytoplankton cells and aggregates, we could not finally use the sample for dry weight and ZOOSCAN processing. However, we used the 120 µm size net for the isotope fractions 80-200 µm and 200-500 µm.

For preparing samples for isotope size fraction analysis, the content of the second 330 µm mesh size net cod end was first processed through the filtration column with the five sieves - 2000 µm, 1000, 500, 200, and 80 µm meshes - and then the filtered samples on the sieves 2000, 1000, 500 µm were collected for isotopes. For the largest size class (> 2000 µm), large organisms such as salps and euphausiids were separated into additional containers. The filtered samples on the mesh 200 µm and 80 µm were kept on the sieves and the filtration column reinstalled for processing the 120 µm net cod-end. Aggregates were blocked by the 2000, 1000 and even 500 µm sieves. Then the filtered samples on the 200 and 80 µm mesh size sieves were collected for isotopes. All samples were placed in small containers and immediately deep-frozen (-80 °C).

2.3 Abundance and biomass using the Zooscan

For each station, the cod end content of a 330 µm mesh size net was processed using ZOOSCAN (www.zooscan.com) to determine the zooplankton community size structure. ZOOSCAN has recently been used to study the zooplankton community in various areas and has been validated by comparisons with traditional sampling methods (Grosjean et al., 2004; Schultes and Lopes, 2009; Gorsky et al., 2010). Our ZOOSCAN setup is similar to the one described by Gorsky et al. (2010), and our sample processing protocol is fully presented in Nowaczyk et al. (2011), following the recommendations of Gorsky et al. (2010).

After homogenization, each sample was quantitatively split with a Motoda box once back in the laboratory and a fraction of each preserved sample containing a minimum of 1000 particles (in general 1/32 or 1/64 of the whole sample) was placed on the glass plate of the ZOOSCAN. Organisms were carefully separated one by one manually with a long wooden needle, in order to avoid overlapping. Each image was then run through the ZooProcess plug-in using the image analysis software Image J (Grosjean et al., 2004; Gorsky et al., 2010). Several measurements of each organism were then computerized. Organism size is given by its equivalent circular diameter (ECD) and can then be converted into biovolume, assuming each organism is an ellipsoid (more details in Grosjean et al., 2004). The lowest ECD detectable by this scanning device is 300 µm. To discriminate between aggregates and

organisms, we used a training set of about 1000 objects which were selected automatically from 40 different scans. This protocol allows discrimination between aggregates and organisms by building the initial training set of images. The biovolume (BV, mm^3) was calculated from the organism image areas and morphometric parameters. In order to estimate the biomass of each organism, we used the same conversion as in Carlotti et al. (2008), each measured biovolume (BV, mm^3) of a zooplankton individual was converted into biomass (W, mg DW) using the following relationship :log (W) = 0.865 log (BV) - 0.887 (Riandey et al., 2005). Carbon content was assumed to be 50% of body dry weight.

In this article, the terms 'ZOOSCAN abundance' and 'ZOOSCAN biomass' will designate the values derived from the laboratory ZOOSCAN processing. The abundance and biomass of organisms were then grouped into four size fractions (<500, 500–1000, 1000–2000, and > 2000 µm) based on their ECD, and summed to deliver the total abundance and biomass per sample over the upper 250 meters.

The choice of the net tow sampling depth was based on mixed layer depth found at the first station of the cruise, and maintained afterwards. Abundance and biomass values are normalized to the volume of water filtered *in situ*. ANOVA test (5% significance level) was used to test differences of abundance and biomass between stations or oceanic areas.

2.4 Taxonomic determination

Common taxa were counted with the binocular microscope for taxonomy. For the 330 µm mesh size net, around 600 organisms were counted from subsamples1/32 or 1/64). For the 120 µm mesh size net around 400 organisms were counted from 1 to 10 /1000 diluted samples. The whole sample was examined for either rare species and/or large organisms (i.e. euphausiids, amphipods). Identification of the copepod community was done down to species level and groups of developmental stage when possible. Species/genus identification was done according to Rose (1933), Tregouboff and Rose (1957) and Razouls et al. (2005–2014). Organisms other than copepods as well as meroplankton were identified down to taxa levels. Identifications were done to genus level for copepods, amphipods, pelagic molluscs, polychaetes, Thaliacea and Cnidarians; and to taxa level for other major holoplanktonic and meroplanktonic groups. To identify which taxonomic groups contribute to the four size fractions defined from ZOOSCAN measurements done on the 330 µm mesh net samples (see above), each observed organism was classified as small, medium, large or very large mesozooplankton, which almost corresponds to the four size fractions determined by ZOOSCAN (see above). Similarly, the organisms observed and counted from the 120µm mesh size net samples were also classified in small and medium size fractions. Distribution in

larger size fractions were not considered from the 120µm mesh size net samples, the large organisms being undersampled.

2.5 Biomass measurement

The subsample of the 330 µm mesh net for bulk biomass measurement was filtered onto pre-weighted and pre-combusted GF/F filter (47 mm) which was quickly rinsed with distilled water and dried in an oven at 60°C for 3 days on board. Dry-weight (mg) of 19 samples was calculated from the difference between the final weight and the weight of the filter and biomass (mg DW m^{-2}) was extrapolated from the total volume sampled by the net.

2.6 Stable isotope analysis

Before processing, identification of the broad taxonomic composition of each sample preserved for isotopic measurements was performed under a binocular microscope. When possible, the main group of organisms in the largest >2000 µm size-fraction were sorted out and processed separately. Then, zooplankton fractions were freeze-dried and ground into a homogeneous powder. As they may contain carbonates, an acidification step was necessary to remove ^{13}C-enriched carbonates (DeNiro and Epstein, 1978; Søreide et al. 2006). A subsample was acidified with 1% HCl, rinsed, dried and used for determination of $\delta^{13}C$ values, while the other untreated subsample was used for determination of nitrogen isotopic composition. Three replicates were performed on each plankton fraction per sampled station for both $\delta^{13}C$ and $\delta^{15}N$ values. Stable isotope measurements were performed with a continuous-flow isotope-ratio mass spectrometer (Delta V Advantage, Thermo Scientific, Bremen, Germany) coupled to an elemental analyzer (Flash EA1112 Thermo Scientific, Milan, Italy). Results are expressed in parts per thousand (‰) relative to Vienna Pee Dee Belemnite and atmospheric N_2 for $\delta^{13}C$ and $\delta^{15}N$, respectively, according to the equation: $\delta X = [(R_{sample}/R_{standard})-1] \times 10^3$, where X is ^{13}C or ^{15}N and R is the isotope ratio $^{13}C/^{12}C$ or $^{15}N/^{14}N$, respectively. Calibration was performed using certified reference materials (USGS-24, IAEA-CH6, -600 for carbon; IAEA-N2, -NO-3, -600 for nitrogen). Analytical precision based on repeated analyses of acetanilide (Thermo Scientific) used as an internal standard was <0.15‰. Percentage of organic C and organic N were obtained using the elemental analyzer and were used to calculate sample C/N ratios.

Lipids are depleted in $\delta^{13}C$ relative to proteins and carbohydrates, and variation in lipid content between organisms can introduce considerable bias into carbon stable isotope analyses (Bodin et al. 2007; Post et al. 2007). Like most polar marine organisms (Lee et al. 2006), KEOPS2 zooplankton fractions could present a high lipid content (up to 20% dry

mass, MHV data not shown), reflected by high C/N values. Thus, $\delta^{13}C$ acidified sample values of fractions >200 µm were corrected according to the formula calculated by Post et al. (2007) for aquatic organisms, using the C/N ratio of each sample: $\delta^{13}C_{normalized} = \delta^{13}C_{acidified} - 3.32 + 0.99 \times C/N$

Acidified $\delta^{13}C$ values of the lowest size-fraction (80-200 µm) were not lipid corrected due to their low lipid content (<5%, MHV data not shown). The resulting $\delta^{13}C_{normalized}$ provides an estimate of $\delta^{13}C$ corrected for the effects of lipid concentration. Lipid correction calculated by Smyntek et al. (2007) for zooplankton give $\delta^{13}C$ values 0.63 ± 0.01‰ lower than those of Post et al. (2007). As $\delta^{13}C$ values provided by Trull et al. (2015) were not lipid normalized, acidified $\delta^{13}C$ values for all zooplankton size-fractions were indicated in Table 2, along with lipid normalized $\delta^{13}C$ values, to allow comparisons between the two data sets.

To consider the relationships between zooplankton and phytoplankton, we used the groups of stations (T-Groups) defined by Trull et al. (2015) based on chemometric measurements of phytoplankton. The HNLC reference station R2 belonged to T-Group1, along with station TEW4. Stations located on the plateau (A3 and E4) and in the eddy (E1 to E5) are included in T-Group 2 and T-Group 3, respectively. The two most easterly stations, located in the open ocean near the polar front (FL and TEW8), belonged to T-Group 5. Trull's T-Group 4 corresponded to coastal stations not sampled for zooplankton analysis.

2.7 Data analysis

The effect of stations and dates (n=12) on zooplankton abundance and biomass was tested statistically using one-way ANOVA with the statistical software Statistica v7. The statistical significance was tested at the 95% confidence level. Community patterns for taxa abundance were explored using the Primer (V6) software package which has been shown to reveal patterns in zooplankton communities (e.g. Clarke and Warwick, 2001; Wishner et al., 2008). Data sets were power transformed (4th root), and the Bray–Curtis dissimilarity index between stations (Bray and Curtis, 1957) was calculated employing all taxonomic categories that contributed at least 1% to any sample in that dataset. Different groups of zooplankton (BC-Groups) were individualized based on their taxonomic composition. Mean C and N stable isotope values among size-fraction and between day and night within each fraction were compared by one-way ANOVAs followed by Tukey post-hoc tests, after testing for normality by Levene test.

3 Results

3.1. Hydrology and trophic conditions

The KEOPS2 campaign was characterized by conditions of complex circulation and rapidly changing phytoplankton biomass (see Trull et al, 2015, their Figs. 1 and 2 and Suppl.). During the survey, the horizontal circulation patterns was dominated by the northernmost branch of the PF (Park et al., 2014) flowing across the plateau in the narrow, mid-depth (1000 m) channel just to the south of Kerguelen Island (Fig. 1). After passing to the east of the plateau, the jet flows outside the shelfbreak northwards and enters in a bathymetrically trapped cyclonic recirculation systems (d'Ovidio et al., 2015; Park et al., 2014a, Trull et al 2015). The variations of the PF position during the KEOPS2 survey are documented in Trull et al. (2015, in supplement). The PF jet separated the central plateau and offshore stations to the south (A3, TNS 10 to TNS 3, TEW3 to TEW6, and E stations) from those to the north and east (TNS1,TNS2, TEW7, TEW8, FL) and to the coast (TEW1 and TEW2).

At the beginning of our study (during the visit to Station A3-1 and to TNS transect), non-significant chlorophyll accumulation was visible from satellite images (see complementary information on satellite-image-derived primary production supplied by Trull et al. 2015, their Fig.2 and supplement), but the sampling at the first visit to station A3 (A3-1, 20th of October) revealed pre-bloom conditions on the plateau and some stations (TNS9, TNS4) of oceanic waters (Jouandet et al, 2015; Lasbleisz et al., 2014). The bloom really started in early November, first massively on the plateau and in coastal waters, and secondly in spatially heterogeneous low biomass in oceanic waters (during our TEW transect and stations E1-E3), with higher chlorophyll values at stations (TEW 7, TEW 8, F-L) downstream polar-front bloom (Lasbleisz et al., 2014; Trull et al. 2015). In mid-November, the central plateau bloom was well-developed (Station A3-2) and afterwards started to decrease slightly, whereas the downstream polar-front bloom was most extensive south of PF and showed its highest biomass there (stations E4-5).

The vertical depth stratification was variable over both space and time (see Trulls et al, their table 4a). Station R2 presented a MLD around 117 m. At station A3, the water column was characterized by a deep mixed layer (around 150 m) during the pre-bloom (station A3-1) and early bloom (station A3-2) surveys, with a range of 120 to 171m (Jouandet et al., 2014). The Chl a concentrations showed a fourfold increase from A3-1 (21 October) to A3-2 (15–17 November), with Chl a concentrations at the surface increasing from 0.5 to 2 mg m−3 (Jouandet et al., 2014, their Figs. 1 and 2). The mixed layer depth of the TNS stations south of the PF decreased northward from around 150 m (TNS10) to 100 m (except TNS6), allowing chlorophyll a concentrations between 0.5 to 1.5 mg m−3 (Lasbleisz et al., 2014, their Fig 3).

During the following visits to the region within the recirculation system in the PF meander (square zoom in Fig 1), the MLD progressively decreased between 50 and 100 m (Stations E1, TEW4 to TEW5), and then below 50 m (stations TEW6, E2 to E5, - except E4 decreasing slightly around 70m) with similar chlorophyll a concentrations between 1.0 to 1.5 mg m−3 (Lasbleisz et al., 2014, their Fig 4). The highest chlorophyll a concentrations (values up to 4.7 mg m−3) were found in the 40 upper meters of the 100m water column of the coastal stations (TEW1 and TEW2, Lasbleisz et al., 2014), whereas the TW3 above the shelf break presented lower chlorophyll a concentrations (<1.0 mg m−3) in its 60 m mixed layer, possibly due to its proximity to the PF jet.

The sampled stations in the Subantarctic Mode Water presented very low chlorophyll a concentration in TNS2 (0.6 5 mg m−3 in the upper 60m), but much higher 10 days later, in TEW-7 and TEW-8 (average above 3 mg m^{-3} in the upper 60m, with peak concentrations up to 5.0 mg m−3; Lasbleisz et al., 2014, their Fig. 4).

3.2 Temporal and spatial variations of zooplankton abundance and biomass

Zooplankton abundances and biomass from ZOOSCAN processed samples of the 330 µm mesh net varied from 14 10^3 to 200 10^3 ind m^{-2} (Fig. 2) and from 0.25 to 4.94 g $C.m^{-2}$ (Fig. 3), respectively. Comparisons of abundance (ind m^{-2}) and biomass (g C m^{-2}) between ZOOSCAN-derived data and direct measurements showed that ZOOSCAN-derived data slightly overestimated direct measurements from regression forced through the origin: slope equal to 1.0015 for abundance (R^2 = 0.75, n = 37, $p<0.01$) and slope equal to 1.1246 for dry weight (R^2 = 0.803, n = 19, $p<0.01$).

Abundance values followed a normal distribution pattern with an average of 7310^3 ind m^{-2} (SD: 42). ANOVA with main effects (stations and dates) without interaction showed clear effect for dates ($p<0.05$) but not for stations. All abundance values plotted against dates (Fig. 4A) showed a general increase, and the linear regression (R2 = 0.42, n = 37) predicted a ratio of 3.7 between abundance at the beginning and at the end of the survey. Highest abundance (above the regression line on Fig. 4A) was observed for oceanic stations within the PF meander, both for the stations of the two transects (Stations TNS4, 5, 7, 8, and TEW 4, 6, 7, 8, and stations E, except for E4-West). By contrast, the lowest abundance was found to the east and north of this PF meander, as well as for the first visit to A3. One exception was station TEW5 which presented the lowest abundance whereas nearby spatial and temporal sampling stations presented much higher abundance. Between the two visits to station A3 at the beginning and the end of the survey, the total abundance was multiplied by 3.5.

The fraction 500-1000 µm (see Fig. 3) presented the most abundant number of organisms (62.0% on average), followed by the < 500 µm fraction (18.8% on average), the 1000-2000 µm fraction (14.2% on average) and the > 2000 µm fraction (5.0 % on average). The contribution of the smaller size fraction (< 500 µm) increased with time from the beginning to the end of the survey (8.1% on average), whereas the 500-1000 µm, 1000-2000 µm, and > 2000 µm decreased to 5.0%, 0.8% and 2.3 %, respectively. However, it was not significant in any of the four regressions due to the variability in size distribution between the stations. In addition, no clear diurnal pattern was observed from the day/night samplings performed at 9 sampling dates.

Log-transformed biomass values followed a normal distribution pattern. As for the abundance, ANOVA with main effects (stations and dates) without interaction for biomass values showed an effect for dates ($p<0.05$) but not for stations. Average biomass was 2.32 g $C.m^{-2}$ (SD: 1.33), and the linear regression against time (not significant) predicted a ratio of 1.7 between biomass values at the beginning and the end of the survey (Fig. 4B). However, the biomass ratio between the two visits at station A3 showed an increase of 2.9, whereas the biomass values at station E (the Lagrangian survey) showed a slightly decreasing trend (with the exception of E4-En). The fraction > 2000 µm represented the highest biomass of organisms (57.1% on average), followed by the 1000-2000 µm, 500-1000 µm and < 500 µm fractions with 22.8%, 18.2% and 1.9% on average, respectively (see Fig. 2). None of the regressions between the percentage value and dates presented a significant correlation, and the slopes of the regression were all near to zero for the intermediate size fractions. From the beginning to the end of the survey, the largest size fraction (> 2000 µm) decreased in its contribution to the biomass (-1.5%), whereas the contribution to the biomass increased with time by 0.1%, 0.5% and 0.9%, respectively, for the 1000-2000 µm, 500-1000 µm and < 500 µm fractions.

The total zooplankton biomass values presented a significant correlation ($p<0.01$) with the average chlorophyll concentrations in the 100 upper meters, as well as with the integrated chlorophyll concentrations in the mixed layer depth (Fig. 5). Only stations TEW1 and TEW2 presented low zooplankton biomass for relative high fluorescence concentrations (>1 µg Chl*a* l^{-1}, Fig. 5A), but not versus the integrated Chla biomass in their narrow (<80 m) mixed layer (Fig. 5B).

3.3 Metazooplankton community composition and distribution

From the 330 µm mesh size net, 65 taxa were identified from net tows for the 37 stations of this study (Table 1) with 26 genera/species of copepods. Copepods contributed the

bulk of the zooplankton community abundance with 78.4 % (SD = 13.13%) of the counted organisms over the whole area, and copepodites represented a little more than half of the counted copepods (mean=52.5%, SD = 8.2%). ANOVA with main effects (stations and dates) without interaction showed no effect either for dates or for stations, either for the percentage of copepods against the whole zooplankton abundance, or for the percentage of copepodites stages against the total copepods abundance. Nauplii represented an average 2% of the total abundance, and showed an increasing abundance with time up to 4%, although they were undersampled with our net. The copepod communities was dominated by *Ctenocalanus citer*, followed by *Oithona similis* and *O. frigida*, *Metridia lucens*, *Scolecithricella minor*, *Calanus simillimus*, *Paraeuchaeta* spp., *Rhincalanus gigas*, and near the coastal area *Drepanopus pectinatus*. Other dominant taxa were the different larval stages of euphausiids (eggs, nauplii, metanauplii, proto et metaozoe), appendicularians (*Oïkopleura* spp., *Fritillaria* spp.), chaetognaths, pteropods (*Limacina retroversa*), amphipods (*Themisto gaudicaudii*, *Hyperia* spp.). Radiolarians and foraminifera were regularly sampled as well. In some stations, other taxa occurred in low numbers, such as salps.

With the 120 µm mesh size net, the number of identified taxa for the 37 stations was reduced to 28 taxa (Table 1), strongly dominated by copepod species. Copepod larval forms as nauplii, undetermined copepod nauplii and copepodites, and copepodid stages of *Oithona* sp., *Oncoea* sp., and *Ctenocalanus citer* represented 20.4 % of organisms in 120 µm mesh size nets. Adult forms (73% of the organisms in nets) were mainly from small and medium size copepods such as *Oithona similis* and *O. frigida, Microsetella rosea, Oncaea* spp.,*Triconia* sp., *Microcalanus pygmaeus* and *Scolecithricella minor.* Other dominant taxa in this net were the different larval stages of euphausiids appendicularians, chaetognaths, pteropods (*Limacina antarctica*), as well as, at a few stations, echinoderm larvae.

Comparison between the community compositions in the two nets clearly showed that some key groups were under-sampled in the 330 µm mesh net: mainly the larval stages of many copepods, small copepods such as *Oithona sp. Microsetella rosea, Oncaea* spp *Triconia* sp., *Microcalanus pygmaeus, Ctenocalanus citer.* The impact of 120 µm mesh size and clogging on the larger planktonic organisms was difficult to assess as many groups were in any case in low density in the 330 µm mesh size net, except for the copepods *Clausocalanus laticeps*, *Calanus simillimus* and *Calanoides acutus.*

The taxonomic distributions are presented in more detail for stations A3 (the two visits A3-1 and A3-2) and for stations E3 and E5 in Figure 6 for the four size fractions from the 330

µm mesh size net sample, and only in the small and medium size fractions from the 120µm mesh size net sample.

The distribution pattern from the 330 µm mesh size net samples is first presented below. The zooplankton community structure in A3-1 was numerically dominated by the medium size fraction (nearly comparable to the fraction 500-1000 µm in total abundance in Fig.2) comprising more than 50% of copepods, characterized by the abundant cyclopoïd *Oithona similis*, along with unspecified calanoid copepodites, and the harpacticoid *Microsetella rosea*. The rest of this fraction included metanauplii of euphausiids, appendicularians, ostracods and small chaetognaths. The fraction 'large size' mesozooplankton, similar to the 1000–2000 µm fraction counted with the ZOOSCAN and representing 10.7% of the total abundance, was composed of 98% copepods with some major taxa (*Ctenocalanus citer*, *Metridia lucens*, *Scolecithricella minor*, *Calanus simillimus*, *Scaphocalanus* spp., *Clausocalanus laticeps*), and early copepodites of *Paraeuchaeta* and of *Calanidae*. The highest size fraction was dominated for more than 75% by *Rhincalanus gigas* and amphipods *Hyperia* spp. and *Themisto gaudicaudii*. It corresponds to the fraction >2000 µm from the ZOOSCAN which contributes to two thirds of the mesozooplancton biomass at station A3-1 (see Fig. 2). The lowest size fraction was mainly composed by euphausiid eggs and nauplii, copepod nauplii, small forms of the pteropod *Limacina retroversa* and in small densities foraminifera and radiolarians. As a whole, the mesozooplancton community in A3-1 was mainly composed by herbivorous species in all fractions, such as the copepods R. *gigas*, *C citer*, *O. similis*, *M. rosea*, but also pteropod *L. retroversa*, appendicularians and different nauplii stages of copepods and euphausiids. In lowest densities, omnivores and detritivores (such as the copepods *M. lucens*, *S. minor*, *C. simillimus*) and carnivores (such as chaetognaths and amphipods, and the copepod *Paraeuchaeta*) were found.

During the second visit to station A3 (A3-2), the size distribution in abundance was dominated by fractions with ECD < 1000 µm (up to 83% of the total abundance, see in Fig 3). The taxa distribution in A3-2 differed from the first visit (station A3-1) both in the "small" size fractions by an increase in copepod nauplii and euphausiid eggs, and in the "medium" size fraction by a large proportion of appendicularians and early copepodid stages of copepods. The two largest fractions ("large" and "very large") were not very different at A3-1 and A3-2 in taxonomic composition and distribution (the only difference being the appearance of late larval stages of euphausiid in the "very large" fraction).

The major features in taxonomic changes between stations E3 (4th November) and E5 (18th November) (Fig. 6) were the increasing contribution of calanoid copepodids in the medium and large size fractions, with the concomitant increase of contribution of these

fractions to the total abundance (see also Fig 2), and the increase of euphausiid larvae in the largest fraction. The smaller fraction presented a rather stable distribution of dominant taxa, with copepod nauplii and *Limacina* as dominant groups (Fig. 6). As a whole, while omnivores, carnivores and scavengers are present, the herbivorous component is strongly dominant with all these larval forms. It is of interest to note that the dominant species for the different fractions at E5 were quite similar to those at A3-1, but with the noticeable difference that many larval stages occurred in all size fractions, inducing the highest observed abundance during the survey (see Fig. 2), although finally representing a lower biomass (see Fig.3).

In the 120 µm mesh size net samples, the taxonomic observation generally delivered the same dominant taxa in the medium size fraction as for the 330 µm mesh size net, but with larger proportions of small copepodid forms and small adult copepods, such as *Oncoea* spp. and *Microsetella rosea*. Copepod nauplii and early copepodid contributed with high abundance (see Table 1) to the small size fraction.

To compare the taxonomic composition between all stations, a cluster dendrogram quantifying the compositional similarity of taxa distributions between the different stations was constructed from the Bray-Curtis coefficient using the 330 µm mesh size net samples which presented the largest number of taxa. **Figure 7** presents the cluster dendrogram and its associated 2D multidimensional scaling plot. This analysis showed a high degree of similarity across the whole region related to the initial phase of zooplankton development. The shelf stations presented the highest level of dissimilarity compared to the other stations.

The cluster dendrogram sliced at 80% similarity distinguished two BC-groups : a first one (BC-Group 1, with more than 80% similarity) grouping the oceanic stations within the PF meander and including eastern stations east of PF (FL and TW7), and a second group of dispersed stations (BC-Group 2, with less than 80 % similarity – differences in day-night samplings not being considered in this analysis), including the R2 station on the western side of the Kerguelen plateau characterized by higher abundance of large calanoid copepods such as *Rhincalanus gigas* and *Paraeuchaeta* spp., the TEW1 and TEW2 stations, near the Kerguelen coast and dominated by *Drepanopus pectinatus* and bivalvia meroplanktonic larvae, the TNS1and TNS2 stations in Sub-Antarctic Surface Water waters dominated by medium size cyclopoid and calanoids and larval forms of euphausiids, the A3 and TNS10 stations in the southern part (see detail below), and stations TEW3, TEW5, TEW8, which were characterized by relative differences in very few taxa compared to other stations of the TEW transect (high density of *Metridida lucens* in TEW3, relatively lower density of *Ctenocalanus citer* in TEW5, and high density of *Triconia* sp. in TEW8).

3.4 Isotopic composition of size-fractionated zooplankton and within zooplankton taxa

A wide range of $\delta^{13}C$ (>8‰) and $\delta^{15}N$ (>4‰) values were recorded among zooplankton size-fractions and stations (Table 2). A slight general increase of $\delta^{13}C$ with increasing size-fraction was observed, while the difference was not significant due to wide differences between sites ($F = 1.818$, $p = 0.132$) (Fig. 8-A). A significant increase in $\delta^{15}N$ with increasing size was observed ($F = 11.67$, $p<0.001$), particularly between the two smallest fractions (80-200 µm and 200-500 µm) and the three largest ones (Fig. 8-B). However, no significant difference in mean $\delta^{15}N$ was apparent between the 500-1000 µm and >2000 µm fractions, while the 1000-2000 µm fraction exhibited a slightly lower $\delta^{15}N$ than the two others. Within each size-fraction, no difference was observed between mean day and night $\delta^{13}C$ and $\delta^{15}N$ values ($p>0.05$ for both), in spite of differences at site level (Table 2). Thus, for both $\delta^{13}C$ and $\delta^{15}N$ values, the main difference occurred between the two smallest size classes (<500 µm) and the three largest ones (>500 µm).

At the station level, mean $\delta^{13}C$ and $\delta^{15}N$ values differed. Station R2 presented the lowest mean $\delta^{13}C$ (-25.26‰) and the highest mean $\delta^{15}N$ (4.49‰), while stations FL, TEW-8 and E4-E were characterized by the highest $\delta^{13}C$ (>-21.2‰) and rather high $\delta^{15}N$ values (>4‰). All the other stations exhibited mean $\delta^{13}C$ values (from -23.26‰ to -21.76‰) and a wide range of mean $\delta^{15}N$ values (from 3.63‰ to 4.25‰).

Differences in mean $\delta^{15}N$ between small (<500 µm) and large (>500 µm) zooplankton size-fractions were low in T-Group 1 (0.3‰), increased in T-Group 5 (0.6‰) and were highest at most stations located in the eddy (Fig. 9). This trend suggested higher food overlap among size-fractions in zooplankton associated with phytoplankton T-Group 1 and T-Group 5, and more partitioned food resources in phytoplankton T-Group 2 and T-Group 3, as indicated by a more even increase in $\delta^{15}N$ with zooplankton size at these stations.

The smaller size-fraction (80-200 µm) was differently composed according to stations, being dominated either by diatoms (A3-2, E-4W), foraminifera (A3-1), small copepods (R2), or a mixture of these groups (most stations). Copepods, eggs, thecosome pteropods foraminifera and small aggregates contributed to 200-500 µm fractions. The following size-fractions (500-1000 µm, 1000-2000 µm and >2000 µm) were all dominated by copepods (60-95%), but amphipods, euphausiids, appendicularians and chaetognaths increased in abundance from the 500-1000 µm to the 1000-2000 µm fractions. The largest size-fraction (>2000 µm) was dominated by *Rhincalanus gigas* and euphausiid larvae or juveniles. Large chaetognaths completed this large fraction.

Thus, differences in specific composition of size-fractions, particularly the smallest and the largest, could result in isotopic differences between stations within a size fraction. For example, when diatoms dominated the 80-200 µm fraction, $\delta^{15}N$ values were lower than when composed of foraminifera or small copepods (2-3‰ and 4-4.5‰, respectively).

Groups of organisms individualized in the >2000 µm fraction presented highly different isotopic signatures according to their main feeding behaviours (Table 3). Filtering salps presented the lowest $\delta^{15}N$ (<4‰), the mostly herbivorous copepods, amphipods, euphausiids, and pteropods intermediate values (4 to 4.6‰), while predatory chaetognaths, fish larvae and polychaetes exhibited higher $\delta^{15}N$ values (>5‰). Thus, $\delta^{15}N$ differences of the >2000 µm fraction between stations resulted mainly from the relative contributions of these groups to bulk samples (ex: higher proportion of salps and euphausiids at A3-2, and large chaetognaths at E5). Accordingly, differences in $\delta^{13}C$ values could be linked to difference in both size and composition of the ingested food. The lower $\delta^{13}C$ recorded in gymnosomes and copepods suggested the consumption of small phytoplankton particles, while the higher $\delta^{13}C$ of euphausiids suggested a consumption of larger-sized phytoplankton. Higher $\delta^{13}C$ in euphausiids compared to copepods was also observed in Arctic seas (Schell et al., 1998).

4. Discussion

4.1 Zooplankton development during the 2011 early spring bloom in and comparison with other seasons

In high latitudes, zooplankton first increase in abundance more than biomass in response to initial phytoplankton spring bloom due to stimulated reproduction of overwintering adults of dominant copepods. This induces a lag-time in the grazing response of herbivorous zooplankton at the beginning of blooms, which further promotes the rapid phytoplankton accumulation. Higher phytoplankton concentrations then stimulate grazing by overwintering stages and new cohorts which results in build-up of zooplankton biomass. With the succession of new cohorts in full bloom conditions (> 0.8 mg Chl*a* m^{-3}), continuous egg production and individual growth induce proportional increase of abundance and biomass.

Such a response of zooplankton to an early phase of the northeastern Kerguelen bloom was observed during the Lagrangian survey within the stationary meander of the Polar Front (stations E1 to E5, except E4-W, Fig. 2, 3 and 4). The average integrated Chl *a* concentrations were rather low (0.49 to 0.77 µg Chl*a* m^{-3}) for these E stations and but slightly higher than the

previous weeks - transects TNS and TEW- (Lasbleisz et al, 2014). The POC was constant in the surface layer up to E3, with an average of 83 mg C m^{-3}, and then slightly increasing at E4 and E5 (with an average up to 109 mg C m^{-3}) (Lasbleisz et al, 2014). Zooplankton densities increased from 60 10^3 ind m^{-2} (E1-d) to 200 10^3 ind m^{-2} (E5-d) whereas biomass gradually decreased (excepted E4-E-n) from 2.3 g C m^{-2} (E1-d) to 1.7 g C m^{-2} (E5-n). Two processes may favor the shift towards smaller size classes. Firstly, the contribution of the larger size classes to biomass decreased with time (Fig. 3) due to the reduction of initial standing stock of overwintering zooplankton by mortality and by investment in egg production. The dominant overwintering copepods (*Ctenocalanus citer*, *Rhincalanus gigas*) are known to be strong seasonal migrants able to spawn in early spring even at low chlorophyll concentrations (Schnack-Schiel, 2001; Atkinson, 1998), i.e. before the full bloom conditions. Moreover, smaller copepod species and copepodids of large copepods may better exploit these low food concentrations (Atkinson et al., 1996), allowing individuals to develop and grow, whereas large copepods are food limited.

The response to chlorophyll increase in waters above the plateau (station A3 in Fig. 4C) was proportional in abundance and biomass (3-fold higher at A3-2 than at A3-1). Lasbleisz et al. (2014) mention that the Chl*a* increase at station A3-2 was accompanied by an increase of the Phaeo:Chl*a* ratio, reflecting a potential higher grazing activity. The mesozooplankton at A3-2 (see Fig. 6) presented a grazer community structure able to feed on a wide spectrum of cells from small diatoms to phytodetritus aggregates, as observed at this station (Lasbleisz et al., 2014; Laurenceau-Cornec et al., 2014), as well as small nano- / microzooplankton (Christaki et al., 2014) and carnivorous zooplankton. Compared to A3-1, the medium size and small size mesozooplankton fractions had a much larger contribution of microphagous organisms (appendicularians, copepod nauplii, etc.) which could quickly remove the smaller planktonic forms (below 20 µm). The larger zooplankton size fractions were a mixture of efficient grazers on large diatoms (> 20 µm), omnivores and detritivores able to feed on aggregates, and carnivores consuming micro- and mesozooplanckton.

The mesozooplankton biomass stocks observed at the beginning of the KEOPS2 cruise (Table 4) were around 1.7 g C m^{-2} above the plateau (A3) and 1.2 g C m^{-2} in oceanic waters (TNS transect). Oceanic biomass slightly increased during the cruise, except the biomass observed in the eastern bloom (station FL) in the Polar Front Zone (above 4 mg C m^{-2}), and station A3 also presented biomass around 4 mg C m^{-2} at the end of the survey. These different results during KEOPS2 suggest that the zooplankton community is able to respond to the growing phytoplankton blooms earlier on the plateau than in the oceanic waters, where

complex mesoscale circulation stimulates initial more or less ephemeral blooms before a broader bloom extension. Due to our constrained sampling for oceanic stations, it was not possible to determine whether the observed zooplankton biomass variability between oceanic stations was linked to enhanced local production (except for stations near the permanent polar front sustaining high level of production). Our results in the quasi-Lagrangian survey within the meander suggests that the heterogeneous primary production linked to oceanic mesoscale activity in the early bloom phase may stimulate the production of new zooplankton cohorts, without sustaining individual growth, slowing down the built-up of new zooplankton biomass. In addition, potential predation on mesozooplankton by euphausiid populations was expected, from observations of the increasing contribution of euphausiid larval stages in our bongo net samples (see Fig. 6) and of long faecal pellets in gel traps (Laurenceau-Cornec et al., 2014).

In contrast, stations FL (Nov. 6th) and A2 (Nov. 16th) presented the highest biomass (maintained below 5 g C m^{-2}) observed in November (Fig 4 and 5) and a similar ratio of abundance to biomass, around 20 10^{3} ind per g C (Fig. 4C) and a lower contribution of smaller size-fractions (ESD < 1000 µm) to total biomass comparatively to station E5. These characteristics could be the results of a phytoplankton-sustained zooplankton development over the previous weeks.

4.2 Comparison with previous results

If we group our observations of KEOPS1 and KEOPS2 (Table 4), the zooplankton seems to continuously increase from mid-October to early February, with a ratio higher on shelf waters (abundance x20 and biomass x9) than in oceanic waters (abundance x3 and biomass x2.5). After early February, the zooplankton community structure remained rather stable (Carlotti et al., 2008). Over the whole spring to summer seasons, the small size fractions (< 500 µm and 500-1000 µm) significantly contribute to the increase in abundance (from 70% to 85 %), with the production of calanoid copepod larval stages and large numbers of cyclopoid copepods, whereas the increase in biomass is mainly due to the fraction 1000-2000 µm with calanoid copepod late larval stages (with a contribution doubling from spring to summer). The taxonomic composition did not show major differences between shelf and oceanic waters, except that the contribution of copepods to the whole mesozooplankton was higher in oceanic waters than on the shelf, and these taxonomic patterns were quite similar between the KEOPS 1 (see Fig. 7 in Carlotti et al. 2008) and KEOPS2 survey (Fig. 6).

The use of different laboratory technologies (Lab OPC during KEOPS1 and ZOOSCAN during KEOPS2) to optically measure and size plankton organisms from net tow samples might be questionable. In their comparative study between LOPC and ZOOSCAN, Schultes and Lopes (2009) found good agreement in the normalized biomass size spectra (NBSS) for particles in the size range of 500 to 1500 µm in equivalent spherical diameter (ESD). Several disparities for smaller and larger particles size range in their study were due both to in situ sampling (LOPC and net have different sampling efficiencies), in situ vs lab counting (LOPC counts any particles, not only zooplankton, with potential overlapping between particles, whereas ZOOSCAN samples are carefully distributed on a scanned window), etc. Our present comparison of estimated abundance and biomass for KEOPS1 and KEOPS2 is based on similar sampling protocols with a 330-µm mesh net on Bongo frame, and in both cases a delicate laboratory protocol. The flow-through system used with the Lab-OPC for KEOPS1 samples was controlled to avoid coincidence of organisms counted by the laser (count rate at 20 particles min^{-1}; see Carlotti et al. 2008) and organisms were carefully separated on the ZOOSCAN window for the KEOPS2 samples. In both studies, a large number of individuals were counted (1000 particles per samples) to correctly count and size larger organisms. Finally, the lower and higher range of counted and measured zooplankton organisms are mainly due to the 330-µm mesh net efficiency, and the abundance and biomass results of both studies might be compared.

In addition to the recent survey of the CPR data for the region (see in Introduction), which shows the strong development of mesozooplankton abundance in October-November, the overall results of KEOPS 1 and 2 in terms of seasonal changes in abundance and biomass values are highly consistent with the information provided by Semelkina (1993) and Razouls et al. (1996, 1998). During the SKALP cruises, all around the Kerguelen Islands (46–52°S, 64–73°E), Semelkina (1993, her Table 1) observed an increase from 62 10^3 ind m^{-2} in July-August 1987 (average values between 0 and 200 m depth for the whole sampled area, nearly double from 0-1000 m) to 570 10^3 ind m^{-2} in February 1988 (values between 0 and 200 m, 100 10^3 ind m^{-2} more in the layer 200-400 m). In terms of biomass, assuming a carbon content to be 50% of body dry weight, the biomass increase in the upper 200 meters was from 2.2 g C m^{-2} to 19 g C m^{-2}. The sampled areas during the SKALP cruises covered a much larger area than that studied during KEOPS2, but these average values corresponded to those observed on eastern side of the Kerguelen Islands (see Semelkina 1993, her Fig. 2). Concerning the taxonomic composition of the mesozooplankton, this author mentioned no seasonal variations but differences in population development and distribution.

Razouls et al. (1998) presented the seasonal changes in copepod distributions at the KERFIX station, a fixed time-series station, situated 60 miles southwest of the Kerguelen Islands (50°40'S, 68°25'E), in 1700 m of water, characteristic of the Permanently Open Ocean Zone (POOZ). The copepod abundance sampled from vertical hauls (300 m – surface) ranged from less than 30 10^3 ind m^{-2} in winter and 45 10^3 ind m^{-2} in October up to 222 10^3 ind m^{-2} in January. The nearest station during KEOPS1 and 2 was station R2 which presented biomasses (respectively abundance densities) of 10.7 g C m^{-2} (272 10^3 ind m^{-2}) in February 2005 and 4.5 g C m^{-2} (80 10^3 ind m^{-2}) in November 2011. Abundances during KEOPS1 and 2 were largely dominated (> 80 %) by copepods (Carlotti et al., 2008, their Fig. 7; distribution not shown for KEOPS2). In addition, during a coastal annual survey in Morbihan Bay at the Kerguelen Islands, Razouls et al. (1996) found a ratio of 10 between winter and spring-summer mesozooplankton density (from 2 to 20 10^3 ind m^{-3}) and a ratio of 20 for the corresponding biomass (from 20 to 400 mg DW m^{-3}).

4.3 Effects of primary production on trophic pathways through mesozooplankton

The KEOPS2 cruise illustrates the complexity of the phytoplankton bloom in spring in the oceanic waters of the Kerguelen Islands, linked to the intense mesoscale activity both in species diversity and spatial production. Comparatively, the mesozooplankton presents initial standing biomass stocks between 1 and 2 g C m^{-2} everywhere in the region, ready to exploit any new phytoplankton production. When this occurs, the initial response is to produce new cohorts which grow further as the bloom builds up, delaying the major grazing impact when these cohorts reach the later stages. Sustained full blooms at plateau stations or permanent fronts favor the highest and longest secondary production rate. The spring period usually shows the greatest increase in grazing pressure on phytoplankton (Razouls et al., 1998).

The comparison of the sinking particle composition at early and advanced stages of the bloom at station A3 (Laurenceau-Cornec et al., 2014 for KEOPS2; Ebersbasch et al., 2008 for KEOPS1) shows that early bloom stage is characterized with particles dominated by phyto-aggregates due to relatively weak grazing pressure on phytoplankton stocks, whereas faecal aggregates characterized the vertical matter flux as soon as zooplankton grazing affects substantially the phytoplankton stock.

The qualitative composition of the bloom had a direct impact in terms of species dominance (mostly herbivorous species) and biochemical composition of the zooplankton

organisms. The spatial differences observed in isotopic signatures of phytoplankton were tracked up to the higher zooplankton levels and showed the impact of the food source.

Differences in the isotopic ratios of zooplankton were observed between stations during the KEOPS2 survey. Station R2 exhibited a 2.2‰ lower $\delta^{13}C$ than stations located on the plateau (A3) or in the eddy (E1 to E5), while $\delta^{13}C$ of stations located in the open ocean (FL) was increased by ~1.5‰ compared to them. A similar increase in carbon isotopic signature was observed by Trull et al. (2015) for phytoplankton, with the lowest $\delta^{13}C$ at the HNLC reference station (R2) and the highest at stations located in the open ocean downstream near the polar front (FL, TEW8) (Fig. 10). The trophic relationship between mesozooplankton and phytoplankton (Trull et al. 2015) was evidenced by the significant positive correlation of their $\delta^{13}C$ values ($\delta^{13}C_{Zooplankton} = 0.745\ \delta^{13}C_{Phytoplankton} - 5.465$, $r = 0.85$, $p < 0.001$). As shown on Fig. 10, mean $\delta^{13}C$ values of zooplankton were related to those of phytoplankton, testifying to the consumption of phytoplankton by zooplankton at station level. The mean trophic fractionation factor from phytoplankton to zooplankton was 0.40 ± 0.71 ‰ for $\delta^{13}C$ and 2.69 ± 0.65 ‰ for $\delta^{15}N$. These values corresponded to a mean increase lower than one trophic level, if we apply the commonly used trophic fractionation factors (1 ‰ for $\delta^{13}C$ and 3.14 ‰ for $\delta^{15}N$) that are in agreement with previous studies on zooplankton (Fry and Quinones, 1994). Such low values again indicated a dominance of herbivory among zooplankton organisms, which confirmed the conclusions based on zooplankton composition. The mean increase in $\delta^{15}N$ in small zooplankton size classes (from 80-200 µm to 500-1000 µm) was higher than among larger size-fractions (from 500-1000 µm to >2000 µm) (1‰ and 0.28‰ respectively). This lower increase in mean $\delta^{15}N$ from 500-1000 µm to >2000 µm suggested a high food overlap among the three largest size-fractions, with a dominance of herbivorous organisms. Within the largest size fraction (>2000 µm), an increase in trophic level ($\delta^{15}N$) was observed from filtering (salps) and mostly herbivorous organisms (copepods, pteropods, etc.) to predatory carnivores (chaetognaths), as observed in other regions (Tarling et al., 2012; Banaru et al., 2014). While different feeding behaviours can be observed among euphausiids (Mauchline, 1980), most euphausiids collected during KEOPS2 survey were mainly herbivores or omnivores with a $\delta^{15}N$ varying between 3.5‰ and 5.0‰ for individuals >2000 µm, a range value already observed in the Southern Ocean (Gurney et al., 2001; Schmidt et al., 2003). High feeding overlap across size-fractionated zooplankton is reported in most studies (Fry and Quiñones, 1994; Bode et al., 2007) and may increase during food shortage (Tarling et al., 2012; Banaru et al., 2014). During KEOPS2, the highest food overlap among zooplankton size-fractions seemed to be associated with phytoplankton T-Group 1 and T-Group 5 in which small-sized cells dominated, while more partitioned food resources among

size-fractions seemed to occur in zooplankton associated with phytoplankton T-Group 2 and T-Group 3, where large phytoplankton cells dominated (Trull et al., 2015).The direct comparison between stable isotope values of size-fractionated zooplankton and their abundance or biomass in water masses is difficult. Zooplankton isotopic values are firstly related to those of the phytoplankton they feed on, themselves linked to water characteristics and nutrient cycling (Trull et al., 2015). The stable isotope values recorded during the KEOPS2 survey suggest a general increase in herbivory in zooplankton during the bloom in accordance with the increase in the abundance of small-sized zooplankton, and corroborate the finding of Lasbleisz et al. (2015) based on the Phaeo : Chl*a* ratio.

5 Conclusions

The complexity of the oceanic processes inducing the large scale phytoplankton bloom in the eastern area of the Kerguelen Islands occurs over scales ranging from the very large (1000s of kilometers) down to the submesoscales (10s of kilometers), marked by intense oceanic–plateau interactions linked to the meandering circulation of the Polar Front (PF) and by a myriad of secondary circulations linked to circulations resulting in a patchy distribution of the new production with different intensity and duration. The KEOPS2 survey addressed the challenge of examining the large-scale phytoplankton bloom that forms over and downstream of the Kerguelen plateau at the most productive season, but also of carrying out observations at a finer resolution in order to understand the influence of spatial and temporal variability of biogeochemical and biological processes on the overall regional ecosystem dynamics and carbon export.

Our results on the mesozooplankton dynamics during KEOPS2 suggest that the zooplankton community maintains relatively high winter stocks both on the plateau and in the oceanic waters, mostly distributed in mesopelagic waters, ready to exploit the early phytoplankton blooms. The timing and intensity of the bloom on the plateau allow an earlier and longer period favorable for zooplankton development and growth compared to the surrounding oceanic waters. A longer lag-time (several weeks) between an initial reproduction phase of the zooplankton organisms and the biomass increase, and thus their grazing impact, was clearly observed in oceanic waters.

References

Atkinson, A.: Life cycle strategies of epipelagic copepods in the Southern Ocean, Journal of Marine Systems, 15, 1-4, 289-311, 1998.

Atkinson, A., Shreeve, R. S., Pakhomov, E. A., Priddle, J., Blight, S. P., and Ward, P.: Zooplankton response to a phytoplankton bloom near South Georgia, Antarctica, Mar. Ecol. Prog. Ser., 144, 195–210, 1996.

Banaru, D., Carlotti, F., Barani, A., Gregory, G., Neffati, N., and Harmelin-Vivien, M.: Seasonal variation of stable isotope ratios of size-fractionated zooplankton in the Bay of Marseille (NW Mediterranean Sea), J. Plankton Res., 36, 145-156, 2014.

Blain, S., Quéguiner, B., Armand, L., Belviso, S., Bombled, B., Bopp, L., Bowie, A., Brunet, C., Brussaard, C., Carlotti, F., Christaki, U., Corbiere, A., Durand, I., Ebersbach, F., Fuda, J. L., Garcia, N., Gerringa, L., Griffiths, B., Guigue, C., Guillerm, C., Jacquet, S., Jeandel, C., Laan, P., Lefèvre, D., Lo Monaco, C., Malits, A., Mosseri, J., Obernosterer, I., Park, Y. H., Picheral, M., Pondaven, P., Remenyi, T., Sandroni, V., Sarthou, G., Savoye, N., Scouarnec, L., Souhaut, M., Thuiller, D., Timmermans, K., Trull, T., Uitz, J., van Beek, P., Veldhuis, M., Vincent, D., Viollier, E., Vong, L., and Wagener, T.: Effect of natural iron fertilization on carbon sequestration in the southern ocean, Nature, 446, 1070–1074, 2007.

Blain, S., Quéguiner, B., and Trull, T.: The natural iron fertilization experiment KEOPS (Kerguelen Ocean and Plateau Compared Study): an overview, Deep-Sea Res. Pt. II, 55, 559–565, 2008.

Blain, S., Renaut, S., Xing, X., Claustre, H., and Guinet, C.: Instrumented elephant seals reveal the seasonality in chloropyll and light-mixing regime in the iron fertilized Southern Ocean, Geophys. Res. Lett., 40, 1–5, 2013.

Bode A., Alvarez-Ossorio, M.T., Cunha, M.E., Garrido, S., Peleteiro, J.B., Porteiro, C., Valdés, L., and Varela, M.: Stable nitrogen isotope studies of the pelagic food web on the Atlantic shelf of the Iberian Peninsula, Prog. Oceanogr., 74, 115-131, 2007.

Bodin, N., Le Loc'h, F., and Hily, C.: Effect of lipid removal on carbon and nitrogen stable isotope ratios in crustacean tissues, J. Exp. Mar. Biol. Ecol., 341, 168–175, 2007.

Bray, J. R. and Curtis, J. T.: An ordination of the upland forest communities of southern Wisconsin, Ecol. Monogr., 27, 273–279, 1957.

Carlotti, F., Botha, D., Nowaczyck, A., and Lefèvre, D.: Structure, biomass, feeding and respiration of the mesozooplankton community during KEOPS, Deep-Sea Res. Pt. II, 55, 720–733, 2008.

Christaki, U., Lefèvre, D., Georges, C., Colombet, J., Catala, P., Sime-Ngando, T., Blain, S., and Obernosterer, I. : Microbial food web dynamics during spring phytoplankton blooms in the naturally iron-fertilized Kerguelen area (Southern Ocean), Biogeosciences, 11, 2014.

Clarke K. R., and Warwick R. M.: Change in Marine Communities: an Approach to Statistical Analysis and Interpretation. 2nd edn. Plymouth, UK: Primer-E Ltd; 2001.

De Niro, M. J., and Epstein, S.: Influence of diet on the distribution of carbon isotopes in animals, Geochim. Cosmochim. Ac., 42, 495–506, 1978.

d'Ovidio, F., Della Penna, A., Trull, T. W., Nencioli, F., Pujol, I., Rio, M. H., Park, Y.-H., Cotté, C., Zhou, M., and Blain, S.: The biogeochemical structuring role of horizontal stirring: Lagrangian 5 perspectives on iron delivery downstream of the Kerguelen plateau, Biogeosciences Discuss., 12, 779–814, doi:10.5194/bgd-12-779-2015, 2015.

Fry, F., and Quiñones, R.. B.: Biomass spectra and stable isotope indicators of trophic level in zooplankton of the northwest Atlantic, Mar. Ecol. Prog. Ser., 112, 201–204, 1994.

Gorsky, G., Ohman, M.D., Picheral, M., Gasparini, S., Stemmann, L., Romagnan, J.B., Cawood, A., Pesant, S., Garcia-Comas, C., and Prejger, F.: Digital zooplankton image analysis using the ZooScan integrated system, J. Plankton Res., 32, 285–303, 2010.

Grosjean, P., Picheral, M., Warembourg, C., and Gorsky, G.: Enumeration, measurement, and identification of net zooplankton samples using the ZOOSCAN digital imaging system, ICES J. Mar. Sci., 61, 518–525, 2004.

Gurney, L.J., Froneman, P.W., Pakhomov, E.A., and McQuaid, C.D.: Trophic positions of three euphausiid species from the Prince Edward Islands (Southern Ocean): implications for the pelagic food web structure, Mar. Ecol. Prog. Ser., 217, 167–174, 2001.

Hindell, M.A., Bost, C.A., Charrassin, J.B., Gales, N., Lea, M.A., Goldsworthy, S., Page, B., Robertson, G., Wienecke, W., O'Toole, M,. and Guinet, C.: Foraging habitats of top predators, and areas of ecological significance, on the Kerguelen Plateau. In: The Kerguelen Plateau: marine ecosystem and fisheries, (Duhamel G. & Welsford D. Eds), Société d'Ichtyologie, 203-215, 2011.

Hosie, G.W., Fukuchi, M., and Kawaguchi, S.. Development of the Southern Ocean Continuous Plankton Recorder Survey, Prog. in Oceanogr., 58, 263–283, 2003.

Hunt, B.P.V., and Hosie, G.W.: Seasonal zooplankton community succession in the Southern Ocean south of Australia, Part I: The Seasonal Ice Zone, Deep-Sea Res. Pt I 53, 1182-1202, 2006a.

Hunt, B.P.V., and Hosie, G.W.: Seasonal zooplankton community succession in the Southern Ocean south of Australia, Part II: The Sub-Antarctic to Polar Frontal Zones, Deep-Sea Res. Pt I, 53, 1203-1223, 2006b.

Jouandet, M-P., Jackson, G., Carlotti, F., Picheral, M., Stemmann, L., Blain, S.: Rapid formation of large aggregates during the spring bloom of Kerguelen Island: observations and model comparisons, Biogeosciences,11, 4949-4993, 2014.

Lasbleiz, M., Leblanc, K., Blain, S., Ras, J., Cornet-Barthaux, V., Helias Nunige, S., and Queguiner, B.: Pigments, elemental composition (C,N,P,Si) and stoichiometry of particulate matter, in the naturally iron fertilized region of Kerguelen in the Southern Ocean, Biogeosciences, 11, 2014.

Laurenceau-Cornec, E., Trull, T. W., Davies, D. M., Bray, S. G., Doran, J., Planchon, F., Carlotti, F., Jouandet, M.-P., Cavagna, A.-J., Waite, A. M., and Blain, S.: Importance of ecosystem structure to carbon export: insights from free-drifting trap deployments in naturally iron-fertilised waters near the Kerguelen plateau, Biogeosciences, 11, 2014.

Lee, R. F., and Hagen, W., and Kattner, G.: Lipid storage in marine zooplankton, Mar. Ecol. Prog. Ser., 307, 273–306, 2006.

Mauchline, J.: The biology of euphausiids, Adv. Mar. Biol., 18, 370–637, 1980.

Motoda S (1959) Device of simple plankton apparatus. Mem Fac Fish Hokkaido Univ 7: 73–94.

Nowaczyk, A., Carlotti, F., Thibault-botha, D., and Pagano, M.: Distribution of epipelagic metazooplankton across the Mediterranean Sea during the summer BOUM cruise, Biogeosciences, 8, 2159-2177, 2011.

Park, Y.-H., Durand, I., Kestenare, E., Rougier, G., Zhou, M., d'Ovidio, F., Cotté, C., and Lee, J.-H..: Polar Front around the Kerguelen Islands: An up-to-date determination and associated circulation of surface/subsurface waters, Journal of Geophysical Research: Oceans, *119*,doi:10.1002/2014JC010061., 2014.

Post, D.M., Layman, C.A., Arrington, D.A., Takimoto, G., Quattrochi, J., and Montaña, C.G.: Getting to the fat of the matter: models, methods and assumptions for dealing with lipids in stable isotope analyses, Oecologia 152, 179–189, 2007.

Quéguiner, B.: Iron fertilization and the structure of planktonic communities in high nutrient regions of the Southern Ocean, Deep-Sea Res. Pt. II, 90, 43–54, 2013.

Razouls C., de Bovée F., Kouwenberg J., and Desreumaux N.: Diversity and Geographic Distribution of Marine Planktonic Copepods, 2014. Available at http://copepodes.obs-banyuls.fr/en

Razouls, S., Du Réau, G., Guillot, P., Maison, J., and Jeandel, C.: Seasonal abundance of copepod assemblage and grazing pressure in the Kerguelen Island area (Southern Ocean), Journal of Plankton Res., 20, 1599–1614, 1998

Razouls, S., Koubbi, P., and Mayzaud, P.: Spatio-temporal distribution of mesozooplankton in a sub-Antarctic coastal basin of the Kerguelen Archipelago (southern Indian Ocean), Polar Biology, 16, 581-587, 1996.

Riandey, V., Champalbert, G., Carlotti, F., Taupier-Letage, I., and Thibault-Botha, D.: Zooplankton distribution related to the hydrodynamic features in the Algerian Basin (western Mediterranean Sea) in summer 1997, Deep-Sea Res. Pt. I, 52, 2029– 2048, 2005.

Rose, M.: Copépodes pélagiques, Faune de France, 26, 374 pp., 1933.

Schell, D. M., Barnett, B. A., and Vinette, K. A.: Carbon and nitrogen isotope ratios in zooplankton of the Bering, Chukchi and Beaufort seas, Mar. Ecol. Prog. Ser., 162, 11–23, 1998.

Schlitzer, R.: Carbon export fluxes in the Southern Ocean: result from inverse modelling and comparison with satellite based estimates. Deep-Sea Research II 49, 1623–1644, 2002.

Schmidt, K., Atkinson, A., Stubing, D., McClelland, J.W., Montoya, J.P., and Voss, M.: Trophic relationships among Southern Ocean copepods and krill: some uses and limitations of a stable isotope approach, Limnol. Oceanogr., 48 (1), 277-289, 2003.

Schnack-Schiel, S.B.: Aspects of the study of the life cycles of Antarctic copepods, Hydrobiol. 453–454, 9–24, 2001.

Schultes, S., and Lopes, R. B.: Laser optical plankton counter and ZooScan intercomparison in tropical and subtropical marine ecosystems, Limnol. Oceanogr. Methods, 7, 771–784, 2009.

Semelkina, A.N.: Development of the zooplankton in the Kerguelen Island region in the years 1987–1988. In: Duhamel, G. (Ed.), Campagnes SKALP 1987 et 1988 aux îles Kerguelen à bord des navires ‘SKIF’ et ‘KALPER’, Institut Français pour la recherche et la technologie polaires, Rapports des campagnes à la mer 93–01, pp. 90–103, 1993.

Soreide J.E., Tamelander T., Hop H., Hobson K.A., and Johansen I.: Sample preparation effects on stable C and N isotope values: a comparison of methods in Arctic marine food web studies, Mar. Ecol. Prog. Ser., 328, 17-28, 2006.

Tarling, G.A., Stowasser, G., Ward, P., Poulton, A.J., Zhou, M., Venables, H.J., McGill, R.A.R., and Murphy, E.J.: Seasonal trophic structure of the Scotia Sea pelagic ecosystem considered through biomass spectra and stable isotope analysis, Deep Sea Res. Pt. II, 59– 60, 222–236, 2012.

Thomalla, S. J., N. Fauchereau, S. Swart, and P. M. S. Monteiro: Regional scale characteristics of the seasonal cycle of chlorophyll in the Southern Ocean, Biogeosciences, 8 (10), 2849–2866, 2011.

Trégouboff, G., and Rose, M.: Manuel de planctonologie méditerranéenne, Centre National de la Recherche Scientifique, 587 pp., 1957.

Trull, T.W. , Davies, D., Dehairs F., Cavagna A.J., Lasbleiz, M., Laurenceau-Cornec, E.C., d'Ovidio, F., Planchon, F., Quéguiner, B., and Blain, S.: Chemometric perspectives on plankton community responses to natural iron fertilization over and downstream of the Kerguelen Plateau in the Southern Ocean, Biogeosciences Discuss., 11, 13841–13903, 2014

Wishner, K. F., Gelfman, C., Gowing, M. M., Outram, D. M., Rapien, M., and Williams, R. L.: Vertical zonation and distributions of calanoid copepods through the lower oxycline of the Arabian Sea oxygen minimum zone, Prog. Oceanogr., 78, 163– 191, 2008.

Zhou, M., Zhu, Y., d'Ovidio, F., Park, Y.-H., Durand, I., Kestenare, E., Sanial, V., Van-Beek,P., Queguiner, B., Carlotti, F., and Blain, S.: Surface currents and upwelling in Kerguelen Plateau regions, Biogeosciences Discuss., 11, 6845--6876, doi: 10.5194/bgd-11-6845-2014, 2014.

Acknowledgements

We thank the project coordinator, S. Blain, and the chief scientist on board, B. Quéguiner, for inviting us to take part in the KEOPS2 project and for supplying data. We thank the captain Bernard Lassiette and crew of the R/V Marion Dufresne for their support aboard. We also thank G. Guillou LIENSs Laboratory at the Université de La Rochelle for processing stable isotope analysis, and Tom Trulls and Marc Pagano for valuable comments on the manuscript. We thank the three reviewers for their comments and suggestions, which significantly contributed to improving the quality of the publication, and M. Paul for English proof-reading. This work was supported by the French Research program of INSU-CNRS LEFE−CYBER ("Les enveloppes fluides et l'environnement –Cycles biogéochimiques, environnement et ressources"), the French ANR (Agence Nationale de la Recherche, ANR-10-BLAN-0614 of SIMI-6 program, and ANR-09-CEXC-006-01 to M. Zhou and F. Carlotti), LABEX OT-MED (n°ANR-11-LABX-0061), the French CNES (Centre National d"Etudes Spatiales) and the French Polar Institute IPEV (Institut Polaire Paul−Emile Victor).

Table 1: List of zooplanktonic taxa collected and identified during the 2011 KEOPS2 cruise (average values for the 37 stations, in ind.m^{-3}). Samples from the 330 µm (left) and 120 µm mesh size (right) nets.

	330 µm mesh size net			120 µm mesh size net		
	Adult forms	**Cope-podites stages**	**Nauplii stages**	**Adult forms**	**Cope-podites stages**	**Nauplii stages**
Copepods						
Oithona similis	**8.1**	**2.8**		**489.8**	**1362.5**	
Oithona frigida	**14.8**			**71.5**		
Microsetella rosea	**1.9**			**79.0**		
Oncaea spp.	**1.3**	**0.1**		**58.2**	**53.6**	
Triconia sp.	**8.7**			**20.7**	**11.4**	
Clausocalanus laticeps	**3.9**	**0.8**		**1.5**	**0.1**	
Ctenocalanus citer	**35,8**	**56.5**		**47.8**	**195.7**	
Microcalanus pygmaeus	**0.7**			**23.2**		
Metridia lucens	**9.6**	**8.9**		**4.6**	**39.3**	
Calanus propinquus	**0.02**					
Calanus simillimus	**6.45**	**1.9**		**1.4**	**1.62**	
Calanoides acutus	**1.1**	**1.6**		**0.2**	**0.41**	
Scolecithricella minor	**9.2**	**6.4**		**10.1**	**8.4**	
Scaphocalanus spp.	**0.7**	**2.5**				
Drepanopus pectinatus	**0.6**	**2.7**		**1.4**	**13.7**	
Pleuromamma robusta	**0.9**	**0.2**		**0.4**		
Candacia maxima	**rare**	**rare**				
Heterorhabdus spp.	**rare**	**rare**				
Aetideus armatus	**rare**	**rare**				
Haloptilus oxycephalus	**rare**	**rare**				
Paraeuchaeta spp.	**0.54**	**14,29**			**14.1**	
Rhincalanus gigas	**2.93**	**7.34**	**3.1**	**1.1**	**7.9**	**26.63**
Subeucalanus longiceps	**0.14**	**0.02**		**0.4**		
Euchirella rostramagna	**rare**	**0.04**			**0.04**	
Gaetanus pungens	**rare**					
Undeuchaeta incisa	**rare**					
Undetermined Nauplii			**2.1**			**1071.7**
Undetermined Copepodites		**22.4**			**253.5**	

Table 1: (continued)

	330 µm mesh size net			120 µm mesh size net		
	Adult forms	**Larval forms**	**Eggs**	**Adult forms**	**Larval forms**	**Eggs**
Euphausiids						
Undetermined species	**0.27**	**6.22**	**32.23**		**6.8**	**32.2**
Ostracods	**2.3**			**7.9**		
Isopods	**0.05**					
Mysid		**rare**				
Decapod		**rare**				
Amphipods						
Themisto gaudicaudii	**0.26**					
Hyperia spp.	**0.86**					
Primno macropa	**0.10**					
Vibilia sp.	**rare**	**0.04**				
Scina sp.	**rare**					
Molluscs						
Limacina retroversa	**3.45**			**33.2**		
Limacina helicina	**rare**					
Spongiobranchaea sp.	**rare**					
Clio sp.	**rare**					
Polychaetes						
Pelagobia sp.	**0.22**					
Tomopteris spp.	**rare**					
Travisiopsis sp.	**rare**					
Undetermined	**rare**	**0.32**		**rare**	**9.28**	
Appendicularians	**8.45**			**149.1**		
Thaliacea						
Salpa thompsoni	**0.07**					
Pyrosomid	**rare**					
Ctenophores	**rare**					
Cnidarians						
Undetermined larvae		**rare**			**rare**	
Undetermined adult	**rare**					
Bougainvillia sp.	**rare**					
Dimophyes arctica	**rare**					
Pyrostephos vanhoeffeni	**rare**					
Rosacea plicata	**rare**					
Muggiaea bargmannae	**rare**					
Solmundella bitentaculata	**rare**					
Pegantha sp.	**rare**					
Chaetognaths	**4.15**			**5.7**		
Radiolarians	**0.93**					
Foraminifera	**0.98**					
Meroplankton						
Cirripedia		**rare**				
Echinodermata		**rare**			**11.4**	
Fish		**0.05**	**rare**			
Mysid		**rare**				
Polychaeta		**rare**				
Bivalvia		**rare**				

Table 2. Isotopic composition of size-fractionated zooplankton (mean and standard deviation). mean $\delta^{13}C$: values of acidified samples, mean $\delta^{13}C$-norm.: lipid-normalised values (except for the lowest size-fraction), mean $\delta^{15}N$: values of untreated samples.

Station Date	Fraction µm	mean $\delta^{13}C$ ‰PDB	sd $\delta^{13}C$ ‰PDB	mean $\delta^{13}C$-norm. ‰PDB	sd $\delta^{13}C$-norm. ‰PDB	mean $\delta^{15}N$ ‰air	sd $\delta^{15}N$ ‰air	C/N mass
A3-1 day	80	-25.52	0.10	**-25.52**	0.06	**4.01**	0.05	**5.31**
20/10/2011	200	-26.48	0.05	**-24.00**	0.08	**4.89**	0.12	**5.85**
	500	-26.20	0.05	**-23.10**	0.07	**4.87**	0.02	**6.48**
	1000	-24.52	0.14	**-22.67**	0.12	**3.21**	0.07	**5.22**
	2000	-25.16	0.03	**-22.97**	0.03	**3.58**	0.05	**5.57**
TNS-7 day	80	-23.26	0.06	**-23.26**	0.07	**3.45**	0.09	**4.55**
22/10/2011	200	-25.18	0.03	**-23.70**	0.02	**3.41**	0.15	**4.85**
	500	-25.74	0.06	**-23.29**	2.00	**4.29**	0.04	**5.82**
	1000	-24.76	0.01	**-23.03**	0.00	**4.21**	0.06	**5.10**
	2000	-25.84	0.03	**-23.21**	0.06	**4.60**	0.22	**6.01**
R-2 day	80	-27.93	0.02	**-27.93**	0.03	**4.36**	0.08	**4.87**
26/10/2011	200	-27.69	0.07	**-25.84**	0.10	**4.97**	0.03	**5.23**
	500	-27.11	0.06	**-24.93**	0.15	**4.79**	0.19	**5.55**
	1000	-26.43	0.10	**-24.52**	0.11	**3.24**	0.06	**5.28**
	2000	-26.15	0.06	**-24.59**	0.03	**5.09**	0.10	**4.93**
E-1 night	80	-23.61	0.05	**-23.61**	0.06	**3.62**	0.23	**4.70**
30/10/2011	200	-25.37	0.04	**-23.91**	0.03	**3.04**	0.05	**4.83**
	500	-25.73	0.03	**-23.26**	0.03	**3.67**	0.03	**5.85**
	1000	-25.26	0.05	**-22.80**	0.05	**3.58**	0.01	**5.84**
	2000	-26.27	0.03	**-22.75**	0.08	**4.69**	0.32	**6.91**
E-2 day	80	-24.62	0.04	**-24.62**	0.12	**3.93**	0.19	**4.89**
01/11/2011	200	-25.86	0.02	**-23.48**	0.09	**3.83**	0.03	**5.75**
	500	-25.70	0.04	**-22.77**	0.08	**4.38**	0.08	**6.32**
	1000	-25.54	0.02	**-22.45**	0.01	**3.65**	0.11	**6.47**
	2000	-26.12	0.02	**-22.74**	0.14	**5.48**	0.15	**6.77**
TEW-4 day	80	-25.15	0.01	**-25.15**	0.05	**4.06**	0.11	**4.67**
01/11/2011	200	-25.98	0.01	**-24.73**	0.02	**3.61**	0.23	**4.62**
	500	-25.23	0.02	**-23.39**	0.02	**3.24**	0.06	**5.21**
	1000	-26.01	0.03	**-21.98**	0.04	**3.50**	0.07	**7.42**
	2000	-27.02	0.06	**-21.52**	0.04	**5.23**	0.03	**8.90**
TEW-8 day	80	-22.58	0.05	**-22.58**	0.04	**3.88**	0.03	**5.24**
02/11/2011	200	-23.29	0.03	**-21.00**	0.04	**3.90**	0.03	**5.67**
	500	-23.73	0.04	**-21.62**	0.07	**4.28**	0.08	**5.48**
	1000	-23.60	0.07	**-21.73**	0.08	**4.30**	0.04	**5.24**
	2000	-23.29	0.05	**-21.61**	0.05	**3.78**	0.02	**5.05**
E-3 night	80	-24.70	0.02	**-24.70**	0.03	**3.02**	0.13	**4.84**
03/11/2011	200	-25.79	0.02	**-23.52**	0.03	**3.50**	0.04	**5.65**
	500	-25.60	0.02	**-23.24**	0.03	**4.14**	0.07	**5.74**
	1000	-25.67	0.03	**-22.63**	0.11	**3.67**	0.02	**6.42**
	2000	-25.62	0.04	**-23.20**	0.03	**4.58**	0.35	**5.79**
E-3 day	80	-24.82	0.04	**-24.82**	0.07	**2.98**	0.10	**4.71**
04/11/2011	200	-25.99	0.02	**-23.51**	0.05	**3.58**	0.06	**5.85**
	500	-26.26	0.02	**-22.79**	0.05	**3.90**	0.04	**6.86**
	1000	-25.57	0.03	**-22.41**	0.03	**3.68**	0.06	**6.54**
	2000	-26.71	0.03	**-22.19**	0.06	**5.23**	0.48	**7.92**

FL day	80	-23.69	0.03	**-23.69**	0.05	**3.66**	0.09	**5.87**
06/11/2011	200	-24.06	0.01	**-21.42**	0.06	**4.20**	0.10	**6.02**
	500	-24.59	0.03	**-21.62**	0.14	**5.08**	0.08	**6.35**
	1000	-24.31	0.03	**-21.58**	0.07	**4.44**	0.10	**6.11**
	2000	-24.64	0.01	**-21.48**	0.04	**5.00**	0.17	**6.55**
FL night	80	-21.77	0.02	**-21.77**	0.01	**4.06**	0.10	**4.80**
06/11/2011	200	-23.41	0.03	**-20.96**	0.03	**3.54**	0.03	**5.83**
	500	-24.67	0.08	**-21.01**	0.12	**4.41**	0.09	**7.05**
	1000	-23.75	0.05	**-21.58**	0.06	**4.06**	0.05	**5.54**
	2000	-22.38	0.01	**-21.53**	0.02	**3.61**	0.06	**4.21**
E-4W day	80	-23.26	0.08	**-23.26**	0.02	**3.17**	0.12	**5.14**
11/11/2011	200	-24.66	0.05	**-22.93**	0.05	**3.43**	0.11	**5.10**
	500	-25.05	0.02	**-22.70**	0.02	**3.85**	0.07	**5.73**
	1000	-24.21	0.02	**-22.31**	0.08	**3.97**	0.08	**5.28**
	2000	-25.01	0.02	**-21.38**	0.01	**4.64**	0.17	**7.53**
E-4W night	80	-23.24	0.03	**-23.24**	0.05	**2.97**	0.29	**4.72**
11/11/2011	200	-24.83	0.07	**-23.10**	0.13	**3.33**	0.06	**5.09**
	500	-25.30	0.06	**-22.81**	0.06	**3.94**	0.02	**5.86**
	1000	-24.83	0.07	**-22.52**	0.04	**3.91**	0.04	**5.68**
	2000	-24.92	0.06	**-22.10**	0.07	**3.85**	0.12	**6.20**
E-4E night	80	-23.47	0.04	**-23.47**	0.04	**2.42**	0.14	**5.14**
12/11/2011	200	-25.24	0.04	**-22.59**	0.10	**3.77**	0.06	**6.03**
	500	-26.07	0.04	**-19.61**	0.06	**4.72**	0.14	**9.88**
	1000	-26.02	0.07	**-18.92**	0.45	**4.82**	0.20	**10.53**
	2000	-27.12	0.11	**-17.64**	0.26	**4.76**	0.59	**12.93**
E-4E day	80	-23.65	0.02	**-23.65**	0.03	**3.17**	0.52	**5.53**
13/11/2011	200	-25.32	0.06	**-21.90**	0.11	**4.02**	0.15	**6.81**
	500	-25.97	0.02	**-20.81**	0.17	**4.40**	0.08	**8.56**
	1000	-25.38	0.08	**-21.06**	0.21	**4.63**	0.08	**7.72**
	2000	-25.76	0.11	**-22.67**	0.09	**3.96**	0.49	**6.48**
A3-2 day	80	-22.82	0.09	**-22.82**	0.22	**1.71**	0.17	**4.49**
16/11/2011	200	-23.58	0.02	**-22.42**	0.05	**3.89**	0.02	**4.53**
	500	-24.19	0.04	**-22.38**	0.15	**5.45**	0.16	**5.19**
	1000	-23.44	0.05	**-21.91**	0.04	**4.66**	0.07	**4.89**
	2000	-23.09	0.04	**-21.42**	0.07	**3.71**	0.20	**5.04**
A3-2 night	80	-22.42	0.02	**-22.42**	0.06	**2.43**	0.09	**4.44**
16/11/2011	200	-23.47	0.04	**-22.31**	0.09	**3.98**	0.16	**4.53**
	500	-23.98	0.05	**-22.33**	0.16	**4.90**	0.06	**5.02**
	1000	-24.99	0.04	**-20.38**	0.10	**5.04**	0.04	**8.01**
	2000	-23.22	0.05	**-21.46**	0.04	**4.11**	0.06	**5.13**
E-5 day	80	-25.88	0.06	**-25.88**	0.09	**2.45**	0.01	**3.71**
18/11/2011	200	-26.64	0.36	**-23.91**	0.30	**3.10**	0.22	**6.11**
	500	-26.01	0.03	**-23.04**	0.04	**3.24**	0.17	**6.35**
	1000	-25.89	0.05	**-23.00**	0.09	**3.30**	0.02	**6.27**
	2000	-27.74	0.01	**-21.59**	0.20	**6.19**	0.14	**9.56**
E-5 night	80	-26.18	0.03	**-26.18**	0.07	**2.87**	0.27	**6.01**
19/11/2011	200	-25.90	0.02	**-22.64**	0.05	**3.45**	0.09	**6.64**
	500	-26.07	0.01	**-22.54**	0.04	**3.61**	0.02	**6.92**
	1000	-25.90	0.04	**-22.71**	0.09	**3.76**	0.05	**6.58**
	2000	-27.39	0.02	**-22.83**	0.10	**4.37**	0.38	**7.97**

Table 3. Mean (± SD) stable isotope values of the main groups of organisms sorted in the largest size fraction (>2000 µm). n = number of samples analysed.

Groups	n	$\delta^{13}C$ (‰)	$\delta^{15}N$ (‰)
Salps	12	-22.36 ± 0.82	3.87 ± 1.29
Copepods	15	-21.98 ± 0.95	4.40 ± 0.54
Euphausiacea	12	-21.03 ± 2.34	4.24 ± 0.63
Amphipods	9	-23.19 ± 0.24	4.14 ± 0.41
Pteropods Gymnosoms	5	-23.44 ± 0.04	4.56 ± 0.09
Chaetognaths	12	-22.94 ± 0.18	5.93 ± 0.60
Polychaetes *Tomopteris*	3	-22.52 ± 0.03	7.72 ± 0.06
Fish larvae	3	-21.60 ± 0.05	5.99 ± 0.08

Table 4: Seasonal variations of zooplankton abundance and biomass from KEOPS2 (15 October – 20 November 2011) and KEOPS1 (January 19- February 13, 2005) surveys with contribution of different size fractions (<500 µm, 500-1000 µm; 1000-2000 µm; > 2000 µm). The reference stations were A3 (shelf waters) and C11 (oceanic waters) for KEOPS1 (see Carlotti et al., 2008, their Figs. 3 and 5) , and A3 (shelf waters) and TNS6-TNS5 and E4E-E5 (oceanic waters) for KEOPS2.

			KEOPS 2		KEOPS 1		
Area	Date		20 -22/X	13-16/XI	22-28/I	4-5/II	12/II
Shelf waters	**Abundance**	**X 10^6. m^{-2}**	**26**	**90**	**600**	**700**	**450**
	Percentages of total abundance	< 500 µm	10 %	34 %	55%	46 %	41 %
		500-1000 µm	60 %	50 %	32 %	35 %	44 %
		10002000 µm	23 %	13 %	12 %	18 %	13.5 %
		>2000 µm	7 %	3 %	1 %	< 1 %	1.5 %
	Biomass	**g C m^{-2}**	**1,7**	**4**	**10**	**15**	**9**
	Percentages of total biomass	< 500 µm	<1 %	4 %	7.5 %	5 %	7 %
		500-1000 µm	12 %	17 %	21.5%	23%	26%
		1000-2000 µm	23 %	28 %	45 %	59%	46 %
		2000 µm	64 %	51 %	26 %	12%	21 %
Oceanic waters	**Abundance**	**X 10^6. m^{-2}**	**70**	**150**	**200**	**100**	**-**
	Percentages of total abundance	< 500 µm	18 %	15 %	50 %	47 %	-
		500-1000 µm	66 %	65 %	40 %	41 %	-
		1000-2000 µm	12 %	15 %	10 %	10 %	-
		>2000 µm	4 %	5 %	< 1 %	2 %	-
	Biomass	**g C m^{-2}**	**1,2**	**2**	**4**	**3**	**-**
	Percentages of total biomass	< 500 µm	1 %	2 %	10 %	5 %	-
		500-1000 µm	16 %	16 %	35 %	25%	-
		1000-2000 µm	18 %	24 %	40 %	40 %	-
		2000 µm	65%	58 %	15 %	30 %	-

Figures

Figure 1: Map of the KEOPS2 study area and station locations. The locations of the stations are marked by color dots. The southern station A3 (red dot) was visited twice at the beginning (station A3-1) and the end (station A3-2) of the KEOPS2 survey. This station A3 situated in the middle of the shelf was the reference station for the shelf bloom observed during KEOP1. The stations of the North-South transect in blue dots are located in oceanic waters and were sampled just after the first visit to A3, from south to north (stations TNS1 to TNS10), from the central plateau (TNS-10) across the recirculation feature (TNS7 toTNS3) and polar front (TNS3-TNS2) and into sub-antarctic waters (TNS-1). Station R-2 (black dot) in the west of the Kerguelen plateau represented a HNLC reference station. The transect TEW (transect east–west) was sampled from west to east from the near coast of Kerguelen Island (TEW1) above the shelf (TEW32) and shelf break (TEW3) across the middle of the recirculation system (TEW4 to TEW6), and beyond the southward meandering polar front (TEW7 and TEW8) in the extreme east of the study region. The survey ended with a quasi-Lagrangian time series (stations E1–E5 in orange dots in the zoom panel), during a progressive phase of the bloom within the recirculation system in the meander of the polar front. In addition, one station (Station F-L) situated in high-biomass waters in the extreme northeast of the study region, near the downstream location of the PF, was sampled within the period of the time series.

Figure 2: Integrated 0–200m mesozooplankton biomass estimated from ZOOSCAN for the different stations sampled during KEOPS2 with size fraction distributions. Size fractions: <500 µm: black; 500–1000 µm: dark gray ; 1000–2000 µm: light gray; >2000 µm: white.

Figure 3: Integrated 0–200m mesozooplankton abundance counted from ZOOSCAN for the different stations sampled during KEOPS2 with size fraction distributions. Size fractions: <500 µm: black; 500–1000 µm: dark gray ; 1000–2000 µm: light gray; >2000 µm: white.

Figure 4. (a) Abundance and **(b) b**iomass values and **(c)** ratio abundance on biomass for the different stations visited during KEOPS2 over sampling dates.
Abundance and biomass values from Figures 2 and 3.

Figure 5. Zooplankton biomass values against average Chl *a* in the upper 100m **(a)** and against the integrated Chl *a* in the mixed layer depth **(b)** for the different stations visited during KEOPS2. Biomass values from Figure 2.

Figure 6. Distributions of main taxa abundance at stations A3-1, A3-2, E3 and E5 from binocular observation. Distributions are presented from left to right for the four stations, and from top to bottom for the four size fractions (four upper bands: small, medium, large, and very large) observed in the 330 µm mesh size net samples (), and for the two lower size fractions (two lower upper bands: small and medium) for the 120 µm mesh size net samples. Distributions are average values between day and night samples. For each size fraction (the four pie charts on the same horizontal band), the color labels for the different taxa are similar.

Figure 7. Dendrogram (A) and MDS plot (B) produced by the clustering of the 37 samples (28 stations, among them 9 stations with day-night sampling) during KEOPS2 based on the density (ind.m^{-3}) of mesozooplankton taxa. Density values were fourth-root transformed prior to analysis of the Bray-Curtis similarity matrix. The stress statistic for the MDS plot is 0,12.

Figure 8: Distribution of $\delta^{13}C$ (A) and $\delta^{15}N$ (B) of zooplankton across size-fractions during KEOPS2. White symbols = day; Black symbols = night.

Figure 9: Distribution of $\delta^{13}C$ (left column, a) and $\delta^{15}N$ (right column, b) values across zooplankton size-fractions for 4 of the 5 T-Groups of stations identified by Trull et al (2015) for phytoplankton. Station E4-E is included here in T-Group 5 instead of T-Group 2. From top to bottom: a1 and b1 = T-Group 1 (diamond), a2 and b2 = T-Group 2 (triangle), a3 and b3 = T-Group 3 (dots), a4 and b4 = T-Group 5 (square). T-Group 4 included coastal stations not sampled for zooplankton analysis.

Figure 10: Mean $\delta^{13}C$ and $\delta^{15}N$ values of phytoplankton (5-210 µm) (Trull et al. 2015) and zooplankton (200->2000 µm) (present study) for stations sampled during KEOPS2 cruise. Symbols correspond to the phytoplankton groups based on chemometric measurements identified by Trull et al. (2015). Diamond = T-Group 1; Triangle = T-Group 2; Dots = T-Group 3; Square = T-Group 5.

Figure 1

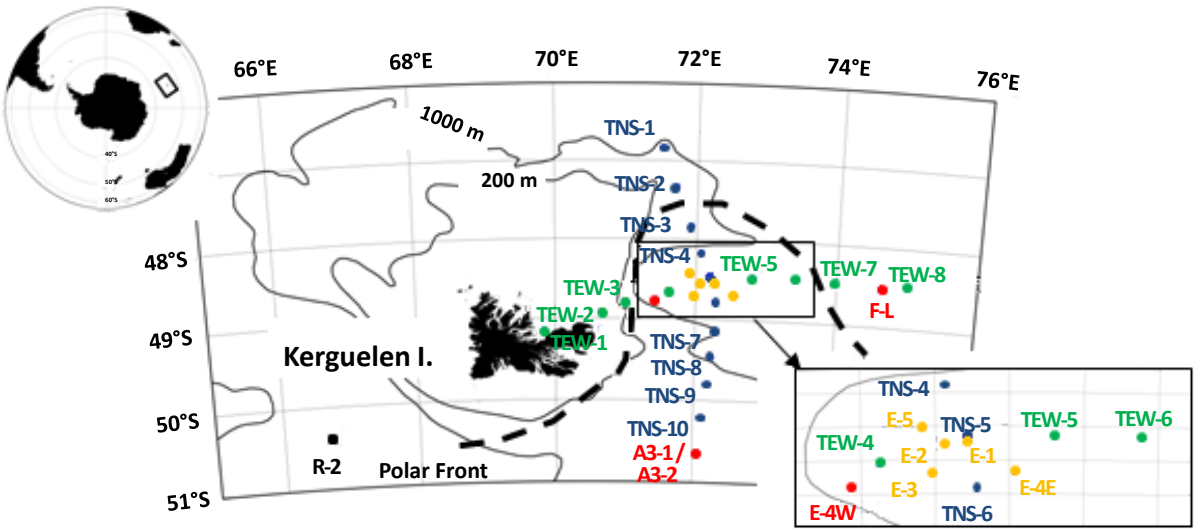
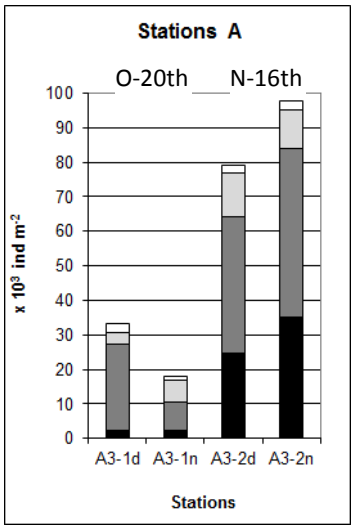
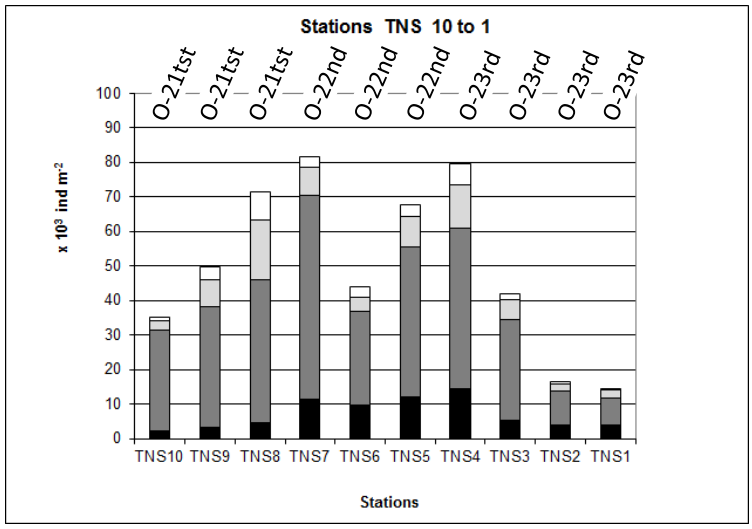
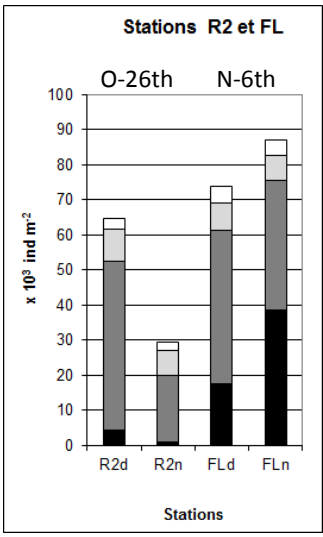
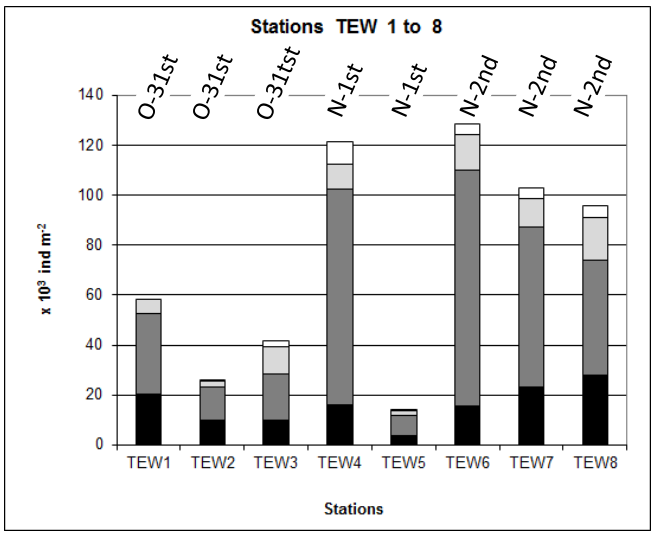

66°E
68°E
70°E
72°E
74°E
76°E
48°S
49°S
50°S
51°S
1000 m
200 m
Kerguelen I.
R-2
Polar Front
TNS-1
TNS-2
TNS-3
TNS-4
TNS-5
TNS-6
TNS-7
TNS-8
TNS-9
TNS-10
TEW-1
TEW-2
TEW-3
TEW-4
TEW-5
TEW-6
TEW-7
TEW-8
F-L
A3-1 / A3-2
E-1
E-2
E-3
E-4E
E-4W
E-5

Figure 2


Stations A
O-20th
N-16th
x 10³ ind m⁻²
A3-1d
A3-1n
A3-2d
A3-2n
Stations


Stations TNS 10 to 1
O-21tst
O-21tst
O-21tst
O-22nd
O-22nd
O-22nd
O-23rd
O-23rd
O-23rd
O-23rd
x 10³ ind m⁻²
TNS10
TNS9
TNS8
TNS7
TNS6
TNS5
TNS4
TNS3
TNS2
TNS1
Stations


Stations R2 et FL
O-26th
N-6th
x 10³ ind m⁻²
R2d
R2n
FLd
FLn
Stations


Stations TEW 1 to 8
O-31st
O-31st
O-31tst
N-1st
N-1st
N-2nd
N-2nd
N-2nd
x 10³ ind m⁻²
TEW1
TEW2
TEW3
TEW4
TEW5
TEW6
TEW7
TEW8
Stations

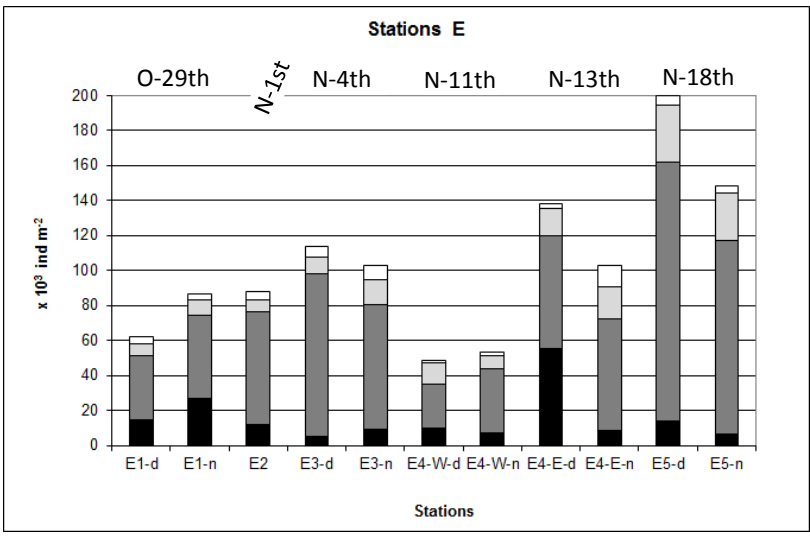

Stations E
O-29th
N-1st
N-4th
N-11th
N-13th
N-18th
x 10³ ind m⁻²
E1-d
E1-n
E2
E3-d
E3-n
E4-W-d
E4-W-n
E4-E-d
E4-E-n
E5-d
E5-n
Stations

Figure 3

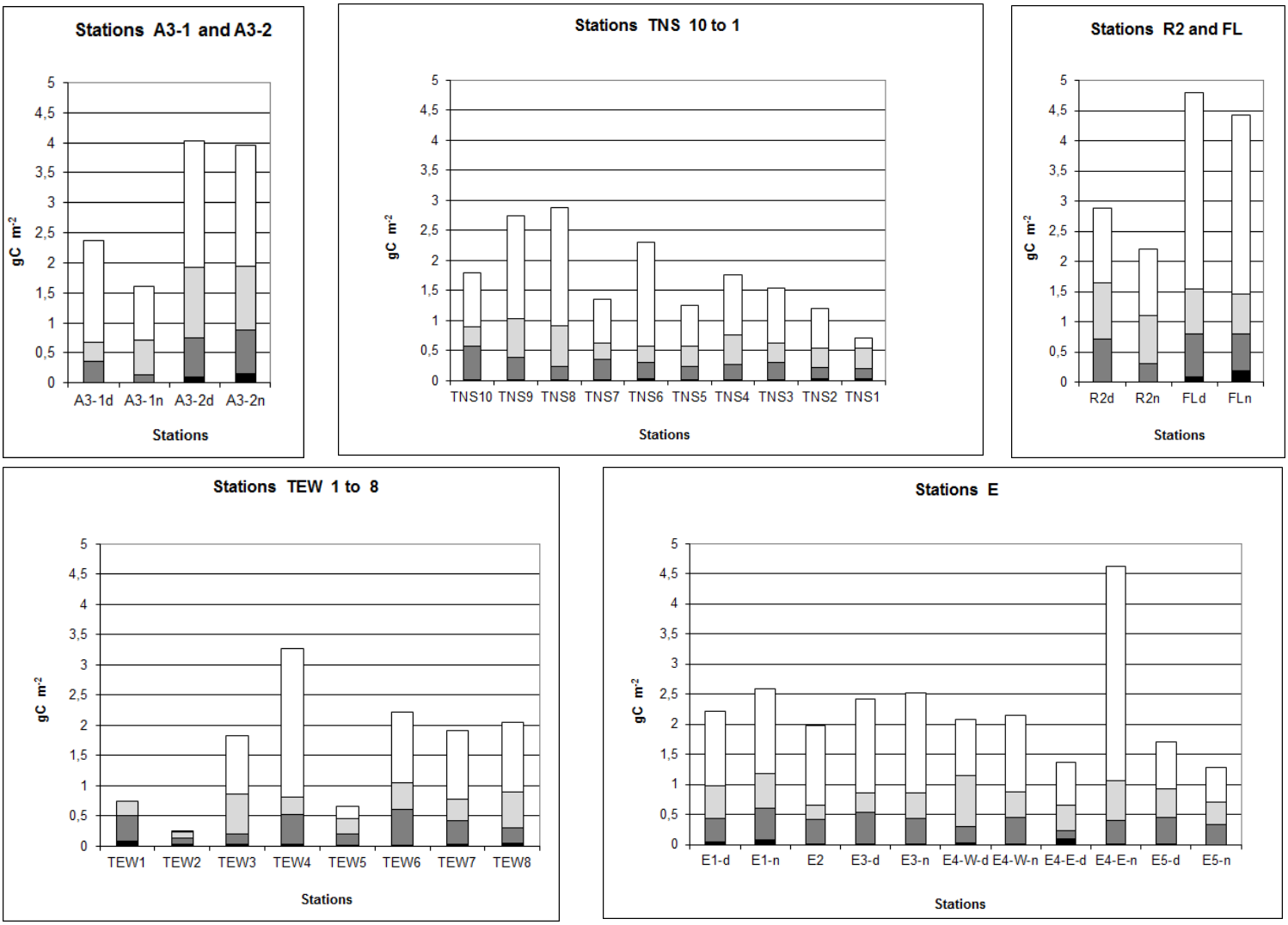

Stations A3-1 and A3-2
gC m-2
A3-1d A3-1n A3-2d A3-2n
Stations
Stations TNS 10 to 1
gC m-2
TNS10 TNS9 TNS8 TNS7 TNS6 TNS5 TNS4 TNS3 TNS2 TNS1
Stations
Stations R2 and FL
gC m-2
R2d R2n FLd FLn
Stations
Stations TEW 1 to 8
gC m-2
TEW1 TEW2 TEW3 TEW4 TEW5 TEW6 TEW7 TEW8
Stations
Stations E
gC m-2
E1-d E1-n E2 E3-d E3-n E4-W-d E4-W-n E4-E-d E4-E-n E5-d E5-n
Stations

Figure 4:

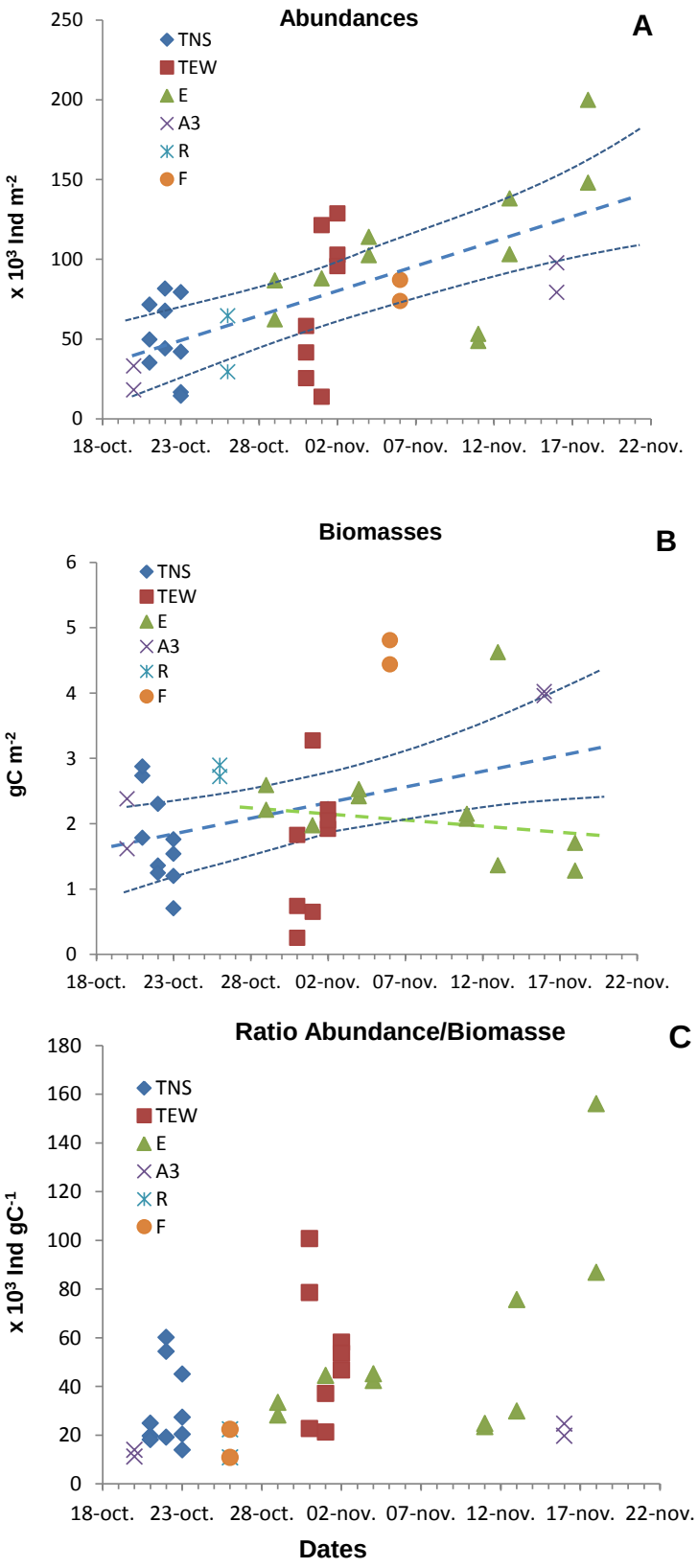

Abundances
A
TNS
TEW
E
A3
R
F
x 10³ Ind m⁻²
250
200
150
100
50
0
18-oct.
23-oct.
28-oct.
02-nov.
07-nov.
12-nov.
17-nov.
22-nov.
Biomasses
B
gC m⁻²
6
5
4
3
2
1
0
Ratio Abundance/Biomasse
C
x 10³ Ind gC⁻¹
180
160
140
120
100
80
60
40
20
0
Dates

Figure 5

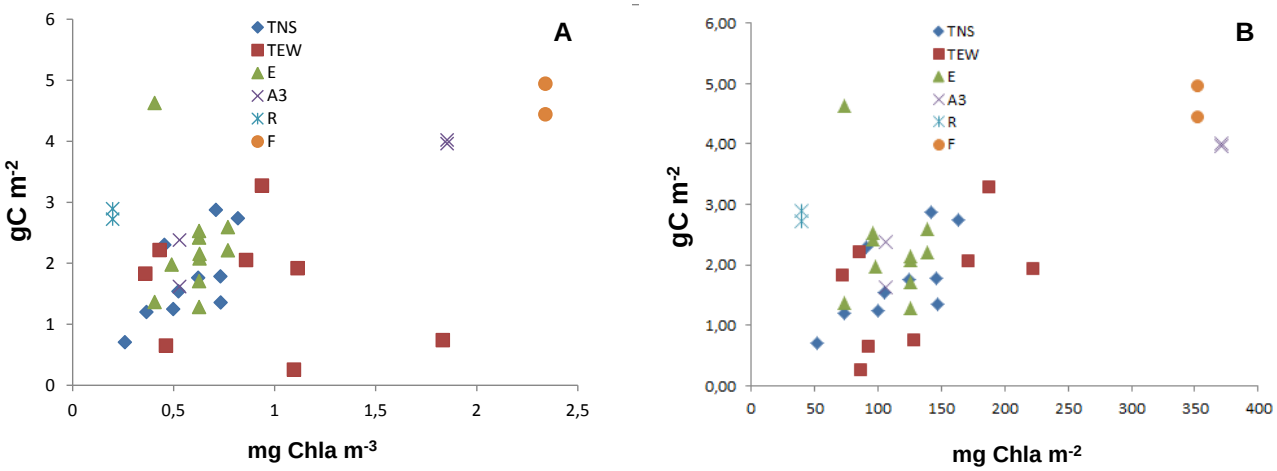

A
TNS
TEW
E
A3
R
F
gC m-2
mg Chla m-3
0
1
2
3
4
5
6
0
0,5
1
1,5
2
2,5
B
TNS
TEW
E
A3
R
F
gC m-2
mg Chla m-2
0,00
1,00
2,00
3,00
4,00
5,00
6,00
0
50
100
150
200
250
300
350
400

Figure 6 .

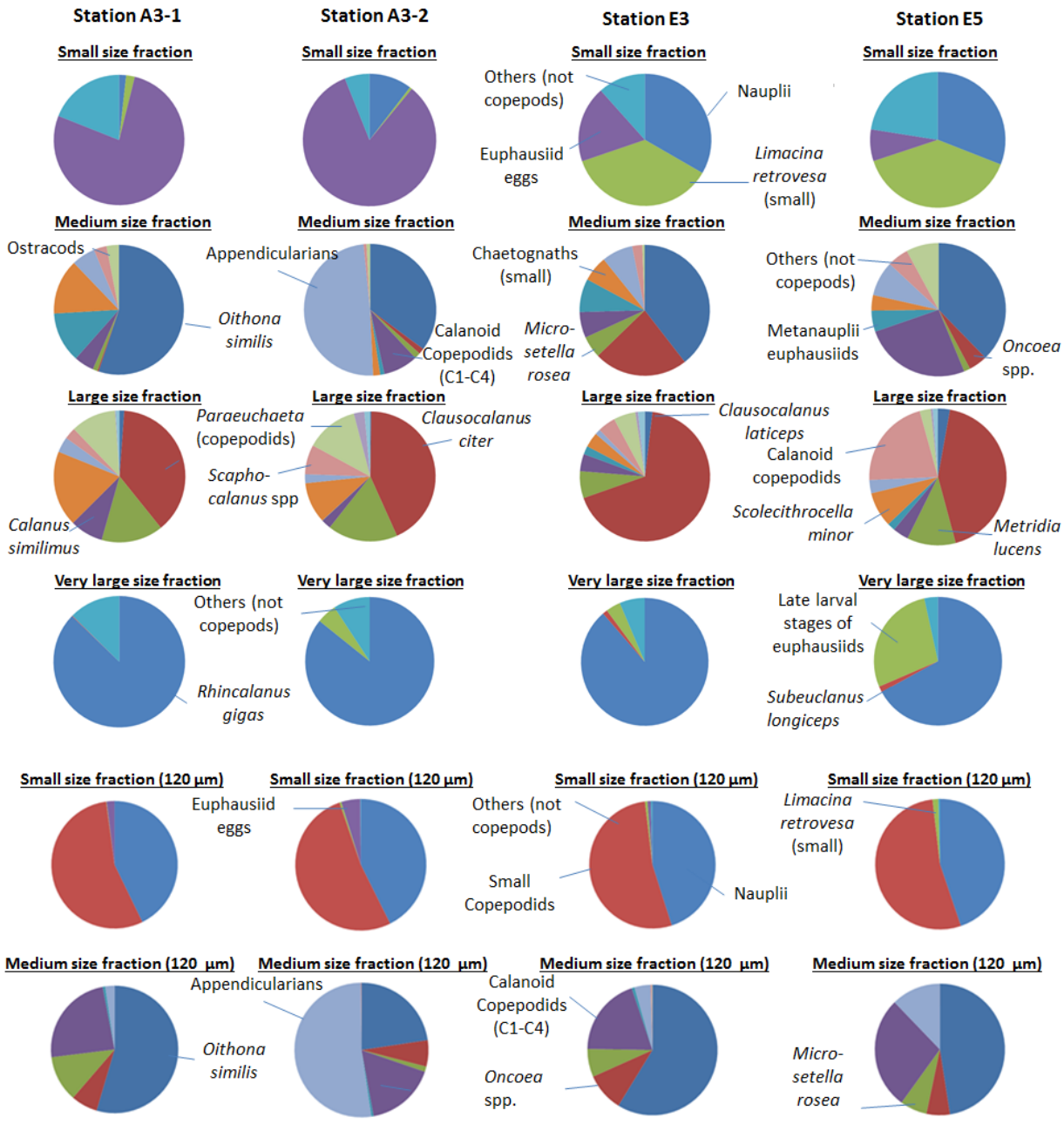

Station A3-1
Small size fraction
Medium size fraction
Ostracods
Oithona similis
Large size fraction
Paraeuchaeta (copepodids)
Scapho-calanus spp
Calanus similimus
Very large size fraction
Others (not copepods)
Rhincalanus gigas
Small size fraction (120 µm)
Euphausiid eggs
Medium size fraction (120 µm)
Appendicularians
Oithona similis
Station A3-2
Small size fraction
Medium size fraction
Appendicularians
Calanoid Copepodids (C1-C4)
Large size fraction
Clausocalanus citer
Very large size fraction
Others (not copepods)
Small size fraction (120 µm)
Medium size fraction (120 µm)
Station E3
Small size fraction
Others (not copepods)
Nauplii
Euphausiid eggs
Limacina retrovesa (small)
Medium size fraction
Chaetognaths (small)
Micro-setella rosea
Large size fraction
Clausocalanus laticeps
Very large size fraction
Small size fraction (120 µm)
Others (not copepods)
Small Copepodids
Nauplii
Medium size fraction (120 µm)
Calanoid Copepodids (C1-C4)
Oncoea spp.
Station E5
Small size fraction
Medium size fraction
Others (not copepods)
Metanauplii euphausiids
Oncoea spp.
Large size fraction
Calanoid copepodids
Scolecithrocella minor
Metridia lucens
Very large size fraction
Late larval stages of euphausiids
Subeuclanus longiceps
Small size fraction (120 µm)
Limacina retrovesa (small)
Medium size fraction (120 µm)
Micro-setella rosea

Figure 7.

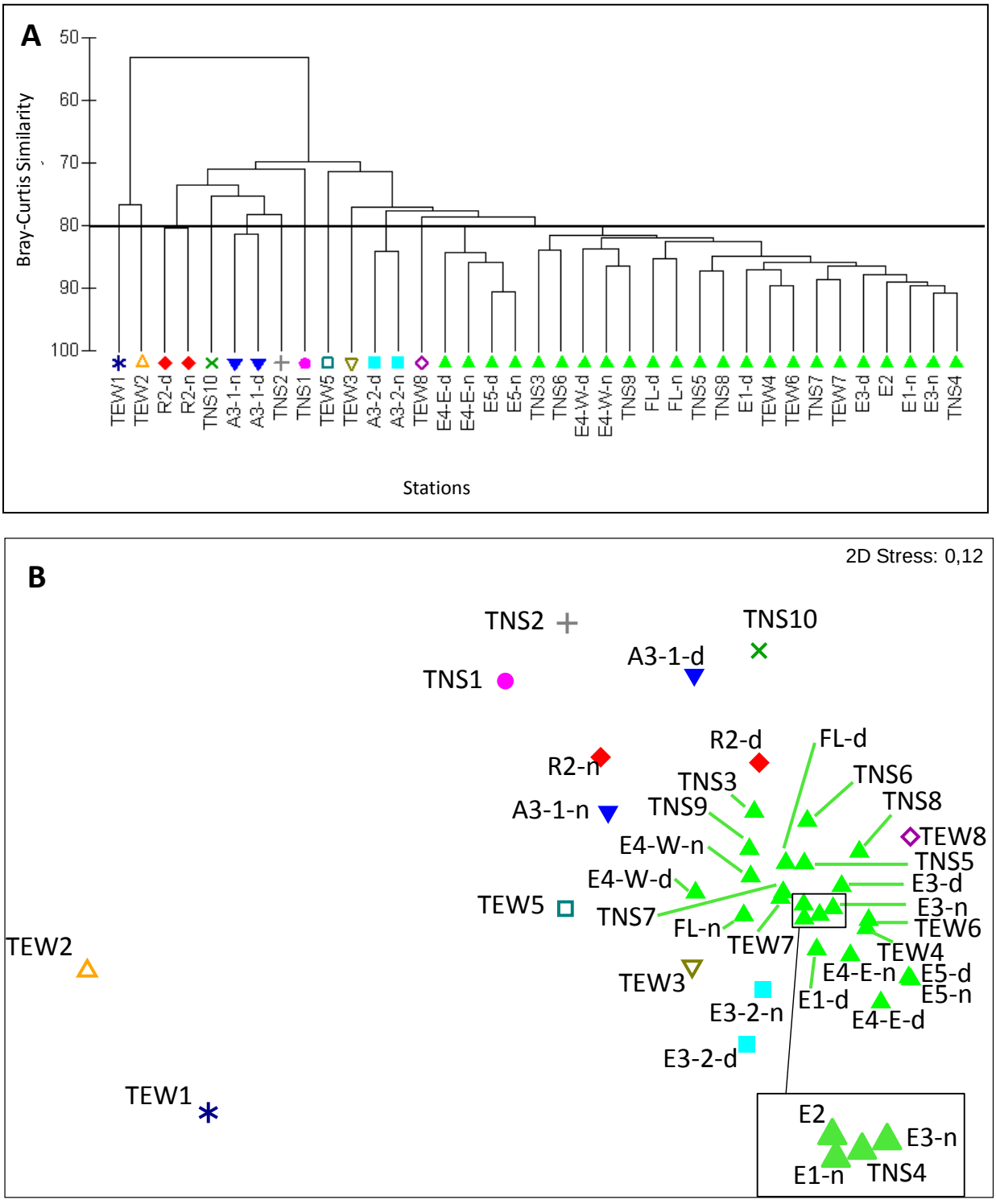

A
Bray-Curtis Similarity
50
60
70
80
90
100
TEW1
TEW2
R2-d
R2-n
TNS10
A3-1-n
A3-1-d
TNS2
TNS1
TEW5
TEW3
A3-2-d
A3-2-n
TEW8
E4-E-d
E4-E-n
E5-d
E5-n
TNS3
TNS6
E4-W-d
E4-W-n
TNS9
FL-d
FL-n
TNS5
TNS8
E1-d
TEW4
TEW6
TNS7
TEW7
E3-d
E2
E1-n
E3-n
TNS4
Stations
B
2D Stress: 0,12
TNS2
TNS10
A3-1-d
TNS1
R2-d
FL-d
R2-n
TNS3
TNS6
TNS8
A3-1-n
TNS9
TEW8
E4-W-n
TNS5
E4-W-d
E3-d
TEW5
TNS7
E3-n
FL-n
TEW6
TEW2
TEW7
TEW4
E4-E-n
E5-d
TEW3
E5-n
E1-d
E3-2-n
E4-E-d
E3-2-d
TEW1
E2
E3-n
E1-n
TNS4

Figure 8

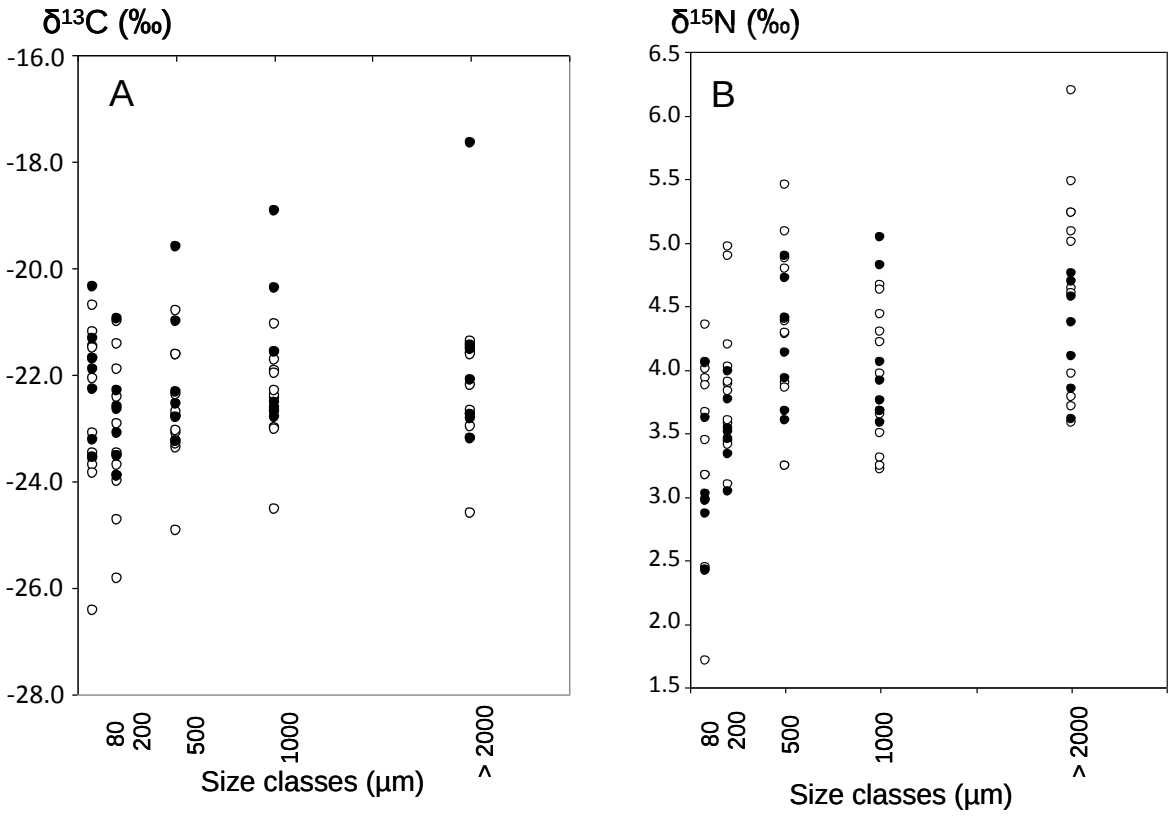

δ13C (‰)
A
-16.0
-18.0
-20.0
-22.0
-24.0
-26.0
-28.0
80
200
500
1000
> 2000
Size classes (µm)
δ15N (‰)
B
6.5
6.0
5.5
5.0
4.5
4.0
3.5
3.0
2.5
2.0
1.5
80
200
500
1000
> 2000
Size classes (µm)

Figure 9

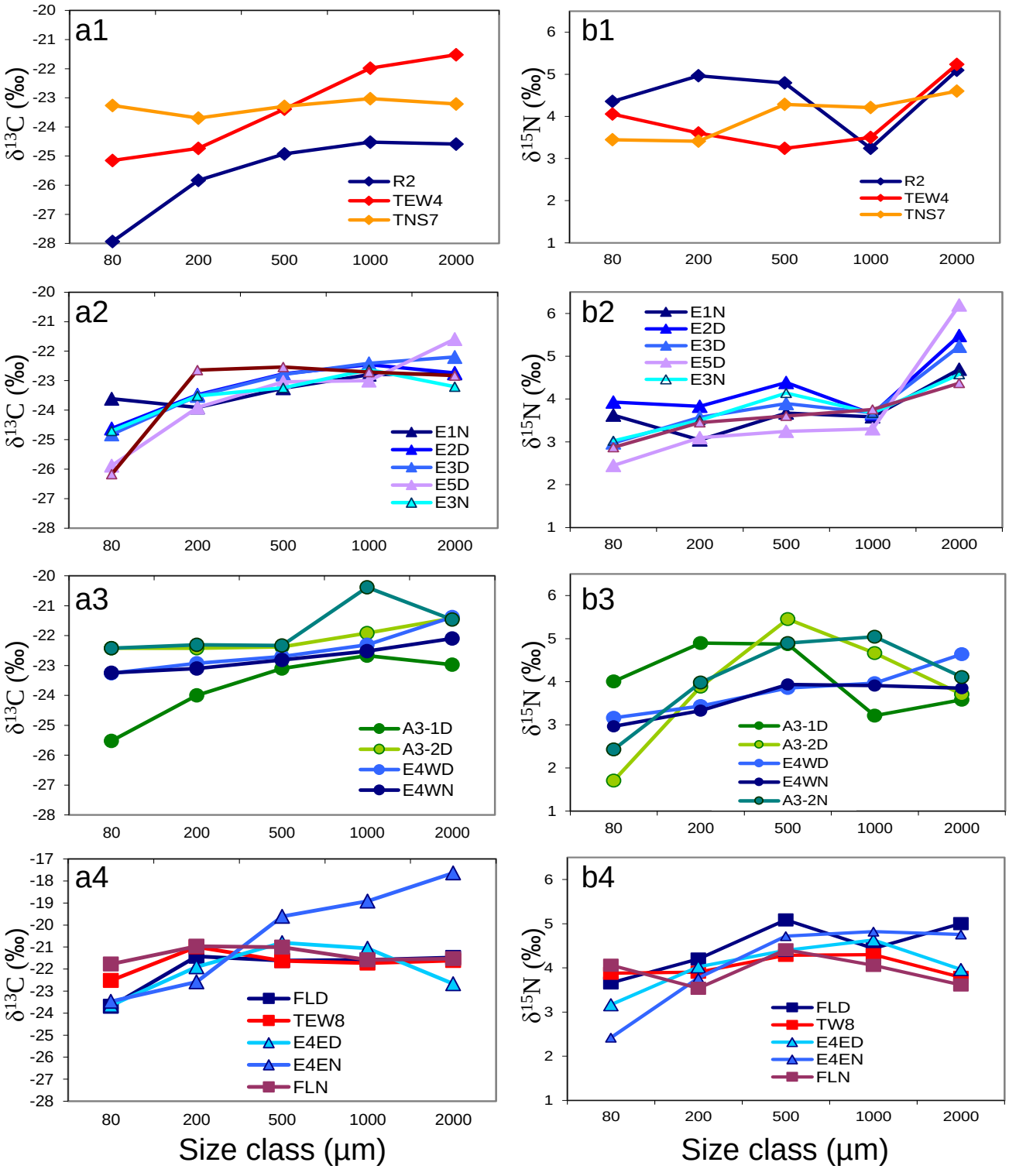
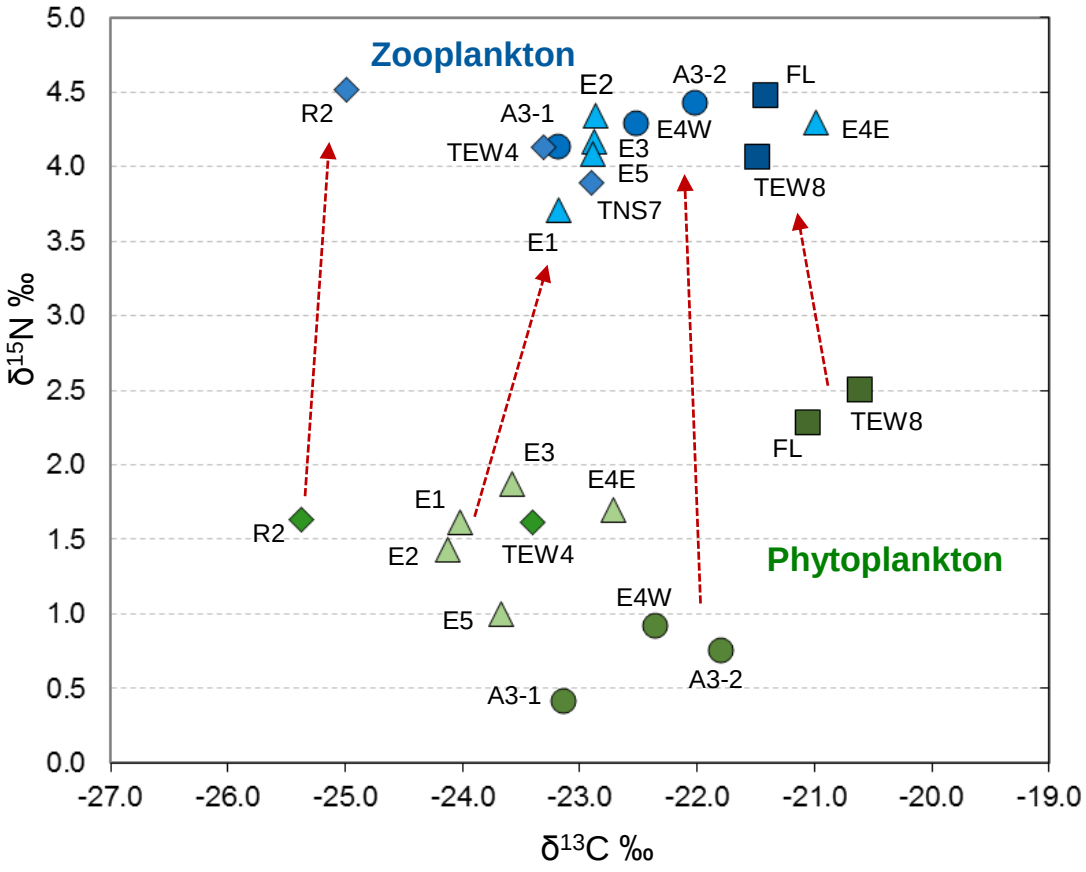

a1
b1
a2
b2
a3
b3
a4
b4
δ13C (‰)
δ15N (‰)
R2
TEW4
TNS7
E1N
E2D
E3D
E5D
E3N
A3-1D
A3-2D
E4WD
E4WN
A3-2N
FLD
TEW8
TW8
E4ED
E4EN
FLN
80
200
500
1000
2000
Size class (µm)

Figure 10


Zooplankton
Phytoplankton
R2
A3-1
E2
A3-2
FL
E4W
E4E
TEW4
E3
E5
TEW8
TNS7
E1
δ15N ‰
δ13C ‰
5.0
4.5
4.0
3.5
3.0
2.5
2.0
1.5
1.0
0.5
0.0
-27.0
-26.0
-25.0
-24.0
-23.0
-22.0
-21.0
-20.0
-19.0