

Dear Editor,

We thank both reviewers for their constructive comments and positive feedback. Both reviewers suggest minor revisions. We have corrected our manuscript following the reviewer's suggestions and address each comment specifically which we document here in detail.

Yours sincerely,

Claire McKay, Jeroen Groeneveld, Helena Filipsson, David Gallego-Torres, Martin Whitehouse, Takashi Toyofuku and Oscar Romero

Reviewer 1: Glock

“One flaw of the manuscript I see is that it's not emphasized in a proper way that *E. exilis* is an infaunal species and that this fact severely affects the Mn/Ca ratios. Usually dissolved Mn²⁺ should be higher in the pore water at the living depth of the foraminifera than in the bottom water, if the oxygen penetration depth is not very deep. An exception would be if the bottom water already is anoxic. Furthermore, infaunal species are able to migrate within the sediment column and probably experience variable pore water conditions within their lifetime. I don't see this fact as a big disadvantage and would rather use infaunal species for this proxy by myself since they probably incorporate more Mn than epifaunal species. These facts are already introduced in the manuscript but discussed only very sparse and very late (page 18; line 18).”

Response: We agree with this statement

Action: We have now emphasized these points in the introduction section (lines 155-160).

“Another point is a major problem with foraminiferal Mn/Ca ratios itself: Diagenetic overprinting by Mn (oxyhydr)oxide coatings. If I look at your data I do not think you have a problem with these, but not every reader is familiar with details of SIMS analyses and, since in the foraminiferal community Mn/Ca ratios are usually used as tracer for contamination, I think it is worth to be discussed in a bit more detail. Regarding this point there is a mistake in a sentence referring to the paper Glock et al. (2012). At page 8, line 19 you wrote “Therefore, we employed a rigorous pre-treatment cleaning technique to remove possible diagenetic coatings following the method of Glock et al. (2012).” Probably I know these details since I'm the author of the paper: If you used the cleaning method from this paper you did not remove possible diagenetic coatings. In this study I just used oxidative cleaning to get rid of organic contamination. Beforehand I showed that the specimens were generally free from diagenetic oxide coatings with element mapping done by EMP. Please correct this sentence! Furthermore, I am not sure if you used reductive cleaning during your flow through analyses, thus it would be nice to provide some more details about this in the methods section. As I already mentioned I do not think that you have a problem with diagenetic coatings if you chose your SIMS spots within the massive centre of the test walls. Furthermore, I guess if there would be an influence of diagenetic coatings on the foraminiferal bulk analyses the results would be much higher than the SIMS results.”

Response: We agree with this statement that not every reader will be familiar with the details of SIMS analyses and acknowledge the mistake.

Action: We have now addressed these points by explaining in more detail how contamination was avoided. and have corrected the mistake regarding organic contamination and stated that the massive centres of the test walls were measured in order to avoid diagenetic coatings (lines 192 – 196 and 253-256). Regarding reductive cleaning, this has now been addressed in lines 278-284.

Page 4, line 3: I would suggest not to write “trace elemental to foraminiferal calcite” ratios and rather write “trace element to calcium ratios”. The trace elements are also incorporated into the foraminiferal calcite and thus part of it.

Response: agreed.

Action: amended (line 61).

Page 4, line 14: “You wrote “... gained more interest is Mn/Ca both as a measure in biogenic foraminiferal calcite and in bulk sediment samples...”. As far as I understood you measured Mn/Al ratios in bulk sediment and not Mn/Ca ratios.

Response: True, Mn/Al was analysed, however in the literature Mn alone has also been analysed in sediments (Lenz et al. 2014). It is true that most geochemical data in bulk sediments are presented as Mn/Al, but eventually, core scanner XRF data (being semiquantitative) might be presented as Mn/Ca (maybe even Mn/Ti, but I am not sure right now if this is frequent at all). In any case, both Al and Ca are used as normalization factor, so it makes sense to use Al for bulk sediment normalization and Ca for foram test normalization.

Action: amended so that there is a clear difference between foraminiferal Mn/Ca and bulk (lines 70-72).

Page 5, line 3: “On the other hand, under anoxic conditions the Mn either diffuses upwards and into the overlying water column or when pore waters become supersaturated with respect to Mn, precipitation of MnCO₃ (rhodochrosite) occurs (Froelich et al., 1979; Pedersen and Price, 1982; Tribovillard et al., 2006).” I do not understand the argument that under these conditions Mn/Ca should be very low in the forams. Even if all Mn diffuses out of the pore water it is available for the forams during that process. Only within the sediment you probably won't find any accumulation.

Response: We now realise this statement is unclear. We have to assume that Mn abundance affects Mn content in *E. exilis* tests, and this is what we want to highlight. Otherwise, all variations in Mn/Ca would be related to vital effect, and our data does not support this.

Action: We have clarified this by stating the expected Mn/Ca under different redox conditions (lines 91-93).

Page 5 line 15: “Mn/Ca signatures of the ambient bottom water are recorded by benthic foraminifera, for instance, culture experiments have confirmed that the species *Ammonia tepida* incorporates Mn into the test (Munsel et al., 2010).” I would suggest to reformulate this sentence. Maybe even divide it into two sentences. Furthermore, it might be good to emphasize that “*Ammonia tepida* incorporates Mn into the test proportional to the concentration in the ambient water masses (Munsel et al., 2010).

Response: agreed

Action: sentence has now been reformulated to clarify our meaning and divided into two sentences to emphasize this point (lines 101-104).

Page 11 line 2: “With cautious positioning of the primary beam on the test walls, such detrital material and potential contaminants were avoided and therefore only the elements actually incorporated into the calcitic tests were measured.” How did you do this? I guess it's easy to avoid macroscopic contaminations which are easy to see optically, but a lot of contaminants you won't see like this. You wrote that you checked the Ca counts and thus assured that you measured within massive calcite which is already good. Do you have any other evidence? Like watching the element distributions during the measurements, for instance (Mn hotspots would probably indicate contaminants), or any pictures which show that you hit the massive centre of the test walls?

Response: Yes we observed the element distributions during measurements and therefore Mn hotspots could be annulled.

Action: these details have now been added to this paragraph (lines 253-256).

Page 14 Line 8: “The slightly higher Mn/Ca determined by FT-ICP-OES in comparison to SIMS derived Mn/Ca perhaps highlights the issue of comparing bulk foraminiferal samples with individual tests comprising of only 6–10 analytical targets. Overall, when a sufficient number (minimum weight of 0.1 mg) of benthic foraminiferal specimens are not available in sediment samples for solution-based analyses (in this case from 35–18 ka), SIMS has the potential to provide reliable results from a few individuals to compensate for this.” I just can say it again: This is a great result! You should emphasize it a bit more (maybe in the conclusions), because it's not trivial that SIMS Mn/Ca ratios on only a few specimens are indeed comparable with foram bulk samples. You won't find this for every element.

Response: agreed.

Action: We have now expanded on this great result and emphasized it in more detail in the conclusion (lines 499-503).

Page 15 line 8: “overlying water column immediately above” I would suggest to remove either “overlying” or “immediately above” because it basically describes the same.

Response: Agreed.

Action: Removed "overlying" (line 371)

Page 15 line 9: “Concurrently, these low Mn/Ca results adhere to the benthic foraminiferal response of a low abundance (ca. 2 specimens cm³) of low oxygen tolerant *E. exilis* (McKay et al., 2014)” Why concurrently? Both proxies indicate into the same direction (higher oxygen concentrations).

Response: Perhaps "concurrently" was not the best word to use.

Action: This has now been amended to clarify that the Mn/Ca results agree with the faunal data (line 372-373).

Line 15, line 22: “Dissolved Mn available for the precipitation of Mn hydroxides” It would be a bit more correct to speak from Mn (oxyhydr)oxides. Please check the paper to keep at one formulation.

Response: agreed

Action: amended throughout the paper where applicable.

Page 15, line 24: “Based on this increase in sedimentary Mn/Al coinciding with relatively low foraminiferal Mn/Ca, we therefore infer that the Mn/Al enrichment occurred immediately below the oxygenrich pore waters during late H3 and throughout the period 30–25 ka, delimiting the oxygen penetration front and the upward diffusion of Mn.” If I understood right you suggest that the oxygen penetration depth is very deep and thus Mn is precipitating already below the living depth of *E. exilis*. What is the typical living depth in the sediment for *B. exilis*? If it doesn't only follow the oxygen gradient maybe it is even possible to reconstruct the minimum oxygen penetration depth like this or at least to give a rough estimate.

Response: Page 15, line 24: Agreed. *E. exilis* has been shown to live at 1-2 cm sediment depth (Caulle et al., 2014) which might give a rough estimate of oxygen penetration front.

Action: A sentence has now been added to this paragraph for this detail (line 390-392).

Page 16, line 11: “We interpret this greater range in Mn/Ca as a relative decrease in oxygen within the pore water from earlier times within the record” What if you indeed see oxygen fluctuations over the lifetime of the specimens or between different specimens? Could it be that oxygen indeed was highly variable during the LGM at this location? Did you check if there was a trend in Mn/Ca from older chambers until the younger ones?

Response: We agree that the greater range in Mn/Ca could be due to oxygen being more variable. We checked if there were any trends in Mn/Ca from the older to the younger chambers, however we found no trends or systematic shifts in values.

Action: The interpretation of the greater range of Mn/Ca has now been amended (lines 401-404).

Page 16, line 24: During H1, the increase in foraminiferal Mn/Ca; both the greater variability within the individual tests (Fig. 3) and the higher average Mn/Ca per sample depth (Fig. 4) indicate lower oxygen conditions in the pore waters (Fig. 5c)” This sentence is hard to read.

Response: This refers to our two lines of evidence for lower oxygen conditions.

Action: We have reworded this sentence and divided it into two sentences to more clearly express our intended meaning (lines 415-417).

Page 17, line 14: “We interpret this similarity in oxygen conditions as being due to comparable....” Being what? I think you forgot a word in this sentence.

Response: there is no word missing

Action: the tense has now been amended to make this sentence clearer (lines 433-434)

Page 18, line 6: “As MnO₂ is rapidly reduced to soluble Mn²⁺ in hypoxic pore waters (Glock et al., 2012)...” I would suggest to give another reference here....

Response: reference has now been amended (line 452).

Page 18, line 8: “causes low bottom water oxygen concentrations or even anoxia within millimetres of the sediment-water interface, we can expect a high accumulation of redox sensitive trace metals” Different redox sensitive elements react different under variable oxygen concentrations. Vanadium vor example accumulates under anoxic conditions, while Manganese does accumulate under oxic conditions. You cannot generalize “redox sensitive trace metals” within this sentence.

Response: Agreed that this statement is too general as different elements precipitate under different conditions.

Action: This sentence has now been amended and made more specific (Mn/Ca opposed to "redox sensitive elements") (line 454).

Page 19, line 18: “Therefore, whilst high phytodetritus export typically causes low bottom water oxygen concentrations and benthic faunal studies are indicative of such a scenario, redox trace elemental test composition presents a more comprehensive interpretation.” I would suggest to rewrite this sentence.

Response: Agreed

Action: This sentence has been made shorter for better clarification (lines 490-492).

Table 1: Just for clarification: Write 1sd instead of just sd. Is it possible to provide the precision as well?

Response: Agreed.

Action: 1 sd has now been written which is the precision.

Figure 4: The figure is hard to read. Maybe it would be better to present it in horizontal format.

Response: Agreed and I would prefer it in landscape format.

Action: The editor will be informed.

Figure 5: This figure is great and provides all the interpretations of your multiproxy approach in one graphic. Unfortunately it is hard to understand if the reader does not jump between the discussion part and figure. I would suggest to extend the figure caption and to give a short explanation for every time interval and the reason for the interpretation.

Response: We agree with this suggestion.

Action: We have now extended the figure caption to provide a better explanation for each time interval to link with our interpretations.

Reviewer 2: Limburg

“Figure 4 is quite difficult to read”

Response: Agreed

Action: as previously stated, figure 4 should be placed in landscape layout so it can be enlarged on a separate page.

“The author’s findings are consistent with recent work on fish otoliths”

Response: we were aware of these publications but since our work is on infaunal foraminifera, we did not directly compare with fish otoliths or other biota living within the water column.

Action: We have however, cited Limburg et al., (2011) when discussing MnO₂ in hypoxic waters (line 449).

“Perhaps eventually there will emerge a consensus about carbonate-based bio proxies of hypoxia?”

Response: we hope that our data here contributes to such a future consensus.

1 **A comparison of benthic foraminiferal Mn/Ca and**
2 **sedimentary Mn/Al as proxies of relative bottom water**
3 **oxygenation in the low latitude NE Atlantic upwelling system**
4

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28 **Abstract**

29 Trace element incorporation into foraminiferal shells (tests) is governed by physical and
30 chemical conditions of the surrounding marine environment and therefore foraminiferal
31 geochemistry provides a means of palaeoceanographic reconstructions. With the availability of
32 high spatial resolution instrumentation with high precision, foraminiferal geochemistry has
33 become a major research topic over recent years. However, reconstructions of past bottom water
34 oxygenation using foraminiferal tests remain in their infancy. In this study we explore the
35 potential of using Mn/Ca determined by Secondary Ion Mass Spectrometry (SIMS) as well as by
36 Flow-Through Inductively Coupled Plasma Optical Emission Spectroscopy (FT-ICP-OES) in the
37 benthic foraminiferal species *Eubulimina exilis* as a proxy for recording changes in bottom
38 water oxygen conditions in the low latitude NE Atlantic upwelling system. Furthermore, we
39 compare the SIMS and FT-ICP-OES results with published Mn sediment bulk measurements
40 from the same sediment core. This is the first time that benthic foraminiferal Mn/Ca is directly
41 compared with Mn bulk measurements, which largely agree on the former oxygen conditions.
42 Samples were selected to include different productivity regimes related to Marine Isotope Stage
43 3 (35-28 ka), the Last Glacial Maximum (28-19 ka), Heinrich Event 1 (18-15.5 ka), Bølling
44 Allerød (15.5-13.5 ka) and the Younger Dryas (13.5-11.5 ka). Foraminiferal Mn/Ca determined
45 by SIMS and FT-ICP-OES are comparable. Mn/Ca was higher during periods with high primary
46 productivity, such as during the Younger Dryas which indicates low oxygen conditions. This is
47 further supported by the benthic foraminiferal faunal composition. Our results highlight the
48 proxy potential of Mn/Ca in benthic foraminifera from upwelling systems for reconstructing past
49 variations in oxygen conditions of the sea floor environment as well as the need to use it in
50 combination with other proxy records such as faunal assemblage data.

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58 1. Introduction

59 Foraminifera, being extensively distributed and highly abundant in most marine environments,
60 are essential proxies for reconstructing the chemical and physical properties of past oceans.
61 **Several trace element to calcium (Ca)** ratios analysed on foraminiferal tests have been developed
62 as proxies in the last decades. Perhaps one of the most conventional approaches is the
63 reconstruction of seawater temperatures using Mg/Ca (e.g. Nürnberg et al., 1996; Elderfield et
64 al., 2006). Other established trace elemental proxies also include Ba/Ca to trace salinity changes
65 due to continental runoff (Lea and Boyle, 1989; Hönisch et al., 2011) and Cd/Ca to reconstruct
66 water masses (Marchitto and Broecker, 2006). Whilst there is a wealth of research applying the
67 geochemistry of foraminiferal calcite for paleoceanographic reconstruction and copious
68 sedimentary redox proxies have been developed (e.g. Gooday et al., 2009), utilising the trace
69 elemental composition of foraminiferal shells (tests) to reconstruct oxygen conditions is still in
70 its infancy. **One redox sensitive element that has recently gained more interest is manganese**
71 **(Mn), both as a trace element in biogenic foraminiferal calcite (Mn/Ca) and in bulk sediment**
72 **samples** (Reichart et al., 2003; Glock et al., 2012, Groeneveld and Filipsson, 2013; Lenz et al.,
73 2014; Koho et al., 2015). Here we aim to explore the potential of Mn/Ca by analysing both
74 benthic foraminiferal tests and comparing it to Mn/Al of bulk sediment samples from an
75 upwelling record to determine if changes in oxygen conditions during different primary
76 productivity regimes are detectable by these methods.

77 At the sediment-water interface, the concentration of dissolved redox sensitive elements such as
78 Mn vary significantly between oxic and hypoxic (hypoxia defined as $<1.42 \text{ ml l}^{-1} \text{ O}_2$ following
79 Levin et al., 2009) settings. In sea water, redox sensitive Mn is mainly present as Mn^{2+} which
80 under oxic conditions precipitates as Mn oxyhydroxide (Burdige, 1993; Glasby, 2006). The Mn
81 flux across the sediment-water interface is driven by reductive dissolution of reactive Mn
82 oxyhydroxide (Froelich et al., 1979).

83 Under oxic conditions, dissolved O_2 is present in the pore waters and thus benthic foraminiferal
84 tests are expected to incorporate less Mn into their test. In contrast, under low oxygen conditions,
85 Mn oxyhydroxide is reduced and the Mn^{2+} concentration increases (Tribovillard et al., 2006),
86 becoming available to be incorporated in the foraminiferal tests. Especially under hypoxic
87 conditions, Mn concentrations will be concentrated in the pore water because the Mn cannot
88 escape into the overlying oxic bottom water meaning that benthic foraminiferal Mn/Ca will be
89 highest. Hence foraminiferal Mn/Ca has potential to be used as a proxy of hypoxic conditions.

90 Accordingly, we expect bulk Mn to be depleted in the sediment and exhibit the opposite trend.
91 On the other hand, under anoxic conditions the Mn is able to diffuse upwards and into the
92 overlying water column or when pore waters become supersaturated with respect to Mn, it is
93 precipitated as MnCO_3 (rhodochrosite) (Froelich et al., 1979; Pedersen and Price, 1982;
94 Tribovillard et al., 2006). Therefore, under low oxygen conditions, foraminiferal Mn/Ca is
95 expected to be higher during hypoxic conditions than during anoxic conditions; but still higher
96 than under oxic conditions.

97 Whilst benthic foraminiferal Mn/Ca has been conventionally used as an indicator of
98 contamination by Mn oxyhydroxide or Mn carbonate (Boyle, 1983; Barker et al., 2003) new
99 studies are pioneering Mn/Ca as a potential proxy of related changes in bottom/pore water
100 oxygen and redox conditions (Ní Fhlaithearta et al., 2010; Glock et al., 2012; Groeneveld and
101 Filipsson, 2013; Koho et al., 2015). Mn/Ca signatures of the ambient bottom water are recorded
102 by benthic foraminifera. For example, culture experiments have confirmed that the species
103 *Ammonia tepida* incorporates Mn into the test proportional to the concentration in the ambient
104 water masses (Munsel et al., 2010). Thus, during benthic foraminiferal calcification under
105 hypoxic conditions, more Mn will be assimilated into their calcite tests, whereas under anoxic
106 conditions or oxic conditions in particular, Mn is expected to decrease, albeit to different
107 amounts (Pena et al., 2005). Hence Mn/Ca used in this study could provide a reliable means of
108 reconstructing the former seafloor oxygen settings at the time of deposition opposed to sediment
109 bulk measurements which can continue to oxidise and be mobilised post deposition.

110 Traditionally, trace element/Ca is analysed on solution based samples containing larger numbers
111 of foraminifera specimens to give a representative result (Groeneveld and Filipsson, 2013).
112 When a sufficient amount of specimens is not available for solution based techniques, or if
113 diagenesis has affected the tests, a micro-analytical technique upon single specimens such as
114 Secondary Ion Mass Spectrometry (SIMS) is a valuable tool. From an analytical perspective
115 SIMS has enhanced our ability to determine how trace elements are distributed within
116 foraminiferal tests at high spatial resolution and precision on individual foraminifera (Allison
117 and Austin, 2003; Bice et al., 2005; Kunioka et al., 2006; Glock et al., 2012). Recently, SIMS
118 determined Mn/Ca of benthic foraminifera has been found to be representative of Mn/Ca in the
119 top cm of the pore water, confirming that the foraminiferal calcite composition relates to the
120 level of oxygen depletion (Glock et al., 2012).

121 Where a sufficient amount of foraminiferal specimens are available, we additionally used Flow-
122 Through Inductively Coupled Plasma Optical Emission Spectroscopy (FT-ICP-OES; Haley and
123 Klinkhammer, 2002). Flow-through analysis is a means of determining elemental composition
124 from samples of foraminiferal tests which permits complete monitoring of the effects of cleaning
125 and dissolution (Haley and Klinkhammer, 2002). However, due to the small size of
126 *Eubuliminella exilis*, a larger number (up to 50 specimens in this case) of foraminiferal tests
127 from the same core sample is required to give a representative average signal. Therefore we
128 utilise both SIMS and FT-ICP-OES to explore the potential of Mn/Ca for interpreting down core
129 oxygen studies.

130 To explore how Mn/Ca works as a potential proxy for bottom/pore water oxygen conditions, we
131 study a site from the low latitude NE Atlantic Upwelling System. Upwelling systems are an ideal
132 environment to test this proxy, as they are renowned for high export rates of labile organic matter
133 from surface waters which provokes severe oxygen depletion in the underlying intermediate
134 waters and at the seafloor (Böning et al., 2004). We study core GeoB7926-2 from the upwelling
135 region off coastal NW Africa (Fig. 1) and compare foraminiferal and sedimentary Mn with
136 published diatom and benthic foraminiferal species composition (Romero et al., 2008; Filipsson
137 et al., 2011; Kim et al., 2012; McKay et al., 2014). In general, coastal upwelling systems are the
138 most productive of the world ocean resulting in vulnerability to oxygen minima within the water
139 column and underlying seafloor (Helly and Levin, 2004; Bakun et al., 2010). At present, the
140 benthic environment of this particular upwelling system is not especially susceptible to low
141 oxygen conditions and is well ventilated with bottom water oxygen of ca. 5 ml l⁻¹ (Goretski and
142 Koltermann, 2004). However, based on benthic foraminiferal faunal studies, there is evidence of
143 previous periods of oxygen depletion at the sea floor during the Younger Dryas (YD, 13.5-11.5
144 ka) and Heinrich Event1 (H1, 18-15.5 ka) in particular. This is inferred from the predominance
145 of the low oxygen tolerant benthic foraminiferal species *Eubuliminella exilis* (synonymised taxa:
146 *Bulimina exilis*) (Filipsson et al., 2011; McKay et al., 2014). Therefore, we selected samples
147 allocated to late Marine Isotope Stage 3 (MIS3, 35-27 ka), the Last Glacial Maximum (LGM),
148 Heinrich Event 1 (H1), Bølling Allerød (BA) and the Younger Dryas (YD) to reconstruct past
149 bottom water oxygen. These climatic intervals were chosen in order to test if Mn/Ca can confirm
150 the low oxygen conditions during different productivity regimes as reported by previous studies
151 from this particular sediment core (Filipsson et al., 2011; McKay et al., 2014). We focus on
152 utilising SIMS and compare this method with FT-ICP-OES where a sufficient number of *E.*

153 *exilis* specimens were available. We also present Mn/Al sediment bulk measurements from the
154 same sediment for further comparison.

155 The low oxygen tolerant, benthic foraminiferal species *Eubuliminella exilis* is an infaunal species
156 and therefore has the ability to migrate within the sediment and experience variable pore water
157 conditions. This migration could affect the Mn/Ca incorporated within the test; however as *E.*
158 *exilis* is so low oxygen tolerant, it possibly migrates less than other infaunal species which are
159 not as tolerant. Therefore *E. exilis* likely incorporates more Mn and is therefore the right
160 recorder, suitable for reconstructing oxygen levels. Furthermore, abundant populations of this
161 taxon are reported from a range of settings, including coastal upwelling sites, sapropels, oxygen
162 minimum zones and other environments for example in the Bering Sea and the Mississippi River
163 mouth (Caralp, 1989; Jorissen, 1999; Jannink et al., 1998; Rasmussen et al., 2002; Khusid et al.,
164 2006). Thus *E. exilis* also has potential to be a useful proxy for other marine environments
165 susceptible to strong oxygen deficiency and high fluxes of organic export. *Eubuliminella exilis*
166 has been found to correlate with diatom accumulation rate and clearly dominates the
167 foraminiferal fauna during very high diatom input (Caralp, 1984; Filipsson et al., 2011; McKay
168 et al., 2014; McKay et al., in revision). Therefore *E. exilis*, being present (albeit in considerably
169 varying abundance) throughout the GeoB7926-2 record is an ideal candidate to record oxygen
170 changes in the environment in which they lived and also provide an opportunity to test if export
171 productivity is causing low oxygen conditions opposed to the dominance of this species merely
172 being a fresh phytodetritus diet signal (Caralp, 1989).

173 We hypothesise that higher foraminiferal Mn/Ca will occur during times of high diatom
174 accumulation rate and lower bottom water oxygen concentrations and accordingly higher *E.*
175 *exilis* abundance, with the opposite effect during times of low surface productivity.

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177 **2. Method**

178 Gravity core GeoB7926-2 from the NE Atlantic Upwelling System (20°13'N, 18°27'E, 2500 m
179 water depth) was recovered during R/V Meteor cruise M53/1 (Meggers and Cruise Participants,
180 2003). The age model for the core was published by Kim et al., (2012) and the timing and
181 duration of the climatic intervals were adopted from the $\delta^{18}\text{O}$ of NGRIP (NGRIP Members,
182 2004) and Sánchez Goñi and Harrison (2010). We selected well-preserved foraminiferal
183 specimens based on the criteria of high and low surface productivity regimes as demonstrated

184 from diatom abundance (Romero et al., 2008). Details of sample preparation for benthic
185 foraminiferal faunal analysis have previously been published (Filipsson et al., 2011; McKay et
186 al., 2014).

187

188 2.1 SIMS analysis

189 From the >150 µm size fraction, a total of 48 specimens of the benthic foraminifera species
190 *Eubuliminella exilis* were hand-picked under a binocular microscope for SIMS analysis (Table
191 1). We acknowledge that the presence of Mn-rich authigenic coatings (e.g. Mn (oxyhydr)oxides
192 and organic matter) can be problematic for trace elemental analysis of foraminifera (Boyle,
193 1983; Pena et al., 2005; Klinkhammer et al., 2009). **Therefore, we employed a rigorous pre-**
194 **treatment cleaning technique to remove possible organic contamination following the method of**
195 **Glock et al., (2012) and avoided potential diagenetic coatings during the SIMS analysis by**
196 **measuring within the massive centre of the test walls (Fig. 2).**

197 For the SIMS analysis, foraminifera from individual sample depths were rinsed over a 63 µm
198 sieve with milliQ water. After this rinsing step, the foraminifera were transferred into vials and
199 sonicated for 20 seconds. Subsequently, the foraminifera were rinsed with methanol and
200 sonicated again for 1 minute. Any residual methanol was then removed with milliQ water. An
201 oxidative cleaning step was performed to remove organic matter which consisted of mixing the
202 following reagents: 100 µl 30% H₂O₂ to 10 ml of 0.1 M NaOH solution. 350 µl of this reagent
203 was added to each individual vial and the vials were put into a water bath at 92°C for 20 minutes.
204 Afterwards another 20 second sonic bath was undertaken; the foraminifera samples were rinsed
205 again with milliQ water in the 63 µm sieve to remove any residues. For the final step, the
206 specimens were transferred back into their respective vials and 250 µl of 0.001 M HNO₃ was
207 added to each vial. The vials were put into a sonic bath for 20 seconds and finally rinsed one last
208 time with milliQ water. After the cleaning procedure, the specimens were checked under a
209 binocular microscope to ensure sufficient cleaning and that the tests remained intact.

210 Foraminifera specimens were embedded in low viscosity epoxy resin at JAMSTEC, Japan. The
211 foraminifera were then ground to expose a cross-section across the test wall using 16 µm silicon
212 carbide paper at the Department of Geosciences, University of Edinburgh, UK. Resin pieces
213 were mounted into low viscosity epoxy resin disks (Struers) at the NORDSIM laboratory,
214 Laboratory for Isotope Geology at the Swedish Museum of Natural history, Stockholm, Sweden.

215 The mounts were polished using a Struers Rotopol-2 at 150 rpm for 1 minute, first with 3 μm
216 diamond suspension and again with 1 μm diamond suspension. Between each grinding and
217 polishing step, mounts were cleaned with ethanol. Each cross-sectioned foraminifera test was
218 examined under high power reflected light microscopy to evaluate the quality of the carbonate
219 and to assist in assessing the progress of polishing until the cross sections were clear.
220 Subsequently, the mounts were cleaned in high purity ethanol and coated in a 20 nm thick high
221 purity Au coat.

222 The reference material used for the SIMS was a polished piece of OKA calcite crystal supplied
223 from Geomar, Kiel University, Germany (E. Hathorne, pers. comm). This standard was obtained
224 from a matrix matched specimen for which Mn/Ca has been reported by solution ICP-MS (Glock
225 et al., 2012). During calibration, the OKA was analysed $n = 16$ times, yielding a high sensitivity
226 with 1 standard deviation repeatability of 1.2% for Mn concentration.

227 The Mn/Ca analyses of the test cross-sections were performed using a Cameca IMS 1280 ion
228 microprobe at the NORDSIM laboratory at the Swedish Museum of Natural history, Stockholm,
229 Sweden. Analysis used a $^{16}\text{O}_2^-$ ion beam accelerated at 23 kV impact energy (-13 kV primary
230 beam, +10 kV secondary beam). It is vital to only analyse Mn which is located internally within
231 the original test wall to attain most representative Mn/Ca for developing it as a redox proxy.
232 Therefore, a 50 μm aperture in the primary column was used to shape a slightly elliptical 5 μm
233 spot on the sample surface, which, together with careful placement, reduced the effects of sample
234 contamination from the test wall outer surface. Prior to each analysis, the analytical location was
235 pre-sputtered for 2 minutes with the ion beam rastered over 10 x 10 μm raster to remove the Au
236 coat and any remaining surface contamination. During the initial pre-sputtering, the ^{44}Ca
237 distribution was monitored using the ion imaging system of the instrument and maximised to
238 ensure high precision beam targeting on the fine foraminiferal test walls. The mass spectrometer
239 was operated at a mass resolution of $M/\Delta M \sim 6000$ to resolve the ^{55}Mn peak from nearby
240 molecular interferences. A 400 μm contrast aperture was employed for maximum transmission
241 together with a 60 μm entrance slit, a 2001 μm field aperture restricting the field of view on the
242 sample to an area of ca. 12 x 12 μm at the transfer magnification of ca. 160x, and a 45 eV wide
243 energy window; all of which combined to yield adequately flat topped peaks on the species of
244 interest. Each analysis comprised of 16 cycles of ^{44}Ca (1s integration/cycle) and ^{55}Mn (2s). Each
245 analysis lasted approximately 9 minutes. Secondary ions were measured using a low noise
246 (<0.01 cps) ion counting electron multiplier. Multiple analysis points were undertaken upon each
247 individual test of *E. exilis* (ca. 6-10 targets per individual specimen) starting from the aperture

248 and taking measurements alternating between the outer wall and internal walls. For the best
249 targets, programming was performed manually to ensure that widest chamber walls and “t”
250 junctions were targeted since they provide a wider test wall for the analyses (Fig. 2).
251 Furthermore, at such high spatial resolution and precision, it is easy to visually observe and
252 avoid encrusting prior to selecting analysis targets via the connected screen and avoid measuring
253 secondary calcite or authigenic clays which would otherwise affect measurements. **With cautious**
254 **positioning of the primary beam on the test walls and observations of the element distributions**
255 **during measurements, such detrital material and potential contaminants were avoided and**
256 **annulled.** Therefore only the elements actually incorporated into the calcitic tests were measured.
257 As an additional prerequisite to this, analyses with Ca values > 500 kcps were classified as being
258 reliable. Mn/Ca was first normalised to those determined in the OKA standard and subsequently
259 converted to the true value in the OKA based on the Glock et al. (2012) value for Mn/Ca of 4920
260 $\mu\text{mol mol}^{-1}$.

261 The advantage of SIMS is that it is non-destructive and as the foraminiferal cross sections are
262 preserved within the mounts, they can be stored for further analyses. Mounts are archived at the
263 NORDSIM laboratory.

264

265 2.2 FT-ICP-OES analysis

266 For FT-ICP-OES, 20-50 specimens per sample depth of *E. exilis* from the GeoB7926-2 record
267 were selected from samples corresponding to H1, BA and the YD for comparisons with the
268 SIMS data. These three climatic intervals encompassed the only samples where a sufficient
269 number of pristine *E. exilis* individuals were present. The tests were gently crushed in a 0.5 ml
270 vial and fragments were transferred into a PTFE filter with 0.45 μm mesh.

271 For analysis, the filters were connected to a Flow-Through – Automated Cleaning Device
272 (Klinkhammer et al., 2004; Haarman et al., 2011). Automatic cleaning prevents the loss of
273 material which occurs with traditional cleaning allowing the analysis of very small samples (~20
274 μg). The Flow-Through was then connected to an ICP-OES (Agilent Technologies, 700 Series
275 with autosampler (ASX-520 Cetac) and micro-nebulizer). Time Resolved Analysis (TRA) was
276 used to analyze the samples at MARUM, University of Bremen, Germany. After an initial rinse
277 (5 minutes) with buffered Seralpur water to remove clays, the samples were slowly dissolved
278 using an acid ramp formed by mixing of Seralpur with 0.3 M QD HNO_3 **(no additional oxidative**

279 and/or reductive cleaning was performed). Starting with pure Seralpur the acid contribution was
280 stepwise increased every minute to 100% acid after 30 minutes. The flow speed of the solution
281 was 250 $\mu\text{l/l}$. Mn/Ca of the foraminiferal calcite was determined by identifying the TRA interval
282 which showed a consistent linear relationship between Mn and Ca counts. Potential diagenetic
283 phases like Mn(oxy)hydroxides are avoided this way, as they would have a different slope, i.e. a
284 significant Mn-signal along with the absence of a Ca- signal. Mn/Ca was then calibrated using
285 the characteristic slope of this linear relationship of a known in-house standard solution analyzed
286 on the same acid ramp. The average standard error on the determination of the slope for Mn/Ca
287 was 0.75%. We analysed an international limestone standard (ECRM752-1) for Mg/Ca to
288 validate the results following the same FT-protocol. The average Mg/Ca of the ECRM752-1 ($n =$
289 4) was 3.76 mmol mol^{-1} , which compares well with average published value of 3.75 mmol mol^{-1}
290 (Greaves et al., 2008).

291

292 2.3 Mn bulk measurements

293 For geochemical bulk analyses, samples were dried and ground in an agate mortar and
294 homogenised. Total dissolution of samples was undertaken using HF and HNO_3 following the
295 standard procedures of Gallego-Torres et al., (2007). Mn and Al content for samples
296 corresponding to 35-10 ka was determined by Atomic Absorption Spectrometry, using Re and
297 Rh as internal standards at the Analytical Facilities at the University of Granada, Spain. Mn
298 being redox sensitive was normalised to Al content in order to correct for detrital variations (Van
299 Der Weijden, 2002). Mn/Al data corresponding to 25-10 ka has previously been published
300 (Gallego-Torres et al., 2014) and here we extend the Mn/Al record to 35 ka.

301

302 3. Results

303 3.1 SIMS and FT-ICP-OES data

304 Overall, Mn/Ca determined by SIMS varied between 2 and 750 $\mu\text{mol mol}^{-1}$. Average values
305 ranged from 6.5 to 260 $\mu\text{mol mol}^{-1}$ throughout the record and displayed a decreasing trend down
306 core (Fig. 3-4, Table 1).

307 Mn/Ca was lowest in foraminiferal tests from Marine Isotope Stage 3 (MIS3) and the Last
308 Glacial Maximum (LGM) with values ranging from 25-68 $\mu\text{mol mol}^{-1}$ (average 50 $\mu\text{mol mol}^{-1}$)

309 and 2-225 $\mu\text{mol mol}^{-1}$ (average 70 $\mu\text{mol mol}^{-1}$) respectively. Foraminifera from samples derived
310 from the Heinrich Event 1 (H1) and the Bølling Allerød (BA) exhibited a slightly elevated range
311 of Mn/Ca at 50-380 $\mu\text{mol mol}^{-1}$ (average 117 $\mu\text{mol mol}^{-1}$) and 27 to 280 $\mu\text{mol mol}^{-1}$ (average
312 133 $\mu\text{mol mol}^{-1}$) respectively. Highest Mn/Ca occurred in foraminiferal tests from the Younger
313 Dryas (YD) (average 175 $\mu\text{mol mol}^{-1}$) period ranging from 23 to 750 $\mu\text{mol mol}^{-1}$ (Fig. 3, Table
314 1).

315 In order to determine statistically significant differences between samples from different
316 productivity regimes, Kruskal-Wallis tests were run and showed a statistically significant
317 difference between the mean Mn/Ca values (per individual foraminifera specimen) between all
318 five climatic intervals ($p = 0.003$). By testing the mean Mn/Ca of each foraminifera between
319 each climatic interval individually in turn, using Post hoc (Mann Whitney U) tests, significant
320 differences lay between climatic intervals with high and low export productivity regimes (based
321 on diatom accumulation rate). Namely, significant differences in Mn/Ca were evident between
322 the YD interval and the LGM and MIS3 (Table 2).

323 Mean SIMS derived Mn/Ca per sample depth compare well with FT-ICP-OES results (Table 1),
324 agreeing with maximum differences of 15 - 24 $\mu\text{mol mol}^{-1}$ (Fig. 4). Mn/Ca from the FT-ICP-
325 OES on bulk foraminiferal samples tended to be slightly higher compared to the mean ratios
326 obtained from the SIMS microanalyses for the same sediment depth (for example 140 $\mu\text{mol mol}^{-1}$
327 1 compared to 116 $\mu\text{mol mol}^{-1}$ during the YD).

328

329 3.2 Mn bulk data

330 Bulk sedimentary Mn/Al showed highest values during MIS3 at 30-24 ka as well as during 19-
331 17.5 ka and the YD. The YD was characterised by a sharp Mn/Al increase at 12.3 ka, coinciding
332 with maximum diatom productivity (Fig. 4). Relatively low Mn values occurred during 35-32 ka
333 and during the LGM Mn was below the level of detection (<0.06%). Between 32 ka and the
334 onset of the LGM, a progressive increased was observed.

335

336 4. Discussion

337 4.1.1 Comparison of analytical methods

338 Our results indicate that Mn/Ca in benthic foraminifera might prove to be a valuable proxy for
339 oxygen in the bottom and pore waters. The downcore variability in foraminiferal Mn/Ca at site
340 GeoB7926-2 displays a consistent agreement between the mean SIMS determined Mn/Ca of
341 each sample depth and the bulk foraminifera Mn/Ca measured by FT-ICP-OES. In general, the
342 agreement in values suggests that the SIMS determined Mn/Ca is likely to be a true signal within
343 our reconstruction. The slightly higher Mn/Ca determined by FT-ICP-OES in comparison to
344 SIMS derived Mn/Ca highlights the issue of comparing bulk foraminiferal samples with
345 individual tests comprising of only 6-10 analytical targets. Overall, when a sufficient number
346 (minimum weight of 0.1 mg) of benthic foraminiferal specimens are not available in sediment
347 samples for solution-based analyses (in this case from 35-18 ka), SIMS has the potential to
348 provide reliable results from a few individuals to compensate for this.

349 Recent culturing experiments on benthic foraminifera demonstrate that calcification can occur
350 even under anoxic conditions (Nardelli et al., 2014). This is key for the discussion of trace
351 elemental data derived from the foraminiferal tests, as not only does the timing of the
352 calcification determine the geochemical signature, it also shows that the signature is recorded in
353 a wide range of oxygen conditions.

354 When comparing foraminiferal Mn/Ca to previously published sediment measurements of Mn/Al
355 (Fig. 4) for site GeoB7926-2 (Gallego-Torres et al., 2014), in general we do not find a
356 continuous relationship in trends throughout the record, but overall they largely agree on the
357 former oxygen conditions. This is most likely due to diagenetic processes and migration of redox
358 fronts through the sediment which redistributes the bulk Mn after deposition, whereas the
359 foraminiferal tests record the Mn concentration at the time of calcification. In fact, bulk sediment
360 Mn concentrations are often interpreted as being related to diagenetic (post-production)
361 oxidation fronts and less often to the syn-sedimentary environment (e.g. Thomson et al., 1995;
362 de Lange et al., 2008). Thus, two different processes govern Mn fixation in sediment and
363 foraminifera tests respectively and consequently we can expect an offset between the two
364 signals.

365

366 4.2 Reconstruction of bottom water oxygen conditions: Late MIS3 to the Younger Dryas
367 (35-11.5 ka)

368 4.2.1 Late MIS3 - late LGM (35 - 19 ka)

369 Foraminiferal Mn/Ca derived from SIMS measurements were comparably low and exhibited less
370 variability within individual tests (Fig. 3) during episodes of low diatom export between 35 and
371 19 ka. The lower Mn/Ca indicates that the pore waters and **water column** immediately above
372 were oxygenated (Fig. 5a). **These low Mn/Ca results are in agreement with the benthic**
373 **foraminiferal response of a low abundance** (ca. 2 specimens cm⁻³) of low oxygen tolerant *E.*
374 *exilis* (McKay et al., 2014) to the extent that not enough well preserved specimens were available
375 for FT-ICP-OES analysis. Such low Mn/Ca and lack of low oxygen dwellers is to be expected
376 since benthic foraminifera living in an environment where productivity export does not fluctuate
377 at great magnitudes are potentially less exposed to a wide range of oxygen conditions and
378 according Mn concentrations in the pore waters during their growth. This stable and relatively
379 low export productivity is not only exhibited by diatom accumulation but also carbonate (CaCO₃
380 = 30-45%) during late MIS3 to late LGM (Romero et al., 2008; McKay et al., 2014).

381

382 However, by late Heinrich Event 3 (H3: 32.5-29 ka), whilst foraminiferal Mn/Ca is low, there is
383 an increase in bulk sediment Mn/Al, suggesting penetration of oxygen-rich bottom waters within
384 the upper centimetres of the sediment column. Dissolved Mn available for the precipitation of
385 Mn (oxyhydr)oxides would most likely be sourced by diffusion from underlying sediments
386 where anoxic conditions prevail (Burdige, 1983). Based on this increase in sedimentary Mn/Al
387 coinciding with relatively low foraminiferal Mn/Ca, we therefore infer that the Mn/Al
388 enrichment occurred immediately below the oxygen-rich pore waters during late H3 and
389 throughout the period 30-25 ka, delimiting the oxygen penetration front and the upward diffusion
390 of Mn (Fig. 5a). **Therefore we suggest that the oxygen penetration depth is deeper, and thus the**
391 **precipitation of Mn too, than the living depth of *E. exilis*, which has been documented to live at**
392 **1-2 cm sediment depth (Caulle et al., 2014).** As both foraminiferal and sedimentary Mn results
393 correspond to low relative abundances (<5%) of the low oxygen indicator *E. exilis* and low
394 diatom input (Romero et al., 2008; McKay et al., 2014), this reinforces our interpretation of more
395 oxygenated conditions at the sea floor during H3. It associates with the scenario that during low
396 primary productivity export, oxygen levels are not lowered by the decomposition of large
397 amounts of fresh phytodetritus.

398 During the LGM, Mn/Ca show a greater range between specimens (three specimens exhibit
399 consistent Mn/Ca in the order of 4 - 16 $\mu\text{mol mol}^{-1}$ whilst others suggest a greater intra-test
400 variability of 16 – 230 $\mu\text{mol mol}^{-1}$) than samples from MIS3 (Fig. 3) despite diatom input being
401 relatively low and stable during this interval. **We interpret the greater range in Mn/Ca as more**

402 variable oxygen levels and a relative decrease within the pore water overall, in comparison to
403 earlier times within the record (Fig. 5b), but not to the extent of hypoxia since *E. exilis*
404 abundance remains low whilst species diversity is high (McKay et al., 2014). Simultaneously,
405 Mn/Al remains low (Mn content <0.06%) which could relate to bottom water masses since Mn
406 can be transported to deep waters via scavenging (Glasby, 2006). In particular, Gallego-Torres et
407 al., (2014) suggest that site GeoB7926-2 was under the influence of Antarctic Bottom Water
408 (AABW) during the LGM. AABW contains lower Mn concentrations relative to North Atlantic
409 water masses (Statham et al., 1998; Idrus, 2013). Mn is scavenged from AABW as it flows north
410 and thus when reaching site GeoB7926-2 is Mn-impoverished. However, we propose that low
411 Mn fixation in the sediment during the LGM is due to low export production leading to relatively
412 more Mn being located in the water column.

413

414 4.2.2 Heinrich Event 1 – Younger Dryas (18 - 11.5 ka)

415 During H1, the increase in foraminiferal Mn/Ca indicates lower oxygen conditions in the pore
416 waters (Fig. 5c). Lower oxygen conditions are evidenced in terms of both the greater variability
417 within the individual tests (Fig. 3) and the higher average Mn/Ca per sample depth (Fig. 4). The
418 comparably lower oxygen conditions are consistent with our hypothesis which stemmed from the
419 benthic foraminiferal faunal assemblage composition (Filipsson et al., 2011; McKay et al., 2014)
420 whereby the dominance of *E. exilis* indicated low oxygen conditions at the seafloor as a result of
421 high primary productivity and export flux.

422 Bulk Mn/Al is moderately low (below average of 3.4×10^{-2} at the corresponding sample depth of
423 the benthic foraminifera analysed by SIMS) when compared to the rest of the record, despite
424 high diatom accumulation rate. The combined interpretation of Mn, Mo and U to Al ratios
425 presented in Gallego-Torres et al., (2014) evidenced that the H1 period experienced suboxic
426 ($<0.1 \text{ ml l}^{-1} \text{ O}_2$ as defined by Morrison et al., 1998) to anoxic bottom water conditions. Under
427 these conditions, the redoxcline would have been very shallow or even located at the sediment-
428 water interface, so that Mn was not preferentially fixed in the sediment. Furthermore, the
429 sedimentary layer might have acted as a source of reduced dissolved Mn to the water column
430 and/or the uppermost pore waters, allowing for higher Mn availability for incorporation into
431 benthic foraminiferal tests.

432 During the BA, foraminiferal Mn/Ca determined by SIMS exhibit a similar range of values and
433 intra-test variability as during H1 (Fig. 3). **We relate this similarity in oxygen conditions to**
434 **comparable diatom accumulation rates during both climatic intervals.** Previously, from the
435 relative decrease in *E. exilis* abundance during the BA, the bottom water oxygen concentrations
436 were interpreted to have increased in comparison to H1 and the YD (Filipsson et al., 2011).
437 Despite this benthic faunal response to productivity export, the diatom input itself was relatively
438 higher at the core depth sampled for SIMS analysis within the BA than the one in H1.
439 Subsequently, our SIMS derived Mn/Ca results do not suggest vast redox shifts between the two
440 climatic intervals generally the values follow the diatom input. This emphasizes that whilst the
441 benthic foraminiferal community composition is a reliable indicator of past environmental
442 conditions, the trace elemental composition of foraminiferal tests has potential to reveal a more
443 detailed palaeoenvironmental interpretation. Furthermore, the foraminiferal Mn/Ca coincides
444 with the lack of Mn enrichment in the bulk sediment and Mn/Al does not substantially differ
445 from the previous climatic interval either. Mo and U suggest prevailing suboxic conditions
446 during the BA (Gallego-Torres et al., 2014) and further, we infer that the redoxcline would be
447 shallow within the sediment which is similar scenario to H1.

448 From both SIMS and FT-ICP-OES, the highest foraminiferal Mn/Ca and greatest Mn variability
449 within individual tests are found during the YD (Fig. 3). This consistent pattern of Mn
450 enrichment coincides with high primary productivity, high export flux and the dominance of low
451 oxygen tolerant *E. exilis*. As MnO₂ is rapidly reduced to soluble Mn²⁺ in hypoxic pore waters
452 **(Burdige 1983; Limburg et al., 2011 and references therein)** and since high phytodetritus export
453 typically causes low bottom water oxygen concentrations or even anoxia within millimetres of
454 the sediment-water interface, **we can expect a high accumulation of Mn** (Hunt, 1983; McKay et
455 al., 2005) as represented in Fig 5d. Furthermore, the sediment accumulation rate during the
456 interval is the highest of the record (350 cm ka⁻¹, Fig. 4) which can also intensify the low oxygen
457 concentrations and promote Mn enrichment (McKay et al. 2005). This confirms our hypothesis
458 that Mn/Ca values in *E. exilis* respond to the oxygen deficiency instigated by the large increase
459 in diatom input. Furthermore, the redox front would also vary in position during this oxygen
460 depleted period with diatom export fluctuating at such great magnitude. Therefore the strong
461 intra-test variability (Fig. 3) may be representative of actual changes in oxygenation of the
462 habitat during test growth, due to amplified seasonal fluctuations in diatom export. Moreover,
463 since *E. exilis* is an infaunal species, it is influenced by the microhabitat of the pore waters.
464 Therefore, the sediment depth at which this species resided and possibly migrated to during

465 calcification in such low oxygen conditions could further explain the greater Mn/Ca variability
466 during the YD.

467 We emphasise that the benthic foraminiferal Mn/Ca seems to represent a more regional signal
468 due to diatom input opposed to being generated by deep bottom water formation and poor
469 ventilation. However, whilst we interpret that the sheer level of diatom input provoked O₂
470 deficiency in the bottom and pore waters, we acknowledge that even in the modern ocean, it is
471 difficult to separate the effects of productivity and deep-water oxygen concentrations since they
472 are inter-related. Gallego-Torres et al., (2014) interpret the YD as a phase of reduced ventilation
473 coinciding with reduced Atlantic Meridional Overturning Circulation (AMOC), promoted by
474 intense export productivity. Both effects provided dissolved Mn available for incorporation into
475 the foraminiferal calcite. Therefore, whilst Mn/Ca has potential for oxygen level reconstruction,
476 supplementary proxies are required in order to determine the precise factor driving the
477 conditions.

478

479 **5. Conclusion**

480 Our study contributes to the development of Mn/Ca in benthic foraminiferal calcite as a proxy
481 for reconstructing past oxygen conditions. The results based on data from the low latitude NE
482 Atlantic Upwelling System indicate that shifts in oxygen levels occurred during different
483 productivity regimes between 35 and 11.5 ka and thus foraminiferal Mn/Ca can assist our
484 understanding of the past environment in the region studied. The foraminiferal Mn/Ca results are
485 supported by benthic foraminiferal faunal data.

486 The highest foraminiferal Mn/Ca and greatest Mn variability within individual tests were
487 obtained during the YD and indicate Mn enrichment which coincides with very high primary
488 productivity and the dominance of low oxygen tolerant benthic foraminifera *Eubulimina*
489 *exilis*. The results confirm our hypothesis that Mn/Ca in *E. exilis* can be applied as a proxy for
490 oxygen deficiency, in this case instigated by the increase in diatom input. **Therefore, whilst the**
491 **benthic faunal abundance data is indicative of such a scenario, foraminiferal Mn/Ca allows a**
492 **more comprehensive interpretation.** Our initial down-core dataset raises the implication of
493 calibrations. Once developed, Mn/Ca determined by the SIMS method in particular may have the
494 potential to be applied to other study regions and foraminiferal species for reconstructing former
495 bottom water oxygen conditions.

496 Furthermore, we conclude that SIMS determined Mn/Ca upon individual tests is comparable
497 with bulk foraminiferal Mn/Ca measured by FT-ICP-OES. However, due to the processing time
498 required to program and target delicately thin foraminiferal test walls, SIMS may not be practical
499 for Mn/Ca studies where large numbers of samples must be processed. **Nevertheless, we**
500 **emphasize that SIMS has great potential to provide reliable Mn/Ca results from just a few**
501 **individual foraminifera. Therefore, SIMS is a robust alternative method to FT-ICP-OES; ideal**
502 **for employing on samples that lack a sufficient abundance of individual benthic foraminiferal**
503 **specimens for solution-based bulk analyses.** Furthermore, SIMS is also non-destructive and thus
504 foraminiferal test cross-sections can even be re-measured.

505 In contrast, foraminiferal Mn/Ca data does not continuously exhibit a consistent trend with
506 Mn/Al determined from bulk sediment measurements. The reason for this discrepancy is that Mn
507 related to redox fronts within the sediment provides a diagenetic signal and thus it continues to
508 react and shift after deposition, whereas foraminiferal tests record the Mn concentration at the
509 time of calcification.

510

511 **Author contribution:**

512 H.L. Filipsson initiated the project; C.L. McKay designed the sampling plan and carried out
513 sample selection and preparation with assistance from T. Toyofuku. J. Groeneveld performed
514 FT-ICP-OES analyses, C.L. McKay and M.J. Whitehouse performed SIMS analyses. D.
515 Gallego-Torres and O.E. Romero provided Mn/Al and diatom data. C.L. McKay prepared and
516 wrote the manuscript with contributions from all co-authors.

517

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725 Table 1. Sample list: climatic intervals, export productivity according to diatom export, average
 726 foraminiferal Mn/Ca for different samples determined by SIMS and FT-ICP-OES.

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Sample no.	Depth (cm)	Age (ka)	Climatic interval	Export productivity	SIMS: average Mn/Ca ($\mu\text{mol mol}^{-1}$)	1 sd ($\mu\text{mol mol}^{-1}$)	SIMS: average Mn/Ca ($\mu\text{mol mol}^{-1}$) per sample depth	FT-ICP-OES : average Mn/Ca ($\mu\text{mol mol}^{-1}$)		
YD A	170	12.2	YD	High	138	62	198	220		
YD B	170	12.2	YD	High	191	116				
YD C	170	12.2	YD	High	321	251				
YD D	170	12.2	YD	High	141	56				
YD E	270	12.7	YD	High	178	82	164	160		
YD F	270	12.7	YD	High	96	53				
YD G	270	12.7	YD	High	117	48				
YD H	270	12.7	YD	High	164	120				
YD I	270	12.7	YD	High	183	51				
YD J	270	12.7	YD	High	111	27				
YD K	270	12.7	YD	High	261	275				
YD L	270	12.7	YD	High	204	78				
BA A	365	13.5	BA	Moderate-high	155	37			Only 1 specimen available	280
BA B	430	15.1	BA	Moderate-high	242	42			130	
BA C	430	15.1	BA	Moderate-high	63	51				
BA D	430	15.1	BA	Moderate-high	217	51				
BA E	430	15.1	BA	Moderate-high	49	18				
BA F	430	15.1	BA	Moderate-high	179	88				
BA G	430	15.1	BA	Moderate-high	175	87				
BA H	430	15.1	BA	Moderate-high	67	149				
BA I	430	15.1	BA	Moderate-high	47	27				
H1 A	500	16.7	H1	High	57	17	117	140		
H1 B	500	16.7	H1	High	154	36				
H1 C	500	16.7	H1	High	186	81				
H1 D	500	16.7	H1	High	191	142				
H1 E	500	16.7	H1	High	76	57				
H1 F	500	16.7	H1	High	92	37				
H1 G	500	16.7	H1	High	62	19				
H1 H	500	16.7	H1	High	116	36				
LGM A	723	20.7	LGM	Low	27	14	74	Insufficient no. of specimens		
LGM B	723	20.7	LGM	Low	146	28				
LGM C	723	20.7	LGM	Low	115	71				
LGM D	723	20.7	LGM	Low	7	4				
LGME	773	22.7	LGM	Low	7	3	65			
LGM F	773	22.7	LGM	Low	10	3				
LGM G	773	22.7	LGM	Low	70	49				

LGM H	773	22.7	LGM	Low	175	91		
MIS3 A	928	29.9	MIS3	Low	38	13	43	Insufficient no. of specimens
MIS3 B	928	29.9	MIS3	Low	48	38		
MIS3 C	928	29.9	MIS3	Low	45	24		
MIS3 D	1058	34.0	MIS3	Low	67	23	61	
MIS3 E	1058	34.0	MIS3	Low	55	62		

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748 Table 2. Post hoc test (Mann Whitney U) z values of the mean foraminiferal Mn/Ca (per
749 individual) between climatic intervals. Significant differences are highlighted in bold.

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	YD	BA	H1	LGM	MIS3
YD					
BA	-1.14				
H1	-1.93	-0.19			
LGM	-2.70	-1.83	-1.68		
MIS3	-3.16	-2.07	0	0	

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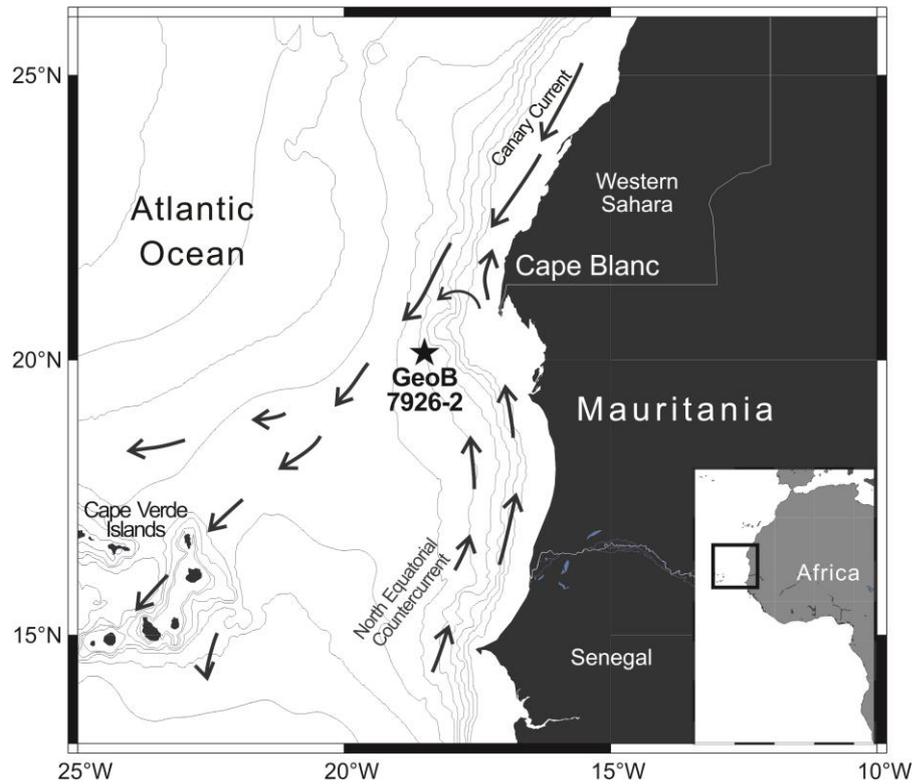
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770 Figure 1. Locality of gravity core GeoB7926-2 (black star) in the low latitude NE Atlantic
 771 upwelling area. Arrows indicate the major oceanic currents in the study area. Inset: Location of
 772 the study area off coastal NW Africa. Modified after Romero et al. (2008).

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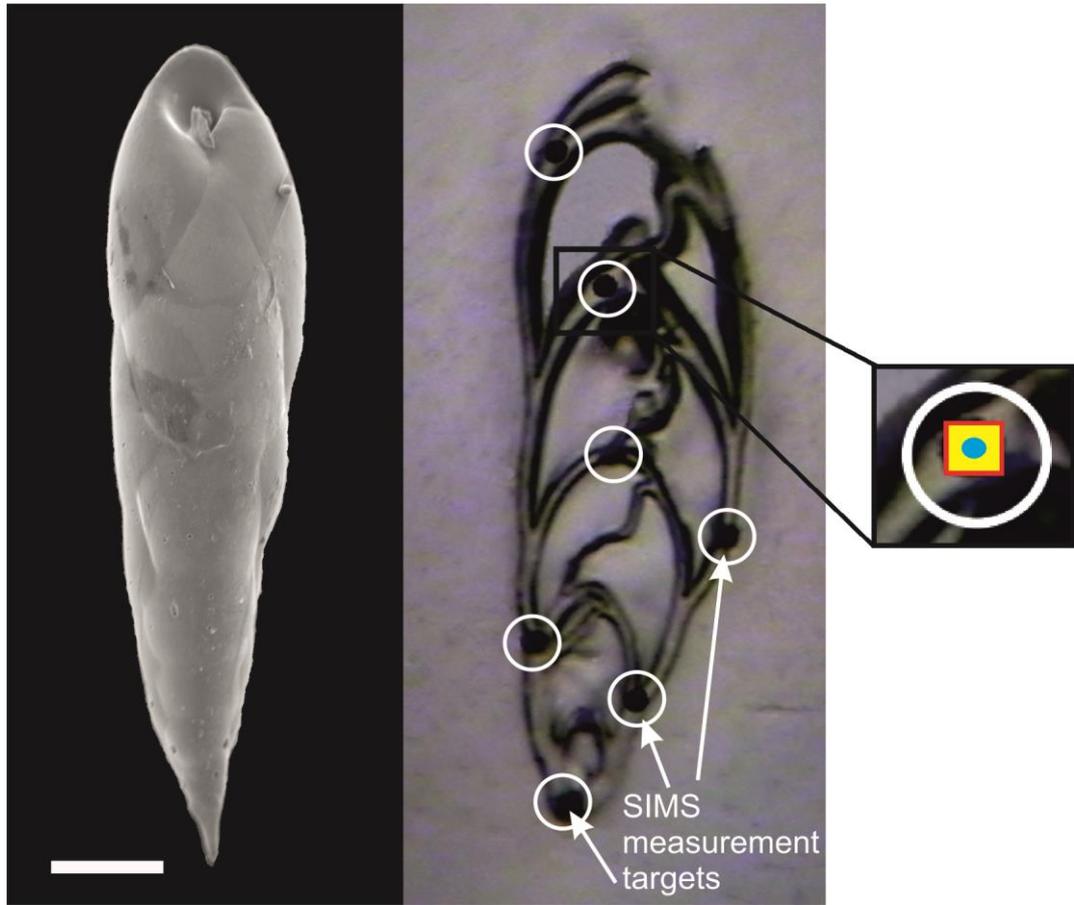
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782 Figure 2. SEM image (scale bar: 100 µm) and cross section image during SIMS analysis of a
 783 single *Eubuliminella exilis* specimen. The white circles highlight the selected spots for SIMS
 784 analyses, measuring 5 µm in diameter. Note that the black areas of the SIMS measurement
 785 targets visible in this image are actually the 5 µm spots plus the 10 µm pre-sputters. Inset is a
 786 close-up of the SIMS targets: the red square is the approximate pre-sputter area (15 x 15 µm, i.e.
 787 5 µm spot + 10 µm raster), the yellow area is the field of view admitted to the mass spectrometer
 788 (controlled by magnification and field aperture) and the blue ellipse is the nominal 5 µm spot.

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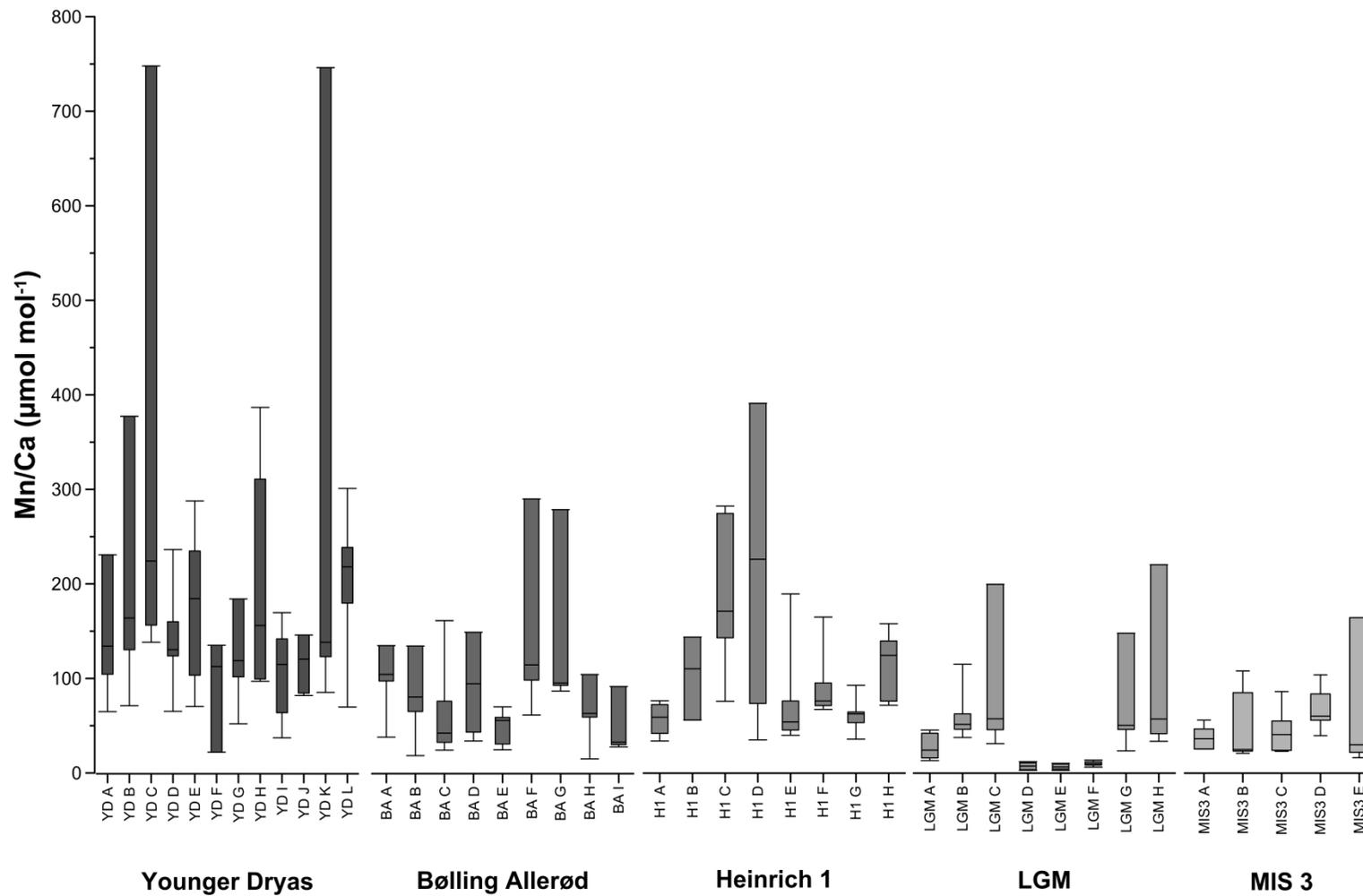
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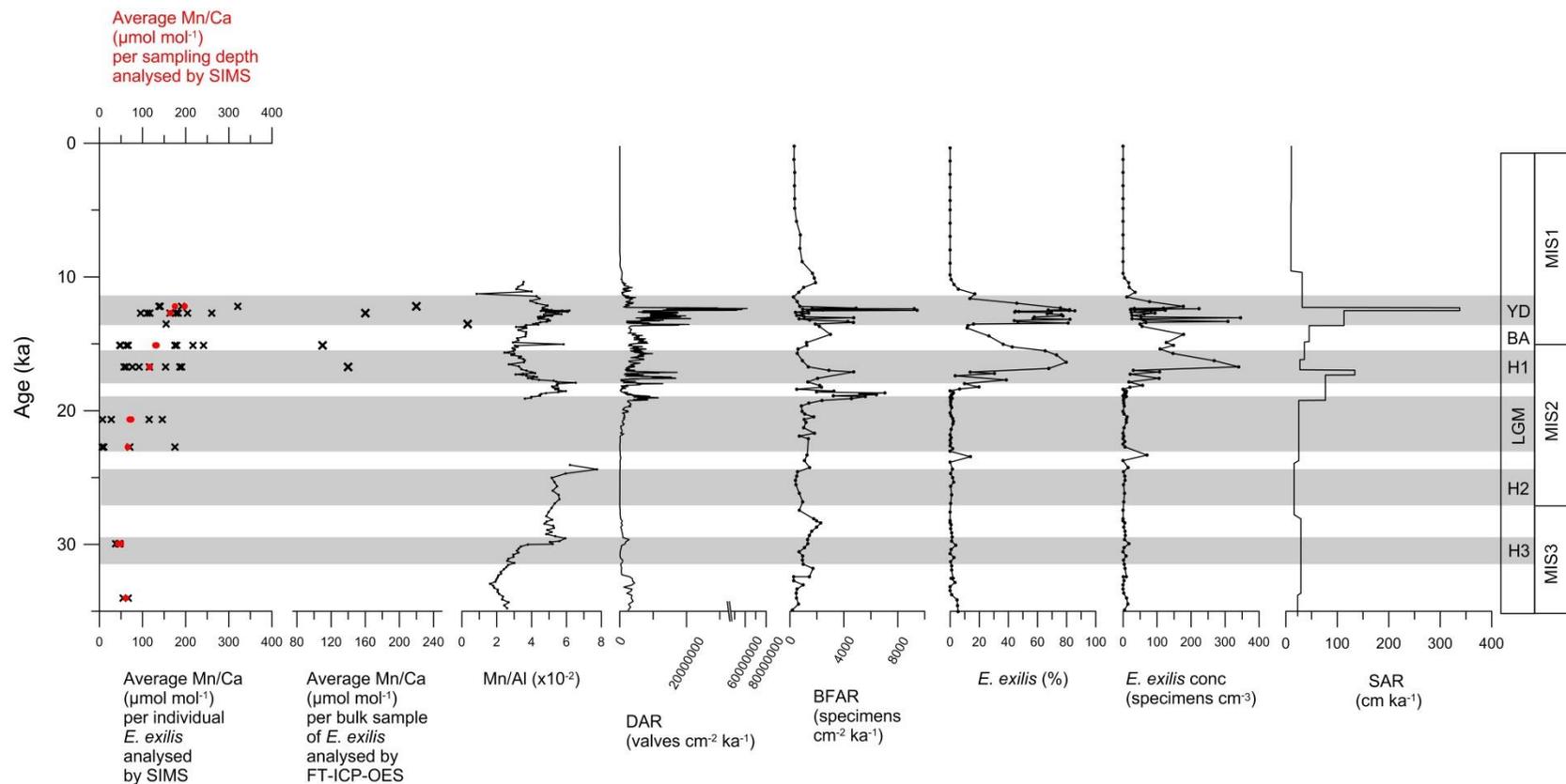


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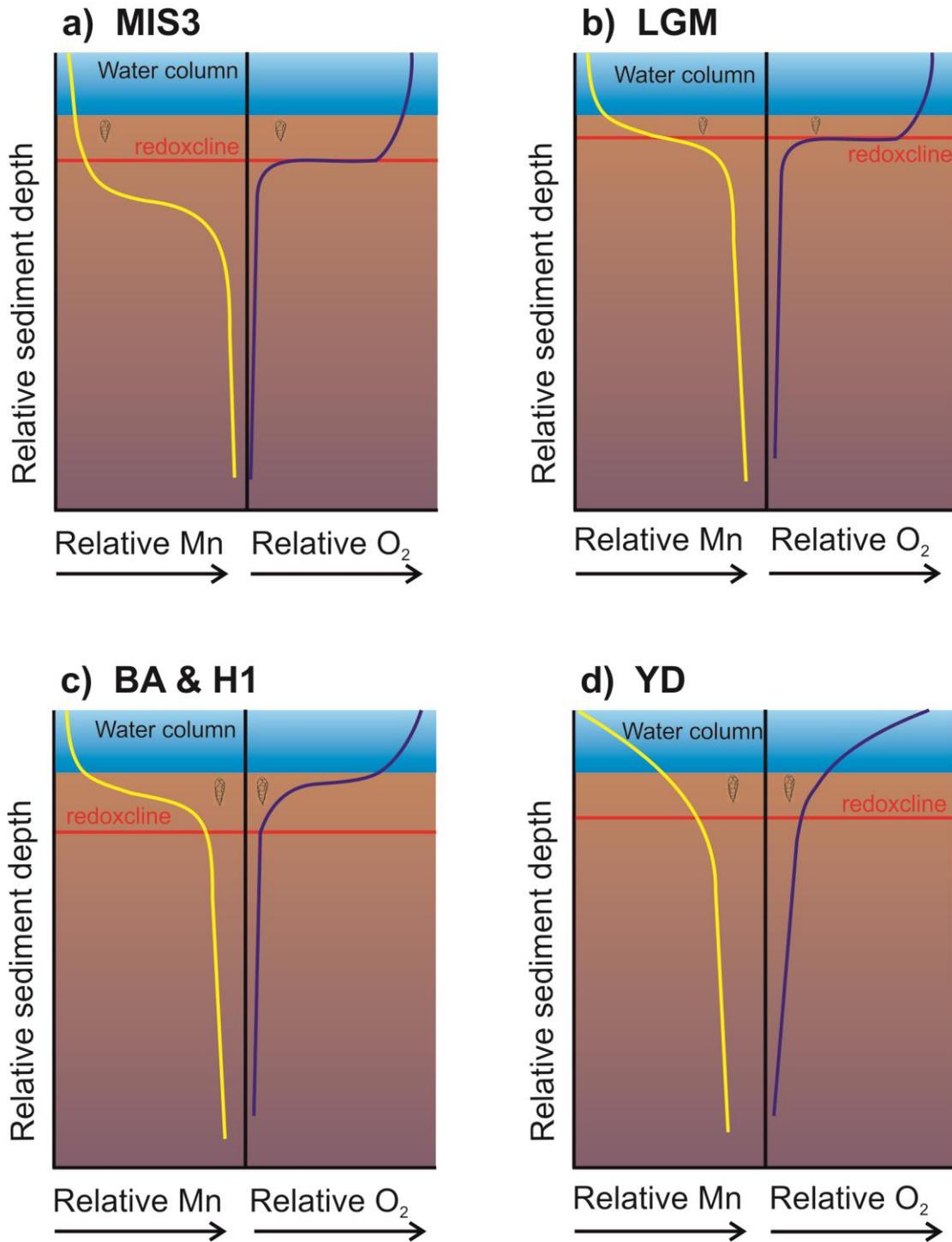
800 Figure 3. The Mn/Ca ($\mu\text{mol mol}^{-1}$) variability within each individual *Ebuliminella exilis* specimen for each climatic interval (labelled on the x-
 801 axis), determined by SIMS.

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804 Figure 4. Foraminiferal Mn/Ca ($\mu\text{mol mol}^{-1}$) determined by SIMS (black crosses: average per specimen, red circles: average per depth) and FT-
 805 ICP-OES, bulk sediment Mn/Al (Gallego-Torres et al., 2014), diatom accumulation rate (DAR; Romero et al., 2008), **Benthic foraminiferal**
 806 **accumulation rate (BFAR; McKay et al., 2014)**, relative abundance and concentration of low oxygen indicating *Eubuliminella exilis* and
 807 sediment accumulation rate (SAR) of core GeoB7926-2.



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811 Figure 5. Idealised schematic pore water Mn (yellow) and oxygen (blue) profiles for the climatic
 812 intervals studied from the GeoB7926-2 record. The red line represents the redoxcline.

813 a) MIS3: Low foraminiferal Mn/Ca and therefore the sedimentary Mn was precipitating below
 814 the living depth of *E. exilis* and more oxygenated conditions prevailed in the bottom and pore

815 waters. b) LGM: foraminiferal Mn/Ca increases and therefore a relative decrease in pore water
816 oxygen is apparent and the redoxcline is shallower. c) BA & H1: foraminiferal Mn/Ca increases
817 further and the moderate-low sedimentary Mn/Al indicates low bottom water conditions. d) YD:
818 highest accumulation of Mn and therefore oxygen deficiency in the bottom and pore waters.

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