

1 **Supplementary information**

2 **1. Possibility of desorption**

3 It has been suggested that CH₄ release from plant materials might result from CH₄ desorption in
4 low-CH₄ environments, e.g., after flushing with CH₄ free air (Kirschbaum et al., 2007).

5 It has previously been shown by Kirschbaum and Walcroft (2008) that desorption does not play
6 a significant role under ambient conditions of moisture, temperature and CH₄ concentrations
7 (Kirschbaum and Walcroft, 2008). However, whereas Kirschbaum and Walcroft used ambient
8 CH₄ levels for adsorption and investigated the desorption at ambient temperatures under low
9 CH₄ conditions, we also investigated adsorption to peat at much higher CH₄ concentrations of
10 12,500 ppm, 100 ppm and 10 ppm and its desorption at 50 °C. For each concentration level
11 vials were prepared and left to rest at room temperature for 3 days to allow the CH₄ to adsorb to
12 the peat surfaces. The samples were then divided into two groups, one being lyophilised
13 overnight, then flushed with CH₄ free air, while one was only flushed with CH₄ free air. A third
14 group of three peat samples, which contained no additional CH₄ but were also lyophilised and
15 flushed with CH₄ free air, served as control. Finally, all three groups were supplemented with
16 water and incubated at 50 °C for 17 h.

17 The objective of this experiment was to determine whether the observed CH₄ emissions were
18 indeed formed during the incubations or were due to an artefact caused by desorption of CH₄,
19 possibly arising from microbial origin, from the material under higher than ambient CH₄ levels
20 in the soil or peat. Moreover, this experiment also enables us to determine if any of the adsorbed
21 CH₄ following the lyophilisation process could account for some of the CH₄ observed in our
22 measurements.

23 The peat samples treated with the highest CH₄ levels (12,500 ppm) showed an increased CH₄
24 release in both CH₄ supplemented groups ($2.2 \pm 0.9 \text{ ng g}^{-1} (\text{dw}) \text{ h}^{-1}$ and $3.3 \pm 0.3 \text{ ng g}^{-1} (\text{dw}) \text{ h}^{-1}$
25 for samples with and without lyophilisation, respectively) compared to the untreated control
26 group ($0.4 \pm 0.1 \text{ ng g}^{-1} (\text{dw}) \text{ h}^{-1}$). However, the samples treated with 100 and 10 ppm CH₄

27 showed no significant increase, both with and without lyophilisation. Furthermore, samples of
28 kaolinite and sea sand were tested for their adsorption potential of CH₄ using 10 ppm CH₄ but
29 again no adsorption/desorption was detected.

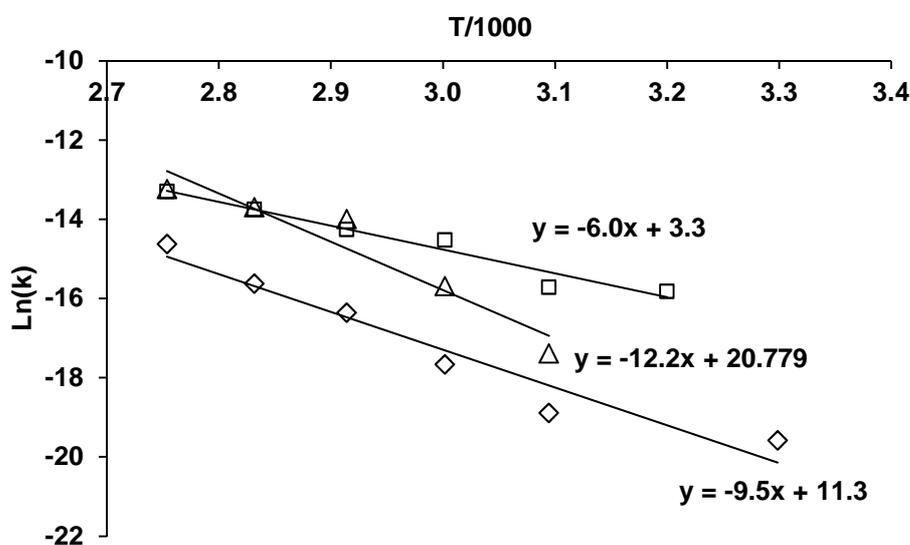
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31 2. Exclusion of methane oxidation by methane consuming bacteria

32 Several experiments from the water dependence study were repeated to investigate the possible
33 influence of methanotrophic bacteria on CH₄ emissions. For the dry samples and those with a
34 sample to water ratio of 2:1, 1:1 and 1:2 the experiment was repeated at 40 °C with the addition
35 of 20 µl difluoromethane (DFM) per vial. DFM was added to inhibit CH₄ oxidation by any
36 methanotrophic bacteria (Miller et al., 1998) possibly present in the non-sterile samples. No
37 significant effect was observed on the emission rates after adding DFM, leading to the
38 conclusion that there were no methanotrophic bacteria active in the lyophilised samples.
39 Measured CH₄ emissions were $0.8 \pm 0.2 \text{ ng g}^{-1} (\text{dw}) \text{ h}^{-1}$ without DFM and $0.4 \pm 0.02 \text{ ng g}^{-1} (\text{dw})$
40 h^{-1} with DFM for the dry samples. The wetted samples showed emissions ranging from 1.6 ± 0.1
41 to $1.9 \pm 0.1 \text{ ng g}^{-1} (\text{dw}) \text{ h}^{-1}$ with and without added DFM.

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43 3. Arrhenius plots



44

45 **Fig S1:** Arrhenius plot for formation of CH₄ in peat PH (◇), soil SL (□) and soil SG (△).

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47 We used the experimental data from samples SL, SG and PH to draw Arrhenius plots for CH₄
48 formation (Fig. S1). For all samples the results were found to follow a linear relationship at
49 temperatures ranging from 30 to 90 °C. The activation energies (E_a) for CH₄ formation for each
50 sample, calculated from the slope of the line, yielded values of 50.1 kJ mol⁻¹, 101.3 kJ mol⁻¹ and
51 79.2 kJ mol⁻¹ for SL, SG and PH, respectively. Again, this is strong supportive evidence of an
52 abiotic underlying process as reactions with activation energies higher than 50 kJ mol⁻¹ are
53 considered to be abiotic (Schönknecht et al., 2008).

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55 **References**

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