Biogeosciences Discuss., 8, 11577–11599, 2011 www.biogeosciences-discuss.net/8/11577/2011/ doi:10.5194/bgd-8-11577-2011 © Author(s) 2011. CC Attribution 3.0 License.



This discussion paper is/has been under review for the journal Biogeosciences (BG). Please refer to the corresponding final paper in BG if available.

The moisture response of soil heterotrophic respiration: interaction with soil properties

F. E. Moyano¹, N. Vasilyeva¹, L. Bouckaert², F. Cook³, J. Craine⁴, J. Curiel Yuste⁵, A. Don⁶, D. Epron⁷, P. Formanek⁸, A. Franzluebbers⁹, U. Ilstedt¹⁰, T. Kätterer¹¹, V. Orchard¹², M. Reichstein¹³, A. Rey¹⁴, L. Ruamps¹, J.-A. Subke¹⁵, I. K. Thomsen¹⁶, and C. Chenu¹

¹CNRS-UPMC-AgroParisTech UMR Bioemco7618, 78850 Thiverval-Grignon, France

²Department of Soil Management and Soil Care, Ghent University, Ghent, Coupure Links 653, Belgium

³Environmental Physicist/Director, Freeman Cook & Associates, Pty Ltd, P.O. Box 948, Mt Ommaney Q4074, Australia

⁴Division of Biology, Kansas State University, Manhattan KS 66506-4901, USA

⁵Museo Nacional de Ciencias Naturales CSIC, Serrano 115 dpdo, 28006 Madrid, Spain

⁶Johann Heinrich von Thünen-Institut, Institut für Agrarrelevante Klimaforschung, Bundesallee 50, 38116 Braunschweig, Germany



7 Nancy-Université, UMR Ecologie et Ecophysiologie Forestières, 54506 Vandoeuvre Les Nancy, France

Mendel University Brno, Department of Geology and Soil Science, Zemedelska 3, 613 00 Brno, Czech Republic

⁹ USDA – Agricultural Research Service, 1420 Experiment Station Road, Watkinsville GA 30677. USA

¹⁰ Department of Forest Ecology and Management, SLU, SE-901 83 Umeå, USA

¹¹ SLU, Dept. Soil and Environment, P.O. Box 7014, 75007 Uppsala, Sweden

¹² Science and Research, ESR, New Zealand

Biogeochemical Model-Data Integration Group, Max-Planck Insititute for Biogeochemistry, 07701 Jena, Germany

¹⁴ Museo Natural De ciencias Naturales (MNCN-CSIC), Serrano 150. 28006 Madrid, Spain

15 University of Stirling, School of Natural Sciences, Biological and Environmental Sciences, Stirling FK9 4LA, Scotland, UK

¹⁶ Department of Agroecology, Organic Matter, Blichers Allé 20, 8830, Tjele, Denmark

Received: 21 October 2011 - Accepted: 2 November 2011 - Published: 2 December 2011

Correspondence to: F. E. Moyano (fernando.moyano@grignon.inra.fr)

Published by Copernicus Publications on behalf of the European Geosciences Union.

Discussion Pap	BGD 8, 11577–11599, 2011							
oer	Moisture response of soil respiration							
Discus	F. E. Moy	ano et al.						
sion P	Title Page							
aper	Abstract	Introduction						
_	Conclusions	References						
Discu	Tables	Figures						
Ission	I	►I						
n Pap	•	•						
ber	Back	Close						
—	Full Scre	en / Esc						
Discussion	Printer-frier Interactive	ndly Version Discussion						
Paper	CCC D							

Abstract

Soil moisture is of primary importance for predicting the evolution of soil carbon stocks and fluxes, both because it strongly controls organic matter decomposition and because it is predicted to change at global scales in the following decades. However, the

- soil functions used to model the heterotrophic respiration response to moisture have limited empirical support and introduce an uncertainty of at least 4 % in global soil carbon stock predictions by 2100. The necessity of improving the representation of this relationship in models has been highlighted in recent studies. Here we present a datadriven analysis of soil moisture-respiration relations based on 90 soils. With the use
- of linear models we show how the relationship between soil heterotrophic respiration and different measures of soil moisture is consistently affected by soil properties. The empirical models derived include main and moisture interaction effects of soil texture, organic carbon content and bulk density. When compared to other functions currently used in different soil biogeochemical models, we observe that our results can correct
- ¹⁵ biases and reconcile differences within and between such functions. Ultimately, accurate predictions of the response of soil carbon to future climate scenarios will require the integration of soil-dependent moisture-respiration functions coupled with realistic representations of soil water dynamics.

1 Introduction

Soil moisture is one of the most important environmental factors driving productivity and carbon cycling in terrestrial ecosystems. Next to temperature, it is a primary determinant of the rate at which soil carbon is mineralized by microbes into carbon dioxide (Greaves and Carter, 1922; Davidson et al., 2000; Davidson and Janssens, 2006; Cook and Orchard, 2008) and the main driver of soil microbial activity in many
 ecosystems (Davidson et al., 2000; Jassal et al., 2008; Liu et al., 2009). It is also expected to change significantly at global scales as a result of climate change in the



coming decades (IPCC, 2007; Burke and Brown, 2010), potentially leading to large scale changes in soil carbon stocks in different regions, such as the amazon basin (Jones et al., 2005; Falloon et al., 2011).

- The relationship between soil moisture and soil respiration is known to be variable. It
 depends not only on the soil type (Franzluebbers, 1999; Vincent et al., 2006) but also on the diversity of measures used to express water conditions in soils, each having a unique relationship with soil microbial activity (Ilstedt et al., 2000; Paul et al., 2003). Soil factors including total pore space, bulk density and texture have been shown to influence this relationship (Franzluebbers, 1999; Thomsen et al., 1999). However, such studies have been few with most concentrating on finding the measure of moisture that best predicts respiration rates (e.g. water potential, water-filled pore space, etc.) or the single function that best describes this relationship (e.g. linear, polynomial, etc.).
- A systematic analysis of variations in response to a wide range of soil types has been lacking.
- ¹⁵ Soil carbon models use soil moisture-respiration functions that, in theory, represent an average response of microbial respiration to soil moisture content; i.e. they do not account for any natural variation in this relationship (Rodrigo et al., 1997). These functions are generally developed and validated using soils from specific sites and, as a consequence, do not well represent a wider range of soil types. Accordingly, the
- few studies comparing different moisture-respiration functions have indicated that the related differences in soil carbon budget predictions can be important (Rodrigo et al., 1997; Bauer et al., 2008; Falloon et al., 2011). Fallon et al. (2011) showed that the divergence in simulations related to the moisture function alone is nearly 4 % of global carbon stocks by 2100. However, since all these functions represent an "average" response, the real uncertainty is probably larger. A better understanding of how this relationship actually depends on soil properties will help to quantify and reduce such

uncertainties (Franzluebbers, 1999; Schjonning et al., 1999; Thomsen et al., 1999). Here we present results from a meta-analysis of multiple soil incubation datasets that describe how soil properties regulate the relationship between soil microbial respiration



and moisture. We use the terms "soil respiration" or simply "respiration" referring to soil CO_2 emissions from heterotrophic microbial activity. The soil respiration response to moisture is the result of several processes – including osmotic stress, diffusion and oxygen limitations – that combined produce a net effect on the rate of carbon decomposition. Consequently, we did not look for a function that acts as the best single average predictor, as it would invariably underperform in most soil types. Instead, we treated the respiration response as a variable that changes freely at different levels of moisture and is explained by moisture itself and other soil properties.

The main outcome of this analysis are statistical models that predict the proportional response of soil respiration to moisture as a soil-type dependent variable. This can then be used to derive relative soil respiration curves for a given soil type. We illustrate the results by comparing the model we derive, using data from soils of England and Wales (Bellamy et al., 2005), with other currently used functions.

2 Materials and methods

5

- Data were assembled from studies where soil carbon dioxide emissions were measured together with variations in soil moisture under controlled laboratory conditions. Treatments varied across studies (e.g. intact vs. homogenized soils) but only homogenous samples with respect to soil properties were used. Incubations with temperatures outside the 10 to 35 °C range were excluded. We converted the measures of soil CO₂
- emissions, moisture and soil properties to the same units and normalized soil respiration to a 0–1 scale. Respiration data from incubations with moisture changing over time were corrected for substrate depletion using data from control samples. Pore space, if not available, was calculated assuming a mineral density of 2.65 and organic matter density of 1.4.
- The resulting database consisted of 90 different soils originating from 42 sites and characterized by a broad range of soil properties (Tables 1 and 2). We defined a dataset as soil respiration data related to one or more out of four ways of expressing



soil moisture. These measures are: mass related or gravimetric moisture (θ_m), volumetric moisture (θ_v), fraction of water saturation (θ_s), and the logarithm of water potential (ψ_{log}). θ_m is a laboratory standard while θ_v is the most widely used field measure, often associated with high frequency carbon flux data. θ_s and ψ_{log} are often considered optimal predictors of microbial respiration as they are related to air space and water energy status, respectively (Orchard and Cook, 1983; Skopp et al., 1990). For convenience, the measure of ψ_{log} used was $(-\log_{10}|\psi|_{kPa})/5+1$, thus obtaining an approximate range of 0 to 1. For other measures we used: $gH_2O \cdot g^{-1}$ dry soil (θ_m), cm³ H₂O cm⁻³ total (θ_v), cm³ H₂O cm⁻³ pore-space (θ_s). When possible, missing moisture measures were derived, e.g. using bulk density or pore space for converting θ_v to θ_m or θ_s , respectively, and vice versa. The following analysis was performed in parallel for each moisture measure using the R statistical software (Supplement: R Code and data files MRD.txt, DD.txt and funs.txt).

We started by assuming that a response to a change in soil moisture is proportional to the value of respiration itself, as normally done in soil carbon models. By using the proportional response we make our results generalizable, avoiding the problem of comparing absolute respiration values which vary largely across soils. Since the respiration response varies along the moisture axis, we defined the Proportional Response of Soil Respiration (PR_{SR}) related to a 0.01 increase in soil moisture as the central unit for analysis. We then tested how PR_{SR} is affected by diverse soil properties.

To obtain PR_{SR} values we used general additive models (GAMs) to fit smooth curves to each of 310 datasets (Supplement: Fig. S1). Linear or polynomial fits were used instead if the number of moisture points in a dataset was less than 4. Respiration values at each 0.01 moisture interval were then derived from the curves between the ²⁵ minimum and maximum of each dataset. The PR_{SR} of each 0.01 increase in moisture was calculated, at moisture *M*, as the average of SR(M)/SR(M - 0.01) and SR(M + 0.01)/SR(M).



To analyze relationships of soil properties with PR_{SB} we used soil pore space, bulk density, soil organic carbon, and sand, silt and clay content. We also tested soil pH and the interaction between organic carbon and clay but found no significant effects. Preliminary results revealed important differences between soils with high and low organic carbon content, so soils with over 50 mg C g^{-1} soil, hereafter referred to as organic 5 soils, were analyzed separately from mineral soils. To isolate the effect of each soil property we used linear regression models of the form:

 $PR_{SB} = \beta_1 M + \beta_2 M^2 + \beta_3 M^3 + \beta_i SP_i + \beta_i M \cdot SP_i + \varepsilon,$

where *M* is soil moisture (either θ_m , θ_v , θ_s or ψ_{log}) and SP are soil properties which can interact with M. Stepwise model selection was applied. Since the proportional 10 increase in respiration tended to be very large at values near 0, producing a strong bias in the models, we excluded outliers defined as any PR_{SR} value further than 3 standard deviations from the mean.

Model simplification led to excluding pore space (strongly correlated with BD), sand, silt and the SOC-M interaction. As bulk density is often not available for use in large 15 scale soil simulations, we fitted a second model for mineral soils including only clay and organic carbon. The final linear models predicting PR_{SR} were:

$$PR_{SR} = \beta_1 M + \beta_2 M^2 + \beta_3 M^3 + \beta 4BD + \beta_5 M \cdot BD + \beta_6 Clay + \beta_7 M \cdot Clay + \beta_8 SOC , \qquad (2)$$

$$PR_{SR} = \beta_1 M + \beta_2 M^2 + \beta_3 M^3 + \beta_6 Clay + \beta_7 M \cdot Clay + \beta_8 SOC , \qquad (3)$$

$$PR_{SR} = \beta_1 M + \beta_2 M^2 + \beta_3 M^3 , \qquad (4)$$

²⁰ PR_{SR} =
$$\beta_1 M + \beta_2 M^2 + \beta_2 M^2$$

25

where M is soil moisture, BD is bulk density, SOC is soil organic carbon and β are model coefficients. Model 1 (Eq. 2) and Model 2 (Eq. 3) are mineral soil models, with the latter excluding bulk density (Table 3). Model 3 (Eq. 4) is for organic soils and has only moisture as a predictor. Few datasets were available for these soils and significant correlations with soil properties were not found (Table 3).



(1)

With the PR_{SR} values obtained from the above models, respiration was predicted using the equation:

$$\mathsf{SR}(M) = \left(\prod_{k=M_0}^M \mathsf{PR}_{\mathsf{SR}_k}\right) \cdot \mathsf{SR}_0 \,,$$

where soil respiration (SR) as a function of soil moisture (M) is equal to an initial respiration value (SR₀) multiplied by the product of all PR_{SR} values at each k 0.01 moisture interval from the initial moisture (M_0) to M (for $M_0 < M$; otherwise the product term divides SR₀). PR_{SR} values at each k interval are predicted with a PR_{SR} model.

Relative respiration curves, which scale respiration from 0 to 1, were obtained in a two-step calculation: 1. Using Eq. (2) to predict PR_{SR} values for each 0.01 moisture interval and 2. Using Eq. (5) to calculate respiration values along the moisture axis (with an arbitrary SR₀ of 1) and dividing all values by the maximum obtained. As data at low moistures extremes was generally missing, regression models did not well reproduce the high PR_{SR} related to respiration values approaching 0. As a result, depending on the soil type, curve intercepts were variably higher than 0. To obtain curves with a 0 intercept we applied a rescaling of respiration from 0 to 1 in the range of 0 to optimum moisture (Supplement: R Code lines 392–396).

In order to compare our results with existing functions, we applied Eq. (3) using θ_s or ψ_{log} to predict respiration curves for 106 soil series from England and Wales covering an area of ca. 50 000 km². Soil organic carbon and clay content in these soils ranged between 0.01–0.05 g g⁻¹ and 80–610 g clay kg⁻¹ soil. We compared these results with functions from six other models using the same moisture measures, plotting the resulting range of respiration values next to θ_s functions belonging to the RothC (Coleman and Jenkinson, 1999; Bauer et al., 2008), CANDY (Franko et al., 1995; Powlson et al., 1996), Bethy (Knorr, 2000) and SimCycle (Ito and Oikawa, 2002) models and ψ_{log} functions from the Daisy (Abrahamsen and Hansen, 2000; Bauer et al., 2008) and SOILCO2 (Šimunek and Suarez, 1993; Bauer et al., 2008) models. To use θ_s with



(5)

the RothC function we followed the same assumption as Bauer and colleagues (Bauer et al., 2008).

3 Results

saturation.

25

- For each dataset, PR_{SR} values are highest at dry conditions and decrease progressively with increasing moisture (Fig. 1), with values below 1 corresponding to a negative trend in respiration rates. Mean PR_{SR} values for θ_s and ψ_{log} decreased monotonically with increasing moisture (Fig. 1c and 1d) while those for θ_m and θ_v showed more discontinuities related to sharp variations in soil types (Fig. 1a,b). All moisture measures had a wide range of soil moisture associated to an optimum for respiration, defined as the point where PR_{SR} crosses 1. Taking θ_s as an example, the PR_{SR} mean value reaches 1 at 0.63 θ_s , consistent with the commonly reported range of 0.6–0.7, but different datasets had values of optimum moisture as low as 0.4 and as high as 0.9 water
- For mineral soils, significant correlations were found between PR_{SR} and all soil properties, with the correlation strength and significance being strongly dependent on the moisture range and type of moisture measure (Fig. 2). With θ_m and θ_v , correlation coefficients of PR_{SR} versus bulk density were negative and tended to increase with increasing moisture. Correlations with pore space, not shown in Fig. 2, were identical but of opposite sign, i.e. positive. Fewer or no significant correlations of these properties were seen for θ_s and ψ_{log} .

Significant PR_{SR} correlations with texture and organic carbon were found for all moisture measures but most importantly for θ_m and θ_v . Correlations were generally negative for sand and positive for clay, silt and organic carbon. Correlations with clay and silt followed a similar pattern that mirrored the behavior of sand. Correlations with organic carbon content were similar to those of clay and silt. In contrast, organic soils showed no significant correlations between PR_{SR} and carbon content (data not shown).



The range of values used for fitting the multiple linear regression models is given in Table 2. Models using θ_m and θ_v showed the largest improvement in their root mean square deviation after adding soil properties to the basic moisture polynomial. However, models using θ_s and ψ_{log} remained the best predictors, with ψ_{log} having a slightly better performance. An analysis of model residuals resulted in no trend or significant correlation with soil incubation temperature and incubation duration.

Relative respiration curves are shown in Fig. 3. The effect of clay content on respiration was mainly at low (aerobic) moisture ranges and strongly affected the spread in the curve. Less clay resulted in a wider range of soil moisture values associated to optimal respiration and a respiration peak at lower water contents. Soil organic carbon

- ¹⁰ optimal respiration and a respiration peak at lower water contents. Soil organic carbon produced a shift in the curve under all moisture measures with the exception of water potential. More carbon content did not affect the spread of the curve but drove the point of maximum respiration towards higher values of moisture. With changes in bulk density, respiration changed relatively little for a constant volumetric moisture (θ_v curve) or
- water potential (ψ_{log} curve) but changed strongly under a constant gravimetric moisture (θ_m curve) or water saturation fraction (θ_s curve).

When compared to currently used models (Fig. 4), results from our model covered much of the range of variability between other functions based on θ_s , which either under- or overestimate average respiration, with a strong tendency towards the latter. Functions using ψ_{log} were comparable to our predictions, where we observed a limited

²⁰ Functions using ψ_{log} were comparable to our predictions, where we observed a limited influence of soil properties, but they showed a general overestimation of respiration values in most of the range of suboptimal moisture conditions.

4 Discussion

5

This comparison of multiple datasets revealed a strong soil-dependent variation of the ²⁵ moisture-respiration relationship, in clear contrast to the simple functions found in all current models. The large range of variability observed (e.g. respiration maximums ranging from 40–100 % water saturation) reflects differences between ecosystems that



are largely ignored in the more common and simplified representations of this relationship. A major difference observed was in the response of organic vs. mineral soils. In the case of organic soils we found little or no effects of soil properties on the PR_{SR} . The models derived for organic soils use moisture as the only predictor and serve as a best approximation. However, they remain rough averages given the limited available data

⁵ approximation. However, they remain rough averages given the limited available data and the often incomplete characterization of soil properties. More data will be needed to better characterize moisture effects in these soils.

For mineral soils (with less than $50 \text{ mg C g soil}^{-1}$) the soil factors having an influence on the moisture-respiration relationship involved aeration and structure (bulk density),

- texture (clay) and composition (carbon content). The models we derived from the data include clay but not silt or sand. This was a result of the large effect of clay and the relatively small influence of silt when including one or the other in the linear regression. This is probably caused by the much larger specific surface of clay particles which affects water retention and availability. The observed increase in the optimum water
 content for respiration with increasing clay fraction has also been observed in field aparticles and the increase of clay and the increase of clay particles with increasing clay fraction has also been observed in field aparticles and the content for respiration with increasing clay fraction has also been observed in field aparticles.
- conditions (Balogh et al., 2011). Water potential is the only measure of soil moisture for which this pattern was not observed.

In accordance with theory (Orchard and Cook, 1983; Orchard et al., 1992), the relation between soil respiration and soil water potential (ψ_{log}) was the least affected by

- ²⁰ soil properties, making this measure the best predictor of respiration rates. However, since large changes in water potential are often associated to small changes in water content, predicting water potential in soils could itself be associated with large errors. Among the models based on measures of water content (θ_s , θ_m or θ_v), those using θ_s resulted in the lowest root mean square deviation and were thus the best predictor
- ²⁵ of respiration rates. Ultimately, finding the measure that performs best in large scale simulations will require a validation of model performance against actual field data.

The data from England and Wales soils used for model comparison represents a large range of properties characteristic of soils in temperate regions. The range of respiration curves we predicted (Fig. 4), related to variations in the properties of



these soils, covered most of the differences between strongly differing functions such as those used in the RothC, CANDY and Bethy models. Thus, the variability in model predictions, associated with different moisture functions, can be largely reduced or eliminated by using a common, but generally valid, soil-dependent moisture-respiration function. Depending on the model, soil and climate, significantly different predictions of

⁵ function. Depending on the model, soil and climate, significantly different predictions of soil carbon decomposition are expected after including these soil-dependent functions. In most cases this will tend towards lower rates of respiration and, consequently, to an increased sequestration of carbon in soils.

5 Conclusions

20

- ¹⁰ Our empirical analysis has shown that the microbial soil respiration response to moisture depends on soil properties in a consistent and largely predictable way. Future studies should concentrate on reducing uncertainties in these relationships and on better representing specific field conditions, such as the depth-dependence of oxygen availability and the dynamics of soil water. It remains unclear if soils will cause a positive or
- ¹⁵ negative feedback to global warming as global changes in climatic patterns affect soil temperature and moisture (IPCC, 2007; Kendon et al., 2009; Burke and Brown, 2010), but the moisture response of soil carbon decomposition will likely have an important role in determining any future evolution.

Supplementary material related to this article is available online at: http://www.biogeosciences-discuss.net/8/11577/2011/ bgd-8-11577-2011-supplement.zip.

Acknowledgements. This work was supported by the Project Carbosoil from GIS Climat-Environnement-Société. England and Wales soils data was used under licence of "Soils Data (c) Cranfield University (NSRI) and for the Controller of HMSO, 2011".



References

20

Abrahamsen, P. and Hansen, S.: Daisy: an open soil-crop-atmosphere system model, Environ. Model. Softw. 15, 313–330, 2000.

Balogh, J., Pintér, K., Fóti, S., Cserhalmi, D., Papp, M., and Nagy, Z.: Dependence of soil respi-

- ration on soil moisture, clay content, soil organic matter, and CO₂ uptake in dry grasslands, Soil Biol. Biochem., 43, 1006–1013, 2011.
 - Bauer, J., Herbst, M., Huisman, J., Weihermuller, L., and Vereecken, H.: Sensitivity of simulated soil heterotrophic respiration to temperature and moisture reduction functions, Geoderma, 145, 17–27, 2008.
- ¹⁰ Bellamy, P. H., Loveland, P. J., Bradley, R. I., Lark, R. M., and Kirk, G. J. D.: Carbon losses from all soils across England and Wales 1978–2003, Nature 437, 245–248, 2005.

Bowden, R., Newkirk, K., and Rullo, G.: Carbon dioxide and methane fluxes by a forest soil under laboratory-controlled moisture and temperature conditions, Soil Biol. Biochem., 30, 1591–1597, 1998.

- ¹⁵ Burke, E. J. and Brown, S. J.: Regional drought over the UK and changes in the future, J. Hydrol., 394, 471–485, 2010.
 - Coleman, K. and Jenkinson, D. S.: RothC-26.3, A Model for the Turnover of Carbon in Soil: Model Description and User's Guide, Lawes Agric. Trust, Harpenden, UK, 1999.

Cook, F. J. and Orchard, V. A.: Relationships between soil respiration and soil moisture, Soil Biol. Biochem. 40, 1013–1018, 2008.

Cook, F. J., Orchard, V. A., and Corderoy, D. M.: Effects of lime and water content on soil respiration, New Zeal. J. Agr. Res., 28, 517–523, 1985.

Craine, J. M. and Gelderman, T. M.: Soil moisture controls on temperature sensitivity of soil organic carbon decomposition for a mesic grassland, Soil Biol. Biochem., 43, 455–457, 2010.

²⁵ Curiel Yuste, J., Baldocchi, D. D., Gershenson, A., Goldstein, A., Misson, L., and Wong, S.: Microbial soil respiration and its dependency on carbon inputs, soil temperature and moisture, Global Change Biol., 13, 2018–2035, 2007.

Davidson, E. A. and Janssens, I. A.: Temperature sensitivity of soil carbon decomposition and feedbacks to climate change, Nature, 440, 165–173, 2006.

³⁰ Davidson, E. A., Verchot, L. V., Cattanio, J. H., Ackerman, I. L., and Carvalho, J. E. M.: Effects of soil water content on soil respiration in forests and cattle pastures of Eastern Amazonia, Biogeochemistry, 48, 53–69, 2000.



- Doran, J. W.: Microbial activity as regulated by soil water-filled pore space, in: Transactions 14th International Congress of Soil Science, Presented at the 14th International Congress of Soil Science, 12–18 August 1990, Kyoto, Japan, pp. 94–99, 1990.
- Falloon, P., Jones, C. D., Ades, M., and Paul, K.: Direct soil moisture controls of future global soil carbon changes: an important source of uncertainty, Global Biogeochem. Cy., 25, 14, 2011.
 - Franko, U., Oelschlagel, B., and Schenk, S.: Modellierung von Bodenprozessen in Agrarlandschaften zur Untersuchung der Auswirkungen möglicher Klimaveränderungen, Sektion Bodenforschung, UFZ-Umweltforschungszentrum Leipzig-Halle GmbH 3, Leipzig, DE, 1995.
- ¹⁰ Franzluebbers, A. J.: Microbial activity in response to water-filled pore space of variably eroded Southern Piedmont soils, Appl. Soil Ecol., 11, 91–101, 1999.
 - Greaves, J. E. and Carter, E. G.: The influence of moisture and soluble salts on the bacterial activities of the soil, Soil Sci., 13, 251–270, 1922.
 - Gulledge, J. and Schimel, J. P.: Moisture control over atmospheric CH₄ consumption and CO₂ production in diverse Alaskan soils, Soil Biol. Biochem., 30, 1127–1132, 1998.

15

Ilstedt, U., Nordgren, A., and Malmer, A.: Optimum soil water for soil respiration before and after amendment with glucose in humid tropical acrisols and a boreal mor layer, Soil Biol. Biochem. 32, 1591–1599, 2000.

IPCC: Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to

- the Fourth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge University Press, Cambridge, UK, New York, USA, 2007.
 - Ito, A. and Oikawa, T.: A simulation model of the carbon cycle in land ecosystems (Sim-CYCLE): a description based on dry-matter production theory and plot-scale validation, Ecol. Model., 151, 143–176, 2002.
- Jassal, R. S., Black, T. A., Novak, M. D., Gaumont-Guay, D., and Nesic, Z.: Effect of soil water stress on soil respiration and its temperature sensitivity in an 18-year-old temperate Douglasfir stand, Global Change Biol., 14, 1305–1318, 2008.
 - Jones, C., McConnell, C., Coleman, K., Cox, P., Falloon, P., Jenkinson, D., and Powlson, D.: Global climate change and soil carbon stocks; predictions from two contrasting models for
- the turnover of organic carbon in soil, Global Change Biol., 11, 154–166, 2005.
 - Kendon, E. J., Rowell, D. P., and Jones, R. G.: Mechanisms and reliability of future projected changes in daily precipitation, Clim. Dynam., 35, 489–509, 2009.

Knorr, W.: Annual and internannual CO2 exchanges of the terrestrial biosphere: process-based



simulations and uncertainties, Global Ecol. Biogeogr., 9, 225-252, 2000.

- Linn, D. and Doran, J.: Effect of water-filled pore-space on carbon-dioxide and nitrous-oxide production in tilled and nontilled soils, Soil Sci. Soc. Am. J., 48, 1267–1272, 1984.
- Liu, W., Zhang, Z., and Wan, S.: Predominant role of water in regulating soil and microbial
- respiration and their responses to climate change in a semiarid grassland, Global Change Biol., 15, 184–195, 2009.
 - Lomander, A., Kätterer, T., and Andrén, O.: Carbon dioxide evolution from top-and subsoil as affected by moisture and constant and fluctuating temperature, Soil Biol. Biochem., 30, 2017–2022, 1998.
- ¹⁰ Nyhan, J. W.: Influence of soil temperature and water tension on the decomposition rate of carbon-14 labeled herbage, Soil Sci., 121, 288, 1976.
 - Orchard, V. A. and Cook, F. J.: Relationship between soil respiration and soil-moisture, Soil Biol. Biochem., 15, 447–453, 1983.

Orchard, V. A., Cook, F. J., and Corderoy, D. M.: Field and laboratory studies on the relationship

- between respiration and moisture for 2 soils of contrasting fertility status, Pedobiologia 36, 21–33, 1992.
 - Paul, K. I., Polglase, P. J., O'Connell, A. M., Carlyle, J. C., Smethurst, P. J., and Khanna, P. K.: Defining the relation between soil water content and net nitrogen mineralization, Eur. J. Soil Sci., 54, 39–48, 2003.
- Powlson, D. S., Smith, P., and Smith, J. U.: Evaluation of Soil Organic Matter Models, NATO ASI Series. Series 1: Global Environmental Change, Vol. 38, pp. 429, Springer, Berlin, DE, 1996.
 - Reichstein, M., Subke, J. A., Angeli, A. C., and Tenhunen, J. D.: Does the temperature sensitivity of decomposition of soil organic matter depend upon water content, soil horizon, or incubation time? Global Change Biol., 11, 1754–1767, 2005.
 - Rey, A., Petsikos, C., Jarvis, P. G., and Grace, J.: Effect of temperature and moisture on rates of carbon mineralization in a Mediterranean oak forest soil under controlled and field conditions, Eur. J. Soil Sci., 56, 589–599, 2005.

25

- Rodrigo, A., Recous, S., Neel, C., and Mary, B.: Modelling temperature and moisture effects on C-N transformations in soils: comparison of nine models, Ecol. Model., 102, 325–339, 1997.
 - Schjonning, P., Thomsen, I., Moberg, J., de Jonge, H., Kristensen, K., and Christensen, B.: Turnover of organic matter in differently textured soils – I. Physical characteristics of struc-



turally disturbed and intact soils, Geoderma 89, 177-198, 1999.

5

Šimunek, J. and Suarez, D. L.: Modeling of carbon dioxide transport and production in soil, 1. Model development, Water Resour. Res., 29, 487–497, 1993.

Skopp, J., Jawson, M., and Doran, J.: Steady-state aerobic microbial activity as a function of soil-water content, Soil Sci. Soc. Am. J., 54, 1619–1625, 1990.

Stott, D. E., Elliott, L., Papendick, R., and Campbell, G.: Low-temperature or low water potential effects on the microbial decomposition of wheat residue, Soil Biol. Biochem., 18, 577–582, 1986.

Thomsen, I., Schjonning, P., Jensen, B., Kristensen, K., and Christensen, B.: Turnover of or-

- ¹⁰ ganic matter in differently textured soils II. Microbial activity as influenced by soil water regimes, Geoderma, 89, 199–218, 1999.
 - Vincent, G., Shahriari, A. R., Lucot, E., Badot, P.-M., and Epron, D.: Spatial and seasonal variations in soil respiration in a temperate deciduous forest with fluctuating water table, Soil Biol. Biochem., 38, 2527–2535, 2006.
- ¹⁵ Wickland, K. P. and Neff, J. C.: Decomposition of soil organic matter from boreal black spruce forest: environmental and chemical controls, Biogeochemistry 87, 29–47, 2008.

Discussion Pa	B(8, 11577–1	BGD 8, 11577–11599, 2011						
per	Moisture r soil res	esponse of piration						
Discu	F. E. Moy	ano et al.						
ssion P	Title	Title Page						
aper	Abstract	Introduction						
_	Conclusions	References						
Discu	Tables	Figures						
noiss	14	►I.						
Par	•	•						
)er	Back	Close						
_	Full Scre	een / Esc						
Discuss	Printer-frie	Printer-friendly Version						
ion F	Interactive	Interactive Discussion						
aper	C	ВУ						

Table 1. Description of the data used in the analysis. Each line represents a specific site or location. DS is number of data sets, representing different soil types related to each site. Site averages of soil properties used in the analysis are shown. BD = bulk density, SOC = soil organic carbon, N = soil nitrogen.

Reference	DS	Country	Ecosystem	BD	SO	N	Clay	Silt	Sand	pН
				(g cm ⁻³)	(mgg^{-1})	(mgg^{-1})	(%)	(%)	(%)	
Bouckaert not published	3	Belgium	Forest	16	44	3.5	18	49	33	54
Bowden et al. (1998)	1	USA	Forest	0.3	NA	NA	NA	NA	NA	3.3
Cook, not published	1	New Zealand	Grassland	1.3	64	4.7	16	6	79	5.4
Cook, not published	1	New Zealand	Grassland	0.9	55	5.5	17	33	50	5.7
Cook, not published	1	New Zealand	Grassland	0.8	71	6.2	52	34	14	6.1
Cook et al. (1985)	1	New Zealand	Grassland	NA	NA	NA	NA	NA	NA	NA
Craine and Gelderman (2010)	8	USA	Grassland	NA	46	3.3	33	60	8	67
Curiel Yuste et al. (2007)	2	USA	Forest	0.9	102	3.5	11	29	60	5.5
Curiel Yuste et al. (2007)	2	USA	Grassland	1.5	28	2.5	14	44	43	6.4
Don, not published	1	Germany	Grassland	1.5	11	1.1	9	10	81	4.5
Doran (1990)	1	USA	Forest	1.1	31	NA	14	16	70	6.8
Doran (1990)	1	USA	Cultivated	1.2	14	NA	18	29	53	6.8
Doran (1990)	1	USA	Cultivated	1.1	21	NA	22	75	3	6.8
Doran (1990)	1	USA	Grassland	12	8	NA	26	19	55	6.8
Doran (1990)	1	USA	Grassland	1.1	22	NA	17	64	19	6.8
Doran (1990)	1	USA	Grassland	1.0	16	NA	46	42	12	6.8
Doran (1990)	1	USA	NA	1.1	35	NA	14	37	49	6.8
Doran (1990)	1	USA	Cultivated	1.2	13	NA	20	51	29	6.8
Doran (1990)	1	USA	Grassland	1.2	7	NA	22	24	54	6.8
Doran (1990)	1	USA	Cultivated	1.1	13	NA	58	35	7	6.8
Doran (1990)	1	USA	Cultivated	11	11	NA	16	68	16	6.8
Skopp et al. (1990)	1	USA	Grassland	1.2	13	NA	24	54	22	6.8
Epron, not published	1	France	Forest	0.8	27	2.5	20	66	14	4.6
Formanek, not published	3	Czech Republic	Forest	NA	318	11.2	NA	NA	NA	4.8
Franzluebbers (1999)	15	USA	Grassland	1.2	16	1.1	19	16	65	6.2
Gulledge and Schimel (1998)	2	USA	Grassland	NA	61	NA	NA	NA	NA	NA
listedt et al. (2000)	3	Malavsia	Forest	0.6	52	4.1	27	32	42	4.7
listedt et al. (2000)	1	Sweden	Forest	0.5	556	NA	NA	NA	NA	4.1
Linn and Doran(1984)	1	USA	Cultivated	NA	21	1.6	34	54	12	5.8
Liu et al. (2009)	1	China	Grassland	NA	16	1.5	17	20	63	6.8
Lomander et al. (1998)	2	Sweden	Cultivated	NA	18	1.9	57	38	5	8.2
Nyhan (1976)	1	USA	NA	1.4	9	NA	NA	NA	NĂ	6.8
Orchard and Cook (1983)	1	New Zealand	Grassland	NA	NĂ	NA	NA	NA	NA	NA
Orchard et al. (1992)	1	New Zealand	Grassland	NA	56	3.5	24	NA	NA	5.2
Orchard et al. (1992)	1	New Zealand	Grassland	NA	NA	NA	NA	NA	NA	NA
Reichstein et al. (2005)	1	Germany	Forest	0.9	45	2.0	10	38	52	2.9
Rev et al. (2005)	2	Italy	Forest	1.0	49	6.0	NA	NA	NA	5.7
Ruamps, not published	1	France	Cultivated	1.5	14	1.2	17	53	30	6.8
Thomsen et al. (1999)	15	Denmark	Cultivated	1.3	15	1.5	23	14	64	6.9
Skopp et al. (1990)	1	USA	Cultivated	1.4	9	NA	3	7	90	6.8
Stott et al. (1986)	1	USA	Cultivated	NA	4	0.6	NĂ	NA	NA	7.0
Wickland and Neff (2008)	3	USA	Forest	NA	318	14.5	NA	NA	NA	NA



Table 2. Range of values for soil moisture (SM), soil organic carbon (SOC), clay, and bulk density (BD) used to fit linear regression models. Soil moisture units depend on the measure used, as described in methods.

Model and	S	М	SOC (g g ⁻¹ soil)	Clay (1	fraction)	BD (g	cm ⁻³)
moisture measure	min	max	min	max	min	max	min	max
Model 1 (mineral soils)								
θ_{m}	0.04	0.65	0.01	0.05	0.09	0.45	0.8	1.5
θ_{v}	0.05	0.60	0.01	0.05	0.03	0.58	0.8	1.5
θ_{s}	0.07	1.00	0.01	0.05	0.03	0.58	0.8	1.5
ψ_{\log}	0.22	1.02	0.01	0.03	0.09	0.45	0.6	1.5
Model 2 (mineral so	oils)							
$ heta_{m}$	0.04	0.66	0.01	0.05	0.09	0.57	NA	NA
θ_{v}	0.05	0.60	0.01	0.05	0.03	0.58	NA	NA
$ heta_{s}$	0.07	1.00	0.01	0.05	0.03	0.58	NA	NA
$\psi_{ m log}$	0.22	1.02	0.01	0.03	0.09	0.45	NA	NA
Model 3 (organic soils)								
$ heta_{m}$	0.05	1.1	0.05	0.40	NA	NA	NA	NA
θ_{v}	0.05	0.85	0.06	0.35	NA	NA	0.24	0.9
θ_{s}	0.07	0.99	0.06	0.35	NA	NA	0.24	0.9
ψ_{log}	0.27	0.97	0.06	0.56	NA	NA	NA	NA

Discussion Paper BGD 8, 11577-11599, 2011 Moisture response of soil respiration **Discussion** Paper F. E. Moyano et al. **Title Page** Abstract Introduction Conclusions References Tables Figures **Discussion** Paper 14 Back Close Full Screen / Esc **Discussion** Paper **Printer-friendly Version** Interactive Discussion

Table 3. Variables and coefficients for linear models predicting the proportional response of soil respiration (PR_{SR}) at 0.01 moisture intervals. SM = soil moisture, BD = bulk density, SOC = soil organic carbon. Coefficient values and standard errors are given under the corresponding moisture measure. All terms are significant at p < 0.001. Values are also shown for the number of datasets used and the model root mean square deviation (RMSD).

Variables	Gravimetric moisture ($\theta_{\rm m}$)	Volumetric moisture (θ_v)	Fraction of saturation (θ_s)	Log water potential (ψ_{log})					
Model 1 (mineral soils)									
<i>n</i> -datasets	<i></i> 50	52	52	42					
RMSD	0.023	0.024	0.014	0.012					
Intercept	1.00 ± 0.02	0.98 ± 0.01	1.02 ± 0.00	1.26 ± 0.02					
SM	-0.80 ± 0.09	-0.48 ± 0.08	-0.29 ± 0.02	-1.36 ± 0.05					
(SM) ²	3.5 ± 0.2	1.8 ± 0.3	0.37 ± 0.04	2.26 ± 0.08					
(SM) ³	-3.1 ± 0.2	-1.6 ± 0.3	-0.19 ± 0.03	-1.12 ± 0.04					
BD (g cm ⁻³)	0.10 ± 0.01	0.1 ± 0.01	0.03 ± 0.00	0.05 ± 0.01					
BD (g cm ^{-3}) H ₂ O	-0.44 ± 0.05	-0.3 ± 0.04	_	-0.09 ± 0.01					
Clay (fraction)	0.33 ± 0.03	0.18 ± 0.02	0.09 ± 0.01	0.17 ± 0.02					
Clay (fraction) H ₂ O	-0.7 ± 0.1	-0.31 ± 0.06	-0.08 ± 0.01	-0.25 ± 0.02					
SOC (ggSoil ⁻¹)	1.5 ± 0.1	1.4 ± 0.09	0.8 ± 0.04	-					
Model 2 (mineral so	ils)								
<i>n</i> -datasets	59	65	66	43					
RMSD	0.025	0.025	0.015	0.013					
Intercept	1.13 ± 0.01	1.11 ± 0.01	1.059 ± 0.003	1.31 ± 0.01					
SM	-1.31 ± 0.05	-0.83 ± 0.07	-0.26 ± 0.02	-1.45 ± 0.05					
(SM) ²	3.0 ± 0.2	1.5 ± 0.3	0.32 ± 0.04	2.18 ± 0.08					
(SM) ³	-2.23 ± 0.2	-1.0 ± 0.3	-0.15 ± 0.03	-1.07 ± 0.04					
Clay (fraction)	0.26 ± 0.02	0.08 ± 0.01	0.08 ± 0.01	0.12 ± 0.01					
Clay (fraction) H ₂ O	-0.39 ± 0.05	-	-0.09 ± 0.01	-0.16 ± 0.02					
SOC (g g Soil ^{-1})	1.07 ± 0.07	1.28 ± 0.08	0.57 ± 0.04	0.19 ± 0.06					
Model 3 (organic soils)									
n-datasets	16	6	6	3					
RMSD	0.020	0.10	0.008	0.014					
Intercept	1.146 ± 0.005	1.178±0.004	1.134 ± 0.003	1.42 ± 0.04					
SM	-0.57 ±0.03	-1.12 ±0.03	-0.67 ± 0.02	-1.9 ± 0.2					
(SM) [∠]	0.79 ± 0.07	2.22 ± 0.09	$\textbf{1.08} \pm \textbf{0.05}$	$\textbf{2.9} \pm \textbf{0.4}$					
(SM) ³	-0.37 ± 0.04	-1.40 ± 0.06	-0.57 ± 0.03	-1.4 ± 0.2					





Fig. 1. Proportional response of soil microbial respiration (PR_{SR}) to moisture. PR_{SR} values correspond to a 0.01 increase in soil moisture. Values are shown for all datasets and for θ_m (gravimetric moisture, **a**), θ_v (volumetric moisture, **b**), θ_s (fraction of saturation, **c**), and ψ_{log} (log water potential, **d**). Units of soil moisture in the *x*-axis are: gH₂O g⁻¹ dry soil (**a**), cm³ H₂O cm⁻³ total (**b**), cm³ H₂O cm⁻³ pore-space (**c**) and $(-\log_{10}|\psi|_{kPa})/5+1$ (**d**). Grey circles represent the PR_{SR} of each 0.01 moisture increase. The black and dashed lines are mean and standard deviations, respectively.

















Fig. 4. Comparison of predicted relative soil heterotrophic respiration as a function of soil moisture. The grey area marks the full range of values obtained with Eqs. (3) and (5) using θ_s (a) or ψ_{log} (b) for 106 soil series from England and Wales ranging from 8 to 50 mg g⁻¹ SOC and 0.08 to 0.61 clay fraction. Other lines are moisture-respiration functions from existing models using either θ_s (a) or ψ_{log} (b) as a predictor. In (a) CANDY model (full line), Bethy model (dashed line), SimCycle model (dotted line), RothC model (dot-dash line). In (b) Daisy model (full line), SOILCO2 model (dashed line).

