Biogeosciences Discuss., 6, C2808–C2810, 2009 www.biogeosciences-discuss.net/6/C2808/2009/ © Author(s) 2009. This work is distributed under the Creative Commons Attribute 3.0 License.



## *Interactive comment on* "The sensitivity of microbial processes in Icelandic soils to increasing temperatures" by R. Guicharnaud et al.

## R. Guicharnaud et al.

rannveig@lbhi.is

Received and published: 2 November 2009

Dear referee #2. We would like to thank for showing interest to our research paper and are thankful for many constructive comments.

We agree with the author that the temperature is the main factor affecting microbial activity. However it is plausible that DOC is the controlling substrate since the highest DOC concentrations are observed at -10°C when the respiration is lowest. Hence lower activity due to low temperature results in highest concentrations of DOC. This sentence will be rephrased in the manuscript for increased clarity and reinforce that rightfully temperature is the main driver.

As per comment 2, we got a similar critic from reviewer 1 and have hence decided

C2808

to remove the sentence from the abstract stating that results are of importance to understand global soil dynamics.

As per comment 3, regarding description of sampling regiments. Before all replicate subsamples were placed in incubators soil was weighted for each measurements and placed in individual containers making them ready for analyses as soon as they where taking out of incubators for analyses. For example, 25 g (which is the amount of soil needed for that particular measurement) of sieved soils (soil where prepared at 4°C to minimise biological activity within samples) was placed in a container before incubation and hence was ready for extractions without affecting the soil microstructure after 2 weeks incubations. This was done for all soils and each measurement conducted in this current study.

Concerning soil bulking, the authors do not agree with comments concerning soil bulking limiting the value of the current study. There are a few reasons for this. Soil bulking is a recognized sampling method and involves sampling cores and bulking into one sample for representing a given area (Parkin and Robinson, 1992; Morton et al., 2000). On the other hand, when studying spatial variability in biological measurements within a given field soil bulking would not be recommended (Officer et al., 2006) as pointed out by the referee and the authors agree. However, the aim of this research paper was not to assess spatial variability within a given field but the direct effect of temperature on soil microbial processes with the aid of soil microcosms in a controlled laboratory study. Microcosms are miniature enclosed ecosystems which allow predetermined environmental factors and treatments to be measured in replicated, controlled conditions (Kampichler et al., 2001) as well as reducing environmental variables and allow predetermined environmental factors (Kampichler et al., 2001) which in this case was temperature. Including spatial variability from each site of this study was therefore believed at add environmental factors not relevant to this study. A future research however studying site specific factors on soils temperature respond would be of great value and interest. The authors have conducted a study investigating the suitability of different sampling methods as well as the spatial variability of biological factors of Icelandic soils within a given field area, which has been submitted for publication elsewhere.

About the sampling size. The authors have tested many different sampling sizes using up to 1 kg of field moist soil as a sample size and gone down to soil cores only 2.5 cm in diameter. In all cases however the variability has always great likely reflecting the high heterogeneity associated with soils.

Technical comments (line 675 line 16 excetera) have all be dealt with accordingly.

References Parkin, T.B., Robinson, J.A., 1992. Analysis of lognormal data. Advances in Soil Science 20, 193-235.

Morton, J.D., Baird, D.B., Manning, M.J., 2000. A soil sampling protocol to minimise the spatial variability in soil test values in new zealand hill country. New Zealand Journal of Agricultural Research 43, 367-375.

Officer, S.J., Tillman, R.W., Palmer, A.S., 2006. Plant available potassium in new zealand steep-land pasture soils. Geoderma 133, 408-420.

Kampichler, C., Bruckner, A., Kandeler, E., 2001. Use of enclosed model ecosystems in soil ecology: A bias towards laboratory research. Soil Biology and Biochemistry 33, 269-275.

Interactive comment on Biogeosciences Discuss., 6, 6749, 2009.

C2810