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**Effects of dust  
deposition on iron  
cycle in the surface  
Mediterranean Sea**

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# Effects of dust deposition on iron cycle in the surface Mediterranean Sea: results from a mesocosm seeding experiment

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## Abstract

Soil dust deposition is recognized as a major source of iron to the open ocean at global and regional scales. However, the processes that control the speciation and cycle of iron in the surface ocean after dust deposition are poorly documented mainly due to the logistical difficulties to investigate in-situ, natural dust events. The development of clean mesocosms in the frame of the DUNE project (a DUst experiment in a low Nutrient low chlorophyll Ecosystem) was a unique opportunity to investigate these processes at the unexplored scale of one dust deposition event. During the DUNE1 mesocosm seeding experiment, iron stocks (dissolved and particulate concentrations in the water column) and fluxes (export of particulate iron in sediment traps) were followed during 8 days after an artificial dust seeding mimicking a wet deposition of  $10 \text{ g m}^{-2}$ . The addition of dust at the surface of the mesocosms was immediately followed by a decrease of dissolved iron [dFe] concentration in the 0–10 m water column. This decrease was likely due to dFe scavenging on settling dust particles and mineral organic aggregates. The scavenging ratio of dissolved iron on dust particles averaged  $0.37 \pm 0.12 \text{ nmol mg}^{-1}$ . Batch dissolution experiments conducted in parallel to the mesocosm experiment showed a increase (up to 600%) in dust iron dissolution capacity in dust-fertilized waters compared to control conditions. This study gives evidences of complex and unexpected effects of dust deposition on surface ocean biogeochemistry: (1) large dust deposition events may be a sink for surface ocean dissolved iron and (2) successive dust deposition events may induce different biogeochemical responses in the surface ocean.

## 1 Introduction

Deposition of atmospheric particles at the ocean atmosphere interface constitutes a flux of chemical elements to the surface ocean. When excluding sea-salt particles, most of the mass flux of atmospheric particles is made of fine mineral particles from

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lithogenic origin (Andreae, 1996). These particles are emitted from arid or semi-arid areas and transported over long distances through atmospheric circulation. The elemental composition of the mineral particles that deposit at the surface ocean is determined by the elemental composition of the continental source area (Claquin et al., 1999; Guieu et al., 2002a) and the chemical processes that affect the particles during transport (Desboeufs et al., 2003; Shi et al., 2009). As the erodible continental crust contains significant amounts of biogeochemically significant elements (Wedepohl 1995), dust deposition is recognized as a significant source of micro - and macro nutrients to the surface ocean (Jickells et al., 2005, Mahowald et al., 2008, Duce et al., 2009).

As often highlighted by impressive satellite pictures (NASA EO, 2010), large dust deposition events are episodic phenomena. This important temporal variability in dust deposition hinders the assessment of the time scales needed to understand the response of the surface ocean (Aumont et al., 2008, Volpe et al 2009). The occurrence of a large dust deposition event can be conceptually investigated as a chemical perturbation to the oceanic system and the episodic nature of the phenomena may certainly lead to non linear biogeochemical responses. Due to the logistical difficulties to investigate the response to a dust pulse at the scale of an event, in situ studies of the processes occurring at this small time scales are quasi undocumented, despite the fact that the iron cycle has received a considerable impetus during the last two decades, due to its role in controlling marine productivity in HNLC areas (Boyd et al., 2007) and to its (co)-limiting role in controlling marine nitrogen fixer organisms in oligotrophic areas (Mills et al., 2004). Moreover, a large scientific debate has taken place on the role of atmospheric dust iron deposition to the surface ocean on the climate variability at millennium time scales (Martin et al., 1990, Watson et al., 2000; Bopp et al., 2003). However, the underlying physico-chemical processes that govern iron cycling at the ocean-atmosphere interface are still poorly understood (Baker and Croot, 2008). Laboratory experiments emphasize the release of iron when dust is in contact with seawater mainly controlled by dissolution processes. However, several studies

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present evidence of inverse processes limiting the dissolved iron concentrations, as re-precipitation (notably for high dust concentrations) (Spokes and Jickels, 1995; Bonnet and Guieu, 2004) or iron adsorption on particles (Zhuang and Duce, 1993).

The project DUNE (a DUst experiment in a low Nutrient low chlorophyll Ecosystem) aims at better understanding the effect of dust deposition on the biogeochemistry of surface waters of the Mediterranean Sea (Guieu et al., 2010). The approach applied in this project was to perform dust addition experiments onto large clean mesocosms. The original design of these mesocosms represented a unique opportunity to study the iron cycle at a scale which has been poorly explored so far. The main objective of this study is to quantify dissolution and adsorption processes of dissolved iron that occurs from (or at) mineral particles surface. These processes modulate the net effect of the input of atmospheric particles as a source or a sink of bioavailable iron to the surface ocean ecosystem.

## 2 Material and methods

### 2.1 Experiment design

The mesocosm experiment design and the accuracy of the strategy developed in the frame of the DUNE project is described in detail in Guieu et al., 2010. In summary, the deployed mesocosms were cylindrical with a diameter of 2.3 m and a volume of 52 m<sup>3</sup>. A system of permanent PVC tubing allowed to sample at three different depths (−0.1, −5 and −10 m) by connecting a Teflon pump. The bottom of the mesocosms was conic and a sediment trap was adapted at the apex in order to collect the exported material. Mesocosms were covered in order to avoid possible additional inputs from natural dust events. The covers were designed in order to let the maximum light reach the water body inside the mesocosm and to allow gas exchanges.

In June 2008, six mesocosms entirely designed in plastic were deployed in the Elbo bay (Scandola Marine preservation area – 8.554° E, 42.374° N), a site characterized by

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oligotrophic conditions in summer. The DUNE-1-P seeding experiment was conducted with the following protocol: after deployment and closing of the mesocosms, sampling was performed to determine the initial conditions. Three mesocosms (D1, D2 and D3, hereafter referred as “DUST-Meso”) were amended with 41.5 g of dust each with a trace metal clean spray. The addition was made with a processed dust (Al:  $4.12 \pm 0.39\%$  mass. and Fe:  $2.31 \pm 0.04\%$  mass.) diluted in 2 liters of ultrapure water in order to mimic a wet deposition event (see Guieu et al., 2010 for details). Dust was obtained through a physico-chemical treatment of fine particles mechanically produced from a dust source areas in southern Tunisia and further processed in the laboratory in order to mimic aging of dust particles by cloud cycling (see Guieu et al., 2010 for details). Three other mesocosms (C1, C2 and C3, hereafter referred as “CONTROL-Meso”) were kept unamended. The time of the dust addition corresponds to the theoretical start of the experiment ( $t_0$ ). Sampling for most of the parameters was performed at a daily scale during 8 days. Every 48 h one cast (at 0.1, 5 and 10 m) was additionally made outside the mesocosms and the sediment traps were recovered and replaced.

All material used for this study was cleaned following trace-metal clean procedures (Bruland et al., 1979) and for analytical work, all manipulations took place under a class 100 laminar flow bench.

## 2.2 Dissolved iron concentration analysis

Samples for dissolved iron concentration [dFe] were filtered during sampling through a  $0.2 \mu\text{m}$  cartridge filter (Sartobran, Sartorius, Germany) adapted on the pumping system and collected in 60 mL polyethylene bottles. Samples were acidified to  $\text{pH} < 2$  with HCl (Merck, Ultrapur, Germany) directly after collection and were analyzed after at least 24 h. [dFe] was analyzed by flow injection with online preconcentration and chemiluminescence detection following exactly the same protocol, instrument and analytical parameters as described in Bonnet and Guieu (2006). Detection limit (DL) was 10 pM and blanks were around 50 pM. An internal acidified seawater standard was measured

every day in order to control the stability of the analysis. The reliability of the method was controlled by analyzing the D2 SAFE seawater standard (Johnson et al., 2007).

### 2.3 Particulate iron [pFe] and aluminium [pAl] concentration analysis in the water column

5 In order to follow the settling of the added mineral particles through the mesocosms, particulate iron [dFe] and aluminium [pAl] were measured during the experiment. An “easy to run” protocol was preferred over common protocols designed to reach low detection limits and requiring larger volumes of filtered water. Samples were collected on 47 mm cellulose acetate filters (Sartorius) previously washed with diluted HCl and  
10 rinsed thoroughly with ultrapure water. Filters were placed in polypropylene filter holders connected to the Teflon pump directly from the mesocosms. For each sample one liter of seawater was filtered. After collection, filters were dried under a laminar flow bench and kept at room temperature until analysis. One half of the collected filters were HNO<sub>3</sub>/HF acid-digested using Suprapur reagents at 150 °C in PTFE vials. After  
15 complete evaporation, samples were diluted in 0.1 M HNO<sub>3</sub> and analyzed for iron and aluminium concentrations by ICP-AES (Jobin Yvon – JY 138 “Ultrace”). A certified marine sediment reference material (GBW from NRCCRM China) was digested following the same protocols to test the reliability of the method (Recovery for Al = 94–96%, for Fe = 99–101%). Reagent blanks and filter blanks were included as control  
20 for possible contamination during the analytical process. Blank values were under the detection limit (3.5 ppb for Fe and 8 ppb for Al corresponding to a detection limit for [pFe] = 0.10 µg L<sup>-1</sup> and [pAl] = 0.24 µg L<sup>-1</sup> in the seawater with the protocol used).

### 2.4 Aluminium [pAl] and iron [dFe] measurements in the sediment trap

The samples collected in the sediment trap were treated following the standard protocol  
25 developed at the national service “Cellule Piegé” of the French INSU-CNRS. Details for this protocol can be found in Guieu et al. (2005). Aluminium and iron were measured on

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dried aliquots of the collected material with the same instrument and the same protocol than described above for particulate iron [pFe] and aluminium [pAl] measurements in seawater.

## 2.5 Dust dissolution experiments during the DUNE-1-P experiment

5 At four selected times during the DUNE-1-P experiment, batch dust dissolution experiments were performed on filtered ( $<0.2\ \mu\text{m}$ ) seawater collected in the mesocosms at 5 m. The aim of this additional “bottle” experiment was to investigate the effect of dust fertilization on the dissolution of additional dust inputs. The protocol used for this dissolution experiments is described in Wagener et al. (2008). Briefly, the same type of  
10 dust as used for seeding the mesocosms was added to 200 mL of filtered seawater with a final concentration of  $5\ \text{mg}\ \text{L}^{-1}$ . Dissolved iron concentration [dFe] was determined after 72 h. As discussed in Wagener et al. (2008), this simple protocol, even if subject to adsorption issues on batch reactor walls, is valuable to assess solubility variations.

## 3 Results

15 All data collected in this study were added to a database which is available as online supplementary information (<http://www.biogeosciences-discuss.net/7/2799/2010/bgd-7-2799-2010-supplement.zip>). All samples were labeled with an incremented cast number, the mesocosm name and the depth of collection. For all samples, time is “tx”, where x are hours since the t0 reference fertilization time.

### 20 3.1 Evolution of iron stocks in the mesocosms

The evolution of [dFe] in the mesocosms is reported on Fig. 1. The initial [dFe] averaged  $2.5\ \text{nM}$  between surface and 10 m. The main trend observed during the eight days of the experiment is consistent among the triplicate mesocosms. In the dust-amended mesocosms (“DUST-Meso”: D1, D2 and D3), [dFe] decreased by  $0.7\ \text{nM}$  6 h after dust addition whereas in the control mesocosms (“CONTROL-Meso”: C1, C2  
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and C3) [dFe] remained constant at 2.5 nM during the experiment with an increase (reaching 3.2 nM for one mesocosm) at 10 meters at the end of the experiment. Values from outside the mesocosms are reported in the supplementary information (<http://www.biogeosciences-discuss.net/7/2799/2010/bgd-7-2799-2010-supplement.zip>) as “OUT” and showed no significant differences with values in “CONTROL-Meso”. A statistical evaluation of the reproducibility of triplicates mesocosms and the coherence between CONTROL-Meso and OUT can be found in Guieu et al. (2010).

The evolution of [pFe] in DUST-Meso is reported on Fig. 2. [pFe] was below  $1 \mu\text{g L}^{-1}$  before the dust addition. Six hours after the dust addition, [pFe] increased to values up to 40 to  $60 \mu\text{g L}^{-1}$  at the surface of the mesocosms. This increase was still detectable after 24 h at 5 and 10 m. A fast decrease in the following three days was observed at all depths. From day four, [pFe] in DUST-Meso remained constant with values slightly higher to those before dust addition. The evolution of [pFe] in CONTROL-Meso was only measured at 5 meters and is reported in the database (supplementary information, <http://www.biogeosciences-discuss.net/7/2799/2010/bgd-7-2799-2010-supplement.zip>). Values were low ( $<1 \mu\text{g L}^{-1}$  or below the detection limit) and constant for the entire duration of the experiment. Variability among the replicates was more important in [pFe] than in [dFe]. The low volume of filtered water (1 liter) is likely too low to integrate the small scale variability in the particulate distribution within a mesocosm.

### 3.2 Particulate iron in the sediment traps

Data on particulate iron that was exported down to the sediment trap of the mesocosms are reported on Table 1. The mass of particulate iron [dFe] per sample and the cumulative mass of pFe for the entire experiment are reported. In the CONTROL-Meso, mass of pFe exported ranged from 0.7 to 8.5 mg with cumulative masses at the end of experiment ranging from 8.3 to 11.5 mg. For DUST-Meso, the mass of pFe collected into a single sample was as high as 340 mg with cumulative masses over the entire experiment ranging from 434 to 538 mg.

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Daily pFe fluxes calculated from [pFe] from each trap ranged between 0.08 and 1.02 mg m<sup>-2</sup> day<sup>-1</sup> for CONTROL-Meso and between 1.46 and 40.4 mg m<sup>-2</sup> day<sup>-1</sup> for DUST-Meso. The highest fluxes for DUST-Meso were recorded during the first 72 h of the experiment. After 120 h, the fluxes in DUST-Meso decreased by one order of magnitude but were still higher than in the CONTROL-Meso. When integrated over the entire duration of the experiment, pFe fluxes were ca. 50 times higher in DUST-Meso (15–18.7 mg m<sup>-2</sup> day<sup>-1</sup>) than in CONTROL-Meso (0.29–0.55 mg m<sup>-2</sup> day<sup>-1</sup>).

### 3.3 Aluminium and iron in the particulate matter

Particulate aluminium concentration [pAl] in the water column and the sediment traps were measured jointly with iron (data available in the database – <http://www.biogeosciences-discuss.net/7/2799/2010/bgd-7-2799-2010-supplement.zip>). The relationship between [pAl] and [pFe] in the water column and in the sediment traps are reported in Fig. 3. [pAl] and [pFe] were significantly correlated both in the water column (slope of 0.44, intercept close to 0,  $n = 68$ ,  $p < 0.001$ ) and in the sediment traps (slope of 0.58, intercept close to 0,  $n = 23$ ,  $p < 0.001$ ). The slopes of both correlations are significantly different. However if the four sample values with [pAl] > 30 µg L<sup>-1</sup> are discarded in the water column dataset, the slope (Fe/Al ratio) is then of 0.53 and hardly differs from the ratio in the sediment trap. The Fe/Al ratio of the dust introduced for the fertilization is  $0.56 \pm 0.06$  (Guieu et al., 2010.).

### 3.4 Dissolution of dust iron in batch experiments

Results of the batch dissolution experiments are presented on Fig. 4. Dissolution at select times of the fertilization is expressed as  $\Delta[\text{dFe}]_{72\text{h}}$  which represents the difference between [dFe] measured after 72 h of contact time between filtered seawater and dust particles in the experimental bottles and [dFe] measured before the addition of dust particles to the batch reactor. This initial [dFe] corresponds to [dFe] measured in the mesocosms at the sampling time of the filtered seawater. A good reproducibility

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was obtained among the replicate mesocosms with coefficient variations ranging from 9 to 22% for  $\Delta[\text{dFe}]_{72\text{h}} > 0.1 \text{ nM}$ .

$\Delta[\text{dFe}]_{72\text{h}}$  remained close to 0 when using water from CONTROL-Meso whereas it reached values between 0.5 and 1 nM when using water from DUST-Meso for the first 120 h after the dust seeding in the mesocosm. Dissolution ( $\Delta[\text{dFe}]_{72\text{h}}$ ) increased slightly in water collected from CONTROL-Meso (0.5 nM) 168 h after the dust seeding in the mesocosm and it increased to 3 nM with water collected in DUST-meso at the same time step (t168).

## 4 Discussion

### 4.1 Initial iron concentration and iron budget during the DUNE-1-P experiment

Dissolved iron concentration ( $[\text{dFe}]$ ) measured outside the mesocosms and inside before the dust addition (2.5 nM) are in the highest range of former measurements at the DYFAMED time series station (43°25' N, 07°52' E) in the western Mediterranean Sea (Sarhou and Jeandel, 2001; Guieu et al., 2002b; Bonnet and Guieu, 2006). Two reasons can explain these higher  $[\text{dFe}]$  values:

1. Saharan dust deposition event associated with dry deposition occurred in the NW Mediterranean Sea two weeks before the beginning of the experiment (26 to 28 May 2008). Associated with this event, a dust flux of  $382 \text{ mg m}^{-2}$  was recorded at Galeria (Corsica, France), close to the mesocosm deployment site (K Desboeufs, personal communication, 2008). This dust deposition event could have increased  $[\text{dFe}]$  in surface stratified waters as has been described by Bonnet and Guieu, 2006 and Sedwick et al., 2005.
2. Rain events occurred twenty to ten days before the start of the experiment (K. Desboeufs, personal communication, 2008) and iron could have been delivered to the study area through surface runoff from surrounding lands. This last

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point would indicate that even if the study site has biogeochemical features typical of open ocean, it may punctually be submitted to coastal influences.

Particulate iron [pFe] before the dust addition or in CONTROL-Meso were in the range of values reported for the surface waters of the NW Mediterranean Sea (Sarthou and Jeandel, 2001) and was likely dominated by lithogenic material. Indeed, even if the cellular content of iron in marine microorganisms is highly variable (e.g. Veldhuis et al., 2005), based on a biogenic Fe/C ratio of 400  $\mu\text{mol/mol-C}$  (highest range of observations), the biogenic Fe derived from particulate organic carbon (data not shown) in the mesocosms would represent 1 to 10% of the measured [pFe] before addition of dust. The clear dominance of iron from lithogenic origin is confirmed by the crustal signal (0.53) of the [pFe]/[pAl] ratio.

All iron reservoirs were measured during the whole duration of the experiment in mesocosms D1 and D2 allowing to construct an iron budget in those two mesocosms (Table 2). The possibility to construct a mass budget based on iron during the experiment may be relevant in order to assess uncertainties in fluxes estimation during the experiment. At each time  $t$ , the iron budget for the mesocosms is the sum of four terms:

1. The “input” term corresponds to the amount of iron added through a dust addition of 41.5 g. The seeding is considered as the only input of iron for the entire experiments, as the mesocosm wall isolates the water mass from further inputs and the mesocosms were covered to avoid any possible additional natural atmospheric deposition.
2. The DFe ( $<0.2 \mu\text{m}$ ) and PFe ( $>0.2 \mu\text{m}$ ) stock in the water column corresponds to the integrated amount of dFe and pFe determined in the mesocosms from [dFe] and [pFe] measurements (See Table 2 for calculations).
3. The “export” term corresponds to the amount of iron collected in the sediment trap.

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It is assumed that the inventory of iron at  $t_0$  is the sum of the iron added by the dust seeding and the stock of iron initially present in the mesocosms in the dissolved ( $<0.2\ \mu\text{m}$ ) and particulate phase ( $>0.2\ \mu\text{m}$ ). After fertilization, the recovery is the percentage of the fraction of iron present at  $t_0$  that is recovered with the different measurements ( $[\text{dFe}]$ ,  $[\text{pFe}]$  and  $\text{pFe}$  in the sediment trap).

The iron budget over the duration of the experiment could not be balanced with the initial  $t_0$  budget: at most, ca. 60% was recovered. Several, non exclusive reasons, can explain this unbalance. First, the stock of  $[\text{pFe}]$  was calculated from discrete measurements at three depths and extrapolated to the whole water column which can lead to important errors. A layer of high dust concentration could be missed between the sampling depths and not be accounted. For instance, this could have happened in the first days of the experiment in the upper part of the mesocosm, or later in the bottom cone. This would underestimate the  $\text{pFe}$  stocks. Besides this interpolation artifact, other reasons related to the design of the mesocosms may explain a part of this unbalanced budget. Adsorption of dissolved iron on plastic surfaces has been demonstrated (Fisher et al., 2007). However, even if the polyethylene (PE) walls of the mesocosms would have absorb half of the dissolved iron in the mesocosms (which has been demonstrated for small PE bottles (Fisher et al., 2007) this, still, only explains a negligible loss of iron. The  $\text{dFe}$  stock is only a small part (less than 1%) of the iron budget (See Table 2). Mesocosm walls represent also an important surface of material where some particulate material could have stick. A visual inspection of the mesocosms at the end of the experiment did not allow establishing a significant loss by particle retention on the vertical walls of the mesocosms, but as described in Guieu et al. (2010), a fraction of the particles could have get lost during the trap changing or have get stuck inside the bottom of the cone.

## 4.2 Impact of the dust addition on dissolved iron concentration

Even if a large debate still exists on the actual bioavailability of different iron species in seawater (e.g. Sunda, 2001), assessing iron dissolution from dust particles through

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[dFe] ( $<0.2 \mu\text{m}$ ) measurements can be assumed to be an indicator of the amount of dust iron that may be available for the biota. Given the importance of dust deposition as a source of iron to the open ocean at a global scale, numerous studies have focused on the release of iron from dust particles in seawater (e.g. Mahowald et al., 2009). The common assumption of these works is that dust releases iron to the dissolved pool once deposited at the sea surface. However, during the DUNE-1-P experiment, after the dust addition, no dissolution of iron from dust particles could be evidenced. On the contrary, the dust addition was immediately followed by a decrease (between 0.7 to 1 nM) of [dFe] (Fig. 5a).

This decrease would correspond to a net sink of up to  $4 \mu\text{mol m}^{-2}$  of dissolved iron in the first 48 h of the experiment (Fig. 5b). This sink of dFe could be due to enhanced biological activity since the addition of dust stimulated the biological activity in the mesocosms (Guieu et al., 2009b). Chlorophyll concentration increased in all “dust” mesocosms from ca  $0.10 \mu\text{g L}^{-1}$  to  $0.25 \mu\text{g L}^{-1}$  within 48 h (Guieu et al., 2009b), and bacterial abundance increased from  $0.5 \cdot 10^6$  to  $1 \cdot 10^6 \text{ cell mL}^{-1}$  (Pulido-Villena et al., 2010.). Although iron consumption by microorganisms was not directly assessed during the experiment, literature values of iron content in cells can be used as estimates. The phytoplankton bloom observed during the experiment could have induced a maximum iron consumption of ca.  $500 \text{ nmol m}^{-2} \text{ d}^{-1}$  based on the highest C/Chla and Fe/C ratio reported by Veldhuis et al. (2005). Concerning heterotrophic bacteria, the maximum iron consumption would be  $55 \text{ nmol m}^{-2} \text{ d}^{-1}$  based on the highest Fe/C ratio reported by Tortell et al. (1996). The biological consumption of iron could then be roughly assumed to be in a range of  $100\text{--}1000 \text{ nmol m}^{-2} \text{ d}^{-1}$ . In the first hours of the experiment the dissolved iron decrease rate in DUST-meso was up to  $400 \text{ nmol m}^{-2} \text{ h}^{-1}$  ( $9600 \text{ nmol m}^{-2} \text{ d}^{-1}$ ), one order of magnitude higher than this overestimated biological consumption of iron, suggesting that enhanced biological activity cannot explain by itself the observed decrease in dissolved iron.

A more plausible explanation for the observed sink of iron is that dust particles scavenged dissolved iron as they settled in the water column. As pointed out in a recent

review (Baker and Croot, 2008), investigation of direct scavenging on dust particles are very limited. However, scavenging of dissolved iron on settling particles represents the highest potential sink of dissolved iron for the ocean (Johnson et al., 1997). Zhuang and Duce (1993), based on radiolabeled  $^{59}\text{Fe}$  studies, have reported values of iron adsorption of  $27 \text{ nmol mg}^{-1}$  on dust particles. Based on the study of pure hematite phases and radiolabeled  $^{59}\text{Fe}$ , Honeyman and Santschi (1991), reported the importance of the colloidal phase formation on the scavenging of trace metals on particulate iron. The design of the DUNE experiment allowed to study this process by following [dFe] variations during the settling of the dust particles in the first 10 meters of the water column.

In order to demonstrate the scavenging of dFe on settling dust particles in the first 48 hours of the experiment, the following terms were estimated based on average values for CONTROL-meso and DUST-meso: (1) The “loss” of dFe stock (dFe\_stock\_loss in  $\mu\text{m m}^{-2}$ ) was estimated in the 0–5 m and 5–10 m water layers as the difference between the dFe stock in CONTROL-meso and DUST-meso and represents the integrated amount of dFe that disappears after dust seeding. (2) The “loss” of pFe stock (pFe\_stock\_loss in  $\text{mg m}^{-2}$ ) was defined as the amount of pFe introduced by the dust seeding that is lost for the 0–5 m layer and the 5–10 m layer at time  $t$ , based on the basic assumption that dust particles have only a vertical motion to the bottom in the mesocosms. Thus pFe\_stock\_loss was calculated at 5 and 10 m by subtracting the stock of pFe determined by [dFe] measurements between 0–5 m and 0–10 m (assuming that all pFe measured comes from the added particles, supported by results in Sect. 3.2.) to the initial stock of pFe introduced by dust (pFe\_stock\_ini =  $231 \text{ mg m}^{-2}$ ). pFe\_stock\_loss is, theoretically, the stock of pFe introduced by the seeding that has been in contact with the stock of dFe between 0–5 m and 5–10 m and that has crossed the “conceptual” 5 and 10 m horizons while settling. dFe\_stock\_loss and pFe\_stock\_loss are presented on Fig. 5c. (3) pFe\_loss\_flux (in  $\text{mg m}^{-2} \text{ h}^{-1}$ ) and dFe\_loss\_flux (in  $\mu\text{mol m}^{-2} \text{ h}^{-1}$ ) correspond to the stock of dFe and pFe lost per unit of time between the different sampling times (Fig. 5c). It can be noted that the pFe\_loss\_flux at 5 and 10 m should be

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proportional to the average settling velocity of dust particles between two sampling times. As expected, pFe\_loss\_flux decreased between 6 and 48 h. Indeed, the largest particles would settle faster in the first hours and then smaller particles would settle with lower velocities.

5 As illustrated on Fig. 6a, the decrease of dFe stock is correlated to the stock of pFe that settles through the water column after the dust addition. Moreover, the flux of dFe loss throughout the water column is even better correlated to the pFe\_loss\_flux (Fig. 6b). This is a robust indication of the scavenging of dissolved iron onto the particulate matter (particularly dust) that settles to the bottom of the mesocosm after the dust addition.  
10 Those results demonstrate that the dust addition in this experiment was a sink rather than a source of bioavailable iron.

The slope of the linear models on Fig. 6 provides an estimation of the dFe scavenging rate on Saharan dust particles settling in the water column. The estimated values of scavenging are  $17 \pm 7$  and  $15 \pm 6$  nmol mg<sup>-1</sup> for particulate iron. By converting the particulate iron into dust concentration, dFe scavenging would be  $0.39 \pm 0.12$  and  $0.35 \pm 0.12$  nmol mg<sup>-1</sup> of dust. These values are two orders of magnitude lower than the values estimated by Zhuang and Duce, (1993) of 27 nmol mg<sup>-1</sup>. The reasons for these differences will be discussed in Sect. 4.5.

### 4.3 Impact of dust seeding on the solubilisation capacity of seawater

20 The goal of the batch dissolution experiments using filtered seawater collected in the mesocosms at different times after the seeding was to evaluate the impact of dust fertilization on the solubilisation capacity of the seawater. Indeed, the ultimate factor controlling dust solubility in seawater is the capacity of seawater to keep iron in solution through the presence of organic ligands (Liu and Millero, 2002; Baker and Croot, 2008).  
25 In a recent study, Wagener et al., (2008) showed that iron dissolution from dust particles in seawater changes over an annual cycle in the Mediterranean Sea and is controlled by the concentration of dissolved organic carbon with higher dissolution in presence of fresh organic matter in the post bloom period.

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Before the dust addition, the solubility of iron dust particles in filtered seawater was very low or even null (Fig. 4) showing that solubility equilibrium between pFe and dFe was reached.

After dust addition, at  $t = 48\text{h}$  and  $t = 120\text{h}$ , the dissolution of iron from dust was detectable in the DUST-meso batch experiments (with  $\Delta [\text{dFe}]_{72\text{h}}$  of ca. 0.6 nM) whereas no detectable dissolution in CONTROL-meso was observed. Two mechanisms can support this increase in dust iron solubility in DUST-meso after 48 h: (1) the production of specific iron binding ligands by phytoplankton (Boye and Van den Berg, 2000) or heterotrophic bacteria (Gledhill et al., 2004) and (2) the scavenging of colloidal iron on dust particles or the re-precipitation of soluble iron favored by the presence of an important amount solid particle after the dust seeding. The addition of new iron by dust particles could then allow the stabilization of new iron in the form of soluble or colloidal iron which would explain the relative increase of [dFe] in DUST-Meso.

At the end of the experiment ( $t = 168\text{h}$ ), there was a clear increase of the iron solubilisation capacity of DUST-meso seawater. This may be related to the formation of organic binding sites induced by increased biological activity. Former studies have demonstrated an increase of iron binding ligands a few days after an increase of biological productivity (Croot et al., 2001). One possible mechanism responsible for the observed increase in the dust dissolution capacity is the production of siderophores by heterotrophic bacteria. This hypothesis is consistent with the decrease in dFe and with the appearance of a new source of non available iron that followed the dust seeding. This hypothesis is also supported by the observation that the diversity of the bacterial community attached to particles revealed some distinct features in the mesocosms amended with dust at the end of the experiment (Laghdass et al., 2010). The increase of zooplankton biovolume in DUST-meso (Guieu et al., 2009b) could also lead to enhanced organic binding sites through the release of porphyrine-like ligands by increased grazing (Vong et al., 2009).

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#### 4.4 Implication for the understanding of iron dust dissolution in seawater

Two distinct protocols have been used in the last years to assess the solubility of aerosols in seawater: flow through protocols (see for ex. Buck et al., 2006) and batch experiments (see for ex. Bonnet and Guieu, 2004). Although these protocols bring different information on the dissolution process (Baker and Croot, 2008) both conclude that a certain amount of iron dissolves from the dust particles. In this study, the addition of dust particles in the mesocosm did not result in a net release of dissolved iron to seawater but in a sink. However, batch dissolution experiments performed in parallel to the mesocosm experiment, did not demonstrate iron scavenging but dissolution when using the water collected in the DUST-Meso where increase in biological activity was evidenced. This study is therefore pointing out the importance of the protocols used for dust iron dissolution measurements.

Flow through protocols could be seen as appropriate to mimic the dynamical process of particle settling in the water column. However, the contact time between particles and seawater is too short (a few seconds) for the scavenging processes to take place. Even if this protocol has the clear advantage to be easy to handle and to bring information on the control of dust iron solubility by aerosols characteristics (Buck et al., 2006), it has clear limitations in terms of marine biogeochemical perspectives, as the complex equilibrium between adsorption and dissolution that occurs during particle settling in the surface ocean is not taken into account.

Batch experiments could be more appropriate to investigate the processes that occur when atmospheric particles are mixed into the oceanic mixed layer. However, this type of experiments may be subject to another artifact because the contact between particles and seawater is constant, that is, the dynamical nature of the dust deposition of particles is not taken into account by this protocol. This could lead to important adsorption on dust particles as demonstrated by Zhuang and Duce (1993) that would not occur with shorter contact times. Slower dissolution processes, that would not have time to occur in a dynamical process where particles are settling, could also lead to wrong dissolution evaluation by this protocol.

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Although mesocosms studies are not easy to handle, they do better represent the natural conditions. In particular, the parametrization obtained integrates the biological compartment and the biogeochemical characteristics of the large body of water enclosed inside the bags do evolve after the introduction of the particles. This allows to account for dynamical equilibrium that occurs in natural conditions: (1) The fertilization induced by the injection of atmospheric nutrient can lead to production of fresh organic matter prone to complex iron and (2) the settling of particles in the mesocosm allow to represent realistically the scavenging of dissolved iron from the water column. For example, we can hypothesize that the presence of dissolved and particulate organic matter allowed the formation of mineral-organic aggregates which have affected the scavenging of dissolved iron and accelerating its export. This hypothesis agrees with recent observations suggesting that organic-Saharan mineral aggregation results, through ballast effect, in a strong POC export in the water column of the NW Mediterranean Sea (Ternon et al., 2010).

## 5 Implications for the biogeochemical functioning of high dust deposition areas

The DUNE-1-P experiment mimicked a strong but realistic dust deposition event ( $10 \text{ g m}^{-2}$ ). Indeed, several strong dust events recently recorded in NW Mediterranean did bring within few hours dust fluxes higher than  $10 \text{ g m}^{-2}$  (Guieu et al., 2009a; Bonnet and Guieu 2006; Ternon et al., 2010). The classical image of dust deposition releasing nutrients (such as Fe) in the surface ocean, fueling biological productivity and thus increasing the biological carbon pump efficiency (e.g. Cassar et al., 2007), is partly revisited in this study. Dust deposition is more complex than just a source of bioavailable iron to the surface ocean: a large dust deposition event can accelerate the export of iron from the water column through scavenging and can be described with a counter-intuitive “dust cleaning effect”.

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We can speculate that the processes described in this study may only apply in the case of important dust deposition events affecting surface seawater with high dFe concentrations. It has been shown that dissolution of atmospheric particulate iron increases during atmospheric transport (e.g. Baker and Jickels, 2006; Chi et al., 2009) and thus the effect of dust deposition could be more important in areas receiving low dust deposition because of the enhanced atmospheric solubility (Fan et al., 2006). This study presents evidence on the opposite effect: in areas receiving large dust deposition events, scavenging processes may be enhanced. This implies that dissolved iron inputs to the surface ocean are not linearly linked to dust deposition: low dust areas are relatively favored in comparison to high dust deposition areas. So, in areas with high seawater dissolved iron and high dust deposition (i.e. oligotrophic areas P- or N-limited such as the Mediterranean Sea or the Tropical Atlantic), a strong individual dust deposition event could act as a sink for dissolved iron.

This study illustrates another potentially important biogeochemical response to dust deposition: the occurrence of successive dust deposition events could have a different effect than one isolated event. In P-limited areas, a first dust deposition event may induce an increase of biological activity triggered by the input of limiting nutrients (e.g. phosphorus (Pulido-Villena et al., 2010)). This could, in turn, induce an increase in the iron binding capacity of seawater. A second deposition event in a short time period (few days) could, thus, induce a much larger release of bioavailable iron.

This work demonstrates that the quantification of the inputs of dissolved iron from dust in the surface ocean cannot be restricted to a simple determination of atmospheric fluxes. Complex atmospheric and oceanic processes at different time scales have to be considered in order to get a realistic picture of the role of dust deposition on marine iron cycle.

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**Table 1.** Fe masses and fluxes exported to the sediment traps during the DUNE-1-P experiment.

Mesocosm	Time <sup>1</sup> (h)	Collection time <sup>2</sup> (h)	Mass Fe <sup>3</sup> (mg)	Mass Fe cum. <sup>4</sup> (mg)	Flux Fe <sup>5</sup> (mg m <sup>-2</sup> d <sup>-1</sup> )	Flux Fe cum. <sup>6</sup> (mg m <sup>-2</sup> d <sup>-1</sup> )
C1	24	24	3.3	3.3	0.8	0.8
	72	48	2.3	5.6	0.28	0.45
	120	48	0.7	6.3	0.08	0.3
	168	48	2	8.3	0.24	0.29
C2	24	24	0.8	0.8	0.19	0.19
	72	48	3.6	4.4	0.43	0.35
	120	48	4.5	8.9	0.54	0.43
	168	48	1	9.9	0.12	0.34
C3	24	24	2.2	2.2	0.53	0.53
	72	48	8.5	10.7	1.02	0.86
	120	48	0.8	11.5	0.1	0.55
	168	48	NA	NA	NA	NA
D1	24	24	122.9	122.9	29.6	29.6
	72	48	340.3	463.2	41	37.2
	120	48	54.4	517.6	6.55	24.9
	168	48	20.4	538.0	2.46	18.5
D2	24	24	87.5	87.5	21.08	21.1
	72	48	235.2	322.7	28.33	25.9
	120	48	144.7	467.4	17.43	22.5
	168	48	23	490.4	2.77	16.9
D3	24	24	167.6	167.6	40.38	40.4
	72	48	168.3	335.9	20.27	27
	120	48	86.8	422.7	10.46	20.4
	168	48	12.1	434.8	1.46	15

<sup>1</sup> Time of recovery of the sediment trap in hours since the fertilization.

<sup>2</sup> Time of deployment and collection of the sediment trap in hours.

<sup>3</sup> Mass of iron recovered in the trap at time  $t$  in mg.

<sup>4</sup> Cumulated mass of iron recovered in the trap since the begin of the experiment in mg.

<sup>5</sup> Flux of iron recovered at time  $t$ . Estimated as the mass of iron, divided through the time of collection and the surface of the mesocosm (4.15 m<sup>2</sup>) in mg m<sup>-2</sup> d<sup>-1</sup>.

<sup>6</sup> Cumulated of iron recovered since the begin of the experiment.

Estimated as the cumulated mass of iron divided through the time  $t$  and the surface of the mesocosm in mg m<sup>-2</sup> d<sup>-1</sup>.

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**Table 2.** Iron budget in the mesocosms at different time *t* in mg.

Time	0	24	120	168	0	24	120	168
Mesocosm	D1				D2			
Input <sup>1</sup>	959	0	0	0	959	0	0	0
DFe Stock <sup>2</sup>	7	6	5	6	7	6	6	5
PFe Stock <sup>3</sup>	41	270	52	54	42	169	67	43
Export <sup>4</sup>	0	123	518	538	0	88	467	490
Sum	1007	399	575	598	1008	263	540	538
% recovery <sup>5</sup>	100	40	57	59	100	26	54	53

<sup>1</sup> Mass of iron introduced during the fertilization.

<sup>2</sup> Mass of iron in the water column in the form of dissolved iron (<0.2 μm). Between 0 and 10 m, the stock is estimated through integration of the concentration between the three sampling depth. From 10 m to the bottom of the mesocosms a constant concentration equal to the concentration is assumed and the stock is estimated with taking into account the conic bottoms of the mesocosms.

<sup>3</sup> Mass of iron in the water column in the form of particulate iron (>0.2 μm). Determined as described at point<sup>2</sup>.

<sup>4</sup> Mass of iron exported in the sediment trap. Correspond to the cumulate mass recovered in the trap (see Table 1).

<sup>5</sup> The recovery is estimated as the percentage of the mass of iron that is recovered relative to the initial mass at t0.

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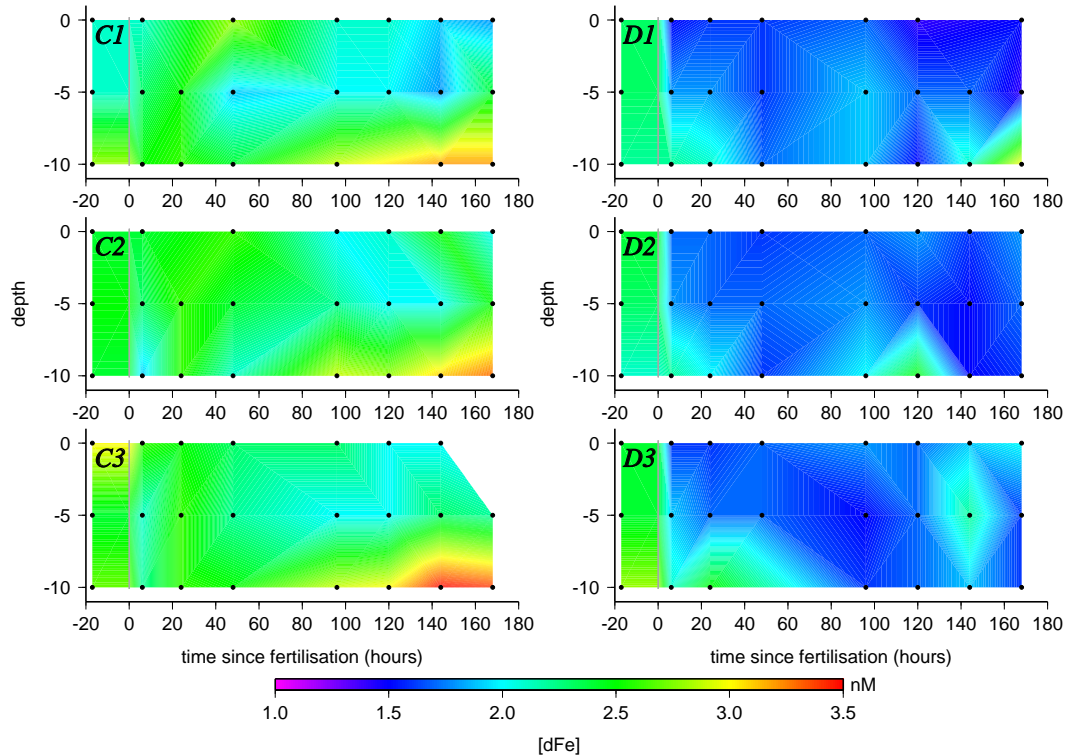
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**Fig. 1.** Evolution of dFe concentrations ([dFe] in nM) during the DUNE-1-P experiment in all mesocosms.

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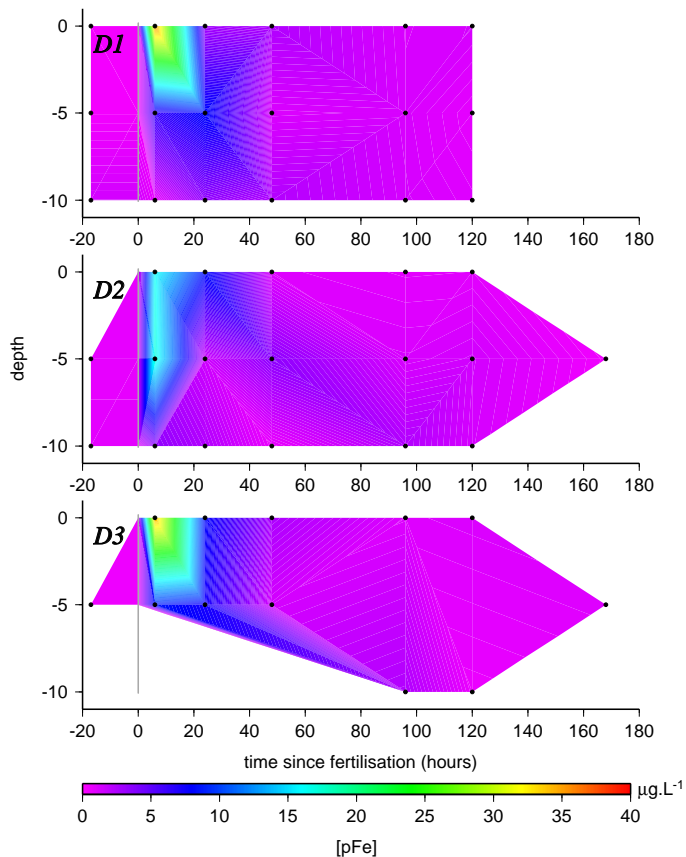
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**Fig. 2.** Evolution of pFe concentrations ( $[pFe]$  in  $\mu\text{g}\text{L}^{-1}$ ) during the DUNE-1-P experiment in mesocosms D1, D2 and D3.

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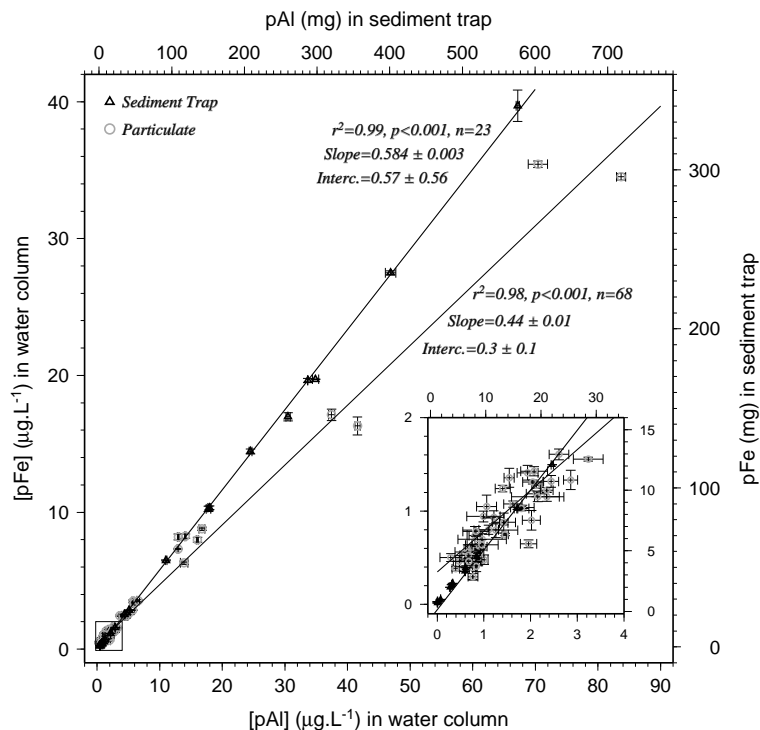
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**Fig. 3.** [pFe] versus [pAl] in sediment trap and in the water column. Black triangles (Right and top axes) are for sediment trap data, in mg. Open circles (Left and bottom axes) are for particulate in the water column, in  $\mu\text{g l}^{-1}$ . Both sets of axis are proportional.

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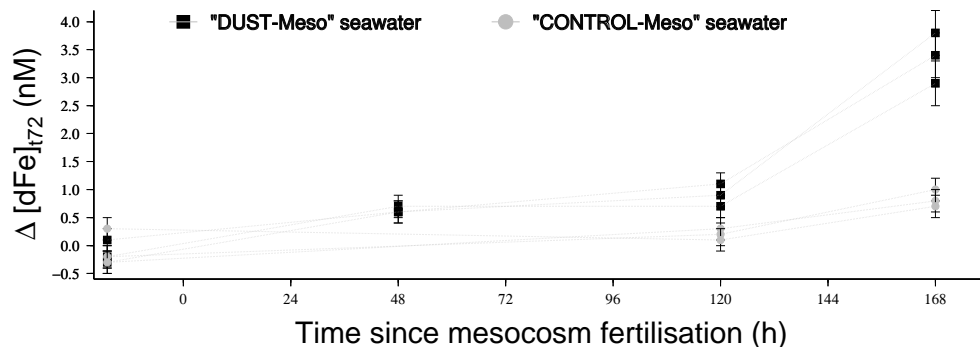
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**Fig. 4.** Results from dust dissolution in batch experiments with filtered seawater collected in the mesocosms. The X axis corresponds to the time of collection of the filtered seawater in the mesocosms and addition of dust to the batch reactor (Teflon bottles). It indicates the time spent since the mesocosm fertilization. The Y axis corresponds to the dissolution measured after 72 h of contact time between dust particles and seawater in the batch reactors. Dissolution is expressed as the difference between [dFe] in seawater after 72 h of contact time and [dFe] at the time of the water collection<sub>72h</sub> in the corresponding mesocosm ( $\Delta[\text{dFe}]_{72\text{h}}$  in nM).

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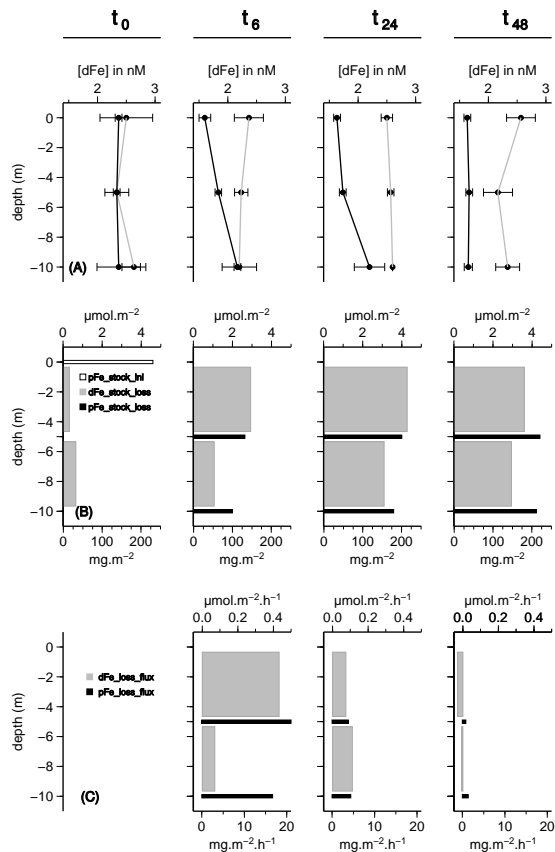
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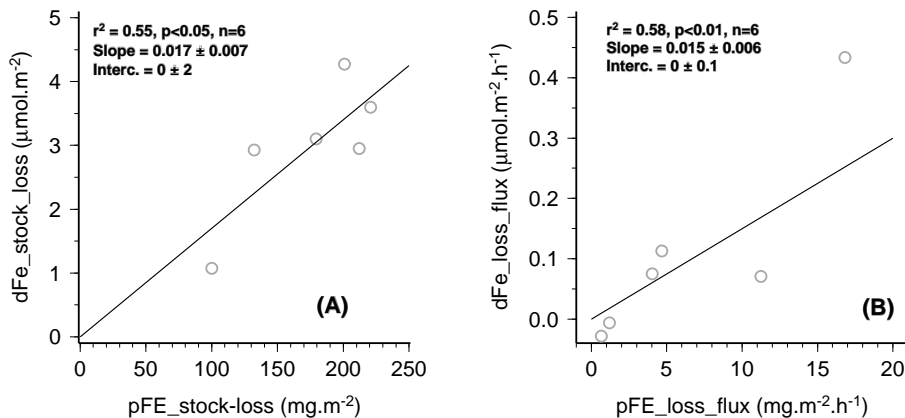


**Fig. 5.** Scavenging of dissolved iron on settling particles during the first 48 h of the DUNE-1-P experiment. All values are the average of the mesocosms triplicates. On Fig. 5a, [dFe] profiles in “CONTROL-meso” (grey line) and “DUST-meso” (black line). On Fig. 5b, loss of dFe stock (dFe.stock.loss, Grey bars, top axis) and loss of pFe stock (pFe.stock.loss, black bars, bottom axis). On Fig. 5c, fluxes of dFe loss (dFe.loss.flux, Grey bars, top axis) and pFe loss (pFe.loss.flux, black bars, bottom axis). The definition and description of the calculation of this parameters can be found in the text (Sect. 4.3.2).

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**Fig. 6.** Correlation between dFe and pFe stock loss (6-A) and fluxes of dFe and pFe loss (6-B). The definition and description of the calculation of this parameters can be found in the text (Sect. 4.3.2).

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