Denitrification and inference of nitrogen sources in the 1 karstic Floridan Aquifer 2

3

J. B. Heffernan^{1,2,*}, A. R. Albertin³, M. L. Fork¹, B.G. Katz⁴, and M. J. Cohen³ 4

[1] {Department of Biological Sciences, Florida International University, Miami, FL, USA } 5

6 [2] {Southeast Environmental Research Center, Florida International University, Miami, FL, 7 USA}

[3] {School of Forest Resources and Conservation, University of Florida, Gainesville, FL, USA } 8

9 [4] {U.S. Geological Survey, Tallahasee, FL, USA}

10 Correspondence to: J. B. Heffernan (jheffer@fiu.edu)

11

12 Abstract

13 Aquifer denitrification is among the most poorly constrained fluxes in global and regional The few direct measurements of denitrification in groundwaters provide 14 nitrogen budgets. limited information about its spatial and temporal variability, particularly at the scale of whole 15 aquifers. Uncertainty in estimates of denitrification may also lead to underestimates of its effect 16 17 on isotopic signatures of inorganic N, and thereby confound the inference of N source from these data. In this study, our objectives are to quantify the magnitude and variability of denitrification 18 19 in the Upper Floridan Aquifer (UFA) and evaluate its effect on N isotopic signatures at the 20 regional scale. Using dual noble gas tracers (Ne, Ar) to generate physical predictions of N₂ gas concentrations for 112 observations from 61 UFA springs, we show that excess (i.e. 21 denitrification-derived) N₂ is highly variable in space and inversely correlated with dissolved 22 oxygen (O₂). Negative relationship between O₂ and $\delta^{15}N_{NO3}$ across a larger dataset of 113 23 springs, well-constrained isotopic fractionation coefficients, and strong ¹⁵N:¹⁸O covariation 24 further support inferences of denitrification in this uniquely organic-matter-poor system. Despite 25 relatively low average rates, denitrification accounted for 32% of estimated aquifer N inputs 26 across all sampled UFA springs. Back-calculations of source $\delta^{15}N_{NO3}$ based on denitrification $\sqrt{5}$ 27

Amit

progression suggest that isotopically-enriched nitrate (NO₃⁻) in many springs of the UFA reflects
groundwater denitrification rather than urban- or animal-derived inputs.

30

31 **1 Introduction**

Anthropogenic increases in reactive nitrogen (N) availability have wide-ranging 32 consequences including eutrophication of aquatic systems, acidification of soils and surface 33 waters, loss of biodiversity, and facilitation of disease transmission (Vitousek, 1994; Galloway et 34 al., 2003; Smith and Schindler, 2009). Denitrification, which reduces NO₃⁻ to N₂ gas, mitigates 35 this enrichment by returning N to long-residence-time atmospheric pools, and is an important 36 37 component of the nitrogen cycle at local, regional, and global scales (David et al., 2006; Seitzinger et al., 2006; Townsend and Davidson, 2006; Schlesinger, 2009; Sigman et al., 2009). 38 39 Although denitrification was once thought to occur only via the oxidation of simple organic compounds, more recent work has demonstrated that NO₃⁻ reduction can involve multiple 40 electron donors and end products (Burgin and Hamilton, 2007). Patchy and ephemeral 41 distribution, diverse reaction modes, and challenges of direct measurement of N₂ all contribute to 42 persistent high uncertainty in local, regional, and global estimates of denitrification (Davidson 43 44 and Seitzinger, 2006; Groffman et al., 2009).

Aquifer denitrification is a potentially large component of regional and global nitrogen 45 (N) budgets, with a recent global estimate of 44 Tg N yr⁻¹ (16% of land-based annual N inputs; 46 (Seitzinger et al., 2006). However, existing estimates are based on simple models and are 47 extremely poorly constrained (range of estimates from Seitzinger et al., [2006]: 0-138 Tg N yr⁻¹), 48 in large part due to the limited number and spatio-temporal extent of available direct 49 measurements of denitrification. Additional uncertainty arises because existing studies of 50 groundwater N concentrations and denitrification are potentially biased by preferential study of 51 aquifers with high N loading and high rates of denitrification (Green et al., 2008; Schlesinger, 52 2009). Moreover, measures of denitrification based on nitrate loss appear to provide much 53 higher estimates than those based on direct measurement of N₂ gas accumulation (Green et al., 54 2008). Despite their limited numbers, directly-measured aquifer denitrification rates nonetheless 55 span several orders of magnitude, and associated reductions in NO₃⁻ range from negligible to 56

complete across aquifer systems (Green et al., 2008). The extent to which these outcomes vary in
space and time within individual aquifers is poorly understood.

59 Estimation of denitrification from N₂ gas concentrations in groundwaters requires 60 determination of physical parameters (recharge temperature [T_{rec}] and excess air [A_{ex}]) that influence the quantity and composition of dissolved gases (Vogel et al., 1981; Wilson and 61 McNeill, 1997; Cey et al., 2009). Recharge temperature, rather than ambient temperature at the 62 63 time of sample collection, is what determines the solubility of atmospheric gases at the time when infiltrating groundwater loses contact with the atmosphere. Depending on the seasonality 64 of temperature, precipitation, and infiltration, as well as other factors, recharge temperatures can 65 diverge substantially from mean annual air temperatures (Hall et al., 2005; Castro et al., 2007). 66 67 Dissolution of excess air occurs when bubbles of atmospheric air are entrained beneath the saturated zone; supersaturation of gases with respect to surface conditions is enabled by 68 69 hydrostatic pressure at depth. Direct simultaneous estimation of these parameters requires two tracers (typically noble gases; Feast et al., 1998; Cey et al., 2009), but most measurements of N₂ 70 are made only in conjunction with Ar (Kana et al., 1994). Studies that estimate denitrification 71 72 via direct measurement of N₂ thus typically rely on assumed constant values of either recharge temperature or excess air entrainment to estimate biologically-derived N₂ (e.g., Green et al., 73 2008). Since recharge temperature and excess air entrainment can vary at broad scales due to 74 variation in climate and geological structure, assessment of denitrification at regional or broader 75 scales requires estimation of these physical parameters for each study site. - prite 76

Across diverse ecosystems, availability of organic matter is the primary driver of 77 denitrification (Taylor and Townsend, 2010). Within ecosystems, spatial and temporal 78 variability in the concentration of organic matter and nitrate and anoxic conditions produce 79 heterogeneous mosaics of denitrifying activity (e.g., Harms and Grimm, 2008). In groundwater 80 environments with strong directional flow, denitrification and other redox processes can follow 81 distinctive spatial patterns reflecting the gradual downstream depletion of electron donors and 82 acceptors (Chapelle et al., 1995; Hedin et al., 1998; Tarits et al., 2006). However, a growing 83 body of research suggests that denitrification in most aquifers depends on matrix-derived, solid-84 phase electron donors (e.g., Fe²⁺, H₂S) rather than surface-derived solutes (Green et al., 2008; 85 Schwientek et al., 2008; Zhang et al., 2009; Torrento et al., 2010; Torrento et al., 2011). As a 86 result, concentrations of dissolved organic matter and other electron donors may be a poor 87

88 indicator of denitrification rates across aquifers, and spatial patterns within aquifers may reflect

- 89 the distribution of these reactants within the aquifer matrix rather than substrate depletion along
- 90 advective flowpaths.

91 Efforts to understand and manage N enrichment of aquatic ecosystems have relied 92 heavily on the distinctive isotopic signatures of potential sources (Kendall, 1998; Kendall et al., 2007), particularly the difference in $\delta^{15}N_{NO3}$ between inorganic fertilizers (typically 0-3‰) and 93 organic N pools (i.e. animal and human waste; typically 9-12‰). More recently, methodological 94 developments that permit determination of both N and O allow greater separation of sources with 95 ¹⁵N signatures (e.g., atmospheric deposition and inorganic fertilizer). 96 overlapping 97 Biogeochemical reactions such as denitrification and assimilation can alter these isotopic signatures during transport along soil-ocean flowpaths, potentially confounding N source 98 99 inference (Kendall et al., 2007). Despite pleas for caution (Bedard-Haughn et al., 2003), observed $\delta^{15}N_{NO3}$ are commonly used to infer N sources and guide management and policy 100 related to point and non-point inputs (Fogg et al., 1998;Survey, 2003; Harrington et al., 2010). 101 102 While the potential effect of denitrification on isotope signatures is widely acknowledged, few 103 studies to date have quantified its influence on source inference at the scale of a regional aquifer.

In addition to their utility in separating potentially counfounded N sources, dual isotopic 104 tracers ($\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$) of NO₃⁻ can also be used to infer nitrogen transformations. 105 Although insufficient to directly estimate rates of denitrification, coupled enrichment of $\delta^{15}N_{NO3}$ 106 and $\delta^{18}O_{NO3}$ are now widely used to infer the occurrence of fractionating processes (Burns et al., 107 108 2009). Among these are recent studies that suggest denitrification in the Upper Floridan Aquifer 109 and other karst groundwater systems (Panno et al., 2001; Albertin et al., 2011). One potential 110 value of dual isotopic measurements is the ability to partition removal into its component processes (e.g. assimilation, denitrification) based on the ratio of ¹⁵N:¹⁸O enrichment. 111 112 Theoretical and laboratory studies have suggested that denitrification results in 2:1 fractionation of ¹⁵N:¹⁸O (Aravena and Robertson, 1998; Lehmann et al., 2003), but other studies have recently 113 suggested a 1:1 ratio (Granger et al., 2008), in which case dual isotopes would be unable to 114 115 distinguish between assimilation and denitrification.

116 In this study, our objectives are (1) to quantify the magnitude and variability of 117 denitrification at the regional scale in a karstic groundwater system (the Upper Floridan Aquifer

[UFA]), and (2) to assess the influence of denitrification on isotopic signatures of nitrate in the 118 UFA and its influence on apparent N sources. To these ends, we measured dissolved gases and 119 other biogeochemical characteristics of 35 Florida springs, conducted a 3 year study (quarterly-120 monthly samples) of 6 springs that feed the Ichetucknee River, and assembled published data 121 describing dissolved gas concentration, nutrient chemistry, and/or nitrate isotope composition 122 from over 100 additional springs. From a subset of 31 of these springs for which dual noble gas 123 tracers (Ne and Ar) were available, we derived statistical predictors of excess air entrainment. 124 These data enable an extensive and robust assessment of denitrification and its influence on 125 nitrate isotopic composition at the scale of the entire UFA. 126

127 2 Methods

Study System 128 2.1

The karstic Upper Floridan Aquifer (UFA) supports the highest density of large natural 129 artesian springs in the world (Fig. 1), and is a major regional and global economic resource 130 (Notholt et al., 1989; Miller, 1990; Bonn and Bell, 2003; Bonn, 2004). Throughout parts of 131 northern Florida, the UFA is confined by low-permeability, high-clay deposits that preclude 132 infiltration except via sinkholes and fractures; these confining layers are largely absent in the 133 central-western portion of the state (Scott et al., 2004). Springs are concentrated along drainage 134 features, especially near boundaries of confining layers. Land use throughout the study region 135 includes variable mixtures of row crop agriculture, urban and suburban development, and 136 secondary forest (Katz et al., 2001). 137

Geochemistry of the UFA can be characterized as a mixture of two end members. Older 138 water, characteristic of matrix porosity and deep flowpaths, is generally anoxic, low in NO₃, and 139 enriched in minerals; younger water characteristic of conduits and shallows flowpaths is 140 generally oxic, enriched in NO3, and sometimes subsaturated in mineral chemistry (Toth and Vague 141 Katz, 2006). Over event-driven and decadal timescales, the contribution of these water sources 142 can vary considerably among springs as changes in flow drive exchange between primary and 143 secondary porosity (i.e., the limestone matrix and karst conduits; Martin and Dean, 2001; 144 Heffernan et al., 2010a,b). Except during runoff and backflow events that deliver organic-145 matter-rich waters to conduits (Gulley et al., 2011), dissolved organic carbon (DOC) levels in 146

5

L

147 UFA springs are among the lowest measured globally (Duarte et al., 2010). In conjunction with 148 oxic conditions of many springs, low DOC concentrations undoubtedly contribute to the 149 prevailing assumption that denitrification is negligible in this system (Katz, 2004).

 NO_3^- concentrations in Florida springs have risen dramatically over the past half-century, 150 151 and springs discharge accounts for a large proportion of the N load to estuarine and coastal 152 waters (Pittman et al., 1997). Despite the perceived vulnerability of the UFA to nutrient 153 enrichment, significant imbalances between inputs to North Florida landscapes and riverine 154 export remain poorly understood (Katz et al., 2009). Landscape-scale mass balance generally suggests inorganic fertilizer as the primary source of N enrichment (Katz et al., 2009), but 155 isotopic studies (that assumed negligible denitrification) have indicated a greater role of organic 156 N from animal or human wastewater (Katz et al., 2001). 157

158 **2.2 Sample collection and analysis**

159 Between June and September 2010, we sampled 33 Floridan Aquifer springs that varied in size, surficial hydrogeology, and NO₃⁻ and O₂ concentrations. At each spring, we measured 160 O₂, temperature, specific conductance, and pH from spring vents using a YSI 556 sonde 161 equipped with an optical or Clark probe. Water samples for laboratory analyses were collected 162 163 using a peristaltic pump with a 5 m weighted intake tube placed as near as possible to the spring vent. We collected 3 replicate samples for nutrient and isotopic analyses in acid-washed pre-164 rinsed polyethylene bottles. During the synoptic survey, we collected 5 replicate field samples 165 for dissolved gas analysis by flushing 300 ml BOD bottles 3 times, sealing with glass stopper, 166 167 and capping with water-filled plastic caps to minimize exchange with atmosphere and to prevent 168 stoppers from becoming dislodged during transport. Dissolved gas samples were stored under 169 ice water until analysis within 36 hours; water samples were frozen until analysis.

We measured dissolved N_2 and Ar using a Membrane Inlet Mass Spectrometer (MIMS: Kana et al., 1994) within 36 hours of collection, over which period our storage protocol exhibited negligible atmospheric contamination. The membrane inlet mass spectrometer was equipped with a copper reduction column heated to 600 °C to remove O_2 and reduce interference with N_2 measurements (Eyre et al., 2002). Standards for N_2 and Ar concentration consisted of atmosphere-equilibrated deionized water in 1 L spherical vessels incubated and stirred in high-

176 precision water baths (\pm 0.01 °C) at their respective temperatures (10, 15, and 20 °C) for at least 24 hours prior to analysis. Gas concentrations in each standard were calculated using 177 temperature-solubility formulas without salinity correction (Hamme and Emerson, 2004). 178 Signal strength for samples and standards was determined as the mean value of the 1st minute 179 following signal stabilization. To account for instrument drift, we ran complete standard curves 180 every 6-8 samples and applied interpolated parameter values from adjacent standard curves (r^2 181 range: 0.997-1.00; mean $r^2 = 0.9997$) to estimate gas concentrations in each sample. A fourth 182 standard equilibrated with pure N₂ gas served as an external source QC. Coefficients of variation 183 for field replicates ranged from 0.22-2.27% (mean: 0.80%; median: 0.49%). 184

We measured nitrate concentrations (expressed in this paper in mg N L⁻¹) in samples from the synoptic survey and Ichetucknee River springs time, series using second-derivative UV spectroscopy (APHA et al., 2005) using an Aquamate UV-Vis spectrometer. Isotopic composition of nitrate ($\delta^{15}N_{NO3}$, $\delta^{18}O_{NO3}$) was measured using the bacterial denitrifier method (Sigman et al., 2001; Casciotti et al., 2002) in the Department of Geological Sciences at the University of Florida (2007-2009) or the UC-Riverside Facility for Isotope Ratio Mass Spectrometry (2010).

Previously collected data both increased spatial coverage and in many cases provided 192 repeated measurements of springs included in our synoptic survey (Fig. 1). Measurements of 193 Ne, Ar, N₂, O₂, NO₃, and nitrate isotopes ($\delta^{15}N_{NO3}$, $\delta^{18}O_{NO3}$) spanning from 1997 to 2008 were 194 195 obtained from published articles (Katz, 2004; Katz et al., 2001, 2004; Toth and Katz, 2006; Knowles et al., 2010) and agency reports (Katz et al., 1999; Phelps, 2004; Phelps et al., 2006; 196 Chasar et al., 2005), or directly from researchers when dissolved gas concentration or other data 197 were not reported directly. Thirty-six archival observations (from 31 springs) included Ne in 198 addition to Ar, O₂, and N₂ (and in 23 cases $\delta^{15}N_{NO3}$). In all, we assembled 112 observations of 199 dissolved gas concentrations (O2, Ar, N2) from 62 distinct spring vents, of which 58 included 200 both $\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$, and 34 others included $\delta^{15}N_{NO3}$ but not $\delta^{18}O_{NO3}$. Excluding the 201 repeated measurements of the Ichetucknee River springs in 2008-2009, our data included 166 202 observations of $\delta^{15}N_{NO3}$ and O₂ and 204 total observations of $\delta^{15}N_{NO3}$. Of the 113 springs 203 represented in the isotope data set, 14 had 4 or more instances of concurrent measurements of 204 both O_2 and $\delta^{15}N_{NO3}$. Observations were drawn from springs in each major drainage in North 205 and Central Florida (Fig. 1), and with discharges ranging from $<0.01 \text{ m}^3 \text{ s}^{-1}$ to $11 \text{ m}^3 \text{ s}^{-1}$. 206

2

All measurements of Ne from prior studies were determined by mass spectrometry at the Lamont-Doherty Earth Observatory Noble Gas Laboratory at Columbia University. NO₃ was generally measured using the cadmium reduction method (Wood et al., 1967). N₂ and Ar from previously published studies were measured using gas chromatography. Recent (2007 and later) measurements of $\delta^{15}N_{NO3}$ were generally conducted using the bacterial denitrifier method and included $\delta^{18}O_{NO3}$ (Sigman et al., 2001; Casciotti et al., 2002). For data prior to 2007, $\delta^{15}N_{NO3}$ was measured via combustion and mass spectrometry (Kendall and Grim, 1990).

214

4 2.3 Springshed characterization

215 To determine hydrogeologic predictors of excess air entrainment and recharge 216 temperature, we characterized each spring by latitude, long-term mean discharge, and springshed 217 hydrogeology as measured by aquifer vulnerability to surface contamination (Arthur et al., 218 2007). We collected discharge records for each spring from online databases of the United States Geological Survey National Water Information System (http://waterdata.usgs.gov/nwis), 219 Southwest Florida Water Management District (http://www.swfwmd.state.fl.us/data/), and the St. 220 221 John's River Water Management District (http://www.sjrwmd.com/toolsGISdata/index.html) where available, since these records were generally the most complete. Where continuous 222 223 records were unavailable, we used the mean of discrete measurements from published studies 224 and agency reports as our estimate of mean long-term discharge. Since discharge variability of 225 Floridan Aquifer springs is extremely low, use of these more limited data to quantify long-term mean discharge is unlikely to have introduced significant error in subsequent analyses. 226

227 Where available, previously delineated boundaries we used (http://www.dep.state.fl.us/geology/programs/hydrogeology/hydro resources.htm) 228 to 229 characterize springshed hydrogeology, and to estimate springshed size and location for un-230 delineated springs. The relationship between discharge and springshed area was determined for those springs with previously delineated springsheds ($A = Q \times 134.9$, where A is springshed area 231 in km² and Q is discharge in m³ s⁻¹; n = 14, r = +0.79, p < 0.001). For springs without a delineated 232 springshed, we estimated the contributing area based on their period-of-record discharge. We 233 234 assumed each springshed was circular, and estimated the springshed orientation based on the regional drainage network such that the springshed was located with one edge at the spring vent, 235 236 and the rest up-gradient from the closest spring-fed river.

237 We used the Floridan Aquifer Vulnerability Assessment (FAVA) as a metric of springshed hydrogeologic characteristics (Arthur et al., 2007). This measure quantifies the 238 intrinsic contamination risk of the Upper Florida Aquifer (UFA) based on local hydrogeologic 239 conditions. Point observations of aquifer properties diagnostic of rapid recharge rates (e.g., 240 nutrient and major element chemistry, high O_2 concentrations) comprise a data set (n = 148) on 241 which aquifer risk was trained using a weights-of-evidence approach based on a variety of 242 spatially extensive data layers. These included surface soil permeability, surface elevation, 243 subsurface stratigraphy, presence of karst features (e.g., sinkholes) at the surface, thickness of a 244 the intermediate aquifer system that regulates hydraulic confinement of the UFA, and the 245 246 potentiometric head difference between the surface and UFA interpolated from a regional well Posterior contamination probabilities were classified as "less vulnerable", 247 network. 248 "vulnerable" and "most vulnerable". None of the springsheds in this study contained more than 3% of their area in the less vulnerable category, so we used the fraction of each springshed area 249 delineated as most vulnerable, typically more than 75% of the area, as a predictor in our model 250 251 of excess air entrainment.

252

2.4 Estimation of excess air, recharge temperature, and excess N₂

253 We used Ne and Ar concentrations to determine recharge temperature (T_{rec}) and excess air (A_{ex}) in the subset (n = 36) of springs for which measurements of both gases were available. 254 We estimated these parameters for each observation air by simultaneously solving the following 255 equations using the Solver function in Microsoft Excel: 256

257
$$(Ne]_{obs} = k_{Ne} \cdot [Ne]_{Trec} + A_{ex} \cdot P_{Ne}$$
(1)

 $\left[\operatorname{Ar}\right]_{obs} = k_{\operatorname{Ar}} \cdot \left[\operatorname{Ar}\right]_{Trec} + A_{\operatorname{ex}} \cdot P_{\operatorname{Ar}}$ (2)

where k_{Ne} and k_{Ar} are coefficients for unit conversion of Ne and Ar from nmol kg⁻¹ (for Ne, k =259 0.02) or μ mol kg⁻¹ (for Ar, k = 0.04; for N₂, k = 0.028) to mg L⁻¹; P_{Ne} and P_{Ar} are the mass 260 proportion of Ne (1.818 \cdot 10⁻⁵) and Ar (9.34 \cdot 10⁻³) in the atmosphere; and [Ne]_{Trec} and [Ar]_{Trec} are 261 the equilibrium concentrations (Hamme and Emerson, 2004) of those gases at the recharge 262 263 temperature as determined by:

264
$$\ln([Ne]_{Tree}) = A_0 + A_1 T_5 + A_2 T_5^2 + A_3 T_5^3$$
 (3)

265
$$\ln([Ar]_{T_{rsc}}) = A_0 + A_1 T_s + A_2 T_s^2 + A_3 T_s^3$$
 (4)

266 where T_S is determined as:

267
$$T_{S} = \ln\left(\frac{298.15 - T_{FEC}}{273.15 + T_{FEC}}\right)$$
(5)

and A_0 - A_3 are compound specific solubility constants (Table 1).

Among this set of springs with Ne data, estimated T_{rec} ranged from 15-22°C, was overwhelmingly determined by Ar rather than Ne, and varied significantly as a function of latitude (Fig. 2a-c). The observed latitudinal variation in T_{rec} is much greater than variation in mean annual air temperature, potentially reflecting regional variation in timing of precipitation and thus temperature of infiltrating water (Schmidt et al., 2001).

A_{ex} ranged from $1.0^{-2.7}$ ml L⁻¹ and was overwhelmingly determined by Ne rather than Ar (Fig. 2a,b). Based on multiple regression analysis, mean discharge over the period of record (Q_{POR}) and springshed vulnerability were strong predictors of excess air (Fig. 2d). Palm spring, whose springshed had no land in the 'most vulnerable' category and was the only value less than 50%, was excluded from this analysis. We used this statistical relationship to estimate A_{ex} in springs for which Ne data were not available, then solved for recharge temperature in those springs using Eq. 1.

We used estimates of T_{rec} and A_{ex} , whether direct or modeled statistically, to determine N₂ concentrations that would be observed based only on those physical processes ([N₂]_{phys}), without any biological N₂ production. Specifically, we calculated expected N₂ concentrations using the same temperature-solubility relations described in Eq. 1-5, but parameterized for N₂:

285
$$[N_2]_{phyz} = k_{N2} \cdot [N_2]_{Trec} + A_{ex} \cdot P_{N2}$$
(6)

where k_{N2} is 0.028; P_{N2} is 0.78084 and $[N_2]_{Trec}$ is the concentration of N_2 at recharge temperature as determined by:

288
$$\ln([N_2]_{Tree}) = A_0 + A_1 T_5 + A_2 T_5^2 + A_3 T_5^3$$
(7)

where T_s is given in Eq. 5 and N_2 -specific values for the A_0 - A_3 are given in Table 1. We estimate the magnitude of denitrification ($[N_2]_{den}$) for each sampling date and time as the difference between observed N_2 concentrations and concentration predicted by physical processes:

292
$$[N_2]_{den} = [N_2]_{obs} - [N_2]_{phys}$$
(8)

To test the hypothesis that $[N_2]_{den}$ reflects the magnitude of denitrification, we used regression analyses to evaluate the relationship between $[N_2]_{den}$ and dissolved O_2 . We used both linear and logarithmic forms to predict $[N_2]_{den}$ from O_2 , for the entire data set and for the subset of observations in which A_{ex} was calculated from Ne data, rather than estimated statistically. We also analyzed the relationship between mean dissolved O_2 and mean $[N_2]_{den}$ from the subset of springs for which 3 or more observations were available.

To evaluate the relative precision and accuracy of $[N_2]_{den}$ estimates based on Ne and statistically modeled excess air, we calculated the mean and standard deviation of $[N_2]_{den}$ estimates for all springs with O₂ greater than 2 mg L⁻¹. Bias in estimates would cause divergence of the mean from zero, assuming that denitrification is negligible in these oxic springs (Bohlke et al., 2002; Green et al., 2008).

304 **2.5 Denitrification progression and isotopic fractionation** $\sim M \swarrow H$

We indirectly evaluated the relationship between denitrification progression and $\delta^{15}N_{NO3}$ 305 via analysis of relationships between dissolved O₂ and $\delta^{15}N_{NO3}$ both within and across springs, 306 reasoning that springs with lower dissolved O2 would have greater depletion of NO3⁻ pools by 307 denitrification than springs with higher O2. We used both linear and logarithmic regression 308 equations to evaluate dissolved O₂ as a predictor of $\delta^{15}N_{NO3}$ across all observations and 309 excluding observations from the Ichetucknee time series collected between July 2007 and 310 November 2009. Inclusion of the entire Ichetucknee data set had a minimal influence on 311 regression parameters, so only the results from the complete (global) data set are reported here. 312 In addition to this global analysis, we used linear regression to evaluate relationships between 313 dissolved O_2 and $\delta^{15}N_{NO3}$ within springs for which 3 or more observations were available. We 314

then used regression analysis to evaluate how the strength (as measured by the correlation coefficient [r]) and slope of these within-spring relationships varied as a function of mean dissolved O₂. This analysis allowed us to evaluate the contribution of variation within and among springs to patterns seen across all observations.

We directly evaluated the relationship between denitrification progression and isotopic 319 composition of NO₃⁻ by determining the fractionation coefficient ($^{15}\varepsilon$) for $\delta^{15}N_{NO3}$ from a cross-320 system analysis that included springs with dissolved gases from both our synoptic survey and 321 previously reported data, and a separate analysis from the Ichetucknee Springs time series (of 322 which most dates did not include dissolved gas measurements). These analyses required 323 estimates of initial NO₃⁻ concentration ([NO₃⁻]_{init}) at the time of recharge, which we estimated 324 using different approaches for springs with dissolved gas data and for the Ichetucknee Springs 325 time series. For analysis of data from the synoptic survey and previous observations that 326 included dissolved gases, we calculated [NO3] init as the sum of [NO3] obs and [N2] den (all in units 327 of mg N L⁻¹). This estimate would include nitrate derived from nitrification in the vadose zone 328 or UFA as part of [NO3]_{init}, and assumes that denitrification is the only sink for NO3 (i.e., that 329 assimilation, dissimilatory nitrate reduction to ammonium [DNRA], etc. are negligible) as 330 indicated by concentrations of ammonium and particulate and dissolved organic nitrogen that are 331 typically below detection limits at spring vents. Effects of these processes on $\delta^{15}N_{NO3}$ are also 332 333 assumed to be zero.

Estimates of $[NO_3^-]_{init}$ allow determination of the progression of denitrification. For each observation, we calculated the proportion of nitrate remaining from the original pool $[NO_3^-]_R$ as:

$$[NO_3^-]_R = \frac{[NO_3^-]_{obs}}{[NO_3^-]_{init}}$$
(9)

where $[NO_3^-]_{obs}$ is measured concentration, and $[NO_3^-]_{init}$ is the initial concentration. We used linear regression to determine the isotopic enrichment factor $(^{15}\varepsilon)$ for $\delta^{15}N_{NO3}$ (assuming Rayleigh distillation kinetics) as the slope of the relationship between $\delta^{15}N_{NO3}$ and $ln([NO_3^-]_R)$ (Mariotti, 1986; Bohlke et al., 2002; Green et al., 2008). We excluded springs with $NO_3^$ concentrations below 0.1 mg L⁻¹ (Juniper, Silver Glen, and Alexander Springs) from this analysis due to the high variability of $[N_2]_{den}$ estimates relative to these lower concentrations.

how can it be estimated

12

For the Ichetucknee Springs time series, we calculated [NO3-]R for each spring and 343 sampling date by assuming that [NO3] init for all springs was equal to [NO3] obs in the Ichetucknee 344 Headspring on the same date. The first assumption implicit in this analysis is that denitrification 345 rates in the Ichetucknee Headspring are negligible. High O_2 concentrations (mean \pm SD: 4.1 \pm 346 0.2 mg O₂ L⁻¹), low values of $[N_2]_{den}$ (which averaged 0.32 mg N L⁻¹ and represented minimal 347 (<30%) depletion of the estimated original nitrate pool), and the low and temporally stable 348 δ^{15} N_{NO3} (mean ± SD: 3.6±0.3, n = 16) observed in the Ichetucknee Headspring all support this 349 assumption. The second implicit assumption, that springsheds of the Ichetucknee springs receive 350 equivalent N loads from sources with identical isotopic signatures, is based on the relative 351 homogeneity of land use in the Ichetucknee springshed, and the predominance of fertilizer 352 application to improved pasture as a source of N to the watershed (Katz et al., 2009). The third 353 assumption, that variation among springs of the Ichetucknee is driven by denitrification, is 354 supported by strong correlations between dissolved O2 and NO3 within and across these systems 355 356 (Fig. 3a).

To further evaluate the latter two assumptions, we used $[NO_3]_{init}$ and $[NO_3]_R$ values for 357 the springs of the Ichetucknee River on three dates when [N₂] den estimates were available. First, 358 we compared [NO₃]_{init} from the Ichetucknee Headspring with the mean value of other springs on 359 the same date. The similarity and covariation of these values (Table 2) is consistent with the 360 assumption that all springs in the Ichetucknee System receive similar N loads. In addition, we 361 assessed the correlation between alternative estimates of [NO3]_R, namely estimates calculated 362 from [N₂]_{den} and NO₃⁻ from reach spring and those estimated from the differences in NO₃⁻ 363 concentration between each spring and the Ichetucknee Headspring (Fig. 3a). The relationship 364 between these estimates (Fig. 3b) suggests that the NO₃⁻ difference approach used in the analysis 365 of the Ichetucknee Springs time series provides a more conservative estimate of the progression 366 of denitrification than those determined using $[N_2]_{den}$, which is to be expected if denitrification 367 has also reduced NO₃⁻ concentrations to a small degree in the Ichetucknee Headspring. More 368 importantly, the correlation between these estimates is consistent with the assumption that 369 variation in NO3⁻ concentration both within and among the springs of the Ichetucknee River is 370 driven at least in part by differences in the progression of denitrification along the flowpaths that 371 372 contribute to these springs.

373

~

374 **3 Results**

For the vast majority of our observations, N2 concentrations exceeded values predicted 375 from recharge temperature and excess air (51 of 61 springs; 94 of 112 observations). [N₂]_{den} 376 ranged from -0.7 to 3.5 mg N₂ L⁻¹ (median: 0.67 mg N₂ L⁻¹; mean \pm SD: 0.82 \pm 0.83 mg N₂ L⁻¹), 377 and was inversely correlated with O₂ (Fig. 4a). Among springs with Ne data, this relationship 378 exhibited a sharp break at ca. 2 mg $O_2 L^{-1}$, above which $[N_2]_{den}$ averaged 0.003 mg $N_2 L^{-1}$ (± 379 0.32; 2 SE); below 2 mg O₂ L⁻¹, $[N_2]_{den}$ averaged 1.5 mg N₂ L⁻¹ (±0.33; 2 SE). Among all 380 springs, this threshold was less distinct, and a linear relationships was a better fit than a 381 382 logarithmic relationship.

Among the 16 springs with 3 or more observations (max = 5) of $[N_2]_{den}$ in our data-set, 383 384 over 90% of the total variation in [N₂]_{den} occurred among rather than within springs (ANOVA; 385 Figure 5a, Table 3). Standard deviations within springs for [N₂]_{den} ranged from 0.05 to 0.65 mg $N_2 L^{-1}$ and averaged 0.31 mg L⁻¹. Among the same set of springs, over 88% of total variation in 386 [NO₃]_R occurred among springs (ANOVA; Fig. 5b, Table 3). Standard deviations within springs 387 388 for $[NO_3]_R$ ranged from <1% to 29% and averaged 10.4%. For both variables, variation among 389 springs was strongly correlated with mean dissolved O₂ from the same set of observations. 390 However, variation in $[N_2]_{den}$ and $[NO_3]_R$ within springs was not correlated with variation within 391 springs in dissolved O₂, presumably due to low sample size

Fluxes of [N2]den from UFA springs were comparable to but uncorrelated with those of 392 NO₃⁻, and the proportion of NO₃⁻ removed by denitrification varied among springs from 0 to as 393 394 high as 97% (mean ± 2 SE: 34 $\pm 9\%$) among springs. Denitrification removed more than 75% of 395 N inputs in 8 of 61 springs, and more than 50% in 20 of 61. Compared to this spatial 396 heterogeneity, temporal variation in [N₂]_{den} among springs was low (Fig. 5, Table 3). We 397 estimate that denitrification reduced total flow-weighted NO₃⁻ flux from sampled UFA springs by 32%, with uncertainty in this estimate primarily driven by the representativeness of our 398 399 sample of springs. Volumetric denitrification rates calculated from N_{2den} and apparent He water age (Green et al. 2008) averaged of 2.75 µmol m⁻³ d⁻¹. Aggregate (i.e. flow-weighted) areal 400 denitrification for all springsheds was 1.22 kg ha⁻¹ y⁻¹, with ca. 20% of springsheds exceeding the 401 estimated global average for groundwater denitrification (3.49 kg ha⁻¹ y⁻¹). 402

Across 292 observations from 103 springs, $\delta^{15}N_{NO3}$ ranged from -0.3 to 23.9%, was 403 inversely correlated with O2, and varied more among low-O2 (<2 mg L⁻¹) than high-O2 springs 404 (Fig. 4b). Among springs with 3 or more observations (max = 18) of $\delta^{15}N_{NO3}$, 86% of the total 405 variation in $\delta^{15}N_{NO3}$ in our data set was accounted for by variation among springs (ANOVA; Fig. 406 6, Table 3). Standard deviations of $\delta^{15}N_{NO3}$ within individual springs ranged from 0.1 to 5.2% 407 and averaged 1.4‰. Unlike $[N_2]_{den}$ and $[NO_3]_R$, variation within springs in $\delta^{15}N_{NO3}$ was also 408 409 correlated with variation in dissolved O₂, particularly for low O₂ springs, which had strong, steeply negative relationships between temporal variation in $\delta^{15}N_{NO3}$ and dissolved O₂ (Fig. 7). 410

The progression of denitrification, as indicated by $[NO_3]_R$, was a strong predictor of 411 $\delta^{15}N_{NO3}$ in both the synoptic survey and Ichetucknee River time series (Fig. 8a,b). Estimated 412 fractionation coefficients were similar between the Ichetucknee River time series and the 413 414 synoptic survey, particularly when only springs with Ne data were considered. Both data sets also exhibited strong relationships between nitrate $\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$, however, the slope of 415 this relationship differed substantially between the synoptic cross-system survey (1.8:1) and the ~416 Y 417 Ichetucknee River time series (1:1; Fig. 8c,d). For the Ichetucknee River springs, relationships between $\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$ within sampling dates were consistently near unity (Table 4). 418

Estimates of source $\delta^{15}N_{NO3}$ from denitrification progression and observed $\delta^{15}N_{NO3}$ values 419 suggest that denitrification alters $\delta^{15}N_{NO3}$ at the regional scale. Among springs with estimates of 420 N_{2-den} , nearly 20% of observed $\delta^{15}N_{NO3}$ values were greater than 9‰, and more than 50% were 421 greater than 6‰ (Fig. 9%c), values used in Florida and elsewhere to delineate inorganic and 422 organic sources and mixtures thereof (Katz 2004, Bohlke 2002). Estimated $\delta^{15}N_{NO3}$ of source N 423 (Fig. 9d) were much lower, with only 5.5% of observations estimated to have original source 424 $\delta^{15}N_{NO3}$ greater than 9‰, and 26% greater than 6‰. Within the Ichetucknee River time series, 425 differences between the distribution of observed and estimated source $\delta^{15}N_{NO3}$ were even greater 426 (Fig. 9(f)) $\delta^{15}N_{NO3}$ values greater than 6‰ were rare among springs with DO greater than 3 mg 427 L^{-1} (Fig. 9a,b). In all three data sets, estimated source $\delta^{15}N_{NO3}$ was both lower on average and 428 429 much less variable than spring water.

430

431 4 Discussion

432

4.1 Evidence for Denitrification in the Upper Floridan Aquifer

Relationships among denitrification-derived N₂ ([N₂]_{den}), O₂, and $\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$ all 433 support the widespread occurrence and significance of denitrification in the Upper Floridan 434 Aquifer. (In springs with low dissolved O_2 , N_2 concentrations exceeded those predicted by 435 physical processes (as measured by noble gas tracers), but closely matched those predictions in 436 high O₂ springs (Fig. 4a). The negative relationship between O₂ concentrations and [N₂]_{den} 437 provides clear evidence of both the accuracy and precision of our approach and the occurrence of 438 denitrification in hypoxic portions of the UFA. Like $[N_2]_{den}$, $\delta^{15}N_{NO3}$ was inversely related to 439 dissolved O₂, with high values observed almost exclusively below 2 mg O₂ L^{-1} . $\delta^{15}N_{NO3}$ values 440 for springs with O₂ greater than 2 mg L⁻¹ were generally consistent with nitrogen derived from 441 predominantly but not exclusively inorganic sources (Fig. 4b). Relationships between the 442 progression of denitrification and $\delta^{15}N_{NO3}$ (Fig. 8a,c) support both the inference of denitrification 443 in the Floridan aquifer and the hypothesis that variation in isotopic signatures is primarily driven 444 by denitrification rather differential contribution from organic and inorganic sources. 445 Fractionation coefficients $(^{15}\varepsilon)$ derived from both the synoptic survey and the Ichetucknee River 446 springs are within the lower range of values reported for other aquifers (Mariotti, 1986; Bohlke 447 448 et al., 2002; Green et al., 2008), and other marine and freshwater environments (Sigman et al., 2005; Granger et al., 2008), but lower than some recent experimental values (Knoller et al., 449 450 2011). These relatively low values may indicate diffusion-constrained NO₃⁻ limitation of 451 denitrification in the UFA (Sebilo et al., 2003).

Covariation between $\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$ confirms the inference of denitrification in the 452 UFA and its influence on isotopic composition of NO3⁻ at the regional scale. Across springs and 453 454 over time within the springs of the Ichetucknee River, we observed strong relationships between $\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$, although the slopes of these relationships differed between the synoptic 455 survey and the temporally intensive study of the Ichetucknee system. The 1:1.7 relationship 456 457 across sites (Fig. 8b) is consistent with theoretical and empirical studies showing 1:2 enrichment 458 by denitrification (Lehmann et al. 2003, Aravena and Robertson 1998). In contrast, covariation 459 within the springs of the Ichetucknee River exhibited slopes near 1:1 on each date and across all 460 dates (Fig. 8d, Table 4). A recent study of isotope dynamics within the Ichetucknee River itself

vielded similar 1:1 fractionation ratio for ¹⁸O:¹⁵N associated with denitrification (Cohen et al., in 461 These relationships, both from studies with independently-constrained, direct 462 revision). estimates of denitrification, are similar to fractionation ratios obtained from laboratory 463 experiments (Granger et al., 2008) and marine systems (Sigman et al., 2005). The divergence of 464 fractionation during denitrification between freshwater and marine ecosystems has been 465 taxonomic or environmental differences between these systems, but our 466 attributed to observations suggest factors varying among watersheds can alter these relationships. Regardless 467 of these differences, strong covariation among $\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$ provide additional evidence 468 that denitrification drives variation in nitrate isotopic composition in the Floridan Aquifer. 469

Temporal patterns within individual springs provide a final line of support for 470 denitrification as a driver of NO3⁻ concentrations and isotopic composition. Within the springs 471 of the Ichetucknee River, O2 concentrations were positively correlated with NO3⁻ concentrations 472 (Fig. 3a). Across all springs, O₂ concentrations were negatively correlated with $\delta^{15}N_{NO3}$, and 473 negative relationships were also observed over time within low-O₂ springs. The apparent 474 absence of O₂-driven variation within springs for $[N_2]_{den}$ and $[NO_3]_R$ is most likely attributable 475 to low power in our data set for those variables and lower precision in estimates of [N₂]_{den} (and 476 thus $[NO_3]_R$ than for $\delta^{15}N_{NO3}$. Given the strength of observed relationships across springs 477 between denitrification progression and $\delta^{15}N_{NO3}$, the most likely explanation for within-spring 478 $\delta^{15}N_{NO3}$ -O₂ relationships (Fig. 7) is that variation in both O₂ and $\delta^{15}N_{NO3}$ reflect differential 479 contributions of young, oxic groundwater, the isotopic signatures of which are unlikely to have 480 been altered by denitrification, and older, anoxic groundwater, for which depletion nitrate by 481 denitrification has produced elevated values of $\delta^{15}N_{NO3}$. A plausible alternative hypothesis is 482 that $O_2 - \delta^{15} N_{NO3}$ relationships reflect the concurrent influence of human- or animal-derived 483 effluent on dissolved O₂ (via increased BOD) and $\delta^{15}N_{NO3}$. However, if coincident BOD and 484 δ^{15} N-enriched NO₃⁻ inputs were responsible for these patterns, then high-O₂ springs would 485 exhibit covariation between O_2 and $\delta^{15}N_{NO3}$, as is observed in low- O_2 springs. Thus, while 486 alternative explanations might plausibly explain some of the pairwise correlations observed in 487 this study (e.g. organic waste inputs as a driver of both O_2 and $\delta^{15}N_{NO3}$), the convergence of 488 multiple lines of evidence clearly indicates that denitrification is a significant process in the 489 UFA, as both a sink for N inputs and as a driver of variation in isotope signatures. 490

492 **4.2** Magnitude and mechanisms of nitrate reduction

An important feature of this study is that springs integrate upgradient N transformations over considerable spatial domains and over the entire duration of water residence in the subsurface. In combination with the large spatial extent of our study springs, this feature of springs enables relatively simple scaling of groundwater denitrification to springshed and regional scales. Studies of aquifer denitrification generally address denitrification along individual flowpaths with relatively small spatial footprints, and as such may not reflect the distribution of rates or residence times within the larger aquifer.

At the scale of the UFA, denitrification appears to be a significant sink for N leaching to 500 the Upper Floridan Aquifer, removing approximately 32% of the total (flow-weighted) N 501 discharging from sampled springs. Average volumetric rates derived from [N₂]_{den} were toward 502 the low end of values obtained from direct measurement of N₂, but were within the range 503 504 reported for agriculturally enriched aquifers (Green et al., 2008). Nonethless, aggregate areal rates of denitrification (122 kg km² y⁻¹) are comparable to the estimated global average for 505 aquifer denitrification (Seitzinger et al., 2006). These averages, however, integrate a high degree 506 of variation among springs. Average areal rates for individual springs ranged from less than 0 to 507 as high as 5300 kg km² y⁻¹, and the depletion of NO_3^- load from 0 to more than 97%. Thus 508 within the Floridan Aquifer we observe variation in denitrification comparable to that observed 509 globally (Seitzinger et al., 2006). Significant spatial heterogeneity of denitrification has been 510 observed in other aquifers, but none to our knowledge have measured denitrification directly 511 over the spatial and temporal extent found in this study. Our results suggest that measurements 512 along individual groundwater flowpaths are unlikely to be applicable to entire regional aquifer 513 514 systems.

The occurrrence and significance of denitrification in the organic-matter-poor Floridan Aquifer is superficially suprising, given the importance of organic matter supply as a constraint on denitrification across diverse ecosystem types. However, several recent studies have found isotopic evidence for denitrification in karstic aquifers, including those in southwestern Illinois, USA (Panno et al., 2001), southern Germany (Einsiedl and Mayer, 2006), and the Floridan Aquifer (Albertin et al., 2011). One feature of karst aquifers that may facilitate denitrification is large difference in hydraulic conductivity, and thus water age and chemistry, between karstic

aquifer matrices and conduits. McCallum et al. (2008) showed that mixing of groundwaters of different ages, NO_3^- concentrations, and redox potentials can promote aquifer denitrification, and such oxic-anoxic interfaces are widely recognized as locations of significant biogeochemical reactivity in surface waters (Dahm et al., 1998). Similar phenomena might be occurring throughout the considerable surface area of the conduit-matrix interface in karstic systems, and its occurrence and distribution might vary sufficiently to generate the observed differences in denitrification among UFA springs.

The source and character of electron donors that fuel nitrate reduction in the UFA are not 529 known. Low DOC concentrations in UFA springs imply that denitrification is fueled by some 530 other source, but availability of labile DOC is likely to be higher in water entering the aquifer 531 than in water discharging from springs. Runoff entering sinkholes provide one pathway for 532 relatively labile carbon to enter karstic aquifers, in either dissolved or particulate form. In some 533 cases backflow from C-rich surface waters influences the chemistry of springs discharge (Gulley 534 et al., 2011), and anthropogenic carbon from septic, municipal, or agricultural waste might also 535 provide labile C to the UFA. The importance of electron donors other than organic C for 536 denitrification in the UFA remains unclear, but has been demonstrated through correlational and 537 experimental studies in other aquifers, including some karstic systems. Although the carbonate 538 Floridan Aquifer matrix itself is generally poor in minerals that might serve as terminal electron 539 donors, Florida springs exhibit significant variation in mineral chemistry (Scott et al., 2004), and 540 many springsheds include the clay-rich Hawthorn Formation (Wicks and Herman 1994). If such 541 alternative electron donors drive variation in denitrification among UFA springs, then 542 concentrations of solutes such as Fe or SO₄ should be correlated with denitrification. The role 543 of different nitrate reduction pathways, as well as broader drivers such as hydrogeology, 544 geochemistry, and land use as factors that influence denitrification in the UFA, are beyond the 545 scope of this paper, but clearly worthy of further investigation. 546

547

548 **4.3 Uncertainty in estimates of denitrification**

549 Use of dual noble gas tracers (Ne, Ar) to estimate recharge temperature and excess air 550 produced estimates that were more precise and more accurate than those derived from statistical

modeling of excess air. Among springs with O2 greater than 2 mg L⁻¹ and thus presumably 551 negligible denitrification, [N2]den estimates based on dual tracers averaged 0.003 mg N L-1 (0.1 552 μ mol N₂ L⁻¹) with a standard deviation of 0.32 mg N L⁻¹(11.6 μ mol N₂ L⁻¹). For observations 553 from high O₂ springs where Ne data were unavailable, [N₂]_{den} estimates based on statistically 554 modeled excess air averaged 0.32 mg N₂ L⁻¹ (11.6 μ mol N₂ L⁻¹) with a standard deviation of 16.8 555 μ mol N₂ L⁻¹. For [N₂]_{den}, [NO₃-]_R, and δ^{15} N_{NO3}, spatial variation among springs was large 556 compared to temporal variation within springs. 557

The stronger relationship between denitrification progression and $\delta^{15}N_{NO3}$ for springs 558 with Ne data vs. all springs provides additional evidence for greater precision of these estimates. 559 Greater bias and lower precision of these estimates most likely reflects the variability of excess 560 air entrainment over time among springs, but may also reflect introduction of excess air during 561 sampling, an artifact for which our statistical approach does not account. Nonetheless, 562 uncertainty of our direct and statistically-derived estimates of denitrification compare favorably 563 with bias (5 μ mol N₂ L⁻¹) and precision (SD = 22 μ mol N₂ L⁻¹) in a previous study of 564 denitrification in agricultural aquifers, in which limited spatial extent permitted assumptions of 565 constant recharge temperature within regions, and calculation of excess air from Ar 566 concentrations (Green et al., 2008). The relatively high precision and minimal bias of [N2]den 567 estimates in this study illustrate both the value of dual isotope tracers and the utility of statistical 568 modeling of physical processes where direct measurements are unavailable. Similar approaches 569 will likely be necessary and useful in evaluating the spatial heterogeneity of denitrification in 570 571 other aquifers.

572 One important implication of the low within-spring variance in $[N_2]_{den}$ and $[NO_3]_R$ (Fig. 5) is that uncertainty in our regional estimate of the magnitude of denitrification and its effect on 573 N loads delivered to surface waters is largely influenced by whether or not sampled springs are 574 representative, rather than by uncertainty of estimates within sampled springs. Our population of 575 springs almost certainly overrepresents large springs, since we include more than half of the first magnitude springs in northern Florida. It is unclear whether our study oversampled N rich or N poor springs, since the distribution of NO₃⁻ concentrations in small emines is small emines in small emines is small emines. 576 577 poor springs, since the distribution of NO_3^- concentrations in small springs is not known. A p^{μ} 578 second source of uncertainty is the magnitude of diffuse groundwater discharge from the UFA, 579 and the comparability of the chemistry of this discharge to that of springs. On an areal basis, we 580 have almost certainly underestimated denitrification in the UFA because estimates only include 581

that hist

discharge from one spring over the area of its springshed. Incorporation of excess N_2 fluxes from springs with overlapping springshed and from diffuse groundwater efflux would increase areal estimates of denitrification, but the magnitude of this bias is not known. It is also unclear whether these unmeasured hydrologic flowpaths have excess N_2 concentrations comparable to those of the measured springs. In light of these uncertainties, our estimate of denitrification in the Floridan Aquifer should be viewed as a first approximation.

reque only individual no overall

588 4.4 Implications for N source inference

589 Concurrent measurements of dissolved gases, nitrate concentrations and isotopes enabled direct estimation of the absolute magnitude and relative progression of denitrification, and their 590 relationship to δ^{15} N_{NO3} enrichment (Fig. 8). The strong negative relationships between the size 591 of residual nitrate pools and $\delta^{15}N_{NO3}$ not only provide evidence for the occurrence of 592 denitrification, but also clearly indicate that denitrification exerts a significant influence on 593 nitrate isotopic composition in the UFA (particularly where dissolved O_2 concentrations are < 2594 mg L^{-1}). These relationships, in turn, allowed us to estimate the isotopic composition of the 595 original source NO₃⁻ for each spring. These estimates suggest that $\delta^{15}N_{NO3}$ of nitrate discharging 596 from UFA springs may in some cases differ substantially from the isotopic signature of the 597 original N source. 598

Observed variation in $\delta^{15}N_{NO3}$ among UFA springs was considerable, ranging from 599 values near zero to more than 20%. $\delta^{15}N_{NO3}$ distributions for springs with denitrification 600 estimates and within the Ichetucknee River springs were similar to that of all springs, but the 601 former was biased towards heavier $\delta^{15}N_{NO3}$ values, and the latter had a smaller range of values 602 and relatively fewer high values. Nonetheless, the similarity of these distributions suggests that 603 interpretation of our source estimates should be applicable to the broader population of springs. 604 In all three data sets, the largest subset of springs had $\delta^{15}N_{NO3}$ less than 6%, but in each case 605 more than one-third of springs had $\delta^{15}N_{NO3}$ greater than 6‰. Observed values in springs with 606 N_{2-den} estimates were higher than the larger data set that included all springs; more than 50% of 607 N_{2-den} springs had $\delta^{15}N_{NO3}$ values greater than 6‰. 608

609 Estimated source $\delta^{15}N_{NO3}$ signatures from springs with dissolved gas data and from the 610 Ichetucknee River springs time series had distributions that differed from observed values in two

4

m

m

0 (4

important respects. First, mean and median values for estimated sources were lower by ca. 2‰ 611 and 1.5%, respectively. Second, the frequency of extremely high $\delta^{15}N_{NO3}$ values was much 612 lower. Significantly, these estimated source values had distributions similar to those of all 613 springs with $O_2 > 3$ mg L⁻¹, where denitrification is presumably negligible. Together, these 614 observations strongly suggest that most $\delta^{15}N_{NO3}$ values in UFA springs somewhat overestimate 615 616 the contribution of organic sources, and in particular that very high values overwhelmingly 617 reflect fractionation resulting from nitrate removal by denitrification, rather than large 618 contributions from organic N sources.

To date, denitrification has largely been assumed to be negligible in the Upper Floridan 619 Aquifer, and as a result, elevated $\delta^{15}N_{NO3}$ values have been interpreted as indicating a significant 620 contribution from organic sources. Despite mass balance studies indicating fertilizer application 621 622 as the dominant N source to springsheds, policy and management efforts have largely focused on reducing N inputs from septic tanks, and agricultural and municipal waste (Loper et al. 2005, 623 Dederkorkut 2005, Mattson et al. 2006), largely on the basis of enriched $\delta^{15}N_{NO3}$ signatures. Our 624 data suggest that interpretation of $\delta^{15}N_{NO3}$ values must account for fractionating N 625 626 transformations within the aquifer.

627 As one example of the effects of fractionation, Wekiwa springs, near Orlando FL, has a heavily urbanized catchment and consistently elevated $\delta^{15}N_{NO3}$ values (mean of 4 observations 628 from 2001-2010: 11.5 \pm 3.1‰), which would initially suggest that elevated NO₃⁻ concentrations (V629 in that system are primarily derived from organic sources. However, $\delta^{18}O_{NO3}$ are also highly 630 enriched (12.4‰), and measurements of N_{2-den} are consistently high (mean of 4 observations 631 from 2001-2010: 3.1 \pm 0.4 mg N₂ L⁻¹). We estimate that that dentrification within the aquifer 632 typically removes ca. 75% of NO3⁻ before discharge from Wekiwa springs, and that the original 633 source of nitrate in Wekiwa Springs had a $\delta^{15}N_{NO3}$ value ca. 6.3‰. While individual estimates of 634 source $\delta^{15}N_{NO3}$ should be viewed with caution, it seems likely that despite its urban setting, N 635 636 enrichment of Wekiwa springs is due primarily to inorganic fertilizers, with contributions from organic sources. In constrast, Wakulla Springs, near Tallahassee, FL, has somewhat enriched 637 $\delta^{15}N_{NO3}$ (6.4-7.9‰) but no excess N₂. Moreover, hydrologic tracer studies have demonstrated 638 639 direct connections between the Tallahassee municipal waste sprayfield and Wakulla Springs (Kincaid et al. 2005). Thus, it seems safe to conclude that elevated $\delta^{15}N_{NO3}$ values in Wakulla 640 springs do in fact reflect the isotopic signature of N sources, which include significant 641

642 contributions from organic sources. However, our observations suggest that this is the exception
643 rather than the rule. Absent direct evidence for substantial organic sources for a specific spring,
644 efforts to reduce N loading to the UFA should focus on fertilizer inputs.

645

646 **5 Conclusions**

The surpsing importance of denitrification for N fluxes and isotopic composition in the 647 UFA has important implications both for management of North Florida landscapes and for 648 broader understanding of groundwater denitrification. Methodologically, this study illustrates 649 the value of multiple lines of inference for assessing denitrification, which are strengthened by 650 direct estimates of the physical processes that influence N₂ concentration using multiple tracers. 651 Significant spatial and temporal variability of denitrification within the UFA suggests that 652 improving regional and global estimates of denitrification will require more extensive (653 measurements in other aquifers. The variability of denitrification in the Floridan Aquifer has 654 implications not only for regional estimates of N removal, but also for values and variability of 655 isotopic signatures of residual nitrate pools at the regional scale. Accurate assessments of the 656 contribution of various sources of N enrichment, in North Florida and elsewhere, must account 657 for the influence of denitrification on N isotope ratios. 658

659

660 Acknowledgements

The authors wish to thank Larry Kohrnak and Chad Foster for their help with field sampling, and Steve Walsh, Sharon Kroenig, and Leel Knowles for sharing data. This research was supported by grants from the St. Johns River Water Management District, the Southwest Florida Water Management District, the Three Rivers Foundation, and the National Science Foundation (EAR 0838390). The conclusions in this paper are those of the authors and do not necessarily reflect the views or policy of any state or federal agency.

667

YAU

668 **References**

Albertin, A. R., Sickman, J. O., Pinowska, A., and Stevenson, R. J.: Identification of nitrogen
sources and transformations within karst springs using isotope tracers of nitrogen,
Biogeochemistry, in press, DOI: 10.1007/s10533-011-9592-0, 2011.

- 672 APHA, AWWA, and WEF: Standard methods for the examination of water and wastewater, 21st
- 673 ed., American Public Health Association, Washington, DC, 2005.
- Aravena, R., and Robertson, W. D.: Use of multiple isotope tracers to evaluate denitrification in
 ground water: Study of nitrate from a large-flux septic system plume, Ground Water, 36, 975982, 1998.
- 677 Arthur, J. D., Wood, H. A. R., Baker, A. E., Cichon, J. R., and Raines, G. L.: Development and
- 678 implementation of a bayesian-based aquifer vulnerability assessment in florida: . Natural
 679 Resources Research, 16, 93-107, 2007.
- Bedard-Haughn, A., van Groenigen, J. W., and van Kessel, C.: Tracing n-15 through landscapes:
 Potential uses and precautions, Journal of Hydrology, 272, 175-190, 2003.
- Bohlke, J. K., Wanty, R., Tuttle, M., Delin, G., and Landon, M.: Denitrification in the recharge
- area and discharge area of a transient agricultural nitrate plume in a glacial outwash sand aquifer,
- 684 Minnesota, Water Resources Research, 38, 110510.1029/2001wr000663, 2002.
- Bonn, M. A., and Bell, F. W.: Economic impact of selected Florida springs on surrounding local
 areas., Florida Department of Environmental Protection, Tallahassee, Florida, USA., 99 pp.,
 2003.
- 688 Bonn, M. A.: Visitor profiles, economic impacts and recreational aesthetic values associated with
- eight priority Florida springs located in the St. Johns River Water Management District, St.
- 690 Johns River Water Management District, Palatka, FL, 136 pp., 2004.
- Burgin, A. J., and Hamilton, S. K.: Have we overemphasized the role of denitrification in aquatic
 ecosystems? A review of nitrate removal pathways, Frontiers in Ecology and the Environment, 5,
- 693 89-96, 2007.

- Burns, D. A., Boyer, E. W., Elliott, E. M., and Kendall, C.: Sources and transformations of nitrate from streams draining varying land uses: Evidence from dual isotope analysis, Journal of Environmental Quality, 38, 1149-1159, 10.2134/jeq2008.0371, 2009.
- Casciotti, K. L., Sigman, D. M., Hastings, M. G., Bohlke, J. K., and Hilkert, A.: Measurement of
 the oxygen isotopic composition of nitrate in seawater and freshwater using the denitrifier
 method, Analytical Chemistry, 74, 4905-4912, 10.1021/ac020113w, 2002.
- Castro, M. C., Hall, C. M., Patriarche, D., Goblet, P., and Ellis, B. R.: A new noble gas
 paleoclimate record in Texas basic assumptions revisited, Earth and Planetary Science Letters,
 257, 170-187, 10.1016/j.epsl.2007.02.030, 2007.
- Cey, B. D., Hudson, G. B., Moran, J. E., and Scanlon, B. R.: Evaluation of noble gas recharge
 temperatures in a shallow unconfined aquifer, Ground Water, 47, 646-659, 10.1111/j.17456584.2009.00562.x, 2009.
- Chapelle, F. H., McMahon, P. B., Dubrovsky, N. M., Fujii, R. F., Oaksford, E. T., and
 Vroblesky, D. A.: Deducing the distribution of terminal electron-accepting processes in
 hydrologically diverse groundwater systems, Water Resources Research, 31, 359-371,
 10.1029/94wr02525, 1995.
- Chasar, L., Katz, B. G., and Griffin, D.: Evaluation of nitrate sources in springs of the Santa Fe
 River basin using natural tracers: Geochemical, specific microbiological, and multiple stable
 isotopic indicators, Alachua County Environmental Protection Department, Gainesville, FL,
 2005.
- 714 Cohen, M., Heffernan, J., Albertin, A., and Martin, J.: Inference of riverine nitrogen processing
- from longitudinal and diel variation in dual nitrate isotopes, Journal of Geophysical Research Biogeosciences, in revision.
- 717 Dahm, C. N., Grimm, N. B., Marmonier, P., Valett, H. M., and Vervier, P.: Nutrient dynamics at
- the interface between surface waters and groundwaters, Freshwater Biology, 40, 427-451, 1998.
- 719 David, M. B., Wall, L. G., Royer, T. V., and Tank, J. L.: Denitrification and the nitrogen budget
- of a reservoir in an agricultural landscape, Ecological Applications, 16, 2177-2190, 2006.
- 721 Davidson, E. A., and Seitzinger, S.: The enigma of progress in denitrification research,
- 722 Ecological Applications, 16, 2057-2063, 2006.

- Dederkorkut, A.: Suwanee River Partnership: Representation instead of regulation, in: Adaptive
 Governance and Water Conflict: New Institutions for Collaborative Planning, edited by: Scholz,
- J. and Stiftel, B., Resources for the Future Press, Washington, DC, pp. 25-39, 2005.
- 726 Duarte, C. M., Prairie, Y. T., Frazer, T. K., Hoyer, M. V., Notestein, S. K., Martinez, R., Dorsett,
- 727 A., and Canfield, D. E.: Rapid accretion of dissolved organic carbon in the springs of Florida:
- The most organic-poor natural waters, Biogeosciences, 7, 4051-4057, 10.5194/bg-7-4051-2010,
 2010.
- 730 Einsiedl, F., and Mayer, B.: Hydrodynamic and microbial processes controlling nitrate in a
- 731 fissured-porous karst aquifer of the Franconian Alb, southern Germany, Environmental Science
- 732 & Technology, 40, 6697-6702, 10.1021/es061129x, 2006.
- 733 Eyre, B. D., Rysgaard, S., Dalsgaard, T., and Christensen, P. B.: Comparison of isotope pairing
- and N-2 : Ar methods for measuring sediment-denitrification-assumptions, modifications, and
 implications, Estuaries, 25, 1077-1087, 2002.
- Feast, N. A., Hiscock, K. M., Dennis, P. F., and Andrews, J. N.: Nitrogen isotope
 hydrochemistry and denitrification within the chalk aquifer system of North Norfolk, UK,
 Journal of Hydrology, 211, 233-252, 1998.
- 739 Fogg, G. E., Rolston, D. E., Decker, D. L., Louie, D. T., and Grismer, M. E.: Spatial variation in
- nitrogen isotope values beneath nitrate contamination sources, Ground Water, 36, 418-426, 1998.
- 741 Galloway, J. N., Aber, J. D., Erisman, J. W., Seitzinger, S. P., Howarth, R. W., Cowling, E. B.,
- and Cosby, B. J.: The nitrogen cascade, Bioscience, 53, 341-356, 2003.
- Granger, J., Sigman, D. M., Lehmann, M. F., and Tortell, P. D.: Nitrogen and oxygen isotope
 fractionation during dissimilatory nitrate reduction by denitrifying bacteria, Limnology and
 Oceanography, 53, 2533-2545, 2008.
- Green, C. T., Puckett, L. J., Bohlke, J. K., Bekins, B. A., Phillips, S. P., Kauffman, L. J., Denver,
- 747 J. M., and Johnson, H. M.: Limited occurrence of denitrification in four shallow aquifers in
- agricultural areas of the United States, Journal of Environmental Quality, 37, 994-1009,
 10.2134/jeq2006.0419, 2008.
- 750 Groffman, P. M., Butterbach-Bahl, K., Fulweiler, R. W., Gold, A. J., Morse, J. L., Stander, E. K.,
- 751 Tague, C., Tonitto, C., and Vidon, P.: Challenges to incorporating spatially and temporally

X

- explicit phenomena (hotspots and hot moments) in denitrification models, Biogeochemistry, 93,
 49-77, 10.1007/s10533-008-9277-5, 2009.
- Gulley, J., Martin, J. B., Screaton, E. J., and Moore, P. J.: River reversals into karst springs: A
- model for cave enlargement in eogenetic karst aquifers, Geological Society of America Bulletin,
- 756 123, 457-467, 10.1130/b30254.1, 2011.
- Hall, C. M., Castro, M. C., Lohmann, K. C., and Ma, L.: Noble gases and stable isotopes in a
 shallow aquifer in southern Michigan: Implications for noble gas paleotemperature
 reconstructions for cool climates, Geophysical Research Letters, 32, 4, L18404,
 10.1029/2005gl023582, 2005.
- Hamme, R. C., and Emerson, S. R.: The solubility of neon, nitrogen and argon in distilled water
 and seawater, Deep-Sea Research Part I-Oceanographic Research Papers, 51, 1517-1528,
 10.1016/j.dsr.2004.06.009, 2004.
- Harms, T. K., and Grimm, N. B.: Hot spots and hot moments of carbon and nitrogen dynamics in
 a semiarid riparian zone, Journal of Geophysical Research-Biogeosciences, 113, G01020, 2008.
- Harrington, D., Maddox, G., and Hicks, R.: Florida springs initiative monitoring network report
 and recognized sources of nitrate, Florida Department of Environmental Protection, Tallahassee,
 FL, 103, 2010.
- Hedin, L. O., von Fischer, J. C., Ostrom, N. E., Kennedy, B. P., Brown, M. G., and Robertson,
 G. P.: Thermodynamic constraints on nitrogen transformations and other biogeochemical
 processes at soil-stream interfaces, Ecology, 79, 684-703, 1998.
- Heffernan, J. B., Cohen, M. J., Frazer, T. K., Thomas, R., Rayfield, T., Gulley, J., Martin J. B.,
 Delfino, J. J., and Graham, W. D.: Hydrologic and biotic influences on nitrate removal in a
 subtropical spring-fed river, Limnology and Oceanography, 55, 249-263, 2010a.
- 775 Heffernan, J.B., Liebowitz, D. L., Frazer, T. K., Evans, J. M., and Cohen, M. J.: Algal blooms
- and the nitrogen-enrichment hypothesis in Florida springs: Evidence, alternatives, and adaptive
- management, Ecological Applications, 20, 816-829, 2010b.
- 778 Kana, T. M., Darkangelo, C., Hunt, M. D., Oldham, J. B., Bennett, G. E., and Cornwell, J. C.:
- 779 Membrane inlet mass-spectrometer for rapid high-precision determination of N-2, O-2, and Ar in
- renvironmental water samples, Analytical Chemistry, 66, 4166-4170, 1994.

Katz, B. G., Hornsby, H. D., Bohlke, J. F., and Mokray, M. F.: Sources and chronology of nitrate
contamination in spring waters, Suwannee River basin, Florida, US Geological Survey, Reston,

783 VA, 1999.

- Katz, B. G., Bohlke, J. K., and Hornsby, H. D.: Timescales for nitrate contamination of spring
 waters, northern Florida, usa, Chemical Geology, 179, 167-186, 2001.
- Katz, B. G.: Sources of nitrate contamination and age of water in large karstic springs of Florida,
 Environmental Geology, 46, 689-706, 2004.
- Katz, B. G., Chelette, A. R., and Pratt, T. R.: Use of chemical and isotopic tracers to assess
 nitrate contamination and ground-water age, Woodville Karst Plain, USA, Journal of Hydrology,
 289, 36-61, 10.1016/j.hydrol.2003.11.001, 2004.
- 791 Katz, B. G., Sepulveda, A. A., and Verdi, R. J.: Estimating nitrogen loading to ground water and
- 792 assessing vulnerability to nitrate contamination in a large karstic springs basin, Florida, Journal
- 793 of the American Water Resources Association, 45, 607-627, 10.1111/j.1752-1688.2009.00309.x,
 794 2009.
- Kendall, C., and Grim, E.: Combustion tube method for measurement of nitrogen isotope ratios
 using calcium-oxide for total removal of carbon-dioxide and water, Analytical Chemistry, 62,
 526-529, 1990.
- Kendall, C.: Tracing nitrogen sources and cycling in catchments, in: Isotope tracers in catchment
 hydrology, edited by: Kendall, C., and McDonnell, J. J., Elsevier Science B.V., Amsterdam, The
 Netherlands, 1998.
- Kendall, C., Elliott, E. M., and Wankel, S. D.: Tracing anthropogenic inputs of nitrogen to
 ecosystems, in: Stable isotopes in ecology and environmental science, edited by: Michener, R.
 H., and Lajtha, K., Wiley Blackwell Publishing, Malden, MA, 375–449, 2007.
- 804 Kincaid, T.R., Hazlett, T. J., and Davies, G.J.: Quantitative Groundwater Tracing and Effective
- 805 Numerical Modeling in Karst: An Example from the Woodville Karst Plain of North Florida,
- 806 ASCE Conf. Proc. Sinkholes and the Engineering and Environmental Impacts of Karst,
- doi:10.1061/40796(177)13, 2005.

- Knoller, K., Vogt, C., Haupt, M., Feisthauer, S., and Richnow, H. H.: Experimental investigation
 of nitrogen and oxygen isotope fractionation in nitrate and nitrite during denitrification,
 Biogeochemistry, 103, 371-384, 10.1007/s10533-010-9483-9, 2011.
- Knowles, L., Katz, B. G., and Toth, D. J.: Using multiple chemical indicators to characterize and
 determine the age of groundwater from selected vents of the Silver Springs group, central
 Florida, USA, Hydrogeology Journal, 18, 1825-1838, 10.1007/s10040-010-0669-y, 2010.
- Lehmann, M. F., Reichert, P., Bernasconi, S. M., Barbieri, A., and McKenzie, J. A.: Modelling
- 815 nitrogen and oxygen isotope fractionation during denitrification in a lacustrine redox-transition
- 816 zone, Geochimica Et Cosmochimica Acta, 67, 2529-2542, 10.1016/s0016-7037(03)00085-1,
- 817 2003.
- 818 Loper, D., Landing, W., Pollman, C., and Chan Hilton, A.: Degradation of water quality at
- 819 Wakulla Springs, FL: Assessment and recommendations, Report of the Peer Review Committee
- 820 on the Workshop Solving Water Pollution Problems in the Wakulla Springshed of North Florida.
- 821 Florida Department of Environmental Protection, Tallahassee, FL, 70 pp, 2005.
- Mariotti, A.: Denitrification in groundwaters, principles and methods for its identification a review, Journal of Hydrology, 88, 1-23, 1986.
- Martin, J. B., and Dean, R. W.: Exchange of water between conduits and matrix in the Floridan Aquifer, Chemical Geology, 179, 145-165, 2001.
- 826 Mattson, R. A., Lowe, E. F., Lippincott, C. L., Di, J., and Battoe, L.: Wekiva River and Rock
- 827 Springs Run Pollutant Load Reduction Goals, Florida Department of Environmental Protection,
- 828 Tallahassee, FL, 69 pp., 2006.
- 829 McCallum, J. E., Ryan, M. C., Mayer, B., and Rodvang, S. J.: Mixing-induced groundwater
- 830 denitrification beneath a manured field in southern Alberta, Canada, Applied Geochemistry, 23,
- 831 2146-2155, 10.1016/j.apgeochem.2008.03.018, 2008.
- Miller, J. A.: Ground water atlas of the United States; Alabama, Florida, Georgia, and South
 Carolina., Hydrologic atlas, U.S. Geological Survey, 1990.
- 834 Notholt, A. J. G., Sheldon, R. P., and Davidson, D. F.: Phosphate rock resources, in: Phosphate
- deposits of the world, Cambridge University Press, Cambridge, UK, xxix + 566 pp, 1989.

- Panno, S. V., Hackley, K. C., Hwang, H. H., and Kelly, W. R.: Determination of the sources of
 nitrate contamination in karst springs using isotopic and chemical indicators, Chemical Geology,
 179, 113-128, 2001.
- Phelps, G. G.: Chemistry of ground water in the Silver Springs basin, Florida, with an emphasis
 on nitrate, U.S. Geological Survey, Reston, VA, 54 p., 2004.
- Phelps, G. G., Walsh, S. J., Gerwig, R. M., and Tate, W. B.: Characterization of the
 hydrology,water chemistry, and aquatic communities of selected springs in the St. Johns River
 Water Management District, Florida, 2004, US Geological Survey, Reston, VA, 51 pp., 2006.
- 844Pittman, J. R., H. H. Hatzell and E. T. Oaksford.: Spring contributions to water quality and845nitrate loads in the Suwannee River during baseflow in July 1995, U. S. Geological Survey,846Reston, VA, 1997.
- Schlesinger, W. H.: On the fate of anthropogenic nitrogen, Proceedings of the National Academy
 of Sciences of the United States of America, 106, 203-208, 10.1073/pnas.0810193105, 2009.
- Schmidt, N., Lipp, E. K., Rose, J. B., and Luther, M. E.: Enso influences on seasonal rainfall and
 river discharge in Florida, Journal of Climate, 14, 615-628, 10.1175/15200442(2001)014<0615:eiosra>2.0.co;2, 2001.
- 852 Schwientek, M., Einsiedl, F., Stichler, W., Stogbauer, A., Strauss, H., and Maloszewski, P.:
- 853 Evidence for denitrification regulated by pyrite oxidation in a heterogeneous porous groundwater
- system, Chemical Geology, 255, 60-67, 10.1016/j.chemgeo.2008.06.005, 2008.
- Scott, T. M., Means, G. H., Meegan, R. P., Means, R. C., Upchurch, S. B., Copeland, R. E.,
 Jones, J., Roberts, T., and Willet, A.: Springs of Florida, Florida Geological Survey, Tallahassee,
 FL, 347 pp., 2004.
- Sebilo, M., Billen, G., Grably, M., and Mariotti, A.: Isotopic composition of nitrate-nitrogen as a
 marker of riparian and benthic denitrification at the scale of the whole Seine River system,
 Biogeochemistry, 63, 35-51, 2003.
- 861 Seitzinger, S., Harrison, J. A., Bohlke, J. K., Bouwman, A. F., Lowrance, R., Peterson, B.,
- Tobias, C., and Van Drecht, G.: Denitrification across landscapes and waterscapes: A synthesis,
 Ecological Applications, 16, 2064-2090, 2006.

- Sigman, D. M., Casciotti, K. L., Andreani, M., Barford, C., Galanter, M., and Bohlke, J. K.: A
 bacterial method for the nitrogen isotopic analysis of nitrate in seawater and freshwater,
 Analytical Chemistry, 73, 4145-4153, 2001.
- 867 Sigman, D. M., Granger, J., DiFiore, P. J., Lehmann, M. M., Ho, R., Cane, G., and van Geen, A.:
- 868 Coupled nitrogen and oxygen isotope measurements of nitrate along the eastern North Pacific
- 869 margin, Global Biogeochemical Cycles, 19, GB4022, 10.1029/2005gb002458, 2005.
- 870 Sigman, D. M., DiFiore, P. J., Hain, M. P., Deutsch, C., Wang, Y., Karl, D. M., Knapp, A. N.,
- 871 Lehmann, M. F., and Pantoja, S.: The dual isotopes of deep nitrate as a constraint on the cycle
- and budget of oceanic fixed nitrogen, Deep-Sea Research Part I-Oceanographic Research Papers,
- 873 56, 1419-1439, 10.1016/j.dsr.2009.04.007, 2009.
- Smith, V. H., and Schindler, D. W.: Eutrophication science: Where do we go from here?, Trends
 in Ecology & Evolution, 24, 201-207, 10.1016/j.tree.2008.11.009, 2009.
- 876 Tarits, C., Aquilina, L., Ayraud, V., Pauwels, H., Davy, P., Touchard, F., and Bour, O.: Oxido-
- reduction sequence related to flux variations of groundwater from a fractured basement aquifer
 (Ploemeur area, France), Applied Geochemistry, 21, 29-47, 10.1016/j.apgeochem.2005.09.004,
 2006.
- Taylor, P. G., and Townsend, A. R.: Stoichiometric control of organic carbon-nitrate
 relationships from soils to the sea, Nature, 464, 1178-1181, 10.1038/nature08985, 2010.
- Torrento, C., Cama, J., Urmeneta, J., Otero, N., and Soler, A.: Denitrification of groundwater with pyrite and thiobacillus denitrificans, Chemical Geology, 278, 80-91, 10.1016/j.chemgeo.2010.09.003, 2010.
- Torrento, C., Urmeneta, J., Otero, N., Soler, A., Vinas, M., and Cama, J.: Enhanced denitrification in groundwater and sediments from a nitrate-contaminated aquifer after addition of pyrite, Chemical Geology, 287, 90-101, 10.1016/j.chemgeo.2011.06.002, 2011.
- Toth, D. J., and Katz, B. G.: Mixing of shallow and deep groundwater as indicated by the chemistry and age of karstic springs, Hydrogeology Journal, 14, 827-847, 10.1007/s10040-005-0478-x, 2006.
- Townsend, A. R., and Davidson, E. A.: Denitrification across landscapes and waterscapes,
 Ecological Applications, 16, 2055-2056, 2006.

31

 \propto

 \times

- 893 USGS: Nutrients in the Upper Mississippi River: Scientific information to support management
 894 decisions, USGS Fact Sheet 6, 2003.
- Vitousek, P. M.: Beyond global warming ecology and global change, Ecology, 75, 1861-1876,
 1994.
- Vogel, J. C., Talma, A. S., and Heaton, T. H. E.: Gaseous nitrogen as evidence for denitrification
 in groundwater, Journal of Hydrology, 50, 191-200, 10.1016/0022-1694(81)90069-x, 1981.
- Wicks, C. M., and Herman, J. S.: The effect of a confining unit on the geochemical evolution of
 groundwater in the Upper Floridan Aquifer system, Journal of Hydrology, 153, 139-155,
 10.1016/0022-1694(94)90189-9, 1994
- Wilson, G. B., and McNeill, G. W.: Noble gas recharge temperatures and the excess air component, Applied Geochemistry, 12, 747-762, 10.1016/s0883-2927(97)00035-8, 1997.
- Wood, E. D., Armstrong, F. A. J., and Richards, F. A.: Determination of nitrate in sea water by cadmium–copper reduction to nitrite, J. Marine Biol. Assoc. UK, 47, 31-43, 1967.
- 206 Zhang, Y. C., Slomp, C. P., Broers, H. P., Passier, H. F., and Van Cappellen, P.: Denitrification
- 907 coupled to pyrite oxidation and changes in groundwater quality in a shallow sandy aquifer,
- 908 Geochimica Et Cosmochimica Acta, 73, 6716-6726, 10.1016/j.gca.2009.08.026, 2009.

Figure 7. Parameters of within-spring relationships between dissolved O_2 and $\delta^{15}N_{NO3}$ as a function of spring mean dissolved oxygen. Large negative values of both (a) correlation coefficient and (b) slope in low- O_2 springs, and their absence in higher- O_2 systems, suggest that isotopically-enriched nitrate pools are associated with old, deeply anoxic flowpaths where denitrification would be most likely to occur. Open symbols indicate springs with 3 observations of O_2 and $\delta^{15}N_{NO3}$. Closed symbols indicated springs with 4 or more observations.

985

Figure 8. Effects of denitrification on isotopic composition of nitrate in Florida springs. Variation in $\delta^{15}N_{NO3}$ was strongly correlated with denitrification progression (a) as estimated from excess N₂ and observed nitrate concentrations across 61 springs, and (b) as estimated from differences between the Ichetucknee Headsprings and other springs in the Ichetucknee River. Positive correlation between $\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$ (c) across springs and (d) over time within the Ichetucknee system are also consistent with denitrification rather than variation in source as a driver of $\delta^{15}N_{NO3}$.

993

Figure 9. Implications of fractionation by denitrification for inference of N sources to Florida 994 springs. The distribution of $\delta^{15}N_{NO3}$ across all observations (A), among springs sampled for 995 dissolved gases in this study (C) and from the Ichetucknee time series (E) all suggest meaningful 996 contributions of organic sources (one third to one half of springs). However, $\delta^{15}N_{NO3}$ values in 997 high DO springs (B) and source $\delta^{15}N_{NO3}$ as back-calculated from isotopic enrichment factor and 998 denitrification progression (D, F) suggest inorganic fertilizers and soil N (from mineralized OM) 999 springs. overwhelming majority of predominant source in the the 1000 as

Figure 1

