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# Organic matter composition and stabilization in a polygonal tundra soil of the Lena-Delta

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## Abstract

This study investigated soil organic matter (OM) composition of differently stabilized soil OM fractions in the active layer of a polygonal tundra soil in the Lena-Delta, Russia by applying density and particle-size fractionation combined with qualitative OM analysis using solid state  $^{13}\text{C}$  nuclear magnetic resonance spectroscopy, and lipid analysis combined with  $^{14}\text{C}$  analysis. Bulk soil OM was mainly composed of plant-derived, little decomposed material with surprisingly low and strongly increasing apparent  $^{14}\text{C}$  ages with active layer depth suggesting slow microbial OM transformation in cold climate. Most soil organic carbon was stored in clay and fine silt fractions ( $< 6.3\ \mu\text{m}$ ), which were composed of little decomposed plant material indicated by the dominance of long *n*-alkane and *n*-fatty acid compounds and low alkyl/O-alkyl C ratios. Organo-mineral associations, which are suggested to be a key mechanism of OM stabilization in temperate soils, seem to be less important in the active layer as the mainly plant-derived clay and fine silt sized OM was surprisingly “young” with  $^{14}\text{C}$  contents similar to the bulk soil values. Furthermore these fractions contained less organic carbon compared to density fractionated OM occluded in soil aggregates – a further important OM stabilization mechanism in temperate soils restricting accessibility of microorganisms. This process seems to be important at greater active layer depth where particulate OM, occluded in soil aggregates, was “older” than free particulate OM.

## 1 Introduction

The Lena Delta–Laptev Sea region is a key area for studies on the Arctic carbon cycle. The area is hosting one of the largest Arctic rivers, the Lena River with a length of 4337 km (Lobbes et al., 2000) and its drainage basin is located almost entirely within the zone of permanent permafrost. About 18% of the total dissolved organic carbon (DOC) and 12–14% of particulate organic carbon (POC) discharged into the Arctic Ocean are transported by the Lena River (Dittmar and Kattner, 2003) with the major

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component being terrestrial organic matter (OM) (Lara et al., 1998; Fahl and Stein, 2007).

Previous studies suggest that much of the terrestrial OM in the Lena River and Laptev Sea shelf derives from permafrost soils and riverbank erosion (Charkin et al., 2011; Gustafsson et al., 2011) including an increasing portion of old, strongly transformed DOC and POC most probably exported from thawing permafrost soils (Guo et al., 2007; van Dongen et al., 2008; Gustafsson et al., 2011). However, most of these studies are identifying terrestrial OM from analyses of river and marine sediments as well as of suspended material from both systems (e.g. Schubert and Stein, 1997; Dittmar and Kattner, 2003; Karlsson et al., 2011; Semiletov et al., 2011) but information on the original source, plant material and soil OM in the hinterland of the Laptev Sea area are missing. Few investigations have been conducted so far of the active layer in permafrost soils, which thaws during summer, as potential source of terrestrial OM to Arctic rivers and the ocean. Due to the expected increase in active layer thickness, which was already proven in some studies (Hinkel and Nelson, 2003; Nelson et al., 2008; Akerman and Johansson, 2008), and which is predicted to accelerate due to global warming (Anisimov and Reneva, 2006; Schaefer et al., 2011), previously frozen OM will become increasingly available for microbial decomposition and export by water (Zimov et al., 2006; Frey and McClelland, 2009). A recent study on polygonal tundra soils in the Lena-Delta showed the importance of the active layer for soil organic carbon storage with on average  $7.6 \text{ kg m}^{-2}$  organic carbon in the active layer (Zubrzycki et al., 2012). However, so far little is known about OM distribution and composition in the active layer and therefore the susceptibility of different OM components to degradation and export into rivers and the Arctic Ocean. Although still discussed controversially, several results from incubation experiments of Arctic soils indicate that microbial activity is temperature sensitive (Guicharnaud et al., 2010) and that after decomposition of labile OM, recalcitrant components will be decomposed at higher soil temperatures due to microbial community changes (Biasi et al., 2005; Knoblauch et al., 2008). However, OM bound to soil minerals may survive biodegradation as suggested

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by Vonk et al. (2010) who measured high  $^{14}\text{C}$  ages of vascular plant-derived lipids in sediments in front of Arctic river mouths (Vonk, 2010; Gustafsson et al., 2011). Physical and chemical stabilization processes, which are considered to play an important role for OM stabilization in soils and sediments (Arnarson and Keil, 2007; Kögel-Knabner et al., 2008; Marschner et al., 2008), have not been investigated in the active layer of permafrost soils.

OM stabilization has been investigated quite intensively in temperate surface soils (Sollins et al., 1996; von Lützow et al., 2007; Kögel-Knabner et al., 2008). Here, physical protection mechanisms were found to be more important than chemical recalcitrance for soil OM stabilisation (Marschner et al., 2008). However, the active layer of permafrost soils differs strongly from temperate soils, e.g. it often contains less mineral substrate, has a high root penetration, and is affected by freezing and thawing processes, which may influence aggregation processes.

Various fractionation methods have been used to separate differently stabilized OM and thereby gain information on stabilization processes (von Lützow et al., 2007) which produce quite heterogeneous OM fractions not representative for distinct functional pools (von Lützow et al., 2007; Trumbore, 2009). Therefore, a combination of  $^{14}\text{C}$  and qualitative analysis of soil physical fractions was proposed, which identifies carbon turnover and the chemical properties of OM on a molecular-level (Quideau et al., 2000; Schöning and Kögel-Knabner, 2006; Trumbore, 2009). This integrative approach has been applied in marine and terrestrial environments giving useful insights into carbon sources and dynamics. For example terrestrial carbon input and turnover has been studied in marine sediments of rivers continental margins (Dickens et al., 2006; Wakeham et al., 2009), while in soils different OM sources, their stabilization and decomposition have been identified (Baldock et al., 1992; von Lützow et al., 2008; Mueller et al., 2009).

Solid state  $^{13}\text{C}$  nuclear magnetic resonance spectroscopy (NMR), recording the relative abundance of different functional groups, can be used to get a non-destructive overview of the chemical OM composition and its decomposition stage (Baldock et al.,

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1992). However, the precise structure of soil OM or specific organic compounds cannot be determined by this method (Kögel-Knabner, 2000). More detailed information is obtained from macromolecular compounds and diagnostic lipids in soil OM (Simpson and Simpson, 2012). The dynamics of organic carbon in physically defined OM fractions are revealed by  $^{14}\text{C}$  analysis.

In this study, we investigated the composition of differently stabilized soil OM compartments in the shallow active layer (ca. 25 cm) and the uppermost still frozen permafrost in a polygonal rim on Samoylov Island (Lena Delta, Siberia). Soil OM from different depth intervals was separated by particle size and density fractionation. The composition of  $^{14}\text{C}$  dated OM in silt and clay fractions as well as in free and occluded particulate OM was characterized by  $^{13}\text{C}$  cross-polarization magic angle spinning nuclear magnetic resonance spectroscopy analysis ( $^{13}\text{C}$ -CPMAS NMR) and lipid analysis.

## 2 Material and methods

### 2.1 Site description and sampling

The polygonal tundra investigated in this study is located on Samoylov Island (72.37° N, 126.48° E), which is part of the active eastern section of the Lena-Delta, Siberia. The Lena delta is the largest (32 000 km<sup>2</sup>) of the Arctic (Are and Reimnitz, 2000) and is located in the zone of continuous permafrost, which has a thickness of 400–600 m in the delta (Romanovskii and Hubberten, 2001). The climate is Arctic continental with low mean annual air temperatures of  $-14,7^{\circ}\text{C}$ , great temperature differences between winter (September to May:  $-30^{\circ}\text{C}$ ) and summer (June to August:  $7^{\circ}\text{C}$ ), and low mean annual precipitation of 190 mm per year. The island is composed mainly by middle Holocene fluvial deposits (Hubberten et al., 2006). Polygonal patterns, dominating the landscape, have formed due to freezing-thawing cycles of ice-wedges (Mueller, 1997). The polygon rims cover about 55 % of the area and the several centimetres lower

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5 polygon centres ca. 45 % (Kutzbach et al., 2004). Glacic Aquiturbels are the dominant soil type of the polygon rims and Typic Historthels dominate the polygon centers (Pfeifer et al., 2002). Vegetation is mainly formed by mosses, grasses and additionally on the polygon rims by lichens, some herbs and small shrubs like *Salix spp.* or *Betula spp.* and in the polygon centres by some sedges (Mueller, 1997).

10 Soil sampling was done during the LENA-2009 and LENA-2010 expeditions of the Alfred-Wegener-Institute, Bremerhaven (Germany; Boike et al., 2009). Samples were taken from different soil layers of the active layer (0–25 cm) and of the uppermost still frozen permafrost (25–30<sub>p</sub> cm) of the polygon rim (Table 1) at the end of the summer season in mid August to beginning of September, when the active layer has reached its maximal depth. Sample material was stored and transported frozen in pre-combusted glass jars at –20 °C and freeze-dried prior to analysis.

## 2.2 Physical fractionation

15 A combined density and particle-size fractionation was applied (Mueller et al., 2009). Briefly, 20 g of soil material were suspended in sodium polytungstate solution (1.8 g cm<sup>-3</sup>) and allowed to settle over night. The floating free particulate organic matter (fPOM) was extracted by sucking via a water jet pump. Sodium polytungstate was removed from the fPOM fraction by washing several times with deionised water on a sieve of 20 µm mesh size. The remaining slurry was dispersed ultrasonically (200 J ml<sup>-1</sup>; Bandelin, Sonopuls HD 2200) in order to break down soil aggregates. The energy input had been tested before to avoid disruption of coarse POM. After sonication the occluded POM (oPOM) was separated from the mineral residue by centrifugation. The fraction < 20 µm (oPOM<sub><20</sub>) was obtained by washing the oPOM<sub>>20</sub> over a sieve of 20 µm mesh size via pressure filtration until the electric conductivity dropped below 5 µS cm<sup>-1</sup>. Sand (63–2000 µm) and coarse silt (20–63 µm) were separated by wet sieving and medium silt (6.3–20 µm), fine silt (2–6.3 µm) and clay (< 2 µm) by sedimentation. All fractions were freeze-dried and weighed before further analysis. Mass recovery of the SOM fractionation was between 97 to 99 %.

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## 2.3 Bulk analysis

Bulk soil samples and soil fractions were manually ground in a porcelain bowl with liquid nitrogen to gain a fine homogeneous material. Total C and N contents were determined with an elemental analyzer (Analysator Vario micro cube, Elementar, Germany). The samples were carbonate-free, therefore total C contents were set equal to total organic carbon content (TOC). Soil pH values were measured in H<sub>2</sub>O (soil water ratio 1 : 2.5) one hour after water addition.

## 2.4 NMR analysis

For <sup>13</sup>C cross-polarization magic angle spinning nuclear magnetic resonance spectroscopy analysis (<sup>13</sup>C-CPMAS NMR) accomplished with a Bruker DSX 200 spectrometer (Bruker BioSpin GmbH, Karlsruhe, Germany) samples were filled into zirconium dioxide rotors and spun in a magic angle-spinning probe at a rotation speed of 6.8 kHz to minimize chemical anisotropy. A ramped <sup>1</sup>H pulse was used during a contact time of 1 ms to prevent Hartmann–Hahn mismatches. The delay times ranged from 400 ms for mineral fractions to 1000 ms for POM fractions. Chemical shifts were referenced to tetramethylsilane (TMS = 0 ppm). For integration, chemical shift regions were used as given: alkyl C (10–45 ppm), O-alkyl C (45–110 ppm), aryl/olefine C (110–160 ppm), and carbonyl/carboxyl/amide C (160–220 ppm). Measurements were done on clay and oPOM<sub><20</sub> fractions for all soil horizons, except for the second horizon (6–11 cm) where only the clay fraction was measured due to the lack of oPOM<sub><20</sub> material.

## 2.5 Extraction of lipids

Lipids were extracted from bulk soil samples by accelerated solvent extraction (Dionex ASE 200, USA) and from soil fractions by sonication. 5 g of bulk soil material was extracted by ASE with dichloromethane : methanol (9 : 1, 75 bar, 120 °C, 20 min). Depending on TOC content, 100–3000 mg of each soil fraction was extracted in an

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ultrasonic bath (room temperature) with a sequence of solvents starting with methanol, dichloromethane : methanol (1 : 1), and finally dichloromethane (25 ml, 5 min each solvent). The three extracts were combined and solvents were rotary evaporated. After saponification of the total lipid extracts with methanolic KOH (0.5 M), neutral lipids were recovered by liquid-liquid phase separation and *n*-fatty acids after acidification with 10% HCl. The neutral fraction was further separated into aliphatic, aromatic and hetero-compound fractions by column chromatography using deactivated SiO<sub>2</sub> (mesh size 60) and elution with hexane, followed by dichloromethane : hexane (2 : 1) and methanol, respectively. The *n*-fatty acid fraction was transesterified with methanolic HCl (95 : 5) to form fatty acid methyl esters (FAMES) and purified over a SiO<sub>2</sub>-Na<sub>2</sub>SO<sub>4</sub> column by eluting with dichloromethane : hexane (2 : 1).

The *n*-alkanes and *n*-fatty acid distributions were measured on a gas chromatography – flame ionization detector (GC-FID, 5890 series II plus, Hewlett Packard, USA equipped with DB-5MS column 50 m, 0.2 mm ID, 0.33 μm df). Identification and quantification of lipids was done using external standard mixtures.

## 2.6 Radiocarbon analysis

Bulk soil samples and soil fractions were pretreated with 1% HCl, which was removed by washing with MilliQ water. After drying (60 °C) the samples were converted to graphite with H<sub>2</sub> over iron as catalyst (Rethemeyer et al., 2012). <sup>14</sup>C contents of soil fraction were measured on a 6 MV Tandem AMS (HVE, The Netherlands) at the University of Cologne (Germany). Bulk soil samples were measured with the MICADAS AMS at the ETH Zurich (Switzerland). <sup>14</sup>C results are reported as percent modern carbon (pMC related to 1950) with one-sigma uncertainties.

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### 3 Results

#### 3.1 Bulk parameters

##### 3.1.1 Bulk soil characteristics

Basic soil parameters of the three depth intervals of the shallow active layer (25 cm) and the uppermost ca. 5 cm still frozen soil are presented in Table 1. The active layer consists of a decomposed organic litter layer (6 cm) and a 19 cm thick mineral soil. Soil OM in the mineral soil contained high amounts of only slightly decomposed roots from mosses, grasses and small shrubs. The soil layer at 6–11 cm depth contained sandy material of aeolian or fluvial origin. Root penetration was high at the surface and decreased rapidly with depth. The pH values in the active layer and in the uppermost permafrost were around 6.5. The four depth intervals yielded relatively high and little variable TOC contents (2.5–3.0 %) except of the 6–11 cm interval containing a sand lens, which had a lower content (1.5 %). Similarly, C/N ratios were at a comparable level in all soils and centred around 18. Bulk soil OM <sup>14</sup>C concentrations decreased strongly in the shallow active layer from 90 to 68 pMC, which is equivalent to an apparent age increase from 866 to 3052 yr BP.

##### 3.1.2 Basic parameters of soil fractions

The upper two depth intervals of the active layer (0–11 cm) and the lowermost intervals (11–30<sub>p</sub> cm) showed a similar particle size distribution. In the upper two depth intervals the sand fraction (63–2000 μm) was the largest fraction with over 62 % of the total soil mass but decreased successive with depth down to ca. 25 % at 25–30<sub>p</sub> cm depth (Table 2). Clay sized material showed a strong increase of 10 % with depth. TOC values were very low for both sand fractions (63–2000 μm: 0.1 %) and coarse silt (20–63 μm: < 0.6 %) from all depth intervals. The medium silt fraction (6.3–20 μm) had still low and decreasing TOC values with soil depth (0.6–2.0 %). Most organic carbon was located

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in fine silt (2–6.3  $\mu\text{m}$ ) and clay ( $< 2 \mu\text{m}$ ) where TOC values were lower only in these fractions from the still frozen soil at 25–30<sub>p</sub> cm depth. TOC storage in the different size fraction related to the mass of each fraction is shown in Fig. 1a revealing that the fine silt and clay fractions contained most organic carbon (7.4–15.3  $\text{mg C g}^{-1}$  soil dry weight – DW). C/N ratios of the silt and clay fractions (20 and 15, respectively) were highest in the surface layer and decreased with particle size as well as with depth to minimal values around 10 in the frozen soil layer.

The three POM fractions made up together between 0.9–2.4 % of the total soil mass (Table 2) and stored together 2.6–7.8  $\text{mg C g}^{-1}$  DW (Fig. 2a). While the mass of fPOM strongly decreased with depth of about 40 %, the oPOM<sub>>20</sub> mass distribution was nearly constant except for the sandy 6–11 cm depth interval. The mass percentage of the oPOM<sub><20</sub> increased and was twice as high in the lowermost depth interval compared to the surface layer (Table 2). Despite the small masses of the POM fractions they had the highest TOC values (21–42 %) in all depths intervals. TOC values were constant over depth for fPOM but increased for oPOM<sub>>20</sub> and oPOM<sub><20</sub>. However, taking the mass of each fraction into account, values for fPOM decreased from 4.7  $\text{mg C g}^{-1}$  DW with depth down to 1.8  $\text{mg C g}^{-1}$  DW, whereas oPOM<sub>>20</sub> had almost constant values, except in the sandy depth interval (Fig. 2a). In contrast the amount of TOC in oPOM<sub><20</sub> increased with depth. The highest C/N ratios of 31 and 34 were measured in the fPOM and oPOM<sub>>20</sub> fraction in the surface soil. The C/N ratios decreased slightly with depth for these two fractions, whereas the oPOM<sub><20</sub> fraction had a lower C/N ratio in the uppermost soil (15), which increased in the three subsoil layers to 21. Generally decreases in C/N ratio with depths were smaller for the POM fractions than for the particle size fractions.

## 3.2 NMR results and lipid concentration

### 3.2.1 Lipid concentration of bulk soil

The *n*-alkane concentrations of the bulk soil showed a clear odd-over-even predominance with the main compounds being C<sub>27</sub> and C<sub>29</sub> (Table 1). Overall concentrations (C<sub>19</sub>–C<sub>33</sub>) were highest in surface soil and in 11–25 cm (17 and 20 µg g<sup>-1</sup> DW) and lower in the sandy 6–11 cm layer and in the uppermost permafrost soil down to 12 µg g<sup>-1</sup> DW. *n*-fatty acid concentrations in bulk SOM showed an even-over-odd-predominance with C<sub>16</sub> and C<sub>24</sub> as main compounds and overall concentrations (C<sub>14</sub>–C<sub>30</sub>) between 52–255 µg g<sup>-1</sup> DW (Table 1). The abundance of C<sub>16</sub> *n*-fatty acid decreased with depth whereas C<sub>24</sub> concentrations increased in deeper intervals of the active layer and in the adjacent permafrost.

### 3.2.2 NMR results and lipid concentration of soil fractions

<sup>13</sup>C-CPMAS NMR spectroscopy was applied to clay sized OM as well as fPOM and oPOM<sub><20</sub>; fPOM was measured only from the uppermost layer due to the lack of material. oPOM<sub><20</sub> and the clay fraction showed intensity differences with depth in their aryl and alkyl regions, while O-alkyl and carboxyl signals were within the same range. The dominant functional group of the oPOM<sub><20</sub> and clay fractions from all depth intervals was O-alkyl C (e.g. carbohydrates, Kögel-Knabner, 2002), which decreased in its abundance with active layer depth (Table 3). Alkyl C (e.g. lipids, suberin, cutin and proteins, Kögel-Knabner, 2002) intensities were slightly higher for the clay fraction (21–28 %) than for the oPOM<sub><20</sub> fraction (18–24 %). Larger signals for the aryl C (aromatic carbon, Baldock et al., 1992) shift region were measured for oPOM<sub><20</sub> (16–24 %) compared to the spectra measured in clay (13–19%). The carboxyl C signals (e.g. fatty acids, fatty ester, Baldock et al., 1992) remained constant over depth. The fPOM fraction, which was only analysed in the 0–6 cm depth interval, showed a considerably

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different composition with highest abundances of O-alkyl C and lowest of alkyl and carboxyl C.

Lipid concentrations were analysed in four particle size fractions only, as sand fractions were not considered to be important for soil OM stabilization. Total extractable lipids were highest in fine silt and clay (Table 2). *n*-alkanes showed an odd-over-even predominance with the main compounds being C<sub>27</sub> and C<sub>29</sub> (Fig. 3). Overall concentrations (C<sub>19</sub>–C<sub>33</sub>) were highest in the surface soil and in 11–25 cm (17 and 20 μg g<sup>-1</sup> DW) and lower in the sandy 6–11 cm layer and in the uppermost permafrost soil (12 μg g<sup>-1</sup> DW). Coarse and medium silt contained extremely low total *n*-alkane concentrations (6 to 17 μg g<sup>-1</sup> DW in 0–6 cm), which decreased further with soil depth. While *n*-alkanes in coarse silt showed no characteristic distribution, medium silt was dominated by C<sub>27</sub> and C<sub>29</sub> *n*-alkanes (Table 2), which decreased in abundance from about 5 μg g<sup>-1</sup> DW (0–6 cm) to 1 μg g<sup>-1</sup> DW (25–30<sub>p</sub> cm; data not shown). Fine silt and clay fractions, which were also dominated by C<sub>27</sub> and C<sub>29</sub> compounds, had higher total *n*-alkane concentrations in all depth intervals than the bulk soil. Medium-chain length *n*-alkanes (C<sub>19</sub>–C<sub>24</sub>) were nearly absent in the coarse and medium silt sized fractions (data not shown) and occurred in relatively low abundances in fine silt and clay.

Similar to the *n*-alkanes, total *n*-fatty acid concentrations (C<sub>14</sub>–C<sub>30</sub>) were extremely low (< 10 μg g<sup>-1</sup> DW) in the coarse and medium silt fractions in all depth intervals with no dominant compounds (data not shown), while higher concentrations were determined in fine silt and clay (45–161 μg g<sup>-1</sup> DW) with C<sub>16</sub> and C<sub>24</sub> *n*-fatty acids being the most abundant compounds (Fig. 4). The dominance of the C<sub>16</sub> *n*-fatty acid in fine silt and clay decreased with depth and shifted to C<sub>24</sub>, which is the most abundant compound in the uppermost permafrost soil. In these two fractions total *n*-fatty acid concentrations decreased with depth within the active layer (0–25 cm) but increase again to concentrations twice as high in the frozen soil layer (25–30<sub>p</sub> cm). This increase mainly resulted from higher abundances of compounds with > C<sub>20</sub> chain-length.

The *n*-alkane concentrations in the fPOM, oPOM<sub>>20</sub> and oPOM<sub><20</sub> fractions were generally higher (15–98 μg g<sup>-1</sup> DW) than in the bulk soil OM and particle size fractions,

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and yielded high abundances of medium-chain length compounds (C<sub>19</sub>–C<sub>24</sub>) representing 33–55 % of the overall concentration. Highest *n*-alkane concentrations were measured in oPOM<sub><20</sub> from all depth intervals. Concentration increased with active layer depth in all three fractions but dropped in the frozen soil layer (25–30<sub>p</sub> cm) in fPOM and oPOM<sub><20</sub> fractions (Fig. 3). fPOM and oPOM<sub><20</sub> were dominated by C<sub>23</sub> and the long-chain compounds C<sub>27</sub> and C<sub>29</sub>, while oPOM<sub>>20</sub> mainly contained *n*-alkanes of medium-chain length centring around C<sub>23</sub>.

*n*-fatty acid concentrations of all density fractions were considerably higher compared to *n*-alkanes particularly in fPOM for the uppermost two depth intervals and had a clear even-over-odd predominance (Fig. 4). Overall concentrations (C<sub>14</sub>–C<sub>30</sub>) varied between 90 and 1129 μg g<sup>-1</sup> DW and increased with depth in the oPOM<sub>>20</sub> fraction (from 103 to 625 μg g<sup>-1</sup> DW). The fPOM and oPOM<sub><20</sub> fractions yielded highest *n*-fatty acid concentrations in the sandy depth interval (6–11 cm) with 1129 and 395 μg g<sup>-1</sup> DW, respectively (Table 2). The dominant compounds in all fractions in the two surface layers were C<sub>16</sub> and C<sub>18</sub> *n*-fatty acids. Compound distributions changed with depth to longer chain length with C<sub>24</sub> being the major compound in oPOM<sub>>20</sub> and oPOM<sub><20</sub> at 11–30<sub>p</sub> cm. In the still frozen depth interval highest total *n*-fatty acid concentration were present in oPOM<sub>>20</sub>, which contained predominantly long-chain compound (C<sub>24</sub>–C<sub>30</sub>).

### 3.3 Radiocarbon contents of soil fractions

The various soil fractions showed decreasing <sup>14</sup>C contents towards larger fraction size and greater soil depth (Figs. 1b and 2b). Fine silt and clay had similar <sup>14</sup>C contents like bulk OM and also comparable values over all depth intervals (Fig. 1b), which were slightly higher (71–94 pMC) than concentrations of bulk OM (68–90 pMC). The medium silt fraction had lower <sup>14</sup>C contents (54–82 pMC) than fine silt and clay fractions and bulk OM. Lowest <sup>14</sup>C contents were measured in the coarse silt fractions, which decreased most strongly with depth down to 24 pMC in the uppermost permafrost soil (25–30<sub>p</sub> cm). This fraction had the lowest contents of all analysed fractions.

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The three density fractions also showed decreasing  $^{14}\text{C}$  contents in the order fPOM > oPOM<sub>>20</sub> > oPOM<sub><20</sub> and a decrease with soil depth (Fig. 2b) but concentrations were slightly higher in both surface layers than in bulk soil OM. The highest  $^{14}\text{C}$  contents and the smallest decrease with depth from 102 to 86 pMC was measured for the fPOM fraction. The oPOM fractions decreased more strongly in  $^{14}\text{C}$  content with depth, except of a higher value for oPOM<sub>>20</sub> from 11–25 cm soil depth, and yielded lowest values in the frozen layer, which are similar to that of the bulk soil.

## 4 Discussion

### 4.1 Distribution and composition of bulk soil organic matter

We found considerable differences of the physical and chemical soil properties in the shallow active layer thawing during summer (0–25 cm) and the uppermost still frozen permafrost (25–30<sub>p</sub> cm) of a polygonal rim on Samoylov Island (Table 2). The distribution of the separated particle size fractions revealed changes in the textural composition from the two near surface intervals (0–6 cm and 6–11 cm) dominated by sand-sized material (60 %) to the two lower horizons (11–25 and 25–30<sub>p</sub> cm), which contained more than 65 % silt and clay-sized material. High sand contents are most probably of aeolian origin eroded from the sandbanks of the Lena-Delta (Hubberten et al., 2006). Differences in soil texture in the surface soil and near the permafrost table may be explained by soil developing processes like physical weathering and vertical translocation of fine-grained particles as well as by cryoturbation processes. Intense freezing-thawing cycles as well as the breakup of particles by cryohydration occurring in cold climates may produce fine-grained particles (Jones et al., 2010).

Changes in chemical soil properties in the active layer are reflected by the elemental composition and, more detailed, by the molecular analyses of organic compounds and structures. The TOC contents in the active layer of the polygonal rim (on average 2.5 %; Table 1) are characteristic for OM in tundra soil developed in Holocene fluvial

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deposits in this area (Zimov et al., 2006; Wagner et al., 2007; Xu et al., 2009). The relatively high C/N ratios around 18 most likely reflect the slow transformation of the OM at cold temperatures and indicate the presence of little decomposed OM. Likewise, the high and rapidly, nearly linearly increasing apparent  $^{14}\text{C}$  ages of bulk OM with depth (866–3052 yr BP; Table 1) suggest slow organic carbon turnover and the accumulation of “old” material, respectively, and no significant mixing by cryoturbation. We did not expect such low  $^{14}\text{C}$  values in the shallow active layer since previous results, which were however determined for OM from permafrost cores from Samoylov Island, were around 2300 yr BP in 289 cm and 7900 yr BP in 557 cm (Wagner et al., 2007). Instead, quite similar  $^{14}\text{C}$  results were determined for OM in the active layer of tundra soils in Russia (66° N, 42° E) and on Svalbard (78–79° N, 13–17° E), Norway, with  $^{14}\text{C}$  concentrations declining from modern values to about 53 pMC equivalent to 5100 yr BP in the upper 40 cm of soils from various sites (Cherkinsky, 1996) and for OM of three active layers (35–50 cm) from sites in Alaska (64–67° N, 147–150° W) with  $^{14}\text{C}$  ages between 585–3363 yr BP (Waldrop et al., 2010).

The *n*-alkane distribution in bulk OM (Table 1) suggests that a major portion in all depth intervals is plant-derived material indicated by  $\text{C}_{27}$  and  $\text{C}_{29}$  *n*-alkanes being the most abundant compounds, which are major compounds of epicuticular waxes (Eglinton and Hamilton, 1967) frequently used as biomarkers for higher plant input to soils and aquatic sediments (e.g. Jambu et al., 1991; Huang et al., 2000). Likewise the *n*-fatty acid distribution maximizing at  $\text{C}_{24}$  reflects the terrestrial plant origin of OM in the two surface soil layers (0–11 cm; data not shown). Since lipid concentrations were found to correlate with TOC contents (Naafs et al., 2004), the strong increase in concentrations of the long-chain *n*-fatty acid compounds in the lowermost active layer interval and in still frozen soil may result from inputs of other sources than plant material at greater depth or more likely, from in-situ secondary production by the transformation of plant wax esters or root-derived polyesters and suberin (Ficken et al., 1998; Bull et al., 2000; Andersson and Meyers, 2012).

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In summary, relatively high C/N values and abundances of long-chain *n*-alkanes in the active layer and adjacent upper permafrost reflected that soil OM consisted of little degraded plant-material. This result is in contrast to the high and increasing apparent <sup>14</sup>C ages with active layer depth, which is generally considered to result from the accumulation of strongly transformed, “old” organic matter (Rumpel and Kögel-Knabner, 2011). However, little degraded, “old” plant material in the depth intervals near the permafrost table may be protected against biodegradation by preservation mechanisms like interaction with clay particles, which increase in abundances in these layers.

## 4.2 Stabilization mechanisms of SOM

Three mechanisms are known to be relevant for OM stability in temperate soils, which are (a) formation of soil aggregates, (b) formation of organo-mineral associations, and (c) the chemical recalcitrance of organic compounds (Sollins et al., 1996; Christensen, 2001; Six et al., 2002; Baldock et al., 2004; Kögel-Knabner et al., 2008). However, so far it is not known if these mechanisms are relevant and if so which of them is of primary importance in the active layer of Arctic soils. Since reduced bioaccessibility of OM in soil aggregates and of organo-mineral assemblages was found to be of major importance for OM preservation in various temperate soils (Sollins et al., 2006; Marschner et al., 2008; Sanaullah et al., 2011), we analyzed the composition of OM in <sup>14</sup>C dated particle size and density fractions from depth intervals of the active layer and uppermost still frozen soil of the polygonal rim to get information about OM sources and protection mechanisms.

### 4.2.1 Organic matter in soil aggregates

The fractions, fPOM, oPOM<sub>>20</sub> and oPOM<sub><20</sub> separated by density fractionation, showed strong differences in their distribution and chemical composition in the active layer (0–25 cm) and the uppermost still frozen permafrost (25–30<sub>p</sub> cm). Similar to previous studies, fPOM material had higher <sup>14</sup>C contents and thus presumably also

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5 a faster turnover than oPOM material (Quideau et al., 2000; Rethemeyer et al., 2005; von Lützow et al., 2007; Trumbore, 2009), which was attributed to stabilization due to occlusion in aggregates and the recalcitrance of organic components in this fraction (Golchin et al., 1997; Poirier et al., 2005). In general, the POM material from all

10 The three fractions however, showed strong compositional differences between each other and with soil depth. The light fPOM fraction, which is not occluded in aggregated soil structures, representing slightly decomposed root and plant material (Golchin et al., 1994), was the most abundant and carbon-rich fraction in the surface soil (0–6 cm) but decreased rapidly in abundance with active layer depth (Fig. 2a). The composition of fPOM material changed with depth from carbohydrate-rich (O-alkyl C; Table 3) material to more strongly decomposed plant litter as suggested by decreasing C/N ratios (Table 2). However, the absence of the dominant long-chain *n*-alkanes (C<sub>27</sub> and C<sub>29</sub>) below 11 cm soil depth may also reflect a change in fPOM sources like decreasing proportions of plant leaf wax compounds and higher amount of still relatively “young” root-derived OM.

20 Organic matter occluded in soil aggregates, which was released upon sonication, increased in importance regarding carbon storage near and below the permafrost table (Fig. 2a). Both oPOM fractions, < 20 μm and > 20 μm, showed a quite similar and stronger <sup>14</sup>C decrease with depth compared to the small <sup>14</sup>C decrease in fPOM material, presumably indicative for better preservation of OM occluded in soil aggregates (Golchin et al., 1997). The high value for oPOM<sub>>20</sub> from 11–25 cm may be biased by the presence of larger plant or root fragments. Despite quite similar <sup>14</sup>C values, both oPOM fractions showed considerable compositional differences. Surprisingly the composition of the large oPOM<sub>>20</sub> fraction obtained after aggregate disruption is quite similar to that of fPOM material, both being little decomposed, plant-derived OM with high C/N ratios around 30 decreasing slightly with depth. The dominance of medium chain-length

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*n*-alkanes and the lack of long-chain compounds in oPOM<sub>>20</sub> (C<sub>27</sub> and C<sub>29</sub>) however suggest either stronger OM transformation or different plant sources contribution to OM in this fraction.

In contrast OM in the small oPOM<sub><20</sub> fraction is apparently derived from other sources since typical leaf wax-derived C<sub>27</sub> and C<sub>29</sub> *n*-alkanes were dominant compounds in most depth intervals. Declining abundances of O-alkyl C (carbohydrate-rich material) and higher proportions of aryl C (aromatic compounds) as well as increasing alkyl C/O-alkyl C (Baldock et al., 1992) and aryl C/O-alkyl C ratios (Quideau et al., 2000) with depth, suggest a stronger transformation of small OM occluded in aggregates, which was also found in temperate soils (Mueller and Koegel-Knabner, 2009).

The lower <sup>14</sup>C values and higher TOC content of oPOM compared to fPOM suggest that aggregate formation may play a considerable role for OM preservation in these Arctic soils, however only at greater active layer depth. The effect seems to be not as prominent as in temperate surface soils under grassland and forest where oPOM fractions were shown to account for around 5% of the soil mass, but contained over 30% of the bulk soil TOC (Mueller and Koegel-Knabner, 2009). In this study the oPOM fractions from the active layer had a cumulative mass of about 1% containing 0.9–3.9 mg C g<sup>-1</sup> DW (8–16% of total soil TOC) (Table 2; Fig. 2a).

#### 4.2.2 Organo-mineral associations

Although the uppermost 11 cm of the active layer were dominated by sand sized material, most organic carbon was stored in the clay and fine silt fractions (< 6.3 μm) containing up to 15.3 mg C g<sup>-1</sup> DW (62% of total soil TOC) and increasing in importance with depth (Fig. 1a; Table 2). This is quite similar to results for temperate soils where clay and silt fractions also had highest TOC contents of above 65% (Flessa et al., 2008) and above 50% (Rumpel et al., 2004). The <sup>14</sup>C contents of the size fractions showed a surprising trend from highest values for fine silt and clay sized OM, which were close to that of bulk OM, to extremely low values for coarse silt (20–63 μm). This is in contrast to results of most previous studies of temperate soils, where often clay to

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silt sized OM (< 63  $\mu\text{m}$ ) both had relatively low  $^{14}\text{C}$  contents while sand sized material (200–2000  $\mu\text{m}$ ) often showed highest  $^{14}\text{C}$  values (Schöning and Kögel-Knabner, 2006; Sollins et al., 2006; von Lützow et al., 2007). Presumably the very low  $^{14}\text{C}$  contents of coarse silt may be affected by preserved POM artefacts in micro-aggregate structures.

More intense transformation of OM with decreasing particles size was observed in the uppermost 11 cm of the active layer where C/N values decreased from sand to clay. However this trend, which was also observed in numerous previous studies of temperate soils and was often attributed to enrichment of microbial reworked products in fine mineral fractions (Sollins et al., 2006; von Lützow et al., 2007; Flessa et al., 2008; Bol et al., 2009; Sollins et al., 2009), is not seen in deeper sampling intervals. Here closely alike C/N ratios of coarse silt to clay-sized material (< 63  $\mu\text{m}$ ) suggest a similar transformed OM. The decomposition degree of OM in fine silt and clay slightly increased with depth as shown by decreasing C/N ratios and increasing alkyl C/O-alkyl C ratios in the clay fraction. These results reflect that the labile O-alkyl C structures like carbohydrates are less stabilized as described already for temperate soils (Rumpel et al., 2004).

Fine silt and clay showed a very similar *n*-alkane and *n*-fatty acid distributions as well as  $^{14}\text{C}$  values in the different depth intervals. The plant source of soil OM in the fine silt and clay fraction did not change significantly over depth as indicated by *n*-alkanes being dominated by leaf wax-derived  $\text{C}_{27}$  and  $\text{C}_{29}$ . The increase of *n*-fatty acid concentration in the uppermost permafrost layer (25–30<sub>p</sub> cm) compared to the concentration in the active layer are most likely the result of in-situ production by the hydrolysis of plant wax esters into its component *n*-alkanols and *n*-fatty acids or contribution of root-derived polyesters and suberin (Ficken et al., 1998).

Despite the lower  $^{14}\text{C}$  contents of coarse and medium silt fractions (6.3–63  $\mu\text{m}$ ) they are less important for organic carbon storage as both fractions represented very little of the total soil TOC (< 2 mgCg<sup>-1</sup> DW, Fig. 1a). The sand fractions (63–200  $\mu\text{m}$  and 200–2000  $\mu\text{m}$ ) were considered to be not important for carbon storage or soil OM stabilization as these fractions contain hardly any carbon (< 0.3 mgCg<sup>-1</sup> DW).

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## 5 Conclusions

Organic matter in the active layer of a polygon rim was dominated by only slightly decomposed, mainly higher plant-derived material with relatively low and strongly increasing apparent  $^{14}\text{C}$  ages. In temperate soils high  $^{14}\text{C}$  ages have been assumed to reflect the accumulation and stabilization of strongly transformed OM with depth. Here, the formation of organo-mineral associations has been thought to be the most important mechanisms for OM stabilization. The results of our study of an Arctic active layer soil suggest that this mechanism is less important for OM preservation in permafrost soils because of relatively higher  $^{14}\text{C}$  contents above values for bulk soil TOC and lower organic carbon contents of clay and fine silt sized OM compared to density fractionated occluded POM (representing organic material occluded in aggregates). Despite the high TOC contents of occluded POM from all depth intervals, protection of OM in soil aggregates seems to be a mechanism, which is important only at greater active layer depth suggested by relatively high  $^{14}\text{C}$  values in the uppermost 6 cm and low values near the permafrost table compared to the free light POM fraction. These results suggest that probably chemical recalcitrance, which was thought to be of minor importance for OM preservation in temperate soils, may play a considerable role in permafrost soil. Therefore, further research, like chemical oxidation treatment analysis, is needed to investigate the importance of the chemical recalcitrance of active layer and permafrost OM.

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**Table 1.** Bulk soil properties in different depths intervals of a polygonal rim on Samoylov Island, Siberia. Total extractable lipids and summed long-chain, plant wax derived *n*-alkanes (C<sub>19</sub>–C<sub>33</sub>) and *n*-fatty acids (C<sub>14</sub>–C<sub>30</sub>) are related to soil dry weight (DW).

Depth [cm]	pH	TOC [%]	C/N	<sup>14</sup> C [pMC]	<i>n</i> -alkanes [ $\mu\text{g g}^{-1}$ DW]		<i>n</i> -fatty acids [ $\mu\text{g g}^{-1}$ DW]	
					total	C <sub>27</sub> +C <sub>29</sub> +C <sub>31</sub>	total	C <sub>24</sub> +C <sub>26</sub> +C <sub>28</sub>
0–6	6.1	3.0	19.5	90	17	11	90	24
6–11	6.4	1.5	17.8	83	14	9	52	15
11–25	6.3	3.0	18.4	78	20	11	255	120
25–30 <sub>p</sub>	6.7	2.5	16.4	68	12	6	178	82

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**Table 2.** Mass and elemental composition of density and particle size soil fractions.

Parameter	Depth [cm]	fPOM	oPOM <sub>&gt;20</sub>	oPOM <sub>&lt;20</sub>	coarse + medium sand (200–2000 μm)	fine sand (63–200 μm)	coarse silt (20–63 μm)	medium silt (6.3–20 μm)	fine silt (2–6.3 μm)	clay (< 2 μm)
mass [%]	0–6	1.3	0.5	0.6	30.2	32.4	16.6	9.6	3.3	5.6
	6–11	0.4	0.1	0.3	31.5	36.2	14.9	8.3	3.1	5.2
	11–25	0.7	0.4	0.7	8.4	24.3	27.6	17.1	7.8	12.9
	25–30 <sub>p</sub>	0.5	0.5	1.2	5.5	19.7	25.8	22.3	9.4	15.0
TOC <sup>1</sup> [%]	0–6	36.9	32.9	21.2	0.1	0.1	0.6	2.0	10.9	9.8
	6–11	38.2	37.6	12.5	0.0	0.1	0.3	1.5	8.6	9.2
	11–25	37.5	40.2	31.9	0.0	0.1	0.3	1.0	8.0	7.0
	25–30 <sub>p</sub>	37.1	42.4	32.1	0.0	0.1	0.2	0.6	5.0	5.0
C/N	0–6	31	34	15	n.d.	n.d.	30	18	20	15
	6–11	25	28	21	n.d.	n.d.	25	17	17	13
	11–25	24	29	21	n.d.	n.d.	14	14	16	12
	25–30 <sub>p</sub>	24	24	21	n.d.	n.d.	9	11	13	11
<i>n</i> -alkanes total	0–6	29	23	69	n.d.	n.d.	6	17	39	21
	6–11	53	68	91	n.d.	n.d.	8	5	40	44
(C <sub>19</sub> –C <sub>33</sub> ) [μg g <sup>-1</sup> DW]	11–25	15	33	98	n.d.	n.d.	3	9	35	40
<i>n</i> -fatty acids total	0–6	394	99	130	n.d.	n.d.	5	10	80	70
(C <sub>14</sub> –C <sub>30</sub> ) [μg g <sup>-1</sup> DW]	11–25	88	386	218	n.d.	n.d.	2	3	58	40
	25–30 <sub>p</sub>	279	620	223	n.d.	n.d.	3	8	105	158

<sup>1</sup> Total organic carbon.

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**Table 3.** Relative contents [%] of chemical structures identified by  $^{13}\text{C}$ -CPMAS NMR spectroscopy in the clay fraction ( $< 2 \mu\text{m}$ ) and in free (fPOM) and occluded particulate organic matter  $< 20 \mu\text{m}$  (oPOM $_{<20}$ ).

Fraction	Depth [cm]	Alkyl C	O-alkyl C	Aryl C	Carboxyl C	Alkyl C/ O-alkyl C	Aryl C/ O-alkyl C
fPOM	0–6	15	56	24	6	0.27	0.42
oPOM $_{<20}$	0–6	21	52	16	12	0.40	0.30
	6–11	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
	11–25	18	47	24	12	0.39	0.51
	25–30 <sub>p</sub>	24	40	24	12	0.59	0.60
clay	0–6	23	51	13	14	0.45	0.25
	6–11	21	50	19	11	0.41	0.37
	11–25	24	49	17	10	0.48	0.34
	25–30 <sub>p</sub>	28	43	15	13	0.65	0.35

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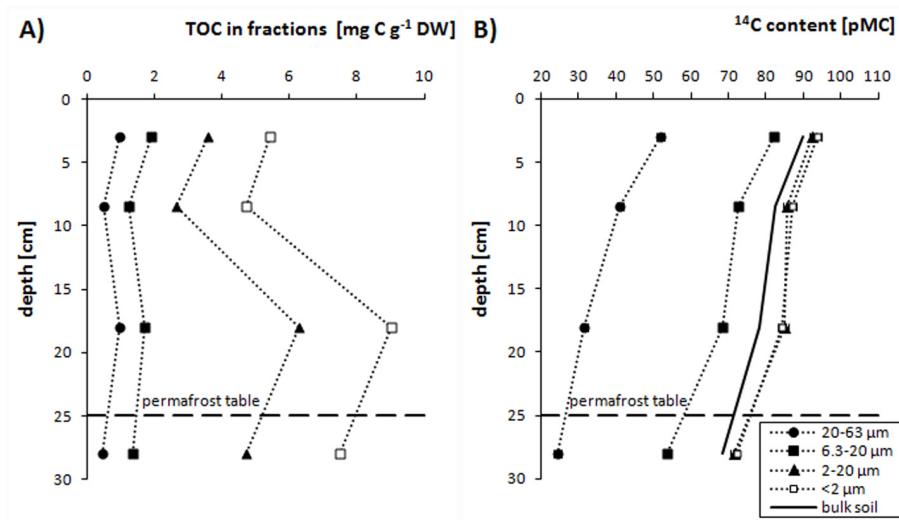


Fig. 1. Distribution of (A) total organic carbon content (TOC) in particle size fractions related to fraction mass and (B) <sup>14</sup>C content (absolute errors are indicated by bars on each point) over depth in the coarse silt (20–63 μm), medium silt (6.3–20 μm), fine silt (2–6.3 μm) and clay (< 2 μm).

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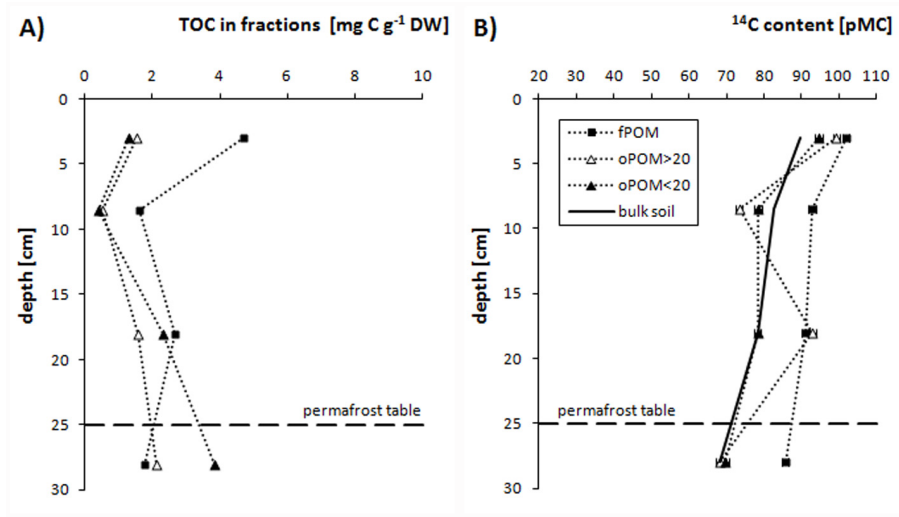
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**Fig. 2.** Distribution of **(A)** organic carbon content (TOC) in density fractions related to fraction mass and **(B)** <sup>14</sup>C content (absolute errors are indicated by bars on each point) over depth.

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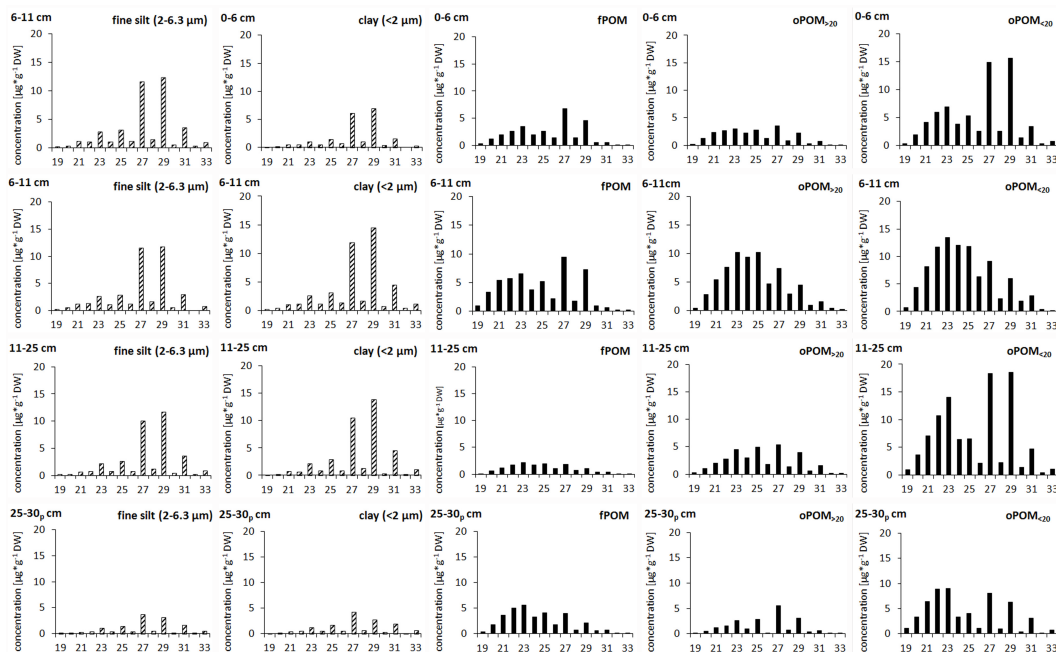
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**Fig. 3.** *n*-alkane concentrations in fine silt and clay (striped bars) and in three density fractions (black bars) from different depths intervals. Only odd *n*-alkanes are labelled on the x-axis.

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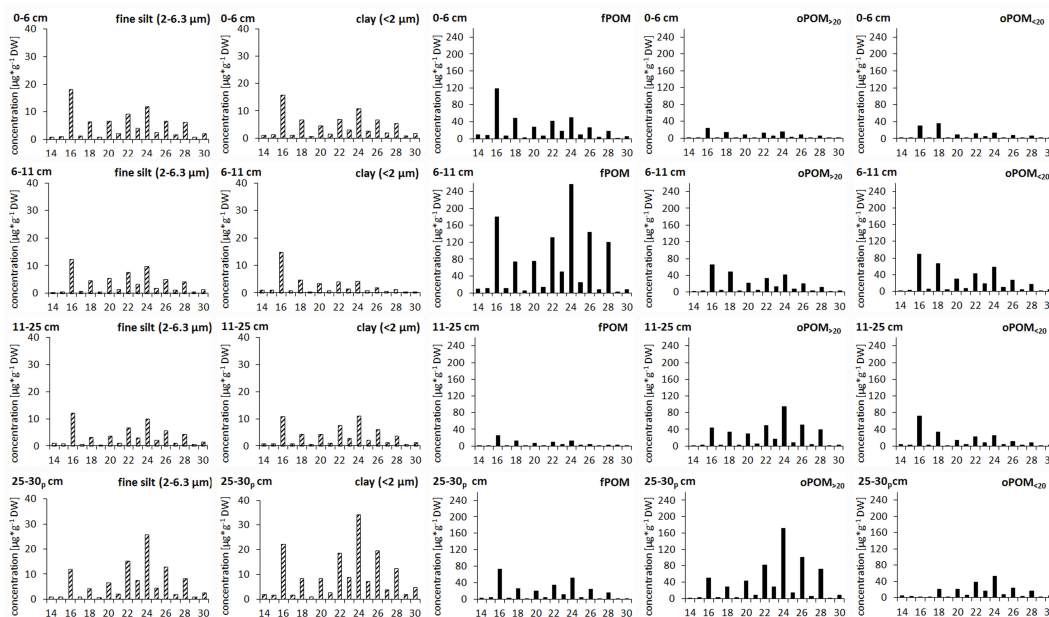
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**Fig. 4.** Distribution of *n*-fatty acid concentrations in fine silt and clay (striped) and in density fractions (black) from different depths intervals. Only even *n*-fatty acids are labelled on the x-axis; note different scaling of y-axis.

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