

1 **Supplementary material**

2 **Fire-derived organic carbon turnover in soils on a centennial scale**

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11 **Methodology**

12 We calculated turnover times based on two data point for each study, the initial stock of  
13 PyC and final PyC remaining at the end of the experiment, using equation (1). Most  
14 studies had only these two data points and intermediate points that were reported in few  
15 studies (Hamer et al., 2004; Brodowski, 2005; Kuzyakov et al., 2009) were not included  
16 for consistency. We observed that turnover time for PyC ranges from <10 to 600 years  
17 (Supplementary Figure 1).

18 
$$C_t = C_0 \cdot \exp(-k \cdot t) \tag{1}$$

19 where  $C_t$  is the remaining stock after time  $t$  (y),  $C_0$  is the initial stock of PyC ( $t=0$ ),  $k$  is  
20 the decay rate ( $y^{-1}$ ) and turnover time  $\tau$  (y) =  $1/k$ .

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22 **Statistical analysis**

23 Non-parametric test for comparison between factors were carried using Wilcoxon rank  
24 sum test. A three way ANOVA was done on the data set using R software to test the  
25 interaction between the variables (supplementary table 2). Therefore, the imbalanced  
26 design of the grouped data (supplementary table 3) does not introduce any significant  
27 error in the interpretation.

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29 **Supplementary Table 1:**

<i>References</i>	Type of BC	Pyrolysis temperature	Soil Type	Climate/ Moisture Temperature	Duration of the experiment	Mineralised PyC (% of the initial PyC)
(Baldock and Smernik, 2002)	Sap wood, <i>Pinus resinosa</i>	150° C	Sand (packed at 1.6 bulk density)	At 25° C and volumetric water content of 0.29 cm <sup>3</sup> water cm <sup>-3</sup> soil	120 days	13%
		≥ 200° C (200-350° C)				<2%
(Hamer et al., 2004)	1. Maize	350° C	Sand + 1ml inoculum + 0.5ml nutrient solution	At 20° C and 60% water holding capacity	60 days	0.78%
	2. Rye	350° C				0.72%
	3. Wood	800° C				0.26%
(Brodowski, 2005) Appendix 7	1. <i>Zea mays</i> (maize) 2. <i>Secale cereale</i> (rye straw)	350° C	Top soil; Ap horizon; 0-25 cm: Haplic Phaeozem	At 20° C and 70% water holding capacity	2 years	Between 16 and 51 %
Cheng et al., 2008	Hardwood (chestnut, hickory, oak and sugar maple)	(similar to wildfires)		30° C and 70° C	1 year	2.86 % incubated at 30° C and 5.5% incubated at 70° C
(Bruun et al., 2008)	<sup>14</sup> C labeled roots of Barley ( <i>Hordeum vulgare</i> )	225° C	Sandy loam	25° C	30 days	1.9%
		300° C				1.4%
		375° C				8.2% (possibly due to Carbonates) of the charcoal-C
(Major et al., 2010)	Mango tree ( <i>Mangifera indica</i> L.)	400-600° C	Isohyperthermic kaolinitic Typic Haplustox sandy clay loam	MAT = 26° C MAP = 2200 mm (95% of precipitation falls between April and December)	2 years	2.2 % by respiration + 1% mobilized by percolating water

(Hilscher et al., 2009)	Rye grass ( <i>Lolium perenne</i> ), Gr Pine wood ( <i>Pinus sylvestris</i> ), P	350° C under oxic condition, 1 min 4 min charring	Bw horizon of Cambisol under Spruce	at 30° C	48 days	Gr1M: 3.1% Gr4M: 2.5% P1M: 0.66% P4M: 0.46%
(Kuzyakov et al., 2009)	<sup>14</sup> C labeled <i>Lolium perenne</i>	13 h charring at 400° C	Ap horizon of a loamy Haplic  luvisol	20° C 70% water holding capacity	1089 days (3.2 years)	4.5 % of the <sup>14</sup> C added as BC was  released as <sup>14</sup> CO <sub>2</sub> during 3 years. First month: 0.016- 0.024% d <sup>-1</sup> After one year: 0.0012-0.0016% d <sup>-1</sup> After 3.2 years, 0.00136% d <sup>-1</sup>
(Zimmerman, 2010)	1. <i>Quercus laurifolia</i> (living wood oak) 2. <i>Pinus taeda</i> (pine) 3. <i>Juniperus virginiana</i> (cedar) 4. <i>Guibourtia demusei</i> (tropical hardwood) 5. <i>Tripsacum dactyloides</i> (mixed stems and blades of gamma grass) 6. Sugarcane baggase	250° C oxic 400° C } 525° C } under 650° C } N <sub>2</sub>	Quartz sand	Incubated in the dark at 32 °C	1 year	0.4-3% (1.4 ± 0.75% average ± stdv) for inoculated biochars C loss 1.0 ± 0.5% for abiotic C loss  10.3 ± 6.2 SD for 100 years and 28.4 ± 20.0 SD for 100 years for microbial incubations
(Nocentini et al., 2010)	Pine needles and pine wood	350° C	Sand	20° C 50% water holding capacity	30 days	0.47% to 0.57% from burned and unburned soil inoculums

(Singh et al., 2010)	<i>Pinus ponderosa</i>	450° C	top Cambisol soil	25° C 60% water holding capacity	45 days	0.14%
(Bird et al., 1999)	<i>Terminalia</i> tree bush savanna	Wildfires	Coarse sand derived from gneissic granite bedrock	Sub-humid MAT = 17.7° C MAP = 630 mm	100 years	50%
(Hammes et al., 2008)	Steppe land cover	Wildfires	Chernozem soil	MAT = 6.6° C (1989-1998) and 5.3° C (1893-1950) MAP = 507.7 mm (1989-1998) and 438.5 mm (1893-1950)	100 years	25%
(Nguyen et al., 2008)	Forest	Wildfires	Humic Nitosols (FAO/UNESCO)	Tropical MAT = 19° C MAP = 2000 mm	100 years	70%
(Cheng et al., 2008)	Hardwood (chestnut, hickory, oak and sugar maple)	Blast furnace temperature	Subsurface soil for incubation	Aged charcoal from areas with MAT ranging between 3.9° C to 17.2° C and MAP between 940 mm to 1500 mm	130 years	22.30%
(Vasilyeva et al., 2010)	Grassland	Wildfires	Chernozem	MAT = +5.5 °C MAP = 600 mm/year	55 years	7%

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32 **Supplementary table 2: Three-way ANOVA for interactions between variables**

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	Df	Sum sq	Mean sq	F value	Pr(>F)
Matrix	2	24838	12419	11.7306	0.0001418 ***
Temperature of pyrolysis	1	35051	35051	33.1074	1.992e-06 ***
Initial biomass	2	3915	1958	1.8490	0.1733280
Medium: Temperature of pyrolysis	1	240	240	0.2263	0.6374453
Medium: initial biomass	1	34	34	0.0321	0.8588546

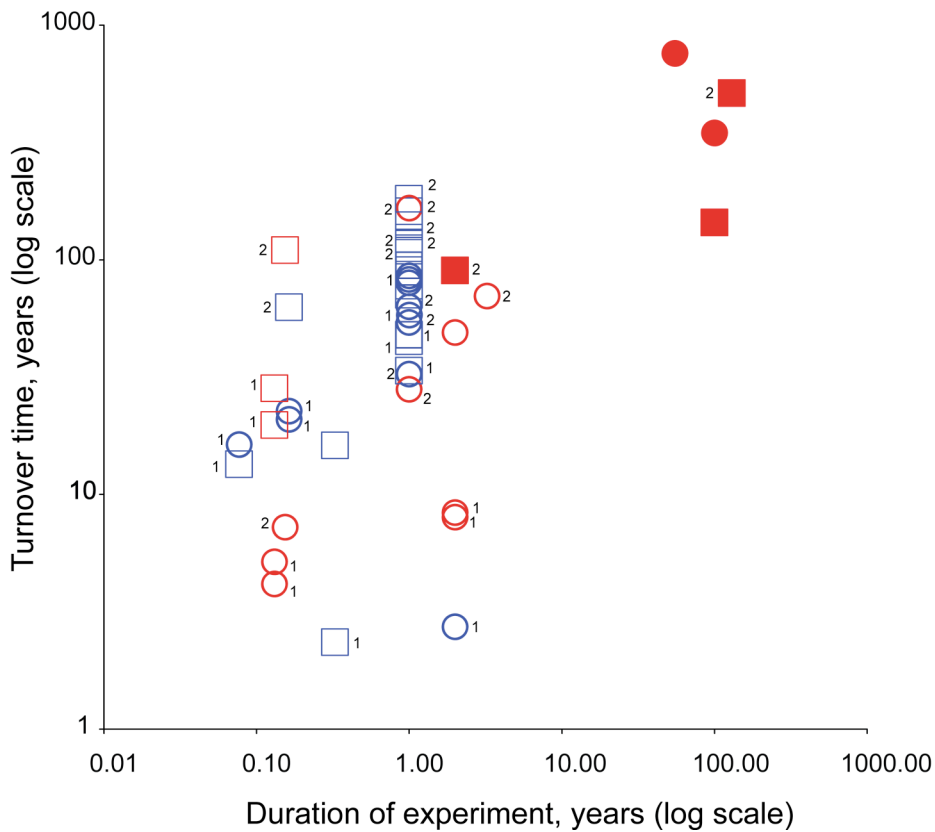
Temperature of pyrolysis: initial biomass	1	3440	3440	3.2488	0.0806158
Residuals	33	34937	1059		

Significance codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

**Supplementary table 3: Number of data point for each factor in the study**

	Incubation	Field study	Grass PyC	Wood PyC	<400°C pyrolysis temperature	≥400°C pyrolysis temperature	Sand medium	Soil medium
<b>Incubation</b>	42	0	17	22	22	20	31	11
<b>Field study</b>	0	5	0	2	0	1	0	5
<b>Grass PyC</b>	17	0	17	0	10	7	12	5
<b>Wood PyC</b>	22	2	0	24	9	15	19	5
<b>&lt;400°C pyrolysis temperature</b>	22	0	10	9	21	0	13	9
<b>≥400°C pyrolysis temperature</b>	20	1	7	15	0	21	18	3
<b>Sand</b>	31	0	12	19	13	18	31	0
<b>Soil</b>	11	5	5	5	9	3	0	12

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61 **References**

- 62 Baldock, J.A., Smernik, R.J., (2002) Chemical composition and bioavailability of  
63 thermally, altered *Pinus resinosa* (Red Pine) wood. *Organic Geochemistry*, 33,  
64 1093-1109.
- 65 Bird, M.I., Moyo, C., Veenendaal, E.M., Lloyd, J., Frost, P., (1999) Stability of elemental  
66 carbon in a savanna soil. *Global Biogeochemical Cycles*, 13, 923-932.
- 67 Brodowski, S., (2005) Origin, function, and reactivity of black carbon in the arable soil  
68 environment. *PhD Thesis, Institut für Bodenkunde, Bonn*, 183.
- 69 Bruun, S., Jensen, E.S., Jensen, L.S., (2008) Microbial mineralization and assimilation of  
70 black carbon: Dependency on degree of thermal alteration. *Organic*  
71 *Geochemistry*, 39, 839-845.
- 72 Cheng, C.H., Lehmann, J., Engelhard, M.H., (2008) Natural oxidation of black carbon in  
73 soils: Changes in molecular form and surface charge along a climosequence.  
74 *Geochimica Et Cosmochimica Acta*, 72, 1598-1610.
- 75 Hamer, U., Marschner, B., Brodowski, S., Amelung, W., (2004) Interactive priming of  
76 black carbon and glucose mineralisation. *Organic Geochemistry*, 35, 823-830.
- 77 Hammes, K., Torn, M.S., Lapenas, A.G., Schmidt, M.W.I., (2008) Centennial black  
78 carbon turnover observed in a Russian steppe soil. *Biogeosciences*, 5, 1339-1350.
- 79 Hilscher, A., Heister, K., Siewert, C., Knicker, H., (2009) Mineralisation and structural  
80 changes during the initial phase of microbial degradation of pyrogenic plant  
81 residues in soil. *Organic Geochemistry*, 40, 332-342.
- 82 Kuzyakov, Y., Subbotina, I., Chen, H.Q., Bogomolova, I., Xu, X.L., (2009) Black carbon  
83 decomposition and incorporation into soil microbial biomass estimated by C-14  
84 labeling. *Soil Biology & Biochemistry*, 41, 210-219.
- 85 Major, J., Lehmann, J., Rondon, M., Goodale, C., (2010) Fate of soil-applied black  
86 carbon: downward migration, leaching and soil respiration. *Global Change*  
87 *Biology*, 16, 1366-1379.
- 88 Nguyen, B.T., Lehmann, J., Kinyangi, J., Smernik, R., Riha, S.J., Engelhard, M.H.,  
89 (2008) Long-term black carbon dynamics in cultivated soil. *Biogeochemistry*, 89,  
90 295-308.
- 91 Nocentini, C., Guenet, B., Di Mattia, E., Certini, G., Bardoux, G., Rumpel, C., (2010)  
92 Charcoal mineralisation potential of microbial inocula from burned and unburned  
93 forest soil with and without substrate addition. *Soil Biology & Biochemistry*, 42,  
94 1472-1478.
- 95 Singh, N., Abiven, S., Schmidt, M.W.I., (2010) Mechanisms of charcoal degradation  
96 during its initial stages of decomposition. In: *European Geosciences Union*  
97 *Meeting*, Vienna, Austria.
- 98 Vasilyeva, N.A., Milanovskiy, E.Y., Abiven, S., Hilf, M., Schmidt, M.W.I., (2010) Black  
99 carbon quantity and quality unchanged after 55 years of organic matter depletion  
100 in a Chernozem. In: *SOM 2010 - Organic matter stabilization and ecosystem*  
101 *functions*.
- 102 Zimmerman, A.R., (2010) Abiotic and Microbial Oxidation of Laboratory-Produced  
103 Black Carbon (Biochar). *Environmental Science & Technology*, 44, 1295-1301.
- 104
- 105