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Improved quantification of microbial CH₄ oxidation efficiency in Arctic wetland soils using carbon isotope fractionation

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Abstract

Permafrost-affected tundra soils are significant sources of the climate-relevant trace gas methane (CH_4). The observed accelerated warming of the Arctic will cause a deeper permafrost thawing followed by increased carbon mineralization and CH_4 formation in water saturated tundra soils which might cause a positive feedback to climate change. Aerobic CH_4 oxidation is regarded as the key process reducing CH_4 emissions from wetlands, but quantification of turnover rates has remained difficult so far. The application of carbon stable isotope fractionation enables the in situ quantification of CH_4 oxidation efficiency in arctic wetland soils. The aim of the current study is to quantify CH_4 oxidation efficiency in permafrost-affected tundra soils in Russia's Lena River Delta based on stable isotope signatures of CH_4 . Therefore, depth profiles of CH_4 concentrations and $\delta^{13}\text{C}_{\text{CH}_4}$ -signatures were measured and the fractionation factors for the processes of oxidation (α_{ox}) and diffusion (α_{diff}) were determined.

Most previous studies employing stable isotope fractionation for the quantification of CH_4 oxidation in soils of other habitats (e.g. landfill cover soils) have assumed a gas transport dominated by advection ($\alpha_{\text{trans}} = 1$). In tundra soils, however, diffusion is the main gas transport mechanism, aside from ebullition. Hence, diffusive stable isotope fractionation has to be considered. For the first time, the stable isotope fractionation of CH_4 diffusion through water-saturated soils was determined with an $\alpha_{\text{diff}} = 1.001 \pm 0.000$ ($n = 3$). CH_4 stable isotope fractionation during diffusion through air-filled pores of the investigated polygonal tundra soils was $\alpha_{\text{diff}} = 1.013 \pm 0.003$ ($n = 18$). Furthermore, it was found that α_{ox} differs widely between sites and horizons (mean $\alpha_{\text{ox}} = 1.017 \pm 0.009$) and needs to be determined individually. The impact of both fractionation factors on the quantification of CH_4 oxidation was analyzed by considering both the potential diffusion rate under saturated and unsaturated conditions and potential oxidation rates. For a submerged organic rich soil, the data indicate a CH_4 oxidation efficiency of 50 % at the anaerobic-aerobic interface in the upper horizon. The improved in situ quantification of CH_4 oxidation in wetlands enables a better

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assessment of current and potential CH₄ sources and sinks in permafrost affected ecosystems and their potential strengths in response to global warming.

1 Introduction

With a global warming potential 25 times higher than carbon dioxide on a century time scale (Forster et al., 2007), methane (CH₄) is an important greenhouse gas in the climate system. Much research effort focuses on identifying the global CH₄ sources and sinks to estimate not only their current strength, but also their potential in response to land-use change and global warming (Walter et al., 2007; Dlugokencky et al., 2009; Keppler et al., 2006). In the focus of this study are arctic wetlands which hold enormous amounts of organic carbon (Tarnocai et al., 2009; Zubrzycki et al., 2012) and are significant sources of CH₄ (Wille et al., 2008; Tagesson et al., 2012). With the observed accelerated warming of the Arctic, a deeper permafrost thawing will cause increased carbon mineralization and CH₄ formation in water saturated tundra soils, bearing the potential to cause a positive feedback to climate change (Anisimov, 2007; Schuur et al., 2009; Åkerman and Johansson, 2008).

One of the key processes regulating wetland CH₄ fluxes is aerobic microbial CH₄ oxidation (Segers, 1998; Whalen, 2005) which is performed by methanotrophic bacteria. CH₄ is formed in the final step of anaerobic microbial degradation of organic matter and is released from wetlands via three transport mechanisms: (1) diffusion along the concentration gradient between wetland soil and atmosphere, (2) ebullition in the form of gas bubbles due to CH₄ supersaturation and (3) plant-mediated transport through gas conducting tissue, the so-called aerenchyma (Joabsson et al., 1999; Lai, 2009; Whalen, 2005; Kutzbach et al., 2004). In contrast to the other two pathways, the slow diffusive flux facilitates extended contact of CH₄ with methanotrophic bacteria. This pathway may allow more than 90 % of the available CH₄ to be oxidized to CO₂ before it reaches the soil surface (Roslev and King, 1996; Sundh et al., 1995). The extent to which the produced CH₄ is oxidized, the CH₄ oxidation efficiency, is controlled by the

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key factors (1) rate of microbial oxidation (Wang et al., 2004) and (2) rate of diffusion of CH₄ (Curry, 2009; Dueñas et al., 1994). These parameters are mainly governed by the abundance and composition of methane oxidizing microbial communities and the environmental factors CH₄ and oxygen (O₂) availability, soil air-filled porosity and soil water content.

To quantify the CH₄ oxidation efficiency several methods including batch or column laboratory experiments and in situ measurements are currently employed, yet each displays different limitations (Huber-Humer et al., 2009). Gas push-pull tests (GPPT) inject and extract a defined volume of a gas mixture of a reactive gas (e.g. CH₄) and a conservative tracer (e.g. argon) into and from the soil and the microbial turnover is quantified by analyzing the break through curves of the gases (Streese-Kleeberg et al., 2011). GPPTs are not easily applicable at sites with low oxidation rates and high water saturation (Gomez et al., 2008; Urmann et al., 2007) such as tundra wetlands and were only successfully applied in near-surface soils with a cylinder driven 50 cm into the soil (Nauer and Schroth, 2010). Furthermore, mass balance calculations using loading and surface flux measurements to determine the fraction of oxidized CH₄ e.g. in biofilters or landfill cover soils (Powelson et al., 2007; Cabral et al., 2010; Gebert et al., 2003) are difficult to apply in wetlands since loading rates cannot be quantified in these open systems.

In addition to the abovementioned methods, studies in landfill cover soils and swamp forests determined the CH₄ oxidation efficiency by measuring the changes in the ratio of the two stable CH₄ isotopologues, ¹³CH₄ and ¹²CH₄ (Nozhevnikova et al., 2003; Chanton et al., 2008a; De Visscher et al., 1999, 2004; Happell et al., 1994; Liptay et al., 1998). The approach utilizes the fact that isotopic fractionation occurs when CH₄ is oxidized: the remaining CH₄ becomes heavier and the produced CO₂ becomes lighter (Barker and Fritz, 1981) as the light isotopologue ¹²CH₄ is oxidized faster by methanotrophic bacteria than the heavier ¹³CH₄. The isotopic fractionation factor α is

defined as the change in isotopic composition between reactant (Q) and product (P):

$$\alpha = \frac{R(^{13}\text{C}/^{12}\text{C})_Q}{R(^{13}\text{C}/^{12}\text{C})_P} \quad (1)$$

where R is the isotope ratio of heavier ^{13}C and lighter ^{12}C . The enrichment of ^{13}C in CH_4 is measured as isotopic abundance, expressed in the δ notation ($\delta^{13}\text{C}$):

$$\delta^{13}\text{C} = \frac{R_{\text{sample}}}{R_{\text{standard}}} - 1 \quad (2)$$

where R_{sample} is the isotope ratio $^{13}\text{C}/^{12}\text{C}$ of the sample and R_{std} is the $^{13}\text{C}/^{12}\text{C}$ ratio of the reference standard VPDB (Vienna Peedee Belemnite; $R_{\text{std}} = 0.0112372$) (McKinney et al., 1950).

Mahieu (2008) showed through a model-based isotope approach that isotopic fractionation by diffusion has to be taken into account as well, given that the faster diffusive transport of the lighter isotope causes an enrichment of the heavier isotope in the remaining gas phase. In air, the diffusion coefficient of $^{12}\text{CH}_4$ exceeds that of $^{13}\text{CH}_4$ by a factor of 1.0195 due to mass differences. No fractionation is expected when advection dominates gas transport (Bergamaschi et al., 1998; Chanton, 2005).

For field applications the so called “open-system equation” by Monson and Hayes (1980) is then applied to determine the CH_4 oxidation efficiency (Mahieu et al., 2008):

$$f_{\text{ox}} = \frac{(\delta_E - \delta_P)}{(\alpha_{\text{ox}} - \alpha_{\text{trans}})} \quad (3)$$

where f_{ox} is the fraction of CH_4 oxidized in the soil; δ_E is the $\delta^{13}\text{C}$ of emitted CH_4 relative to VPDB; δ_P is the $\delta^{13}\text{C}$ of produced CH_4 relative to VPDB; α_{ox} is the isotopic fractionation factor of oxidation; α_{trans} is the isotopic fractionation factor of transport.

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The Lena River Delta belongs to the area of continuous permafrost with an Arctic continental climate characterized by both low temperatures and precipitation (Boike et al., 2008). The soils only thaw in the upper part (< 60 cm) during the summer (active layer) and are rich in organic matter (Zubrzycki et al., 2012). Samples were taken during two expeditions in 2009 and 2010 in the eastern part of the island which is characterized by a wet polygonal tundra, a typical patterned ground of permafrost landscapes (French, 1996). Its microrelief consists of homogeneously spread soil units of depressed centers of low-centered ice wedge polygons (hereinafter “polygon center”) and their elevated surrounding rims. In this study four polygon centers with differing water table positions were investigated, one characterized by a permanent water level above soil surface (hereinafter “polygonal pond”), two by a changing water level close to the soil surface (hereinafter “saturated polygon center”) and the fourth by a distinctly lower water level (hereinafter “unsaturated polygon center”).

2.2 Soil sampling, storage and analysis

Soil samples were taken from pits opened to the frozen ground of four polygon centers. Mixed soil samples were collected in plastic bags, refrozen in the field and kept frozen until arrival in the laboratory. In addition, three undisturbed soil cores (100 cm³, height 4 cm) were retrieved from each horizon, stored either cooled (samples 2009) or frozen (samples 2010) until further analysis.

Prior to analysis of mixed organic samples, all living roots and plant material was removed. All mixed soil samples were air dried. The dried organic samples were cut into 2–5 mm pieces, the mineral samples sieved (to < 2 mm). Subsequently the samples were milled and dried at 105 °C. Total carbon and nitrogen were measured with an elemental analyzer (VarioMAX; Elementar, Hanau, Germany). Electrical conductivity (LF 90, WTW) and soil pH (CG 820, Schott) were determined in a suspension of 10 g of fresh soil in 50 mL of distilled water.

2.3 Vegetation analysis

Plant species were investigated according to the approach of Braun-Blanquet (1964) in three plots of 0.25 m² at each site. Species dominance of *Carex aquatilis* was estimated as the percentage of the basal area covering the plots.

2.4 Pore-water sampling, storage and gas concentration analyses

For CH₄ concentration and stable isotope (SI) profiles pore-water samples were taken at several depths via perforated stainless steel tubes (ø 1/8"), sampling 5 mL for concentration and 50 mL for SI analyses. Samples were conserved in vials or serum bottles flushed with nitrogen and containing sodium chloride, thus forming a saturated saline solution after injection. To liberate dissolved CO₂, 100 µL of 10% HCl solution was added afterwards.

Gas analyses were carried out at the field station on Samoylov Island and in the laboratory in Germany with gas chromatographs (both GC 7890, Agilent Technologies, Germany) equipped with a Porapak Q column (2 mm ID, 1.8 m length) separating CH₄ and CO₂. CH₄ concentration was measured with a flame ionization detector (FID) and concentration of CO₂ with a thermal conductivity detector (TCD). Oven, injection, FID and TCD temperatures were 40, 75, 250 and 180 °C, respectively. Helium served as carrier and make-up gas. Injection volume was 200 µL and samples were measured twice. Pore water gas concentrations were calculated from the concentration measured and the headspace volume and pressure (pressure gauge LEO1, Fa. Keller, Switzerland) applying Henry's Law and corrected for the partition of CH₄ between the aqueous and the gaseous phase by using the solubility coefficient $\beta = 0.00867 \text{ mL mL}^{-1}$ for solubility of CH₄ in a saturated saline solution at 20 °C; (Yamamoto et al., 1976; Kutzbach et al., 2004; Seibt et al., 2000). For calibration CH₄ standard gases of 1.7 and 200 ppmv, 1, 10 and 50 vol. % were used. Uncertainty due to manual injection onto the column is < 1 % for the standards > 200 ppmv and < 18 % for the 1.7 ppmv standard.

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Oxygen depth-profiles were measured with a Fibox 3-trace v3 planar trace oxygen minisensor (Presens, Regensburg, Germany).

2.5 Isotope ratio mass spectrometry

Samples were analyzed in duplicate by gas chromatography isotope ratio mass spectroscopy (GC-IRMS, Delta Plus, ThermoScientific, Dreieich, Germany) with a 25 m capillary column (Poraplot, 0.32 mm ID). Analytical replicate precision generally was < 0.2‰. For samples with near-atmospheric CH₄ concentrations a pre-concentration system (PreCon, ThermoScientific, Dreieich, Germany) was used (Brand, 1995) with standard error of replicate measurements generally less than 0.5‰. Injected sample volumes varied with sample concentrations (0.01–6 mL). Values are expressed relative to VPDB (Vienna Pee Dee Belemnite Standard) using the reference standard NGS3 8561 ($\delta^{13}\text{C} = -73.27\text{‰ VPDB}$; NIST, Gaithersburg, USA).

2.6 Potential CH₄ oxidation rates

Potential CH₄ oxidation rates were determined for horizons of one saturated polygon center (saturated polygon center A) and the polygonal pond in batch cultures. Homogenized soil material (cut to < 2 mm, 4 g) with in situ water content was distributed in a thin layer over the side wall in flat-walled culture bottles (50 mL) to prevent substrate limitation effects. The flasks were closed with gas-tight butyl rubber stoppers and CH₄ was added giving an initial concentration of $1.5 \pm 0.3\%$. Three flasks per sample were incubated horizontally in the dark at 4 °C for a few hours up to several weeks, depending on the oxidation rate.

CH₄ concentration in the headspace was measured over time by gas chromatography and oxidation rates were calculated from the declining CH₄ amount in the incubation by linear regression analysis using 6–8 measurement points ($R^2 > 0.81$, $p < 0.01$).

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2.7 Analysis of soil gas diffusivity

To analyze the effective diffusion coefficient for each soil horizon, the water content in the three undisturbed soil cores collected from each horizon of the polygon centers were adjusted to field capacity on a sand bath. Afterwards they were installed on top of cylindrical metal chambers of approximately 3 L volume, (Rolston 1986). The chambers were filled with an initial CH₄ concentration of 3.5 ± 0.3 mmol L⁻¹ (experiment runs > 10 h) and 6.7 ± 0.3 mmol L⁻¹ (experiment runs > 10 h) which could only escape via diffusion through the soil core cylinders. Methanotrophic activity was blocked by addition of 0.8 mmol L⁻¹ acetylene. Diffusion was monitored by gas chromatography and rates were calculated via the decrease of CH₄ concentration inside the chamber over time. The inhibition of CH₄ oxidation by acetylene was verified by placing a soil core into a jar with an atmosphere of 3.5 mmol L⁻¹ CH₄ and 0.8 mmol L⁻¹ acetylene. No CH₄ concentration change was detected over a period of three days.

To study the effect of pore-size distribution on diffusivity, samples of the unsaturated polygon center were consecutively drained in a pressure-plate apparatus (Richards and Fireman, 1943) using pressure heads of 6 kPa (drainage of macropores; > 50 μm ø), 30 and 100 kPa (drainage of mesopores; 50–10 μm and ≤ 10 μm ø) rerunning the experiment at each water content. Fick's first law was transformed to calculate the effective diffusion coefficient D_{eff} [m² s⁻¹] as follows:

$$D_{\text{eff}} = -J_{\text{CH}_4} \left(\frac{dc}{dx} \right)^{-1} \quad (4)$$

where J_{CH_4} is the diffusive CH₄ flux (mol m⁻² s⁻¹); dx is the distance of occurring diffusion, i.e. height of cylinder (m); dc is the concentration difference between chamber and atmosphere (mol m⁻³).

The final value of D_{eff} for each soil core was calculated as an average of six individual measurements. Experiments were either carried out at room temperature or, when run for more than one day, in an incubator at 20 °C and 98–100 % relative humidity to

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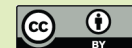
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minimize evaporation effects. Molar concentrations were corrected for temperature and pressure according to the ideal gas law. Soil cores were weighed at each dewatering stage to determine the water content and air-filled porosity. The chambers were tested for leaks with the first experimental set-up using a resin casted core.

5 The experimental set-up was modified with a second chamber to determine the diffusion through water-saturated soils. First of all, the diffusion chamber was filled with distilled water adjusted to pH2 with phosphoric acid and initially containing 1.44 mmol L⁻¹ CH₄ and 0.8 mmol L⁻¹ acetylene. Three water saturated soil cores of the uppermost horizon of the saturated polygon center B were installed on top of the chamber. Then, a second chamber (1.4 L volume) was installed on top of the cylinder containing the undisturbed soil sample. The top chamber was subsequently filled with distilled water at pH2. With this experimental setup, CH₄ diffusion from the bottom chamber through the water-saturated soil sample into the top chamber could be measured. Within 4 h the solution of the bottom chamber was sampled 3–4 times by collecting 3 mL water with a syringe and a hypodermic needle through a rubber stopper at one side of the chamber and simultaneously injecting 3 mL of the initial solution at the other side of the chamber. Samples were conserved in vials flushed with nitrogen and containing sodium chloride. Experiments were run consecutively and at 20 °C to minimize expansion effects of the solutions. Gas diffusivity was calculated from the decreasing gas concentration in the bottom chamber.

To prevent CH₄ production in the water-saturated soil samples during diffusion measurements, the undisturbed soil samples were set into a solution of 10 mmol L⁻¹ 2-bromoethane-sulfonate (an inhibitor of methanogenesis) dissolved in distilled water for more than five days prior to the experiment. The inhibition of methanogenesis and CH₄ oxidation in the second experimental set-up was verified by placing the treated soil cores into jars with distilled water adjusted to pH2 and 1.44 mmol L⁻¹ CH₄ and 0.8 mmol L⁻¹ acetylene. Neither a decrease nor increase of CH₄ was detected.

After the diffusion experiments, core samples were dried to a constant weight at 105 °C and the total porosity was determined by helium pycnometry (AccuPyc II 1340,

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Micromeritics, Norcross, USA). The volumetric water content was subtracted from the total porosity to obtain the air-filled porosity.

2.8 Determination of α_{ox} and α_{diff}

To determine the fractionation factors for oxidation and diffusion, gas samples from the batch oxidation experiment measurements and gas or water samples from the diffusion chambers were analyzed for $\delta^{13}\text{CH}_4$ composition.

Both experimental set-ups are closed systems where a limited supply of reactant, CH_4 , undergoes an irreversible conversion to a product, CO_2 , which is either constantly removed (in the diffusion experiment) or remains in the system (in the batch experiment) without further reacting with the reactant. In this respect closed system kinetic fractionation behaves like open system fractionation, where CH_4 is constantly removed. Assuming Rayleigh (1896) open system fractionation, the isotopic fractionation factor was calculated based on the approach described in Coleman et al. (1981)

$$\delta^{13}\text{C}_t \cong \left(\left(\frac{1}{\alpha} \right) - 1 \right) \times \ln \left(M_t / M_0 \right) + \delta^{13}\text{C}_0 \quad (5)$$

where M_0 is the concentration of CH_4 at time 0; M_t is the concentration of CH_4 at time t , $\delta^{13}\text{C}_0$ is the $\delta^{13}\text{C}$ value of CH_4 at time 0; $\delta^{13}\text{C}_t$ is the $\delta^{13}\text{C}$ value of CH_4 at time t . From the slope (m) of the linear regression between the differences in CH_4 isotope values ($\delta^{13}\text{C}_t - \delta^{13}\text{C}_0$) and the fraction of the remaining CH_4 concentration ($\ln(M_t/M_0)$) the isotopic fractionation factor can be derived as

$$\alpha = \frac{1}{(m+1)}. \quad (6)$$

Fractionation factors were determined for three replicates each with at least five gas samples. The fractionation factor for diffusion at water saturation was determined for three replicates with 3–4 water samples.

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2.9 CH₄ oxidation efficiency calculations

The determined isotopic fractionation factors α_{ox} and α_{diff} were used to calculate the CH₄ oxidation efficiency from the $\delta^{13}\text{C}_{\text{CH}_4}$ isotopic signatures at different soil depths of the saturated polygon center A sampled on 19 July 2009. Calculations were made for horizons where both a decrease in concentration and an enrichment of ¹³C in CH₄ were observed and diffusion was assumed to be the main occurring transport mechanism ($\alpha_{\text{trans}} = \alpha_{\text{diff}}$) using Eq. (3). Furthermore, to determine the CH₄ oxidation efficiency assuming that no fractionation by transport occurs, calculations were repeated with $\alpha_{\text{trans}} = 1$. In addition, calculations were conducted with the mean, minimum and maximum of both fractionation factors of all investigated sites to estimate their impact on f_{ox} .

To account for a potential impact of temperature on the isotopic fractionation during CH₄ oxidation, calculations were additionally conducted with a temperature-dependent correction for α_{ox} , decreasing with rising temperature by $3.9 \times 10^{-4} \text{ } ^\circ\text{C}^{-1}$ (Chanton et al., 2008b).

2.10 Statistical analyses

Statistical analyses were performed using OriginPro 8G (OriginLab Corporation, USA). The relationship between air-filled porosity and soil gas diffusion was curve-fitted and examined with a one-way ANOVA. Correlations between oxidation rate and α_{ox} and between diffusion coefficients and α_{diff} were tested with Pearson's correlation analysis. Isotopic fractionation factors of different sites were compared with one-way ANOVAs and post-hoc Tukey's Honestly Significant Differences test (Tukey's).

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3 Results

3.1 Soil and vegetation characteristics

In the depressed polygon centers drainage was impeded by the underlying permafrost. Thus, the soils of the polygon centers were mostly water-saturated with a varying water level close to the surface. All polygon centers were characterized by reducing conditions facilitating anaerobic degradation of organic matter. The two saturated polygon centers and the unsaturated polygon center showed a very high organic carbon content in the upper horizons ($> 12\%$ wt % C_{org} , designated as O_i according to US Soil Taxonomy (2010), Table 1). Subjacent horizons (A, O_i) showed an accumulation of humified organic matter mixed with fine sand bands and hydromorphical features (Bg). According to the US Soil Taxonomy the soils of these three polygon centers were classified as *Typic Historthels*. (USDA, 2010) and as *Histic Crysols* according to the WRB (WRB, 2006).

The saturated polygonal pond was characterized by a more uniform accumulation of organic carbon across the profile (ranging around 6% wt % C_{org}), containing fine sand and showing features of gleying. The soil of this polygon center was classified as *Typic Aquorthel*. (USDA, 2010) and *Typic Cryosol* (WRB, 2006).

The total porosities of the upper horizons ranged around 90% , decreasing within the profile to 50% . Accordingly, air-filled porosities at field capacity were high in the top horizons ($> 18\%$) and bulk densities low ($< 0.3\text{ g cm}^{-3}$) in comparison to the more mineral horizons above the frozen ground.

The vegetation of all polygon centers was dominated by the hydrophilic sedge *Carex aquatilis* covering $25 \pm 3\%$ of the basal area of the saturated polygon center A, $17 \pm 4\%$ of the saturated polygon center B, $27 \pm 10\%$ of the unsaturated polygon center and $6 \pm 1\%$ of the polygonal pond. The unsaturated polygon center was further covered by the mosses *Limprichtia revolvens* and *Meesia longiseta* and the polygonal pond by the submerged brown moss *Scorpidium scorpioides*.

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3.2 Potential CH₄ oxidation rates and isotopic fractionation associated with oxidation

High potential CH₄ oxidation rates occurred in samples from the two organic rich top horizons of the saturated polygon center (Oi: $31.7 \pm 2.3 \text{ nmol h}^{-1} \text{ g dw}^{-1}$; AOi: $18.8 \pm 8.4 \text{ nmol h}^{-1} \text{ g dw}^{-1}$, Table 2). In comparison, the upper horizons of the polygonal pond featured lower potential CH₄ oxidation rates (A: $4.4 \pm 0.3 \text{ nmol h}^{-1} \text{ g dw}^{-1}$; Ag1: $6.1 \pm 4.4 \text{ nmol h}^{-1} \text{ g dw}^{-1}$). The lowest horizon of the polygonal pond showed a high potential CH₄ oxidation rate of $49.2 \pm 7.7 \text{ nmol h}^{-1} \text{ g dw}^{-1}$.

The isotopic fractionation during oxidation was highest in the top horizons of the saturated polygon center (Oi: $\alpha_{\text{ox}} = 1.031 \pm 0.002$, Table 2). Low isotopic fractionation was detected for the two upper soil horizons of the polygonal pond (A: $\alpha_{\text{ox}} = 1.005 \pm 0.001$; Ag1: $\alpha_{\text{ox}} = 1.009 \pm 0.007$) corresponding with low oxidation activities found in this experimental set-up. In comparison, the calculated isotopic fractionation factors of the lower horizons were higher (Ag2: $\alpha_{\text{ox}} = 1.017 \pm 0.001$; ABg: $\alpha_{\text{ox}} = 1.020 \pm 0.002$). There was a significant positive correlation between oxidation rates and α_{ox} (Pearson's correlation coefficient $r = 0.6$; $p < 0.01$, $n = 18$). Furthermore, isotopic fractionation factors associated with oxidation differed significantly between sites (ANOVA, Tukey's, $p < 0.01$, $n = 18$).

3.3 Analysis of soil gas diffusivity and isotopic fractionation associated with diffusion

Under water-saturated conditions the potential diffusion rates were low with $0.5 \pm 0.3 \text{ nmol h}^{-1} \text{ m}^{-2}$ ($n = 3$; Table 2). At field capacity, the polygonal pond was characterized by higher potential diffusion rates than the saturated polygon center with the highest rate found in the upper horizon ($39.6 \pm 26.9 \text{ nmol h}^{-1} \text{ m}^{-2}$). Potential diffusion rates decreased from upper to lower horizons with an exception in the lowest horizon of the polygonal pond.

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Diffusion tests under different water contents showed that diffusion predominantly took place through macropores in each horizon of both sites (see Table 3). Once the macropores were drained (6 kPa), the diffusion was faster than at field capacity, but did not change strongly during further drainage. The lowest horizon was characterized by the lowest diffusion coefficient in each case. Diffusivity measurements at different water contents showed that the diffusion coefficient exponentially increased with an increasing volume of air-filled pore space ($R^2 = 0.9$, $p < 0.001$, $n = 64$, $D_{\text{eff}} = -8.27237 \times 10^{-7} + 8.89081 \times 10^{-7} \times e^{(2.71241 \times \Phi_a)}$ where Φ_a is the air-filled pore space, Fig. 1).

Isotopic fractionation by diffusion under unsaturated conditions ranged between 1.007 and 1.018 (Table 2). Values for α_{diff} did not correlate significantly with diffusion coefficients (Pearson's correlation coefficient $r = -0.1$; $p > 0.05$, $n = 18$) and did not differ significantly between sites (ANOVA, Tukey's, $p > 0.05$, $n = 18$) with a mean $\alpha_{\text{diff}} = 1.013 \pm 0.003$. Almost no isotopic fractionation by diffusion was detected under water-saturated conditions with $\alpha_{\text{diff}} = 1.001 \pm 0.000$ ($n = 3$).

3.4 CH₄ oxidation efficiency calculations

To investigate the potential impacts of the measured isotopic fractionation factors on oxidation efficiency estimates, the CH₄ oxidation efficiency was calculated based on the stable carbon isotope signatures of CH₄ sampled in the soil-water phase of the saturated polygon center A on 19 July 2009 (see Fig. 2). During sampling, the *Typical Historthel* featured a water-level 9 cm above soil surface and a thawing depth of 30 cm.

The highest CH₄ concentration was found close to the frozen ground and decreased from there to 9 cm by 88%. A CH₄ concentration peak at 6.5 cm was followed by a further decrease to near atmospheric concentrations towards the water surface (4 cm above soil surface in the pond water). Concurrently, the $\delta^{13}\text{C}$ values of CH₄ fluctuated between the frozen ground and 9 cm within 2.5‰ (-56.8 ± 1.1 ‰), decreasing between 6.5 and 1.5 cm from -53.8 ‰ to -58.3 ‰ and then increasing towards the water surface (-44.9 ‰). Two O₂ profile measurements during the same month (Fig. 2) showed

that O₂ was depleted (< 3 μmol L⁻¹) within the first horizon (Oi), thus the main part of oxidation presumably only occurred close to the soil surface.

Employing the isotopic fractionation factors (Oi $\alpha_{\text{ox}} = 1.031$; $\alpha_{\text{trans}} = \alpha_{\text{diff}} = 1.001$ for water-saturated conditions) a CH₄ oxidation efficiency of $f_{\text{ox}} = 45\%$ was assessed between 1.5 and -4 cm by assuming diffusion to be the sole transport mechanism (see Table 4). These values of CH₄ oxidation efficiency approximated those assuming that no CH₄ isotope fractionation associated with transport mechanisms took place $\alpha_{\text{trans}} = 1$, e.g. in case that ebullition or plant-mediated transport dominate. The fraction of oxidized CH₄ (f_{ox}) in this case would account for 43% of the produced CH₄.

Assuming unsaturated conditions with a water content at field capacity ($\alpha_{\text{trans}} = \alpha_{\text{diff}} 0.3 \text{ kPa} = 1.014$) the same $\delta^{13}\text{C}$ profile would result in a CH₄ oxidation efficiency almost twice as high as under water-saturated conditions ($f_{\text{ox}} = 79\%$ between 1.5 and -4 cm; Table 4) when using the determined α_{ox} of 1.031. Calculations with the mean, maximum and minimum of both fractionation factors of all sites measured under unsaturated conditions showed highly varying results (Table 4). Moreover, the CH₄ oxidation efficiency estimates negative values when $\alpha_{\text{diff}} > \alpha_{\text{ox}}$. However, calculations using α_{ox} of the saturated polygon center and the mean α_{diff} of all sites showed results close to the calculations using α_{diff} at field capacity of the saturated polygon center with $f_{\text{ox}} = 74\%$ (Table 4).

While α_{ox} was determined in the experiments at 4 °C, in situ temperature profiles indicated higher temperatures in the significant soil horizon on 19 July 2009 (1.5 cm: 12.4 °C). Applying the temperature-dependent correction by Chanton et al. (2008b), the corrected isotopic fractionation factor of Oi was $\alpha_{\text{ox}} = 1.028$, resulting in a CH₄ oxidation efficiency of $f_{\text{ox}} = 50\%$ in the topmost horizon (Table 4).

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4 Discussion

4.1 Potential CH₄ oxidation rates and isotopic fractionation associated with oxidation

Wetland CH₄ fluxes are mainly regulated by aerobic microbial CH₄ oxidation (Segers, 1998; Whalen, 2005). Similar to previous measurements in another polygon center in the Lena River Delta (Knoblauch et al., 2008), the surface horizon of the saturated polygon center holds a high potential methanotrophic activity with $31.7 \pm 2.3 \text{ nmol h}^{-1} \text{ g dw}^{-1}$, in spite of being prevalently water-saturated. Measurements have shown dissolved O₂ concentrations of up to 100% air saturation in this horizon even if the water table lies 10 cm above the soil surface. This might be explained by the high average density of *Carex aquatilis* of 25% at this site, a vascular plant with an internal gas-space ventilation system (aerenchyma) which is able to draw O₂ from the atmosphere to roots and rhizomes in the anoxic zone (Kutzbach et al., 2004) making it available for methane-oxidizing bacteria (Laanbroek, 2010).

In comparison, in the polygonal pond the highest potential oxidation rates are not found in the soil, but in the submerged brown moss layer ($201 \pm 41 \text{ nmol h}^{-1} \text{ g dw}^{-1}$ (Liebner et al., 2011)). In contrast to the saturated polygon center, the average density of *Carex aquatilis* is much lower (6%). Thus, the density of aerenchymatous sedges is likely to be a key factor for CH₄ oxidation rates in upper soil horizons of polygon centers under in situ conditions. The high potential methanotrophic activity of the thick, submerged brown moss layer of *Scorpidium scorpioides* in the polygonal pond has been explained with a mutualistic symbiosis of the moss with methanotrophic bacteria (Liebner et al., 2011).

With 1.031 ± 0.001 the mean value of α_{ox} of the top horizon of the polygon center A (Oi) is as high as values of landfill cover soils determined for this temperature (Chan-ton et al., 1999), where CH₄ oxidation rates are several orders higher in magnitude (Scheutz et al., 2009). While Teh et al. (2006) found α_{ox} to be inversely proportional to

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the CH₄ oxidation rate ($r^2 = 0.86$, $p < 0.001$, $n = 9$) in tropical rain forest soils with maximum oxidation rates between 8.2 and 11.3 nmol h⁻¹ g dw⁻¹, Pearson's regression analyses found a positive correlation of oxidation rates with α_{ox} ($r = 0.6$; $p < 0.01$, $n = 18$) across the two polygon centers and their horizons. Moreover, α_{ox} differed significantly between the polygon centers (mean $\alpha_{ox} = 1.017 \pm 0.009$). Thus, the different sites probably host different methanotrophic communities with different α_{ox} . Methanotrophs are generally divided into the three main groups type I, type II and type X, based on phylogeny and formaldehyde assimilation pathways, internal membrane arrangement and other biochemical characteristics (Kamal and Varma, 2008).

Moreover, isotopic fractionation associated with methanotrophic activity presumably occurs in the submerged brown moss layer of *Scorpidium scorpioides* in the polygonal pond as it has shown high oxidation rates in previous studies. Hence, not only fractionation processes in the soil have to be considered in CH₄ oxidation efficiency calculations, but also those of mosses when common.

4.2 Analysis of soil gas diffusivity and isotopic fractionation associated with diffusion

Since CH₄ diffusion alters the isotopic signature of the remaining gas phase, isotopic fractionation associated by diffusion needs to be taken into account in CH₄ efficiency calculations (Mahieu et al., 2008). Factors determining the soil gas diffusivity comprise air-filled porosity, the interconnectedness of the pore system and tortuosity. Results showed that diffusion occurred mainly through macropores. The exponential relationship between air-filled porosity and the diffusion coefficient is related to an increasing interconnectivity of pores with an increasing share of air-filled pores. Latter effect has been observed in the same magnitude for mineral soils at lower air-filled porosities (Gebert et al. (2011), $D_{eff} = 1.319 \times 10^{-7} \times e^{(\Phi^a/0.116)} - 1.477 \times 10^{-7}$), but is less pronounced at higher porosities in comparison to mineral soils where the effects of tortuosity play a larger role. At this, soils with a larger air-filled porosity warrant higher

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diffusive gas supply of both O₂ into the uppermost soil horizon and CH₄ escaping from lower horizons.

This is in line with the low diffusion coefficients of lowest horizon of the unsaturated polygon center (Table 3) which is characterized by higher bulk density and less air-filled porosity (Table 1). The higher potential diffusion rates at field capacity in the polygonal pond compared to the saturated polygon center might derive from a higher mineral content in the polygonal pond resulting in lower tortuosity.

Furthermore, the soil water content strongly controls the diffusivity through determining the pore space available for gas phase transport and thus the fractionation by diffusion. The determined diffusion coefficient for water-saturated soils ($1.03 \pm 0.25 \times 10^{-6} \text{ cm}^2 \text{ s}^{-1}$) was one order of magnitude lower than the theoretical value of CH₄ diffusion in pore water at 20 °C ($1.62 \times 10^{-5} \text{ cm}^2 \text{ s}^{-1}$) found by Hornibrook et al. (2009). The CH₄ diffusion through organic tortuous soil is thus much slower than through pure water where molecules face less resistance. Under water-saturated conditions almost no isotopic fractionation occurred ($\alpha_{\text{diff}} = 1.001 \pm 0.000$). Under unsaturated conditions the isotopic fractionation by diffusion ranged between the theoretical maximum value in air $\alpha_{\text{diff}} = 1.0195$ and the low isotopic fractionation during air-water gas transfer of $\alpha_{\text{diff}} = 1.0008$ (Knox et al., 1992). The mean $\alpha_{\text{diff}} = 1.013 \pm 0.003$ of the study sites at field capacity is lower than the maximum value of $\alpha_{\text{diff}} = 1.0178 \pm 0.001$ assumed for sandy landfill cover soils (De Visscher et al., 2004) which is slightly less than the theoretical value of $\alpha_{\text{diff}} = 1.0195$. De Visscher et al. (2004) used glass beads to determine the fractionation factor which presumably feature both a lower tortuosity and a higher pore interconnectedness than the peaty tundra soils. The higher diffusion coefficient in the porous medium ($5.54 \times 10^{-6} \text{ m}^2 \text{ s}^{-1}$) could facilitate an isotope fractionation by diffusion. Measurements of this study showed no correlation between diffusion coefficients and α_{diff} , but might show a different relationship with higher concentration gradients as found in landfill habitats where CH₄ concentrations up to 60 % are found at the bottom of cover soils (Cabral et al., 2010).

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4.3 CH₄ oxidation efficiency calculations

The observed decrease in CH₄ concentration between the lower and the upper horizons of the saturated polygon center A cannot be explained by oxidation since no oxygen was available in these depths and no change in isotopic signatures occurs between these depths. It must originate from the different transport mechanisms (diffusive, advective or ebullitive with $\alpha_{\text{trans}} = 1$). Moreover, the increase of CH₄ concentration and lower $\delta^{13}\text{CH}_4$ values between 6.5 and 1.5 cm imply further CH₄ production in a horizon where fresh organic material is available for degradation. In this horizon CH₄ production and oxidation occur in close proximity.

Calculations indicated a CH₄ oxidation efficiency of up to 50 % in the first 1.5 cm of the saturated polygon center where oxygen is present. The highest methanotrophic activity expectedly occurs at the anaerobic-aerobic interface, since the ratio of CH₄ to O₂ is optimal here (Dedysh, 2002). An oxidation efficiency of this magnitude seems reasonable and has been described before e.g. for peat cores from a fresh water marsh soils with up to 32 % under water-saturated conditions (Roslev and King, 1996). Since the potential diffusion rates are much lower than the potential oxidation rates at this site, a complete oxidation of CH₄ might be expected during this transport mechanism. The non-oxidized share of CH₄ is presumed to be transported by different mechanisms to the atmosphere (ebullition, plant-mediated transport and diffusion), but cannot be accounted for with this method.

Wetlands inhabited by vascular plants show plant-mediated CH₄ transport as the predominant transport mechanism (Van Der Nat and Middelburg, 1998) which may account for up to two-thirds of the total flux in a water-saturated polygon center of the Siberian tundra (Kutzbach et al., 2004). The site used for the current study is characterized by an average density of *Carex aquatilis* of 25 %. The downward transport of O₂ of these plants is accompanied by an upward diffusion of CH₄ from the rhizosphere along the concentration gradient (Lai, 2009). This passive transport mechanism is accompanied by isotopic fractionation resulting in the emission of lighter CH₄ (Chanton

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and Whiting, 1996; Chasar et al., 1999). However, plant-mediated transport does not affect the CH₄ efficiency calculations as the CH₄ bypasses the aerobic layer and is not available for oxidation.

Fractionation associated with diffusion can obscure calculating the balances of CH₄ production and oxidation. Since the effects of diffusion and oxidation on the isotopic signature of the remaining gas phase are of opposite direction, neglecting diffusive fractionation by setting α_{trans} to 1 causes an underestimation of CH₄ oxidation: a lighter isotopic signature is observed which could misleadingly be interpreted as less oxidation efficiency. Therefore, the isotopic fractionation factor of transport is subtracted from the fractionation of oxidation in the CH₄ efficiency calculation. As a result, the calculated efficiency increases, since the shift in $\delta^{13}\text{C}$ values is caused by a smaller difference between α_{ox} and α_{trans} . However, results show that isotope fractionation by diffusion only plays a substantial role under unsaturated conditions. Thus, under water-saturated conditions α_{trans} can be assumed to be 1. For the presented study sites of the polygonal tundra in the Siberian Lena River Delta it seems plausible to use the mean $\alpha_{\text{diff}} = 1.013$ under unsaturated conditions for CH₄ oxidation efficiency calculations when diffusion is the predominant transport mechanism, since α_{diff} did not differ significantly between sites. On the contrary, isotopic fractionation factors associated with oxidation need to be determined for the oxic horizons of the sites of interest as they differ strongly. The isotopic fractionation factors presented in the current study might be higher than under in situ conditions, since they were determined in laboratory experiments applying high initial CH₄ concentrations (Templeton et al., 2006).

As previously mentioned by Nihous (2010), the calculated oxidation efficiency is only as reliable as the knowledge of the isotopic fractionation factors since slight variations in the adopted α_{ox} and α_{trans} change the outcome strongly. Especially under unsaturated conditions, the predominant transport mechanism has to be considered to obtain trustworthy oxidation efficiency values. Here, CH₄ emission measurements might help.

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Results showed that the applied method meets its limitations when $\alpha_{\text{diff}} > \alpha_{\text{ox}}$. In this case fractionation processes are predominantly attributed to diffusion not to oxidation and Eq. (3) cannot be used for CH₄ oxidation efficiency calculations.

Including temperature-dependent corrections for the isotopic fractionation factors into the oxidation efficiency calculations, results in higher oxidation efficiencies. Tyler et al. (1994) showed that the correlation between temperature and isotopic fractionation factor decreased with soil depth ranging between $4.3\text{--}5.0 \times 10^{-4} \text{ } ^\circ\text{C}^{-1}$. Further, Knoblauch et al. (2008) found with SI probing of microbial PLFAs that the community active in situ is dominated by type I methanotrophs and that rising temperatures increase the importance of type II in soils of the same area. Type II bacteria show a lower CH₄ oxidation activity and a lower α_{ox} than type I (Zyakun and Zakharchenko, 1998). Differences in the carbon isotopic fractionation are due to the type of methane monooxygenase (MMO) expressed by the cells, the mechanism for assimilation of cell-carbon and type of cellular physiology (Jahnke et al., 1999). Each process of the first CH₄ oxidation step (adsorption and desorption from the cell wall and conversion to methanol) may precede at a specific rate with a specific isotopic fractionation (Nihous, 2010). Thus, it is assumed that microbial communities of different ecosystems react unequally to temperature and universal applications of correction factors seem problematic. Nonetheless it is likely that α_{ox} is directly influenced by soil temperature and neglecting might underestimate the CH₄ oxidation efficiency.

The presented study shows that isotope fractionation by diffusion could play an important role in wetland soils under unsaturated conditions resulting in higher calculated CH₄ oxidation efficiencies. However, these calculations have been conducted with $\delta^{13}\text{C}$ profiles of water-saturated conditions. Thus, future measurements and CH₄ oxidation efficiency calculations need to be conducted for different water level conditions of the studied sites as they drive the strength of oxidation processes.

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The isotopic fractionation factors presented here enable calculations to estimate the potential CH₄ oxidation efficiency in arctic wetland soils. To perform CH₄ oxidation efficiency calculations, the isotopic fractionation factors associated with oxidation need to be determined for the oxic horizons of sites of interest, since it strongly differs from site to site and horizon to horizon. Furthermore, the contribution of diffusion to other simultaneously occurring transport mechanisms has to be estimated by means of the interpretation of unsaturated/water-saturated conditions and both the CH₄ concentration and SI soil profiles. A mean value of $\alpha_{\text{diff}} = 1.013$ may be applied for the investigated polygonal tundra sites under unsaturated conditions, while under water saturation $\alpha_{\text{diff}} = 1.001$ can be used. The experimental set-up to determine the potential CH₄ oxidation efficiency at 4 °C gives conservative estimates. If feasible, isotopic fractionation factors should be determined at in situ temperatures. To deepen the understanding of CH₄ oxidation efficiencies of northern wetland soils, it is essential to study soils of different hydrological regimes. These calculations could then provide the basis for an improved estimation of current CH₄ sources and sinks and their potential strength in response to environmental change and global warming, especially in permafrost-affected soils which bear the potential to cause a positive feedback to climate change.

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Table 1. Selected physical soil properties. Horizon designation according to US Soil Taxonomy (USDA, 2010).

Site	Horizon	Mean depth below soil surface (cm)	pH	Electrical conductivity (μS)	Total porosity (%)	Air-filled porosity at field capacity (%)	Water content at field capacity (vol %)	Bulk density (g cm^{-3})	C_{org} content (wt%)	Soil texture & further characteristics
Saturated polygon center A	Oi	2.5	5.6	90.7	90.2 \pm 0.6	22.2 \pm 2.8	68.0 \pm 2.4	0.22 \pm 0.01	17.70	Slightly decomposed plant material, <i>Carex</i> rhizomes, no mineral material
	AOi	7.5	5.5	37.8	91.6 \pm 0.6	27.7 \pm 6.1	63.9 \pm 5.5	0.19 \pm 0.01	10.80	Slightly humified plant material, <i>Carex</i> rhizomes, pure fine sand
	A1	14.5	5.5	49.4	69.0 \pm 1.8	3.8*	65.7 \pm 0.9	0.79 \pm 0.05	3.0	Humified organic matter, slightly silty fine sand
	A2	25	5.6	48.7	80.8 \pm 1.7	1.8 \pm 1.3**	79.9 \pm 1.7	0.46 \pm 0.04	4.2	Humified organic matter, fine sand bands, frozen ground
Saturated polygon center B	Oi	2.5	6.0	83.8	88.24**	n.a.	n.a.	0.27**	20.17	Slightly decomposed plant material, <i>Carex</i> rhizomes, no mineral material
Unsaturated polygon center	Oi1	1.5	5.6	153.9	93.4 \pm 2.4	32.8 \pm 10.2	60.6 \pm 8.0	0.15 \pm 0.05	12.46	Slightly decomposed plant material, <i>Carex</i> rhizomes, no mineral material
	Oi2	8	5.6	73.3	94.9*	18.32*	72.0 \pm 6.8	0.10 \pm 0.00	14.98	Slightly decomposed plant material, <i>Carex</i> rhizomes, no mineral material
	A	17.5	5.6	40.8	92.3 \pm 0.4	32.0 \pm 2.6	60.3 \pm 2.4	0.16 \pm 0.01	8.74	Humified organic matter, slightly silty fine sand,
	Bg	26.5	5.9	34.8	54.4 \pm 2.9	1.7*	56.0 \pm 0.3	1.19 \pm 0.08	1.53	Medium silty fine sand, frozen ground
Polygonal pond	A	3.5	5.4	30.6	89.7 \pm 0.6	33.2 \pm 7.4	56.4 \pm 6.8	0.23 \pm 0.01	5.98	Slightly decomposed plant material, containing fine sand bands
	Ag1	12.5	5.4	28.6	85.8 \pm 1.5	22.0 \pm 3.6	63.8 \pm 2.2	0.34 \pm 0.04	6.46	Humified organic matter, fine sand bands, gleying
	Ag2	25	6.0	24.2	77.1 \pm 1.2	6.8 \pm 1.1	70.3 \pm 2.3	0.57 \pm 0.04	6.09	Humified organic matter, fine sand bands, gleying
	ABg (2010)	33	6.1	86.2	65.4 \pm 1.8	5.8 \pm 1.3	59.5 \pm 2.4	0.88 \pm 0.05	5.41	Humified organic matter, slightly silty fine sand bands, gleying, frozen ground

** One replicate only, * two replicates only, n.a. = not analyzed.

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Table 2. Potential methanotrophic activity, potential rates of CH₄ diffusion and isotopic fractionation factors for diffusion and oxidation calculated for the different horizons of the studied sites. Potential diffusion rates of the saturated polygon center A and the polygonal pond were determined at field capacity (0.3 kPa) and of the saturated polygon center B at water saturation. n.a. = not analyzed.

Water potential	Site	Horizon	Mean depth below soil surface in cm	Potential CH ₄ oxidation rate in nmol h ⁻¹ g dw ⁻¹ (mean ± SD)	α_{ox} (mean ± SD)	Potential CH ₄ diffusion rate in nmol h ⁻¹ m ⁻² (mean ± SD)	α_{diff} (mean ± SD)
Water saturated	Saturated polygon center B	Oi	2.5	n.a.	n.a.	0.5 ± 0.3	1.001 ± 0.000
0.3 kPa	Saturated polygon center A	Oi	2.5	31.7 ± 2.3	1.031 ± 0.002	9.8 ± 6.2	1.014 ± 0.003
		AOi	7.5	18.8 ± 8.4	1.023 ± 0.002	8.8 ± 4.2	1.014 ± 0.001
	Polygonal pond	A	3.5	4.4 ± 0.3	1.005 ± 0.001	39.6 ± 26.9	1.010 ± 0.003
		Ag1	12.5	6.1 ± 4.4	1.009 ± 0.007	17.0 ± 11.3	1.013 ± 0.002
		Ag2	25	7.3 ± 1.8	1.017 ± 0.001	2.9 ± 0.7	1.011 ± 0.000
ABg (2010)	33	49.2 ± 7.7	1.020 ± 0.002	9.6 ± 1.5	1.017 ± 0.001		

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Table 3. Diffusion coefficients of an unsaturated polygon center at the different dewatering levels 0.3, 6, 30 and 100 kPa at different soil depths (mean \pm std, n = 3).

Site	Horizon	Mean depth below soil surface in cm	Diffusion coefficients in $10^{-6} \times \text{m}^2 \text{s}^{-1}$ (mean \pm SD) at dewatering levels of			
			0.3 kPa	6 kPa	30 kPa	100 kPa
Unsaturated polygon cente	Oi1	1.5	0.78 \pm 0.46	5.52 \pm 2.29	6.72 \pm 2.14	7.15 \pm 2.08
	Oi2	8	0.40 \pm 0.15	5.22 \pm 1.02	7.15 \pm 1.03	7.48 \pm 1.30
	A	17.5	0.67 \pm 0.18	4.98 \pm 0.69	6.37 \pm 0.85	6.80 \pm 1.11
	Bg	26.5	0.24 \pm 0.17	0.75 \pm 0.33	2.18 \pm 0.77	3.15 \pm 0.99

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Table 4. Calculated CH₄ oxidation efficiency f_{ox} in per cent for the saturated polygon center A (“saturated pc”) between 1.5 cm below (δ_P) and 4 cm above (δ_E) soil surface according to Eq. (3) applying the fractionation factors α_{ox} and α_{diff} of the respective horizon (horizon Oi). Calculations were further conducted with the mean, minimum and maximum of both fractionation factors of all investigated sites, with α_{ox} corrected for temperature according to Chanton et al. (2008b) and with $\alpha_{trans} = 1$ (no fractionation due to transport).

Saturated polygon center A			α_{trans}						
			no fractionation	α_{diff}					
δ_E	–4 cm:	–44.9‰		saturated pc Oi			all sites		
			δ_P	1.5 cm:	–58.3‰	Water saturated	Field capacity	mean	min
						1.000	1.001	1.014	1.013
α_{ox}	all sites	mean	1.017	79	84	447	335	134	–1340
		min	1.004	335	447	–134	–149	–447	–96
		max	1.032	42	43	74	71	54	96
	saturated pc	Oi temp corr	1.031	43	45	79	74	56	103
			1.028	48	50	96	89	64	134

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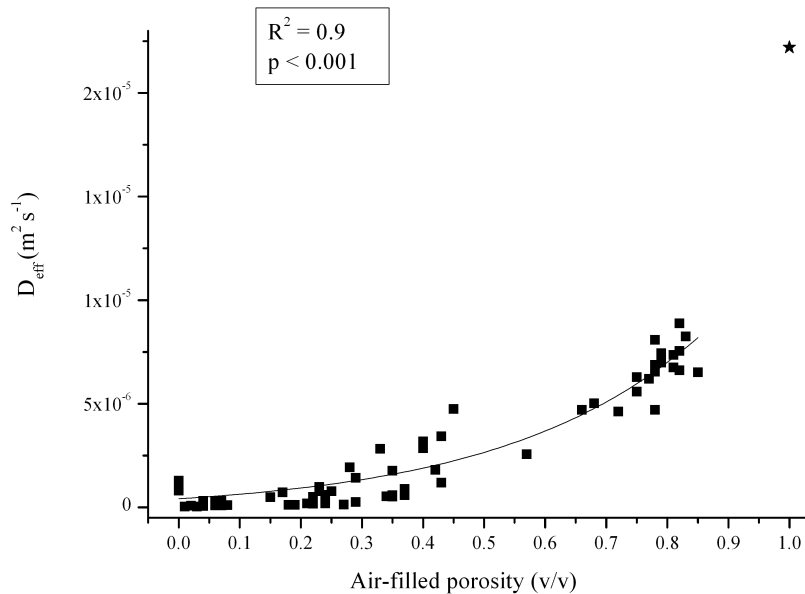


Fig. 1. Relationship between air-filled porosity and soil gas diffusion (effective diffusion coefficient) with exponential fit, $n = 64$. Star marks diffusion coefficient of CH_4 ($D = 2.2 \times 10^{-5} \text{ m}^2 \text{ s}^{-1}$) in free air at 20°C and 101.33 kPa given by Coward and Georgeson (1937).

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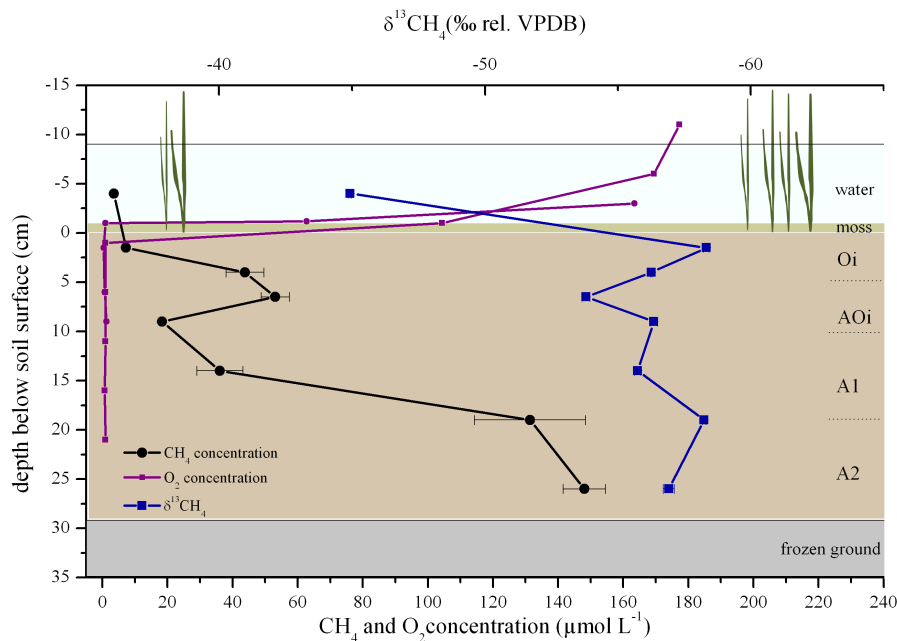


Fig. 2. Depth profiles of CH₄ concentration (black line, circles) and δ¹³C (blue line, squares) of saturated polygon center A on 19 July 2009 and depth profiles of O₂ concentration of the same polygon center on 8 July 2009 (purple line, squares) and 24 July 2009 (purple line, circles). Error bars represent the standard deviations of the means of two analytical replicates.