

Drivers and uncertainties of future global marine primary production in marine ecosystem models

Charlotte Laufkötter¹, Meike Vogt¹, Nicolas Gruber¹, Maki Aita-Noguchi⁷, Olivier Aumont², Laurent Bopp³, Erik Buitenhuis⁴, Scott C. Doney⁵, John Dunne⁶, Taketo Hashioka⁷, Judith Hauck⁸, Taka Hirata⁹, Jasmin John⁶, Corinne Le Quéré¹⁴, Ivan D. Lima⁵, Hideyuki Nakano¹³, Roland Seferian¹⁵, Ian Totterdell¹⁰, Marcello Vichi^{11,12}, and Christoph Völker⁸

¹Environmental Physics, Institute of Biogeochemistry and Pollutant Dynamics, ETH Zürich, Zürich, Switzerland

²Laboratoire de Physique des Océans, Centre IRD de Bretagne, Plouzane, France

³Laboratoire des sciences du climat et de l'Environnement (LSCE), IPSL, CEA-UVSQ-CNRS, UMR8212, Gif-sur-Yvette, France

⁴Tyndall Centre for Climate Change Research, School of Environmental Sciences, University of East Anglia, Norwich, UK

⁵Woods Hole Oceanographic Institution, Department of Marine Chemistry & Geochemistry, Woods Hole MA, USA

⁶NOAA Geophysical Fluid Dynamics Laboratory, Princeton, New Jersey, USA

⁷Core Research for Evolutional Science and Technology, Japan Science and Technology Agency, Tokyo, Japan

⁸Alfred Wegener Institute, Helmholtz Centre for Polar and Marine Research, Bremerhaven, Germany

⁹Faculty of Environmental Earth Science, Hokkaido University, Japan

¹⁰Met Office, Exeter, UK

¹¹Centro Euro-Mediterraneo sui Cambiamenti Climatici (CMCC), Bologna, Italy

¹²Department of Oceanography, University of Cape Town (UCT), South Africa

¹³Meteorological Research Institute, Tsukuba, Ibaraki, Japan

¹⁴Tyndall Centre for Climate Change Research, University of East Anglia, Norwich Research Park, Norwich NR47TJ, UK

¹⁵CNRM-GAME, Centre National de Recherche Mtorologique-Groupe d'Etude de l'Atmosphère Mtorologique, Mto-France/CNRS, 42 Avenue Gaspard Coriolis, 31100 Toulouse, France

Correspondence to: Charlotte Laufkötter, charlotte.laufkoetter@env.ethz.ch

Abstract.

Past model studies have projected a global decrease in marine net primary production (NPP) over the 21st century, but these studies focused on the multi-model mean rather than on the large inter-model differences. Here, we analyze model simulated changes of NPP for the 21st century under IPCC's high emission scenario RCP8.5. We use a suite of nine coupled carbon-climate Earth System Models with embedded marine ecosystem models and focus on the spread between the different models and the underlying reasons. Globally, NPP decreases in five out of the nine models over the course of the 21st century, while three show no significant trend and one even simulates an increase. The largest model spread occurs in the low latitudes (between 30°S and 30°N), with individual models simulating relative changes between -25% and +40%. Of the seven models diagnosing a net decrease in NPP in the low latitudes, only three simulate this to be a consequence of the classical

interpretation, i.e., a stronger nutrient limitation due to increased stratification leading to reduced
15 phytoplankton growth. In the other four, warming-induced increases in phytoplankton growth out-
balance the stronger nutrient limitation. However, temperature-driven increases in grazing and other
loss processes cause a net decrease in phytoplankton biomass and reduce NPP despite higher growth
rates. One model projects a strong increase in NPP in the low latitudes, caused by an intensification
of the microbial loop, while NPP in the remaining model changes less than 0.5%. While models
20 consistently project increases NPP in the Southern Ocean, the regional inter-model range is also
very substantial. In most models, this increase in NPP is driven by temperature, but it is also mod-
ulated by changes in light, macronutrients and iron as well as grazing. Overall, current projections
of future changes in global marine NPP are subject to large uncertainties and necessitate a dedicated
and sustained effort to improve the models and the concepts and data that guide their development.

25 **1 Introduction**

By producing organic matter, marine phytoplankton form the base of the marine food web, control
the amount of food available for higher trophic levels, and drive the majority of the ocean's
biogeochemical cycles, particularly that of carbon. The net formation rate of organic carbon by
phytoplankton, i.e., net primary production, NPP, is a key determinant for the export of organic car-
30 bon from the surface ocean, thereby governing how ocean biology impacts the ocean/atmosphere
exchange of CO₂ (Falkowski et al., 2003; Sarmiento and Gruber, 2006). Accurate projections of fu-
ture patterns of NPP may be crucial not only to estimate the potential impacts of climate change on
marine ecosystems and fishery yields, but also to properly assess the evolution of the ocean carbon
sink under anthropogenic climate change.

35 Several authors have analyzed trends in future NPP and the underlying drivers, using models of
strongly varying complexity and spatial resolution with regard to both the physical and the ecosys-
tem components and also investigating different climate change scenarios. In the majority of these
studies, global marine NPP was projected to decrease in response to future climate change (Bopp
et al., 2001; Boyd and Doney, 2002; Steinacher et al., 2010; Bopp et al., 2013; Marinov et al., 2013;
40 Cabré et al., 2014). The main mechanism suggested to explain this decrease in NPP was a decrease in
the upward supply of nutrients in the low latitudes because of increased vertical stratification (Bopp
et al., 2001; Steinacher et al., 2010) and reduced upwelling. Lower nutrient availability resulted then
in a decrease in phytoplankton growth and therefore reduced NPP.

However, a few studies produced contradicting results, i.e., they reported increases in global
45 NPP as climate change progresses over the 21st century (Sarmiento et al., 2004; Schmittner et al.,
2008). Taucher and Oschlies (2011) showed that in the case of the model used by Schmittner et al.
(2008), the simulated increase in NPP is caused by the warmer temperatures enhancing phytoplank-
ton growth, overcoming the suppression of their growth owing to stronger nutrient stress. But this

result cannot be easily generalized, since some models used in Steinacher et al. (2010) still project a
50 decrease in NPP even though they have a stronger temperature dependence of the growth rate than
that used in the model by Schmittner et al. (2008).

The past century provides very little experimental constraint on the impact of long-term climate
change on marine productivity, largely because of the lack of long-term (>50 yrs) observations.
Using a combination of in-situ observations of chlorophyll and of ocean transparency, Boyce et al.
55 (2010) suggested a substantial decrease in phytoplankton biomass over the last 50 years, imply-
ing a very strong response of phytoplankton to ocean warming. This result has been met with a
lot of scepticism (e.g. Rykaczewski and Dunne, 2011), especially because an independent assess-
ment of long-term trends in ocean color by Wernand et al. (2013) implied no overall global trend.
Smaller decreases in NPP (-6% over 50 years) were suggested by a hindcast simulation, where a
60 marine ecosystem model coupled to an ocean general circulation model was forced with observed
atmospheric variability and changes over the last 50 years (Laufkötter et al., 2013). The satellite ob-
servations since late 1997 suggest a negative correlation between sea surface temperature and NPP
(Behrenfeld et al., 2006), but the observation period is clearly too short to distinguish natural fluctu-
ations from an anthropogenically driven trend in global marine NPP (Henson et al., 2011; Antoine
65 et al., 2005; Gregg, 2003).

Far less work has been done regarding future trends in the biomass of specific plankton functional
types (PFT), despite their importance in shaping ecosystem structure and function (Le Quéré et al.,
2005). Experiments have revealed a negative relationship between warmer waters and phytoplank-
ton cell size, suggesting that future warming may tend to favor small phytoplankton (Morán et al.,
70 2010). Moreover, using year-to-year variability associated with the North Atlantic Oscillation and
the Southern Annular Mode, Alvain et al. (2013) found that more stagnant conditions and warmer
temperatures tend to disfavor diatoms, suggesting that diatoms will become less prevalent in the
future. The few modeling studies available support this view, i.e., they reported global decreases in
the diatom fraction and a shift towards smaller size classes (Bopp et al., 2005; Marinov et al., 2010,
75 2013; Dutkiewicz et al., 2013). In these models, this shift was driven by increased nutrient limitation
that affected diatoms more strongly than small phytoplankton.

While published studies emphasized the role of changes in bottom-up factors in explaining the
changes in NPP, top-down control by zooplankton grazing may also drive future changes in total
NPP or phytoplankton composition. This mechanism is intriguing, since top-down control was
80 recently identified as one of the main drivers of phytoplankton competition during blooms in several
ecosystem models (Hashioka et al., 2013; Prowe et al., 2011). Further, top-down control affects the
onset of the spring bloom (Behrenfeld, 2010; Behrenfeld et al., 2013), influences primary production
in a trait-based ecosystem model (Prowe et al., 2012) and affects NPP and EP changes on regional
scales (Bopp et al., 2001).

85 Previous efforts in comparing different models with regard to future trends in NPP have analyzed

the multi-model mean response and focused on identifying regions of consistent changes and mechanisms among models (Steinacher et al., 2010; Bopp et al., 2013; Cabré et al., 2014). By largely disregarding the regions of large inconsistencies, this focus may have underestimated the uncertainty associated with current projections of future marine NPP changes. This is well illustrated by
90 the most recent model comparison study by Bopp et al. (2013), where the spread in the global NPP change between the 10 investigated global models for a given climate change scenario was larger (-20% and +2%) than the NPP difference between the different scenarios for the multi-model mean (-9% to -2%), demonstrating that the model uncertainty is larger than the scenario uncertainty.

Reasons why models differ are seldom investigated in model comparison studies. In particular, it
95 is often not readily clear whether the large spread in model projections is mainly caused by differences in the underlying ocean circulation model, by differences in the complexity of the ecosystem models or by differences in the parameterizations leading to differing sensitivities to e.g., changes in temperature, nutrients and light. Such information is needed, however, in order to improve the existing models and eventually obtain reliable future projections.

100 In this work we go beyond the basic analysis of the multi-model mean and the identification of regions of model consistency. Our aim is to identify where models differ and by how much, and then determine why they do so, i.e., identify the underlying drivers of change. To this end, we use results from a set of 8 global marine ecosystem models coupled to or forced with 9 coupled carbon-climate Earth System Models, which have simulated the future evolution of marine NPP under the
105 Intergovernmental Panel on Climate Change (IPCC) Representative Concentration Pathways (RCP) 8.5 (van Vuuren et al., 2011). We decompose the long-term changes in NPP into the contributions of the different phytoplankton functional types and then identify the relative importance and uncertainty of the main drivers. We demonstrate that i) current marine ecosystem models are revealing more spread with regard to future changes of NPP than shown previously, and (ii) even where the models
110 simulate consistent changes, the underlying drivers are quite different. In particular, we highlight the critical, but not well quantified role of temperature change in determining the future changes in NPP.

2 Methods

2.1 Model descriptions

115 We use projections for the 2012-2100 period of nine model simulations for IPCC's RCP8.5 scenario from either the "MARine Ecosystem Model Intercomparison Project" (MAREMIP, <http://pft.ees.hokudai.ac.jp/maremip/index.shtml>, (Vogt et al., 2013; Sailley et al., 2013; Hashioka et al., 2013)) and/or the "Coupled Model Intercomparison Project 5" (CMIP5, Taylor et al. (2012)). As we perform an analysis of the effect of PFT composition on NPP changes, we included only data
120 from those models that possess at least 2 phytoplankton PFTs and at least one zooplankton PFT. For

the models taken from the CMIP5 archive, only the first ensemble member (r1i1p1) was used.

These criteria led us to use data from eight different marine ecosystem models: diat-HadOCC, BEC, TOPAZ, PISCES, MEM, PELAGOS, REcoM2 and PlankTOM5.3 (Table 9 lists the model acronyms, their main references, and further information, e.g., on spin-up times). Since the same
125 ocean ecosystem model PISCES was used in two different Earth System Models, we analyze a total of 9 different simulations. In most simulations, the ecosystem model was embedded into a coupled climate model and integrated over thousands of years in order to spin-up the model under pre-industrial conditions (see Table 9). In two simulations (REcoM2 and PlankTOM5.3), the ecosystem model was used within a forced ocean model and was initialized with observed climatologies. In
130 these simulations, a control run showed considerably smaller drift than the climate change response. We do not correct the small drift in these models to keep the internal mechanisms in the models consistent.

We describe the most important features of the ecosystem models in the following, and give the full equations and parameters for the offline calculations shown in this work in the Appendix. The
135 ocean ecosystem models used in this study are structurally similar, but they differ substantially in their details (see Table 10 for an overview of the model structures). Within our selection, all models simulate at least two phytoplankton PFTs, usually representing diatoms and a nanophytoplankton type, and one zooplankton PFT. BEC and TOPAZ have an additional diazotrophic phytoplankton PFT. Moreover, TOPAZ differentiates between diatoms and other large phytoplankton depending
140 on the availability of silicic acid. In PELAGOS, the nanophytoplankton type is further divided into flagellates and picophytoplankton. PlankTOM5.3 includes an explicit coccolithophore type, while in most other models coccolithophores are modeled implicitly as a fraction of nanophytoplankton. Regarding zooplankton PFTs, TOPAZ only has implicit zooplankton activity, diat-HadOCC, BEC, and REcoM2 have one zooplankton type, while PISCES and PlankTOM5.3 differentiate between
145 micro- and mesozooplankton. MEM and PELAGOS have three zooplankton types, i.e., in addition to the micro- and mesozooplankton, they include predatory zooplankton in MEM and heterotrophic flagellates in PELAGOS. Finally, PELAGOS is the only model that includes heterotrophic bacteria explicitly.

2.2 Analysis of NPP and its drivers

A change in NPP can be driven by (i) a change in the biomass specific rate of photosynthesis, (ii) changes in autotrophic respiration, or (iii) changes in phytoplankton biomass through, e.g., zooplankton grazing, sinking and other loss processes of phytoplankton. However, only PELAGOS and REcoM2 model photosynthesis (gross primary production, GPP) and autotrophic respiration separately. Rather, most models calculate NPP directly as the product of the growth rate μ and biomass of phytoplankton, P . In these latter models, changes in marine NPP can thus result only from (i) changes in the phytoplankton growth rate and (ii) changes in phytoplankton biomass. In order to

disentangle these two main classes of drivers, it is helpful to consider the full mass balance equation for any phytoplankton type P_i :

$$\Gamma(P_i) = (\mu_i \times P_i) - \text{grazing} - \text{sinking} - \text{other losses} \quad (1)$$

150 where Γ is the sum of the time rate of change and the physical processes of advection, convection, and diffusion, and where the first term on the right hand side is NPP. We consider any driver that alters the growth rate μ_i as a bottom-up driver, while those that alter P , i.e., grazing, sinking, and other losses we consider as top-down drivers, even though only grazing is strictly speaking a top down process.

In all models, the growth rate of phytoplankton is parameterized using a multiplicative function of a maximum growth rate μ_{\max} , the temperature limitation T_f^P and the nutrient and light limitation factors $N_{\text{lim}}, L_{\text{lim}}$, i.e.,

$$\mu = \mu_{\max} \times T_f^P \times N_{\text{lim}} \times L_{\text{lim}} \quad (2)$$

155 In all eight models except for diat-HadOCC, the temperature dependence of phytoplankton growth, i.e., T_f^P is described using an exponential function based on Eppley (1972), albeit with rather different temperature sensitivities (i.e., Q_{10} values, see also Table 11). In diat-HadOCC, phytoplankton growth is independent of temperature. While in most models the same Q_{10} value is used for all phytoplankton PFTs, mesozooplankton has a higher Q_{10} in PISCES and PELAGOS and each PFT and process has its own Q_{10} value in PlankTOM5.3, derived from observations. In REcoM2 an Arrhenius function is used which results in a Q_{10} that decreases with temperature.

165 The nutrient and light limitation factors have dimensionless values between 0 and 1, with higher values promoting higher growth. All models consider limitation by multiple nutrients, with six out of the eight models applying Liebig's law of the minimum (Liebig, 1840), such that the value of the strongest limiting nutrient sets the total nutrient limitation. Thus, these models do not consider nutrient co-limitation. Exceptions to this are PELAGOS and diat-HadOCC, where nutrient limitation is multiplicative. In all models, nanophytoplankton growth is limited by nitrate and iron, while diatoms are additionally limited by silicic acid. In several models, limitation with respect to phosphate and ammonia is additionally considered (see Table 10). The limitation regarding a specific nutrient is 170 calculated either with Michaelis-Menten functions (Michaelis and Menten, 1913), following optimal uptake kinetics (Smith et al., 2009), or using a cell quota representation of nutrient deficiency, often with strong differences in half-saturation constants. The values of the half-saturation constants and the equations are given in the Appendix, Table 12 lists the type of nutrient limitation for the different models.

For diat-HadOCC, the full model equations are not available, therefore we cannot describe the light limitation. In all other models light limitation is parameterized based on the work of Geider et al. (1998); Webb et al. (1974) and Platt et al. (1980). Most models (except for MEM) use the

following equation:

$$L_{\text{lim}} = 1 - e^{\left(-\frac{\alpha \times \theta^{\text{chl:c}} \times \text{PAR}}{\mu_{\text{T,N}}}\right)} \quad (3)$$

175 where the constant parameter α denotes the initial slope of the photosynthesis-irradiance curve, $\theta^{\text{chl:c}}$ is the chlorophyll to carbon ratio, PAR is the photosynthetically available radiation and $\mu_{\text{T,N}}$ is the maximum growth rate multiplied with the temperature effect and nutrient limitation. PISCES models an additional strengthening in light limitation when the mixed layer depth is deeper than the euphotic zone. In PELAGOS, $\mu_{\text{T,N}}$ is replaced by a constant p_r for the maximum specific photosynthetic rate.
180 TOPAZ replaces the instantaneous chlorophyll to carbon ratio with a variable ratio that depends additionally on the memory of irradiance over the scale of 24 hours (see Appendix).

MEM uses the light limitation function from Platt et al. (1980):

$$L_{\text{lim}} = \frac{\left(1 - e^{\left(-\frac{\alpha \times \text{PAR}}{p_r}\right)}\right) \times e^{\frac{\beta \times \text{PAR}}{p_r}}}{\frac{\alpha}{\alpha + \beta} \times \frac{\beta}{\alpha + \beta}} \quad (4)$$

where β is a photo-inhibition index and α, p_r, PAR as above.

Note that in most models, temperature and nutrient status influence also the light limitation, such
185 that in addition to the direct effects of temperature and nutrients on the growth rate, there is an additional indirect way through their effect on light limitation (Geider et al., 1998).

Since PELAGOS does not compute NPP directly and also uses a different formulation for the growth limitation terms, it requires a separate analysis: In this model, NPP is calculated for each phytoplankton type by subtracting autotrophic respiration and other loss processes from its GPP,
190 i.e., $\text{NPP}_i = \text{GPP}_i - \text{exudation}_i - \text{respiration}_i - \text{lysis}_i$. GPP is calculated in PELAGOS in analogy to how NPP is calculated in the other models, i.e., using the product of biomass, maximum growth rate, temperature, light limitation and iron and silicic acid limitation. Nitrate and phosphate limitation are accounted for in the phytoplankton exudation and lysis terms. The reason for this differentiation between the various limiting nutrients is to account for internal storage capabilities of the phytoplankton cells (Vichi et al., 2007). To be able to compare PELAGOS to all other ecosystem models within
195 a common framework, we estimated a multiplicative nutrient limitation factor out of temperature, light limitation and the growth rate that was given in the PELAGOS output: $N_{\text{lim}} = \frac{\mu}{\mu_{\text{max}} \times T_f^P \times L_{\text{lim}}}$.

Regarding the loss terms for phytoplankton biomass, grazing is considered in all models. But given the large diversity in the complexity and parameterizations associated with the modeling of
200 zooplankton, the role of grazing may differ substantially among the considered models (Sailley et al., 2013).

Grazing of zooplankton Z on phytoplankton P is calculated as

$$\text{grazing}(Z, P) = g_{Z, \text{max}}^P * T_f^Z * \text{food dependence} * Z \quad (5)$$

in all models except TOPAZ, where g_Z^P is the maximum grazing rate of zooplankton Z on phytoplankton P and T_f^Z is the temperature limitation of zooplankton feeding. TOPAZ simulates the

effects of zooplankton implicitly and the representation of grazing is based on Dunne et al. (2005).

205 Most models employ the same temperature sensitivity for zooplankton as they use for phytoplankton, i.e., $T_f^Z = T_f^P$, with the exception of PISCES and PELAGOS, where the mesozooplankton has a higher temperature sensitivity, and PlankTOM5.3, where each PFT has a different Q_{10} value. The food dependence is modeled differently in each model and is shown in Table 13.

2.3 Data processing

210 Our analysis is based on monthly mean output for all surface ocean variables for the period 2012-2100. In order to facilitate direct comparisons, we regrid the model to a common 1x1 degree grid using the Earth System Modeling Framework (ESMF) regrid routines included in the NCAR Command Language (NCL) version 6.1.2, with interpolation method set to bilinear.

All models provided in the output vertically (0-100 m) integrated net primary production (NPP) and biomass (in carbon units) of all PFTs. Primary production by diatoms and small phytoplankton was not available for PlankTOM5.3, MEM and PELAGOS, and was estimated offline using the product of biomass and growth rate. The temperature limitations and growth rates were recalculated for all models except for PELAGOS and TOPAZ, where the growth rates were given in the model output. The nutrient and light limitation factors were included in the output of BEC, REcoM2 and 220 TOPAZ, while they were recalculated from the monthly mean data for all other models using the original (not interpolated) data. The equations used for the recalculations are given in the Appendix. A comparison of recalculated and true values in the BEC showed that the error in the recalculation is on the order of less than 10%.

Changes for all properties are computed by first averaging the data for two 20 year periods, i.e., 225 2012-2031 and 2081-2100, and then taking the difference. For the growth limitation factors, we show the ratio changes, i.e., for any limitation factor x , we show the ratio $\frac{\langle x \rangle_{(t=2081-2100)}}{\langle x \rangle_{(t=2012-2031)}}$, where the chevrons denote temporal averages. This is because the product of the relative changes in the temperature, light and nutrient limitation results approximately in the relative change in growth rate, and the factor with the strongest change also has the strongest effect on the change in growth rate.

230 3 Model evaluation

Most of the models analyzed in this study have been evaluated individually in their respective documenting publications (see references in Table 9). Therefore, we restrict ourselves to an evaluation of the variables that are most relevant for this work, i.e., vertically integrated net primary production (NPP), chlorophyll (chl), surface NO_3 , surface PO_4 and surface SiOH_4 (Fig. 1 and Tables 14, 15).

235 We compare modeled NPP, using a 1998-2007 climatology for each model, with results from the updated Carbon-based Production Model-2 algorithm derived from SeaWiFS satellite data (Westberry et al., 2008), downloaded from <http://www.science.oregonstate.edu/ocean.productivity/index.php>. For

chlorophyll, we use chlorophyll-a from the Sea-viewing Wide Field-of-view Sensor (SeaWiFS) Project generated by the NASA Goddard Space Flight Centre (<ftp://oceans.gsfc.nasa.gov>). We used
240 monthly means computed from Level 3 binned daily products. For both NPP and chlorophyll data we removed coastal values (depth < 500m) prior to the calculations. For the nutrients, we used the respective objectively analyzed climatologies from the World Ocean Atlas 2013 (Garcia et al., 2014) and compared it to model output for the 1990-1999 period.

On a global scale, the model simulated nitrate fields correlate reasonably well with the observa-
245 tions, with all models showing correlations between 0.62 and 0.85 and normalized standard deviations (NStdDev) between 0.86 and 1.10. However, the bias is rather large, with values between -4.24 and +4.89 mmol N m⁻³, corresponding to a bias of approximately ± 70% of the global average. For phosphate (not shown), the results are very similar to those of nitrate, but for silicic acid the models perform less successfully. The correlations are lower and between 0.45 and 0.76, the normalized
250 standard deviations scatter more, and the biases are larger (see Table 14).

The correlations for chlorophyll are mostly between 0.5 and 0.72, however the normalized standard deviations are rather low (most models have NStdDev values < 0.5). The higher standard deviation in the observations stems mostly from the coastal ocean (standard deviation decreases from 1.8 mg Chl m⁻³ to 0.5 mg Chl m⁻³ when removing coastal areas with water depths < 500 m).
255 Most models capture the lower open ocean variability, however in the two models that have a variability comparable to the observations (diat-HadOCC and PlankTOM.3), the variability arises from the open ocean and is therefore significantly higher than the observed open ocean variability.

Least well simulated is the distribution of NPP. The correlations are relatively low (0.18 - 0.69), the range of normalized standard deviation is as large as that of silicic acid (0.78 to 1.49), and in
260 some of the models, the bias is very large (-8.8 - +6.8 mol C m⁻² yr⁻²). Global annual NPP ranges between 17 and 83 Pg C yr⁻¹ (40.1 Pg C yr⁻¹ in the multi-model mean), compared to on average 50.7 Pg C yr⁻¹ in the satellite-based estimates (Carr et al., 2006) and 58 ± 7 based on ¹⁴C NPP (Buitenhuis et al., 2013a).

However, global correlations in nutrients and NPP are strongly influenced by the globally dominant gradient between the Southern Ocean and the low latitudes. While this gradient is generally
265 well reproduced by the models, the model skill in reproducing the regional nutrient and NPP patterns is considerably lower (not shown).

4 Results

4.1 21st century changes in primary production

270 Starting from very different levels, the models simulate global net primary production (NPP) to change under the RCP8.5 scenario anywhere from -15% to +30% (-4.3 to +10 Pg C yr⁻¹) over the 2012 to 2100 period (Fig. 2). One model shows an increase, five models show a decrease and three

models project changes less than 1%, which are not significant (p -value > 0.05) when compared to the level of inter-annual variability. The models suggest a median decrease of -7.2% with an inter quartile range (IQR) of 13.4% (-2 Pg C yr^{-1} with an IQR of 4.5 Pg C yr^{-1}). This is comparable to 275 the results reported by Bopp et al. (2013) using 10 Earth System Models from the CMIP5 project under RCP8.5 ($-8.6\% \pm 7.9\%$) and also to another recent multi-model comparison conducted by Steinacher et al. (2010) under SRES A2 emission scenario ($-10 \pm 3\%$, $-2.9 \pm 1.4 \text{ Gt C yr}^{-1}$). However, the range of projections covered by our study with respect to NPP (45%, 16% without 280 PlankTOM5.3) is higher than the 14% and 6% reported by Bopp et al. (2013) and Steinacher et al. (2010), respectively.

The regional pattern of the multi-model median change in NPP (Fig. 3b) shows distinct regional differences. The multi-model median suggest NPP increases in the Southern Ocean (south of 40°S , +10%), in the Arctic Ocean (+40%), in the southern Indian Ocean and in the southern subtropical 285 Pacific, while decreases by $-10.9 \pm 23.5\%$ are projected for the low latitudes (30°S - 30°N), with strongest decreases in the North Atlantic (-30%) and along the Equator in all basins. The range of NPP projections in different regions is given in Table 16. In most models as well as in the multi-model median, the decreases in the low latitudes are stronger than the increases in the high latitudes, resulting in the global decrease in NPP. This partial regional compensation was equally noted by 290 Bopp et al. (2013) and Steinacher et al. (2010). However, these changes are spatially heterogeneous and the multi-model mean masks differences between the individual models.

To illustrate these inter-model differences, we show the IQR (Fig. 3c) of the absolute change in NPP at each location. The IQR of NPP is around $1 \text{ mol C m}^{-2}\text{yr}^{-1}$ in the high and intermediate latitudes, which is of the same magnitude as the trends in the multi-model median changes. In the low 295 latitudes the IQR is significantly higher with values between 3 and $5 \text{ mol C m}^{-2} \text{ yr}^{-1}$, exceeding the multi-model median substantially. Thus, the model projections lack consistency, confounding direct interpretation of the multi-model median response.

4.2 Changes in bottom up versus top down control

The changes in NPP in the different models can be driven either by changes in the growth rates 300 (bottom-up) or phytoplankton biomass (top-down control) (see section 2.2 above). In order to obtain a first impression of the potential reasons underlying the NPP changes, we split the change in NPP into a component representing the change in the biomass of the whole phytoplankton community and a component representing the whole community growth rate. As the growth rates are only available at the surface in many models, we calculate the components for surface NPP changes. We computed 305 these two components by first calculating a first order Taylor decomposition of NPP into the changes in growth rate weighted with biomass and the changes in biomass weighted with growth rate within each model and for each phytoplankton PFT j :

$$\frac{\delta \text{NPP}}{\delta t} = \sum_j \left(\frac{\delta \text{Biomass}}{\delta t} \Delta \text{Growth} \right)_j + \sum_j \left(\frac{\delta \text{Growth}}{\delta t} \Delta \text{Biomass} \right)_j + \text{Residual}$$

We then determine the median across all models (Fig. 4). We find that the multi-model median growth rates increase nearly everywhere, while the median biomass decreases in the low latitudes, but increases in the Southern Ocean, mimicking the changes in NPP. As was the case for NPP, the model spread is large for both factors driving NPP, and particularly so in the low-latitudes (not shown).

We focus next on the drivers affecting the growth rates, i.e., the bottom up factors temperature, light, and nutrients and afterwards discuss the factors affecting phytoplankton biomass, i.e. the top-down control, and do so from a global perspective. We then extend the analysis to the level of individual phytoplankton PFTs, which is best done at the regional scale, across which the responses are relatively homogeneous in contrast to the global scale.

4.3 Global analysis of bottom up factors

Fig. 5 shows the projected changes in sea surface temperature, photosynthetically active radiation (PAR) and surface concentrations of NO_3 and Fe as a zonal average for all models. Fig. 6 shows the resulting relative changes in the growth rates and the limitation factors for temperature, light and nutrients for all models where the equations describing the limitation factors were available. Note that an increase in any limitation factor corresponds to an alleviation of this limitation i.e., a positive impact on the growth rate. To simplify the plot, for each model only the values for the phytoplankton PFT with the strongest temperature (or light or nutrient limitation factor or growth rate) response is shown, and minor phytoplankton PFTs like diazotrophs are not included.

In the low latitudes, sea surface temperature is projected to warm by about 2-3°C with some model variance (Fig. 5a). In the Southern Ocean, the warming is less pronounced and even more consistent among models ($+1 \pm 1^\circ\text{C}$), while in the Arctic Ocean, the warming is not only stronger but also differs strongly among the models (projections range between no change and $+4^\circ\text{C}$). This surface ocean warming stimulates phytoplankton growth everywhere and in all models, although given the different temperature sensitivities and the different levels of warming, the spread is large (Fig. 6a). In the low latitudes, the temperature limitation factor is simulated to increase by +10% and +30% (corresponding to weaker limitation). In the Southern Ocean the increase remains small (0-10%) reflecting the small temperature changes, while in the Northern high latitudes the temperature limitation factor increases by up to 40%.

In contrast to the large changes in temperature, the photosynthetically active radiation (PAR) at the surface changes little globally, with the important exception of the high latitudes (Fig. 5b), where light availability is affected by changes in sea-ice. In the Arctic, PAR is modeled to increase (projections range between 2 and 18 W m^{-2} increase), while in the Southern Ocean, models disagree even

on the direction of change, reflecting the divergent trends in sea-ice (Mahlstein et al., 2013). Consequently, most models show little changes and also little spread in the surface light limitation term between 60°N-60°S (Fig. 6b). In the high latitudes the spread is generally larger, with projections in light limitation factor ranging between -10% and +40%. However, in all but one model, relative
345 changes in light limitation are of similar magnitude as the relative changes in temperature limitation in the high latitudes.

The iron concentrations are projected to change in a latitudinally relatively uniform manner with changes between -0.05 and +0.2 $\mu\text{mol Fe m}^{-3}$, with one exception (diat-HadOCC), where a strong increase is simulated (+0.5 $\mu\text{mol Fe m}^{-3}$) in the Arctic. These generally small and uniform changes
350 are reflecting the constant dust deposition in all models. Regionally, models differ most in the change in iron concentration in the equatorial Pacific (not shown), potentially related to differences in the transport of iron rich water by the Equatorial Undercurrent (Vichi et al., 2011a; Ruggio et al., 2013). There is little agreement among the models with regard to the direction of changes in the surface concentration of nitrate, with decreases and increases of up to $\pm 3 \text{ mmol N m}^{-3}$. Similar changes
355 are modeled for phosphate (not shown). The large range of projected trends leads to very wide ranges for the relative changes in the nutrient limitation factor. In fact, with increases and decreases of up to $\pm 90\%$ in the low latitudes, $\pm 15\%$ in the Southern Ocean and 0 and -40% in the region north of 30°N, the nutrient limitation factor is changing the most.

In nearly all models, the magnitude of the nutrient limitation term is determined solely by the most
360 limiting nutrient (Liebig limitation, see Section 2). Except for PlankTOM5.3, the limitation patterns for different PFTs within the same model are rather similar, but the differences between models are large. Therefore, we show in Fig. 7 the limitation pattern only for diatoms. In the Southern Ocean, most models agree on iron limiting phytoplankton growth in the annual mean, while PlankTOM5.3 only simulates iron limitation in parts of the Southern Ocean and near the Antarctic continent in
365 summer. In the low latitudes, models show substantial differences in the equatorial upwelling region in the Pacific. Only some models capture the iron limitation shown in data (Moore et al., 2013a). There is substantial variation in the extent of the iron limited region and also the direction of change in iron concentration. As this is a region with high NPP values in the annual mean (see Table 16), uncertainties in this region significantly affect the range in NPP projections. In the remaining low
370 latitudes, models show either phosphate or nitrate limitation.

As half of the models use specified N:P Redfield ratios instead of modeling an explicit PO_4 tracer, nitrate and phosphate limitation cannot be distinguished in these models. However, as nitrate and phosphate are usually highly correlated, a differentiation between nitrate and phosphate limitation might not significantly increase the uncertainty in nutrient limitation projections. Most models agree
375 on stronger nutrient limitation (a decrease in the nutrient limitation factor of between -0.01 and -0.05) in the low latitudes excluding the equatorial upwelling region. The exceptions are REcoM2 and PlankTOM5.3, which simulate weaker nutrient limitation.

In summary, the changes in nutrients and temperature emerge as the most important determinants for the changes in the growth rates, with light generally playing a lesser role, except for the very high latitudes, particularly the Arctic. The changes in the bottom-up factors determine the changes in phytoplankton growth rate, which are shown in Fig. 6d, again the PFT with the strongest changes is shown. In two models (CNRM-PISCES and IPSL-PISCES), the growth rate is decreasing in all latitudes except for the Southern Ocean, resulting from the high nutrient stress in these two models. However, all other models predominantly simulate increases in growth rates, owing to the temperature effect outweighing the decrease in nutrient availability in these models. The decreases in low latitude NPP in the six models that show increases in growth rates is thus not bottom-up driven, but caused by a loss of biomass. Future changes in phytoplankton biomass can be caused by several top-down factors which we will discuss in the following.

4.4 Analysis of top-down control

Possible reasons for the simulated phytoplankton biomass decrease between 2000 and 2100 are 1) changes in circulation or mixing leading to a stronger lateral/vertical loss of biomass, 2) increased aggregation or mortality of phytoplankton if explicitly modeled or 3) a higher grazing pressure. Unfortunately, none of these fluxes have been stored by most models. Further, recalculated values are not precise enough to analyze the difference between NPP and loss processes. Therefore, we cannot quantitatively differentiate our analysis into the changes in grazing loss, aggregation and physical biomass loss across all models. We nevertheless try to shine some light into this critically important issue by using qualitative arguments and the partial information we have from those models that were able to provide the phytoplankton grazing loss.

We hypothesize that the loss of biomass caused by physical transport does not significantly increase, as all models show an increase in stratification over the next century. Furthermore, phytoplankton aggregation (and mortality) depend exponentially (linearly) on biomass but are temperature independent, so neither aggregation nor mortality losses can increase at lower biomass levels, eliminating this set of processes as well. This leaves us with increased grazing pressure as the most likely driver of the simulated biomass loss in the low and intermediate latitudes and the high northern latitudes. This hypothesis is supported by the fact that in all five models, for which the grazing fluxes were available, the fraction of grazed NPP increases throughout the 21st century north of 50°S (Fig. 8), i.e., the grazing pressure increases. In TOPAZ the increase is comparatively small (+0.1%). However, grazing is the only loss process in this model and changes in the ratio between grazing and NPP ratio have a direct and strong impact on phytoplankton biomass. In the models where aggregation and mortality are explicitly modeled, the increase in the grazed fraction of NPP is stronger (+5±3%). This larger change in the grazed fraction in these models can be understood when considering that as biomass decreases, the aggregation losses decrease as well. This automatically leads to a shift of the loss pathways toward grazing, even though the grazing pressure per se

does not change. The fact, however, that biomass is decreasing in the first place, strongly indicates
 415 that increases in grazing pressure are the driver for the phytoplankton biomass losses diagnosed in
 the models.

To better understand the potential drivers for this increase in the grazing pressure, we analyze the
 fraction of NPP that is grazed by zooplankton, given by:

$$\frac{\text{grazing}}{\text{NPP}} = \frac{g_Z^P \times T_f^Z \times \text{P-dependence} \times Z}{\mu_{\max} \times N_{\text{lim}} \times L_{\text{lim}} \times T_f^P \times P} \quad (6)$$

Here, g_Z^P is the grazing rate, T_f^Z and T_f^P are the temperature limitation for phytoplankton and zoo-
 plankton, respectively, P and Z denote phytoplankton and zooplankton biomass and μ_{\max} the maxi-
 mum phytoplankton growth rate, as introduced in Eq. 2 and 5. P-dependence is the dependence on
 420 prey concentration as shown in Table 13.

Climate change affects the ratio between grazing and NPP via temperature and also via changes
 in nutrient and light limitation. Furthermore, the grazing:NPP ratio is affected by changes in zoo-
 plankton biomass, i.e., increases in total grazing and zooplankton mortality indirectly play a role.
 In the models where the same temperature function for both phytoplankton growth and zooplankton
 425 grazing is used (i.e. $T_f^P = T_f^Z$, see Table 11), the temperature limitation cancels out. Still, with a
 higher temperature the total grazing intensifies due to an increase in zooplankton growth rate and
 thus a larger zooplankton biomass, which will intensify grazing (see Equation 6). On the other hand,
 the grazing:NPP ratio can increase through a decrease in the phytoplankton growth rate μ , because
 of stronger light or nutrient limitation, thus decreasing NPP in the equation above.

430 Lacking the model output from the three dimensional models, we use a one-box model to explore
 the sensitivity of the grazing:NPP ratio to changes in temperature and nutrient limitation instead. To
 this end we consider only one phytoplankton and one zooplankton group, using a simplified form of
 the equations and parameterizations of the BEC model. We did not include further phytoplankton
 loss terms like aggregation or mortality and used a quadratic temperature-independent mortality as
 435 loss process for zooplankton. We performed a spin-up until the model reached an equilibrium state
 under conditions representative for the low latitudes (temperature limitation of 0.8 corresponding
 to about 27°C, strong nutrient limitation of 0.1 corresponding to less than 0.5 mmol NO₃ m⁻³ and
 weak light limitation). As grazing is the only loss process of phytoplankton, 100% of NPP are
 grazed in the equilibrium state. To test the sensitivity of grazing pressure to temperature changes,
 440 we increased the temperature from 27 to 30°C over a time period of 10 years but kept light and
 nutrient limitation constant. The experiment showed an 8% decrease in phytoplankton biomass
 within the 10 simulation years even though the phytoplankton growth rate was increasing, caused by
 a temperature-driven increase in zooplankton biomass and thus grazing. On average, about 101% of
 NPP was grazed per month during the 10 year period.

445 To test the sensitivity of grazing pressure to nutrient changes, we enhanced nutrient limitation by
 30% (nutrient limitation factor decreases from 0.1 to 0.07) over 10 years while keeping tempera-

ture constant at 27°C. In this experiment, phytoplankton biomass decreased by 15%. Besides the decrease in phytoplankton growth in this experiment compared to the equilibrium state and the first experiment, 102.5% of NPP was grazed on average each month, indicating that the change in nutrient limitation has a similar effect on grazing like the temperature increase. These results indicate that the grazing pressure can be increased by both stronger nutrient limitation and higher temperatures. As the basic structure of the NPP and grazing equations is similar in most models, this mechanism might explain the observed biomass loss in the low and northern high latitudes. However, the specific grazing parameterizations and also the zooplankton mortality parameterizations differ substantially between models, such that the strength of the grazing response and the magnitude of the biomass loss is most likely different between models.

4.5 Regional changes in phytoplankton community structure

In the following we will refine our analysis to include differences in PFT responses and focus on two example regions: the low latitudes (30°S - 30°N) and the Southern Ocean (50°S - 90°S). The low latitudes have been chosen because they explain a large part of the global NPP change (Table 16). Moreover, they exhibit the largest interquartile range (Fig. 3c) and are therefore the main reason for the large range in global NPP projections. The Southern Ocean has been chosen to demonstrate the mechanisms underlying NPP changes for a region where NPP increases in the multi-model median. This is a also region where diatoms form a significant fraction of biomass and drive NPP changes in several models. The drivers of the NPP changes in the North Atlantic and North Pacific will be described briefly at the end of this Section.

4.5.1 Low latitude phytoplankton community changes

All models analyzed in this study except one agree that NPP decreases between the 2012-2031 and the 2081-2100 in the low latitudes (Fig. 9), albeit with different magnitudes (between -0.004 and -0.09 molC m⁻³yr⁻¹). The exception is PlankTOM5.3 that shows a strong increase of 0.1 molC m⁻³yr⁻¹, on average. It will be discussed separately below. In three models the trend is caused by similar decreases in both diatom and nanophytoplankton NPP (BEC, TOPAZ and diat-HadOCC). Diatom changes contribute about a third of total NPP changes in both PISCES simulations and the decrease is mainly driven by a decrease of the NPP by nano- or picophytoplankton in PELAGOS and MEM, with little changes in diatom NPP. In REcoM2, diatoms and nanophytoplankton trends almost fully compensate each other. Changes in diazotrophs (modeled in BEC and TOPAZ) and large non-diatom phytoplankton contribute less than 10% to the total trend.

Fig. 10 shows the relative change in temperature, light and nutrient limitation, growth rate, biomass and NPP for diatoms, nano- or picophytoplankton and coccolithophores in the low latitudes. Diat-HadOCC could not be included in the figure as the equations for the limitation factors are not available. In three models (BEC, MEM, REcoM2) diatoms show a stronger response to

nutrient limitation than nanophytoplankton, which translates to a smaller increase or even decrease in growth. However, in all models except MEM diatoms show larger relative biomass and NPP losses than nanophytoplankton, indicating that in TOPAZ, PELAGOS and the PISCES simulations
485 top-down control is the main reason for the decrease in diatom relative contribution to biomass.

The PlankTOM5.3 trend is caused by an increase in coccolithophore NPP ($+0.14 \text{ molC m}^{-3}\text{yr}^{-1}$), partly compensated by a decrease in nanophytoplankton NPP ($-0.04 \text{ molC m}^{-3}\text{yr}^{-1}$). We note that export production changes do not follow the increase in NPP but decreases strongly (not shown), indicating a very large increase in the recycling efficiency in this model. This is caused by a strong
490 increase in microzooplankton biomass and their grazing on phytoplankton, with rapid recycling of the nutrients back to their inorganic forms explaining the increase in nutrient availability. This greatly enhances regenerated production, even as new production decreases.

4.5.2 Southern Ocean phytoplankton community changes

All models simulate an increase in surface NPP in the Southern Ocean south of 50°S , but the
495 magnitude of the change varies by several orders of magnitude ($+0.006$ and $+0.11 \text{ molC m}^{-3}\text{yr}^{-1}$) (Fig. 11). Furthermore, the contributions of the different phytoplankton PFTs to these NPP trends differ strongly between the different models. Four models show a stronger increase in the NPP by nanophytoplankton compared to that by diatoms (PlankTOM5.3, MEM, TOPAZ, CNRM/PISCES), three models show an exclusively diatom-driven NPP change (BEC, PELAGOS, REcoM2) and
500 two models show similar changes in the NPP by diatoms and nanophytoplankton (diat-HadOCC, IPSL/PISCES). Only one model shows a significant decrease in diatom NPP (PlankTOM5.3).

In seven out of eight models, surface ocean warming is the most important driver for the increase in phytoplankton growth for both diatoms and nanophytoplankton. All but the CNRM/PISCES and PELAGOS model show a relief from nutrient stress for all phytoplankton types, i.e., an increase in
505 the nutrient limitation factor (1-15% increase), although these models remain iron limited throughout the 21st century. Diatoms respond more strongly to changes in nutrient concentrations than nanophytoplankton in all models except for PlankTOM5.3. In addition, in many models a stronger top-down control of nanophytoplankton than diatoms becomes apparent, indicated by differences in biomass changes despite similar growth rate changes. Only in MEM and PlankTOM5.3 diatoms
510 seem to be stronger top-down controlled. In PELAGOS the diatom fraction is almost 100% south of 50°S , and shows little changes. The final result is a stronger increase in diatom NPP compared to nanophytoplankton NPP in BEC, TOPAZ, IPSL, CNRM and REcoM2, and a weaker increase or even decrease in diatom NPP in MEM and PlankTOM5.3.

5 Discussion

515 5.1 NPP changes and their drivers

Our finding of the key role of temperature in defining the response of NPP to future climate change contrasts with the conclusion of the majority of the past studies, which attributed the decrease in NPP to a decrease in nutrient availability, particularly in the low latitudes (Bopp et al., 2001; Moore et al., 2002; Steinacher et al., 2010; Marinov et al., 2010; Cabré et al., 2014). To explain this discrepancy, we focus on the temperature functions in the models used in the studies above. First, several of the earlier models had either no temperature dependence of phytoplankton growth at all (the model HAMOCC5.1), or the temperature sensitivity was rather weak with a Q_{10} value of 1.13 for temperatures higher than 15°C (in the models HAMOCC3 and NCAR CSM1.4-carbon). It is thus not surprising that ocean warming did not significantly affect global productivity in these model simulations compared to the models analyzed in this study that have a Q_{10} of at least 1.68. A further model analyzed in several of these studies is the IPSL model with PISCES as the ecological/biogeochemical component. A later version of this model is analyzed in our study. Consistent with previous authors we find that changes in nutrient limitation are the main driver of NPP changes in PISCES, mostly because decreases in nutrient limitation are significantly stronger compared to other models. Finally, several authors attribute projected decreases in NPP to concurrent decreases in macronutrient availability (Steinacher et al., 2010; Marinov et al., 2013; Cabré et al., 2014). But our analysis shows that in many models the global NPP decrease, and particularly that in the low latitudes, is not caused by decreasing growth rates, such as one would expect from increasing nutrient limitation. Rather the decrease in NPP is caused by biomass losses, presumably a result of a warming-induced increase in grazing pressure. We conclude that the temperature effect and top-down control might have been underestimated in several earlier studies.

Our identified importance of warming for future NPP is more in line with another group of studies, where global NPP was projected to increase with climate change, and a temperature-driven increase in metabolic rates was identified as the cause (Schmittner and Galbraith, 2008; Sarmiento et al., 2004; Taucher and Oschlies, 2011). This agreement might be somewhat fortuitous however, as Schmittner and Galbraith (2008) and Taucher and Oschlies (2011) considered only the temperature dependence of phytoplankton growth and remineralization, while the growth of zooplankton and hence the grazing pressure on phytoplankton were independent of temperature. Likewise, the algorithm used to estimate chlorophyll in Sarmiento et al. (2004) is based on the assumption that chlorophyll is purely bottom-up controlled.

Finally, Dutkiewicz et al. (2013) aimed to separate the direct temperature effect from the altered nutrient input and light availability caused by stratification. In their study, temperature, nutrient and light changes compensate each other nearly perfectly, resulting in very little change in global NPP. Still, the importance of temperature for phytoplankton growth and zooplankton grazing shown by

550 most models in our study indicates that temperature might play a major role in the response of NPP
to climate change.

While we emphasize here the role of temperature in the models, our understanding of how temper-
ature controls the most important ecological and biogeochemical processes in real marine ecosys-
tems is not well established. Most models base their parameterizations of temperature effects on
555 laboratory studies that show - within favourable thermal ranges - an exponential increase of growth
with increasing temperature (Eppley, 1972; Bissinger et al., 2008). However, there are major un-
certainties in quantifying the temperature sensitivities of different physiological processes and of
functional types (Ikeda et al., 2001; Lomas et al., 2002; Hirst and Bunker, 2003; Hancke and Glud,
2004; Sand-Jensen et al., 2007). Several authors suggest a stronger temperature response of het-
560 erotrophs than autotrophs (López-Urrutia et al., 2006; Rose and Caron, 2007), which would lead to
major consequences for the metabolic balance of the oceans under rising temperatures (Duarte et al.,
2013; Williams et al., 2013; Ducklow and Doney, 2013; García-Corral et al., 2014). Furthermore,
in current implementations both phyto- and zooplankton grow faster with increasing temperatures
without any upper thermal limit beyond which growth rates may come down. The underlying as-
565 sumption is that if the temperature rises to values outside the optimal range of a certain species, the
species will be replaced by another species with a higher temperature tolerance. However, particu-
larly in the tropics, it is unclear if this assumption holds. Thomas et al. (2012) show that warming
might lead to a decrease in diversity in the tropics, which could potentially lower NPP due to the loss
of highly productive species. Finally, due to the lack of measurements, synergistic effects of multiple
570 stressors are barely considered in current models. Recently, temperature sensitivity has been shown
to be reduced under nutrient limitation (Staehr and Sand-Jensen, 2006; Tadonlécé, 2010; Marañón
et al., 2014) which would result in an overestimation of temperature sensitivity and therefore NPP in
the oligotrophic regions of the ocean. Overall, the temperature assumptions on which current model
projections are based are afflicted with high uncertainties.

575 **5.2 Changes in phytoplankton community**

Seven out of nine models in our study show a global decrease in the relative abundance of diatoms
with decreases in low latitudes but increases in the Southern Ocean, confirming results reported
by Bopp et al. (2005); Marinov et al. (2010); Dutkiewicz et al. (2013); Manizza et al. (2010) and
Marinov et al. (2013). The difference between the diatom and nanophytoplankton nutrient response
580 has been identified as the primary driver of the decrease in diatom fraction in Bopp et al. (2005);
Marinov et al. (2010) and Marinov et al. (2013). Our results show that while models currently agree
on a global decrease in diatom fraction, there is no agreement on regional changes and models do not
agree whether a stronger nutrient response or a higher susceptibility to grazing pressure is the cause.
As diatom biomass tends to be overestimated by several of the models (Vogt et al., 2013; Hashioka
585 et al., 2013), the relative importance of changes in diatom biomass may constitute an upper bound

for future global NPP changes.

6 Identifying and Reducing Uncertainties

The spread in globally integrated NPP projections in our study is 45%, with the PlankTOM5.3 model causing 25% of it alone. Given this wide spread in NPP projections, we attempt to identify
590 the different sources of uncertainty in the following and then investigate whether there is a way to narrow the uncertainty of the projections using emergent constraints.

6.1 Sources of Model Uncertainties

The biogeochemical and biological parameterizations that contribute the largest uncertainties are:

- 595 – Initial nutrient concentrations: Models (except PlankTOM5.3) agree on similar decreases in nutrient concentration in the low latitudes, and also on the total change in the nutrient limitation factor between ± 0.1 , despite disparities with regard to the identification of the most limiting nutrient. However, the differences in relative nutrient limitation change are very large ($\pm 90\%$, see Fig. 6). Particularly the PISCES simulations show a strong relative decrease in nutrient limitation, which is caused by it having (too) low nutrient concentrations at the
600 beginning of the simulation (see Section 3). On the other hand, a positive bias in nutrients as observed in other models might lead to a too weak response in nutrient limitation.
- 605 – The relative importance of iron versus nitrate limitation and projections for iron concentrations: Increases in iron availability allow the small global increase in nanophytoplankton NPP in REcoM2 and attenuate or even outbalance the low latitude NPP decrease in BEC and TOPAZ. This is of particular relevance in the equatorial upwelling region in the Pacific (see Fig. 3), which is iron limited according to observations (Moore et al., 2013a) and is responsible for 14-33% of global NPP at present in the different models (Table 16). The differences in the projected changes in iron concentration in the equatorial upwelling region in the Pacific are potentially related to differences in circulation: According to Vichi et al. (2011b) and Ruggio
610 et al. (2013), the Equatorial Undercurrent may intensify and shoal with climate change and this may bring more iron to the eastern equatorial upwelling, partly off-setting the reduced nutrient input due to the warming surface. Note that the dust deposition is held constant in current projections. Variable iron forcing in future simulations will lead to more realistic NPP projections but might further increase this uncertainty.
- 615 – Different Q_{10} values (between 1.68 and 2.08) and different projections for SST increase (+2,+3°C) together result in a high uncertainty of the temperature response of both phytoplankton growth and zooplankton grazing. Further uncertainty is introduced by the stronger temperature response of zooplankton types parameterized in some models.

- 620 – The relative importance of the response of top-down controls versus that of the microbial loop, potentially related to different Q_{10} values and differences in the partitioning of the grazed material.
- 625 – There is no agreement with regard to the direction of change in light limitation in the Southern Ocean, reflecting the wide range in projected sea-ice changes and other factors influencing surface light such as cloud cover. However, light limitation introduces currently only a minor uncertainty compared to the nutrient and temperature effects, at least for surface NPP.

In order to reduce the spread in NPP projections, we also need to understand how much of the uncertainty arises from the underlying physical forcing (transport, mixing, and temperature) and how much is caused by the different ecosystem parameterizations. Unfortunately, the design of our study does not allow for a clear distinction between uncertainty from physical forcing and from the use of different ecosystems, as this would require to compare projections from different Earth System Models using the same ecosystem model with projections of one Earth System Model coupled to different ecosystem models. Results from Sinha et al. (2010), where they compared two different circulation models coupled to the same biogeochemical model indicate that differences in the underlying physics lead to substantial differences in PFT biogeography, but only small effects on total NPP. A more ambitious program is currently undertaken, where a larger group of ecosystem models are being coupled to the same circulation model (the iMarNet project, Kwiatkowski et al., 2014). The outcome of this project will help to better separate the ecosystem model uncertainty from the uncertainty introduced by different physical models.

6.2 Constraining NPP projections

640 The concept of emergent constraints (e.g. Allen and Ingram, 2002) has been used with success to reduce uncertainties for future projections. The basic premise is that models that provide a better fit to a specific set of current constraints provide a better estimate for the future changes. The emergent constraint is usually established by finding a good correlation between an observable parameter for the present and the future change in NPP. We have tested for correlations between the different models' skill to predict current NPP and their projected changes, using both the 2012-2031 average of globally integrated NPP and the slope between chlorophyll a and sea surface temperature as a measure for model skill. Although chlorophyll a is a poor indicator for biomass in the low latitudes (Siegel et al., 2013), it can be used as indicator for model skill and is comparatively well constrained by observations. As the metric for the projected changes we used the change in NPP defined as the difference between the 2012-2031 average and the 2081-2100 average and the change in NPP weighted with the temperature increase. Moreover, as regions with positive and negative changes might cancel each other out, leading to little net NPP changes despite strong local changes, we also tested for a relation between the mean of absolute NPP changes and model skill.

We did not find any significant correlation between model skill and NPP changes, neither on regional nor global scales, and the relation is weak at best between globally integrated NPP and the absolute change in NPP (Fig. 13). We hypothesize that the cause for this lack of relationship is the uncertainty in the relative importance of the net effect of temperature on NPP and on nutrient limitation. This hypothesis is supported by results from Taucher and Oschlies (2011) who compared two simulations, one temperature dependent and one independent. Both simulations fitted equally well to observations, but the direction of NPP change was opposite. It seems that matching the current observations is not sufficient to estimate which sign of future NPP change is more realistic. Thus we need a better understanding of the mechanisms in order to reduce the uncertainty in projections. Efforts to extend the amount of data that is available for model parameterisation and evaluation (Buitenhuis et al., 2013b) will hopefully help achieve that goal.

7 Conclusions

In this work we present a multi-model comparison of nine model simulations with regard to NPP and its underlying drivers. We show projected changes in global NPP between -15% and +30% by the end of this century for the high emission scenario RCP8.5, with the largest inter-model discrepancies stemming from the low latitudes. All but one model simulate either a decrease in NPP or changes less than 0.5% in the low latitudes, but for very different reasons. The main drivers are warming-induced enhancement of phytoplankton growth, increased nutrient limitation and decreases in phytoplankton biomass, which are most likely caused by temperature-enhanced grazing by zooplankton. Only three models show reduced phytoplankton growth rates due to increased nutrient limitation. Thus in this set of models, temperature and nutrient concentrations are at least equally important drivers for changes in NPP, contradicting many prior studies that emphasized the sole importance of a stronger nutrient limitation.

One major difficulty faced in this study is the limited availability of model output variables related to ecosystem growth and loss rates, particularly limitation factors and grazing rates. The changes in growth rate, temperature limitation, light and nutrient limitation reported in this work have been recalculated in six out of nine models using surface monthly mean fields. The obtained results are therefore an approximation of the original values. We have compared recalculated values with original values in the models where the limitation factors were given, and we estimate the error to be less than 10%. We conclude that while the absolute values reported might be inaccurate, the relative importance of nutrient vs. temperature limitation shown in this work is correct. Furthermore, we can discuss only surface NPP changes. For the models where 3D limitation factors were available (BEC, REcoM2), we compared our results for the surface with the 100m average, and we can confirm that the same mechanisms that govern the surface changes also hold for the 100m average. In addition, the changes in surface NPP correlate with the changes in integrated NPP in all models, except for

Symbol	Meaning
T_f	Temperature limitation factor
N_{lim}	Nutrient limitation factor
L_{lim}	Light limitation factor
μ_i	Growth rate of phytoplankton i
T	Temperature in °C
$\theta_{Chl/C}$	Chlorophyll to Carbon ratio
I_{PAR}	Photosynthetically active radiation
Fe	Iron concentration
PO_4	Phosphate concentration
NH_4	Ammonium concentration
NO_3	Nitrate concentration
$SiOH_4$	Silicate concentration

Table 1: Symbols used in the model equations

the Arctic Ocean. It therefore seems likely that our surface drivers also describe the changes in
690 integrated NPP. To ease future studies of NPP changes, we recommend the inclusion of mixed layer
averages of growth rate, light and nutrient limitation and grazing fluxes in the standard model output
of future model inter-comparison projects. The availability of changes in growth rates could prevent
common misinterpretations of drivers by analysing univariate correlations with only one of several
possible drivers.

695 To reduce the ecosystem model uncertainty afflicted with NPP projections, the representation of
present day nutrient concentrations and resulting limitation patterns should be further improved.
Particularly a bias in present-day nutrient concentration strongly affects relative changes in nutrient
limitation and therefore NPP projections. Furthermore, given the importance of top-down control
shown in this work, we need a better understanding of zooplankton mortality and further potential
700 drivers of zooplankton biomass like phenological or trophic mismatches, diseases or changes in pre-
dation from higher trophic levels. Finally, a better understanding of the temperature dependency of
all key ecological/biogeochemical processes is needed. In particular, this includes the determination
of the different temperature response functions for the different PFTs and trophic levels.

Appendix A Model equations and Parameters

705 A1 BEC

Growth rate of phytoplankton PFT i :

$$\mu^i = \mu_{max}^i \times T_f \times N_{lim}^i \times L_{lim}^i$$

Parameter	Value	Unit	Description
Q_{10}	2		temperature dependence factor
T_{ref}	30	$^{\circ}\text{C}$	reference temperature
α^{diat}	0.3	$\text{mmol C m}^2 (\text{mg Chl W d})^{-1}$	initial slope of P-I curve
α^{nano}	0.3	$\text{mmol C m}^2 (\text{mg Chl W d})^{-1}$	initial slope of P-I curve
μ_{max}^{diat}	3.0	d^{-1}	max. phytoplankton growth rate at T_{ref}
μ_{max}^{nano}	3.0	d^{-1}	max. phytoplankton growth rate at T_{ref}
$K_{\text{NH}_4}^{diat}$	0.08	mmol N m^{-3}	NH_4 half saturation coefficient
$K_{\text{NH}_4}^{nano}$	0.005	mmol N m^{-3}	NH_4 half saturation coefficient
$K_{\text{NO}_3}^{diat}$	2.5	mmol N m^{-3}	NO_3 half saturation coefficient
$K_{\text{NO}_3}^{nano}$	0.5	mmol N m^{-3}	NO_3 half saturation coefficient
$K_{\text{PO}_4}^{diat}$	0.005	$\text{mmol PO}_4 \text{ m}^{-3}$	PO_4 half saturation coefficient
$K_{\text{PO}_4}^{nano}$	3.125 e^{-4}	$\text{mmol PO}_4 \text{ m}^{-3}$	PO_4 half saturation coefficient
K_{Fe}^{diat}	1.5 e^{-4}	mmol Fe m^{-3}	Fe half saturation coefficient
K_{Fe}^{nano}	6 e^{-5}	mmol Fe m^{-3}	Fe half saturation coefficient
$K_{\text{SiOH}_4}^{diat}$	1.0	$\text{mmol SiOH}_4 \text{ m}^{-3}$	SiOH_4 half saturation coefficient

Table 2: BEC parameters

Temperature function (for all PFTs):

$$T_f = Q_{10}^{\frac{T - T_{ref}}{10}}$$

710 **Total nutrient limitation:**

$$N_{lim}^{nano} = \min(N_{Fe}^{nano}, N_{\text{PO}_4}^{nano}, N_{\text{NO}_3 + \text{NH}_4}^{nano})$$

$$N_{lim}^{diat} = \min(N_{Fe}^{diat}, N_{\text{PO}_4}^{diat}, N_{\text{NO}_3 + \text{NH}_4}^{diat}, N_{\text{SiOH}_4}^{diat})$$

Iron limitation of PFT i:

$$N_{Fe}^i = \frac{\text{Fe}}{\text{Fe} + K_{Fe}^i}$$

715 **Phosphate limitation of PFT i:**

$$N_{\text{PO}_4}^i = \frac{\text{PO}_4}{\text{PO}_4 + K_{\text{PO}_4}^i}$$

Silicate limitation of diatoms:

$$N_{\text{SO}_3}^{diat} = \frac{\text{SO}_3}{\text{SO}_3 + K_{\text{SO}_3}^{diat}}$$

Nitrate and Ammonium limitation of PFT i:

$$720 \quad N_{\text{NO}_3 + \text{NH}_4}^i = \frac{\text{NO}_3}{K_{\text{NO}_3}^i \times (1 + \frac{\text{NO}_3}{K_{\text{NO}_3}^i} + \frac{\text{NH}_4}{K_{\text{NH}_4}^i})} + \frac{\text{NH}_4}{K_{\text{NH}_4}^i \times (1 + \frac{\text{NO}_3}{K_{\text{NO}_3}^i} + \frac{\text{NH}_4}{K_{\text{NH}_4}^i})}$$

Light limitation of PFT i:

$$L_{lim}^i = 1 - e^{-\frac{\alpha^i \times \theta^i \times I_{PAR}}{\mu_{max}^i \times T_f \times N_{lim}^i}}$$

Grazing:

$$G_{nano} = u_{max}^{nano} \times T_f \times \frac{P_{nano}^2}{P_{nano}^2 + g^2} \times Z$$

$$725 \quad G_{diat} = u_{max}^{diat} \times T_f \times \frac{P_{diat}^2}{P_{diat}^2 + g^2 \times f_z^{diat}} \times Z$$

A2 TOPAZ

Growth rate of phytoplankton PFT i:

$$\mu^i = \frac{\mu_{\max}^i}{1.0 + \zeta} \times T_f \times N_{\lim}^i \times L_{\lim}^i$$

Temperature function (for all PFTs):

$$730 \quad T_f = e^{k_{\text{Eppley}} \times T}$$

Total nutrient limitation:

$$N_{\lim}^{\text{nano}} = \min(N_{\text{Fe}}^{\text{nano}}, N_{\text{PO}_4}^{\text{nano}}, N_{\text{NO}_3 + \text{NH}_4}^{\text{nano}})$$

$$N_{\lim}^{\text{large}} = \min(N_{\text{Fe}}^{\text{large}}, N_{\text{PO}_4}^{\text{large}}, N_{\text{NO}_3 + \text{NH}_4}^{\text{large}})$$

diatoms of large phytoplankton depends on the silicate concentration:

$$735 \quad P_{\text{Diatoms}} = P_{\text{Large}} \times N_{\text{SO}_3}^{\text{diat}}$$

Iron limitation of PFT i:

$$N_{\text{Fe}}^i = \frac{(Q_{\text{Fe/N}}^i)^2}{(K_{\text{Fe/N}}^i)^2 + (Q_{\text{Fe/N}}^i)^2}, \text{ with } Q_{\text{Fe/N}}^i = \min(Q_{(\text{Fe/N}, \max)}^i, \theta_{\text{Fe/N}}^i)$$

Phosphate limitation of PFT i:

$$N_{\text{PO}_4}^i = \frac{Q_{\text{P/N}}^i}{Q_{(\text{P/N}, \max)}^i}, \text{ with } Q_{\text{P/N}}^i = \min(Q_{(\text{P/N}, \max)}^i, \theta_{\text{P/N}}^i)$$

740 Nitrate and Ammonium limitation of PFT i:

$$N_{\text{NO}_3 + \text{NH}_4}^i = \frac{\text{NO}_3}{(K_{\text{NO}_3}^i + \text{NO}_3)} \times \frac{1 + \text{NH}_4}{K_{\text{NH}_4}^i}$$

Silicate limitation of diatoms:

$$N_{\text{SO}_3}^{\text{diat}} = \frac{\text{SO}_3}{\text{SO}_3 + K_{\text{SO}_3}^{\text{diat}}}$$

Light limitation of PFT i:

$$745 \quad L_{\lim}^i = 1 - e^{-\frac{\alpha^i \times Q_{\text{Chl/C}}^i \times I_{\text{PAR}}}{\mu_{\max}^i \times T_f \times N_{\lim}^i}}$$

$$\text{with } Q_{\text{Chl/C}}^i = \frac{Q_{\max}^i - Q_{\min}^i}{1.0 + (Q_{\max}^i - Q_{\min}^i) \times \alpha^i \times I_{\text{Mem}} \times \frac{0.5}{\mu_{\max} \times N_{\lim} \times T_f}} + Q_{\min}^i$$

$$\text{and } Q_{\min}^i = \max(0, Q_{\min}^{\text{no lim}} - Q_{\min}^{\text{lim}}) \times N_{\lim}^i \times Q_{\min}^{\text{lim}}$$

I_{Mem} is the memory of irradiance over the scale of 24 hours and was provided in the model output.

Grazing:

$$750 \quad G_{\text{nano}} = \min(k_{\text{graz}, \max}, u_{\max} \times T_f \times \frac{P_{\text{nano}}}{P^*}) \times \frac{P_{\text{nano}}^2}{P_{\text{nano}} + P_{\min}}$$

$$G_{\text{large}} = \min(k_{\text{graz}, \max}, u_{\max} \times T_f \times \{N_{\text{large}}^{\text{graz}}\}) \times P_{\text{large}}$$

$$\{N_{\text{large}}^{\text{graz}}\} = \left[\frac{P_{\text{large}} + P_{\text{diaz}}}{P^*} \right]^{\frac{1}{3}} \times \frac{P_{\text{large}} + P_{\text{diaz}}}{P_{\text{large}} + P_{\text{diaz}} + P_{\min}} \times (P_{\text{large}}^2 + P_{\text{diaz}}^2)^{\frac{1}{2}}$$

A3 PISCES

Growth rate of phytoplankton PFT i:

$$755 \quad \mu^i = \mu_{\max} \times T_f \times N_{\lim}^i \times L_{\lim}^i$$

Temperature function (for nanophytoplankton, diatoms and microzooplankton):

$$T_f = e^{k_{\text{Eppley}} \times T}$$

Temperature function (for mesozooplankton):

$$T_{f, \text{meso}} = e^{k_{\text{Eppley}, \text{meso}} \times T}$$

760 Total nutrient limitation:

Parameter	Value	Unit	Description
ζ	0.1		photorespiration loss
K_{Eppley}	0.063	$^{\circ}\text{C}^{-1}$	temperature dependence factor
α^{diat}	$\frac{2.4e^{-5} \times 2.77e^{18}}{6.022e^{17}}$	$\text{g C m}^2 (\text{g Chl W s})^{-1}$	initial slope of P-I curve
α^{nano}	$\frac{2.4e^{-5} \times 2.77e^{18}}{6.022e^{17}}$	$\text{g C m}^2 (\text{g Chl W s})^{-1}$	initial slope of P-I curve
$\mu_{\text{max}}^{\text{diat}}$	$1.5e^{-5}$	s^{-1}	max. phytoplankton growth rate at 0°C
$\mu_{\text{max}}^{\text{nano}}$	$1.5e^{-5}$	s^{-1}	max. phytoplankton growth rate at 0°C
$Q_{\text{min}}^{\text{nolim}}$	0.01	gChl (gC)^{-1}	min. Chl:C without nutrient limitation
$Q_{\text{min}}^{\text{lim}}$	0.001	gChl (gC)^{-1}	min. Chl:C with complete nutrient limitation
$Q_{\text{max}}^{\text{nano}}$	0.04	gChl (gC)^{-1}	max. Chl:C
$Q_{\text{max}}^{\text{large}}$	0.06	gChl (gC)^{-1}	max. Chl:C
$K_{\text{NH}_4}^{\text{large}}$	$2e^{-7}$	molN (kg)^{-1}	NH_4 half saturation coefficient
$K_{\text{NH}_4}^{\text{nano}}$	$6e^{-7}$	molN (kg)^{-1}	NH_4 half saturation coefficient
$K_{\text{NO}_3}^{\text{large}}$	$6e^{-6}$	molN (kg)^{-1}	NO_3 half saturation coefficient
$K_{\text{NO}_3}^{\text{nano}}$	$2e^{-6}$	molN (kg)^{-1}	NO_3 half saturation coefficient
$Q_{\text{Fe:N, max}}^{\text{large}}$	$666e^{-6} \times \frac{106}{16}$	$\text{mol Fe (mol N)}^{-1}$	maximum Fe:N limit
$Q_{\text{Fe:N, max}}^{\text{nano}}$	$46e^{-6} \times \frac{106}{16}$	$\text{mol Fe (mol N)}^{-1}$	maximum Fe:N limit
$Q_{\text{P:N, max}}^{\text{large}}$	0.1236	$\text{mol P (mol N)}^{-1}$	maximum P:N limit
$Q_{\text{P:N, max}}^{\text{nano}}$	0.1458	$\text{mol P (mol N)}^{-1}$	maximum P:N limit
u_{max}	0.19/86400	s^{-1}	grazing rate at 0°C
P^*	$1.9e^{-6} \times \frac{16}{106}$	molN kg^{-1}	pivot phyto concentration for grazing allometry
P_{min}	$1e^{-10}$	molN kg^{-1}	min. phyto concentration threshold for grazing

Table 3: TOPAZ parameters

$$N_{\text{lim}}^{\text{nano}} = \min(N_{\text{Fe}}^{\text{nano}}, N_{\text{PO}_4}^{\text{nano}}, N_{\text{NO}_3 + \text{NH}_4}^{\text{nano}})$$

$$N_{\text{lim}}^{\text{diat}} = \min(N_{\text{Fe}}^{\text{diat}}, N_{\text{PO}_4}^{\text{diat}}, N_{\text{NO}_3 + \text{NH}_4}^{\text{diat}}, N_{\text{SiOH}_4}^{\text{diat}})$$

Iron limitation of PFT i:

$$N_{\text{Fe}}^i = \frac{\text{Fe}}{\text{Fe} + K_{\text{Fe,variable}}^i}$$

765 where $K_{\text{Fe,variable}}^i = \max \left\{ \begin{array}{l} K_{\text{Fe,min}}^i \\ \frac{\text{Diat}^* \times K_{\text{Fe,min}}^i + \text{Nano}^* \times K_{\text{Fe}}^i}{P_i} \end{array} \right.$

and $\text{Diat}^* = \min \left\{ \begin{array}{l} P_{\text{diat}} \\ 5e^{-7} \end{array} \right.$ and $\text{Nano}^* = \min \left\{ \begin{array}{l} P_{\text{nano}} \\ 1e^{-6} \end{array} \right.$

Phosphate limitation of PFT i:

$$N_{\text{PO}_4}^i = \frac{\text{PO}_4}{\text{PO}_4 + K_{\text{PO}_4}^i}$$

Silicate limitation of diatoms:

770 $N_{\text{SO}_3}^{\text{diat}} = \frac{\text{SO}_3}{\text{SO}_3 + K_{\text{SO}_3}^{\text{diat}}}$

Nitrate and Ammonium limitation of PFT i:

$$N_{\text{NO}_3 + \text{NH}_4}^i = \frac{K_{\text{NH}_4}^i \times \text{NO}_3 + K_{\text{NO}_3}^i \times \text{NH}_4}{K_{\text{NH}_4}^i K_{\text{NO}_3}^i + K_{\text{NH}_4}^i \text{NO}_3 + K_{\text{NO}_3}^i \text{NH}_4}$$

Light limitation of PFT i:

$$L_{\text{lim}}^i = \left(1 - \frac{\{\text{MXL influence}\}}{\beta_i + \{\text{MXL influence}\}}\right) \times \left(1 - e^{-\frac{-\alpha \times \theta^{\frac{z}{H}} \times I_{\text{PAR}}}{\mu_{\text{max}}^i \times N_{\text{lim}}^i}}\right)$$

$$775 \quad \text{with } \{\text{MXL influence}\} = \begin{cases} \text{MXL} - \text{Heup} & \text{if } \text{MXL} > \text{Heup} \\ 0 & \text{otherwise} \end{cases}$$

where MXL denotes the mixed layer depth and Heup the depth of the euphotic zone

Microzooplankton grazing:

$$G_{\text{micro} \rightarrow \text{nano}} = u_{\text{max}}^{\text{micro} \rightarrow \text{nano}} \times T_f \times \frac{\Psi_{\text{nano}}^{\text{micro}} P_{\text{nano}}}{\sum_I \Psi_{\text{nano}}^{\text{micro}} \times I} \times \frac{P_{\text{nano}}}{K_G + \sum_I (\Psi_I^{\text{micro}} \times I)}$$

I denotes the food options and consists of diatoms and nanophytoplankton for microzooplankton.

780 Grazing on diatoms is calculated accordingly.

Mesozooplankton grazing:

$$G_{\text{meso} \rightarrow \text{nano}} = u_{\text{max}}^{\text{meso} \rightarrow \text{nano}} \times T_{f,\text{meso}} \times \frac{\Psi_{\text{nano}}^{\text{meso}} P_{\text{nano}}}{K_G + \sum_I \Psi_{\text{nano}}^{\text{meso}} \times I} \times Z_{\text{meso}}$$

The food options I for mesozooplankton are nanophytoplankton, diatoms and microzooplankton.

A4 MEM

785 **Growth rate of phytoplankton PFT i:**

$$\mu^i = \mu_{\text{max}}^i \times T_f \times N_{\text{lim}}^i \times L_{\text{lim}}^i$$

Temperature function (for all PFTs):

$$T_f = e^{k_{\text{Eppley}} \times T}$$

Total nutrient limitation:

$$790 \quad N_{\text{lim}}^{\text{nano}} = \min(N_{\text{Fe}}^{\text{nano}}, N_{\text{NO}_3 + \text{NH}_4}^{\text{nano}})$$

$$N_{\text{lim}}^{\text{diat}} = \min(N_{\text{Fe}}^{\text{diat}}, N_{\text{NO}_3 + \text{NH}_4}^{\text{diat}}, N_{\text{SiOH}_4}^{\text{diat}})$$

Iron limitation of PFT i:

$$N_{\text{Fe}}^i = \frac{\text{Fe}}{\frac{\text{Fe}}{1 - f_A^i} + \frac{\mu_{\text{max}}^i}{f_A^i A_{\text{Fe}}^i}}$$

with

$$795 \quad f_A^i = \max \left\{ \left(1 + \sqrt{\frac{\max(A_{\text{NO}_3}^i \times \text{NO}_3, A_{\text{NH}_4}^i \times \text{NH}_4)}{\mu_{\text{max}}^i}}\right)^{-1}, \left(1 + \sqrt{\frac{A_{\text{Fe}}^i \times \text{Fe}}{\mu_{\text{max}}^i}}\right)^{-1} \right\}$$

Silicate limitation of diatoms:

$$N_{\text{SO}_3}^{\text{diat}} = \frac{\text{SO}_3}{\frac{\text{SO}_3}{1 - f_A^{\text{diat}}} + \frac{\mu_{\text{max}}^{\text{diat}}}{f_A^{\text{diat}} \times A_{\text{Si}}^{\text{diat}}}}$$

Nitrate and Ammonium limitation of PFT i:

$$N_{\text{NO}_3 + \text{NH}_4}^i = \frac{\text{NO}_3}{\frac{\text{NO}_3}{1 - f_A^i} + \frac{\mu_{\text{max}}^i}{f_A^i \times A_{\text{NO}_3}^i}} \left(1 - \frac{\text{NH}_4}{\text{NH}_4 + K_{\text{NH}_4}^i}\right) + \frac{\text{NH}_4}{\frac{\text{NH}_4}{1 - f_A^i} + \frac{\mu_{\text{max}}^i}{f_A^i \times A_{\text{NH}_4}^i}}$$

800 **Light limitation of PFT i:**

$$L_{\text{lim}} = \frac{(1 - e^{-\frac{\alpha_i \times \text{PAR}}{p_r}}) \times e^{-\frac{\beta \times \text{PAR}}{p_s}}}{\frac{\alpha_i}{\alpha_i + \beta} \times \frac{\beta}{\alpha_i + \beta_i} \times \frac{\beta}{\alpha_i}}$$

Parameter	Value	Unit	Description
k_{Eppley}	0.063913	$^{\circ}\text{C}^{-1}$	temperature dependence factor
$k_{\text{Eppley, meso}}$	00.07608	$^{\circ}\text{C}^{-1}$	temp. dependence factor mesozooplankton
α	3.0	$(\text{W m}^2)^{-1} \text{d}^{-1}$	initial slope of P-I curve
β^{nano}	1.0	m	Coefficient for mixed layer depth influence
β^{diat}	3.0	m	Coefficient for mixed layer depth influence
μ_{max}	0.6	d^{-1}	max. phytoplankton growth rate
$K_{\text{NH}_4}^{\text{diat}}$	5e^{-7}	mol N l^{-1}	NH_4 half saturation coefficient
$K_{\text{NH}_4}^{\text{nano}}$	1e^{-7}	mol N l^{-1}	NH_4 half saturation coefficient
$K_{\text{NO}_3}^{\text{diat}}$	10e^{-6}	mol N l^{-1}	NO_3 half saturation coefficient
$K_{\text{NO}_3}^{\text{nano}}$	2e^{-6}	mol N l^{-1}	NO_3 half saturation coefficient
$K_{\text{PO}_4}^{\text{diat}}$	1e^{-7}	$\text{mol PO}_4 \text{l}^{-1}$	PO_4 half saturation coefficient
$K_{\text{PO}_4}^{\text{nano}}$	1e^{-7}	$\text{mol PO}_4 \text{l}^{-1}$	PO_4 half saturation coefficient
$K_{\text{Fe,min}}^{\text{diat}}$	1e^{-10}	mol Fe l^{-1}	min. Fe half saturation coefficient
$K_{\text{Fe,min}}^{\text{nano}}$	2e^{-11}	mol Fe l^{-1}	min. Fe half saturation coefficient
$K_{\text{Fe}}^{\text{diat}}$	4e^{-10}	mol Fe l^{-1}	Fe half saturation coefficient
$K_{\text{Fe}}^{\text{nano}}$	8e^{-11}	mol Fe l^{-1}	Fe half saturation coefficient
$K_{\text{SiO}_3}^{\text{diat}}$	3.33e^{-6}	$\text{mol SiOH}_4 \text{l}^{-1}$	SiOH_4 half saturation coefficient
$u_{\text{max}}^{\text{meso} \rightarrow \text{nano}}$	0.7	d^{-1}	max. meso zoo. growth rate on nanos
$u_{\text{max}}^{\text{micro} \rightarrow \text{nano}}$	4.0	d^{-1}	max. micro zoo. growth rate on nanos
K_G	20e^{-6}	molC l^{-1}	half-saturation constant for grazing
$\Psi_{\text{nano}}^{\text{micro}}$	0.5		preference coefficient for micro grazing on nanos
$\Psi_{\text{diat}}^{\text{micro}}$	0.5		preference coefficient for micro grazing on diatoms
$\Psi_{\text{nano}}^{\text{meso}}$	0.2		preference coefficient for meso grazing on nanos
$\Psi_{\text{diat}}^{\text{meso}}$	1.0		preference coefficient for meso grazing on diatoms
$\Psi_{\text{micro}}^{\text{meso}}$	1.0		preference coefficient for meso grazing on micro

Table 4: PISCES parameters

Grazing:

$$G_{\text{micro} \rightarrow \text{nano}} = u_{\text{max}}^{\text{micro} \rightarrow \text{nano}} \times T_f \times Z_{\text{micro}} \times \max \begin{cases} 0 \\ 1 - e^{\lambda(T_{\text{nano}}^{\text{micro}} - P_{\text{nano}})} \end{cases}$$

805 $G_{\text{meso} \rightarrow \text{nano}}, G_{\text{meso} \rightarrow \text{diat}}, G_{\text{meso} \rightarrow \text{micro}}, G_{\text{pred} \rightarrow \text{diat}}$ are all calculated using the same equation but different parameters.

$$G_{\text{pred} \rightarrow \text{micro}} = u_{\text{max}}^{\text{pred} \rightarrow \text{micro}} \times T_f \times Z_{\text{pred}} \times \max \begin{cases} 0 \\ 1 - e^{\lambda(T_{\text{micro}}^{\text{pred}} - P_{\text{micro}})} \end{cases} \times e^{-\Psi_{\text{micro}} Z_{\text{pred}}}$$

$G_{\text{pred} \rightarrow \text{meso}}$ analog.

A5 PELAGOS

Growth rate of phytoplankton PFT i:

810
$$\mu^i = \mu_{\text{max}}^i \times T_f \times \{N_{\text{lim, Fe}}^i \times N_{\text{lim, Si}}^i\} \times L_{\text{lim}}^i$$

Nutrient limitation with respect to phosphate and nitrate is not included in the phytoplankton growth rate, but acts through the exudation and lysis terms. The exudation and lysis terms have not been recalculated in this work, instead we estimated a multiplicative nutrient limitation factor (see Section 9). We refer to Vichi et al. (2007) for a full description of the nutrient limitation in PELAGOS.

Temperature function for PFT i:

$$T_f = (Q_{10}^i)^{\frac{T-10}{10}}$$

Light limitation of PFT i:

820
$$L_{\text{lim}}^i = 1 - e^{-\frac{-\alpha^i \times \theta^i \times I_{\text{PAR}}}{\mu_{\text{max}}^i \times C_{\text{chl}} \times I_{\text{PAR}}}}$$

Grazing:

Grazing of zooplankton type i on phytoplankton type j is calculated as:

$$G^{i \rightarrow j} = u_{\text{max}}^i \times T_f \times \frac{\delta_j^i \times e_j^i \times P_j}{F} \times \frac{F}{F + K_{1/2}^F} \times Z_i$$

where F denotes the total food available and is calculated as:

825
$$F = \sum_j \delta_j^i \times e_j^i \times P_j$$

e_j^i denotes the capture efficiency of zooplankton i when grazing on phytoplankton j , and is set so 1.0 for mesozooplankton, but depends on prey density for microzooplankton and heterotrophic flagellates:

$$e_j^{\text{micro, flagellates}} = \frac{P_j}{P_j + \mu_{\text{micro, flagellates}}}$$

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Growth rate of phytoplankton PFT i:

$$\mu^i = \mu_{\text{max}}^i \times T_f \times N_{\text{lim}}^i \times L_{\text{lim}}^i$$

Temperature function for PFT i:

Parameter	Value	Unit	Description
k_{Eppley}	0.0639	$^{\circ}\text{C}^{-1}$	temperature dependence factor
μ_{\max}^{diat}	1.2	d^{-1}	max. phytoplankton growth rate at T_{ref}
μ_{\max}^{nano}	0.6	d^{-1}	max. phytoplankton growth rate at T_{ref}
α^{diat}	0.045	$(\text{W m}^{-2})^{-1} \text{d}^{-1}$	Initial slope of P-I curve
α^{nano}	0.013	$(\text{W m}^{-2})^{-1} \text{d}^{-1}$	Initial slope of P-I curve
β	1.4e^{-15}	$(\text{W m}^{-2})^{-1} \text{d}^{-1}$	Photoinhibition index
P_S^{diat}	1.4	d^{-1}	Potential maximum light saturated photosynthetic rate
P_S^{nano}	0.4	d^{-1}	Potential maximum light saturated photosynthetic rate
$K_{\text{NH}_4}^{\text{diat}}$	0.3	$\mu\text{mol l}^{-1}$	NH_4 Half-saturation coefficient
$K_{\text{NH}_4}^{\text{nano}}$	0.1e^{-6}	$\mu\text{mol l}^{-1}$	NH_4 half-saturation coefficient
$A_{\text{NO}_3}^{\text{diat}}$	10.0	$(\text{mol N})^{-1} \text{s}^{-1}$	Potential maximum affinity for NO_3
$A_{\text{NO}_3}^{\text{nano}}$	30.0	$(\text{mol N})^{-1} \text{s}^{-1}$	Potential maximum affinity for NO_3
$A_{\text{NH}_4}^{\text{diat}}$	100.0	$(\text{mol N})^{-1} \text{s}^{-1}$	Potential maximum affinity for NH_4
$A_{\text{NH}_4}^{\text{nano}}$	300.0	$(\text{mol N})^{-1} \text{s}^{-1}$	Potential maximum affinity for NH_4
$A_{\text{Fe}}^{\text{diat}}$	1.111e^{-5}	$(\text{mol Fe})^{-1} \text{s}^{-1}$	Potential maximum affinity for Fe
$A_{\text{Fe}}^{\text{nano}}$	2.5e^{-5}	$(\text{mol Fe})^{-1} \text{s}^{-1}$	Potential maximum affinity for Fe
$A_{\text{SiO}_3}^{\text{diat}}$	1.6666	$(\text{mol SiOH}_4)^{-1} \text{s}^{-1}$	Potential maximum affinity for SiOH_4
$u_{\max}^{\text{micro} \rightarrow \text{nano}}$	0.4	d^{-1}	Max. micro zoo. growth rate on nanos at 0°C
$u_{\max}^{\text{meso} \rightarrow \text{nano}}$	0.1	d^{-1}	Max. meso zoo. growth rate on nanos at 0°C
$u_{\max}^{\text{meso} \rightarrow \text{diat}}$	0.4	d^{-1}	Max. meso zoo. growth rate on diatoms at 0°C
$u_{\max}^{\text{meso} \rightarrow \text{micro}}$	0.4	d^{-1}	Max. meso zoo. growth rate on micro zoo. at 0°C
$u_{\max}^{\text{pred} \rightarrow \text{diat}}$	0.2	d^{-1}	Max. pred zoo. growth rate on diatoms at 0°C
$u_{\max}^{\text{pred} \rightarrow \text{micro}}$	0.2	d^{-1}	Max. pred zoo. growth rate on micro zoo. at 0°C
$u_{\max}^{\text{pred} \rightarrow \text{meso}}$	0.4	d^{-1}	Max. pred zoo. growth rate on meso zoo. at 0°C
$T_{\text{nano}}^{\text{micro}}$	0.043	$\mu\text{molN l}^{-1}$	Threshold value for micro. zoo. grazing on nanos
$T_{\text{nano}}^{\text{meso}}$	0.04	$\mu\text{molN l}^{-1}$	Threshold value for meso. zoo. grazing on nanos
$T_{\text{diat}}^{\text{meso}}$	0.04	$\mu\text{molN l}^{-1}$	Threshold value for meso. zoo. grazing on diatoms
$T_{\text{micro}}^{\text{meso}}$	0.04	$\mu\text{molN l}^{-1}$	Threshold value for meso. zoo. grazing on micro zoo.
$T_{\text{diat}}^{\text{pred}}$	0.04	$\mu\text{molN l}^{-1}$	Threshold value for pred. zoo. grazing on diatoms
$T_{\text{micro}}^{\text{pred}}$	0.04	$\mu\text{molN l}^{-1}$	Threshold value for pred. zoo. grazing on micro zoo
$T_{\text{meso}}^{\text{pred}}$	0.04	$\mu\text{molN l}^{-1}$	Threshold value for pred. zoo. grazing on meso zoo
λ	1.4	$1 \mu\text{molN}^{-1}$	Ivlev constant (all zoo PFTs)
Ψ_{micro}	3.01	$1 \mu\text{molN}^{-1}$	Preference coefficient for predation on micro zoo
Ψ_{meso}	4.605	$1 \mu\text{molN}^{-1}$	Preference coefficient for predation on meso zoo

Table 5: MEM parameters

Parameter	Value	Unit	Description
μ_{\max}^{diat}	3.0	d^{-1}	Max. phytoplankton growth rate
μ_{\max}^{nano}	3.0	d^{-1}	Max. phytoplankton growth rate
μ_{\max}^{pico}	3.0	d^{-1}	Max. phytoplankton growth rate
Q_{10}	2		temperature dependence factor
$Q_{10, \text{meso}}$	3		temperature dependence factor
α^{diat}	$1.38e^{-5}$	$\text{mgC (mg Chl)}^{-1} \mu\text{E}^{-1} \text{m}^2 \text{s}$	Maximal light utilization coefficient
α^{nano}	$0.46e^{-5}$	$\text{mgC (mg Chl)}^{-1} \mu\text{E}^{-1} \text{m}^2 \text{s}$	Maximal light utilization coefficient
α^{pico}	$1.52e^{-5}$	$\text{mgC (mg Chl)}^{-1} \mu\text{E}^{-1} \text{m}^2 \text{s}$	Maximal light utilization coefficient
u^{meso}	2.0	d^{-1}	Max. zoo. growth rate rate
u^{micro}	2.0	d^{-1}	Max. zoo. growth rate rate
$u^{\text{flagellates}}$	10.0	d^{-1}	Max. zoo. growth rate rate
$\delta_{\text{diat}}^{\text{micro}}$	0.2		Prey availability
$\delta_{\text{nano}}^{\text{micro}}$	1.0		Prey availability
$\delta_{\text{pico}}^{\text{micro}}$	0.1		Prey availability
$\delta_{\text{micro}}^{\text{micro}}$	1.0		Prey availability
$\delta_{\text{flagellates}}^{\text{micro}}$	0.8		Prey availability
$\delta_{\text{diat}}^{\text{meso}}$	1.0		Prey availability
$\delta_{\text{meso}}^{\text{meso}}$	1.0		Prey availability
$\delta_{\text{micro}}^{\text{meso}}$	1.0		Prey availability
$\delta_{\text{pico}}^{\text{flagellates}}$	0.9		Prey availability
$\delta_{\text{flagellates}}^{\text{flagellates}}$	0.2		Prey availability
e^{meso}	1		Capture efficiency
μ_{micro}	20.0	mg C m^{-3}	Feeding threshold
$\mu_{\text{flagellates}}$	20.0	mg C m^{-3}	Feeding threshold
$K_{1/2}^{\text{F,meso}}$	80	mg C m^{-3}	Grazing half-saturation constant
$K_{1/2}^{\text{F,micro}}$	20	mg C m^{-3}	Grazing half-saturation constant
$K_{1/2}^{\text{F,flagellates}}$	20	mg C m^{-3}	Grazing half-saturation constant

Table 6: PELAGOS parameters

$$T_f = (Q_{10}^i)^{\frac{T}{10}}$$

835 **Total nutrient limitation:**

$$N_{\text{lim}}^{\text{nano}} = \min(N_{\text{Fe}}^{\text{nano}}, N_{\text{NO}_3}^{\text{nano}})$$

$$N_{\text{lim}}^{\text{diat}} = \min(N_{\text{Fe}}^{\text{diat}}, N_{\text{NO}_3}^{\text{diat}}, N_{\text{SiOH}_4}^{\text{diat}})$$

Iron limitation of PFT i:

$$N_{\text{Fe}}^i = \frac{\theta_{\text{Fe/C}}^i - \theta_{\text{Fe/C,min}}^i}{\theta_{\text{Fe/C,opt}}^i - \theta_{\text{Fe/C,min}}^i}$$

840 **Silicate limitation of diatoms:**

$$N_{\text{SO}_3}^{\text{diat}} = \frac{\text{SO}_3}{\text{SO}_3 + K_{\text{SO}_3}^{\text{diat}}}$$

Nitrate and Ammonium limitation of PFT i:

$$N_{\text{NO}_3}^i = \frac{\text{NO}_3}{\text{NO}_3 + K_{\text{NO}_3}^i}$$

Light limitation of PFT i:

845
$$L_{\text{lim}}^i = 1 - e^{-\frac{\alpha^i \times \theta_{\text{chl/C}}^i \times I_{\text{PAR}}}{\mu_{\text{max}}^i \times T_f \times N_{\text{lim}}^i}}$$

Grazing:

$$G_{\text{micro}} = u_{\text{max}}^{\text{micro}} \times T_f \times \frac{\Psi_{\text{micro}}^i P_i}{K_{1/2}^{\text{micro}} + \sum_{i \in F} \Psi_{\text{micro}}^i P_i} \times Z_{\text{micro}}$$

The food sources F for microzooplankton are small phytoplankton, diatoms, coccolithophores and small particulate organic carbon.

850
$$G_{\text{meso}} = u_{\text{max}}^{\text{meso}} \times T_{f,\text{meso}} \times \frac{\Psi_{\text{meso}}^i P_i}{K_{1/2}^{\text{meso}} + \sum_{i \in F} \Psi_{\text{meso}}^i P_i} \times Z_{\text{meso}}$$

The food sources F for mesozooplankton are small phytoplankton, diatoms, coccolithophores and small particulate organic carbon.

A7 REcoM2

Growth rate of phytoplankton PFT i:

855
$$\mu^i = \mu_{\text{max}}^i \times T_f \times N_{\text{lim}}^i \times L_{\text{lim}}^i$$

Temperature function for all PFTs:

$$T_f = e^{-4500 \times \left(\frac{1}{T} - \frac{1}{T_{\text{ref}}} \right)}$$

Total nutrient limitation:

$$N_{\text{lim}}^{\text{nano}} = \min(N_{\text{Fe}}^{\text{nano}}, N_{\text{N}}^{\text{nano}})$$

860
$$N_{\text{lim}}^{\text{diat}} = \min(N_{\text{Fe}}^{\text{diat}}, N_{\text{N}}^{\text{diat}}, N_{\text{SiOH}_4}^{\text{diat}})$$

Iron limitation of PFT i:

$$N_{\text{Fe}}^i = \frac{\text{Fe}}{\text{Fe} + K_{\text{Fe}}^i}$$

Silicate limitation of diatoms:

$$N_{\text{SO}_3}^{\text{diat}} = \begin{cases} 1 - e^{-4\theta_{\text{min}}^{\text{SiO}_3} (\theta_{\text{Si/C,min}} - \theta_{\text{Si/C}}^{\text{diat}})^2} & \theta_{\text{Si/C,min}} < \theta_{\text{Si/C}}^{\text{diat}} \\ 0 & \theta_{\text{Si/C,min}} \geq \theta_{\text{Si/C}}^{\text{diat}} \end{cases}$$

865 **Nitrate and Ammonium limitation of PFT i:**

$$N_{\text{N}}^i = \begin{cases} 1 - e^{-4\theta_{\text{min}}^{\text{N}} (\theta_{\text{N/C,min}} - \theta_{\text{N/C}}^i)^2} & \theta_{\text{N/C,min}} < \theta_{\text{N/C}}^i \\ 0 & \theta_{\text{N/C,min}} \geq \theta_{\text{N/C}}^i \end{cases}$$

Light limitation of PFT i:

Parameter	Value	Unit	Description
μ_{\max}^{diat}	0.33	d^{-1}	Max. phytoplankton growth rate at 0°C
μ_{\max}^{nano}	0.16	d^{-1}	Max. phytoplankton growth rate at 0°C
$\mu_{\max}^{\text{cocco}}$	0.23	d^{-1}	Max. phytoplankton growth rate at 0°C
Q_{10}^{diat}	1.93		Temperature dependence factor
Q_{10}^{nano}	2.08		Temperature dependence factor
Q_{10}^{cocco}	1.68		Temperature dependence factor
Q_{10}^{micro}	1.71		Temperature dependence factor
Q_{10}^{meso}	3.18		Temperature dependence factor
α^{diat}	0.79e^{-6}	$\text{mol C m}^2 (\text{g Chl W d})^{-1}$	Initial slope of P-I curve
α^{nano}	0.83e^{-6}	$\text{mol C m}^2 (\text{g Chl W d})^{-1}$	Initial slope of P-I curve
α^{cocco}	1.25e^{-6}	$\text{mol C m}^2 (\text{g Chl W d})^{-1}$	Initial slope of P-I curve
$K_{\text{NO}_3}^{\text{diat}}$	50.0e^{-6}	mol N m^{-3}	NO_3 Half saturation coefficient
$K_{\text{NO}_3}^{\text{nano}}$	9.2e^{-6}	mol N m^{-3}	NO_3 Half saturation coefficient
$K_{\text{NO}_3}^{\text{cocco}}$	3.0e^{-6}	mol N m^{-3}	NO_3 Half saturation coefficient
$\theta_{\text{Fe/C,min}}^{\text{diat}}$	2.5e^{-6}	$\text{mol Fe} (\text{mol C})^{-1}$	Minimum Fe:C ratio
$\theta_{\text{Fe/C,min}}^{\text{nano}}$	2.0e^{-6}	$\text{mol Fe} (\text{mol C})^{-1}$	Minimum Fe:C ratio
$\theta_{\text{Fe/C,min}}^{\text{cocco}}$	3.7e^{-6}	$\text{mol Fe} (\text{mol C})^{-1}$	Minimum Fe:C ratio
$\theta_{\text{Fe/C,opt}}^{\text{diat}}$	3.2e^{-6}	$\text{mol Fe} (\text{mol C})^{-1}$	Optimal Fe:C ratio
$\theta_{\text{Fe/C,opt}}^{\text{nano}}$	3.0e^{-6}	$\text{mol Fe} (\text{mol C})^{-1}$	Optimal Fe:C ratio
$\theta_{\text{Fe/C,opt}}^{\text{cocco}}$	5.9e^{-6}	$\text{mol Fe} (\text{mol C})^{-1}$	Optimal Fe:C ratio
$K_{\text{SiO}_3}^{\text{diat}}$	4.0e^{-6}	$\text{mol SiOH}_4 \text{ m}^{-3}$	SiOH_4 Half saturation coefficient
u_{\max}^{micro}	0.3	d^{-1}	Max. micro zoo. growth rate at 0°C
u_{\max}^{meso}	0.26	d^{-1}	Max. meso zoo. growth rate at 0°C

Table 7: PlankTOM5.3 parameters

$$L_{\text{lim}}^i = 1 - e^{-\frac{\alpha^i \times \theta_{\text{Chl/C}}^i \times I_{\text{PAR}}}{\mu_{\max}^i \times T_f \times N_{\text{lim}}^i}}$$

Grazing:

$$870 \quad G_{\text{nano}} = u_{\max} \times T_f \times \frac{(P_{\text{nano}} + \Psi_{\text{diat}} P_{\text{diat}})}{K_{\text{zoo}} + (P_{\text{nano}} + \Psi_{\text{diat}} P_{\text{diat}})^2} \times P_{\text{nano}} \times Z$$

$$G_{\text{diat}} = u_{\max} \times T_f \times \frac{(P_{\text{nano}} + \Psi_{\text{diat}} P_{\text{diat}})}{K_{\text{zoo}} + (P_{\text{nano}} + \Psi_{\text{diat}} P_{\text{diat}})^2} \times \Psi_{\text{diat}} P_{\text{diat}} \times Z$$

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Parameter	Value	Unit	Description
μ_{\max}^{diat}	3.5	d^{-1}	Max. phytoplankton growth rate at 0°C
μ_{\max}^{nano}	3.0	d^{-1}	Max. phytoplankton growth rate at 0°C
T_{ref}	288.15	$^{\circ}\text{K}$	Temperature dependence factor
α^{diat}	0.19	$\text{mmol C (mg Chl)}^{-1} (\text{W m}^{-2} \text{d})^{-1}$	Initial slope of P-I curve
α^{nano}	0.14	$\text{mmol C (mg Chl)}^{-1} (\text{W m}^{-2} \text{d})^{-1}$	Initial slope of P-I curve
$K_{\text{Fe}}^{\text{diat}}$	0.12	$\mu\text{mol Fe m}^{-3}$	Fe Half saturation coefficient
$K_{\text{Fe}}^{\text{nano}}$	0.02	$\mu\text{mol Fe m}^{-3}$	Fe Half saturation coefficient
$\theta_{\text{N/C,min}}$	0.04	$\text{mol N (mol C)}^{-1}$	Minimum N:C ratio
$\theta_{\text{Si/C,min}}$	0.04	$\text{mol Si (mol C)}^{-1}$	Minimum Si:C ratio
θ_{\min}^{N}	50		regulation slope
$\theta_{\min}^{\text{Si}}$	1000		regulation slope
K_{Zoo}	0.35	$(\text{mmolN m}^{-3})^2$	half-saturation constant for grazing
u_{\max}	2.4	d^{-1}	max. micro zoo. growth rate on nanos
Ψ_{diat}	0.5		preference coefficient for grazing on diatoms

Table 8: REcoM2 parameters

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References

- Allen, M. R. and Ingram, W. J.: Constraints on future changes in climate and the hydrologic cycle., *Nature*, 419, 224–32, doi:10.1038/nature01092, 2002.
- Alvain, S., Le Quéré, C., Bopp, L., Racault, M.-F., Beaugrand, G., Dessailly, D., and Buitenhuis, E. T.: Rapid
885 climatic driven shifts of diatoms at high latitudes, *Remote Sensing of Environment*, 132, 195–201, doi: 10.1016/j.rse.2013.01.014, 2013.
- Antoine, D., Morel, A., Gordon, H. R., Banzon, V. F., and Evans, R. H.: Bridging ocean color observations of the 1980s and 2000s in search of long-term trends, *Journal of Geophysical Research*, 110, C06009, doi: 10.1029/2004JC002620, 2005.
- 890 Aumont, O. and Bopp, L.: Globalizing results from ocean *in situ* iron fertilization studies, *Global Biogeochemical Cycles*, 20, 1–15, doi:10.1029/2005GB002591, 2006.
- Behrenfeld, M. J.: Abandoning Sverdrup’s Critical Depth Hypothesis on phytoplankton blooms., *Ecology*, 91, 977–89, 2010.
- Behrenfeld, M. J. and Falkowski, P. G.: Photosynthetic rates derived from satellite-based chlorophyll concentration, *Limnology and Oceanography*, 42, 1–20, 1997.
- 895 Behrenfeld, M. J., O’Malley, R. T., Siegel, D. A., McClain, C. R., Sarmiento, J. L., Feldman, G. C., Milligan, A. J., Falkowski, P. G., Letelier, R. M., and Boss, E. S.: Climate-driven trends in contemporary ocean productivity., *Nature*, 444, 752–755, doi:10.1038/nature05317, 2006.
- Behrenfeld, M. J., Doney, S. C., Lima, I., Boss, E. S., and Siegel, D. A.: Annual cycles of ecological disturbance
900 and recovery underlying the subarctic Atlantic spring plankton bloom, *Global Biogeochemical Cycles*, 27, 526–540, doi:10.1002/gbc.20050, 2013.
- Bissinger, J. E., Montagnes, D. J. S., Sharples, J., and Atkinson, D.: Predicting marine phytoplankton maximum growth rates from temperature: improving on the Eppley curve using quantile regression, *Limnology and oceanography*, 53, 487–493, doi:10.4319/lo.2008.53.2.0487, 2008.
- 905 Bopp, L., Monfray, P., Aumont, O., Dufresne, J., Le Treut, H., Madec, G., Terray, L., and Orr, J.: Potential impact of climate change on marine export production, *Global Biogeochemical Cycles*, 15, 81–100, 2001.
- Bopp, L., Aumont, O., Cadule, P., Alvain, S., and Gehlen, M.: Response of diatoms distribution to global warming and potential implications: A global model study, *Geophysical Research Letters*, 32, 2–5, doi: 10.1029/2005GL023653, 2005.
- 910 Bopp, L., Resplandy, L., Orr, J. C., Doney, S. C., Dunne, J. P., Gehlen, M., Halloran, P., Heinze, C., Ilyina, T., Séférian, R., Tjiputra, J., and Vichi, M.: Multiple stressors of ocean ecosystems in the 21st century: projections with CMIP5 models, *Biogeosciences*, 10, 6225–6245, doi:10.5194/bg-10-6225-2013, 2013.
- Boyce, D. G., Lewis, M. R., and Worm, B.: Global phytoplankton decline over the past century, *Nature*, 466, 591–596, doi:10.1038/nature09268, 2010.
- 915 Boyd, P. W. and Doney, S. C.: Modelling regional responses by marine pelagic ecosystems to global climate change, *Geophysical Research Letters*, 29, 1–4, doi:10.1029/2001GL014130, 2002.
- Buitenhuis, E. T., Hashioka, T., and Le Quéré, C.: Combined constraints on global ocean primary production using observations and models, *Global Biogeochemical Cycles*, 27, 847–858, doi:10.1002/gbc.20074, 2013a.
- 920 Buitenhuis, E. T., Vogt, M., Moriarty, R., Bednaršek, N., Doney, S. C., Leblanc, K., Le Quéré, C., Luo, Y.-W.,

- O'Brien, C., O'Brien, T., Peloquin, J., Schiebel, R., and Swan, C.: MAREDAT: towards a world atlas of MARine Ecosystem DATA, *Earth System Science Data*, 5, 227–239, doi:10.5194/essd-5-227-2013, 2013b.
- Cabré, A., Marinov, I., and Leung, S.: Consistent global responses of marine ecosystems to future climate change across the IPCC AR5 earth system models, *Climate Dynamics*, doi:10.1007/s00382-014-2374-3, 925 2014.
- Cagnazzo, C., Manzini, E., Fogli, P. G., Vichi, M., and Davini, P.: Role of stratospheric dynamics in the ozone–carbon connection in the Southern Hemisphere, *Climate dynamics*, 41, 3039–3054, 2013.
- Carr, M., Friedrichs, M., Schmeltz, M., Noguchiaita, M., Antoine, D., Arrigo, K., Asanuma, I., Aumont, O., Barber, R., and Behrenfeld, M.: A comparison of global estimates of marine primary production from ocean color, *Deep Sea Research Part II: Topical Studies in Oceanography*, 53, 741–770, doi:10.1016/j.dsr2.2006.01.028, 2006.
- 930 Collins, W. J., Bellouin, N., Doutriaux-Boucher, M., Gedney, N., Halloran, P., Hinton, T., Hughes, J., Jones, C. D., Joshi, M., Liddicoat, S., Martin, G., O'Connor, F., Rae, J., Senior, C., Sitch, S., Totterdell, I., Wiltshire, a., and Woodward, S.: Development and evaluation of an Earth-System model HadGEM2, *Geoscientific Model Development*, 4, 1051–1075, doi:10.5194/gmd-4-1051-2011, 2011.
- 935 Duarte, C. M., Regaudie-de Gioux, A., Arrieta, J. M., Delgado-Huertas, A., and Agustí, S.: The oligotrophic ocean is heterotrophic., *Annual review of marine science*, 5, 551–69, doi:10.1146/annurev-marine-121211-172337, 2013.
- Ducklow, H. W. and Doney, S. C.: What is the metabolic state of the oligotrophic ocean? A debate., *Annual review of marine science*, 5, 525–33, doi:10.1146/annurev-marine-121211-172331, 2013.
- 940 Dufresne, J.-L., Foujols, M.-A., Denvil, S., Caubel, A., Marti, O., Aumont, O., Balkanski, Y., Bekki, S., Belenger, H., Benshila, R., Bony, S., Bopp, L., Braconnot, P., Brockmann, P., Cadule, P., Cheruy, F., Codron, F., Cozic, A., Cugnet, D., Noblet, N., Duvel, J.-P., Ethé, C., Fairhead, L., Fichet, T., Flavoni, S., Friedlingstein, P., Grandpeix, J.-Y., Guez, L., Guilyardi, E., Hauglustaine, D., Hourdin, F., Idelkadi, A., Ghattas, J., 945 Joussaume, S., Kageyama, M., Krinner, G., Labetoulle, S., Lahellec, A., Lefebvre, M.-P., Lefevre, F., Levy, C., Li, Z. X., Lloyd, J., Lott, F., Madec, G., Mancip, M., Marchand, M., Masson, S., Meurdesoif, Y., Mignot, J., Musat, I., Parouty, S., Polcher, J., Rio, C., Schulz, M., Swingedouw, D., Szopa, S., Talandier, C., Terray, P., Viovy, N., and Vuichard, N.: Climate change projections using the IPSL-CM5 Earth System Model: from CMIP3 to CMIP5, *Climate Dynamics*, 40, 2123–2165, doi:10.1007/s00382-012-1636-1, 2013.
- 950 Dunne, J. P., Armstrong, R. A., Gnanadesikan, A., and Sarmiento, J. L.: Empirical and mechanistic models for the particle export ratio, *Global Biogeochemical Cycles*, 19, 1–16, doi:10.1029/2004GB002390, 2005.
- Dunne, J. P., Hales, B., and Toggweiler, J. R.: Global calcite cycling constrained by sediment preservation controls, *Global Biogeochemical Cycles*, 26, GB3023, doi:10.1029/2010GB003935, 2012.
- Dunne, J. P., John, J. G., Shevliakova, E., Stouffer, R. J., Krasting, J. P., Malyshev, S. L., Milly, P. C. D., 955 Sentman, L. T., Adcroft, A. J., Cooke, W., Dunne, K. A., Griffies, S. M., Hallberg, R. W., Harrison, M. J., Levy, H., Wittenberg, A. T., Phillips, P. J., and Zadeh, N.: GFDLs ESM2 Global Coupled Climate-Carbon Earth System Models. Part II: Carbon System Formulation and Baseline Simulation Characteristics*, *Journal of Climate*, 26, 2247–2267, doi:10.1175/JCLI-D-12-00150.1, 2013.
- Dutkiewicz, S., Scott, J. R., and Follows, M. J.: Winners and losers: Ecological and biogeochemical changes 960 in a warming ocean, *Global Biogeochemical Cycles*, 27, 463–477, doi:10.1002/gbc.20042, 2013.

- Eppley, R.: Temperature and phytoplankton growth in the sea, *Fishery Bulletin*, 70, 1063–1085, 1972.
- Falkowski, P. G., Laws, E. A., Barber, R. T., and Murray, J. W.: Phytoplankton and their role in primary, new, and export production, in: *Ocean Biogeochemistry: The Role of the Ocean Carbon Cycle in Global Change*, edited by Fasham, M. J. R., The IGBP Series, chap. 4, pp. 99–121, Springer, Berlin, 2003.
- 965 Garcia, H., Locarnini, R. A., Boyer, T. P., Antonov, J. I., Baranova, O., Zweng, M., Reagan, J., and Johnson, D.: *World Ocean Atlas 2013, Volume 4: Dissolved Inorganic Nutrients (phosphate, nitrate, silicate)*, vol. 4, 2014.
- García-Corral, L. S., Barber, E., Regaudie-de Gioux, a., Sal, S., Holding, J. M., Agustí, S., Navarro, N., Serret, P., Mozetič, P., and Duarte, C. M.: Temperature dependence of planktonic metabolism in the subtropical
970 North Atlantic Ocean, *Biogeosciences*, 11, 4529–4540, doi:10.5194/bg-11-4529-2014, 2014.
- Geider, R. J., MacIntyre, H. L., and Kana, T. M.: A dynamic regulatory model of phytoplanktonic acclimation to light, nutrients, and temperature, *Limnology and Oceanography*, 43, 679–694, doi:10.4319/lo.1998.43.4.0679., 1998.
- Gregg, W. W.: Ocean primary production and climate: Global decadal changes, *Geophysical Research Letters*,
975 30, 10–13, doi:10.1029/2003GL016889, 2003.
- HadGEM Team, Martin, G., Bellouin, N., Collins, W. J., Culverwell, I. D., Halloran, P. R., Hardiman, S. C., Hinton, T. J., Jones, C. D., McDonald, R. E., McLaren, a. J., O'Connor, F. M., Roberts, M. J., Rodriguez, J. M., Woodward, S., Best, M. J., Brooks, M. E., Brown, a. R., Butchart, N., Dearden, C., Derbyshire, S. H., Dharssi, I., Doutriaux-Boucher, M., Edwards, J. M., Falloon, P. D., Gedney, N., Gray, L. J., Hewitt,
980 H. T., Hobson, M., Huddleston, M. R., Hughes, J., Ineson, S., Ingram, W. J., James, P. M., Johns, T. C., Johnson, C. E., Jones, A., Jones, C. P., Joshi, M. M., Keen, a. B., Liddicoat, S., Lock, a. P., Maidens, a. V., Manners, J. C., Milton, S. F., Rae, J. G. L., Ridley, J. K., Sellar, A., Senior, C. a., Totterdell, I. J., Verhoef, A., Vidale, P. L., and Wiltshire, A.: The HadGEM2 family of Met Office Unified Model climate configurations, *Geoscientific Model Development*, 4, 723–757, doi:10.5194/gmd-4-723-2011, 2011.
- 985 Hancke, K. and Glud, R.: Temperature effects on respiration and photosynthesis in three diatom-dominated benthic communities, *Aquatic Microbial Ecology*, 37, 265–281, doi:10.3354/ame037265, 2004.
- Hashioka, T., Vogt, M., Yamanaka, Y., Le Quéré, C., Buitenhuis, E. T., Aita, M. N., Alvain, S., Bopp, L., Hirata, T., Lima, I., Sailley, S., and Doney, S. C.: Phytoplankton competition during the spring bloom in four plankton functional type models, *Biogeosciences*, 10, 6833–6850, doi:10.5194/bg-10-6833-2013, 2013.
- 990 Hauck, J., Völker, C., Wang, T., Hoppema, M., Losch, M., and Wolf-Gladrow, D. A.: Seasonally different carbon flux changes in the Southern Ocean in response to the southern annular mode, *Global Biogeochemical Cycles*, 27, 1236–1245, doi:10.1002/2013GB004600, 2013.
- Henson, S. A., Sanders, R., Madsen, E., Morris, P. J., Le Moigne, F., and Quartly, G. D.: A reduced estimate of the strength of the ocean's biological carbon pump, *Geophysical Research Letters*, 38, L04606, doi:
995 10.1029/2011GL046735, 2011.
- Hirst, A. G. and Bunker, A. J.: Growth of marine planktonic copepods: Global rates and patterns in relation to chlorophyll a, temperature, and body weight, *Limnology and Oceanography*, 48, 1988–2010, doi:10.4319/lo.2003.48.5.1988, 2003.
- Hurrell, J. W., Holland, M. M., Gent, P. R., Ghan, S., Kay, J. E., Kushner, P. J., Lamarque, J.-F., Large, W. G.,
1000 Lawrence, D., Lindsay, K., Lipscomb, W. H., Long, M. C., Mahowald, N., Marsh, D. R., Neale, R. B.,

- Rasch, P., Vavrus, S., Vertenstein, M., Bader, D., Collins, W. D., Hack, J. J., Kiehl, J., and Marshall, S.: The Community Earth System Model: A Framework for Collaborative Research, *Bulletin of the American Meteorological Society*, 94, 1339–1360, doi:10.1175/BAMS-D-12-00121.1, 2013.
- Ikeda, T., Kanno, Y., Ozaki, K., and Shinada, A.: Metabolic rates of epipelagic marine copepods as a function of body mass and temperature, *Marine Biology*, pp. 587–596, doi:10.1007/s002270100608, 2001.
- Kwiatkowski, L., Yool, A., Allen, J. I., Anderson, T. R., Barciela, R., de Mora, L., Buitenhuis, E. T., Butenschön, M., Enright, C., Quéré, C. L., Racault, M.-F., and Sinha, B.: iMarNet : an ocean biogeochemistry model inter-comparison project within a common physical ocean modelling framework, *Biogeosciences*, 11, 7291–7304, doi:10.5194/bg-11-7291-2014, 2014.
- Laufkötter, C., Vogt, M., and Gruber, N.: Long-term trends in ocean plankton production and particle export between 1960–2006, *Biogeosciences*, 10, 7373–7393, doi:10.5194/bg-10-7373-2013, 2013.
- Le Quéré, C., Harrison, S. P., Prentice, I., Buitenhuis, E. T., Aumont, O., Bopp, L., Claustre, H., Da Cunha, L. C., Geider, R., Giraud, X., Klaas, C., Kohfeld, K. E., Legendre, L., Manizza, M., Platt, T., Rivkin, R. B., Sathyendranath, S., Uitz, J., Watson, A. J., and Wolf-Gladrow, D.: Ecosystem dynamics based on plankton functional types for global ocean biogeochemistry models, *Global Change Biology*, 11, 2016–2040, doi:10.1111/j.1365-2486.2005.01004.x, 2005.
- Liebig, J.: *Chemistry and its Applications to Agriculture and Physiology: On Chemical Processes in the Nutrition of Vegetables*, Taylor and Walton, London, 1840.
- Lindsay, K., Bonan, G. B., Doney, S. C., Hoffman, F. M., Lawrence, D. M., Long, M. C., Mahowald, N. M., Moore, J. K., Randerson, J. T., and Thornton, P. E.: Preindustrial Control and 20th Century Carbon Cycle Experiments with the Earth System Model CESM1(BGC), *Journal of Climate*, 1, 141006111735 008, doi:10.1175/JCLI-D-12-00565.1, 2014.
- Lomas, M. W., Glibert, P. M., Shiah, F.-K., and Smith, E. M.: Microbial processes and temperature in Chesapeake Bay: current relationships and potential impacts of regional warming, *Global Change Biology*, 8, 51–70, doi:10.1046/j.1365-2486.2002.00454.x, 2002.
- López-Urrutia, A., San Martín, E., Harris, R. P., and Irigoien, X.: Scaling the metabolic balance of the oceans., *Proceedings of the National Academy of Sciences of the United States of America*, 103, 8739–44, doi:10.1073/pnas.0601137103, 2006.
- Mahlstein, I., Gent, P. R., and Solomon, S.: Historical Antarctic mean sea ice area, sea ice trends, and winds in CMIP5 simulations, *Journal of Geophysical Research: Atmospheres*, 118, 5105–5110, doi:10.1002/jgrd.50443, 2013.
- Manizza, M., Buitenhuis, E. T., and Le Quéré, C.: Sensitivity of global ocean biogeochemical dynamics to ecosystem structure in a future climate, *Geophysical Research Letters*, 37, 3–7, doi:10.1029/2010GL043360, 2010.
- Marañón, E., Cermeño, P., Huete-Ortega, M., López-Sandoval, D. C., Mouríño Carballido, B., and Rodríguez-Ramos, T.: Resource supply overrides temperature as a controlling factor of marine phytoplankton growth, *PLoS ONE*, 9, 20–23, doi:10.1371/journal.pone.0099312, 2014.
- Marinov, I., Doney, S. C., and Lima, I. D.: Response of ocean phytoplankton community structure to climate change over the 21st century: partitioning the effects of nutrients, temperature and light, *Biogeosciences*, 7, 3941–3959, doi:10.5194/bg-7-3941-2010, 2010.

- Marinov, I., Doney, S. C., Lima, I. D., Lindsay, K., Moore, J. K., and Mahowald, N.: North-South asymmetry in the modeled phytoplankton community response to climate change over the 21st century, *Global Biogeochemical Cycles*, 27, GB004 599, doi:10.1002/2013GB004599, 2013.
- Michaelis, L. and Menten, M.: Die Kinetik der Invertinwirkung, *Biochem. Z.*, 49, 333–369, 1913.
- 1045 Moore, C. M., Mills, M. M., Arrigo, K. R., Berman-Frank, I., Bopp, L., Boyd, P. W., Galbraith, E. D., Geider, R. J., Guieu, C., Jaccard, S. L., Jickells, T. D., La Roche, J., Lenton, T. M., Mahowald, N. M., Marañón, E., Marinov, I., Moore, J. K., Nakatsuka, T., Oschlies, A., Saito, M. A., Thingstad, T. F., Tsuda, A., and Ulloa, O.: Processes and patterns of oceanic nutrient limitation, *Nature Geoscience*, 6, 701–710, doi:10.1038/ngeo1765, 2013a.
- 1050 Moore, J., Doney, S., Kleypas, J., Glover, D., and Fung, I.: An intermediate complexity marine ecosystem model for the global domain, *Deep Sea Research Part II: Topical Studies in Oceanography*, 49, 403–462, doi:10.1016/S0967-0645(01)00108-4, 2002.
- Moore, J. K., Lindsay, K., Doney, S. C., Long, M. C., and Misumi, K.: Marine Ecosystem Dynamics and Biogeochemical Cycling in the Community Earth System Model [CESM1(BGC)]: Comparison of the 1990s
 1055 with the 2090s under the RCP4.5 and RCP8.5 Scenarios, *Journal of Climate*, 26, 9291–9312, doi:10.1175/JCLI-D-12-00566.1, 2013b.
- Morán, X. A. G., López-Urrutia, A., Calvo-Díaz, A., and Li, W. K. W.: Increasing importance of small phytoplankton in a warmer ocean, *Global Change Biology*, 16, 1137–1144, 2010.
- Platt, T., Gallegos, C., and Harrison, W.: Photoinhibition of photosynthesis in natural assemblages of marine
 1060 phytoplankton in the Arctic, *Deep Sea Research*, 29, 1159–1170, 1980.
- Prowe, A. F., Pahlow, M., Dutkiewicz, S., Follows, M., and Oschlies, A.: Top-down control of marine phytoplankton diversity in a global ecosystem model, *Progress In Oceanography*, doi:10.1016/j.pocean.2011.11.016, 2011.
- Prowe, A. F., Pahlow, M., and Oschlies, A.: Controls on the diversityproductivity relationship in a marine
 1065 ecosystem model, *Ecological Modelling*, 225, 167–176, doi:10.1016/j.ecolmodel.2011.11.018, 2012.
- Rose, J. M. and Caron, D. A.: Does low temperature constrain the growth rates of heterotrophic protists? Evidence and implications for algal blooms in cold waters, *Limnology and Oceanography*, 52, 886–895, doi:10.4319/lo.2007.52.2.0886, 2007.
- Ruggio, R., Vichi, M., Paparella, F., and Masina, S.: Climatic trends of the equatorial undercurrent: A backup
 1070 mechanism for sustaining the equatorial Pacific production, *Journal of Marine Systems*, 121–122, 11–23, doi:10.1016/j.jmarsys.2013.04.001, 2013.
- Rykaczewski, R. R. and Dunne, J. P.: A measured look at ocean chlorophyll trends, *Nature*, 472, E5–6, doi:10.1038/nature09952, 2011.
- Sailley, S., Vogt, M., Doney, S., Aita, M., Bopp, L., Buitenhuis, E., Hashioka, T., Lima, I., Le Quéré, C., and
 1075 Yamanaka, Y.: Comparing food web structures and dynamics across a suite of global marine ecosystem models, *Ecological Modelling*, 261–262, 43–57, doi:10.1016/j.ecolmodel.2013.04.006, 2013.
- Sand-Jensen, K., Pedersen, N. L., and Søndergaard, M.: Bacterial metabolism in small temperate streams under contemporary and future climates, *Freshwater Biology*, 52, 2340–2353, doi:10.1111/j.1365-2427.2007.01852.x, 2007.
- 1080 Sarmiento, J. L. and Gruber, N.: *Ocean Biogeochemical Dynamics*, Princeton University Press, Princeton, 2006.

- Sarmiento, J. L., Slater, R., Barber, R., Bopp, L., Doney, S. C., Hirst, A., Kleypas, J., Matear, R., Mikolajewicz, U., Monfray, P., Soldatov, V., Spall, S., and Stouffer, R.: Response of ocean ecosystems to climate warming, *Global Biogeochemical Cycles*, 18, GB3003, doi:10.1029/2003GB002134, 2004.
- 1085 Schmittner, A. and Galbraith, E. D.: Glacial greenhouse-gas fluctuations controlled by ocean circulation changes., *Nature*, 456, 373–6, doi:10.1038/nature07531, 2008.
- Schmittner, A., Oschlies, A., Matthews, H. D., and Galbraith, E. D.: Future changes in climate, ocean circulation, ecosystems, and biogeochemical cycling simulated for a business-as-usual CO₂ emission scenario until year 4000 AD, *Global Biogeochemical Cycles*, 22, GB1013, doi:10.1029/2007GB002953, 2008.
- 1090 S  f  rian, R., Bopp, L., Gehlen, M., Orr, J. C., Eth  , C., Cadule, P., Aumont, O., Salas y M  lia, D., Voltaire, A., and Madec, G.: Skill assessment of three earth system models with common marine biogeochemistry, *Climate Dynamics*, 40, 2549–2573, doi:10.1007/s00382-012-1362-8, 2013.
- Shigemitsu, M., Okunishi, T., Nishioka, J., Sumata, H., Hashioka, T., Aita, M. N., Smith, S. L., Yoshie, N., Okada, N., and Yamanaka, Y.: Development of a one-dimensional ecosystem model including the iron cycle applied to the Oyashio region, western subarctic Pacific, *Journal of Geophysical Research*, 117, C06021, doi:10.1029/2011JC007689, 2012.
- 1095 Siegel, D., Behrenfeld, M., Maritorena, S., McClain, C., Antoine, D., Bailey, S., Bontempi, P., Boss, E., Dierssen, H., Doney, S., Eplee, R., Evans, R., Feldman, G., Fields, E., Franz, B., Kuring, N., Mengelt, C., Nelson, N., Patt, F., Robinson, W., Sarmiento, J., Swan, C., Werdell, P., Westberry, T., Wilding, J., and Yoder, J.: Regional to global assessments of phytoplankton dynamics from the SeaWiFS mission, *Remote Sensing of Environment*, 135, 77–91, doi:10.1016/j.rse.2013.03.025, 2013.
- 1100 Sinha, B., Buitenhuis, E. T., Qu  r  , C. L., and Anderson, T. R.: Comparison of the emergent behavior of a complex ecosystem model in two ocean general circulation models, *Progress In Oceanography*, 84, 204–224, doi:10.1016/j.pocean.2009.10.003, 2010.
- Smith, S., Yamanaka, Y., Pahlow, M., and Oschlies, A.: Optimal uptake kinetics: physiological acclimation explains the pattern of nitrate uptake by phytoplankton in the ocean, *Marine Ecology Progress Series*, 384, 1–12, doi:10.3354/meps08022, 2009.
- 1105 Staehr, P. A. and Sand-Jensen, K. A. J.: Seasonal changes in temperature and nutrient control of photosynthesis, respiration and growth of natural phytoplankton communities, *Freshwater Biology*, 51, 249–262, doi:10.1111/j.1365-2427.2005.01490.x, 2006.
- 1110 Steinacher, M., Joos, F., Fr  licher, T. L., Bopp, L., Cadule, P., Doney, S. C., Gehlen, M., Schneider, B., and Segschneider, J.: Projected 21st century decrease in marine productivity: a multi-model analysis, *Biogeosciences*, 6, 979–1005, 2010.
- Tadonl  k  , R.: Evidence of warming effects on phytoplankton productivity rates and their dependence on eutrophication status, *Limnology and Oceanography*, 55, 973–982, doi:10.4319/lo.2010.55.3.0973, 2010.
- 1115 Taucher, J. and Oschlies, A.: Can we predict the direction of marine primary production change under global warming?, *Geophysical Research Letters*, 38, 1–6, doi:10.1029/2010GL045934, 2011.
- Taylor, K. E., Stouffer, R. J., and Meehl, G. A.: An Overview of CMIP5 and the Experiment Design, *Bulletin of the American Meteorological Society*, 93, 485–498, doi:10.1175/BAMS-D-11-00094.1, 2012.
- 1120 Thomas, M. K., Kremer, C. T., Klausmeier, C. A., and Litchman, E.: A Global Pattern of Thermal Adaptation in Marine Phytoplankton, *Science*, 338, 1085–1088, doi:10.1023/A, 2012.

- Totterdell, I.: Description of Diat-HadOCC, Tech. rep., MetOffice, 2013.
- 1125 van Vuuren, D. P., Edmonds, J., Kainuma, M., Riahi, K., Thomson, A., Hibbard, K., Hurtt, G. C., Kram, T., Krey, V., Lamarque, J.-F., Masui, T., Meinshausen, M., Nakicenovic, N., Smith, S. J., and Rose, S. K.: The representative concentration pathways: an overview, *Climatic Change*, 109, 5–31, doi: 10.1007/s10584-011-0148-z, 2011.
- Vichi, M., Pinardi, N., and Masina, S.: A generalized model of pelagic biogeochemistry for the global ocean ecosystem. Part I: Theory, *Journal of Marine Systems*, 64, 89–109, doi:10.1016/j.jmarsys.2006.03.006, 2007.
- 1130 Vichi, M., Allen, J. I., Masina, S., and Hardman-Mountford, N. J.: The emergence of ocean biogeochemical provinces: A quantitative assessment and a diagnostic for model evaluation, *Global Biogeochemical Cycles*, 25, 1–17, doi:10.1029/2010GB003867, 2011a.
- Vichi, M., Manzini, E., Fogli, P. G., Alessandri, A., Patara, L., Scoccimarro, E., Masina, S., and Navarra, A.: Global and regional ocean carbon uptake and climate change: sensitivity to a substantial mitigation scenario, *Climate Dynamics*, 37, 1929–1947, doi:10.1007/s00382-011-1079-0, 2011b.
- 1135 Vogt, M., Hashioka, T., Payne, M. R., Buitenhuis, E. T., Quéré, C. L., Alvain, S., Aita, M. N., Bopp, L., Doney, S. C., Hirata, T., Lima, I., Sailley, S., and Yamanaka, Y.: The distribution, dominance patterns and ecological niches of plankton functional types in Dynamic Green Ocean Models and satellite estimates, *Biogeosciences Discussions*, 10, 17 193–17 247, doi:10.5194/bgd-10-17193-2013, 2013.
- 1140 Voldoire, A., Sanchez-Gomez, E., Salas y Méliá, D., Decharme, B., Cassou, C., Sénési, S., Valcke, S., Beau, I., Alias, A., Chevallier, M., Déqué, M., Deshayes, J., Douville, H., Fernandez, E., Madec, G., Maisonnave, E., Moine, M.-P., Planton, S., Saint-Martin, D., Szopa, S., Tyteca, S., Alkama, R., Belamari, S., Braun, A., Coquart, L., and Chauvin, F.: The CNRM-CM5.1 global climate model: description and basic evaluation, *Climate Dynamics*, 40, 2091–2121, doi:10.1007/s00382-011-1259-y, 2012.
- 1145 Watanabe, S., Hajima, T., Sudo, K., Nagashima, T., Takemura, T., Okajima, H., Nozawa, T., Kawase, H., Abe, M., Yokohata, T., Ise, T., Sato, H., Kato, E., Takata, K., Emori, S., and Kawamiya, M.: MIROC-ESM: model description and basic results of CMIP5-20c3m experiments, *Geoscientific Model Development Discussions*, 4, 1063–1128, doi:10.5194/gmdd-4-1063-2011, 2011.
- Webb, W. L., Newton, M., and Starr, D.: Carbon dioxide exchange of *Alnus rubra*, *Oecologia*, 17, 281–291, doi:10.1007/BF00345747, 1974.
- 1150 Wernand, M. R., van der Woerd, H. J., and Gieskes, W. W. C.: Trends in ocean colour and chlorophyll concentration from 1889 to 2000, worldwide., *PLOS one*, 8, e63 766, doi:10.1371/journal.pone.0063766, 2013.
- Westberry, T., Behrenfeld, M. J., Siegel, D. A., and Boss, E.: Carbon-based primary productivity modeling with vertically resolved photoacclimation, *Global Biogeochemical Cycles*, 22, GB2024, doi:10.1029/2007GB003078, 2008.
- 1155 Williams, P. J. L. B., Quay, P. D., Westberry, T. K., and Behrenfeld, M. J.: The oligotrophic ocean is autotrophic., *Annual review of marine science*, 5, 535–49, doi:10.1146/annurev-marine-121211-172335, 2013.

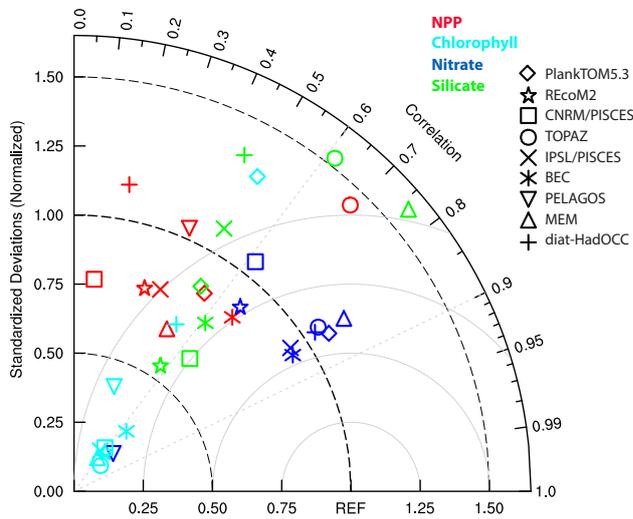


Fig. 1: Taylor diagram showing the model-data correspondence for NPP (red), surface chlorophyll (light blue), NO_3 (dark blue) and SiOH_4 (green). The data-based estimates are from WOA2013 for NO_3 and SiOH_4 , from the SeaWiFS Project for chlorophyll and from Westberry et al. (2008) for NPP. We compare nutrients for the 1990-1999 period while chlorophyll and NPP data are from 1997-2006. The angular coordinate shows the correlation coefficient, the distance from the origin denotes the normalized standard deviation and the distance from point [1,1] describes the root mean squared error.

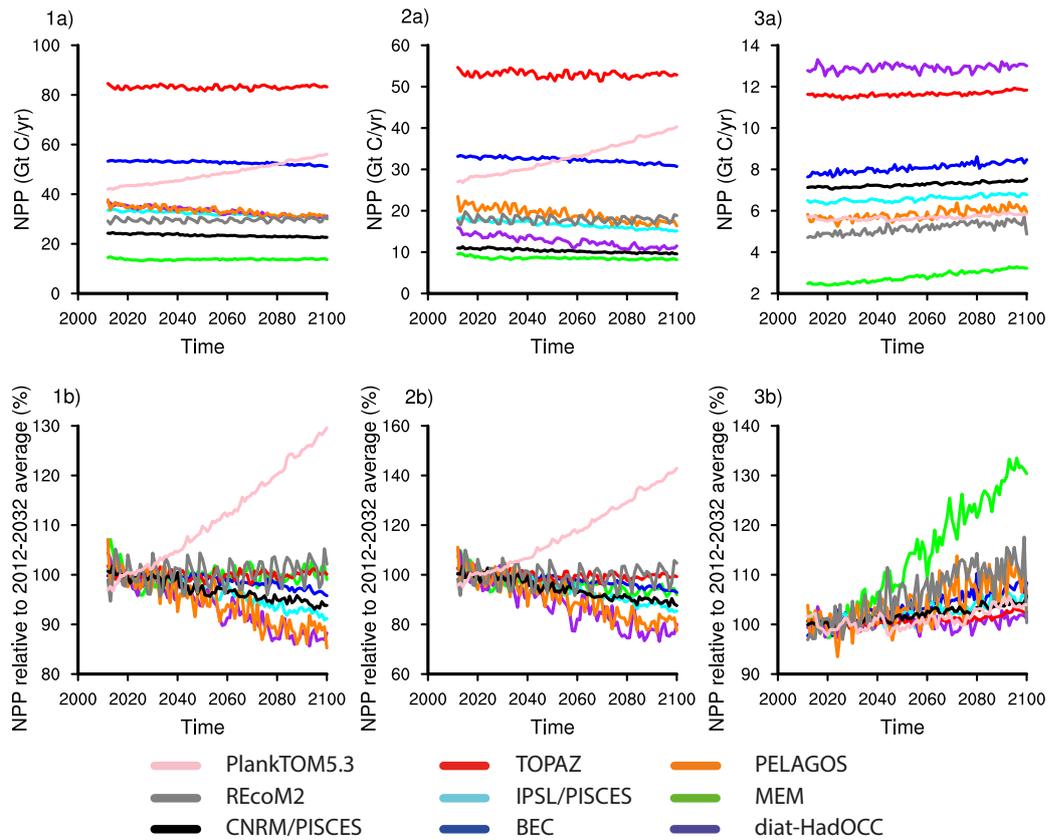


Fig. 2: Projected trends in annual mean integrated net primary production (NPP) for the 2012-2100 period under RCP8.5, shown both in GtC/year (Panels a) and in percent (Panels b). Panels 1a,b show global values, Panels 2a,b and 3a,b show low latitudes (30S-30N) and Southern Ocean (90-50S), respectively.

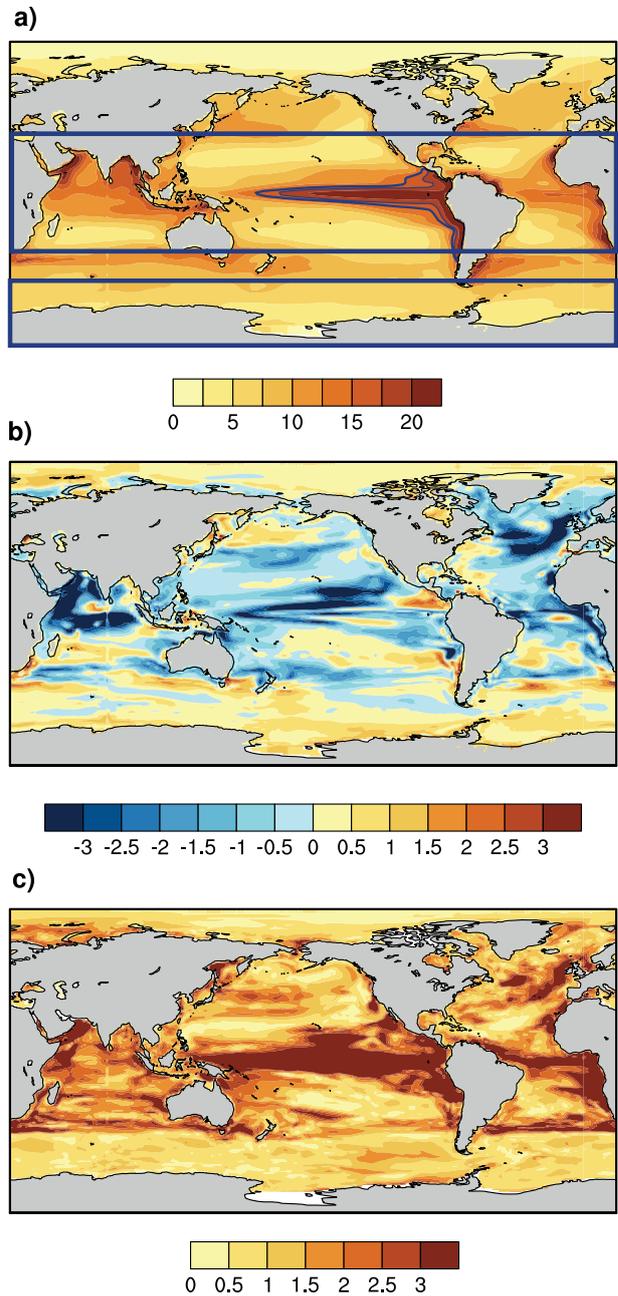


Fig. 3: Spatial patterns of multi-model annual mean integrated net primary production (NPP) for a) the 2012-2031 average, b) changes between 2081-2100 and 2012-2031 under RCP8.5 and c) interquartile range of the changes in NPP projections. The unit is $\text{mol C m}^{-2} \text{ yr}^{-1}$. The blue boxes in Panel a) mark the regions which are discussed in more detail in this work, namely the Southern Ocean south of 50° South, the low latitudes ($30^\circ\text{S} - 30^\circ\text{N}$) and the equatorial upwelling region in the Pacific.

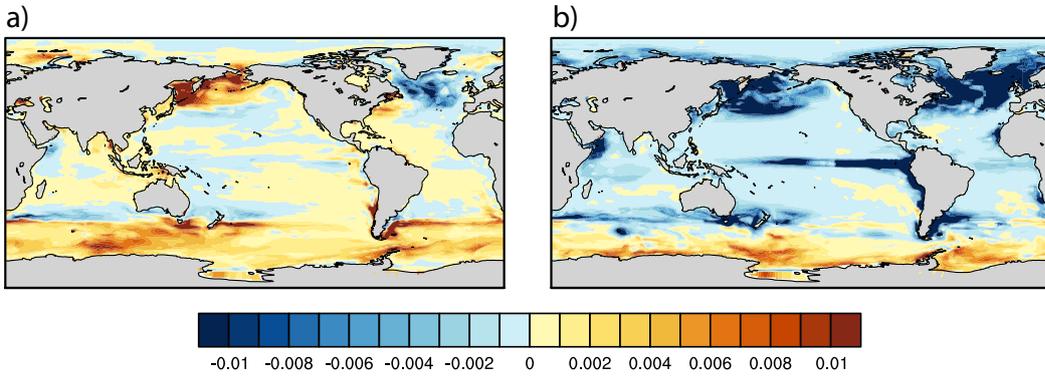


Fig. 4: First order Taylor decomposition of the surface NPP changes between 2012-2032 and 2080-2100 in a) biomass-weighted changes in growth and b) growth-weighted changes in biomass. The unit is mol C m⁻³ yr⁻¹.

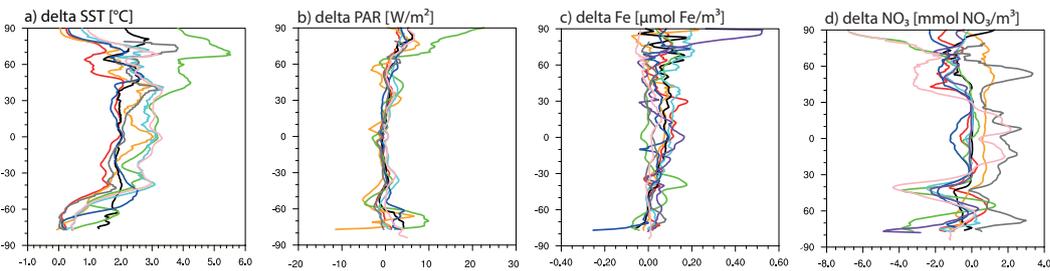


Fig. 5: Zonal mean of projected sea surface temperature change, photosynthetically active radiation (PAR) change and change in surface Fe and NO₃ concentrations. We calculate the change as the difference between the 2012-2031 average and the 2081-2100 average. Different line colors denote different models as in the legend of Fig. 2.

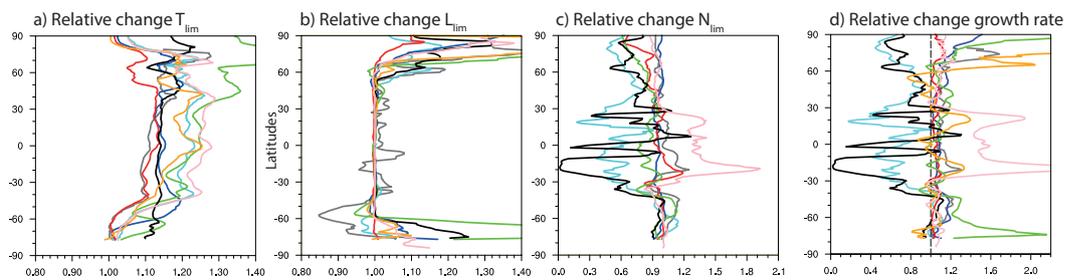


Fig. 6: Zonal mean of the relative change in temperature, nutrient and light limitation and growth rate. For each model only the values for the phytoplankton PFT with the strongest temperature (or light or nutrient limitation factor or growth rate) response is shown. We calculate the relative change as $\frac{2081-2100 \text{ average}}{2012-2031 \text{ average}}$. A value of 1 means no change and is indicated by the dotted line. Different line colors denote different models as in the legend of Fig. 2.

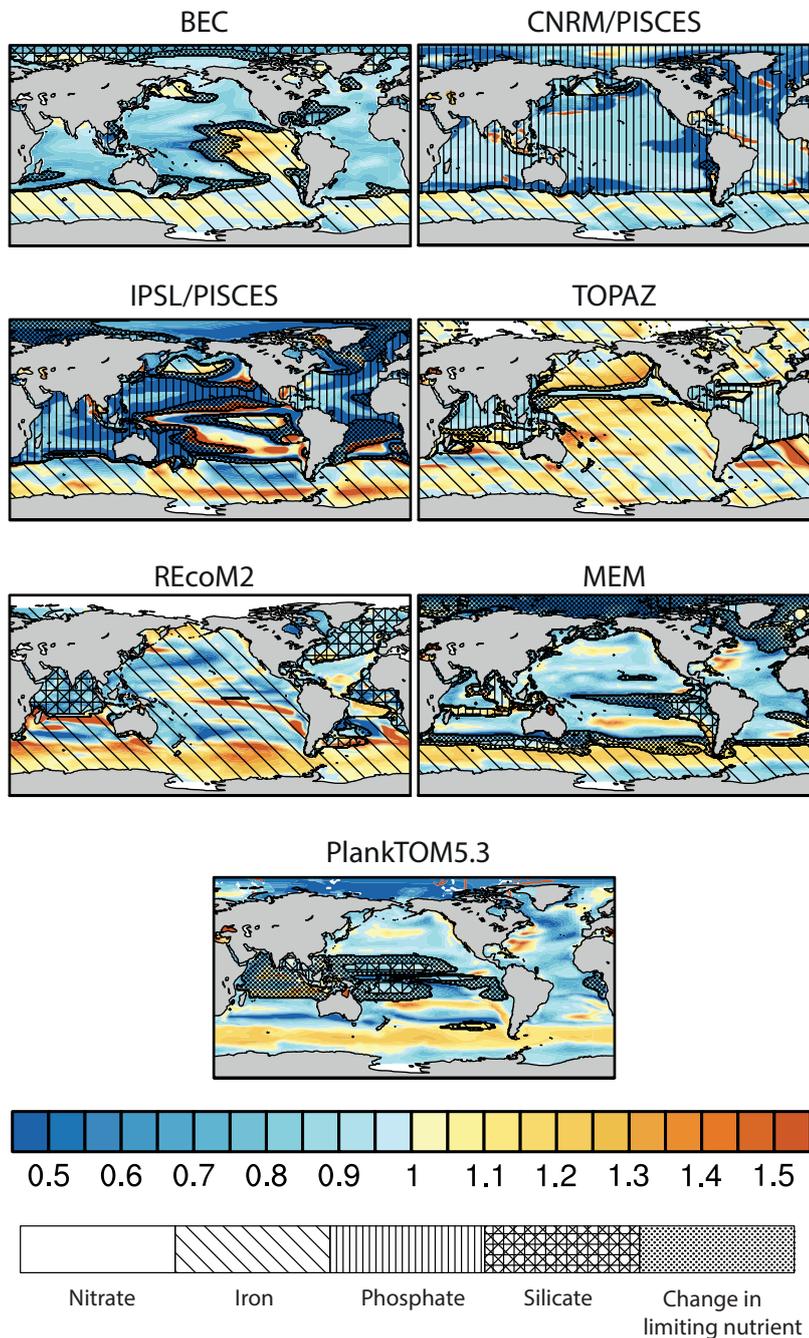


Fig. 7: Changes in relative diatom nutrient limitation (calculated as the 2081-2100 average divided by the 2012-2031 average) in all models that use Liebig limitation (smallest individual nutrient limitation term determines total nutrient limitation). The colors indicate changes in the nutrient limitation value, with positive values indicating an increase in nutrient limitation factor which is equivalent to lower nutrient limitation and an increase in growth. The hatching indicates the limiting nutrient. A change in limiting nutrient during the simulation period is shown with dots. REcoM2 does not simulate the Arctic, these missing values are shown in white.

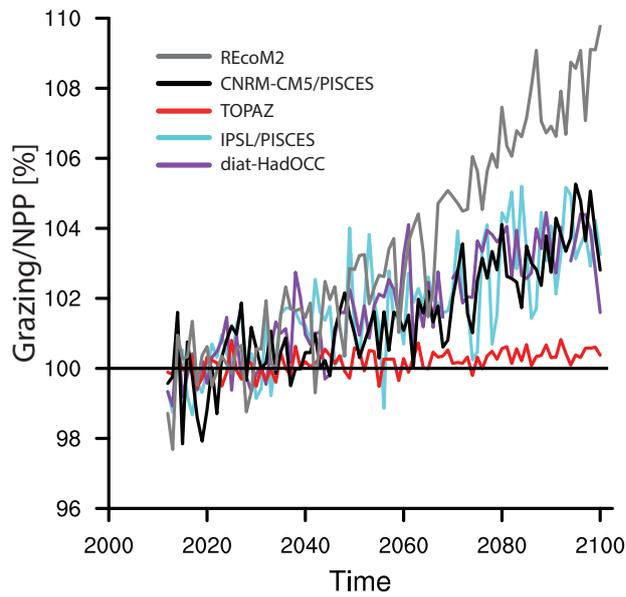


Fig. 8: Fraction of NPP that is grazed (Grazing/NPP) normalized to the 2012-2031 average in the surface of the low and high northern latitudes ($-50^{\circ}\text{S} - 90^{\circ}\text{N}$). This plot shows data from all models where total grazing on phytoplankton is available in the output.

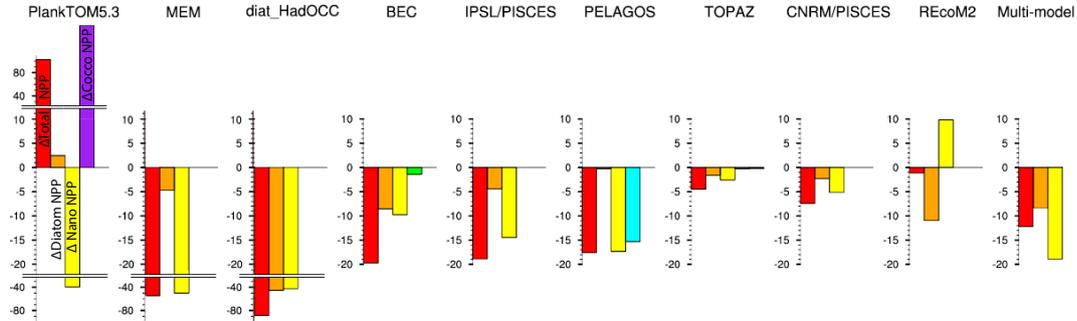


Fig. 9: Decomposition of annual mean area-averaged low latitude surface NPP changes between 2012-2032 and 2080-2100 (red bar, in $\text{mol C m}^{-2} \text{ yr}^{-1}$) into change in nanophytoplankton (yellow) and diatom (orange) surface NPP. Changes in diazotrophs (green) and picophytoplankton (light blue) have been included in the bar indicating nanophytoplankton changes for the models that simulate these functional types. For TOPAZ, changes in large non-diatom phytoplankton (dark blue) are included in the bar indicating diatom changes. Changes in coccolithophore NPP are shown in purple. Note the change in scale between the first three plots (models with large surface NPP changes) and the remaining 6 plots. While for diat-HadOCC, BEC, IPSL/PISCES, CNRM/PISCES, REcoM2 and TOPAZ the surface NPP of the PFTs was included in the model output, we show recalculated values for PlankTOM5.3, MEM and PELAGOS.

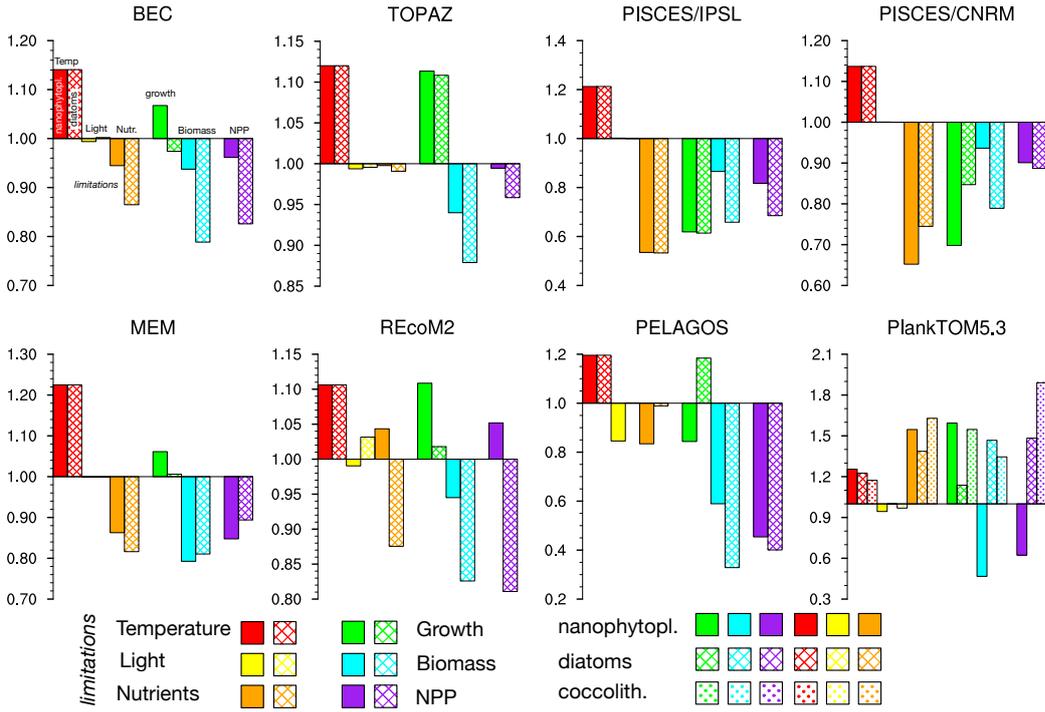


Fig. 10: Relative change in temperature limitation factor (red), light limitation factor (yellow), nutrient limitation factor (orange), growth rate (green), biomass (light blue) and NPP (purple) for nanophytoplankton (full), diatoms (hatched) and coccolithophores (dotted) in the surface of the low latitudes, for all models where the full equations were available. An increase in limitation factor denotes weaker limitation, which leads to stronger growth. The relative change of a variable is the ratio between the 2081-2100 average and the 2012-2031 average. A value of one means no change, 1.2 corresponds to a 20% increase, 0.8 corresponds to a 20% decrease. The product of the relative change in temperature, light and nutrient limitation results approximately in the relative change in growth rate. See main text for further details.

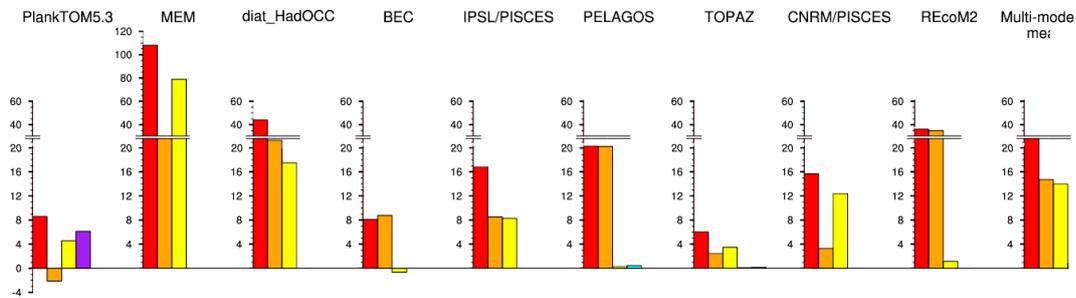


Fig. 11: Decomposition of Southern Ocean (50-90 deg S) surface NPP changes between 2012-2032 and 2080-2100 (red bar, in $\text{mol C m}^{-2} \text{ yr}^{-1}$) into change in nanophytoplankton (yellow) and diatom (orange) surface NPP. Changes in diazotrophs (green) and picophytoplankton (light blue) have been included in the bar indicating nanophytoplankton changes for the models that simulate these functional types. For TOPAZ, changes in large non-diatom phytoplankton (dark blue) are included in the bar indicating diatom changes. Changes in coccolithophore NPP are shown in purple. Note the change in scale between the first three plots (models with large surface NPP changes) and the remaining 6 plots. While for diat-HadOCC, BEC, IPSL/PISCES, CNRM/PISCES, RECOM2 and TOPAZ the surface NPP of the PFTs was included in the model output, we show recalculated values for PlankTOM5.3, MEM and PELAGOS.

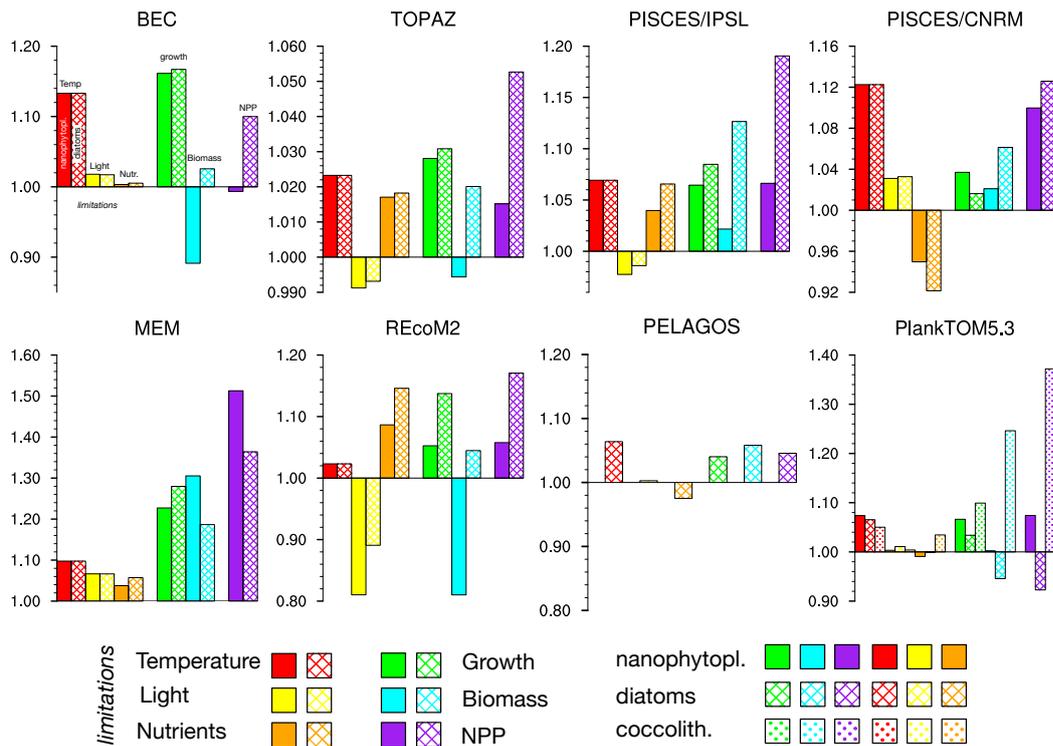


Fig. 12: Relative changes between 2012-2032 and 2080-2100 in annual mean temperature limitation factor (red), light limitation factor (yellow), nutrient limitation factor (orange), growth rate (green), biomass (light blue) and NPP (purple) for nanophytoplankton (full), diatoms (hatched) and coccolithophores (dotted) in the surface of the Southern Ocean (50-90°S). An increase in limitation factor denotes weaker limitation, which leads to stronger growth. PELAGOS has a relative diatom contribution of more than 95% of total biomass, therefore we show only results for diatoms.

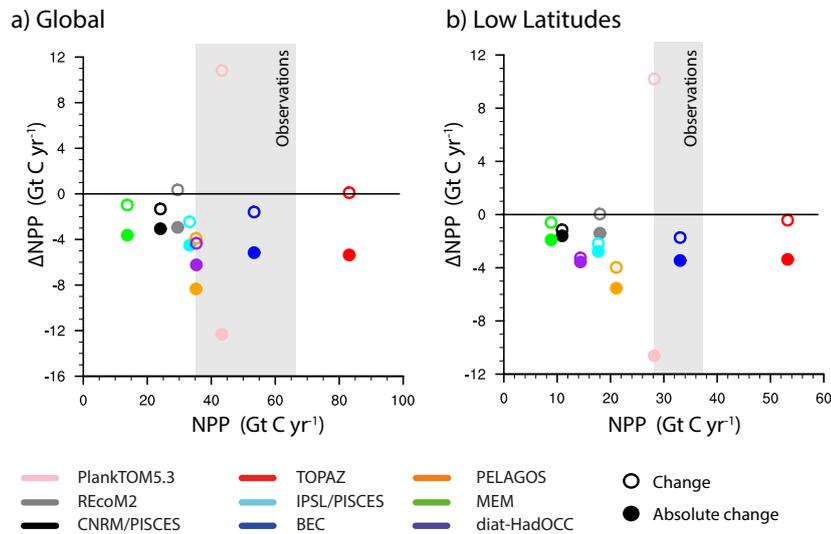


Fig. 13: Relationship between the change in NPP and the 2012-2031 average NPP for all models. Change in NPP has been calculated as the sum of the differences between the 2012-2031 average and the 2081-2100 average for each grid cell (open dots). We additionally show the negative absolute differences of the changes (full dots), calculated by taking the sum of the negative absolute differences between the 2012-2031 average and the 2081-2100 average for each grid cell. Each color represents a model, panel a) shows global values and panel b) shows the low latitudes. The gray area marks the range of current observational NPP estimates. For global values we show the observed NPP range as reported by Carr et al. (2006), for the low latitudes we give the observed NPP range spanned by the estimates of Behrenfeld and Falkowski (1997) and Westberry et al. (2008).

Table 9: Overview of model simulations used in this work

Earth System Model	Reference	ocean model	Ecosystem model	Reference	Spin-up (years, offline + online)	Project	Coupling
HadGEM2-ES	Collins et al. (2011), HadGEM Team et al. (2011)	MetUM	diat-HadOCC	Totterdell (2013)	CMIP3 + 500 + 100	CMIP5	fully coupled
CESM1	Hurrell et al. (2013), Lindsay et al. (2014)	POP	BEC	Moore et al. (2013b)	1025 + 150	MAREMIP	fully coupled
GFDL-ESM2M	Dunne et al. (2012, 2013)	MOM	TOPAZ	Dunne et al. (2013)	1 + 1000	MAREMIP	fully coupled
IPSL-CM5A-LR	Dufresne et al. (2013)	NEMO	PISCES	Aumont and Bopp (2006)	3000 + 300	CMIP5	fully coupled
CNRM-CM5	Voldoire et al. (2012)	NEMO	PISCES	Aumont and Bopp (2006) ¹	3000 + 300	CMIP5	fully coupled
MIROC5	Watanabe et al. (2011)	MRI.COM	MEM	Shigemitsu et al. (2012)	1245 + 480	MAREMIP	ocean only
CMCC-CESM	Vichi et al. (2011a) Cagnazzo et al. (2013)	NEMO	PELAGOS	Vichi et al. (2007)	1 + 450 ²	CMIP5	fully coupled
MIROC5	Watanabe et al. (2011)	MITgcm	REcoM2	Hauck et al. (2013)	0 + 112	MAREMIP	ocean only
IPSL-CM5A-LR	Dufresne et al. (2013)	NEMO	PlankTOM5.3	Buitenhuis et al. (2013a)	0 + 6	MAREMIP	ocean only

1) For differences between the two PISCES simulations see Séférian et al. (2013)

2) Land and ocean carbon pools have been adjusted to the atmospheric preindustrial CO₂ with an acceleration method described in Vichi et al. (2011a)

Table 10: Overview of ecosystem models used in this work, extended from Bopp et al. (2013)

Ecosystem model	Nutrients	Phytoplankton types	Zooplankton types	Redfield/Variable Stoichiometry
diat-HadOCC	4 (NO ₃ , NH ₄ , SiOH ₄ , Fe)	2 (diatom, non-diatom); implicit calcification	1	R(C:N), V(Si, Fe)
REcoM2	3 (NO ₃ , SiOH ₄ , Fe)	2 (diatom, nano-, implicit calcification)	1	V(C,N,Si,Chl), (C:Fe) fix
BEC	5 (NO ₃ , NH ₄ , PO ₄ , SiOH ₄ , Fe)	3 (diatom, nano-, diazotroph, implicit calcification)	1	R(C:N:P), V(Si,Chl,Fe)
TOPAZ	5 (NO ₃ , NH ₄ , PO ₄ , SiOH ₄ , Fe)	3 (large separated into diatoms and other eukaryotes, nano-, diazotrophs, implicit calcification)	(implicit)	R(C:N), V(P, Si, Chl, Fe)
PISCES	5 (NO ₃ , NH ₄ , PO ₄ , SiOH ₄ , Fe)	2 (diatom, nano-, implicit calcification)	2 (micro- and mesozooplankton)	R(C:N:P), V(Si, Chl, Fe)
MEM	4 (NO ₃ , NH ₄ , SiOH ₄ , Fe)	2 (diatom, nanophytoplankton)	3 (micro-, meso-, predatory zooplankton)	R(C:N:P), Chl, Si, Fe fix
PELAGOS	5 (NO ₃ , NH ₄ , PO ₄ , SiOH ₄ , Fe)	3 (diatoms, flagellates, picophytoplankton)	3 (micro-, mesozooplankton, heterotrophic nanoflagellates)	V(N,P,Si,Chl,Fe)
PlankTOM5.3	3 (NO ₃ , SiOH ₄ , Fe)	3 (diatoms, nanophytoplankton, coccolithophores)	2 (micro-, mesozooplankton)	R(C:N), V(Si, Chl, Fe)

Table 11: Comparison of temperature limitations in ecosystem models

Ecosystem model	Q_{10}
Diat-HadOCC	None
REcoM2	15 to 25°C: 1.69, 0 to 10°C:1.79
BEC	2.0
TOPAZ	1.87
PISCES	1.89, mesozooplankton: 2.14
MEM	2.0
PELAGOS	2.0, mesozooplankton: 3.0
PlankTOM5.3	cocco: 1.68, diatoms: 1.93, nano: 2.08 micro: 1.77, meso: 1.71

Table 12: Comparison of nutrient limitation of phytoplankton growth in ecosystem models

Ecosystem model	Nutrient limitation
Diat-HadOCC	Michaelis-Menten, multiplicative iron limitation
REcoM2	Cell quota, Fe with Michaelis-Menten
BEC	Michaelis-Menten
TOPAZ	Cell quota
PISCES	Michaelis-Menten
MEM	Optimal uptake kinetics
PELAGOS	Multiplicative, cell quota, included in exudation term
PlankTOM5.3	Fe with cell quota, Si, NO ₃ Michaelis-Menten and μ_{\max} depends on Fe and Chl cell quota

Ecosystem model	Prey dependence
Diat-HadOCC	Holling Type II
REcoM2	Holling Type III
BEC	Holling Type III
TOPAZ	implicit zoo, see Appendix
PISCES	Holling Type II
MEM	Ivlev
PELAGOS	Holling Type II
PlankTOM5.3	Holling Type II

Table 13: Comparison of prey dependence of grazing. For the full equations see Appendix.

Table 14: Model skill in representing global NPP, measured in Spearman’s rank correlation, normalized standard deviation (NStdDev) and bias. The NPP data is from Westberry et al. (2008), the average global NPP value is $12.6 \text{ molC m}^{-2} \text{ yr}^{-1}$. The chlorophyll data is from the SeaWiFS Project, the average global chlorophyll value is $0.28 \text{ mg Chl m}^{-3}$.

Model simulation	Correlation	NStdDev	Bias
Integrated NPP (bias in [molC/m2/yr])			
Diat-HadOCC	0.18	1.20	-4.5
REcoM2	0.33	0.84	-6.54
BEC	0.67	0.95	-0.56
TOPAZ	0.69	1.49	6.80
CNRM/PISCES	0.09	0.78	-7.94
IPSL/PISCES	0.39	0.80	-4.65
MEM	0.49	1.14	-8.87
PELAGOS	0.40	1.04	-4.47
PlankTOM5.3	0.54	0.92	-0.47
Surface chlorophyll (bias in [mg Chl/m3])			
Diat-HadOCC	0.52	0.62	0.61
REcoM2	0.62	0.19	-0.02
BEC	0.66	0.40	0.01
TOPAZ	0.72	0.14	0.04
CNRM/PISCES	0.58	0.23	-0.13
IPSL/PISCES	0.54	0.19	-0.09
MEM	0.58	0.14	-0.03
PELAGOS	0.36	0.44	-0.02
PlankTOM5.3	0.50	1.35	2.69

Table 15: Model skill in representing surface nutrients, measured in Pearson correlation, normalized standard deviation (NStdDev) and bias. Nutrient data from Garcia et al. (2014). The average global values for NO_3 , SiOH_4 and PO_4 are $6.7 \text{ mmol NO}_3 \text{ m}^{-3}$, $10.6 \text{ mmol SiOH}_4 \text{ m}^{-3}$, and $0.66 \text{ mmol PO}_4 \text{ m}^{-3}$, respectively.

Model	Correlation	NStdDev	Bias
Surface NO_3 (bias in [mmol N/m ³])			
Diat-HadOCC	0.83	1.01	-0.51
REcoM2	0.67	0.86	3.60
BEC	0.84	0.91	0.23
TOPAZ	0.83	0.99	1.43
CNRM/PISCES	0.62	1.10	4.89
IPSL/PISCES	0.83	0.91	-0.69
MEM	0.84	1.10	0.82
PELAGOS	0.72	0.19	-4.24
PlankTOM5.3	0.85	1.01	3.23
Surface SiOH_4 (bias in [mmol Si/m ³])			
Diat-HadOCC	0.45	0.83	45.11
REcoM2	0.56	0.62	-5.24
BEC	0.61	0.75	-0.17
TOPAZ	0.62	1.36	4.63
CNRM/PISCES	0.66	0.60	-0.97
IPSL/PISCES	0.50	1.01	2.75
MEM	0.76	1.47	6.58
PELAGOS			
PlankTOM5.3	0.51	0.85	-6.87
Surface PO_4 (bias in [mmol P/m ³])			
BEC	0.87	0.93	0.03
TOPAZ	0.83	0.99	-0.10
CNRM/PISCES	0.82	0.99	-0.32
IPSL/PISCES	0.85	1.05	-0.17
PELAGOS	0.77	13.51	5.20

Table 16: NPP changes (total and in % of the global value) in different regions. The Pacific upwelling region is shown in Fig. 3. Changes describe the difference between the 2012-2031 and the 2081-2100 average.

Region	Area in %	NPP in %	multimodel median	Change (Gt C yr ⁻¹)	multimodel median change (Gt C yr ⁻¹)
global	100	100	100	-4.3 - +10	-0.2
low latitudes	53	40 - 65	58	-3.9 - +9.9	-0.3
Pacific upwelling	15	14 - 33	20	-2.2 - +2.3	-0.36
S. Ocean (<50S)	12	6.5 - 19	9	-0.01 - +0.4	+0.24
S. Intermediate (30 - 50S)	18	13 - 27	18	-0.7 - +0.27	-0.01
N. Hemisphere (30 - 90N)	16	11 - 17	14	-0.6 - +0.39	-0.1